

Analyst

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

Qualitative SERS Analysis of G-Quadruplex DNAs Using Selective Stabilising Ligands

K. Gracie^a, V. Dhamodharan^b, P. I. Pradeepkumar^b, K. Faulds^{a*}, D. Graham^{a*}

^aCentre of Molecular Nanometrology, WestChem, Department of Pure and Applied Chemistry, University of Strathclyde, 295 Cathedral Street, Glasgow, G1 1XL, UK

^bDepartment of Chemistry, Indian Institute of Technology Bombay, Mumbai 400076, India

Abstract

Nucleic acids are of key biological importance due to their range of functions and ability to form various different structures, with an example of emerging significance being quadruplexes formed by guanine-rich sequences. These guanine rich sequences are found in different regions of the genome such as telomeres, gene promoters and introns and UTRs of mRNAs. Here a new approach has been developed that utilises surface enhanced Raman scattering (SERS) for the detection of the formation of G-quadruplexes. Three G-quadruplex stabilising ligands that each have their own unique SERS response were used in this study and their ability to act as reporters assessed. A SERS response was only obtained from the ligands in the absence of G-quadruplex formation. This resulted in an “on/off” method which was successfully used to qualitatively detect the formation of G-quadruplex using quadruplex-forming sequences such as human telomeric and *C-MYC* promoter DNAs. The unique SERS spectra of each stabilising ligand offer the potential for use of SERS to study higher order DNA structures. This work shows that the ligands used can act simultaneously as a potential therapeutic stabilising agent and a SERS reporter, therefore allowing the use of SERS as a method of analysis of the formation of G-quadruplex DNAs.

Introduction

G-quadruplexes are guanine rich sequences that are capable of forming stable structures under physiological conditions *in vivo*.^{1,2} They are composed of G-quartets that are a result of Hoogsteen hydrogen-bonding between guanines, which stack on top of each other producing four-stranded helical structures held together by π - π interactions.^{3,4} The structure and stability of G-quadruplexes have been investigated using methods such as gel electrophoresis, circular dichroism and UV melting analysis.⁵ The structure and stability of G-quadruplexes are known to be dependent on various factors including the nature of the sequence, flanking nucleotides, loop length and concentration as well as the presence of monovalent cations such as K^+ and Na^+ .^{6,7,8} The specific cation can have an

1
2
3 overall effect on the topology of the G-quadruplex. For example, in the presence of Na^+ , human
4 telomeric DNA sequence, $\text{AG}_3(\text{T}_2\text{AG}_3)_3$, folds into anti-parallel structures, whereas in the presence of
5 K^+ , TTAGGG repeats can fold into at least five distinct quadruplex structures.^{9,10}
6
7

8
9 Numerous biochemical and structural studies have shown that guanine rich DNA sequences can
10 spontaneously fold into G-quadruplex DNA structures *in vitro* and more recent studies have shown
11 that quadruplexes have certain functions *in vivo*.² G-quadruplexes are involved in regulating
12 telomere maintenance, as well as transcription, replication and translation.³ Telomeres are
13 nucleoprotein complexes and their main function is to protect the end of eukaryotic chromosomes
14 containing repetitive guanine-rich DNA.¹ Telomeres can be structurally complex, in somatic cells
15 telomeres are shortened by 50-200 bases after each successive cycle of cell division. This is due to
16 the inability of DNA polymerase to replicate the telomeric DNA strand due to the unavailability of
17 3'OH group at the extreme end of the chromosome (end replication problem). When telomeres are
18 shortened critically, they reach the hayflick limit and cells become senescent and induce apoptotic
19 signals. Nevertheless, telomere length is maintained in ~85% cancers, by the enzyme, telomerase,
20 which is notably absent in somatic cells. The formation of a G-quadruplex structure is an ideal
21 approach to the inhibition of telomerase activity because it is capable of isolating the substrate of
22 telomerase, a single stranded telomeric DNA sequence.¹¹
23
24

25
26 Recently, there has been extensive research on the development of G-quadruplex ligands,
27 particularly ones that target the human telomeric repeat $[\text{d}(\text{T}_2\text{AG}_3)_4]$ and would consequently block
28 the action of telomerase.¹²⁻¹⁴ G-quadruplexes have a larger π -surface area compared to DNA as a
29 result of the four coplanar bases compared to two in duplexes. The majority of small molecules that
30 bind to these structures also have large π -surfaces in order to maximise the π - π interactions that will
31 form.¹⁵ Small organic molecules can lead to cell death by displacing the protective telomeric
32 components or sequestering the substrate needed for telomeric activity.^{16, 17} Telomestatin is a
33 natural product that was isolated from the bacterium *Streptomyces anulatus* and has been shown to
34 be a very potent telomerase inhibitor.¹⁸
35
36

37
38 There are several spectroscopic techniques that can be used to investigate the structure and stability
39 of G-quadruplexes, such as UV molecular absorption¹⁹, circular dichroism (CD)²⁰, fluorescence
40 (FRET)²¹ and NMR²². UV/CD techniques have the most sensitive instrumental response to the
41 interaction between the G-quadruplex strands, additionally CD can be used to determine if the
42 quadruplex has parallel or anti-parallel configuration.²³ Raman spectroscopy is an alternative
43 technique which can be used to analyse the structure of macromolecules such as DNA. It was
44 successfully applied to the investigation of the phosphodiester backbone conformation and the
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3 hydrogen-bonding interactions of quadruplexes.²⁴ Miura *et al* identified specific Raman markers as
4 indicators of the presence of parallel or anti-parallel G-quadruplexes.²⁵ Raman spectroscopy is
5 limited by the relatively low cross section of the Raman scattering process compared to other
6 techniques such as fluorescence and absorption spectroscopy. Surface enhanced Raman scattering
7 (SERS) has now become an appealing alternative technique as it overcomes the limitations
8 associated with Raman spectroscopy.²⁶ The high sensitivity achieved when using SERS is due to the
9 enhancement of the Raman cross section when the analyte is in close proximity to a metal surface,
10 which is usually in the form of a thin metallic film or a colloidal suspension. There have been several
11 studies involving the direct detection of DNA bases or short DNA sequences utilizing SERS. Kneipp *et al*
12 *et al* applied SERS to the detection of adenine bases, and emphasising the capability of SERS to be used
13 as a method for single molecule detection.²⁷ Bell *et al* successfully used SERS for the label free
14 detection of DNA bases further emphasising the capability of SERS to be used for DNA detection.²⁸
15 The detection of DNA structures has proved to be more difficult due to the enhancement of Raman
16 scattering having a high dependence on the orientation of the DNA with respect to the metallic
17 surface. However, Breuzard *et al* demonstrated that it is possible to use SERS to analyse DNA
18 structures by using the technique to investigate the stability of G-quadruplexes.²⁹ Results revealed
19 that there was a decrease in ethidium bromide SERS signal when it was bound to a quadruplex
20 structure. Rusciano *et al* then further developed the application of SERS as a method to determine
21 the stability of G-quadruplexes by using direct SERS analysis to gain information on the structure of
22 guanine-rich DNA sequences.³⁰ Different G-quadruplex structures were analysed and their relative
23 stabilities were compared. Specific Raman features of G-quadruplex DNAs were identified in the
24 SERS spectra and subsequent analysis on the phosphate backbone conformation and the strength of
25 the Hoogsteen-like hydrogen bonding was performed.³⁰ This demonstrated for the first time that
26 SERS can be an extremely useful technique for the analysis of the stabilities of different G-
27 quadruplex structures.

28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48 G-quadruplexes can be used as potential therapeutic targets for anti-cancer therapy.³¹ The ligand
49 2,6-diamidoanthraquinone was shown to stabilise a quadruplex structure and inhibit telomerase
50 activity.³² Since then, many classes of ligands have been reported as G-quadruplex stabilising agents,
51 however very few of these have shown specificity towards the targeted quadruplex structure over
52 other DNA topologies. Some ligands have also been shown to be indirect inhibitors of telomerase by
53 displacing the telomere binding proteins that then induce a DNA damage response.^{33,34} Therefore,
54 there has been great interest in developing strong and selective quadruplex binding agents that can
55 detect the presence of quadruplex structures *in vivo*.¹⁷
56
57
58
59
60

1
2
3
4
5 Here, we report the use of SERS for the qualitative detection of the formation of G-quadruplex
6 structures. SERS is used as a method to monitor the formation of G-quadruplexes that is based on
7 the unique SERS response given by three G-quadruplex stabilising ligands. When the ligand is added
8 to a solution containing duplex DNA, it remains unbound and free, upon addition of silver
9 nanoparticles aggregation occurs resulting in a strong SERS response. However, when the ligand is
10 added to a solution of G-quadruplex DNAs, the ligand binds strongly to this DNA structure therefore
11 when silver nanoparticles are added; the ligand is no longer free to induce aggregation or a SERS
12 response (Figure 1). The ligands used here are 360A previously reported by Granotier *et al*,³⁵ and the
13 other two ligands, 3AQN and 6AQN, are reported by us and were shown previously to stabilise
14 various quadruplexes (telomeric, *C-KIT1*, *C-KIT2* and *C-MYC*) selectively over duplex DNA (Figure 2).³⁶
15 For the first time we report the use of SERS as the analysis method due to the ability of the ligands
16 to act as Raman reporters. Various guanine-rich DNA sequences were used in this study, including
17 the human telomeric sequence and the sequence from promoter of *C-MYC* gene. We have
18 demonstrated the first report of these stabilising ligands being used to analyse the formation of G-
19 quadruplex DNAs over duplex DNA by SERS, which has previously been done using fluorescence
20 based methods.
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

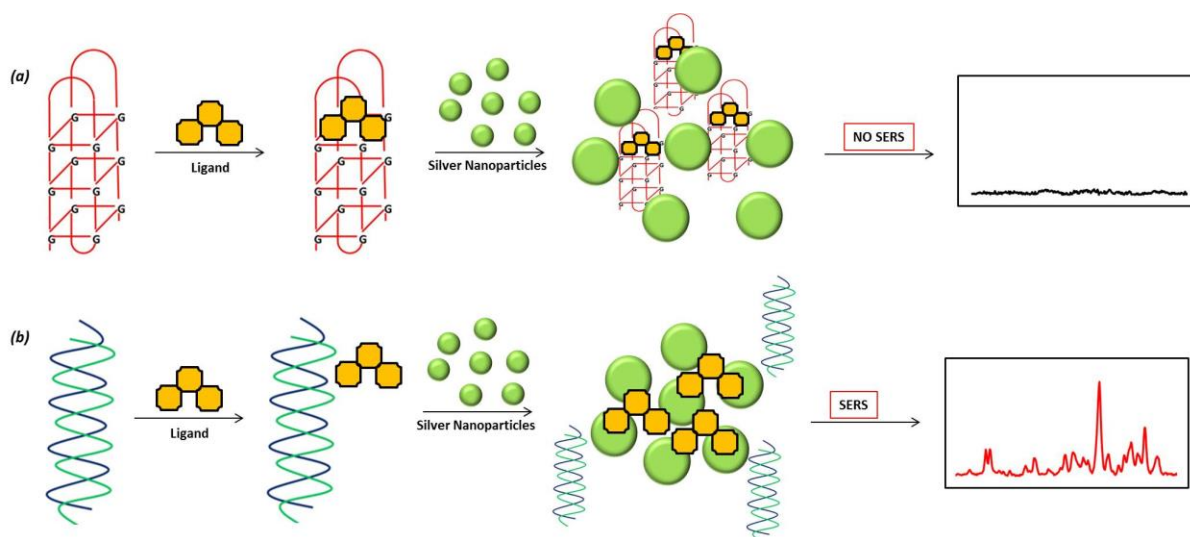


Figure 1 Schematic overview of the qualitative detection of G-quadruplex DNA. (a) When the stabilising ligand is added to a solution containing G-quadruplex DNAs, the ligand strongly binds to the G-quadruplex and when silver nanoparticles are added, the ligand is not free to induce aggregation and a unique SERS response. (b) When the stabilising ligand is added to a solution containing duplex DNAs, the ligand does not bind to this DNA structure, therefore in the presence of silver nanoparticles, aggregation occurs resulting in a SERS response from the ligand.

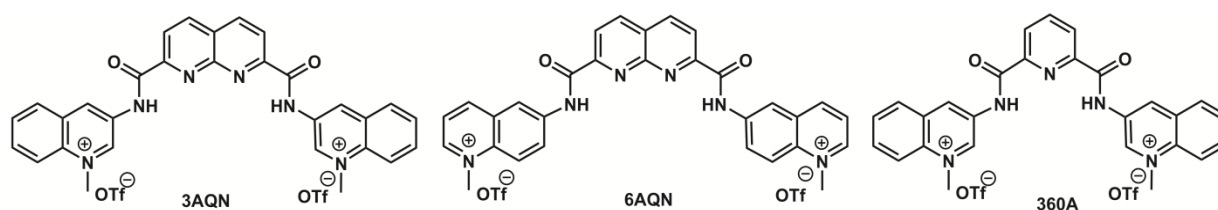


Figure 2 Structures of the three selective G-quadruplex binding ligands. From left: 3AQN (3-amino-quinolinium naphthridine); 6AQN (6-amino-quinolinium naphthridine) and 360A (3-amino-quinolinium pyridine).

Materials and Methods

Ligand Synthesis

The three ligands (3AQN, 6AQN and 360A) used in this study were synthesized as reported previously.³⁶

Colloid Synthesis

Silver nanoparticles were synthesised using a modified Lee and Meisel method.³⁷ Silver nitrate (90 mg) was dissolved in distilled water (500 mL). The solution was heated rapidly to boiling with continuous stirring. Once boiling, an aqueous solution of sodium citrate (1 %, 10 mL) was added quickly. The heat was reduced and the solution was left to boil gently for 90 min with stirring. The colloid was then analysed by UV-vis spectroscopy and the λ_{\max} was 403 nm with the full width half-height (FWHH) measured to be 124 nm. The concentration of the colloid was calculated to be 0.2 nM.

Oligonucleotides

All DNAs were purchased on a 0.2 μ mole scale with HPLC purification from Eurofins MWG (Ebersberg, Germany).

Name	Description	Sequence (5' – 3')
Peroxidase Deoxyribozyme	Quadruplex	GTGGGTAGGGCGGGTTGG
Human Telomeric DNA	Quadruplex	AGGGTTAGGGTTAGGGTTAGGG
C-MYC Promoter	Quadruplex	TGAGGGTGGGTAGGGTGGGTAA
Control	Duplex (Self complementary)	CAATCGGATCGAATTCGATCCGATTG

Table 1 Oligonucleotides used in this study.

SERS Analysis

SERS analysis was performed using an Avalon Plate Reader (Belfast, UK), with an excitation wavelength of 532 nm. A 96 well plate was placed onto a stage and the instrument's software was used to automatically move the stage so that spectra could be recorded from each well. The accumulation time was 10 s. A single well can hold 300 μL . Prior to SERS analysis, the ligand (5×10^{-5} M, 20 μL) and the specific DNA (5×10^{-5} M, 20 μL) were premixed at room temperature for 30 minutes. This mixture was then added to the well that contained citrate reduced silver nanoparticles (0.2 nM, 260 μL). In the studies involving the presence or absence of monovalent cations, either NaCl (100 mM, 20 μL) or KCl (100 mM, 20 μL) was used. The samples were then analysed immediately. For peak intensity studies, three replicate samples were scanned 5 times each, with the peak intensities then being averaged to calculate the differences. Spectra were baseline corrected using a multipoint polynomial fit and level and zero mode using Grams software (AI 7.00).

Results and Discussion

Bisquinolinium ligands such as 360A are known to selectively bind and stabilise quadruplex nucleic acids over duplex DNA.³⁸⁻⁴⁰ Recently we reported two new derivatives of bisquinolinium carboxamide having 1,8-naphthyridine as the central core³⁶ (Figure 2). Various biophysical and biochemical studies were used to investigate the interaction between the ligands and different nucleic acids. Circular dichroism (CD) melting assays indicated that ligands can impart higher stabilisation to quadruplex DNA over duplex DNA. Furthermore, CD studies were used to investigate the potential of these ligands to induce quadruplex formation in different DNA sequences. As observed with telomestatin, in the absence of monovalent cations, the ligands induced the formation of a quadruplex structure. A Fluorescent Intercalator Displacement (FID) assay demonstrated the ligands (3AQN, 6AQN and 360A) have high binding affinity and selectivity for quadruplex DNA over duplex DNA. Here, we report the use of surface enhanced Raman scattering (SERS) for the direct detection of the interaction of these ligands with G-quadruplex DNA that does not require the additional use of any other fluorescent agent. This is possible as the ligands themselves give a unique SERS response therefore the interaction between the ligand and DNA can be directly measured. This has potential future implications for the detection of the formation of G-quadruplexes *in vivo* for use in diagnostic applications.

Initial experiments were performed to determine if the three ligands gave a SERS response by addition of the ligands directly to a suspension of silver nanoparticles. As can be seen in Figure 3, each ligand had a unique SERS response due to their structural variations. A SERS response was obtained due to the ability of the ligands at the concentration used (50 μM) to aggregate the nanoparticles causing the SERS enhancement.

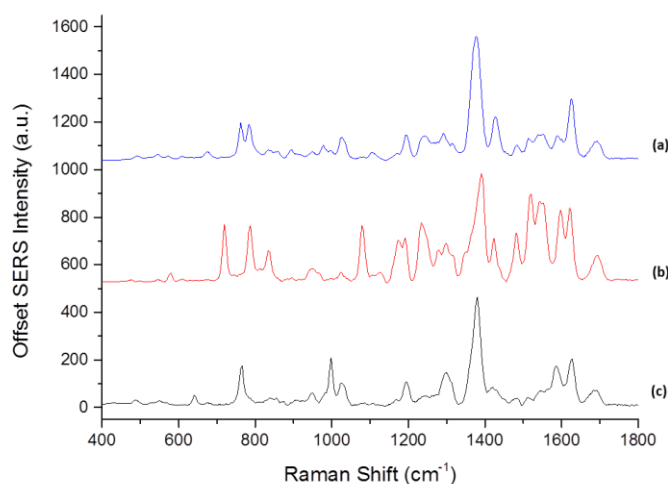


Figure 3 SERS spectra obtained for each stabilising ligand using citrate reduced silver nanoparticles. (a) SERS spectrum of 3AQN, (b) SERS spectrum of 6AQN and (c) SERS spectrum of 360A. Spectra were obtained using 532 nm laser excitation and an accumulation time of 1 second.

With the knowledge that each of the ligands gave a unique SERS response, they were then incubated for 30 minutes at room temperature with the peroxidase deoxyribozyme quadruplex sequence (Table 1), to study the SERS signal when the ligand was bound to G-quadruplex DNAs. As can be seen in Figure 4, there is a significant reduction in SERS signal when each of the three ligands is bound to the G-quadruplex sequence. This indicates that the ligand was now complexed into the G-quadruplex DNAs, which prevented the ligands from interacting with the surface of the silver nanoparticles, resulting in the reduced SERS intensity observed.

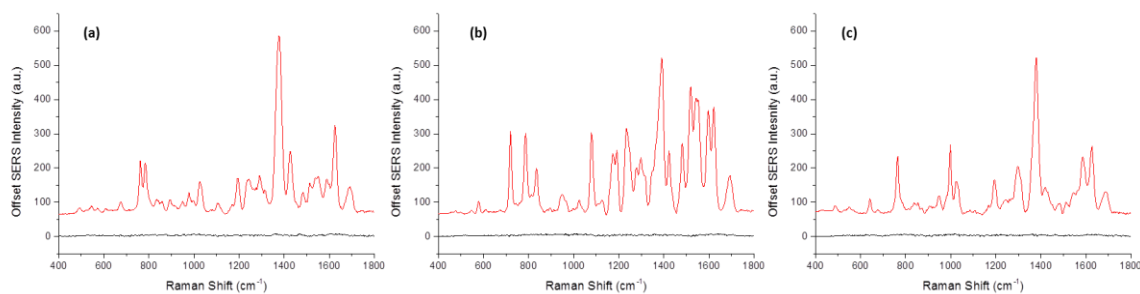


Figure 4 SERS spectra of free ligand (red) and peroxidase deoxyribozyme complex (black): (a) spectra of 3AQN free and complexed to G-quadruplex DNA, (b) spectra of 6AQN free and complexed to G-quadruplex sequence and (c) spectra of 360A free and complexed to G-quadruplex sequence. Spectra were obtained using 532 nm laser excitation and an accumulation time of 1 second.

To demonstrate the specificity of the binding ligands towards G-quadruplexes, the SERS signal was obtained from the interaction of the ligands with a duplex DNA compared to a G-quadruplex structure (Table 1), the results are shown in Figure 5. For each ligand, SERS signals were obtained when the duplex control sequence was used in place of the G-quadruplex sequence. This suggests that the ligands do not bind strongly to the duplex allowing them to remain free in solution and to come into close proximity to the surface of the silver nanoparticles resulting in a SERS signal. However, when a G-quadruplex is present a much reduced SERS signal is observed (Figure 1). This confirms the specificity that each of the ligands has towards G-quadruplex DNAs compared to other DNA structures and the strong binding affinity that the ligands have for a G-quadruplex structure. As mentioned previously, the ligands are able to cause nanoparticle aggregation, which means that no aggregating agent was used.

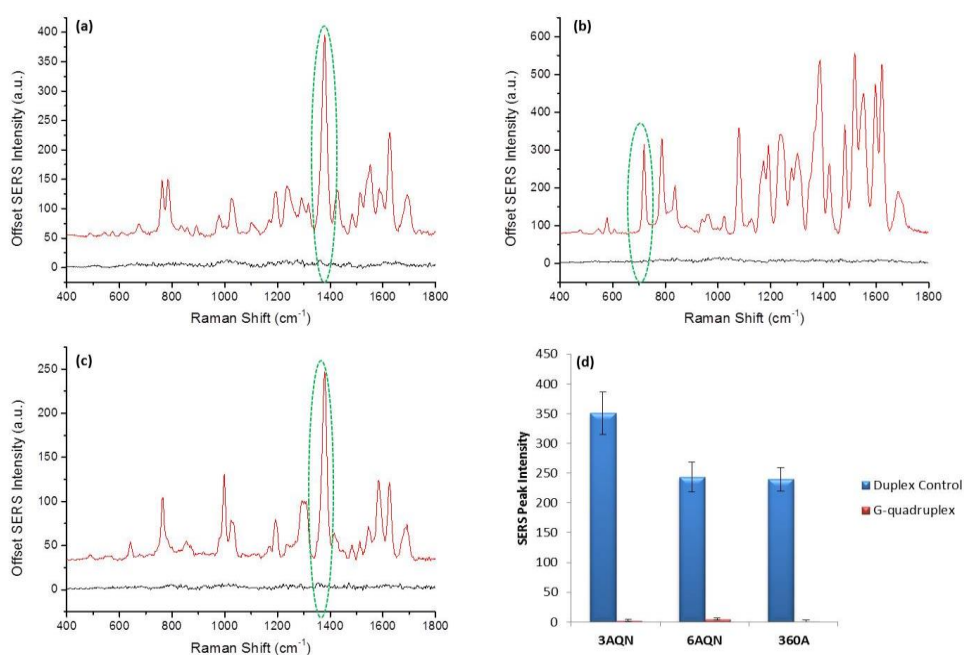


Figure 5 SERS spectra of each ligand binding to the peroxidase deoxyribozyme quadruplex (black) and the spectra obtained when the ligand is in the presence of duplex DNA control sequence (red): (a) spectra of 3AQN binding events; (b) spectra of 6AQN binding events and (c) spectra of 360A binding events. All three ligands and G-quadruplex were used at 50 μM . The bar chart, (d), is a comparison of the peak intensities of unique peaks of each ligand, highlighted by the green circle. In the spectrum of 3AQN, the peak at 1380 cm^{-1} was used for comparison studies, the peak at 719 cm^{-1} in the 6AQN spectrum was chosen and for 360A peak intensity studies, the unique peak at 1379 cm^{-1} was used. Spectra were obtained using 532 nm laser excitation and an accumulation time of 1 second. All peak intensities were obtained by scanning 3 replicate samples 5 times with an accumulation time of 1 second. Averages are shown and error bars are \pm one standard deviation.

These results were obtained when the ligands and the G-quadruplex DNA in equimolar concentrations (50 μM). An investigation was carried out to determine what effect reducing the concentration of G-quadruplex, whilst keeping the concentration of ligand constant, would have on the SERS response. Figure 6 shows the SERS results obtained when the G-quadruplex concentration was reduced from 50 μM to 5 nM. It can be seen that when the G-quadruplex concentration was reduced to below 1 μM a SERS response was obtained due to insufficient G-quadruplex being present to sequester all of the ligand resulting in signal being obtained. This demonstrates that, as expected, the ratio of ligand to G-quadruplex is vital. For example, if the formation of a G-quadruplex was to be monitored using a lower concentration of DNA then a corresponding lower concentration of ligand should be used however, it will be at this point an aggregating agent will be required as the amount of ligand present would not cause the aggregation previously obtained. It can be clearly seen in Figure 6 that by using the correct ratio of DNA to ligand results in a clear, sharp “on to off” signal being obtained upon formation of the G-quadruplex.

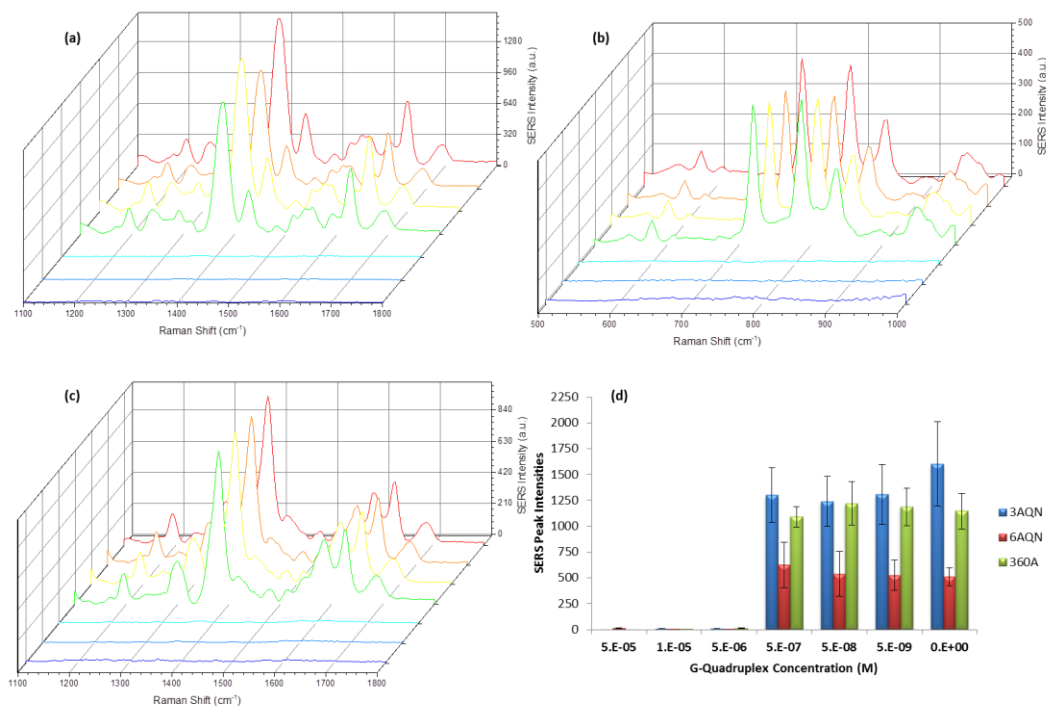


Figure 6 SERS spectra obtained from G-quadruplex concentration studies using each of the three ligands. The G-quadruplex concentration was reduced from 50 μM (dark blue), 10 μM (light blue), 5 μM (turquoise), 0.5 μM (green), 50 nM (yellow), 5 nM (orange) and no G-quadruplex present (red). The concentration of the ligands remained constant at 50 μM . (a) SERS spectra of 3AQN obtained when reducing the G-quadruplex concentration. (b) SERS spectra of 6AQN obtained when decreasing the G-quadruplex concentration. (c) SERS spectra of 360A obtained when decreasing the G-quadruplex concentration. (d) Peak intensity comparisons for each of the three ligands using; 1380 cm^{-1} for 3AQN comparisons, 719 cm^{-1} for 6AQN comparisons and 1379 cm^{-1} peak intensity comparisons. Spectra were obtained using 532 nm laser excitation and an accumulation time of 1 second. All peak intensities were obtained by scanning 3 replicate samples 5 times with an accumulation time of 1 second. Averages are shown and error bars are \pm one standard deviation.

Subsequent studies focussed on the ligand 3AQN and its behaviour towards various G-quadruplex DNAs and in different salt conditions. Results obtained thus far indicated that in each situation the ligand is bound to the G-quadruplex DNAs, i.e. when the ligand is no longer “free” there is a marked decrease in SERS intensity. For a G-quadruplex structure to form, it is reported that the presence of a monovalent ion, either K^+ or Na^+ , is essential.^{3,5} Experiments for this study were originally carried out using either 100 mM KCl or 100 mM NaCl to provide these monovalent ions for quadruplex formation. However, it was reported that when using certain stabilising agents such as the ligand 3AQN, neither salt is needed for G-quadruplex formation; the cation presence actually influences the orientation of the quadruplex structure, whether it is parallel or antiparallel.^{16,36,41} This is due to the ability of 3AQN itself, to induce the formation of a G-quadruplex reported by us.³⁶ Not only does it allow the formation of the desired structure, it also stabilises the quadruplex, which can be exploited to detect stable quadruplex structures *in vivo*.¹⁷ Figure 7(a-c) shows the SERS spectra obtained when NaCl or KCl is present and when salt is replaced by water in the reaction mixture and further analysis

of the intensity of the peak at 1380 cm^{-1} was performed (Fig 7d). There was a decrease in the absolute SERS intensity when the monovalent ions were absent due to the reduced aggregation of the nanoparticles that would have been caused by the salt, however the discrimination between the double stranded control DNA sequence and the G-quadruplex DNAs was still very significant. SERS signals were only observed when the G-quadruplex DNA was absent; no SERS signals were observed when the ligand was bound to a G-quadruplex sequence, regardless of the salt conditions. This further highlights the potential of using 3AQN as a stabilising agent for the formation of G-quadruplex DNAs as SERS has shown that it stabilises the G-quadruplex in the absence of monovalent cations.

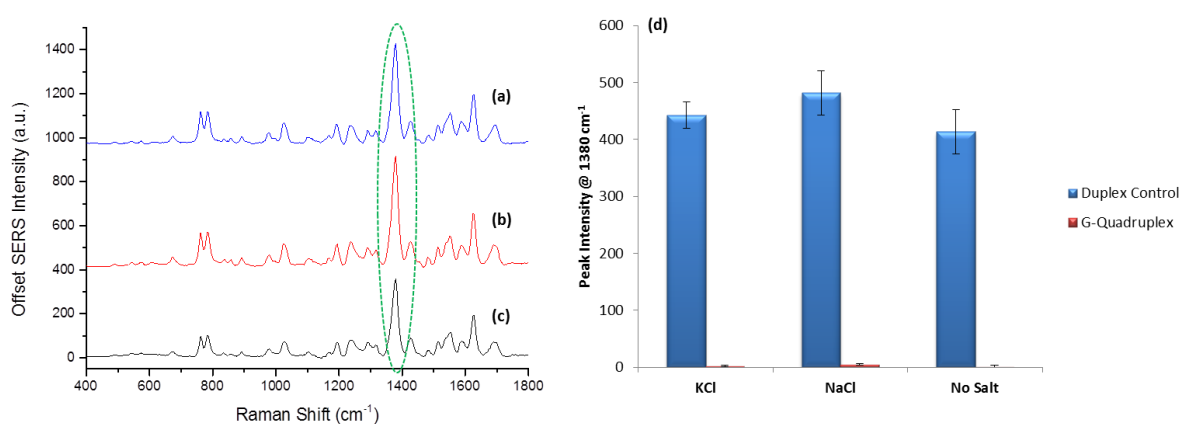


Figure 7 SERS spectra of 3AQN complexed to the duplex DNA control sequence in the presence of (a) Na^+ ions, (b) K^+ ions and (c) no monovalent ions. Spectra were also recorded in the different salt conditions when 3AQN was complexed to the peroxidase deoxyribozyme quadruplex sequence. (d) Comparison of peak intensities using the unique peak at 1380 cm^{-1} . Spectra were obtained using 532 nm laser excitation and an accumulation time of 1 second. All peak intensities were obtained by scanning 3 replicate samples 5 times with an accumulation time of 1 second. Averages are shown and error bars are \pm one standard deviation.

G-quadruplex forming sequences are also found in the promoter regions of oncogenes such as *C-MYC*. The *C-MYC* oncogene is the most common malfunctioning gene in human cancers; therefore it is an appealing target for anti-cancer therapies.⁴² The human *C-MYC* gene is highly regulated and any changes in its expression are a crucial point in cancer progression. Detection of these guanine-rich sequences is extremely beneficial to the field of cancer diagnostics. Results shown so far have proved that 3AQN can be used to successfully determine if a G-quadruplex sequence is present or absent. Therefore, studies were carried out to determine if 3AQN would bind to a G-quadruplex structure present in telomeric or in a proto-oncogene region such as *C-MYC*.

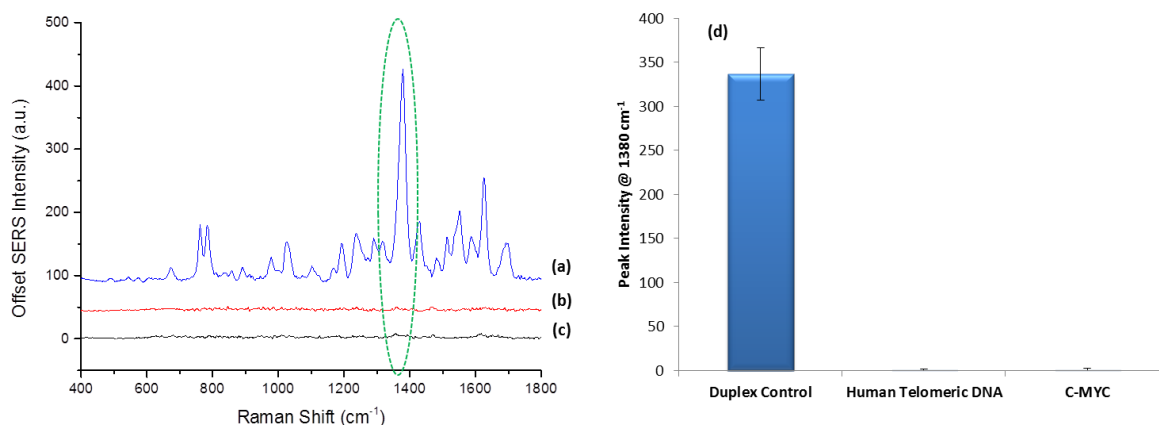


Figure 8 SERS spectra of 3AQN in the presence of (a) control duplex DNA, (b) human telomeric DNA, (c) *C-MYC* DNA and (d) a comparison of peak intensity at 1380 cm⁻¹. Spectra were obtained using 532 nm laser excitation and an accumulation time of 1 second. All peak intensities were obtained using the 1380 cm⁻¹ peak by scanning 3 replicate samples 5 times with an accumulation time of 1 second. Averages are shown and error bars are \pm one standard deviation.

Figure 8(a-c) shows that ligand peaks are only observed in the absence of a guanine rich DNA sequence, regardless of whether it is present in a telomeric end or in a proto-oncogene such as *C-MYC*. This demonstrates the strong affinity the ligand 3AQN has for a G-quadruplex topology, regardless of where it is found in a biological system and in the absence of the essential monovalent ions. Figure 8d is a comparison of the peak intensities at 1380 cm⁻¹ and as can clearly be seen there is a large discrimination between the ligand in the presence of the duplex control DNA and when the ligand is complexed to G-quadruplex DNAs. These results reaffirm the concept of using SERS and a G-quadruplex binding ligand such as 3AQN as a specific, efficient and reliable qualitative detection method of telomeric sequences or cancer promoting genes such as *C-MYC*.

Conclusions

Detection of the formation of G-quadruplex DNAs found in the peroxidase deoxyribozyme, telomeric region and in the proto-oncogene *C-MYC* have been demonstrated by a SERS based approach. Studies were carried out involving three quadruplex-binding ligands; 360A, 6AQN and 3AQN. Each produced successful results allowing detection of the formation of G-quadruplexes with high specificity. Further studies were performed using 3AQN that demonstrated the added stabilising affect offered by this ligand, due to the fact that the monovalent ions usually required for G-quadruplex formation are not needed when using 3AQN, can be directly detected using SERS without adding additional fluorescent agents and even in a salt-deficient condition. This is due to the

1
2
3 ability of this ligand itself to both induce the formation of the G-quadruplex structure and give a
4 unique SERS response. Since the ligands used here are known to bind/stabilise various quadruplexes,
5 this method may be extended to any quadruplex forming sequences present in the biological
6 landscape. This study also implies that SERS could be used to distinguish a particular quadruplex
7 topology over other quadruplex structures by employing quadruplex-specific ligands. By using SERS,
8 reliable results were obtained in a short time frame without the need for extensive laboratory work
9 and additional assays. The absence of an aggregating agent resulted in the lack of DNA peaks in the
10 SERS spectra, meaning the peaks observed result from the presence of the G-quadruplex stabilising
11 ligands. This is a new and information rich approach to higher order DNA structural analysis and
12 offers significant opportunities for use in studies involving G-quadruplexes as key biological targets.
13
14
15
16
17
18
19
20
21
22
23

24 Acknowledgements

25
26 KF and KG wish to thank the RSC Analytical Trust Fund and EPSRC for funding through the award of
27 DTG funding to KG. DG thanks the Royal Society for support from a Wolfson Research Merit award.
28 P.I.P. thank Council of Scientific and Industrial Research (CSIR), and Department of Atomic Energy-
29 Board of Research in Nuclear Sciences (BRNS-DAE), Government of India for the research grants.
30 V.D. thanks CSIR the for the fellowship. We also thank Prof. R. Murugavel, IIT Bombay for catalyzing
31 IIT Bombay-Strathclyde collaborations.
32
33
34
35
36
37
38
39
40

41 References

- 42 1. I. A. Pedroso, L. F. Duarte, G. Yanez, K. Burkewitz and T. M. Fletcher, *Biopolymers*, 2007, **87**,
43 74-84.
- 44 2. E. Y. Lam, D. Beraldi, D. Tannahill and S. Balasubramanian, *Nature Communications*, 2013, **4**,
45 1796-1796.
- 46 3. H. J. Lipps and D. Rhodes, *Trends in cell biology*, 2009, **19**, 414-422.
- 47 4. G. N. Parkinson, M. P. H. Lee and S. Neidle, *Nature*, 2002, **417**, 876-880.
- 48 5. H.-Q. Yu, D. Miyoshi and N. Sugimoto, *Journal of the American Chemical Society*, 2006, **128**,
49 15461-15468.
- 50 6. Y. Chen and D. Yang, in *Current Protocols in Nucleic Acid Chemistry*, John Wiley & Sons, Inc.,
51 2001.
- 52 7. S. Burge, G. N. Parkinson, P. Hazel, A. K. Todd and S. Neidle, *Nucleic Acids Research*, 2006,
53 **34**, 5402-5415.
- 54 8. N. V. Hud, F. W. Smith, F. A. L. Anet and J. Feigon, *Biochemistry*, 1996, **35**, 15383-15390.
- 55 9. B. Heddi and A. T. Phan, *Journal of the American Chemical Society*, 2011, **133**, 9824-9833.
- 56 10. A. T. Phan, *FEBS Journal*, 2010, **277**, 1107-1117.
- 57 11. M. Y. Kim, M. Gleason-Guzman, E. Izicka, D. Nishioka and L. H. Hurley, *Cancer Research*,
58 2003, **63**, 3247-3256.
- 59
60

12. S. Neidle and G. Parkinson, *Nature Reviews Drug Discovery*, 2002, **1**, 383-393.
13. L. Oganessian and T. M. Bryan, *Bioessays*, 2007, **29**, 155-165.
14. D. J. Patel, A. T. Phan and V. Kuryavyi, *Nucleic Acids Research*, 2007, **35**, 7429-7455.
15. J. L. Huppert, *Chemical Society Reviews*, 2008, **37**, 1375-1384.
16. M. Y. Kim, H. Vankayalapati, S. Kazuo, K. Wierzba and L. H. Hurley, *Journal of the American Chemical Society*, 2002, **124**, 2098-2099.
17. G. Biffi, D. Tannahill, J. McCafferty and S. Balasubramanian, *Nature Chemistry*, 2013, **5**, 182-186.
18. K. Shin-ya, K. Wierzba, K.-i. Matsuo, T. Ohtani, Y. Yamada, K. Furihata, Y. Hayakawa and H. Seto, *Journal of the American Chemical Society*, 2001, **123**, 1262-1263.
19. M. Fialova, J. Kypr and M. Vorlickova, *Biochemical and Biophysical Research Communications*, 2006, **344**, 50-54.
20. Y. Xu, Y. Noguchi and H. Sugiyama, *Bioorganic & Medicinal Chemistry*, 2006, **14**, 5584-5591.
21. F. He, Y. Tang, S. Wang, Y. Li and D. Zhu, *Journal of the American Chemical Society*, 2005, **127**, 12343-12346.
22. O. Y. Fedoroff, M. Salazar, H. Han, V. V. Chemeris, S. M. Kerwin and L. H. Hurley, *Biochemistry*, 1998, **37**, 12367-12374.
23. V. Viglasky, L. Bauer and K. Tluckova, *Biochemistry*, 2010, **49**, 2110-2120.
24. C. Wei, G. Jia, J. Yuan, Z. Feng and C. Li, *Biochemistry*, 2006, **45**, 6681-6691.
25. T. Miura and G. J. Thomas, *Biochemistry*, 1994, **33**, 7848-7856.
26. K. Kneipp, H. Kneipp, I. Itzkan, R. R. Dasari and M. S. Feld, *Chemical Reviews*, 1999, **99**, 2957-2975.
27. K. Kneipp, H. Kneipp, V. B. Kartha, R. Manoharan, G. Deinum, I. Itzkan, R. R. Dasari and M. S. Feld, *Physical Review E*, 1998, **57**, R6281-R6284.
28. S. E. J. Bell and N. M. S. Sirimuthu, *Journal of the American Chemical Society*, 2006, **128**, 15580-15581.
29. G. Breuzard, J. M. Millot, J. F. Riou and M. Manfait, *Analytical Chemistry*, 2003, **75**, 4305-4311.
30. G. Rusciano, A. C. De Luca, G. Pesce, A. Sasso, G. Oliviero, J. Amato, N. Borbone, S. D'Errico, V. Piccialli, G. Piccialli and L. Mayol, *Analytical Chemistry*, 2011, **83**, 6849-6855.
31. S. Balasubramanian, L. H. Hurley and S. Neidle, *Nature Reviews Drug Discovery*, 2011, **10**, 261-275.
32. D. Y. Sun, B. Thompson, B. E. Cathers, M. Salazar, S. M. Kerwin, J. O. Trent, T. C. Jenkins, S. Neidle and L. H. Hurley, *J. Med. Chem.*, 1997, **40**, 2113-2116.
33. S. Neidle, *Febs Journal*, 2010, **277**, 1118-1125.
34. S. Balasubramanian and S. Neidle, *Current Opinion in Chemical Biology*, 2009, **13**, 345-353.
35. C. Granotier, G. Pennarun, L. Riou, F. Hoffschir, L. R. Gauthier, A. De Cian, D. Gomez, E. Mandine, J. F. Riou, J. L. Mergny, P. Mailliet, B. Dutrillaux and F. D. Boussin, *Nucleic Acids Research*, 2005, **33**, 4182-4190.
36. V. Dhamodharan, S. Hari Krishna, C. Jagadeeswaran, K. Halder and P. I. Pradeepkumar, *J. Org. Chem.*, 2012, **77**, 229-242.
37. P. C. Lee and D. Meisel, *Journal of Physical Chemistry*, 1982, **86**, 3391-3395.
38. J. F. Riou, L. Guittat, P. Mailliet, A. Laoui, E. Renou, O. Petitgenet, F. Mégnin-Chanet, C. Hélène and J. L. Mergny, *Proceedings of the National Academy of Sciences*, 2002, **99**, 2672-2677.
39. G. Pennarun, C. Granotier, L. R. Gauthier, D. Gomez, F. Hoffschir, E. Mandine, J. F. Riou, J. L. Mergny, P. Mailliet and F. D. Boussin, *Oncogene*, 2005, **24**, 2917-2928.
40. A. De Cian, E. DeLemos, J.-L. Mergny, M.-P. Teulade-Fichou and D. Monchaud, *Journal of the American Chemical Society*, 2007, **129**, 1856-1857.
41. E. Izbicka, R. T. Wheelhouse, E. Raymond, K. K. Davidson, R. A. Lawrence, D. Y. Sun, B. E. Windle, L. H. Hurley and D. D. Von Hoff, *Cancer Research*, 1999, **59**, 639-644.

- 1
2
3 42. A. Siddiqui-Jain, C. L. Grand, D. J. Bearss and L. H. Hurley, *Proceedings of the National*
4 *Academy of Sciences*, 2002, **99**, 11593-11598.
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4
5 A SERS-based method has been developed for the analysis of G-quadruplex DNAs using three novel
6 stabilising ligands.
7
8
9
10

