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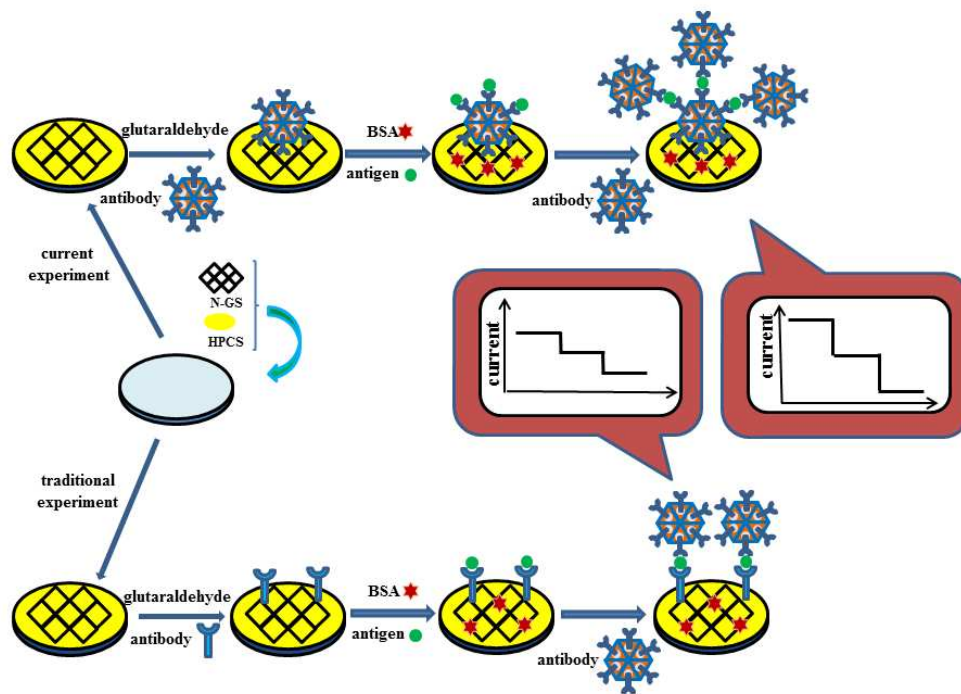
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1 **Ultrasensitive dual amplification sandwich immunosensor**
2 **for breast cancer susceptibility gene based on sheet materials**

3
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23 **Abstract**

24 A new electrochemical dual amplification sandwich immunosensor
25 (DASI) for ultrasensitive and accurate detection of breast cancer
26 susceptibility gene was designed based on the combination of N-doped
27 graphene, hydroxypropyl chitosan and Co_3O_4 mesoporous nanosheet.
28 N-doped graphene has better electroconductibility than traditional
29 graphene. It is an ideal electrochemistry material with large specific
30 surface area and little resistance. Hydroxypropyl chitosan replaces the
31 pure chitosan in the immobilization of the sensor to realize the sensitivity
32 increasement. Co_3O_4 mesoporous nanosheet can enhance the effective
33 area of the immunoreaction. This kind of dual amplification sandwich
34 immunosensor was first used to the detection of breast cancer
35 susceptibility gene. It has a wide linear response range of 0.001-35 ng/mL
36 and a low detection limit of 0.33 pg/mL. It demonstrated that the stability,
37 selectivity and reproducibility of the sensor were acceptable. The
38 fabricated immunosensor shows great potential application in early
39 disease diagnosis.

1. Introduction

The earlier clinical diagnosis of tumor makers can be detected, the easier cancer can be cured successfully. Recently, the reliable and effective detection of tumor maker is current studies¹⁻³. Breast cancer susceptibility gene (BRCA1), can be detected in the population of pancreatic cancer, stomach cancer and colon cancer⁴. BRCA1 mutation, caused about 40%-50% of hereditary breast cancers, is closely combined with the occurrence of the family ovary and breast cancer⁵. Therefore it is extremely urgent for the high-risk to develop a new type method to detect the BRCA1 tumor makers. Traditional ways in detecting BRCA1 include high performance liquid chromatography⁶, enzymatic mutation detection⁷ and protein truncation test⁸. However, these methods are complex and time-consuming, and the test processing requires large and expensive apparatus. Therefore, to develop a rapid, sensitive and cheap method for BRCA1 detection is a great challenge.

Electrochemical immunosensor is a kind of new method for the trace detection. It has been used in many fields such as food⁹, molecular imprinting¹⁰, tumor maker¹¹, virus analysis¹² and environmental estrogen analysis¹³. The sandwich immunosensor, compared with label-free or competitive type, exhibits a higher sensitivity and lower detection limit^{11, 14, 15}. That may attribute to the fact that dual amplification sandwich immunosensor (DASI) can capture more antigens. In previous study,

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4 64 sandwich immunosensor was used extensively due to its excellent
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6 65 payload capability like enzyme, metal ions or electron mediator². But in
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9 66 this research, the DASI was fabricated by initial antibody and second
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11 67 antibody all incubated with mesoporous materials, thus leading to a dual
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13 68 amplification effect. The incubated initial antibody can capture more
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15 69 antigens to enhance the sensitivity, and the incubated second antibody
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17 70 which attached more noble metal nanoparticles can strengthen payload
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19 71 capability to amplify the reaction signals.
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24 72 Graphene sheets (GS), a two-dimensional hexagonal lattice structure,
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26 73 has attracted a great deal of interest due to its high conductivity, large
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28 74 specific surface area, good biocompatibility and potential applications in
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30 75 immunosensors^{16, 17}. Recently, N-doped graphene sheets (N-GS) have
31
32 76 been used in many fields such as electrochemiluminescence, molecular
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34 77 imprinting, battery and electrocatalysis^{10, 18-20}. Typically, GS is a kind of
35
36 78 good conductor, while the nitrogen replaces the carbon atom of the
37
38 79 hexagonal lattice skeleton structure. Hence, the obtained N-GS exhibit
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40 80 excellent more electroconductibility than the GS. That can be explained
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42 81 as follows²⁰⁻²²: there exist six electrons outside of the carbon atom, and
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44 82 four in the six electrons are in four p orbital separately of the nuclear
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46 83 structure, so three of the four p orbital can be bonding each other due to
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48 84 the hexagonal lattice skeleton structure, which may lead to a hanging p
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50 85 orbital (one electron) unpaired. When the nitrogen replaces the carbon
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4 86 atom of GS, there exist a p orbital with two electrons owing to the N has
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6 87 seven electrons and five of them are filled with four p orbital of the
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9 88 nuclear structure. According to the bond of hexagonal lattice skeleton
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11 89 structure of the GS, the remaining one is the two electrons p orbital,
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13 90 which can increase the amount of the unpaired electrons. And the
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15 91 electroconductibility of the GS derive from the electrons. In addition,
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17 92 such doped N atoms can decorate the graphene planar sheet and introduce
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19 93 a change in the Fermi level, engendering the doping effects and opening
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21 94 the band gap of the graphene. Therefore, the great electroconductibility of
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23 95 N-GS exceed GS.

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29 96 Chitosan is widely used in immunosensors in view of its wonderful
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31 97 film-forming property²³. Nowadays, many chitosan derivatives have been
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33 98 applied in electroanalytical chemistry²⁴. Here, hydroxypropyl chitosan
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35 99 (HPCS), which is full of amino and can combine more biomolecule than
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39 100 chitosan, was used to fabricate the immunosensor. Mesoporous materials
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41 101 were used in immunosensors extensively, such as Fe_3O_4 ²⁵, SiO_2 ²⁶,
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43 102 mesoporous noble metal²⁷ and alloy^{28, 29}. Many mesoporous materials are
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46 103 spherical or other block shapes with large specific surface area and
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49 104 abundant of pores and canals. However, a lot of pores and canals of the
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51 105 materials are inside of them, which may decrease the quantity of noble
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54 106 metal attached on the inner pores and block the reaction of immunizing
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57 107 conjugation. Therefore, mesoporous sheet materials are the perfect
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4 108 candidate for the immunosensor. Here, mesoporous nano- Co_3O_4 sheet
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6 109 materials were synthesized for the DASI. The aminated Co_3O_4 is
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9 110 combined with silver nanoparticles to form the $\text{Ag}@\text{Co}_3\text{O}_4$. The initial
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11 111 antibody and second antibody are all incubated with it to the formation of
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13 112 $\text{Ab-Ag}@\text{Co}_3\text{O}_4$ because of the combination of Ag nanoparticles and
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16 113 amino.

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19 114 Herein, a novel electrochemical DASI for the detection of BRCA1 has
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21 115 been fabricated. The immune reaction based on the specific binding of
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23 116 antibody and antigen was performed on the sheet materials. N-GS can
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25 117 increase the electroconductibility significantly and decrease the protein
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27 118 resistance. $\text{Ag}@\text{Co}_3\text{O}_4$ can enlarge the effective specific surface area for
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29 119 the sensitive detection. This immunosensing method is novel and
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31 120 effective, which may provide a potential application in clinical tests.
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34 121 **Experimental section**

35 122 *Materials and reagents*

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41 123 Rabbit anti-BRCA1 (primary antibody), BRCA1 and BSA were
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43 124 purchased from Sigma-Aldrich. HPCS was gained from Lvshen
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45 125 Bioengineering Co., Ltd. (Nantong, China). Glutaraldehyde (25%),
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47 126 ammonia, sodium borohydride, silver nitrate, graphite, cobalt chloride
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49 127 hexahydrate and ethylene glycol were obtained from Sinopharm
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51 128 Chemical Reagent Co., Ltd. (Beijing, China). Phosphate buffer solutions
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53 129 (PBS) were prepared by compounding the solutions of KH_2PO_4 (0.067
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4 130 mol/L) and Na_2HPO_4 (0.067 mol/L) to appropriate pH value. PBS was
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6 131 used as electrolyte for all electrochemistry measurements. Ultrapure
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8 132 water was used throughout the experiment. All other chemicals were of
9
10 133 analytical reagents grade and used without further purification.

134 *Apparatus*

135 All electrochemical measurements were carried out on an
136 electrochemical workstation (CHI760D, Chenhua Instrument Shanghai
137 Co., Ltd., China). A conventional three-electrode configuration was used.
138 Working electrode is glassy carbon electrode (GCE, 4 mm diameter);
139 reference electrode is a saturated calomel electrode (SCE) and Pt as the
140 counter electrode. Surface area measurements were performed on
141 Micromeritics ASAP 2020 surface area and porosity analyzer
142 (Quantachrome, United States). X-ray powder diffraction (XRD) patterns
143 were acquired from a Bruker D8 Focus diffractometer (Germany) using
144 $\text{CuK}\alpha$ radiation (40 kV, 30 mA) of wavelength 0.154 nm. Transmission
145 electron microscope (TEM) images were recorded from a JEOL
146 JEM-1400 microscope (Japan). Scanning electron microscope (SEM) and
147 Energy Dispersive X-Ray Spectroscopy (EDS) were obtained by JEOL
148 JSM-6700F microscope (Japan)

149 *Synthesis of materials*

150 The synthesis of N-GS can be divided into two steps: preparation of
151 graphene oxide (GO) and process of doping nitrogen. GO was prepared

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4 152 according to a reported method³⁰. In detail, graphite powder (0.3 g) and
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6 153 potassium permanganate (1.8 g) were added into a solution of mixed
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9 154 acid (36 mL H₂SO₄ and 4 mL H₃PO₄). Then the mixture was kept stirring
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11 155 at 50 °C for 12 h. After that, the mixture was poured slowly onto a
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14 156 pre-prepared ice (~ 400 mL) and 6 mL H₂O₂ was added into it dropwise.
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16 157 Subsequently, the filtrate was centrifuged (8000 rpm, 30 min). The
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19 158 remaining solid was washed in succession with 200 mL of water, 200 mL
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21 159 of 30% HCl, and 200 mL of ethanol for 3 times. And the resulting solid
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24 160 was dried at vacuum at the temperature of 35 °C for 24 h. GO was
25
26 161 synthesized successfully. N-GS was prepared simply based on GO³¹. The
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29 162 as-synthesized GO was firstly dispersed in DMF (0.5 mg/mL) under
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31 163 ultrasonically treatment for 30 min, and then heated in an oil bath (153 °C)
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34 164 for 1 h. The N-GS was synthesized successfully.

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36 165 Mesoporous nano-Co₃O₄ was prepared as follows³². 10 mmol
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39 166 CoCl₂·6H₂O was dissolved into 50 mL ultrapure water to form a
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42 167 homogeneous solution under magnetic stirring. An appropriate amount of
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44 168 sodium hydroxide dissolved in 20 ml distilled water was dropwise into
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47 169 the above solution. The brown precipitation was centrifuged, washed with
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50 170 distilled water and absolute alcohol three times, and dried at 60 °C in
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52 171 vacuum for 12 h. Finally, the precipitation was heated at 400 °C for 1 h in
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55 172 the air and slowly cooled to room temperature. And the Co₃O₄ was
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57 173 obtained for further usage.
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4 174 Ag nanoparticles (Ag NPs) were prepared by a classical approach³³.
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6 175 Briefly, AgNO₃ (1 mL, 50 mM) and trisodium citrate (1 mL, 5% (w/w))
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9 176 were mixed with ultrapure water under vigorous stirring for 1 h. Then 0.1
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11 177 mM of NaBH₄ solid was added into above mixture. Under continuous
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13 178 stirring for about 20 min, the mixture's color changed to brown-yellow
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16 179 and indicated the successful synthesis of Ag NPs. Finally, the solution
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19 180 was continuously stirred until the color not change.

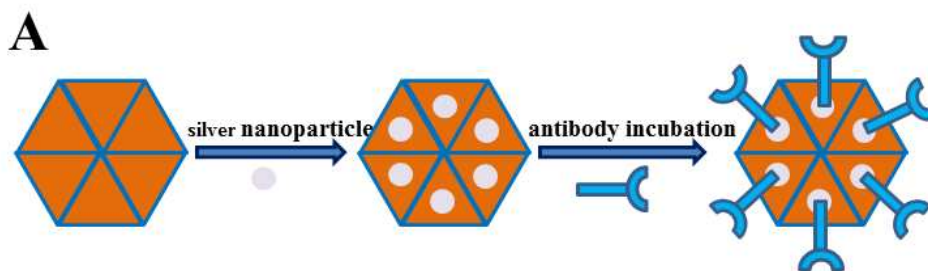
20
21 181 Aminated Co₃O₄ was synthesized from some reported methods^{34, 35}.
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23 182 The as-synthesized Co₃O₄ (0.5 g) was dispersed in anhydrous toluene (50
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25 183 mL) in a flask and heated to 70 °C. Then, 3-aminopropyltriethoxysilane
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27 184 (500 μL) was dropped to the solution rapidly and the reaction refluxed 3
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29 185 h at constant temperature 70 °C. After centrifuged, the solid product was
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31 186 washed 3 times with absolute alcohol and dried at 30 °C for 24 h. A
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33 187 free-flowing powdery material was obtained. The aminated Co₃O₄ was
34
35 188 shown to contain -NH₂ by ninhydrin test³⁶.

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37 189 Ag@Co₃O₄ was prepared by mixing of Ag NPs and aminated Co₃O₄.
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39 190 Ag NPs can connect with -NH₂ tightly³⁷. Thus, the aminated Co₃O₄ was
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41 191 put into Ag NPs solution and centrifuged to obtain Ag@Co₃O₄ until the
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43 192 supernate was a little yellow which indicated the Co₃O₄ adsorbed Ag NPs
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45 193 sufficiently.

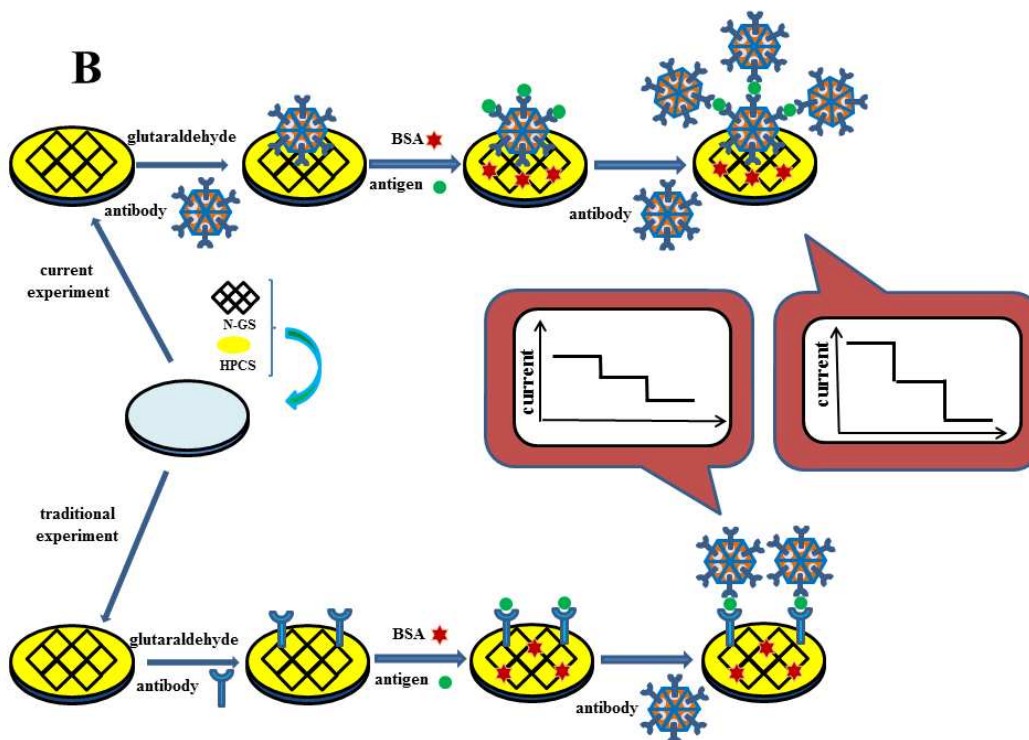
46 194 *Fabrication of the immunosensor*

47 195 Scheme 1 illustrates the process of the proposed method. It can be
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4 196 divided into two sections: incubation of the Ab-Ag@Co₃O₄ and
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6 197 fabrication of the immunosensor. From Scheme 1A, aminated Co₃O₄ is
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9 198 covered with Ag nanoparticles, and BRCA1 antibodies can connect on it.
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11 199 From Scheme 1B, in current experiment, HPCS is mixed with N-GS
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13 200 under ultrasonication to form uniform solution and the 6 μL of the
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15 201 prepared solution (1.0 mg/mL) is dropped on the surface of the polished
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17 202 electrode. Then 6μL of the incubated Ab-Ag@Co₃O₄ is connected on it
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19 203 through glutaraldehyde and 3 μL of bovine serum albumin (BSA) is
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21 204 covered on the former layer to block the nonspecific active sites.
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23 205 Subsequently, BRCA1 antigens are adsorbed on the antibodies of
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25 206 Ab-Ag@Co₃O₄ through specific binding of the immune substance.
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27 207 Finally, 6 μL of the Ab-Ag@Co₃O₄ is dropped on the electrode. That is
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29 208 the preparation of the DASI. And the DASI achieve the effect of space
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31 209 amplification. In traditional method, the previous sandwich
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33 210 immunosensor is fabricated in a similar way^{26, 38}. However, in the current
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35 211 research, the primary antibody was incubated with Ag@Co₃O₄ to amplify
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37 212 the antigen loading capacity instead of pure antibody. Therefore this kind
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39 213 is plane amplification. And in this way, the DASI (current experiment)
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41 214 can enhance the sensitivity than the common sandwich immunosensor
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43 215 (traditional experiment) due to the large amplification of the spatial
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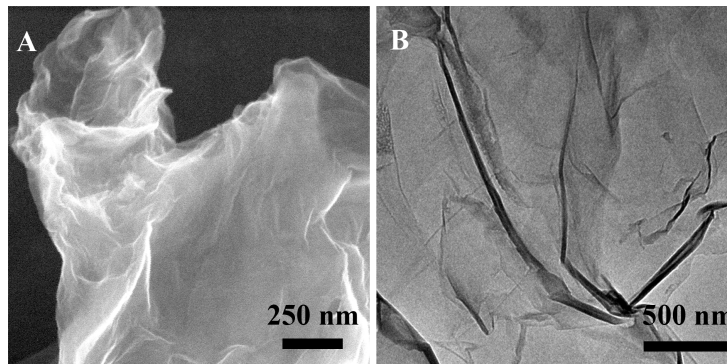
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219 Scheme 1 the incubation of Ab-Ag@Co₃O₄ (A); fabrication of the DASI in both

220 current experiment and traditional experiment (B)

221 **Results and Discussion**222 *Characterization of the materials*

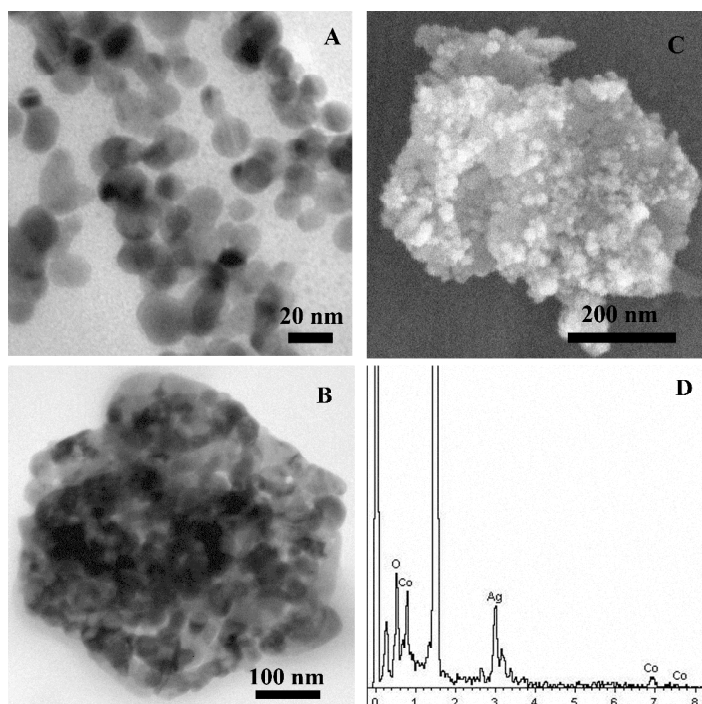
223 Fig. 1 shows the basic characterization of the N-GS. The base
224 material of the sensor, exhibits the wrinkled paper morphology in the
225 SEM image (Fig. 1A) and TEM image (Fig. 1B).



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Fig. 1 SEM image (A) and TEM image (B) of N-GS



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Fig. 2 TEM image (A) of Ag NPs, TEM image (D) of mesoporous Co_3O_4 nanosheet;

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SEM image (E) and EDS diagram (F) of the $\text{Ag}@ \text{Co}_3\text{O}_4$.

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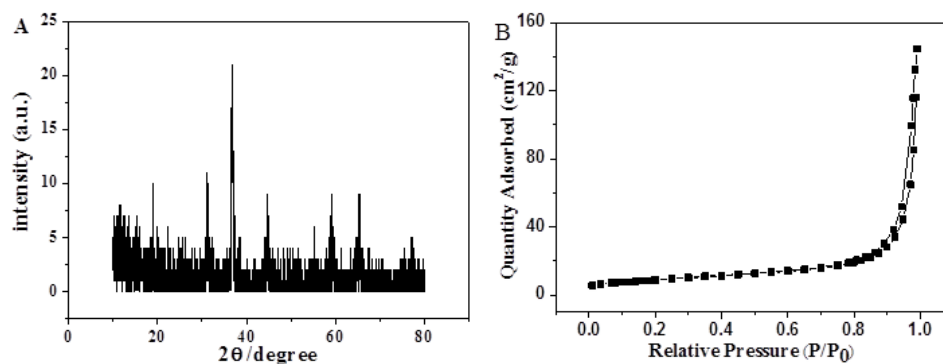
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From Fig. 2A, we can clearly observe the Ag NPs are about 15 nm in diameter. The mesoporous structure of the hexagonal Co_3O_4 nanosheets can be seen from Fig. 2B, which demonstrates the holes of Co_3O_4 are irregular and without standard size. In this research, $\text{Ag}@ \text{Co}_3\text{O}_4$ is used to prepare the sensor, and $\text{Ag}@ \text{Co}_3\text{O}_4$ is also characterized by SEM (Fig. 2C)

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4 236 and EDS (Fig. 2D). In Fig. 2C, there exists a mass of bright points
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6 237 attached on the surface and holes of hexagon. The bright points are the
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9 238 Ag nanoparticles combined on the aminated Co_3O_4 . And EDS diagram
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11 239 further proves the $\text{Ag}@\text{Co}_3\text{O}_4$ synthesized successfully.

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14 240 The morphology about the Co_3O_4 is detailed description above. But
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16 241 its specific information can be concluded from the XRD and BET (Fig. 3).
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18 242 The XRD patterns of cubic phase Co_3O_4 JCPDS card is No. 73-1701. And
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20 243 its diffraction pattern is in accordance with Fig. 3A. By Debye-Scherrer
21
22 244 analysis, the lattice parameters $a=b=c=8.08 \text{ \AA}$ accord with the standard
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24 245 values of cubic Co_3O_4 . BET can reflect the specific information about the
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26 246 mesoporous nanosheet. Its BET surface area is $31.7745 \text{ m}^2/\text{g}$; pore
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28 247 volume (single point adsorption) is $0.1004 \text{ cm}^3/\text{g}$ and the average pore
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30 248 size is 12.64 nm . In Fig. 3B, we can see clearly the adsorption line (the
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32 249 lower curve) and the desorption line (the upper curve) are nearly
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34 250 coincident, which demonstrates the sheet material has much effect outside
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36 251 specific surface area.

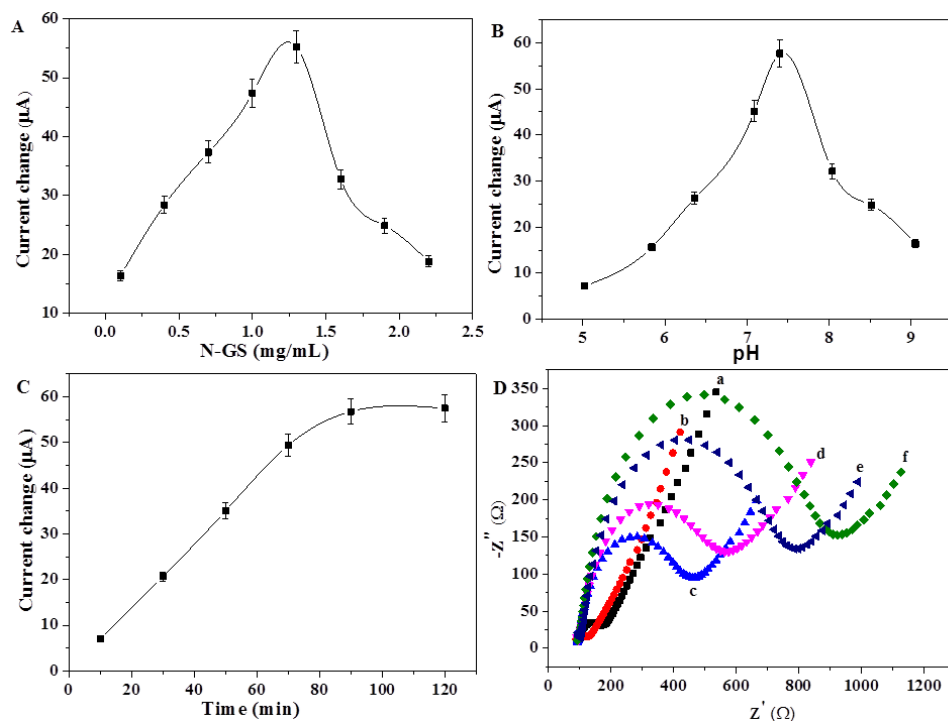


253 Fig. 3 XRD patterns of Co_3O_4 (A); BET analysis of Co_3O_4 (B)

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4 254 *Characterization of the immunosensor*

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6 255 To characterize the layer by layer self-assembly successfully,
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9 256 electrochemical impedance spectroscopy (EIS) was employed in the
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11 257 research using 2.5 mmol/L $\text{Fe}(\text{CN})_6^{3-}/\text{Fe}(\text{CN})_6^{4-}$ containing 0.1 mmol/L
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14 258 KCl solution. The EIS curves can be divided in two parts: the high
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16 259 frequency region and the low frequency region. The high frequency
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18 260 region is like a semicircle, which is related to the redox probe $\text{Fe}(\text{CN})_6^{3-/4-}$.
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21 261 And the low frequency is a Warburg line corresponding to the diffusion
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23 262 step of overall process³⁹. The resistance of the immunosensor can be
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25 263 calculated from the semicircle diameter in the nyquist plots of the EIS. In
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27 264 Fig. 4D, the nyquist plots of the EIS can be seen clearly. Curve a stands
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29 265 for the EIS of bare glassy carbon electrode (GCE) with a quite small
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31 266 semicircle diameter and a straight line, indicating the GCE polished well
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33 267 and held little resistance. Curve b represents the HPCS@N-GS modified
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35 268 layer, exhibiting a much small semicircle diameter than bare GCE that
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37 269 may be due to the N-GS super electroconductibility. Then the
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39 270 Ab-Ag@Co₃O₄ layer is described by curve c, which shows a big arc in
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41 271 high frequency because of the resistance of the BRCA1 antibody.
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43 272 Subsequently, BSA and BRCA1 antigen are dropped on the electrode, and
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45 273 their semicircle diameters of EIS plots (curve d and curve e) enhance
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47 274 gradually. Last, Ab-Ag@Co₃O₄ is connected again with a large diameter
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49 275 EIS plots. All the EIS plots indicate the layer by layer self-assembly was
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276 effective.



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278 Fig. 4 A): effect of N-GS concentration on the response of the DASI (pH=7.4,

279 BRCA1 antigen 20 ng/mL, $n=5$); B): effect of pH on the response of the DASI (N-GS

280 1.3 mg/mL, BRCA1 antigen 20 ng/mL, $n=5$); C): effect of incubation time on the

281 response of the DASI (pH=7.4, N-GS 1.3 mg/mL, BRCA1 antigen 20 ng/mL, $n=5$);

282 D): EIS obtained for different modified electrodes: (a) GCE, (b) GCE/HPCS@N-GS,

283 (c) GCE/HPCS@N-GS/Ab-Ag@Co₃O₄, (d)

284 GCE/HPCS@N-GS/Ab-Ag@Co₃O₄/BSA, (e)

285 GCE/HPCS@N-GS/Ab-Ag@Co₃O₄/BSA/BRCA1, (f)

286 GCE/HPCS@N-GS/Ab-Ag@Co₃O₄/BSA/BRCA1/ Ab-Ag@Co₃O₄.

287 *Optimization of experimental conditions*

288 In order to obtain the best analytical performance, some testing

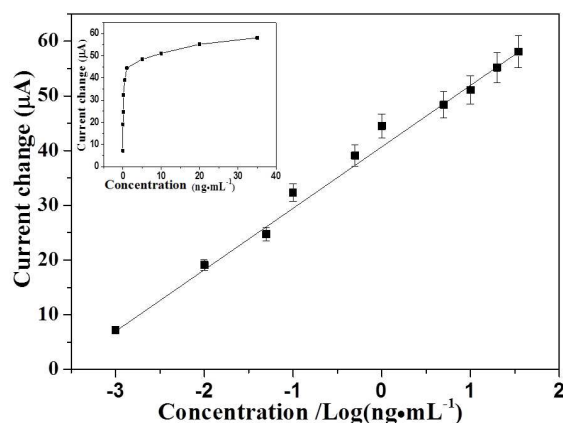
289 conditions have been optimized. N-GS are modified on the electrode to

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4 290 amplify the area of the GCE without inducing great resistance. So the
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6 291 concentration of N-GS requires to be selected. In Fig. 4A, it is clear that
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9 292 the current change of the DASI firstly increases and then decreases, and
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11 293 the top point is 1.3 mg/mL. This phenomenon can be explained as follows:
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14 294 within low concentration, N-GS can increase the electroconductibility
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16 295 and enlarge contacting area, but in high concentration, N-GS
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18 296 electroconductibility cannot enough to offset overmuch layer resistance
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21 297 of the sheets. The best concentration is 1.3 mg/mL. The pH value is an
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24 298 important factor in the electrochemistry test as the acid-base condition
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26 299 may exhibit different results. In this research, pH=7.4 PBS (phosphate
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29 300 buffer solution) is selected as the best testing environment (Fig. 4B). That
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31 301 may be due to two reasons: First, neutral solution is suitable for the
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34 302 activity of biomolecule since the antibody-antigen linkage would be
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36 303 broken under alkalinity or acidity conditions; Second, in this research,
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39 304 H₂O₂ is selected as the signal source and Ag@Co₃O₄ exhibits obvious
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41 305 catalytic power to H₂O₂ in pH=7.4 PBS. Hence pH=7.4 PBS is chosen as
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44 306 the suitable buffer solution. Incubation time is also an important factor in
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47 307 the fabricating process. The incubation time can affect the loading
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49 308 quantity of the BRCA1 antigen and the activity of the biomolecule. From
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51 309 Fig. 4C, 90 min is selected as the adaptive incubation time with high
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54 310 signal response.

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56 311 *Analysis and detection*
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4 312 Under optimum testing conditions, the DASI was used to detect
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6 313 different concentrations of BRCA1 antigen. DASI was fabricated in
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9 314 Scheme 1 and H₂O₂ was used as the redox probe to record the electron
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11 315 transfer generated from H₂O₂ to the electrode when the immunological
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14 316 reaction occurs. And the best potential to catalyze H₂O₂ was from -0.4 V
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16 317 to -0.2 V. The sensitivity of Ag@Co₃O₄ modified electrode toward H₂O₂
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18 318 increased with the negative increase of detection potential. At higher
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21 319 detection potentials, such as -0.6 V, the background of the electrode is
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24 320 also increased and the O₂ reduction will interfere with H₂O₂ detection⁴⁰.
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26 321 Thus, in this research, cyclic voltammograms were performed from -0.6
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29 322 V to 0.2 V to show the electrochemical response and -0.4 V was selected
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31 323 the suitable reduction potential (Supplementary material, Fig. S1). After
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34 324 that, 5 mmol/L H₂O₂ was added into the buffer solution, and the current
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36 325 change was recorded (Supplementary material, Fig. S2). According to the
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39 326 current changes, calibration curve was drawn in Fig. 5. The current
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41 327 changed linearly with logarithm of the antigen concentrations. The equation
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44 328 of the calibration curve is $Y = 41.43 + 11.39 X$, $r = 0.9978$. And the linear
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46 329 range is 0.001-35 ng/mL with a low detection limit of 0.33 pg/mL at a
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49 330 signal to noise ratio of 3σ (where σ is the standard deviation of a blank
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51 331 solution, $n = 11$, $r = 0.9978$). And this method for the detection of
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54 332 BRCA1 has a better result than some reported researches (Table 1)⁴¹⁻⁴³.
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56 333 The low detection limit can be attributed to three factors: First, N-GS can
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334 enhance the electroconductibility and specific surface area markedly;
 335 second, DASI can adsorb more antigens to amplify the signals or even in
 336 low antigen concentration, more signal source substance can connect on
 337 the only antigen; third, Co_3O_4 nanosheet can enlarge the effective contact
 338 area to magnify biological fixation loads.



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340 Fig. 5 Calibration curve of the prepared DASI towards different concentrations of

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BRCA1, error bar = RSD (n = 5).

342 scientific research

Linear range

Detection limit

Reference

343 Present research

0.001-35 ng/mL

0.33 pg/mL (10^{-15} mol/L)

344 Biochip (molecular

 $2 \times 10^{-5} \sim 5 \times 10^{-6}$ mol/L 7×10^{-8} mol/L

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345 beacon detection scheme)

346 Monolithic silicon

 $1 \times 10^{-9} \sim 5 \times 10^{-7}$ mol/L 9×10^{-10} mol/L

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347 optocoupler array

348 Enzyme-free amplified

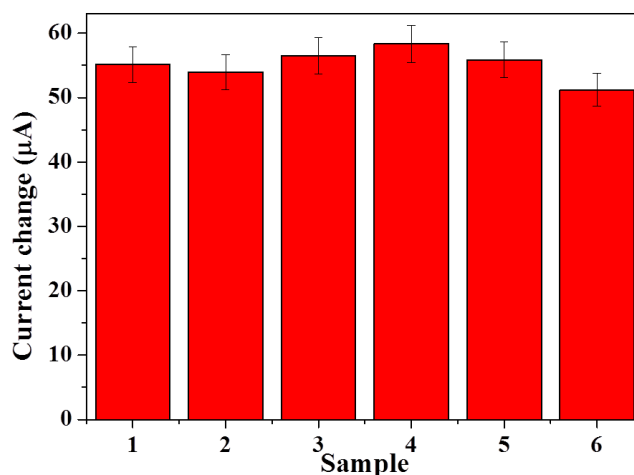
 $1 \times 10^{-6} \sim 1 \times 10^{-13}$ mol/L 1×10^{-13} mol/L

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detection platform

Table 1 Comparison of the present research and other reports

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4 349 Selectivity of DASi plays an essential role in the determination of
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6 350 analyzing biological samples⁴⁴. Some else tumor makers were used to
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9 351 carry out the selectivity research. In Fig. 6, six DASi were fabricated as
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11 352 the former way with 20 ng/mL of BRCA1, and five of them contain 200
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13 353 ng/mL of different interfering antigens (alpha fetal protein, BSA,
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16 354 carcino-embryonic antigen, human immune globulin, melanoma antigen).
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18 355 The result displays that the current changes of the DASi were all less than
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21 356 6.2% with adding interfering antigens which demonstrated the selectivity
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24 357 of DASi was acceptable. Stability is another factor of the DASi in
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26 358 potential practical application. Some fabricated electrodes were stored in
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29 359 refrigerator at 4 °C. 10 days later, the signal strength decreased to 98% of
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31 360 initial value. And one month later, compared with the initial, 95% signal
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34 361 strength was obtained. Reproducibility is also operated in this research. 5
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36 362 prepared DASi were tested in identical conditions, and the relative
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39 363 standard deviation (RSD) of the measurement was 4.8%, suggesting the
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41 364 precision of the DASi was fairly good for the detection of BRCA1. The
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44 365 selectivity, stability and reproducibility of the DASi exhibit tiny current
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46 366 change, indicating the result was satisfactory and acceptable.
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368 Fig. 6 The column of interference (1) 20 ng/mL BRCA1; (2) 20 ng/mL BRCA1+200
 369 ng/mL alpha fetal protein; (2) 20 ng/mL BRCA1+200 ng/mL BSA, (3) 20 ng/mL
 370 BRCA1+200 ng/mL carcino-embryonic antigen; (4) 20 ng/mL BRCA1+200 ng/mL
 371 human immune globulin; (5) 20 ng/mL BRCA1+200 ng/mL melanoma antigen. Error
 372 bar=RSD ($n=5$).

Human serum sample (ng/mL)	The addition content (ng/mL)	The detection Content (ng/mL)	RSD (% , $n=5$)	Recovery (%)
0.83	1	1.81 ± 0.04	1.60	101.8
0.83	5	5.86 ± 0.07	5.41	100.5
0.83	10	10.92 ± 0.08	6.95	100.9

373 Table 2 Results for the detection of BRCA1 in human serum sample by DASI.

374 In this research, in order to evaluate the performance of the sensor,
 375 human serum sample tests are carried out on DASI with standard addition
 376 method to verify the precision. The relative standard deviations (RSD) for

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4 377 1 ng/mL, 5 ng/mL and 10 ng/mL are found to be 1.60%, 5.41%, 6.95%,
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6 378 respectively. And the achieved BRCA1 recoveries were 101.8%, 100.5%
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9 379 and 100.9%, respectively (Table 2).

380 **Conclusion**

381 In conclusion, we reported an Ag@Co₃O₄-based DASI for the
382 sensitive detection of BRCA1. The strategy combines the advantages of
383 N-GS and Ag@Co₃O₄ nanosheet. And HPCS was used in the sensor to
384 enhance the biological adsorption. This novel immunosensor achieved a
385 low detection limit of 0.33 pg/mL and a wide linear range of 0.001-35
386 ng/mL. This DASI had wonderful property of selectivity, stability and
387 reproducibility, and human serum testing achieved a decent result. In
388 addition, this DASI was easily manipulated and low cost. And the
389 proposed method can test other tumor makers or it has potential
390 application in clinical diagnosis.

391 **Acknowledge**

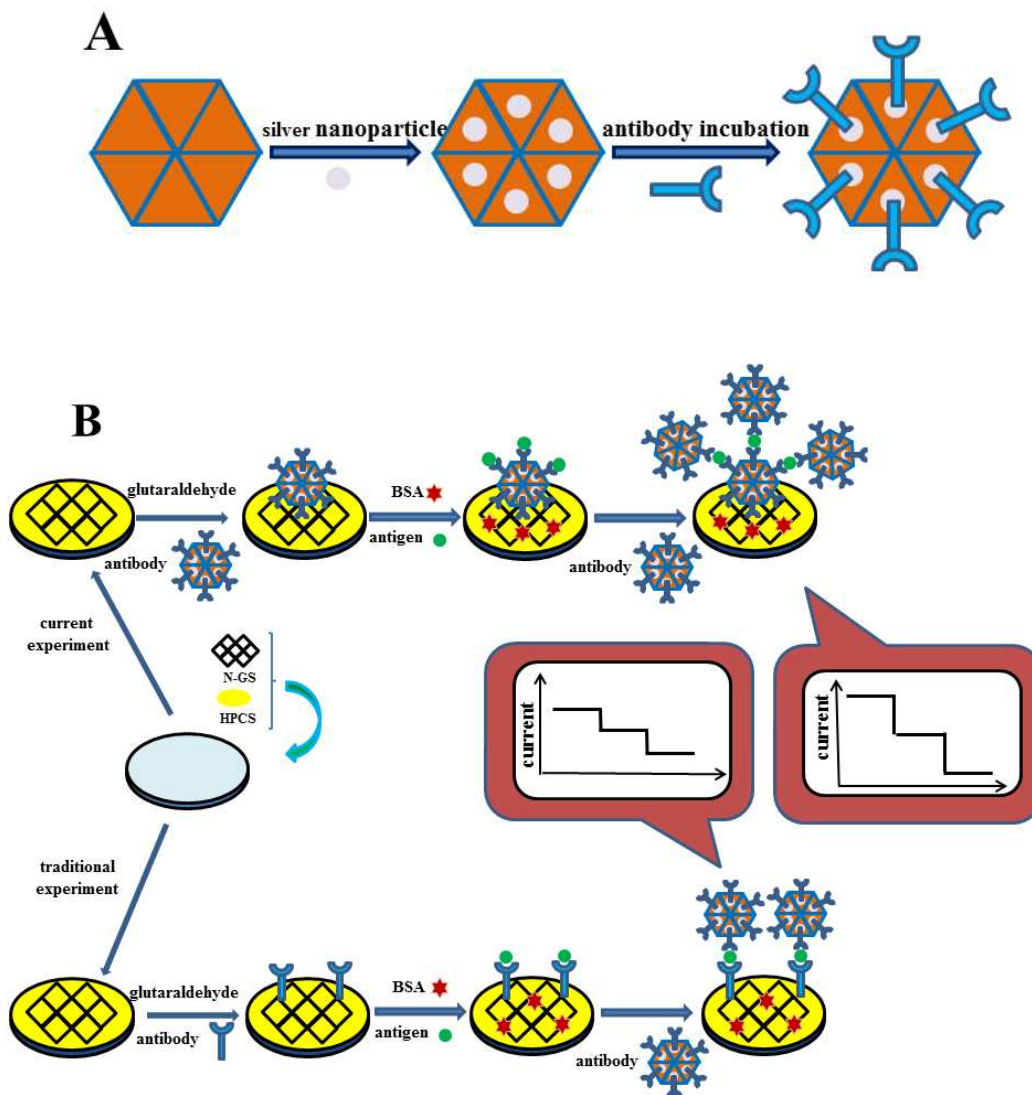
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398 **References**

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Scheme 1 the incubation of Ab-Ag@Co₃O₄ (A); fabrication of the DASI in both current experiment and traditional experiment (B)

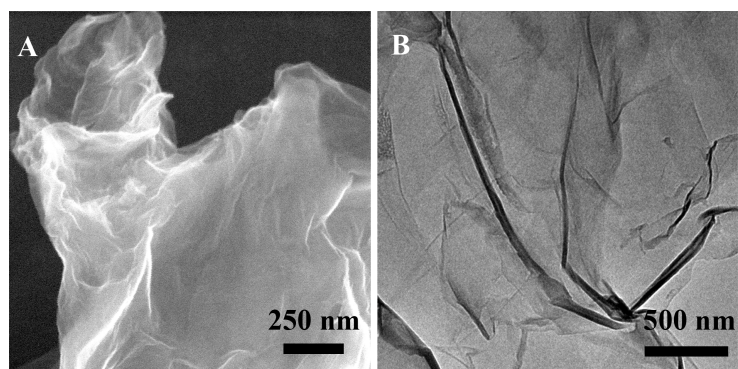


Fig. 1 SEM image (A) and TEM image (B) of N-GS

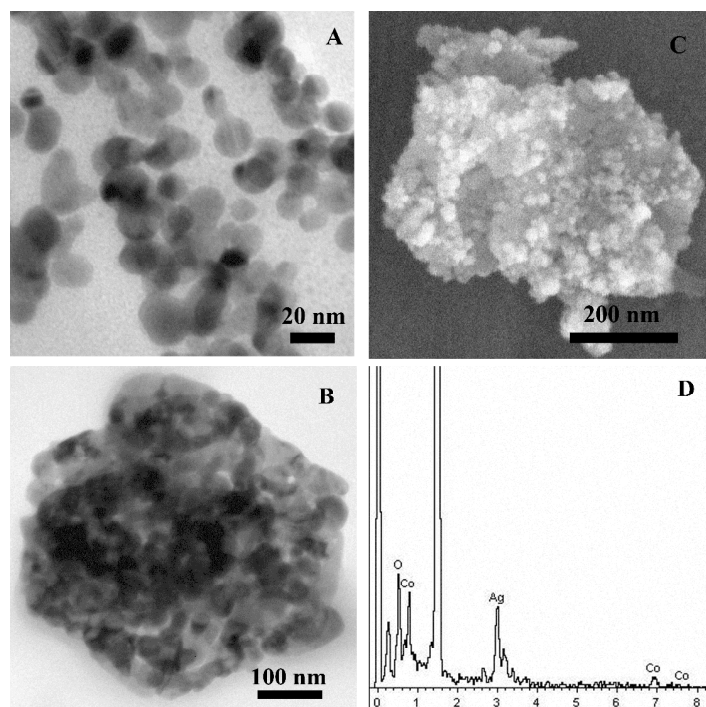


Fig. 2 TEM image (A) of Ag NPs, TEM image (D) of mesoporous Co_3O_4 nanosheet;

SEM image (E) and EDS diagram (F) of the $\text{Ag}@\text{Co}_3\text{O}_4$.

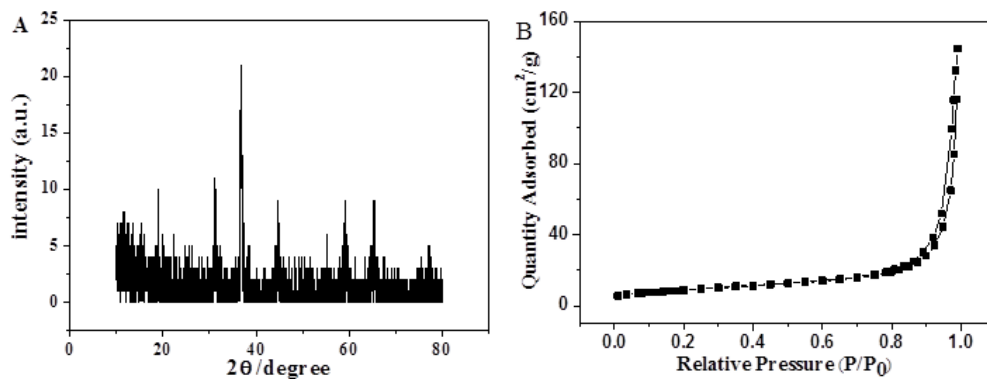


Fig. 3 XRD patterns of Co_3O_4 (A); BET analysis of Co_3O_4 (B)

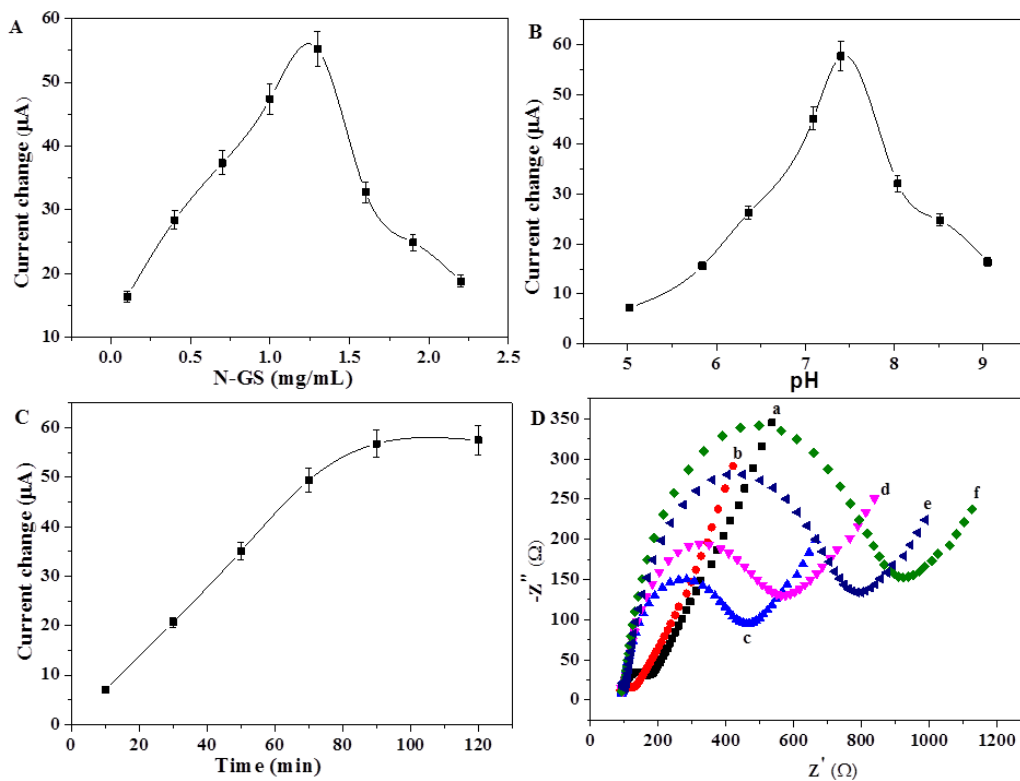
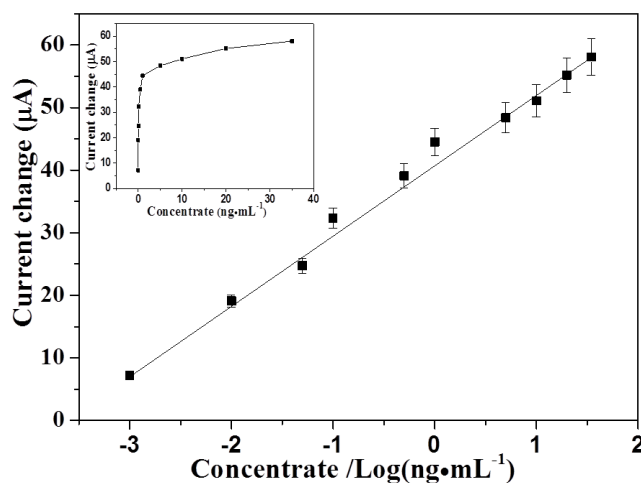


Fig. 4 A): effect of N-GS concentration on the response of the DASI (pH=7.4, BRCA1 antigen 20 ng/mL); B): effect of pH on the response of the DASI (N-GS 1.3 mg/mL , BRCA1 antigen 20 ng/mL); C): effect of incubation time on the response of

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4 the DASI (pH=7.4, N-GS 1.3 mg/mL, BRCA1 antigen 20 ng/mL); D): EIS obtained
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6 for different modified electrodes: (a) GCE, (b) GCE/HPCS@N-GS, (c)
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8 GCE/HPCS@N-GS/Ab-Ag@Co₃O₄, (d) GCE/HPCS@N-GS/Ab-Ag@Co₃O₄/BSA, (e)
9
10 GCE/HPCS@N-GS/Ab-Ag@Co₃O₄/BSA/BRCA1, (f) GCE/HPCS@N-GS/
11
12 Ab-Ag@Co₃O₄/BSA/BRCA1/ Ab-Ag@Co₃O₄.



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40 Fig. 5 Calibration curve of the prepared DASI towards different concentrations of
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42 BRCA1, error bar = RSD ($n = 5$).

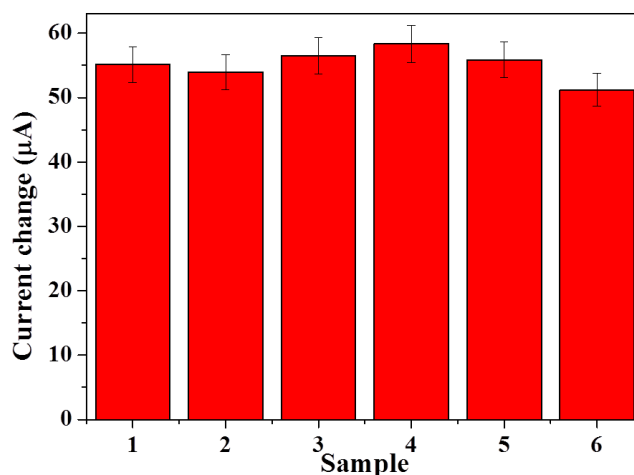


Fig. 6 The column of interference (1) 20 ng/mL BRCA1; (2) 20 ng/mL BRCA1+200 ng/mL alpha fetal protein; (3) 20 ng/mL BRCA1+200 ng/mL BSA, (4) 20 ng/mL BRCA1+200 ng/mL carcino-embryonic antigen; (5) 20 ng/mL BRCA1+200 ng/mL human immune globulin; (6) 20 ng/mL BRCA1+200 ng/mL melanoma antigen. Error bar=RSD ($n=5$).

scientific research	Linear range	Detection limit	Reference
Present research	0.001-35 ng/mL	0.33 pg/mL(10^{-15} mol/L)	
Biochip (molecular beacon detection scheme)	$2 \times 10^{-5} \sim 5 \times 10^{-6}$ mol/L	7×10^{-8} mol/L	41
monolithic silicon optocoupler array	$1 \times 10^{-9} \sim 5 \times 10^{-7}$ mol/L	9×10^{-10} mol/L	42
Enzyme-free amplified detection platform	$1 \times 10^{-6} \sim 1 \times 10^{-13}$ mol/L	1×10^{-13} mol/L	43

Table 1 Comparison of the present research and other reports

Human serum sample (ng/mL)	The addition content (ng/mL)	The detection Content (ng/mL)	RSD (% , n=5)	Recovery (%)
0.83	1	1.81 ± 0.04	1.60	101.8
0.83	5	5.86 ± 0.07	5.41	100.5
0.83	10	10.92 ± 0.08	6.95	100.9

Table 2 Results for the detection of BRCA1 in human serum sample by DASi.

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