Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/foodfunction

- 1 High hydrostatic pressure treatment provides persimmon good characteristics to
- 2 formulate milk-based beverages with enhanced functionality
- 3 M. Hernández-Carrión¹*, A. Tárrega², I. Hernando¹, S. M. Fiszman², A. Quiles¹
- ⁴ ¹Research Group of Food Microstructure and Chemistry. Department of Food
- 5 Technology. Universitat Politècnica de València. Camino de Vera s/n, 46022 Valencia,
- 6 Spain.
- ⁷ ²Instituto de Agroquímica y Tecnología de Alimentos (IATA-CSIC). Avda. Agustín
- 8 Escardino 7, 46980, Paterna, Valencia (Spain).
- 9 *Corresponding author: María Hernández Carrión
- 10 E-mail address: maherca2@upvnet.upv.es
- 11 Camino de Vera s/n, 46022 Valencia (Spain)
- 12 Tel: +34 96 387 70 00 (Ext. 78230) Fax: +34 96 387 93 60

Food & Function Accepted Manuscript

Abstract 13

High hydrostatic pressure (HHP) applied during food processing can improve the 14 retention of food quality attributes and nutritional values in comparison with 15 pasteurization. Persimmon is a good source of bioactive compounds but they are a 16 17 seasonal fruit that cannot be consumed throughout the year. The aim of this work was to compare the HHP and pasteurization treatments to formulate milk-based beverages 18 containing this carotenoid rich ingredient and to evaluate their performance in these 19 20 beverages. The carotenoid and tannin content of persimmon were determined and the 21 microstructure and rheology of the new milk-based persimmon beverages were 22 examined. The results showed that HHP treatment favoured the release of carotenoids from the fruit matrix and precipitation of the tannins. The milk-based beverages 23 24 prepared with the high-pressure persimmon presented the best rheological properties 25 because unlike the untreated and pasteurized persimmon milk-based beverages, they 26 did not form a gel-like structure or separate out. 27

28 Keywords: carotenoids, high hydrostatic pressure, microstructure, persimmon, 29 rheological properties, tannins.

30

Introduction 31

32 Persimmon (*Diospyros kaki* L. f.) is an important and widespread fruit crop in China. In Europe, Spain is one of the major producers reaching in Valencia more than 130 000 33 tonnes in 2012¹. Persimmons are among the fruits with the highest levels of bioactive 34 antioxidant compounds², such as carotenoids and polyphenols. The main polyphenols 35 in persimmons are tannins, which have degenerative disease prevention effects ³, but 36 37 when they are in their soluble form they can impart astringency to the persimmon fruits ⁴. Carotenoids such as lycopene, lutein, and zeaxanthin have considerable antioxidant 38 potential ⁵⁻⁶. Another important role of carotenes, especially β -carotene, is provitamin A 39

2

activity ⁷⁻⁸. A major variety of persimmon grown in Spain is 'Rojo Brillante', an 40 41 astringent cultivar which requires postharvest deastringency treatment before the fruit 42 can be marketed such as exposure to carbon dioxide in high concentrations, appropriate ethylene treatment or drying after peeling ⁹. Exposure to high levels of 43 carbon dioxide (950 g kg⁻¹ for 24 h at 20 °C) has proven to be the most effective way to 44 remove astringency while maintaining fruit firmness ¹⁰. 45 46 Persimmons are normally sold in fresh form but are a seasonal fruit that cannot be 47 consumed throughout the year. Therefore, it would be useful to develop industrial processes that prolong their shelf life and that make it possible to produce persimmon 48 derivatives for fruit juice mixtures, jams, yoghurts or ice creams from astringent 49 50 varieties, in order to obtain products with high nutritional value. Until now, thermal 51 processing has been the method most commonly employed because of its ability to inactivate microorganisms and spoilage enzymes (polyphenoloxidase, 52 53 pectinmethylesterase, etc.). However, heat may induce chemical and physical changes 54 that damage organoleptic properties and may reduce the content or bioavailability of some bioactive compounds, particularly if applied under severe conditions ¹¹⁻¹³. 55 According to Rawson et al.¹⁴, most authors have reported that pasteurization leads to 56 57 a decrease in the bioactive content (carotenoid content, phenolic content and antioxidant activity) of some fruits and vegetables such as exotic fruits ¹⁵, mangos ¹⁶⁻¹⁸ 58 and mulberry fruit extract, durian juice, pineapple juice, and cashew apple juice ¹⁹⁻²². 59 60 High hydrostatic pressure (HHP) processing would appear to be a suitable alternative to thermal processing as it is considered one of the most economically viable of the 61 62 non-thermal technologies and makes it possible to obtain products with high nutritional and quality parameters compared to conventional thermal processing ^{14, 23}. HHP 63 64 treatment is expected to be less harmful to low molecular weight food compounds such as flavouring agents, pigments and vitamins and preserves the nutritional value of 65 treated food better than heat due to its limited effects on covalent bonds ^{14, 24}. HHP 66

3

treatment at ambient temperature is reported to have minimal effect on the bioactive
 content of various fruits and vegetables ²⁴.

69 Both HHP treatment and pasteurization can cause changes in the structure of the plant 70 tissue which could be related to modifications in the bioavailability of some bioactive compounds. So, the microstructural study of food products could clarify the relationship 71 72 between structure and functionality. In this sense, confocal laser scanning microscopy 73 (CLSM) could be a suitable technique to study the interactions among components 74 using specific dyes and their fluorescent excitation-emission features. CLSM has been 75 employed to study the microstructure of some dairy products such as the interaction between carrageenan and milk proteins ²⁵, the distribution of fat and protein in different 76 dairy products ²⁶, the structure dairy products with different compositions ²⁷, or the 77 effect of whey protein addition on the structural properties of stirred yoghurt systems at 78 different protein and fat content ²⁸. 79

The aim of this work was to study the effect of HHP and pasteurization processes on some bioactive compounds of the persimmon, as carotenoids and tannins, in order to formulate milk-based beverages with improved nutritional and functional properties.

The microstructure and rheological behaviour of these milk-based beverages were alsoexamined.

85

86 Materials and methods

87 Materials and milk based beverages preparation

Persimmon fruits (cv. Rojo Brillante) were harvested in Carlet (Valencia, Spain) at the beginning of November 2012. The maturity index was selected following the method of Salvador et al. ²⁹, which defines six maturity stages based on the external colour ranging from I (yellow green) to VI (orange red); fruits on their commercial maturity stage (stage IV) were used in the present work. Cubes (15 mm) were taken from the equatorial area of the fruits and heat-sealed in 110 x 220 mm plastic bags (Doypack type, Amcor, Spain). Each bag contained approximately 80 g of persimmon. One-third

119

Food & Function

95 of the bags were not subjected to any treatment (untreated persimmon). Another third 96 of the bags (HHP-treated persimmon) was placed inside a hydrostatic pressure unit 97 with a 2350 mL capacity (GEC Alsthom ACB 900 HP, type ACIP 665, Nantes, France) 98 using water as the pressure medium; the pressure employed was 200 MPa for 6 min at 25 °C (energy consumption 244 kJ kg⁻¹), based on previous studies where HHP 99 treatment at 200 MPa was applied for 1, 3, and 6 min and the treatment at 200 MPa for 100 6 min gave the highest level of carotenoid compound extraction ³⁰. The last third of the 101 bags was pasteurized in a water bath at 70 °C for 15 min with an energy consumption 102 of 3600 kJ kg⁻¹ (pasteurized persimmon). Each type of persimmon was homogenised 103 during 90 s and freeze-dried during 120 h at -45 °C and 1.3.10⁻³ mPa in a freeze-drier 104 (Lioalfa-6®, Telstar, Terrassa, Spain) before their use in the milk based beverages. 105 106 Nine different milk-based persimmon beverages were studied varying the type of milk 107 and the treatment of the persimmon. Three different types of milk were used: whole milk (36 g L⁻¹ fat content), semi-skimmed milk (15.5 g L⁻¹ fat content), and skimmed 108 milk (2.5 g L⁻¹ fat content) from Central Lechera Asturiana, Siero, Spain. Three different 109 110 types of freeze-dried persimmon were employed: untreated, HHP-treated and 111 pasteurized persimmon. The quantity of persimmon used in the formulation of 1 L of 112 milk-based beverage was calculated to have the same carotenoid content to that in 200 113 g of fresh persimmon (0.743 mg β -carotene/100 g fresh weight). The milk-based beverages were prepared by placing the corresponding amount of 114 freeze-dried untreated persimmon (115.9 g kg⁻¹), freeze-dried HHP-treated persimmon 115 (52.8 g kg^{-1}) or freeze-dried pasteurized persimmon (95.1 g kg $^{-1}$) in a food processor 116 117 (Thermomix TM31, Wuppertal, Germany) and stirring at increasing agitation speeds 118 (1100 rpm, 3250 rpm and 10200 rpm), for 10 s at each speed, in order to reduce the

following the same procedure (1100 rpm, 3250 rpm and 10200 rpm, for 10 s at each

particle size of the freeze-dried persimmon samples. The milk was added and stirred

121 speed). All the milk-based beverages were kept at 4-5 °C until their analysis. The

microstructure, rheological properties and loss of stability were analysed within 24 h ofmilk-based beverage preparation.

124

125 Extraction and quantification of carotenoids

126 The total carotenoids were determined according to the method described by Hornero-Méndez & Mínguez-Mosquera³¹ with modifications. Freeze-dried persimmon (5 g) was 127 128 extracted with 25 mL of cool acetone using a homogenizer (IKA T25 Basic Ultra-129 Turrax) and vacuum filtered until no more colour was extracted. The extract was added gradually to 50 mL ethyl ether in a decanting funnel. With each addition of extract, 130 enough NaCl solution (100 g L⁻¹) was added to separate the phases and transfer the 131 pigments to the ether phase; the aqueous phase was removed. This process was 132 133 carried out in several steps to ensure maximum elimination of the aqueous phase. The 134 ether phase was treated several times with anhydrous Na₂SO₄ to remove residual 135 water and finally evaporated to dryness in a rotary evaporator (model RII; Büchi 136 Labortechnik, Flawil, Switzerland) at a temperature below 35 °C. Finally, the pigments 137 were collected with acetone to a volume of 100 mL and the absorbance was measured 138 at 450 nm using a spectrophotometer (model Helios Zeta UV Visible; Thermo Fisher 139 Scientific Inc., Cambridge, UK). The calibration curve was constructed with different concentrations of β carotene (Sigma Aldrich, Madrid, Spain) in acetone (Panreac, 140 Barcelona, Spain). The results were expressed as mg β carotene/100 g of dry weight 141 freeze-dried persimmon. Three separate carotenoid extractions were made for each 142 143 type of persimmon treatment and for untreated persimmon and the measurements 144 were performed in triplicate.

145

146 Total soluble tannin content

The total soluble tannin content of the freeze-dried persimmons was determined with a
spectrophotometer (Helios Zeta UV Visible) using the Folin Denis colorimetric method
as described by Arnal and Del Río ³². Freeze-dried persimmon (5 g) was homogenised

with 25 mL of 800 g kg⁻¹ methanol-water blend in a homogenizer (IKA T25 Basic Ultra-150 Turrax). The homogenate was centrifuged (14500 rpm, 20 min, 4 °C) and filtered. The 151 supernatant was kept. More supernatant was extracted from the pellet with 25 mL of 152 800 g kg⁻¹ methanol and added to the first supernatant. The total supernatant was 153 brought up to 100 mL with 800 g kg⁻¹ methanol. In a test tube, 1 mL of the extract and 6 154 mL of distilled water were mixed and vortexed, then 0.5 mL of Folin Ciocalteu reagent 155 156 (Panreac, Barcelona, Spain), was added. After 3 min, 1 mL saturated Na₂CO₃ was 157 added, the mixture was vortexed and 1.5 mL distilled water was added. Absorbance was measured after 90 min at 725 nm. The calibration curve was constructed with 158 different concentrations of gallic acid (Panreac, Barcelona, Spain) in 800 g kg⁻¹ 159 160 methanol. The results were expressed as g gallic acid/100 g of dry weight. Three separate total soluble tannin extractions were made for each persimmon treatment and 161 162 for untreated persimmon and the measurements were performed in triplicate.

163

164 Confocal Laser Scanning Microscopy (CLSM)

Equipment and dyes. CLSM was selected as the most appropriate microscopy 165 technique for studying the microstructure of the freeze- dried persimmons and the milk-166 167 based beverages, due to the ability of carotenoid compounds to emit fluorescence when excited by a laser line. This makes it possible to locate them using CLSM without 168 169 staining the sample. Moreover, protein and fat can also be identified by using specific dves such as Rhodamine and Nile Red, respectively. A Nikon confocal microscope C1 170 171 unit fitted on a Nikon Eclipse E800 microscope (Nikon, Tokyo, Japan) was used. An Ar 172 laser line (488 nm) was employed as the light source to excite the Rhodamine B and Nile Red fluorescent dyes. Rhodamine B (Fluka, Sigma-Aldrich, Missouri, USA) with λ_{ex} 173 _{max} 488 nm and $\lambda_{em max}$ 580 nm was dissolved in distilled water at 2 g L⁻¹. This dye was 174 175 used to stain proteins and carbohydrates. Nile Red (Fluka, Sigma-Aldrich, Missouri, USA) with $\lambda_{ex max}$ 488 nm and $\lambda_{em max}$ 515 nm was dissolved in polyethylene glycol 176 (PEG) 200 at 0.1 g L⁻¹ and was used to stain fat. The autofluorescence of the samples 177

178 was observed using the Ar laser line without any dye. A 60x/1.40NA/Oil/ Plan Apo VC179 Nikon objective lens was used.

180

Sample viewing. A drop of freeze-dried persimmon or milk- based beverages was placed on a slide and 20 μ L of Rhodamine B solution and 20 μ L of Nile Red solution were added. The observations were made 10 min after diffusion of the dyes into the sample or beverage. The images were obtained and stored with 1024 x 1024 pixel resolution using the microscope software (EZ-C1 v.3.40, Nikon, Tokyo, Japan).

186

187 Rheological measurements

Both the flow behaviour and the viscoelastic properties of each milk-based beverage 188 189 were measured in triplicate. The measurements were carried out in a RS1 controlled 190 stress rheometer (Thermo Haake, Karlsruhe, Germany), using a parallel plate geometry of 6 cm in diameter with a 1mm gap, and monitored by the RheoWin 191 192 software package (version 2.93, Thermo Haake). A temperature of 10 ± 1°C, selected 193 as representative of the usual consumption temperature of dairy desserts; it was 194 maintained throughout the measurements by means of a Phoenix P1 Circulator device 195 (Thermo Haake). The milk-based beverages were allowed to rest on the rheometer 196 plate for 5 min before measurement and a fresh milk-based beverage was loaded for 197 each measurement.

198

Flow behaviour. The flow of milk and milk-based beverages was measured by recording the shear stress values when shearing them with a linear increasing shear rate from 0 to 200 s⁻¹ for a period of 60 s and in reverse sequence for the same time. The areas under the upstream data point curve (A_{up}) and under the downstream data point curve (A_{down}), as well as the hysteresis area ($A_{thix} = A_{up} - A_{down}$), were obtained using RheoWin Pro software (version 2.93, Thermo Haake). The data from the

205	ascending segment of the shear cycle were fitted to the Ostwald-de Waele model (Eq.
206	(1)) using RheoWin Pro software (version 2.93, Thermo Haake):
207	$\sigma = \mathbf{K} \cdot \gamma^{n} \tag{1}$
208	where K (Pa s^n) is the consistency index and n is the flow index. In addition, apparent
209	viscosity values at 10 s ⁻¹ and 100 s ⁻¹ (η_{10} and η_{100} , respectively) were also calculated as
210	follows.
211	$\eta_{app} = \mathbf{K} \cdot \gamma^{n-1} $ (2)
212	
213	Viscoelastic properties. In order to determine the linear viscoelastic region (LVR),
214	stress sweeps (0.01–100 Pa) were run at 1 Hz. Frequency sweeps were then
215	performed within the LVR over the range $f = 0.01-10$ Hz; the values of the storage
216	modulus (G') and the loss modulus (G'') as a function of frequency (mechanical
217	spectra) were obtained using RheoWin Pro software (version 2.93, Thermo Haake).
218	
219	Sedimentation
220	Sedimentation is very negative for the quality of food products and may cause
221	consumers to reject the product. The loss of stability (sedimentation) of the milk-based
222	beverages was measured by placing the milk-based beverages in 10 mL test tubes and
223	leaving them to stand until the amount of sediment remained constant. This state was
224	reached after 90 min. The % of sedimentation was calculated as (volume of
225	sediment/total volume) x 100.
226	
227	Statistical analysis
228	The effect of the persimmon treatment (untreated, HHP treatment and pasteurization)
229	on the carotenoid and total soluble tannin content was analysed by one-way ANOVA.
230	The effects of the type of milk (whole milk, semi-skimmed milk, and skimmed milk) on
231	the flow parameters (K, n, η_{10} , η_{100} , and A_{thix}) and viscoelastic parameters (G' and G") at

1Hz were analysed by one-way ANOVA for each type of milk-based beverage (prepared with freeze-dried untreated, freeze-dried HHP-treated, and freeze-dried pasteurized persimmon). The least significant difference (LSD) test with a 95% confidence interval was used to compare the mean values obtained. All the calculations were made with Statgraphics Plus 5.1 software.

237

247

238 **Results and discussion**

239 Carotenoid and total soluble tannin content of the freeze-dried persimmons.

Fig. 1A shows the carotenoid content of the different persimmons analysed in this study. The untreated and the pasteurized persimmons did not show statistically significant differences (P > 0.05) in the extracted carotenoid content. However, the HHP-treated persimmons presented a significantly higher extracted carotenoid content (P < 0.05). Similar results have been published for persimmon and orange juice, in which the authors reported increases in total carotenoids extracted after HHP processing $^{30, 33-34}$.



Fig. 1. Carotenoid content and total soluble tannin content of untreated, HHP-treated
and pasteurized freeze-dried persimmon. Different letters indicate significant
differences (P < 0.05) between the samples.

No statistically significant differences in total soluble tannin content were found between the untreated and pasteurized persimmons (P > 0.05). The HHP-treated persimmons presented significantly lower total soluble tannin content (P < 0.05). It has been proved that this treatment decreases the total soluble tannin content in

255 persimmon, as this treatment favours the precipitation of soluble tannins ³⁵. The fact 256 that HHP treatment seems to favour tannin precipitation could be related to a 257 decreased astringency of the HHP-treated fruit. So, this treatment could be used to 258 develop new, less astringent and more versatile persimmon derivatives.

259

260 Confocal laser scanning microscopy (CLSM)

261 Freeze-dried persimmon. In order to understand the structure of the freeze-dried 262 persimmon without the interference of the milk structure, samples were prepared by 263 dispersing the corresponding amount of freeze-dried persimmon in deionised water 264 and their microstructure was analysed. Fig. 2 shows the CLSM images of the freezedried persimmon dispersed in water. All the persimmon samples showed significant 265 autofluorescence (Figs. 2A, 2E, and 2I) due to the presence of carotenoids. According 266 to Vázquez-Gutiérrez et al.³⁶, carotenoids are associated mainly with cell walls, 267 forming spherical bodies (chromoplasts). In the untreated samples (Fig. 2A) most of the 268 269 carotenoids tended to aggregate into interconnected clusters, forming a network. In the 270 HHP-treated and pasteurized samples (Fig. 2E and 2I) the carotenoid aggregates were 271 smaller than in the untreated ones, probably because the carotenoids were more 272 dispersed throughout the plant tissue. HHP treatments are known to induce morphological changes in plant cells which result in the rupture of cell walls ³⁶⁻³⁸. These 273 structural modifications could cause leaching of cellular constituents into the food 274 275 matrix, and so the spread of carotenoids throughout the tissue. Other authors have 276 established that thermal processing can affect functionalities such as carotenoid 277 bioaccessibility due to its effect on the barrier properties of the cell wall polysaccharide network ³⁹. When the samples were stained with Nile Red, a specific dye for fat, (Figs. 278 279 2B, 2F and 2J) a green network was observed, consisting of unstructured fat from the plant persimmon tissues and carotenoid compounds, as these pigments were excited 280 by the same wavelength as Nile Red. This network was denser and more widely-281 dispersed in the HHP and pasteurized samples (Figs. 2F and 2J) than in the untreated 282

283 ones (Fig. 2B); this fact could be related to the structural modifications occurred on the treated tissue (HHP and pasteurized) in comparison with the untreated one. Staining 284 285 with Rhodamine dye (Figs. 2C, 2G and 2K) made it possible to view the plant 286 persimmon structures in the samples because Rhodamine allows visualizing proteins 287 and carbohydrates as those present in the cell wall. When Rhodamine and Nile Red (Figs. 2D, 2H and 2L) were used to stain protein and fat respectively, the samples 288 289 showed a protein network (red) in which carotenoid pigments (green) were dispersed. 290 The HHP-treated samples seemed to present the highest carotenoid content (Fig. 2H), 291 as reported above (Fig. 1A).



292 293

Confocal Laser Scanning Microscopy (CLSM). Samples prepared with Fig. 2. 294 untreated, HHP-treated and pasteurized freeze-dried persimmon dispersed in water. 295 Green: carotenoids and fat globules. Red: proteins and carbohydrates. FG: Fat 296 globules. Magnification 60x.

297 Milk. Fig. 3 shows the CLSM images of the three different types of milk (whole, semi-

skimmed and skimmed) employed for formulating the milk-based persimmon 298

299 beverages. The fat globules did not display intrinsic fluorescence, despite β -carotene being normally present in their composition (Figs. 3A, 3E and 3I). To study these 300 different milks by fluorescence, an extrinsic fluorescent dye was therefore required 40. 301 302 Figs. 3B, 3F and 3J show the fat globules coloured green due to Nile Red staining. In 303 Figs. 3C, 3G and 3K, the proteins can be seen as a continuous phase stained red by 304 Rhodamine. Fig. 3D shows how the protein and lipid phases interacted, forming a 305 homogenous network in whole milk. In semi-skimmed milk (Fig. 3H) this mixture was 306 not as homogenous as in whole milk, as in these milks the protein acted as a 307 continuous phase and the fat globules as a dispersed phase. In skimmed milk (Fig. 3L) 308 the distinction between the two phases was more evident.



309

Fig. 3. Confocal Laser Scanning Microscopy (CLSM). Different milk matrixes (whole, 310 311 semi-skimmed, and skimmed milk) used in the formulation of the milk-based 312 persimmon beverages. Green: carotenoids and fat globules. Red: proteins and 313 carbohydrates. Magnification 60x.

314

315 Milk-based persimmon beverage. Fig. 4 shows the CLSM images of the untreated persimmon milk-based beverages prepared with milk with different fat contents. All the 316 317 beverages studied presented considerable intrinsic autofluorescence (Figs. 4A, 4E and 4I) due to the presence of carotenoids, which appeared as groups of spherical bodies. 318 319 These bodies appeared to be linked up and arranged in a pattern. The milk-based beverages stained with Nile Red (Figs. 4B, 4F and 4J) showed a green network made 320 321 up of fat globules and unstructured fat from the milk and unstructured fat and 322 carotenoid compounds from the persimmon. Consequently, the fat from the milk was 323 found to form two networks: one made up of globules connected together forming 324 bright green clusters and another made up of unstructured fat and carotenoids 325 coloured dark green.



Fig. 4. Confocal Laser Scanning Microscopy (CLSM). Milk-based beverages prepared
with freeze-dried untreated persimmon dispersed in whole, semi-skimmed and
skimmed milk. Green: carotenoids and fat globules. Red: proteins and carbohydrates.
FG: Fat globules, PT: Plant tissue. Magnification 60x.

331 In the milk-based beverages stained with Rhodamine (Figs. 4C, 4G and 4K), a red protein network was observed. This distribution was similar in all the milk-based 332 333 persimmon beverages studied. This protein network, in which part of the plant tissue is 334 fused, seemed to behave as a continuous phase that kept some fragments of plant 335 tissue dispersed throughout. In the micrographs of the milk-based persimmon 336 beverages with a high fat content stained with Nile Red and Rhodamine (Figs. 4D and 337 4H), two different phases could be observed: a continuous phase consisting of a dense 338 lipoprotein network stained a yellow-green colour and a dispersed phase formed by 339 green groups, probably carotenoids and fragments of plant tissue, with red contours. 340 When skimmed milk was used in the formulations (Fig. 4L) the continuous phase was 341 coloured orange and was mainly composed of protein, while the dispersed phase 342 consisted of plant material, with red contours.

343

344 Effect of the persimmon treatment. The milk-based persimmon beverages prepared 345 with freeze-dried HHP-treated persimmon (Fig. 5) differed from the milk-based 346 beverages prepared with freeze-dried untreated persimmon (Fig. 4). The former 347 presented higher autofluorescence (Figs. 5A, 5E and 5I) than the latter despite 348 containing a lower quantity of persimmon in their formulation. This high 349 autofluorescence made it possible to see fragments of persimmon, probably due to the 350 dispersion of carotenoids throughout the tissue. Consequently, HHP treatment could 351 encourage the release and extraction of carotenoids. Nile Red staining (Figs. 5B, 5F 352 and 5J) showed a network made up of fat from the milk matrix and fat and carotenoids 353 from the plant material. This network formed a continuous film in the HHP-treated milk-354 based beverages and clusters in the untreated ones which could be related, again, to 355 higher carotenoid extractability obtained in the HHP-treated persimmons compared to the untreated ones. When Rhodamine (Figs. 5C, 5G and 5K) was used to stain the 356 357 milk-based beverages a continuous and homogeneous red-coloured protein phase was 358 observed, with red- and black-coloured fragments of plant tissue dispersed throughout

15

359 the milk matrix. Milk-based beverages with a high fat content stained with Nile Red and Rhodamine (Figs. 5D and 5H) showed two different phases, a continuous phase 360 361 consisting of a dense lipoprotein network coloured yellow-green and a dispersed 362 green-stained phase formed by carotenoids and fat with dark fragments of plant tissue. 363 When skimmed milk was used in the formulation of the milk-based beverages (Fig. 5L) 364 the continuous phase was composed basically of protein. The carotenoid compounds 365 were better identified in these milk-based beverages (Figs. 5D, 5H and 5L) than in 366 those made with freeze-dried untreated persimmon (Figs. 4D, 4H and 4L).



367 368

Fig. 5. Confocal Laser Scanning Microscopy (CLSM). Milk-based beverages prepared 369 with freeze-dried HHP-treated persimmon dispersed in whole, semi-skimmed and 370 skimmed milk. Green: carotenoids and fat globules. Red: proteins and carbohydrates. 371 FG: Fat globules, PT: Plant tissue. Magnification 60x.

372 When the milk-based beverages were prepared with freeze-dried pasteurized

persimmon (Fig. 6) considerable autofluorescence was also observed (Figs. 6A, 6E 373

and 6l), although less than in the milk-based beverages prepared with freeze-dried 374

HHP-treated persimmon (Figs. 5A, 5E and 5I). It should be noted that the formulation
of these milk-based beverages contained less persimmon than in those made with
freeze-dried untreated persimmon, but more than when prepared with the freeze-dried
HHP-treated persimmon, although containing the same carotenoid content. The milkbased beverages stained with Nile Red (Figs. 6B, 6F and 6J) showed a less dense
network of fat and carotenoids than those with the freeze-dried HHP-treated

381 persimmon.



382

Fig. 6. Confocal Laser Scanning Microscopy (CLSM). Milk-based beverages prepared
 with freeze-dried pasteurized persimmon dispersed in whole, semi-skimmed and
 skimmed milk. Green: carotenoids and fat globules. Red: proteins and carbohydrates.
 FG: Fat globules, PT: Plant tissue. Magnification 60x.

387 When Rhodamine was used to stain the milk-based beverages (Figs. 6C, 6G and 6K),

red- and black-coloured fragments of plant tissue were observed as a dispersed phase.

389 When the milk-based beverages were stained with Nile Red and Rhodamine (Figs. 6D,

390 6H and 6L), the carotenoids appeared as a green dispersed phase. They were

- 391 observed more clearly than in the beverages with freeze-dried untreated persimmon
- 392 (Figs. 4D, 4H and 4L) but less so than in those made with the freeze-dried HHP-treated
- 393 persimmon (Figs. 5D, 5H and 5L).
- 394

395 Rheological behaviour of milk-based persimmon beverages

- 396 Flow behaviour. Fig. 7 shows the flow curves for the different milk-based persimmon
- 397 beverages analysed in this study. The milk-based beverages prepared with the
- 398 different types of persimmon presented different flow behaviour.



399 400

Fig. 7. Flow behaviour of milk-based persimmon beverages prepared with: freeze-dried untreated persimmon, freeze-dried HHP-treated persimmon, and freeze-dried pasteurized persimmon. Whole milk ($\langle \rangle$), semi-skimmed milk (\blacktriangle) and skimmed milk (\bigcirc). Lines represent the curves fits to Ostwald models.

Those made with freeze-dried untreated persimmon were semisolid systems which 404 could be attributed to the gelation of pectins in the presence of divalent ions like Ca⁺² 405 406 from the persimmon fruit and the milk. The gelation mechanism of persimmon has 407 been studied previously in acidified and natural persimmon purée with and without the addition of ethylenediaminetetraacetic acid disodium salt 2-hydrate (EDTA)⁴. Table 1 408 and 2 show the flow parameters of milk without persimmon and milk-based persimmon 409 410 beverages, respectively. The flow curves for the milk-based beverages made with freeze-dried untreated persimmon, regardless of the type of milk used, showed 411 412 pseudoplastic, time-dependent flow behaviour that was characterised by determining Ostwald-de Waele parameters and the thixotropic area (Table 2). The milk-based 413 beverages prepared with freeze-dried untreated persimmon presented higher 414 415 consistency, pseudoplasticity and thixotropy than milk without persimmon (Table 1). No 416 statistically significant differences (P > 0.05) were observed in the consistency index (K), flow index (n), apparent viscosity at 10 s⁻¹ (η_{10}), or thixotropic area (A_{thix}) when 417 different types of milk were used (Table 2). It would therefore seem to be the addition 418 419 of persimmon rather than the fat content of the milk that defines the flow behaviour of the milk-based beverages. However, the apparent viscosity at 100 s⁻¹ (η_{100}) decreased 420 significantly when skimmed milk was used (P < 0.05). 421

Table 1. Ostwald de Waele fit of milks used to formulate the milk-based persimmon beverages (R²>0.9979)

()			
Milk type	K (Pa⋅s ⁿ)	n	η ₁₀ (Pa⋅s)
Whole milk	0.0079	0.8254	0.0053
	(0.0002)	(0.0005)	(0.0001)
Semi-skimmed milk	0.0066	0.8459	0.0046
	(0.0010)	(0.0260)	(0.0004)
Skimmed milk	0.0046	0.9015	0.0037
	(0.0004)	(0.0191)	(0.0001)
Values between parentheses are the	standard doviations		

Values between parentheses are the standard deviation

122	Table 2. Ostv	vald de Waele fit of milk	-based persimme	on beverages (0.	915 < R ² < 0.9957	())	
123		Milk type	K (Pa⋅s ⁿ)	n	η ₁₀ (Pa⋅s)	η ₁₀₀ (Pa⋅s)	A _{thix} (Pa·s ⁻¹)
121	Untreated	Whole milk	28.275 ^a	0.191 ^a	4.313 ^a	0.664 ^a	3860.500 ^a
27			(8.719)	(0.058)	(0.770)	(0.030)	(962.372)
25	Untreated	Semi-skimmed milk	31.630 ^a	0.146 ^a	4.423 ^a	0.619 ^ª	4258.500 ^a
25			(3.847)	(0.001)	(0.544)	(0.077)	(656.902)
	Untreated	Skimmed milk	20.710 ^a	0.125 ^a	2.764 ^a	0.369 [°]	2597.000 ^a
26			(2.093)	(0.013)	(0.362)	(0.059)	(576.999)
	HHP	Whole milk	1.258 ^a	0.253 ^a	0.225 ^a	0.040 ^a	148.850 ^a
27			(0.177)	(0.007)	(0.028)	(0.004)	(2.333)
	HHP	Semi-skimmed milk	0.730 ^b	0.353 ^b	0.164 ^a	0.037 ^a	139.700 ^a
28			(0.139)	(0.029)	(0.020)	(0.002)	(26.022)
20	HHP	Skimmed milk	0.225 ^c	0.458 ^c	0.065 ⁶	0.019 ⁶	34.055 ⁶
20			(0.011)	(0.012)	(0.005)	(0.002)	(21.970)
29	Pasteurized	Whole milk	0.050 ^á	0.813 ^á	0.032 ^á	0.021 ^a	9.639 ^a
			(0.011)	(0.023)	(0.005)	(0.002)	(4.145)
30	Pasteurized	Semi-skimmed milk	0.090 ^á	0.670 ^á	0.046 ^á	0.022 ^á	27.675 ^{°a}
			(0.001)	(0.003)	(0.001)	(0.000)	(2.001)
31	Pasteurized	Skimmed milk	0.065 ^á	0.714 ^á	0.032 ^á	0.016 ^á	17.620 ^{°a}
			(0.032)	(0.085)	(0.010)	(0.002)	(7.750)
.32	Values between pa	arentheses are the standard devia	tions.	· · · · /	//	/	\/

Table 2 Ostwald de Waele fit of milk-based persimmon beverages (0.915 $< R^2 < 0.9957$)

433

All values shown are averages of three measurements. For each milk-based beverage, values within a column with different letters are significantly different

434 The HHP-treated persimmon milk-based beverages (Fig. 7B) were liquid systems that also showed pseudoplastic and thixotropic behaviour. As in the case of freeze-dried 435 436 untreated persimmon, when compared the milks without persimmon (Table 1) with 437 those of the milk-based beverage elaborated with freeze-dried HHP-treated 438 persimmon, an increase in the consistency, pseudoplasticity and thixotropy could be 439 observed (Table 2). However, these effects were much smaller than those observed 440 with the freeze-dried untreated persimmon, indicating that gelation did not occur or was 441 less intense with the persimmon obtained by HHP treatment. So the freeze-dried HHP-442 treated persimmon was more appropriate for formulating this kind of product since a 443 gel-like texture would be not adequate for beverages. The ANOVA results showed that 444 with the HPP-treated persimmon the flow parameter values varied significantly 445 depending on the type of milk. As expected, as the fat content of the milk decreased, 446 the consistency index decreased significantly (P < 0.05) and the milk-based beverages formulated with skimmed milk presented significantly the lowest values (P < 0.05). The 447 448 opposite occurred with the flow index: as the fat content decreased, the flow index increased significantly (P < 0.05), which corresponded to a more Newtonian flow. No 449 significant differences (P > 0.05) in apparent viscosity at 10 s⁻¹ and 100 s⁻¹, and 450 451 thixotropic area were found between the HHP-treated milk-based beverages made with 452 whole and semi-skimmed milk, but the milk-based beverages formulated with skimmed 453 milk presented significantly the lowest values (P < 0.05). It therefore seems that the 454 flow behaviour of these milk-based beverages is governed by both the addition of persimmon and the fat content of the milk used in the formulation. Freeze-dried 455 456 untreated persimmon has a gelling ability very high; so the presence of fat globules in 457 the system structure does not significantly affect the response to deformation. In contrast, in the case of freeze-dried HHP-treated persimmon, which has less gelling 458 ability, the presence of fat or not in the structure, does influence its response to 459 460 deformation.

461 When freeze-dried pasteurized persimmon (Fig. 7C) was used in the formulation of the milk-based beverages, non-homogeneous dispersion of persimmon particles in a liquid 462 463 matrix were obtained. This fact could probably correspond to the persimmon already 464 being gellified and particles of persimmon gel remaining undissolved. As commented 465 above, gelation of the persimmon could have taken place during pasteurization: the gel formation of pectins caused by heating process has been observed previously by other 466 authors in olives ⁴¹ and persimmons ⁴. A noticeable sedimentation was observed in 467 468 these milk-based beverages $(43 \pm 1\%)$ for whole milk; $45 \pm 4\%$ for semi-skimmed milk, 469 and $42 \pm 3\%$ for skimmed milk). This sedimentation could be responsible for the lower consistency index (K) and apparent viscosity at 10 s⁻¹ and 100 s⁻¹ obtained for these 470 milk-based beverages compared to the values obtained when freeze-dried untreated 471 and HHP-treated persimmon were used to prepare the milk-based beverages (Table 472 473 2). Before flow measurement, the milk-based beverages that showed sedimentation 474 were stirred manually to favour homogenisation. In these milk-based beverages, 475 Newtonian-like flow was observed, with more linear flow curves, without thixotropy (Fig. 476 7C) and flow index (n) values closer to 1 (Table 2) compared to the milk-based beverages prepared with freeze-dried untreated and HHP-treated persimmon. No 477 478 statistically significant differences (P > 0.05) were observed (Table 2) in any of the flow 479 parameters studied when different types of milk were used. The heterogeneous final 480 product, with sedimented particles, could explain the fact that the type of milk used did 481 not affect the rheological behaviour of the milk-based beverages. The flow response of 482 the milk-based beverages made with freeze-dried pasteurized persimmon therefore seems to be mainly governed by the gelified persimmon particles. 483

Viscoelastic properties. Fig. 8 shows the storage modulus (G') and loss modulus (G'')
values as a function of frequency. The mechanical spectra of the untreated persimmon
milk-based beverages showed a response typical of weak gels, with G' higher than G''
(Fig. 8A). Although small differences in rheological parameters were noticed when

- using different types of milk (Table 3), these differences were not significant (P > 0.05). 488
- As with flow behaviour, it was observed that the fat content did not affect the 489
- 490 viscoelastic properties of the milk-based beverages.





Table 3. Storage modulus (G') and loss modulus (G'') values at 1 Hz for milk-based

empty symbols, whole milk, $\langle \rangle$; semi-skimmed milk, \triangle ; skimmed milk, \bigcirc . 493

495	persimmon beverages						
496		Milk type	G' (Pa)	G"(Pa)			
	Untreated	Whole milk	138.300 ^a	30.950 ^a			
497			(31.961)	(8.910)			
	Untreated	Semi-skimmed milk	113.095 ^{°a}	26.420 ^a			
498			(22.210)	(2.291)			
	Untreated	Skimmed milk	62.435 ^a	14.220 ^a			
499			(4.250)	(0.325)			
135	HHP	Whole milk	23.230 ^a	4.360 ^a			
500			(1.428)	(0.074)			
300	HHP	Semi-skimmed milk	17.080 ^b	3.344 ^b			
504			(0.792)	(0.227)			
501	HHP	Skimmed milk	16.315 [⊳]	2.766 ^b			
			(0.884)	(0.346)			
502	Values between parent	heses are the standard deviations.					

All values shown are averages of three measurements. For each milk-based persimmon beverage, values within a column with different letters are significantly different.

504

503

494

505 The mechanical spectra of the HHP-treated milk-based beverages (Fig. 8B) also 506 showed G' values higher than G" ones, but both of them were much lower than those obtained with freeze-dried untreated persimmon and G" showed higher frequency 507 508 dependence, indicating a much weaker structure and therefore confirming the lower 509 gelling ability of these milk-based beverages. No statistically significant differences (P > 0.05) in either G' or G' at 1 Hz were found between the milk-based beverages made 510

with semi-skimmed and skimmed milk (Table 3). However, the milk-based beverages
formulated with whole milk presented significantly the highest G' and G" values (P <
0.05).

514 Due to the heterogeneity and low viscosity of the milk-based beverages made with 515 freeze-dried pasteurized persimmon, the linear viscoelastic region could be not found 516 and viscoelastic properties for these milk-based beverages could not be determined. 517

518 Conclusions

519 HHP treatment would encourage the release of carotenoids from the plant material 520 matrix and hence increase their extractability as could be seen via confocal microscopy 521 and when quantifying the carotenoid content. In addition, HHP treatment seems to 522 favour tannin precipitation and could therefore decrease the astringency of the fruit. 523 Freeze-drying processing demonstrate to be useful to extend the shelf life of 524 persimmon and to obtain derivatives that could be incorporated in formulations with 525 new functional features. HHP treatment provides persimmon to formulate milk-based 526 beverages with high carotenoid content using smaller quantities of freeze-dried persimmon. These beverages possess suitable rheological properties because they do 527 528 not form a gel-like structure, unlike the milk-based beverages with freeze-dried 529 untreated persimmon, and do not present sedimentation, unlike the milk-based 530 beverages formulated with freeze-dried pasteurized persimmon. Further research 531 should be conducted to evaluate the sensory properties and consumers' liking of these 532 milk-based persimmon beverages prior to their commercial release.

533 Acknowledgments

The authors wish to acknowledge the Spanish Ministry of Science and Innovation for the financial support (project AGL2011-30064-C02) and to the Universitat Politècnica de València (UPV) for the FPI grant given to María Hernández Carrión. The authors also thank to the 'Agrupación Nacional de Exportación de Cooperativas Citrícolas' Conselleria de Presidencia y Agricultura, Pesca, Alimentación y Agua, Boletín de

538 (ANECOOP) for the supply of the samples. Moreover, the authors wish to thank Mary

539 Georgina Hardinge for assistance with the English manuscript.

540 **References**

1.

541

542 información agraria (num. 190), Valencia, 2013. 543 2. S. T. Jung, Y. S. Park, Z. Zachwieja, M. Folta, H. Barton, J. Piotrowicz, E. Katrich, S. 544 Trakhtenberg and S. Gorinstein, Int. J. Food Sci. Nutr., 2005, 56, 105-113. 545 3. Y. Achiwa, H. Hibasami, H. Katsuzaki, K. Imai and T. Komiya, Biosci., Biotechnol., Biochem., 1997, 61, 1099-1101. 546 547 A. Tárrega, M. Carmen Gurrea, J. Navarro and J. Carbonell, Food Bioprocess Technol., 4. 548 2013, 6, 2399-2405. 549 5. W. Stahl and H. Sies, Biochimica et Biophysica Acta (BBA) - Molecular Basis of Disease, 550 2005, 1740, 101-107. 551 6. N. J. I. E. Shami and E. A. M. Moreira, *Revista de Nutrição*, 2004, 17, 227-236. 552 7. D. B. Rodriguez-Amaya, J. Micronutr. Anal., 1989, 5, 191-225. 553 N. J. Miller, J. Sampson, L. P. Candeias, P. M. Bramley and C. A. Rice-Evans, FEBS 8. 554 Letters, 1996, 384, 240-242. 555 9. X. Wei, F. Liu, Z. Qiu, Y. Shao and Y. He, Food Bioprocess Technol. (in press), 2013, 1-10. 556 10. L. Arnal and M. A. Del Río, J. Food Sci., 2003, 68, 1516-1518. 557 11. A. Patras, N. Brunton, S. Da Pieve, F. Butler and G. Downey, Innov. Food Sci. Emerg. 558 Technol., 2009, 10, 16-22. 559 12. A. Patras, N. P. Brunton, C. O'Donnell and B. K. Tiwari, Trends Food Sci. Technol., 2010, 560 **21**, 3-11. 561 13. A. Rawson, A. Koidis, D. K. Rai, M. Tuohy and N. Brunton, J. Agric. Food Chem., 2010, 562 **58**, 7740-7747. 563 14. A. Rawson, A. Patras, B. K. Tiwari, F. Noci, T. Koutchma and N. Brunton, Food Res. Int., 564 2011, 44, 1875-1887. 565 15. P. Elez-Martínez, I. Aguiló-Aguayo and O. Martín-Belloso, J. Sci. Food Agric., 2006, 86, 566 71-81. 567 T. Djioua, F. Charles, F. Lopez-Lauri, H. Filgueiras, A. Coudret, M. F. Jr, M.-N. Ducamp-16. 568 Collin and H. Sallanon, Postharvest Biol. Technol., 2009, 52, 221-226. 569 17. Y. Kim, A. J. Lounds-Singleton and S. T. Talcott, Food Chem., 2009, 115, 989-993. 570 18. A. L. Vásquez-Caicedo, S. Schilling, R. Carle and S. Neidhart, Food Chem., 2007, 102, 571 1172-1186. 572 19. P. Aramwit, N. Bang and T. Srichana, Food Res. Int., 2010, 43, 1093-1097. 573 20. S. T. Chin, S. A. Hamid Nazimah, S. Y. Quek, Y. B. Che Man, R. A. Rahman and D. M. 574 Hashim, LWT - Food Sci. Technol., 2010, 43, 856-861. 575 21. M. Rattanathanalerk, N. Chiewchan and W. Srichumpoung, J. Food Eng., 2005, 66, 259-576 265. 577 22. L. Q. Zepka and A. Z. Mercadante, Food Chem., 2009, 117, 28-34. 578 23. T. Norton and D. W. Sun, Food Bioprocess Technol., 2008, 1, 2-34. 579 24. I. Oey, I. Van der Plancken, A. Van Loey and M. Hendrickx, Trends Food Sci. Technol., 580 2008, 19, 300-308. 581 25. D. Arltoft, R. Ipsen, F. Madsen and J. de Vries, *Biomacromolecules*, 2007, **8**, 729-736. 582 26. M. A. Auty, M. Twomey, T. P. Guinee and D. M. Mulvihill, J. Dairy Res., 2001, 68, 417-583 427. 584 27. M. Panouillé, A. Saint-Eve, C. de Loubens, I. Déléris and I. Souchon, Food Hydrocolloids, 585 2011, **25**, 716-723.

586	28.	A. Krzeminski, K. Großhable and J. Hinrichs, LWT - Food Sci. Technol., 2011, 44, 2134-
587		2140.
588	29.	A. Salvador, L. Arnal, C. Besada, V. Larrea, A. Quiles and I. Pérez-Munuera, Postharvest
589		Biol. Technol., 2007, 46 , 181-188.
590	30.	L. Plaza, C. Colina, B. De Ancos, C. Sánchez-Moreno and M. P. Cano, Food Chem., 2012,
591		130 , 591-597.
592	31.	D. Hornero-Méndez and M. I. Mínguez Mosquera, Journal of Agricultural and Food
593		Chemistry, 2001, 49 , 3584-3588.
594	32.	L. Arnal and M. A. Del Río, <i>Span. J. Agric. Res.</i> , 2004, 2 , 243-247.
595	33.	B. De Ancos, E. Gonzalez and M. P. Cano, J. Agric. Food Chem., 2000, 48, 3542-3548.
596	34.	C. Sanchez-Moreno, L. Plaza, B. De Ancos and M. P. Cano, J. Agric. Food Chem., 2003,
597		51 , 647-653.
598	35.	J. Vázquez-Gutiérrez, I. Hernando and A. Quiles, Eur. Food Res. Technol., 2013, 237, 9-
599		17.
600	36.	J. L. Vázquez-Gutiérrez, A. Quiles, I. Hernando and I. Pérez-Munuera, Postharvest Biol.
601		Technol., 2011, 61 , 137-144.
602	37.	J. L. Vázquez-Gutiérrez, M. Hernández-Carrión, A. Quiles, I. Hernando and I. Pérez-
603		Munuera, Food Res. Int., 2012, 47, 218-222.
604	38.	J. L. Vázquez-Gutiérrez, L. Plaza, I. Hernando, C. Sanchez-Moreno, A. Quiles, B. de
605		Ancos and M. P. Cano, <i>Food Funct.</i> , 2013, 4 , 586-591.
606	39.	A. Ribas-Agustí, S. Van Buggenhout, P. Palmero, M. Hendrickx and A. Van Loey, Innov.
607		Food Sci. Emerg. Technol. (in press), 2013
608	40.	S. Gallier, D. Gragson, R. Jimenez-Flores and D. Everett, J. Agric. Food Chem., 2010, 58,
609		4250-4257.
610	41.	C. M. Galanakis, E. Tornberg and V. Gekas, LWT - Food Sci. Technol., 2010, 43, 1001-
611		1008.
612		