

Dalton Transactions

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

ARTICLE

Ligand-based Molecular Recognition and Dioxygen Splitting: An Endo Epoxide Ending†

Cite this: DOI: 10.1039/x0xx00000x

Peter E. Sues^a, Matthew W. Forbes^a, Alan J. Lough^a, and Robert H. Morris^{a*}Received 00th January 2013,
Accepted 00th January 2013

DOI: 10.1039/x0xx00000x

www.rsc.org/

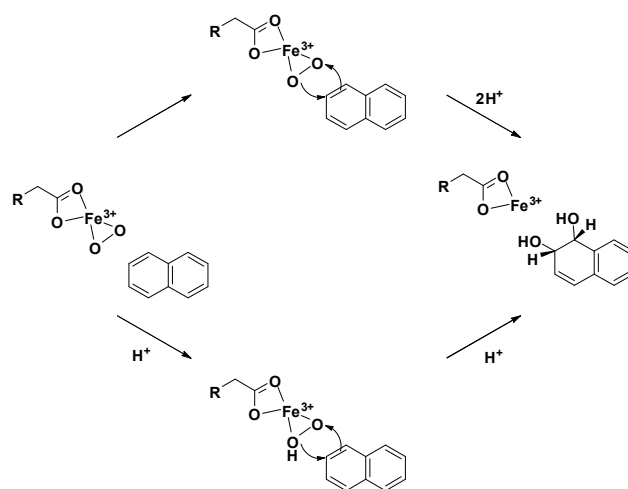
The phosphido complex $\text{RuCp}^*(\text{PPh}_2\text{CH}=\text{CHPPh}_2)(\text{PPh}_2)$ (**1**) was exposed to number of small molecules and was found to recognize and activate molecular oxygen in an unprecedented fashion: the ruthenium species split O_2 in a ligand-based 4-electron reduction to produce an endo epoxide, as well as a phosphinito ligand. Based on XRD data, VT NMR studies, cyclooctene trapping studies, and crossover experiments it was determined that the reaction proceeded through an intramolecular mechanism in which initial oxidation of the phosphido ligand generated an end-on peroxo intermediate. This mechanism was also supported by computational studies and electrochemical experiments. In contrast, an analogue of **1**, $\text{RuCp}^*(\text{Ph}_2\text{P}(\textit{ortho}\text{-C}_6\text{H}_4)\text{PPh}_2)(\text{PPh}_2)$ (**3**), reacted in an intermolecular fashion to generate two phosphinito ligands.

Introduction

The direct use of molecular oxygen in chemical transformations, as a source of oxygen atoms, is an attractive prospect for synthetic and industrial chemists because it is an abundant, cheap and completely green reagent. The problem with using dioxygen, however, is that it is often difficult to activate without resorting to highly reducing species.^[1, 2] In addition, once oxygen has been activated and partially reduced, the intermediate species are often more oxidising than O_2 itself, and, thus, are extremely reactive.^[1, 2] This can lead to uncontrolled reactions, and a lack of selectivity.^[1, 2] It is therefore important to harness the oxidizing power of dioxygen in a measured and directed fashion in order to obtain a desired product.

Although difficult to control, there are some examples in the literature of using molecular oxygen directly as an oxygen source for epoxidation reactions.^[2-11] In an industrially relevant process that is carried out on multimillion tonne scales every year, ethylene oxide is generated from ethylene and dioxygen, where the product epoxide is an important building block in the synthesis of polyethyleneglycol.^[3, 4] Biomimetic systems with iron heme complexes are also known to catalyse the epoxidation of alkenes.^[9, 10] In addition, there are a number of copper based systems that utilize aldehyde reductants that can affect this transformation.^[5, 12] Despite these successes, most of these reactions produce free peroxide species in solution (although in the biomimetic systems these are iron-oxo species). These long lived intermediates can be problematic with respect to selectivity, especially if there are multiple reactive sites in a substrate of interest.

Unlike synthetic and industrial chemists, nature is adept at selectively using dioxygen as an oxygen source for a number of aerobic transformations. In particular, mononuclear non-heme iron enzymes (MNHEs) can catalyse monooxygenation, dioxygenation and epoxidation reactions.^[13-16] The enzymatic active site contains a high spin iron(II) core that can activate molecular oxygen and generate reactive iron-oxo species. These oxidizing intermediates can then attack nearby substrates and facilitate selective reactions.^[13-21] A key feature of this process is the positioning of the substrates close to the reactive species, which splits O_2 in a reliable and controlled manner, as seen for naphthalene dioxygenase (Scheme 1).^[13-21] This molecular recognition constrains the reactive intermediates and directs their oxidizing potential towards a compound of choice.



Scheme 1. Proposed mechanism for naphthalene dioxygenase.^[21]

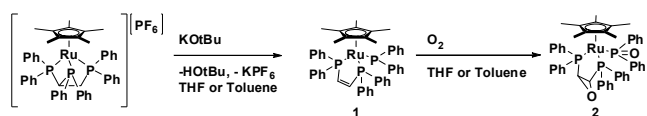
Moreover, O₂ splitting is a 4-electron reduction process. In Scheme 1 the naphthalene provides two electrons, while enzymatic co-factors and the iron active site provide the other reducing equivalents.^[13-21] The availability of all 4 electrons required to reduce O₂ is crucial for preventing the formation and release of highly-reactive, partially-reduced oxygen species, which can degrade the catalytic system. Studies examining oxygen reduction catalysts by Collman *et al.* and Love *et al.* have demonstrated this important concept in further detail.^[22-27]

We have recently described the reaction of ruthenium pentamethylcyclopentadienyl (Cp*) complexes supporting 1,1,2-tris(diphenylphosphino)ethane (tpe) ligands with KOtBu in aprotic solvents.^[28, 29] The strong base lead to the fragmentation of the tpe ligand in an elimination reaction, producing a terminal diphenylphosphido ligand, as well as 1,2-bis(diphenylphosphino)ethane (dppen); RuCp*(PPh₂CH=CHPPh₂)(PPh₂), **1** (Scheme 2).^[28] In the crystal structure of **1**, the reactive phosphido functionality was in close proximity to the alkene backbone, which suggested that small molecules could be activated in this cleft.^[28] Herein, we report the reaction of **1** with molecular oxygen in a 4 electron reduction process and explore the mechanism of this transformation.

Results and Discussion

Reactions with Dioxigen

Complex **1** was subjected to a number of small molecules, including H₂ and N₂ gas, but was unreactive towards these substrates. When **1** was exposed to air, on the other hand, a colour change from the red to yellow was seen. The ³¹P{¹H} NMR indicated that oxidation of the phosphido ligand had taken place as the phosphorus resonance shifted downfield from 23.2 ppm in **1** to 71.0 ppm.^[28] Formation of a P=O bond, however, was not the only oxidation event that had occurred. In the ¹H NMR, the absence of alkene signals between 7.4 and 7.2 ppm and the presence of aliphatic protons at 3.7 ppm hinted that activation of dppen backbone had also taken place.^[28] This observation was supported by HRMS, m/z⁺ 848.2, [C₄₈H₄₈O₂P₃Ru]⁺, which indicated that two oxygen atoms had been incorporated into the metal complex, **2**. These results revealed that epoxidation of the dppen backbone, as well as oxidation of the phosphido ligand had occurred (Scheme 2).



Scheme 2. Synthesis and oxidation of **1** using molecular oxygen.

The crystal structure of **2** confirmed these results, as both a phosphinito and an epoxide were evident (Figure 1). The C(1)-C(2) bond length was indicative of a single bond, 1.473(9) Å, while the C(1)-O(1) and C(2)-O(1) bonds were within the normal range for epoxides, 1.439(9) and 1.437(9) Å,

respectively.^[30] Furthermore, the P(3)-O(2) bond length was diagnostic for a P-O double bond, 1.519(5) Å (for other notable bond lengths and angles see Table S1).^[31] It is interesting to note that only the “endo” isomer was generated, with the epoxide oxygen tucked under the ruthenium centre (also only one species in solution was seen by ³¹P NMR). This suggested an intramolecular, ligand-based mechanism for O₂ splitting.

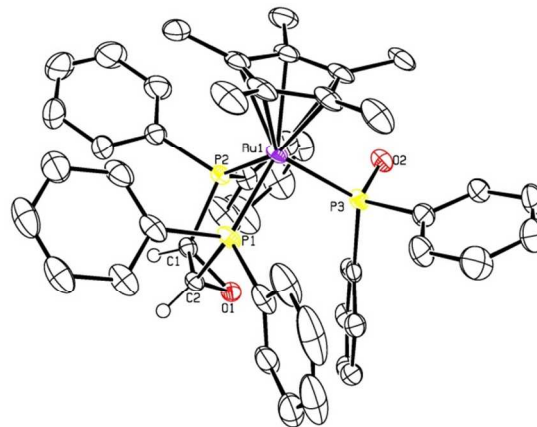


Figure 1. ORTEP3 representation (thermal ellipsoids at 50% probability; most of the hydrogens are omitted for clarity) and atom numbering for **2**.

In order to probe the nature of the epoxidation process, VT NMR studies were conducted. A ³¹P{¹H} NMR of the reaction mixture at -80°C revealed three species in solution: starting material, product, and small amounts of another species with broad signals around 77.6 and 60.3 ppm. Upon heating the sample to -60°C the intermediate disappeared while the product grew in intensity (see Supporting Information, Figure S1). The chemical shift of the peak at 60.3 ppm suggested that the phosphido ligand had been oxidized (shifted downfield from 23.2 ppm in **1**), while the dppen phosphorus nuclei were mainly undisturbed (77.6 ppm versus 82.0 ppm for **1**).^[28] Trapping studies with cyclooctene were also performed to determine if the reaction was occurring in an intermolecular fashion. Complex **1** was dissolved in almost neat cyclooctene (some THF-d₈ was added to solubilize the ruthenium species) and air was injected into the sample. The ³¹P{¹H} NMR showed that **2** had formed, while the ¹H NMR contained no cyclooctene oxide peaks.

Crossover experiments utilizing isotopically enriched ¹⁸O₂ gas, as well as pure ¹⁶O₂, were also conducted. Complex **1** was dissolved in THF and the reaction vial was sealed with a rubber septum. Half an equivalent of ¹⁸O₂, with respect to ruthenium starting material, as well as half an equivalent of ¹⁶O₂ were then simultaneously injected into the sealed vial. The reaction was allowed to proceed to completion and a mass spectrum was obtained of the resulting isotopomers. The product distribution was very close to an ideal 50:50 mixture of the all ¹⁸O species and the all ¹⁶O species (Figure 2a). This conclusively demonstrated that the oxygen splitting reaction proceeded through an intramolecular reaction, as significant amounts of the mixed ¹⁸O¹⁶O species could not be detected (Figure 2b).

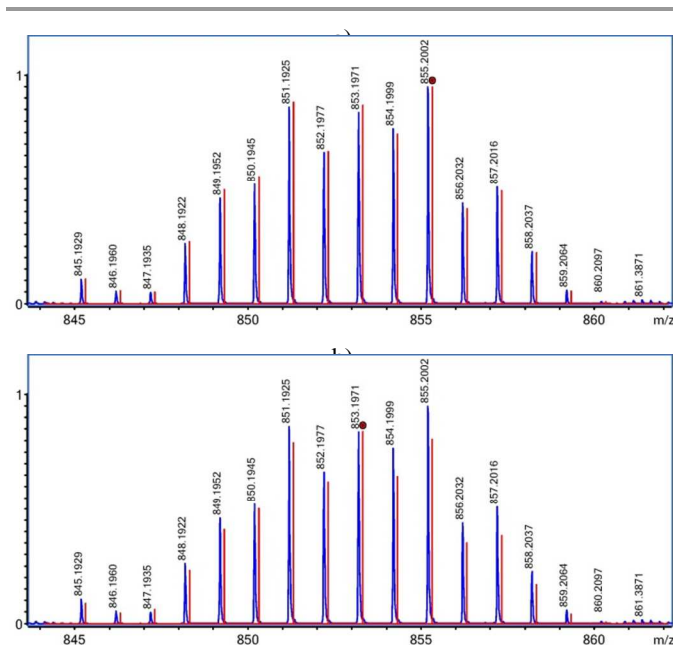


Figure 2. The experimentally determined ESI⁺ spectrum of the crossover experiment (blue) versus a simulated spectrum (red) of: a) a 48:52 mixture of the C₄₈H₄₉¹⁸O₂P₃Ru⁺ and C₄₈H₄₉¹⁶O₂P₃Ru⁺ isotopomers; b) a 45:45:10 mixture of the C₄₈H₄₉¹⁸O₂P₃Ru⁺, C₄₈H₄₉¹⁶O₂P₃Ru⁺, and C₄₈H₄₉¹⁸O¹⁶OP₃Ru⁺ isotopomers, respectively. Note that the red asterisk denotes the major peak in the simulated spectrum.

Based on the XRD data, the VT NMR studies, the cyclooctene trapping studies, and the crossover experiment we propose an intramolecular mechanism in which initial oxidation of the phosphido ligand produces an end-on peroxy species (Figure 3). This reactive phosphorus-peroxy moiety, which we predict is the intermediate that was observed by NMR, is then perfectly positioned next to the unsaturated dppen backbone, such that it can react in a selective fashion to produce an “endo” epoxide. We believe that the internal attack on the alkene backbone is so fast, and the lifetime of the intermediate is so short, that it cannot be intercepted by external alkenes. This type of mechanism is known in biological systems, where heme peroxidases, such as cytochrome P450, react with O₂ to produce end-on iron peroxy species, which can be used to generate epoxides.^[32-35] Moreover, the positioning of the substrate close to the reactive, partially reduced oxygen intermediate is somewhat reminiscent of the naphthalene dioxygenase enzyme mentioned previously, in that the proximity of the substrate, in this case the alkene backbone, quenches the reactivity of the peroxy species, thus preventing uncontrolled side reactions. Furthermore, much like the natural system, all 4-electrons necessary to fully reduce O₂ are present, which also explains why the reaction is so selective.

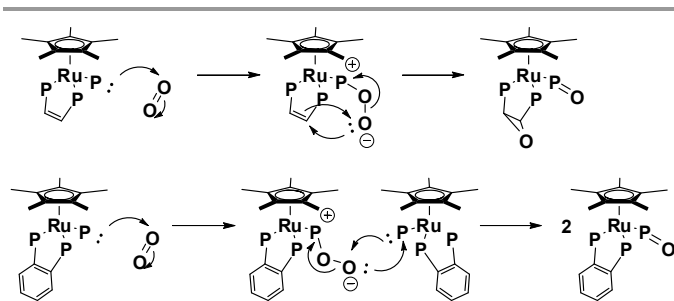
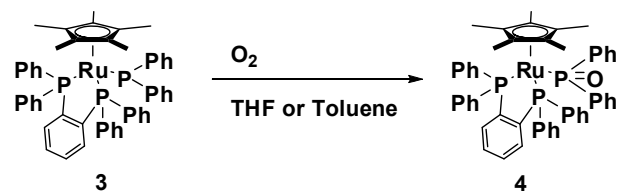


Figure 3. Proposed inter- and intramolecular mechanisms for **2** and **4**. Note that the phenyl groups have been omitted for clarity.

Similar oxidation studies were performed with the previously reported ortho-phenylene analogue of **1**; RuCp*(Ph₂PC₆H₄PPh₂)(PPh₂) (**3**).^[28] Unlike the dppen variant, the 1,2-bis(diphenylphosphino)benzene (dppbz) ligand did not get oxidized, as shown by DART HRMS (m/z⁺ 885.2, [C₅₂H₅₀OP₃Ru]⁺) and ¹H NMR (aromatic backbone protons detected, Scheme 3).

This indicated that the oxidation of **3** must have proceeded through a different mechanism than the intramolecular process discussed earlier for **1**, and that an alternative intermolecular reduction of O₂ needed to be considered in its place (Figure 3). We propose initial formation of an end-on phosphorus-peroxy species, much like the proposed intermediate for the oxidation of the dppen system, except that the peroxy species in this case cannot oxidise the dppbz backbone, but instead reacts with another equivalent of **3**. In this way, two equivalents of **3** are required to split one equivalent of O₂, which was reflected experimentally by the fact that only half an equivalent of dioxygen (half the volume of air that was necessary to fully oxidize an equivalent amount of **1**) was needed to fully oxidize **3** and generate **4**. Moreover, trapping studies with cyclooctene revealed small amounts of cyclooctene oxide in the ¹H NMR, thus supporting an intermolecular mechanism.



Scheme 3. Reactivity of complex **3** with O₂.

The alternative process seen for the oxidation of **3** further reinforces the parallels between the epoxidation seen in the generation of **2** and the molecular recognition that is vital for naphthalene dioxygenase activity. If the alkene in **1** were not perfectly positioned to react with the phosphorus-oxo intermediate, then it is likely that an intermolecular reaction would have occurred instead, and only the phosphido ligands would have been oxidized, as seen with **3**.

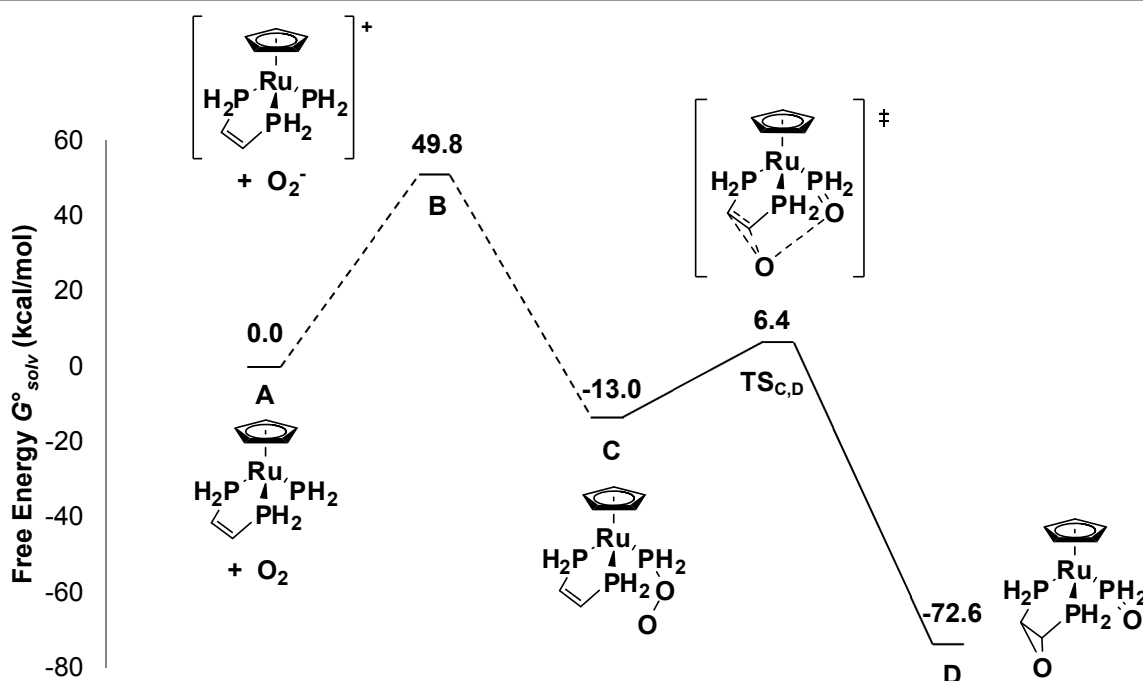


Figure 4. Energy profile for the oxygen splitting reaction (m06/6-31++G(d,p)/SDD-Ru/IEF-PCM + SMD(THF)). All energies are relative to **A** and O_2 .

DFT Studies

Detailed computational studies using the model complex **A**, $[Ru(\eta^5-C_5H_5)((PH_2)_2CHCH_2PH_2)]^+$ (all the phenyl and methyl groups were removed to reduce computational costs), were conducted in order to gain further insight into the O_2 splitting mechanism (Figures 4, 5). The studies were based on the initial presence of **A** and molecular oxygen, and as such all energies were compared to these species. From this starting point, outer-sphere electron reduction of O_2 was considered (**A** \rightarrow **B**), followed by rebound to form an end-on peroxo species (**B** \rightarrow

C), and then splitting of O_2 to produce the epoxide product (**C** \rightarrow **D**).

Calculations showed that the outer-sphere electron reduction of O_2 to generate O_2^- and **B** was very energetically demanding: 49.8 kcal/mol. On-going from the ruthenium(III) intermediate to the end-on peroxo species **C**, however, there was a large drop in energy down to -13.0 kcal/mol. A moderate activation barrier of 19.4 kcal/mol for **C** \rightarrow **D** led to the transition state $TS_{C,D}$ where the O-O bond was broken and a P-O double bond, as well as two C-O single bonds, were made. The final product **D** was found to be very stable at -72.6 kcal/mol (selected ground state and transition state geometries are shown in Figure 5).

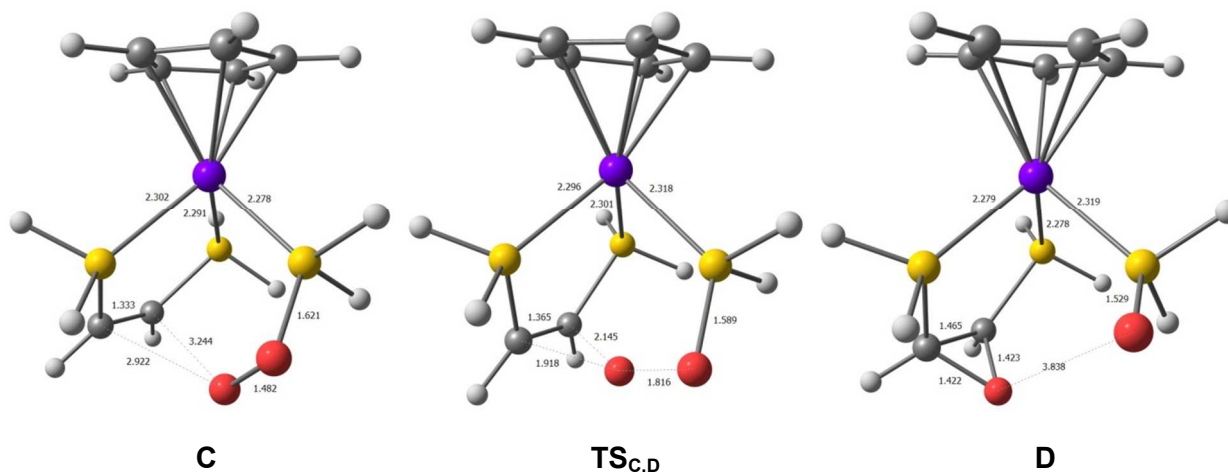


Figure 5. Optimized structures (m06/6-31++G(d,p)/SDD-Ru/IEF-PCM + SMD(THF)) as well as selected bond lengths (Å) of **C**, $TS_{C,D}$ (363i cm^{-1}), and **D**.

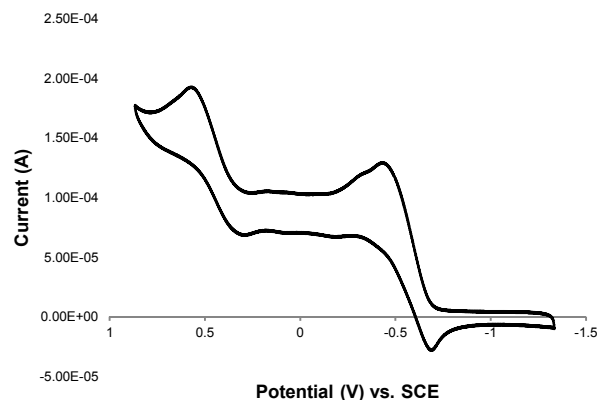
Despite the unreasonably high energy calculated for **B**, our computational studies do support our oxygen-splitting mechanism in that a low energy end-on phosphorus-peroxo intermediate was found. In addition, **C** led directly to the product epoxide with a reasonable activation barrier, giving theoretical evidence for the structure of the proposed species. With respect to the outer sphere reduction of O₂, it should be noted that the system was greatly simplified, and, if all the methyl and phenyl groups were included in the calculations, the energy of the ruthenium(III) complex could be significantly lower. Furthermore, DFT methods are capable of accurately modelling and comparing the energies of charged species or neutral species, but often have difficulty combining the two.^[36] This could also account for the disproportionately high energy calculated for the cationic complex **B**, in comparison to the other neutral species.

Electrochemical Studies

Following the DFT studies, an electrochemical analysis of **1** in THF was conducted in order to verify that the neutral ruthenium species was capable of reducing O₂ to superoxide. Scan rates of 100, 150, 200, and 400 mVs⁻¹ were carried out in both positive and negative directions, with no detectable difference between the two routes (scan rates lower than 100

mVs⁻¹ did not generate reproducible traces and lead to rapid decomposition of the phosphido complex). Two quasi-reversible redox couples were detected with an E_{pa} and E_{pc} of 0.51V and 0.25V, as well as -0.48V and -0.73V vs. SCE, which we tentatively assign as metal- and ligand-based, respectively (Figure 6 and Supporting Information). The E_{pa} for the most reducing redox couple (1) was very close to the literature value for the E_{1/2} of superoxide in CH₃CN vs. SCE (2), and as such the overall redox reaction had a modest potential of -0.22V (3).^[37] This corresponds to ΔG = 5.1 kcal/mol, which is considerably lower in energy than the value from DFT (Figure 4 vs Figure 6b).

Although these are rough calculations (the redox potential for superoxide was not measured in THF and the electrochemical analysis of **1** could not be run in CH₃CN because the ruthenium species decomposes in solution) they qualitatively show that the outer sphere-electron reduction of O₂ is not as unreasonable as the DFT studies suggest. Moreover, the E_{1/2} for the outer sphere reduction of molecular oxygen by heme proteins can be as high as -0.37V, which gives ΔG = 8.5 kcal/mol. Despite being a non-spontaneous reaction, an outer sphere mechanism is commonly invoked to explain the reactivity and kinetics of these iron systems.^[14, 15, 37-41]



a)

b)

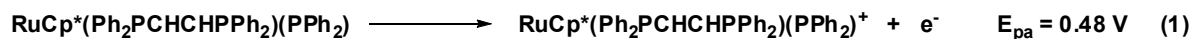
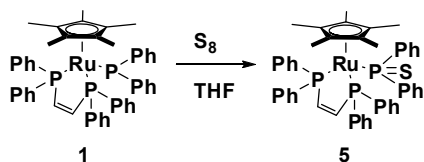


Figure 6. a) Cyclic voltammogram of **1** in THF, 0.1M [nBu₄N][PF₆], scan rate; 100 mVs⁻¹ with a scanning window of -1.34 to 0.86V; scanning in a window of -1.34 to 0.00V produced redox features in this region identical to those shown here; b) calculation of the redox potential for the reduction of oxygen by **1** to superoxide.

Reactions with Elemental Sulphur

In addition to O₂, the reactivity of complex **1** with respect to molecular sulphur was explored. Much like the reaction of **3** with dioxygen, only the phosphido ligand was oxidized, while the bidentate backbone remained untouched (Scheme 4). This is most likely due to the lower oxidation potential of sulphur in comparison to that of oxygen, such that any reactive intermediates may not have been oxidizing enough to attack the alkene backbone.



Scheme 4. Oxidation of **1** using elemental sulphur.

Moreover, assuming that the elemental sulphur was sufficiently oxidizing, the crystal structure of the sulphonated product, **5**, indicates that the alkene backbone would most likely not have formed a thioepoxide, notwithstanding. The space-filling model shows that the ruthenium complex is too crowded to accommodate two sterically demanding sulphur atoms (Figure 7).

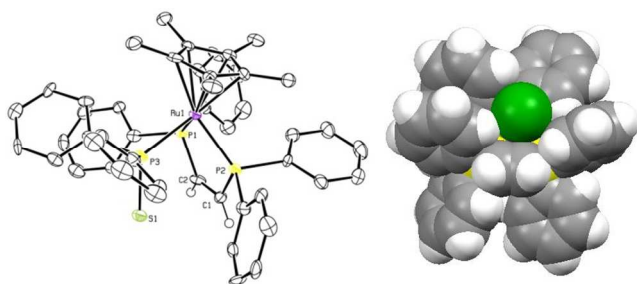


Figure 7. ORTEP3 representation (thermal ellipsoids at 50% probability; most of the hydrogens are omitted for clarity) and atom numbering for **5** (left) as well as a space-filling model of the structure (right).

The increased steric demand of the sulphur atom is reflected in the NMR spectra of the oxidized product. Crystals of **5** were dissolved in deuterated THF and two separate species could be detected in solution. We attribute this to limited rotation about the ruthenium thiophosphinito bond on the same timescale as the NMR experiment.

It seems that the cleft between the alkene backbone and the phosphido ligand is the optimal size for splitting O₂, but cannot accommodate larger substrates. This further reinforces the idea of molecular recognition with respect to complex **2** and molecular oxygen.

Conclusions

In summary, we have shown the reaction of complex **1** with molecular oxygen in a rare, ligand-based process, of which there are few related examples in the literature,^[40, 42-47] where both oxygen atoms are utilized, in this case, to generate a phosphinito ligand and an epoxide. The reaction is selective for

the reduction of O₂, as larger, less oxidizing substrates, such as S₈, and small molecules that do not undergo a 4-electron reduction process did not demonstrate the same reactivity. Although not catalytic, this system demonstrates a completely synthetic approach to using air directly in the production of epoxides; a process that generally requires peroxides as oxidizing agents or catalysts that can generate free peroxides *in situ*.^[4, 48-58] The direct use of oxygen in air, however, is much more attractive, as it is an abundant, cheap, and completely green reagent. A crucial aspect of future catalyst design is the concept of molecular recognition and the arrangement of substrates and reactive species in specialized clefts or “active sites”, such that a desired reaction proceeds in a controlled and selective manner.

Experimental

General Considerations. All procedures and manipulations were performed under an argon or nitrogen atmosphere using standard Schlenk-line and glove box techniques unless stated otherwise. All solvents were degassed and dried using standard procedures prior to all manipulations and reactions unless stated otherwise. The syntheses of complexes **1** and **3** were described previously.^[28] Deuterated solvents were purchased from Cambridge Isotope Laboratories or Sigma Aldrich, degassed, and dried over activated molecular sieves prior to use. All other reagents were purchased from commercial sources and utilized without further purification. The ESI-MS data was collected on an AB/Sciex QStar mass spectrometer with an ESI source, the EI-MS data was collected on a Waters GC ToF mass spectrometer with an EI/CI source, and the DART-MS data was collected on a JOEL AccuTOF-DART mass spectrometer with a DART-ion source (no solvent is required). NMR spectra were recorded at ambient temperature and pressure using a Varian Gemini 400 MHz spectrometer (400 MHz for ¹H, 100 MHz for ¹³C, 376 MHz for ¹⁹F, and 161 MHz for ³¹P) or an Agilent DD2-600 MHz spectrometer (600 MHz for ¹H, 151 MHz for ¹³C, 564 MHz for ¹⁹F, and 243 MHz for ³¹P) unless stated otherwise. The ¹H and ¹³C NMR were measured relative to partially deuterated solvent peaks but are reported relative to tetramethylsilane (TMS). All ³¹P chemical shifts were measured relative to 85% phosphoric acid as an external reference. The elemental analyses were performed at the Department of Chemistry, University of Toronto, on a Perkin-Elmer 2400 CHN elemental analyser. Single-crystal X-ray diffraction data were collected using a Nonius Kappa-CCD or Bruker Kappa APEX DUO diffractometer with Mo K α radiation ($\lambda = 0.710 \text{ \AA}$) or Cu K α radiation ($\lambda = 2.29 \text{ \AA}$). The CCD data were integrated and scaled using the Denzo-SMN package. The structures were solved and refined using SHELXTL V6.1. Refinement was by full-matrix least squares on F² using all data.

Computational Details. Density functional theory calculations were performed using the Gaussian09 package⁵⁹ and the M06 hybrid functional.⁶⁰⁻⁶² Ruthenium was treated with the SDD relativistic effective core potential and associated basic set,^{63, 64} while all other atoms were treated with the 6-31++G(d,p).⁶⁵⁻⁶⁷ A pruned (99,590) integration grid was used throughout

(Grid=UltraFine). The substituents on phosphorus and Cp* were replaced with hydrogen atoms to reduce computational cost. Optimizations were performed in THF using the integral equation formalism polarizable continuum model (IEF-PCM)^{68, 69} with radii and non-electrostatic terms from the SMD solvation model.⁷⁰ Ground states were connected to their transition states by performing intrinsic reaction coordinate (IRC) calculations,⁷¹ and stationary points were characterized by normal-mode analysis. Open-shell triplet-state optimizations were performed for structures **C** and **D**, but were considerably higher in energy than their singlet-state free energies (by 48.0 and 42.0 kcal/mol, respectively). In addition, a triplet transition state connecting **C** and **D** could not be found, and a high-spin ground state for **B** could also not be found. All structures except O₂ (triplet state), O₂⁻ (doublet state), and **B** (doublet state) were modelled as closed-shell singlet states. Full vibrational and thermochemical analyses (1 atm, 298 K) were performed on optimized structures to obtain solvent-corrected free energies (G°) and enthalpies (H°). Optimized ground states were found to have zero imaginary frequencies, while transition states were found to have one imaginary frequency. Three dimensional visualizations of calculated structures were generated by ChemCraft.

Electrochemical Details. Cyclic voltammetry was performed with a SP-200 Bio-Logic-Science Instruments potentiostat. A standard three-electrode cell equipped with a 2mm Pt mesh working electrode, a silver wire pseudo-reference electrode, and a Pt wire counter-electrode was used. The electrolyte was a 0.1 M solution of [nBu₄N][PF₆] in THF. Reported potentials are referenced to the ferrocenium/ferrocene couple versus the standard calomel electrode, SCE (0.50 V).

Crossover Experiment Mass Spectrometry. Electrospray ionization (ESI) mass spectra were acquired in positive ion mode using a 6538 UHD model quadrupole time-of-flight mass spectrometer (Agilent Technologies, Santa Clara, CA). Samples were introduced to the ESI source via loop injection into a mobile phase having composition 1:1 (v/v) methanol : 0.1% aqueous formic acid and flowing at a rate of 100 μL/min. Neat samples were diluted first in MeOH and subsequently in mobile phase to appropriate concentrations from which aliquots of 1-2 μL were taken for injection.

MS Data Analysis and Isotopic Abundance Simulations. Mass spectrometry data was processed using the MassHunter Qualitative Analysis software package version B.00.6 SP1 (Agilent Technologies, Santa Clara, CA). Isotopic abundance distributions were computed using the Mass Mountaineer software package version 1.00.58 (©2012 R.B. Cody).

Synthesis of complex 2 (RuCp*(Ph₂PCHOCHPPH₂)(OPPh₂)). A small amount of **1** (0.020 g, 0.023 mmol) was dissolved in THF (5 mL) in a sealed vial. Approximately 3 mL of air was added to the vial and the solution was stirred for 20 min. The colour changed from deep red to yellow, and the solvent was removed under reduced pressure. The resulting yellow solid was washed with a small

amount of cold toluene (~1 mL) and dried overnight. Yield: 86.7% (0.018 g). Crystals suitable for X-ray diffraction studies were grown from the slow diffusion of oxygen into a toluene solution of complex **1**. ¹H NMR (400 MHz, CD₂Cl₂) δ: 7.95 (br s, 4H, Ar-CH), 7.64-7.65 (m, 6H, Ar-CH), 7.36 (t, 2H, Ar-CH, *J* = 7.1 Hz), 7.28 (t, 4H, Ar-CH, *J* = 7.3 Hz), 7.18-7.11 (m, 4H, Ar-CH), 6.89 (t, 2H, Ar-CH, *J* = 7.1 Hz), 6.77 (t, 4H, Ar-CH, *J* = 7.3 Hz), 6.57 (t, 4H, Ar-CH, *J* = 8.0 Hz), 3.74 (t, 2H, O-CH, *J* = 10.4 Hz), and 1.35 (s, 15H, Cp*-CH₃) ppm. ³¹P{¹H} NMR (161 MHz, CD₂Cl₂) δ: 70.98 (t, *J* = 42.3 Hz), and 67.57 (br d, *J* = 42.3 Hz) ppm. ¹³C{¹H} NMR (100 MHz, CD₂Cl₂) δ: 139.54-139.43 (m, C-P), 137.04-136.93 (m, C-P), 134.12-133.65 (m, Ar-CH), 133.27-133.12 (m, Ar-CH), 130.77 (d, Ar-CH, *J* = 9.9 Hz), 130.04 (s, Ar-CH), 129.51 (s, Ar-CH), 127.86-127.75 (m, Ar-CH), 127.60-127.47 (m, Ar-CH), 125.94-125.85 (m, Ar-CH), 125.54 (d, Ar-CH, *J* = 8.7 Hz), 96.82-96.75 (m, Cp*-C), 58.72-58.56 (m, O-C), and 10.50 (s, Cp*CH₃) ppm. Anal. Calcd for [C₄₈H₄₇O₂P₃Ru][C₄H₁₀O]: C, 67.59; H, 6.22. Found: C, 67.49; H, 6.59. MS (DART, DCM; *m/z*⁺): 848.2 [C₄₈H₄₈O₂P₃Ru]⁺.

Synthesis of complex 4 (RuCp*(Ph₂PC₆H₄PPh₂)(OPPh₂)). A small amount of **3** (0.040 g, 0.046 mmol) was dissolved in THF (3 mL) in a sealed vial. Approximately 3 mL of air was added to the vial and the solution was stirred for 30 min. The colour gradually changed from deep red to yellow, and the solvent was removed under reduced pressure. The resulting yellow solid was dissolved in toluene (~2 mL) and cooled to -33 °C in a freezer overnight. The solution was filtered through Celite and the solvent was removed under reduced pressure to give a yellow powder. Yield: 89.3% (0.036 g). ¹H NMR (400 MHz, C₄D₈O) δ: 8.06-7.98 (m, 4H, Ar-CH), 7.34-7.38 (m, 4H, Ar-CH), 7.27-7.17 (m, 4H, Ar-CH), 7.17-7.07 (m, 12H, Ar-CH), 6.78-6.72 (m, 2H, Ar-CH), 6.65-6.58 (m, 4H, Ar-CH), 6.54-6.46 (m, 4H, Ar-CH), and 1.28 (d, 15H, Cp*-CH₃, *J* = 1.2 Hz) ppm. ³¹P{¹H} NMR (161 MHz, C₄D₈O) δ: 90.79 (br s), and 75.15 (d, *J* = 45.3 Hz) ppm. ¹³C{¹H} NMR (100 MHz, C₄D₈O) δ: 148.60 (d, P-C, *J* = 38.4 Hz), 145.95 (t, P-C, *J* = 43.5 Hz), 140.20-138.48 (m, P-C), 135.72 (dd, C-P, *J* = 23.5, 3.4 Hz), 135.43-135.15 (m, Ar-CH), 133.29 (t, Ar-CH, *J* = 5.0 Hz), 132.11 (d, Ar-CH, *J* = 10.2 Hz), 132.04 (t, Ar-CH, *J* = 8.3 Hz), 128.39 (br s, Ar-CH), 128.32 (br s, Ar-CH), 126.92-126.68 (m, Ar-CH), 126.50 (d, Ar-CH, *J* = 2.1 Hz), 125.30 (d, Ar-CH, *J* = 8.7 Hz), 94.90 (d, Cp*-C, *J* = 1.6 Hz), and 9.89 (s, Cp*-CH₃, *J* = 4.7 Hz) ppm. Anal. Calcd for [C₅₂H₄₉OP₃Ru]0.66[C₅H₁₂]: C, 71.31; H, 6.16. Found: C, 71.25; H, 6.65. MS (DART, no solvent; *m/z*⁺): 885.2 [C₅₂H₅₀OP₃Ru]⁺.

Synthesis of complex 5 (RuCp*(Ph₂PCHCHPPh₂)(SPPH₂)). A small amount of **1** (0.020 g, 0.023 mmol) was dissolved in THF (5 mL). Elemental sulphur (0.006 g, 0.023 mmol) was added and the solution was stirred for 15 min. The colour rapidly changed from deep red to yellow, and the solvent was removed under reduced pressure. The resulting yellow solid was dissolved in toluene (~2 mL) and cooled to -33 °C in a freezer overnight. The solution was filtered through Celite and

the solvent removed under reduced pressure to give a yellow powder. Yield: 92.1% (0.019 g). Crystals suitable for X-ray diffraction studies were grown from the slow evaporation of a THF solution of complex **5**. **Isomer 1**: ^1H NMR (600 MHz, $\text{C}_4\text{D}_8\text{O}$, -35°C) δ : 7.96 (t, 4H, Ar-CH, $J = 8.5$ Hz), 7.77-7.64 (m, 2H, CH=CH), 7.33-7.27 (m, 6H, Ar-CH), 7.22 (t, 4H, Ar-CH, $J = 8.1$ Hz), 7.12 (t, 4H, Ar-CH, $J = 8.5$ Hz), 6.84-6.79 (m, 6H, Ar-CH), 6.69 (t, 6H, Ar-CH, $J = 7.2$ Hz), and 1.27 (s, 15H, Cp*-CH₃) ppm. $^{31}\text{P}\{^1\text{H}\}$ NMR (242 MHz, $\text{C}_4\text{D}_8\text{O}$, -35°C) δ : 71.86 (d, $J = 38.2$ Hz), and 60.64 (t, $J = 38.2$ Hz) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (150 MHz, $\text{C}_4\text{D}_8\text{O}$, -35°C) δ : 145.82 (d, CH-P, $J = 32.8$ Hz), 138.20 (d, C-P, $J = 39.9$ Hz), 135.22 (d, C-P, $J = 45.8$ Hz), 133.76 (d, Ar-CH, $J = 7.8$ Hz), 133.27 (d, Ar-CH, $J = 8.7$ Hz), 133.08 (d, Ar-CH, $J = 11.3$ Hz), 128.93 (s, Ar-CH), 128.40 (s, Ar-CH), 127.26 (d, Ar-CH, $J = 7.8$ Hz), 126.83 (d, Ar-CH, $J = 8.9$ Hz), 126.11 (s, Ar-CH), 125.10 (d, Ar-CH, $J = 9.9$ Hz), 94.19 (s, Cp*-C), and 9.91 (s, Cp*CH₃) ppm. **Isomer 2**: ^1H NMR (600 MHz, $\text{C}_4\text{D}_8\text{O}$, -35°C) δ : 7.64-7.60 (m, 2H, Ar-CH), 7.59-7.50 (m, 4H, Ar-CH), 7.48-7.36 (m, 6H, Ar-CH), 7.36-7.32 (m, 2H, Ar-CH, $J = 7.3$ Hz), 7.08-6.99 (m, 4H, Ar-CH), 6.97-6.91 (m, 2H, Ar-CH), 6.90-6.84 (m, 4H, Ar-CH), 6.45-6.37 (m, 2H, Ar-CH, $J = 8.0$ Hz), and 1.55 (s, 15H, Cp*-CH₃) ppm. $^{31}\text{P}\{^1\text{H}\}$ NMR (242 MHz, $\text{C}_4\text{D}_8\text{O}$, -35°C) δ : 70.16 (br d, $J = 36.4$ Hz), 65.34 (br d, $J = 36.4$ Hz), and 62.23 (t, $J = 36.4$ Hz) ppm. $^{13}\text{C}\{^1\text{H}\}$ NMR (150 MHz, $\text{C}_4\text{D}_8\text{O}$, -35°C) δ : 145.42-145.14 (m, CH-P), 145.10-144.83 (m, CH-P), 143.00-142.35 (m, C-P), 138.12-137.61 (m, C-P), 136.84-136.45 (m, C-P), 134.05-133.81 (m, Ar-CH), 132.98-132.77 (m, Ar-CH), 132.68-132.37 (m, Ar-CH), 129.57-129.30 (m, Ar-CH), 129.24-129.02 (m, Ar-CH), 128.20-128.05 (m, Ar-CH), 127.19-126.87 (m, Ar-CH), 126.77-126.52 (m, Ar-CH), 125.80-125.59 (m, Ar-CH), 125.35-125.09 (m, Ar-CH), 95.58 (s, Cp*-C), and 10.53 (s, Cp*CH₃) ppm. Anal. Calcd for $[\text{C}_{48}\text{H}_{47}\text{P}_3\text{RuS}]_0.33[\text{C}_7\text{H}_8]_0.33[\text{C}_4\text{H}_8\text{O}]$: C, 68.59; H, 5.83. Found: C, 68.54; H, 5.83. MS (ESI⁺, DCM; m/z^+): 851.2 $[\text{C}_{48}\text{H}_{48}\text{P}_3\text{RuS}]^+$.

$^{18}\text{O}_2/^{16}\text{O}_2$ Crossover Experiment. A small amount of **1** (0.010 g, 0.012 mmol) was dissolved in 2 mL THF in a vial sealed with a rubber septum. The two oxygen isotopomers, $^{18}\text{O}_2$ (0.15 mL, 0.06 mmol) and $^{16}\text{O}_2$ (0.15 mL, 0.06 mmol) were then simultaneously injected into the vial. The solution was stirred for 30 min. and the deep red solution turned bright yellow. The solvent was removed *in vacuo*, and the yellow solid was used for MS experiments.

Acknowledgements

We thank NSERC Canada for a Discovery grant to R. H. M., and Tyler Schon from the Seferos group for his assistance with cyclic voltammetry.

Notes and References

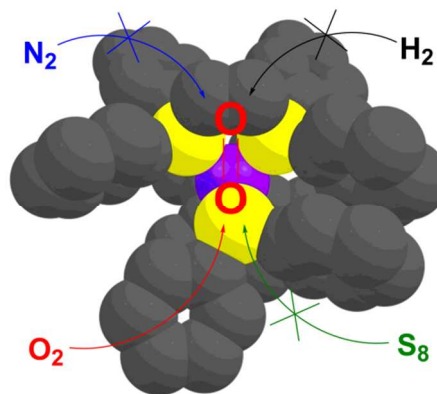
^a Department of Chemistry, University of Toronto, Toronto, Ontario, M5S 3H6, Canada. E-mail: rmmorris@chem.utoronto.ca

† Electronic Supplementary Information (ESI) available: Extended experimental section. Selected bond lengths and angles for **2** and **5**. Crystallographic data tables for **2** and **5** (including CIF files). Discussion of the VT NMR studies. Discussion of the origin of the quasi-reversible redox couples for **2**. Tables giving Cartesian coordinates and free energies for optimized structures. Text giving the complete ref 59. See DOI: 10.1039/b000000x/

- Verkhovskiy, M. I.; Morgan, J. E.; Wikstroem, M. *Biochemistry* **1994**, *33*, 3079-3086.
- Shi, Z.; Zhang, C.; Tang, C.; Jiao, N. *Chem. Soc. Rev.* **2012**, *41*, 3381-3430.
- Rebsdat, S.; Mayer, D. Ethylene Oxide. In *Ullmann's Encyclopedia of Industrial Chemistry*, Wiley-VCH: Weinheim, 2000.
- Kahlich, D.; Wiechern, U.; Lindner, J. Propylene Oxide. In *Ullmann's Encyclopedia of Industrial Chemistry*, Wiley-VCH: Weinheim, 2000.
- Allen, S. E.; Walvoord, R. R.; Padilla-Salinas, R.; Kozlowski, M. C. *Chem. Rev.* **2013**, *113*, 6234-6458.
- Jira, R. *Angew. Chem. Int. Ed.* **2009**, *48*, 9034-9037.
- Hosseini-Monfared, H.; Meyer, H.; Janiak, C. *J. Mol. Catal. A: Chem.* **2013**, *372*, 72-78.
- Punniyamurthy, T.; Velusamy, S.; Iqbal, J. *Chem. Rev.* **2005**, *105*, 2329-2364.
- Piera, J.; Bäckvall, J.-E. *Angew. Chem. Int. Ed.* **2008**, *47*, 3506-3523.
- Que, L.; Tolman, W. B. *Nature* **2008**, *455*, 333-340.
- Sorokin, A. B. *Chem. Rev.* **2013**, *113*, 8152-8191.
- Punniyamurthy, T.; Rout, L. *Coord. Chem. Rev.* **2008**, *252*, 134-154.
- Kovaleva, E. G.; Lipscomb, J. D. *Nat. Chem. Biol.* **2008**, *4*, 186-193.
- Bugg, T. D. H. *Tetrahedron* **2003**, *59*, 7075-7101.
- Bugg, T. D. H.; Ramaswamy, S. *Curr. Opin. Chem. Biol.* **2008**, *12*, 134-140.
- Lange, S. J.; Que, L. *Curr. Opin. Chem. Biol.* **1998**, *2*, 159-172.
- Kovaleva, E. G.; Neiberger, M. B.; Chakrabarty, S.; Lipscomb, J. D. *Acc. Chem. Res.* **2007**, *40*, 475-483.
- Costas, M.; Mehn, M. P.; Jensen, M. P.; Que, L. *Chem. Rev.* **2004**, *104*, 939-986.
- Solomon, E. I.; Wong, S. D.; Liu, L. V.; Decker, A.; Chow, M. S. *Curr. Opin. Chem. Biol.* **2009**, *13*, 99-113.
- Buongiorno, D.; Straganz, G. D. *Coord. Chem. Rev.* **2013**, *257*, 541-563.
- Karlsson, A.; Parales, J. V.; Parales, R. E.; Gibson, D. T.; Eklund, H.; Ramaswamy, S. *Science* **2003**, *299*, 1039-1042.
- Collman, J. P.; Devaraj, N. K.; Decréau, R. A.; Yang, Y.; Yan, Y.-L.; Ebina, W.; Eberspacher, T. A.; Chidsey, C. E. D. *Science* **2007**, *315*, 1565-1568.
- Collman, J. P.; Kaplun, M.; Decréau, R. A. *Dalton Trans.* **2006**, *0*, 554-559.
- Chang, C. J.; Loh, Z.-H.; Shi, C.; Anson, F. C.; Nocera, D. G., *J. Am. Chem. Soc.* **2004**, *126*, 10013-10020.
- Devoille, A. M. J.; Love, J. B. *Dalton Trans.* **2012**, *41*, 65-72.
- Boulatov, R.; Collman, J. P.; Shiryayeva, I. M.; Sunderland, C. J. *J. Am. Chem. Soc.* **2002**, *124*, 11923-11935.
- Askarizadeh, E.; Yaghoob, S. B.; Boghaei, D. M.; Slawin, A. M. Z.; Love, J. B. *Chem. Commun.* **2010**, *46*, 710-712.
- Sues, P. E.; Lough, A. J.; Morris, R. H. **2013**, *submitted*.

29. Sues, P. E.; Lough, A. J.; Morris, R. H. *Organometallics* **2012**, *31*, 6589-6594.
30. Illa, O.; Gornitzka, H.; Branchadell, V.; Baceiredo, A.; Bertrand, G.; Ortuño, Rosa M. *Eur. J. Org. Chem.* **2003**, *2003*, 3147-3152.
31. Sues, P. E.; Lough, A. J.; Morris, R. H. *Inorg. Chem.* **2012**, *51*, 9322-9332.
32. Wang, D.; Thiel, W. *J. Mol. Struct. (Theochem.)* **2009**, *898*, 90-96.
33. Chen, H.; Ikeda-Saito, M.; Shaik, S. *J. Am. Chem. Soc.* **2008**, *130*, 14778-14790.
34. van Rantwijk, F.; Sheldon, R. A. *Curr. Opin. Biotechnol.* **2000**, *11*, 554-564.
35. Wilson, S. A.; Kroll, T.; Decréau, R. A.; Hocking, R. K.; Lundberg, M.; Hedman, B.; Hodgson, K. O.; Solomon, E. I. *J. Am. Chem. Soc.* **2013**, *135*, 1124-1136.
36. Wu, Q.; Ayers, P. W.; Yang, W. *J. Chem. Phys.* **2003**, *119*, 2978-2990.
37. Kryatov, S. V.; Taktak, S.; Korendovych, I. V.; Rybak-Akimova, E. V.; Kaizer, J. Z.; Torelli, S. P.; Shan, X.; Mandal, S.; MacMurdo, V. L.; Mairata i Payeras, A.; Que, L. *Inorg. Chem.* **2004**, *44*, 85-99.
38. Gonzalez, G.; Gilles-Gonzalez, M. A.; Rybak-Akimova, E. V.; Buchalova, M.; Busch, D. H. *Biochemistry* **1998**, *37*, 10188-10194.
39. Shikama, K. *Chem. Rev.* **1998**, *98*, 1357-1374.
40. Tejel, C.; Del Río, M. P.; Ciriano, M. A.; Reijerse, E. J.; Hartl, F.; Zális, S.; Hettterscheid, D. G. H.; Spithas, N. T. I.; De Bruin, B. *Chem. - Eur. J.* **2009**, *15*, 11878-11889.
41. Purdy, M. M.; Koo, L. S.; Ortiz de Montellano, P. R.; Klinman, J. P., *Biochemistry* **2006**, *45*, 15793-15806.
42. Chohan, B. S.; Maroney, M. J. *Inorg. Chem.* **2006**, *45*, 1906-1908.
43. Lee, C.-M.; Hsieh, C.-H.; Dutta, A.; Lee, G.-H.; Liaw, W.-F. *J. Am. Chem. Soc.* **2003**, *125*, 11492-11493.
44. McQuilken, A. C.; Jiang, Y.; Siegler, M. A.; Goldberg, D. P. *J. Am. Chem. Soc.* **2012**, *134*, 8758-8761.
45. Videla, M.; Roncaroli, F.; Slep, L. D.; Olabe, J. A. *J. Am. Chem. Soc.* **2006**, *129*, 278-279.
46. Ho, D. G.; Gao, R.; Celaje, J.; Chung, H.-Y.; Selke, M. *Science* **2003**, *302*, 259-262.
47. Mastroilli, P.; Latronico, M.; Nobile, C. F.; Suranna, G. P.; Fanizzi, F. P.; Englert, U.; Ciccarella, G. *Dalton Trans.* **2004**, 1117-1119.
48. Prileschajew, N. *Ber. Dtsch. Chem. Ges.* **1909**, *42*, 4811-4815.
49. Ishii, Y.; Sakaguchi, S.; Iwahama, T. *Adv. Synth. Catal.* **2001**, *343*, 393-427.
50. Habibi, D.; Faraji, A. R.; Arshadi, M.; Heydaria, S.; Gil, A. *Appl. Catal. A* **2013**, *466*, 282-292.
51. Tada, M.; Muratsugu, S.; Kinoshita, M.; Sasaki, T.; Iwasawa, Y. *J. Am. Chem. Soc.* **2009**, *132*, 713-724.
52. Tada, M.; Coquet, R.; Yoshida, J.; Kinoshita, M.; Iwasawa, Y. *Angew. Chem. Int. Ed.* **2007**, *46*, 7220-7223.
53. Tada, M.; Akatsuka, Y.; Yang, Y.; Sasaki, T.; Kinoshita, M.; Motokura, K.; Iwasawa, Y. *Angew. Chem. Int. Ed.* **2008**, *47*, 9252-9255.
54. Nishiyama, H.; Motoyama, Y. *Chem. Commun.* **1997**, 1863-1864.
55. Liu, L.-L.; Li, H.-X.; Wan, L.-M.; Ren, Z.-G.; Wang, H.-F.; Lang, J.-P. *Chem. Commun.* **2011**, *47*, 11146-11148.
56. Koya, S.; Nishioka, Y.; Mizoguchi, H.; Uchida, T.; Katsuki, T. *Angew. Chem. Int. Ed.* **2012**, *51*, 8243-8246.
57. Goberna-Ferrón, S.; Lillo, V.; Galán-Mascarós, J. R. N. *Catal. Commun.* **2012**, *23*, 30-33.
58. Oloo, W. N.; Fielding, A. J.; Que, L. *J. Am. Chem. Soc.* **2013**, *135*, 6438-6441.
59. Frisch, M. J. *et. al.*, Gaussian 09, Revision B.01; Gaussian, Inc.: Wallingford CT, **2010**. For the full reference, see the Supporting Information.
60. Kulkarni, A. D.; Truhlar, D. G. *J. Chemical Theory Comput.* **2011**, *7*, 2325-2332.
61. Zhao, Y.; Truhlar, D. *Theor. Chem. Acc.* **2008**, *120*, 215-241.
62. Zhao, Y.; Truhlar, D. G. *J. Chem. Phys.* **2006**, *125*, 194101-194118.
63. Leininger, T.; Nicklass, A.; Stoll, H.; Dolg, M.; Schwerdtfeger, P. *Journal Chem. Phys.* **1996**, *105*, 1052-1059.
64. Andrae, D.; Häußermann, U.; Dolg, M.; Stoll, H.; Preuß, H. *Theor. Ahim. Acta* **1990**, *77*, 123-141.
65. Clark, T.; Chandrasekhar, J.; Spitznagel, G. W.; Schleyer, P. V. R. *J. Comput. Chem.* **1983**, *4*, 294-301.
66. Lynch, B. J.; Zhao, Y.; Truhlar, D. G. *J. Phys. Chem. A* **2003**, *107*, 1384-1388.
67. Frisch, M. J.; Pople, J. A.; Binkley, J. S. *J. Chem. Phys.* **1984**, *80*, 3265-3269.
68. Tomasi, J.; Mennucci, B.; Cammi, R. *Chem. Rev.* **2005**, *105*, 2999-3094.
69. Tomasi, J.; Mennucci, B.; Cancès, E. *J. Mol. Struct. (THEOCHEM)* **1999**, *464*, 211-226.
70. Marenich, A. V.; Cramer, C. J.; Truhlar, D. G. *J. Phys. Chem. B* **2009**, *113*, 6378-6396.
71. Fukui, K. *Acc. Chem. Res.* **1981**, *14*, 363-368.

TOC Graphic



Text: Dioxygen adds selectively at the cleft created by diphenylphosphide and an unsaturated chelating diphosphine on ruthenium to produce an endo epoxide complex.