PCCP



Q1 Q2

10

15

20

25

30

35

40

45

50

1

10

5

Cite this: DOI: 10.1039/c3cp54913e

15

20

40

45

50

Received 20th November 2013, Accepted 23rd December 2013

DOI: 10.1039/c3cp54913e

25 www.rsc.org/pccp

Introduction

30 Bioactive glasses (BGs) represent a key reference in the field of materials for biomedicine. Traditional clinical uses of BGs exploit the ability of the glass to form bonds with existing tissues, thus providing a stable matrix that promotes the

Antonio Tilocca*

35 Department of Chemistry, University College London, 20 Gordon Street, London WC1H 0AJ, UK. E-mail: a.tilocca@ucl.ac.uk

Antonio Tilocca

Antonio Tilocca holds a Royal Society University Research Fellowship (RS-URF) at University College London. He received his PhD in Physical Chemistry from University of Sassari, Italy, in 2000. He then held postdoctoral positions at University of Insubria at Como (Italy), Princeton University, and University College London, before taking up the RS-URF in 2006. His research is focused on rationalising the behaviour of

materials at the atomic scale. He employs classical and ab initio simulations to model structure, dynamics, and reactivity of 55 materials including crystalline and amorphous oxides with applications in biomedicine and (photo)catalysis.

Atomic-scale computer simulations have emerged as a powerful tool to probe at a very high resolution, the structural and dynamical properties of amorphous and crystalline biomaterials with a direct impact on their biological activity. In particular, bioactive glasses (BGs) represent a target of high strategic importance for the simulations, due to the central role that they play in the broad arena of materials for repairing and regenerating tissues. Simulations aimed at understanding the properties of bioactive glasses thus reveal the potential, and also the limitations, of computational approaches to support the rational development of biomaterials. This perspective article examines several key challenges that computer simulations of BGs are currently dealing with and that will need to be effectively tackled in order to achieve further substantial progress in this field. Relevant examples are the identification of new structural descriptors, the modelling of ion migration, and the simulation of nanosized samples, which are discussed in relation to the underlying issues, such as the limited space and time scales that can be probed using simulations.

Current challenges in atomistic simulations of

glasses for biomedical applications

growth of new bone and supports bone repair.¹ Subsequent developments are attempting to capitalise on the ability of BGs to promote regeneration of tissues away from the implant interface, thanks to the osteogenic properties of the ionic products released at and around the implant site by the glass dissolution.^{2,3} Reflecting this shift of perspective, while bioactivity was sometimes associated only with the bonebonding ability of a material, nowadays the term tends to be used in a broader context, to indicate materials capable of inducing a favourable response from the body.³ The bone bonding and tissue regeneration abilities both depend on the fast release of soluble species such as calcium, silica and phosphates from the implanted glass. This establishes critical concentrations of these species in the environment surrounding the implant, which are then capable of activating bone-bonding processes at the glass/tissue interface and also triggering osteogenic mechanisms at the cellular level.³ The proangiogenic and anti-osteoporotic properties of these glasses, recently emerged, also depend on the delivery and release in situ of other ions, such as nickel, cobalt and strontium from suitably doped bioglasses.4-6

The key role of the glass dissolution in controlling its biological activity underpins current attempts at rationalising the performances of these materials based on structural and dynamical features with a direct impact on the dissolution process itself.^{7,8} The typical size and time scales of these features, involving structural units covering few Å and elementary dynamical events lasting few picoseconds, make atomistic computer simulations such as Molecular Dynamics (MD) particularly suitable to investigate them. Whereas promising

1 progress has been made in some respects (for instance, the identification of direct structure-bioactivity links⁹) progress has been slower in other directions, such as in the explicit simulation of dynamical transformation and reactivity at the interface.^{10,11}

5

In this Perspective, several challenges faced today by atomistic computer modelling approaches applied to rationalise the behaviour of bioactive glasses are discussed, in relation to the successes and limitations of the simulations in each case. Even

10 though the discussion is focused on BGs, the established role of these systems in the field is such that many considerations made here can be considered relevant in the broader area of atomistic simulations of biomaterials.

15

50

55

Current challenges for atomic-scale simulations

The physicochemical behaviour of a bioactive glass following 20 its implant in a physiological environment can be investigated and understood using computer simulations targeting different aspects:

(1) Bulk and surface structure. The rationalisation of the compositional dependence of several macroscopic properties 25 of glasses based on the underlying microstructure is a very active field.^{12–15} In particular, several structural features of the bulk glass have previously been associated with the glass stability, solubility and bioactivity.¹⁶⁻¹⁸ The simulations can directly determine these and other microscopic features, assess 30 their correlation with available experimental data on the

solubility, and extrapolate these findings to predict the behaviour of new glasses.¹⁹ Furthermore, reactive models of the surface region exposed to molecular probes such as water and ammonia^{11,20,21} allow one to integrate the information 35 above with additional details about specific surface sites (not necessarily stable in the bulk glass structure) that contribute to the interaction of the biomaterial with the host.

(2) Ion migration dynamics. The release of a biologically active ionic species (e.g., a modifier cation, but also a soluble phos-40phate or a silicate chain, etc.) from the glass depends on the rate at which the species itself can migrate through the bulk and gain access to the interface region. Indirect conjectures about the expected ion migration behaviour can be based on the static local structure (coordination environment) of the mobile species, which obviously influences its dynamical 45 behaviour.^{5,22,23} However, MD simulations also allow one to directly observe the diffusive process with very high space and time resolution, so that the diffusive mechanism of different soluble species can be fully analysed and linked to the biological behaviour.²⁴

(3) Reduced size. Once their effect on the glass behaviour has been fully understood, adjusting the structural and dynamical features mentioned above is the key to achieve specific resorption rates and bioreactivity, tailored to a specific application. The emerging new behaviour exhibited by BG systems of sub-

micrometre dimensions, such as BG nanoparticles (NPs) and

10

15

40

1

5

Bulk structure

New structural descriptors

further progress in each case.

The largest fraction of computational work in the field of bioactive glasses has been focused on their bulk structure. This is because the computational methods to obtain a realistic model of a bulk melt-derived glass are relatively well developed, also for more complex, multicomponent compositions.^{7,30,31}

nanofibres,^{25–28} points to the reduced size as a powerful way to

achieve this target. Simulations can provide answers to the

question of how shape and morphology of the glass substrate

In the following, I will summarize the main findings of

affect the above properties and thus control its performance.

recent simulations performed in the three areas above, and

discuss some important challenges that must be faced to make

For instance, Reverse Monte Carlo (RMC) modelling enables the constrained fitting of experimental neutron or X-ray diffrac-20 tion patterns with an atomic-scale 3D structure, which can then be used to isolate short-range features such as the local coordination of a species.³² The quantitative reproduction of medium-range features, however, is not necessarily as accurate, and further independent experimental and theoretical data 25 may be needed for refining the RMC hypothetical structure,³³ which cannot then be completely unbiased. On the other hand, MD simulations directly yield an unbiased structural picture of the glass, which does not rely on the availability of additional structural data, and whose reliability only depends on the 30 quality of the underlying force field. Accurate force fields are now available that allow one to access quite large system sizes, on the order of $\sim 10^4$ atoms, with a high level of accuracy.³⁴ Models of this size, spanning lengths between 2 and 10 nm, are necessary to extract structural properties relevant for the glass 35 dissolution with a high statistical accuracy. In fact, ion dissolution rates of glasses indirectly reflect medium-range bulk features such as network connectivity,¹⁶ ion clustering and nanosegregation⁹ or organisation in chain and ring nanostructures.¹⁷ Whereas the effect of these features on the bioactivity is now relatively well established,⁷ it has also been emerging that they cannot provide a complete description, allowing accurate predictive estimates of the glass behaviour in different cases. Further work is needed to discover additional structural descriptors that exhibit significant correlation with the glass 45 durability and can then be employed to expand and complement the set of structural parameters discussed above. This task requires experimental reference data on the compositional trends of the glass biodegradation rate.³⁵ By fitting a suitable predictive model to the experimental dataset, one can identify 50 new structural parameters whose inclusion in the set of descriptors improves the fitting, and discard those descriptors that do not. Following this approach, we were recently able to identify a few new structural descriptors that affect the solubility and thus the performances of glasses used as radioisotope 55 vectors for *in situ* radiotherapy,^{19,29} an application which also

PCCP

20

25

30

35

40

45

50



Fig. 1 Nanostructures taken from the model of a Y-doped bioglass, showing a modifier ion (the sphere at the centre of the loop, left: Ca, right: Y) cross-linking two tetrahedra belonging to the same chain fragment: the modifier ion can be thought as holding the whole O3SiO- $(SiO_2)_n$ -OSiO₃ unit in place and thus increasing the network durability by hindering its release. Adapted from ref. 29 with permission from The Royal 15 Society of Chemistry.

critically depends on the glass durability. The additional structural descriptors that were identified complement standard 20 parameters such as the network connectivity in quantitatively expressing the strength of the glass network and its resistance to dissolution. For instance, non-covalent cross-links between separate portions of the silicate network, bridged by a central modifier cation, play a key role in this context. Phosphosilicate chains

- 25 whose end members are coordinated to the same modifier cation can be considered as held together (and thus, to some extent - depending on the specific strength of the cation-glass network interaction - prevented from dissolving) by that same cation^{29,36} (Fig. 1). It turned out that a higher ionic field
- 30 strength (such as that of Y^{3+} , used as β -emitter in radiotherapy) increases the ratio of inter- to intra-tetrahedral linkages in the ion's coordination shell, and leads to a greater ability of the higher-field strength cation to bind together several spatiallyseparated fragments of the glass network (Fig. 1).^{29,37} Because 35 these effects work together against the break-up and dissolu-
- tion of the glass matrix, quantitative measurements of the corresponding features, extracted from the models, can be successfully included into the set of structural descriptors that a solubility-predictive model is built upon.¹⁹ The key challenges 40here are represented by the need to obtain experimental
- datasets covering the biodegradation of a wide range of relevant compositions, and the task of devising structural descriptors such as those above that can help to reproduce, and then eventually predict, the experimental trends.
- 45

Cooling-rate and size effects

50

A key issue to be assessed is the possible effect of the necessarily limited time and space scales common to all MD-based computational procedures to generate a model of a glass, on the accuracy of key structural descriptors such as those discussed above. The MD procedures typically involve a melt-andquench approach that mimics the experimental one, but over much faster cooling rates and with much smaller samples. The 55 thorough assessment of size and cooling rate effects is conceptually straightforward, but it represents a significant



Fig. 2 Main panel: distribution of the lengths of (phospho)silicate chains identified in MD models of 45S5 Bioglass[®] of different sizes.³⁸ Inset: average chain length of models obtained at different cooling rates.

challenge in practice, because of the implicit difficulty in evaluating the convergence of the structure over simulation conditions varied over very different time and space scales, covering several orders of magnitude. For instance, the 10³ time scaling factor involved when switching from a conventional (10 K ps^{-1}) to a considerably slower (0.01 K ps^{-1}) cooling rate leads to a corresponding massive increase in the simulation time required to complete the simulation.† This difficulty is vastly amplified for large sample sizes, with the result that even with powerful state-of-the-art computer resources at hand, combining large systems ($N > 10^5$ atoms) with slow cooling rates ($<10^{-2}$ K ps⁻¹) still represents an arduous task.

However, recent investigations^{38,39} show that the weight of size and cooling rate effects is indeed small, not only for shortrange properties (which is expected), but particularly for key structural properties such as Q^n speciation, chain/ring structure, and spatial distribution of modifier cations. For instance, Fig. 2 illustrates the chain structure of MD models of 45S5 Bioglass[®] obtained under different simulation conditions.³⁸ The 45S5 structure appears dominated by short silicate chains containing 2 to 4 monomers: the inset of Fig. 2 confirms that the average chain length is around 3, essentially unaffected by the cooling rate, besides statistical fluctuations. The distributions of chain lengths, shown in the main panel of the figure, retain a very similar trend in a range of system sizes varied by a factor of 32, the only apparent effect of a smaller size being a slightly higher fraction of the smallest (dimer and trimer) chain fragments. This is consistent with the essentially constant Q^n distribution recently found for sodium silicate glass models containing up to one million atoms (Fig. 3).³⁹

[†] For instance, a total of 270 ns, or >1.3 billion MD time steps (a typical time step is 2×10^{-16} s in simulations performed with a shell model potential³⁰) would be 55 needed to cool down to room temperature a melt pre-equilibrated at 3000 K, as done in common MD procedures.



Fig. 3 Q^n distribution for several sodium silicate glasses, obtained from samples of different sizes, from 3000 to 10^6 atoms. Reprinted from ref. 39, with permission from Elsevier.

- 20 The important message, then, is that medium-range structural features extracted from models of silicate and phosphate glasses obtained through conventional MD setups⁴⁰⁻⁴³ are generally reliable, so that CPU-demanding procedures to access more challenging conditions are in most cases unnecessary.
- 25 Essentially, the unrealistic (compared to practical synthesis) MD conditions employed to obtain the models do not appear to have a negative impact on the accuracy of the main structural features that affect the glass durability, and then on key conclusions based on the analysis of those features. It should
- 30 be noted, however, that the different performances of polarisable vs. non-polarisable force fields (with the former generally leading to a more accurate description of the Q^n speciation^{34,44}) means that the convergence of medium-range structural features should ideally be evaluated on a case-by-case basis, 35 especially for non-polarisable force fields whose convergence could be slower.

Dynamical properties

- 40 MD simulations are naturally suited to follow dynamical processes in condensed phases, and many MD studies have indeed investigated the migration of modifier ions in silicate and phosphate glasses.⁴⁵⁻⁵⁰ Not as many studies, however, have concerned ion migration in bioactive compositions. Even though some general features identified for conventional, bio-inactive glasses could also describe the migration of network-modifier cations in bioactive compositions, the same structural peculiarities that enhance the biological response of these glasses can also determine a different dynamical behaviour
- ⁵⁰ for an ion moving in the bioactive matrix, and this different behaviour can in turn further affect the activity of the biomaterial. For instance, it is well known that the fast dissolution of highly bioactive compositions such as 45S5 Bioglass[®] arises from a highly fragmented, open silicate backbone.⁷ It turns out
- 55 that this fragmentation enables ion migration pathways not favourable in the denser network of common higher-silica



Fig. 4 3D traces of the MD trajectory of key Na and Ca ions, illustrating the migration of an individual Na⁺ (red) and the correlated displacements of several other ions (Na, yellow and Ca, cyan). Reprinted with permission from ref. 24, Copyright 2010 American Institute of Physics.

(bio-inactive) glasses: recent simulations²⁴ have highlighted how an ion migrating in the fragmented 45S5 matrix can travel through vacant transient sites created by temporary displacements of another Na or a Ca cation (Fig. 4). The formation of these temporary sites, even if still possible, would not be as favourable in the more rigid network of a higher-silica common glass.⁴⁸

The main challenge to be faced in order to apply standard MD approaches to model ion migration in bioactive glasses has to do with the infrequent nature of the hopping events that compose the diffusive process. The "slow" (relative to typical MD time scales) character of ion migration in glasses entails that prohibitively long trajectories would be needed in order to gain a reasonably accurate sampling of the diffusive event at room temperature. Most MD studies of diffusion in glasses of biomedical interest to date have adopted an effective strategy to cope with this problem, wherein the simulations are run at a high temperature, below the glass transition.^{24,51,52} This approach should ensure (although this condition must be directly verified in each case) that the modifier ions move in a static silicate/ phosphate network whose average configuration and energy landscape match the ones stable at room temperature, so that the description of the diffusive phenomenon at the higher temperature is still representative of practical conditions.

Another potential difficulty is represented by the possible inadequacy of force fields employed in classical MD runs: being normally parameterized by fitting structural and (less frequently) elastic properties of crystalline phases related to the target glass, a potential that provides a good description of the glass structure does not necessarily perform equally well in the reproduction of diffusive processes. Whereas classical potentials have been employed with good results to model diffusive processes in bioglasses,^{51,52} a safer solution would undoubtedly be represented by parameter-free *ab initio* MD (AIMD) approaches.⁵³ The higher computational demands of the latter, however, limit the AIMD trajectory length to below the nanosecond range, with the consequence that a straightforward investigation of the migration of slow-moving cations is complicated, even with the highertemperature strategy described above. For instance, using AIMD it has been possible to characterise sodium migration in 45S5 Bioglass[®] (Fig. 4), but not enough calcium migration events

PCCP

1

5

20

25

30

35

40

45

50

PCCP

¹ were observed during the simulation time to yield an equally clear picture of the (slower) diffusive process of Ca.²⁴

A more rigorous approach for tackling the timescale problem affecting MD simulations of migration in glasses could be

- ⁵ represented by enhanced-sampling methods.^{54–56} These have been developed to accelerate the sampling of processes that, due to high energy barriers or for other reasons, proceed too slowly in configuration space to provide accurate statistics over typical MD runs. Whereas the application of these methods to
- 10 study ion migration in crystalline solids, often characterised by well-defined deep energy minima separated by high barriers, does not present particular difficulties,^{57–59} the application of AIMD combined with enhanced-sampling to the more complex energy landscapes experienced by ions migrating in multi-
- 15 component bioactive glasses is less straightforward, and represents an intriguing challenge for the future.

Reduced size

20

Several recent examples show the reliability of MD simulations for modelling crystalline and amorphous nanoparticles (NPs).⁶⁰⁻⁶² The number of atoms contained, for instance, in an isolated BG nanoparticle of 5–15 nm is of the order of 10⁴–10⁵,
a manageable size for classical MD simulations. The latter can then provide, in a relatively straightforward way, an atomistic-resolution picture of the actual nanosized substrates⁶³ (Fig. 5). A suitable computational procedure in this case involves fast

- quenching a liquid mixture constrained in an isolated sphere of
 the desired size, roughly replicating the flame synthesis used to
 prepare small BG nanoparticles in a high-temperature environment.⁶⁴ Models of a 45S5 particle obtained in this way recently
 provided some preliminary indications about which structural and dynamical effects of the reduced size can be relevant for the
 biological behaviour.⁶⁰ It turns out that some of the key properties of bioactive glasses most beneficial for their bioactive behaviour
- are further enhanced when the size of the glass substrate is



Fig. 5 Models of 45S5 Bioglass[®] nanoparticles of 3–10 nm size, obtained
 using Molecular Dynamics.⁶⁰ The largest particle contains ~40 000 atoms.



Fig. 6 Silicon speciation in the external region of 45S5 nanoparticles, compared to the flat surface and to the bulk structure. Adapted from ref. 60 with permission from The Royal Society of Chemistry.

reduced: the high fragmentation of the silicate network further decreases on the surface of a 45S5 nanoparticle, compared not only to the bulk glass but, most importantly, to the virtually flat surface of a corresponding larger glass substrate (Fig. 6). Moreover, the mobility of modifier cations and the density of three-membered silicate rings – key features to support rapid dissolution and bone bonding processes at the surface⁶⁵ – are also enhanced at the nanoparticle surface compared to samples of larger size.

Whereas models of dry nanoparticles are the necessary starting point to gather information on these systems, they do not take into account the additional perturbation induced by the surrounding fluids that come into contact with the particle in a biological environment. An important future step thus must involve modelling of the explicit interface between the nanoparticle and a suitable aqueous medium, and assess the effects of this interaction on the properties discussed above. The main challenges that will have to be faced concern: (i) the need of accurate force fields to model the additional interactions at the biomaterial interface (the considerable size of the models prevents the straightforward application of AIMD approaches in this case); (ii) the significantly increased computational demands of the simulations of solvated systems compared to the dry cases, especially for the largest NPs.

Conclusions

The direct observation, through computer simulations, of the chemical processes that follow the implantation of a biomaterial in a biological environment is complicated by the characteristic space and time scales that rule these phenomena. In principle, a process such as the biodegradation of a bioactive glass scaffold for tissue engineering can only be explicitly modelled through very large-scale (coarse-grained, mesoscale and Finite Elements) numerical approaches.^{66–68} The coarsegrained nature of all these approaches, while allowing one to reach more realistic space and time scales, averages out the chemical details of the biomaterial and its biointerfaces, such

Perspective

25

20

15

30

40

35

45

50

Perspective

- 1 as short- and medium-range structural features and elementary dynamical (ion migration and reaction) steps. The latter aspects can only be modelled through the higher resolution of atomistic simulations. These atomic-level details provide the foundation
- for a rational design of the core biomedical materials, whose 5 performances can be understood and optimised based on the insight extracted from the atomistic models. While there is some important degree of complementarity between very different information provided by the macroscopic and atomic-scale
- 10 methods, an important future challenge will involve filling the substantial gap that separates them, for instance by further extending the space and time scales that can be accessed in atomistic simulations, but also in mapping the fundamental information provided by the latter to develop and refine the macroscopic approaches.

15

This is particularly important from the perspective of modelling nanosized biomaterials of practical sizes, and of simulating infrequent dynamical processes such as activated chemical reactions and ion migration on those substrates. The

20 combined application of enhanced-sampling approaches with AIMD on high-end parallel supercomputers represents a very promising tool in this context.

Acknowledgements 25

Financial support (University Research Fellowship) from the Royal Society is gratefully acknowledged. The author would also like to thank Alastair Cormack for useful discussions.

30

35

40

45

Notes and references

- 1 L. L. Hench, J. Am. Ceram. Soc., 1998, 81, 1705-1728.
- 2 I. D. Xynos, A. J. Edgar, L. D. K. Buttery, L. L. Hench and
- J. M. Polak, J. Biomed. Mater. Res., 2001, 55, 151-157.
 - 3 J. R. Jones, Acta Biomater., 2013, 9, 4457-4486.
 - 4 J. M. Smith, R. A. Martin, G. J. Cuello and R. J. Newport, J. Mater. Chem. B, 2013, 1, 1296-1303.
 - 5 R. A. Martin, H. L. Twyman, G. J. Rees, E. R. Barney,
- R. M. Moss, J. M. Smith, R. G. Hill, G. Cibin, T. Charpentier, M. E. Smith, J. V. Hanna and R. J. Newport, J. Mater. Chem., 2012, 22, 22212-22223.
- 6 C. Bonhomme, C. Gervais, N. Folliet, F. Pourpoint, C. C. Diogo, J. Lao, E. Jallot, J. Lacroix, J.-M. Nedelec,
- D. Iuga, J. V. Hanna, M. E. Smith, Y. Xiang, J. Du and D. Laurencin, J. Am. Chem. Soc., 2012, 134, 12611-12628.
- 7 A. Tilocca, J. Mater. Chem., 2010, 20, 6848-6858.
- 8 A. Hoppe, N. S. Güldal and A. R. Boccaccini, Biomaterials, 2011, 32, 2757-2774.
- 50 9 A. Tilocca and A. N. Cormack, J. Phys. Chem. B, 2007, 111, 14256-14264.
 - 10 A. Tilocca and A. N. Cormack, Langmuir, 2010, 26, 545-551.
 - 11 E. Berardo, A. Pedone, P. Ugliengo and M. Corno, Langmuir, 2013, 29, 5749-5759.
- 55 12 G. A. Yang, B. Bureau, T. Rouxel, Y. Gueguen, O. Gulbiten, C. Roiland, E. Soignard, J. L. Yarger, J. Troles,

J. C. Sangleboeuf and P. Lucas, Phys. Rev. B: Condens. Matter **Q3** 1 Mater. Phys., 2010, 82.

- 13 M. M. Smedskjaer, J. C. Mauro, J. Kjeldsen and Y. Z. Yue, J. Am. Ceram. Soc., 2013, 96, 1436-1443.
- 14 T. Le, V. C. Epa, F. R. Burden and D. A. Winkler, Chem. Rev., 2012, 112, 2889-2919.
- 15 C. Duée, F. Désanglois, I. Lebecq and C. Follet-Houttemane, J. Non-Cryst. Solids, 2012, 358, 1083-1090.
- 16 Z. Strnad, Biomaterials, 1992, 13, 317-321.
- 17 D. Arcos, D. C. Greenspan and M. Vallet-Regi, Chem. Mater., 10 2002, 14, 1515-1522.
- 18 Q. Zheng, M. M. Smedskjaer, R. E. Youngman, M. Potuzak, J. C. Mauro and Y. Yue, Appl. Phys. Lett., 2012, 101.
- 19 J. K. Christie and A. Tilocca, J. Phys. Chem. B, 2012, 116, 12614-12620.
- 20 A. Rimola and P. Ugliengo, J. Chem. Phys., 2008, 128.
- 21 A. Tilocca and A. N. Cormack, J. Phys. Chem. C, 2008, 112, 11936-11945.
- 22 J. K. Christie, A. Pedone, M. C. Menziani and A. Tilocca, J. Phys. Chem. B, 2011, 115.
- 23 Y. Xiang and J. Du, Chem. Mater., 2011, 23, 2703-2717.
- 24 A. Tilocca, J. Chem. Phys., 2010, 133, 014701-014710.
- 25 F. Quintero, J. Pou, R. Comesana, F. Lusquinos, A. Riveiro, A. B. Mann, R. G. Hill, Z. Y. Wu and J. R. Jones, Adv. Funct. Mater., 2009, 19, 3084-3090.
- 26 S. Labbaf, O. Tsigkou, K. H. Mueller, M. M. Stevens, A. E. Porter and J. R. Jones, Biomaterials, 2011, 32, 1010-1018.
- 27 M. Vollenweider, T. J. Brunner, S. Knecht, R. N. Grass, M. Zehnder, T. Imfeld and W. J. Stark, Acta Biomater., 30 2007, 3, 936-943.
- 28 S. K. Misra, D. Mohn, T. J. Brunner, W. J. Stark, S. E. Philip, I. Roy, V. Salih, J. C. Knowles and A. R. Boccaccini, Biomaterials, 2008, 29, 1750-1761.
- 29 J. K. Christie and A. Tilocca, J. Mater. Chem, 2012, 22, 35 12023-12031.
- 30 A. Tilocca, in Nanotechnology in Regenerative Medicine, Methods in Molecular Biology, ed. J. Navarro and J. A. Planell, Humana Press Springer, USA, 2012, vol. 811, p. 285.
- 31 A. Takada and A. N. Cormack, Phys. Chem. Glasses, 2008, 49, 40 127-135.
- 32 L. B. Skinner, A. C. Barnes, P. S. Salmon, H. E. Fischer, J. W. E. Drewitt and V. Honkimaki, Phys. Rev. B: Condens. Matter Mater. Phys., 2012, 85.
- 33 M. Schuch, R. Christensen, C. Trott, P. Maass and 45 S. W. Martin, J. Phys. Chem. C, 2011, 116, 1503-1511.
- 34 A. Tilocca, J. Chem. Phys., 2008, 129, 084504.
- 35 E. M. Erbe and D. E. Day, J. Biomed. Mater. Res., 1993, 27, 1301-1308.
- 36 Y. Vaills, Y. Luspin and G. Hauret, Mater. Sci. Eng., B, 1996, 50 40, 199-202.
- 37 J. K. Christie, J. Malik and A. Tilocca, Phys. Chem. Chem. Phys., 2011, 13, 17749-17755.
- 38 A. Tilocca, J. Chem. Phys., 2013, 139, 114501.
- 39 L. Adkins and A. Cormack, J. Non-Cryst. Solids, 2011, 357, 55 2538-2541.

5

15

20

Phys. Chem. Chem. Phys., 2014, 00, 1-7 | 7

1

5

20

25

30

35

40

45

50

55

- 40 K. Vollmayr, W. Kob and K. Binder, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 1996, **54**, 15808–15827.
- 41 J. C. Du, L. Kokou, J. L. Rygel, Y. S. Chen, C. G. Pantano, R. Woodman and J. Belcher, *J. Am. Ceram. Soc.*, 2011, 94, 2393–2401.
- 42 G. Malavasi, A. Pedone and M. C. Menziani, *J. Phys. Chem. B*, 2013, **117**, 4142–4150.
- 43 R. A. Martin, G. Mountjoy and R. J. Newport, *J. Phys.:* Condens. Matter, 2009, 21, 075102.
- 10 44 J. Malik and A. Tilocca, J. Phys. Chem. B, 2013, 117, 14518–14528.
 - 45 M. Vogel, Phys. Rev. B: Condens. Matter Mater. Phys., 2003, 68.
 - 46 J. Habasaki and K. L. Ngai, *Phys. Chem. Chem. Phys.*, 2007, 9, 4673–4689.
- 15 47 A. Heuer, M. Kunow, M. Vogel and R. D. Banhatti, *Phys. Chem. Chem. Phys.*, 2002, 4, 3185–3192.
 - 48 A. N. Cormack, J. Du and T. R. Zeitler, *Phys. Chem. Chem. Phys.*, 2002, 4, 3193–3197.
 - 49 H. Lammert, M. Kunow and A. Heuer, Phys. Rev. Lett., 2003,
- 20

90

- 50 W. Smith, T. R. Forester, G. N. Greaves, S. Hayter and M. J. Gillan, *J. Mater. Chem.*, 1997, 7, 331–336.
- 51 J. Du and Y. Xiang, J. Non-Cryst. Solids, 2012, 358, 1059–1071.
- 52 A. Pedone, G. Malavasi, M. C. Menziani, U. Segre and
- A. N. Cormack, J. Phys. Chem. C, 2008, 112, 11034-11041.
- 53 R. Car and M. Parrinello, *Phys. Rev. Lett.*, 1985, 55, 2471–2474.

- 54 A. Laio and M. Parrinello, PNAS, 2002, 99, 12562-12566.
- 55 M. Cogoni, B. P. Uberuaga, A. F. Voter and L. Colombo, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 2005, **71**.
- 56 Y. Kanai, A. Tilocca, A. Selloni and R. Car, *J. Chem. Phys.*, 2004, **121**, 3359–3367.
- 57 K. Jug, N. N. Nair and T. Bredow, *Phys. Chem. Chem. Phys.*, 2005, 7, 2616–2621.
- 58 U. Aschauer, P. Bowen and S. C. Parker, *Acta Mater.*, 2009, 57, 4765–4772.
- 59 H. Cheng and A. Selloni, *Phys. Rev. B: Condens. Matter Mater.* 10 *Phys.*, 2009, **79**.
- 60 A. Tilocca, J. Mater. Chem., 2011, 21, 12660-12667.
- 61 N. N. Linh and V. Van Hoang, J. Phys.: Condens. Matter, 2008, 20.
- 62 D. Spagnoli, J. P. Allen and S. C. Parker, *Langmuir*, 2011, 27, 15 1821–1829.
- 63 S. Lin, C. Ionescu, K. J. Pike, M. E. Smith and J. R. Jones, *J. Mater. Chem.*, 2009, **19**, 1276–1282.
- 64 T. J. Brunner, R. N. Grass and W. J. Stark, *Chem. Commun.*, 2006, 1384–1386.
- 65 N. Sahai and M. Anseau, *Biomaterials*, 2005, 26, 5763-5770.
- 66 J. A. Sanz-Herrera and A. R. Boccaccini, *Int. J. Solids Struct.*, 2011, 48, 257–268.
- 67 S. O. Nielsen, C. F. Lopez, G. Srinivas and M. L. Klein, J. Phys.: Condens. Matter, 2004, 16, R481.
- 68 J. L. Suter, R. L. Anderson, H. C. Greenwell and P. V. Coveney, *J. Mater. Chem.*, 2009, **19**, 2482–2493.

30

35

40

45

50

25

1

5

This journal is © the Owner Societies 2014