

ChemComm

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

Early amyloid β -protein aggregation precedes conformational change[†]

Bogdan Barz,^a Olujide O. Olubiye,^a and Birgit Strodel^{a,b,*}

Received Xth XXXXXXXXXX 20XX, Accepted Xth XXXXXXXXXX 20XX

First published on the web Xth XXXXXXXXXX 200X

DOI: 10.1039/b000000x

The aggregation of amyloid- β protein (1-42) is studied at experimental concentrations using all-atom molecular dynamics simulations. We observe a fast aggregation into oligomers without significant changes in the internal structure of individual proteins. The aggregation process is characterized in terms of transition networks.

Oligomers of amyloid- β protein (A β) are considered one of the main causes of neurotoxicity and are thus highly associated with the onset of Alzheimer's disease.¹ Experimental methods are able to identify some characteristics of aggregating proteins such as the oligomer size distribution or cellular toxicity,² but due to the fast conversion of oligomers into fibrils, the elucidation of their structure at molecular level is challenging. Computational methods have the advantage of atomistic detail but generally lack the size and time scales available in experiment. While there is a plethora of molecular dynamics (MD) simulations describing the structure of the full-length A β monomers, dimers and small oligomers at both atomistic³⁻⁶ and coarse grained level,⁶⁻⁸ the aggregation from a large number of spatially separated monomers has mainly been studied by coarse-grained simulations with implicit solvent.⁹ The studies of monomers and dimers are very insightful, but the structure of small A β oligomers observed in experiments during early aggregation might be considerably different due to their interaction with other oligomeric species. In addition, the structure of oligomers could also be influenced by the solute concentration due to different lifetimes of oligomeric species at different concentrations. Generally, in computational studies the solute concentration is one or even two orders of magnitude higher than in experiments, which will influence the aggregation process. Here we report the early assembly of A β 42 proteins at experimental concentrations using all-atom MD simulations in implicit solvent,

which were initiated from 20 isolated A β 42 monomers.¹⁰ To describe the assembly process we derived a maximum flow transition network (MTN) based on aggregation states defined by $N1|N2|N3$, where $N1$ represents the oligomeric size, $N2$ is the average number of hydrogen-bonds between individual chains from the oligomer, and $N3$ is the average number of amino acids in β -strand conformation per peptide in the oligomer. Detailed information about the methods are provided in the supplement (ESI[†]text).

The MTN in Fig. 1 displays a complex aggregation process in which initial monomers assemble into oligomers up to 18-mers during 200 ns. The aggregation states with $N1 = 1$ are distributed linearly with a gradual increase in β -strand content from right to left. The monomers with more β -strands more readily aggregate than the others: state 1|0|17 is the main connection node to oligomers 2|2|15, 3|4|16, 4|6|13, 5|6|17, 12|7|13 and 17|8|13. These states are also the central connection to the other states with the same $N1$ value. In addition, state 1|0|20 is in direct contact with state 18|9|13 and thus to the rest of the 18-mer cluster. A representative snapshot of the 18-mer at the end of the simulation indicates an elongated conformation rather than a globular one as observed for the 8-mer (Fig. 1). The central aggregation states for hexamer and heptamers are 6|9|15 and 7|7|16, respectively, which are preferentially formed from trimers. Dimers are directly connected to aggregation state 8|7|12 and thus to the other $N1 = 8$ states, indicating that octamers are largely formed by the addition of dimers to either tetramers or hexamers. The oligomer mass distribution in Fig. 2a reveals a higher population for dimers, tetramers, hexamers, octamers, 12-mers and 18-mers. Previous computational⁹ and experimental studies² of A β aggregation report significant peaks in pentamers/hexamers and 12/13-mers, in agreement with our results.

To describe structural changes during the assembly process we derived the time evolution of the secondary structure propensities (Fig. 2b). The initial β -strand propensity ($\sim 10\%$) decreases slightly throughout the simulation and has an average of $7.6 \pm 2.2\%$. The average helical propensity is around $10.1 \pm 4.0\%$ and the average coil propensity starts around 80% and increases slightly, having an average value of

^a Forschungszentrum Jülich GmbH, Institute of Complex Systems: Structural Biochemistry (ICS-6), 52425 Jülich, Germany Fax: +49 2461 618766; Tel: +49 2461 613670; E-mail: b.strodel@fz-juelich.de

^b Institute of Theoretical and Computational Chemistry, Heinrich Heine University Düsseldorf, 40225 Düsseldorf, Germany

[†] Electronic Supplementary Information (ESI) available: Methods Section, and additional figures. See DOI: 10.1039/b000000x/

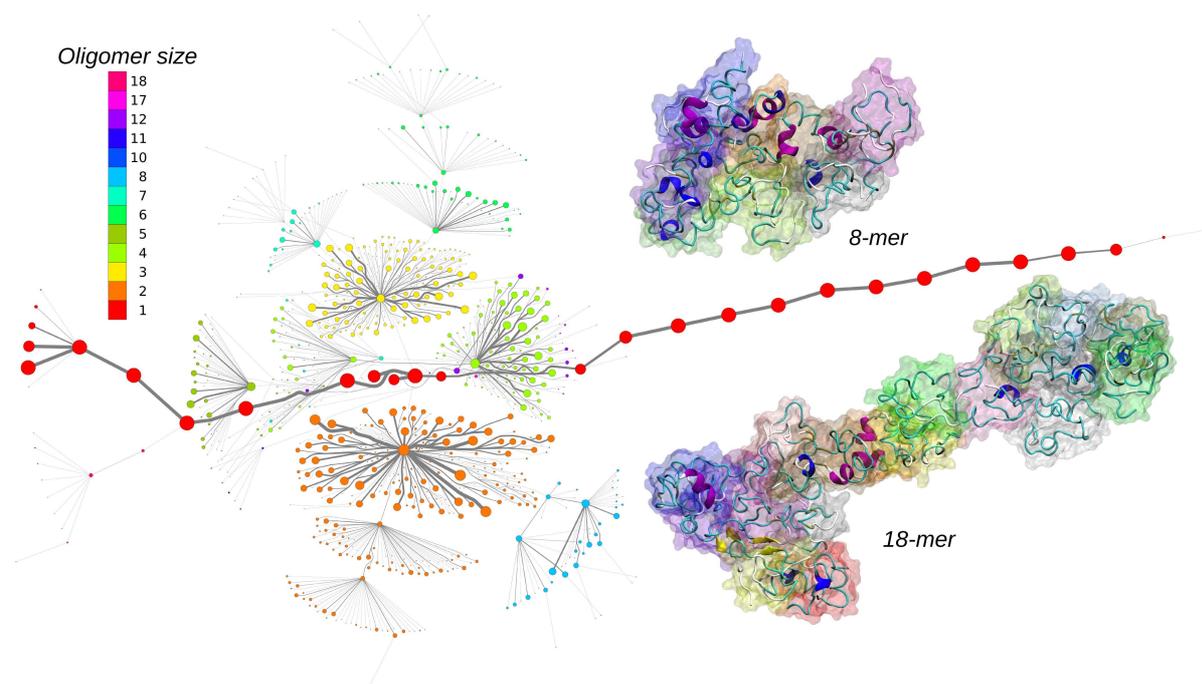


Fig. 1 Maximum flow transition network. The node area is proportional to the state population and the color corresponds to the oligomer size. The edge thickness correlates with the number of transitions. Two representative oligomer structures for the octamer and 18-mer are shown in cartoon and surface representation.

82.2±4.3%. This indicates little change in the overall secondary structure. Coarse-grained computational studies of A β aggregation showed an increase in the β -strand propensity upon dimer formation from ~10% to ~15%.⁹ However, when converted to all-atom models, these values change to 6–7% for both monomers and dimers.⁵ Experimental studies² indicate 13–20% β -sheet for A β 42 and A β 40 while a more recent study shows a jump from ~25% to ~45% when converting from monomers to tetramers of A β 40.¹¹ Another experimental study reports A β 42 oligomers without β -sheet structure, in close agreement with our findings, which are on-pathway intermediates for fibril formation.¹² It is important to mention the high turn propensity of residues V36 and G37 (67.1±0.1% and 50.5±0.1%, respectively), which has been recently shown to play an important role in the increased toxicity of A β 42 relative to A β 40 oligomers.¹³ In addition to the secondary structure analysis we calculated the RMSD of individual monomers throughout the simulation with respect to their conformations at the end of the simulation (Fig. S1, ESI \dagger). This quantity reveals that besides an initial change in the monomeric conformation most of the proteins were quite stable following oligomerization with RMSD values below 0.5 nm. This result in combination with the small changes in the secondary structure suggest an aggregation process with few structural changes of individual peptides due to oligomer formation. To

investigate the key amino acids involved in the assembly process we calculated contact maps for the interface between any two proteins that are part of an oligomer (Fig. 2c). An important result from the inter-molecular contact map is the proximity of hydrophobic regions from the C-terminus including L31–A42. In addition, region L33–V35 is in close proximity to region L17–F20. On the basis of solid-state NMR spectroscopy it has been shown that the C-terminal region is buried inside disc-shaped oligomers (pentamers and 10-mers) with strong contacts between F19 and L34.¹² The 8-mer shown in Fig. 1 matches the description of the low-order oligomers in non- β -sheet secondary structure observed by Ahmed et al.¹² The increased interaction between the C-termini observed in Fig. 2c is in agreement with the study of Urbanc et al.⁹ who proposed that oligomer formation of A β 42 proceeds through interactions between the C-termini while for A β 40 through interactions between the central hydrophobic regions (L17–A21). In addition, the 18-mer from Fig. 1 resembles the elongated protofibrillar assemblies (28-mer) reported by Urbanc et al. To gain additional information regarding the effect of the amino acids I41–A42 on the aggregation process we performed an additional 200 ns simulation with 20 A β 40 peptides and computed the difference between A β 42 and A β 40 contact maps (Fig. S2, ESI \dagger). We observe that A β 42 forms more contacts between the C-termini than A β 40, and exhibits many

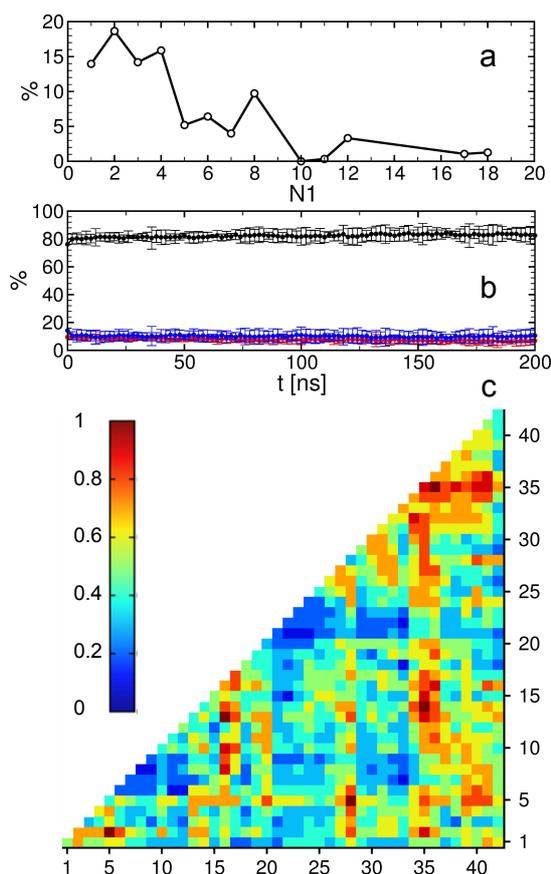


Fig. 2 Analysis of A β oligomers. (a) Relative oligomer mass distribution during aggregation. The probability was determined as the population of $N1$ multiplied by $N1$. (b) Secondary structure propensity as a function of time. Black corresponds to coil, red to β -strand, and blue to helix. (c) Inter-molecular contact map for all oligomers based on a cutoff distance of 0.75 nm. Color coding corresponds to the normalized number of contacts.

interactions between the charged C-terminus at A42 and positively charged residues, which in A β 40 are formed with V40 instead. The extension of A β 42 by I41 and A42 increases the hydrophobicity of V39–I41 compared to A β 40 as the C-terminal charge is shifted by two residues, explaining the increased number of contacts between the C-termini in A β 42. Electrostatic interactions contribute to oligomer formation by strong contacts formed by amino acid K16 with region S8–Q15 containing the negatively charged residue E11, and by K28 with region D1–D7 containing the negatively charged residues D1, E3, and D7.

Our results suggest that early A β aggregation precedes β -sheet formation. This is an important observation considering the low A β concentration in the current study (~ 0.8 mM), close to experimental concentrations (~ 30 μ M–1 mM),² and smaller than concentrations from other computational stud-

ies of 3.4 mM.⁹ At the same time, the structurally stable oligomers and the small changes in their secondary structure indicate a fast assembly process relative to the time scale corresponding to their conformational reorganization. Whether the oligomers observed here during a very early aggregation stage are similar to the toxic species observed in experiments is difficult to assess. While we find similarities with experimentally observed A β 42 oligomers in terms of size and structure,^{9,12} there is still a large debate regarding which A β oligomers are the toxic ones.¹⁴ Some groups consider small oligomers with high content of β -sheet as toxic species,¹¹ others suggest a second nucleation process where amyloid fibrils are present with small oligomer species as the source for toxic oligomers,¹⁵ and others propose that toxic A β oligomers have cross- β structure.¹⁶ In our future studies we plan to follow the further growth and structural conversion of the oligomers obtained here and study their interactions with membranes. The aim should be that experiments probe at the same time the size, secondary structure and toxicity of low-order oligomers, allowing to directly relate simulation and experimental results.

References

- 1 W. L. Klein, G. A. Krafft and C. E. Finch, *Trends Neurosci.*, 2001, **24**, 219–224.
- 2 G. Bitan, M. D. Kirkitadze, A. Lomakin, S. S. Vollers, G. B. Benedek and D. B. Teplow, *Proc. Natl. Acad. Sci. USA*, 2003, **100**, 330–335.
- 3 A. Baumketner, S. L. Bernstein, T. Wyttenbach, G. Bitan, D. B. Teplow, M. T. Bowers and J.-E. Shea, *Protein Sci.*, 2006, **15**, 420–428.
- 4 T. Takeda and D. Klimov, *J. Phys. Chem. B*, 2009, **113**, 6692–6702.
- 5 B. Barz and B. Urbanc, *PLoS ONE*, 2012, **7**, e34345.
- 6 B. Urbanc, L. Cruz, D. B. Teplow and H. E. Stanley, *Curr. Alzheimer Res.*, 2006, **3**, 493–504.
- 7 S. Ct. R. Laghaei, P. Derreumaux and N. Mousseau, *J. Phys. Chem. B*, 2012, **116**, 4043–4055.
- 8 A. Melquiond, X. Dong, N. Mousseau and P. Derreumaux, *Curr. Alzheimer Res.*, 2008, **5**, 244–250.
- 9 B. Urbanc, M. Betnel, L. Cruz, G. Bitan and D. B. Teplow, *J. Am. Chem. Soc.*, 2010, **132**, 4266–4280.
- 10 O. O. Olubiyi and B. Strodel, *J. Phys. Chem. B*, 2012, **116**, 3280–3291.
- 11 K. Ono, M. M. Condron and D. B. Teplow, *Proc. Natl. Acad. Sci. USA*, 2009, **106**, 14745–14750.
- 12 M. Ahmed, J. Davis, D. Aucoin, T. Sato, S. Ahuja, S. Aimoto, J. I. Elliott, W. E. Van Nostrand and S. O. Smith, *Nat. Struct. Mol. Biol.*, 2010, **17**, 561–567.
- 13 R. Roychaudhuri, M. Yang, A. Deshpande, G. M. Cole, S. Frautschy, A. Lomakin, G. B. Benedek and D. B. Teplow, *J. Mol. Biol.*, 2013, **425**, 292–308.
- 14 I. Benilova, E. Karran and B. De Strooper, *Nat. Neurosci.*, 2012, **15**, 349–357.
- 15 S. I. A. Cohen, S. Linse, L. M. Luheshi, E. Hellstrand, D. A. White, L. Rajah, D. E. Otzen, M. Vendruscolo, C. M. Dobson and T. P. J. Knowles, *Proc. Natl. Acad. Sci. USA*, 2013, 201218402.
- 16 J. C. Stroud, C. Liu, P. K. Teng and D. Eisenberg, *Proc. Natl. Acad. Sci. USA*, 2012, **109**, 7717–7722.