

Analytical Methods

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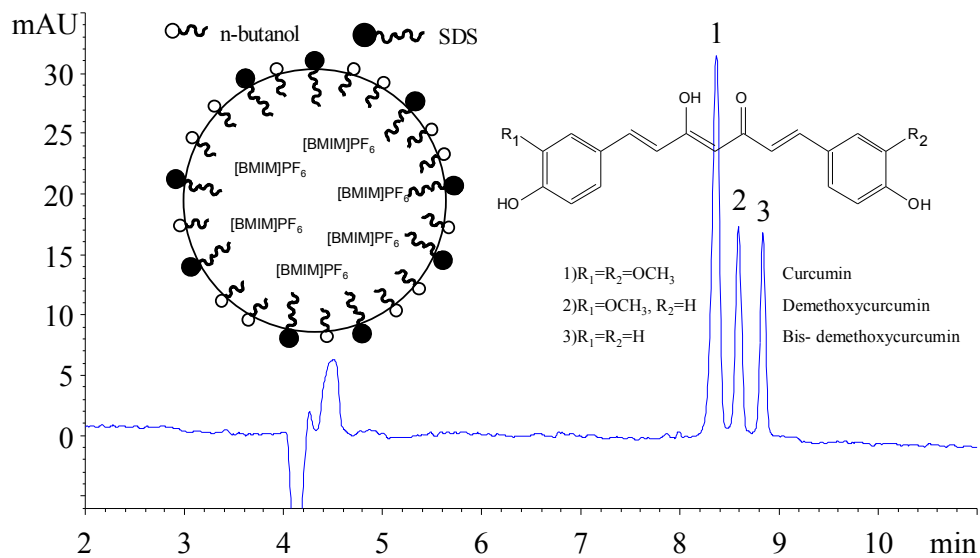
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Graphical abstract



**Determination of three curcuminoids in *Curcuma longa* by
microemulsion electrokinetic chromatography with protective
effect on the analytes**

Feng Li, Rui Liu, Fengqing Yang*, Wen Xiao, Cheng Chen, Zhining Xia
School of Chemistry and Chemical Engineering, Chongqing University,
Chongqing 400030, China

*Author to whom correspondence should be addressed:

Asso. Prof. Feng-Qing Yang

Tel/Fax: +86-23-6510-6615; E-mail: fengqingyang@cqu.edu.cn

1 Abstract

2 Alkaline degradation of curcuminoids was observed when they were exposed to
3 alkaline solution, which are usually occur during capillary electrophoresis (CE)
4 analysis with alkaline buffers. In the present study, the stability of curcuminoids
5 (curcumin, demethoxycurcumin, bis-demethoxycurcumin) in different solutions
6 (boric acid buffer, microemulsion with *n*-octane as oil phase+boric acid buffer,
7 microemulsion with ionic liquid [BMIM]PF₆ as oil phase+boric acid buffer) were
8 investigated. The results indicated that microemulsion systems have significant
9 protective effect on the analytes even under alkaline conditions as compared to
10 boric acid buffer only, while microemulsion with ionic liquid showed the best
11 protecting effect. Therefore, a stable and rapid microemulsion electrokinetic
12 chromatography (MEEKC) method with ionic liquid as oil phase was developed
13 for simultaneous determination of the three curcuminoids in rhizome (*Jianghuang*)
14 and tuberous root (*Yujin*) of *Curcuma longa*. Experimental parameters including the
15 microemulsion compositions (surfactant, co-surfactant and oil phase), pH,
16 concentration of borate buffer and capillary temperature were intensively
17 investigated. Finally, the investigated curcuminoids were well separated within 9
18 min using the running buffer containing 100 mM SDS, 1.6 M *n*-butanol, 20 mM
19 [BMIM]PF₆ in 10 mM borate buffer of pH 10.2. The developed method was fully
20 validated (LOD, LOQ, intra-, inter-day precision and recovery) and successfully
21 applied to determine the contents of investigated curcuminoids in four real
22 samples (*Yujin* and *Jianghuang* from Chengdu and Leshan). All of the results
23 showed that the MEEKC with ionic liquid as oil phase should be an ideal choice for
24 the analysis of curcuminoids by CE.

25

1 Introduction

2 Capillary electrophoresis (CE) has been widely applied to analyze curcuminoids
3 using different modes including capillary zone electrophoresis (CZE) [1-3],
4 micellar electrokinetic chromatography (MEKC) [4], microemulsion electrokinetic
5 chromatography (MEEKC) [5]. However, alkaline degradation of curcuminoids
6 was observed when they were exposed to alkaline solutions [6-8], which were
7 usually used in CZE and MEKC analysis. Thumnoon Nhujak, et al [5] analyzed
8 those curcuminoids by MEEKC with acidic buffer (pH 2.5), the alkaline
9 degradation could be avoided to some extent, but the solubility of curcuminoids is
10 very low under acidic and neutral solutions [5, 9]. Therefore, to develop a MEEKC
11 method with protective effect on the curcuminoids under alkaline conditions
12 should be of interest for analyzing those compounds.

13 Ionic liquids are molten salts which have strong dissolution ability for most
14 organic and inorganic compounds, with very low volatility and high thermal
15 stability and have been testified that they are more environmentally attractive than
16 organic solvents [10-11]. Actually, ionic liquids have been used as the constituents
17 (surfactant, co-surfactant or oil phase) to prepare microemulsion [12-17]. In this
18 study, the stability of curcuminoids in different solutions [boric acid buffer,
19 microemulsion with *n*-octane as oil phase+boric acid buffer, microemulsion with
20 ionic liquid ([BMIM]PF₆) as oil phase+boric acid buffer] were preliminarily
21 investigated by measuring the UV absorbance at 420 nm (Fig.1). Compared with
22 the background electrolyte, the microemulsion could significantly suppress the
23 alkaline degradation of curcuminoids. The protective effect of microemulsion may
24 due to the inclusion of analytes into the oil phase, which is similar to that of
25 cyclodextrin [9, 18]. Furthermore, it was reported that the stability of curcuminoids
26 could be greatly increased by the presence of some reducing agents like GSH,

1 ascorbic acid [6, 7]. Therefore, the microemulsion with [BMIM]PF₆ had a better
2 protection effect (even under alkaline conditions, Fig. 1) on curcuminoids than the
3 microemulsion with octane as oil phase, which may due to the reducibility of
4 [BMIM]PF₆ [19-20]. Herein, microemulsion with [BMIM]PF₆ as oil phase is
5 undoubtedly a good choice for MEEKC analysis of curcuminoids.

6 *Curcuma longa* is widely used as a traditional Chinese medicine, natural pigment
7 and spice [21]. Its rhizome (*Jianghuang*) and tuberous root (*Yujin*) are reported to
8 mainly contain three curcuminoids, namely curcumin, demethoxycurcumin and
9 bis-demethoxycurcumin (Fig.2). Curcumin possesses multiple pharmacological
10 activities, including cholesterol-lowering [22], antidiabetic [23], anti-inflammatory
11 [24], anti-oxidant [25], antimicrobial [26], immune activation [27] and
12 antidepressant [28] activity. Demethoxycurcumin is reported to own practically the
13 same activity as curcumin [29], while different from the cholesterol-lowering
14 activity of the former two curcuminoids, bis-demethoxycurcumin is reported to
15 have strong choleric effect [30]. Therefore, analysis of those three curcuminoids
16 in *C. longa* is helpful for its quality control and pharmaceutical studies.

17 In the present study, a MEEKC method with protective effects even under
18 alkaline conditions on curcuminoids was developed by using [BMIM]PF₆ as oil
19 phase, and the developed method was fully validated and applied to determine
20 those curuminoids in *C. longa*.

21 **Materials and methods**

22 **Chemicals and reagents**

23 Sodium Dodecyl Sulfonate (SDS), *n*-octane and *n*-butanol were purchased from
24 Chongqing Chuandong Chemical (Group) Co., Ltd. (Chongqing, China). Sodium
25 hydroxide and boric acid were obtained from Chengdu Kelong Chemical Works
26 (Chengdu, China), 1-Butyl-3-methylimidazolium hexafluorophosphate

1 ([BMIM]PF₆) was purchased from Lanzhou Institute of Chemical Physics (Lanzhou,
2 China). All reagents above were of analytical grade, and reverse osmosis (RO)
3 water used throughout was prepared by an AKWL-IV-16 water purification
4 system (Chengdu Tang's Kangning Science and Technology Development Co., Ltd.,
5 Chengdu, China).

6 Curcumin, demethoxycurcumin and bis-demethoxycurcumin with the purity
7 higher than 99% were purchased from Chengdu Must Bio-technology Co., Ltd.
8 (Chengdu, China). The rhizome (*Jianghuang*) and tuberous root (*Yujin*) samples
9 from the same plant of *C. longa* were collected from Chengdu and Leshan, Sichuan
10 province, respectively. The voucher specimens of *C. longa* were deposited at the
11 Department of Pharmaceutics, School of Chemistry and Chemical Engineering,
12 Chongqing University, Chongqing, China.

13 **Apparatus and procedures**

14 All CE separations were performed on an Agilent 7100 3D CE system (Agilent
15 Technologies, Palo Alto, CA, USA) equipped with a DAD and an Agilent
16 ChemStation software. The uncoated fused-silica capillary (Hebei Yongnian
17 Ruifeng Chromatographic Implements, Hebei, China) was of 50 μm i.d and had a
18 total length of 54 cm (46 cm effective length). All the analyses in the present study
19 were performed on the same capillary. The UV-2450 UV-visible spectrophotometer
20 (Shimadzu UV-2450 spectrophotometer, Tokyo, Japan) was used for measurement
21 of UV absorbance. The KQ-100B ultrasonic cleaner (Kunshan ultrasonic
22 instruments Co., Ltd., Jiangsu, China) was applied for the preparation of
23 microemulsion and samples. A Delta 320 pH meter (Mettler-Toledo Instruments,
24 Shanghai, China) was used for measuring the pH of running buffer.

25 The running buffer was prepared by adding of 100 mM SDS, 1.6 M *n*-butanol
26 and 20 mM [BMIM]PF₆ to 10 mM borate buffer of pH 10.2 (adjusted by 1 M NaOH

1 solution). The mixture was then ultrasonicated for 20 min. The freshly prepared
2 microemulsion running buffer was stood for at least one hour at ambient
3 temperature (about 25 °C) prior to use.

4 Curcumin, demethoxycurcumin and bis-demethoxycurcumin were accurately
5 weighted and dissolved in ethanol to prepare the mixture of standard solutions
6 (about 1.5 mg mL⁻¹), then the solutions were further diluted to different
7 concentrations by adding desired amount of microemulsion. For preparation of
8 sample solution, a desired amount (1.0 g) of crude material powders was extracted
9 with 10 mL absolute ethanol in a 50 mL flask by ultrasonication for 40 min. After
10 extraction, the extract was cooled down to the room temperature (25 °C), and made
11 up the lost weight with absolute ethanol, then centrifuged for 5 min at 4.2×10³ g,
12 and the supernatant was filtered through a 0.45 µm nylon membrane (Auto science
13 instrument Co., Ltd., Tianjing, China) before use.

14 The capillary was flushed daily with 0.1 M sodium hydroxide for 15 min,
15 followed by RO water for 5 min and finally with running buffer for 15 min.
16 Between consecutive analyses, the capillary was rinsed with 0.1 M sodium
17 hydroxide (2 min), then with RO water (2 min), and finally with the running buffer
18 (5 min). The operating conditions were: injection pressure 50 mbar; injection time 3
19 s and detection wavelength at 420 nm.

20 **Results and discussion**

21 **Optimization of separation conditions**

22 **Effect of SDS concentration**

23 It is known that the concentration of SDS has a remarkable effect on the separation
24 achieved in MEEKC for it has effect on the oil droplet charge and size, the level
25 and direction of the EOF, and the level of any ion-pairing of microemulsion
26 droplets with charged solutes [31]. Different concentrations (60, 80, 100, 120, 140,

1 and 160 mM) of SDS were investigated by using resolution (R_s) of curcumin *vs*
2 demethoxycurcumin (R_{s1}), demethoxycurcumin *vs* bis-demethoxycurcumin (R_{s2})
3 and the retention time of bis-demethoxycurcumin (RT_b) as markers (the other
4 conditions were: 1.4 M *n*-butanol, 20 mM [BMIM]PF₆, pH 10.0, 20 mM borate
5 buffer, 20 kV and 25 °C). As shown in Fig.3A, the migration time of analytes
6 increased with the increase of concentration of SDS. When the concentration of
7 SDS was higher than 120 mM, some baseline drift was observed due to the high
8 level of current. Therefore, in order to achieve better separation with good
9 repeatability in a shorter analysis time, 100 mM of SDS was used in the subsequent
10 experiments.

11 **Effect of *n*-butanol concentration**

12 *n*-butanol was reported to stabilize the microemulsion and affect the selectivity of
13 the analysis [32]. Furthermore, the concentration of co-surfactant could change the
14 viscosity of the solution, and therefore affected the EOF. In the present study,
15 different concentrations (1.2, 1.4, 1.6, 1.8 and 2.0 M) of *n*-butanol were investigated
16 (the other conditions were: 100 mM SDS, 20 mM [BMIM]PF₆, pH 10.0, 20 mM
17 borate buffer, 20 kV and 25 °C). As it was shown in Fig.3B, with the concentration
18 of *n*-butanol increased from 1.2 M to 1.6 M, both the resolution of analytes and
19 migration time increased, which may be caused by the increased viscosity of
20 *n*-butanol. While further increase of *n*-butanol concentration led to the decrease of
21 resolution and migration time. The excess *n*-butanol in the aqueous phase might be
22 adsorbed and shield the negative-charged micelle, consequently increased the
23 viscosity of micelle and shortened the migration time [33]. So 1.6 M *n*-butanol was
24 selected for its best separation efficiency ($R_{s1} > 1.35$ and $R_{s2} > 1.56$).

25 **Effect of [BMIM]PF₆ concentration**

26 Different concentrations of [BMIM]PF₆ (0, 10, 20, 30, 40 and 50 mM) were compared
27 (the other conditions were: 100 mM SDS, 1.6 M *n*-butanol, pH 10.0, 20 mM borate

1 buffer, 20 kV and 25 °C). The results (Fig.3C) indicated that the concentrations of
2 [BMIM]PF₆ had not significant effect on the separation of analytes, and the best
3 resolution ($R_{s1} > 1.40$ and $R_{s2} > 1.62$) for analytes can be obtained at the
4 concentration of 20 mM. Furthermore, when the concentration of [BMIM]PF₆
5 increased from 0 mM to 20 mM, the cation of [BMIM]PF₆ could shield the anion of
6 SDS, so the EOF increased and migration time of analytes shortened. However, if
7 the concentration of [BMIM]PF₆ increased from 20 mM to 50 mM, the adsorption
8 effect of cation of [BMIM]PF₆ to capillary wall played the dominant role and led to
9 the prolonging of migration time of analytes. Hence, 20 mM [BMIM]PF₆ was used.

10 **Effect of pH, buffer concentration and temperature**

11 As one of the most important parameters in MEEKC separation, pH is directly
12 related to the degree of solute ionization and EOF velocity [34]. In this study,
13 different pH (9.0, 9.5, 10.0, 10.2, 10.5, 11.0) were evaluated (the other conditions
14 were: 100 mM SDS, 1.6 M *n*-butanol, 20 mM [BMIM]PF₆, 20 mM borate buffer, 20
15 kV and 25 °C). The results in Fig.3D showed that pH played a crucial role on the
16 separation. With the pH increased from 9.0 to 11.0, both R_{s1} and R_{s2} increased
17 accordingly, when the pH reached at 10.2, those three analytes could be baseline
18 separated ($R_{s1} > 1.8$, $R_{s2} > 2.3$). Therefore, pH 10.2 was chosen to achieve
19 acceptable resolution within a short migration time (11.65 min).

20 It has been shown previously that using low-ionic-strength (5 - 10 mM) borate or
21 phosphate buffers generated relatively low currents and high EOF value [16]. In
22 present study, different concentrations (5 - 50 mM) borate buffer were investigated
23 (the other conditions were: 100 mM SDS, 1.6 M *n*-butanol, 20 mM [BMIM]PF₆, pH
24 10.2, 20 kV and 25 °C). The results (not shown) revealed that baseline separation
25 could be achieved only when the concentration of borate buffer was above 10 mM.
26 In general, use of high buffer concentrations should be prevented because the high
27 electric current generated led to low efficiency and poor reproducibility [32]. So 10

1 mM borate buffer was used for its good separation within the shortest analysis
2 time. Furthermore, the effect of temperature was also investigated. The migration
3 time decreased with the temperature increased. Finally 25 °C was used for its best
4 separation efficiency ($R_{s1} > 1.7$, $R_{s2} > 2.1$, $RT_b < 8.9$ min).

5 Based on the optimization, the best conditions for the analysis were: running
6 buffer composed of 100 mM SDS, 1.6 M *n*-butanol, 20 mM [BMIM]PF₆ in 10 mM
7 borate buffer of pH 10.2. Applied voltage was 20 kV and capillary temperature was
8 25 °C.

9 **Method Validation**

10 **Linearity, LOD, and LOQ**

11 Stock solution containing three standard compounds (curcumin,
12 demethoxycurcumin and bis-demethoxycurcumin) was prepared and diluted to
13 appropriate concentrations for the construction of calibration curves. At least five
14 concentrations of the solution were analyzed in triplicates, and then the calibration
15 curves were constructed by plotting the peak area of individual standard versus
16 the concentration of each analyte. The limits of detection (LOD) and quantification
17 (LOQ) under the optimum conditions were determined based on the
18 signal-to-noise (S/N) ratio of 3 and 10 respectively. Table 1 shows the linear
19 regression data, LOD and LOQ.

20 **Precision and recovery**

21 Intra- and inter-day variations were chosen to determine the precision of the
22 developed assay. The known concentrations of the three investigated compounds
23 were tested. For intra-day variability test, the mixed standards solutions were
24 analyzed for six replicates within one day, while for inter-day variability test, the
25 solutions were examined in duplicates for consecutive three days. Variations were
26 expressed by relative standard deviation (RSD).

1 The recovery was performed by adding a known amount of individual
2 standards into a certain amount (0.50 g) of crude material (*Jianghaung* from Leshan)
3 powder. The mixture was extracted and analyzed using the method mentioned
4 above. Three replicates were performed for the test. [Table 2](#) shows the precision
5 and recovery results.

6 **Determination of investigated compounds in four different samples**

7 Under the optimum conditions, the ethanol extracts of the four samples (*Yujin*
8 from Chengdu and Leshan, *Jianghuang* from Chengdu and Leshan) were analyzed.
9 The electrochromatograms are shown in [Fig.4](#). The identifications of compounds
10 were done by comparing their retention time with those of standards, as well as
11 adding the individual standard to the samples. By using the calibration curve of
12 each standard, the contents of investigated compounds in those three TCMs were
13 determined. The quantification results were shown in [Table 3](#). Except for *Yujin*
14 from Chengdu, all the analytes could be detected in other samples, while the
15 content of bis-demethoxycurcumin in *Yujin* from Leshan was under the LOQ.
16 Therefore, the contents of curcuminoids in the rhizome (*Jianghuang*) are
17 significantly higher than in the tuberous root (*Yujin*) of *C. longa*. Furthermore, the
18 content of curcumin was higher than that of demethoxycurcumin and
19 bis-demethoxycurcumin in all samples.

20 **Conclusion**

21 An MEEKC method with protective effect on the analytes was developed to
22 simultaneously determine the contents of curcumin, demethoxycurcumin and
23 bis-demethoxycurcumin in *C. longa*. The curcuminoids are stable in the
24 microemulsion system, which ensure the accuracy of the quantitative results of
25 MEEKC method. Therefore, the developed MEEKC with ionic liquid as oil phase is
26 an ideal method for the analysis of curcuminoids by CE. In addition, the developed

1 MEEKC method may also be applied in analyzing other analytes which are
2 unstable during CZE analysis.

3

4 **Acknowledgement**

5 This work was supported by the National Natural Science Foundation of China
6 (21175159, 21275169 and 81202886), the International Cooperation Project of
7 Ministry of Science and Technology (2010DFA32680).

8

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2 **Legend of Figures:**3 **Fig. 1**

4 The stability tests of three curcuminoids dissolved in 20 mM boric acid (\blacklozenge), 3.3%
5 SDS (w/v), 6.6% *n*-butanol and 0.8% *n*-octane in 20 mM boric acid buffer (\blacksquare), and
6 3.3% SDS (w/v), 6.6% *n*-butanol and 0.8% [BMIM]PF₆ in 20 mM boric acid buffer
7 (\blacktriangle), detected by UV absorbance (420 nm).

8

9 **Fig. 2**

10 Chemical structures of three curcuminoids.

11

12 **Fig. 3**

13 Effects of SDS concentration (A), *n*-butanol concentration (B), [BMIM]PF₆
14 concentration (C) and buffer pH (D) on the *Rs1* (\blacklozenge), *RS2* (\blacksquare) and *RTb* (\blacktriangle)

15

16 **Fig. 4**

17 Electrochromatograms for the mixture of standards (A), ethanol extracts of
18 *Jianghuang* from Chengdu (B), *Yujin* from Leshan (C) and *Jianghuang* from Leshan
19 (D).

20 1, curcumin 2, demethoxycurcumin 3, bis-demethoxycurcumin U, unknown

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Table 1 Linear regression data, LOD and LOQ of three curcuminoids

Analytes	Calibration curves	Linear range ($\mu\text{g mL}^{-1}$)	R ²	LOD ($\mu\text{g mL}^{-1}$)	LOQ ($\mu\text{g mL}^{-1}$)
Curcumin	$y = 331.00x - 4.21$	41.87-670.00	0.9962	9.30	25.04
Demethoxycurcumin	$y = 565.12x + 0.91$	25.63-410.00	0.9961	3.80	12.50
Bis-demethoxycurcumin	$y = 584.75x + 7.45$	25.63-410.00	0.9978	3.20	11.00

3

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Table 2 Intra- and Inter-day precision and recovery of the investigated compounds

Analytes	Intra-day ($n = 6$, RSD %)	Inter-day ($n = 6$, RSD %)	Recovery ($n=3$, %)
Curcumin	3.56	3.97	98.87
Demethoxycurcumin	4.90	5.86	95.54
Bis-demethoxycurcumin	3.45	4.20	103.42

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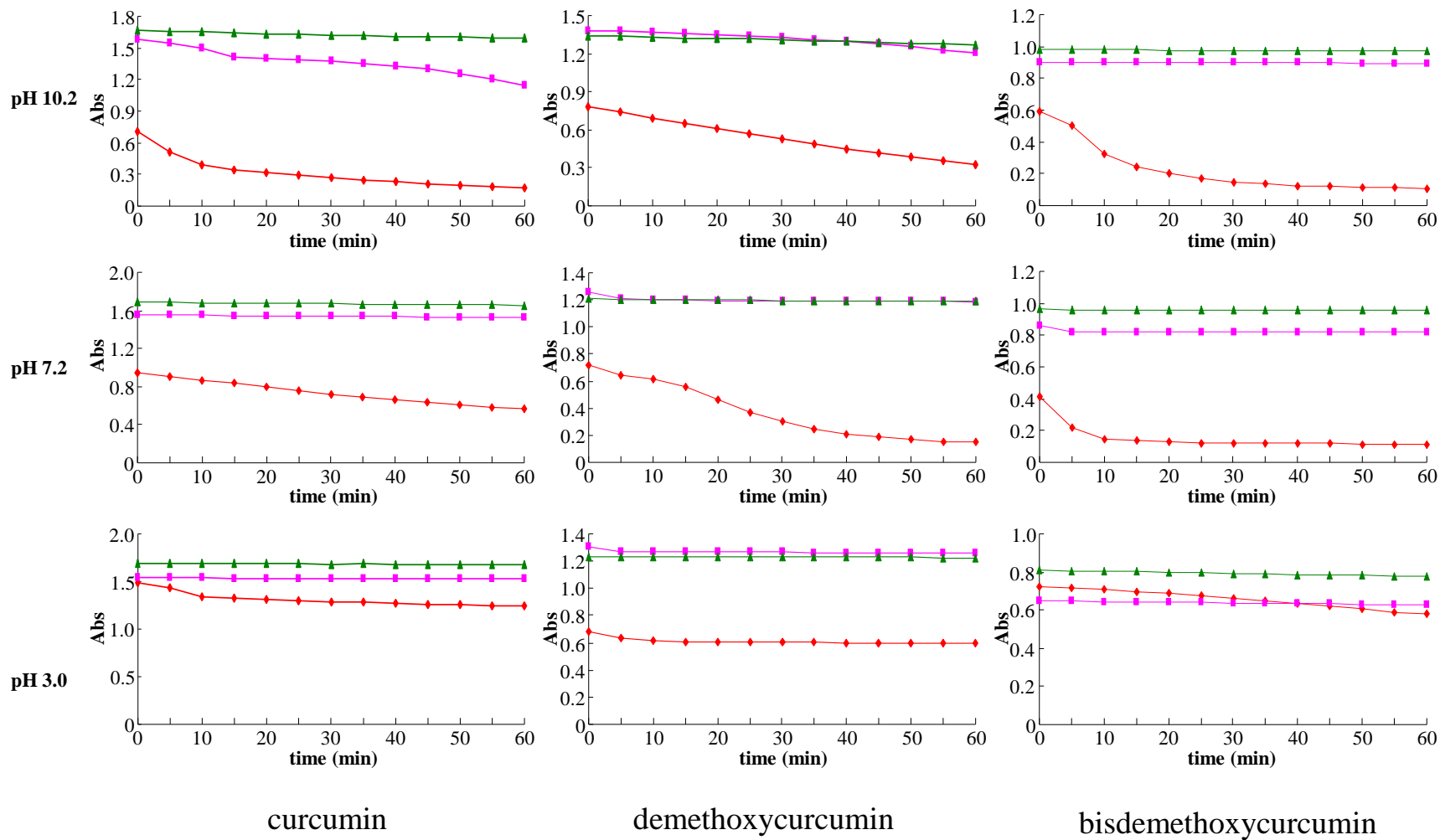
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Table 3 Contents (mg g^{-1}) of investigated compounds in four samples

Analytes	Yujin (CD ^a)	Jianghuang (CD)	Yujin (LS)	Jianghuang (LS)
Curcumin	- ^b	9.52	1.02	7.93
Demethoxycurcumin	-	2.27	0.12	1.78
Bis-demethoxycurcumin	-	2.73	+ ^c	1.80

7

a, CD, Chengdu, LS, Leshan; b, under the limit of detection; c, under the limit of qualification



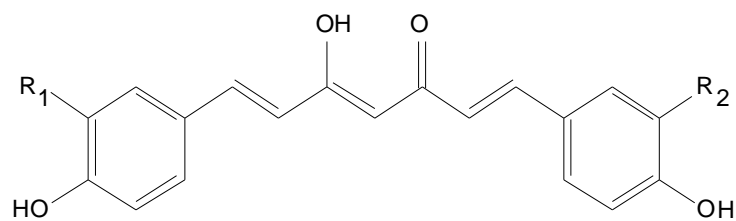
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Fig. 1

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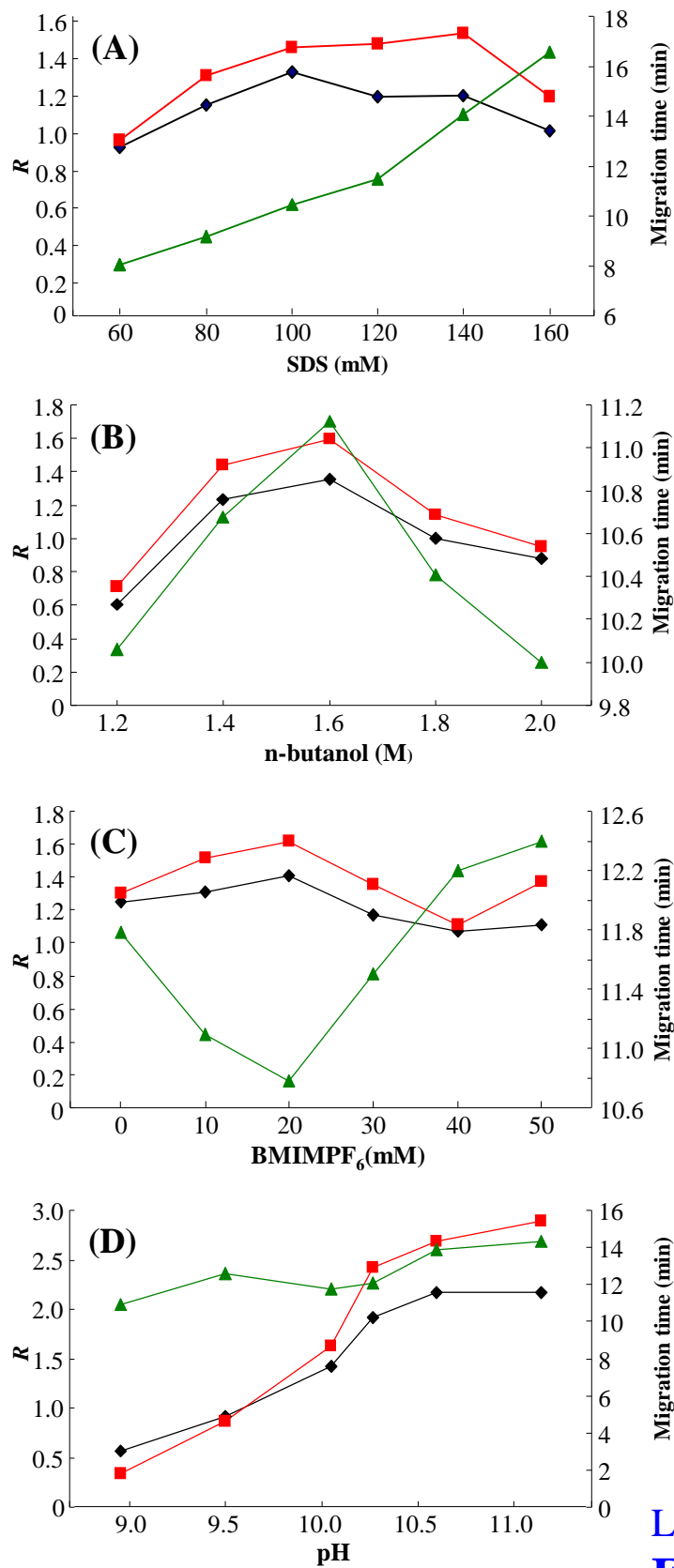


- 1) R₁=R₂=OCH₃ Curcumin
2) R₁=OCH₃, R₂=H Demethoxycurcumin
3) R₁=R₂=H Bis- demethoxycurcumin

4

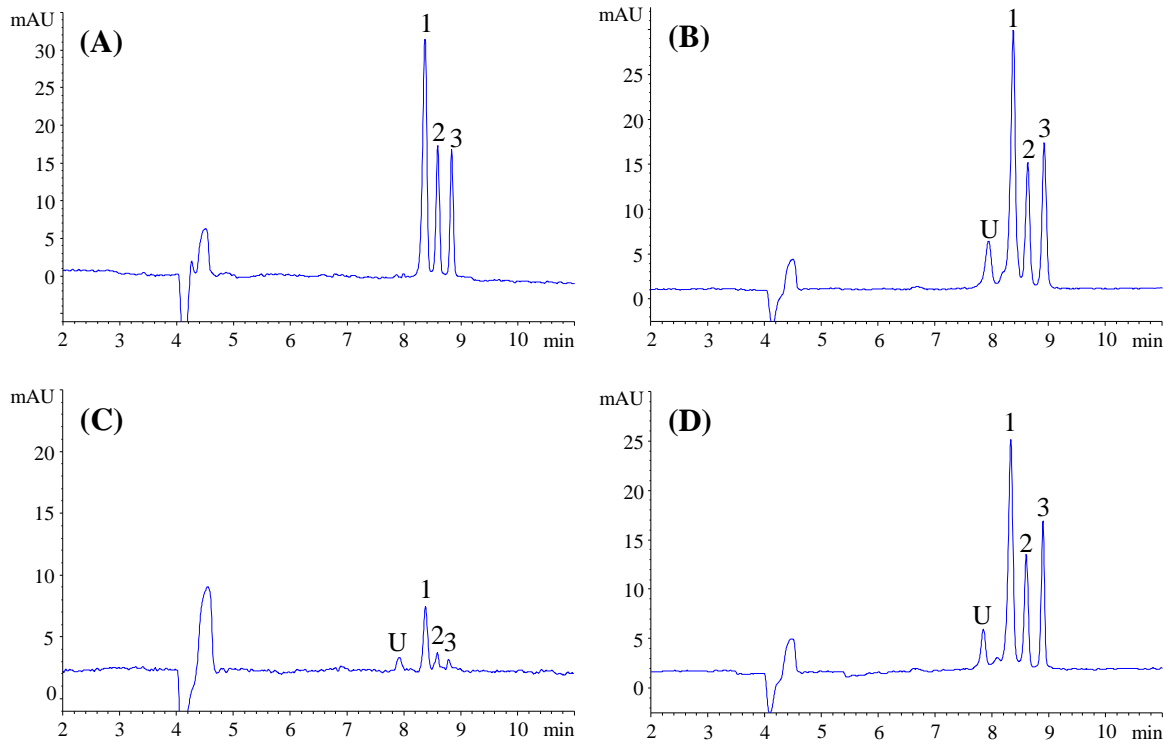
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Li *et al.*, 2013
Fig. 2



Li *et al.*, 2013
Fig. 3

1

2
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Li *et al.*, 2013
Fig. 4