

Analytical Methods

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the RSC Publishing peer review process and has been accepted for publication.

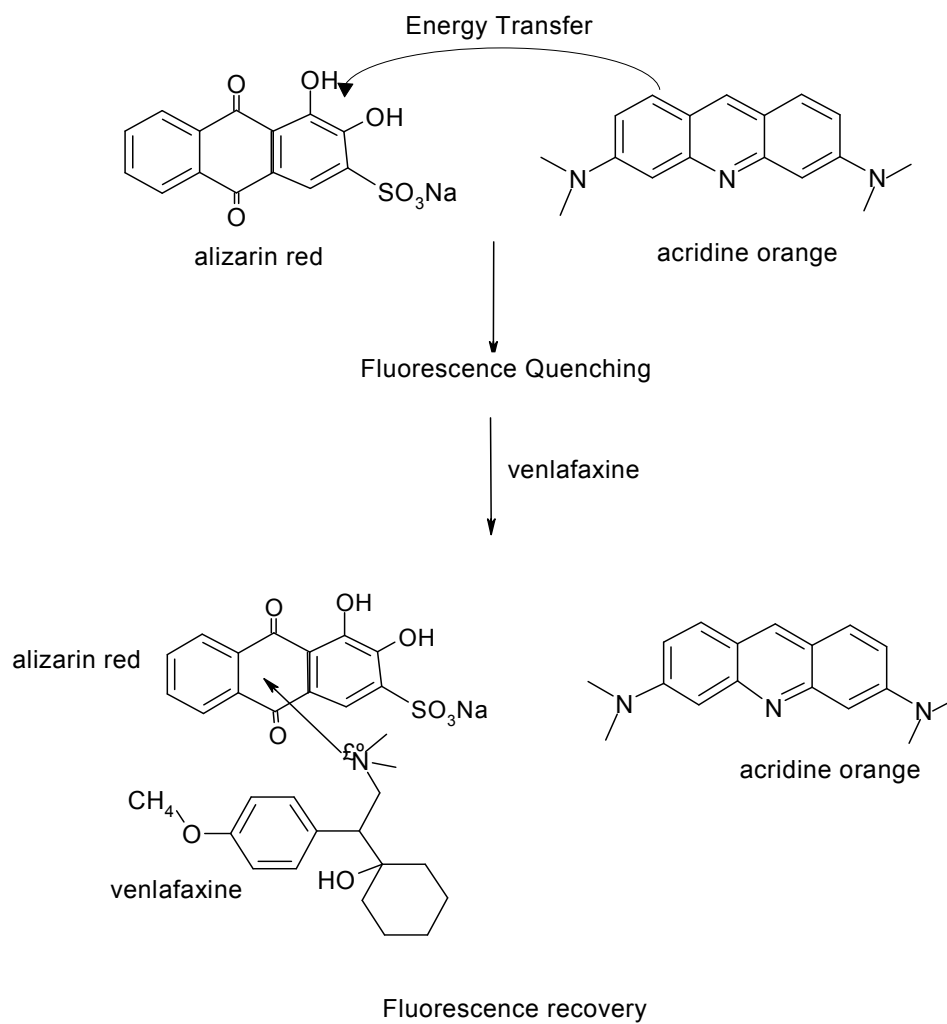
Accepted Manuscripts are published online shortly after acceptance, which is prior to technical editing, formatting and proof reading. This free service from RSC Publishing allows authors to make their results available to the community, in citable form, before publication of the edited article. This *Accepted Manuscript* will be replaced by the edited and formatted *Advance Article* as soon as this is available.

To cite this manuscript please use its permanent Digital Object Identifier (DOI®), which is identical for all formats of publication.

More information about *Accepted Manuscripts* can be found in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics contained in the manuscript submitted by the author(s) which may alter content, and that the standard [Terms & Conditions](#) and the [ethical guidelines](#) that apply to the journal are still applicable. In no event shall the RSC be held responsible for any errors or omissions in these *Accepted Manuscript* manuscripts or any consequences arising from the use of any information contained in them.

Graphical abstract



A Fluorescence Method for the Determination of Venlafaxine Hydrochloride

Liangwei Du, Xiaoling Wei*, Xiangrong Lei, Lisheng Wang, Qi Gong, Xiaojun Wang

College of Chemistry and Chemical Engineering, Guangxi University, Nanning, 530004, China

Abstract: In this study, fluorescence quenching of acridine orange was observed when the effective energy transfer happened between alizarin red and acridine orange under aqueous conditions with a pH of 6.8. However, the quenched fluorescence in this aqueous solution could be partially recovered with the addition of an appropriate amount of venlafaxine hydrochloride. And the recovery of fluorescence intensity had a good linear relationship with the amount of added venlafaxine hydrochloride. Based on this, a novel fluorescence recovery method for the determination of venlafaxine hydrochloride was established. The experimental results showed that under optimal conditions, the linear range of this method was 3.49–31.4 mg·L⁻¹, the detection limit was 1.65 mg·L⁻¹, and the precision was 1.16%. The method was used for the determination of venlafaxine hydrochloride in sustained-release tablets and capsules with satisfactory results, and the recoveries were in the range of 95.5%–105%.

Keywords: venlafaxine hydrochloride, fluorescence recovery, acridine orange, alizarin red

*Corresponding Author

E-mail: wx11651@163.com

22 1 Introduction

23 Venlafaxine hydrochloride (VLX), a second-generation non-tricyclic antidepressant,
24 was used clinically to treat depression and certain types of anxiety disorders with mild
25 side effect in recent years. Currently, some methods for the determination of
26 venlafaxine hydrochloride have been reported, such as high performance liquid
27 chromatography (HPLC) [1–12], liquid chromatography-mass spectrometry [13–19],
28 visible spectrophotometry [20, 21], ultraviolet spectrophotometry [22, 23], electrical
29 analysis [24–26], and charge-transfer spectrophotometry [27] and so on. These
30 methods all have its relative merits, but there are still some unsatisfactory aspects. For
31 instance, high performance liquid chromatography and liquid chromatography-mass
32 spectrometry methods require complicated sample pretreatment procedure, expensive
33 instrument and high operative expenses. Spectrophotometry and electroanalysis
34 methods have low selectivity and were easy to be interfered by the excipients.

35 In this paper, it was found that venlafaxine could recover the quenched
36 fluorescence of acridine orange-alizarin red (AO-AR) system through charge-transfer
37 reaction between venlafaxine and alizarin red. Accordingly, a novel fluorescence
38 recovery method for the determination of venlafaxine hydrochloride was established.
39 Experimental results showed that this method was simple, fast, low cost and suitable
40 for the batch analysis of venlafaxine hydrochloride with good sensitivity and
41 selectivity, which had important significance for the quality control of pharmaceutical
42 production. To our knowledge, no paper using fluorescent recovery for the
43 determination of venlafaxine hydrochloride has been reported.

44 **2 Experimental**

45 **2.1 Apparatus**

46 All fluorescence spectra were measured with a RF-5301 PC fluorescence
47 spectrophotometer (Shimadzu Corp., Japan). The UV-vis absorption spectra were
48 carried out on a UV-2102 PCS UV-vis spectrophotometer (Unico Shanghai
49 Instrument Co., Ltd., China). All pH measurements were conducted with Ray
50 magnetic PHS-3C pH meter (Shanghai Jingke Industrial Co., Ltd., China). The
51 temperature was controlled by HH-38 thermostat water bath (Zhengzhou Changcheng
52 Technology and Business Co., Ltd., China).

53 **2.2 Reagents**

54 All reagents were used as received without further experimental purification.
55 Acridine orange of analytical grade was purchased from Blue Season Science and
56 Technology Development Co., Ltd. (Shanghai, China). Alizarin red of analytical pure
57 was obtained from XinZhong Chemical Plant (Shanghai, China). Venlafaxine
58 hydrochloride was standard compound with the purity of 99.9% from National
59 Institutions for Food and Drug Control (Beijing, China). Analytical pure sodium
60 hydroxide (NaOH) was purchased from Shanghai Chemical Reagent Co., Ltd.
61 (Shanghai, China). Hydrochloric acid (HCl) of analytical grade was from Xilong
62 Chemical Plant (Guangdong, China). All other reagents were of analytical grade. The
63 water for the experiment was deionized water.

64 **2.3 Preparation of stock solutions**

65 Three stock solutions with the concentration of $1.00 \times 10^{-2} \text{ mol} \cdot \text{L}^{-1}$ were prepared

66 by dissolving 0.3700 g acridine orange, 0.3603 g alizarin red and 0.3139 g
67 venlafaxine hydrochloride in deionized water with constant volume to 100 mL,
68 respectively. They were diluted to $5.00 \times 10^{-4} \text{ mol} \cdot \text{L}^{-1}$ with deionized water for the
69 experiment. 0.1 M $\text{KH}_2\text{PO}_4\text{-Na}_2\text{HPO}_4$ buffer solution with $\text{pH}=6.8$ was also
70 prepared.

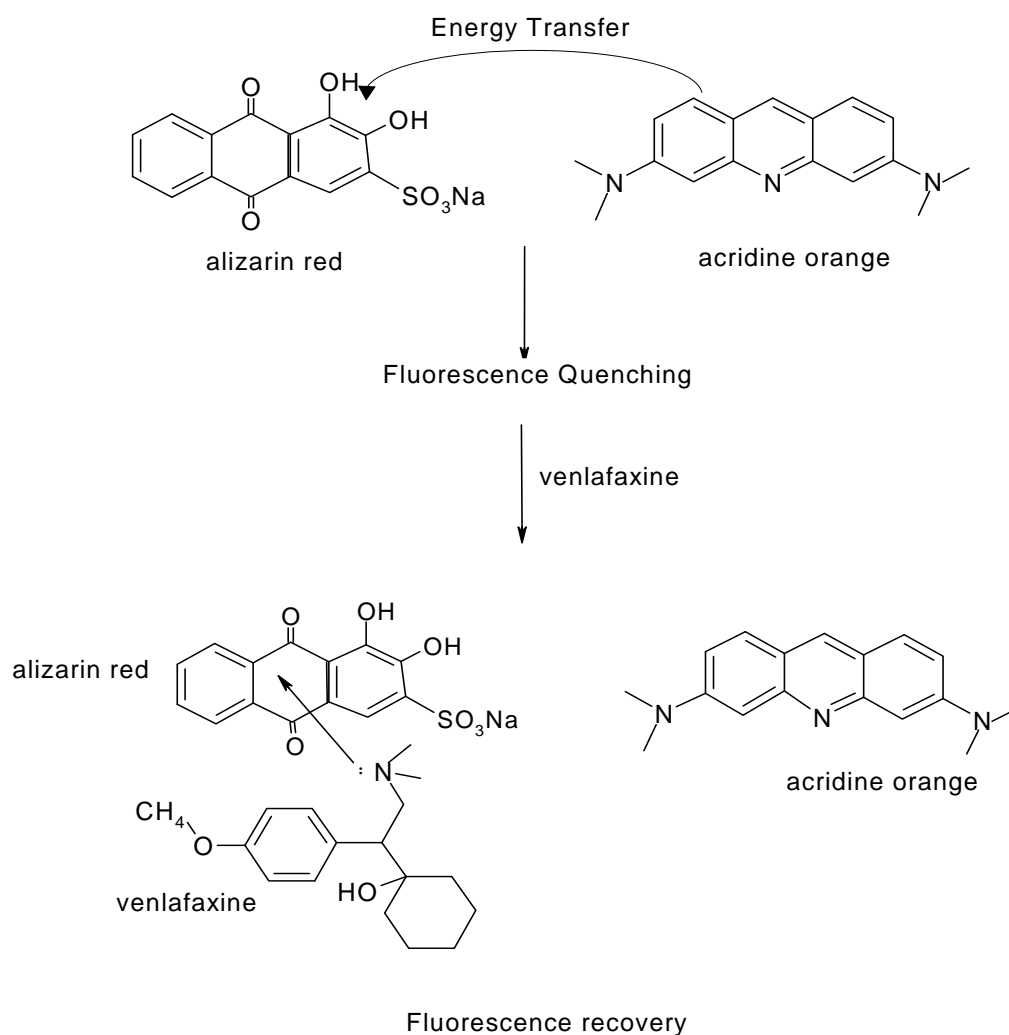
71 **2.4 Experimental procedure**

72 1.50 mL $5.00 \times 10^{-4} \text{ mol} \cdot \text{L}^{-1}$ alizarin red solution, 1.00 mL of $\text{KH}_2\text{PO}_4\text{-Na}_2\text{HPO}_4$
73 buffer solution ($\text{pH}=6.8$) and 1.70 mL $5.00 \times 10^{-4} \text{ mol} \cdot \text{L}^{-1}$ acridine orange solution
74 were added into a 10 mL volumetric flask, mixed well and reacted under stationary
75 condition for 5 min. After that, a certain amount of venlafaxine hydrochloride solution
76 was added, then diluted to the mark with deionized water and mixed well. The
77 volumetric flask with the mixed solution was placed in a constant temperature water
78 bath for 40 min at 30 °C. The solution's fluorescence intensity denoted as F_1 was
79 measured with 495 nm for excitation wavelength and 530 nm for emission
80 wavelength. The fluorescence intensity of blank solution without the addition of
81 venlafaxine hydrochloride was denoted as F_0 and was measured under the same
82 conditions. The intensity difference between them was calculated using $\Delta F = F_1 - F_0$.

83 **2.5 Method principle**

84 Under the conditions of pH from 6 to 7, alizarin red had a strong absorption peak
85 centered at 525 nm. Acridine orange, a strong fluorescent material with tricyclic
86 aromatic plane structure, has a maximum emission wavelength between 526 and 530
87 nm [28]. In the solution of $\text{pH}=6\text{--}7$, because the maximum absorption wavelength of

88 alizarin red is relatively consistent with the maximum emission wavelength of
89 acridine orange, efficient energy transfer can occur between them, which results in the
90 fluorescence quenching of acridine orange. Alizarin red, an anthraquinone compound,
91 contains electron-withdrawing groups such as carbonyl and sulfonic acid with
92 electron-deficient molecular structure of planar conjugated large π bond [29]. Thus,
93 charge transfer can occur with electron donor. When venlafaxine hydrochloride was
94 added to the solution, hydrochloric acid was neutralized in this near neutral solution,
95 then venlafaxine containing nitrogen group which has lone electron pair could behave
96 as electron donor. Charge transfer occurred with alizarin red can inhibit energy
97 transfer between acridine orange and alizarine red, so that the quenched fluorescence
98 of acridine orange was released. The process of fluorescence quenching and recovery
99 was illustrated in Fig. 1. There was a good linear relationship between the
100 fluorescence recovery and the concentration of venlafaxine hydrochloride in the
101 solution. Accordingly, a new fluorescence recovery method for the indirect
102 determination of venlafaxine hydrochloride was established.



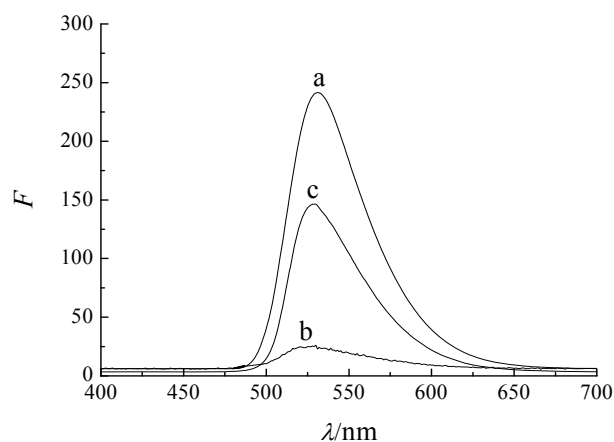
103

104 Fig.1 The illustration for the method principle of fluorescence recovery in this system.

105 **3 Results and discussion**106 **3.1 Fluorescence spectra of different solutions**

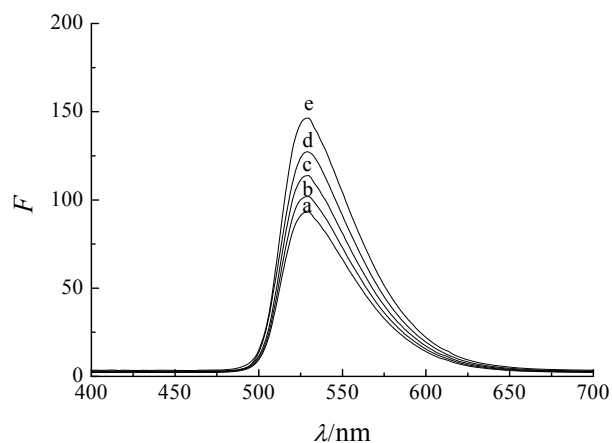
107 In accordance with the experimental method, fluorescence spectra of acridine
 108 orange, acridine orange-alizarin red and acridine orange-alizarin red-venlafaxine
 109 hydrochloride solutions containing 1.00 mL KH₂PO₄-Na₂HPO₄ buffer were obtained
 110 and shown in Fig. 2a, b and c, respectively. It can be seen from Fig. 2a that acridine
 111 orange has an intense emission peak with the maximum wavelength at 530 nm.
 112 Observed from Fig. 2b and Fig. 2c, alizarin red quenches significantly the

113 fluorescence of acridine orange and venlafaxine hydrochloride anti-quenches the
114 fluorescence of acridine orange-alizarin red solution.



115
116 Fig. 2 Fluorescence spectra of different solutions: (a) $8.5 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$ AO; (b) a+ 7.5×10^{-5}
117 $\text{mol} \cdot \text{L}^{-1}$ AR; (c) b+ $6.0 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$ VLX.

118 In addition, fixed acridine orange concentration of $8.5 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$, alizarin red
119 concentrations of $7.5 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$, and added different amounts of venlafaxine
120 hydrochloride solution, the fluorescence spectra of these mixed solutions were
121 obtained and shown in Fig. 3. It is illustrated that the fluorescence intensity enhanced
122 with the increasing concentration of venlafaxine hydrochloride. There was
123 proportional relationship between the enhanced fluorescence and the concentration of
124 venlafaxine hydrochloride. Based on this relationship, venlafaxine hydrochloride can
125 be measured quantitatively.



126

127 Fig. 3 Fluorescence spectra of VLX with different concentration in the solution of 7.5×10^{-5}

128 $\text{mol} \cdot \text{L}^{-1}$ AR and $8.5 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$ AO, in which the concentrations were 4.71, 9.42, 14.12, 18.83

129 and $23.54 \text{ mg} \cdot \text{L}^{-1}$ from curve a to curve e, respectively.

130 3.2 Optimization of the determination conditions

131 3.2.1 Choice of the solution acidity and buffer system

132 According to the experimental method, fixed other conditions, adjusted pH value of

133 solutions with $0.01 \text{ mol} \cdot \text{L}^{-1}$ HCl and NaOH, the effects of solution acidity on ΔF of

134 the reaction system was investigated and shown in Fig. 4. It can be seen that under

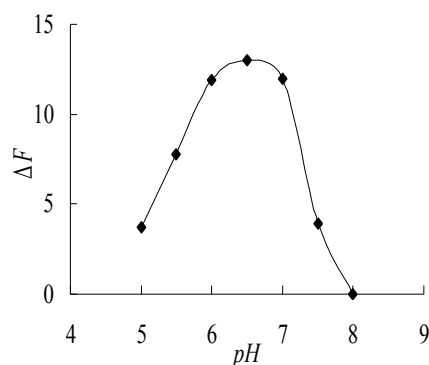
135 conditions of pH between 6 and 7, ΔF value reached maximum and became stable.

136 Because hydrochloric acid in venlafaxine hydrochloride was neutralized under near

137 neutral conditions, a stable charge transfer complex could be formed by reacting

138 between venlafaxine and alizarin red. Therefore, the pH of this experiment was

139 controlled between 6 and 7.



140

141 Fig. 4 Influence of pH on ΔF , in which the solution contains $7.5 \times 10^{-5} \text{ mol}\cdot\text{L}^{-1}$ AR, 8.5×10^{-5} 142 $\text{mol}\cdot\text{L}^{-1}$ AO and $15.7 \text{ mg}\cdot\text{L}^{-1}$ VLX143 In this experiment, buffer solutions with pH of 6.8 including $\text{KH}_2\text{PO}_4\text{-Na}_2\text{HPO}_4$,144 $\text{NaH}_2\text{PO}_4\text{-Na}_2\text{HPO}_4$, citric acid- Na_2HPO_4 and citric acid-sodium citrate were selected

145 to investigate the influence of buffer solution with different compositions on the

146 reaction system. The experimental results showed that when $\text{KH}_2\text{PO}_4\text{-Na}_2\text{HPO}_4$ buffer147 solution was selected, ΔF value of the system reached maximum and stable. It may

148 be that citric acid and sodium citrate as organic molecules have unfavorable influence

149 on the organic reaction system. Thus, $\text{KH}_2\text{PO}_4\text{-Na}_2\text{HPO}_4$ was selected as buffer

150 solution in the experiment.

151 According to the experimental method, the amount of the buffer solution was also

152 investigated. It was found that when the amount of buffer solution was less than 0.8

153 mL, ΔF value enhanced with the increasing amount of buffer solution and when the154 amount of buffer solution was in the range of 0.8–1.2 mL, ΔF value reached

155 maximum and substantially constant. In this study, the amount of buffer solution was

156 1.0 mL.

157

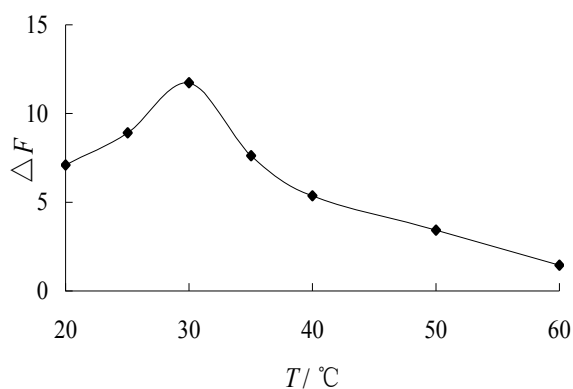
3.2.2 The adding sequence of reagents

158 The effect of adding sequence on the reaction system was also investigated. Five

159 kinds of adding sequences were tested, i.e. VLX \rightarrow buffer solution \rightarrow AR \rightarrow AO, AR
160 \rightarrow buffer solution \rightarrow VLX \rightarrow AO, AR \rightarrow buffer solution \rightarrow AO \rightarrow VLX, AO \rightarrow AR
161 \rightarrow buffer solution \rightarrow VLX and AO \rightarrow buffer solution \rightarrow AR \rightarrow VLX. Finally, it was
162 found that when the adding sequence was AR \rightarrow buffer solution \rightarrow AO \rightarrow VLX, ΔF
163 value of the system was maximum and stable. Thus, it was chosen as the optimal
164 adding sequence in this research.

165 3.2.3 Choice of the reaction temperature

166 Within the temperature range of 20–60 °C, the influence of reaction temperature on
167 ΔF value was investigated and shown in Fig. 5. Observed from Fig. 5, when
168 temperature was below 30 °C, ΔF value enhanced with the increase of temperature
169 and reached maximum at 30 °C; however, ΔF value decreased with the increasing
170 temperature when temperature was above 30 °C. We presume that at first the
171 increasing temperature is helpful to the formation of charge transfer complex and
172 energy transfer in this system, then the promoted dissociation of complex is
173 unfavorable for energy transfer with the continuous increase of temperature. Thus, 30
174 °C was chosen as optimum temperature for this experiment.



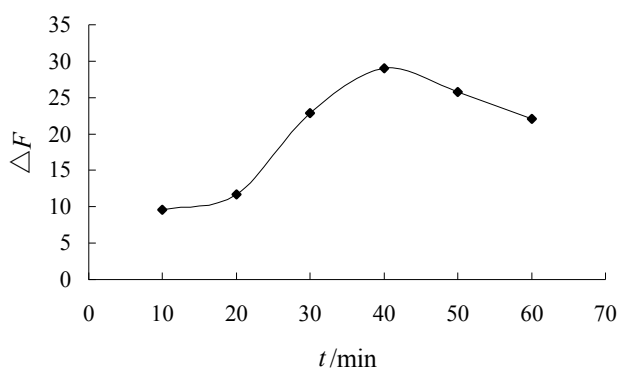
175

176 Fig. 5 Influence of temperature on ΔF , in which the solution contains $7.5 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$ AR ,

177 $8.5 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$ AO and $15.7 \text{ mg} \cdot \text{L}^{-1}$ VLX

178 3.2.4 Choice of the reaction time

179 With the temperature at $30 \text{ }^\circ\text{C}$, the effect of reaction time from 10 to 60 min on ΔF
180 was illustrated in Fig. 6. As shown in Fig. 6, at first, ΔF value of the system was
181 increasing with the increase of reaction time, and then ΔF value reached maximum
182 when the system reacted for 40 min, finally, ΔF value gradually decreased with the
183 reaction time more than 40 min. This result showed that too long reaction time could
184 cause the decomposition of charge transfer complex. In this paper, reaction time of 40
185 min was selected.



186

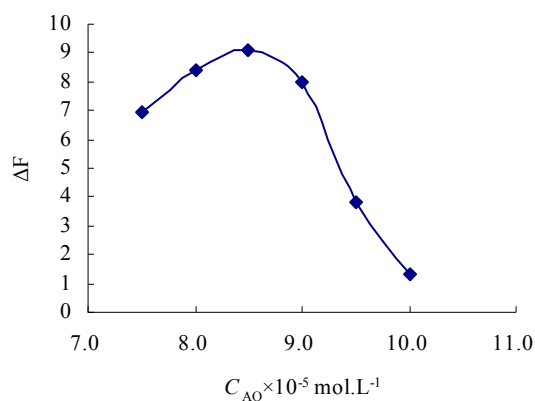
187 Fig. 6 Influence of reaction time on ΔF , in which the solution contains $7.5 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$ AR,

188 $8.5 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$ AO and $20.0 \text{ mg} \cdot \text{L}^{-1}$ VLX

189 3.2.5 Optimization of the amount of acridine orange

190 In order to investigate the effect of the amount of acridine orange on ΔF , fixed other
191 conditions, changed the amount of acridine orange, a series of ΔF values were
192 measured according to the experimental method. The results shown in Fig. 7 indicated
193 that when the amount of acridine orange solution was within
194 $8.0 \times 10^{-5} \sim 9.0 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$, ΔF value reached maximum and relatively stable, then

195 with the continuous increase of the concentration of acridine orange, ΔF value
 196 decreased. Thus, the best proportion of alizarin red, acridine orange and venlafaxine
 197 hydrochloride should be controlled at 1.5:1.7:≤ 1 in this reaction system.



198

199 Fig. 7 Effect of the amount of AO on ΔF , in which the different amount of solution was added into the
 200 reaction solution containing $7.5 \times 10^{-5} \text{ mol}\cdot\text{L}^{-1}$ AR and $15.7 \text{ mg}\cdot\text{L}^{-1}$ VLX.

201 3.3 Calibration curve and detection limit of the method

202 Under optimum conditions, fixed the amount of other solutions, changed the
 203 amount of venlafaxine hydrochloride solution, a series of reaction solutions were
 204 prepared. In accordance with the test method, ΔF values of the solutions were
 205 measured. A standard curve was obtained by taking ΔF as the ordinate and mass
 206 concentration (mg/L) of venlafaxine hydrochloride as the abscissa. The regression
 207 equation was $\Delta F = 0.4997m_{VLX} + 0.745$ with the correlation coefficient R of 0.9976.
 208 The results indicated that there was a good linear relationship between ΔF value and
 209 the concentration of venlafaxine hydrochloride in the range of 3.49–31.4 $\text{mg}\cdot\text{L}^{-1}$. The
 210 detection limit (3σ) obtained according to IUPAC regulation was $1.65 \text{ mg}\cdot\text{L}^{-1}$.

211 3.4 Precision and accuracy

212 To assess the precision and accuracy of the method, the determination of 20.0
213 $\text{mg}\cdot\text{L}^{-1}$ venlafaxine hydrochloride was carried out for eleven times. The average value
214 for these determinations was $20.1 \text{ mg}\cdot\text{L}^{-1}$ with the relative standard deviation of
215 1.16%, which indicated that this method had good accuracy and precision.

216 **3.5 Interferences of co-existing foreign substances**

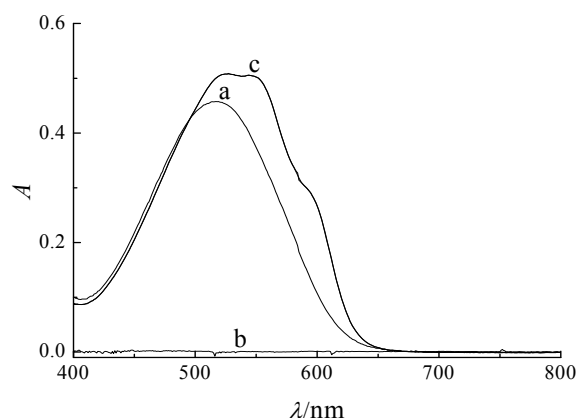
217 In order to investigate the possibility of practical application in the determination of
218 pharmaceutical preparation, interferences from excipients which were often contained
219 in tablets and capsules, such as glycerol, lactose, starch and stearic acid, etc. were
220 tested under optimum conditions. The results showed that the largest allowable
221 amount of foreign substances was 500 times of K^+ and Na^+ , 400 times of glucose and
222 ethanol, 300 times of stearic acid, 200 times of lactose, 100 times of surfactants
223 OP-10, 50 times of dextrin and 10 times of starch based on the impact on fluorescence
224 intensity was not more than $\pm 5\%$. However, the impact of amino acids and proteins on the
225 fluorescence intensity measurements is greater.

226

227 **3.6 Investigation on reaction mechanism**

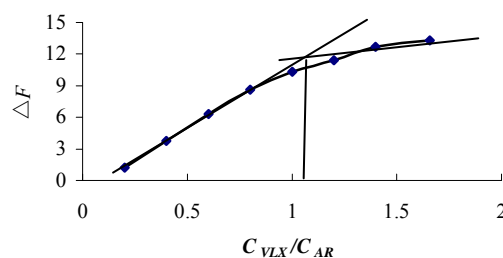
228 The UV-vis absorption spectra of alizarin red, venlafaxine hydrochloride and
229 alizarin red-venlafaxine hydrochloride in $\text{KH}_2\text{PO}_4\text{-Na}_2\text{HPO}_4$ buffer solution ($\text{pH}=6.8$)
230 were investigated and the results were shown in Fig. 8. It can be seen that
231 venlafaxine hydrochloride has no absorption in the visible region (curve b), alizarin
232 red has the maximum absorption peak at ca. 522 nm (curve a), the maximum
233 absorption peak of alizarin red-venlafaxine hydrochloride is around 550 nm (curve c).

234 Compared with curve a, peak shape and height of curve c has significantly changed.
 235 This indicated that the charge transfer reaction between alizarin red and venlafaxine
 236 hydrochloride occurred to form a stable charge transfer complex under the conditions
 237 of pH=6.8.



238
 239 Fig. 8 Absorption spectra of AR, VLX and AR+VLX in pH=6.8 buffer solution with the
 240 concentration of $7.5 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$ illustrated in curves a, b and c, respectively.

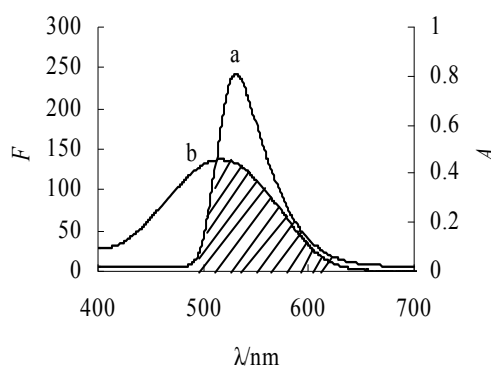
241 The mole ratio method was used to determine the composition ratio of charge
 242 transfer complex. The results showed that the composition ratio of charge transfer
 243 complex between venlafaxine hydrochloride and alizarin red was 1:1, as illustrated in
 244 Fig. 9.



245
 246 Fig. 9 The stoichiometry of charge transfer complex determined by mole ratio method.

247 The necessary condition for the occurrence of energy transfer between substance
 248 molecules is that the emission spectrum of energy donor has a certain degree of

249 overlap with the absorption spectrum of energy receptor [30]. UV-vis spectrum of AR
250 and fluorescence emission spectrum of AO were shown in Fig. 10. The figure showed
251 that under conditions of $\text{KH}_2\text{PO}_4\text{-Na}_2\text{HPO}_4$ buffer solution (pH=6.8), fluorescence
252 emission peak of acridine orange centers around 530 nm, and maximum absorption
253 wavelength of alizarin red is at 522 nm. There exist 8 nm differences between the two
254 peaks and two spectra overlap partially. Thus, it is possible that energy transfer will
255 occur between acridine orange as energy donor and alizarin red as energy acceptor.



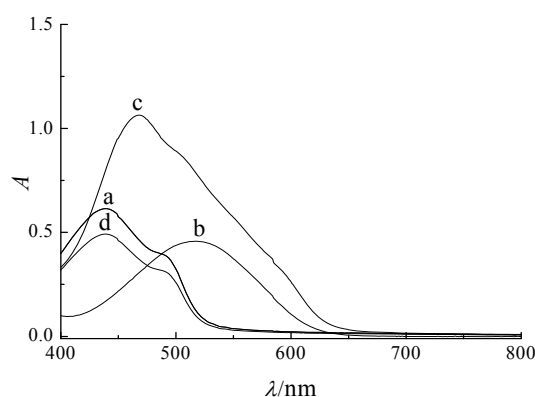
256

257 Fig. 10 (a) Fluorencence emission spectrum of AO ($7.5 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$); (b) absorption spectrum of
258 AR ($7.5 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$) in $\text{KH}_2\text{PO}_4\text{-Na}_2\text{HPO}_4$ buffer solution with pH=6.8.

259

260 In this paper, UV-vis absorption spectra of acridine orange, alizarin red, acridine
261 orange-alizarin red and acridine orange-alizarin red-venlafaxine hydrochloride were
262 also investigated. The results were shown in Fig. 11. It can be seen that acridine
263 orange has a maximum absorption at 440 nm and the maximum absorption peak of
264 alizarin red is at 522 nm, but when the two substances are mixed, the maximum
265 absorption peak moves to 470 nm. This indicated that the effective energy transfer
266 between acridine orange and alizarin red has occurred. When venlafaxine

267 hydrochloride was added to this system, the absorption peak at 470 nm disappeared
268 and the characteristic absorption peak of acridine orange appeared again at 440 nm.
269 This showed that the energy transfer between acridine orange and alizarin red was
270 inhibited due to the charge transfer reaction of venlafaxine hydrochloride with alizarin
271 red.



272
273 Fig. 11 Absorption spectra of different solutions: (a) AO ($8.5 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$); (b) AR (7.5×10^{-5}
274 $\text{mol} \cdot \text{L}^{-1}$); (c) a+b; (d) c+VLX ($6.0 \times 10^{-5} \text{ mol} \cdot \text{L}^{-1}$) in KH_2PO_4 - Na_2HPO_4 buffer solution with
275 pH=6.8.

276 A series of exploring experimental results mentioned above proved that the reaction
277 principle described in section 2.5 is reasonable.

278 3.7 Analytical application

279 To investigate the possibility of practical application, the concentration of
280 venlafaxine hydrochloride in tablets and capsules was determined by this method. The
281 relative standard deviation was lower than 2.5%. The recovery of the method was
282 obtained through detecting three samples with adding different concentrations of
283 venlafaxine hydrochloride. The results were illustrated in Table 1 and the average
284 recovery was between 95.5% and 105%. HPLC [31] was also performed as

285 comparative method. It was found that the result of determining venlafaxine
 286 hydrochloride using this method was almost the same with HPLC. Thus, this method
 287 was suitable to detect venlafaxine hydrochloride in tablets and capsules.

288

289

290 Table 1 Determination results of samples and recovery of the method (n=5).

Number	Labeled amount (mg·L ⁻¹)	Found (mg·L ⁻¹)	RSD (%)	Added (mg·L ⁻¹)	Found (mg·L ⁻¹)	Average recovery (%)	HPLC (mg·L ⁻¹)
1	0.150	0.150	0.99	0.0314	0.180	95.5	0.149
				0.0627	0.212	98.9	
2	0.150	0.149	2.3	0.0314	0.182	105	0.148
				0.0627	0.212	100	
3	0.150	0.150	1.2	0.0314	0.181	98.7	0.151
				0.0627	0.212	98.9	

291 *Note: 1. Venlafaxine hydrochloride sustained-release tablets (manufacturers: Chengdu Nakasone*

292 *Pharmaceutical Group Co., Ltd., China; batch number: 100904; labeled amount: 75 mg/tablet).*

293 *2. Venlafaxine hydrochloride sustained-release tablets (manufacturers: Chengdu Nakasone*

294 *Pharmaceutical Group Co., Ltd., China; batch number: 100804; labeled amount: 75 mg/tablet).*

295 *3. Venlafaxine hydrochloride extended-release capsules (manufacturer: Wyeth Medica Ireland,*

296 *Ireland; batch number: 0901058; labeled amount: 75 mg/capsule).*

297 **4 Conclusions**

298 A novel and convenient technique for venlafaxine hydrochloride analysis has been

299 developed by fluorescence recovery on acridine orange-alizarin red system. Using this

300 method, venlafaxine hydrochloride can be measured directly in aqueous solution and
301 the reagents required in this method are simple and easy to obtain. When this method
302 was used for the determination of venlafaxine hydrochloride in capsule and tablet
303 samples, the results were in excellent agreement with the labeled amount and that
304 measured by the HPLC [31] method. Comparing methods [1-19] mentioned in the
305 introduction, this method has the advantages of the simple sample pretreatment, measured at a
306 lower cost and no need to use the organic solvents. This method also has better sensitivity and
307 selectivity than methods [20-23] mentioned in the references [20-23].

308

309 **Acknowledgements**

310 This work was supported by the National Natural Science Foundation of China (No. 20942005).

311

312

313

314

315

316

317

318

319

320

321

322

323

324

325

326

327

328

329

330 **References**331 [1] H.D. Li, D.R. Ding, B.K. Zhang, H.Y. Yuan, *Chin. J. Pharm. Anal.* 21 (2001)

332 240–242.

333 [2] J. Meng, X.L. Sun, X.Y. Qin, X.Y. Li, A.L. Zhang, A.D. Wen, *Chin. J. Pharm.*334 *Anal.* 28 (2008) 1442–1445.335 [3] H.S. Li, M. Zhao, C. Liang, C.J. Zhao, *China Pharm.* 19 (2010) 27–28.336 [4] R. Mandrioli, L. Mercolini, R. Cesta, S. Fanali, M. Amore, M.A. Raggi, *J.*337 *Chromatogr. B* 856 (2007) 88–94.338 [5] Y. Huang, M.Z. Liang, Y.G. Zou, Q. Yu, Y.P. Qin, *Chin. Hosp. Pharm. J.* 22 (2002)

339 335–337.

340 [6] R. Waschgler, W. Moll, P. König, A. Conca, *Int. J. Clin. Pharmacol. Ther.* 42 (2004)

341 724–728.

342 [7] U.K. Chhalotiya, H.B. Patel, K.K. Bhatt, *Indian J. Pharm. Sci.* 72 (2010) 814–818.343 [8] K. Jaspreet, K.K. Srinivasan, J. Alex, *J. Pharm. Bioall. Sci.* 2 (2010) 22–26.344 [9] L.S. Bernardi, P.R. Oliveira, F.S. Murakami, *J. Chrom. Sci.* 47 (2008) 770–776.345 [10] S.L. Baldania, K.K. Bhatt, R.S. Mehta, *Indian j. Pharm. Sci.* 70 (2008) 124–128.346 [11] S. Victoria, N. Chrysa, K. Leda, *Bioanal.* 3 (2011) 1713–1718.347 [12] Z. Wei, B.R. Xiang, C.Y. Wang, *Biomed. Chrom.* 21 (2007) 266–272.

- 348 [13] J. He, Z.L. Zhou, H.D. Li, *Chin. J. Pharm. Anal.* 25 (2005) 1428–1432.
- 349 [14] J. Bhatt, A. Jangid, G. Venkatesh, G. Subbaiah, S. Singh, *J. Chromatogr. B* 829
350 (2005) 75–81.
- 351 [15] W. Liu, Y.C. Dai, N. Deng, *Biomed. Chromatogr.* 25 (2011) 412–416.
- 352 [16] G. R. Shah, B.T. Thaker, K.R. Surati, *Anal. Sci.* 25 (2009) 1207–1210.
- 353 [17] W. Liu, H.L. Cai, H.D. Li, *J. Chromatogr. B* 850 (2007) 405–411.
- 354 [18] J. He, Z.L. Zhou, H.D. Li, *J. Chromatogr. B* 820 (2005) 33–39.
- 355 [19] B.N. Patel, S. Naveen, S. Mallika, *J. Pharm. Boimed. Anal.* 47 (2008) 603–611.
- 356 [20] K. Raghubabu, L.S. Swarup, B. Kalyanaramu, *E-J. Chem.* 9 (2012) 1645–1654.
- 357 [21] D.G. Sankar, B. Snehalatha, K. Vijayasri, *Asian J. Chem.* 15 (2003) 509–511.
- 358 [22] S. Radha, J. Suman. *Asian J. Chem.* 21 (2009) 7440–7442.
- 359 [23] D.G. Sankar, K. Vijayasri, B. Snehalatha, *Asian J. Chem.* 14 (2002) 1779–1781.
- 360 [24] B.J. Sanghavi, A.K. Srivastava, *Electrochim. Acta.* 56 (2011) 4188–4196.
- 361 [25] A.A. Ensafi, R. Faridfar, A.R. Allafchian, *Sensor Lett.* 9 (2011) 479–484.
- 362 [26] A.X. Yang, G.D. Jin, J.L. Ge, *Chin. J. Pharm. Anal.* 29 (2009) 1134–1137.
- 363 [27] U.S. Tatar, *Opt. Spectrosc.* 110 (2011) 508–512.
- 364 [28] Y.X. Luan, G.Y. Xu, Y.M. Li, C.X. Sun, S.E. Song, J. Liu, *Dyes Pigments* 67
365 (2005) 223–228.
- 366 [29] F. Gul, A.M. Khan, S.S. Shah, M.F. Nazar, *Color. Technol.* 126 (2010) 109–113.
- 367 [30] B.S. Liu, J. Gao, G.L. Yang, *Spectrosc. Spect. Anal.* 25 (2005) 1080–1082.
- 368 [31] N. Du, Y.L. Zhou. *Chinese J. North. Pharm.* 9(2012) 4-5.
- 369