



Cite this: DOI: 10.1039/d6sc01847e

 All publication charges for this article have been paid for by the Royal Society of ChemistryReceived 4th March 2026  
Accepted 5th May 2026DOI: 10.1039/d6sc01847e  
rsc.li/chemical-science

## Introduction

Recent observations of temperature ( $T$ ) independence (or weak dependence) of kinetic isotope effects (KIEs) in various enzyme-catalyzed hydrogen (H) transfer reactions have been ascribed by several research groups to the  $T$ -independence of narrowly distributed donor-acceptor distances (DADs) for H-tunneling processes.<sup>1–14</sup> Therefore, such observations are often cited in support of the proposed role of strong protein vibrational dynamics in facilitating short DAD sampling,<sup>1,15–20</sup> and further promoting enzyme catalysis.<sup>8,12,20–31</sup> Use of this approach to propose the new physical origin of enzyme catalysis is currently, however, debated.<sup>32–36</sup> Central to the debate is whether  $T$ -dependence of KIEs reliably reflects DAD fluctuations or overall system rigidity. To address this question, biochemists have designed experiments to probe enzyme system rigidity and its correlation with the  $T$ -dependence of KIEs.<sup>6,9,11,12,37–39</sup> Theoreticians have refined or developed H-tunneling models to simulate the observations in attempts to test whether there is such a correlation.<sup>14,32,33,40–43</sup> For the same purpose, our group has

# Free energy relationship analysis for temperature dependence of hydride kinetic isotope effects of NADH/NAD<sup>+</sup> model reactions: implication for barrier compression by enzyme dynamics

Ava Austin-Kloppe,<sup>†</sup> Nicholas DeGroot,<sup>†</sup> Bikram Dhakal, Jessica Sager, Lauren Phan, Seyedmehrad Poormoghim and Yun Lu \*

The observed shift from temperature ( $T$ )-independence of hydrogen kinetic isotope effects (KIEs) in wild-type enzymes to  $T$ -dependence of KIEs in enzyme mutants has been explained as evidence for the role of protein dynamics in compressing donor (Don)–acceptor (Acc) distances (DADs) for catalysis. To test this explanation, correlation analysis of free energy changes ( $\Delta G^\circ = -44.3$  to  $6.7$  kcal mol<sup>-1</sup>) that simulate system rigidities and  $T$ -dependence of KIEs (represented by  $\Delta E_a = E_{aD} - E_{aH}$ ) was carried out for 34 hydride-tunneling reactions of NADH/NAD<sup>+</sup> models in acetonitrile. For exergonic reactions,  $\Delta E_a$  increases as  $\Delta G^\circ$  approaches zero, with the linear trend appearing to reverse for endergonic reactions. Both  $\Delta E_a$  and KIEs reach their maximum near thermoneutral reactions, where the charge-transfer (CT) complexation vibration is the weakest and DAD is the longest. A small portion of the free energy change drives the CT complexation vibrations and thus the DAD sampling that correlates with KIEs and their  $T$ -dependences. The results support the role of protein dynamics in barrier compression for catalysis. The new physical-organic linear  $\Delta E_a$ – $\Delta G^\circ$  relationship will contribute to the development of future H-tunneling models as well as updated theories for enzyme catalysis.

studied model reactions in solution to explore the potential relationship between the structure and  $T$ -dependence of hydride KIEs.<sup>24,25,27,29,44–50</sup> The solution-phase H-transfer reactions permit systematic variation of the molecular structure and solvent environment, enabling more controlled evaluation of how structural rigidity influences the  $T$ -dependence of KIEs in a broader context.

$T$ -Dependence of KIEs reflects the isotopic activation energy difference, represented as  $\Delta E_a (= E_{aD} - E_{aH})$  for hydrogen/deuterium transfer reactions. Semi-classically,  $\Delta E_a$  falls within the range between 1.0 and 1.2 kcal mol<sup>-1</sup>, but its relationship with structure is unpredictable.  $\Delta E_a$  outside of this range has been used to suggest an H-tunneling mechanism. Notably, a shift from  $\Delta E_a \sim 0$  in wild-type enzymes to  $\Delta E_a > 0$  (often exceeding the semi-classical limit) in mutant variants has been frequently observed. This trend has prompted the application of existing H-tunneling models, as well as the development of new theoretical frameworks, to account for these observations and to further elucidate the mechanisms of H-transfer chemistry.<sup>2,5–9,11,12,18,31,42,51–58</sup> Among these, the recently proposed vibration-assisted activated H-tunneling (VA-AHT) model appears to be able to explain the unusual KIE behaviors.<sup>2,5,8,20,59</sup>

The VA-AHT model incorporates two orthogonal activation processes: (1) heavy atom motions bring the reactants (donor-H and acceptor) and products to degenerate energy states at which

Department of Chemistry, Southern Illinois University Edwardsville, Edwardsville, Illinois 62026, USA. E-mail: yulu@siue.edu

<sup>†</sup> A. A. and N. D. contributed equally to both the experiments and writing of the manuscript.



H-wave functions overlap ( $[\text{Don-H} \leftrightarrow \text{H-Acc}]^\ddagger$ ), *i.e.*, tunnelling ready states (TRS's); and (2) more rapid heavy atom motions, also called promoting vibrations,<sup>19,60</sup> sample short DADs for efficient tunneling to occur. Since tunneling of a D-isotope requires a smaller average DAD due to its shorter de Broglie wavelength, a higher activation energy is needed leading to  $E_{\text{aD}} > E_{\text{aH}}$  (assuming that the first activation process is isotope insensitive). In wild-type enzymes, however, strong protein vibrations compress the donor/acceptor closely together, thereby facilitating the formation of short DADs that are extremely densely populated, eliminating the possibility for further short DAD sampling for D-tunneling and making  $\Delta E_{\text{a}} \sim 0$ . In enzyme mutants, the constructive vibrations are disrupted, and thermal sampling of shorter DADs is allowed so that  $T$ -dependence of DADs/KIEs ( $\Delta E_{\text{a}} > 0$ ) emerges. The phenomenological model has been claimed by some researchers to be able to explain all of the hydrogen KIEs.<sup>5,6,8,12</sup>

Computational simulations of  $T$ -dependence of KIEs followed various theoretical models including the VA-AHT model to investigate the proposed role of fast thermal dynamics in sampling short DADs. Other models include ensemble-averaged variational TS theory with multi-dimensional H-tunneling<sup>32,41,61</sup> as well as approaches using the empirical valence bond theory.<sup>33,34,62</sup> While the VA-AHT model has successfully reproduced the  $\Delta E_{\text{a}}$  in nonadiabatic reactions to support the thermally activated DAD sampling concept,<sup>63–65</sup> simulations for more adiabatic hydride/proton transfer reactions have been challenging as there is no direct mathematical relationship yet between DADs and  $\Delta E_{\text{a}}$  for this type of reaction.<sup>30,43</sup> On the other hand, theoretical replications of the observations using other H-tunneling models, especially the huge KIEs, have encountered difficulty.<sup>10,32,36,66</sup> Nevertheless, even some of the latter computational results sometimes show that  $\Delta E_{\text{a}} \sim 0$  in some reactions results from the insensitivity of the DAD to temperature,<sup>13,34,41,63</sup> whereas other researchers have shown that it could also result from the effect of temperature on the microscopic mechanism, for example, on the position of the TS and shape of the potential barrier.<sup>32,34</sup> The potential relationship between DAD distributions and  $\Delta E_{\text{a}}$  magnitudes warrants further investigation to search for a potential mathematical relationship, if it exists, for building future H-tunnelling theoretical frameworks.

To address the “DAD– $\Delta E_{\text{a}}$ ” relationship, additional data, including from model systems, are needed. Over the years, we have designed hydride transfer reactions of NADH/NAD<sup>+</sup> models in solution to tackle the problem. Our hypothesis, based on enzyme observations, is that a more rigid system with densely populated DADs gives rise to a smaller  $\Delta E_{\text{a}}$ .<sup>25</sup> Systems have been designed to study the electronic, steric, and solvent effects as well as effects of remote heavy group vibrations and mechanisms on the  $T$ -dependence of KIEs.<sup>25,27,29,44–50</sup> Current results show that a tighter charge-transfer (CT) complexation between NADH/NAD<sup>+</sup> model structures exhibits a smaller  $\Delta E_{\text{a}}$  value, supporting the hypothesis.

As the project progressed, we seemed to have identified a trend indicating that a reaction with a more negative free energy change ( $\Delta G^\circ$ ) tends to show a smaller  $\Delta E_{\text{a}}$  value.<sup>29,44,46,48</sup> This appears consistent with our hypothesis as a stronger

hydride donor/acceptor would form stronger CT complexation vibrations and thus more rigid donor–acceptor centers. In this context, a larger negative  $\Delta G^\circ$  corresponds to a tighter CT complexation and, consequently, more densely populated smaller DADs, which in turn result in smaller  $\Delta E_{\text{a}}$  values. Based on this reasoning, we envisioned that there might be a free energy relationship with  $T$ -dependence of KIEs for this type of reaction. As a matter of fact, a VA-AHT-inspired model can be formulated in which a portion of the free energy ( $\Delta G_{\text{TRS}}^\circ$ ) drives the structural and solvent rearrangement required to reach a TRS, while the remaining portion ( $\Delta G_{\text{DAD}}^\circ$ ) modulates DAD sampling toward shorter distances; together, both contributions enable H-tunneling. While the overall free energy change ( $\Delta G^\circ = \Delta G_{\text{TRS}}^\circ + \Delta G_{\text{DAD}}^\circ$ ) is expected to linearly correlate with the logarithm of the observed rates ( $\ln(k)$ ),  $\Delta E_{\text{a}}$  reflects (but is not equal to) the thermal energy required for DAD sampling and is expected to correlate linearly with the  $\Delta G_{\text{DAD}}^\circ$ . Although  $\Delta G_{\text{DAD}}^\circ$  cannot be directly determined, establishing a  $\Delta E_{\text{a}}-\Delta G^\circ$  relationship would provide an indirect means to test our DAD– $\Delta E_{\text{a}}$  hypothesis, clarify the driving force underlying  $\Delta E_{\text{a}}$ , and deepen our understanding of DAD sampling as well as the enzymatic KIE behavior.

In this work, we examined 34 hydride transfer reactions of NADH/NAD<sup>+</sup> models in acetonitrile, with  $\Delta G^\circ$  values spanning from  $-44.3$  to  $6.7$  kcal mol<sup>-1</sup>, to investigate the previously unreported  $\Delta E_{\text{a}}-\Delta G^\circ$  relationship and further test our hypothesis concerning the DAD– $\Delta E_{\text{a}}$  relationship. We also compare the  $\Delta E_{\text{a}}-\Delta G^\circ$  relationship with the  $\ln(k)-\Delta G^\circ$  and  $\ln(\text{KIE})-\Delta G^\circ$  relationships. Furthermore, this study represents the first application of our hypothesis to endergonic reactions. The dataset reveals how the trends of these relationships evolve when transitioning from exergonic to endergonic regions. These insights enable testing of the DAD sampling mechanism in the VA-AHT(-inspired) model, evaluating other current H-transfer theories, as well as informing the development of future theoretical frameworks for both general H-transfer/tunneling reactions and enzyme catalysis.

## Results

The hydride donor and acceptor structures are shown in Fig. 1. Kinetics were determined spectroscopically on a stopped-flow UV-vis spectrophotometer. The  $\Delta G^\circ$ , second-order rate constants ( $k_2$ ) and KIEs at 25 °C, as well as the  $\Delta E_{\text{a}}$  values are listed in Table

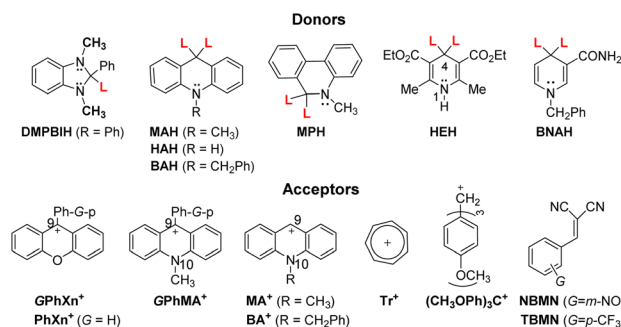


Fig. 1 Hydride donors and acceptors (counter-ions: BF<sub>4</sub><sup>-</sup>) (L = H or D) of the reactions in acetonitrile.



Table 1 Hydride affinities ( $-\Delta G_{\text{H}}^{\circ}$ ), free energy changes ( $\Delta G^{\circ}$ ), and hydride transfer kinetic data in acetonitrile

Rxns	Donors (Don-H)	Hydride acceptors (Acc-BF <sub>4</sub> <sup>-</sup> )	$-\Delta G_{\text{H}}^{\circ}$ (Don) <sup>y</sup> (kcal mol <sup>-1</sup> )	$-\Delta G_{\text{H}}^{\circ}$ (Acc) <sup>y</sup> (kcal mol <sup>-1</sup> )	$\Delta G^{\circ}$ (kcal mol <sup>-1</sup> )	$k_{\text{2H}}$ (25 °C) <sup>z</sup> (M <sup>-1</sup> s <sup>-1</sup> )	KIE <sup>m</sup> (25 °C)	$\Delta E_{\text{a}}^m$ (kcal mol <sup>-1</sup> )
<b>Exergonic reactions</b>								
1 <sup>a</sup>	DMPBIH	CF <sub>3</sub> PhXn <sup>+</sup>	49.2	93.5	-44.3	1.44 (0.01) × 10 <sup>5</sup>	2.56 (0.03)	0.03 (0.07)
2 <sup>a</sup>	DMPBIH	BrPhXn <sup>+</sup>	49.2	92.5	-43.3	9.86 (0.09) × 10 <sup>4</sup>	2.55 (0.03)	0.07 (0.07)
3 <sup>a</sup>	DMPBIH	PhXn <sup>+</sup>	49.2	91.6	-42.4	4.54 (0.05) × 10 <sup>4</sup>	2.68 (0.04)	0.27 (0.06)
4 <sup>a</sup>	DMPBIH	CH <sub>3</sub> OPhXn <sup>+</sup>	49.2	90.2	-41.0	2.08 (0.02) × 10 <sup>4</sup>	2.74 (0.03)	0.55 (0.06)
5 <sup>a</sup>	DMPBIH	(CH <sub>3</sub> ) <sub>2</sub> NPhXn <sup>+</sup>	49.2	86.7	-37.5	6.34 (0.04) × 10 <sup>2</sup>	2.89 (0.06)	0.50 (0.08)
6 <sup>b</sup>	DMPBIH	MA <sup>+</sup>	49.2	76.2	-27.0	2.12 (0.01) × 10 <sup>2</sup>	3.57 (0.03)	0.43 (0.15)
7 <sup>a</sup>	MAH	CF <sub>3</sub> PhXn <sup>+</sup>	76.2	93.5	-17.3	1.03 (0.01) × 10 <sup>3</sup>	4.06 (0.04)	0.89 (0.07)
8 <sup>a</sup>	MAH	BrPhXn <sup>+</sup>	76.2	92.5	-16.3	6.45 (0.03) × 10 <sup>2</sup>	4.04 (0.03)	0.89 (0.07)
9 <sup>a</sup>	MAH	PhXn <sup>+</sup>	76.2	91.6	-15.4	4.10 (0.03) × 10 <sup>2</sup>	4.08 (0.03)	0.88 (0.05)
10 <sup>a</sup>	MAH	CH <sub>3</sub> OPhXn <sup>+</sup>	76.2	90.2	-14.0	1.56 (0.01) × 10 <sup>2</sup>	4.18 (0.04)	0.92 (0.16)
11 <sup>a</sup>	MAH	(CH <sub>3</sub> ) <sub>2</sub> NPhXn <sup>+</sup>	76.2	86.7	-10.5	4.34(0.03)	4.45 (0.05)	0.96 (0.18)
12 <sup>c</sup>	MAH	Tt <sup>+</sup>	76.2	83.0	-8.8	4.01 (0.02)	4.95 (0.12)	1.14 (0.10)
13 <sup>c</sup>	HAH	Tt <sup>+</sup>	74.9	83.0	-8.1	7.33 (0.05) × 10	5.34 (0.04)	1.27 (0.09)
14 <sup>c</sup>	HAH	PhXn <sup>+</sup>	74.9	91.6	-16.7	1.15 (0.01) × 10 <sup>3</sup>	4.19 (0.03)	0.88 (0.05)
15 <sup>c</sup>	BAH	PhXn <sup>+</sup>	77.4	91.6	-14.2	3.79 (0.02) × 10 <sup>2</sup>	4.26 (0.03)	0.89 (0.05)
16 <sup>c</sup>	MPH	PhXn <sup>+</sup>	65.7	91.6	-25.9	3.74 (0.02) × 10 <sup>3</sup>	3.18 (0.03)	0.71 (0.05)
17 <sup>c</sup>	MPH	CH <sub>3</sub> OPhXn <sup>+</sup>	65.7	90.2	-24.5	1.65 (0.01) × 10 <sup>3</sup>	3.28 (0.03)	0.63 (0.05)
18 <sup>c,d</sup>	BNAH	MA <sup>+</sup>	59.3	76.2	-16.9	7.16 (0.07) × 10	4.59 (0.06)	1.14 (0.17)
19 <sup>c</sup>	BNAH	BA <sup>+</sup>	59.3	77.4	-18.1	2.48 (0.02) × 10 <sup>2</sup>	3.75 (0.03)	0.82 (0.07)
20 <sup>c</sup>	BNAH	(CH <sub>3</sub> OPh) <sub>3</sub> C <sup>+</sup>	59.3	88.6	-29.3	1.66 (0.01) × 10 <sup>5</sup>	2.80 (0.03)	0.75 (0.05)
21 <sup>e</sup>	BNAH	(CH <sub>3</sub> ) <sub>2</sub> NPhXn <sup>+</sup>	59.3	86.7	-27.4	5.62 (0.04) × 10 <sup>4</sup>	3.19 (0.03)	0.82 (0.04)
22 <sup>e</sup>	BNAH	(CH <sub>3</sub> ) <sub>2</sub> NPhMA <sup>+</sup>	59.3	67.4	-8.1	8.37 (0.08) × 10 <sup>-1</sup>	4.79 (0.06)	1.14 (0.21)
23 <sup>b</sup>	HEH	MA <sup>+</sup>	64.4	76.2	-11.8	1.56 (0.01) × 10 <sup>2</sup>	4.92 (0.04)	0.99 (0.11)
24 <sup>f</sup>	HEH	BA <sup>+</sup>	64.4	77.4	-13.0	5.73 (0.03) × 10 <sup>2</sup>	4.53 (0.04)	1.01 (0.12)
25 <sup>c</sup>	HEH	(CH <sub>3</sub> OPh) <sub>3</sub> C <sup>+</sup>	64.4	88.6	-24.2	5.66 (0.03) × 10 <sup>3</sup>	3.64 (0.03)	0.88 (0.06)
26 <sup>g</sup>	HEH	CF <sub>3</sub> PhMA <sup>+</sup>	64.4	78.4	-14.0	2.74 (0.03) × 10	5.23 (0.05)	1.13 (0.19)
27 <sup>g</sup>	HEH	BrPhMA <sup>+</sup>	64.4	75.8	-11.4	2.09 (0.03) × 10	5.20 (0.08)	1.32 (0.09)
28 <sup>g</sup>	HEH	PhMA <sup>+</sup>	64.4	74.1	-9.7	1.37 (0.08) × 10	5.31 (0.04)	1.19 (0.07)
29 <sup>g</sup>	HEH	CH <sub>3</sub> PhMA <sup>+</sup>	64.4	72.8	-8.4	1.13 (0.02) × 10	5.30 (0.10)	1.33 (0.10)
30 <sup>g</sup>	HEH	CH <sub>3</sub> OPhMA <sup>+</sup>	64.4	71.7	-7.3	1.00 (0.01) × 10	5.11 (0.08)	1.31 (0.10)
31 <sup>g</sup>	HEH	(CH <sub>3</sub> ) <sub>2</sub> NPhMA <sup>+</sup>	64.4	67.4	-3.0	4.19 (0.03)	5.09 (0.06)	1.27 (0.14)
32 <sup>g</sup>	HEH	(CH <sub>3</sub> ) <sub>2</sub> NPhXn <sup>+</sup>	64.4	86.7	-22.3	8.87 (0.05) × 10 <sup>4</sup>	3.56 (0.02)	0.86 (0.08)
<b>Endergonic reactions</b>								
33 <sup>c</sup>	HEH	NBMN	64.4	58.1	6.3	1.14 (0.01)	5.13 (0.06)	1.17 (0.15)
34 <sup>c</sup>	HEH	TBMN	64.4	57.7	6.7	6.07 (0.02) × 10 <sup>-1</sup>	5.27 (0.06)	1.19 (0.16)
35 <sup>h</sup>	BnOH	PhXn <sup>+</sup>	—	91.6	—	6.73 (0.29) × 10 <sup>-5k</sup>	4.83 (0.21) <sup>k</sup>	1.00 (0.26)
36 <sup>i</sup>	i-PrOH	PhXn <sup>+</sup>	—	91.6	—	2.02 (0.05) × 10 <sup>-5k</sup>	3.63 (0.23) <sup>k</sup>	0.83 (0.27)
37 <sup>i</sup>	i-PrOH-β,β-d <sub>6</sub>	PhXn <sup>+</sup>	—	91.6	—	2.01 (0.05) × 10 <sup>-5k</sup>	3.64 (0.16) <sup>k</sup>	0.80 (0.19)
38 <sup>i</sup>	cyclo-HexOH	PhXn <sup>+</sup>	—	91.6	—	2.68 (0.01) × 10 <sup>-5k</sup>	3.68 (0.16) <sup>k</sup>	0.90 (0.27)

<sup>a</sup> Ref. 44. <sup>b</sup> Ref. 25. <sup>c</sup> This work. <sup>d</sup>  $\Delta E_{\text{a}} = 1.53$  (0.15) was reported by us before.<sup>25</sup> This was determined with temperatures from 4.5–29.5 °C, using the Hi-Tech Scientific SFA-20 fast kinetic determination kit interfaced to a UV-vis spectrophotometer. The conclusions from that publication are not changed using the numbers obtained in this work. <sup>e</sup> Ref. 48. <sup>f</sup> Ref. 29. <sup>g</sup> Ref. 47. <sup>h</sup> Ref. 24. <sup>i</sup> Ref. 29. <sup>j</sup> Unreported. <sup>k</sup> For 22 °C. <sup>l</sup> Ref. 68. <sup>m</sup> Numbers in parentheses are pooled standard deviations (standard deviations of the average values from different days of measurements in this work are listed in Tables S1 to S10 for comparison).



1 (for Rxns 1–34). As noted, some data were taken from our previous publications. Although kinetic study of the endergonic reactions is challenging due to the limited extent of the reaction and slow kinetics, in this work we were able to conduct a study on two such hydride transfer reactions: from Hantzsch 1,4-dihydropyridine (HEH) to benzylidenemalononitriles (BMNs), NBMN ( $\Delta G^\circ = 6.3 \text{ kcal mol}^{-1}$ ) and TBMN ( $6.7 \text{ kcal mol}^{-1}$ ) (Rxns 33 and 34 in Table 1; see structures in Fig. 1). The reactions proceed to completion due to subsequent rapid irreversible transfer of a proton from the N–H site of the HE<sup>+</sup> product to the dicyanomethanide anion product, as illustrated below,<sup>67</sup>

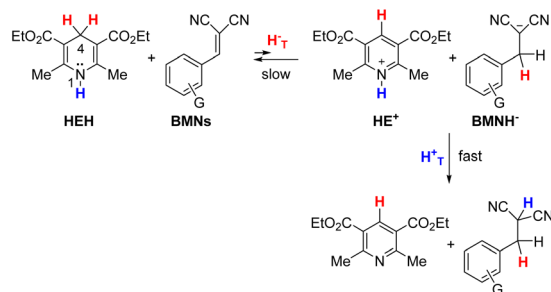
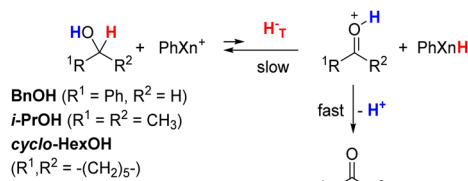


Table 1 also summarizes the relevant kinetic results for our previous study of the unfavorable hydride transfer reactions from alcohols ( $\text{R}^1\text{CH}(\text{OH})\text{R}^2$ ) to  $\text{PhXn}^+$  in acetonitrile (Rxns 35–38).<sup>24,29</sup>



In these reactions, hydride-transfer from the  $\alpha$ -position of the alcohol generates an  $\alpha$ -hydroxy carbocation intermediate ( $\text{R}^1\text{C}^+(\text{OH})\text{R}^2$ ). This process is subsequently followed by rapid proton transfer of the OH group to basic species in solution (*e.g.*, excess alcohol substrate or acetonitrile solvent), producing the corresponding oxidized carbonyl product ( $\text{R}^1\text{C}(\text{O})\text{R}^2$ ).<sup>24,29,69–71</sup>

While these extremely slow hydride transfer processes are expected to be endergonic, the corresponding free energy data are not available in the literature, limiting quantitative free energy relationship analysis. Notably, these reactions proceed significantly more slowly than the endergonic reactions of HEH with BMNs (more than  $10^4$  times slower, see Table 1), suggesting that they would be even more endergonic. Estimated  $\Delta G^\circ$  values have therefore been obtained by extrapolating the linear  $\ln(k_2)$ – $\Delta G^\circ$  correlation derived from reactions 1–34 (see the Discussion section below). These estimates are included to support the discussion of the trends observed across this limited set of endergonic reactions.

## Discussion

Although the KIEs at 25 °C are all below 6, a majority of the  $\Delta E_a$  values fall outside of the semiclassical range from 1.0 to 1.2 kcal  $\text{mol}^{-1}$  (Table 1). These results are consistent with observations

from enzyme-catalyzed hydride transfer reactions involving NADH/NAD<sup>+</sup> coenzymes. In both enzymatic and solution systems, such findings have been analyzed to suggest tunneling mechanisms.<sup>5,48,72–74</sup>

### Free energy relationship analysis for *T*-dependence of KIEs: the $\Delta$ -shaped $\Delta E_a$ – $\Delta G^\circ$ correlation

We first performed a linear correlation analysis of all 34  $\ln(k_2)$ – $\Delta G^\circ$  data points (Fig. 2A). Although the fit is poor ( $R^2 = 0.748$ ), it reveals a clear trend that the reaction rate decreases as  $\Delta G^\circ$  becomes less negative within the exergonic region and continues to decrease as the reactions become endergonic. Analysis of the  $\Delta E_a$ – $\Delta G^\circ$  relationship shows a relatively good linear correlation for the 32 exergonic reactions (Fig. 2B for Rxns 1–32,  $\Delta G^\circ = -44.3$  to  $-3.0 \text{ kcal mol}^{-1}$ ,  $R^2 = 0.852$ ), whereas the two endergonic reactions appear as significant outliers. Within the exergonic region, the  $\Delta E_a$  increases as  $\Delta G^\circ$  approaches zero. This new observation is consistent with our expectation; a more negative  $\Delta G^\circ$  corresponds to a more rigid system and a smaller  $\Delta E_a$ , with a portion of  $\Delta G^\circ$  ( $\Delta G^\circ_{\text{DAD}}$ ) contributing to the activation process associated with DAD sampling.

Therefore, the two endergonic reactions appear to align with the 32 exergonic reactions in the linear  $\ln(k_2)$ – $\Delta G^\circ$  correlation, whereas they appear as significant outliers relative to the linear  $\Delta E_a$ – $\Delta G^\circ$  correlation for the 32 exergonic reactions. In the latter case,  $\Delta E_a$  reaches a maximum when the negative  $\Delta G^\circ$  approaches zero, but upon entering the endergonic region, it begins to decrease. One may think that this behavior resulted from the experimental error or from structural diversity among the reactants, where factors other than electronic effects, such as steric effects, can affect the CT complexation strength and, consequently, the DAD distributions. While we already restricted the structures to rings containing C, N, and O that involve only 2p orbitals in CT complexation and the reactions to only one-step hydride transfers between two carbons, to further minimize the structural variability factor, we replotted the  $\ln(k_2)$ – $\Delta G^\circ$  and  $\Delta E_a$ – $\Delta G^\circ$  correlations using only the 12

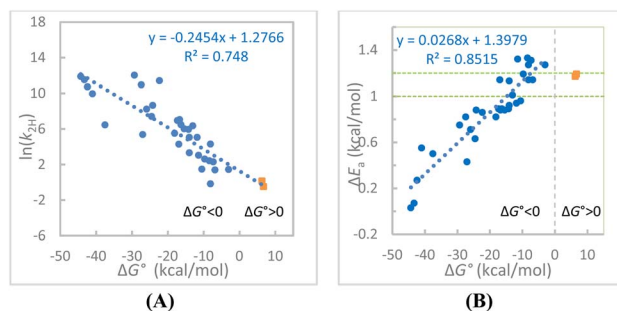


Fig. 2 (A) The linear fit of the  $\ln(k_2)$ – $\Delta G^\circ$  data for the 34 hydride transfer reactions (Rxns 1–34). (B) The linear fit of the  $\Delta E_a$ – $\Delta G^\circ$  data for the 32 exergonic hydride transfer reactions in acetonitrile (Rxns 1–32). The trend appears to turn to the opposite direction for endergonic reactions of BMNs (Rxns 33–34). Area between the two horizontal green dotted lines represents the semiclassical range. The reactions 35–38 are not included in both plots (see the text).



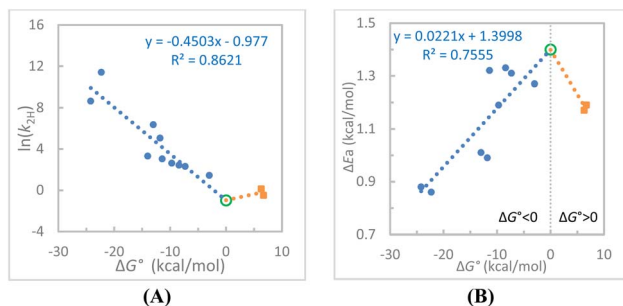


Fig. 3 (A) The linear fit of the  $\ln(k_{2H})-\Delta G^\circ$  data for the 12 hydride transfer reactions with a common HEH hydride donor (Rxnns 23–34). (B) The linear fit of the  $\Delta E_a-\Delta G^\circ$  data for the same 12 exergonic hydride transfer reactions. In both plots, the trend appears to turn to the opposite direction for endergonic reactions 33–34. The empty green circle point in each plot is an extrapolated point at  $\Delta G^\circ = 0$  derived from the linear equation from the exergonic reactions.

reactions with a common hydride donor, HEH (Fig. 3A and B, respectively). In both plots, the endergonic data points are clearly outliers.

As shown in Fig. 3A, the two endergonic reactions appear to turn the linear  $\ln(k_2)-\Delta G^\circ$  correlation derived from the exergonic reactions to the opposite direction. As shown in Fig. 3B, the two endergonic reactions clearly turn the linear  $\Delta E_a-\Delta G^\circ$  correlation for the exergonic reactions to the opposite direction. Using the extrapolated  $\Delta G^\circ = 0$  point from the exergonic reaction data (the circled points in Fig. 3A and B), the corresponding linear correlations with the two data points from the endergonic reactions were drawn, which show a “V-shaped”  $\ln(k_2)-\Delta G^\circ$  relationship and a “Λ-shaped”  $\Delta E_a-\Delta G^\circ$  relationship.

We first discuss about the  $\Delta E_a-\Delta G^\circ$  relationship (Fig. 2B and 3B). Although the current analysis includes only two endergonic reaction data points, this trend could be reinforced by considering additional potential endergonic reactions 35–38. While the  $\Delta G^\circ$  values are not available for these reactions, a qualitative assessment can still be performed. These  $\Delta G^\circ$  values could be approximately derived by substituting the corresponding  $k_2$  values into the linear  $\ln(k_2)-\Delta G^\circ$  correlation equation for the reactions 1–34 (Fig. 2A). For one example, the  $k_2$  for the reaction of BnOH with  $\text{PhXn}^+$  in acetonitrile at 22 °C is  $6.73 \times 10^{-5} \text{ M}^{-1} \text{ s}^{-1}$  (Table 1). Using our reported rate data at other temperatures<sup>24</sup> for an Arrhenius analysis, the rate constant at 25 °C is estimated to be  $8.59 \times 10^{-5} \text{ M}^{-1} \text{ s}^{-1}$ . Substituting the number in the linear fit equation from Fig. 2A yields an estimated  $\Delta G^\circ = 43.4 \text{ kcal mol}^{-1}$ , indicating a highly endergonic process. We recognize that such an estimate could carry large uncertainty as the fit equation used depends upon many factors, such as dataset size, structural similarity among reactions, as well as experiment errors. Therefore, these estimated  $\Delta G^\circ$  values will not be used for quantitative analysis of the  $\Delta E_a-\Delta G^\circ$  relationship. Nevertheless, the  $\Delta E_a$  values for reactions 35–38 (0.80–1.00  $\text{kcal mol}^{-1}$ ) are significantly lower than those of the two endergonic reactions (33–34;  $\sim 1.2 \text{ kcal mol}^{-1}$ ). These observations support the conclusion that the  $\Delta E_a-\Delta G^\circ$  trend for endergonic reactions is opposite to that observed for exergonic reactions.

The key question is why the clear linear  $\Delta E_a-\Delta G^\circ$  plot for the exergonic reactions reverses direction at  $\Delta G^\circ \sim 0$ . This observation is reminiscent of the Marcus inverted region. In fact, the weakest CT complexation (*i.e.*, the longest average DAD) at the TRS is expected from thermoneutral reactions ( $\Delta G = \Delta G_{\text{DAD}}^\circ = 0$ ). This inference is based on Hammond’s postulate. For an exergonic reaction, a more negative  $\Delta G^\circ$  would correspond with a tighter TRS that resembles more reactive reactant structures, whereas for an endergonic reaction, the more positive  $\Delta G^\circ$  would also correspond with a tighter TRS that, however, resembles more reactive products. Therefore, within the VA-AHT-inspired model,  $\Delta G_{\text{DAD}}^\circ$  can be a proxy for TRS rigidity and a Λ-shaped  $\Delta E_a-\Delta G^\circ$  relationship is expected, with the maximum  $\Delta E_a$  occurring near  $\Delta G^\circ = 0$  where the DAD is longest. Our experimental observations (Fig. 2B and considering the  $\Delta E_a$  values of the endergonic reactions 35–38) are largely consistent with this expectation. Therefore, not only the  $\Delta E_a-\Delta G^\circ$  correlation derived from exergonic reactions but the reversed  $\Delta E_a-\Delta G^\circ$  relationship for endergonic reactions also supports our proposed DAD- $\Delta E_a$  relationship. That is, a more flexible system with a longer average DAD and a  $\Delta G^\circ$  closer to zero exhibits a larger  $\Delta E_a$  value, regardless of whether the reaction is exergonic or endergonic.

It should be noted that steric effects can influence the DAD distributions and, consequently, the  $\Delta E_a$  values. Increased steric crowding may enhance system rigidity, leading to a decrease in  $\Delta E_a$ .<sup>25</sup> Conversely, steric hindrance may also physically separate the donor and acceptor, enhancing system flexibility and resulting in higher  $\Delta E_a$  values. Therefore, variations in steric effects across different systems may contribute to the observed scatter in the correlation, in addition to experimental errors.

### The $\ln(k_2)-\Delta G^\circ$ correlation is consistent with the Λ-shaped $\Delta E_a-\Delta G^\circ$ relationship

Another question arises as to why the  $\Delta E_a$  values of the two endergonic reactions decrease (Fig. 3A) but their rates increase (Fig. 3B) as compared to their corresponding values at  $\Delta G^\circ = 0$ . The question can also be answered by considering the two orthogonal but coupled activation processes in the VA-AHT-inspired model, the TRS formation and DAD sampling. Note that TRS formation involves both structural (hybridization and charge) and solvation reorganizations. The observed  $k_2$  is determined by both processes (driven by both  $\Delta G_{\text{TRS}}^\circ$  and  $\Delta G_{\text{DAD}}^\circ$ ) and can be expressed as  $k_2 = (k_{2,\text{TRS}} \times k_{2,\text{DAD}})^{1/2}$  (or more generally, as a weighted geometric mean), where  $k_{2,\text{TRS}}$  is the rate of reaching the TRS with correct donor-acceptor alignment and  $k_{2,\text{DAD}}$  is the rate of reaching the TRS with short DADs; both structural features of the TRS are required for tunneling. Theoretically,  $\ln(k_{\text{TRS}})$  would correlate linearly with the  $\Delta G_{\text{TRS}}^\circ$  across both the exergonic and endergonic regions, which is,  $\ln(k_{\text{TRS}})$  decreases as  $\Delta G_{\text{TRS}}^\circ$  increases from negative through zero to positive values (Fig. 4A). In contrast,  $\ln(k_{\text{DAD}})$  is expected to decrease as exergonic reactions approach thermoneutral reactions due to increasing DADs, but then increase as the reactions become more endergonic due to, however, decreasing



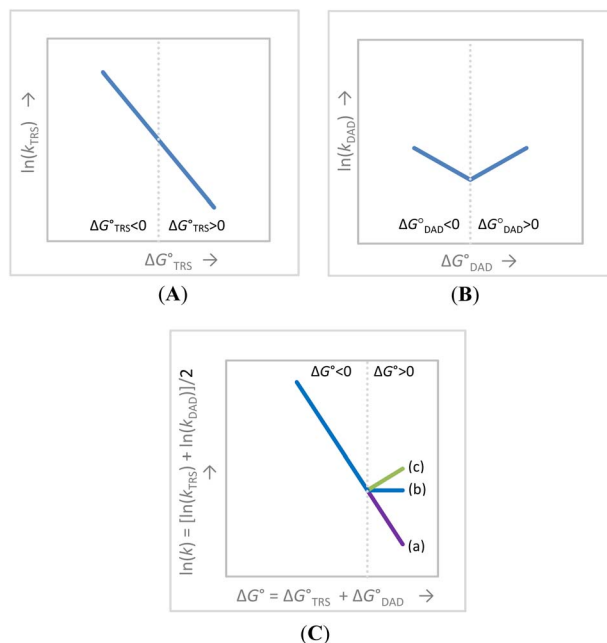


Fig. 4 Predictions from the VA-AHT-inspired model for H-tunneling reactions. (A) The linear  $\ln(k_{\text{TRS}}) - \Delta G^{\circ}_{\text{TRS}}$  correlation involving both endergonic and exergonic reactions. (B) The V-shaped  $\ln(k_{\text{DAD}}) - \Delta G^{\circ}_{\text{DAD}}$  correlation with the turning point at  $\Delta G^{\circ}_{\text{DAD}} = 0$ . (C) The overall  $\ln(k) - \Delta G^{\circ}$  correlation that reflects the combined assumed equal contributions of the plots (A) and (B); case (a): (A) is a steeper line or (B) is a shallow V-shape or both; from cases (b) to (c): (A) becomes flatter or (B) becomes deeper or both. Note that the  $\Lambda$ -shaped  $\Delta E_a - \Delta G^{\circ}_{\text{TRS}}$  relationship is another prediction from the model, mirroring the V-shaped  $\ln(k_{\text{DAD}}) - \Delta G^{\circ}_{\text{DAD}}$  correlation (but  $k_{\text{DAD}}$  is not directly experimentally accessible).

DADs (by Hammond's postulate, as discussed above). In other words, a V-shaped  $\ln(k_{\text{DAD}}) - \Delta G^{\circ}_{\text{DAD}}$  relationship is expected (Fig. 4B). Since the observed  $\ln(k_2) - \Delta G^{\circ}$  correlation is the "sum" of the two relationships (A and B), the observed  $\ln(k_2) - \Delta G^{\circ}$  correlation is expected to show a broken-line profile with a breaking point at  $\Delta G^{\circ} = 0$  (Fig. 4C).

Theoretically, the pattern of the  $\ln(k_2) - \Delta G^{\circ}$  correlation across exergonic and endergonic reactions depends upon the nature of the reaction systems, specifically, the steepness of the linear  $\ln(k_{\text{TRS}}) - \Delta G^{\circ}_{\text{TRS}}$  correlation and the depth of the V-shaped  $\ln(k_{\text{DAD}}) - \Delta G^{\circ}_{\text{DAD}}$  correlation. Fig. 4C(a)–(c) describe three representative patterns. In (a), the  $\ln(k_{\text{TRS}}) - \Delta G^{\circ}_{\text{TRS}}$  linear correlation is steep while the V-shaped  $\ln(k_{\text{DAD}}) - \Delta G^{\circ}_{\text{DAD}}$  correlation is shallow, leading to only a slight deviation at  $\Delta G^{\circ} = 0$ . From (b) to (c), the former correlation becomes less steep and/or the latter becomes deeper, leading to a more evident break and, ultimately a distinctly V-shaped profile. Therefore, all three patterns (a) to (c) are possible. Fig. 3A presents the V-shaped  $\ln(k_2) - \Delta G^{\circ}$  correlation (type (c) pattern). This correlation, along with the  $\Lambda$ -shaped  $\Delta E_a - \Delta G^{\circ}$  correlation in Fig. 3B, appears to support the two-coordinate mechanism proposed in the VA-AHT-inspired model.

It should be noted that patterns (b) and (c) in Fig. 4C also resemble the Marcus inverted region. It is important, however,

to note that relying solely on the  $\ln(k) - \Delta G^{\circ}$  correlation to identify the DAD sampling mechanism needs caution as the bent or V-shaped  $\ln(k_{\text{DAD}}) - \Delta G^{\circ}_{\text{DAD}}$  relationship may be masked by the usually dominant and thus much steeper  $\ln(k_{\text{TRS}}) - \Delta G^{\circ}_{\text{TRS}}$  relationship. The latter relationship reflects structural rehybridization, charge redistribution, and solvent reorganization (pattern (a)), and thus contribute significantly to the activation process. Furthermore, the "bentness" of the  $\ln(k) - \Delta G^{\circ}$  correlation depends on the relative linear correlations in between exergonic and endergonic regions. Each slope is influenced by factors such as electronic and steric properties, structural dynamics, the statistical size of the dataset, as well as experimental uncertainties. Consequently, failure to observe a bent or V-shaped  $\ln(k) - \Delta G^{\circ}$  correlation cannot preclude the existence of a DAD sampling mechanism. Therefore, the seemingly linear  $\ln(k) - \Delta G^{\circ}$  correlation observed for all of the 34 reactions in Fig. 2A cannot be taken as evidence against the existence of the DAD sampling mechanism. Nevertheless, simultaneous observation of both the  $\Lambda$ -shaped  $\Delta E_a - \Delta G^{\circ}$  relationship and V-shaped  $\ln(k) - \Delta G^{\circ}$  relationship for the 12 reactions of HEH (Fig. 3B versus 3A) provides strong support for our DAD- $\Delta E_a$  relationship hypothesis.

Therefore, the  $\Lambda$ -shaped  $\Delta E_a - \Delta G^{\circ}$  correlation and various patterns of the  $\ln(k_2) - \Delta G^{\circ}$  correlation are predicted by the VA-AHT-inspired model, and our experiments are consistent with these predictions. Note that observation of a  $\Lambda$ -shaped  $\Delta E_a - \Delta G^{\circ}$  correlation does not guarantee that an evidently bent or V-shaped  $\ln(k) - \Delta G^{\circ}$  correlation will also be observed. Likewise, a single bent or V-shaped  $\ln(k_2) - \Delta G^{\circ}$  correlation should also be used cautiously to determine the DAD sampling mechanism.

### The $\Lambda$ -shaped $\ln(\text{KIE}) - \Delta G^{\circ}$ correlation is also consistent with the $\Lambda$ -shaped $\Delta E_a - \Delta G^{\circ}$ relationship

With the KIE data in hand, we are also able to study the  $\text{KIE} - \Delta G^{\circ}$  correlation. Study of this relationship has had a long history. In 1960, Melander and Westheimer proposed that the maximum KIE should occur at  $\Delta G^{\circ} = 0$  where the transition state (TS) is symmetric.<sup>75,76</sup> Later, in 1980, Kresge deduced a parabolic relationship linking KIEs to  $(\Delta G^{\circ})^2$  from the Marcus rate theory.<sup>77</sup> In the 1970s, Bell provided an alternative by introducing a tunnel correction to the one-dimensional energy barrier in the TS theory. In his model, tunneling probability is the greatest at a symmetrical TS, leading to the highest KIE, and it decreases as the TS becomes reactant- or product-like.<sup>78</sup> Since the 1980s, Kreevoy and coworkers studied the structure-KIE relationship for the hydride transfer reactions of NADH models in isopropanol/water using the Marcus rate theory that attributes a fraction of the KIE to the corner-cutting tunneling mechanism.<sup>72,74,79–81</sup> Kil and Lee later found that KIEs of several such exergonic hydride transfer reactions increase as  $\Delta G^{\circ}$  becomes less negative close to zero.<sup>82</sup> They used Kreevoy's treatment to explain their results and further predicted that the  $\text{KIE} - \Delta G^{\circ}$  relationship would reverse upon entering the endergonic region, which has, however, not yet been experimentally supported due to lack of experiment data. Thus, this latter model also predicts a similar bell-shaped (or "upward/



downward”) dependence of KIEs on  $\Delta G^\circ$ , with maximal tunneling and thus the largest KIE occurring near  $\Delta G^\circ \sim 0$ , where the effective barrier is highest.

Within the VA-AHT(-inspired) model, the KIE is defined differently. It is primarily dependent upon the difference in isotopic wave-function overlaps at the TRSs over a spectrum of DADs.<sup>6,8,40,59</sup> Because the vibrational wave-packet of H is more diffuse than that of the D isotope, the H-overlap is more than the D-overlap so that  $\text{KIE} > 1$ <sup>40,83</sup> (in other words, because D-tunneling requires shorter DADs than H-tunneling,<sup>40,83–86</sup>  $k_{\text{D,DAD}} < k_{\text{H,DAD}}$ ). Moreover, since the overlap for D-transfer decreases more rapidly than that for H-transfer with increasing DAD, the KIE increases with DAD (in other words, the difference between  $k_{\text{D,DAD}}$  and  $k_{\text{H,DAD}}$  becomes larger as the DAD increases). Consequently, the KIE is predicted to be the largest at  $\Delta G^\circ = 0$  where the DAD is the longest, and to decrease as the TRS becomes reactant- or product-like where the DAD shortens. Since the  $\ln(k_{\text{DAD}}) - \Delta G^\circ_{\text{DAD}}$  relationship is V-shaped (Fig. 4B), and  $k_{\text{TRS}}$  is mostly isotope insensitive, the  $\ln(\text{KIE}) - \Delta G^\circ_{\text{DAD}}$  relationship is expected to be  $\Lambda$ -shaped.

We examine the  $\ln(\text{KIE}) - \Delta G^\circ$  correlation to indirectly test the predicted  $\Lambda$ -shaped  $\ln(\text{KIE}) - \Delta G^\circ_{\text{DAD}}$  relationship. The 34  $\ln(\text{KIE}) - \Delta G^\circ$  data points and the 12 such data points for the reactions of HEH only, are plotted in Fig. 5A and B, respectively. Linear fits were applied separately to the exergonic and endergonic regions, with the  $\Delta G^\circ = 0$  point extrapolated from the exergonic data. In both plots, the endergonic reactions appear to reverse the trend observed for the exergonic reactions. The reverse trend could be further supported by including the smaller KIE values (3.63–4.83) from the endergonic reactions between alcohols and  $\text{PhXn}^+$  (Rxnns 35–38), but again because of lack of the corresponding  $\Delta G^\circ$  values these KIE data cannot be used for quantitative fitting analysis.

The  $\Lambda$ -shaped  $\ln(\text{KIE}) - \Delta G^\circ$  relationship is consistent with the  $\Lambda$ -shaped  $\Delta E_a - \Delta G^\circ$  relationship in that both  $\Delta E_a$ s and KIEs reach maxima near thermoneutral conditions (Fig. 5 versus 3B). Importantly, both relationships can be uniformly explained using the DAD coordinate mechanism within the VA-AHT-inspired model. According to this model, the KIE increases with

the DAD. Note that previous studies on enzymatic reactions as well as  $\text{NADH}/\text{NAD}^+$  model reactions have showcased this KIE–DAD relationship for exergonic reactions (ref. 48 and references cited therein). Therefore, the maxima in both the KIE and  $\Delta E_a$  occurring near  $\Delta G^\circ = 0$  correspond to the longest DADs in H-tunneling mechanisms.

We noticed that the slopes of the  $\Delta E_a - \Delta G^\circ$  (0.0268) and  $\ln(\text{KIE}) - \Delta G^\circ$  (0.0196) correlations for the exergonic reactions are significantly small (Fig. 2B and 5A). This likely partly reflects the fact that  $\Delta E_a$  and  $\ln(\text{KIE})$  correlate only with the  $\Delta G^\circ_{\text{DAD}}$ , which constitutes only a small fraction of the overall  $\Delta G^\circ$  in this particular class of reactions, so that they are much less sensitive to the overall  $\Delta G^\circ$ .

## Conclusions

Free-energy dependences of  $\Delta E_a$ , rates ( $\ln(k)$ ), and  $\ln(\text{KIE})$  were analysed with 34 hydride tunneling reactions of  $\text{NADH}/\text{NAD}^+$  models between two carbons in acetonitrile, which span both exergonic and endergonic regions. A clear linear  $\Delta E_a - \Delta G^\circ$  relationship was, for the first time, observed for the exergonic reactions, with  $\Delta E_a$  increasing as  $\Delta G^\circ$  approaches zero. A quantitative relationship has not yet been obtained for the limited number of endergonic reactions, but reversal of the trend from the exergonic reactions including reactions 35–38 has been clearly shown. This is consistent with our hypothesis regarding the DAD– $\Delta E_a$  relationship.  $\Delta G^\circ$  (strictly,  $\Delta G^\circ_{\text{DAD}}$ ) acts as an indicator of system rigidity (or DAD distributions), where thermoneutral reactions correspond to the most flexible and longest DADs. Overall, a portion of  $\Delta G^\circ$  of the reactions modulates the CT complexation vibrations and thus DAD sampling, which is directly related to the  $T$ -dependence of KIEs. This resulting  $\Lambda$ -shaped  $\Delta E_a - \Delta G^\circ$  relationship agrees with the prediction from the VA-AHT-inspired model that involves both the TRS formation and DAD sampling processes and attributes the KIE to only the latter DAD sampling process.

The second prediction from the same model includes the bent- or V-shaped  $\ln(k) - \Delta G^\circ$  relationship with the breaking/turning point at  $\Delta G^\circ = 0$ . It was found that the  $\ln(k) - \Delta G^\circ$  plot is linear across the 34 reactions, but when using the data from the 12 reactions with a common hydride donor HEH, the V-shaped  $\ln(k) - \Delta G^\circ$  relationship emerges, which mirrors the  $\Lambda$ -shaped  $\Delta E_a - \Delta G^\circ$  relationship found from the same series of the reactions. The latter also appears to agree with the DAD sampling mechanism within the VA-AHT-inspired model.

The third prediction from the model includes the  $\Lambda$ -shaped  $\ln(\text{KIE}) - \Delta G^\circ$  relationship. It was found that the  $\ln(\text{KIE}) - \Delta G^\circ$  plot is linear across the 32 exergonic reactions, but when using the endergonic reactions, the correlation is reversed. This  $\Lambda$ -shaped trend is the same when using the 12 reactions of HEH. These results are consistent with the prediction. Therefore, all of the three predictions from the VA-AHT-inspired model, including the V-shaped  $\ln(k) - \Delta G^\circ$ ,  $\Lambda$ -shaped  $\Delta E_a - \Delta G^\circ$ , and  $\Lambda$ -shaped  $\ln(\text{KIE}) - \Delta G^\circ$  relationships, were simultaneously found in the 12 reactions of HEH, which support our DAD– $\Delta E_a$  relationship hypothesis.

Caution is required when using the  $\ln(k) - \Delta G^\circ$  correlation alone to identify the DAD sampling mechanism as the expected

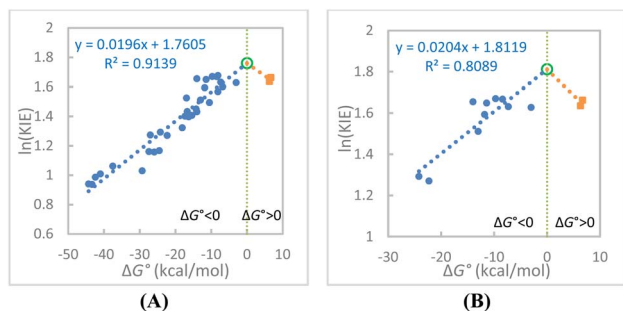


Fig. 5 (A) The linear fit of the  $\ln(\text{KIE}(25\text{ }^\circ\text{C})) - \Delta G^\circ$  data for the 34 hydride transfer reactions. The reactions 35–38 are not included (see the text). (B) The linear fit of the  $\ln(\text{KIE}(25\text{ }^\circ\text{C})) - \Delta G^\circ$  data for the 12 hydride transfer reactions of a common hydride donor HEH (Rxnns 23–34). The empty green circle point in each plot is an extrapolated point at  $\Delta G^\circ = 0$  derived from the linear equation from the exergonic reactions.



bent- or V-shaped correlations can be largely masked by the dominant contribution from TRS formation, as well as by system selection and experimental uncertainties. Conversely, observation of the bent- or V-shaped  $\ln(k)-\Delta G^\circ$  correlations alone should also be cautiously used to propose the DAD sampling mechanism, since  $\Delta G_{\text{DAD}}^\circ$  could account for a small portion of the  $\Delta G^\circ$  so that system selection and experimental uncertainty need to be carefully considered. Also, a  $\Lambda$ -shaped  $\ln(\text{KIE})-\Delta G^\circ$  relationship alone should not be considered definitive evidence for DAD sampling either, since similar trends can arise from the Marcus theory combined with the corner-cutting tunneling mechanism process, or even possibly from the classical Melander–Westheimer's "bond-stretching" model. In sharp contrast, the newly identified  $\Lambda$ -shaped  $\Delta E_{\text{a}}-\Delta G^\circ$  relationship appearing to arise only from the DAD sampling could provide evidence for the DAD sampling mechanism; however, whether or not this particular relationship is explainable by other tunnelling models remains unclear.

The small slopes of the  $\Lambda$ -shaped  $\Delta E_{\text{a}}-\Delta G^\circ$  and  $\ln(\text{KIE})-\Delta G^\circ$  relationships may suggest that only a small portion of the  $\Delta G^\circ$  contributes to the activated DAD sampling process. The DAD increases as  $\Delta G^\circ$  of the exergonic reactions becomes less negative, becomes the largest with thermoneutral reactions, and decreases as  $\Delta G^\circ$  becomes more positive in the endergonic reactions. This  $\Lambda$ -shaped  $\Delta E_{\text{a}}-\Delta G^\circ$  relationship should be useful for evaluating the existing H-tunneling models and help develop future H-transfer/tunneling theories.

Overall, our  $\Delta E_{\text{a}}-\Delta G^\circ$  correlation results provide support for the DAD– $\Delta E_{\text{a}}$  relationship in both the VA-AHT and VA-AHT-inspired models; systems with more densely distributed small DADs give rise to smaller  $T$ -dependence of KIEs. Side-by-side comparison of solution-phase and enzymatic reactions on KIEs and their  $T$ -dependences suggests that frequently observed  $T$ -independence of KIEs from enzymes can be attributed to strong constructive (fast) protein dynamics (modulated by  $\Delta G_{\text{DAD}}^\circ$ ) that compress the donor and acceptor close to each other for H-tunneling to occur. Therefore, there appears a synergy between C–H activation (for TRS formation and DAD sampling) and diverse protein dynamics (encoded in  $\Delta G_{\text{TRS}}^\circ$  and  $\Delta G_{\text{DAD}}^\circ$ , respectively) cooperatively functioning in enzymatic reactions.

## Author contributions

A. A.-K., N. D., B. D. and J. S. performed organic synthesis and kinetic measurements. A. A.-K. and N. D. wrote part of the paper. B. D. analyzed the data. L. P. and S. P. performed organic synthesis. Y. L. designed research and wrote the manuscript.

## Conflicts of interest

The authors declare no competing financial interests.

## Data availability

The data underlying this study are available in the published article and its supplementary information (SI). Supplementary information: synthesis, procedures to determine the rate

constants,  $T$ -dependence of KIE plots, tabulated raw rate constant data, and the primary kinetic experiment data. See DOI: <https://doi.org/10.1039/d6sc01847e>.

## Acknowledgements

Acknowledgment is made to the donor of the National Institutes of Health (NIH R15 GM148951). We thank Praticchhya Adhikari (Southern Illinois University Edwardsville) for helping collect part of the kinetic data for the reaction of MPH with  $p$ -MeOPhXn<sup>+</sup>, and Binita Maharaj and Peter Maness (from the same institution) for initial kinetic study of the reaction of MAH with Tr<sup>+</sup>.

## References

- 1 A. Kohen, R. Cannio, S. Bartolucci and J. P. Klinman, Enzyme dynamics and hydrogen tunnelling in a thermophilic alcohol dehydrogenase, *Nature*, 1999, **399**, 496–499.
- 2 M. J. Knapp, K. Rickert and J. P. Klinman, Temperature-dependent isotope effects in soybean lipoxygenase-1: Correlating hydrogen tunneling with protein dynamics, *J. Am. Chem. Soc.*, 2002, **124**, 3865–3874.
- 3 R. S. Sikorski, L. Wang, K. A. Markham, P. T. R. Rajagopalan, S. J. Benkovic and A. Kohen, Tunneling and coupled motion in the *E. coli* dihydrofolate reductase catalysis, *J. Am. Chem. Soc.*, 2004, **126**, 4778–4779.
- 4 E. J. Loveridge, R. M. Evans and R. K. Allemann, Solvent Effects on Environmentally Coupled Hydrogen Tunnelling During Catalysis by Dihydrofolate Reductase from *Thermotoga maritima*, *Chem.–Eur. J.*, 2008, **14**(34), 10782–10788.
- 5 Z. D. Nagel and J. P. Klinman, Update 1 of: Tunneling and dynamics in enzymatic hydride transfer, *Chem. Rev.*, 2010, **110**, PR41–PR67.
- 6 C. R. Pudney, O. L. Johannissen, M. J. Sutcliffe, S. Hay and N. S. Scrutton, Direct Analysis of Donor-Acceptor Distance and Relationship to Isotope Effects and the Force Constant for Barrier Compression in Enzymatic H-Tunneling Reactions, *J. Am. Chem. Soc.*, 2010, **132**, 11329–11335.
- 7 Z. Wang, P. N. Singh, M. C. Czekster, A. Kohen and V. L. Schramm, Protein Mass-Modulated Effects in the Catalytic Mechanism of Dihydrofolate Reductase: Beyond Promoting Vibrations, *J. Am. Chem. Soc.*, 2014, **136**, 8333–8341.
- 8 A. Kohen, Role of Dynamics in Enzyme Catalysis: Substantial vs. Semantic Controversies, *Acc. Chem. Res.*, 2015, **48**, 466–473.
- 9 E. Romero, S. T. Ladani, D. Hamelberg and G. Gadda, Solvent-Slaved Motions in the Hydride Tunneling Reaction Catalyzed by Human Glycolate Oxidase, *ACS Catal.*, 2016, **6**, 2113–2120.
- 10 J. P. Klinman and A. R. Offenbacher, Understanding Biological Hydrogen Transfer Through the Lens of Temperature Dependent Kinetic Isotope Effects, *Acc. Chem. Res.*, 2018, **51**, 1966–1974.



- 11 G. W. Howe and W. A. van der Donk, Temperature-Independent Kinetic Isotope Effects as Evidence for a Marcus-like Model of Hydride Tunneling in Phosphite Dehydrogenase, *Biochemistry*, 2019, **58**, 4260–4268.
- 12 P. Singh, A. Vandemeulebroucke, J. Li, C. Schulenburg, G. Fortunato, A. Kohen, D. Hilvert and C. M. Cheatum, Evolution of the Chemical Step in Enzyme Catalysis, *ACS Catal.*, 2021, **11**, 6726–6732.
- 13 E. Hatcher, A. V. Soudackov and A. Hammes-Schiffer, Proton-Coupled Electron Transfer in Soybean Lipoxygenase: Dynamical Behavior and Temperature Dependence of Kinetic Isotope Effects, *J. Am. Chem. Soc.*, 2007, **129**, 187–196.
- 14 S. Hu, A. V. Soudackov, S. Hammes-Schiffer and J. P. Klinman, Enhanced Rigidification within a Double Mutant of Soybean Lipoxygenase Provides Experimental Support for Vibronically Nonadiabatic Proton-Coupled Electron Transfer Models, *ACS Catal.*, 2017, **7**, 3569–3574.
- 15 S. Hammes-Schiffer and S. J. Benkovic, Relating protein motion to catalysis, *Annu. Rev. Biochem.*, 2006, **75**, 519–541.
- 16 K. A. Henzler-Wildman, M. Lei, V. Thai, S. J. Kerns, M. Karplus and D. Kern, A Hierarchy of Timescales in Protein Dynamics Is Linked to Enzyme Catalysis, *Nature*, 2007, **450**, 913–916.
- 17 S. Hay and N. S. Scrutton, Good vibrations in enzyme-catalysed reactions, *Nat. Chem.*, 2012, **4**, 161–168.
- 18 J. P. Klinman and A. Kohen, Hydrogen Tunneling Links Protein Dynamics to Enzyme Catalysis, *Annu. Rev. Biochem.*, 2013, **82**, 471–496.
- 19 V. L. Schramm and S. D. Schwartz, Promoting vibrations and the function of enzymes. Emerging theoretical and experimental convergence, *Biochemistry*, 2018, **57**, 3299–3308.
- 20 J. P. Klinman, S. M. Miller and N. G. J. Richards, A Foundational Shift in Models for Enzyme Function, *J. Am. Chem. Soc.*, 2025, **147**, 14884–14904.
- 21 S. Hay, M. J. Sutcliffe and N. S. Scrutton, Promoting motions in enzyme catalysis probed by pressure studies of kinetic isotope effects, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 507–512.
- 22 J. Bandaria, S. Dutta, M. W. Nydegger, W. Rock, A. Kohen and C. Cheatum, Characterizing the dynamics of functionally relevant complexes of formate dehydrogenase, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 17974–17979.
- 23 E. J. Loveridge, L.-H. Tey and R. K. Allemann, Solvent Effects on Catalysis by *Escherichia coli* Dihydrofolate Reductase, *J. Am. Chem. Soc.*, 2010, **132**, 1137–1143.
- 24 Q. Liu, Y. Zhao, B. Hammann, J. Eilers, Y. Lu and A. Kohen, A Model Reaction Assesses Contribution of H-Tunneling and Coupled Motions to Enzyme Catalysis, *J. Org. Chem.*, 2012, **77**, 6825–6833.
- 25 Y. Lu, S. Wilhelm, M. Bai, P. Maness and L. Ma, Replication of the Enzymatic Temperature Dependency of the Primary Hydride Kinetic Isotope Effects in Solution: Caused by the Protein Controlled Rigidity of the Donor-Acceptor Centers?, *Biochemistry*, 2019, **58**, 4035–4046.
- 26 D. Antoniou and S. D. Schwartz, Role of protein motions in catalysis by Formate Dehydrogenase, *J. Phys. Chem. B*, 2020, **124**, 9483–9489.
- 27 M. Bai, P. Rijal, S. Salarvand and Y. Lu, Correlation of Temperature Dependence of Hydride Kinetic Isotope Effects with Donor-Acceptor Distances in Two Solvents of Different Polarities, *Org. Biomol. Chem.*, 2023, **21**, 5090–5097.
- 28 S. D. Schwartz, Protein Dynamics and Enzymatic Catalysis, *J. Phys. Chem. B*, 2023, **127**, 2649–2660.
- 29 G. Singh, A. Austin, M. Bai, J. Bradshaw, B. A. Hammann, D. E. K. Kabotso and Y. Lu, Study of the Effects of Remote Heavy Group Vibrations on the Temperature Dependence of Hydride Kinetic Isotope Effects of the NADH/NAD<sup>+</sup> Model Reactions, *ACS Omega*, 2024, **9**, 20593–20600.
- 30 S. Hammes-Schiffer, Explaining Kinetic Isotope Effects in Proton-Coupled Electron Transfer Reactions, *Acc. Chem. Res.*, 2025, **58**, 1335–1344.
- 31 S. Hu, A. R. Offenbacher, E. M. Thompson, C. L. Gee, J. Wilcoxon, C. A. M. Carr, D. M. Prigozhin, V. Yang, T. Alber, R. D. Britt, J. S. Fraser and J. P. Klinman, Biophysical Characterization of a Disabled Double Mutant of Soybean Lipoxygenase: The “Undoing” of Precise Substrate Positioning Relative to Metal Cofactor and an Identified Dynamical Network, *J. Am. Chem. Soc.*, 2019, **141**, 1555–1567.
- 32 J. Pu, S. Ma, J. Gao and D. G. Truhlar, Small temperature dependence of the kinetic isotope effect for the hydride transfer reaction catalyzed by *Escherichia coli* dihydrofolate reductase, *J. Phys. Chem. B*, 2005, **109**, 8551–8556.
- 33 H. Liu and A. Warshel, Origin of the Temperature Dependence of Isotope Effects in Enzymatic Reactions: The Case of Dihydrofolate Reductase, *J. Phys. Chem. B*, 2007, **111**, 7852–7861.
- 34 S. C. L. Kamerlin and A. Warshel, An analysis of all the relevant facts and arguments indicates that enzyme catalysis does not involve large contributions from nuclear tunneling, *J. Phys. Org. Chem.*, 2010, **23**, 677–684.
- 35 R. A. More O’Ferrall, Introduction to a symposium in print on tunnelling, *J. Phys. Org. Chem.*, 2010, **23**, 559–560.
- 36 G. D. Truhlar, Tunneling in enzymatic and nonenzymatic hydrogen transfer reactions, *J. Phys. Org. Chem.*, 2010, **23**, 660–676.
- 37 Z. Wang and A. Kohen, Thymidylate synthase catalyzed H-transfers: Two chapters in one tale, *J. Am. Chem. Soc.*, 2010, **132**, 9820–9825.
- 38 C. Ranasinghe, P. Pagano, P. J. Sapienza, A. L. Lee, A. Kohen and C. M. Cheatum, Isotopic Labeling of Formate Dehydrogenase Perturbs the Protein Dynamics, *J. Phys. Chem. B*, 2019, **123**, 10403–10409.
- 39 M. Horitani, A. R. Offenbacher, C. A. Marcus Carr, T. Yu, V. Hoeke, G. E. Cutsail, S. Hammes-Schiffer, J. P. Klinman and B. M. Hoffman, <sup>13</sup>C ENDOR Spectroscopy of Lipoxygenase–Substrate Complexes Reveals the Structural Basis for C–H Activation by Tunneling, *J. Am. Chem. Soc.*, 2017, **139**, 1984–1997.



- 40 D. Roston and A. Kohen, Elusive transition state of alcohol dehydrogenase unveiled, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 9572–9577.
- 41 N. Kanaan, S. Ferrer, S. Martí, M. Garcia-Viloca, A. Kohen and V. Moliner, Temperature Dependence of the Kinetic Isotope Effects in Thymidylate Synthase. A Theoretical Study, *J. Am. Chem. Soc.*, 2011, **133**, 6692–6702.
- 42 A. R. Mhashal and D. T. Major, Temperature-Dependent Kinetic Isotope Effects in R67 Dihydrofolate Reductase from Path-Integral Simulations, *J. Phys. Chem. B*, 2021, **125**, 1369–1377.
- 43 J. P. Layfield and S. Hammes-Schiffer, Hydrogen tunneling in enzymes and biomimetic models, *Chem. Rev.*, 2014, **114**, 3466–3494.
- 44 P. Maness, S. Koirala, P. Adhikari, N. Salimraftar and Y. Lu, Substituent Effects on Temperature Dependence of Kinetic Isotope Effects in Hydride-Transfer Reactions of NADH/NAD<sup>+</sup> Analogues in Solution: Reaction Center Rigidity Is the Key, *Org. Lett.*, 2020, **22**, 5963–5967.
- 45 M. Bai, S. Koirala and Y. Lu, Direct Correlation Between Donor-Acceptor Distance and Temperature Dependence of Kinetic Isotope Effects in Hydride-Tunneling Reactions of NADH/NAD<sup>+</sup> Analogues, *J. Org. Chem.*, 2021, **86**, 7500–7507.
- 46 P. Adhikari, M. Song, M. Bai, P. Rijal, N. DeGroot and Y. Lu, Solvent Effects on the Temperature Dependence of Hydride Kinetic Isotope Effects: Correlation to the Donor-Acceptor Distances, *J. Phys. Chem. A*, 2022, **126**, 7675–7686.
- 47 A. Beach, P. Adhikari, G. Singh, M. Song, N. DeGroot and Y. Lu, Structural Effects on the Temperature Dependence of Hydride Kinetic Isotope Effects of the NADH/NAD<sup>+</sup> Model Reactions in Acetonitrile: Charge-Transfer Complex Tightness Is a Key, *J. Org. Chem.*, 2024, **89**, 3184–3193.
- 48 A. Austin, J. Sager, L. Phan and Y. Lu, Structural Effects on the Hydride-Tunneling Kinetic Isotope Effects of NADH/NAD<sup>+</sup> Model Reactions: Relating to the Donor-Acceptor Distances, *J. Org. Chem.*, 2025, **90**, 3110–3115.
- 49 M. Bai, G. Singh and Y. Lu, Rigidity Analysis of Hydride Tunneling Ready States from Secondary Kinetic Isotope Effects and Hammett Correlations: Relating to the Temperature Dependence of Kinetic Isotope Effects, *J. Phys. Org. Chem.*, 2025, **38**, DOI: [10.1002/poc.70002](https://doi.org/10.1002/poc.70002).
- 50 B. Pokhrel, J. Sager, P. Adhikari, G. Singh, B. Dhakal and Y. Lu, Large Temperature Dependence of Large Kinetic Isotope Effects of Multistep Hydride Reduction of p-Chloranil by NADH Models in Acetonitrile: Proton Tunneling within Loose Radical Ion-Pairs, *J. Org. Chem.*, 2025, 16912–16917.
- 51 D. Roston, C. M. Cheatum and A. Kohen, Hydrogen Donor-Acceptor Fluctuations from Kinetic Isotope Effects: A Phenomenological Model, *Biochemistry*, 2012, **51**, 6860–6870.
- 52 V. Stojković, L. Perissinotti, D. Willmer, S. Benkovic and A. Kohen, Effects of the donor acceptor distance and dynamics on hydride tunneling in the dihydrofolate reductase catalyzed reaction, *J. Am. Chem. Soc.*, 2012, **134**, 1738–1745.
- 53 P. Pagano, Q. Guo, C. Ranasinghe, E. Schroeder, K. Robben, F. Häse, H. Ye, K. Wickersham, A. Aspuru-Guzik, D. T. Major, L. Gakhar, A. Kohen and C. M. Cheatum, Oscillatory Active-Site Motions Correlate with Kinetic Isotope Effects in Formate Dehydrogenase, *ACS Catal.*, 2019, **9**, 11199–11206.
- 54 M. J. Knapp and J. P. Klinman, Environmentally coupled hydrogen tunneling. Linking catalysis to dynamics, *Eur. J. Biochem.*, 2002, **269**, 3113–3121.
- 55 N. Agrawal, B. Hong, C. Mihai and A. Kohen, Vibrationally Enhanced Hydrogen Tunneling in the *Escherichia coli* Thymidylate Synthase Catalyzed Reaction, *Biochemistry*, 2004, **43**, 1998–2006.
- 56 Z. D. Nagel, C. W. Meadows, M. Dong, B. J. Bahnson and J. P. Klinman, Active Site Hydrophobic Residues Impact Hydrogen Tunneling Differently in a Thermophilic Alcohol Dehydrogenase at Optimal versus Nonoptimal Temperatures, *Biochemistry*, 2012, **51**, 4147–4156.
- 57 K. Francis, P. Sapienza, A. Lee and A. Kohen, The Effect of Protein Mass Modulation on Human Dihydrofolate Reductase, *Biochemistry*, 2016, **55**, 1100–1106.
- 58 A. Geddes, C. E. Paul, S. Hay, F. Hollmann and N. S. Scrutton, Donor–Acceptor Distance Sampling Enhances the Performance of “Better than Nature” Nicotinamide Coenzyme Biomimetics, *J. Am. Chem. Soc.*, 2016, **138**, 11089–11092.
- 59 J. P. Klinman, A new model for the origin of kinetic hydrogen isotope effects, *J. Phys. Org. Chem.*, 2010, **23**, 606–612.
- 60 R. K. Roy, D. Antoniou and S. D. Schwartz, The Shaping of Enzymatic Free Energy Barriers through the Creation of Rate-Promoting Vibrations via Inter-Residue Cross-Talk on Multiple Time Scales, *J. Phys. Chem. B*, 2025, **129**, 9973–9982.
- 61 M. Delgado, S. Görlich, J. E. Longbotham, N. S. Scrutton, S. Hay, V. Moliner and I. Tuñón, Convergence of Theory and Experiment on the Role of Preorganization, Quantum Tunneling, and Enzyme Motions into Flavoenzyme-Catalyzed Hydride Transfer, *ACS Catal.*, 2017, **7**, 3190–3198.
- 62 P. K. Agarwal, S. R. Billeter, P. T. R. Rajagopalan, S. J. Benkovic and S. Hammes-Schiffer, Network of coupled promoting motions in enzyme catalysis, *Proc. Natl. Acad. Sci. U. S. A.*, 2002, **99**, 2794–2799.
- 63 E. Hatcher, A. V. Soudackov and S. Hammes-Schiffer, Proton-coupled electron transfer in soybean lipoxygenase, *J. Am. Chem. Soc.*, 2004, **126**, 5763–5775.
- 64 B. Salna, A. Benabbas, D. Russo and P. M. Champion, Tunneling kinetics and nonadiabatic proton-coupled electron transfer in proteins: The effect of electric fields and anharmonic donor-acceptor interactions, *J. Phys. Chem. B*, 2017, **121**, 6869–6881.
- 65 P. Li, A. V. Soudackov and S. Hammes-Schiffer, Fundamental insights into proton-coupled electron transfer in soybean lipoxygenase from quantum mechanical/molecular mechanics free energy simulations, *J. Am. Chem. Soc.*, 2018, **140**, 3068–3076.
- 66 J. Pang, J. Pu, J. Gao, D. G. Truhlar and R. K. Allemann, Hydride Transfer Reaction Catalyzed by Hyperthermophilic Dihydrofolate Reductase Is Dominated by Quantum Mechanical Tunneling and Is Promoted by Both Inter- and



- Intramonomeric Correlated Motions, *J. Am. Chem. Soc.*, 2006, **128**, 8015–8023.
- 67 X.-Q. Zhu, H.-L. Zou, P.-W. Yuan, Y. Liu, L. Cao and J.-P. Cheng, A detailed investigation into the oxidation mechanism of Hantzsch 1,4-dihydropyridines by ethyl -cyanocinnamates and benzylidenemalononitriles, *J. Chem. Soc., Perkin Trans. 2*, 2000, 1857–1861.
- 68 G. B. Shen, K. Xia, X. T. Li, J. L. Li, Y. H. Fu, L. Yuan and X. Q. Zhu, Prediction of Kinetic Isotope Effects for Various Hydride Transfer Reactions Using a New Kinetic Model, *J. Phys. Chem. A*, 2016, **120**, 1779–1799.
- 69 Y. Lu, F. Qu, B. Moore, D. Endicott and W. Kuester, Hydride Reduction of NAD<sup>+</sup> Analogues by Isopropyl Alcohol: Kinetics, Deuterium Isotope Effects and Mechanism, *J. Org. Chem.*, 2008, **73**, 4763–4770.
- 70 Y. Lu, F. Qu, Y. Zhao, A. M. Small, J. Bradshaw and B. Moore, Kinetics of the hydride reduction of an NAD(+) analogue by isopropyl alcohol in aqueous and acetonitrile solutions: solvent effects, deuterium isotope effects, and mechanism, *J. Org. Chem.*, 2009, **74**, 6503–6510.
- 71 Y. Lu, J. Bradshaw, Y. Zhao, W. Kuester and D. Kabotso, Structure–reactivity relationship for alcohol oxidations via hydride transfer to a carbocationic oxidizing agent, *J. Phys. Org. Chem.*, 2011, **24**, 1172–1178.
- 72 M. M. Kreevoy, D. Ostovic and D. G. Truhlar, Phenomenological manifestations of large-curvature tunneling in hydride-transfer reactions, *J. Phys. Chem.*, 1986, **90**, 3766–3774.
- 73 A. Kohen and J. P. Klinman, Enzyme catalysis: beyond classical paradigms, *Acc. Chem. Res.*, 1998, **31**, 397–404.
- 74 I.-S. Han Lee, E. H. Jeoung and M. M. Kreevoy, Primary Kinetic Isotope Effects on Hydride Transfer from 1,3-Dimethyl-2-phenylbenzimidazole to NAD<sup>+</sup> Analogues, *J. Am. Chem. Soc.*, 2001, **123**, 7492–7496.
- 75 L. Melander, in *Isotope Effects on Reaction Rates*, Ronald Press, New York, 1960, pp. 24–32.
- 76 F. H. Westheimer, THE magnitude of the Primary kinetic isotope effect for compounds of hydrogen and deuterium, *Chem. Rev.*, 1961, **61**, 265–273.
- 77 M. M. Kreevoy and S. W. Oh, Relation between rate and equilibrium constants for proton-transfer reactions, *J. Am. Chem. Soc.*, 1973, **95**, 4805–4810.
- 78 R. P. Bell, Liversidge lecture. Recent advances in the study of kinetic hydrogen isotope effects, *Chem. Soc. Rev.*, 1974, **3**, 513–544.
- 79 Y. Kim, D. G. Truhlar and M. M. Kreevoy, An experimentally based family of potential energy surfaces for hydride transfer between NAD<sup>+</sup> analogs, *J. Am. Chem. Soc.*, 1991, **113**, 7837–7847.
- 80 Y. Kim and M. M. Kreevoy, The experimental manifestations of corner-cutting tunneling, *J. Am. Chem. Soc.*, 1992, **114**, 7116–7123.
- 81 I.-S. H. Lee, E. H. Jeoung and M. M. Kreevoy, Marcus Theory of a Parallel Effect on R for Hydride Transfer Reaction between NAD<sup>+</sup> Analogues, *J. Am. Chem. Soc.*, 1997, **119**, 2722–2728.
- 82 H. J. Kil and I.-S. H. Lee, Primary Kinetic Isotope Effects on Hydride Transfer from Heterocyclic Compounds to NAD + Analogues, *J. Phys. Chem. A*, 2009, **113**, 10704–10709.
- 83 D. Roston and A. Kohen, A Critical Test of the “Tunneling and Coupled Motion” Concept in Enzymatic Alcohol Oxidation, *J. Am. Chem. Soc.*, 2013, **135**, 13624–13627.
- 84 S. Kashefolgheta, M. Razzaghi, B. Hammann, J. Eilers, D. Roston and Y. Lu, Computational Replication of the Abnormal Secondary Kinetic Isotope Effects in a Hydride Transfer Reaction in Solution with a Motion Assisted H-Tunneling Model, *J. Org. Chem.*, 2014, **79**, 1989–1994.
- 85 B. Maharjan, M. R. Boroujeni, J. Lefton, O. R. White, M. Razzaghi, B. A. Hammann, M. Derakhshani-Molayousefi, J. E. Eilers and Y. Lu, Steric Effects on the Primary Isotope Dependence of Secondary Kinetic Isotope Effects in Hydride Transfer Reactions in Solution: Caused by the Isotopically Different Tunneling Ready State Conformations?, *J. Am. Chem. Soc.*, 2015, **137**, 6653–6661.
- 86 M. Derakhshani-Molayousefi, S. Kashefolgheta, J. E. Eilers and Y. Lu, Computational Replication of the Primary Isotope Dependence of Secondary Kinetic Isotope Effects in Solution Hydride-Transfer Reactions: Supporting the Isotopically Different Tunneling Ready State Conformations, *J. Phys. Chem. A*, 2016, **120**, 4277–4284.

