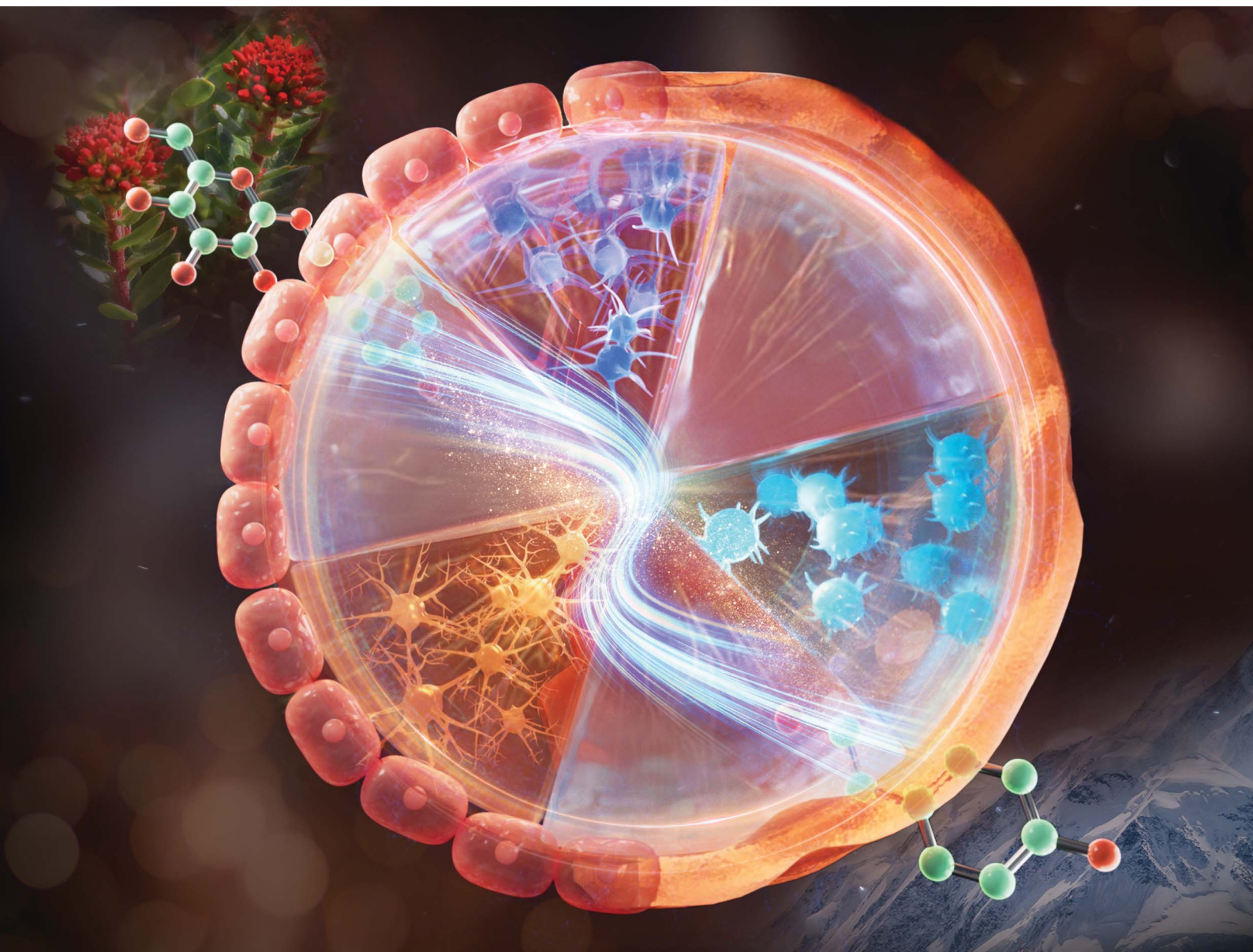


Chemical Science

Volume 17
Number 13
1 April 2026
Pages 6245–6788

rsc.li/chemical-science



ISSN 2041-6539

EDGE ARTICLE

Zengnan Wu, Xianli Meng, Jin-Ming Lin *et al.*
Microfluidic-mass spectrometry analysis of blood-brain
barrier transport using engineered microparticle interfaces

Cite this: *Chem. Sci.*, 2026, 17, 6371 All publication charges for this article have been paid for by the Royal Society of Chemistry

Microfluidic-mass spectrometry analysis of blood–brain barrier transport using engineered microparticle interfaces

Shiyu Chen,^{ab} Yingrui Zhang,^{bd} Zengnan Wu,^{id}*^b Tianze Xie,^b Tong Xu,^{id}^d Yi Zhang,^d Xianli Meng*^c and Jin-Ming Lin^{id}*^b

The blood–brain barrier (BBB) is vital for maintaining central nervous system (CNS) homeostasis but represents a formidable obstacle to drug delivery, underscoring the need for physiologically relevant *in vitro* models for CNS drug screening. However, existing *in vitro* BBB models remain limited in their ability to simultaneously recapitulate multicellular architecture, spatial anisotropy, and dynamic transport behavior while enabling real-time, quantitative analysis. Here, we present an integrated microfluidic-mass spectrometry platform that utilizes engineered BBB particles (BBBps) as a functional analytical interface to enable dynamic, *in situ* monitoring of BBB transport and metabolism. Using a microfluidic-aerosol hybrid process, we generate multicompartamental microparticles featuring a Matrigel-modified rough surface that enhances endothelial adhesion and tight junction formation, while their cores co-encapsulate microglia, neurons, and astrocytes to establish controlled neurovascular interactions. The BBBps exhibit size- and lipophilicity-dependent permeability, hypoxia-responsive antioxidative protection, and barrier-limited metabolism of salidroside. Real-time analysis with a Chip-MS system further reveals attenuated efflux of glutamate and lactate, confirming barrier integrity and metabolic regulation. This layered design of the micro-blood–brain barrier, combined with Chip-MS analysis, provides a scalable and functionally faithful *in vitro* model platform for studying drug transport, metabolism, and neurovascular studies.

Received 24th December 2025

Accepted 20th February 2026

DOI: 10.1039/d5sc10112c

rsc.li/chemical-science

Introduction

The blood–brain barrier (BBB), composed of brain microvascular endothelial cells, a basement membrane, and astrocytes, serves as a highly selective physiological barrier that maintains central nervous system (CNS) homeostasis by tightly regulating molecular transport between the blood and neural tissue.^{1–3} While this barrier protects the brain from neurotoxic insults, its tightly controlled transport complicates CNS drug development and mechanistic studies of neurovascular function. In this context, the BBB is not only a biological obstacle but also an analytically challenging system.^{4–6} Therefore, there is a pressing

need for analytical platforms that can quantify transport and metabolism across BBB-like interfaces dynamically, *in situ*, and with high sensitivity.

Conventional *in vitro* BBB models, such as transwell co-culture systems, have advanced the understanding of endothelial permeability and intercellular signaling.^{7,8} However, their planar architecture and reliance on multi-layer assembly hinder spatial fidelity, throughput, and reproducibility, particularly when mimicking the three-dimensional (3D) neurovascular unit (NVU). Although microfabrication approaches including soft lithography and 3D bioprinting have improved structural complexity, they often require complex fabrication workflows and remain challenging to scale or couple seamlessly with continuous detection methods such as mass spectrometry (MS).^{9–13}

In contrast, microfluidic droplet-derived hydrogel microparticles offer advantages such as uniformity, customizability, monodispersity, and high throughput.^{14,15} These microparticles exhibit excellent extracellular matrix (ECM)-mimicking properties, enabling effective multi-component cell co-culture and establishing them as ideal cellular carriers.¹⁶ Their monodispersity, tunable composition, and high-throughput production enable controlled mass transport and reproducible microenvironments, while their extracellular-matrix (ECM)-

^aSchool of Health and Rehabilitation, Chengdu University of Traditional Chinese Medicine, Chengdu, China

^bBeijing Key Laboratory of Microanalytical Methods and Instrumentation, Key Laboratory of Bioorganic Phosphorus Chemistry & Chemical Biology (Ministry of Education), Department of Chemistry, Tsinghua University, Beijing, 100084, China. E-mail: wzn@mail.tsinghua.edu.cn; jmlin@mail.tsinghua.edu.cn

^cChinese Medicine Germplasm Resources Innovation and Effective Uses Key Laboratory of Sichuan Province, Institute of High Altitude Multimorbidity/TCM-Integrated High Altitude Medicine Center, Hospital of Chengdu University of Traditional Chinese Medicine, Chengdu, China. E-mail: xlm999@cdutcm.edu.cn

^dSchool of Ethnic Medicine, Chengdu University of Traditional Chinese Medicine, Chengdu, 611137, China



mimicking properties support multicellular co-culture. Diverse architectures, such as core-shell,^{17,18} non-spherical,^{19,20} and multicompartmental particles,²¹ have been explored for tissue modeling and cell delivery. Nevertheless, most existing micro-particle systems lack the anisotropic architecture and defined cellular organization required to recapitulate BBB barrier functions, limiting their use for spatially resolved chemical analysis.²² An effective analytical strategy would thus require (i) precisely engineered physical boundaries for compartmentalized organization of BBB-relevant cell populations, (ii) anisotropic interfacial microenvironments that guide heterotypic cellular responses and regulate diffusion pathways, and (iii) modular architectures that are inherently compatible with online analysis to enable continuous monitoring of barrier transport and metabolism.

Here, we introduce an integrated fabrication-to-analysis workflow that couples microfluidic-aerosol-engineered BBB hydrogel microparticles (BBBps) with Chip-MS, establishing a multicellular barrier-mimetic analytical interface for real-time interrogation of BBB permeability and metabolism (Fig. 1). The fabrication process yields anisotropic, multicompartmental microparticles with a Matrigel-modified rough surface that supports endothelial adhesion and enhances tight-junction formation, while the inner compartments co-encapsulate neurons, astrocytes, and microglia to establish metabolically active NVU-like crosstalk. The multicompartmental 3D architecture provides controlled spatial positioning and proximity among encapsulated cell types while allowing inter-compartment exchange of paracrine factors and metabolites, thereby supporting localized NVU-like interactions. In contrast to conventional microfluidic BBB models that rely on planar, membrane-based architectures and predominantly end-point sampling, the proposed BBB interface is directly compatible with microfluidic handling and online MS coupling, enabling continuous, time-resolved chemical measurement of transport and metabolism under controlled flow. Using this platform, we quantified *trans*-barrier transport for twelve compounds

spanning a range of physicochemical properties, characterized salivroside metabolism, and monitored endogenous metabolite fluxes *via* real-time Chip-MS coupling. Collectively, this work introduces a microparticle-enabled analytical interface that links multicellular barrier architecture with continuous MS detection, providing a versatile approach for dynamic analysis and compound screening in BBB-like systems.

Results and discussion

Generation of multicompartmental hydrogel microparticles

The lab-made microfluidic system for the generation of multicompartmental microparticles includes a microfluidic chip for microparticle layout customization, gas-shearing coaxial needles for particle breakup, and an aerosol ejector for microparticle surface processing (Fig. S1). During the experiment, a 2% sodium alginate solution was used to prepare the multicompartmental microparticles. The chip features six asymmetrically arranged microchannels, allowing spatially controlled introduction of sodium alginate mixed with three different cell types (Fig. S1a). The central part of the chip connects to the inner needle of the droplet generator, while the nitrogen outlet is linked to the space between the inner needle and outer shell (Fig. S1b). Nascent multicompartmental alginate droplets were formed under nitrogen shear force. These droplets then passed through a CaCl₂ atmosphere produced by the aerosol ejectors, where their surfaces were impacted by ultra-fine aerosol sprays. This process induced an uneven aggregation of calcium alginate, freezing the transient topological morphology and preventing the contraction and disappearance of surface structures due to the gravitational potential energy from fluid displacement. The droplets were then fully solidified in a downstream collection bath, resulting in rough-surfaced multicompartmental alginate microparticles, which were further modified by soaking in 5% Matrigel.

Under microscopic observation, the untreated microparticles have a rough surface texture (Fig. S2a). The coefficient of

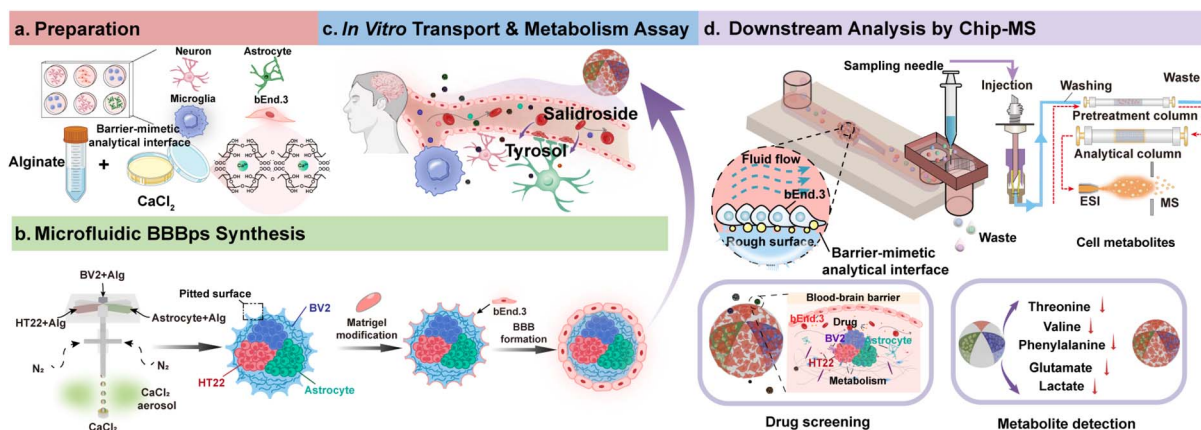


Fig. 1 A hydrogel microparticle engineered with hierarchical tissue architecture and BBB functionality to simulate the neurovascular unit *in vivo*. (a) Preparation of cells and reagents. (b) Schematic diagram of the BBBps construction process. (c) Schematic diagram of the scale of the neurovascular unit in the brain. (d) Schematic diagram of drug screening and metabolism across the BBB, including the Chip-MS online detection system and endogenous metabolism and drug metabolism of BBB-related cells.



variation (CV) of size was under 5%, indicating good monodispersity, which is favorable for uniform cell assembly (Fig. S2b). Furthermore, the stability of the multi-compartmental microparticles was assessed by incorporating fluorescent nanoparticles of different colors into the sodium alginate solution. Fluorescence intensity analysis indicated that the fluorescence occupancy of each compartment was similar, confirming clear segregation between particle regions (Fig. S2c).

Construction of BBB microparticles

The above-mentioned multicompartmental microparticles were loaded with three BBB-relevant types (BV2 microglia, HT22 neurons, and astrocytes), each confined to a separate compartment to facilitate downstream tracking and analysis of individual cell populations, while bEnd.3 endothelial cells were cultured on the particle surface using the hanging drop method.²³ Although alginate microparticles are generally considered biopolymers with low cell adhesion properties, their rough surface structure and matrix modification provided a supportive microenvironment that promoted cell attachment.^{24,25} Previous research has shown that materials with surface roughness and chemical modifications can promote cell adhesion through interactions with cell membrane receptors and proteins.²⁴ As a result, we observe that cells were firmly attached to the particle surfaces (Fig. 2a).

To assess the functionality of the BBBps, we evaluated the viability and function of the loaded cells. Cell growth and morphology were monitored using the Calcein-AM/EthD-1 double-staining kit. On the third day of culture, 3D imaging showed that the cells had spread across the microparticle surface, forming a uniform and complete barrier while maintaining high growth activity for 7 days (Fig. 2b). Interestingly, during the culture process, we observed that this barrier layer hinders the entry of small molecule substances such as fluorescent reagents. Therefore, in order to observe the viability of cells inside the microparticles, we performed precise micro-perforation using a syringe needle, allowing fluorescent reagents to penetrate. Over a course of 7 days, a large number of cells were observed to proliferate, while the cells both inside

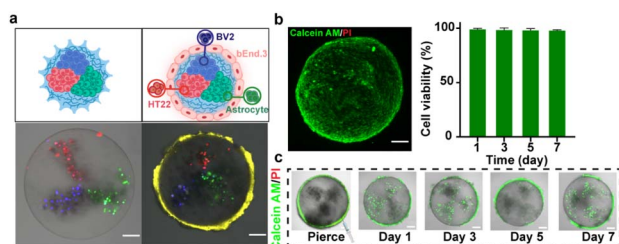


Fig. 2 The preparation of BBBps. (a) Cell tracker characterization of internal compartments and external cell adhesion on BBBps. (b) The 3D layer scan image of BBBps; the surface barrier maintains high activity within 7 days ($n = 4$). (c) External and internal cells in BBBps after 7 days maintained high viability. The dense barrier structure formed by bEnd.3 on the surface of microparticles hindered the entry of fluorescent dyes, which was pierced to allow staining of internal cells. Scale bars = 100 μm .

and on the surface of the microparticles maintained high viability (>98%) (Fig. 2c).

Function of BBB microparticles

To further verify the feasibility of the constructed BBBps, immunofluorescence staining was employed to confirm the formation of tight junctions between adjacent cells, validating the creation of the BBB. After 3 days of culture, CD31 (a marker of endothelial cells,²⁶ indicating vascularized-like expression) (Fig. 3a) and Claudin-5 (a key tight junction protein in the BBB)²⁷ (Fig. 3b) were both highly expressed, demonstrating good vascularized-like characteristics and tight junction formation. VE-cadherin (a key component of intercellular adhesion junctions) staining showed a continuous filamentous pattern, which further confirmed that endothelial cells formed a continuous monolayer on the surface (Fig. 3c).²⁸ Double staining for β -catenin (essential for regulating and maintaining the barrier by connecting cadherins to the actin cytoskeleton)²⁹ and ZO-1 (contributing to barrier integrity and controlling intercellular permeability)³⁰ further supported the integrity of the barrier and the maintenance of tissue homeostasis (Fig. 3d). The tight junctions between brain vascular endothelial cells restrict the entry of hydrophilic macromolecules into the brain. Thus, the selective permeability of hydrophilic non-ionic tracers is commonly used to assess the integrity of tight junction formation between endothelial cells.³¹ Here, both BBBps and non-BBB microparticles (non-BBBps) were exposed to solutions containing fluorescein isothiocyanate (FITC) and FITC-labeled dextran of 10 kDa (Fig. 3e). The fluorescence distribution inside and around the particles was measured to assess barrier function. In the non-BBBps group, FITC diffusion reaches equilibrium rapidly within 5 min, resulting in similar

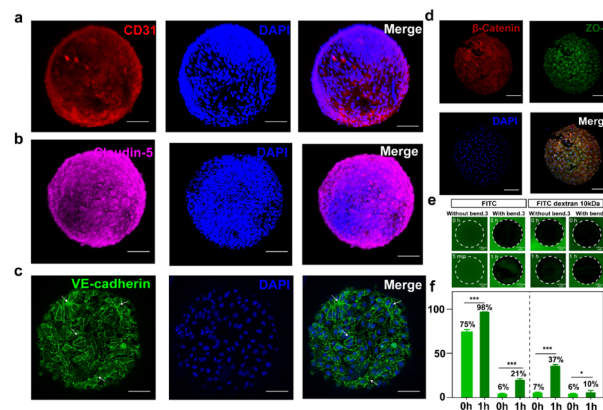


Fig. 3 Fluorescence images of proteins related to barrier formation on the surface of BBBps and evaluating the barrier function by the permeability of BBBps and non-BBBps (microspheres without bEnd.3 on the surface). (a) CD31. (b) Claudin-5. (c) VE-cadherin. (d) Dual staining images of β -catenin and ZO-1. (e) Fluorescence imaging of FITC and 10 kDa FITC-dextran entering the internal space through BBB-like microspheres. (f) The penetration efficiency of non-BBBps and BBBps. The penetration efficiency is the ratio of the fluorescence intensity of tracers inside particles to the surroundings. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, one-way ANOVA with Tukey's multiple comparison test. Data are presented as mean values \pm SD. Scale bars = 100 μm .



fluorescence intensities inside and outside the particles. In contrast, the BBBps significantly inhibited FITC diffusion, with remarkably reduced diffusion even after 1 h. This trend was consistently observed with another tracer, confirming the successful establishment of selective permeability in BBBps (Fig. 3f).

Evaluation of BBB protective function in BBBps

The BBB selectively permits the passage of specific substances to protect the brain from toxins and large molecules. Factors such as oxidative stress and neuroinflammation can lead to BBB damage.^{32,33} Nitric oxide (NO) and reactive oxygen species (ROS) are key signaling molecules involved in the body's hypoxic response,^{34,35} and their excessive accumulation can result in acute brain damage and impaired BBB function.³⁶ The production levels of NO and ROS were measured to evaluate BBB damage under hypoxic conditions induced by deferoxamine (DFO).^{37,38} After 12 hours of DFO exposure, NO and ROS levels inside BBBps and non-BBBps were assessed using DCFH-DA and DAF-FM fluorescent probes. The results showed that BBBps exhibited significantly suppressed production levels of NO (Fig. 4a and b) and ROS (Fig. 4c and d) compared to non-BBBps, demonstrating the protective effect of the BBB structure. These findings demonstrate that BBBps successfully mimicked the structural and functional characteristics of the BBB on the single-particle level, exhibiting protective capabilities under hypoxic stimulation.

Assessment of drug permeability across the BBB using BBBps

Next, we tested the permeability of three small molecule drugs on the BBBps model: (1) caffeine, a lipophilic small molecule

that can rapidly permeate the BBB through simple diffusion and carrier-mediated transport; (2) cimetidine, a moderately permeable molecule transported by efflux transporters and organic cation transporters; (3) doxorubicin, known for its poor BBB penetration. ESI-MS was used to quantify the drugs, detecting caffeine (m/z 194.1 \rightarrow 138.2), cimetidine (m/z 253.0 \rightarrow 159.1), and doxorubicin (m/z 543.9 \rightarrow 361.2) (Table S1). Calibration curves were constructed by spiking different concentrations of each drug (Fig. S3). Compared with the *in vivo in situ* perfusion data reported in the literature, these results in BBBps closely resemble the *in vivo* situations (Table S2),^{39,40} indicating the formation of a functional high-fidelity BBB unit in the BBBps model.⁴¹ This suggests its suitability for subsequent experiments on drug permeability across the BBB (Fig. 5a). Subsequently, nine active ingredients from traditional Chinese medicine of different molecular weights were further tested, including resveratrol (m/z 227.1 \rightarrow 143.2), osthole (m/z 244.9 \rightarrow 131.2), matrine (m/z 249.1 \rightarrow 148.3), wogonin (m/z 284.8 \rightarrow 270.1), salidroside (m/z 299.0 \rightarrow 119.0), quercetin (m/z 302.8 \rightarrow 165.2), scutellarin (m/z 461.0 \rightarrow 285.0), paeoniflorin (m/z 525.0 \rightarrow 121.2), and ginsenoside Rb1 (m/z 1107.4 \rightarrow 783.5) (Table S1). The permeability coefficients for each drug showed that substance permeation through the BBB was closely related to molecular weight, with larger molecules facing greater difficulty in crossing the BBB. Additionally, the physical and chemical properties of compounds influenced their BBB permeability (Fig. 5b and c and Table S3). Lastly, we investigated the relationship between the octanol/water partition coefficient (K_{ow}) and the permeability of five active ingredients (Paeoniflorin, Quercetin, Wogonin, Resveratrol, and Osthole) across the BBB (Fig. 5d). Studies confirmed that the BBB, as a highly selective and dynamic lipophilic barrier, favors substances with higher K_{ow} values, indicating that greater lipid solubility facilitates BBB penetration.

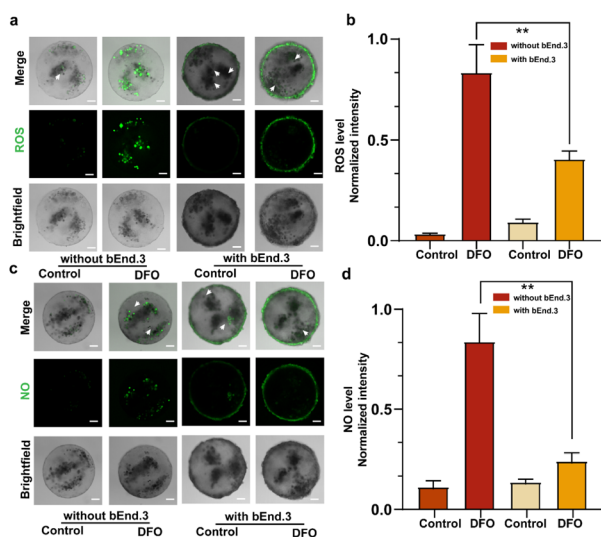


Fig. 4 Permeability of active ingredients of different molecular weights on BBBps. Evaluation of the BBB protective function of BBBps against ROS and NO. (a and b) The ROS level induced by DFO in non-BBBps and BBBps ($n = 5$). (c and d) The NO level induced by DFO in non-BBBps and BBBps ($n = 5$). * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, one-way ANOVA with Tukey's multiple comparison test. Data are presented as mean values \pm SD. Scale bars = 100 μ m.

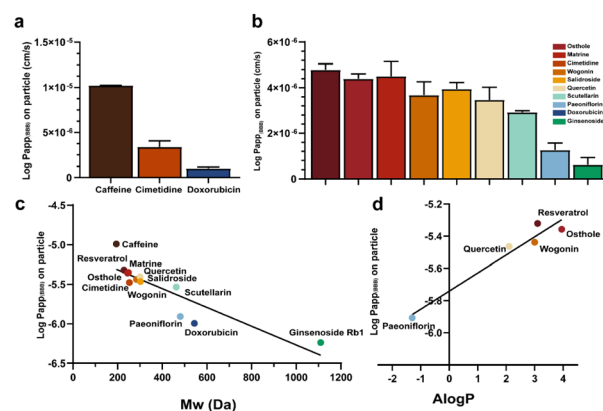


Fig. 5 Permeability of active ingredients of different molecular weights on BBBps. (a) Permeability of caffeine, cimetidine, and doxorubicin in the BBBps model. (b) Permeability of 12 active substances in the BBBps model. (c) Linear relationship between the molecular weight and permeability of 12 active substances in the BBBps model ($R^2 = 0.7433$). (d) Relationship between the octanol/water partition coefficients and P_{app} (BBB) of resveratrol, osthole, wogonin, quercetin, and paeoniflorin ($R^2 = 0.8903$).



Quantification of salidroside and tyrosol on the BBBPs

To evaluate the functionality of the BBBPs constructs, we assessed the metabolic activation of salidroside, the primary component of the Tibetan medicine *Rhodiola rosea* rhizoma et radix. Salidroside is known to mitigate hypoxia-induced metabolic disturbances and neuronal damage by preserving mitochondrial function.^{42,43} Its metabolite, tyrosol, also exhibits anti-hypoxic and anti-inflammatory properties. After crossing the barrier to reach the internal structure, salidroside began to metabolize into tyrosol (Fig. 6a). After treating BBBPs with 50 μ M salidroside for 24 hours, identification and quantification of salidroside and tyrosol were conducted using negative mode MRM (Table S4). A standard curve was drawn by preparing samples with different concentration gradients using the standard substance, and the concentrations of salidroside and tyrosol in the samples were calculated (Fig. S4). Quantitative analysis revealed that BBBPs exhibited significantly lower levels of tyrosol (Fig. 6b) and a reduced metabolic conversion rate of salidroside compared to non-BBBPs (Fig. 6c). These results further demonstrate that the BBBPs model successfully simulates the *in vivo* phenomenon of diminished drug metabolic activity due to BBB interference.

Effects of the BBB on amino acid metabolism in BBBPs

We evaluated the impact of the barrier on the levels of specific intracellular endogenous metabolites, including amino acids and lactate, which serve as indicators of BBB functionality. The evaluation was performed by comparative analysis of secreted metabolite concentrations between BBBPs and non-BBBPs. Emerging evidence indicates that metabolites such as

threonine, valine, phenylalanine, glutamate and lactate are critically involved in the modulation of BBB function.^{44–46} Among them, threonine, valine, and phenylalanine need to cross the BBB. When their concentrations are excessively elevated, they competitively inhibit the transport of other amino acids, leading to cerebral amino acid imbalance that disrupts neurotransmitter synthesis and protein metabolism. Notably, the BBB regulates intracerebral concentrations of threonine, valine, phenylalanine, glutamate, and lactate through specialized transporters and selective permeability mechanisms, thereby modulating their metabolic pathways and neurological functions.^{47,48} In our study, after 12-hour cultivation of BBBPs and non-BBBPs models, culture supernatants were collected to quantify five intracellular endogenous metabolites: threonine, valine, phenylalanine, glutamate, and lactate. The results demonstrated that threonine, phenylalanine, glutamate and lactate exhibited elevated efflux under compromised barrier integrity, attributable to increased BBB permeability (Fig. 7a). The intact BBB leads to a decrease in the efflux of these metabolites, with the reduction exceeding 20%. To further validate these findings, we dynamically monitored the 12 h concentration profiles of glutamate and lactate using a Chip-MS system for real-time metabolic flux analysis. The microfluidic chip design and internal instrument of Chip-MS are shown in Fig. S5. Fragmentation ions and patterns of glutamate and lactate were analyzed using reference standards in product ion scanning mode, with representative spectra shown in Fig. S5a and b. Detailed analytical parameters are provided in Table S5. Subsequently, calibration curves were established for reliable

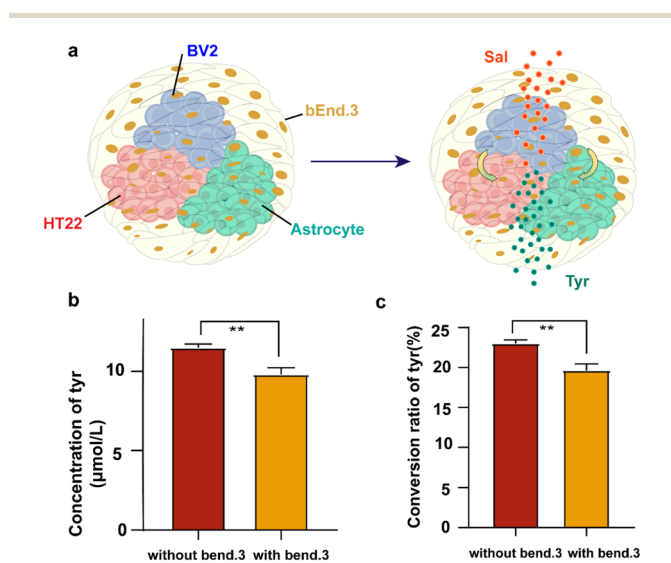


Fig. 6 Assessment of the conversion of salidroside to the drug-reactive metabolite tyrosol on BBBPs. (a) Schematic of salidroside transformation in the BBBPs. (b) The yield of the metabolic product tyrosol in cell culture medium after the non-BBBPs and BBBPs were treated with salidroside for 24 h. (c) Calculated percentage conversion of salidroside to tyrosol. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, one-way ANOVA with Tukey's multiple comparison test. Data are presented as mean values \pm SD.

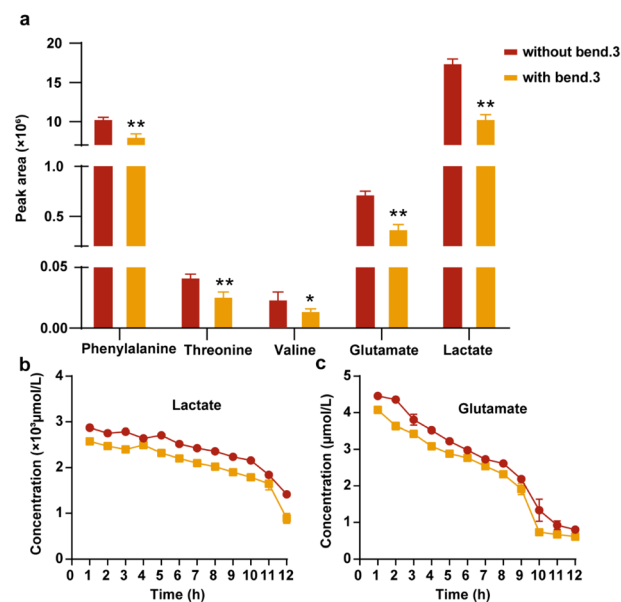


Fig. 7 Dynamic monitoring of amino acid and lactate metabolites based on Chip-MS between non-BBBPs and BBBPs. (a) Differences in the changes of BBB marker amino acid and lactate metabolites. (b) Real-time quantitative analysis of lactate over 12 h. (c) Real-time quantitative analysis of glutamate over 12 h. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, one-way ANOVA with Tukey's multiple comparison test. Data are presented as mean values \pm SD.



quantification of both metabolites (Fig. S5c and d), enabling hourly monitoring of their concentrations in real-time throughout the 12-hour dynamic cultivation process. The results demonstrated that compared to the non-BBBps group, our BBBps model attenuated the glutamate and lactate accumulation caused by increased BBB permeability, effectively suppressing their efflux to maintain cerebral metabolic homeostasis (Fig. 7b and c). These findings align with trends reported in prior literature.^{44,45,47} Overall, our results confirm that we have successfully established an *in vitro* BBB model on single particles and simulated the effects of the BBB on endogenous metabolites.

Conclusions

In this study, we established an integrated BBBps-Chip-MS analytical platform in which microfluidic-engineered hydrogel microparticles serve as standardized multicellular barrier interfaces for BBB interrogation. This particle-based design enables spatially defined co-culture of brain-relevant cell types, programmable surface modification to support endothelial adhesion and tight-junction formation, and coupling with online mass spectrometry for dynamic and quantitative chemical analysis. The resulting BBBps exhibited key barrier phenotypes, including tight-junction protein expression and selective molecular permeability, and maintained barrier responsiveness under hypoxic stress. Leveraging continuous Chip-MS coupling, we quantified drug transport across the interface and observed permeability trends governed by molecular size and lipophilicity that are consistent with *in vivo* behavior. Importantly, the platform enabled dynamic, *in situ* monitoring of endogenous metabolite exchange, revealing markedly reduced efflux of neuroactive metabolites (*e.g.*, glutamate and lactate) under intact barrier conditions. Moreover, real-time metabolic profiling captured salidroside conversion and its dependence on barrier integrity, demonstrating the capability to resolve coupled transport-metabolism processes within a controlled microenvironment. Collectively, this work introduces a scalable and versatile analytical strategy that couples engineered multicellular barrier interfaces with online mass spectrometry, offering broad potential for quantitative CNS permeability assessment, metabolism studies, and barrier-associated phenotyping.

Looking forward, the platform can be further improved by enhancing batch-to-batch consistency in BBBp production, increasing the number of parallel channels for higher screening capacity, and strengthening multi-channel operational robustness for synchronized dosing, perfusion, and data acquisition. These advances will further improve scalability and reliability and broaden the utility of the BBBps-Chip-MS strategy for compound profiling and CNS-relevant phenotyping.

Author contributions

Z. W., Y. Z., X. M. and J.-M. L. obtained the funds and supervised the project; S. C., Z. W. and J.-M. L. conceived the idea; S. C., Y. Z., and Z. W. designed the experiments; S. C. and Y. Z.

performed experiments; S. C., Y. Z. and T. X. analyzed the data; and S. C. wrote the original manuscript. All authors revised and approved the manuscript.

Conflicts of interest

There are no conflicts to declare.

Data availability

The data supporting this article have been included as part of the supplementary information (SI). Supplementary information: materials and methods; the homemade microfluidic system for generating multicompartment particles; characterization of rough-surfaced multicompartment hydrogel microspheres; chromatograms, calibration curves, and ion fragmentation spectra of active ingredients with different molecular weights; fragment spectra and calibration curves for salidroside and tyrosol; photographs and schematics of the Chip-MS device; calibration information for glutamate and lactate; MRM parameters for target compounds and metabolites; permeability comparisons of model drugs (caffeine, cimetidine, and doxorubicin) in BBBps, transwell, and *in vivo* models; molecular weights and P_{app} data for 12 active ingredients; and LC gradient elution conditions for the target compounds and metabolites. See DOI: <https://doi.org/10.1039/d5sc10112c>.

Acknowledgements

This work was supported by the National Key R&D Program of China (No. 2024YFB4607800), the Key Program of the National Natural Science Foundation of China (No. U23A20520), the National Natural Science Foundation of China (No. 22034005 and 22404094), the Natural Science Foundation of Sichuan Province (No. 2024NSFSC0701) and the Chengdu Municipal Health Commission & Chengdu University of Traditional Chinese Medicine 2024 Annual Joint Commission-University Innovation Fund (WXLH202403076).

Notes and references

- 1 B. Zierfuss, C. Laroche and A. Prat, Blood-brain barrier dysfunction in multiple sclerosis: causes, consequences, and potential effects of therapies, *Lancet Neurol.*, 2024, **23**, 95–109.
- 2 J. R. Wu, Y. Hernandez, K. F. Miyasaki and E. J. Kwon, Engineered nanomaterials that exploit blood-brain barrier dysfunction for delivery to the brain, *Adv. Drug Delivery Rev.*, 2023, **197**, 114820.
- 3 C. Hajal, G. S. Offeddu, Y. Shin, S. Zhang, O. Morozova, D. Hickman, C. G. Knutson and R. D. Kamm, Engineered human blood-brain barrier microfluidic model for vascular permeability analyses, *Nat. Protoc.*, 2022, **17**, 95–128.
- 4 B. Li, L. Y. Zhang, Y. Yin, A. Q. Chen, B. R. Seo, J. Z. Lou, D. J. Mooney and D. A. Weitz, Stiff hydrogel encapsulation



- retains mesenchymal stem cell stemness for regenerative medicine, *Matter*, 2024, 7, 3447–3468.
- 5 D. L. Liu, P. Liao, H. Li, S. H. Tong, B. Q. Wang, Y. F. Lu, Y. S. Gao, Y. G. Huang, H. Zhou, L. J. Shi, J. Papadimitriou, Y. Zong, J. Yuan, P. L. Chen, Z. M. Chen, P. Ding, Y. Q. Zheng, C. Q. Zhang, M. H. Zheng and J. J. Gao, Regulation of blood–brain barrier integrity by -expressing astrocytes through mitochondrial transfer, *Sci. Adv.*, 2024, 10, eadk2913.
 - 6 L. Dao, Z. You, L. Lu, T. Y. Xu, A. K. Sarkar, H. Zhu, M. Liu, R. Calandrelli, G. Yoshida, P. Lin, Y. F. Miao, S. Mierke, S. Kalva, H. N. Zhu, M. X. Gu, S. Vadivelu, S. Zhong, L. F. Huang and Z. Y. Guo, Modeling blood–brain barrier formation and cerebral cavernous malformations in human PSC-derived organoids, *Cell Stem Cell*, 2024, 31, 818–833.
 - 7 D. Liu, M. Y. Zhu, Y. Lin, M. M. Li, R. L. Huang, L. Yang, Y. L. Song, Y. Diao and C. Y. Yang, LY6E protein facilitates adeno-associated virus crossing in a biomimetic chip model of the human blood–brain barrier, *Lab Chip*, 2022, 22, 4180–4190.
 - 8 R. Song, K. Pekrun, T. A. Khan, F. J. Zhang, S. P. Pasca and M. A. Kay, Selection of rAAV vectors that cross the human blood–brain barrier and target the central nervous system using a transwell model, *Mol. Ther.–Methods Clin. Dev.*, 2022, 27, 73–88.
 - 9 S. Massa, M. A. Sakr, J. Seo, P. Bandaru, A. Arneri, S. Bersini, E. Zare-Eelanjegh, E. Jalilian, B. H. Cha, S. Antona, A. Enrico, Y. Gao, S. Hassan, J. P. Acevedo, M. R. Dokmeci, Y. S. Zhang, A. Khademhosseini and S. R. Shin, Bioprinted 3D vascularized tissue model for drug toxicity analysis, *Biomicrofluidics*, 2017, 11, 044109.
 - 10 Y. J. Wang, Z. G. Di, M. L. Qin, S. M. Qu, W. B. Zhong, L. F. Yuan, J. Zhang, J. M. Hibberd and Z. Y. Yu, Advancing Engineered Plant Living Materials through Tobacco BY-2 Cell Growth and Transfection within Tailored Granular Hydrogel Scaffolds, *ACS Cent. Sci.*, 2024, 10, 1094–1104.
 - 11 B. Peng, S. P. Hao, Z. Q. Tong, H. Bai, S. J. Pan, K. L. Lim, L. Li, N. H. Voelcker and W. Huang, Blood–brain barrier (BBB)-on-a-chip: a promising breakthrough in brain disease research, *Lab Chip*, 2022, 22, 3579–3602.
 - 12 S. Fengler, B. Kurkowsky, S. K. Kaushalya, W. Roth, E. Fava and P. Denner, Human iPSC-derived brain endothelial microvessels in a multi-well format enable permeability screens of anti-inflammatory drugs, *Biomaterials*, 2022, 286, 121525.
 - 13 W. Y. Bi, S. L. Cai, T. Lei and L. Wang, Implementation of blood–brain barrier on microfluidic chip: Recent advance and future prospects, *Ageing Res. Rev.*, 2023, 87, 101921.
 - 14 Y. J. Zheng, Z. N. Wu, L. Lin, X. N. Zheng, Y. Hou and J. M. Lin, Microfluidic droplet-based functional materials for cell manipulation, *Lab Chip*, 2021, 21, 4311–4329.
 - 15 Z. N. Wu, Y. J. Zheng, L. Lin, G. W. Xing, T. Z. Xie, J. X. Lin, X. R. Wang and J. M. Lin, Construction of Multiplexed Assays on Single Anisotropic Particles Using Microfluidics, *ACS Cent. Sci.*, 2025, 11, 294–301.
 - 16 Z. Q. Zhu, T. A. Chen, Y. Q. Zhu, F. S. Huang, K. Mu, T. Si and R. X. Xu, Programmable pulsed aerodynamic printing for multi-interface composite manufacturing, *Matter*, 2023, 6, 2034–2051.
 - 17 Y. X. Li, H. R. Yao, S. L. Chen, N. Xu and J. M. Lin, Effect of Tryptophan Metabolites on Cell Damage Revealed by Bacteria-Cell Interactions in Hydrogel Microspheres, *Anal. Chem.*, 2023, 95, 1402–1408.
 - 18 Z. N. Wu, Y. J. Zheng, J. M. Lin, Y. X. Li, Y. N. Lin, X. R. Wang and L. Lin, On-microparticle construction of endothelialized liver microtissues for drug testing, *Chem. Eng. J.*, 2024, 481, 148403.
 - 19 H. H. Zheng, W. Du, Y. Y. Duan, K. Y. Geng, J. Deng and C. Y. Gao, Biodegradable Anisotropic Microparticles for Stepwise Cell Adhesion and Preparation of Janus Cell Microparticles, *ACS Appl. Mater. Interfaces*, 2018, 10, 36776–36785.
 - 20 Y. T. Wang, L. R. Shang, G. P. Chen, C. M. Shao, Y. X. Liu, P. H. Lu, F. Rong and Y. J. Zhao, Pollen-inspired microparticles with strong adhesion for drug delivery, *Appl. Mater. Today*, 2018, 13, 303–309.
 - 21 Z. N. Wu, Y. J. Zheng, L. Lin, Y. N. Lin, T. Z. Xie, W. J. Liao, S. Y. Chen, Y. R. Zhang and J. M. Lin, Open aerosol microfluidics enable orthogonal compartmentalized functionalization of hydrogel particles, *Matter*, 2024, 7, 3645–3657.
 - 22 K. O. Rojek, M. Cwiklinska, J. Kuczak and J. Guzowski, Microfluidic Formulation of Topological Hydrogels for Microtissue Engineering, *Chem. Rev.*, 2022, 122, 16839–16909.
 - 23 F. G. Kong, X. Jiang, R. C. Wang, S. Y. Zhai, Y. Z. Zhang and D. Wang, Forsythoside B attenuates memory impairment and neuroinflammation via inhibition on NF- κ B signaling in Alzheimer's disease, *J. Neuroinflammation*, 2020, 17, 305.
 - 24 K. Rechendorff, M. B. Hovgaard, M. Foss, V. P. Zhdanov and F. Besenbacher, Enhancement of protein adsorption induced by surface roughness, *Langmuir*, 2006, 22, 10885–10888.
 - 25 D. H. Cho and J. I. Hahm, Protein-Polymer Interaction Characteristics Unique to Nanoscale Interfaces: A Perspective on Recent Insights, *J. Phys. Chem. B*, 2021, 125, 6040–6057.
 - 26 T. Fu, D. P. Sullivan, A. M. Gonzalez, M. E. Haynes, P. J. Dalal, N. S. Rutledge, A. L. Tierney, J. A. Yescas, E. W. Weber and W. A. Muller, Mechanotransduction via endothelial adhesion molecule CD31 initiates transmigration and reveals a role for VEGFR2 in diapedesis, *Immunity*, 2023, 56, 2311–2324.
 - 27 Z. G. Yang, P. P. Lin, B. Chen, X. Q. Zhang, W. Xiao, S. L. Wu, C. N. Huang, D. Feng, W. Q. Zhang and J. J. Zhang, Autophagy alleviates hypoxia-induced blood–brain barrier injury via regulation of CLDN5 (claudin 5), *Autophagy*, 2021, 17, 3048–3067.
 - 28 J. S. Tang, Y. Y. Kang, Y. J. Zhou, N. Y. Shang, X. N. Li, H. Y. Wang, J. Q. Lan, S. Wang, L. Wu and Y. Peng, TIMP2 ameliorates blood–brain barrier disruption in traumatic



- brain injury by inhibiting Src-dependent VE-cadherin internalization, *J. Clin. Invest.*, 2024, **134**, e164199.
- 29 S. S. Song, H. C. Huang, X. D. Guan, V. Fiesler, M. I. H. Bhuiyan, R. J. Liu, S. Jalali, M. N. Hasan, A. K. Tai, A. Chattopadhyay, S. Chaparala, M. Sun, D. B. Stolz, P. N. He, D. Agalliu, D. D. Sun and G. Begum, Activation of endothelial Wnt/ β -catenin signaling by protective astrocytes repairs BBB damage in ischemic stroke, *Prog. Neurobiol.*, 2021, **199**, 101963.
- 30 Y. Y. Li, B. W. Liu, T. T. Zhao, X. P. Quan, Y. Han, Y. X. Cheng, Y. L. Chen, X. Shen, Y. Zheng and Y. H. Zhao, Comparative study of extracellular vesicles derived from mesenchymal stem cells and brain endothelial cells attenuating blood–brain barrier permeability *via* regulating Caveolin-1-dependent ZO-1 and Claudin-5 endocytosis in acute ischemic stroke, *J. Nanobiotechnol.*, 2023, **21**, 70.
- 31 M. Ryma, H. Genç, A. Nadernezhad, I. Paulus, D. Schneiderreit, O. Friedrich, K. Anđelović, S. Lyer, C. Alexiou, I. Cicha and J. Groll, A Print-and-Fuse Strategy for Sacrificial Filaments Enables Biomimetically Structured Perfusable Microvascular Networks with Functional Endothelium Inside 3D Hydrogels, *Adv. Mater.*, 2022, **34**, e2200653.
- 32 Y. I. Wang, H. E. Abaci and M. L. Shuler, Microfluidic Blood–Brain Barrier Model Provides In Vivo-Like Barrier Properties for Drug Permeability Screening, *Biotechnol. Bioeng.*, 2017, **114**, 184–194.
- 33 E. Candelario-Jalil, R. M. Dijkhuizen and T. Magnus, Neuroinflammation, Stroke, Blood–Brain Barrier Dysfunction, and Imaging Modalities, *Stroke*, 2022, **53**, 1473–1486.
- 34 H. S. Chen, Y. C. He, S. Chen, S. H. Qi and J. G. Shen, Therapeutic targets of oxidative/nitrosative stress and neuroinflammation in ischemic stroke: Applications for natural product efficacy with omics and systemic biology, *Pharmacol. Res.*, 2020, **158**, 104877.
- 35 J. Feng, Y. L. Zheng, M. Y. Guo, I. Ares, M. Martínez, B. Lopez-Torres, M. R. Martínez-Larranaga, X. Wang, A. Anadón and M. A. Martínez, Oxidative stress, the blood–brain barrier and neurodegenerative diseases: The critical beneficial role of dietary antioxidants, *Acta Pharm. Sin. B*, 2023, **13**, 3988–4024.
- 36 S. P. Chen, L. Z. Li, C. Peng, C. J. Bian, P. E. Ocak, J. H. Zhang, Y. Yang, D. Zhou, G. Z. Chen and Y. M. Luo, Targeting Oxidative Stress and Inflammatory Response for Blood–Brain Barrier Protection in Intracerebral Hemorrhage, *Antioxid. Redox Signaling*, 2022, **37**, 115–134.
- 37 N. J. Abbott, L. Ronnback and E. Hansson, Astrocyte–endothelial interactions at the blood–brain barrier, *Nat. Rev. Neurosci.*, 2006, **7**, 41–53.
- 38 S. Chen, F. Fan, Y. Zhang, J. Zeng, Y. Li, N. Xu, Y. Zhang, X. L. Meng and J. M. Lin, Metabolites from scutellarin alleviating deferoxamine-induced hypoxia injury in BV2 cells cultured on microfluidic chip combined with a mass spectrometer, *Talanta*, 2023, **259**, 124478.
- 39 A. Avdeef, Physicochemical profiling (solubility, permeability and charge state), *Curr. Top. Med. Chem.*, 2001, **1**, 277–351.
- 40 L. Y. Shi, M. Zeng, Y. Sun and B. M. M. Fu, Quantification of Blood–Brain Barrier Solute Permeability and Brain Transport by Multiphoton Microscopy, *J. Biomech. Eng.*, 2014, **136**, 031005.
- 41 W. Kim, J. Kim, S. Y. Lee, H. M. Kim, H. T. Jung, K. M. Joo and D. Nam, Functional validation of the simplified 3D Co-culture based BBB model, *Biochem. Biophys. Res. Commun.*, 2022, **625**, 128–133.
- 42 S. Chen, Y. Li, Y. Zhang, H. Yao, T. Xu, Y. Zhang, J. M. Lin and X. L. Meng, Establishment of microneurovascular units in alginate hydrogel microspheres to reveal the anti-hypoxic effect of salidroside, *J. Pharm. Anal.*, 2024, **14**, 100967.
- 43 Y. Hou, Y. T. Zhang, S. N. Jiang, N. Xie, Y. Zhang, X. L. Meng and X. B. Wang, Salidroside intensifies mitochondrial function of CoCl₂-damaged HT22 cells by stimulating PI3K-AKT-MAPK signaling pathway, *Phytomedicine*, 2023, **109**, 154568.
- 44 D. C. Tarlunganu, E. Deliu, C. P. Dotter, M. Kara, P. C. Janiesch, M. Scalise, M. Galluccio, M. Tesulov, E. Morelli, F. M. Sonmez, K. Bilguvar, R. Ohgaki, Y. Kanai, A. Johansen, S. Esharif, T. Ben-Omran, M. Topcu, A. Schlessinger, C. Indiveri, K. E. Duncan, A. O. Caglayan, M. Gunel, J. G. Gleeson and G. Novarino, Impaired Amino Acid Transport at the Blood–Brain Barrier Is a Cause of Autism Spectrum Disorder, *Cell*, 2016, **167**, 1481–1494.
- 45 S. K. Adla, H. Virtanen, T. Thongsodsang and K. M. Huttunen, Amino acid transporters in neurological disorders and neuroprotective effects of cysteine derivatives, *Neurochem. Int.*, 2024, **177**, 105771.
- 46 J. Albrecht and M. Zielinska, Exchange-mode glutamine transport across CNS cell membranes, *Neuropharmacology*, 2019, **161**, 107560.
- 47 W. H. Oldendorf and J. Szabo, Amino acid assignment to one of three blood–brain barrier amino acid carriers, *Am. J. Physiol.*, 1976, **230**, 94–98.
- 48 Q. R. Smith, Transport of glutamate and other amino acids at the blood–brain barrier, *J. Nutr.*, 2000, **130**, 1016S–1022S.

