


 Cite this: *RSC Adv.*, 2026, 16, 4776

# High-yielding and scalable synthesis of furfural acetals using protonated $\kappa$ -carrageenan as a biorenewable acid catalyst

 Rachitha S. Natraj and Saikat Dutta \*

The acetals of carbohydrate-derived furfurals have potential applications as fuel oxygenates and chemical intermediates. This work reports the use of protonated  $\kappa$ -carrageenan (kCH) as a marine biomass-derived renewable acid catalyst for the acetalization of furfural and 5-methylfurfural with ethylene glycol, 1,2-propanediol, and 1,3-propanediol. Since these glycols have promising catalytic production routes from biomass, the corresponding acetals are biorenewable to their entirety. Cyclohexane was used as the water removal agent by azeotropic distillation in a Dean–Stark apparatus. The isolated yields of the acetals were excellent (>90%), and the 2-(furan-2-yl)-1,3-dioxolane was conveniently prepared on a 10 g scale. The broad substrate scope of the catalyst was demonstrated by the successful preparation of acetals from benzaldehyde in excellent isolated yields. The kCH catalyst was conveniently recovered after the reaction and reused without catastrophic loss in catalytic activity. The effect of the extent of proton exchange on the activity, thermal stability, and recyclability of the kCH catalyst was also explored.

Received 30th December 2025

Accepted 10th January 2026

DOI: 10.1039/d5ra10090a

[rsc.li/rsc-advances](https://rsc.li/rsc-advances)

## 1 Introduction

The organic chemical manufacturing industries are undergoing a transition to integrate biomass as a renewable carbon-based feedstock alongside traditional fossil-based exhaustible resources (*e.g.*, petroleum).<sup>1</sup> Using renewable biomass as the starting material for liquid transportation fuels, organic chemicals, and synthetic polymers has clear societal, economic, and environmental advantages.<sup>2</sup> Among the various biomass value-addition technologies, the catalytic pathway has received particular attention. The catalytic transformation of biomolecules into targeted organic molecules is fast, scalable, highly selective, energy-efficient, and reagent-economical.<sup>3</sup> Moreover, the processes can be conveniently incorporated into the existing petrorefinery infrastructure. The structurally diverse polymeric biomolecules can be transformed into functionalized organic molecules in a two-step approach. The biomolecules are initially transformed into a handful of functionalized organic molecules, known as platform chemicals.<sup>4</sup> Synthetic upgrading of the platform molecules in the second step leads to molecules of desired structural characteristics and properties.<sup>5</sup>

Carbohydrates constitute the major fraction of most biomasses, and they have received much attention as feedstock for synthesizing biofuels and biochemicals.<sup>6,7</sup> Cellulose and hemicellulose are abundant carbohydrates in the second-generation biomass, whereas abundant carbohydrates in the third-generation marine biomass include carrageenans, ulvans,

and alginates.<sup>8</sup> Furfural (**FF**) and 5-(hydroxymethyl)furfural (**HMF**) are at the forefront of biorefinery research as carbohydrate-derived platform chemicals.<sup>9,10</sup> **FF** and **HMF** have been known in the literature for more than a century, and they have received renewed attention over the past three decades due to the commercial value of many of their derivatives.<sup>11</sup> **FF** is produced by the acid-catalyzed dehydration of pentose sugars, such as xylose and arabinose, in the hemicellulose fraction of lignocellulosic biomass.<sup>12</sup> **HMF** is produced by the dehydration of hexose sugars (*e.g.*, glucose, fructose) and polymeric carbohydrates containing hexoses (*e.g.*, starch, cellulose).<sup>13</sup> **HMF** has also been produced from marine carbohydrates, such as carrageenans.<sup>14</sup> **FF** and **HMF** retain all the biogenic carbon atoms of the parent sugar molecule, as well as some reactive functionalities. Moreover, the catalytic dehydration process produces water as the sole innocuous byproduct. The functionalities present in **FF** and **HMF** have been exploited for their synthetic value addition to virtually all classes of organic chemicals of industrial value.<sup>15</sup> 5-Methylfurfural (**MF**) is an important furanic platform chemical produced from hexose sugars and polymeric carbohydrates.<sup>16</sup> The catalytic hydrogenation reaction can produce **MF** by partially reducing isolated **HMF**.<sup>17</sup> **MF** can be produced from hexoses in a one-pot process without isolating the **HMF** as an intermediate.<sup>18</sup> **MF** has also been produced by reducing the hydrophobic analogs of **HMF**, such as 5-(chloromethyl)furfural.<sup>19</sup> **MF** can also directly be produced by the acid-catalyzed dehydration of specific sugars (*e.g.*, L-rhamnose).<sup>20,21</sup>

Acetalization is a prominent reaction for aldehyde functionality in **FF** and **MF**. The acetals of **FF** and **MF** have direct

Department of Chemistry, National Institute of Technology Karnataka (NITK), Surathkal, Mangalore 575025, Karnataka, India. E-mail: [sdutta@nitk.edu.in](mailto:sdutta@nitk.edu.in)

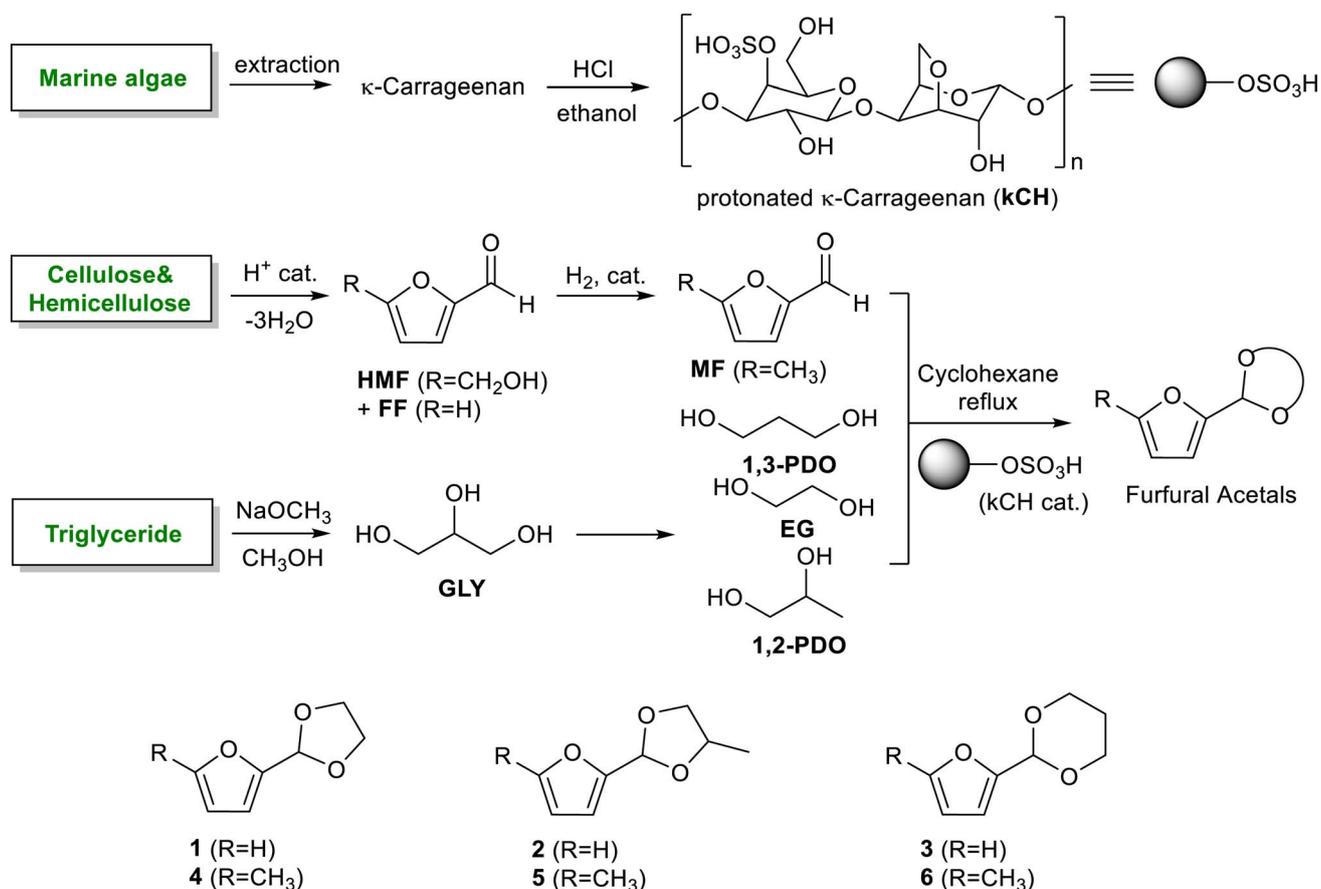


applications as potential fuel oxygenates.<sup>22</sup> Moreover, the acetals can act as chemical intermediates for downstream synthetic value addition pathways, as well as polymeric applications.<sup>23</sup> Acetalization of the aldehyde group in **FF** or **MF** increases the electron density on the furan ring, allowing it to participate in specific transformations, such as the [4 + 2] Diels–Alder reaction, to synthesize polyfunctionalized bioaromatics.<sup>24</sup>

The cyclic acetals of **FF** and **MF** have received particular interest, as they exhibit significantly better hydrolytic and storage stability than their acyclic counterparts. Ethylene glycol (**EG**), 1,2-propanediol (**1,2-PDO**), and 1,3-propanediol (**1,3-PDO**) are routinely used for synthesizing cyclic acetals of **FF** and **MF**.<sup>25,26</sup> These compounds have established renewable production routes. For example, **EG** can be produced by oxidizing ethylene, which is derived from the dehydration of bioethanol.<sup>27</sup> Alternatively, the retro-Diels–Alder reaction of glucose under hydrothermal conditions in the presence of a suitable Lewis acid catalyst can lead to glycolaldehyde, which, on hydrogenation, forms **EG**.<sup>28</sup> The selective catalytic reduction of triglyceride-derived glycerol (**GLY**) can produce **1,2-PDO** and **1,3-PDO**.<sup>29,30</sup> **EG** can be produced **GLY** through the C–C bond-cleavage reaction under hydrothermal conditions.<sup>31</sup> Therefore, the acetals formed by **FF** or **MF** with these diols and triols are biorenewable to their entirety (Scheme 1).

Numerous homogeneous and heterogeneous acid catalysts have been developed for the acetalization reaction of aromatic aldehydes, including furfurals.<sup>32–34</sup> There is still enormous scope for developing new catalysts that are robust, inexpensive, recyclable, eco-friendly, possessing a broad substrate scope, working under moderate conditions, and affording high selectivity and yield of catalysts. Catalysts sourced from biomass have garnered attention due to their renewable and environmentally friendly characteristics.<sup>35,36</sup> The use of biomass-derived catalysts for synthesizing these renewable molecules would certainly expand the scope of introducing carbon atoms of biogenic origin in synthesizing value-added organic chemicals, especially in the context of a carbohydrate-centric biorefinery.<sup>37</sup> Polymeric carbohydrates, functionalized with appropriate functionalities, have been used as catalysts for various organic transformations.<sup>38</sup> We had reported using cellulose sulfuric acid (CSA), produced by sulfonating semi-crystalline cellulose, as the heterogeneous acid catalyst for the ketalization of levulinic esters, which are also carbohydrate-derived platform chemicals.<sup>39</sup>  $\kappa$ -Carrageenan (**kC**) is a naturally occurring sulfated carbohydrate that is found in red seaweed (e.g., *Kappaphycus alvarezii*), which can be protonated by treating with a strong mineral acid.<sup>14</sup>

In this work, we report the use of protonated  $\kappa$ -carrageenan (**kCH**) as a novel heterogeneous acid catalyst for synthesizing



Scheme 1 Acetalization of carbohydrate-derived furfurals and glycols using protonated  $\kappa$ -carrageenan as the biomass-derived renewable heterogeneous catalyst.



the cyclic acetals of **FF** and **MF** using **EG**, **1,2-PDO**, and **1,3-PDO**. The reaction was performed in a Dean–Stark apparatus using cyclohexane as the solvent and azeotropic water removal. The effects of various reaction parameters, including the molar ratio of reagents and catalyst loading, on the selectivity and isolated yields of acetals were investigated. The **kCH** catalysts (both fresh and recycled) were characterized extensively using various analytical techniques, including FTIR, PXRD, FESEM, and TGA.

## 2 Materials, instruments, and experimental procedures

### 2.1 Materials and methods

Furfural (99%) was purchased from Spectrochem Pvt. Ltd. 5-Methylfurfural (**MF**, 99%) and Amberlyst-15 were purchased from Sigma. Ethyl acetate (99%) was purchased from Finar Limited. Cyclohexane (99.5%) was purchased from Merck. Ethylene glycol (99%), 1,3-propanediol (98%), 1,2-propanediol (99%), sodium sulfate (anhydrous, 99%), and HCl (35%, aq.) were purchased from Loba Chemie Pvt. Ltd. Furfural was distilled and refrigerated in an airtight, amber-colored glass container. All other chemicals are used as received without further purification. Thin-layer chromatography (TLC) plates and silica gel pre-coated on aluminum sheets were purchased from Merck (TLC Silica Gel 60, F254).

### 2.2 Instruments and characterization of the catalyst

The synthesized acetal products were characterized by spectroscopic techniques. The Fourier transform infrared (FTIR) spectra of the organic compounds were collected using a Bruker Alpha II FTIR 4000 instrument equipped with the attenuated total reflectance (ATR) technique. FTIR spectra of the materials (**kC**, fresh **kCH-1**, recycled **kCH-1**, and fresh **kCH-2**) were collected using the same instrument by pelletizing them with anhydrous KBr. The crystallinity of the samples was analyzed from Bragg's diffraction angles ( $2\theta$ ) using an Empyrean 3rd Gen powder X-ray diffractometer (PXRD) from Malvern PANalytical, Netherlands. Field-emission scanning electron microscopy (FE-SEM) images were collected using Thermo Fisher, FEI QUANTA 250 FEG. Samples were prepared by spin-coating the solutions onto a silicon substrate and dried under vacuum at room temperature. Samples were sputtered with gold for FE-SEM measurements. Elemental mapping of carbon, oxygen, potassium, and sulfur was done using a Thermo Fisher, FEI QUANTA 250 FEG (EDS). The Brunauer–Emmett–Teller (BET) was used for surface area analysis using Autosorb IQ-XR-XR, Anton Paar, Austria. Thermogravimetry analysis (TGA) was performed using a TGA 4000 from PerkinElmer, Singapore. TA instrument in the temperature range of 50–800 °C under the N<sub>2</sub> atmosphere at the temperature ramp of 10 °C min<sup>-1</sup>.

### 2.3 Synthetic procedures

**2.3.1 Preparation of the catalyst.** Two separate processes were employed to prepare **kCH** catalysts of varying acidity for comparison of their activity, stability, and recyclability.

*Process A.* The commercial  $\kappa$ -carrageenan (white powder, 2.00 g) was taken in a 100 mL beaker. A mixture of 35% aqueous HCl (2 mL) and absolute ethanol (25 mL) was added sequentially to the **kC** and stirred vigorously at room temperature for 1 h. The suspension was filtered through a Whatman filter paper (grade 5), and the solid residue was washed multiple times with ethanol to remove excess acid. The final product was dried under vacuum for 5 h at RT, yielding **kCH-1** as an off-white powder (2.01 g).

*Process B.* Dried  $\kappa$ -carrageenan (2.00 g) was taken in a 100 mL beaker, 27 mL of 0.84 M (concentrated adjusted for comparison with Process A) aq. HCl is added to the **kC** and kept under vigorous magnetic stirring for 1 h at room temperature. The suspension was filtered through a Whatman filter paper (grade 5), and the solid residue was washed with ethanol to remove excess acid. The final product was dried under vacuum for 5 h at RT, yielding **kCH-2** as an off-white powder (1.89 g).

**2.3.2 General procedure for the synthesis of furfural acetals.** Acetalization of biomass-derived furfurals was performed using suitable diols in the presence of **kCH-1** as a heterogeneous acid catalyst. For the synthesis of 2-(furan-2-yl)-1,3-dioxolane (**1**), furfural (1.001 g, 0.010 mol), ethylene glycol (0.700 g, 0.011 mol), **kCH-1** catalyst (0.200 g, 20 wt% of furfural), and cyclohexane (15 mL) were introduced into a round-bottomed flask (100 mL). The flask was fitted with a Dean–Stark apparatus and a water-cooled reflux condenser. The flask was then placed in an oil bath maintained at 110 °C and stirred continuously. During the reaction, water formed as a byproduct distilled off from the reaction vessel, forming an azeotrope with cyclohexane (b.p. 69.8 °C,  $\chi(\text{H}_2\text{O}) = 0.3$ ). The collected water in the Dean–Stark trap was removed at regular intervals. The reaction was monitored by thin-layer chromatography (TLC) using TLC plates (silica gel coated on aluminium plates) and petroleum ether (60–80 °C)–ethyl acetate (9 : 1, v/v) as the eluent. The KMnO<sub>4</sub> or 2,4-dinitrophenylhydrazine (DNP) stains were used for visualizing the spots (Fig. S39, SI). After 5 h of reflux, complete conversion of furfural was confirmed by TLC, and the reaction mixture was allowed to cool to room temperature. The reaction mixture was transferred into a centrifuge tube, and the solid **kCH-1** catalyst was separated by centrifugation (1000 rpm, 10 min). The liquid was carefully decanted, washed twice with distilled water (10 mL each), and dried over anhydrous sodium sulfate. The solvent was then removed under reduced pressure in a rotary evaporator to afford the acetal product as a light-yellow oil in over 92% yield. The recovered catalyst was washed with ethyl acetate, dried overnight at 60 °C, and reused in subsequent runs.

When the **kCH-2** catalyst was used, the reaction kinetics and isolated yield of **1** were comparable. However, the catalyst visibly deteriorated, including partial carbonization, during the reaction.

## 3 Results and discussion

### 3.1 Characterization of the **kCH** catalysts

The **kCH-1** and **kCH-2** catalysts were characterised using FTIR, PXRD, BET, and FE-SEM coupled with EDS. The **kCH-1** catalyst



demonstrated greater structural stability and recyclability than the kCH-2 catalyst, as evidenced by the higher reaction completeness sustained throughout numerous catalytic cycles. Therefore, the catalyst recyclability studies were performed on the kCH-1 system. The fresh and recycled kCH-1 catalysts were characterised to understand their deactivation pathways.

The FTIR spectrum (Fig. 1a) showed all characteristic bands of kC, where a broad peak at  $3573\text{ cm}^{-1}$  is due to the OH stretching mode from polysaccharide backbones. The peaks at  $1260\text{ cm}^{-1}$  and  $845\text{ cm}^{-1}$  represent the characteristic peaks of kC, corresponding to the  $\text{O}=\text{S}=\text{O}$  stretching vibration mode and the  $\text{O}-\text{SO}_3$  stretching vibration mode at the C4 position of galactose, respectively. The absorbance peak displays the bridge  $\text{C}-\text{O}$  stretching mode at  $1158\text{ cm}^{-1}$ . The intense peak at  $1068\text{ cm}^{-1}$  represents the  $\text{C}-\text{O}$  stretching mode. The characteristic of the  $\text{C}-\text{O}-\text{C}$  vibration mode of the 3,6-anhydrogalactose residue absorbance peak appears at  $928\text{ cm}^{-1}$ .  $845\text{ cm}^{-1}$  is the characteristic peak of the galactose 4-sulfate.<sup>40</sup> The signals at  $1441\text{ cm}^{-1}$  and  $1377\text{ cm}^{-1}$  are due to the  $\text{C}-\text{O}-\text{H}$  in-plane bending vibration mode and  $\text{C}-\text{H}$  bending vibration mode, respectively.<sup>41</sup> Fresh kCH-1 catalyst showed a peak shift from  $3573\text{ cm}^{-1}$  to  $3437\text{ cm}^{-1}$ , indicating enhanced hydrogen bonding due to protonation. The shift of the asymmetric  $\text{S}=\text{O}$  peak from  $1260\text{ cm}^{-1}$  to  $1254\text{ cm}^{-1}$  is also possible due to protonation of the sulfate oxygen, resulting in a reduction of electron density. Comparison of the FTIR spectra of kCH-1 and kCH-2 (Fig. 1b) confirms that both samples retain the  $\kappa$ -carrageenan framework, as evident from the fingerprint region. However, noticeable shifts in the  $\text{O}-\text{H}$  and sulfate bands reflect different extents of protonation and hydrogen bonding. The recycled kCH-1 catalyst exhibited a broad peak at  $3418\text{ cm}^{-1}$ , indicating the retention of hydrogen bonding after recycling. The sulfate ester peak at  $1254\text{ cm}^{-1}$  was retained. The presence of 3,6-anhydrogalactose was hinted at by the peak at  $925\text{ cm}^{-1}$ , suggesting that the core polysaccharide backbone remained intact.

Powder X-ray diffraction (PXRD) analysis of kC, fresh kCH-1, and recycled kCH-1 showed a broad hump in the range of  $15.8-$

$25.2^\circ$  (Fig. 2b), which signifies the semicrystalline characteristic of the sulfated polysaccharide. Sharp peaks at approximately  $28.4^\circ$ ,  $40.5^\circ$ , and  $50.2^\circ$  are likely due to inorganic components, such as potassium salts, sodium salts, or other residual minerals.<sup>42</sup> The sharp peak at  $10^\circ$  in kC arises from regular lateral packing of double helices, which is stabilized by the counter ions, such as  $\text{K}^+$  and  $\text{Na}^+$ . The peak disappeared in fresh and recycled kCH-1 due to disruption of the helix aggregation.<sup>43,44</sup> The intense peak at  $28.4^\circ$  and peaks formed around  $28.4^\circ$  are likely due to interaction between kC and aq. HCl during the preparation of the catalyst. The process adopted for proton exchange of kC by HCl treatment significantly impacts the extent of proton exchange, resulting in a notable difference in the morphology of the catalytic materials. There is a significant difference in the amount of inorganic salts between the two samples. The kCH-1 sample had larger and sharper signature peaks for KCl. In contrast, the recycled kCH-1 catalyst exhibited less intense peaks compared to the fresh kCH-1 catalyst, possibly due to the loss of crystallinity or the formation of amorphous deposits and chemical degradation during the recycling process.

The FE-SEM images depicted in Fig. 3 clarify the distinct morphological transformations upon protonation under different acid-treatment conditions. The FE-SEM images of kC showed dense, large, and relatively smooth structures with compact surface morphology.<sup>45,46</sup> The fresh kCH-1 catalyst, prepared using a mixture of 35% HCl and ethanol, exhibits a more fragmented and loosely packed structure, with increased surface roughness, cracks, and pores compared to kC. These changes were likely due to the cation exchange during protonation, which disrupted the dense polymer matrix of kC. The recycled kCH-1 catalyst exhibited a more compact surface with a smoother region and showed irregular crystalline deposits, indicating partial structural degradation, which led to morphological changes and reduced porosity. The kCH-2 catalyst prepared by using a purely aqueous 0.84 M HCl solution shows a more fibrous, highly entangled, and densely aggregated

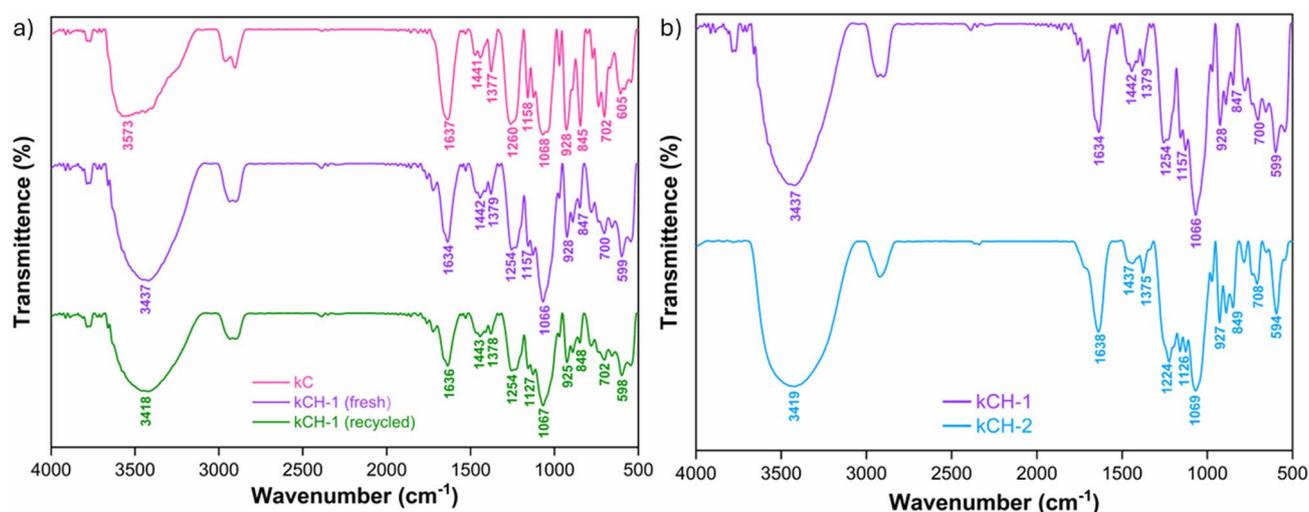


Fig. 1 (a) FTIR spectrum of kC, fresh kCH-1, and recycled kCH-1. (b) FTIR spectrum of kCH-1 and kCH-2.

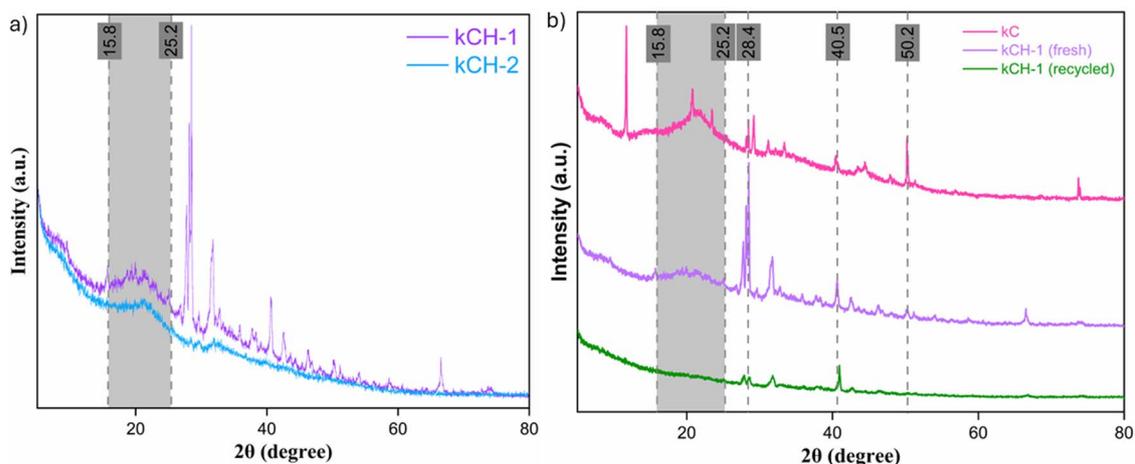


Fig. 2 (a) PXRD diffractogram of the two acid-treated catalysts kCH-1 and kCH-2. (b) PXRD diffraction pattern of kC, fresh kCH-1, and recycled kCH-1.

morphology, which suggests a stronger acid-induced restructuring of the carrageenan matrix.

The EDS analysis (Fig. 4) of kC, fresh kCH-1, and recycled kCH-1 revealed dominant carbon and oxygen peaks, which are expected from an organic polysaccharide. The sulfur content pointed to the presence of sulfate groups. The elemental composition changed noticeably after HCl treatment and after recycling. In kC, the corresponding peaks for sodium and potassium ions are noticeable, since these counterions balance the sulfate groups in kC. The potassium content was expected to be reduced in the kCH-1 catalyst compared to kC due to protonation. However, a higher amount of K was observed, likely due to the co-precipitation of KCl on the surface of kCH-1 during its preparation. Crystalline KCl gives a strong EDS signal and a sharp XRD diffraction at  $28.3^\circ$  (Fig. 2). In contrast, kCH-2, prepared without ethanol, displayed significantly lower Cl and

K contents than kCH-1, suggesting the presence of a lesser amount of KCl. The EDS confirms that the extent of protonation and mineral deposition depends significantly on the catalyst preparation.

Volumetric titration with 0.01 N KOH was used to assess the acid site density of kC, which was observed to steadily rise following HCl treatment. The acidity of the kC was  $0.05 \text{ mmol g}^{-1}$ , while the acidity of the kCH-1 catalyst was  $0.13 \text{ mmol g}^{-1}$ . The acid site density in kCH-2 was significantly higher with a value of  $1.12 \text{ mmol g}^{-1}$ , indicating a nearly tenfold increase in acid sites. The creation of strongly acidic sulfonic acid through the proton exchange of native  $\text{K}^+/\text{Na}^+$  ions introduces acidity into the kCH catalysts. Whereas recycled kCH-1 showed  $0.091 \text{ mmol g}^{-1}$ .

The ammonia temperature-programmed desorption ( $\text{NH}_3$ -TPD) analysis of kC, kCH-1, and kCH-2 was performed to check the acid value. However, these materials started to decompose

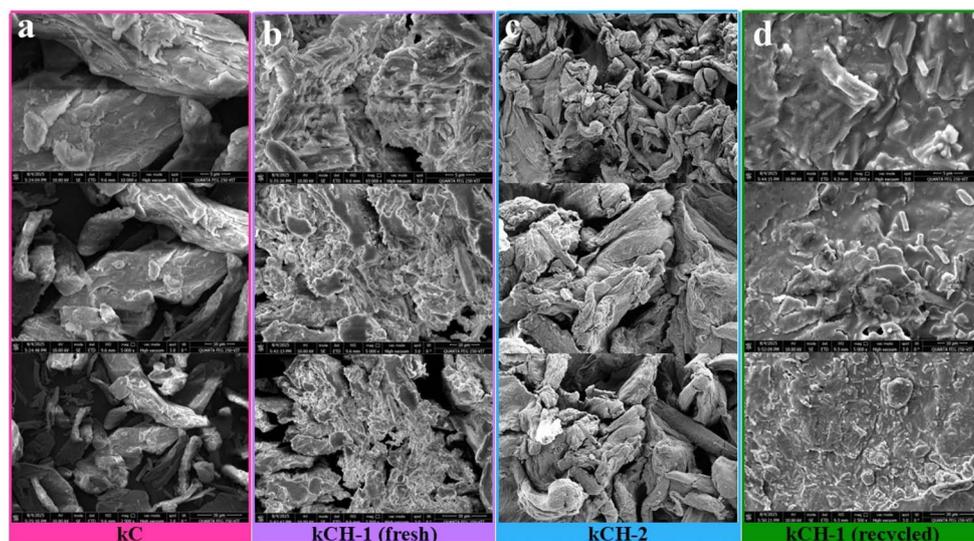


Fig. 3 FE-SEM images of: (a) kC, (b) fresh kCH-1, (c) fresh kCH-2, and (d) recycled kCH-1 at the magnification of  $5 \mu\text{m}$  (top),  $10 \mu\text{m}$  (middle), and  $30 \mu\text{m}$  (bottom).



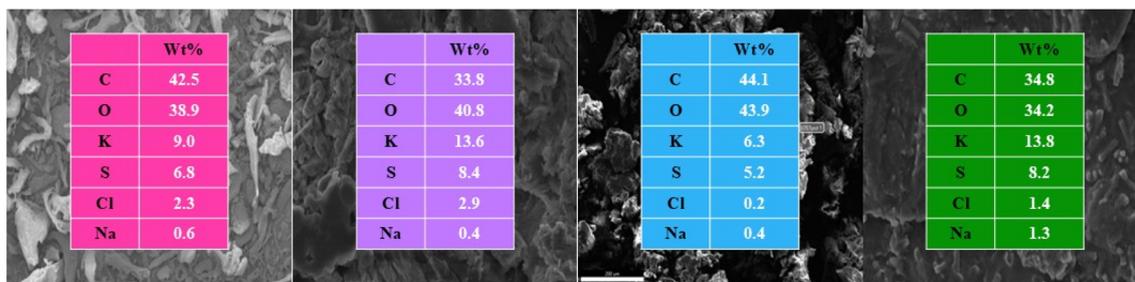


Fig. 4 EDS analysis of kC (pink), fresh kCH-1 (purple), kCH-2 (blue), and recycled kCH-1 (green).

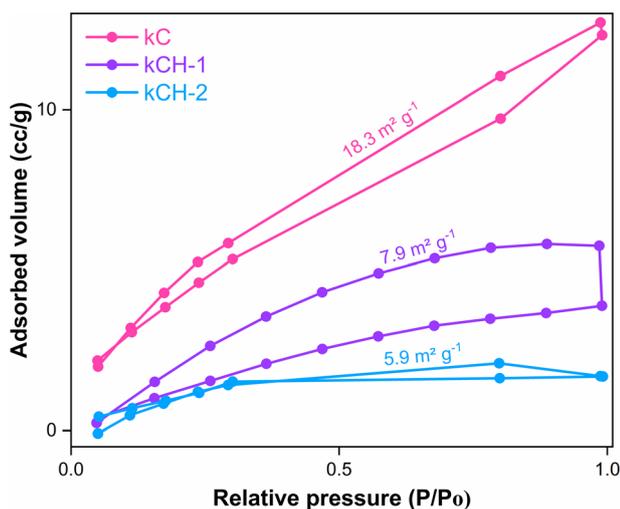


Fig. 5 BET analysis of kC, kCH-1, and the kCH-2 catalyst.

inside the cell during the analysis, resulting in irregular peaks on the graph. However, the acid sites present in kCH-1 and kCH-2 can be observed, whereas they were absent in kC (Fig. S29, SI).

The textural properties of the catalysts were investigated by nitrogen adsorption measurements at 77 K. The specific surface area was calculated using the BET method. The BET surface area of kC (pink) was found to be  $18.3 \text{ m}^2 \text{ g}^{-1}$ , which decreased to  $7.9 \text{ m}^2 \text{ g}^{-1}$  for kCH-1 (purple) and further to  $5.9 \text{ m}^2 \text{ g}^{-1}$  for kCH-2 (blue) after acid treatment (Fig. 5). This gradual reduction in surface area upon HCl protonation can be attributed to polymer chain rearrangement, partial pore collapse, and aggregation induced by strong hydrogen bonding. The total pore volume was determined at a relative pressure ( $p/p_0$ ) of approximately 0.99. The average pore diameter was obtained from nitrogen adsorption data based

on the average pore radius provided by the ASiQwin software (Table 1).

In both the kCH-1 and kCH-2 catalysts, the TGA data indicated the presence of two major mass-loss events (Fig. 6). The initial event is attributed to the removal of adsorbed and bound water or volatile species, which occurred at temperatures of approximately  $87.9 \text{ }^\circ\text{C}$  (kCH-1, 10.7% loss) and  $103 \text{ }^\circ\text{C}$  (kCH-2, 10.3% loss). The temperature is higher for kCH-2 compared to kCH-1, indicating that water is more strongly bound by the former. The second, much larger event is related to the thermolysis of the polysaccharide backbone (and possible loss of sulphate groups and low-molecular fragments) in the case of kCH-1 (24.93% loss) and in the case of kCH-2 (24.42% loss) at  $232 \text{ }^\circ\text{C}$  and  $161 \text{ }^\circ\text{C}$ , respectively. The higher decomposition temperature of kCH-1 as compared to that of kCH-2 is due to the difference in the extent of proton exchange. A greater extent of proton exchange in kCH-2 autocatalyzes its heat and acid-promoted depolymerization and dehydration reactions. Therefore, kCH-1 exhibited satisfactory thermal stability, which assures its reusability in the acetalization reaction under refluxing cyclohexane.

The thermogravimetric analysis (TGA) of kC, fresh kCH-1, and recycled kCH-1 shows different thermal stability (Fig. 7). The native kC displayed two large degradation events: loss of physisorbed and bound water at  $80.4 \text{ }^\circ\text{C}$  (weight loss, 9.23%), and cleavage of sulphate groups and the carrageenan backbone resulted in a second major event at  $253.3 \text{ }^\circ\text{C}$  (weight loss, 20.37%).<sup>41,47</sup> On protonation, fresh kCH-1 also underwent a two-stage degradation, with the first peak at  $87.9 \text{ }^\circ\text{C}$  (10.7% weight loss) and a second, stronger one at  $232.1 \text{ }^\circ\text{C}$  (24.93% weight loss), indicating that the protonated matrix was more sensitive to thermal conditions. On the other hand, the recycled kCH-1 showed a higher decomposition temperature at  $256.5 \text{ }^\circ\text{C}$ , which resulted in 31.87% weight loss, after an initial dehydration peak at  $72.4 \text{ }^\circ\text{C}$  (9.7% weight loss). The observed behaviour indicates

Table 1 The BET surface area, total pore volume, and average pore diameter of the kC, kCH-1, and kCH-2 catalyst

Sample	BET area ( $\text{m}^2 \text{ g}^{-1}$ )	Total pore volume ( $\text{cm}^3 \text{ g}^{-1}$ )	Average pore diameter (nm)
kC	18.3	0.006	3.0
kCH-1	7.9	0.002	4.1
kCH-2	5.9	0.003	1.7



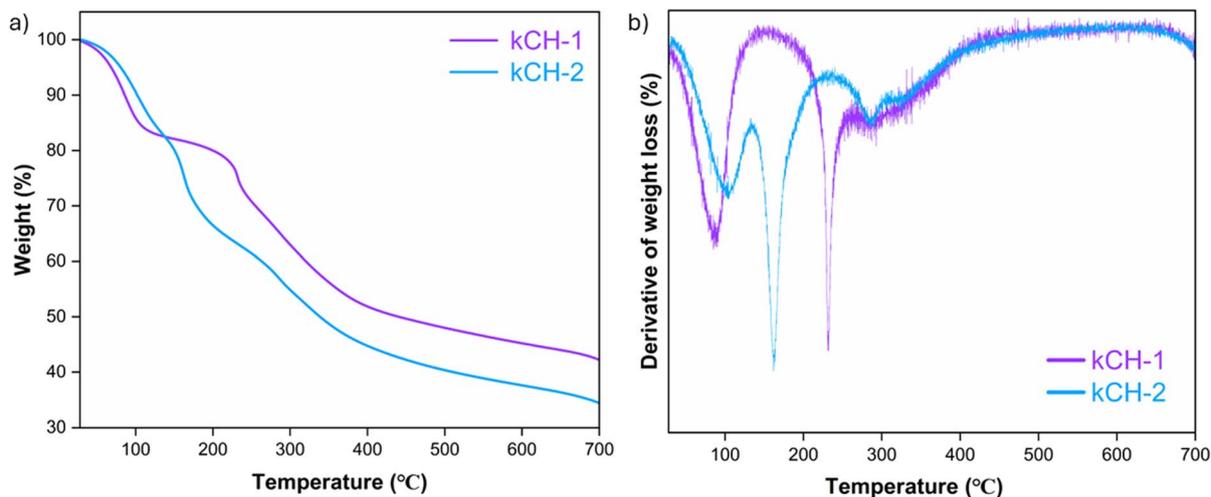


Fig. 6 (a) The thermogram of kCH-1 and kCH-2, and (b) DTG plot of kCH-1 and kCH-2.

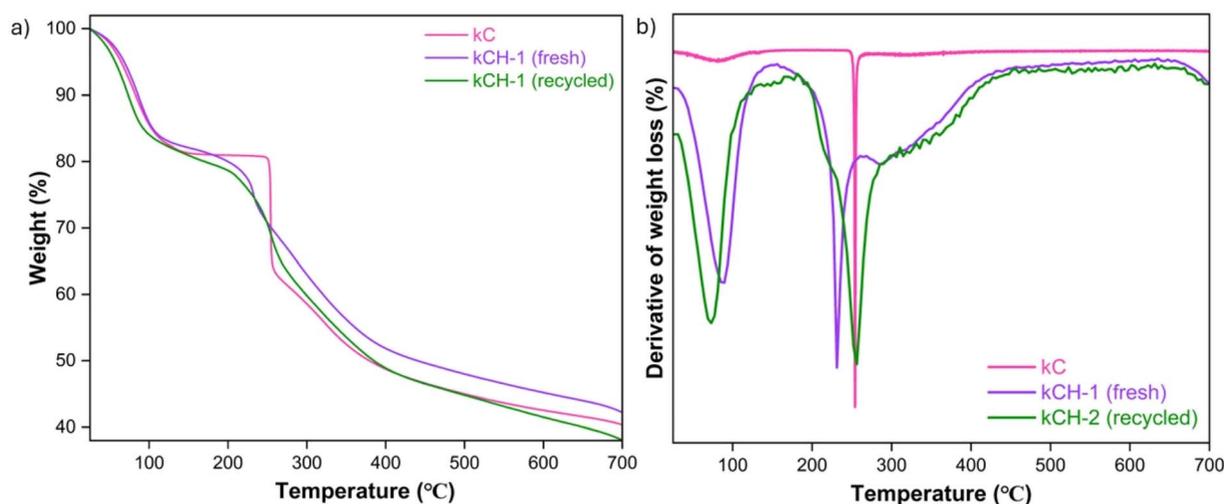


Fig. 7 (a) TGA analysis of kC, fresh kCH-1, and recycled kCH-1 catalyst. (b) DTG of kC, fresh kCH-1, and recycled kCH-1 catalyst.

that, even though kCH-1 maintains its general thermal pattern after recycling, the loss of acidity and partial carbonization are responsible for its apparent higher thermal stability.

In a typical process, FF, EG, and kCH-1 were added in cyclohexane, and the suspension was refluxed in a round-bottomed flask. The flask was connected to a Dean-Stark apparatus for azeotropic removal of water. The progress of the reaction was monitored by thin-layer chromatography (TLC) for the disappearance of FF. When the complete conversion of FF was achieved, the reaction mixture was cooled down to RT, filtered to remove the catalyst, washed with saturated NaHCO<sub>3</sub>, and the cyclohexane layer was separated. 2-(Furan-2-yl)-1,3-dioxolane (**1**) was isolated by evaporating cyclohexane under reduced pressure. Cyclohexane was the most effective of the various water removal agents.

The acetalization reaction in toluene requires a higher temperature due to the higher boiling point of toluene and the toluene–water azeotrope. Higher temperature degrades the

kCH-1 catalyst, making it challenging to recycle. Since cyclohexane is a greener solvent than benzene and toluene and afforded a slightly better yield of acetal **1**, it was considered the most suitable solvent and water removal agent for the reaction (Fig. 8).

Under optimized conditions, other heterogeneous catalysts with comparable acidity (*i.e.*, sulfonic acid), such as Amberlyst-15 and CSA, were also used, and their catalytic activity was compared with that of kCH-1 (Fig. 9). CSA and kCH-1 catalysts showed good catalytic activity for acetals. However, the conversion of FF was not complete even after extending the reaction duration to 8 h when CSA was used as a catalyst. However, kCH-1 was preferred since its synthesis is more eco-friendly than that of CSA. More specifically, kCH-1 was produced by the simple protonation of kC at ambient conditions, whereas CSA requires the sulfonation of cellulose using concentrated H<sub>2</sub>SO<sub>4</sub> or chlorosulfonic acid. The control reaction



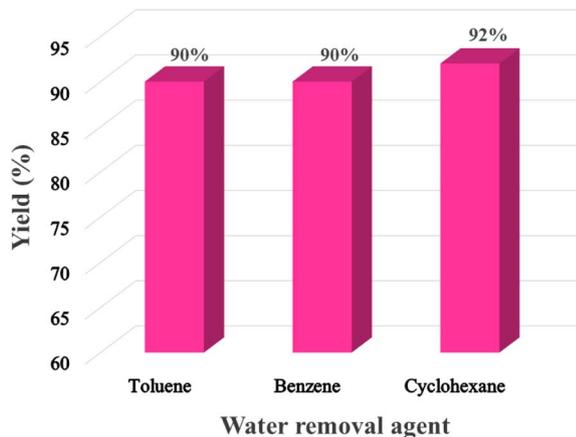


Fig. 8 Effect of water removal agent on the preparation of **1**.

(*i.e.*, without an acid catalyst) yielded only a trace amount of **1** after 8 h at 110 °C.

The molar ratio between **FF** and **EG** was varied for faster kinetics and better yields of **1**. Typically, an excess of the alcohol reagent is used for the acetalization reaction to favor the equilibrium towards the acetal. However, excess **EG** complicates the separation of acetal and mandates recycling for better process economy. Therefore, an optimal amount of **EG** must be used to make the process more reagent-economical without compromising reaction kinetics and product selectivity. When **FF** and **EG** were used in an equimolar ratio (1 : 1), a small amount of **FF** remained unreacted even after 7 h, indicating slower kinetics. In contrast, when the **FF** : **EG** molar ratio was increased to 1 : 1.5 with a 20 wt% catalyst loading, the reaction was completed within 5 h, indicating that a slight excess of **EG** significantly reduces the reaction time. The catalyst loading of **kCH-1** was also varied to see the effect on reaction kinetics and product selectivity. At a 10 wt% **kCH-1** loading (compared to the mass of **FF**), the reaction required about 8 h to ensure a quantitative conversion of **FF**, with the yield of acetal remaining almost unchanged compared to higher loadings. However, when the catalyst loading was increased to 20 wt%, the reaction completed within 5 h, indicating that a higher catalyst

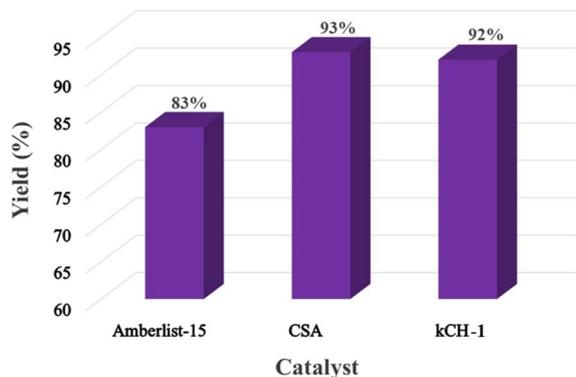


Fig. 9 Screening of sulfonic acid-based heterogeneous acid catalysts for preparing **1**.

concentration accelerates the reaction kinetics without significantly affecting the final yield.

Table 2 shows the molecular structure and isolated yield of the acetals (**1–9**) starting from **FF**, **MF**, and benzaldehyde (**BZ**) under the optimized reaction conditions. The acetals of **FF** with **EG**, **1,2-PDO**, and **1,3-PDO** afforded excellent isolated yields of the corresponding cyclic acetals (entries 1–3, Table 2). The acetals of **MF** (entries 4–6, Table 2) were also obtained in satisfactory isolated yields. The substrate scope of the acetalization process was explored by using **BZ** as the substrate, and no significant different in the selectivity or yield was observed (entries 7–9, Table 2).

The protonated form of ι-carrageenan was also screened for its catalytic activities. We envisioned that the acid density of ι-carrageenan would be significantly higher than that of **kC** since the former has two sulfate groups in the repeating unit compared to only one in the latter. The results showed comparable kinetics, yield, and recyclability of protonated ι-carrageenan to those of **kCH-1**. When purified **HMF** was used as a substrate for the acetalization reaction with **EG**, significant decomposition of **HMF** into insoluble humin was noted, resulting in significant material loss, and only a trace amount of the equivalent acetal was obtained. The observation is commensurate with the poor thermal and acid stability of **HMF**, which is well-documented in the literature.

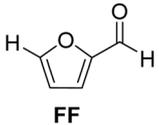
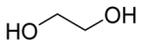
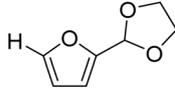
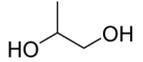
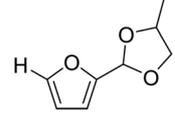
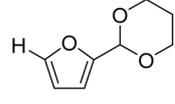
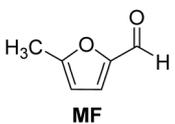
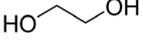
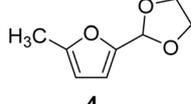
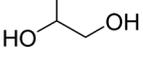
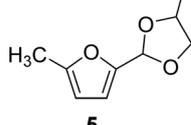
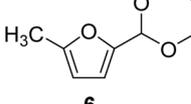
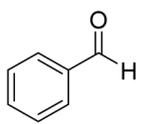
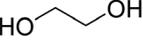
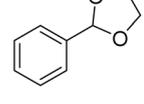
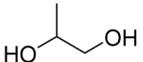
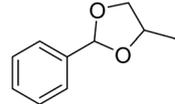
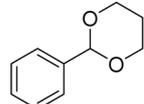
## 4 Recyclability of the **kCH-1** catalyst

Recyclability of the **kCH-1** (Fig. 10) catalyst was investigated over four consecutive cycles in the preparation of 2-(furan-2-yl)-1,3-dioxolane (**1**). The reaction kinetics remained relatively unaltered until the third cycle, indicating that the catalytic efficiency was maintained. However, the conversion of **FF** was not complete in the fourth cycle, even after an extended duration, hinting at a reduced catalyst activity. However, the yield of **1** at the fourth cycle was still satisfactory (85%). With each recycling, the **kCH-1** catalyst became increasingly colored and sticky (semi-solid), indicating partial depolymerization and decomposition.

The recycling of the **kCH-2** catalyst was investigated, revealing noticeably lower stability compared to the **kCH-1** catalyst. Partial decomposition of the **kCH-2** catalyst resulted in incomplete conversion of **FF** in the second cycle. Therefore, further recycling attempts of the **kCH-2** catalyst were not undertaken. An attempt was made to initially prepare the **kCH-2** catalyst by treating **kC** with 35% aqueous **HCl** (2 g **kC** with 6 mL **HCl**) and washing the filtered solid with cold water to remove excess **HCl**. However, the catalyst began to carbonize even at room temperature during the storage and drying process. Therefore, **kCH-2** was prepared using dil. **HCl** and the recyclability of the **kCH-2** catalyst were investigated. However, the **kCH-2** catalyst prepared in this way showed noticeably lower stability compared to the **kCH-1** catalyst. Partial decomposition of the **kCH-2** catalyst resulted in incomplete conversion of **FF** in the second cycle. Therefore, further recycling attempts of the **kCH-2** catalyst were not undertaken.



**Table 2** Synthesis of the cyclic acetals of FF, MF, and BZ using protonated  $\kappa$ -carrageenan as the heterogeneous acid catalyst. Reaction conditions: aldehyde (10 mmol), Diol (11 mmol), cyclohexane (15 mL), kCH-1 catalyst (20 wt% of aldehyde), 110 °C, 5 h

Entry	Aldehyde	Polyol	Cyclic acetal	Yield (%)
1	 FF	 EG	 1	92%
2		 1,2-PDO	 2	90%
3		 1,3-PDO	 3	89%
4	 MF	 EG	 4	91%
6		 1,2-PDO	 5	88%
7		 1,3-PDO	 6	88%
9	 BZ	 EG	 7	91%
10		 1,2-PDO	 8	91%
11		 1,3-PDO	 9	90%

The impact of the extent of protonation of kC on its stability and recyclability was further confirmed. Specifically, the kCH-2 catalyst was treated with an aqueous solution of KCl and then dried. The storage stability improved, and no color change was observed even after 2 days of storage at RT. The recyclability of the catalyst was evaluated over four cycles, yielding a similar

pattern to that of kCH-1 (Fig. S33, SI). The observation can be explained by the lowering of the acidity of kCH-2 by  $K^+$  in KCl, which replaces some protons, thereby increasing storage and thermal stability while improving recyclability. However, attempts to dry the catalyst at 60 °C resulted in partial



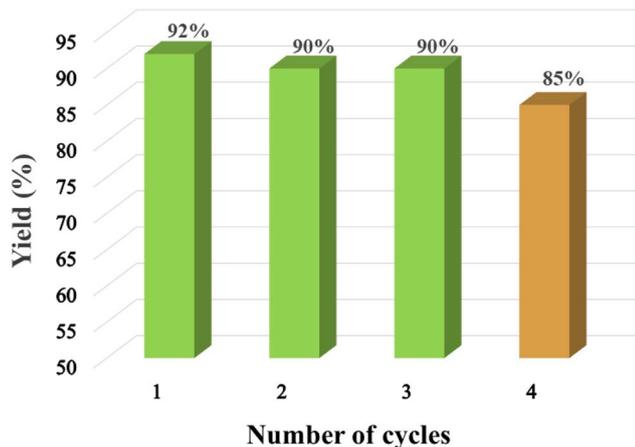


Fig. 10 Recyclability of the kCH-1 catalyst for synthesizing 1 (110 °C, 5 h).

decomposition, leading to a darkening of the color (Fig. S34, SI). Therefore, the catalyst was instead dried under vacuum at RT.

## 5 Conclusion

In conclusion, protonated  $\kappa$ -carrageenan was used as a marine biomass-derived renewable biopolymer-derived heterogeneous acid catalyst for the acetalization of carbohydrate-derived furfural and 5-methylfurfural. Ethylene glycol, 1,2-propanediol, and 1,3-propanediol were used as the diol reagents. Since the diols mentioned above can be renewably sourced from carbohydrates, this work expands the scope of employing biogenic carbon atoms in synthesizing value-added organic chemicals in a carbohydrate-centric biorefinery, where the substrate, reagent, and catalyst are sourced from carbohydrates. The reactions were performed in a Dean–Stark apparatus using cyclohexane as the solvent for water removal. Excellent isolated yields of the corresponding acetals (>90%) were obtained under optimized conditions. The broad substrate scope of the synthetic protocol was successfully demonstrated by preparing the acetals of benzaldehyde. Moreover, the catalyst was successfully recovered and reused for multiple cycles without a catastrophic drop in its catalytic efficiency. The future work will emphasize on the energy efficiency and detailed life-cycle analysis of the feedstock, reagents, and solvents employed in the synthetic process for quantitative evaluation of the environmental sustainability.

## Author contributions

Rachitha S. N. performed the experiments and analyzed the synthetic and spectroscopic data. Saikat Dutta conceptualized the idea, supervised the work, and wrote the original manuscript.

## Conflicts of interest

The authors declare no competing interests.

## Abbreviations

NH <sub>3</sub> -TPD	Ammonia temperature-programmed desorption
BET	Brunauer–Emmett–Teller
kC	$\kappa$ -Carrageenan
CSA	Cellulose sulfuric acid
EDS	Energy dispersive spectroscopy
EG	Ethylene glycol
FE-SEM	Field-emission scanning electron microscopy
FTIR	Fourier transform infrared
FF	Furfural
GLY	Glycerol
HMF	5-(Hydroxymethyl)furfural
MF	5-Methylfurfural
NMR	Nuclear magnetic resonance
PXRD	Powder X-ray diffraction
1,2-PDO	1,2-Propanediol
1,3-PDO	1,3-Propanediol
kCH	Protonated $\kappa$ -carrageenan
TLC	Thin-layer chromatography

## Data availability

Supplementary information (SI): The spectroscopic (FTIR, <sup>1</sup>H-NMR, and <sup>13</sup>C-NMR) characterization of all synthesized compounds reported in this manuscript. Photographic images of the reaction setup, EDS data, NH<sub>3</sub>-TPD data, TLC images, and images of the fresh and recycled catalysts have also been added. See DOI: <https://doi.org/10.1039/d5ra10090a>.

## Acknowledgements

This research received funding from DST-SERB, India, under the Core Research Grant (CRG) scheme (File no. CRG/2022/009346). The authors thank the Central Research Facility (CRF), NITK, Surathkal, for their assistance in the NMR characterization of the synthesized compounds and the characterization of the kCH-1 and kCH-2 catalysts. The authors thank Dr Navya Subray Bhat and Dr Navneet Kumar Gupta from Indian Institute of Science (IISc), Bangalore, for their assistance in collecting the NH<sub>3</sub>-TPD data.

## References

- G. Lopez, D. Keiner, M. Fasihi, T. Koiranen and C. Breyer, From Fossil to Green Chemicals: Sustainable Pathways and New Carbon Feedstocks for the Global Chemical Industry, *Energy Environ. Sci.*, 2023, **16**(7), 2879–2909, DOI: [10.1039/D3EE00478C](https://doi.org/10.1039/D3EE00478C).
- A. Arias, G. Feijoo and M. T. Moreira, Biorefineries as a Driver for Sustainability: Key Aspects, Actual Development and Future Prospects, *J. Clean. Prod.*, 2023, **418**, 137925, DOI: [10.1016/j.jclepro.2023.137925](https://doi.org/10.1016/j.jclepro.2023.137925).
- J. C. Serrano-Ruiz, R. M. West and J. A. Dumesic, Catalytic Conversion of Renewable Biomass Resources to Fuels and



- Chemicals, *Annu. Rev. Chem. Biomol. Eng.*, 2010, **1**(1), 79–100, DOI: [10.1146/annurev-chembioeng-073009-100935](https://doi.org/10.1146/annurev-chembioeng-073009-100935).
- 4 S. Takkellapati, T. Li and M. A. Gonzalez, An Overview of Biorefinery Derived Platform Chemicals from a Cellulose and Hemicellulose Biorefinery, *Clean Technol. Environ. Policy*, 2018, **20**(7), 1615–1630, DOI: [10.1007/s10098-018-1568-5](https://doi.org/10.1007/s10098-018-1568-5).
- 5 R. Bielski and G. Gryniewicz, Furan Platform Chemicals beyond Fuels and Plastics, *Green Chem.*, 2021, **23**(19), 7458–7487, DOI: [10.1039/D1GC02402G](https://doi.org/10.1039/D1GC02402G).
- 6 M. Dusselier, M. Mascal and B. F. Sels, Top Chemical Opportunities from Carbohydrate Biomass: A Chemist's View of the Biorefinery, in *Selective Catalysis for Renewable Feedstocks and Chemicals*, ed. K. M. Nicholas, Springer International Publishing, Cham, 2014, Topics in Current Chemistry, vol. 353, pp. 1–40, DOI: [10.1007/128\\_2014\\_544](https://doi.org/10.1007/128_2014_544).
- 7 A. Bayu, A. Abudula and G. Guan, Reaction Pathways and Selectivity in Chemo-Catalytic Conversion of Biomass-Derived Carbohydrates to High-Value Chemicals: A Review, *Fuel Process. Technol.*, 2019, **196**, 106162, DOI: [10.1016/j.fuproc.2019.106162](https://doi.org/10.1016/j.fuproc.2019.106162).
- 8 P. Sarkar, T. K. Bandyopadhyay, K. Gopikrishna, O. N. Tiwari, B. Bhunia and M. Muthuraj, Algal Carbohydrates: Sources, Biosynthetic Pathway, Production, and Applications, *Bioresour. Technol.*, 2024, **413**, 131489, DOI: [10.1016/j.biortech.2024.131489](https://doi.org/10.1016/j.biortech.2024.131489).
- 9 G. Shen, B. Andrioletti and Y. Queneau, Furfural and 5-(Hydroxymethyl)Furfural: Two Pivotal Intermediates for Bio-Based Chemistry, *Curr. Opin. Green Sustainable Chem.*, 2020, **26**, 100384, DOI: [10.1016/j.cogsc.2020.100384](https://doi.org/10.1016/j.cogsc.2020.100384).
- 10 X. Zhang, S. Xu, Q. Li, G. Zhou and H. Xia, Recent Advances in the Conversion of Furfural into Bio-Chemicals through Chemo- and Bio-Catalysis, *RSC Adv.*, 2021, **11**(43), 27042–27058, DOI: [10.1039/D1RA04633K](https://doi.org/10.1039/D1RA04633K).
- 11 A. E. Eseyin and P. H. Steele, An Overview of the Applications of Furfural and Its Derivatives, *Int. J. Adv. Chem.*, 2015, **3**(2), 42–47, DOI: [10.14419/ijac.v3i2.5048](https://doi.org/10.14419/ijac.v3i2.5048).
- 12 Y. Bao, Z. Du, X. Liu, H. Liu, J. Tang, C. Qin, C. Liang, C. Huang and S. Yao, Furfural Production from Lignocellulosic Biomass: One-Step and Two-Step Strategies and Techno-Economic Evaluation, *Green Chem.*, 2024, **26**(11), 6318–6338, DOI: [10.1039/D4GC00883A](https://doi.org/10.1039/D4GC00883A).
- 13 Q.-S. Kong, X.-L. Li, H.-J. Xu and Y. Fu, Conversion of 5-Hydroxymethylfurfural to Chemicals: A Review of Catalytic Routes and Product Applications, *Fuel Process. Technol.*, 2020, **209**, 106528, DOI: [10.1016/j.fuproc.2020.106528](https://doi.org/10.1016/j.fuproc.2020.106528).
- 14 A. S. Wagh, T. M. Ukarde, P. H. Pandey, A. M. Lali and H. S. Pawar, Self-Catalyzed Deconstruction of Acid-Modified  $\kappa$ -Carrageenan for Production of 5-Hydroxymethyl Furfural, *ACS Sustain. Chem. Eng.*, 2019, **7**(16), 13932–13940, DOI: [10.1021/acssuschemeng.9b02186](https://doi.org/10.1021/acssuschemeng.9b02186).
- 15 A. Jaswal, P. P. Singh and T. Mondal, Furfural – a Versatile, Biomass-Derived Platform Chemical for the Production of Renewable Chemicals, *Green Chem.*, 2022, **24**(2), 510–551, DOI: [10.1039/D1GC03278J](https://doi.org/10.1039/D1GC03278J).
- 16 K. I. Galkin and V. P. Ananikov, Towards Improved Biorefinery Technologies: 5-Methylfurfural as a Versatile C<sub>6</sub> Platform for Biofuels Development, *ChemSusChem*, 2019, **12**(1), 185–189, DOI: [10.1002/cssc.201802126](https://doi.org/10.1002/cssc.201802126).
- 17 L. Dong, J. Morales-Vidal, L. Mu, L. Li, N. López, J. Pérez-Ramírez and Z. Chen, Selective Hydrogenolysis of 5-Hydroxymethylfurfural to 5-Methylfurfural over Au/TiO<sub>2</sub>, *Appl. Catal., B*, 2023, **335**, 122893, DOI: [10.1016/j.apcatb.2023.122893](https://doi.org/10.1016/j.apcatb.2023.122893).
- 18 W. Yang and A. Sen, Direct Catalytic Synthesis of 5-Methylfurfural from Biomass-Derived Carbohydrates, *ChemSusChem*, 2011, **4**(3), 349–352, DOI: [10.1002/cssc.201000369](https://doi.org/10.1002/cssc.201000369).
- 19 S. Dutta and M. Mascal, Novel Pathways to 2,5-Dimethylfuran via Biomass-Derived 5-(Chloromethyl) Furfural, *ChemSusChem*, 2014, **7**(11), 3028–3030, DOI: [10.1002/cssc.201402702](https://doi.org/10.1002/cssc.201402702).
- 20 Z. He, P. Jiang, Q. Cui, Z. Wang, Y. Wei, C. Luo, J. Guo, C. Liu and W. Zhang, Efficient Method for the Synthesis of 5-Methylfurfural from l-Rhamnose Using a Biphasic System, *Catalysts*, 2025, **15**(5), 465, DOI: [10.3390/catal15050465](https://doi.org/10.3390/catal15050465).
- 21 Y. Feng, Z. Li, S. Long, Y. Sun, X. Tang, X. Zeng and L. Lin, Direct Conversion of Biomass Derived l-Rhamnose to 5-Methylfurfural in Water in High Yield, *Green Chem.*, 2020, **22**(18), 5984–5988, DOI: [10.1039/D0GC02105A](https://doi.org/10.1039/D0GC02105A).
- 22 J. He, Q. Qiang, L. Bai, W. Su, H. Yu, S. Liu and C. Li, Acetalization Strategy in Biomass Valorization: A Review, *Ind. Chem. Mater.*, 2024, **2**(1), 30–56, DOI: [10.1039/D3IM00050H](https://doi.org/10.1039/D3IM00050H).
- 23 S. Kirchhecker, A. Dell'Acqua, A. Angenvoort, A. Spannenberg, K. Ito, S. Tin, A. Taden and J. G. de Vries, HMF–Glycerol Acetals as Additives for the Debonding of Polyurethane Adhesives, *Green Chem.*, 2021, **23**(2), 957–965, DOI: [10.1039/D0GC04093B](https://doi.org/10.1039/D0GC04093B).
- 24 J. M. J. M. Ravasco and R. F. A. Gomes, Recent Advances on Diels-Alder-Driven Preparation of Bio-Based Aromatics, *ChemSusChem*, 2021, **14**(15), 3047–3053, DOI: [10.1002/cssc.202100813](https://doi.org/10.1002/cssc.202100813).
- 25 F. Guerrero-Ruiz, E. Yara-Varón, M. Dolores González, M. Torres, P. Salagre, R. Canela-Garayoa and Y. Cesteros, Use of Biobased Crude Glycerol, Obtained Biocatalytically, to Obtain Biofuel Additives by Catalytic Acetalization of Furfural Using SAPO Catalysts, *Fuel*, 2022, **319**, 123803, DOI: [10.1016/j.fuel.2022.123803](https://doi.org/10.1016/j.fuel.2022.123803).
- 26 H. Song, F. Jin, Q. Liu and H. Liu, Zeolite-Catalyzed Acetalization Reaction of Furfural with Alcohol under Solvent-Free Conditions, *Mol. Catal.*, 2021, **513**, 111752, DOI: [10.1016/j.mcat.2021.111752](https://doi.org/10.1016/j.mcat.2021.111752).
- 27 A. Mohsenzadeh, A. Zamani and M. J. Taherzadeh, Bioethylene Production from Ethanol: A Review and Techno-economical Evaluation, *ChemBioEng Rev.*, 2017, **4**(2), 75–91, DOI: [10.1002/cben.201600025](https://doi.org/10.1002/cben.201600025).
- 28 M. K. Wong, S. S. M. Lock, Y. H. Chan, S. J. Yeoh and I. S. Tan, Towards Sustainable Production of Bio-Based Ethylene Glycol: Progress, Perspective and Challenges in Catalytic Conversion and Purification, *Chem. Eng. J.*, 2023, **468**, 143699, DOI: [10.1016/j.cej.2023.143699](https://doi.org/10.1016/j.cej.2023.143699).
- 29 D. K. Pandey and P. Biswas, Continuous Production of Propylene Glycol (1,2-Propanediol) by the Hydrogenolysis



- of Glycerol over a Bi-Functional Cu–Ru/MgO Catalyst, *React. Chem. Eng.*, 2020, 5(12), 2221–2235, DOI: [10.1039/D0RE00285B](https://doi.org/10.1039/D0RE00285B).
- 30 A. D. da Silva Ruy, R. M. de Brito Alves, T. L. Reis Hower, D. de Aguiar Pontes, L. S. Gomes Teixeira and L. A. Magalhães Pontes, Catalysts for Glycerol Hydrogenolysis to 1,3-Propanediol: A Review of Chemical Routes and Market, *Catal. Today*, 2021, 381, 243–253, DOI: [10.1016/j.cattod.2020.06.035](https://doi.org/10.1016/j.cattod.2020.06.035).
- 31 H. Chi, Z. Liang, S. Kuang, Y. Jin, M. Li, T. Yan, J. Lin, S. Wang, S. Zhang and X. Ma, Electrosynthesis of Ethylene Glycol from Biomass Glycerol, *Nat. Commun.*, 2025, 16(1), 979, DOI: [10.1038/s41467-025-56104-5](https://doi.org/10.1038/s41467-025-56104-5).
- 32 L. Bai, M. Wu, H. Yu, S. Liu, K. Song, X. Zhou, J. Guo and J. He, Catalyst-Free Acetalization of Biobased Furfural into Cyclic Acetal Fuel Additives with Biogenic Ethylene Glycol, *ACS Sustain. Chem. Eng.*, 2023, 11(43), 15743–15753, DOI: [10.1021/acssuschemeng.3c05408](https://doi.org/10.1021/acssuschemeng.3c05408).
- 33 I. Kopa, O. Yevdokimova, M. E. Martínez-Klimov, M. Kurmach, D. Y. Murzin and N. Shcherban, Furfural Acetalization with Ethanol over Hierarchical vs. Conventional Beta Zeolites, *ChemistrySelect*, 2024, 9(15), e202304754, DOI: [10.1002/slct.202304754](https://doi.org/10.1002/slct.202304754).
- 34 M. J. da Silva, M. G. Teixeira and R. Natalino, Highly Selective Synthesis under Benign Reaction Conditions of Furfural Dialkyl Acetal Using SnCl<sub>2</sub> as a Recyclable Catalyst, *New J. Chem.*, 2019, 43(22), 8606–8612, DOI: [10.1039/C9NJ01284B](https://doi.org/10.1039/C9NJ01284B).
- 35 P. R. Yaashikaa, P. S. Kumar and S. Karishma, Bio-Derived Catalysts for Production of Biodiesel: A Review on Feedstock, Oil Extraction Methodologies, Reactors and Lifecycle Assessment of Biodiesel, *Fuel*, 2022, 316, 123379, DOI: [10.1016/j.fuel.2022.123379](https://doi.org/10.1016/j.fuel.2022.123379).
- 36 L. Bai, F. Deng, S. Wu, S. Liu, K. Song, X. Zhou, J. Guo, J. He, X. Liu and C. Li, Construction of Lignin-Based Heterogeneous Acid Catalysts with an Excellent Performance for the Acetalization of Biomass-Based Furfural and Glycerol, *ACS Sustain. Chem. Eng.*, 2025, 13(20), 7631–7644, DOI: [10.1021/acssuschemeng.5c02613](https://doi.org/10.1021/acssuschemeng.5c02613).
- 37 M. Belluati, S. Tabasso, E. Calcio Gaudino, G. Cravotto and M. Manzoli, Biomass-Derived Carbon-Based Catalysts for Lignocellulosic Biomass and Waste Valorisation: A Circular Approach, *Green Chem.*, 2024, 26(15), 8642–8668, DOI: [10.1039/D4GC00606B](https://doi.org/10.1039/D4GC00606B).
- 38 V. J. Kolcsár and G. Szöllösi, Chitosan as a Chiral Ligand and Organocatalyst: Preparation Conditions–Property–Catalytic Performance Relationships, *Catal. Sci. Technol.*, 2021, 11(23), 7652–7666, DOI: [10.1039/D1CY01674A](https://doi.org/10.1039/D1CY01674A).
- 39 P. S. Prabhakar and S. Dutta, Ketalization of Carbohydrate-Derived Levulinic Esters Using Cellulose Sulfuric Acid as a Heterogeneous Catalyst: A Closed-Loop Biorefinery Approach, *RSC Adv.*, 2025, 15(15), 11301–11307, DOI: [10.1039/D5RA00610D](https://doi.org/10.1039/D5RA00610D).
- 40 P. Volery, R. Besson and C. Schaffer-Lequart, Characterization of Commercial Carrageenans by Fourier Transform Infrared Spectroscopy Using Single-Reflection Attenuated Total Reflection, *J. Agric. Food Chem.*, 2004, 52(25), 7457–7463, DOI: [10.1021/jf040229o](https://doi.org/10.1021/jf040229o).
- 41 J. W. Y. Liew, K. S. Loh, A. Ahmad, K. L. Lim and W. R. Wan Daud, Synthesis and Characterization of Modified κ-Carrageenan for Enhanced Proton Conductivity as Polymer Electrolyte Membrane, *PLoS One*, 2017, 12(9), e0185313, DOI: [10.1371/journal.pone.0185313](https://doi.org/10.1371/journal.pone.0185313).
- 42 F. Adam, J. Jamaludin, S. H. Abu Bakar, R. Abdul Rasid and Z. Hassan, Evaluation of Hard Capsule Application from Seaweed: Gum Arabic-Kappa Carrageenan Biocomposite Films, *Cogent Eng.*, 2020, 7(1), 1765682, DOI: [10.1080/23311916.2020.1765682](https://doi.org/10.1080/23311916.2020.1765682).
- 43 R. P. Millane, R. Chandrasekaran, S. Arnott and I. C. M. Dea, The Molecular Structure of Kappa-Carrageenan and Comparison with Iota-Carrageenan, *Carbohydr. Res.*, 1988, 182(1), 1–17, DOI: [10.1016/0008-6215\(88\)84087-4](https://doi.org/10.1016/0008-6215(88)84087-4).
- 44 N. S. Anderson, J. W. Campbell, M. M. Harding, D. A. Rees and J. W. B. Samuel, X-Ray Diffraction Studies of Polysaccharide Sulphates: Double Helix Models for κ- and ι-Carrageenans, *J. Mol. Biol.*, 1969, 45(1), 85–97, DOI: [10.1016/0022-2836\(69\)90211-3](https://doi.org/10.1016/0022-2836(69)90211-3).
- 45 E. N. Dewi, R. Ibrahim and S. Suharto, Morphological Structure Characteristic and Quality of Semi Refined Carrageenan Processed by Different Drying Methods, *Procedia Environ. Sci.*, 2015, 23, 116–122, DOI: [10.1016/j.proenv.2015.01.018](https://doi.org/10.1016/j.proenv.2015.01.018).
- 46 M. Zhu, L. Ge, Y. Lyu, Y. Zi, X. Li, D. Li and C. Mu, Preparation, Characterization and Antibacterial Activity of Oxidized κ-Carrageenan, *Carbohydr. Polym.*, 2017, 174, 1051–1058, DOI: [10.1016/j.carbpol.2017.07.029](https://doi.org/10.1016/j.carbpol.2017.07.029).
- 47 F. Zhou, D. Wang, J. Zhang, J. Li, D. Lai, S. Lin and J. Hu, Preparation and Characterization of Biodegradable κ-Carrageenan Based Anti-Bacterial Film Functionalized with Wells-Dawson Polyoxometalate, *Foods*, 2022, 11(4), 586, DOI: [10.3390/foods11040586](https://doi.org/10.3390/foods11040586).

