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# Towards greener reduced graphene oxide: a critical review of environmentally driven reduction strategies

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The synthesis of graphene-based materials has attracted immense interest due to their exceptional properties. However, graphene oxide (GO), a common precursor, contains oxygen-containing functional groups that disrupt its  $sp^2$  carbon network, thereby limiting its electrical conductivity and other key properties. The reduction of GO to reduced graphene oxide (rGO) is therefore a crucial step in restoring these properties. Traditional reduction methods often use toxic, hazardous chemical reagents, such as hydrazine, which pose significant environmental and health risks. Consequently, there is a pressing need for environmentally benign, sustainable, and cost-effective reduction strategies. This review provides a critical examination of green reduction methods for GO, focusing on plant extracts, microorganisms, and isolated biomolecules as sustainable reducing agents. It moves beyond a simple summary of existing literature to offer a comparative analysis of these methods, evaluating their reduction efficacy based on key material properties, such as the C/O ratio, electrical conductivity, and structural integrity, as determined by spectroscopic and microscopic techniques (UV-vis, XRD, Raman, XPS, SEM, TEM). The central focus of this review is to establish a clear link between the choice of green reduction strategy, the resulting physicochemical properties of the rGO, and its performance in specific technological applications, including energy storage, sensing, environmental remediation, and biomedicine. By analyzing reaction mechanisms, scalability, and application-specific outcomes, this review identifies current research gaps and provides a forward-looking perspective on the rational design of green-synthesized rGO for advanced, sustainable technologies.

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## 1 Introduction

Graphene, a two-dimensional monolayer of  $sp^2$ -hybridized carbon atoms arranged in a hexagonal lattice, was first isolated in 2004, marking the beginning of a new era in materials science.<sup>1</sup> Its extraordinary electrical, thermal, mechanical, and optical properties have positioned it as a transformative material for applications in electronics, drug delivery, sensors, and energy storage.<sup>2–6</sup> One of the most viable routes to large-scale production of graphene-based materials is chemical exfoliation of graphite. This top-down approach, however, yields graphene oxide (GO) rather than pristine graphene.<sup>7–10</sup> GO is a graphene sheet decorated with oxygen-containing functional groups, primarily hydroxyl and epoxide groups on its basal plane, and carboxyl and carbonyl groups at its edges (Fig. 1).<sup>11</sup>

The synthesis of GO is typically achieved through strong oxidation of graphite, with methods developed by Brodie, Staudenmaier, and Hummers being foundational.<sup>12–15</sup> The modified Hummers' method is now widely used as it improves safety and efficiency.<sup>16</sup> While the oxygen functional groups render GO hydrophilic and dispersible, they disrupt the  $\pi$ -conjugated network, making it electrically insulating and unsuitable for many applications. To restore the graphene-like properties, GO must be reduced to rGO. The primary goal of this reduction is to remove the oxygen-containing groups, thereby recovering the  $sp^2$ -conjugated structure and significantly enhancing electrical and thermal conductivity.<sup>17</sup> The effectiveness of any reduction process is typically quantified by the increase in the carbon-to-oxygen (C/O) ratio and the corresponding improvement in electrical conductivity.<sup>18</sup>

Conventional reduction methods often rely on highly toxic and hazardous chemicals, such as hydrazine hydrate and sodium borohydride, or on energy-intensive thermal annealing processes.<sup>19,20</sup> These approaches raise significant environmental and safety concerns. In response, the field has shifted towards "green" reduction strategies that utilize non-toxic, sustainable, and cost-effective reducing agents derived from

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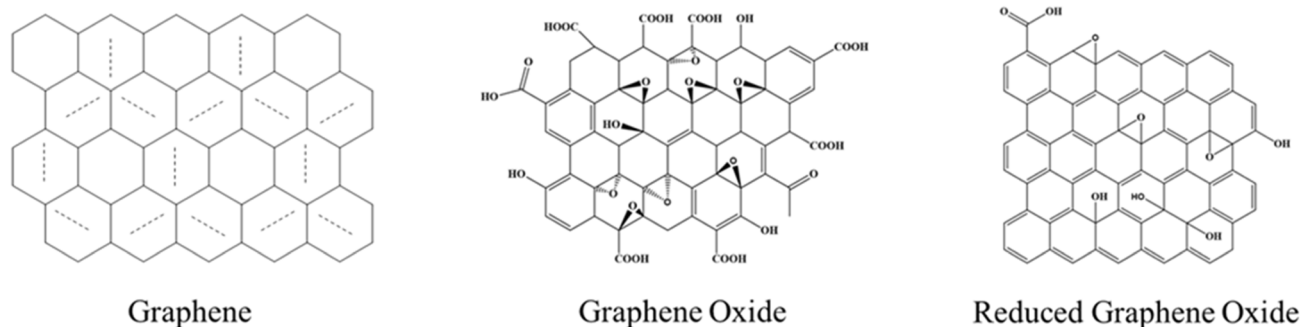



Fig. 1 Structures of graphene (G), graphene oxide (GO), and reduced graphene oxide (rGO), illustrating the removal of oxygen functional groups during reduction.

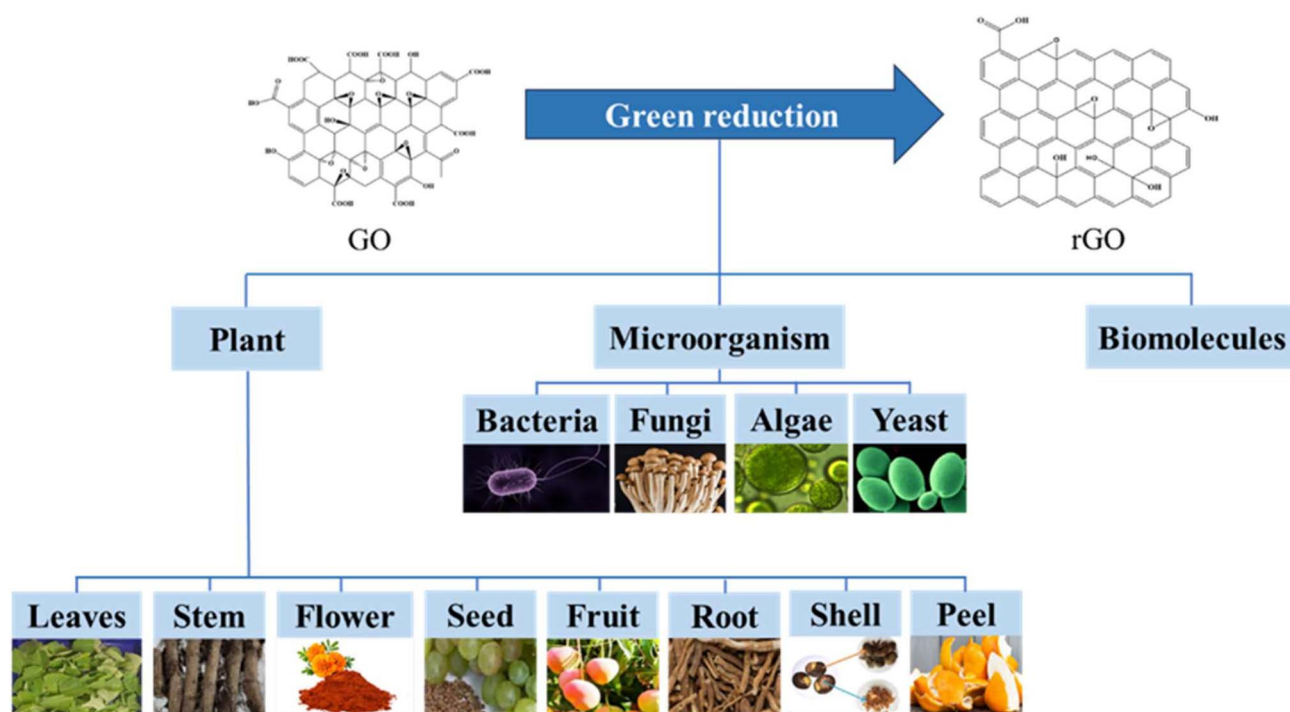


Fig. 2 Classification of green reductants for the reduction of GO into three main categories: plants, microorganisms, and biomolecules, with further subdivisions.

natural sources. As illustrated in Fig. 2, these green reductants can be broadly classified into three categories: plant extracts, microorganisms (*e.g.*, bacteria, fungi, yeast), and isolated biomolecules (*e.g.*, vitamins, amino acids, sugars).<sup>21–24</sup> The phytochemicals, enzymes, and other bioactive compounds within these sources not only reduce GO but often act as stabilizing agents, preventing the agglomeration of rGO sheets.<sup>25</sup>

While several works have explored the green synthesis of rGO, they have served mainly as summaries of published reports. This review aims to provide a more critical and analytical perspective. This work not only surveys the various green reduction strategies but also critically evaluates their efficacy by directly linking the choice of reductant and reaction conditions to the final material properties. By integrating

discussions of synthesis, characterization, and application, this review aims to establish a clearer understanding of the structure–property–application relationships for green-synthesized rGO. The objective is to move beyond mere cataloging and provide deeper insights into mechanistic understanding, identify specific research gaps, and offer a forward-looking perspective to guide the rational design of rGO for targeted, high-impact applications.

## 2 Fundamental properties of GO and rGO: the motivation for reduction

The transformation from GO to rGO involves a fundamental change in material properties, driven by the removal of oxygen functional groups and the restoration of the  $sp^2$  carbon lattice.

Understanding these changes is crucial to appreciating the importance of the reduction process.

## 2.1 Electrical conductivity

Pristine graphene is a semi-metal with an electrical conductivity that can reach up to  $2000 \text{ S cm}^{-1}$ .<sup>26</sup> In contrast, GO is an electrical insulator, with sheet resistance values often exceeding  $10^{12} \Omega \text{ sq}^{-1}$  and conductivity as low as  $0.0486 \text{ S m}^{-1}$ .<sup>27–29</sup> The insulating nature is a direct consequence of the  $\text{sp}^3 \text{ C-O}$  bonds that disrupt the percolating pathways for charge carriers among the remaining  $\text{sp}^2$  carbon clusters. The reduction process systematically removes these  $\text{sp}^3$  defects, restoring the conjugated  $\text{sp}^2$  network and dramatically increasing electrical conductivity. The final conductivity of rGO is highly dependent on the degree of reduction. For instance, rGO synthesized using gallic acid exhibits a modest conductivity of  $0.358 \text{ S m}^{-1}$ ,<sup>30</sup> whereas microbial reduction with *Shewanella oneidensis* yields a conductivity of  $55.32 \text{ S m}^{-1}$ .<sup>31</sup> While these values represent a significant improvement over GO, they remain lower than those of rGO produced via hazardous chemical reductants, such as hydrazine ( $216.56 \text{ S m}^{-1}$ ),<sup>31</sup> highlighting the ongoing challenge of achieving complete reduction using green methods.

## 2.2 Thermal conductivity

Graphene possesses an exceptionally high thermal conductivity ( $4840\text{--}5300 \text{ W m}^{-1} \text{ K}^{-1}$ ), making it ideal for thermal management applications.<sup>32</sup> The oxidation process introduces structural defects and vacancies that act as phonon scattering sites, causing the thermal conductivity of GO to plummet to around  $2.90 \text{ W m}^{-1} \text{ K}^{-1}$ . The reduction to rGO partially restores the lattice integrity, leading to a significant recovery in thermal conductivity. For example, an rGO film produced using dopamine as a reductant exhibited a thermal conductivity of  $13.42 \text{ W m}^{-1} \text{ K}^{-1}$ ,<sup>33</sup> and values as high as  $500 \text{ W m}^{-1} \text{ K}^{-1}$  have been reported for well-reduced, annealed films.<sup>34</sup>

## 2.3 Mechanical strength

While GO possesses impressive mechanical strength, it is inherently lower than that of pristine graphene due to the structural defects introduced by the oxygen functional groups. The reduction process, by restoring the  $\text{sp}^2$ -conjugated structure, generally enhances mechanical properties. The Young's modulus of an rGO film can be significantly higher than that of a GO film. For example, one study reported that the Young's modulus increased from  $27.3 \text{ GPa}$  for an unannealed rGO film to  $158.0 \text{ GPa}$  after annealing, demonstrating the improved rigidity that comes with a more ordered, graphene-like structure.<sup>35</sup>

## 2.4 Chemical stability and dispersibility

The abundant oxygen functional groups on GO make it highly reactive and hydrophilic, allowing it to form stable colloidal dispersions in water. This property is advantageous for solution-based processing. However, upon reduction, the removal of these polar groups restores the hydrophobic nature of the carbon lattice. Consequently, rGO sheets have a strong

tendency to agglomerate and restack in aqueous solutions due to  $\pi$ - $\pi$  interactions and van der Waals forces, making them less dispersible.<sup>36,37</sup> A key advantage of many green reduction methods is that the phytochemicals or biomolecules used for reduction often co-functionalize the rGO surface, acting as stabilizing or capping agents that improve dispersibility without compromising the restored electronic properties.

## 2.5 Surface area

Theoretically, GO can have a surface area as high as  $736.6 \text{ m}^2 \text{ g}^{-1}$ .<sup>38</sup> However, in practice, the wrinkling and distortion of the sheets caused by functional groups lead to much lower values (e.g.,  $18 \text{ m}^2 \text{ g}^{-1}$ ).<sup>17</sup> The reduction process typically increases the specific surface area by removing the bulky oxygen groups and creating a more porous, accessible structure. The final surface area is highly dependent on the reductant. rGO produced with lemon juice showed a surface area of  $159 \text{ m}^2 \text{ g}^{-1}$ ,<sup>17</sup> while microbial reduction with *Shewanella* achieved  $243.24 \text{ m}^2 \text{ g}^{-1}$ .<sup>39</sup> These values, while substantial, are often lower than those achieved with chemical reductants like hydrazine ( $400\text{--}700 \text{ m}^2 \text{ g}^{-1}$ ),<sup>40</sup> indicating that green methods can sometimes leave behind residues or cause more sheet aggregation, affecting the final morphology.

# 3 Green reduction strategies: a critical and integrated analysis

A critical analysis of the three primary green reduction strategies is provided in this section. For each method, the general methodology is discussed, the reduction efficacy is evaluated by synthesizing data from the literature, the proposed chemical mechanisms are compared, and direct links are made between the resulting material properties and their performance in specific, high-value applications.

## 3.1 Reduction of GO using plant extracts

Plant extracts have emerged as one of the most popular choices for green GO reduction due to their low cost, ready availability, and rich composition of phytochemicals, including polyphenols (flavonoids, tannins), alkaloids, and vitamins, which can act as potent reducing agents.<sup>41–44</sup>

**3.1.1 General methodology and reduction efficacy.** The typical procedure involves preparing an aqueous extract from a specific plant part (leaves, roots, fruit, etc.), mixing it with a GO dispersion, and heating the mixture, often under reflux, until the color changes from brown to black, signifying the formation of rGO (Fig. 3).<sup>45,46</sup> The resulting rGO is then washed and dried.

A critical evaluation of the extensive literature reveals a wide range of reduction efficacy depending on the plant source and reaction conditions. A meta-analysis of the studies compiled in Table 1 shows several key trends. Firstly, extracts derived from sources known for high concentrations of potent antioxidants, such as artemisinin and wild carrot root, achieve exceptionally high C/O ratios of 11.7 and 11.9, respectively.<sup>47,48</sup> These values are superior to many other green methods and are competitive with the C/O ratio of 9.82 obtained using hydrazine.<sup>49</sup> This suggests that the specific chemical nature of the reductant is more critical than its general



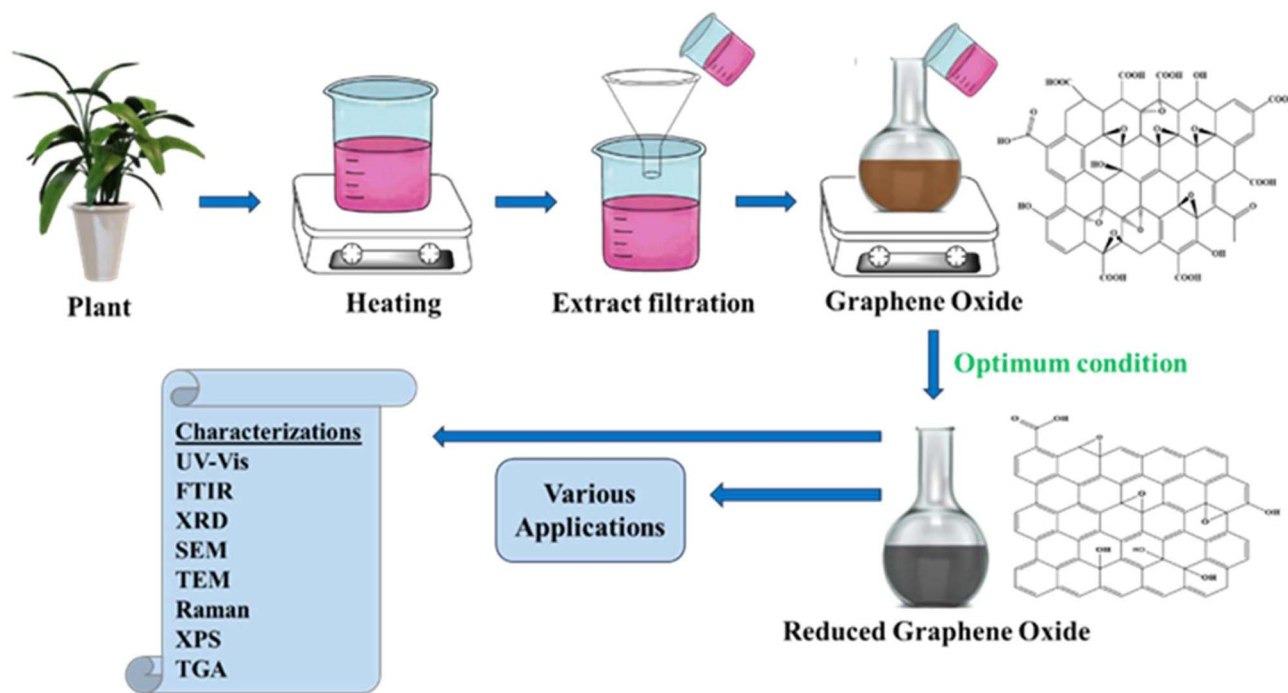


Fig. 3 Schematic diagram illustrating the typical process for the green reduction of GO using plant extracts, followed by characterization and application.

classification as a “phytochemical.” Secondly, reaction conditions play a crucial role; methods that employ heating or reflux consistently outperform those conducted at room temperature, indicating that thermal energy is necessary to overcome the activation barrier for deoxygenation. For instance, rGO produced from banana peel extract under reflux yielded a C/O ratio of 3.80, a value significantly better than those reported in many room-temperature syntheses.<sup>50</sup> However, the complexity of plant extracts, which contain dozens of compounds, leads to a lack of selectivity in functionalization and can result in batch-to-batch variability, posing a challenge for scalable and reproducible manufacturing.

**3.1.2 Proposed reduction mechanisms.** The most widely accepted mechanism for GO reduction by polyphenol-rich plant extracts involves a nucleophilic attack. As shown in Fig. 4, the hydroxyl groups of polyphenols (like luteolin) are deprotonated, and the resulting oxygen anion acts as a nucleophile, attacking the electrophilic carbon of an epoxide group on the GO basal plane. It leads to a ring-opening reaction. Subsequent dehydration restores the  $sp^2$  C=C bond, forming rGO, while the polyphenol is oxidized to a quinone-type structure.<sup>44</sup> However, other mechanisms are also proposed. For artemisinin, a free-radical-driven mechanism is proposed, in which heating the endoperoxide bridge generates hydroxyl radicals that aggressively attack and remove all types of oxygen functional groups.<sup>47</sup> This radical pathway may explain the superior deoxygenation (C/O ratio of 11.7) compared to the more selective nucleophilic pathway of polyphenols. In another case, the amine groups in histamine and serotonin from nettle extract were proposed to reduce GO *via* a mechanism similar to hydrazine, involving a nucleophilic attack followed by elimination (Fig. 5).<sup>42</sup> The diversity of these mechanisms highlights that the “plant

extract” category is not monolithic; the specific chemistry of the dominant phytochemical dictates the reduction pathway and, ultimately, the quality of the final rGO.

**3.1.3 Application-specific properties and outlook.** The properties of plant-extract-synthesized rGO make it suitable for a range of applications, particularly in environmental remediation and energy storage.

**3.1.3.1 Environmental remediation.** The reduction process with plant extracts is often incomplete, leaving residual oxygen functional groups on the rGO surface. While detrimental for conductivity, these groups act as excellent binding sites for adsorbing heavy metal ions and organic dyes. For instance, rGO produced with *Citrus hystrix* peel extract showed an impressive dye-removal efficacy with a maximum adsorption capacity ( $q_{\max}$ ) of  $276.06 \text{ mg g}^{-1}$  at room temperature,<sup>83</sup> while tulsi green tea extract-derived rGO could remove MG with a maximum adsorption capacity of  $416.7 \text{ mg g}^{-1}$ .<sup>101</sup> On the other hand, Mahmoud, *et al.*<sup>84</sup> utilized *Tecoma stans* extracts to synthesize rGO for removing Ni(II) with a maximum uptake capacity ( $q_{\max}$ ) of  $69 \text{ mg g}^{-1}$ . The combination of a restored  $\pi$ -system (for  $\pi$ - $\pi$  stacking interactions with aromatic dyes) and residual polar groups (for electrostatic and hydrogen bonding interactions) makes this type of rGO a highly effective adsorbent.

**3.1.3.2 Energy storage.** For supercapacitor applications, a high specific surface area is paramount for enabling ion adsorption at the electrode-electrolyte interface. The use of certain plant extracts, such as lemon juice, yields rGO with a moderate degree of reduction but a high specific surface area ( $159 \text{ m}^2 \text{ g}^{-1}$ ). Such property is particularly advantageous for supercapacitors, as demonstrated by the specific capacitance of  $124 \text{ F g}^{-1}$  reported by Joshi, *et al.*<sup>17</sup> Moreover, an excellent





Table 1 Summary of plant extracts utilized in the reduction of GO, with key characterization data<sup>a</sup>

| Plants/plant extracts                       | Part used      | Reduction conditions                              | Characteristics  | Application  | Ref. |
|---|----------------|---|--|--|------|
| <i>Camellia oleifera</i>                    | Shell          | Water bath, 80 °C for 3 h                         | 2θ: 24.9°, $I_D/I_G$ (rGO): 1.01 > $I_D/I_G$ (GO): 0.90                          | Adsorption of copper(II)                             | 51   |
| Carrot                                      | Root           | Stir, 48 h at 150 rpm and reflux, 100 °C for 24 h | 2θ: 24.2°, C/O ratio: 3.91, $I_D/I_G$ (rGO): 0.94 > $I_D/I_G$ (GO): 0.83         | Supercapacitor                                       | 52   |
| Lemon juice                                 | Fruit          | Stir, 24 h at 150 rpm and reflux, 100 °C for 8 h  | 2θ: 25.1°, C/O ratio: 4.91, $I_D/I_G$ (rGO): 0.96 > $I_D/I_G$ (GO): 0.83         | Supercapacitor                                       | 52   |
| <i>Persea americana</i>                     | Seed           | Stir, 100 °C for 10 h                             | UV-vis peak: 280 nm, $I_D/I_G$ (rGO): 0.89 > $I_D/I_G$ (GO): 0.76                | Antibacterial activity                               | 53   |
| <i>Plectranthus amboinicus</i>              | Leaves         | Autoclave, 100 °C for 12 h                        | 2θ: 25°, $I_D/I_G$ (rGO): 1.297 > $I_D/I_G$ (GO): 1.07                           | Supercapacitor                                       | 20   |
| <i>Cinnamomum zeylanicum</i>                | Bark           | Reflux, 45 min                                    | UV-vis peak: 280 nm, 2θ: 23°   | Dye elimination and antioxidant activity             | 54   |
| <i>Tithonia diversifolia</i>                | Flower         | Stir, 80 °C for 12 h                              | UV-vis peak: 265 nm, 2θ: 24°–26°   | Cytotoxicity   | 55   |
| <i>Lantana camara</i>                       | Leaves         | Reflux, 24 h                                      | UV-vis peak: 273 nm, 2θ: 21.9°, $I_D/I_G$ (rGO): 0.37 < $I_D/I_G$ (GO): 0.98     | Antibacterial, antioxidant and cytotoxicity activity | 25   |
| <i>Phyllanthus emblica</i>                  | Fruit          | Reflux, 95 °C for 3 h                             | UV-vis peak: 270 nm, 2θ: 23.11°, $I_D/I_G$ (rGO): 1.11 < $I_D/I_G$ (GO): 1.29    | Photovoltaic activity                                | 56   |
| Banana                                      | Peel           | Reflux, 90 °C for 48 h                            | C/O ratio: 81.0 : 19.0, $I_{2D}/I_G$ (rGO): 0.84117 > $I_{2D}/I_G$ (GO): 0.00005 | n.d.   | 45   |
| Banana                                      | Fruit          | Reflux, 90 °C for 48 h                            | C/O ratio: 78.1 : 21.9, $I_{2D}/I_G$ (rGO): 0.42773 > $I_{2D}/I_G$ (GO): 0.00005 | n.d.   | 45   |
| <i>Caesalpinia sappan</i> L.                | Flower         | Autoclave, 100 °C for 6 h                         | UV-vis peak: 259 nm, 2θ: 25.67°, $I_D/I_G$ (rGO): 1.29 < $I_D/I_G$ (GO): 1.60    | n.d.   | 57   |
| Artemisinin                                 | Leaves         | Water bath, 95 °C for 24 h                        | C/O ratio: 11.7, $I_D/I_G$ (rGO): 1.32 > $I_D/I_G$ (GO): 0.90                    | n.d.   | 47   |
| <i>Chenopodium album</i>                    | Vegetable      | Reflux, 100 °C for 12 h                           | UV-vis peak: 263 nm, 2θ: 22.50°  | Antimicrobial and anticancer activity                | 58   |
| Eucalyptus                                  | Leaves         | Stir, 80 °C for 8 h                               | UV-vis peak: 273.5 nm  | Dye removal  | 59   |
| Lemon juice                                 | Fruit          | Reflux, RT for 45 min                             | UV-vis peak: 259 nm, 2θ: 30°   | Antimicrobial potency                                | 60   |
| <i>Clinacanthus nutans</i>                  | Leaves         | Stir and reflux, (60–100) °C for 1–6 h            | UV-vis peak: 270 nm, 2θ: 22.12°, $I_D/I_G$ (rGO): 1.08 > $I_D/I_G$ (GO): 1.01    | n.d.   | 61   |
| <i>Acorus calamus</i>                       | Rhizome        | Sonication, 1–2 h                                 | UV-vis peak: 278 nm, 2θ: 26.4°, dense, compact structure                         | Antibacterial efficacy                               | 62   |
| <i>Terminalia bellirica</i>                 | Fruit and seed | Sonication, 1–2 h                                 | UV-vis peak: 262 nm, 2θ: 26.4°, layered structure                                | Antibacterial efficacy                               | 62   |
| <i>Helicteres isora</i>                     | Fruit and seed | Sonication, 1–2 h                                 | UV-vis peak: 268 nm, 2θ: 26.4°, layered structure                                | Antibacterial efficacy                               | 62   |
| <i>Quercus infectoria</i>                   | Fruit and seed | Sonication, 1–2 h                                 | UV-vis peak: 263 nm, 2θ: 26.4°, staked, crumpled and flaky                       | Antibacterial efficacy                               | 62   |
| <i>Turbinella pyrum</i>                     | Shell          | Sonication, 1–2 h                                 | UV-vis peak: 264 nm, 2θ: 26.4°, layered structure                                | Antibacterial efficacy                               | 62   |
| <i>Vitis vinifera</i>                       | Fruit          | Reflux, 95 °C for 1–6 h                           | UV-vis peak: 270 nm, 2θ: 23.7°   | Removal of dye                                       | 63   |
| <i>Murraya koenigii</i>                     | Leaves         | Autoclave, 100 °C for 12 h                        | UV-vis peak: 270 nm, 2θ: 26.25°, $I_D/I_G$ (rGO): 1.14 > $I_D/I_G$ (GO): 1.02    | Photocatalysis                                       | 64   |
| <i>Catharanthus roseus</i>                  | Roots          | Stir, 24 h  | $I_D/I_G$ (rGO): 1.20 > $I_D/I_G$ (GO): 0.93                                     | n.d.   | 65   |
| <i>Phyllanthus madagascariense</i> K. Schum | Leaves         | Stir, 24 h  | $I_D/I_G$ (rGO): 1.17 > $I_D/I_G$ (GO): 1.01                                     | n.d.   | 65   |
| <i>Cinnamomum camphora</i> cineoliferum     | Leaves         | Stir, 24 h  | $I_D/I_G$ (rGO): 1.21 > $I_D/I_G$ (GO): 1.01                                     | n.d.   | 65   |
| <i>Cedrelopsis grevei</i> Baill             | Barks          | Stir, 24 h  | $I_D/I_G$ (rGO): 1.26 > $I_D/I_G$ (GO): 0.93                                     | n.d.   | 65   |
| Lemon juice                                 | Fruit          | Stir, 80 °C for 2 h                               | UV-vis peak: 272 nm, 2θ: 24.26°, $I_D/I_G$ (rGO): 1.05 > $I_D/I_G$ (GO): 1.04    | Adsorption of methylene blue                         | 66   |



Table 1 (Contd.)

| Plants/plant extracts         | Part used  | Reduction conditions            | Characteristics   | Application                              | Ref. |
|-------------------------------|------------|---------------------------------|---|--|------|
| Mango                         | Leaves     | Stir and reflux, 90 °C for 24 h | UV-vis peak: 272 nm, $2\theta = 25.53^\circ$ , C/O ratio: 3.75  | n.d.                                     | 50   |
| Potato                        | Vegetable  | Stir and reflux, 90 °C for 24 h | UV-vis peak: 277 nm, $2\theta = 21.34^\circ$ , C/O ratio: 3.77  | n.d.                                     | 50   |
| Banana                        | Peel       | Stir and reflux, 90 °C for 24 h | UV-vis peak: 280 nm, $2\theta = 22.89^\circ$ , C/O ratio: 3.80  | n.d.                                     | 50   |
| Rose water                    | Flower     | Stir, RT for (70–100) °C        | $2\theta = 24^\circ$ , C/O ratio: 2.97  | n.d.                                     | 67   |
| Wild carrot                   | Roots      | Stir, RT for 48 h               | $2\theta = 23.96^\circ$ , C/O ratio: 11.9, $I_D/I_G$ (rGO): $1.06 > I_D/I_G$ (GO): 0.80               | n.d.                                     | 48   |
| Palm                          | Leaves     | Reflux, 100 °C for 3 h          | $2\theta = 24.5^\circ$  | n.d.                                     | 68   |
| <i>Hibiscus sabdariffa</i> L. | Flower     | Stir, RT for 1 h                | UV-vis peak: 262.8 nm, $2\theta = 25.0^\circ$ , $I_D/I_G$ (rGO): $1.24 > I_D/I_G$ (GO): 1.01          | Supercapacitor                           | 69   |
| <i>Ficus carica</i>           | Leaves     | Reflux, 98 °C for 1–30 h        | UV-vis peak: 270 nm, $2\theta = 24.50^\circ$ and $43^\circ$   | n.d.                                     | 70   |
| <i>Phragmites australis</i>   | Leaves     | Reflux, 98 °C for 1–30 h        | UV-vis peak: 267 nm, $2\theta = 24.50^\circ$ and $43^\circ$   | n.d.                                     | 70   |
| Sweet potato                  | Vegetable  | Reflux, 80 °C for 3 h           | UV-vis peak: 269 nm, $I_D/I_G$ (rGO): $0.97 > I_D/I_G$ (GO): 0.94                                     | n.d.                                     | 71   |
| <i>Bougainvillea glabra</i>   | Flower     | Stir, 95 °C for 5 h             | UV-vis peak: 270 nm, C/O ratio: Sensing 4.6   |  | 72   |
| <i>Citrus grandis</i>         | Fruit      | Reflux, 95 °C for 12 h          | UV-vis peak: 270 nm, $2\theta = 24.5^\circ$ , $I_D/I_G$ (rGO): $1.14 > I_D/I_G$ (GO): 0.86            | Supercapacitor                           | 73   |
| <i>Tamarindus indica</i>      | Fruit      | Reflux, 95 °C for 12 h          | UV-vis peak: 275 nm, $2\theta = 24.9^\circ$ , $I_D/I_G$ (rGO): $1.16 > I_D/I_G$ (GO): 0.86            | Supercapacitor                           | 73   |
| <i>Chrysanthemum</i>          | Flower     | Water bath, 95 °C for 24 h      | $2\theta = 24.6^\circ$ , C/O ratio: 4.96, $I_D/I_G$ (rGO): $1.14 > I_D/I_G$ (GO): 0.896               | n.d.                                     | 41   |
| <i>Lycium barbarum</i>        | Fruit      | Water bath, 95 °C for 24 h      | $2\theta = 26^\circ$ , C/O ratio: 4.96, $I_D/I_G$ (rGO): $1.05 > I_D/I_G$ (GO): 0.896                 | n.d.                                     | 74   |
| Tea                           | Leaves     | Stir, 80 °C for 1 h             | C/O ratio: 3.88, $I_D/I_G$ (rGO): $1.02 > I_D/I_G$ (GO): 1.005  | n.d.                                     | 75   |
| <i>Syzygium samarangense</i>  | Fruit      | Stir, 60 °C for 40 h            | $2\theta = 23.78^\circ$ , $I_D/I_G$ (rGO): $1.17 > I_D/I_G$ (GO): 0.92                                | n.d.                                     | 76   |
| Sugarcane bagasse             | Agro waste | Stir, 95 °C for 12 h            | UV-vis peak: 270 nm, C/O ratio: Removal of cadmium 4.27, $I_D/I_G$ (rGO): $1.16 > I_D/I_G$ (GO): 0.98 |  | 77   |
| <i>Larrea tridentata</i>      | Flower     | Reflux, 80 °C for 12 h          | UV-vis peak: 280 nm, $I_D/I_G$ (rGO): $0.983 < I_D/I_G$ (GO): 0.99                                    | Photocatalysis                           | 78   |
| <i>Capsicum chinense</i>      | Vegetable  | Reflux, 80 °C for 12 h          | UV-vis peak: 260 nm, $I_D/I_G$ (rGO): $0.987 < I_D/I_G$ (GO): 0.99                                    | Photocatalysis                           | 78   |
| <i>Ocimum sanctum</i> L.      | Leaves     | Stir, 70 °C for 4 h             | UV-vis peak: 267.8 nm, C/O ratio: 3.10  | n.d.                                     | 79   |
| <i>Acalypha indica</i>        | Leaves     | Autoclave, 100 °C for 12 h      | UV-vis peak: 272 nm, $I_D/I_G$ (rGO): $1.22 > I_D/I_G$ (GO): 1.02                                     | Cytotoxicity                             | 80   |
| <i>Raphanus sativus</i>       | Root       | Autoclave, 100 °C for 12 h      | UV-vis peak: 282 nm, $I_D/I_G$ (rGO): $1.15 > I_D/I_G$ (GO): 1.02                                     | Cytotoxicity                             | 80   |
| <i>Aloe vera</i>              | Leaves     | Stir, 95 °C for 24 h            | UV-vis peak: 259 nm   | Electrochemical analysis and dye removal | 81   |
| <i>Salvadora persica</i> L.   | Root       | Reflux, 98 °C for 24 h          | UV-vis peak: 280 nm, $2\theta = 22.4^\circ$   | n.d.                                     | 82   |
| <i>Citrus hystrix</i>         | Peel       | Stir, RT for 8 h                | UV-vis peak: 300 nm, $2\theta = 8.75^\circ$ and $26.34^\circ$   | Methylene blue adsorption                | 83   |
| <i>Tecoma stans</i>           | Leaves     | Stir, 70 °C for 12 h            | UV-vis peak: 280 nm   | Removal of Ni(II)                        | 84   |
| <i>Salvia spinosa</i>         | Leaves     | Reflux, 95 °C for 12 h          | UV-vis peak: 274 nm, $2\theta = 26.2^\circ$ , $I_D/I_G$ (rGO): $0.91 < I_D/I_G$ (GO): 0.95            | Evaluation of photothermal effect        | 85   |
| <i>Mangifera indica</i>       | Leaves     | Reflux, 70–80 °C for 12 h       | UV-vis peak: 259 nm, $2\theta = 21.87^\circ$ , $I_D/I_G$ (rGO): $1.024 > I_D/I_G$ (GO): 0.846         | Electrical conductivity analysis         | 29   |

Table 1 (Contd.)

| Plants/plant extracts        | Part used             | Reduction conditions                     | Characteristics   | Application                                | Ref. |
|------------------------------|-----------------------|--|---|--|------|
| <i>Solanum tuberosum</i> L.  | Vegetable             | Reflux, 70–80 °C for 12 h                | UV-vis peak: 265 nm, $2\theta = 21.86^\circ$ , $I_D/I_G$ (rGO): $1.066 > I_D/I_G$ (GO): 0.846 | Electrical conductivity analysis           | 29   |
| <i>Tinospora cordifolia</i>  | Stem                  | Reflux, 85 °C for 3 h                    | UV-vis peak: 263 nm, $2\theta = 22.81^\circ$  | Dye degradation and antibacterial activity | 86   |
| <i>Ocimum sanctum</i>        | Leaves                | Reflux, 100 °C for 10 h                  | $2\theta = 25^\circ$  | Cytotoxicity                               | 87   |
| Spinach                      | Leaves                | Reflux, RT for 30 min                    | UV-vis peak: 282 nm, $2\theta = 26^\circ$   | Antioxidant and dye adsorption             | 88   |
| <i>Citrus hystrix</i>        | Peel                  | Stir, RT for 8 h                         | C–C/C–O ratio: 1.07, $2\theta = 10.15^\circ$  | Methylene blue adsorption                  | 89   |
| <i>Punica granatum</i> L.    | Seed                  | Reflux, 98 °C for 8 h                    | UV-vis peak: 280 nm   | Antioxidant                                | 90   |
| <i>Prunus serrulata</i>      | Leaves                | Reflux, 95 °C for 12 h                   | UV-vis peak: 272 nm, C/O ratio: n.d.<br>$5.10$ , $2\theta = 26.2^\circ$                       |  | 91   |
| <i>Magnolia kobus</i>        | Leaves                | Reflux, 95 °C for 12 h                   | C/O ratio: 4.40   | n.d.                                       | 91   |
| <i>Platanus orientalis</i>   | Leaves                | Reflux, 95 °C for 12 h                   | C/O ratio: 4.96   | n.d.                                       | 91   |
| <i>Eclipta prostrata</i>     | Leaves                | Stir, RT for 4 h                         | C/O ratio: 2.70   | n.d.                                       | 92   |
| <i>Eichhornia crassipes</i>  | Whole except the root | Reflux                                   | UV-vis peak: 274 nm, $2\theta = 26^\circ$   | n.d.                                       | 93   |
| <i>Pulicaria glutinosa</i>   | Whole plant           | Stir, 98 °C for 24 h                     | UV-vis peak: 280 nm, $2\theta = 22.4^\circ$   | n.d.                                       | 94   |
| <i>Rhus coriaria</i>         | Fruit                 | Reflux, 95 °C for 12 h                   | UV-vis peak: 282 nm, $2\theta = 26.91^\circ$ , $I_D/I_G$ (rGO): $1.04 > I_D/I_G$ (GO): 0.84   | Cytotoxicity                               | 95   |
| Olive                        | Leaves                | Reflux, 100 °C for 10 h                  | UV-vis peak: 270 nm, $2\theta = 24.6^\circ$   | n.d.                                       | 46   |
| <i>Annona squamosa</i>       | Leaves                | Reflux, 100 °C for 12 h                  | UV-vis peak: 276 nm, $2\theta = 23^\circ$   | n.d.                                       | 96   |
| Green coffee bean            | Fruit                 | Stir, 80 °C for 12 h                     | UV-vis peak: 275 nm, $2\theta = 22^\circ$ , $I_D/I_G$ (rGO): $1.02 < I_D/I_G$ (GO): 1.04      | Dye removal                                | 97   |
| <i>Mangifera indica</i>      | Leaves                | Stir, 50 °C for 24 h                     | UV-vis peak: 266 nm, $I_D/I_G$ (rGO): $1.21 > I_D/I_G$ (GO): 1.10                             | Highly conductive film                     | 98   |
| <i>Ficus religiosa</i>       | Leaves                | Stir, 50 °C for 24 h                     | $I_D/I_G$ (rGO): $1.12 > I_D/I_G$ (GO): 1.10  | Highly conductive film                     | 98   |
| <i>Polyalthia longifolia</i> | Leaves                | Stir, 50 °C for 24 h                     | $I_D/I_G$ (rGO): $1.18 > I_D/I_G$ (GO): 1.10  | Highly conductive film                     | 98   |
| Ginger                       | Root                  | Stir, 90 °C for 24 h                     | $2\theta = 24.34^\circ$ , $I_D/I_G$ (rGO): $0.91 < I_D/I_G$ (GO): 1.14                        | Supercapacitor                             | 99   |
| <i>Urtica dioica</i>         | Leaves                | Ultra-sonication, 90 °C for 1 h          | UV-vis peak: 259 nm, $I_D/I_G$ (rGO): $1.13 > I_D/I_G$ (GO): 0.91                             | Antioxidant                                | 42   |
| <i>Colocasia esculenta</i>   | Leaves                | Reflux, 5 h                              | UV-vis peak: 270.9 nm   | n.d.                                       | 44   |
| <i>Mesua ferrea</i> Linn.    | Leaves                | Reflux, 8 h                              | UV-vis peak: 268 nm   | n.d.                                       | 44   |
| <i>Terminalia chebula</i>    | Seed                  | Reflux in a water bath, 90 °C for 24 h   | UV-vis peak: 275 nm, $2\theta = 26.6^\circ$ , $I_D/I_G$ (rGO): $I_D/I_G$ (GO)                 | n.d.                                       | 52   |
| Eucalyptus                   | Bark                  | Reflux, 80–85 °C for 24 h                | UV-vis peak: 270 nm, $2\theta = 25^\circ$ , $I_D/I_G$ (rGO): $1.15 > I_D/I_G$ (GO): 0.98      | Supercapacitor                             | 100  |
| Tulsi (holy basil) green tea | Leaves                | Microwave irradiation at 800 W for 1 min | UV-vis peak: 270 nm, $2\theta = 26.35^\circ$ , $I_D/I_G$ (rGO): $1.40 > I_D/I_G$ (GO): 1.08   | Supercapacitor and dye removal             | 101  |

<sup>a</sup> n.d. = not done.

specific capacitance of 239 F g<sup>−1</sup> was observed for the rGO synthesized using eucalyptus bark extract.<sup>100</sup> This highlights how the morphological properties imparted by certain green reductants can be tailored for specific energy storage needs.

### 3.2 Reduction of GO using microorganisms

Microorganisms, including bacteria, fungi, and yeast, offer a sustainable and biocompatible route for rGO synthesis. These

methods leverage the metabolic and enzymatic machinery of microbes to reduce GO under mild conditions, often at room temperature or slightly above (Fig. 6).

**3.2.1 General methodology and reduction efficacy.** In a typical microbial reduction, a GO dispersion is introduced into a microbial culture (e.g., bacteria in a nutrient broth) and incubated for a period ranging from hours to several days. The microbial cells or their secreted enzymes interact with the GO



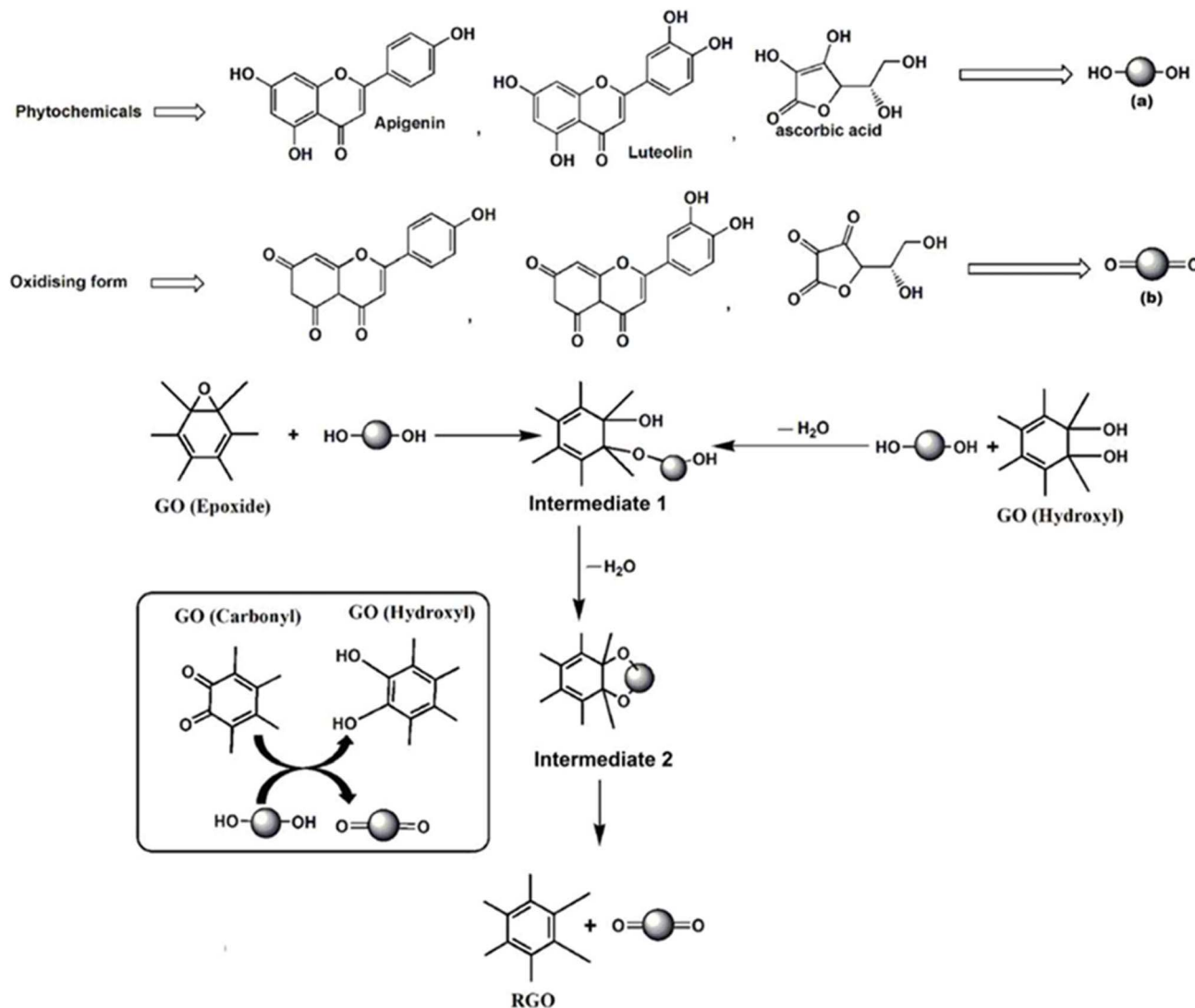


Fig. 4 Proposed reaction mechanism for the chemical reduction of GO by polyphenols (e.g., luteolin, apigenin), proceeding via a nucleophilic attack on epoxide and hydroxyl groups followed by dehydration. Reproduced from ref. 44 (Thakur and Karak) with permission from Elsevier, copyright 2012.

sheets, facilitating electron transfer and reduction. The bacterial species *Shewanella* has been extensively studied for this purpose, as it can reduce GO under both anaerobic and aerobic conditions.<sup>31,102,103</sup> The reduction efficacy is generally good, with C/O ratios reaching up to 5.9 for yeast-mediated reduction<sup>104</sup> and electrical conductivity of 55.32 S m<sup>-1</sup> for *Shewanella*-reduced GO.<sup>31</sup> The main drawback of microbial methods is the long reaction time (often 24–168 h) and the need for sterile culture conditions, which can complicate scalability. A summary of representative studies is provided in Table 2.

**3.2.2 Proposed reduction mechanisms.** Microbial reduction of GO can occur through several distinct pathways, primarily involving electron transfer from the cell's respiratory chain.

**3.2.2.1 Direct electron transfer.** Electrochemically active bacteria like *Shewanella* possess outer-membrane cytochromes (e.g., the Mtr pathway) that can directly transfer electrons to GO when it acts as a terminal electron acceptor, analogous to how they reduce metal oxides (Fig. 7, path 2).<sup>102,126</sup>

**3.2.2.2 Indirect electron transfer via mediators.** Some bacteria secrete redox-active molecules (electron shuttles) like flavins, which can accept electrons from the cell and subsequently shuttle them to external GO sheets (Fig. 7, path 1).<sup>31</sup>

**3.2.2.3 Enzymatic reduction.** Specific enzymes, such as nitrogenase from *Azotobacter chroococcum*, have been proposed to directly catalyze the reduction of GO through a series of proton and electron additions followed by dehydration.<sup>108</sup>

**3.2.2.4 Chemical oxidation of cellular components.** An alternative pathway suggests that the antibacterial properties of GO itself can induce oxidative stress and membrane damage in microbes. It leads to the leakage of intracellular reducing components (e.g., NADH, glutathione) that then chemically reduce the GO (Fig. 7, paths 4 and 5).<sup>106</sup>

**3.2.3 Application-specific properties and outlook.** The inherent biocompatibility of microbially synthesized rGO makes it exceptionally well-suited for biomedical applications.



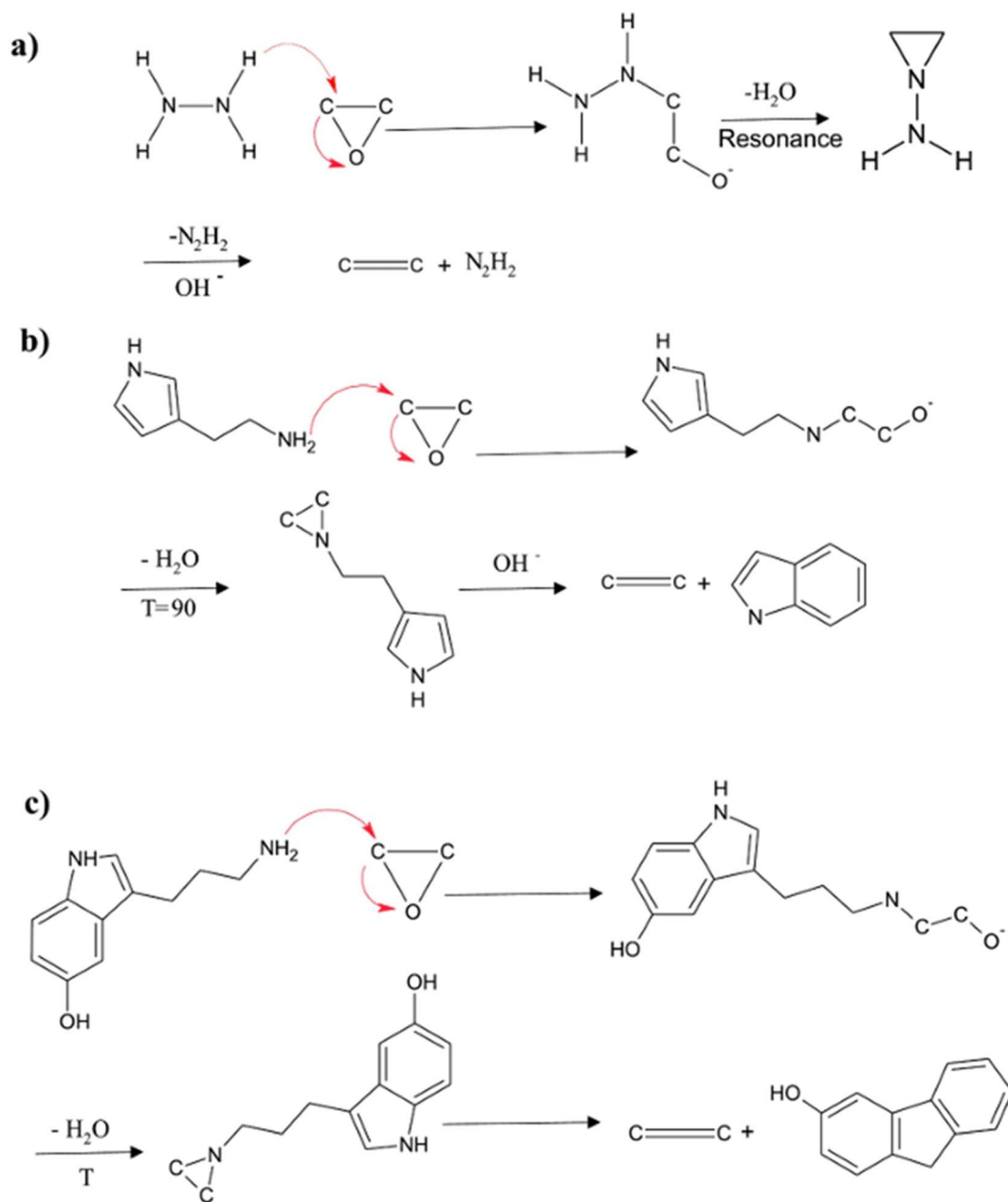


Fig. 5 Comparison of the proposed reduction mechanism for epoxide groups on GO by (a) hydrazine, (b) histamine, and (c) serotonin, all involving a nucleophilic attack by the amine group. Reproduced from ref. 42 (Mahmudzadeh, *et al.*) with permission from Elsevier, copyright 2019.

**3.2.3.1 Biomedical applications.** Because the reduction occurs in a biological environment and often involves biomolecules that remain on the rGO surface, the resulting material exhibits low cytotoxicity. Utkan, *et al.*<sup>105</sup> demonstrated that rGO produced by *Lactococcus lactis* and *Lactobacillus plantarum* exhibited minimal cytotoxicity against human cell lines, making it a safe candidate for biomedical use. Furthermore, the rGO itself often exhibits potent antibacterial activity. Akhavan and Ghaderi<sup>127</sup> showed that *E. coli* reduces GO to a bactericidal form of graphene, likely due to the sharp edges of the restored nanosheets, which induce membrane stress. This dual function—biocompatible synthesis leading to an antimicrobial product—is a unique advantage of this method.

**3.2.3.2 Anti-biocorrosion coatings.** rGO's impermeability and hydrophobicity make it an excellent barrier against corrosive agents. Song, *et al.*<sup>109</sup> demonstrated that *D. desulfuricans*, a bacterium responsible for biocorrosion, could be used to reduce a GO coating on a copper surface. The resulting *in situ*-formed rGO layer then acted as a protective barrier, inhibiting further corrosion by the same bacteria.

### 3.3 Reduction of GO using biomolecules

Using well-defined, pure biomolecules as reducing agents offers a “best of both worlds” approach, combining the eco-friendliness of natural compounds with the precision and



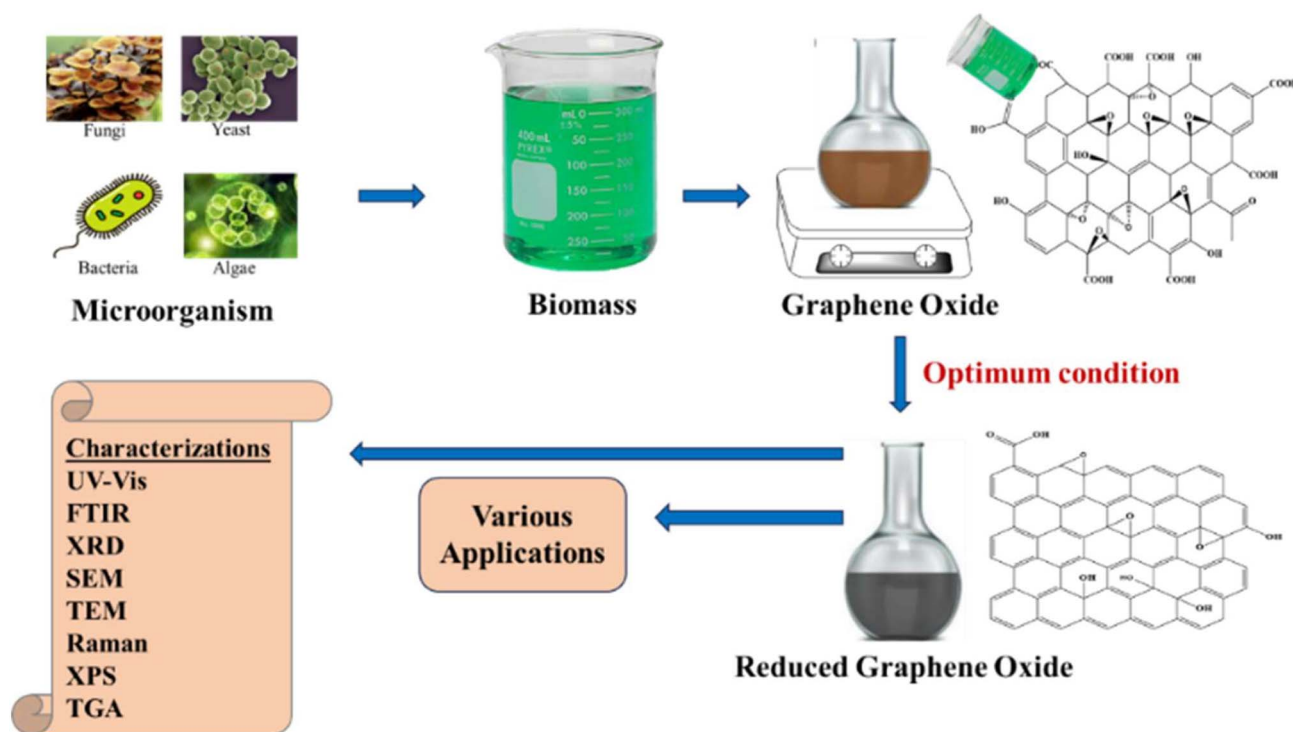


Fig. 6 Schematic illustration of the general process for microbial reduction of GO.

control of synthetic chemistry. This strategy allows for better reproducibility and the ability to tune the properties of the final rGO product.

**3.3.1 General methodology and reduction efficacy.** This method is experimentally straightforward, involving the addition of a pure biomolecule solution (*e.g.*, ascorbic acid, L-cysteine, glucose) to a GO dispersion, typically with mild heating. Ascorbic acid (vitamin C) is one of the most effective and widely studied green reductants. A seminal study by Fernández-Merino, *et al.*<sup>128</sup> showed that ascorbic acid could achieve a degree of reduction comparable to hydrazine, yielding an excellent O/C ratio of 0.08 (equivalent to a C/O ratio of 12.5). The results demonstrated that green reductants could match the performance of hazardous chemicals. Other biomolecules like gallic acid (C/O ratio: 3.89)<sup>30</sup> and caffeic acid (C/O ratio: 7.15)<sup>40</sup> are also highly effective. The use of pure compounds provides excellent control over the reaction, but the cost of purified biomolecules can be a limiting factor for large-scale production compared to crude plant extracts. A summary of various biomolecules used as reductants is presented in Table 3.

**3.3.2 Proposed reduction mechanisms.** The reduction mechanisms for biomolecules are specific to their chemical structures. For ascorbic acid, the reduction is believed to proceed *via* a nucleophilic attack on the epoxide groups, similar to that of polyphenols, followed by dehydration.<sup>136,147</sup> For amino acids containing thiol groups, such as L-cysteine, the thiol group (–SH) is deprotonated and attacks the oxygen functional groups. In this process, the thiol is oxidized to form a disulfide (–S–S–) bond, while GO is reduced to rGO.<sup>148</sup> For biomolecules with conjugated systems like  $\beta$ -carotene, the proposed mechanism

involves the formation of an epoxide on  $\beta$ -carotene itself, which is then hydrolyzed to a diol. The resulting oxygen anions act as nucleophiles, deoxygenating GO (Fig. 8).<sup>131</sup> The ability to propose such specific chemical pathways is a significant advantage of using pure biomolecules, as it allows for a more rational design of the reduction process.

**3.3.3 Application-specific properties and outlook.** The high degree of reduction and precise surface chemistry achievable with biomolecules make this rGO ideal for electronics and sensing applications.

**3.3.3.1 Sensors and electronics.** The excellent C/O ratios and high conductivity achieved with reductants like ascorbic acid and humanin ( $105 \text{ S m}^{-1}$ )<sup>146</sup> make this rGO a prime candidate for conductive films and sensors. Bo, *et al.*<sup>40</sup> used caffeic acid-reduced rGO to fabricate a gas sensor that exhibited rapid, sensitive responses to  $\text{NO}_2$  and  $\text{NH}_3$ . The gas-sensing mechanism relies on charge transfer between gas molecules and the rGO surface, a process that requires a highly conductive material with a low defect density. The ability of biomolecules to produce high-quality, electronically active rGO is a key advantage for these applications.

**3.3.3.2 Flexible films.** The self-polymerization of specific biomolecules, like dopamine, can be exploited to create robust materials. Luo, *et al.*<sup>33</sup> used dopamine to both reduce GO and serve as a polymeric binder. As the solvent evaporated, the polydopamine-coated rGO sheets self-assembled into a flexible, layer-by-layer film with a tensile strength of 25 MPa and excellent flame-retardant properties. The outcome demonstrates an intelligent synthesis strategy in which the reducing agent serves a dual purpose, leading to a functional macroscopic material.



Table 2 Summary of microorganism-mediated reduction of GO, with key characterization data<sup>a</sup>

| Species                                | Reduction conditions                                   | Characteristics  | Applications                              | Ref. |
|--|--|--|---|------|
| <b>Bacteria</b>                        |  |  |   |      |
| <i>Lactococcus lactis</i>              | Aerobic, 30 °C for 7 days                              | $2\theta = 26.5^\circ$ , C/O ratio: 3.70, $I_D/I_G$ (rGO): $1.35 < I_D/I_G$ (GO): 2.41     | Cytotoxicity                              | 105  |
| <i>Lactobacillus plantarum</i>         | Aerobic, 30 °C for 7 days                              | $2\theta = 26.5^\circ$ , C/O ratio: 2.98, $I_D/I_G$ (rGO): $1.46 < I_D/I_G$ (GO): 2.41     | Cytotoxicity                              | 105  |
| <i>Escherichia coli</i>                | Aerobic, 37 °C for 7 days                              | $2\theta = 26.5^\circ$ , C/O ratio: 2.80, $I_D/I_G$ (rGO): $1.26 < I_D/I_G$ (GO): 2.41     | Cytotoxicity                              | 105  |
| <i>Shewanella oneidensis</i>           | Aerobic and anaerobic, RT for 2 days                   | $I_D/I_G$ (rGO): $1.00 \pm 0.09 > I_D/I_G$ (GO): $0.85 \pm 0.03$                           | Creation of conductive graphene materials | 31   |
| <i>Shewanella oneidensis</i> MR-1      | Aerobic and anaerobic, RT for different time intervals | C/O ratio: increasing over time  | n.d.                                      | 103  |
| <i>Enterobacter cloacae</i>            | Aerobic, 20–25 °C for 3 days                           | UV-vis peak: 270 nm, $I_D/I_G$ (rGO): $1.17 > I_D/I_G$ (GO): 1.09                          | n.d.                                      | 106  |
| <i>Bacillus</i> sp.                    | Aerobic, 20–25 °C for 3 days                           | UV-vis peak: 270 nm, $I_D/I_G$ (rGO): $1.20 > I_D/I_G$ (GO): 1.09                          | n.d.                                      | 106  |
| <i>Shewanella baltica</i>              | Aerobic, 20–25 °C for 3 days                           | UV-vis peak: 270 nm, $I_D/I_G$ (rGO): $0.99 < I_D/I_G$ (GO): 1.09                          | n.d.                                      | 106  |
| <i>Shewanella oneidensis</i> MR-1      | Anaerobic, RT for 3 days                               | % C–C: 56%   | n.d.                                      | 102  |
| <i>Shewanella putrefaciens</i> CN32    | Anaerobic, RT for 3 days                               | % C–C: 91%   | n.d.                                      | 102  |
| <i>Shewanella amazonensis</i> SB2B     | Anaerobic, RT for 3 days                               | % C–C: 75%   | n.d.                                      | 102  |
| <i>Shewanella putrefaciens</i> W3-18-1 | Anaerobic, RT for 3 days                               | % C–C > 95%  | n.d.                                      | 102  |
| <i>Shewanella baltica</i> 10735        | Anaerobic, RT for 3 days                               | % C–C: 54%   | n.d.                                      | 102  |
| <i>Escherichia coli</i>                | 37 °C for 3 days                                       | UV-vis peak: 267 nm, $2\theta = 24^\circ$  | n.d.                                      | 22   |
| <i>Bacillus sphaericus</i>             | 30 °C for 2 days                                       | UV-vis peak: 261 nm, C/O ratio: 2.62, $I_D/I_G$ (rGO): $1.17 > I_D/I_G$ (GO): 0.99         | n.d.                                      | 107  |
| <i>Azotobacter chroococcum</i>         | RT for 72 h  | C/O ratio: 4.18, $2\theta = 17\text{--}24^\circ$   | n.d.                                      | 108  |
| <i>Desulfovibrio desulfuricans</i>     | 25 °C for 24 h   | $2\theta = 17\text{--}24^\circ$ , $I_D/I_G$ (rGO): $1.13 > I_D/I_G$ (GO): 0.92             | Anti-biocorrosion                         | 109  |
| <i>Escherichia coli</i>                | Aerobic, 37 °C for 0.5 h                               | $2\theta = 26.6^\circ$ , C/O ratio: 5.78, $I_D/I_G$ (rGO): $0.72 < I_D/I_G$ (GO): 0.95     | Superoxide formation                      | 110  |
| <i>Shewanella</i> sp. CF8-6            | Facultative anaerobic, 25 °C for 12 h                  | $2\theta = 23.1^\circ$ , $I_D/I_G$ (rGO): $1.26 > I_D/I_G$ (GO): 1.11                      | Dye adsorption                            | 39   |
| <i>Shigella dysenteriae</i>            | 37 °C for 10 h   | $2\theta = 20\text{--}23^\circ$ , $I_D/I_G$ (rGO): $1.15 > I_D/I_G$ (GO): 0.84             | n.d.                                      | 111  |
| <i>G. sulfurreducens</i>               | 30 °C for 9 days                                       | O/C ratio: 0.49, $I_D/I_G$ (rGO): $1.324 > I_D/I_G$ (GO): 0.945                            | n.d.                                      | 112  |
| <i>Bacillus subtilis</i> 168           | $25 \pm 2$ °C for time intervals                       | $2\theta = 24.18^\circ$ , $I_D/I_G$ (rGO): $1.01 > I_D/I_G$ (GO): 0.92                     | n.d.                                      | 113  |
| <i>Shewanella decolorationis</i> NTOU1 | 35 °C for 24 h   | C/O ratio: 3.0   | n.d.                                      | 114  |
| <i>Escherichia coli</i> strain E-NO.7  | 37 °C for 72 h   | $2\theta = 26.5^\circ$   | n.d.                                      | 115  |
| <i>Lactobacillus plantarum</i>         | 30 °C for 7 days                                       | C/O ratio: 3.3, $I_D/I_G$ (rGO): $0.92 < I_D/I_G$ (GO): 0.94                               | n.d.                                      | 116  |
| <i>Lactococcus lactis</i>              | 30 °C for 3–4 days                                     | $2\theta = 23\text{--}26^\circ$ , $I_D/I_G$ (rGO): $0.97 < I_D/I_G$ (GO): 2.15             | n.d.                                      | 117  |
| <i>Bacillus clausii</i>                | 37 °C for 72 h   | UV-vis peak: 268 nm, $2\theta = 24.5^\circ$  | Against MDR uropathogenic isolates        | 118  |
| <i>Pseudoalteromonas</i> sp. CF10-13   | Facultative anaerobic, 25 °C for 12 h                  | $2\theta = 21.9^\circ$ , $I_D/I_G$ (rGO): $1.3 > I_D/I_G$ (GO): 1.03                       | n.d.                                      | 119  |
| <i>Gluconobacter roseus</i>            | 37 °C for 24 h   | UV-vis peak: 280 nm, $I_D/I_G$ (rGO): $0.87 < I_D/I_G$ (GO): 1.12                          | Electrochemical study                     | 120  |
| <i>Escherichia fergusonii</i>          | 37 °C for 72 h   | UV-vis peak: 267 nm, $2\theta = 25.6^\circ$ , $I_D/I_G$ (rGO): $1.96 > I_D/I_G$ (GO): 1.58 | n.d.                                      | 121  |
| <b>Fungi</b>                           |  |  |   |      |
| <i>Rhizopus oryzae</i>                 | Shaking, 37 °C for 24 h                                | $2\theta = 26.07^\circ$ , $I_D/I_G$ (rGO): $1.17 > I_D/I_G$ (GO): 0.96                     | Antimicrobial coating for medical devices | 23   |
| <i>Ganoderma</i> spp.                  | Ultrasonicated, 40 °C for 24 h                         | UV-vis peak: 265 nm, $2\theta = 26.5^\circ$ , $I_D/I_G$ (rGO): $2.1 > I_D/I_G$ (GO): 1.8   | Cytotoxicity                              | 122  |
| <i>Aspergillus</i> sp.                 | Static, 40 °C for 72 h                                 | UV-vis peak: 270 nm, $I_D/I_G$ (rGO): $1.06 > I_D/I_G$ (GO): 1.01                          | Antibacterial study                       | 123  |
| <i>Ganoderma lucidum</i>               | Water bath, 85 °C for 16 h                             | UV-vis peak: 260 nm, $2\theta = 24.0^\circ$ , $I_D/I_G$ (rGO): $0.99 > I_D/I_G$ (GO): 0.94 | n.d.                                      | 124  |



Table 2 (Contd.)

| Species                  | Reduction conditions       | Characteristics   | Applications            | Ref. |
|--------------------------|----------------------------|---|-------------------------|------|
| <b>Algae</b>             |                            |   |                         |      |
| <i>Chlorella</i> sp.     | Water bath, 90 °C for 96 h | UV-vis peak: 267 nm, $I_D/I_G$ (rGO): 0.935 > $I_D/I_G$ (GO): 0.853                                       | Biophotovoltaic devices | 24   |
| <i>Turbinaria ornata</i> | Water bath, 60 °C for NA   | UV-vis peak: 267 nm, $2\theta = 26.4^\circ$   | Cytotoxicity            | 125  |
| <b>Yeast</b>             |                            |   |                         |      |
| Baker's yeast            | Stir, 35–40 °C for 72 h    | UV-vis peak: 264 nm, $2\theta = 23.5^\circ$ , C/O ratio: 5.9, $I_D/I_G$ (rGO): 1.44 > $I_D/I_G$ (GO): 0.8 | n.d.                    | 104  |

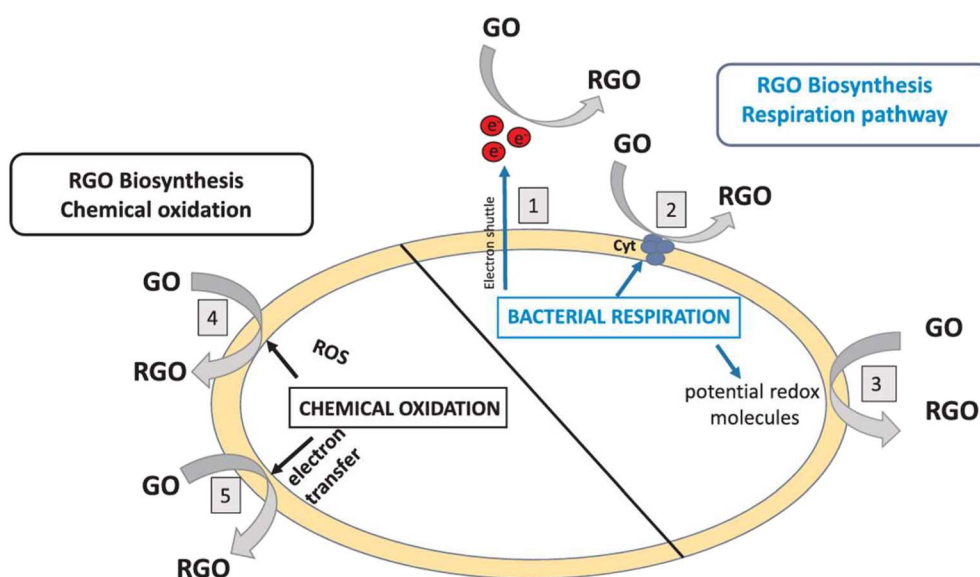
<sup>a</sup> n.d. = not done.

Fig. 7 Proposed bacterial reduction strategies for GO, mediated by either bacterial respiration (paths 1–3) or chemical oxidation via lysis and release of intracellular components (paths 4 and 5). Reproduced from ref. 106 (Vargas, et al.) with permission from Elsevier, copyright 2019.

## 4 Comparative analysis, challenges, and future outlook

While green reduction methods offer a sustainable alternative to conventional techniques, choosing a method involves trade-offs among cost, scalability, reaction efficiency, and the final properties of rGO. A quantitative comparison of these strategies is essential for guiding future research and application-driven synthesis (Table 4).

### 4.1 Current challenges and specific research gaps

Despite the significant progress, several challenges must be addressed to advance the field:

**4.1.1 Standardization of natural extracts.** Plant extracts are complex mixtures whose composition can vary with season, geography, and extraction method. A key research gap is the need for analytical techniques to identify and quantify the primary active reducing agents in these extracts, ensuring reproducibility and process control.

**4.1.2 Reaction kinetics and mechanism.** The kinetics of most green reduction processes are poorly understood. Detailed kinetic studies are needed to optimize reaction times and temperatures. Furthermore, a deeper mechanistic understanding is required to explain why certain reductants (e.g., artemisinin) are far more effective than others.

**4.1.3 Scalability and integration.** While many methods work well at the lab scale, scaling them for industrial production remains a significant hurdle, especially for microbial processes. Research is needed on continuous-flow reactors and process optimization to bridge the gap between laboratory synthesis and industrial application.

### 4.2 Future outlook

The future of rGO synthesis lies in the rational design of materials for specific applications. Key future directions include:

**4.2.1 Application-driven synthesis.** Instead of a one-size-fits-all approach, synthesis methods will be tailored to the end-use. For electronics, the focus will be on achieving the



Table 3 Summary of biomolecule-mediated reduction of GO, with key characterization data<sup>a</sup>

| Biomolecules                                    | Reduction conditions  | Characteristics   | Applications                                | Ref. |
|---|---|---|---|------|
| Glucose   | Ultrasonic bath, RT for 60 min                                  | UV-vis peak: 270 nm, $2\theta$ : 20.78°, C/O ratio: 2.2   | n.d.  | 129  |
| Ascorbic acid and sodium citrate binary mixture | Ultrasonic bath, RT for 24 h                                    | UV-vis peak: 260 nm, $2\theta$ : 23.54°   | n.d.  | 130  |
| $\beta$ -Carotene                               | Reflux, 95 °C for 24 h  | UV-vis peak: 270 nm, $2\theta$ : 23.13°, $I_D/I_G$ (rGO): 1.01 > $I_D/I_G$ (GO): 0.86               | Supercapacitor                              | 131  |
| Gallic acid                                     | Stir, RT for 24 h   | UV-vis peak: 270 nm, $2\theta$ : 26°, C/O ratio: 3.89, $I_D/I_G$ (rGO): 1.92 > $I_D/I_G$ (GO): 1.74 | n.d.  | 30   |
| Tannic acid                                     | Sonication, 80 °C for 10 h                                      | UV-vis peak: 274 nm, C/O ratio: 1.21, $I_D/I_G$ (rGO): 1.18 > $I_D/I_G$ (GO): 0.97                  | n.d.  | 132  |
| Honeycomb flavone chrysin                       | Stir, 90 °C for 1 h   | $2\theta$ : 24.6°, $I_D/I_G$ (rGO): 1.755 > $I_D/I_G$ (GO): 1.518                                   | Improved bactericidal and skin regeneration | 133  |
| Starch  | Reflux, 80 °C for 3 h   | UV-vis peak: 269 nm, $2\theta$ : 21.3°, $I_D/I_G$ (rGO): 0.97 > $I_D/I_G$ (GO): 0.94                | n.d.  | 71   |
| Ascorbic acid                                   | Stir, 95 °C for 1 h   | $2\theta$ : 23.8°   | n.d.  | 134  |
| Ascorbic acid                                   | Stir, 60 °C for 12 h  | UV-vis peak: 308 nm, $2\theta$ : 24.10°   | n.d.  | 135  |
| Ascorbic acid                                   | Spray, 50 °C for 48 h   | $2\theta$ : 25.39°  | n.d.  | 136  |
| Ascorbic acid                                   | 95 °C for 30 min  | UV-vis peak: 266 nm, O/C ratio: 0.08  | n.d.  | 128  |
| Dopamine  | Stir, 60 °C for 2 h   | $2\theta$ : 21.88°, $I_D/I_G$ (rGO): 1.06 > $I_D/I_G$ (GO): 0.87                                    | Flexible film                               | 33   |
| Uric acid                                       | Incubate, 40 °C for 1 h and stir, 90 °C for 1 h                 | UV-vis peak: 260 nm, $2\theta$ : 25.9°, $I_D/I_G$ (rGO): 2.02 > $I_D/I_G$ (GO): 1.5                 | Anticancer agent                            | 137  |
| Citric acid                                     | Ultrasonic bath, 92 °C for 1.5 h                                | UV-vis peak: 268 nm, $I_D/I_G$ (rGO): 1.29 > $I_D/I_G$ (GO): 1.09                                   | Adsorption of dye                           | 138  |
| Ethanol   | Reflux, 150 °C for 8 h  | $2\theta$ : 24.40°, C/O ratio: 2.72   | Superconductor                              | 139  |
| Caffeic acid                                    | Stir, 95 °C for 24 h  | $2\theta$ : 24.89°, C/O ratio: 7.15, $I_D/I_G$ (rGO): 1.15 > $I_D/I_G$ (GO): 0.86                   | Sensing and energy storage                  | 40   |
| Alanine   | Stir, 85 °C for 24 h  | UV-vis peak: 258 nm, $I_D/I_G$ (rGO): 0.996 > $I_D/I_G$ (GO): 0.943                                 | n.d.  | 140  |
| Extracellular polymeric substances              | Stir, 40 °C for 24 h  | C/O ratio: 3.18, $I_D/I_G$ (rGO): 1.0183 < $I_D/I_G$ (GO): 1.0375                                   | n.d.  | 141  |
| Enhanced green fluorescent protein              | Ultrasonication, 40 °C for 15 min and water bath, 90 °C for 1 h | UV-vis peak: 258 nm, $2\theta$ : 25.8°, $I_D/I_G$ (rGO): 2.149 > $I_D/I_G$ (GO)                     | n.d.  | 142  |
| Nicotinamide                                    | Stir, 40 °C for 6 h   | UV-vis peak: 260 nm, $2\theta$ : 26.2°, $I_D/I_G$ (rGO): 1.74 > $I_D/I_G$ (GO): 1.01                | Cytotoxicity                                | 143  |
| L-Glutathione                                   | Ultrasonication for 1 h and 50 °C for 6 h                       | $2\theta$ : 24.7°   | n.d.  | 144  |
| Lignin  | Autoclave, 180 °C for 12 h                                      | UV-vis peak: 270 nm, O/C ratio: 0.286   | Electrochemical property                    | 35   |
| Melatonin                                       | Ultrasonication, 80 °C for 3 h                                  | UV-vis peak: 269 nm, $I_D/I_G$ (rGO): 1.07 < $I_D/I_G$ (GO): 1.21                                   | Antioxidant                                 | 145  |
| Humanin   | Ultrasonication for 15 min and 40 °C for 1 h                    | UV-vis peak: 265 nm, $2\theta$ : 26.4°, $I_D/I_G$ (rGO): 2.3 > $I_D/I_G$ (GO): 1.4                  | n.d.  | 146  |

<sup>a</sup> n.d. = not done.

highest possible C/O ratio and conductivity using potent biomolecules. For environmental applications, the goal may be to attain partial reduction to retain functional groups for adsorption. For biomedical uses, microbial methods that ensure biocompatibility will be prioritized.

**4.2.2 Structural and functional control.** Advanced techniques will enable precise control over the number and type of residual functional groups. The incorporation of heteroatoms (e.g., nitrogen, sulfur) during the green reduction process,





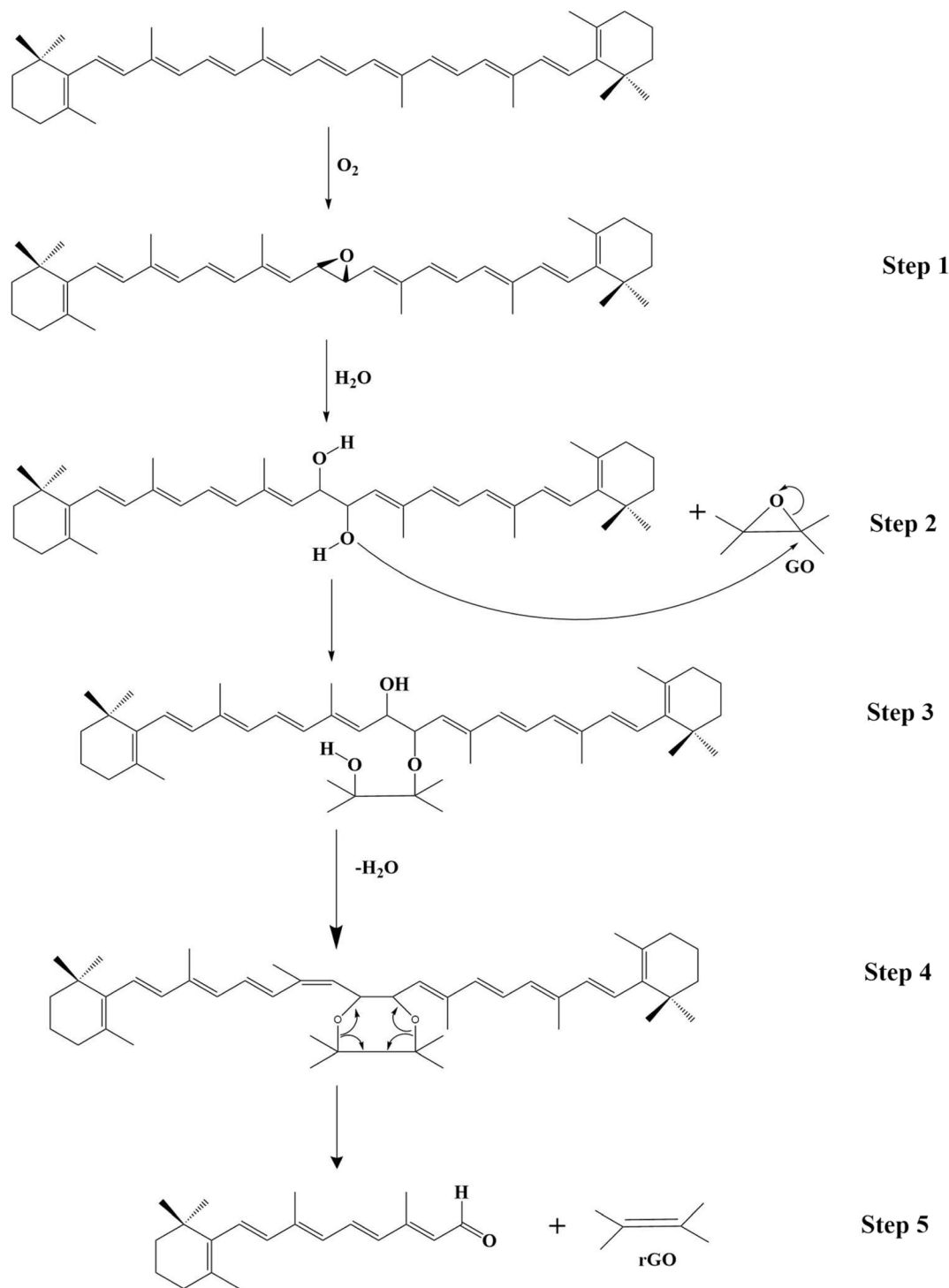


Fig. 8 Proposed multi-step reduction mechanism of GO to rGO by  $\beta$ -carotene, involving epoxidation, hydrolysis, and nucleophilic attack. Adapted from ref. 131 (Zaid, *et al.*) with permission under the terms of the Open Access Creative Commons CC-BY-NC-ND license, Elsevier (*Arabian Journal of Chemistry*), copyright 2014.

a technique known as doping, will be explored to tune the electronic and catalytic properties of rGO.

**4.2.3 Hybrid methods.** Combining the advantages of different techniques, such as a microwave-assisted reduction using a plant extract, could dramatically shorten reaction times while maintaining the benefits of a green process.

**4.2.4 Integration with artificial intelligence.** Machine learning algorithms could be used to screen vast libraries of natural compounds to predict their reduction potential and to optimize reaction parameters, accelerating the discovery of new, highly efficient green reduction pathways.

Table 4 A quantitative comparison of green reduction strategies

| Criteria   | Plant extract-mediated                        | Microorganism-mediated                         | Biomolecule-mediated                               |
|--|---|--|--|
| Reduction efficiency (C/O ratio)                     | Moderate to high (typical range: 3.8–11.9)    | Moderate (typical range: 2.6–5.9)              | High (typical range: 3.9–12.5, esp. ascorbic acid) |
| Reaction time  | Fast (0.5–24 h)                               | Slow (24–168 h)                                | Moderate (1–24 h)                                  |
| Typical electrical conductivity (S m <sup>-1</sup> ) | Low to moderate (e.g., 0.358 for gallic acid) | Moderate (e.g., 55.32 for <i>Shewanella</i> )  | Moderate to high (e.g., 105 for humanin)           |
| Scalability  | High (simple equipment, abundant materials)   | Low (requires bioreactors, sterile conditions) | Moderate (depends on biomolecule cost)             |
| Control over functionalization                       | Low (complex mixture of phytochemicals)       | Moderate (bio-functionalization possible)      | High (well-defined reductant molecule)             |
| Cost   | Low   | High (culture media, incubation energy)        | Variable (low for glucose, high for proteins)      |

## 5 Conclusion

This review critically examines significant progress in the green reduction of graphene oxide using plant extracts, microorganisms, and biomolecules. These environmentally friendly strategies offer sustainable and often low-cost alternatives to conventional methods that rely on hazardous chemicals. By moving beyond a descriptive summary, we have provided a comparative analysis that links the choice of green reductant to the resulting material properties and its ultimate performance in technological applications. The study reveals that while plant extracts offer a scalable and straightforward route, biomolecules provide superior control and higher reduction efficiency, and microorganisms yield products with exceptional biocompatibility. For instance, the use of potent biomolecules, such as ascorbic acid, offers the most promising path to high-conductivity electronics, while microbially reduced rGO is the clear-cut choice for biocompatible medical applications. Substantial challenges remain, particularly in standardizing natural extracts, understanding reaction kinetics, and scaling up production. However, the immense biodiversity of the natural world offers a vast, largely untapped resource for discovering novel reducing agents. Future research should focus on a more rational, application-driven approach to synthesis, aiming to precisely control the structural and chemical properties of rGO to meet the specific demands of advanced applications in energy, environmental science, and medicine. By addressing current research gaps, green-synthesized rGO is poised to become a cornerstone material in the development of next-generation sustainable technologies.

## Consent to publish

All authors agreed with the manuscript's content and gave explicit permission to publish the work.

## Conflicts of interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## Data availability

No primary research results, software or code have been included and no new data were generated or analysed as part of this review.

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