



Cite this: *Org. Biomol. Chem.*, 2026, **24**, 1672

## Unusual C-2 selectivity of Grignard additions to *N*-Boc-protected isatin 3-imines

Alexandra Mary Plakas,<sup>a,b</sup> Kamogelo Rosinah Butsi,<sup>a,b</sup> Sahil Lala,<sup>b,c</sup> Robyn Lynne van Zyl,<sup>b,c</sup> Manuel Antonio Fernandes,<sup>b</sup> Songeziwe Ntsimango,<sup>a</sup> Moira Leanne Bode<sup>b</sup> and Amanda Louise Rousseau<sup>\*a,b</sup>

A series of *N*-*tert*-butoxycarbonyl-protected isatin 3-imines underwent an unexpected C-2 selective addition reaction with Grignard reagents, affording 3-imino-2-phenethylindolin-2-ols. The protecting group plays a role in this observed chemoselectivity, with *N*-benzyl and *N*-*p*-methoxybenzyl-protected isatin 3-imines undergoing C-3 addition under Grignard reaction conditions, affording 3-amino-3-phenethylindolin-2-ones. Both C-2 and C-3 addition products were assessed for antiplasmodial activity *in vitro*, with four compounds displaying activity in the sub-micromolar range against both drug-sensitive and drug-resistant *Plasmodium falciparum* strains.

Received 5th December 2025,  
Accepted 27th January 2026

DOI: 10.1039/d5ob01909e

rsc.li/obc

## Introduction

The oxindole scaffold is present in numerous natural and synthetic products that display a wide range of biological activities.<sup>1–5</sup> In particular, 3,3-disubstituted 2-oxindoles have shown promise in the clinic, with selected examples including cipargamin, a potent antimalarial drug candidate currently in clinical trials,<sup>6</sup> and nelivaptan, a vasopressin V1b receptor antagonist for the treatment of depression and anxiety<sup>7</sup> (although suboptimal pharmacokinetic properties have halted further development) (Fig. 1).

The pharmacological activity of this class of compounds has attracted the interest of synthetic and medicinal chemists, and many synthetic methods have been described for the synthesis of compounds containing an oxindole core.<sup>8–13</sup>

Previously, we reported the synthesis and antiplasmodial activity of a series of racemic spiroindolones **1** inspired by cipargamin, prepared by the Povarov reaction of ketimines with electron-rich alkenes (Fig. 2).<sup>14</sup> We found that only spiroindolones bearing a methyl or a halogen substituent *para* to the ring nitrogen (*i.e.* **1**, R<sup>1</sup> = Me or Hal) showed moderate antiplasmodial activity *in vitro* (1.31–4.20 μM), with all other compounds being inactive in this assay. Although these spiroindolones displayed only moderate activity *in vitro*, the interesting

structure–activity relationships that were observed prompted us to consider the design of second-generation analogues. As the mode of action was not known, we considered strategies that would allow us to explore additional chemical space. One

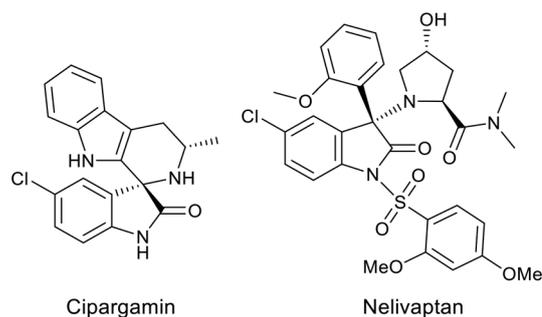


Fig. 1 Oxindoles with potent biological activity, cipargamin and nelivaptan.

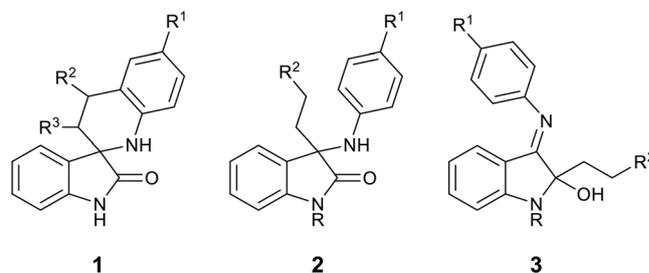


Fig. 2 Spiroindolones **1** prepared previously,<sup>14</sup> ring-opened analogues **2** and indolin-2-ols **3**.

<sup>a</sup>Molecular Sciences Institute, School of Chemistry, University of the Witwatersrand, Private Bag 3, PO WITS, 2050, South Africa. E-mail: Amanda.Rousseau@wits.ac.za

<sup>b</sup>WITS Research Institute for Malaria (WRIM), Faculty of Health Sciences, University of the Witwatersrand, 7 York Road, Parktown 2193, South Africa

<sup>c</sup>Pharmacology Division, Department of Pharmacy and Pharmacology, Faculty of Health Sciences, University of the Witwatersrand, 7 York Road, Parktown 2193, South Africa



such strategy was to consider the virtual ring opening of the tetrahydroisoquinoline ring, affording oxindoles **2** with increased flexibility, while retaining the *para*-substituent R<sup>1</sup>. Herein, we report our synthetic approaches to a series of ring opened analogues **2**, and the unexpected synthesis of indolin-2-ols **3**.

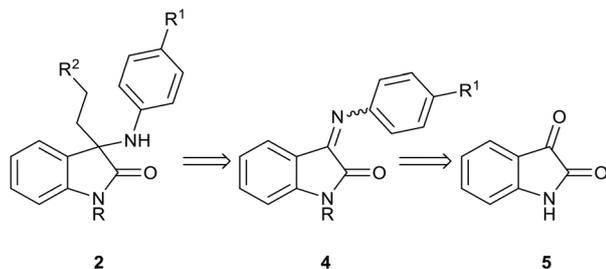
## Results and discussion

We envisaged preparing ring-opened analogues **2** as racemates initially, by reaction of ketimines **4** (derived from commercially available isatins **5**), with suitably substituted Grignard reagents (Scheme 1).

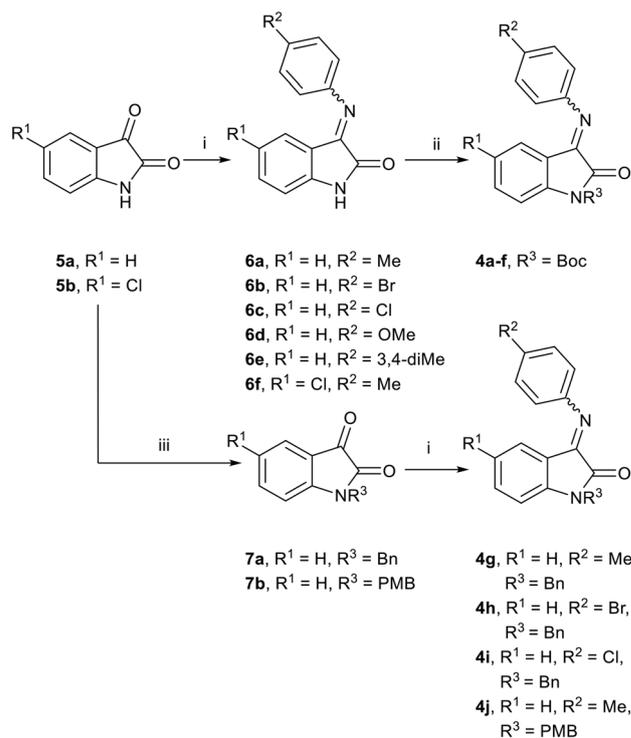
Our initial approach involved isatin *N*-Boc-protection, followed by reaction with *para*-substituted anilines to prepare ketimines **4**. However, under these conditions, only the hemiaminal was isolated in low yields in each case. Although there is literature precedent for the use of hemiaminals in Grignard addition reactions,<sup>15</sup> this also proved to be low-yielding in our hands. Furthermore, the *N*-Boc-protected isatin was found to be unstable at room temperature. By first reacting isatins **5** with *para*-substituted anilines to afford intermediate ketimines **6a–f**, followed by *N*-Boc-protection, the protected ketimines **4a–f** were isolated in reasonable yields (Scheme 2).<sup>16</sup> In each case, the *E*-isomer was the major isomer formed, and in some cases only the *E*-isomer was isolated after purification.

We then reacted the *N*-Boc-protected ketimines **4a–f** with Grignard reagents derived from substituted phenethyl bromides, in an attempt to form oxindoles **2** (Scheme 3). This approach was based on the work of Lesma *et al.*, where various Grignard reagents were successfully added to isatin-derived alkyl- and sulfinyl-imines, achieving yields ranging from 49–77%.<sup>15</sup> However, to our surprise, Grignard addition to our ketimine substrates **4a–f** occurred at the C-2 carbonyl of the indolin-2-one core, and not the expected C-3 imine, affording 3-imino-2-phenethylindolin-2-ols **3** instead (Scheme 3 and Table 1).

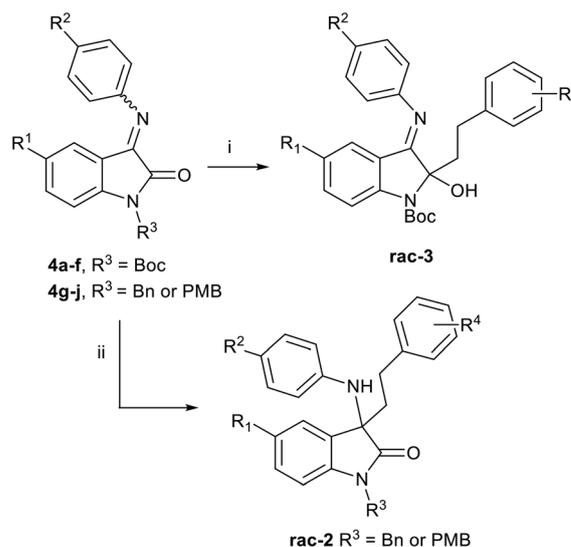
This was confirmed by single-crystal X-ray diffraction on two analogues, one of which is shown in Fig. 3a. The reaction was repeated on ketimines substituted with both electron-donating and electron-withdrawing groups using a range of substituted phenethylmagnesium bromides, affording analogues **3a–3aa** in varying yields of 10–53%.



**Scheme 1** Planned approach for the synthesis of oxindoles **2**.



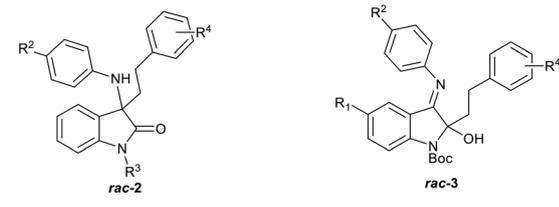
**Scheme 2** Reaction conditions: (i) substituted aniline, EtOH, AcOH, reflux, 5 h, 61–91%; (ii) Boc<sub>2</sub>O, THF, DMAP, rt, 3 h, 71–98%; (iii) (a) BnBr (for **7a**) or 4-(OCH<sub>3</sub>)C<sub>6</sub>H<sub>4</sub>-CH<sub>2</sub>Br (for **7b**), K<sub>2</sub>CO<sub>3</sub>, KI, DMF, reflux, 5 h, 55–74%.



**Scheme 3** Reaction conditions: (i) substituted phenethylmagnesium bromide, MgBr<sub>2</sub>, THF, –40 °C–rt, 18 h, 10–53%; (ii) substituted phenethylmagnesium bromide, MgBr<sub>2</sub>, DCM, –40 °C–rt, 18 h, 2–12%.

The yield of isolated product was found to be low in most cases and attempts to improve the yield of the reaction by varying the solvent, temperature and number of equivalents of the Grignard reagent added, were not successful. In every case,



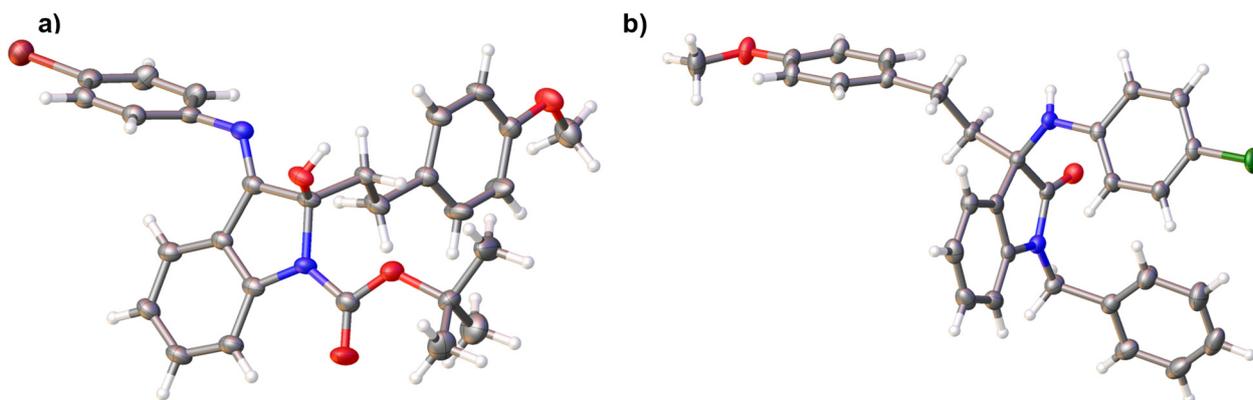
**Table 1** Yields of oxindoles **2** and 3-imino-2-phenethylindolin-2-ols **3** prepared as described in Scheme 3


Compound	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	R <sup>4</sup>	Yield %
2a	—	Me	Bn	2-Cl	5
2b	—	Br	Bn	4-OMe	6
2c	—	Cl	Bn	4-OMe	12
2d	—	Me	PMB	2-OMe	2
3a	H	Me	—	4-OMe	35
3b	H	Me	—	4-Cl	38
3c	H	Me	—	3-Cl	19
3d	H	Me	—	2,4-diCl	10
3e	H	Br	—	4-OMe	45
3f	H	Br	—	4-Cl	34
3g	H	Br	—	3-Cl	39
3h	H	Br	—	2,4-diCl	27
3i	H	Cl	—	4-OMe	50
3j	H	Cl	—	4-Cl	53
3k	H	Cl	—	3-Cl	48
3l	H	Cl	—	2,4-diCl	15
3m	H	OMe	—	4-OMe	32
3n	H	OMe	—	4-Cl	39
3o	H	OMe	—	3-Cl	40
3p	H	OMe	—	2,4-diCl	33
3q	H	3,4-diMe	—	4-Cl	15
3r	5-Cl	3,4-diMe	—	3-Cl	18
3s	5-Cl	Me	—	4-OMe	13
3t	5-Cl	Me	—	3-OMe	12
3u	5-Cl	Me	—	2-OMe	19
3v	5-Cl	Me	—	4-Cl	37
3w	5-Cl	Me	—	3-Cl	49
3x	5-Cl	Me	—	2-Cl	42
3y	5-Cl	Me	—	2,4-diCl	22
3z	5-Cl	Me	—	4-F	48
3aa	5-Cl	Me	—	2-F	23

both starting material and several side products were isolated, including the *N*-Boc deprotected starting material **6**, as well as isatin **5**. The product of Wurtz homocoupling of the phenethyl bromides used to prepare the Grignard reagent was also identified in some cases in low yields (3–7%), however, the low yield of desired products is not likely to be due to inefficient preparation of the phenethylmagnesium bromide derivatives as the Grignard reagent was prepared in excess. Interestingly, the  $\beta$ -hydride elimination product was not isolated from these reactions. However, we did isolate secondary amines in some instances, which could have formed by the reaction of either the  $\beta$ -hydride elimination product or unreacted phenethyl bromide with aniline formed *in situ* by imine hydrolysis.

To the best of our knowledge, this unexpected C-2 chemoselectivity in reactions of *N*-Boc isatin-derived imines has not been reported previously. However, similar reactivity has been observed in *N*-acyl isatins with nucleophiles, typically leading to ring opening by cleavage of the isatin C-2–N bond.<sup>17–20</sup>

By comparison, isatin and *N*-alkyl isatins have been shown to undergo nucleophilic addition at the C-3 carbonyl with hard amine and alcohol nucleophiles.<sup>18</sup> We therefore speculated that the *N*-protecting group may have played a role in the unexpected chemoselectivity of the reaction. To further investigate this, we prepared *N*-benzyl- and *N*-*p*-methoxybenzyl-protected ketimines **4g–j** (Scheme 2) and subjected these to Grignard reaction conditions. Interestingly, these reactions did not proceed at all in tetrahydrofuran, however, very low yields of product were isolated when the reaction was carried out in dichloromethane, affording analogues **2** (Scheme 3, four examples, **2a–d**), with starting material recovered in each case. Further attempts to improve the yield of the reaction by the addition of LiCl to the Grignard reagent, or by the addition of CuI to the ketimine prior to the addition of the Grignard reagent, were not successful. Despite the very low yields obtained for these reactions, we were able to obtain a single crystal X-ray structure for one analogue, which confirmed that in these instances, Grignard addition had taken place at the C-3 imine carbon to afford *rac*-**2c** (Fig. 3b). When the Grignard

**Fig. 3** ORTEP diagrams of the X-ray crystal structures of (a) *tert*-butyl 3-((4-bromophenyl)imino)-2-hydroxy-2-(4-methoxyphenethyl)indoline-1-carboxylate **3e** and (b) 1-benzyl-3-((4-chlorophenyl)amino)-3-(4-methoxyphenethyl)indolin-2-one **2c**. Thermal ellipsoids are shown at the 50% probability level.

reaction was tested on the unprotected imines **6**, no reaction was observed.

Apart from the *N*-protecting group, scaffolds **2** and **3** have similar electronic functionalities; C2=O/C3=N and C3=N/C2=O, and therefore, only give rise to minor <sup>13</sup>C NMR spectroscopic chemical shift differences. 3-Amino-3-phenethylindolin-2-ones **2** contain characteristic C2=O and C3=N functionalities, which give rise to signals in their <sup>13</sup>C NMR spectra at 177.4–177.8 ppm and 64.0–65.3 ppm, respectively. In contrast, 2-phenethyl-3-imino-indolin-2-ols **3** contain characteristic C3=N and C2=O moieties, which give rise to signals appearing at 165.1–171.1 ppm and 91.6–95.6 ppm in the <sup>13</sup>C NMR spectra, respectively.

To explore this unusual chemoselectivity further, we tested the addition of allylmagnesium bromide to both *N*-benzyl-protected ketimine (**4i**) and *N*-Boc-protected ketimine (**4c**), however, only the benzyl-protected ketimine (**4i**) afforded the expected addition product upon reaction with allylmagnesium bromide (see SI). Reaction of **4c** with allylmagnesium bromide gave the deprotected imine **6c** as the major product. We did not confirm the mechanism by which this deprotection occurred, however, it is possible that chelation with the Lewis acid may have promoted hydrolysis upon work-up, or that Grignard addition may have occurred at the Boc-carbonyl carbon instead of the isatin C-2 carbonyl.

We then postulated that Grignard additions to *N*-Boc-protected benzylidene ketimines **10** would result in C-2 addition. To this end, we prepared benzylidene isatin **10** via a Knoevenagel condensation of oxindole and *p*-tolualdehyde (Scheme 4). To our surprise, treatment of benzylidene ketimines **10** under the same Grignard reaction conditions shown in Scheme 3 were unsuccessful, with only starting material recovered in each case. This suggests that the C-3 electron-withdrawing group present in the *N*-Boc-protected ketimines **4a–f** is also essential for activating the C-2 carbon towards nucleophilic attack.

We therefore deduce that in the case of *N*-alkyl protected substrates, the C-2 carbonyl participates in amide resonance, reducing the electrophilicity of the C-2 carbon while enhancing the electrophilicity of the C-3 carbon (Fig. 4a). In contrast, for *N*-acyl protected substrates, while the resonance form described in Fig. 4a is possible, there is an additional form in which the nitrogen engages in resonance with the acyl carbonyl (Fig. 4b). We speculate that this, together with the electron-

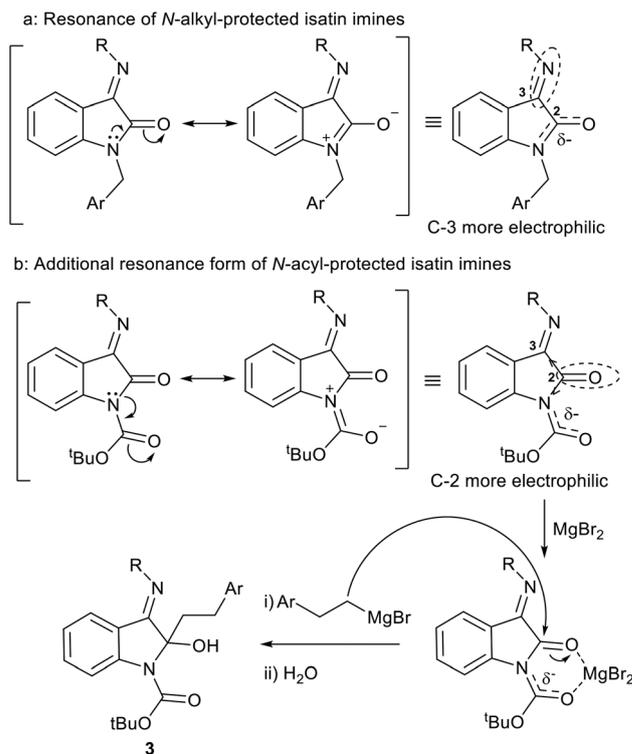
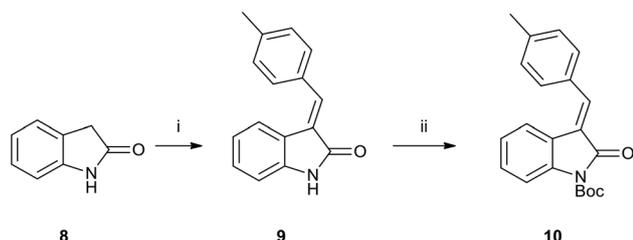


Fig. 4 Comparison of the resonance behaviour in (a) *N*-alkyl and (b) *N*-acyl protected isatin imines and implications for chemoselectivity.

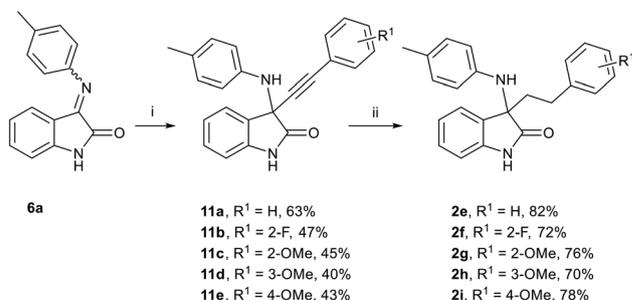
withdrawing effect of the C-3 imine, increases the electrophilicity of the C-2 carbonyl carbon. In the presence of the Lewis acid MgBr<sub>2</sub>, which can coordinate to both the *N*-Boc carbonyl and the isatin C-2 carbonyl, this facilitates reaction at C-2 with phenethylmagnesium bromides, affording 3-imino-2-phenethylindolin-2-ols **3** after work-up (Fig. 4b).

Finally, we set about preparing the original targeted compounds **2** by reacting isatin-derived imine **6a** with substituted phenylacetylene nucleophiles (Scheme 5) as demonstrated by Xie *et al.*<sup>21</sup> The desired C-3 addition products, 3-amino-3-ethynylindolin-2-ones **11a–e**, were afforded in yields of 40–63%.

Initial attempts to reduce substrates **11a–e** with traditional Pd/C catalysed hydrogenation conditions led to the removal of the aniline moiety. However, the desired 3-amino-3-phenethyl-

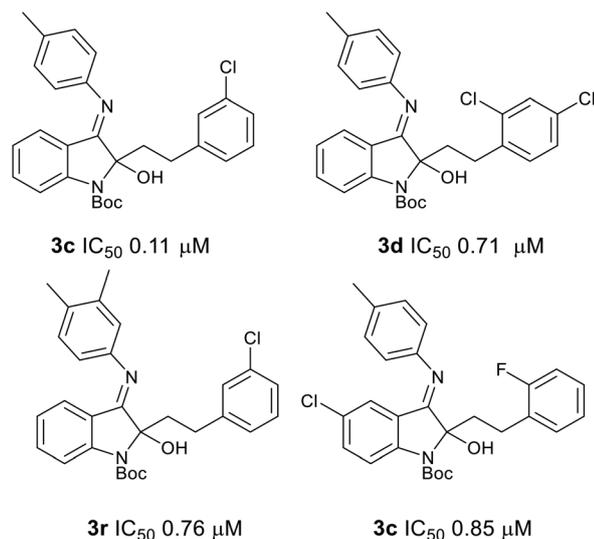


Scheme 4 Reaction conditions: (i) *p*-tolualdehyde, piperidine, EtOH, under Ar, reflux, 4 h., 24 h, 80%; (ii) Boc<sub>2</sub>O, DMAP, THF, under Ar, rt, 24 h, 73%.



Scheme 5 Reaction conditions: (i) HC≡C-PhR<sup>1</sup>, KOtBu, CuI, THF, r.t., 24 h, 40–63%; (ii) H<sub>2</sub>, Pd/C (5%), Na<sub>2</sub>CO<sub>3</sub>, EtOH, r.t., 200 mbar, 70–82%.





**Fig. 5** Most potent compounds identified from the *P. falciparum* whole cell assay.

indoline-2-ones **2e–i** were afforded in yields of 70–82% by the addition of sodium carbonate. C-3 addition was confirmed by NMR spectroscopy, where the characteristic C2=O and C3–N <sup>13</sup>C NMR spectroscopic chemical shifts were observed at 179.0–179.2 ppm and 64.0–64.2 ppm, respectively.

We assessed a selection of the Boc-protected 2-phenethylindolin-2-ols **3** and 3-amino-3-phenethylindoline-2-ones **2** for antiparasitic activity in a whole cell *P. falciparum* screen against a drug-sensitive parasite strain (NF54). Those compounds that were found to inhibit parasite growth by 60% or more at a concentration of 10 μM were then assessed further and IC<sub>50</sub> values obtained. From these assays, we were able to determine that the 3-imino-2-phenethylindolin-2-ols **3** were more potent inhibitors of parasite growth than the 3-amino-3-phenethylindolin-2-ones **2**, as only one of the 3-amino-3-phenethylindolin-2-ones **2** tested inhibited parasite growth by more than 50% at a concentration of 10 μM (see SI). By comparison, all but one of the 2-phenethylindolin-2-ols **3** inhibited parasite growth by more than 50% at a concentration of 10 μM, with four compounds displaying sub-micromolar activity against the NF54 strain *in vitro* (IC<sub>50</sub> 0.11–0.85 μM, Fig. 5). The remaining compounds were active in the low micromolar range (1.03–7.83 μM, see SI). A similar trend was observed against the chloroquine-resistant FCR3 strain, with four compounds displaying moderate activity (IC<sub>50</sub> 0.35–0.63 μM). Compounds were also tested for cytotoxicity against the human embryonic kidney epithelial cell line HEK293 in an MTT cell viability assay, with no significant cytotoxicity observed.

## Conclusions

Attempts to prepare a series of 3-amino-3-phenethylindolin-2-ones *via* Grignard addition to *N*-Boc-protected isatin-derived

imines led to the discovery of an interesting chemoselectivity, yielding 3-imino-2-phenethylindolin-2-ols instead. While the Grignard addition reactions to *N*-Boc-protected isatin imines were C-2 selective, those to *N*-Bn- and *N*-PMB-protected isatin imines favoured C-3 addition, establishing the importance of the *N*-protecting group in this observed chemoselectivity. Furthermore, Grignard additions to benzylidene indolin-2-one substrates lacking the imine moiety did not proceed, confirming the imine as essential for C-2 selectivity. This Grignard-based approach offers a novel strategy for functionalising the C-2 carbonyl of 3-iminoindolin-2-ones, yielding the biologically relevant indolin-2-ol scaffold. The target 3-amino-3-phenethylindolin-2-ones were subsequently prepared *via* alkynylation and hydrogenation of isatin-derived imines in yields up to 52%. Compounds from each series were evaluated for antiparasitic activity against the drug-sensitive NF54 strain of the parasite. Four compounds exhibited moderate activity in the sub-micromolar range against both drug-sensitive and drug-resistant strains of *Plasmodium falciparum*. In future studies, the synthesis of chiral oxindole products could be considered.

## Author contributions

Conceptualisation, ALR and AMP; synthesis of compounds KRB and AMP; crystallography, MAF; biological data, SL and RLvZ; writing, ALR and AMP; co-supervision of AMP and KRB, SN and MLB.

## Conflicts of interest

There are no conflicts to declare.

## Data availability

The data supporting this article have been included as part of the supplementary information (SI). Supplementary information: experimental procedures, characterization data and copies of NMR spectra. See DOI: <https://doi.org/10.1039/d5ob01909e>.

Ref. 22–39 appear in the SI.

CCDC 2513396 and 2513397 contain the supplementary crystallographic data for this paper.<sup>40a,b</sup>

## Acknowledgements

We thank the NRF and the University of the Witwatersrand for generous funding to enable the purchase of a dual-wavelength hybrid diamond anode X-ray diffractometer (Bruker D8 Discovery Bio equipped with Mo and Cu X-ray sources) under NEP Grant No 129920. We acknowledge the WITS Pharmacology Division McGill Trust for research funding support.



## References

- 1 F. Salim and R. Ahmad, *Stud. Nat. Prod. Chem.*, 2015, **45**, 485–525.
- 2 B. Yu, D.-Q. Yu and H.-M. Liu, *Eur. J. Med. Chem.*, 2015, **97**, 673–698.
- 3 Y.-T. Yang, J.-F. Zhu, G. Liao, H.-J. Xu and B. Yu, *Curr. Med. Chem.*, 2018, **25**, 2233–2244.
- 4 S. S. Panda, R. A. Jones, P. Bachawala and P. P. Mohapatra, *Mini-Rev. Med. Chem.*, 2017, **17**, 1515–1536.
- 5 M. Kaur, M. Singh, N. Chadha and O. Silakari, *Eur. J. Med. Chem.*, 2016, **123**, 858–894.
- 6 M. Rottmann, C. McNamara, B. K. S. Yeung, M. C. S. Lee, B. Zou, B. Russell, P. Seitz, D. M. Plouffe, N. V. Dharia, J. Tan, S. B. Cohen, K. R. Spencer, G. E. González-Páez, S. B. Lakshminarayana, A. Goh, R. Suwanarusk, T. Jegla, E. K. Schmitt, H.-P. Beck, R. Brun, F. Nosten, L. Renia, V. Dartois, T. H. Keller, D. A. Fidock, E. A. Winzeler and T. T. Diagana, *Science*, 2010, **329**, 1175–1180.
- 7 M. Schönberger, C. Leggett, S. W. Kim and J. M. Hooker, *Bioorg. Med. Chem. Lett.*, 2010, **20**, 3103–3106.
- 8 G. M. Ziarani, R. Moradi and N. Lashgari, *Tetrahedron*, 2018, **74**, 1323–1353.
- 9 B. Yu, H. Xing, D.-Q. Yu and H.-M. Liu, *Beilstein J. Org. Chem.*, 2016, **12**, 1000–1039.
- 10 J.-S. Yu, F. Zhou, Y.-L. Liu and J. Zhou, *Synlett*, 2015, 2491–2504.
- 11 G. M. Ziarani, P. Gholamzadeh, N. Lashgari and P. Hajiabbasi, *ARKIVOC*, 2013, **2013**, 470–535.
- 12 M. Zhao, N.-K. Li, Y.-F. Zhang, F.-F. Pan and X.-W. Wang, *Tetrahedron*, 2016, **72**, 1406–1414.
- 13 H. Pellissier, *Synthesis*, 2019, 1311–1318.
- 14 B. Mathebula, K. R. Butsi, R. L. van Zyl, N. C. Jansen van Vuuren, H. C. Hoppe, J. P. Michael, C. B. de Koning and A. L. Rousseau, *Chem. Biol. Drug Des.*, 2019, **94**, 1849–1858.
- 15 G. Lesma, N. Landoni, T. Pilati, A. Sacchetti and A. Silvani, *J. Org. Chem.*, 2009, **74**, 4537–4541.
- 16 Z. Al Marhoon, A. Abdel-Megeed, E. N. Sholkamy, M. Rafiq, H. Siddiqui and A. El-Faham, *Asian J. Chem.*, 2014, **26**, 7665–7672.
- 17 J. F. M. da Silva, S. J. Garden and A. C. Pinto, *J. Braz. Chem. Soc.*, 2001, **12**, 273–324.
- 18 W. C. Cheah, K. Wood, D. S. Black and N. Kumar, *Tetrahedron*, 2011, **67**, 7603–7610.
- 19 A. El-Faham, S. N. Khattab, H. A. Ghabbour, H.-K. Fun and M. R. H. Siddiqui, *Chem. Cent. J.*, 2014, **8**, 27.
- 20 V. Suryanti, R. Zhang, V. Aldilla, M. Bhadbhade, N. Kumar and D. S. Black, *Molecules*, 2019, **24**, 4343.
- 21 D. Xie, X. Xu, S. Long, X.-Y. Tang and L. Wang, *J. Org. Chem.*, 2022, **87**, 7852–7863.
- 22 I. Allous, S. Comesse, M. Sanselme and A. Daïch, *Eur. J. Org. Chem.*, 2011, 5303–5310.
- 23 P. K. Prasad, R. G. Kalschetti, R. N. Reddi, S. P. Kamble and A. Sudalai, *Org. Biomol. Chem.*, 2016, **14**, 3027–3030.
- 24 X. Guo, P. Zhang, M. Chen, T. Li, C. Hou, X. Que, L. Xu, Z. Zhou, Q. Wang and Z. Wang, *Bioorg. Chem.*, 2024, **153**, 107757.
- 25 J. Tian, C. Li and H. Lv, *J. Org. Chem.*, 2025, **90**, 12023–12028.
- 26 S. K. Sridhar, M. Saravanan and A. Ramesh, *Eur. J. Med. Chem.*, 2001, **36**, 615–625.
- 27 F. Yu, P. Li, B. Wang and K. Han, *J. Am. Chem. Soc.*, 2013, **135**, 7674–7680.
- 28 K. Gopalaiyah and A. Tiwari, *Eur. J. Org. Chem.*, 2020, 7229–7237.
- 29 P. You, M. Liu, K. Zhang, F. Yang, Z. Tan and F. Chen, *Org. Biomol. Chem.*, 2024, **22**, 4466–4471.
- 30 W. Trager and J. B. Jensen, *Science*, 1976, **193**, 673–675.
- 31 C. Lambros and J. P. Vanderberg, *J. Parasitol.*, 1979, **65**, 418–420.
- 32 M. T. Makler and D. J. Hinrichs, *Am. J. Trop. Med. Hyg.*, 1993, **48**, 205–210.
- 33 R. L. van Zyl, A. M. Viljoen and A. K. Jäger, *S. Afr. J. Bot.*, 2002, **68**, 106–110.
- 34 T. Mosmann, *J. Immunol. Methods*, 1983, **65**, 55–63.
- 35 R. L. Van Zyl, S. T. Seatlholo and A. M. Viljoen, *S. Afr. J. Bot.*, 2010, **76**, 662–667.
- 36 Bruker, *APEX4 version 2021.4-1 data collection software which includes SAINT version 8.40B, SADABS-2016/2, and XPREP version 2014/2*, Bruker AXS Inc., Madison, Wisconsin, USA, 2021.
- 37 O. V. Dolomanov, L. J. Bourhis, R. J. Gildea, J. A. K. Howard and H. Puschmann, *J. Appl. Crystallogr.*, 2009, **42**, 339–341.
- 38 G. Sheldrick, *Acta Crystallogr., Sect. A: Found. Adv.*, 2015, **71**, 3–8.
- 39 G. Sheldrick, *Acta Crystallogr., Sect. C: Struct. Chem.*, 2015, **71**, 3–8.
- 40 (a) CCDC 2513396: Experimental Crystal Structure Determination, 2026, DOI: [10.5517/ccdc.csd.cc2qcd9t](https://doi.org/10.5517/ccdc.csd.cc2qcd9t); (b) CCDC 2513397: Experimental Crystal Structure Determination, 2026, DOI: [10.5517/ccdc.csd.cc2qcdbv](https://doi.org/10.5517/ccdc.csd.cc2qcdbv).

