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The ammonia-Ugi reaction employing ammonium carboxylates of N-protected amino acids (or peptides), ketones, and  $\alpha$ -isocyano esters enabled N-to-C peptide elongation, together with *in situ* construction of  $\alpha$ , $\alpha$ -disubstituted amino acid residues. This method offered an effective synthetic method of novel elastin-like short peptides, which exhibited highly potent self-assembling properties.

Peptide synthesis is typically performed via stepwise coupling of N-protected amino acids to a growing peptide chain anchored on a solid support using condensation agents.<sup>3</sup> Although such a solid-phase peptide synthesis (SPPS) works well with proteinogenic and other  $\alpha$ -monosubstituted amino acids, it often results in poor yields with sterically hindered  $\alpha,\alpha$ -disubstituted amino acids, requiring excessive amounts of coupling agents under harsh conditions such as heating or microwave irradiation.<sup>4</sup> Moreover, preparation of N-protected  $\alpha,\alpha$ -disubstituted amino acids generally requires multi-step and tedious processes. Alternatively, highly reactive condensation agents<sup>5</sup> and catalytic reaction systems<sup>6</sup> have offered efficient methods for unnatural peptide synthesis; however, these still use expensive unnatural  $\alpha,\alpha$ -disubstituted amino acids as substrates. These synthetic challenges have long

posed significant barriers to the discovery and development of bioactive peptides.

The ammonia-Ugi reaction, a variant of the Ugi reaction, <sup>7,8</sup> is a four-component coupling reaction that involves ammonia, an aldehyde or a ketone, a carboxylic acid, and an isocyanide, enabling a straightforward synthesis of peptides. 9 Although the ammonia-Ugi reaction had long been considered impractical and unsuccessful, 10 we recently reported an efficient synthetic protocol of unnatural dipeptides using the ammonia-Ugi reaction: by stirring ammonium carboxylates derived from N-protected amino acids 1, ketones, and isocyanides in trifluoroethanol (TFE) at ambient temperature, a variety of dipeptides 2 were obtained in good yields (Scheme 1a).11 Of note, these dipeptides 2 contained unnatural α,α-disubstituted amino acids, which were constructed in situ during the ammonia-Ugi reaction from readily available ketones as building blocks. Mechanistically, ammonium (NH<sub>4</sub><sup>+</sup>) dissociates into NH3 and H+, which in turn facilitates the thermodynamically unfavourable N-unsubstituted imine formation (Scheme 1a). 12 With its success, however, the products 2 were obtained as amides at their C-termini, rendering them unsuitable for further N-to-C peptide elongation. To the best of our knowledge, neither the ammonia-Ugi reaction nor the Ugi reaction has ever been applied for N-to-C peptide chain elongation. 13,14 Here, this study proposes a novel strategy for N-to-C elongation of unnatural peptides by the ammonia-Ugi reaction employing α-isocyano ester 3 as an elongation unit (Scheme 1b). The expected ammonia-Ugi adducts 4 possess ester moieties at their C-termini, providing a versatile platform for subsequent peptide chain elongation.

Using a commercially available ethyl isocyanoacetate (3, CN-Gly-OEt), a series of unnatural tripeptides 4 were synthesized (Scheme 2). The key ammonium carboxylates of N-protected amino acids 1 were prepared by stirring N-protected amino acids with aqueous ammonia in acetonitrile or THF at 0 °C (Table S1). Then, the ammonia-Ugi reaction using Boc-Phe-ONH<sub>4</sub>, cyclopentanone, and CN-Gly-OEt (3) afforded the tripeptide Boc-Phe-Ac<sub>5</sub>c-Gly-OEt (4a) in

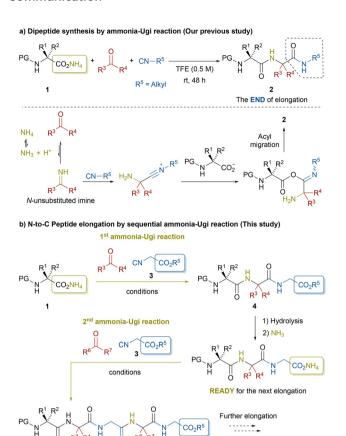
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**Scheme 1** (a) Dipeptide synthesis by ammonia-Ugi reaction, (b) N-to-C peptide elongation by sequential ammonia-Ugi reaction (this study).

93% yield, with no detectable racemization of the chiral α-carbon of phenylalanine (>99.5:0.5 er). Herein, the unnatural 1-aminocyclopentane-1-carboxylic acid (Ac5c) residue in 4a was constructed in situ from cyclopentanone as a substrate. In general, diethylglycine (Deg) is a challenging substrate to be incorporated into peptides under conventional SPPS conditions because of the steric hindrance, <sup>15</sup> whereas the present ammonia-Ugi reaction successfully delivered Boc-Phe-Deg-Gly-OEt (4b) in 87% yield. Again, no racemization was observed during this process (>99.5 : 0.5 er). The chiral  $\alpha$ -carbon of serine is known to be susceptible to racemization during peptide synthesis;<sup>16</sup> however, the present reaction conditions afforded 4c in an excellent yield (97%) with a perfect stereochemical integrity (>99.5:0.5 er). Even the sterically demanding Boc-Aib was compatible with the present reaction conditions, affording the sterically congested tripeptide 4d, composed of contiguous α,α-disubstituted amino acids {α-aminoisobutyric acid (Aib) and  $\alpha$ -methylhomophenylalanine [( $\alpha$ Me)Hph]}, in 78% yield. In addition to Boc, various N-protecting groups, including Cbz, Bz, and formyl (For), were well tolerated, giving the tripeptides 4e, 4f, and 4g in 83%–99% yields. Unfortunately, the stereochemical integrity of Bz-Phe was slightly lost (98:2 er), whereas that of Ac-Phe remained intact under the same conditions.<sup>11</sup> It is worth noting that the N-formyl group in 4g can

**Scheme 2** Synthesis of tripeptides **4a–4g** by ammonia-Ugi reaction using ammonium carboxylates of *N*-protected amino acids. <sup>a</sup> 5.4 mmol, <sup>b</sup> 3.5 mmol, <sup>c</sup> 1 mmol, <sup>d</sup> 4.5 mmol.

serve as a precursor of an isocyano group, potentially enabling inverse C-to-N peptide elongation. Scale-up experiments (up to 5.4 mmol) proceeded smoothly without any detrimental effects on the reaction system, providing the corresponding products **4a**, **4b**, and **4g** in excellent yields (83%–93%). Overall, the ammonia-Ugi reaction using  $\alpha$ -isocyano ester and N-protected amino acids enabled the efficient synthesis of unnatural tripeptides containing  $\alpha$ , $\alpha$ -disubstituted amino acids. These tripeptides possess ester groups at their C-termini, offering a platform for subsequent N-to-C peptide chain elongation.

Peptides containing glycine are frequently found in biomaterials<sup>17</sup> such as elastin, <sup>18</sup> collagen, <sup>19</sup> and silk fibroin. <sup>20</sup> Owing to glycine's small size and conformational flexibility, it plays a crucial role in modulating peptide structure and function. The development of efficient synthetic strategies for unnatural analogues of glycine-rich peptides is therefore of significant interest in both synthetic and biomedical research. In this study, we developed an efficient N-to-C peptide elongation method for synthesizing glycine-containing unnatural peptides by the ammonia-Ugi reaction employing CN-Gly-OEt (3) (Scheme 3). The C-terminal ester moieties of tripeptides 4a and 4b were hydrolyzed under basic conditions and subsequently treated with aqueous ammonia to give the corresponding ammonium carboxylates 5a and 5b in excellent yields. Under the present

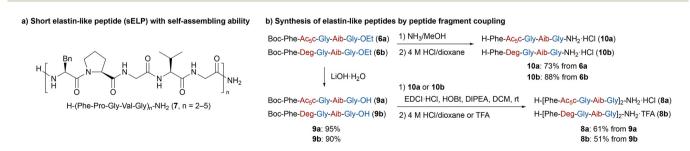
Scheme 3 Synthesis of pentapeptides 6a and 6b by ammonia-Uqi reaction starting from ammonium carboxylates of N-protected trippeptides 4a and 4b.

ammonia-Ugi reaction conditions, both tripeptides were successfully elongated into the pentapeptides Boc-Phe-Xaa-Gly-Aib-Gly-OEt 6a and 6b in good yields, together with in situ construction of Aib residue from acetone. The resulting peptides again possess ester moieties at C-termini, potentially providing a versatile platform for further N-to-C elongation or other chemical modifications. Totally, starting from N-protected amino acids 1, the first ammonia-Ugi reaction furnished tripeptides 4 (Scheme 2), and the second ammonia-Ugi reaction extended them to pentapeptides 6 (Scheme 3). Each step elongated peptides by two amino acid residues at a time while constructing  $\alpha,\alpha$ -disubstituted amino acids in situ. This streamlined approach requires no condensation agents, thus providing an environmentally friendly synthetic method of glycine-containing unnatural peptides.

The obtained pentapeptides 6a and 6b share the repeating amino acid sequence of a short elastin-like peptide (sELP) H-(Phe-Pro-Gly-Val-Gly)<sub>n</sub>-NH<sub>2</sub> (7, Scheme 4a).  $^{21}$  sELP exhibits reversible lower critical solution temperature (LCST)-type behaviour, being soluble at low temperatures and insoluble at high temperatures.<sup>22</sup> Such reversible temperature-responsive and self-assembling properties make sELP a promising candidate for drug delivery applications. The development of unnatural analogs with enhanced self-assembling properties is therefore of considerable interest. Herein, we designed a novel unnatural analogue, H-(Phe-Ac<sub>5</sub>c-Gly-Aib-Gly)<sub>2</sub>-NH<sub>2</sub> (8a), by modifying the original sELP sequence (Scheme 4b). Specifically, the Pro-2 in 7 was replaced with Ac<sub>5</sub>c, a noncanonical amino acid structurally related to Pro, and the Val-4, located at a guest position in the original sequence, 23 was substituted with Aib, a noncanonical amino acid analogous to Val.

The decapeptide 8a was synthesized in a good yield via segment coupling between Boc-Phe-Ac5c-Gly-Aib-Gly-OH (9a) and H-Phe-Ac5c-Gly-Aib-Gly-NH2 (10a) under standard condensation conditions, followed by Boc deprotection under acidic conditions. We also designed and synthesized a structurally related analogue, H-(Phe-Deg-Gly-Val-Gly)2-NH2 (8b), in which Ac<sub>5</sub>c residue in 8a was replaced with Deg, a noncyclic analogue of Ac5c. Both 8a and 8b were obtained as TFA salts in pure form after purification by reversed-phase (RP)-HPLC (Fig. S1). Additionally, we designed and synthesized H-(Phe-Pro-Gly-Aib-Gly)<sub>2</sub>-NH<sub>2</sub> (11) as an Aib analogue of the original sELP 7 via a standard SPPS protocol (see, SI).

The self-assembling properties of peptides 8a, 8b, and 11 were then investigated under buffered aqueous conditions. Each peptide was dissolved in phosphate buffer containing NaCl (pH 7.4; 27.4 mM Na<sub>2</sub>HPO<sub>4</sub>, 17.8 mM NaH<sub>2</sub>PO<sub>4</sub>, and 3M NaCl), and their turbidities at 400 nm were recorded upon increasing and decreasing temperatures (Fig. 1a, b and Table S2). The results revealed that these synthetic peptides 8a and 8b exhibited reversible LCST behaviour at concentrations of approximately 1-3.5 mM. The control peptide 11 showed the similar behaviour; however, it required significantly higher concentrations to initiate aggregation compared to the peptides 8a and 8b (Fig. 1c). Herein, the transition temperature  $(T_t)$  was defined as the temperature at which turbidity reached half of its maximum value during heating. The relationship between  $T_t$  and peptide concentration of 8a and 8b fitted well to a power function  $(T_t = aC^b)$ , while that of **11** followed a conventional logarithmic function  $(T_t = a \log(C) + b)$  as previously demonstrated for ELPs, where C is peptide concentration and a and b are constants (Fig. 1d). A clear correlation between



Scheme 4 (a) Chemical structure of short elastin-like peptide (sELP) 7, (b) synthesis of unnatural sELP analogues 8a and 8b by conventional peptide segment coupling.

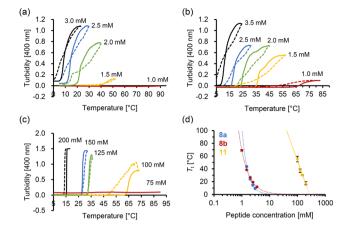


Fig. 1 Turbidity measurements of peptides 8a (a), 8b (b), and 11 (c), and the correlation between  $T_t$  and concentration (d). In (a)-(c), the solid and dashed lines represent the turbidity profiles upon heating and cooling, respectively. Measured  $T_{\rm t}$  values of peptides 8a and 8b were fitted with power functions, while that of 11 was fitted with conventional logarithmic functions.  $T_t$  values are shown with standard error.

peptide concentration and  $T_t$  demonstrated that the selfassembling capabilities of unnatural analogues 8a and 8b were approximately 100-fold stronger than that of reference peptide 11 (Fig. 1d). The reversibility of structural changes in 8a and 8b was roughly investigated by circular dichroism measurements (Fig. S2 and S3). Furthermore, the self-assembling behaviours were confirmed by bright-field microscopy (Fig. S4) and dynamic light scattering measurements (Fig. S5). Taken together, both peptides 8a and 8b were found to form aggregates above their respective  $T_t$  values. Ultra performance liquid chromatography (UPLC)-MS analysis showed peptides 8a and 8b were more hydrophobic than peptide 11 (Fig. S1), suggesting that the increased hydrophobicity of 8a and 8b may contribute to their enhanced self-assembling properties.

#### Conclusions

This study, although preliminary, developed a streamlined N-to-C peptide elongation method for synthesizing unnatural peptides using the ammonia-Ugi reaction. Starting from readily available N-protected amino acids, ketones, and α-isocyano ester as building blocks, a series of tripeptides 4a-4g were synthesized in high yields and with excellent stereochemical integrity. Notably, the sterically demanding residues such as Aib,  $Ac_5c$ , ( $\alpha$ Me)Hph, and Deg were constructed in situ from simple ketones, and successfully incorporated into peptides, overcoming limitations of conventional peptide synthesis. The tripeptides 4 were further elongated into pentapeptides 6 via the second ammonia-Ugi reaction, demonstrating the method's versatility. Interestingly, both 8a and 8b exhibited reversible LCST behaviour at approximately 100-fold lower concentrations than 11, due probably to the increased hydrophobicity. Future work will focus on synthesis of longer peptides and more sterically congested peptides using α-isocyano

 $\alpha,\alpha$ -disubstituted amino acids. Extensions in these directions will be reported from our laboratory in due course.

#### Author contributions

KT, conceptualization, investigation, methodology, and writing - original draft; KT, NT, YK, MU, MA, HK, investigation; HN and TN, supervision and writing - review & editing.

#### Conflicts of interest

There are no conflicts to declare.

# Data availability

The data supporting this article have been included as part of the supplementary information (SI). Supplementary information: experimental procedures, supplementary figures and tables, NMR spectra of all new compounds, and HPLC charts. See DOI: https://doi.org/10.1039/d5ob01834j.

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#### References

- 1 For selected reviews, see: S. Li, J. C. M. van Hest, R. Xing and X. Yan, Chem. Commun., 2025, 61, 13841; K. Sharma, K. K. Sharma, A. Sharma and R. Jain, Drug Discovery Today, 2023, 28, 103464; A. D. Cunningham, N. Qvit and D. Mochly-Rosen, Curr. Opin. Chem. Biol., 2017, 44, 59; L. Nevola and E. Giralt, Chem. Commun., 2015, 51, 3302.
- 2 For selected reviews, see: T. Misawa, N. Ohoka, M. Oba, H. Yamashita, M. Tanaka, M. Naito and Y. Demizu, Chem. Commun., 2019, 54, 7792; B. C. Doak, J. Zheng, D. Dobritzsch and J. Kihlberg, J. Med. Chem., 2016, 59, 2312
- 3 R. B. Merrifield, J. Am. Chem. Soc., 1963, 85, 2149.
- 4 F. Zieleniewski, D. N. Woolfson and J. Clayden, Chem. Commun., 2020, 56, 12049.
- 5 For recent examples, see: T. Liu, Z. Peng, M. Lai, L. Hu and J. Zhao, J. Am. Chem. Soc., 2024, 146, 4270; D. Uehara, S. Adachi, A. Tsubouchi, Y. Okada, V. V. Zhdankin,

- This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.
- A. Yoshimura and A. Saito, Chem. Commun., 2024, 60, 956; J. Yang, D. Zhang, Y. Chang, B. Zhang, P. Shen, C. Han and J. Zhao, Org. Chem. Front., 2024, 11, 5422; Z. Wang, X. Wang, P. Wang and J. Zhao, J. Am. Chem. Soc., 2021, 143, 10374.
- 6 M. Todorovic and D. M. Perrin, Pept. Sci., 2020, 112, e24210.
- 7 I. Ugi, Angew. Chem., Int. Ed. Engl., 1962, 1, 8; A. Dömling and I. Ugi, Angew. Chem., Int. Ed., 2000, 39, 3168.
- 8 For recent examples, see: J. Zhang, P. Yu, S.-Y. Li, H. Sun, S.-H. Xiang, J. Wang, K. N. Houk and B. Tan, Science, 2018, 361, eaas8707; K. Tomohara, N. Ohashi, T. Uchida and T. Nose, Sci. Rep., 2022, 12, 15568.
- 9 T. M. Vishwanatha, K. Kurpiewska, J. Kalinowska-Tłuścik and A. Dömling, J. Org. Chem., 2017, 82, 9585; A. L. Brown, Q. I. Churches and C. A. Hutton, J. Org. Chem., 2015, 80, 9831; S. de Castro, M.-J. Camarasa, J. Balzarini and S. Velázquez, Eur. J. Med. Chem., 2014, 83, 174; M. Abbas and L. A. Wessjohann, Org. Biomol. Chem., 2012, 10, 9330; S. Ackermann, H.-G. Lerchen, D. Häbich, A. Ullrich and U. Kazmaier, Beilstein J. Org. Chem., 2012, 8, 1652; R. Pick, M. Bauer, U. Kazmaier and C. Hebach, Synlett, 2005, 757; U. Kazmaier and C. Hebach, Synlett, 2003, 1591; M. Waki and J. Meienhofer, J. Am. Chem. Soc., 1977, 99, 6075.
- 10 I. P. Bhela, M. Serafini, E. D. Grosso, G. C. Tron and T. Pirali, Org. Lett., 2021, 23, 3610.
- 11 K. Tomohara, S. Kusaba, M. Masui, T. Uchida, H. Nambu and T. Nose, Org. Biomol. Chem., 2024, 22, 6999; K. Tomohara, T. Nose and H. Nambu, Synlett, 2025, 2671.
- 12 Y. Kondo, H. Morimoto and T. Ohshima, Synlett, 2024, 379.
- 13 K. Ghosh and W. D. Lubell, J. Pept. Sci., 2025, 31, e70019.
- 14 Lubell and co-workers reported inverse C-to-N peptide elongation by the ammonia-Ugi reaction. F. M. Mir, M. Crisma, C. Toniolo and W. D. Lubell, Chem. Sci., 2019, 10, 6908.
- 15 M. D. Zotti and J. Clayden, Org. Lett., 2019, 21, 2209.
- 16 Y. E. Jad, S. N. Khattab, B. G. de la Torre, T. Govender, H. G. Kruger, A. El-Faham and F. Albericio, Org. Biomol.

- Chem., 2014, 12, 8379; W. Van Den Nest, S. Yuval and F. Albericio, J. Pept. Sci., 2001, 7, 115; A. D. Fenza, M. Tancredi, C. Galoppini and P. Rovero, Tetrahedron Lett., 1998, 39, 8529.
- 17 M. V. Katti, R. Sami-Subbu, P. K. Ranjekar and V. S. Gupta, Protein Sci., 2000, 9, 1203.
- 18 R. M. Senior, G. L. Griffin, R. P. Mecham, D. S. Wrenn, K. U. Prasad and D. W. Urry, J. Cell Biol., 1984, 99, 870; D. W. Urry, J. Protein Chem., 1984, 3, 403.
- 19 J. A. M. Ramshaw, N. K. Shah and B. Brodsky, J. Struct. Biol., 1998, 122, 86.
- 20 C.-Z. Zhou, F. Confalonieri, M. Jacquet, R. Perasso, Z.-G. Li and J. Janin, Proteins, 2001, 44, 119.
- 21 S. Taniguchi, N. Watanabe, T. Nose and I. Maeda, J. Pept. Sci., 2016, 22, 36; I. Maeda, S. Taniguchi, N. Watanabe, A. Inoue, Y. Yamasaki and T. Nose, Protein Pept. Lett., 2015, 22, 934.
- 22 K. Suyama, E. N. Mai, I. Maeda and T. Nose, ACS Synth. Biol., 2025, 14, 2999; N. Tanaka, K. Suvama, K. Tomohara and T. Nose, Biomacromolecules, 2024, 25, 7156; N. Tanaka, K. Suyama, K. Tomohara, I. Maeda and T. Nose, J. Pept. Sci., 2023, 29, e3449; D. Tatsubo, K. Suyama, N. Sakamoto, K. Tomohara, S. Taniguchi, I. Maeda and T. Nose, Biochemistry, 2023, 62, 2559; S. Sumiyoshi, K. Suyama, Tanaka, T. Andoh, A. Nagata, K. Tomohara, S. Taniguchi, I. Maeda and T. Nose, Sci. Rep., 2022, 12, 19414; K. Suyama, M. Shimizu, I. Maeda and T. Nose, Biopolymers, 2022, 113, e23521; K. Suyama, M. Mawatari, D. Tatsubo, I. Maeda and T. Nose, ACS Omega, 2021, 6, 5705; K. Suyama, D. Tatsubo, W. Iwasaki, M. Miyazaki, Y. Kiyota, I. Takahashi, I. Maeda and T. Nose, Biomacromolecules, 2018, 19, 3201; D. Tatsubo, K. Suyama, M. Miyazaki, I. Maeda and T. Nose, Biochemistry, 2018, 57, 1582; K. Suyama, S. Taniguchi, D. Tatsubo, I. Maeda and T. Nose, J. Pept. Sci., 2016, 22, 236.
- 23 D. L. Barreiro, I. J. Minten, J. C. Thies and C. M. J. Sagt, ACS Biomater. Sci. Eng., 2023, 9, 3796.
- 24 D. E. Meyer and A. Chilkoti, Biomacromolecules, 2004, 5, 846.