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The ammonia-Ugi reaction employing ammonium carboxylates of *N*-protected amino acids (or peptides), ketones, and α -isocyano esters enabled N-to-C peptide elongation, together with *in situ* construction of α,α -disubstituted amino acid residues. This method offered an effective synthetic method of novel elastin-like short peptides, which exhibited highly potent self-assembling properties.

Peptides possess considerable potential both as drug delivery vehicles and therapeutic agents against pharmacologically diverse and otherwise intractable biological targets.¹ Incorporation of unnatural α,α -disubstituted amino acid residues into peptides can significantly impact their metabolic resistance, hydrophobicity, membrane permeability, biological activity and selectivity, and immunogenicity, ultimately enhancing the druggability of peptide-based therapeutics.²

Peptide synthesis is typically performed *via* stepwise coupling of *N*-protected amino acids to a growing peptide chain anchored on a solid support using condensation agents.³ Although such a solid-phase peptide synthesis (SPPS) works well with proteinogenic and other α -monosubstituted amino acids, it often results in poor yields with sterically hindered α,α -disubstituted amino acids, requiring excessive amounts of coupling agents under harsh conditions such as heating or microwave irradiation.⁴ Moreover, preparation of *N*-protected α,α -disubstituted amino acids generally requires multi-step and tedious processes. Alternatively, highly reactive condensation agents⁵ and catalytic reaction systems⁶ have offered efficient methods for unnatural peptide synthesis; however, these still use expensive unnatural α,α -disubstituted amino acids as substrates. These synthetic challenges have long

posed significant barriers to the discovery and development of bioactive peptides.

The ammonia-Ugi reaction, a variant of the Ugi reaction,^{7,8} is a four-component coupling reaction that involves ammonia, an aldehyde or a ketone, a carboxylic acid, and an isocyanide, enabling a straightforward synthesis of peptides.⁹ Although the ammonia-Ugi reaction had long been considered impractical and unsuccessful,¹⁰ we recently reported an efficient synthetic protocol of unnatural dipeptides using the ammonia-Ugi reaction: by stirring ammonium carboxylates derived from *N*-protected amino acids 1, ketones, and isocyanides in trifluoroethanol (TFE) at ambient temperature, a variety of dipeptides 2 were obtained in good yields (Scheme 1a).¹¹ Of note, these dipeptides 2 contained unnatural α,α -disubstituted amino acids, which were constructed *in situ* during the ammonia-Ugi reaction from readily available ketones as building blocks. Mechanistically, ammonium (NH_4^+) dissociates into NH_3 and H^+ , which in turn facilitates the thermodynamically unfavourable *N*-unsubstituted imine formation (Scheme 1a).¹² With its success, however, the products 2 were obtained as amides at their C-termini, rendering them unsuitable for further N-to-C peptide elongation. To the best of our knowledge, neither the ammonia-Ugi reaction nor the Ugi reaction has ever been applied for N-to-C peptide chain elongation.^{13,14} Here, this study proposes a novel strategy for N-to-C elongation of unnatural peptides by the ammonia-Ugi reaction employing α -isocyano ester 3 as an elongation unit (Scheme 1b). The expected ammonia-Ugi adducts 4 possess ester moieties at their C-termini, providing a versatile platform for subsequent peptide chain elongation.

Using a commercially available ethyl isocyanoacetate (3, CN-Gly-OEt), a series of unnatural tripeptides 4 were synthesized (Scheme 2). The key ammonium carboxylates of *N*-protected amino acids 1 were prepared by stirring *N*-protected amino acids with aqueous ammonia in acetonitrile or THF at 0 °C (Table S1).¹¹ Then, the ammonia-Ugi reaction using Boc-Phe-ONH₄, cyclopentanone, and CN-Gly-OEt (3) afforded the tripeptide Boc-Phe-Ac₅c-Gly-OEt (4a) in

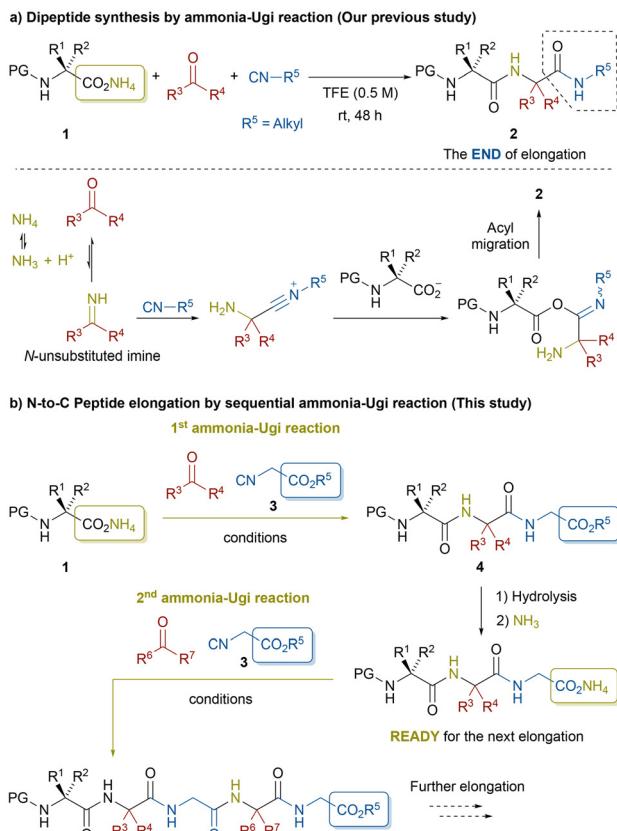
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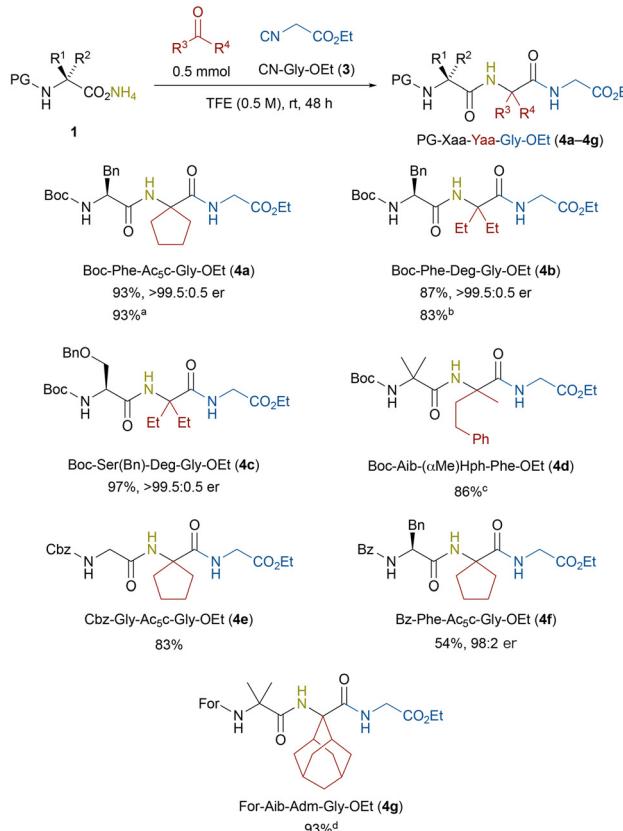
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Scheme 1 (a) Dipeptide synthesis by ammonia-Ugi reaction, (b) N-to-C peptide elongation by sequential ammonia-Ugi reaction (this study).

93% yield, with no detectable racemization of the chiral α -carbon of phenylalanine ($>99.5:0.5$ er). Herein, the unnatural 1-aminocyclopentane-1-carboxylic acid (Ac₅c) residue in **4a** was constructed *in situ* from cyclopentanone as a substrate. In general, diethylglycine (Deg) is a challenging substrate to be incorporated into peptides under conventional SPPS conditions because of the steric hindrance,¹⁵ whereas the present ammonia-Ugi reaction successfully delivered Boc-Phe-Deg-Gly-OEt (**4b**) in 87% yield. Again, no racemization was observed during this process ($>99.5:0.5$ er). The chiral α -carbon of serine is known to be susceptible to racemization during peptide synthesis;¹⁶ however, the present reaction conditions afforded **4c** in an excellent yield (97%) with a perfect stereochemical integrity ($>99.5:0.5$ er). Even the sterically demanding Boc-Aib was compatible with the present reaction conditions, affording the sterically congested tripeptide **4d**, composed of contiguous α,α -disubstituted amino acids [α -aminoisobutyric acid (Aib) and α -methylhomophenylalanine [α (Me)Hph]], in 78% yield. In addition to Boc, various *N*-protecting groups, including Cbz, Bz, and formyl (For), were well tolerated, giving the tripeptides **4e**, **4f**, and **4g** in 83%–99% yields. Unfortunately, the stereochemical integrity of Bz-Phe was slightly lost (98:2 er), whereas that of Ac-Phe remained intact under the same conditions.¹¹ It is worth noting that the *N*-formyl group in **4g** can

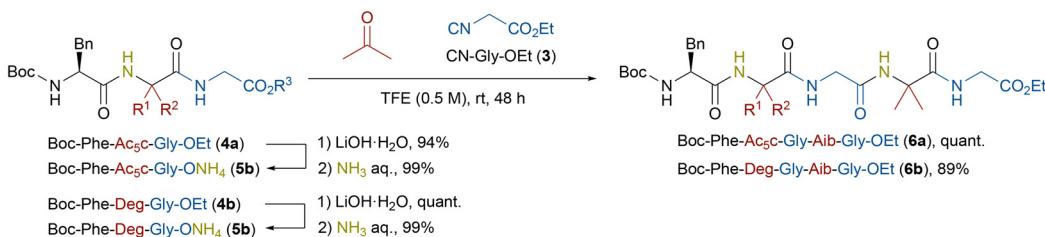


Scheme 2 Synthesis of tripeptides **4a–4g** by ammonia-Ugi reaction using ammonium carboxylates of *N*-protected amino acids. ^a 5.4 mmol, ^b 3.5 mmol, ^c 1 mmol, ^d 4.5 mmol.

serve as a precursor of an isocyano group, potentially enabling inverse C-to-N peptide elongation.¹⁴ Scale-up experiments (up to 5.4 mmol) proceeded smoothly without any detrimental effects on the reaction system, providing the corresponding products **4a**, **4b**, and **4g** in excellent yields (83%–93%). Overall, the ammonia-Ugi reaction using α -isocyano ester and *N*-protected amino acids enabled the efficient synthesis of unnatural tripeptides containing α,α -disubstituted amino acids. These tripeptides possess ester groups at their C-termini, offering a platform for subsequent N-to-C peptide chain elongation.

Peptides containing glycine are frequently found in biomaterials¹⁷ such as elastin,¹⁸ collagen,¹⁹ and silk fibroin.²⁰ Owing to glycine's small size and conformational flexibility, it plays a crucial role in modulating peptide structure and function. The development of efficient synthetic strategies for unnatural analogues of glycine-rich peptides is therefore of significant interest in both synthetic and biomedical research. In this study, we developed an efficient N-to-C peptide elongation method for synthesizing glycine-containing unnatural peptides by the ammonia-Ugi reaction employing CN-Gly-OEt (3) (Scheme 3). The C-terminal ester moieties of tripeptides **4a** and **4b** were hydrolyzed under basic conditions and subsequently treated with aqueous ammonia to give the corresponding ammonium carboxylates **5a** and **5b** in excellent yields. Under the present





Scheme 3 Synthesis of pentapeptides **6a** and **6b** by ammonia-Ugi reaction starting from ammonium carboxylates of *N*-protected tripeptides **4a** and **4b**.

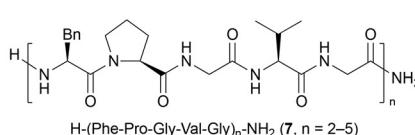
ammonia-Ugi reaction conditions, both tripeptides were successfully elongated into the pentapeptides Boc-Phe-Xaa-Gly-Aib-Gly-OEt **6a** and **6b** in good yields, together with *in situ* construction of Aib residue from acetone. The resulting peptides again possess ester moieties at C-termini, potentially providing a versatile platform for further N-to-C elongation or other chemical modifications. Totally, starting from *N*-protected amino acids **1**, the first ammonia-Ugi reaction furnished tripeptides **4** (Scheme 2), and the second ammonia-Ugi reaction extended them to pentapeptides **6** (Scheme 3). Each step elongated peptides by two amino acid residues at a time while constructing α,α -disubstituted amino acids *in situ*. This streamlined approach requires no condensation agents, thus providing an environmentally friendly synthetic method of glycine-containing unnatural peptides.

The obtained pentapeptides **6a** and **6b** share the repeating amino acid sequence of a short elastin-like peptide (sELP) H-(Phe-Pro-Gly-Val-Gly)_n-NH₂ (**7**, Scheme 4a).²¹ sELP exhibits reversible lower critical solution temperature (LCST)-type behaviour, being soluble at low temperatures and insoluble at high temperatures.²² Such reversible temperature-responsive and self-assembling properties make sELP a promising candidate for drug delivery applications. The development of unnatural analogs with enhanced self-assembling properties is therefore of considerable interest. Herein, we designed a novel unnatural analogue, H-(Phe-Ac_{5c}-Gly-Aib-Gly)₂-NH₂ (**8a**), by modifying the original sELP sequence (Scheme 4b). Specifically, the Pro-2 in **7** was replaced with Ac_{5c}, a noncanonical amino acid structurally related to Pro, and the Val-4, located at a guest position in the original sequence,²³ was substituted with Aib, a noncanonical amino acid analogous to Val.

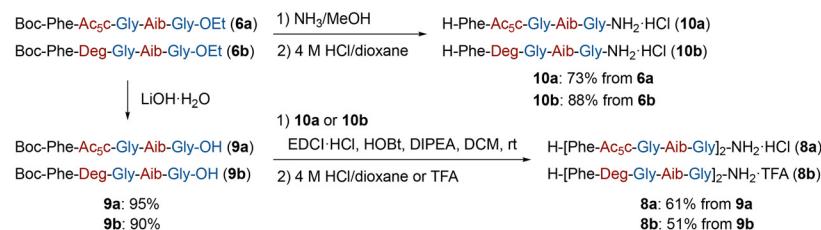
The decapeptide **8a** was synthesized in a good yield *via* segment coupling between Boc-Phe-Ac_{5c}-Gly-Aib-Gly-OH (**9a**) and H-Phe-Ac_{5c}-Gly-Aib-Gly-NH₂ (**10a**) under standard condensation conditions, followed by Boc deprotection under acidic conditions. We also designed and synthesized a structurally related analogue, H-(Phe-Deg-Gly-Val-Gly)₂-NH₂ (**8b**), in which Ac_{5c} residue in **8a** was replaced with Deg, a noncyclic analogue of Ac_{5c}. Both **8a** and **8b** were obtained as TFA salts in pure form after purification by reversed-phase (RP)-HPLC (Fig. S1). Additionally, we designed and synthesized H-(Phe-Pro-Gly-Aib-Gly)₂-NH₂ (**11**) as an Aib analogue of the original sELP **7** *via* a standard SPPS protocol (see, SI).

The self-assembling properties of peptides **8a**, **8b**, and **11** were then investigated under buffered aqueous conditions. Each peptide was dissolved in phosphate buffer containing NaCl (pH 7.4; 27.4 mM Na₂HPO₄, 17.8 mM NaH₂PO₄, and 3M NaCl), and their turbidities at 400 nm were recorded upon increasing and decreasing temperatures (Fig. 1a, b and Table S2). The results revealed that these synthetic peptides **8a** and **8b** exhibited reversible LCST behaviour at concentrations of approximately 1–3.5 mM. The control peptide **11** showed the similar behaviour; however, it required significantly higher concentrations to initiate aggregation compared to the peptides **8a** and **8b** (Fig. 1c). Herein, the transition temperature (*T_t*) was defined as the temperature at which turbidity reached half of its maximum value during heating. The relationship between *T_t* and peptide concentration of **8a** and **8b** fitted well to a power function ($T_t = aC^b$), while that of **11** followed a conventional logarithmic function ($T_t = a \log(C) + b$) as previously demonstrated for ELPs, where *C* is peptide concentration and *a* and *b* are constants (Fig. 1d).²⁴ A clear correlation between

a) Short elastin-like peptide (sELP) with self-assembling ability



b) Synthesis of elastin-like peptides by peptide fragment coupling



Scheme 4 (a) Chemical structure of short elastin-like peptide (sELP) **7**, (b) synthesis of unnatural sELP analogues **8a** and **8b** by conventional peptide segment coupling.



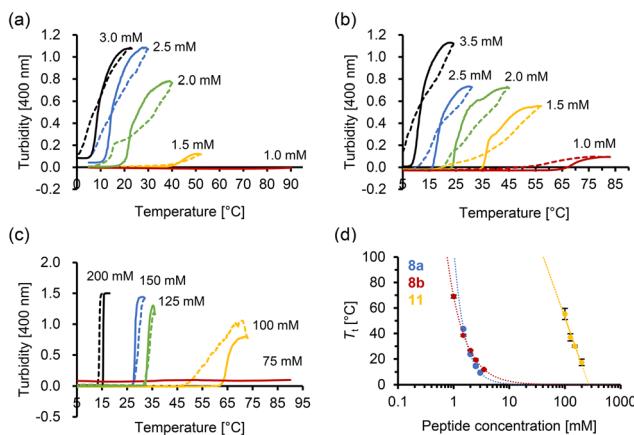


Fig. 1 Turbidity measurements of peptides **8a** (a), **8b** (b), and **11** (c), and the correlation between T_t and concentration (d). In (a)–(c), the solid and dashed lines represent the turbidity profiles upon heating and cooling, respectively. Measured T_t values of peptides **8a** and **8b** were fitted with power functions, while that of **11** was fitted with conventional logarithmic functions. T_t values are shown with standard error.

peptide concentration and T_t demonstrated that the self-assembling capabilities of unnatural analogues **8a** and **8b** were approximately 100-fold stronger than that of reference peptide **11** (Fig. 1d). The reversibility of structural changes in **8a** and **8b** was roughly investigated by circular dichroism measurements (Fig. S2 and S3). Furthermore, the self-assembling behaviours were confirmed by bright-field microscopy (Fig. S4) and dynamic light scattering measurements (Fig. S5). Taken together, both peptides **8a** and **8b** were found to form aggregates above their respective T_t values. Ultra performance liquid chromatography (UPLC)-MS analysis showed peptides **8a** and **8b** were more hydrophobic than peptide **11** (Fig. S1), suggesting that the increased hydrophobicity of **8a** and **8b** may contribute to their enhanced self-assembling properties.

Conclusions

This study, although preliminary, developed a streamlined N-to-C peptide elongation method for synthesizing unnatural peptides using the ammonia-Ugi reaction. Starting from readily available *N*-protected amino acids, ketones, and α -isocyano ester as building blocks, a series of tripeptides **4a**–**4g** were synthesized in high yields and with excellent stereochemical integrity. Notably, the sterically demanding residues such as Aib, Ac₅c, (α Me)Hph, and Deg were constructed *in situ* from simple ketones, and successfully incorporated into peptides, overcoming limitations of conventional peptide synthesis. The tripeptides **4** were further elongated into pentapeptides **6** *via* the second ammonia-Ugi reaction, demonstrating the method's versatility. Interestingly, both **8a** and **8b** exhibited reversible LCST behaviour at approximately 100-fold lower concentrations than **11**, due probably to the increased hydrophobicity. Future work will focus on synthesis of longer peptides and more sterically congested peptides using α -isocyano

α,α -disubstituted amino acids. Extensions in these directions will be reported from our laboratory in due course.

Author contributions

KT, conceptualization, investigation, methodology, and writing – original draft; KT, NT, YK, MU, MA, HK, investigation; HN and TN, supervision and writing – review & editing.

Conflicts of interest

There are no conflicts to declare.

Data availability

The data supporting this article have been included as part of the supplementary information (SI). Supplementary information: experimental procedures, supplementary figures and tables, NMR spectra of all new compounds, and HPLC charts. See DOI: <https://doi.org/10.1039/d5ob01834j>.

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References

- For selected reviews, see: S. Li, J. C. M. van Hest, R. Xing and X. Yan, *Chem. Commun.*, 2025, **61**, 13841; K. Sharma, K. K. Sharma, A. Sharma and R. Jain, *Drug Discovery Today*, 2023, **28**, 103464; A. D. Cunningham, N. Qvit and D. Mochly-Rosen, *Curr. Opin. Chem. Biol.*, 2017, **44**, 59; L. Nevola and E. Giralt, *Chem. Commun.*, 2015, **51**, 3302.
- For selected reviews, see: T. Misawa, N. Ohoka, M. Oba, H. Yamashita, M. Tanaka, M. Naito and Y. Demizu, *Chem. Commun.*, 2019, **54**, 7792; B. C. Doak, J. Zheng, D. Dobritzsch and J. Kihlberg, *J. Med. Chem.*, 2016, **59**, 2312.
- R. B. Merrifield, *J. Am. Chem. Soc.*, 1963, **85**, 2149.
- F. Zieleniewski, D. N. Woolfson and J. Clayden, *Chem. Commun.*, 2020, **56**, 12049.
- For recent examples, see: T. Liu, Z. Peng, M. Lai, L. Hu and J. Zhao, *J. Am. Chem. Soc.*, 2024, **146**, 4270; D. Uehara, S. Adachi, A. Tsubouchi, Y. Okada, V. V. Zhdankin,



A. Yoshimura and A. Saito, *Chem. Commun.*, 2024, **60**, 956; J. Yang, D. Zhang, Y. Chang, B. Zhang, P. Shen, C. Han and J. Zhao, *Org. Chem. Front.*, 2024, **11**, 5422; Z. Wang, X. Wang, P. Wang and J. Zhao, *J. Am. Chem. Soc.*, 2021, **143**, 10374.

6 M. Todorovic and D. M. Perrin, *Pept. Sci.*, 2020, **112**, e24210.

7 I. Ugi, *Angew. Chem., Int. Ed. Engl.*, 1962, **1**, 8; A. Dömling and I. Ugi, *Angew. Chem., Int. Ed.*, 2000, **39**, 3168.

8 For recent examples, see: J. Zhang, P. Yu, S.-Y. Li, H. Sun, S.-H. Xiang, J. Wang, K. N. Houk and B. Tan, *Science*, 2018, **361**, eaas8707; K. Tomohara, N. Ohashi, T. Uchida and T. Nose, *Sci. Rep.*, 2022, **12**, 15568.

9 T. M. Vishwanatha, K. Kurpiewska, J. Kalinowska-Tłuścik and A. Dömling, *J. Org. Chem.*, 2017, **82**, 9585; A. L. Brown, Q. I. Churches and C. A. Hutton, *J. Org. Chem.*, 2015, **80**, 9831; S. de Castro, M.-J. Camarasa, J. Balzarini and S. Velázquez, *Eur. J. Med. Chem.*, 2014, **83**, 174; M. Abbas and L. A. Wessjohann, *Org. Biomol. Chem.*, 2012, **10**, 9330; S. Ackermann, H.-G. Lerchen, D. Häbich, A. Ullrich and U. Kazmaier, *Beilstein J. Org. Chem.*, 2012, **8**, 1652; R. Pick, M. Bauer, U. Kazmaier and C. Hebach, *Synlett*, 2005, 757; U. Kazmaier and C. Hebach, *Synlett*, 2003, 1591; M. Waki and J. Meienhofer, *J. Am. Chem. Soc.*, 1977, **99**, 6075.

10 I. P. Bhela, M. Serafini, E. D. Grossi, G. C. Tron and T. Pirali, *Org. Lett.*, 2021, **23**, 3610.

11 K. Tomohara, S. Kusaba, M. Masui, T. Uchida, H. Nambu and T. Nose, *Org. Biomol. Chem.*, 2024, **22**, 6999; K. Tomohara, T. Nose and H. Nambu, *Synlett*, 2025, 2671.

12 Y. Kondo, H. Morimoto and T. Ohshima, *Synlett*, 2024, 379.

13 K. Ghosh and W. D. Lubell, *J. Pept. Sci.*, 2025, **31**, e70019.

14 Lubell and co-workers reported inverse C-to-N peptide elongation by the ammonia-Ugi reaction. F. M. Mir, M. Crisma, C. Toniolo and W. D. Lubell, *Chem. Sci.*, 2019, **10**, 6908.

15 M. D. Zotti and J. Clayden, *Org. Lett.*, 2019, **21**, 2209.

16 Y. E. Jad, S. N. Khattab, B. G. de la Torre, T. Govender, H. G. Kruger, A. El-Faham and F. Albericio, *Org. Biomol. Chem.*, 2014, **12**, 8379; W. Van Den Nest, S. Yuval and F. Albericio, *J. Pept. Sci.*, 2001, **7**, 115; A. D. Fenza, M. Tancredi, C. Galoppini and P. Rovero, *Tetrahedron Lett.*, 1998, **39**, 8529.

17 M. V. Katti, R. Sami-Subbu, P. K. Ranjekar and V. S. Gupta, *Protein Sci.*, 2000, **9**, 1203.

18 R. M. Senior, G. L. Griffin, R. P. Mecham, D. S. Wrenn, K. U. Prasad and D. W. Urry, *J. Cell Biol.*, 1984, **99**, 870; D. W. Urry, *J. Protein Chem.*, 1984, **3**, 403.

19 J. A. M. Ramshaw, N. K. Shah and B. Brodsky, *J. Struct. Biol.*, 1998, **122**, 86.

20 C.-Z. Zhou, F. Confalonieri, M. Jacquet, R. Perasso, Z.-G. Li and J. Janin, *Proteins*, 2001, **44**, 119.

21 S. Taniguchi, N. Watanabe, T. Nose and I. Maeda, *J. Pept. Sci.*, 2016, **22**, 36; I. Maeda, S. Taniguchi, N. Watanabe, A. Inoue, Y. Yamasaki and T. Nose, *Protein Pept. Lett.*, 2015, **22**, 934.

22 K. Suyama, E. N. Mai, I. Maeda and T. Nose, *ACS Synth. Biol.*, 2025, **14**, 2999; N. Tanaka, K. Suyama, K. Tomohara and T. Nose, *Biomacromolecules*, 2024, **25**, 7156; N. Tanaka, K. Suyama, K. Tomohara, I. Maeda and T. Nose, *J. Pept. Sci.*, 2023, **29**, e3449; D. Tatsumi, K. Suyama, N. Sakamoto, K. Tomohara, S. Taniguchi, I. Maeda and T. Nose, *Biochemistry*, 2023, **62**, 2559; S. Sumiyoshi, K. Suyama, N. Tanaka, T. Andoh, A. Nagata, K. Tomohara, S. Taniguchi, I. Maeda and T. Nose, *Sci. Rep.*, 2022, **12**, 19414; K. Suyama, M. Shimizu, I. Maeda and T. Nose, *Biopolymers*, 2022, **113**, e23521; K. Suyama, M. Mawatari, D. Tatsumi, I. Maeda and T. Nose, *ACS Omega*, 2021, **6**, 5705; K. Suyama, D. Tatsumi, W. Iwasaki, M. Miyazaki, Y. Kiyota, I. Takahashi, I. Maeda and T. Nose, *Biomacromolecules*, 2018, **19**, 3201; D. Tatsumi, K. Suyama, M. Miyazaki, I. Maeda and T. Nose, *Biochemistry*, 2018, **57**, 1582; K. Suyama, S. Taniguchi, D. Tatsumi, I. Maeda and T. Nose, *J. Pept. Sci.*, 2016, **22**, 236.

23 D. L. Barreiro, I. J. Minten, J. C. Thies and C. M. J. Sagt, *ACS Biomater. Sci. Eng.*, 2023, **9**, 3796.

24 D. E. Meyer and A. Chilkoti, *Biomacromolecules*, 2004, **5**, 846.