






Cite this: DOI: 10.1039/d5ob01802a

Synthesis and evaluation of carbagalactosyl 1,2-aziridines and -epoxides as glycosidase inhibitors

Yuqing Tian, Hong-Ming Chen, Lyann Sim, Seyed A. Nasseri,  Jolene P. Reid  and Stephen G. Withers *

The natural product cyclophellitol is a cyclitol epoxide that mimics a β -glucoside and functions as a mechanism-based inhibitor for retaining β -glucosidases by forming a stable covalent intermediate with the enzyme's catalytic nucleophile. The epoxide is located across the carbons equivalent to C1 and O5 of glucose, and multiple congeners have been synthesised, as well as the corresponding aziridines, and shown to inactivate their cognate glycosidases. In a quest for a potent covalent inactivator of an α -galactosaminidase that is used in the conversion of A type blood to O type through modification of the antigen, we synthesised the galacto-configured 1,2- α -aziridine in which the aziridine ring is located across the carbons equivalent to C1 and C2. While neither this nor the corresponding epoxide inactivated our enzyme or any of the α -galactosidases tested, the β -configured analogues were shown by mass spectrometry to function as covalent inactivators of several β -glycosidases and their inactivation behaviour characterised kinetically. Molecular modeling indicates that 1,2- α -aziridines adopt an unfavorable conformation in solution, preventing binding to the enzyme active site. The synthesis of these compounds and their activities as inhibitors and inactivators of glycosidases of different families will direct future attempts to develop inactivators beyond those of the more common cyclophellitol class.

Received 14th November 2025,
 Accepted 15th December 2025

DOI: 10.1039/d5ob01802a

rsc.li/obc

Introduction

The antigens that define our ABO blood type on the surface of our red blood cells and tissues are oligosaccharides based upon the fucosyl- α -1,2-galactose (Fuc- α -1,2-Gal) terminal disaccharide of the H antigen found on O type cells. Extension of this structure with *N*-acetylgalactosamine attached α -1,3 to the galactose generates the A antigen, while equivalent extension with galactose yields the B-antigen. The ABO type of blood or organs must be carefully matched between donor and recipient to avoid serious immune responses due to anti-A or anti-B antibodies in non-matched recipient blood. The only exceptions are O type red blood cells or organs which can be donated to persons of essentially all types since antibodies against the H antigen are not present in A or B type persons. One consequence of this is that O-type blood and organs are always in short supply.

One approach to improving the supply of O type blood and organs might be to enzymatically convert the A or B antigen to the H antigen using suitably active and specific glycosidases.

Our search of human gut microbiome-derived metagenomic libraries for enzymes that will remove the terminal GalNAc of the A-antigen recently uncovered a pair of enzymes that work efficiently together. The first of these is a highly specific *N*-acetylgalactosamine deacetylase, which forms a terminal galactosamine that is then removed by a highly efficient α -galactosaminidase.¹ Interestingly this latter enzyme, *FpGalNase*, also has a weak α -galactosidase activity. As part of our characterisation of this enzyme we solved its three-dimensional structure by X-ray crystallography and showed that the active site contains a cobalt that coordinates the substrate amine moiety.² Most other enzymes of the CAZy GH36 family to which this enzyme belongs are α -galactosidases that have very similar active sites, but do not contain a metal.³ Enzymes from this family carry out hydrolysis with net retention of anomeric configuration through a β -glycosyl enzyme intermediate.⁴

In order to better characterise active site interactions in this enzyme, with a view to engineering its activity, we were interested in developing inhibitors that form a stable covalent intermediate that could be structurally characterised. A strategy used widely for trapping intermediates on glycosidases has involved 2-fluorosugars containing a reactive leaving group.⁵ The fluorine inductively destabilises the oxocarbenium ion

Department of Chemistry, University of British Columbia, Vancouver, V6T 1Z1, Canada. E-mail: withers@chem.ubc.ca



transition state, slowing both formation and hydrolysis of this intermediate while the reactive leaving group ensures relatively rapid intermediate formation. However, that strategy is not applicable here due to the need for the 2-amine substituent and also because the strategy does not work well for α -glycosidases.⁶ While an equivalent approach using a 5-fluoroglycosyl fluoride might seem attractive,^{7,8} the presence of a free amine at the 2-position would likely render it unstable or at least very synthetically challenging.

An alternative approach is to use suitably substituted cyclophellitol epoxides or aziridines of the cyclophellitol family. Cyclophellitol is an epoxide-containing natural product that functions as a time-dependent covalent mechanism-based inhibitor of β -glucosidases⁹ (Fig. 1A). Many epimers of this molecule have been synthesised and shown, in most cases, to function as covalent inhibitors of their cognate glycosidases, including α -glycosidases. The equivalent aziridines (not yet known as natural products) have also been synthesised and shown to be similarly, indeed often more, potent as covalent inhibitors. However inhibitors of this general class will not form covalent bonds with glycosidases employing a neighbouring group assistance mechanism, such as those of GH20.¹⁰ Considerable work has been performed on these classes of molecules, primarily by the Overkleeft group in conjunction with the group of Davies. References to this work are found in the recent review.¹¹

The epoxide or aziridine in these “cyclophellitols” is located across carbons equivalent to C1 and O5 in the sugar being mimicked. We questioned whether, for our enzyme, a particularly effective inactivator might be an “ α ”-aziridine that bridged the carbons equivalent to C1 and C2 of galactosamine.

In that case the nucleophilic carboxylate of the enzyme should attack C1 in the normal manner, and the amine moiety would end up coordinated by those residues ordinarily ligating the normal substrate amine (Fig. 1B). Compounds of this class in the *gluco* configuration have been synthesised previously (Shing,¹² Overkleeft¹³) and generally found to be poorer inhibitors. Disaccharide versions containing α - and β -configured epoxides and aziridines were also synthesised as part of a mechanistic investigation of a GH99 *endo*-mannosidase, providing valuable support for an epoxide intermediate in the enzymatic reaction.^{14,15} However, no such studies had been performed with hexosaminidases which, as noted above, would seem to be particularly suitable target enzymes for this aziridine class of reagent. In this study we synthesise a full set of *galacto* configured “ α ” and “ β ”-1,2-epoxides and aziridines and explore their potential as inactivators of an α -galactosaminidase and a series of α - and β -galactosidases.

Results and discussion

Synthetic procedures

The most logical route for synthesis of the 1,2-epoxide and aziridine analogues of *galacto*-configured “cyclophellitol” involves preparation of the alkene (7) shown below as a key intermediate. Various routes to this compound class are published. The Shing synthesis accessed the alkene in 19% yield *via* a ten-step route from quinic acid.⁸ Overkleeft, on the other hand, used an eight-step route involving Claisen rearrangement of a protected glycal.⁹ A related Claisen rearrangement route was used for the disaccharide inhibitor by the Sollogoub group.¹⁴ We chose to access the “galacto” version through an asymmetric Diels Alder approach developed by Nishikawa.¹⁶ In this way (Scheme 1), the key intermediate (7) was synthesized in seven steps in 13% overall yield.

Epoxidation of the double bond in 7 was performed with *m*-CPBA in DCM, affording “ α ”-epoxide **8a** in 44% and “ β ”-epoxide **8b** in 50% yield, respectively.¹³ At this stage, it was very difficult to determine the configurations of the two anomers by NMR since the chemical shifts, coupling constants and NOEs are very similar for the two compounds thus we continued the synthesis with each compound separately expecting to assign configurations later. Deprotection of the sample comprising compound **8a** was achieved by treatment with Pd/C, affording **9a** in 58% yield, along with by-products of the over-reduced triol and the methyl ether formed upon attack on the epoxide by methanol. By contrast, only the over-reduced by-products were detected when compound **8b** was treated under the same conditions. In order to access product **9b** the reduction was performed using Pearlman's catalyst Pd(OH)₂/C in a mixed solvent system (MeOH/H₂O/dioxane), leading to the product with comparable yields to that of the other isomer.¹⁷

Azidolysis of the benzyl-protected α -epoxide **8a** (Scheme 2) was achieved by stirring the compound with sodium azide for 48 h at 100 °C, affording **10a** as the major product in 55% yield along with **10a'** as the minor product in 9% yield.

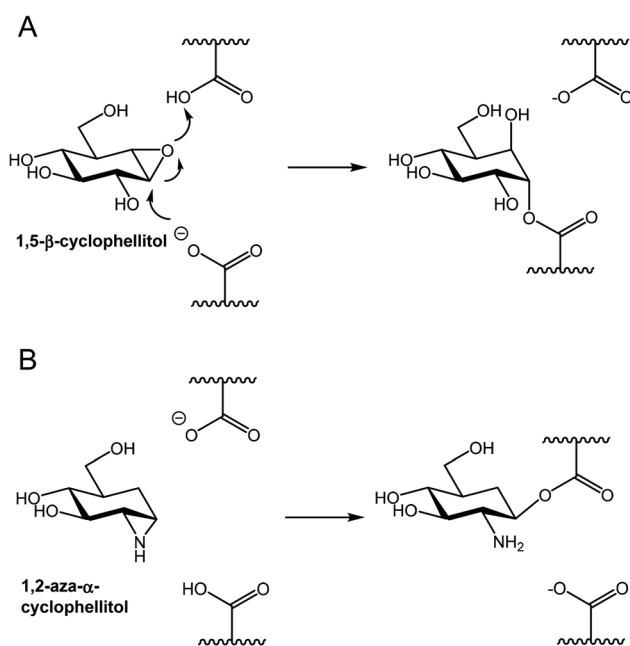
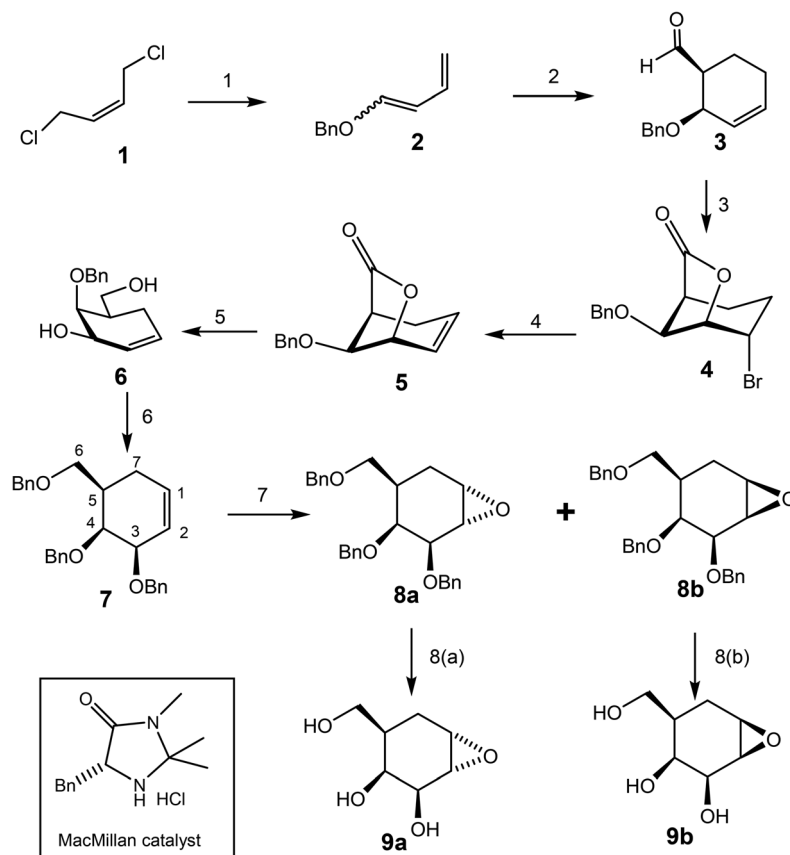


Fig. 1 Examples of (A) 1,5-cyclophellitol and (B) 1,2-aza- α -cyclophellitol acting as mechanism-based inhibitors of, respectively, retaining β - and α -glycosidases.





Scheme 1 Synthesis of epoxides (**9a** & **9b**) Reagents and conditions: (1) DMSO, 4,4'-thiobis(2-*tert*-butyl-5-methylphenol), NaH, BnOH, rt–50 °C, 82%. (2) MeOH, H₂O, Acrolein, MacMillan catalyst, –10 °C, 22 h, 38%. (3) DMF, Oxone, rt; DCM, NaHCO₃, NBS, rt, 63%. (4) THF, DBU, 50 °C, 86%. (5) Et₂O, LiAlH₄, –78 to –20 °C, 92%. (6) DMF, NaH, BnBr, 0 °C–rt, 92%. (7). DCM, *m*-CPBA, 0 °C–rt, **8a**: 44%; **8b**: 50%. (8(a)) MeOH/EtOAc = 5/1, Pd/C (10%), H₂, rt, 24 h, 58%. (8(b)). MeOH/H₂O/dioxane = 1/1/1, Pd(OH)₂/C (20% wet), H₂, rt, 2 h, 52%.

Treatment of β -epoxide **8b** with sodium azide under similar conditions (stirring for 22 h at 100 °C), afforded **10b** in 92% yield.^{17,18} Attack of azide on the α -epoxide **8a** necessarily occurred from the top face, which is very hindered. Indeed hindrance was such that ring opening occurred in a trans diequatorial fashion *via* attack at C1 rather than di-axial through attack at C-2. By contrast attack of azide on the “ β ”-epoxide ring of **8b** occurred at C-1 from the less hindered bottom face resulting in the trans-diaxial compound as the sole significant product. Notably this reaction was complete in a shorter period of time, consistent with the lesser hindrance. These reaction outcomes support the assignments of **8a** and **8b**.

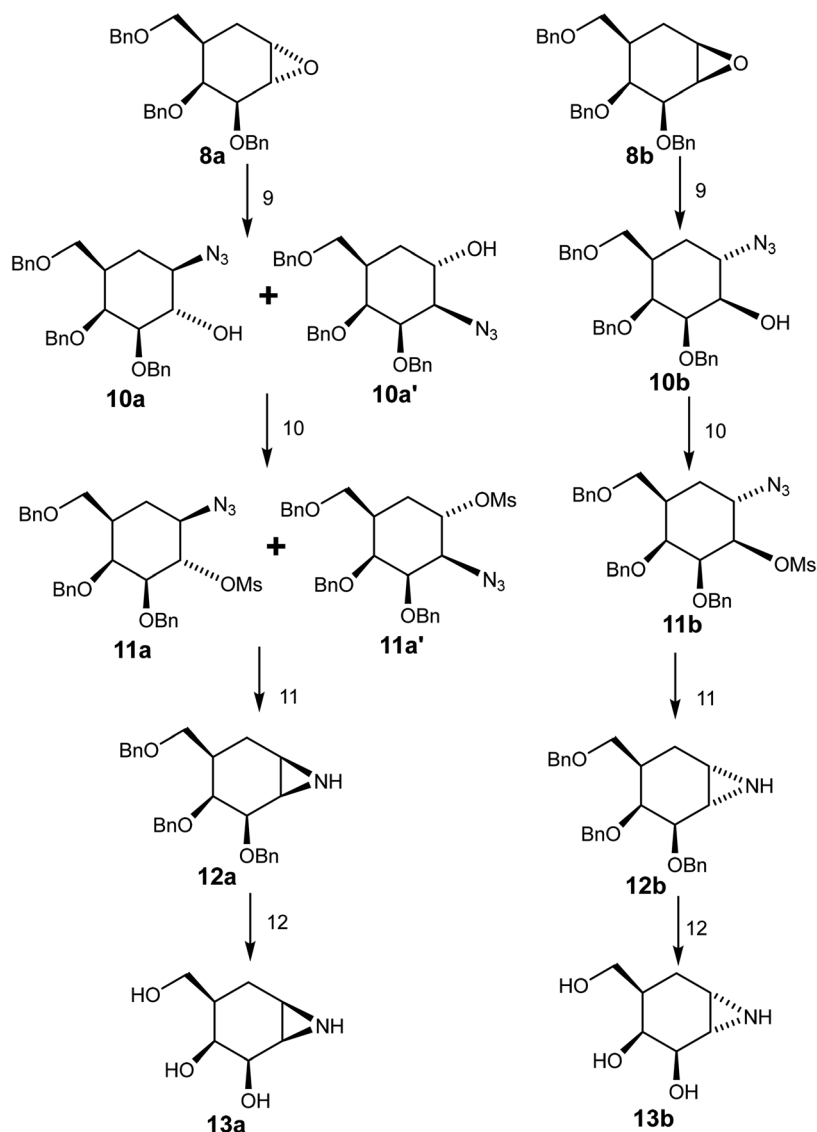
Reaction of compounds **10a**, **10a'** and **10b** with methanesulfonyl chloride afforded the desired products **11a**, **11a'** and **11b** in good yields.¹⁹ The NMR spectra of these compounds provided further support for the assignments of **8a** and **8b** since the coupling constant $J_{2,3}$ for **11a** was 9.9 Hz (trans) while that for **11b** was 2.8 Hz (*cis*). Selective reduction of the azide in compounds **11a** and **11a'** with LiAlH₄ followed by ring closure through *in situ* intramolecular displacement of the amino mesylates gave the same product **12a**, the “ β ” aziridine. Similarly, **11b** was converted to the “ α ” aziridine **12b**. Removal of the benzyl protecting group was accomplished using a Birch

reduction (Na, NH₃) yielding the target aziridines **13a** and **13b** (Scheme 2), both of which proved to be unstable under neutral conditions. For stable storage it proved necessary to dissolve them in a basic (NaOH or NH₄OH) aqueous solution and then lyophilize them.

Inactivation of galactosidases

The epoxides and aziridines synthesised (**9a**, **9b**, **13a**, **13b**) were tested as inactivators and as reversible inhibitors for a range of α - and β -galactosidases from different families, with the results shown in Table 1 and Fig. S15–S19. Testing for inactivation was carried out by incubating the relevant reagent with each enzyme individually, removing aliquots at time intervals, and assaying for residual enzyme activity using the substrates indicated. To our considerable disappointment, none of the “ α ”-1,2-epoxides or aziridines functioned as time-dependent inactivators. While it would be tempting to ascribe this to the need for less favourable 1,2-di-equatorial ring opening in these cases, it is important to note that the equivalent “ α ”-1,5a-epoxides and aziridines, which would also need to open in a di-equatorial fashion, are indeed inactivators of α -glycosidases.¹¹ Other factors must therefore be considered.





Scheme 2 Synthesis of aziridines (**13a** & **13b**) Reagents and conditions: 9. DMF, LiClO₄, NaN₃, 100 °C, 48 h, **10a**: 55%, **10a'**: 9%; **10b**, 22 h, 92%. 10. THF (dry), MsCl, TEA, rt, **11a**: 100%; **11a'**: 94%; **11b**: 100%. 11. THF (dry), LiAlH₄, 0 °C–rt, **12a**: 50%; **12b**: 84%. 12. NH₃, Na, THF (dry), –78 °C; **13a**: 84%; **13b**: 92%.

Table 1 Testing for inhibition of galactosaminidase and galactosidases

Enzyme	Family	Substrate	Epoxide		Aziridine	
			α	β	α	β
<i>F. plautii</i> α-galactosaminidase (<i>FpGalNase</i>) ²	GH36	GalN-α-MU	No	No	No	Reversible
<i>T. maritima</i> α-galactosidase (<i>TmGalA</i>) ⁴	GH36	Gal-α-pNP	No	No	No	Reversible
Coffee bean α-galactosidase ²⁰	GH27	Gal-α-pNP	No	No	No	No
<i>Agrobacter</i> sp. β-glucosidase/galactosidase (<i>Abg</i>) ²¹	GH1	Glc-β-pNP	No	Covalent	No	Covalent
<i>E. coli</i> (LacZ) β-galactosidase ²²	GH2	Gal-β-pNP	No	Covalent	No	Reversible

Data for all assays can be found in Fig. 2 and Fig. S15–19. Compounds denoted as “Reversible” inhibitors do not cause any time dependent inactivation, but inhibit the enzyme in a concentration dependent manner, with inhibition constants projected to be higher than 5 mM.

Inspired by a recent report by the Overkleeft group on *gluco*-configured aziridines and epoxides, which they found to adopt an unreactive conformation,¹³ we consider it most

likely that our *galacto*-configured counterparts also adopt such a conformation. To probe this possibility, we calculated the most stable conformations of the “α”- and “β”-1,2-



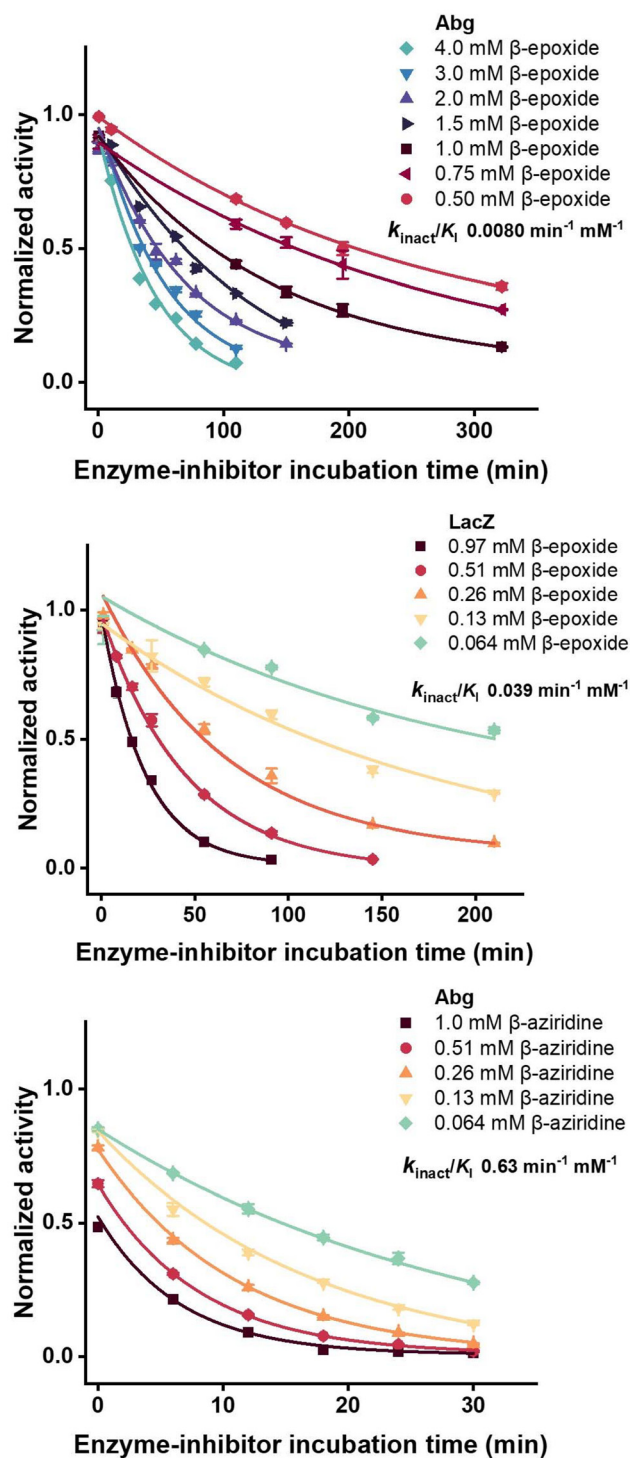
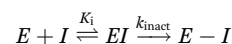


Fig. 2 Time-dependent inhibition of “ β ”-1,2-galacto-configured epoxide and aziridine. Residual enzyme activities were measured after incubation with varying concentrations of the compounds for different time periods. During the incubation, protein concentrations were approximately 1.9 nM for Abg (with 0.1 mg mL⁻¹ BSA) and 0.5 nM for LacZ β -galactosidase (with 0.1 mg mL⁻¹ BSA). For activity assays, 200 μ M Glc- β -pNP was used as the substrate for Abg in HEPES buffer (pH 7.4), and 100 μ M Gal- β -pNP was used as the substrate for LacZ in HEPES buffer (pH 7.4) supplemented with 1 mM Mg²⁺.

galacto-configured aziridines at the M062X/def2TZVP level of theory (Fig. S20). The lowest-energy DFT minima, which we refer to as the most stable conformations, correspond to a ⁴H₅ conformation for the “ α ” *galacto*-configured aziridine both in its neutral and protonated forms while the “ β ” *galacto*-configured aziridine appears to adopt a conformation between ⁴E and ⁴H₅. These are quite distinct from the ⁴H₃ conformation adopted by the substrate at the glycosylation transition state, and also seen for the 1,5a-aziridines and epoxides.¹³ Although Ofman *et al.*¹³ used metadynamics simulations to compute full free-energy landscapes for the *gluco*-configured analogues, their lowest-energy regions show the same conformational preferences for the *gluco*-configured aziridines and epoxides. Their analysis further indicates that these 1,2 configured structures are more rigid and therefore less able to undergo the conformational changes necessary for effective reaction.

By contrast the *Agrobacterium* sp. β -glucosidase/galactosidase (Abg) was inactivated by both the “ β ”-1,2-galacto-configured aziridine and epoxide, while the *Escherichia coli* (LacZ) β -galactosidase was inactivated only by the “ β ”-1,2-galacto-configured epoxide (Fig. 2).

Consistent with the covalent nature of the reactions, inactivation occurred in a time-dependent manner, allowing determination of rate constants for inactivation (k_{inact}) and dissociation constants (K_i) for binding according to the simple kinetic model below.



The formation of a covalent linkage was confirmed for Abg by intact mass electrospray ionisation mass spectrometry as shown in Fig. 3.

Conclusions

In summary, although the 1,2-configured cyclophellitol analogues are highly attractive candidates as mechanism-based inactivators of retaining glycosidases, no inactivation of α -galactosidases by the α -configured derivatives was seen. More unexpectedly, the α -configured aziridine analogue also failed to inhibit the α -galactosaminidase. Computational studies suggest that this is due to an unfavourable conformation imposed by the three membered rings, that makes binding in the enzyme active site energetically unfavorable. However, time-dependent, covalent inhibition of β -galactosidases by the β -configured analogues is shown by mass spectrometric analysis and kinetic studies. Very similar findings were reported recently for the equivalent *gluco*-configured versions, highlighting the importance of ring conformation to ligand binding.^{11,13}

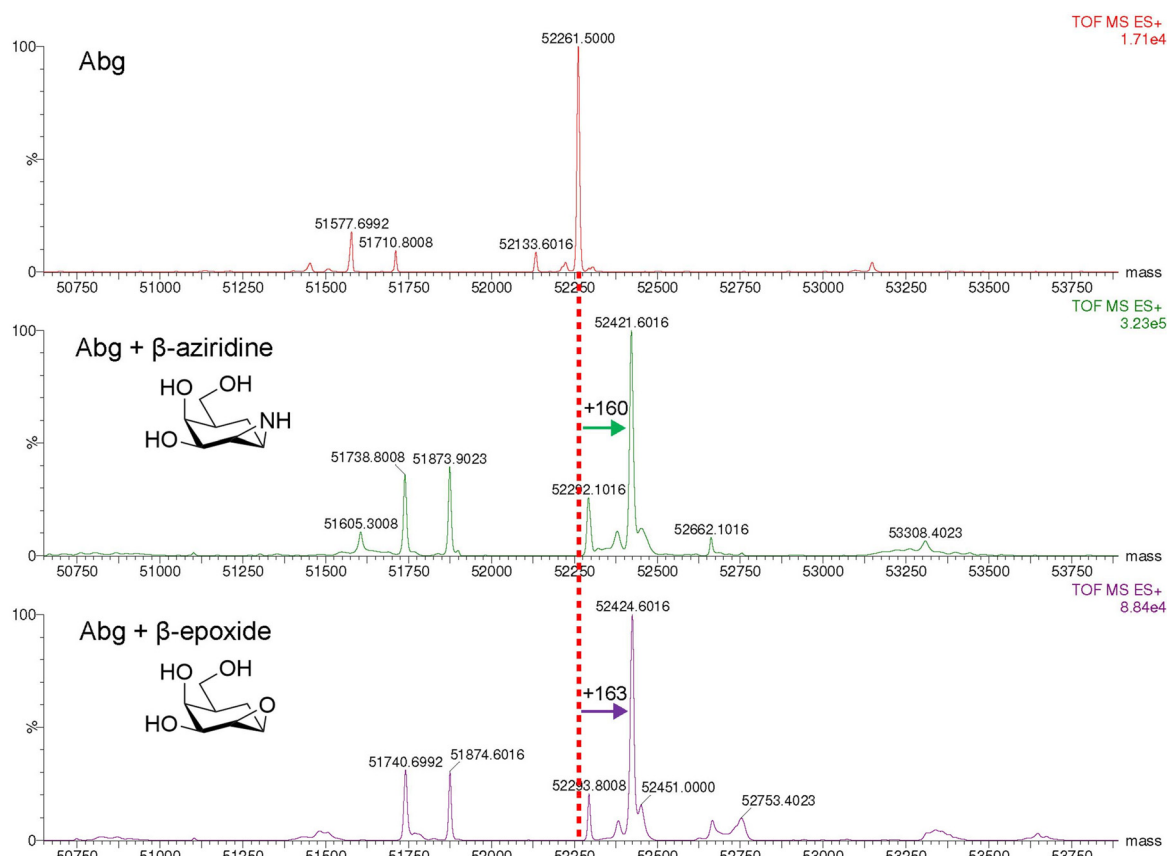


Fig. 3 Intact protein mass spectrometry of inactivated Abg. Observed increases in mass (Da) relative to apo enzyme are labeled on figures. Expected mass increases = 159 (aziridine), 160 (epoxide).

Experimental

Synthesis and characterization of compounds

General information. All chemicals were of analytical grade and purchased from the Sigma-Aldrich company, unless otherwise stated. All solvents were BOC standard grade and dried before use. Dichloromethane was distilled from calcium hydride. Methanol was distilled from magnesium. THF was distilled from sodium. DMF and DMSO were dried and stored over 4 Å molecular sieves. Triethylamine (TEA) was dried with KOH. Analytical thin-layer chromatography (TLC) was performed on aluminum-backed sheets of Silica Gel 60F₂₅₄ (E. Merck) of thickness 0.2 mm. The plates were visualised using UV light (254 nm) and/or by exposure to 10% ammonium molybdate (2 M in H₂SO₄) followed by charring. Flash column chromatography was carried out using Silicycle silica gel (230–400 mesh). Proton and carbon NMR spectra were recorded on Bruker Avance 400inv, 400dir Fourier Transform spectrometers fitted with a 5 mm BBI-Z probe. All spectra were recorded using an internal deuterium lock and are referenced internally using the residual solvent peak. Carbon and proton chemical shifts are quoted in parts per million (ppm) downfield of tetramethylsilane. Coupling constants (*J*) are given in Hertz (Hz). Carbon NMR spectra were measured with broad-

band proton decoupling and were recorded with DEPT. Mass spectra were measured on a Waters/Micromass LCT instrument in methanol using electrospray ionisation (ESI) and recorded using the Time-Of-Flight (TOF) method.

General procedure (GP1) for the preparation of azido alcohol (10a, 10a' & 10b). To a solution of benzyl protected epoxides (1.0 equiv.) in dry DMF (0.1 M), NaN₃ (10 equiv.) and LiClO₄ (20 equiv.) were added, and the reaction mixture was heated at 100 °C under an inert atmosphere. The reaction mixture was cooled down to room temperature, and water was added. The crude product was extracted with EtOAc (3 × 50 mL). The organic phase was washed with water (2 × 50 mL) and brine (50 mL), dried over MgSO₄, filtered and evaporated under reduced pressure. The resulting residue was purified by silica gel flash column chromatography (petroleum ether/EtOAc = 6/1 & 3/1) to afford the desired products.

General procedure (GP2) for the preparation of azido mesylates (11a, 11a' & 11b). To a solution of benzyl protected azido alcohols (1.0 equiv.) in dry THF (0.04 M) was added TEA (8.0 equiv.), and methanesulfonyl chloride (3.3 equiv.) was added dropwise with stirring under N₂. The reaction mixture was stirred for 2 h at room temperature, then solvent evaporated under reduced pressure. The crude product was redissolved in EtOAc (50 mL), washed with water (2 × 25 mL) and brine



(25 mL), dried over MgSO_4 , filtered and concentrated. The resulting residue was purified by silica gel flash column chromatography (petroleum ether/EtOAc = 9/1 & 5/1) to afford the desired products.

General procedure (GP3) for the preparation of aziridines (12a & 12b). A suspension of LiAlH_4 (10 equiv.) in dry THF (0.7 M) was cooled with an ice bath under N_2 . A solution of the mesylates (1 equiv.) in dry THF (0.07 M) was added dropwise to the above suspension with stirring. The reaction mixture was stirred for 2 h at 0 °C, gradually warmed to room temperature then stirred continually overnight. The reaction was quenched carefully by addition of EtOAc and water, and more water was added. The crude product was extracted with EtOAc (4 × 25 mL), then the organic phase was washed with water (50 mL) and brine (50 mL), dried over MgSO_4 , filtered and concentrated. The resulting residue was purified by silica gel flash column chromatography (petroleum ether/EtOAc = 1/1, 1/2 & 1/3 with 1% TEA) to afford the desired products.

General procedure (GP4) for the preparation of unprotected aziridines (13a & 13b). Ammonia (0.02 M) was condensed at −78 °C under N_2 . Sodium (60 equiv., cut in small pieces) was added and the mixture was stirred under the same conditions until all the sodium was completely dissolved (0.5 h). To the resulting dark blue solution, a solution of benzyl protected aziridines (1 equiv.) in dry THF (0.06 M) was added dropwise slowly to maintain a dark blue color. The reaction mixture was stirred for 2 h at −78 °C before being quenched with MilliQ- H_2O , warmed to room temperature and stirred overnight to evolve all ammonia. After evaporation under reduced pressure, the crude product was redissolved in MilliQ- H_2O , and purified by Amberlite 120 (NH_4^+) column eluted with 1 M NH_4OH solution. The combined fractions were evaporated, redissolved in 0.1 M NaOH or 1 M NH_4OH and lyophilized to afford 13a and 13b. Both aziridines have to be kept under basic conditions to avoid decomposition.

3,4,6-Tri-*O*-benzyl- α -carbagalactosyl & β -carbatalosyl epoxides (8a & 8b). To a solution of benzylated cyclohexene (7, 2.17 g, 5.24 mmol) in dry DCM (125 mL), cooled with an ice-bath, was added *m*-CPBA (2.72 g, 77%, 12.18 mmol, 2.3 equiv.) as one portion. The reaction mixture was stirred overnight under N_2 , and gradually warmed to room temperature. To the reaction mixture were added DCM (80 mL), 1 M $\text{Na}_2\text{S}_2\text{O}_3$ (60 mL) and saturated NaHCO_3 (60 mL), then the mixture was stirred vigorously for 15 minutes at room temperature. The two layers were separated and the water layer was extracted with DCM (3 × 100 mL). The combined DCM solution was washed with saturated NaHCO_3 (2 × 300 mL) and brine (300 mL), dried over MgSO_4 , filtered and concentrated. The resulting residue was purified by silica gel flash column chromatography (petroleum ether/EtOAc = 9/1 & 4/1) to afford the α -epimer (8a) as a white solid (1.118 g, 50%) and the β -epimer (8b) as a colorless oil (1.0 g, 44%), respectively. **α -Epimer 8a:** ^1H NMR (d_6 -acetone, 400 MHz): δ 7.46–7.23 (m, 15 H, Ar-H), 4.91 (d, 1 H, J = 11.8 Hz, ArCH_2), 4.83 (d, 1 H, J = 12.1 Hz, ArCH_2), 4.78 (d, 1 H, J = 12.1 Hz, ArCH_2), 4.51 (d, 1 H, J = 11.8 Hz, ArCH_2), 4.47 (d, 1 H, J = 12.0 Hz, ArCH_2), 4.43 (d, 1 H, J = 12.0 Hz, ArCH_2),

4.02 (brs, 1 H, H-4), 3.92 (dd, 1 H, J = 2.4 & 3.8 Hz, H-3), 3.49 (dd, 1 H, J = 8.0 Hz & 8.9 Hz, H-6), 3.39 (dd, 1 H, J = 6.3 & 8.9 Hz, H-6), 3.26 (brs, 1 H, H-2), 3.20 (t, 1 H, J = 4.1 Hz, H-1), 1.91 (m, 1 H, H-5), 1.79 (m, 2 H, H-7). ^{13}C NMR (CDCl_3 , 100 MHz): δ 139.5, 138.6, 138.4, 128.6, 128.5, 128.3, 128.0, 127.95, 127.82, 127.8, 127.7, 127.4, 77.1, 74.4, 74.0, 73.3, 71.1, 70.9, 52.5, 38.6, 23.0. ESI-HRMS: calcd for $[\text{C}_{28}\text{H}_{30}\text{O}_4 + \text{H}]^+$: 431.2222; found m/z : 431.2220. **β -Epimer 8b:** ^1H NMR (d_6 -acetone, 400 MHz): δ 7.47–7.23 (m, 15 H, Ar-H), 4.95 (d, 1 H, J = 11.5 Hz, ArCH_2), 4.84 (d, 1 H, J = 12.0 Hz, ArCH_2), 4.76 (d, 1 H, J = 12.0 Hz, ArCH_2), 4.54 (d, 1 H, J = 11.5 Hz, ArCH_2), 4.45 (d, 1 H, J = 12.0 Hz, ArCH_2), 4.39 (d, 1 H, J = 12.0 Hz, ArCH_2), 4.00 (brs, 1 H, H-4), 3.58 (brd, 1 H, J = 3.2 Hz, H-3), 3.47 (t, 1 H, J = 8.6 Hz, H-6), 3.31 (dd, 1 H, J = 4.8 & 8.6 Hz, H-6), 3.24 (brd, 1 H, J = 3.7 Hz, H-2), 3.15 (dd, 1 H, J = 1.8 & 3.7 Hz, H-1), 1.85 (brd, 1 H, J = 5.2 Hz, H-7), 1.79 (m, 2 H, H-5 & H-7). ^{13}C NMR (CDCl_3 , 100 MHz): δ 139.5, 138.43, 138.39, 128.6, 128.56, 128.3, 128.0, 127.9, 127.8, 127.6, 127.5, 78.4, 74.8, 73.2, 72.9, 71.7, 70.8, 54.1, 53.7, 33.8, 23.1. ESI-HRMS: calcd for $[\text{C}_{28}\text{H}_{30}\text{O}_4 + \text{H}]^+$: 431.2222; found m/z : 431.2219.

α -Carbagalactosyl epoxide (9a). A solution of 8a (70 mg, 0.16 mmol) in MeOH/EtOAc (5/1, 6 mL) was treated with Pd/C (10%, 28 mg) and H_2 . The suspension was stirred for 24 h at room temperature, filtered through Celite and washed with MeOH. The solvents were evaporated under reduced pressure and the resulting residue was purified by silica gel flash column chromatography (DCM/MeOH = 15/1) to afford (9a) as a white solid (15 mg, 58%). ^1H NMR (d_4 -methanol, 400 MHz): 3.85 (dd, 1 H, J = 1.6 Hz, H-3), 3.62 (d, 1 H, J = 4.0 Hz, H-4), 3.60 (dd, 1 H, J = 8.0 Hz, H-6), 3.48 (dd, 1 H, J = 8.0 & 10.6 Hz, H-6), 3.29 (t, 1 H, J = 1.6 Hz, H-1), 2.98 (dd, 1 H, J = 1.6 & 3.7 Hz, H-2), 1.88 (ddd, 1 H, J = 1.2, 5.0 & 14.6 Hz, H-7), 1.74 (ddd, 1 H, J = 1.9, 12.0 & 14.6 Hz, H-7), 1.53 (m, 1 H, H-5). ^{13}C NMR (d_4 -methanol, 100 MHz): δ 68.6, 66.2, 62.9, 55.1, 53.5, 34.7, 21.7. ESI-HRMS: calcd for $[\text{C}_7\text{H}_{12}\text{O}_4 + \text{Na}]^+$: 183.0633; found m/z : 183.0628.

β -Carbatalosyl epoxide (9b). A solution of 8b (50 mg, 0.12 mmol) in MeOH/ H_2O /dioxane (1/1/1, 7 mL) was treated with $\text{Pd}(\text{OH})_2/\text{C}$ (20%, wet, 10 mg) and H_2 . The suspension was stirred for 2 h at room temperature, filtered through Celite and washed with MeOH. The solvents were evaporated under reduced pressure and the resulting residue was purified by silica gel flash column chromatography (DCM/MeOH = 9/1) to afford (9b) as a white solid (9.6 mg, 52%). ^1H NMR (d_4 -methanol, 400 MHz): 3.89 (m, 2 H, H-3 & H-4), 3.59 (dd, 1 H, J = 7.4 & 10.6 Hz, H-6), 3.44 (m, 2 H, H-1 & H-6), 3.22 (m, 1 H, H-2), 2.00 (m, 1 H, H-7), 1.69 (m, 2 H, H-5 & H-7). ^{13}C NMR (d_4 -methanol, 100 MHz): δ 69.7, 68.6, 62.7, 57.3, 54.0, 40.5, 21.4. ESI-HRMS: calcd for $[\text{C}_7\text{H}_{12}\text{O}_4 + \text{H}]^+$: 161.0814; found m/z : 161.0809.

1-Azido-1-deoxy-3,4,6-tri-*O*-benzyl- β -carbagalactose (10a) & 2-azido-2-deoxy-3,4,6-tri-*O*-benzyl- α -carbatalose (10a'). Starting from 8a (985 mg, 2.29 mmol), the reaction mixture was stirred for 48 h and processed following GP1 to afford 10a (583 mg, 55%) and 10a' (99 mg, 9%). **10a:** ^1H NMR (CDCl_3 , 400 MHz): δ 7.37–7.27 (m, 15 H, Ar-H), 4.92 (d, 1 H, J = 11.4 Hz, ArCH_2),



4.80 (d, 1 H, $J = 11.6$ Hz, ArCH_2), 4.57 (d, 1 H, $J = 11.6$ Hz, ArCH_2), 4.52 (d, 1 H, $J = 11.4$ Hz, ArCH_2), 4.46 (s, 2 H, ArCH_2), 4.15 (brs, 1 H, H-4), 4.01 (t, 1 H, $J = 9.5$ Hz, H-2), 3.51 (t, 1 H, $J = 8.8$ Hz, H-6), 3.36 (m, 2 H, H-1 & H-6), 3.28 (dd, 1 H, $J = 2.1$ & 9.5 Hz, H-3), 2.49 (brs, 1 H, OH), 1.85 (m, 1 H, H-5), 1.70 (dt, 1 H, $J = 4.0$ & 12.8 Hz, H-7), 1.58 (t, 1 H, H-7). ^{13}C NMR (CDCl_3 , 100 MHz): δ 139.0, 138.1, 138.0, 128.8, 128.6, 128.4, 128.1, 128.04, 128.0, 127.9, 127.7, 127.6, 84.5, 74.6, 73.6, 73.5, 73.1, 72.5, 70.5, 63.1, 38.6, 27.5. ESI-HRMS: calcd for $[\text{C}_{28}\text{H}_{31}\text{N}_3\text{O}_4 + \text{Na}]^+$: 496.2212; found m/z : 496.2206. **10a'**: ^1H NMR (d_6 -acetone, 400 MHz): δ 7.33–7.26 (m, 15 H, Ar-H), 4.88 (d, 1 H, $J = 11.6$ Hz, ArCH_2), 4.82 (d, 1 H, $J = 11.3$ Hz, ArCH_2), 4.73 (d, 1 H, $J = 11.7$ Hz, ArCH_2), 4.64 (d, 1 H, $J = 11.6$ Hz, ArCH_2), 4.48 (s, 2 H, ArCH_2), 4.25 (d, 1 H, $J = 3.6$ Hz, OH), 4.10 (t, 1 H, $J = 2.8$ Hz, H-3), 4.02 (brs, 2 H, H-1 & H-4), 3.72 (m, 2 H, H-2 & H-6), 3.55 (t, 1 H, $J = 8.8$ Hz, H-6), 2.44 (m, 1 H, H-5), 2.04 (m, 1 H, H-7), 1.44 (ddd, 1 H, $J = 4.0$, 6.8 & 13.4 Hz, H-7). ^{13}C NMR (d_6 -acetone, 100 MHz): δ 139.6, 139.3, 139.0, 128.34, 128.31, 128.3, 127.7, 127.67, 127.65, 127.5, 127.44, 127.42, 127.3, 79.4, 77.3, 73.0, 72.8, 72.7, 70.1, 66.8, 64.3, 36.4, 28.8. ESI-HRMS: calcd for $[\text{C}_{28}\text{H}_{31}\text{N}_3\text{O}_4 + \text{Na}]^+$: 496.2212; found m/z : 496.2208.

1-Azido-1-deoxy-3,4,6-tri-*O*-benzyl- α -carbatalose (**10b**)

Starting from **8b** (966 mg, 2.25 mmol), the reaction was carried out following GP1 stirring for 22 h to afford **10b** (975 mg, 92%). **10b**: ^1H NMR (d_6 -acetone, 400 MHz): δ 7.44–7.27 (m, 15 H, Ar-H), 4.84 (d, 1 H, $J = 11.1$ Hz, ArCH_2), 4.78 (d, 1 H, $J = 11.8$ Hz, ArCH_2), 4.74 (d, 1 H, $J = 11.8$ Hz, ArCH_2), 4.59 (d, 1 H, $J = 11.1$ Hz, ArCH_2), 4.50 (s, 2 H, ArCH_2), 4.23 (d, 1 H, $J = 11.2$ Hz, OH), 4.09 (brs, 1 H, H-4), 3.91 (brs, 2 H, H-1 & H-2), 3.80 (brs, 1 H, H-3), 3.67 (dd, 1 H, $J = 7.0$ & 9.2 Hz, H-6), 3.55 (dd, 1 H, $J = 7.6$ & 9.2 Hz, H-6), 2.28 (m, 1 H, H-5), 2.03 (m, 1 H, H-7), 1.47 (m, 1 H, H-7). ^{13}C NMR (d_6 -acetone, 100 MHz): δ 139.2, 139.1, 138.8, 128.5, 128.4, 128.35, 127.9, 127.88, 127.85, 127.8, 127.75, 127.6, 127.5, 127.48, 78.5, 77.3, 73.9, 72.7, 71.6, 71.4, 69.6, 61.4, 36.5, 24.2. ESI-HRMS: calcd for $[\text{C}_{28}\text{H}_{31}\text{N}_3\text{O}_4 + \text{Na}]^+$: 496.2212; found m/z : 496.2206.

1-Azido-1-deoxy-3,4,6-tri-*O*-benzyl- β -carbagalactosyl mesylate (11a**)**. Starting from **10a** (575 mg, 1.22 mmol), the reaction was carried out following GP2 to afford **11a** (670 mg, 100%) as a colorless oil. **11a**: ^1H NMR (d_6 -acetone, 400 MHz): δ 7.49–7.26 (m, 15 H, Ar-H), 4.96 (d, 1 H, $J = 11.4$ Hz, ArCH_2), 4.85 (d, 1 H, $J = 11.4$ Hz, ArCH_2), 4.83 (t, 1 H, $J = 9.9$ Hz, H-2), 4.74 (d, 1 H, $J = 11.4$ Hz, ArCH_2), 4.57 (d, 1 H, $J = 11.4$ Hz, ArCH_2), 4.50 (d, 1 H, $J = 12.0$ Hz, ArCH_2), 4.45 (d, 1 H, $J = 12.0$ Hz, ArCH_2), 4.29 (brs, 1 H, H-4), 3.74 (dd, 1 H, $J = 2.3$ & 9.9 Hz, H-3), 3.70 (m, 1 H, H-1), 3.54 (t, 1 H, $J = 8.8$ Hz, H-6), 3.42 (dd, 1 H, $J = 2.0$ & 9.0 Hz, H-6), 3.03 (s, 3 H, CH_3), 2.05 (m, 1 H, H-5), 1.94 (dt, 1 H, $J = 4.2$, 8.6 & 12.6 Hz, H-7), 1.57 (dd, 1 H, $J = 12.7$ Hz, H-7). ^{13}C NMR (d_6 -acetone, 100 MHz): δ 139.3, 138.8, 138.2, 128.5, 128.4, 128.3, 127.9, 127.8, 127.7, 127.6, 127.5, 83.4, 81.8, 74.7, 74.4, 72.7, 72.4, 70.2, 61.7, 38.6, 37.7, 27.6. ESI-HRMS: calcd for $[\text{C}_{29}\text{H}_{33}\text{N}_3\text{O}_6\text{S} + \text{Na}]^+$: 574.1988; found m/z : 574.1983.

2-Azido-2-deoxy-3,4,6-tri-*O*-benzyl- α -carbatalosyl mesylate (11a'**)**. Starting from **10a'** (105 mg, 0.22 mmol), the reaction was carried out following GP2 to afford **11a'** (115 mg, 94%) as a colorless oil. **11a'**: ^1H NMR (d_6 -acetone, 400 MHz): δ

7.42–7.26 (m, 15 H, Ar-H), 4.90 (d, 1 H, $J = 11.5$ Hz, ArCH_2), 4.85 (d, 1 H, $J = 11.6$ Hz, ArCH_2), 4.85 (m, 1 H, H-1), 4.80 (d, 1 H, $J = 11.6$ Hz, ArCH_2), 4.64 (d, 1 H, $J = 11.5$ Hz, ArCH_2), 4.49 (s, 2 H, ArCH_2), 4.41 (brs, 3 H, H-2, H-3 & H-4), 3.70 (dd, 1 H, $J = 6.4$ & 9.3 Hz, H-6), 3.55 (dd, 1 H, $J = 7.9$ & 9.1 Hz, H-6), 3.16 (s, 3 H, CH_3), 2.39 (m, 1 H, H-5), 2.17 (m, 1 H, H-7), 1.72 (ddd, 1 H, $J = 4.1$, 6.4 & 14.0 Hz, H-7). ^{13}C NMR (d_6 -acetone, 100 MHz): δ 139.3, 139.1, 138.6, 128.4, 128.3, 127.7, 127.64, 127.63, 127.5, 127.4, 78.8, 77.7, 76.2, 73.4, 72.8 (2C), 69.6, 61.1, 37.8, 36.4, 26.6. ESI-HRMS: calcd for $[\text{C}_{29}\text{H}_{33}\text{N}_3\text{O}_6\text{S} + \text{Na}]^+$: 574.1988; found m/z : 574.1982.

1-Azido-1-deoxy-3,4,6-tri-*O*-benzyl- α -carbatalosyl mesylate (11b**)**. Starting from **10b** (960 mg, 2.03 mmol), the reaction was carried out following GP2 to afford **11b** (1.118 g, 100%): ^1H NMR (d_6 -acetone, 400 MHz): δ 7.40–7.27 (m, 15 H, Ar-H), 4.81 (d, 1 H, $J = 11.4$ Hz, ArCH_2), 4.77 (d, 1 H, $J = 11.7$ Hz, ArCH_2), 4.74 (d, 1 H, $J = 12.0$ Hz, ArCH_2), 4.69 (dd, 1 H, $J = 2.8$ & 7.9 Hz, H-2), 4.64 (d, 1 H, $J = 11.7$ Hz, ArCH_2), 4.51 (d, 1 H, $J = 12.0$ Hz, ArCH_2), 4.46 (d, 1 H, $J = 12.0$ Hz, ArCH_2), 4.17 (brs, 1 H, H-3), 4.04 (ddd, 1 H, $J = 4.2$, 8.6 & 12.5 Hz, H-1), 3.98 (dd, 1 H, $J = 2.4$ & 4.4 Hz, H-4), 3.77 (dd, 1 H, $J = 4.6$ & 9.6 Hz, H-6), 3.66 (t, 1 H, $J = 9.3$ Hz, H-6), 3.11 (s, 3 H, CH_3), 2.43 (m, 1 H, H-5), 2.25 (ddd, 1 H, $J = 4.4$, 5.8 & 13.9 Hz, H-7), 1.55 (ddd, 1 H, $J = 4.3$, 9.1 & 13.6 Hz, H-7). ^{13}C NMR (CDCl_3 , 100 MHz): δ 139.2, 139.1, 138.7, 128.3, 127.8, 127.78, 127.6, 127.54, 127.46, 127.5, 80.7, 78.1, 76.8, 74.0, 72.6, 71.9, 68.6, 57.7, 37.8, 36.8, 26.3. Calcd for $[\text{C}_{29}\text{H}_{33}\text{N}_3\text{O}_6\text{S} + \text{Na}]^+$: 574.1988; found m/z : 574.1982.

3,4,6-Tri-*O*-benzyl- β -carbatalosyl aziridine (12a**)**. Starting from **11a** and **11a'** (670 mg, 1.22 mmol), the reaction was carried out following GP3 to afford **12a** (261 mg, 50%): ^1H NMR (CDCl_3 , 400 MHz): δ 7.38–7.27 (m, 15 H, Ar-H), 5.01 (d, 1 H, $J = 11.2$ Hz, ArCH_2), 4.92 (d, 1 H, $J = 12.0$ Hz, ArCH_2), 4.77 (d, 1 H, $J = 12.0$ Hz, ArCH_2), 4.43 (m, 3 H, ArCH_2), 4.14 (brs, 1 H, H-4), 3.95 (t, 1 H, $J = 3.2$ Hz, H-3), 3.38 (t, 1 H, $J = 8.8$ Hz, H-6), 3.28 (dd, 1 H, $J = 5.0$ & 8.9 Hz, H-6), 2.36 (brs, 1 H, H-2), 2.27 (t, 1 H, $J = 5.8$ Hz, H-1), 1.92 (dt, 1 H, $J = 6.8$ & 13.9 Hz, H-7), 1.84 (m, 1 H, H-5), 1.59 (t, 1 H, $J = 12.9$ Hz, H-7). ^{13}C NMR (CDCl_3 , 100 MHz): δ 138.9, 138.8, 138.3, 128.61, 128.59, 128.4, 128.1, 128.0, 127.9, 127.7, 127.67, 77.5, 75.7, 75.5, 73.3, 70.7, 70.5, 39.2, 33.5, 31.6, 22.3. ESI-HRMS: calcd for $[\text{C}_{28}\text{H}_{31}\text{NO}_3 + \text{H}]^+$: 452.2202; found m/z : 452.2198.

3,4,6-Tri-*O*-benzyl- α -carbagalactosyl aziridine (12b**)**. Starting from **11b** (1.118 g, 2.03 mmol), the reaction was carried out following GP3 to afford **12b** (766 mg, 88%). **12b**: ^1H NMR (CDCl_3 , 400 MHz): δ 7.35–7.25 (m, 15 H, Ar-H), 4.98 (d, 1 H, $J = 11.9$ Hz, ArCH_2), 4.80 (d, 1 H, $J = 12.0$ Hz, ArCH_2), 4.71 (d, 1 H, $J = 12.0$ Hz, ArCH_2), 4.55 (d, 1 H, $J = 11.9$ Hz, ArCH_2), 4.36 (s, 2 H, ArCH_2), 3.95 (d, 1 H, $J = 2.8$ Hz, H-3), 3.53 (d, 1 H, $J = 3.0$ Hz, H-4), 3.40 (t, 1 H, $J = 8.5$ Hz, H-6), 3.25 (dd, 1 H, $J = 4.0$ & 8.8 Hz, H-6), 2.44 (d, 1 H, $J = 6.2$ Hz, H-1), 2.34 (d, 1 H, $J = 6.2$ Hz, H-2), 1.70 (m, 3 H, H-5 & 2 \times H-7). ^{13}C NMR (d_6 -acetone, 100 MHz): δ 139.8, 138.8, 138.6, 128.51, 128.47, 128.3, 127.9, 127.8, 127.7, 127.6, 127.5, 127.3, 80.3, 74.6, 73.5, 73.0, 71.5, 71.1, 33.8, 32.3, 30.6, 22.9. ESI-HRMS: calcd for $[\text{C}_{28}\text{H}_{31}\text{NO}_3 + \text{Na}]^+$: 430.2382; found m/z : 452.2377.



β -Carbatalosyl aziridine (13a). Starting from **12a** (48 mg, 0.11 mmol), the reaction was carried out following GP4 to afford **13a** (15 mg, 84%): ^1H NMR (D_2O , 400 MHz): δ 3.88 (dd, 1 H, J = 3.2 & 3.5 Hz, H-3), 3.83 (brs, 1 H, H-4), 3.48 (dd, 1 H, J = 7.9 & 10.9 Hz, H-6), 3.39 (dd, 1 H, J = 6.3 & 10.9 Hz, H-6), 2.43 (t, 1 H, J = 6.4 Hz, H-1), 2.35 (brs, 1 H, H-2), 1.91 (dd, 1 H, J = 6.9 & 14.8 Hz, H-7), 1.65 (m, 1 H, H-5), 1.39 (dd, 1 H, J = 12.1 & 14.8 Hz, H-7). ^{13}C NMR (D_2O , 100 MHz): δ 70.5, 68.1, 63.0, 40.3, 35.5, 30.8, 21.2. ESI-HRMS: calcd for $[\text{C}_7\text{H}_{13}\text{NO}_3 + \text{H}]^+$: 160.0974; found m/z : 160.0968.

α -Carbagalactosyl aziridine (13b). Starting from **12b** (130 mg, 0.3 mmol), the reaction was carried out following GP4 to afford **13b** (44 mg, 92%): ^1H NMR (D_2O , 400 MHz): δ 3.82 (d, 1 H, J = 3.2 Hz, H-3), 3.63 (d, 1 H, J = 3.6 Hz, H-4), 3.57 (dd, 1 H, J = 7.7 & 11.0 Hz, H-6), 3.48 (dd, 1 H, J = 5.8 & 11.0 Hz, H-6), 2.43 (dd, 1 H, J = 3.8 & 6.1 Hz, H-1), 2.10 (d, 1 H, J = 6.2 Hz, H-2), 1.80 (dd, 1 H, J = 4.9 & 14.0 Hz, H-7), 1.55 (dt, 1 H, J = 3.6 & 12.6 Hz, H-7), 1.44 (m, 1 H, H-5). ^{13}C NMR (D_2O , 100 MHz): δ 69.8, 67.1, 63.1, 34.9, 33.0, 30.4, 20.8. ESI-HRMS: calcd for $[\text{C}_7\text{H}_{13}\text{NO}_3 + \text{H}]^+$: 160.0974; found m/z : 160.0968.

Inhibition kinetics determination

Protein expression and purification. Recombinant enzymes, including GH1 β -glucosidase from *Agrobacterium* sp. (Abg),²¹ GH36 α -galactosidase from *Thermotoga maritima* (TmGalA),⁴ GH36 α -galactosaminidase from *Flavonofractor plautii* (FpGalNase),¹ and *Escherichia coli* (LacZ) GH2 β -galactosidase²² were overexpressed in *E. coli* BL21(DE3) cells. Cultures were grown in ZY5052 auto-induction medium²³ or LB medium supplemented with appropriate antibiotic (50 mg L^{-1} kanamycin or 100 mg L^{-1} ampicillin). Each culture was inoculated with 0.5 mL of LB pre-culture and incubated at 37 °C for 16 h. Cells were harvested by centrifugation (4000 rpm, 30 min, 4 °C), and the pellets were resuspended in lysis buffer (50 mM HEPES, 150 mM NaCl, 20 mM imidazole, 5 mM MgCl_2 ; pH 7.4) containing 2 U mL^{-1} Benzonase (Novagen), 0.2 g L^{-1} lysozyme, and one protease inhibitor tablet per 50 mL buffer (Pierce). The suspension was lysed by sonication on ice (3 min total; 5 s on/15 s off cycles at 35% amplitude). After centrifugation (15 000 rpm, 30 min, 4 °C), the supernatant was loaded onto a nickel affinity column. Bound proteins were eluted using a gradient of 10–100% elution buffer (50 mM HEPES, 150 mM NaCl, 400 mM imidazole; pH 7.4), and fractions were analyzed by SDS-PAGE. Imidazole was removed and the buffer exchanged to the appropriate assay buffer using Amicon Ultra centrifugal filters (MWCO 30 kDa or 10 kDa, Millipore). α -galactosidase from green coffee beans was purchased from Sigma-Aldrich.

Inhibition assay. The inhibitory effects of compounds were evaluated using a time-course assay, in which the enzyme was first incubated with each inhibitor, and then sampled at specific time points to measure the residual enzymatic activity using the corresponding substrate.

For each inhibitor, sets of 85 μL PCR tubes were prepared containing the inhibitor at 2 \times the final assay concentration in the appropriate assay buffer. In parallel, another set of PCR

tubes was prepared with the enzyme solution at 2 \times the final assay concentration in the same buffer. All tubes were pre-warmed at 25 °C for 10 minutes. Prior to initiating the inhibition reaction, substrate mixtures were prepared in a half-area 96-well plate, black plate for methylumbelliferyl (MU) substrate or clear plate for *para*-nitrophenyl (*p*NP) substrate. Each well contained 80 μL of substrate at 1.25 \times the final assay concentration in the appropriate buffer. The plate was pre-warmed at 25 °C for 5 minutes.

To initiate the reaction ($t = 0$), 85 μL of enzyme solution was added to 85 μL of inhibitor solution, mixed thoroughly (yielding a 170 μL mixture), and immediately, 20 μL of this mixture was transferred into 80 μL of substrate solution in the plate. The release of *p*NP or MU was monitored in real time for 5 minutes using a BioTek Synergy H1 plate reader. The initial reaction rate (V) was calculated from the linear portion of the curve. At subsequent incubation time points, additional 20 μL aliquots were taken from the enzyme-inhibitor mixture and added to fresh wells containing 80 μL of pre-warmed 1.25 \times substrate solution. All reaction rates were normalized to the relative activity of the $t = 0$ control sample to which no additional inhibitor compound was added. The inhibitor concentrations reported in the result figures refer to the concentrations during the 170 μL incubation phase in PCR tubes. When 20 μL of the enzyme-inhibitor mixture was added into 80 μL of substrate solution, the inhibitor concentration was 5 times diluted. All assays were performed in duplicate.

Intact protein mass spectrometry. To assess covalent inhibition, intact mass spectrometry was performed on Abg. The protein was prepared at 1 mg mL^{-1} in 50 mM HEPES buffer (pH 7.4) and treated with either 12.5 mM β -aziridine or β -epoxide. A control sample contained the same concentration of Abg in the same buffer without inhibitor. After incubation for 30 min, the reaction mixtures were washed three times with 5 mM HEPES buffer (pH 7.4) using 10 kDa MWCO VivaSpin columns and centrifuged at 8000g to remove unbound compounds and reduce salt concentration prior to MS analysis. The samples were then diluted to 10 ng μL^{-1} in 50% acetonitrile containing 0.2% formic acid. Mass spectra were acquired by direct injection of the sample at a flow rate of 10 $\mu\text{L min}^{-1}$ onto a Waters Xevo G2-S QToF mass spectrometer. Data were processed using the MaxEnt1 algorithm in Waters MassLynx 4.1 software.

Conflicts of interest

There are no conflicts to declare.

Data availability

Supplementary information (SI): cartesian coordinates of the lowest-energy structures and full NMR spectra. See DOI: <https://doi.org/10.1039/d5ob01802a>.



Acknowledgements

S. G. W. thanks the Natural Sciences and Engineering Research Council of Canada (NSERC) and the Canadian Glycomics Network, GlycoNet, for financial support. Financial support to J. P. R. was provided by the University of British Columbia, the Natural Sciences and Engineering Research Council of Canada (NSERC) and the CFI John R. Evans Leaders Fund. Computational resources were provided from the Digital Research Alliance of Canada and the Advanced Research Computing (ARC) center at the University of British Columbia. We thank Dr Shinya Fushinobu for providing access to his Cremer-Pople parameters calculator (<https://enzyme13.bt.a.u-tokyo.ac.jp/CP/>).

References

- 1 P. Rahfeld, L. Sim, H. Moon, I. Constantinescu, C. Morgan-Lang, S. J. Hallam, J. N. Kizhakkedathu and S. G. Withers, *Nat. Microbiol.*, 2019, **4**, 1475–1485.
- 2 Y. Tian, L. J. Worrall, L. Sim, F. Liu, S. A. Nasser, P. Rahfeld, W. Mu, J. N. Kizhakkedathu, N. C. J. Strynadka and S. G. Withers, *ACS Catal.*, 2024, **14**, 13497–13508.
- 3 F. Fredslund, M. A. Hachem, R. J. Larsen, P. G. Sorensen, P. M. Coutinho, L. Lo Leggio and B. Svensson, *J. Mol. Biol.*, 2011, **412**, 466–480.
- 4 D. A. Comfort, K. S. Bobrov, D. R. Ivanen, K. A. Shabalin, J. M. Harris, A. A. Kulminkaya, H. Brumer and R. M. Kelly, *Biochemistry*, 2007, **46**, 3319–3330.
- 5 I. P. Street, J. B. Kempton and S. G. Withers, *Biochemistry*, 1992, **31**, 9970–9978.
- 6 R. M. Mosi and S. G. Withers, in *Methods Enzymol.*, ed. D. L. Purich, Academic Press, 2002, vol. 354, pp. 64–84.
- 7 J. D. McCarter and S. G. Withers, *J. Am. Chem. Soc.*, 1996, **118**, 241–242.
- 8 D. J. Vocadlo, C. Mayer, S. M. He and S. G. Withers, *Biochemistry*, 2000, **39**, 117–126.
- 9 S. G. Withers and K. Umezawa, *Biochem. Biophys. Res. Commun.*, 1991, **177**, 532–537.
- 10 D. J. Vocadlo and G. J. Davies, *Curr. Opin. Chem. Biol.*, 2008, **12**, 539–555.
- 11 M. Artola, J. Aerts, G. A. van der Marel, C. Rovira, J. D. C. Codée, G. J. Davies and H. S. Overkleeft, *J. Am. Chem. Soc.*, 2024, **146**, 24729–24741.
- 12 V. W.-F. Tai, P. H. Fung, Y. S. Wong and T. K. M. Shing, *Tetrahedron: Asymmetry*, 1994, **5**, 1353–1362.
- 13 T. P. Ofman, J. J. A. Heming, A. Nin-Hill, F. Küllmer, E. Moran, M. Bennett, R. Steneker, A.-M. Klein, G. Ruijgrok, K. Kok, Z. W. B. Armstrong, J. M. F. G. Aerts, G. A. van der Marel, C. Rovira, G. J. Davies, M. Artola, J. D. C. Codée and H. S. Overkleeft, *Chem. – Eur. J.*, 2024, **30**, e202400723.
- 14 D. Lu, S. Zhu, L. F. Sobala, G. Bernardo-Seisdedos, O. Millet, Y. M. Zhang, J. Jiménez-Barbero, G. J. Davies and M. Sollogoub, *Org. Lett.*, 2018, **20**, 7488–7492.
- 15 L. F. Sobala, G. Speciale, S. Zhu, L. Raich, N. Sannikova, A. J. Thompson, Z. Hakki, D. Lu, S. S. K. Abadi, A. R. Lewis, V. Rojas-Cervellera, G. Bernardo-Seisdedos, Y. M. Zhang, O. Millet, J. Jiménez-Barbero, A. J. Bennet, M. Sollogoub, C. Rovira, G. J. Davies and S. J. Williams, *ACS Cent. Sci.*, 2020, **6**, 760–770.
- 16 N. Ushida, N. Nagai, M. Adachi and T. Nishikawa, *Synlett*, 2019, **30**, 977–981.
- 17 S. P. Schröder, R. Petracca, H. Minnee, M. Artola, J. M. F. G. Aerts, J. D. C. Codée, G. A. van der Marel and H. S. Overkleeft, *Eur. J. Org. Chem.*, 2016, 4787–4794.
- 18 D. Lu, S. Zhu, L. F. Sobala, G. Bernardo-Seisdedos, O. Millet, Y. Zhang, J. Jiménez-Barbero, G. J. Davies and M. Sollogoub, *Org. Lett.*, 2018, **20**, 7488–7492.
- 19 P. Serrano, A. Llebaria and A. Delgado, *J. Org. Chem.*, 2005, **70**, 7829–7840.
- 20 A. Zhu, Z.-K. Wang and J. Goldstein, *Biochim. Biophys. Acta*, 1995, **1247**, 260–264.
- 21 J. B. Kempton and S. G. Withers, *Biochemistry*, 1992, **31**, 9961–9969.
- 22 D. H. Juers, T. D. Heightman, A. Vasella, J. D. McCarter, L. Mackenzie, S. G. Withers and B. W. Matthews, *Biochemistry*, 2001, **40**, 14781–14794.
- 23 F. W. Studier, *Protein Expression Purif.*, 2005, **41**, 207–234.

