



Marine natural products

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Covering: January to the end of December 2024

This review covers the literature published in 2024 for marine natural products (MNPs), with 617 citations (578 for the period January to December 2024) referring to compounds isolated from marine microorganisms and phytoplankton, green, brown and red algae, sponges, cnidarians, bryozoans, molluscs, tunicates, echinoderms, the submerged parts of mangroves and other intertidal plants. The emphasis is on new compounds (1256 in 336 papers for 2024), together with the relevant biological activities, source organisms and country of origin. Pertinent reviews, biosynthetic studies, first syntheses, and syntheses that led to the revision of structures or stereochemistries, have been included. An analysis of the role of artificial intelligence in marine natural products research is discussed.

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1 Introduction

This review is of the literature for 2024 and describes 1256 new compounds from 336 papers, compared to 1220 new compounds in 340 papers reported for 2023.¹ In addition, 30 known NPs were reported from a marine source for the first time, three artefacts were identified, and 48 known MNPs had their structures revised. Only new MNP structures or previously reported compounds where there has been a structural revision, or a newly established stereochemistry are shown in this review. The review also covers previously reported MNPs with significant new bioactivities or ones that have been synthesised for the first time, but their structures are generally not shown. A † symbol on the identifying diagram number is used to distinguish structures where the absolute configuration has been determined for all stereogenic centres, axes and/or planes in a compound. Reports of new MNPs that were identified based solely on a combination of biosynthetic gene cluster (BGC) information, MS/MS data and/or Global Natural Products Social (GNPS)-based molecular networking, with compounds not isolated and no NMR data recorded, are excluded from the review. Only a selection of highlighted structures (54) is shown. Compound numbers for structures not highlighted in the review are *italicised*, and all structures are available for viewing, along with their names, taxonomic origins, collection locations, and biological activities, in an associated SI document. Over the past 20 years we have discussed a variety of topics relevant to

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MNP research in the conclusion section. Given that these treatises introduce new data derived from analysis of historical trends, we feel that it is more appropriate to include this

discussion in a stand-alone section rather than the conclusion section. Therefore, this year we introduce a new “Perspective” section to the review, leaving the conclusion section to sum-up the overall insights gained from documenting the years MNP research outputs. Access to the curated MNP data held in the Marinlit database² provides all the structural and literature data used to prepare this review.



Anthony R. Carroll

Anthony (Tony) Carroll initially studied the alkaloid and lignan chemistry of rainforest plants (BSc (Hons) and PhD, Prof Wal Taylor, Sydney University) but marine natural products became a major focus after postdoctoral fellowships at the University of Hawaii with Paul Scheuer and at James Cook University, Australia with John Coll and Bruce Bowden. Fifteen years as head of natural products chemistry for the AstraZeneca/Griffith University

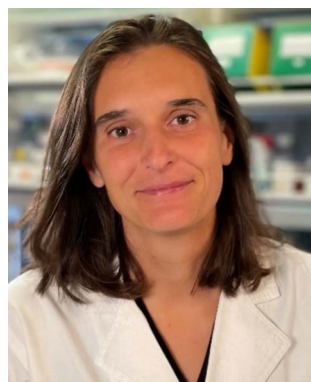
drug discovery project expanded his interests to include high throughput purification and structure determination techniques and cheminformatics. Since 2008 he has held a faculty position at Griffith University, Gold Coast where he is currently a Professor.



Brent R. Copp

Brent Copp received his BSc (Hons) and PhD degrees from the University of Canterbury, where he studied the isolation, structure elucidation and structure-activity relationships of biologically active marine natural products under the guidance of Professors John Blunt and Murray Munro. Two postdoctoral positions with Jon Clardy at Cornell and Chris Ireland at the University of Utah were then followed with a period spent

working in industry as an isolation chemist with Xenova Plc. In 1993 Brent returned to New Zealand to take a lectureship at the University of Auckland, where he is currently a Professor.



Tanja Grkovic

Tanja Grkovic received her MSc and PhD degrees from the University of Auckland under the supervision of Professor Brent Copp. She then carried out postdoctoral research at the National Cancer Institute with Kirk Gustafson, and Griffith University with Professor Ron Quinn. She is currently a Staff Scientist at the Natural Products Branch and the Molecular Targets Program at the National Cancer Institute where her research is focused on the

generation of prefractionated natural product libraries as well as the isolation and structure elucidation of natural products sourced from marine, plant, and microbial biota.



Robert A. Keyzers

Rob Keyzers carried out his BSc (Hons) and PhD studies at Victoria University of Wellington. His thesis research, carried out under the guidance of Assoc. Prof. Peter Northcote, a former contributor to this review, focused on spectroscopy-guided isolation of sponge metabolites. He then carried out post-doctoral research with Mike Davies-Coleman (Rhodes University, South Africa) and Raymond Andersen (University of British

Columbia, Canada) before a short role as a flavour and aroma chemist at CSIRO in Adelaide, Australia. He was appointed to the faculty at his alma mater in 2009 where he is currently an Associate Professor.



Michèle R. Prinsep

Michèle Prinsep received her BSc (Hons) and PhD degrees from the University of Canterbury, where she studied the isolation and structural elucidation of biologically active secondary metabolites from sponges and bryozoans under the supervision of Professors Blunt and Munro. She undertook postdoctoral research on cyanobacteria with Richard Moore at the University of Hawaii before returning to New Zealand to take up a lectureship at the University of Waikato, where she is currently a Professor.



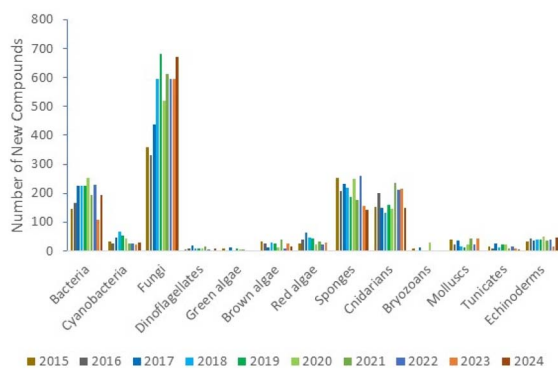


Fig. 1 Trends in new MNPs. The bars represent the total number of new MNPs reported each year over the last 10 years. Fungi MNPs only includes those isolated from submerged sources.

the decadal average. In contrast, numbers of MNPs isolated from fungi continue to rise and based on our changed reporting criteria, that now only includes fungi isolated from submerged environments, it is likely that 2024 had the highest number of new fungal MNPs reported over the 42-year history of this review. New MNPs reported from bacteria resurged to a decadal average level (Fig. 1).

2 Marine microorganisms and phytoplankton

2.1 Marine-sourced bacteria

Actinobacteria continue to be the most prolific source of MNPs reported from bacteria (with 165 new structures), even though they represent approximately 10% of total marine bacterial biodiversity. A sediment-derived *Actinomadura* sp. yielded five phenyl polyenes, maduraflavacins A–E **1–5**.³ Two rare genera of bacteria yielded MNPs for the first time in 2024, linear peptides cellulamides A **6** and B **7** were reported from *Cellulosimicrobium funkei*,⁴ and glycolipids testacosides A–D **8–11** were the first MNPs reported from *Microbacterium testaceum*.⁵ A sponge-derived *Microbispora* sp. strain yielded okichromanone, a chromanone NP isolated as a mixture of two interconverting epimers **12** and **13** due to hemiketal ring opening and recyclicalisation.⁶

The genus *Micromonospora* yielded two anthracyclines **14** and **15** and one anthraquinone **16**,⁷ three nona-2,7-dienoic acid derivatives apocimycins A–C **17–19**,⁸ as well as three spiroketal-containing macrolides **20–22**, and a diketopiperazine **23**.⁹ X-ray diffraction (XRD) analysis was used to define the absolute configuration of the co-occurring known MNP IB96212 **24** for the first time.⁹ A large scale (96 L) fermentation of *Nocardioopsis maritima* yielded two cyclic hexapeptides, maritiamides A **25** and B **26**, which showed moderate activity against *S. epidermidis* and weak activity against *E. coli* and *P. fluorescens*.¹⁰ Two α -pyrone polyketides, nocardioapyrones D **27** and E **28**, as well as an alkaloid, nocarterphenyl I **29**, were reported from a sediment-derived *Nocardioopsis* sp.¹¹ Nocarterphenyl I **29** possesses a rare 2,2'-bithiazole-*p*-terphenyl scaffold, the structure of which

was confirmed *via* XRD, and showed weak to moderate activity against four bacterial strains with MIC values ranging from 0.8 to 1.6 μM . Nocarterphenyl I showed no cytotoxicity against methicillin-resistant *S. aureus* (MRSA) or four human tumour cell lines (HTCLs).

The genus *Saccharomonospora* yielded four pyridine-, thiazole-, and chloroquinoline-containing alkaloids lodopyridones D–G **30–33**,¹² and two prenylated indole alkaloids, penipaline D **34** and *N*-acetyl-6-dimethylallyl-L-tryptophan **35**.¹³ Three sulfur-bridged angucycline dimers, spongisulfins A–C **36**, **37** and **38**, and one monomer, rubiginone A3 **39**, were reported from a sponge-derived *Spongiactinospora rosea*, with the structures of **37** and **39** confirmed *via* XRD.¹⁴

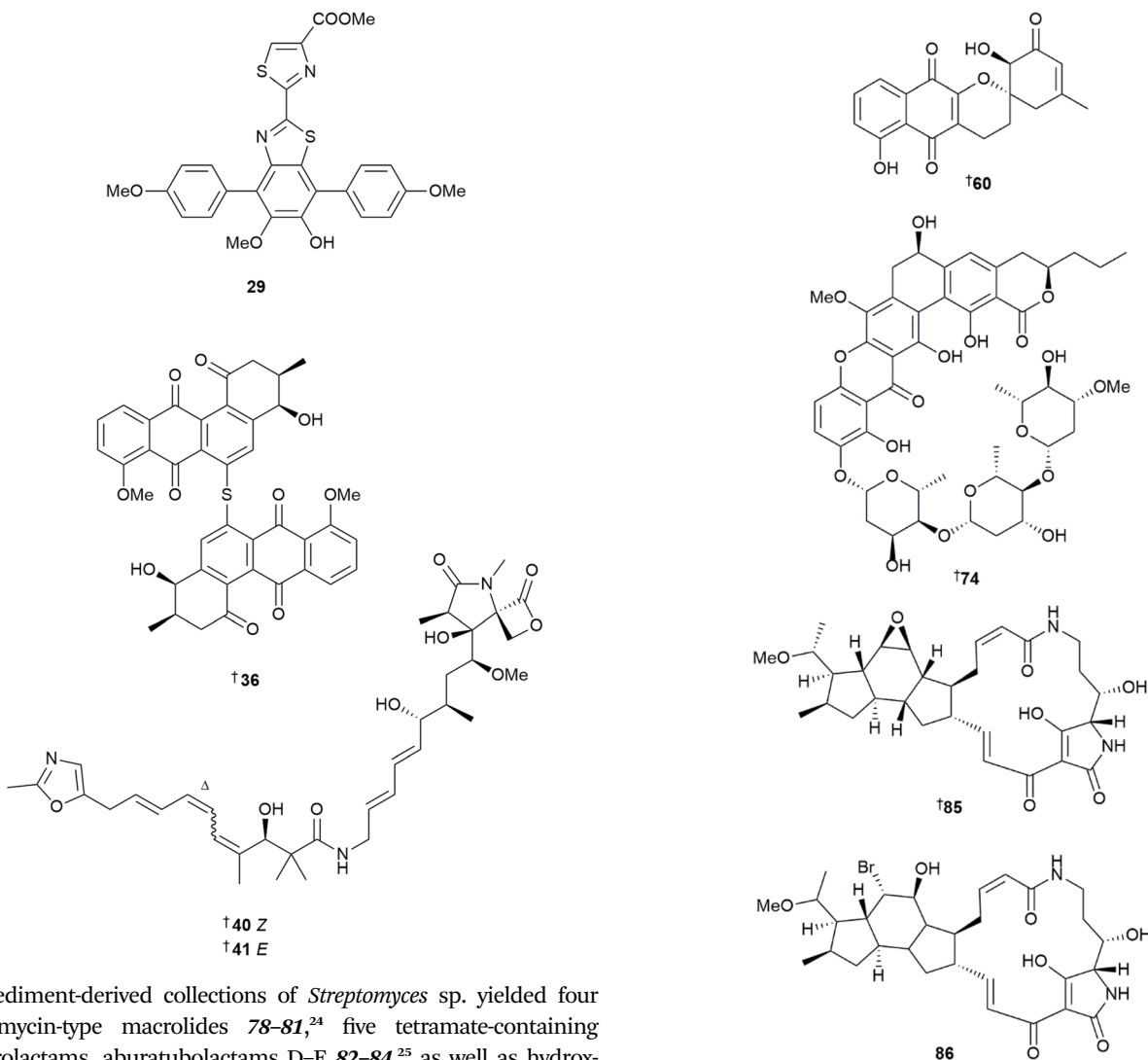
As for previous years, the genus *Streptomyces* was the most studied, with 125 new MNPs reported. A discovery campaign targeting the identification of terminal oxazole-bearing NPs using PCR screening for oxazole cyclase encoding genes coupled with NMR-based screening measuring $^1J_{\text{CH}}$ one-bond coupling constants specific to oxazole rings, identified two polyketide MNPs, methyl-oxazolomycins A **40** and B **41**.¹⁵ The two compounds showed moderate cytotoxicity against two estrogen receptor (ER) positive breast cancer cell lines MCF7 and T-47D, but no activity against ER-negative cell line MDA-MB-231.

Two linear polyketides, alpiniamides H **42** and I **43** were isolated from a sediment-derived collection of *Streptomyces* sp.,¹⁶ and a coral-derived *Streptomyces* sp. yielded polyketide chromone **44**, polyketide **45** and fatty acids **46** and **47**, and **48** and **49**, each as racemates but purified to separate each enantiomeric pair.¹⁷

A series of methylene-bridged, dimeric, 4-hydroxy-2*H*-pyran-2-one (HPO)-containing pyrones, the phaeochromycins M–U **50–59** were produced by a large-scale culture of sediment-derived *Streptomyces* sp.¹⁸ The authors showed that simple HPO-containing molecules can react with formaldehyde to generate methylene-bridged dimers, proposing a chemical defence mechanism where HPO-producing bacteria can sequester, and effectively neutralise formaldehyde from the environment for competitive gain. Cloning and heterologous expression of a type II polyketide BGC *spi1* sourced from a marine sediment-derived *Streptomyces* sp. into a *S. coelicolor* host, led to the isolation and identification of six angucyclines **60**, **61–65**, including spirocyclinone A **60** with a rare oxaspiro [5,5]undecane motif.¹⁹ Madeiron **66**, a new mixed terpene- and polyketide-derived MNP, was reported from a sediment-derived *Streptomyces aculeoletus*.²⁰

Glycosylated angucyclines reported from *Streptomyces* sp. included seven *C*-glycosides, chrysomycins F–J **67–71**,²¹ and grincamycins V **72** and U **73**.²² A mangrove rhizosphere-derived *Streptomyces* sp. yielded three glycosylated polycyclic xanthenes, kebanmycins A–C **74**, **75** and **76**, and the aglycone kebanmycin D **77**.²³ Kebanmycin A showed potent activity against *S. aureus*, including four MRSA isolates, with MIC values ranging from 0.125 to 0.5 $\mu\text{g mL}^{-1}$, but did not inhibit *B. subtilis* growth. Based on the annotations from the genome mining tool antiSMASH, the candidate BGC *keb* was proposed to be responsible for assembly of the compounds.





Sediment-derived collections of *Streptomyces* sp. yielded four ansamycin-type macrolides **78–81**,²⁴ five tetramate-containing macrolactams, aburatubolactams D–E **82–84**,²⁵ as well as hydroxycapsimycin **85** and brokamycin **86**.²⁶ Two 6,6-spiroketal-containing MNPs, streptospirodienoic acids D **87** and E **88** were reported from a coral-derived *S. cavourensis* together with two known compounds bafilomycins P **89**, and Q **90** whose absolute configurations were revised based on nOe correlation analysis, DP4+ calculations, and XRD data.²⁷ Compounds **89** and **90** showed moderate activity against the A-549 and HCT-116 HTCLs. Five oligomycins **91–95** were reported from a sediment-derived *Streptomyces* sp.,²⁸ four polyene macrolides **96–99**, were isolated from a mangrove-derived *S. hiroshimensis*,²⁹ and a sponge-derived *Streptomyces* sp. yielded a tricyclic polyene macrolactam, weddellamycin **100**.³⁰

Albusamides A–G, **101–107**, a series of new long-chain hydroxylated acetamide MNPs, were reported from *S. albus*,³¹ and a sediment-derived collection of *Streptomyces* sp. yielded additional acetamide-containing MNPs, streptothiomycin F **108** and *N*-(5-nitropentyl)acetamide **109**, the latter reported as a first time MNP.³² Albusamide C **103** displayed moderate cytotoxicity against a panel of fourteen HTCLs, but with no selectivity margin against the normal B lymphocyte cell line RPMI-1788. A genome-based mining strategy targeting the presence of the carpatamide BCG *ctd*, identified five acylated arylamines, carpatamides I–M **110–114** from *S. parvus*.³³

A series of shorter chain aryl amines **115–119**, a phenylpropanoid **120**, unsaturated fatty acids **121**, **122**, and two thiazinone-containing MNPs **123**, **124** were reported from a sediment-derived *Streptomyces* sp., with **119** and **121–124** representing new MNPs and others, known compounds reported from the marine environment for the first time.³⁴ A mudflat-derived *Streptomyces* sp. yielded a phenylthiazole anti-thiactin D **125**,³⁵ and heterologous expression of an NRPS BGC *grsc*, sourced from a marine sediment-derived *Streptomyces* sp. into three different *Streptomyces* surrogate hosts, yielded eight new thiazole-containing MNPs, grisechelins E **126**, F **127**, and I–N **128–133**, as well as two quinoline derivatives, grisechelins G **134** and H **135**.³⁶ Notably, grisechelin E **127** showed moderate antimycobacterial activity, with an MIC of 8 $\mu\text{g mL}^{-1}$ against *M. tuberculosis*.

A pyrrolo-pyrido-benzoquinone, streptoquinoneazaindole A **136**, a urea-containing pyrrole alkaloid, streptoureipyrrole A **137** and 6-hydroxybenzothiazole **138**, reported as a MNP for the first time, were isolated from a deep-sea sediment-derived *Streptomyces* sp.³⁷ A thioether-linked quinoline-quinazoline, quinosumycin **139** and a chromone, chromycone **140**, were reported from a sponge-derived *S. diastaticus*.³⁸ A dithiolopyrrolone, **141**



was isolated from a tunicate-derived *Streptomyces* sp.³⁹ and a staurosporine analogue, streptomholyrine A **142** was reported from a sediment-derived *Streptomyces* sp.⁴⁰

Pattern-based genome mining, combining mass spectrometry (MS) isotopic signature and biosynthetic sequence data, led to the identification of two chlorinated pyrrolketoidanes, indanopyrroles A **143** and B **144**, with their candidate BGC *idp* identified and biosynthetic assembly proposed.⁴¹ The trichlorinated indanopyrrole A showed moderate to potent activity against several Gram-positive and -negative bacteria, while the dichlorinated analogue indanopyrrole B was inactive.

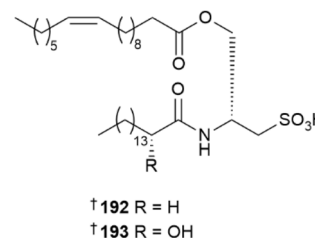
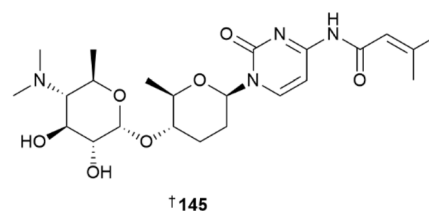
In a high-throughput screening (HTS) discovery campaign against *Mycobacterium avium* complex (MAC) pathogens, seven disaccharide pyrimidine nucleosides, mavintramycins A–G **145**, **146–151** were identified from a sediment-derived *Streptomyces* sp.⁴² Mavintramycin A **145** showed potent activity against two different MAC strains, with only moderate activity against two TB-associated mycobacterial strains and no cytotoxicity against other bacterial and fungal pathogens and one mammalian cell line. Moreover, **145** was found to be active against 40 clinically isolated *M. avium* strains, including those that were multidrug-resistant. Mechanism of action studies found that mavintramycin A binds to 23S ribosomal DNA and inhibits protein synthesis. The structure of **145** was later unequivocally confirmed *via* total synthesis.⁴³

A Korean sediment-derived *Streptomyces* sp. yielded six linear depsipeptides, homiamides A–C **152–154**,⁴⁴ and cavomycins A–C **155–157**,⁴⁵ all of which contained a core amino acid sequence of α -hydroxyisovaleric acid (Hiv)-valine (Val)-lactic acid (Lac)-valine (Val) residues like the valinomycin family of cyclic peptides. Three linear lipopeptides, albugamycins A–C **158–160** were isolated from a tunicate-derived *S. albidoflavus*,⁴⁶ with the structure of **158** confirmed *via* total synthesis. A sediment-derived *Streptomyces* sp. yielded *N*-acylated cyclic octapeptides, acyl-surugamides A1–A4 **161–164**,⁴⁷ with the structure of acyl-surugamide A2 **162** also reported, but not fully characterised, from a tunicate-derived *S. albidoflavus*.⁴⁸ A sponge-derived *Streptomyces* sp. yielded four glycosylated cyclopeptides, pyridapeptides F–I **165–168**,⁴⁹ and three cyclic lipopeptides olenamidinins A–C **169–171** were reported from a deep-sea derived *S. olivaceus*.⁵⁰ Six terpenoid structures reported from *Streptomyces* spp. included the iridoid lucknolide A **172**, reported as a MNP for the first time,⁵¹ and five labdanes, chlorolabdans A–C **173–175**, and epoxylabdane A **176** and B **177**.⁵²

Only one MNP from the phylum Bacteroidota was reported in 2024; an alga-derived *Algoriphagus* sp. strain yielded a new carotenoid, 2-hydroxyflexixanthin **178**.⁵³ Five MNPs were identified from the phylum Fimicutes, which included three glycosylated, 24-membered polyene macrolides, amylo-macrolactines A–C **179–181** sourced from a sediment-derived *Bacillus amylo-liquefaciens*,⁵⁴ and a new 25-membered polyene macrolide, macrolactin XY **182** and a new fatty alcohol **183** from a sponge-derived *B. subtilis*.⁵⁵

Fourteen MNPs were reported from the phylum Proteobacteria in 2024. A deep-sea sediment-derived *Alcanivorax dieselolei* strain yielded two *p*-aminoacetophenonic acid analogues, mohangic acids E **184** and F **185**,⁵⁶ and four lipopeptides,

bokeelamides A–D **186–189** were isolated from snail egg mass-derived *Ectopseudomonas khazarica*.⁵⁷ Two ureido-containing linear peptides, alteropeptilides A **190** and B **191** were reported from an arthropod-derived *Pseudoalteromonas flavipulchra*.⁵⁸ Structures of sulfur-containing amino lipids cysteinolides A **192** and B **193** were reported from bacterial species *Ruegeria pomeroyi* and *R. meonggei* and represent the first members of the sulfonolipid structural class of MNPs that have been fully characterised by NMR spectroscopy and their structures confirmed by total synthesis.⁵⁹ A seawater-derived *Vibrio ruber* yielded a new pyrrolidine, vibripyrrolidine A **194**, a piperazine vibriopiperazine A **195**, and two diazinanes, vibridiazinanes A **196** and B **197**, with **196** showing potent activity against *S. aureus* with a MIC value of 0.98 $\mu\text{g mL}^{-1}$ and no inhibitory activity against *E. coli*, *K. pneumoniae*, *P. aeruginosa*, or a panel of three HTCLs.⁶⁰



As in previous years, some MNPs reported from marine bacteria did not have adequate spectrometric or NMR spectroscopic data in full support of the proposed structures and were omitted from this review.^{61–64} Total synthesis of the reported structure of marinoaziridine B⁶⁵ revealed significant inconsistencies in the ¹H and ¹³C NMR spectroscopic data, suggesting the structure of this MNP should be revised. Total syntheses of other bacteria-sourced MNPs reported in 2024 included bacillimidazole B and discolins A, B, and E,⁶⁶ bacilotetrin C,⁶⁷ das-sonmycins A and B,⁶⁸ caerulomycin K,⁶⁹ cihunamide B,⁷⁰ cyclomarin,⁷¹ discoipyrroles A–C,⁷² (+)-dixiamycin,⁷³ homo-seongomycin,⁷⁴ iedomycins A and B,⁷⁵ incarnatapeptins A and B,⁷⁶ iturin A,⁷⁷ lorneic acids C and D,⁷⁸ marformycins A and D,⁷⁹ mavintramycin A,⁴³ moiramide B,⁸⁰ octalactins A and B,⁸¹ rufo-mycins 1, 2, 4, 11, 22, and 24,⁸² salimabromide,⁸³ and seal-utomicin C.⁸⁴

Reviews focused on bacterial MNPs published during 2024 included a summary of metagenomic strategies for the discovery of MNPs,⁸⁵ and actinomycete promoter engineering approaches.⁸⁶ The bacterium *B. subtilis* was reviewed as a versatile host for heterologous expression of NP BGCs,⁸⁷ as was the regulation of acyl homoserine lactone-based quorum



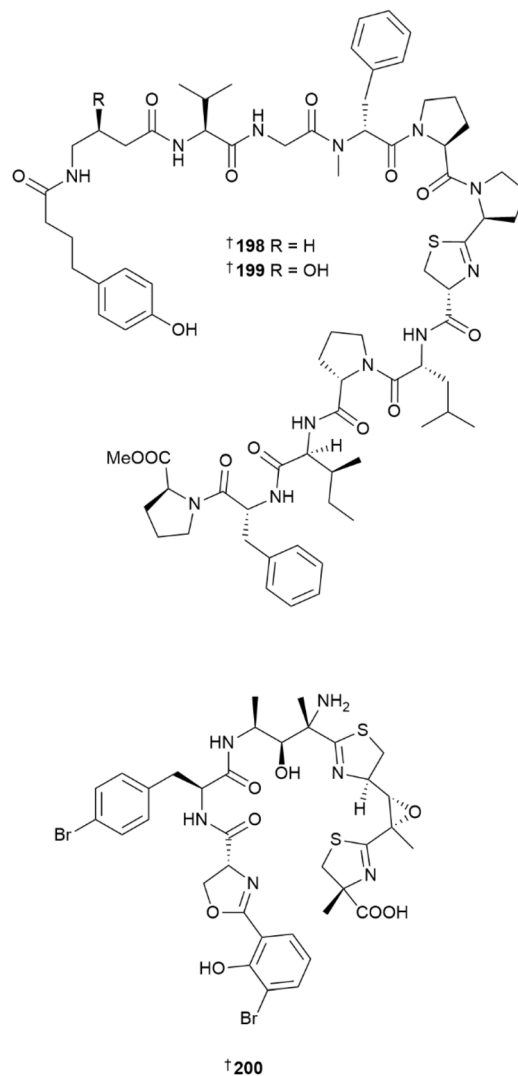
sensing effects on the biosynthesis of bioactive molecules in bacteria.⁸⁸

Structural classes of bacterial NPs reviewed included polyacetylenes,⁸⁹ alkaloids,⁹⁰ and γ -pyrones.⁹¹ Groups of marine bacteria reviewed for biosynthetic potential and the diversity and activity of MNPs produced included the MAR4 clade of the genus *Streptomyces*,⁹² and marine-sourced *Pseudomonas* sp.⁹³ Biological activity studies of previously reported bacteria-sourced MNP structures included *in vitro* and *in vivo* anti-*Toxoplasma gondii* activity assessment of a series of marinoquinolines,⁹⁴ mechanism of action (MoA) studies of marinopyrrole, pentachloropseudilin, and pentabromopseudilin,⁹⁵ tartrolon D,⁹⁶ and tunicamycins,⁹⁷ and structure–activity relationship (SAR) reports of thiomarinol A,⁹⁸ and sunshinamide.⁹⁹

2.2 Cyanobacteria

Two linear lipopeptides, pemuchiamides A **198** and B **199** were isolated from a relatively large-scale (3 kg) Japanese collection of the filamentous cyanobacterium *Hormoscilla* sp.¹⁰⁰ The compounds contain 13 amino acid residues including four prolines and several unusual ones such as 4-aminobutanoic acid, 4-(4-hydroxyphenyl)butanoic acid and *N*-methyl-*D*-phenylalanine. Pemuchiamide A showed moderate activity against *Trypanosoma brucei rhodesiense* (IC₅₀ = 0.63 μ M), while the hydroxylated analogue pemuchiamide B was an order of magnitude less active. This indicated that the 4-aminobutanoic acid moiety may be an important pharmacophore feature involved in the antitrypanosomal activity for this peptide series. Laboratory cultures (some up to six months in length) of three geographically dispersed collections of *Lepthotoe* strains yielded three new PKS/NRPS hybrid linear peptide metallophones, leptochelins A–C **200**, **201** and **202**.¹⁰¹ The leptochelin structures were challenging to elucidate since they contain modified amino acids and isolated stereogenic centres. Their structures were deduced using a combination of tandem mass spectrometry, NMR spectroscopy, and bioinformatic analyses of the putative leptochelin BGC *lec*. The leptochelins bind to zinc, copper, iron, and cobalt and were moderately cytotoxic against four HTCLs. They are also the first reported MNPs from the genus *Lepthotoe*.

Studies on *Moorena* spp. yielded seven MNPs. A glycosylated macrolide, moorenaside **203** was reported from a Florida collection of *Moorena* sp.¹⁰² and a Guam-sourced collection yielded two polyketides **204** and **205**.¹⁰³ A co-culture of *M. producens* with *Candida albicans* led to significant upregulation of the production of a linear depsipeptide hectoramide B **206**. Its putative BGC *hca* was identified and its biosynthetic assembly annotated.¹⁰⁴ Even though the presence of a fungal competitor was crucial to produce hectoramide in liquid culture, when tested against two *Candida* strains, **206** did not show any antifungal activity. *M. bouillonii* collected in India, yielded a linear lipodepsipeptide, kavaratamide A **207**,¹⁰⁵ and a cyclic depsipeptide, wajepeptin **208** was isolated from a Japanese collection of *Moorena* sp.¹⁰⁶ A benthic collection of *Moorena* sp. (annotated as *Lyngbya* sp.) in Indonesia yielded a cyclic peptide PM170453 **209**, the structure and absolute configuration of

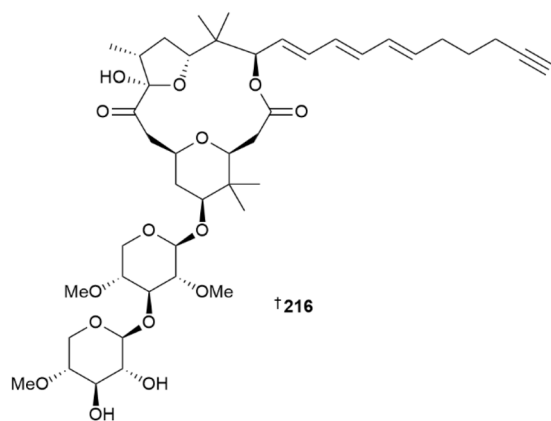


which was unequivocally confirmed *via* total synthesis.¹⁰⁷ Six lipopeptides, noducyclamides A1–A4 **210–213** and noducyclamides B1 **214** and B2 **215**, were reported from a laboratory culture of *Nodularia* sp.¹⁰⁸ The dodecapeptide noducyclamide B1 was weakly cytotoxic against the MCF7 breast cancer cell line.

The genus *Okeania* yielded six new MNPs. A glycosylated, 16-membered macrolide, polycavernoside E **216**, was reported from a coastal Okinawan collection of *Okeania* sp.¹⁰⁹ Related analogues of this MNP, the polycavernosides A–C were originally reported from the red alga *Polycavernosa tsudaii*^{110–112} and were implicated in human illness and death from consumption of this edible seaweed in Guam. An additional congener, polycavernoside D, was later identified from a Caribbean collection of the cyanobacterium *Okeania* sp.,¹¹³ and this latest report of **216** provides increasing evidence that polycavernosides are produced by cyanobacteria rather than red algae. Polycavernoside D did not show significant *in vitro* cytotoxicity against normal human fibroblast WI-38 cells, was weakly active against *T. b. rhodesiense*,¹⁰⁹ but demonstrated potent *in vivo* toxicity in mice with an ip. lethal dose of 0.81 mg kg⁻¹.¹¹⁴ Additional *Okeania* sp. collections yielded a lipopeptide, *N*-



desmethylmajusculamide B **217**,¹¹⁵ cyclic peptide okeaniazole A **218**,¹¹⁶ and a linear lipopeptide, amantamide C **219**.¹¹⁷ Terpenoids continue to be reported from cyanobacterial sources, with kagimminols A **220** and B **221** representing the first examples of cembrene diterpenoids isolated from the genus *Okeania*.¹¹⁸



An unidentified cyanobacterium with morphological resemblance to *Symploca* sp. collected in Guam, yielded three enyne-containing chlorinated fatty acid amides, taveuniamides L–N **222–224** and the absolute configuration of the C-8 stereogenic centre was reported for this MNP class for the first time.¹¹⁹ Using genome-mining and metabolomic strain prioritization strategies, two dialkylresorcinol polyketides, 29-dechloro-29-hydroxybartoloside A **225** and 17-dechloro-17-hydroxybartoloside A **226**, were isolated from *Synechocystis salina*,¹²⁰ with hydroxylation along the aliphatic side-chain in both compounds representing a novel structural modification of the bartoloside family of polyketides. Finally, a methyl-branched fatty acid, jobosic acid **227** was reported from an unidentified bacterial biomass collected in Puerto Rico.¹²¹ Cyanobacteria remain one of the few marine taxa where reports of new MNPs from mixed assemblages or incomplete taxonomic identifications continue to be accepted for publication.

The total synthesis of the cyclic depsipeptide lagunamide C was achieved,¹²² resulting in the revision of the proposed structure from a 27-membered macrocycle originally published¹²³ to the 26-membered odoamide,¹²⁴ a known cyanobacterial MNP. Other cyanobacterial MNPs with reported total syntheses included dragocins A–C,¹²⁵ irijimaside A,¹²⁶ jamaicamide B,¹²⁷ majusculamide D,¹²⁸ rivularin A,¹²⁹ serinolamides A and B,¹³⁰ and trichophycin-I.¹³¹

Reviews included summaries of anti-infective,¹³² anti-protozoal,¹³³ and cancer-related¹³⁴ activities of cyanobacterial MNPs, as well as a comprehensive review of the diversity of NP structures and biological activities reported from the genus *Oscillatoria*.¹³⁵ Notable work on the biosynthesis of cyanobacterial MNPs included the identification and characterisation of the putative BGC for the assembly of dolastatin, reported from *Caldora penicillata*,¹³⁶ and SAR studies of known cyanobacterial compounds biselyngbyolide B,¹³⁷ carmaphycin B¹³⁸ iezoside,¹³⁹ and majusculamide D.¹⁴⁰ Environmental reports of cyanobacterial MNPs with broad implications to human health included a review on impacts of cyanobacterial blooms on fish

farming,¹⁴¹ and the first account of bioaccumulation of lyngbyatoxin A in edible shellfish along the coast of New Zealand.¹⁴² This, together with a report of acute toxicity of a new poly-cavernoside analogue **216** in mice, accentuates the importance of accurate annotation, full structural characterisation, and monitoring of the occurrence of cyanobacterial MNPs that can impact marine ecosystems and human health.

2.3 Marine-sourced fungi

Co-culture of an *Acremonium* species with heat-killed *Pseudomonas aeruginosa*, resulted in isolation of acremosides A–G **228–234**, linear sesquiterpenoids with a 2-phenyl acetoxy substituent conjugated to a sugar alcohol.¹⁴³ Other *Acremonium* strains were the source of the meroterpenoids, acremochlorins A **235** and B **236**, pyridine alkaloid acremopyridone A **237** and cyclopentenone derivative acremocketene A **238**,¹⁴⁴ sorbicillin-like compound **239** and acremonilactone **240**,¹⁴⁵ and ascochlorin glycoside **241**.¹⁴⁶

Fermentation of *Albifimbria verrucaria* yielded the nonenes, verrucanonenones A–C,¹⁴⁷ **242–244** and tetrahydrofuran-containing polyketide derivatives¹⁴⁸ **245–248**, the butanolide derivative, alterbutenolide **249** was obtained from *Alternaria alternata*,¹⁴⁹ and *Arthrinium arundinis* cultures, respectively derived from a sea anemone and a sponge, yielded arthrinic acid **250**, hexylaconitic anhydride methyl ester **251**, nonanoic acid derivative **252**, arthripenoids G **253** and H **254**,¹⁵⁰ and alternapyrone derivatives, alternapyrones G **255** and H **256**. The BGC for alternapyrone biosynthesis was identified and heterologous expression led to isolation of an additional alternapyrone, alterpyranone I **257**.¹⁵¹

As has been the case in many previous years, the genus *Aspergillus* was the most common source of fungal metabolites. Co-culture of deep-sea-derived *Aspergillus aculeatinus* with a mangrove-derived *Penicillium* strain yielded the paraherquamides, aculeaquamides A **258** and B **259**,¹⁵² while addition of sodium bromide to growth media of *A. alliaceus* resulted in production of brominated anthraquinone monomer **260** and dimers **261–263**.¹⁵³ *p*-Terphenyl **264**, **265** and diphenyl **266**, **267** ether derivatives were obtained from *A. candidus*,¹⁵⁴ whilst the benzophenone derivatives, carneusones A–F **268–273** were isolated from a culture of *A. carneus*.¹⁵⁵ *A. chevalieri* and *A. fischeri* respectively yielded prenylated indole diketopiperazine **274** and prenylated indole derivative **275**,^{156,157} ascidian-derived *A. clavatus* was the source of 12*S*-deoxynortryptoquivaline **276**,¹⁵⁸ co-culture of *A. insulacola* with a mangrove-derived *Alternaria* strain led to isolation of cyclic tetrapeptides, violacetoides B–E, **277–280**,¹⁵⁹ miniaturised cultivation profiling enabled isolation of 2,6-diketopiperazines, noonazines A–C **281–283**, and azaphilone, noonaphilone A **284**,¹⁶⁰ and a nitrobenzoyl sesquiterpenoid **285** was obtained from culture of *A. ochraceopetaliformis*.¹⁶¹

Stephachratidin A **286**, an unusual stephacidin-asperochratide hybrid, was isolated from culture of deep-sea-derived *A. ochraceus*.¹⁶² Several deep-sea sediment-derived *Aspergillus* strains yielded a range of metabolites. The sesquiterpenoid malfilanol C **287** was isolated from a culture of *A.*



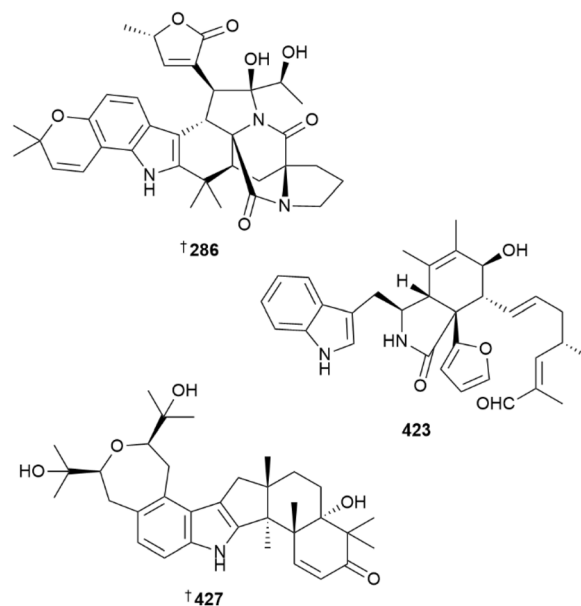
punicus, along with known terrestrial but new MNP, maliflanol B **288**,¹⁶³ phenol derivatives **289–298**, (the last a known synthetic compound but new NP), were obtained from *A. subversicolor*,¹⁶⁴ and separate cultures of *A. sydowii* yielded sesquiterpenoid **299** and sydowimide A **300**,^{165,166} the latter by epigenetic modification. Another culture of *A. sydowii* was the source of xanthone-alkaloids sydoxanthones F–M **301–311**,¹⁶⁷ and a prenylated indole alkaloid, aspertaichamide A **312** was obtained from *A. taichungensis*.¹⁶⁸

Cultures of *A. terreus* were the source of a range of metabolites including caryophyllene-type sesquiterpenoids, asperporonins A **313** and B **314**,¹⁶⁹ ergone derivatives **315–324**,¹⁷⁰ (all deep-sea-derived), sulfur containing metabolites asperterretals L **325** and M **326** and terreins A **327** and B **328**,¹⁷¹ brominated aromatic butenolides **329–342**,¹⁷² meroterpenoids **343–348**,¹⁷³ butenolide derivatives **349** and **350** (via epigenetic manipulation and the latter a known synthetic compound but new NP)¹⁷⁴ and a suite of azaphilones, **351–361**, which were produced through activation of a cryptic BGC.¹⁷⁵

Some polyketides **362–365**, and a nucleoside **366** were isolated from a culture of *A. versicolor*,¹⁷⁶ while a range of metabolites were obtained from *Aspergillus* strains from a variety of sources. Sponge-derived *Aspergillus* strains yielded ergostane-type steroid **367**,¹⁷⁷ terminal olefin-containing unsaturated fatty acid **368**, 2-carboxy-2'-methyl-azobenzene **369** and 1,3,9-trimethyluric acid **370** (the last two known synthetic compounds but new NPs).¹⁷⁸ Sediment-derived *Aspergillus* strains yielded aspergilalkaloid A **371**,¹⁷⁹ asperindopiperazines A–C **372–374**, 5-methoxy-8,9-dihydroxy-8,9-deoxyaspyrone **375**, and 12*S*-aspetetranone D **376**,¹⁸⁰ emericelactones F **377** and G **378**, preshamixanthones **379** and **380**, 25-*O*-methylarugosin A enantiomers **381** and **382**, furanones, 9-hydroxymicroperfurane **383** and microperfurane **384**, (these last four known terrestrial but new MNPs), phthalimidinic acids **385** and **386**, aspergilol G **387**, anthraquinone **388**, and propionic acid lactone derivative **389**,¹⁸¹ (the last a known synthetic compound but new NP). A seawater strain of *Aspergillus* yielded ceramide **390** while a coral-derived *Aspergillus* strain yielded megastigmanones A–C **391–393** and prenylterphenyllin H **394** and a starfish-derived strain yielded brominated isocoumarin **395**.^{182–184} The production of **395** resulted from addition of bromide salts to the culture medium.¹⁸⁴ *Aspergillus* species from mangrove sediments have yielded a nor-diterpenoid acid, asperbrunneo acid **396**,¹⁸⁵ indole alkaloids asperdinones A–H **397–404**,¹⁸⁶ isoprenyl phenol **405**,¹⁸⁷ and alkyl resorcinols **406–408**.¹⁸⁸

Culture of *Aureobasidium melanogenum* yielded aliphatic δ -lactones **409–411** and fatty acid methyl esters **412–414**,¹⁸⁹ sterol **415** was isolated from a culture of *Beauveria* sp.¹⁹⁰ and triterpenoid **416** was obtained from shellfish-derived *Ceriporia lacerata*.¹⁹¹ Sesterterpenoids with a 5/8/6/5 tetracyclic ring system, sesterchaetins A **417** and B **418** and diepoxide polyketides **419** and **420**, were isolated from deep sea sediment-derived *Chaetomium globosum*,¹⁹² while sponge-derived *C. globosum* yielded six new cytochalasins, marcytoglobosins A **421** and B **422** and marchaeoglobins A–D **423**, **424–426**, of which **423** is the first furan-containing cytochalasin.^{193,194}

Aseptepenedol A **427**, an indole sesquiterpenoid obtained from culture of a *Chloridium* strain, contains a seven-membered ether ring as part of a 7/6/5/5/6/6 hexacyclic framework.¹⁹⁵



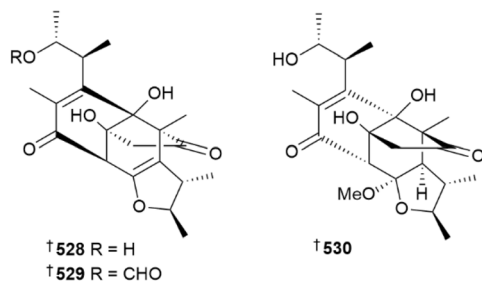
Cladosporium strains were the source of a range of new metabolites. Dodecanoic acid derivatives **428–431** (the last two known as semi-synthetic compounds) were isolated from a culture of *C. cladosporioides*,¹⁹⁶ *C. halotolerans* yielded pyrone derivatives **432** and **433**,¹⁹⁷ drimane sesquiterpenoids **434**, isocoumarin **435**, coumarin **436** and known synthetic compound, 9-formylfluorene **437** as a new NP,¹⁹⁸ and a further *Cladosporium* strain was the source of isochromanone **438**.¹⁹⁹ *Curvularia aeria* and *C. lunata* respectively yielded tricyclic aromatic polyketide **439** (as a racemic mixture) and resorcylic acid lactones **440–443** and isolation of curdepsidone A from a *Curvularia* strain led to revision of the structure to **444**.^{200–202}

A one strain many compound (OSMAC) approach to cultivation of *Emericellopsis maritima* resulted in isolation of eremophilane sesquiterpenoids **445–447**,²⁰³ 6/7/6 tricyclic diterpenoid eutyditerpenoid A **448** and cytochalasin **449** were obtained from a culture of *Eutypella scoparia*,²⁰⁴ whilst another *Eutypella* strain yielded fifteen new sesquiterpenoids, eutypelides A–O **450–464**.²⁰⁵ Sesquiterpenoids, including eremophilane derivatives fureremophilanes A–D **465–468** and acorane analogues furacoranes A **469** and B **470**, were isolated from culture of *Furcaterigmium furcatum*.²⁰⁶ *Fusarium* strains were the source of fusarochromanone derivatives **471** and **472**, hydroxyphenylacetic acid **473–476** and hydroxyphenylethanol derivatives **477** and polyketide **478**.^{207–209} The cyclic pentapeptides, avellanins D–O **479–490** were obtained from culture of *Hamigera ingelheimensis*.²¹⁰ An *Irpex* strain produced the heptaketides irpetones A **491** and B **492**,²¹¹ catechol derivatives, meirols A–C **493–495** were obtained by fermentation of a *Meira* strain,²¹² whilst a *Microascus* strain yielded eight decahydrofluorene alkaloids, **496–503**.²¹³ Azaphilones **504–506** and a dihydroisocoumarin **507** were isolated from a strain of



Neopetalotiopsis,²¹⁴ mangrove sediment-derived *Nigrospora oryzae* contained four aniline derivatives **508–511**,²¹⁵ spiro trione compound **512** and asparvenone derivatives **513–516** were obtained from a culture of *Paraconiothyrium sporulosum*,²¹⁶ and a culture of *Parengyodontium album* was the source of aromatic polyketide alternaphenol B2 **517**.²¹⁷

The *Penicillium* genus has proven once again to be a prolific source of new metabolites. A phenyl 6,7-dihydroxygeranyl ether derivative **518** was obtained from a *P. arabicum* culture,²¹⁸ *P. brasilianum* yielded diketopiperazine alkaloids **519** and **520**,²¹⁹ soft coral-derived strains of *P. chrysogenum* were the source of aromatic heterocycles **521–525**, and a sorbicillinoid **526**,²²⁰ and of anthranilic acid-peptide derivative **527**.²²¹ Dicitrinols A–C **528–530**, citrinin derivatives with a 6/5/7/5 core, were obtained from *P. citrinum* derived from hydrothermal vent sediment,²²² a culture of *P. corylophilum* yielded enantiomeric hydroxyphenylacetic acid derivatives **531–534** and α -pyrone analogue **535**,²²³ and co-culture of deep-sea sediment-derived *P. crustosum* with mangrove-derived *P. citrinum* resulted in production of the dihydropyrones, rhytismatones C **536**, and D **537**.²²⁴



The relative and absolute configuration of the known indolone, notoamide X **538** was assigned for the first time. It was obtained from the mangrove sediment-derived *P. janthinellum* along with three other indolones with proposed new structures. However, the structures of these additional compounds were only proposed based on MS data and so are not included here.²²⁵ Culture of *P. oxalicum* derived from a red alga yielded prenylated indole alkaloids **539** and **540** and polyoxygenated steroid **541**,^{226,227} sediment-derived strains, *P. pancosmium* and *P. rubens* respectively yielded meroterpenoids **542** and **543** (*P. pancosmium*)²²⁸ and polyketide derivatives **544** and **545** and amino-bis-tetrahydrofuran derivatives **546** and **547** (*P. rubens*),²²⁹ while a coral-derived *P. sclerotium* culture was the source of azaphilone pigments **548–559**.²³⁰ Highly oxygenated meroterpenoids, penisimplinoids A–K **560–570**, were isolated from a *P. simplicissimum* culture,²³¹ steroid **571** and polyketides **572** and **573** were obtained from *P. variable*²³² and seagrass-derived cultures of *P. velutinum* and *P. yezoense* respectively yielded piperazine derivative **574**,²³³ and decalin polyketides **575–584**.²³⁴ Meroterpenoids, including **585–591** and penicianstinoids F–K **592–597**, indole diterpenoids, including **598–604** and penpaxilloids F **605** and G **606** and diketopiperazine alkaloid **607** (the last as a racemate), were isolated from cultures of various *Penicillium* strains.^{235–239} Sediment-derived strains were the source of azaphilones **608–**

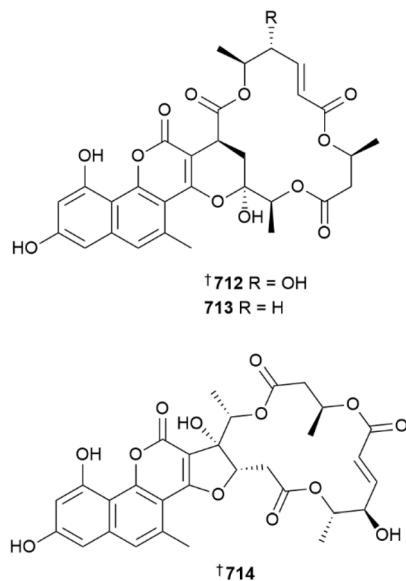
612 and clavatul derivative **613**,^{240,241} and *Penicillium* strains derived from seawater yielded racemic polyketides **614** and **615** and enantiomeric hydroxyphenylacetic acid derivatives **616** and **617**,^{242,243} which were separated by chiral chromatography. Cultures of various *Penicillium* strains also resulted in production of racemic phenalenone **618**,²⁴⁴ quinolone-citrinin hybrids **619** and **620** and speradines I **621** and J **622**.^{245,246}

Mangrove sediment or root-derived *Penicillium* spp. have also yielded pyridyl derivatives **623** and **624**, indole **625**,²⁴⁷ isobenzofuranone derivative **626**, α -pyrone **627**, quinone **628**, phenol derivative **629**,²⁴⁸ steroids **630** and **631**, benzopyran **632**,²⁴⁹ diterpenoid indole alkaloid derivatives **633** and **634**,²⁵⁰ sesquiterpenoids eupenicisirenins C–O **635–647**,²⁵¹ aryl and decalin polyketide acids **648–669**,²⁵² and alkyl resorcinols **670–673**.²⁵³

Peniotrinins A–F, comprising cytochalasin derivatives **674** and **675**, citrinin derivatives **676–678** and tetramic acid derivative **679**, were isolated from a culture of a *Peniophora* strain²⁵⁴ and merocytochalasans, perochoalasin A **680**, and inseparable C20 epimeric pairs **681** and **682** were obtained from a *Peroneutypa* species. The BGC responsible for cytochalasin biosynthesis was identified through full genome sequencing and the biosynthesis elucidated through feeding experiments utilising ¹⁵N and deuterium labelled hydroxylamine.²⁵⁵ *Pestalotiopsis* strains were the source of rearranged pimarane diterpenoid **683** and known terrestrial but new MNPs, nodulisporones A **684** and B **685**,²⁵⁶ and polyketide derivatives, pestalotiopols E–J **686–691** (the last two as a racemic mixture).²⁵⁷ Cultures of deep-sea sediment-derived *Phomopsis* strains yielded benzoic acid derivative **692** and 3-carboxy-indole derivatives **693–695**.^{258,259} Co-culture of two *Phomopsis* strains obtained from the roots of the Chinese mangrove *Rhizophora mangle* yielded pyrazine **696**, steroid **697** and α -pyrone **698** (from *P. asparagi*)²⁶⁰ and bis-chromones phomoxanthenes L–N **699–701** (from *Phomopsis* sp.),²⁶¹ none of which were produced in monoculture of the individual strains.

Sediment-derived fungal strains have yielded the sesquiterpenoids **702–711** (from *Pseudallescheria boydii*)²⁶² naphthopyrone-macrolide hybrids, gymnoasins A–C **712–714** and glucolipid **715** (from *Pseudogymnoascus* strains) and sesquiterpenoids **716** and **717** (from *Pyrroderma noxium*).^{263–265} Isolated from the Antarctic, the gymnoasins A–C **712–714** are the first naturally occurring compounds of their type and a biomimetic synthesis of **712** was achieved.²⁶³ Deep-sea sediment-derived fungal strains were the source of bisabolene sesquiterpenoid **718** (from *Retroconis fusiformis*),²⁶⁶ sesquiterpenoid-hydroquinone hybrids, saromacrophorins A–C **719–721** (from *Sarocladium terricola*)²⁶⁷ and 4-hydroxyphenyl derivative **722** (from *Scedosporium apiospermum*), which displayed moderate to potent activity against a panel of *Candida auris* strains,²⁶⁸ while mangrove sediment-derived *Roussouella* sp. contained two sesquiterpenoids **723** and **724** and a shikimate **725**.²⁶⁹





Dihydroisocoumarins, exserolides L **726** and M **727** were obtained from culture of a *Setosphaeria* species,²⁷⁰ while various *Simplicillium* strains yielded a range of metabolites including nortriterpenoids **728–730**,²⁷¹ peptaibiotics **731–741**,²⁷² dipeptide **742** and acylated valine derivative, siamysin **743** (the last discovered *via* heterologous expression of a cryptic BGC).^{273,274} Eleven brominated depsidones **744–754** were isolated from a culture of *Spiromastix* sp.,²⁷⁵ *Stachybotrys* strains yielded atranones V–Z **755–759** and dolabellane diterpenoids **760–762**,²⁷⁶ and phenylspirodrimanes, stachybotrins K **763** and L, **764**,²⁷⁷ and cyclic tetrapeptides, endolides E **765** and F **766** were obtained from a *Stachylidium* strain. The endolides contain the unusual amino acid, *N*-methyl-3-(3-furyl)-alanine which was used in fragment pattern searches of tandem mass spectrometry data, in conjunction with molecular networking, to assist both isolation and structural elucidation of these metabolites.²⁷⁸

Culture of *Talaromyces* strains isolated from sediment have yielded a diverse array of metabolites, including oligophenalenone dimers **767–775**,²⁷⁹ phenylhydrazone alkaloids **776–779**,²⁸⁰ highly oxygenated *seco*-terpenoids **780–785**,²⁸¹ macrolactin **786**,²⁸² highly oxygenated phenol derivatives **787–789**, isocoumarin glycosides **790** and **791** and naphthalene glycoside **792**.²⁸³ Culture of *T. purpureogenus* yielded six prenylated indole diketopiperazine alkaloids, talaromyines A–F **793–798**,²⁸⁴ cyclic heptapeptides **799** and **800**, nor-diterpenoids **801** and **802**, diterpenoid acids **803** and **804** and triterpenoid **805** were obtained from cultivation of *T. scorteus*,²⁸⁵ *T. siglerae* yielded cyclic heptapeptides, talaromides A–C **806–808** and *T. variabilis* was the source of hybrid pentaketide sesquiterpenoids **809–811**.^{286,287} Seawater-derived *Talaromyces* strains yielded meroterpenoids **812–814** and benzoquinone **815** and furopyridinone **816** derivatives,^{288,289} a strain derived from sediment was the source of ring-opened azaphilone derivative **817**,²⁹⁰ thirteen new polyketide derivatives **818–830**, (6-hydroxy-4-methoxycoumarin **829** a known synthetic compound but new NP) were obtained from a deep-sea mussel-derived strain,²⁹¹ and duclauxin analogues, taladuxins A–N **831–844** were isolated from a culture of a coral-derived *Talaromyces* strain from which

verruculosin B was also isolated and the configuration revised to *1S* as in **845**.²⁹² Investigations of mangrove sediment-derived *Talaromyces* spp. have resulted in isolation of γ -lactams, talarolactams A–D **846–849**,²⁹³ sesquiterpenoids, talaroterpenoids A–D **850–853**,²⁹⁴ *p*-terphenyl derivatives, talaroterphenyls A–D **854–857**,²⁹⁵ diketopiperazines, **858–860**, a cyclopentenone **861**, an α -pyrone **862**,²⁹⁶ and linear polyketides **863–868**.²⁹⁷ Two of the talaroterphenyls **855** and **856** potentially inhibited the enzyme phosphodiesterase-4 (PDE4) linked to inflammation.

Tetramic acid alkaloids, tolypyridones I **869** and J **870** were isolated from a culture of *Tolypocladium cylindrosporium*,²⁹⁸ *Trichoderma* strains were the source of sesquiterpenoid derivatives **871–874**,²⁹⁹ bergamotene-derived sesquiterpenoids **875–882**, (through heterologous expression of a cryptic BGC),³⁰⁰ cyclohexane sesquiterpenoid **883**,³⁰¹ γ -butyrolactones **884–889** and monomeric sorbicillinoids **890–893**.^{302,303} Steroid derivatives **894** and **895** and linear polyketide **896** have been isolated from mangrove sediment-derived *Trichoderma* spp.^{304,305} and culture of *Xylaria acuta* yielded a range of metabolites; lactones **897**, **898**, **899**, glucopyranosides **900** and **901** and mannopyranoside **902**, epoxychothalasins **903** and **904** and butylitaconic acid **905**.³⁰⁶

Total synthesis of acremolides A and B has been achieved and the absolute configurations established as **906** and **907**,³⁰⁷ while asymmetric syntheses of aspilactonol F and aspiketo-lactonol were achieved in eight and ten steps respectively and their absolute configurations determined as **908** and **909**.³⁰⁸ (\pm)-Notoamide N was prepared in a biomimetic approach *via* synthesis of (\pm)-stephacidin A,³⁰⁹ and several prenylated indole alkaloids of the aspersiamide family, namely aspersiamides B, D, E, G and J and dihydrocarneamide A, were prepared in biogenetically patterned total syntheses. The structure originally proposed for aspersiamide A was shown to be incorrect with a revised structure proposed but not yet proven.³¹⁰ A unified approach was utilised to prepare cyclic tetrapeptide JM-47,³¹¹ cyclocondensation of an electrophilic homoserine lactone was employed in the total syntheses of polonimides A and C³¹² and synthesis of bipenicilisorin was achieved through initial preparation of its monomer penicilisorin and the absolute configuration was established as **910**.³¹³ Penostatin D was prepared in an asymmetric synthesis from *L*-ascorbic acid,³¹⁴ 12 β -hydroxy conidiogenone C was also obtained in an asymmetric total synthesis³¹⁵ and total synthesis of the cyclic peptide, talarolide A showed that it (and by inference, talarolides C and D), occur naturally as atropisomers.³¹⁶ Racemic total synthesis of phomopsol B has been reported³¹⁷ and syntheses of aspersiocoumarin A and fusarimatin C were achieved from commercially available starting materials.³¹⁸

Alternariol monomethyl ether was shown to relieve food allergy symptoms in mice and reduce histamine release into blood serum,³¹⁹ the verticillin analogue, leptosin A showed potent inhibition of the parasite *Toxoplasma gondii*,³²⁰ brevianamide F exhibited antithrombotic activity in a zebrafish model³²¹ and agonodepside B showed potential as a photoprotection agent.³²² Fumetremorgin derivatives 12*R*,13*S*-dihydroxyfumitremorgin C and tryprostatin A displayed anti-hypercholesterolemia activity in an *in vivo* model,³²³ terphenyl



HN-001 was shown to attenuate obesity in mice through induction of thermogenesis in adipose tissue³²⁴ and to suppress lipotoxicity from metabolic-associated fatty liver disease through inhibition of phospholipase A₂ (PLA2)³²⁵ and linoleic acid ameliorated metabolic dysfunction-associated steatotic liver disease in mice by targeting fatty acid binding protein 4.³²⁶ The mechanism underlying the antithrombotic activity of the peptide isaridin E was shown to be associated with inhibition of von Willebrand Factor secretion from activated platelets and this leads to protection against sepsis,³²⁷ butyrolactone-I alleviated intestinal heat stress *in vivo* through inhibition of two key enzymes which are upregulated in heat stress³²⁸ and pannorin was shown to be a selective, if relatively weak (IC₅₀ 1.73 μM) monoamine oxidase inhibitor.³²⁹

The BGC responsible for the biosynthesis of the quinolone alkaloids, the asperalins in *Aspergillus alabamensis*, was identified and the biosynthesis elucidated through heterologous expression.³³⁰

'Omics'-based methods for isolation and cultivation of marine fungi have been reviewed³³¹ as have genome mining and biosynthetic pathways of biologically active marine fungal metabolites.³³²

2.4 Dinoflagellates

The number of new compounds reported from dinoflagellates and other microalgae is slightly lower than the decadal average.¹ *Prorocentrum lima* was the source of polyketides prorocentin-5 **911** and three okadaic acid derivatives **912–914**, although surprisingly, the latter compounds were not assessed for bioactivity.^{333,334} An indole methyl ether **915** and a biaryl ester **916** were reported from a Vietnamese *Aurantiochytrium* strain, although **915** is a likely artefact of isolation.³³⁵

Ectoine plays an important role in controlling osmotic responses in marine planktonic species. Using a hydrophilic interaction liquid chromatography (HILIC) based molecular networking approach, new analogue 2-homoectoine **917** was detected in multiple harmful algal bloom genera including *Alexandrium*, *Karenia*, and *Prorocentrum*. 2-Homoectoine plays a similar osmoadaptive role as archetype ectoine, for the producing strains. The synthesis of **917** was also achieved.³³⁶

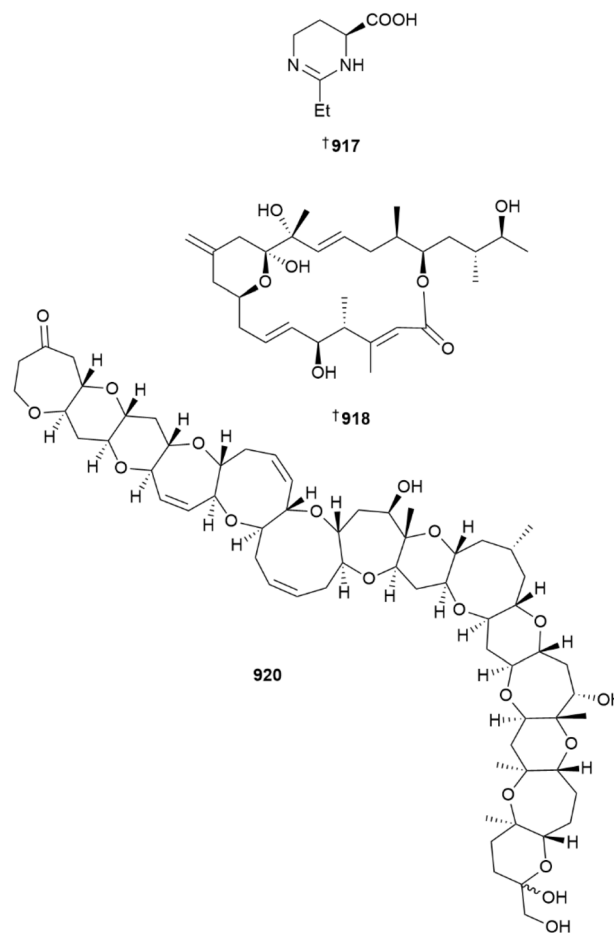
Other structures were suggested from MS and transcriptomics data but are not shown here.^{337,338}

Although only isolated in 2023, the total synthesis of diatom sex pheromone peptide SIP¹ has been accomplished,³³⁹ as has the production of *seco*-steroid gibbosterol A.³⁴⁰ The original structure of iriomoteolide-1a was disproven by synthesis in 2010, but efforts have not revealed the correct constitution of the molecule until now. The corrected structures of iriomoteolides-1a **918** and 1b **919** have now been determined following an extensive integration of DFT calculations, NMR analysis, and total syntheses.³⁴¹

A summary of the biomedical application of *Alexandrium* spp. NPs has been published.³⁴² Compound specific reviews focused on amphidinolides,³⁴³ azaspiracids,³⁴⁴ and ciguatera poisoning, especially related to ciguatoxins.³⁴⁵ The structure of ciguatoxin-5 **920**, originally proposed based solely upon MS

data, has been confirmed by NMR spectroscopic analysis of only 40 μg of isolated material.³⁴⁶ The ambiguity between synthetic and naturally occurring portimine B has been resolved, in part by use of the new i-HMBC NMR experiment that can differentiate ²J_{C,H} vs. ³J_{C,H} correlations. Differences in the NMR spectra observed were attributed to the formation of a transient hemiaminal hydrate, facilitated by acid modifiers added to aid HPLC separation. The authors advise researchers to be careful when adding acid or base to their chromatography solvents, especially when comparing spectroscopic data to literature values.³⁴⁷

The structure of 44-methylgambierone **921** was revised following extensive NMR analysis. The compound was also found to be unstable in DMSO-*d*₆ used for spectroscopic analysis hence a microstudy was undertaken to determine its stability towards various solvents over the course of eight months. The effects of the solvent chosen for NMR analysis, and potential storage, should be considered before committing an entire sample.³⁴⁸ A study of the accumulation of domoic acid (DA), the causative agent of amnesic shellfish poisoning, in five species of invertebrate including commercially valuable shellfish, has shown that DA concentrates in scallops (*Pecten maximus*) at levels 20-fold higher than in clams, slippersnails and sea squirts. The *Pectenid* species showed a distinctive ability to accumulate, transform and distribute DA within their body tissue.³⁴⁹ Transcriptomics analysis of saxitoxin and non-



saxitoxin producing species of the prymnesins, super-carbon chain ladder polyethers produced by *Prymnesium parvum*, are some of the largest known natural products yet their biosynthetic origins have remained elusive until now. The megadalton PKZILLA-1 and -2 polyketide BGC's encode for 140 and 99 enzyme domains, respectively, that match the predicted "pre-prymnesin" linear backbone. A smaller PKZILLA-B1 encodes for a smaller analogue, prymnesin-B1. This work expands the state of knowledge of large biosynthetic genes and enzymatic pathways to important biomolecules.³⁵⁰

3 Green algae

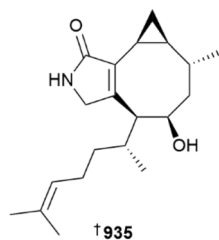
A Chinese *Caulerpa racemosa* var. *turbinata* yielded caulespiros A **922** and B **923**, with neither being active against various bacterial strains.³⁵¹ Mebamamide C **924** was discovered from a Japanese specimen of *Bryopsis*, but also from the algal gastropod predator *Elysia marginata*.³⁵²

Two notable reviews of green algal metabolites were published in 2024. One focused on the allelopathic compounds from *Ulva prolifera*, the algal species that creates "green tides",³⁵³ while the other summarised the metabolites and bioactivities of compounds from *Chlorella vulgaris*.³⁵⁴

4 Brown algae

Glyceroether **925** and polyunsaturated glycolipid **926** metabolites were reported from *Sargassum* sp. (China) and *Hizikia fujiformis* (South Korea), respectively.^{355,356} Macrocarquinoid J **927** and sargasilols J–N **928–931** are meroterpenoid chromanes from *Sargassum* spp., although all are weak or inactive as anti-inflammatory agents.^{357,358} An Indian *Stoechospermum marginatum* was the source of three new spatane diterpenoids **932**, **933**, **934** that showed weak cytotoxicity against four HTCLs.³⁵⁹

Coriaceumins A–D **935**, **936–938** are rare examples of nitrogenous xenicane diterpenoids. Two diterpenoid lactams, dictyolactams C **939** and D **940** were also isolated from the same algal species, *Dictyota coriacea*, collected at Nanji Island, Zhejiang Province, China, although the lactams are artefacts of isolation as proven by extraction using different alcoholic solvents.³⁶⁰



The total syntheses of ecklonialactones C and D, and of eiseniachloride A, have been achieved for the first time.³⁶¹ Notable reviews include a summary of the phytochemicals produced by *Halopteris scoparia*,³⁶² and those from the Sargassaceae between 1975 and August 2023.³⁶³ Marine carotenoid

fucoxanthin has been shown to reduce the effects of autism spectrum disorder induced by valproate treatment in a mouse model in a dose dependent manner. This reduction is induced by alterations to AKT/GSK-3 β signalling.³⁶⁴

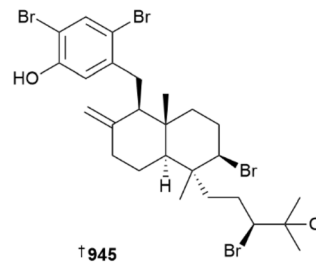
5 Red algae

An Okinawan *Portieria* sample yielded a new halogenated monoterpenoid **941**, which also necessitated the reassignment of related compound **942**. Both metabolites exhibited anti-fouling activity at 0.16 $\mu\text{mol cm}^{-2}$.³⁶⁵ The only other report of new red algal metabolites was of diastereomeric sesquiterpenoids laurenolides A **943** and B **944** from a Vietnamese sample of *Palisada intermedia*.³⁶⁶

The same group has reported the total synthesis of pigments borolithochromes A, D, G, H1, H2, I1 and I2, metabolites of fossilised putative red alga *Solenopora jurassica*, via two separate publications.^{367,368}

A review of the metabolites of *Gelidium corneum* including mycosporine-like amino acids, carotenoids and polyphenols, along with their associated bioactivities, has been published.³⁶⁹ The current status of Japanese *Laurencia* complex focusing on chemical diversity and morphological characters for identification has been reviewed.³⁷⁰ A comprehensive review of the metabolites of *Asparagopsis* and their use for chemotaxonomic analysis of the genus has also been released.³⁷¹

A study has analysed the metabolic profile of different races of *Laurencia nipponica* considering their cytotoxicity, genotoxicity and their antifungal properties as it all relates to chemotaxonomy. Ultimately, with the aid of multivariate statistics, three different races can be differentiated using four different compounds, laurenin, laurallene, prepacifenol and laureatin.³⁷² An innovative approach to establishing the absolute configuration of metabolites with vicinal bromo-chloro substituents using chiroptical methods has been published. By applying van't Hoff's principle of optical superposition, and with the aid of chiroptical measurements of suitably well characterised model compounds for comparison, the contributions of different chromophores within a compound to the overall molar rotations can be mathematically disentangled to provide the absolute configuration of the metabolites in question. Use of this method has necessitated the revision of the structures of the halogenated meroditerpenoids callyphycols A **945** and B **946** and suggested that the structures of other metabolites within the class should be reinvestigated.³⁷³

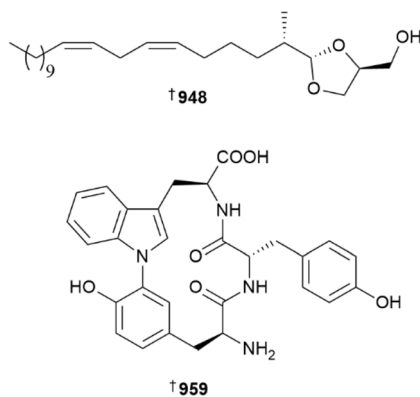


6 Sponges

Aurantioside L **947** is an anti-leishmanial tetramic acid glycoside from *Siliquariaspongia japonica*.³⁷⁴ A *Leucetta* sponge from Bohol Province, the Philippines, was the source of five dioxolane ether lipids **948**, **949–952**. To probe the relative and absolute configurations of their dioxolane cores, model compounds were prepared and their NMR data compared with that of the isolated metabolites. Even then, some compounds were indistinguishable, so the authors utilised europium-based lanthanide chiral shift reagents to ultimately discriminate all the compounds. None of the metabolites exhibited antibacterial activity.³⁷⁵

Endoperoxides, plakortides V **953** and W **954**, and related gracilioether M **955**, 11,12-dihydrogracilioether M **956**, and 9,10-dihydrogracilioether G **957**, were reported from *Plakinastrella* and *Plakortis* sponges, respectively.^{376,377}

Somewhat surprisingly, only two new peptides from sponges were reported. Halichondamide A **958** was isolated from a USA-sourced *Halichondria bowerbanki*, with molecular dynamics simulation used to help delineate the biosynthetic formation of its disulfide bonds; note the name halichondamide should not be confused with that of the much more commonly found halichondramide macrolide family.³⁷⁸ Neopetromin **959** is a cyclic tripeptide that contains a rare C–N biaryl link. Although inactive against two HTCLs, various enzyme targets, and as an antimicrobial, it does cause vacuole fragmentation in an actin-independent manner in tobacco BY-2 cells, like other actin polymerisation inhibitors but without disrupting cellular actin-dynamics. Studies of the impact of this unique bioactivity upon plant physiology are underway.³⁷⁹



PM742 **960** is the first example of a polyketide α -pyrone coupled to an oxime-containing, non-ribosomal peptide. Isolated from a *Discodermia* sponge, PM742 is potently cytotoxic against eight cancer cell lines. Mode of action studies indicate it binds at the colchicine site, causing mitotic arrest *via* inhibition of microtubule dynamics at the G2/M checkpoint.³⁸⁰ A synthetic SAR study identified an analogue, PM534, that covers 80% of the centres within the pharmacophore model of the colchicine binding site. PM534 has demonstrated a potent ability to reduce tumour growth in mouse xenograft models and has entered phase 1 clinical trials for the treatment of non-small cell lung cancer.³⁸¹

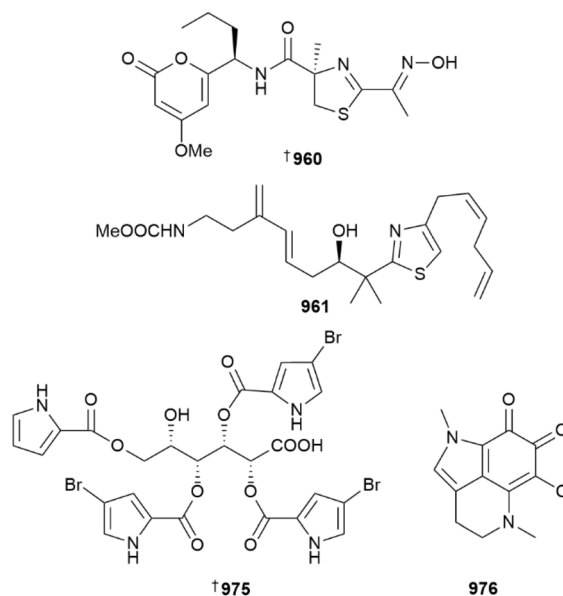
A new mycothiazole congener **961** was reported from a sample of *Cacospongia mycofijiensis* collected in Vanuatu.

Although a potent cytotoxin, the new analogue is still 400- to 700-fold less active than mycothiazole itself which is cytotoxic in the pM-range. Nematodes (*Caenorhabditis elegans*) treated with sub-lethal doses of **961** exhibited an extended mitochondrial lifespan. SAR analysis shows that *Z*- Δ^5 alkene geometry is critical for pM levels of cytotoxicity but does not impact the mitochondrial life extension bioactivity.³⁸²

Polybrominated **962** and lipidated **963** phenols have been reported from *Lendenfeldia* and *Hemimycale* sponges, respectively.^{383,384} Several lactam- (**964–967**) and pyridine-based (**968** and **969**) alkaloids have been reported from a Chinese (Xisha Is.) *Pseudospongosorites suberitoides*. All are likely artefacts of extraction with methanol and are inactive as antivirals.³⁸⁵ A pair of enantiomeric indolinone alkaloids, **970** and **971**, were sourced from a *Tedania* sponge.³⁸⁶ Neopetrotaurines A–C **972–974** are taurine-bridged isoquinoline dimer alkaloids that are weak suppressors of oncogenic fusion protein PAX3-FOXO-1 driven transcription. Neopetrotaurine C is a likely artefact of isolation.³⁸⁷

A new bromopyrrole glycoside has been reported from a deep sea *Lissodendoryx papillosa* specimen. Named in honour of the major contributions of Prof. Valentin Stonik to the field of MNPs, stonikacidin A **975** contains the unusual sugar, L-idonic acid, at its core. Although inactive against numerous mammalian cell lines, **975** has some antibacterial activity and it inhibits biofilm formation in *S. aureus*.³⁸⁸

Pyrroloiminoquinone alkaloids are commonly encountered in *Latrunculia* sponges. A dredged *Latrunculia* specimen yielded 6-chlorodamirone A **976** and its brominated analogue **977**; over time the latter converts to the former when exposed to chloride in seawater in a non-enzymatic reaction. The total synthesis of both compounds was also achieved.³⁸⁹



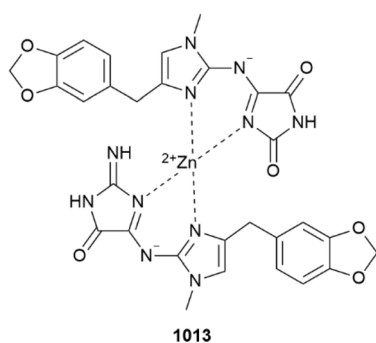
The PIP-HSQC-MBC-IPAP NMR pulse sequence was instrumental in helping identify a series of discorhabdin D congeners **978–982** reported from New Zealand *Latrunculia* spp.³⁹⁰

Multiple pyrrololactam alkaloids and their related metabolites (**983–1007**) have been reported from *Phakellia*, *Stylissa*, and



Pseudospongosorites sponges collected around Xisha Is. in the South China Sea.^{391–393} A New Zealand *Dictyodendrilla* c.f. *dendyi* yielded five new sulfated denigrin metabolites **1008–1012** with one rearranging spontaneously upon short term storage in an NMR tube.³⁹⁴

A diverse series of metabolites, including multiple 2-aminoimidazolone derivatives, were isolated from calcareous sponge *Pericharax heteroraphis* collected from a lagoon in Wallis and Futuna Is.; a new Zn²⁺-containing, heterodimeric clathridine A – clathridimine complex **1013** was one of the isolates. Although inactive as a stimulant of endochondrial ossification, the structure of the compound was confirmed by total synthesis as were those of several other congeners.³⁹⁵



New bromotyrosine derivatives were isolated from *Suberea* **1014–1017**, *Ircinia* **1018–1021**, and *Pseudoceratina* **1022** sponges collected in Australia, the Philippines, and Okinawa, respectively.^{396–398} Merosesquiterpenoids have been reported from *Pseudoceratina purpurea* (**1023**, **1024**, and **1025–1036**) and *Dysidea arenaria* (**1037–1042**), although all the isolates were only weakly active or inactive altogether in a variety of bioassays.^{399,400} Further meroterpenoids were isolated from *Hyrtilis* (**1043** and **1044**) and *Hippospongia* (**1045–1052**) sponges,^{401,402} while a number of norditerpenoid cyclic peroxides **1053–1062** were reported from a Bohol Is. (the Philippines) specimen of *Diacarnus spinipoculum*.⁴⁰³ A *Hemimycale* sponge was the source of *seco*-diterpenoid **1063**, which was not assessed for bioactivity.⁴⁰⁴ Isolation of ircinialactams **1064–1066** from a Mexican *Ircinia felix* also provided an opportunity to revise the structure of related variabilin metabolite **1067**; the total synthesis of **1064** was also achieved.⁴⁰⁵ An Antarctic *Suberites* sponge yielded suberitenone sesterterpenoids **1068–1072** although none showed activity as antivirals.⁴⁰⁶ Given how common scalarane sesterterpenoids are in marine sponges, surprisingly only one, lendenfeldarane V **1073**, was reported in 2024,⁴⁰⁷ while two nor-sterol epoxide saponins **1074** and **1075** were obtained from a *Petrosia* sponge collected in the Philippines.⁴⁰⁸ *Seco*-Sterols were reported from *Cliona* (**1076**), *Hippospongia* (**1077**, **1078**) and *Spongia* (**1079**) sponges, respectively.^{409–411} A series of 12 nor-terpenoid peroxides, **1080–1091**, all derived from oxidative cleavage of larger metabolites, were isolated from a *Diacarnus* sponge and all with varying levels of antibacterial and cytotoxic activities.⁴¹² Other structures have been claimed but are inconsistent with the spectroscopic data provided, or were only

detected by GNPS-based molecular networking, and hence are not shown.^{384,404,413}

Numerous first total syntheses were reported and these included halisphingosine A **1092**, which necessitated a revision of structure,⁴¹⁴ cyclic peroxide ethyl plakortide Z,⁴¹⁵ peptide neopetromin,⁴¹⁶ and macrolide salarin C, which also validated a biosynthetic oxidation – Wasserman rearrangement to give salarin A in a biomimetic approach.⁴¹⁷ A multiplexed strategy using 96-well plates to optimise conditions for reactions between fatty dialdehydes, a nitrogen source and acrolein has been used to probe and verify the Whitehead – Baldwin biosynthetic proposal for the manzamines.⁴¹⁸ Indole alkaloids dragmacidins G and H, and tulongicin A, have been prepared for the first time,^{419,420} as has aminophenol eribusinone, leading to its revised structure **1093**.⁴²¹ The first total synthesis of spirodactylone has been accompanied by the synthesis of several related denigrin metabolites that has indicated the published structure of denigrin C is incorrect, although no alternative was proposed.⁴²² Guanidine alkaloids (–)monanchoradin A and (–)crambescin A2 **393** have been prepared for the first time,⁴²³ as have makaluvamines H, L, M and N.^{424,425} A series of BF₂-complexes of ageladine A have been prepared as BODIPY mimics, with their fluorescence being heavily dependent on both pH and halogenation of the imidazole ring.⁴²⁶ Psammaphysins M, O and Q, along with ceratinamide A, have been synthesised as racemates,⁴²⁷ as have sesquiterpenoids halichonadins A, B and C.⁴²⁸

A review of sponge-derived MNPs with activity against lung cancer has been published.⁴²⁹ Taxa-based reviews have focused on compounds from the *Spongia* and *Neopetrosia* genera, respectively,^{430,431} while compound specific reviews have summarised the literature pertaining to the convoluted structure elucidation, total synthesis and biological evaluation of PUFA-derivative (–)mucosin,⁴³² and also of the immunomodulatory effects of halichondrin and its derivatives.⁴³³ Other compound specific reviews have concentrated on aaptamine and its derivatives,⁴³⁴ the anti-cancer potential of marine bis-indoles such as the dragmacidins and topsentins,⁴³⁵ and of bromotyrosine-derivatives, mainly from Verongiid sponges.⁴³⁶ A compendium of 413 sponge-derived alkaloids published between 2000–2023 has been released,⁴³⁷ with a similar summary of diterpenoids isolated from sponges in 2009–2022 covering 228 structures.⁴³⁸

The inseparable endoperoxide pair, plakortinic acids C and D, isolated from the mutualistic symbiotic sponge pair *Plakortia symbiotica* and *Xestospongia deweerdtiae*, have antiplasmodial activity without causing haemolysis. They exhibit promising absorption, distribution, metabolism, excretion, and toxicity (ADMET) properties making them potential antimalarial leads.⁴³⁹ Chemoproteomic analysis has indicated that ubiquitin carboxyl-terminal lyase 5 (USP5) is the molecular target of endoperoxide gracilioether A (*Plakinastrella mamillaris*). USP5 is heavily involved in chromatin maintenance, catalyses degradation of abnormal proteins, and is over-expressed in multiple cancer types. Inhibitors of USP5 therefore have promise as anticancer agents.⁴⁴⁰

Peptide stylissatin A and its analogues inhibit membranous neuraminidase-1 functioning by modulating the chaperone



activity of cathepsin A, thereby having application in preventing the formation of adipose tissue.⁴⁴¹ Chemoproteomics has also been used to detect agents that protect against inflammatory acute lung injury. Naamidine J interacts with His745 and Phe903 in chromosome segregation 1-like protein (CSE1L) which inhibits nuclear translocation and transcriptional activity of SP1, suppressing inflammation. This study also provided more naturally occurring material which facilitated the re-assignment of naamidines A, G and J **1094–1096** through X-ray crystallographic analyses.⁴⁴² Although only weak inhibitors of *M. tuberculosis* under standard culturing conditions, axinellamines A and B have moderate to potent activity against the pathogen when grown with limited protein supplementation, mimicking *in vivo* conditions. Both compounds are inactive against the RAW264.7 cell line as a model host.⁴⁴³ Several bromotyrosine metabolites sourced from Verongiid sponges were found to be weak to moderate inhibitors of prion PrP^{Sc} protein, which in turn reduces ER stress and also the spread of the pathogen in eukaryotic cells. This work provides a lead series of structures for development against a currently untreatable, and often fatal, disease.⁴⁴⁴

A nematocidal screen of over ten thousand fractions from approximately two thousand extracts held in the NatureBank repository highlighted seven extracts for further investigation. Deeper study of two chemotypes with activity against the sheep nematode *Haemonchus contortus* led to halaminol A and several agelasines as the dominant bioactive metabolites, but only agelasine B showed weak activity against the model roundworm *Caenorhabditis elegans*, suggesting either a minor component is responsible for the broader activity profile, or there is some instability in the isolated compounds.⁴⁴⁵ Sterol sokotrasterol sulfate (*Topsentia* sp.) was found to inhibit IFN- γ -induced expression of programmed cell death – ligand 1 (PD-L1) protein *via* targeting Janus kinase with a downstream activation of signal transducer and activator of transcription (STAT) signaling, making it a new immunomodulatory lead.⁴⁴⁶

Assignment of the absolute configuration of antifungal depsipeptide cyclolithistide A **1097** has been completed by the synthesis of all four stereoisomers of the rare amino acid, 4-chloroisoleucine, and then use of Marfey's method to establish the configuration of the natural residue within the metabolite.⁴⁴⁷

An important cautionary observation regarding isolation of artefacts has been made. Pyrrole-containing MNPs including makaluvamines, discorhabdins, tambjamins and brominated tryptamines, were found to be trideuteromethylated when exposed to DMSO-*d*₆ NMR solvent in the presence of water, methanol, TFA and trace amounts of iron. Mechanistic investigation indicates that the reaction is caused by the presence of trideuteromethyl radical species generated by Fenton-type chemistry catalysed by the presence of iron or TFA.⁴⁴⁸

Finally, a study has explored the state of knowledge of the currently underexplored terpenoid biosynthesis in Dictyoceratid sponges, which, for example, are well known producers of spongian diterpenoids. While bioinformatic analysis detected analogues of known terrestrial

prenyltransferases for one sesterterpenoid and multiple diterpenoid, steroid and carotenoid biosyntheses across various species, a lack of relatives to known terpenoid cyclase enzymes suggests the existence of divergent and unknown biosynthetic pathways in sponges. This study provides a start-point for exploring these currently cryptic biosynthetic genes and enzymes.⁴⁴⁹

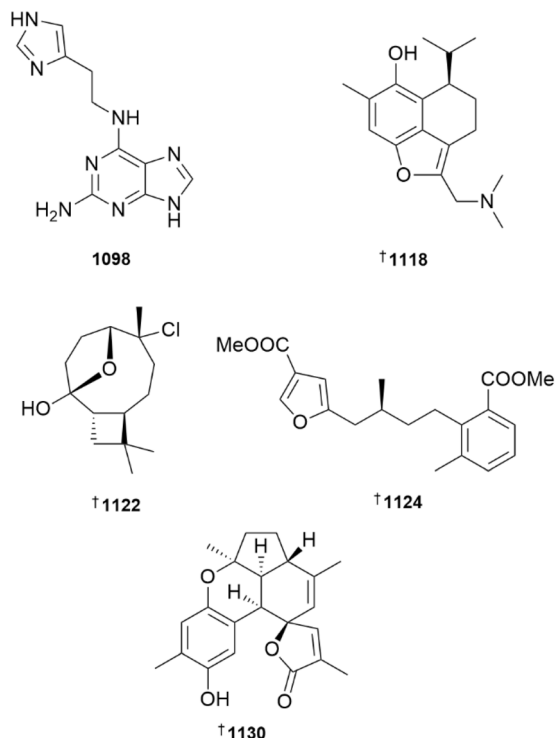
7 Cnidarians

Tubeastriane, sourced from the hard coral *Tubeastrea tagusensis*, acts as an antioxidant to reduce levels of ROS, which is able to, in turn prevent neuron-like cell death caused by the oligomeric amyloid-beta peptide α A β 42.⁴⁵⁰ While acroamine A **1098**, a 2-amino adenine alkaloid reported from the soft coral *Acrozoanthus australiae*, was essentially inactive as an inhibitor of cAMP-dependent protein kinase A, several semi-synthetic analogues including di- and tri-bromo derivatives were active.⁴⁵¹ One of the more common family of alkaloids reported from cnidaria are the zoanthamines, isolated from *Zoanthus* sp. A new, efficient synthetic route to norzoanthamine has been reported, notable for including three photochemical reactions used at key steps.⁴⁵² Of a set of thirteen prostanoids isolated from *Clavularia* sp., two (**1099**, **1100**) were new, with the former demonstrating weak *in vitro* cytotoxicity towards the Ca9-22 oral cancer cell line.⁴⁵³ A new stereoselective approach to the synthesis of prostaglandin A₂ epimers (15*S*)-PGA₂ and (15*R*)-PGA₂ has been reported that makes use of only catalytic amounts of heavy metals.⁴⁵⁴ The study also revealed that (15*R*)-PGA₂ is a weak inhibitor of GABA_A receptors in *Xenopus laevis* oocytes, while the (15*S*)-epimer was inactive.

While the structures of sesquiterpenoids **1101–1103**, isolated from *Lemnalia* sp., were established using a combination of NMR and MS techniques,⁴⁵⁵ the structures of six of seven new sesquiterpenoids, bellissinanes A–G **1104–1110** (*Stereonephthya bellissima*) and each of lemneolemnanes A–D **1111–1114** (*Lemnalia* sp.) were secured using XRD.^{456,457} Two new examples of isodene-type sesquiterpenoids, **1115** and **1116** were isolated from the soft coral *Heteroxenia* sp., with both MNPs being inactive against SAR-CoV-2 and *Leishmania major*.⁴⁵⁸ A deep-sea collection of *Anthothela grandiflora* afforded unusual dimethylamino substituted cadinene-type sesquiterpenoids, anthoteibinanes A–E **1117**, **1118**, **1119–1121**.⁴⁵⁹ Anthoteibinane B exhibited weak activity towards respiratory syncytial virus *in vitro*. Asymmetric syntheses of the sesquiterpenoids (–)-lemnalemnane A and (+)-lemnardosinane A, with both routes starting from (*S*)-carvone,^{460,461} and a racemic synthesis of paralemnolin A have been reported.⁴⁶² Using a synthetic route starting from (–)- β -caryophyllene oxide, the structure of the chlorinated norsesquiterpenoid rumphellatin B has been confirmed, while those of rumphellatins A and C have been revised to **1122** and **1123** respectively.⁴⁶³ During the study, it was observed that reaction of rumphelloilide C with HCl in dioxane afforded rumphellatin B, with the authors speculating that the simple transformation may occur in soft coral species.



The structures of meroterpenoids sinudenoids **F 1124** and **G 1125**, isolated from *Sinularia densa*, are unusual due to the presence of a methylbenzoate core⁴⁶⁴ while more classically functionalized meroterpenoids bearing hydroquinol or naphthoquinone cores **1126–1129** were isolated from *Litophyton brassicum*.⁴⁶⁵ An asymmetric total synthesis of the pentacyclic benzosesquiterpenoid (+)-verrubenzospinolactone **1130** made use of a late-stage, spontaneous, intramolecular Diels–Alder reaction.⁴⁶⁶



Four examples of diterpenoids embodying three different skeletons, herpetopanone **B 1131** bellissimain **A 1132** elisabethadienol **B 1133** and elisabethadienol **C 1134** were reported from extracts of *Stereonephthya bellissima*, with none showing activity in a pro-angiogenesis assay in zebrafish.⁴⁵⁶ Further examples of biflorane-type diterpenoids, biofloranates **E–I 1135–1139**, in addition to two glycosides lemnaboursides **H 1140** and **I 1141**, were reported from a South China Sea collection of *Lemnalia bournei*.⁴⁶⁷ Of the set, only lemnabourside **H** exhibited antibacterial activity, weakly inhibiting the growth of *E. coli*. The structures and relative configurations of *seco*-diterpenoids kallopterolides **A–I 1142–1150**, isolated from the sea plume *Antillologorgia kallos*, were secured by combinations of NMR analysis and DFT DU8ML calculations.^{468,469} Structures were assigned to polyoxygenated casbane-type diterpenoids sinueracasbanones **E–O 1151–1161**, isolated from *Sinularia erecta*, by extensive use of NMR data in conjunction with DFT DP4+ analysis, while absolute configurations were assigned by the use of Mosher's method and TDDFT-ECD calculations.⁴⁷⁰ Sinueracasbanones **I** and **O** exhibited weak anti-inflammatory activity. The capnosane-type diterpenoid **1162**, isolated from *Lobophytum pauciflorum*, was inactive in a zebrafish model of inflammation.⁴⁷¹ Three

cycloamphilectane-type diterpenoids, sinucycloamtins **A–C 1163, 1164** and **1165** were isolated from a South China Sea collection of *Sinularia brassica*.⁴⁷² Purification of sinucycloamtins **B** and **C** was hampered by the apparent ability of the two compounds to undergo intramolecular ester exchange. Subsequent saponification afforded a diol product, the structure and absolute configuration of which were established by XRD. Pseudopteroxazole and pseudopterosein **G** are antibacterial diterpenoids previously reported from the Caribbean soft coral *Antillologorgia* (formerly *Pseudopterogorgia*) *elisabethae*. Using an unbiased proteomics approach, the antibacterial mechanism of action of both MNPs appears to be associated with disruption of membrane-associated steps involved in cell wall biosynthesis.⁴⁷³

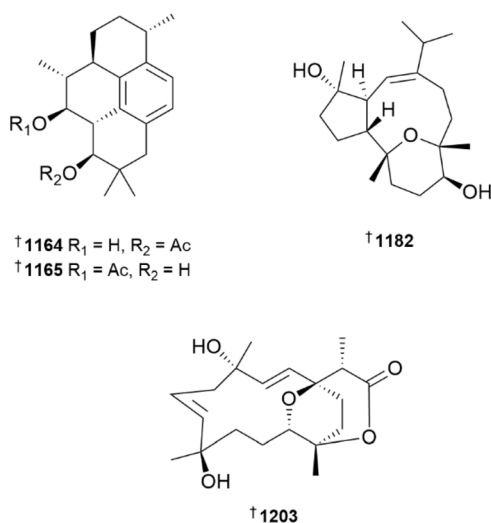
With over sixty new examples, cembranoids remain the dominant sub-structural class of diterpenoids reported from soft corals. Of a set of eleven cembranoids isolated from a South China Sea collection of *Litophyton amentaceum*, litoamentenes **A–K 1166–1176**, only litoamentene **I** exhibited cytotoxicity, being weakly active against the NCI-H446 cell line.⁴⁷⁴ In addition to sterols (see later), an extract of *Lobophytum catalai* afforded the cembrane-type diterpenoids lobocalines **A–E 1177–1181**.⁴⁷⁵ While XRD analysis provided the structure and absolute configuration of lobocaline **A**, interpretation of NMR data, including DP4+ probability analysis, and TDDFT-ECD calculations, secured the structures and configuration of the remaining examples. The structure of co-metabolite lobophytol **B**, previously reported as a bicyclic terpenoid, was revised on the basis of XRD analysis to **1182**.

The 3D structure of 7*S*,8*R*-dihydrodepoxy sarcophytoxide **1183** has been secured by XRD analysis.⁴⁷⁶ The findings call into doubt the structure and/or stereochemistry previously proposed for (+)-7β,8β-dihydroxydepoxy sarcophytoxide. Tetrahydropyran ring-containing cembranoid **1184** and a 4-hydroperoxy derivative of sarcophine **1185** were reported from an extract of *Sarcophyton glaucum*, collected off the coast of Egypt.⁴⁷⁷ Four sarcophine analogues, sarcoeleganolides **H–K 1186–1189** were isolated from a South China Sea collection of *Sarcophyton elegans*,⁴⁷⁸ while *S. crassocaulis* was the source of meijicrassolins **A–E 1190–1194**.⁴⁷⁹

A series of capnosane-type cembranoids, sarcocinerenoids **A–H 1195–1202** and two unusual variants, sarcocinerenoids **I 1203** and **J 1204**, were reported from extracts of a Xisha Is., South China Sea collection of *Sarcophyton cinereum*.⁴⁸⁰ XRD analysis was used to secure the structure and absolute configuration of sarcocinerenoids **A–C, F, G** and **I**. In a separate publication, details of the characterisation of a further nine cembranoid-type diterpenoids, sarcocinerenolides **A–I 1205–1213** from the same *S. cinereum* biomass were reported.⁴⁸¹ Amongst these structures, sarcocinerenolide **A** is unusual in being a ring-opened variant, while the remaining congeners are more typical of cembranoids, displaying variation in the location of ether linkages. A structurally-related sarcophine derivative bearing a C-1 to C-12 peroxy linkage, sarcocutumolid **A 1214** was reported from the extract of a Red Sea collection of *Sarcophyton acutum*.⁴⁸²



A Ximao Is., South China Sea collection of *Sinularia pedunculata* afforded a structurally-diverse set of cembranoid diterpenoids including sinupedunolides A-E **1215–1219** and sinupedunols A **1220** and B **1221**.^{483,484} The structures of sinupedunolide A and sinupedunols A and B were secured by XRD analysis. Of the five cembrane diterpenoids, sinudenoids H–L **1222, 1223–1226** isolated from *Sinularia densa*, sinudenoid H is an unusual example of a bisnorfuranocembrenolide skeleton.⁴⁶⁴ The tricyclic structures of 4 α -hydroxy-chatancin **1227** and sarcotoroid **1228**, isolated from *Sarcophyton tortuosum*, were proposed to be derived from cembranoid precursors that had undergone transannular Diels–Alder cycloaddition reactions.⁴⁸⁵



The possible role of photochemistry in the biosynthesis of the cyclobutane ring of the cembranoid diterpenoid (+)-provindencin **1229** has been supported by its semisynthesis from (–)-bipinnatin E **1230**, which was isolated from a Floridian specimen of *Antillogorgia kallos*.⁴⁸⁶ The Norrish–Yang cyclisation step also afforded a C-14 regioisomeric product, which is likely an anticipated NP. Absolute configuration was assigned to all three compounds based upon XRD analysis of (–)-bipinnatin E. Two accounts of the synthesis of the revised structure of scabrolide B were reported in 2024. In the first publication, (–)-scabrolide B was synthesised in 19 steps (longest linear sequence), and in a simple set of reactions was converted into (–)-sinuscalide C **1231**, (+)-ineleganolide and (+)-horiolide **1232**.⁴⁸⁷ A second route to (–)-scabrolide B was reported by another research group, with simple conversions again affording related diterpenoids (–)-sinuscalide C and (+)-ineleganolide, as well as a third MNP, (+)-fragilolide A **1233**.⁴⁸⁸ A full account of the synthesis of (–)-scabrolide F has been reported, with weak anti-fouling activity observed for the NP and synthetic intermediates.⁴⁸⁹

New biological activities were reported for cnidarian-derived cembranoids including the finding that 8-*epi*-sarcophinone and *ent*-sarcophine exhibit anti-fouling activity when included in marine paint preparations,⁴⁹⁰ sinulariolide exhibits anti-inflammatory activity in a rheumatoid arthritis model,⁴⁹¹ 7,8-epoxy-11-sinulariolide acetate also exhibits anti-inflammatory

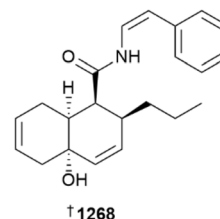
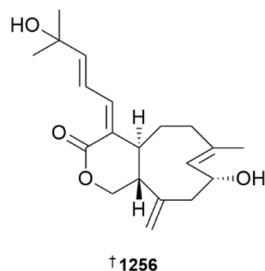
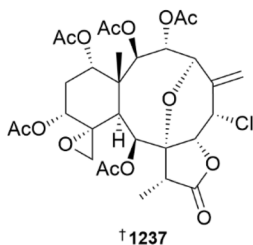
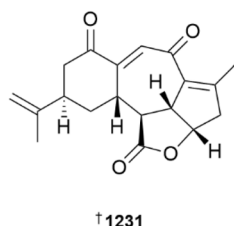
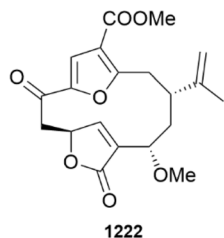
as well as inhibition of osteoclastogenesis,⁴⁹² sinularin induces apoptosis in prostate cancer cells *via* a mechanism related to up-regulation of FOXO3 forkhead transcription factor,⁴⁹³ and 4-carbomethoxy-10-*epi*-gyrosanolide E inhibits oral cancer cell migration and invasion by inhibition of the MAPK pathway.⁴⁹⁴ Sarcotrocheliol exhibits weak cytotoxicity, antimicrobial and larvicidal activities.⁴⁹⁵ An aquaculture specimen of *Briareum stechei* afforded the epoxybriarane briastecholide O **1234**, while wild-type specimens of the same organism yielded the related analogue briastecholide P **1235**.⁴⁹⁶

XRD analysis was used to secure the structure and absolute configuration of not only briastecholide P, but also of the known co-metabolite briaexcavatin X **1236**. In two separate accounts, a large array of new and known polyoxygenated briaranes were isolated from a collection of *Junceella fragilis* collected off the coast of Southern Taiwan. In addition to two new chlorine-containing congeners, **1237** and **1238**, two previously reported analogues gemmacolide X **1239** and frajunolide I **1240** were re-isolated.⁴⁹⁷ Absolute configurations of the latter two diterpenoids were determined for the first time. In the second publication, one new briarane example was reported, fragilide Z **1241**, while the structures and absolute configurations of known co-metabolites 12-*epi*-fragilide G **1242**, fragilide P **1243**, junceollolide D **1244**, junceollonoid A **1245** and juncin ZI **1246** were secured by XRD analysis.⁴⁹⁸ Excavatulide B and two semisynthetic analogues, one being an ester linked piperidine salt and the second being an oxime-linked terminal alkyne, inhibit inflammation and angiogenesis in models of atopic dermatitis.⁴⁹⁹

As part of a SAR study of transient receptor potential melastatin 7 (TRPM7) ion channel inhibitors, six new xenicane diterpenoids 7S,8S-epoxywaixenicin A **1247**, 7S,8S-epoxywaixenicin B **1248**, 12-deacetylwaixenicin A **1249**, waixenicin E **1250**, waixenicin F **1251** and 20-acetoxyxenafaraunol B **1252** were isolated from a Hawaiian collection of *Sarcothelia edmondsoni*.⁵⁰⁰ The authors suggested that the epoxide-containing analogues 7S,8S-epoxywaixenicin A and 7S,8S-epoxywaixenicin B may be artefactual. Preliminary elements of an SAR were proposed, indicating the requirement of the 12-acetoxy group in combination with a dihydropyran ring for activity, and that bioactive compounds may act as latent electrophiles. Three examples of xenicane diterpenoids, miolenol **1253**, epoxy miolenol **1254** and epoxy coraxeniolide A **1255**, were reported from a deep-sea collection of the soft coral *Paragorgia arborea*.⁵⁰¹

The first asymmetric total synthesis of the xenicane isoxeniolide A **1256** has been reported, confirming the structure and defining the absolute configuration.⁵⁰² Ten new sterols were reported from cnidarians in 2024, including lobocaloids A–D **1257–1260** from *Lobophytum catalai*,⁴⁷⁵ cholesterol **1261** and **1262** from *Litophyton mollis*,⁵⁰³ capnesterones A **1263** and B **1264** from *Capnella imbricata*,⁵⁰⁴ and cholestene-7-ones **1265** and **1266** from *Lobophytum durum* and *Capnella imbricata*, respectively.^{505,506} The study of *Lobophytum durum* also led to the re-isolation of the known ergosterol derivative **1267**, the relative configuration of which was determined by XRD analysis, providing clarification on the configuration of the 24-methyl group.⁵⁰⁵





A secogorgostenone steroid, originally sourced from *Sinularia leptocladus*, exerts anti-inflammatory activity and brain-protective effects by attenuating matrix metalloproteinase-mediated events.⁵⁰⁷ The MNP chemistry of cnidarians belonging to the genus *Capnella*, covering the period 1974 to May 2024, has been reviewed.⁵⁰⁸ The background history of the palytoxins, originally isolated from the zoanthid *Palythoa* sp., as well as their biological activities and mechanism of action, has also been reviewed.⁵⁰⁹ The chemistry and biological activities of ketosteroids, a number of which are isolated from soft corals, have been reviewed.⁵¹⁰ A recombinant variant of Tst2, a 38-residue peptide identified by transcriptomic and proteomic analysis of the Australian sea anemone *Telmatactis stephensoni*, is an inhibitor of the transient receptor potential subfamily V member 1 (TRPV1) ion channel.⁵¹¹ HCIQ2c1, a Kunitz-peptide identified in cDNA analysis of the sea anemone *Heteractis crispa*, binds to the transient potential ankyrin 1 (TRPA1) calcium-permeable channel, appearing to stabilise the channel in the open conformation, and reduces pain and inflammation *in vivo*.⁵¹² Bioassay-directed fractionation of toxins from a Japanese collection of the sea anemone *Heteractis aurora* led to the identification of one novel class of toxin, denoted Hau I, with as-yet undetermined target(s), two new examples of toxins from the boundless β -hairpin family (Hau II and III) and a new example of a sea anemone type 1 sodium channel toxin (Hau IV).⁵¹³

8 Bryozoans

Brominated indole alkaloids, alternatamides B and D have been synthesised from 3,4-dibromoaniline *via* the intermediate 5,6-dibromotryptamine⁵¹⁴ and bryostatin 4 has been shown to inhibit the transforming growth factor (TGF)- β signalling pathway, responsible for disease progression in acute erthroleukaemia.⁵¹⁵

9 Molluscs

Compared to previous years, there was a significant drop in new MNPs reported from molluscs in 2024, with the two examples

being bursatamide A **1268** and bursatellin B **1269** isolated from the sea hare *Bursatella leachii*.⁵¹⁶ A general review covering the nutraceutical and medical potential of marine molluscs has been published.⁵¹⁷ The potential of dolastatin MNPs and synthetic analogues as anticancer agents has been reviewed.⁵¹⁸ Exploration of the SAR of (–)-dolicolide A led to the identification of some analogues with similar levels of cytotoxicity as the MNP towards the HCT-116 cell line.⁵¹⁹ Furodysin, isolated from the nudibranch *Hypselodoris tryoni*, exhibited weak *in vitro* activity against wild-type *Plasmodium berghei*, but was considerable less active against an atovaquone-resistant strain.⁵²⁰

Tambjamine analogues show interesting results as anti-leishmanials, with potent intracellular activity against *Leishmania mexicana* and *L. donovani*, and with one analogue demonstrating partial protection in a rodent *in vivo* model.⁵²¹ The structures, synthetic approaches and SARs of the lamellarin alkaloids have been reviewed.^{522–524}

The global occurrence, toxicity and risk assessment of cyclic imine shellfish toxins have been reviewed.⁵²⁵ Analysis of scallops *Argopecten irradians* exposed to paralytic shellfish toxin producing dinoflagellate *Gymnodinium catenatum* identified that the toxins accumulate in the adductor muscle, leading to levels exceeding regulatory limits.⁵²⁶ Diatoms of the genus *Pseudo-nitzschia* produce the neurotoxin (DA), which in the case of king scallop contamination, can take many months to clear from the mollusc tissue. DA immunodetection has been used to examine the time course of subcellular location of the toxin, finding preliminary evidence of accumulation in autophagosomes in the digestive gland, which may go some way to explain the toxin's persistence.⁵²⁷ Two fatty acid desaturase genes were identified in the dwarf surf clam *Mulinia lateralis*, the expression levels of which varied during embryonic development.⁵²⁸

Investigation of the transcriptome of the carnivorous snail *Raphitoma purpurea* has identified putative conotoxin-like peptides in both salivary gland and venom duct tissue, proving further implication for venom evolution in molluscs of the family Conidae.⁵²⁹ The predatory and defensive venoms associated with 16 different cone snail species have been reviewed,⁵³⁰ while another review has focused more on the effects of κ -conotoxins on potassium channels.⁵³¹ A cyclic octapeptide Am931, identified from analysis of the venom duct transcriptome of *Conus amadis*, appears to act as a catalyst promoting formation of the natively folded disulfide form of α -conotoxin ImI.⁵³² Replacement of L-lysine or L-arginine residues in α -conotoxin RgIA with their corresponding D-enantiomers has afforded weakly antifungal analogues.⁵³³

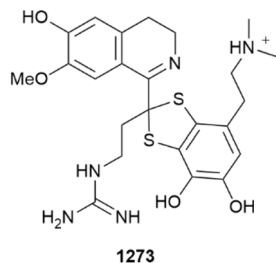
Six synthetic α -conotoxins, sequences of which were derived from transcriptome sequencing of venom of *C. quercinus*,



exhibited potent cytotoxicity towards fall army worm cells as well as anti-larval toxicity.⁵³⁴ In two separate studies by the same research group, α -O-conotoxin GeXIVA[1,2] was shown to both reduce cancer chemotherapy induced neuropathic pain in a mouse model and to exhibit *in vivo* tumour growth suppression of triple-negative breast cancer.^{535,536} A disulfide bond deleted and truncated conotoxin analogue, based on the sequences of α -O-conotoxins GeXIVA and GeXXVIIA, demonstrated retained potency of human $\alpha 9\alpha 10$ nAChR inhibitory activity, inhibited Ca_v2.2 channels and alleviated pain in a mouse model.⁵³⁷ Conotoxin kM-RIIIJ has been used to explore the physiological functions of heterodimeric K_v1.2 channels, revealing that the receptors are highly expressed in proprioceptive (sensory motor) neurons in the dorsal root ganglion.⁵³⁸

10 Tunicates (ascidians)

Further investigation of an extract of an unidentified Didemnidae ascidian collected in Palau, afforded guanidine alkaloids 1-carboxydopargimine **1270**, mellpaladines D–F **1271**, **1272** and **1273**, serodopalgimine **1274** and a 4,4'-dimer of serotonin **1275**.⁵³⁹ Sulfoxide-containing mellpaladines D and E were isolated in equal amounts, with the authors suggesting they may be products of autoxidation. Mellpaladines D–F were evaluated for effects on behavioural activities in mice, with mellpaladine F being found to induce whole body convulsion.



The original proposed disulfide, homocoupled, cyclic hexapeptide structures of antatollamides A and B have been revised by total synthesis to be simpler monomeric cyclic peptides **1276** and **1277**, respectively.⁵⁴⁰ The first enantioselective synthesis of (+)-cylindricine B has been reported.⁵⁴¹ The synthesis and multidrug resistance reversing activities of the pyrrole-containing ningalin MNPs have been reviewed,⁵⁴² as have the cytotoxic properties of cephalostatin and ritterazine dimeric steroidal pyrazine alkaloids.⁵⁴³ The human triggering receptor expressed on myeloid cells 2 (TREM2) is an important regulator of immunological responses. *N*-Deacetylshermilamine B, as well as a partially characterised mixture of glucosylceramides, isolated from the ascidian *Cystodytes dellechiaiei*, bind to TREM2, with the pyridoacridine alkaloid also exhibiting weak *in vitro* cytotoxicity towards two multiple myeloma cell lines.⁵⁴⁴

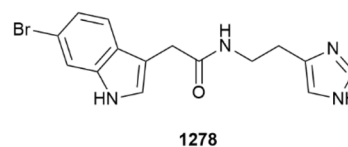
Phosphoeleganin and one semisynthetic analogue decrease interleukin 6 secretion, suggestive of this compound class having potential in the management of diabetes.⁵⁴⁵ A further SAR study of phosphoeleganin has identified the importance of the phosphate and glycine groups, in combination with backbone carbinol stereochemistry, for inhibition of h-15-LOX-1

enzyme.⁵⁴⁶ Of a library of synthetic analogues of rubrolide-related γ -butenolides, a non-halogenated example exhibited weak to moderate cytotoxicity towards a panel of HTCLs, induced ROS production leading to apoptosis, arrested cell cycle at G₂/M and exhibited *in vivo* activity in a HeLa xenograft assay.⁵⁴⁷

A structurally simplified analogue of diazonamide A exhibited potent antiproliferative activity towards a panel of HTCLs which appeared to be related to inhibition of tubulin assembly, and the simplified analogue was less toxic to noncancerous cell lines.⁵⁴⁸ Screening a library of marine bacterium *Tistrella* strains revealed the majority were capable of producing didemnin B.⁵⁴⁹ The introduction of a second copy of the didemnin B BGC afforded a modified strain with excellent production capability (75 mg L⁻¹), enabling semisynthesis of a series of new analogues. A cytotoxic photoaffinity probe successfully enriched a large number of potential protein targets, including the previously noted depalmitoylating enzyme PPT1. Before being discontinued due to a lack of COVID-19 related hospitalisations, a Phase III trial of plitidepsin showed it to be generally well tolerated and suggested a two-day improvement in the median time to withdrawal of supplementary patient oxygen support.⁵⁵⁰ Screening a library of marine-derived compounds identified polycarpine as a potent inhibitor of the main protease M^{Pro} of coronaviruses, also inhibiting SARS-CoV-2 and PEDV (porcine epidemic diarrhoea virus) viral replication in Vero-E6 cells.⁵⁵¹ The uses of the tetrahydroquinoline alkaloid trabectedin and its related analogue lurbinctedin in oncology, mechanisms of action, and uses in combination therapies have been reviewed.⁵⁵²

11 Echinoderms

Extracts of the arms of the starfish *Thromidia catalai* afforded three indole derivatives, catalindoles A–C **1278**, **1279** and **1280**, the structures of which were confirmed by synthesis.⁵⁵³ When collected, the starfish were found feeding on the sponge *Theonella swinohei* – LCMS analysis of extracts of the starfish digestive gland identified the presence of *Theonella*-sourced NPs while none were located in the arms, suggesting that the starfish does not accumulate sponge metabolites.



Steroidal disulfates **1281–1284** were isolated from a Far Eastern collection of the slime sea-star *Pteraster marsippus*.⁵⁵⁴ The biological activities of saponins isolated from sea-stars has been reviewed.⁵⁵⁵ Several examples of polyhydroxylated steroidal saccharides were also reported from starfish, including pectiniferosides A–J **1285–1294** from *Patiria* (= *Asterina*) *pectinifera*,⁵⁵⁶ spiculiferosides A–D **1295–1298** from *Henricia leviuscula spiculifera*,⁵⁵⁷ and ceramasterosides A, B, D and E **1299–1302**, from *Ceramaster patagonicus*.⁵⁵⁸ Of these MNPs, only the latter, ceramasteroside D, exhibited a notable



biological activity, of moderate ability to inhibit the release of nitrite from LPS stimulated microglial cells.

Ten examples of triterpenoid glycosides were reported from sea cucumber species, including pacificosides L–Q **1303–1308** from *Solaster pacificus*,⁵⁵⁹ peronioside A **1309** from *Psolus peronii*,⁵⁶⁰ conicospermiumosides A₃-1, A₃-2, A₃-3, A₇-1 and A₇-2 **1310–1314** from *Cucumaria conicospermium*,⁵⁶¹ and **1315–1318** from *Apostichopus japonicus*.^{562,563} Investigation of extracts of the sea urchin *Chlypeaster humilis* afforded sulfate **1319**, methane sulfonate **1320** and boldine **1321**, the latter two compounds being reported from a marine organism for the first time,⁵⁶⁴ while bibenzochromenones phanogracilins A–C **1322–1324** were isolated from the crinoid *Phanogenia gracilis*.⁵⁶⁵ Additional biological properties have been reported for echinochrome A, including anti-asthmatic activity⁵⁶⁶ and the ability to modulate migration of non-small cell lung cancer cells.⁵⁶⁷ 2-Butoxytetrahydrofuran, previously reported from *Holothuria scabra*, attenuates the aggregation and oxidative properties of α -synuclein, lessening its toxicity in a *C. elegans* model of Parkinson's disease.⁵⁶⁸

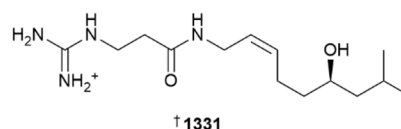
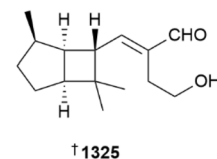
The effect of the saponin frondoside A on inflammation associated with tooth replantation has been investigated.⁵⁶⁹ Holothurin B and related saponins are able to prevent mast cell degranulation, as measured by the release of β -hexosaminidase, and to alleviate the action of a number of pro-inflammatory cytokines.⁵⁷⁰ The chemistry and biological activities of MNPs reported from the sea cucumber *Apostichopus japonicus* (2000 to late 2023) have been reviewed.⁵⁷¹ Holotoxin A₁, isolated from *A. japonicus*, inhibits candidiasis by a wide range of mechanisms including increased intracellular accumulation of ROS, mitochondrial membrane depolarisation, and inhibition of mitochondrial complex I.⁵⁷² Further examination of the triterpenoid glycoside cucumarioside A₂-2 has identified it to be active against prostate cancer cell lines, while cucumarioside A₀-1 and djakonovioside A are active against triple-negative breast cancer cell lines.^{573,574}

12 Miscellaneous

The structure of the bicycloheptane derivative (–)-raikovenal **1325**, originally isolated from the marine ciliate *Euplotes raikovi*, has been confirmed by total synthesis, with the study also assigning absolute configuration.⁵⁷⁵ The potential of seagrass meadows as a source of bioactive molecules has been reviewed,⁵⁷⁶ while species specific reviews of *Halophila stipulacea*⁵⁷⁷ and *Halodule uninervis*⁵⁷⁸ covered the phytochemistry and pharmacological activities of metabolites. Further investigation of the diarylheptanoid heterodimer zosterabisphenone B determined it can induce apoptosis in HCT116 cells and reduce the growth of human colon tumour xenografts in mice.⁵⁷⁹

Five pyridyl and dihydroisoquinoline alkaloids, **1326–1330**, were reported from the nemertean worm *Amphiporus angulatus*.⁵⁸⁰ Collections of the intertidal sipunculid worm *Phascolosoma granulatum* afforded phascolosomines A–F **1331–1336**, characterised as linear guanidine amides.⁵⁸¹ A SAR study of the antimicrobial peptide capitellacin, originally isolated from the annelid worm *Capitella teleta*, identified the importance of one

of the disulfide linkages for activity, that the linear peptide was devoid of activity, the D-enantiomer had reduced activity and that most analogues only exhibited weak, at best, activity towards *E. coli*.⁵⁸² Bioinformatic searching of the transcriptome of the lugworm *Arenicola marina* identified a new BRICHOS-related antimicrobial peptide, AmbRI-aaa, that was prepared using heterologous expression in *E. coli*.⁵⁸³ The 44 residue peptide exhibited weak to moderate activity towards some Gram-positive bacterial strains and whole genome sequencing of a resistant mutant identified targets associated with cell wall biosynthesis.



The ribbon worm *Cephalothrix simula*, known to contain tetrodotoxin, may be a source of toxin in British farmed Pacific oysters *Magallana gigas*.⁵⁸⁴ A truncated variant of a peptide, bolespleenin,^{334–347} derived from a gene product of the mudskipper fish *Boleophthalmus pectinirostris*, exhibited broad-spectrum antibacterial activity, inhibited biofilm formation and was active in a mouse model of skin infection.⁵⁸⁵ Further investigation of the peptide epinecidin-1, originally isolated from the orange-spotted grouper *Epinephelus coioides*, has determined its ability to suppress lipoteichoic acid-enhanced proliferation of non-small cell lung cancer cells and induce necrotic cell death *via* a mechanism involving mitochondrial damage and elevated levels of ROS.⁵⁸⁶ The 22-residue peptide piscidin-1, derived from mast cells of the hybrid striped bass *Morone saxatilis* \times *M. chrysops*, induces apoptosis in oral squamous cell carcinoma cells, and also increases ROS levels leading to mitochondrial dysfunction.⁵⁸⁷

GC/electron capture negative ion-MS analysis of seventeen species of fish from the Seychelles in the Western Indian Ocean identified the predominant polyhalogenated compounds present were halogenated NPs of the methoxylated diphenyl ether class, rather than PCB or DDT anthropogenic persistent pollutants.⁵⁸⁸

13 Perspective – the emerging role of artificial intelligence in MNP research

The rapid uptake of artificial intelligence (AI) in commerce, health, transportation and communications is stunning. In the sciences, AI is equally transformative. AI is used to analyse large amounts of different kinds of data, to find patterns and relationships, and use those to make predictions, decisions or generate “new” content. Machine learning (ML) is the most



widely used approach, and it identifies patterns in data without relying on rules-based programming, allowing tasks to be performed automatically. When dealing with more complex, high-dimensional, unstructured data, a specialized subset of ML called deep-learning (DL) employs multilayered neural networks to identify intricate patterns.

Given that computer assisted analysis have been used for decades in MNP research, how much has the introduction of AI improved productivity in MNP research and increased the discovery of novel and/or potentially active MNPs in the last decade?

Two main applications of AI are generative (GAI) and predictive (PAI). GAI is based on large language models (LLM) mostly trained on online data and analysed using ML or DL algorithms to produce content copied from existing data but stitched together in different ways to construct a “new” amalgam as text, image or video that mimics the training data. GAI can aid researchers to write code for software packages such as Python and R, produce images for graphical abstracts or presentations, help construct grammatically correct text, and provide suggestions for areas to investigate when initiating research on an unfamiliar topic. But GAI needs to be used with caution because it has a propensity to generate wildly inaccurate results. For instance, creating images of chemical structures using GAI is particularly unreliable (Fig. 2).⁵⁸⁹

PAI by contrast can analyse large amounts of orthogonal data to identify patterns that are often hidden or difficult to discern and predict new findings based on this pattern recognition.

The application of AI in the molecular sciences is mostly predictive rather than generative and this approach has resulted in important breakthroughs. The development of AlphaFold, an AI model to predict 3D protein structures based on amino acid sequence, for example, is transformative. Using a transformer neural network architecture trained on ~180 000 experimentally determined 3D protein structures, AlphaFold can now predict the 3D structures of over 200 million proteins.⁵⁹⁰ These predictions are now being incorporated into the drug discovery process.

Computational methods such as TDDFT calculations to assign absolute configurations to molecules, spectroscopic simulations, computer-assisted structure elucidation using rules-based algorithms, compound similarity and substructure

searches, dereplication using spectral database matching tools and virtual screening of compound libraries through *in silico* protein docking are all computer assisted (but not AI) methods developed since the 1990s. Many of these methods are still widely used in NP research and most involve complex calculations that require supercomputers to do time-consuming analyses. The arrival of ML has created opportunities to massively reduce the time required to generate similar results without using supercomputers.^{591–594}

ML is now used as an aid in MS-based dereplication by predicting MS fragmentation patterns for NPs found in databases to augment the annotation of molecules in molecular networks that were previously only identified through matching experimental spectrometric data.⁵⁹⁵ Augmenting lower level DFT calculations with ML generated parametric corrections has resulted in more accurate predictions of NMR data in minutes rather than days.^{596,597} ML assisted genome mining has resulted in the discovery of BGCs and the prediction of their products.^{598,599}

MNP researchers still face many bottlenecks that impede productivity. These include access to biota, accurate taxonomic identification of biota, identification of “interesting” NPs especially those present in complex mixtures, separation and structure identification of NPs, identifying a NP's bioactivity and understanding its mode of action, identifying the genes and proteins responsible for NP biosynthesis and synthesis of the NP for sustainable production or to generate analogues.

We analysed 13 582 papers and reviews published in the field of MNP (obtained from MarinLit) over the last 10 years (January 2015–December 2024) to help understand how AI has contributed to solving these MNP research bottlenecks. Keyword searches (Table S2) of all abstracts allowed a connectivity map (using VOSviewer)⁶⁰⁰ to be generated by creating a thesaurus of keywords that we grouped into ontologies based on relatedness of the words. The limitation of this approach is that some abstracts do not necessarily include information regarding the use of AI, but the keyword connectivity maps can help to infer AI usage based on a combination of ontologies. A network from genome mining to bacteria, BGC, molecular network, predict, MS, bioinformatic, model and database for example infers AI is being used in microbial MNP research even though there is no explicit association between the use of AI terms and genome mining in the abstracts (Fig. 3).

We found that very few studies have used AI to aid in the discovery of new MNPs or bioactivities over the last decade. Overall, the rate of discovery of new MNPs (Fig. 1) and potentially active MNPs has remained static over the decade.⁶⁰¹

AI has only been explicitly referred to in 82 MNP papers over this period (Fig. 4 and 5) and is most highly associated with the prediction of bioactivity through either *in silico* docking of MNPs to proteins or inferred bioactivity based on structural similarities. ML augmentation of DFT calculations for more accurate NMR chemical shift prediction (DU8ML),⁵⁹⁶ pattern recognition within 2D NMR spectra to predict structures (SMART2.0/DeepSAT),^{602,603} pattern recognition within structures to predict NP structure classes (NPclassifier),⁶⁰⁴ prediction of peptide quaternary structures (conotoxins),⁶⁰⁵ and genome

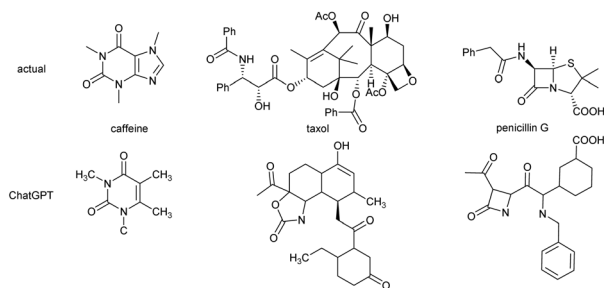


Fig. 2 Structures of three ubiquitous NPs (caffeine, taxol and penicillin G) and their “structures” generated by ChatGPT. The “structures” generated by Gemini and DeepSeek are presented in the SI (Fig S1).



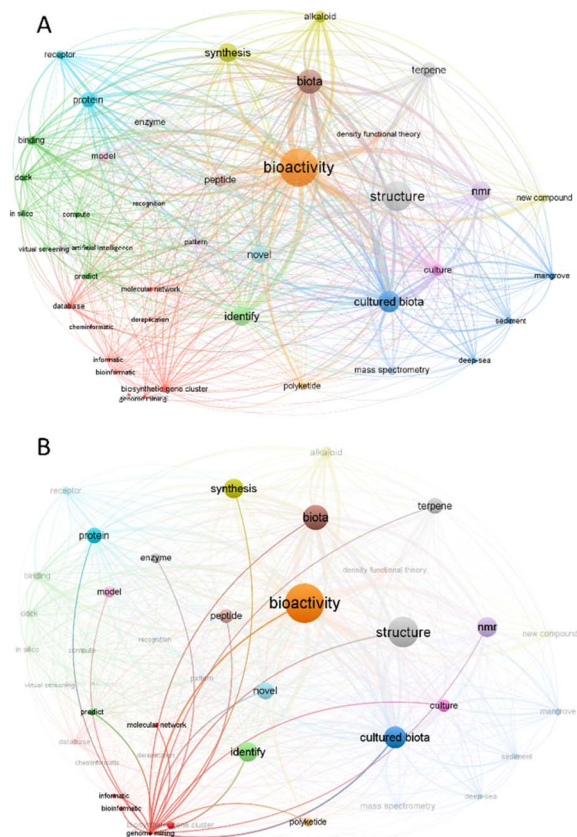


Fig. 3 Connectivity map (created in VOSviewer)⁶⁰⁰ of network linkages between keywords extracted from MNP paper abstracts. The size of a keyword node is proportional to the number of papers that contain that ontology and the width of the linkage to another node is proportional to the number of instances that the linkage occurs. Each of the top 21 nodes has a individual colour. (A) Total network; (B) highlighted network from “genome mining”.

mining to generate hypothetical virtual libraries of potential structures of undiscovered molecules for MS molecular network matching (HypoRiPPAtlas)⁵⁹⁹ are examples of other uses. To date, none of these methods have been widely adopted.

In silico screening methods (233 papers) have steadily increased over the decade. There have been several studies that have used databases to predict MNPs active against specific targets (such as the SARS-COV 19 virus) through virtual screening. This approach previously used the knowledge of the protein structure (obtained mostly by XRD) and *in silico* docking techniques to identify molecules that interacted with the protein. However, AI methods now tend to use the knowledge gained from molecules that have been shown experimentally (or virtually through *in silico* methods) to interact with a specific target protein (or a structurally related protein) and these interactions are used to train a model to identify other compounds that will also interact. This pattern recognition approach is potentially quicker to implement and can be used to analyse large datasets. Unfortunately, where this approach has been adopted using MNPs, the verification of “hits” obtained mostly relies on virtual screening of the “hit” molecules

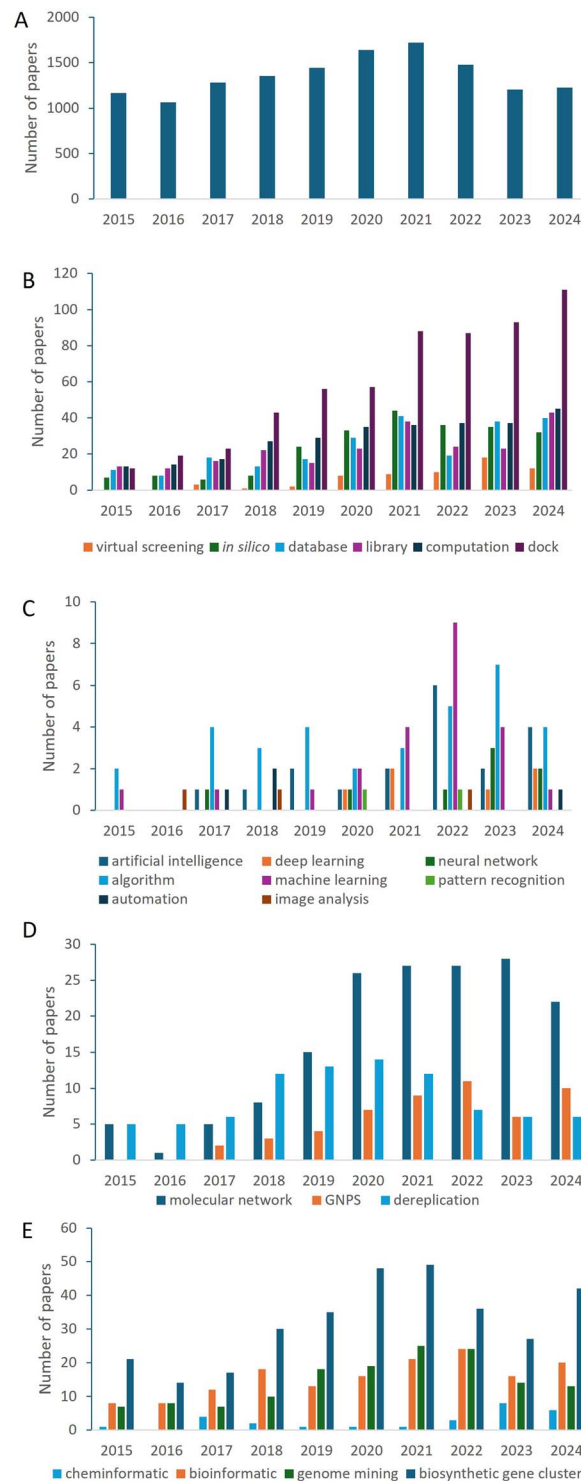


Fig. 4 Trends in number of papers using keywords in their abstracts from January 2015 to December 2024. (A) Total MNP papers, (B) papers discussing virtual screening and interactions of MNPs with proteins; (C) papers discussing AI; (D) papers discussing MS molecular networking; (E) bioinformatics application in MNP research. The bars represent the total number papers.

through molecular docking experiments. This seems to be a circular approach given that in some cases, the training data was originally obtained through virtual screening. There has



sciences, ecological studies and metagenomics for ML might help to uncover specific nutrient requirements that could enhance the isolation and cultivability of truly marine species.

MNP discovery continues to rely on serendipity. There is no evidence so far that AI has been applied to the biota selection process. At its most basic, MNP researchers have used prior knowledge to target specific species for study (soft bodied marine invertebrates, algae, fungi, bacteria, cyanobacteria). AI could aid in this process. However, we know from experience that the random collection of an invertebrate that was collected simply because it looked different or was found in an unusual place, has led to some of the most unusual molecules we have discovered. Perhaps an AI assistant on a SCUBA dive could help find that elusive organism.

Genome mining helps to identify the vast potential of BGCs (most of which are silent) in microorganisms, but what about the NPs that are not made by enzymes encoded by clustered genes? Pertinent examples are many types of alkaloids which represent a substantial proportion of MNPs and drugs derived from NPs. ML methods using AlphaFold could provide insights to predict proteins from genomic data for non-clustered biosynthesis.

The structure determination of NPs has always been challenging, and continued reports of MNP structure revisions show that improved methods to aid in structure determination are always welcomed. Computationally demanding DFT methods to predict NMR chemical shifts have been augmented with parametric rules obtained through ML approaches using NMR data obtained from large libraries of published structures. This has resulted in much faster predictions and resulted in two MNP structure revisions.⁵⁹⁶ The tool can also be used to establish if a proposed structure is correct prior to publication. SMART2.0 is an image-based AI tool that predicts the structure of a compound based on the similarity of its HSQC image to those generated from a training set of over 53 000 experimental HSQC images or HSQC images generated through predictions of H and C chemical shifts for compounds lacking experimental data.⁶⁰² The four MNP studies that have used SMART2.0 to guide prioritisation all resulted in the isolation of either close analogues of known compounds or a chimeric molecule containing features from two known molecules. Overall, these ML NMR methods can improve the structure determination workflow, but do not necessarily result in greater novelty of structures being published. Hopefully, it will reduce the number of incorrect structures appearing in the literature (if used judiciously at the time of peer review prior to publication).

Open Molecules 2025 (OMol25), released in mid-2025 by Meta and the US Department of Energy's Lawrence Berkeley National Laboratory, contains 100 million DFT calculations covering a huge variety of molecular systems that can now be used in ML applications.⁶⁰⁹ This has the potential to massively reduce the time required to predict NMR and ECD data with estimates suggesting a 10-thousand-fold reduction in cpu time to generate equivalent quality predictions to those generated by DFT calculations. This resource has the potential, through ML algorithms, to predict NMR and ECD data for all known MNPs with comparable accuracy to quantum mechanical calculations

virtually instantaneously on a laptop or smart phone. Linking this potential capability with other AI platforms such as 2D NMR image recognition, MS² data recognition and NP structure prediction from genomic data and NP bioactivity predictions could provide a paradigm shift in NP discovery efforts.

Since ML methods are trained on known data (such as molecules, NMR spectra, MS² data, genes) there is an obvious bias to match unknowns that mimic knowns. "Rare" structures might not feature in a training set, and undiscovered novel molecules clearly cannot be there at all. Therefore, important potential matches might be missing in the model. This is analogous to the inherent bias used by Amazon to recruit IT staff where female applicants were culled early in the automated selection process because the data used to train the algorithm did not include applications from women.⁶¹⁰ Training models to highlight and prioritise the unknowns will help to counter these biases.

There is a danger when AI is used as a "black box" method by non-experts who accept the results as gospel without any understanding or questioning because of a lack informed knowledge. If these unverified AI results are subsequently used to train new models, a misinformation loop is exacerbated, rendering future models even less accurate than their predecessors. There is, therefore, an ongoing need for orthogonal testing of results obtained from AI tools in MNP research. In structure determination, this means that XRD and total synthesis will continue to be important verification tools. Laboratory-based bioassays need to be used to confirm or reject an AI computational result and MS² fragmentation matching for structure annotation will still rely on real MS data from standards.

There have been highly innovative AI methods developed, and some have been widely adopted while others have not. It is somewhat baffling to know why this is the case. For instance, molecular networking is now widely adopted with platforms such as GNPS and applications such as MZmine providing a simple computational engine for upload of raw data and download of clustered and annotated results.^{611,612} The barrier to adopting this technology is low and labs using these platforms benefit from their simplicity to access. Equally as innovative is the SMART2.0/DeepSAT platform for interpretation of 2D NMR data to match known compounds and to predict structure classes for unknown compounds. However, the adoption of these methods by the MNP community is low (only four papers refer to SMART2.0 in their abstract). One reason may be that many MNP research groups have NMR "gurus" who can identify a known structure from limited experimental data extremely quickly and do not need to rely on computational methods. However, the most likely reason for slow uptake is that these methods still do not work well with NMR of mixtures (apart from matching major components). The time involved to purify an extract and then obtain HSQC data on individual molecules compared to injecting a minute quantity of an extract into an HPLC column and generating MS² data for thousands of compounds in minutes is likely to be a serious impediment.

Negative results are a major gap in published knowledge. For example, many MNP groups have large inventories of marine



species collected or cultured randomly. These collections invariably contain individual samples that have been studied but either produce no MNPs or only known MNPs or the isolated MNPs showed low or no bioactivity in a range of bioassays. These results do not generally make it into research papers but are a gold mine of information that could be used in AI analysis. To make the most of all research outputs therefore requires an acceptable mechanism to capture open access, validated negative research results as well as positive ones.

Linking datasets will be key to seeing the most productive uptake and application of AI in MNP research. The World Register of Marine Species (WoRMS) contains up-to-date taxonomic classifications, collection records and distributions for over 500 000 species,⁶¹³ the Ocean Biodiversity Information System contains 167 million geolocation records for over 200 000 marine species and 27 million gene sequences,⁶¹⁴ citizen science initiatives such as iNaturalist have images of species linked to taxonomic data and geolocation with close to 280 million observations linked to 536 000 species (including 4041 observations from 1280 marine invertebrate species),⁶¹⁵ the Lotus initiative provides taxonomic links to over 750 000 NP structure/organism pairs,⁶¹⁶ the newly released Minimum Information about a Tailoring Enzyme (MITE) links biosynthetic enzymes with reaction type and precursor and product structures,⁶¹⁷ SMART2.0 uses a database of over 50 000 HSQC images to link with structures, GNPS and other MS based platforms link MS² data to structures. Linking these disparate datasets could provide opportunities to guide new discoveries and to unlock novelty in plain sight.

This perspective has highlighted that MNP researchers are only beginning to embrace the power of AI and it will be fascinating to see where MNPs research in the AI era evolves over the coming decade.

14 Conclusion

New MNP discoveries in 2024 have shown an increasing prevalence of MNPs compounds derived from marine fungi and a decreasing trend in macro-organism studies. Although close to half of all new MNPs were reported from marine fungi in 2024, the majority are minor variations on already well represented structure classes. Actinomycetes are a minor contributor to overall marine procaryote biodiversity, but they continue to be the major source of new bacterial MNPs.

BCGs associated with MNPs obtained from bacteria, cyanobacteria and to a lesser extent fungi continue to be characterised, and genome mining is increasingly being used to find new bacterial MNPs. In some cases, these genomic approaches have resulted in proposed new MNP structures without comprehensive and convincing spectroscopic evidence and from our perspective these compounds have not been structurally verified. Rigorous spectroscopic characterisation including NMR spectroscopy and or XRD analysis should complement MS-based evidence in future studies.

The bioactivities reported for select MNPs continue to be a highlight, with the clinical development of a synthetic derivative of the Indonesian sponge-derived alkaloid PM742 holding

promise for lung cancer patients. Synthetic studies continue to be an important technique to support or refute MNP structures and in 2024, over 120 first total syntheses of MNPs were accomplished, leading to several structure revisions and over 10 absolute configurational assignments being established.

New sponge-derived MNPs had fewer than expected examples of peptides and alkaloids based on previous trends. Cnidarian studies were predominantly associated with organisms collected from Chinese, Vietnamese and Taiwanese waters in the South China Sea pointing to reduced emphasis on cnidarian biodiversity hotspots. Molluscs, ascidians, bryozoans, red, green and brown algae were, in total, only represented by a handful of studies, while echinoderms were mainly studied from Russian collections. These observations point to diminished geospatial and phylogenetic diversity of MNP discoveries. If these biodiversity trends continue, the overall relevance of MNP research will be diminished. That notwithstanding, the contributions of new MNPs to the structural diversity of organic chemistry remains high and they represent a valuable resource for biodiscovery.

15 Conflicts of interest

There are no conflicts to declare.

16 Data availability

Supplementary information (SI): all structures along with their names, taxonomic origins, collection locations, and biological activities. See DOI: <https://doi.org/10.1039/d5np00080g>.

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18 References

- 1 A. R. Carroll, B. R. Copp, T. Grkovic, R. A. Keyzers and M. R. Prinsep, *Nat. Prod. Rep.*, 2025, **42**, 257–297, DOI: [10.1039/D4NP00067F](https://doi.org/10.1039/D4NP00067F).
- 2 <https://pubs.rsc.org/marinlit>, accessed July 2025.
- 3 Y. Zou, J. Shi, J. L. Sun, L. Y. Li, Z. Y. Yan, Z. K. Guo and R. H. Jiao, *J. Nat. Prod.*, 2024, **87**, 2530–2536, DOI: [10.1021/acs.jnatprod.4c00836](https://doi.org/10.1021/acs.jnatprod.4c00836).



- 4 M. Girão, J. Murillo-Alba, J. Martín, I. Pérez-Victoria, R. B. Leite, R. Urbatzka, P. N. Leão, M. F. Carvalho and F. Reyes, *Mar. Drugs*, 2024, 22, 268, DOI: [10.3390/md22060268](https://doi.org/10.3390/md22060268).
- 5 J. I. Quintana-Bulla, L. A. C. Tonon, L. F. Michaliski, E. Hajdu, A. G. Ferreira and R. G. S. Berlinck, *Appl. Microbiol. Biotechnol.*, 2024, 108, 112, DOI: [10.1007/s00253-023-12870-0](https://doi.org/10.1007/s00253-023-12870-0).
- 6 M. Elsbaey, T. Jomori, J. Tanaka, N. Oku and Y. Igarashi, *J. Antibiot.*, 2024, 77, 389–392, DOI: [10.1038/s41429-024-00718-2](https://doi.org/10.1038/s41429-024-00718-2).
- 7 H. P. S. de Freitas Cesário, F. das Chagas Lima Pinto, K. M. Canuto, E. R. Silveira, D. V. Wilke, E. G. Ferreira, A. M. da Fonseca, M. A. de Vasconcelos, E. H. Teixeira and O. D. L. Pessoa, *Chem. Biodiversity*, 2024, 21, e202301771, DOI: [10.1002/cbdv.202301771](https://doi.org/10.1002/cbdv.202301771).
- 8 M. Wang, Z. Zheng, Z. Hu, B. Fan, J. Liu and Q. Xu, *Nat. Prod. Res.*, 2024, 38, 2681–2687, DOI: [10.1080/14786419.2023.2200183](https://doi.org/10.1080/14786419.2023.2200183).
- 9 W. Zhao, H. Jiang, Y. Ge, C. Zhou, Y. Ma, J. Zhou, Y. Xie, Y. Wang and B. Wu, *Fitoterapia*, 2024, 175, 105946, DOI: [10.1016/j.fitote.2024.105946](https://doi.org/10.1016/j.fitote.2024.105946).
- 10 J. Lee, S. Um, E.-H. Kim and S. H. Kim, *J. Nat. Prod.*, 2024, 87, 733–742, DOI: [10.1021/acs.jnatprod.3c00843](https://doi.org/10.1021/acs.jnatprod.3c00843).
- 11 L. Zhou, Y. Chang, S. Yang, X. Huang, J. Wang, C. Jiang, T. Zhu, D. Li and Q. Che, *J. Antibiot.*, 2024, 77, 201, DOI: [10.1038/s41429-023-00698-9](https://doi.org/10.1038/s41429-023-00698-9).
- 12 P. F. Hillman, J. Y. Lee, J. Lee, J. Lee, D.-C. Oh, W. Fenical and S.-J. Nam, *Nat. Prod. Res.*, 2024, 38, 2357–2362, DOI: [10.1080/14786419.2023.2173192](https://doi.org/10.1080/14786419.2023.2173192).
- 13 S. Um, H. Jeong, J.-E. Park, J. Seo, S. H. Jung, M. Bae, K.-T. Lee and K. Moon, *Mar. Drugs*, 2024, 22, 539, DOI: [10.3390/md22120539](https://doi.org/10.3390/md22120539).
- 14 W. Tang, D. Zhang, J. Xu, S. Wang, B. Wei and L. Li, *Arab. J. Chem.*, 2024, 17, 105687, DOI: [10.1016/j.arabjc.2024.105687](https://doi.org/10.1016/j.arabjc.2024.105687).
- 15 J. Park, Y. Shin, S. Hwang, J. Kim, D. H. Moon, I. Kang, Y. Ko, B. Chung, H. Nam, S. Kim, K. Moon, K. Oh, J. Cho, S. K. Lee and D. Oh, *Angew. Chem., Int. Ed.*, 2024, 63, e202402465, DOI: [10.1002/anie.202402465](https://doi.org/10.1002/anie.202402465).
- 16 F. Pu, J. Fang, W. Li, B. Zhang, X. Hong, L. Xu, X. Li and Y. Jiang, *Chem. Biodiversity*, 2024, 21, e202400029, DOI: [10.1002/cbdv.202400029](https://doi.org/10.1002/cbdv.202400029).
- 17 W. Ding, Y. Li, X. Li, J. Yin, S. Shi, X. Tian, S. Zhang and H. Yin, *Mar. Drugs*, 2024, 22, 181, DOI: [10.3390/md22040181](https://doi.org/10.3390/md22040181).
- 18 W. Wang, J. Shang, S. Yin, K. Xia, R. Ma, W. Zhu and P. Fu, *J. Nat. Prod.*, 2024, 87, 2580–2591, DOI: [10.1021/acs.jnatprod.4c00702](https://doi.org/10.1021/acs.jnatprod.4c00702).
- 19 X. Xu, Y. Chang, Y. Chen, L. Zhou, F. Zhang, C. Ma, Q. Che, T. Zhu, B. A. Pfeifer, G. Zhang and D. Li, *Org. Lett.*, 2024, 26, 7489–7494, DOI: [10.1021/acs.orglett.4c02074](https://doi.org/10.1021/acs.orglett.4c02074).
- 20 J. L. Wissner, J. R. Almeida, I. R. Grilo, J. F. Oliveira, C. Brizida, W. Escobedo-Hinojosa, P. Pissaridou, M. I. Vasquez, I. Cunha, R. G. Sobral, V. Vasconcelos and S. P. Gaudêncio, *Front. Chem.*, 2024, 12, 1425953, DOI: [10.3389/fchem.2024.1425953](https://doi.org/10.3389/fchem.2024.1425953).
- 21 J. Yu, H. Guo, J. Zhang, J. Hu, H. He, C. Chen, N. Yang, F. Yang, Z. Lin, H. Dai, L. Ouyang, C. Liu, X. Lei, L. Zhang, G. Zhu and F. Song, *Mar. Drugs*, 2024, 22, 259, DOI: [10.3390/md22060259](https://doi.org/10.3390/md22060259).
- 22 H. Huang, Y. Zhang, Y. Song, C. Ling, S. Peng, B. Ding, Y. Tao and J. Ju, *Mar. Drugs*, 2024, 22, 32, DOI: [10.3390/md22010032](https://doi.org/10.3390/md22010032).
- 23 M. Zhao, W. Zhang, C. Yang, L. Zhang, H. Huang, Y. Zhu, D. Ratnasekera and C. Zhang, *J. Nat. Prod.*, 2024, 87, 1591–1600, DOI: [10.1021/acs.jnatprod.4c00232](https://doi.org/10.1021/acs.jnatprod.4c00232).
- 24 K.-X. Yi, Q.-Y. Xie, Q.-Y. Ma, L. Yang, H.-F. Dai, Y.-X. Zhao and Y.-E. Hao, *Fitoterapia*, 2024, 173, 105814, DOI: [10.1016/j.fitote.2023.105814](https://doi.org/10.1016/j.fitote.2023.105814).
- 25 Y. Chen, H. Jin, W. Xiong, Z. Fang, W. Sun, Y. Zhu, L. Zhang, Y. Zhang, W. Zhang and C. Zhang, *Org. Lett.*, 2024, 26, 1677–1682, DOI: [10.1021/acs.orglett.4c00168](https://doi.org/10.1021/acs.orglett.4c00168).
- 26 S. Shigeno, M. Kadowaki, K. Nagai, K. Hosoda, T. Terahara, T. Nishimura, N. Hasegawa, H. Tomoda and T. Ohshiro, *J. Antibiot.*, 2024, 77, 265–271, DOI: [10.1038/s41429-024-00710-w](https://doi.org/10.1038/s41429-024-00710-w).
- 27 W. Ding, Y. Li, X. Li, S. Shi, J. Yin, X. Tian, M. Xiao, S. Zhang and H. Yin, *Phytochemistry*, 2024, 222, 114101, DOI: [10.1016/j.phytochem.2024.114101](https://doi.org/10.1016/j.phytochem.2024.114101).
- 28 X.-Y. Feng, J.-H. Li, R.-J. Li, S.-Z. Yuan, Y.-J. Sun, X.-P. Peng, H. Dong, H.-X. Lou and G. Li, *J. Agric. Food Chem.*, 2024, 72, 1082–1095, DOI: [10.1021/acs.jafc.3c06307](https://doi.org/10.1021/acs.jafc.3c06307).
- 29 Z. Wang, J. Yin, M. Bai, J. Yang, C. Jiang, X. Yi, Y. Liu and C. Gao, *Mar. Drugs*, 2024, 22, 38, DOI: [10.3390/md22010038](https://doi.org/10.3390/md22010038).
- 30 L. Chen, K. Liu, J. Hong, Z. Cui, W. He, Y. Wang, Z. Deng and M. Tao, *Mar. Drugs*, 2024, 22, 189, DOI: [10.3390/md22040189](https://doi.org/10.3390/md22040189).
- 31 H. J. Shin, S.-Y. Jung, J. S. Kang, C.-S. Heo and S. J. Park, *J. Nat. Prod.*, 2024, 87, 2432–2440, DOI: [10.1021/acs.jnatprod.4c00683](https://doi.org/10.1021/acs.jnatprod.4c00683).
- 32 Y. Xu, F. Pu, Y. Cui, Y. Wu, Z. Lei and Y. Jiang, *Chem. Biodiversity*, 2024, 21, e202401154, DOI: [10.1002/cbdv.202401154](https://doi.org/10.1002/cbdv.202401154).
- 33 S.-M. Shen, Y.-C. Xie, L.-R. Tu, M.-E. Wu, Y.-M. Wang, C.-H. Song, Y.-H. Sun and M.-H. Luo, *Mar. Drugs*, 2024, 22, 521, DOI: [10.3390/md22110521](https://doi.org/10.3390/md22110521).
- 34 Q. Dai, M. Ma, N. Wang, Y. Zhou and Z. Zhang, *Fitoterapia*, 2024, 178, 106176, DOI: [10.1016/j.fitote.2024.106176](https://doi.org/10.1016/j.fitote.2024.106176).
- 35 S. Pulat, I. Yang, J. Lee, S. Hwang, R. Zhou, C. D. B. Gamage, M. Varlı, İ. Taş, Y. Yang, S.-Y. Park, A. Hong, J.-H. Kim, D.-C. Oh, H. Kim, S.-J. Nam and H. Kang, *Mar. Drugs*, 2024, 22, 88, DOI: [10.3390/md22020088](https://doi.org/10.3390/md22020088).
- 36 W. Liu, S. Zhai, L. Zhang, Y. Chen, Z. Liu, W. Ma, T. Zhang, W. Zhang, L. Ma, C. Zhang and W. Zhang, *J. Nat. Prod.*, 2024, 87, 371–380, DOI: [10.1021/acs.jnatprod.3c01132](https://doi.org/10.1021/acs.jnatprod.3c01132).
- 37 S. Liu, C. Sun, Y. Ha, M. Ma, N. Wang, Y. Zhou and Z. Zhang, *Tetrahedron Lett.*, 2024, 137, 154935, DOI: [10.1016/j.tetlet.2024.154935](https://doi.org/10.1016/j.tetlet.2024.154935).
- 38 S. Tan, Y. Liu, H. Fu, Y. Xue, F. Feng, J. Li, X. Yan, S. Wu, S. He and L. Ding, *J. Oceanol. Limnol.*, 2024, 42, 1991–1997, DOI: [10.1007/s00343-024-3195-9](https://doi.org/10.1007/s00343-024-3195-9).
- 39 H. P. S. d. F. Cesário, F. C. O. Silva, M. K. A. Ferreira, J. E. S. A. de Menezes, H. S. dos Santos, A. M. da Fonseca,



- C. E. S. Nogueira, M. M. Marinho, E. S. Marinho, A. M. R. Teixeira, E. R. Silveira and O. D. L. Pessoa, *J. Biomol. Struct. Dyn.*, 2024, **42**, 445–460, DOI: [10.1080/07391102.2023.2193988](https://doi.org/10.1080/07391102.2023.2193988).
- 40 L. Xu, R. Liu, J. Fang, N. Zhang, F. Pu, Z. Lei, W. Ding and Y. Jiang, *Chem. Biodiversity*, 2024, **21**, e202301712, DOI: [10.1002/cbdv.202301712](https://doi.org/10.1002/cbdv.202301712).
- 41 D. Sweeney, A. Bogdanov, A. B. Chase, G. Castro-Falcón, A. Trinidad-Javier, S. Dahesh, V. Nizet and P. R. Jensen, *J. Nat. Prod.*, 2024, **87**, 2768–2778, DOI: [10.1021/acs.jnatprod.4c00934](https://doi.org/10.1021/acs.jnatprod.4c00934).
- 42 K. Hosoda, N. Koyama, S. Shigeno, T. Nishimura, N. Hasegawa, A. Kanamoto, T. Ohshiro, H. Tomoda and C. A. Arias, *Antimicrob. Agents Chemother.*, 2024, **68**, e00917–e00923, DOI: [10.1128/aac.00917-23](https://doi.org/10.1128/aac.00917-23).
- 43 H. Satoh, D. Lee, S. Arima, K. Hosoda, S. Shigeno, H. Tomoda, T. Ohshiro and T. Nagamitsu, *Tetrahedron*, 2024, **168**, 134299, DOI: [10.1016/j.tet.2024.134299](https://doi.org/10.1016/j.tet.2024.134299).
- 44 J.-H. Kim, J. Y. Lee, J. Lee, P. F. Hillman, J. Lee, B. Choi, M.-J. Paik, S. Lee and S.-J. Nam, *Molecules*, 2024, **29**, 5539, DOI: [10.3390/molecules29235539](https://doi.org/10.3390/molecules29235539).
- 45 W. Wang, J. Lee, E. Roh, G. Shetye, J. Cao, J. McAlpine, G. Pauli, S. Franzblau, T. Hanh Nguyen Vu, N. T. Quach, E. Oh, K.-H. Park, C. Park, Y. Cho, H. Jang, S. Han, H. Kim, S. Cho, Q.-T. Phi and H. Kang, *J. Nat. Prod.*, 2024, **87**, 976–983, DOI: [10.1021/acs.jnatprod.3c01275](https://doi.org/10.1021/acs.jnatprod.3c01275).
- 46 Z. A. Maw, A. L. Grunwald, B. A. Haltli, C. Cartmell and R. G. Kerr, *J. Nat. Prod.*, 2024, **87**, 1682–1693, DOI: [10.1021/acs.jnatprod.3c01234](https://doi.org/10.1021/acs.jnatprod.3c01234).
- 47 T. Wu, W. M. Hussein, K. Samarasekera, Y. Zhu, Z. G. Khalil, S. Jin, D. F. Bruhn, Y. Moreno, A. A. Salim and R. J. Capon, *Mar. Drugs*, 2024, **22**, 312, DOI: [10.3390/md22070312](https://doi.org/10.3390/md22070312).
- 48 Z. A. Maw, B. Haltli, J. J. Guo, D. M. Baldisseri, C. Cartmell and R. G. Kerr, *Molecules*, 2024, **29**, 1482, DOI: [10.3390/molecules29071482](https://doi.org/10.3390/molecules29071482).
- 49 S. Zhao, P. Yan, P. Liu, H. Liu, N. Li, P. Fu and W. Zhu, *Chin. Chem. Lett.*, 2024, **35**, 108950, DOI: [10.1016/j.cclet.2023.108950](https://doi.org/10.1016/j.cclet.2023.108950).
- 50 Q. Sun, D. Yu, X. Zhang, F. Xiao and W. Li, *Mar. Drugs*, 2024, **22**, 262, DOI: [10.3390/md22060262](https://doi.org/10.3390/md22060262).
- 51 J. H. Lee, B.-K. Choi, M. Kim, H. J. Shin and S. J. Park, *Mar. Drugs*, 2024, **22**, 533, DOI: [10.3390/md22120533](https://doi.org/10.3390/md22120533).
- 52 C.-S. Heo, J. S. Kang, J.-W. Yang, M. A. Lee, H.-S. Lee and H. J. Shin, *Int. J. Mol. Sci.*, 2024, **25**, 3311, DOI: [10.3390/ijms25063311](https://doi.org/10.3390/ijms25063311).
- 53 N. Takatani, T. Maoka, T. Sawabe, F. Beppu and M. Hosokawa, *Appl. Microbiol. Biotechnol.*, 2024, **108**, 102, DOI: [10.1007/s00253-023-12995-2](https://doi.org/10.1007/s00253-023-12995-2).
- 54 Y. Song, Y. Zhou, M. Cong, S. Deng, Y. Chen, X. Pang, Y. Liu, L. Liao, L. Yang and J. Wang, *Mar. Drugs*, 2024, **22**, 484, DOI: [10.3390/md22110484](https://doi.org/10.3390/md22110484).
- 55 Y. Xu, Y. Song, Y. Ning, S. Li, Y. Qu, B. Jiao and X. Lu, *Mar. Drugs*, 2024, **22**, 331, DOI: [10.3390/md22080331](https://doi.org/10.3390/md22080331).
- 56 H.-W. Zhao, Y. Yi, J.-Q. Li and J.-H. Wang, *Nat. Prod. Res.*, 2024, **38**, 206–210, DOI: [10.1080/14786419.2022.2112578](https://doi.org/10.1080/14786419.2022.2112578).
- 57 R. Campbell, L. Kyei, K. Piedl, Z. Zhang, M. Chen and E. Mevers, *Org. Lett.*, 2024, **26**, 9693–9697, DOI: [10.1021/acs.orglett.4c03470](https://doi.org/10.1021/acs.orglett.4c03470).
- 58 R. Feng, Q. Dai, M. Ma, S. Zhao, Z. Wang, H. Wang, Y. Zhang, L. Huo and F. Yan, *J. Nat. Prod.*, 2024, **87**, 2801–2809, DOI: [10.1021/acs.jnatprod.4c00965](https://doi.org/10.1021/acs.jnatprod.4c00965).
- 59 D. Roman, P. Meisinger, R. Guillonneau, C. Peng, L. K. Peltner, P. M. Jordan, V. Haensch, S. Götz, O. Werz, C. Hertweck, Y. Chen and C. Beemelmans, *Angew. Chem., Int. Ed.*, 2024, **63**, e202401195, DOI: [10.1002/anie.202401195](https://doi.org/10.1002/anie.202401195).
- 60 X. Zha, Y. Li, H. Zhao, Y. Tan and S. Zhou, *Molecules*, 2024, **29**, 4446, DOI: [10.3390/molecules29184446](https://doi.org/10.3390/molecules29184446).
- 61 N. Sermkaew, A. Atipairin, S. Krobthong, C. Aonbangkhen, Y. Yingchutrakul, J. Uchiyama and N. Songnaka, *Antibiotics*, 2024, **13**, 846, DOI: [10.3390/antibiotics13090846](https://doi.org/10.3390/antibiotics13090846).
- 62 M. Girão, S. Freitas, T. P. Martins, R. Urbatzka, M. F. Carvalho and P. N. Leão, *Front. Pharmacol.*, 2024, **15**, 1347485, DOI: [10.3389/fphar.2024.1347485](https://doi.org/10.3389/fphar.2024.1347485).
- 63 P. Li, Y. Wu, S. He, H. Lu, S. He and L. Liu, *Chem. Nat. Compd.*, 2024, **60**, 831–834, DOI: [10.1007/s10600-024-04458-5](https://doi.org/10.1007/s10600-024-04458-5).
- 64 R. Ahmad, M. I. Tousif, M. Nazir, A. Yaqoob, S. A. Ali Shah, G. Zengin, A. I. Uba and S. Muhammad, *Food Biosci.*, 2024, **57**, 103600, DOI: [10.1016/j.fbio.2024.103600](https://doi.org/10.1016/j.fbio.2024.103600).
- 65 A. Buljan, V. Stepanić, A. Čikoš, S. B. Brčić, K. Bojanić and M. Roje, *Mar. Drugs*, 2024, **22**, 310, DOI: [10.3390/md22070310](https://doi.org/10.3390/md22070310).
- 66 R. R. Khatua, S. A. Z. Ahmad, S. Das, R. Anindya and F. A. Khan, *Synlett*, 2024, **35**, 1872–1876, DOI: [10.1055/a-2256-5509](https://doi.org/10.1055/a-2256-5509).
- 67 S. S. Auddy, S. Gupta, S. Mandi, H. Sharma, S. Sinha and R. K. Goswami, *ACS Med. Chem. Lett.*, 2024, **15**, 1340–1350, DOI: [10.1021/acsmedchemlett.4c00237](https://doi.org/10.1021/acsmedchemlett.4c00237).
- 68 T. C. Murgatroyd, T. Söhnel and J. Sperry, *Org. Lett.*, 2024, **26**, 10041–10045, DOI: [10.1021/acs.orglett.4c03937](https://doi.org/10.1021/acs.orglett.4c03937).
- 69 A. Dimasi, M. Failla, A. Montoli, A. Citarella, P. Ronchi, D. Passarella and V. Fasano, *RSC Adv.*, 2024, **14**, 5542–5546, DOI: [10.1039/D4RA00589A](https://doi.org/10.1039/D4RA00589A).
- 70 L. Yu, Y. Nagata and H. Nakamura, *J. Am. Chem. Soc.*, 2024, **146**, 2549–2555, DOI: [10.1021/jacs.3c11016](https://doi.org/10.1021/jacs.3c11016).
- 71 F. Fei, S. Lun, A. Saxena, M. Raghavan, J. L. DeRisi, W. R. Bishai and J. K. Sello, *Org. Lett.*, 2024, **26**, 9698–9703, DOI: [10.1021/acs.orglett.4c03473](https://doi.org/10.1021/acs.orglett.4c03473).
- 72 G. G. Dake and K. P. Kaliappan, *J. Org. Chem.*, 2024, **89**, 5825–5832, DOI: [10.1021/acs.joc.4c00462](https://doi.org/10.1021/acs.joc.4c00462).
- 73 S. Niyogi, A. Mondal, M. Nandy, S. Pal, A. Khatua and A. Bisai, *Org. Lett.*, 2024, **26**, 8643–8647, DOI: [10.1021/acs.orglett.4c02436](https://doi.org/10.1021/acs.orglett.4c02436).
- 74 G. Petruncio, Z. Shellnutt, L. L. Young, M. Girgis, W. K. Strangman, R. T. Williamson, K. Kehn-Hall and M. Paige, *ACS Omega*, 2024, **9**, 30993–30997, DOI: [10.1021/acsomega.4c04249](https://doi.org/10.1021/acsomega.4c04249).
- 75 D.-B. Sung, D.-B. Choi, J. H. Seol, N. Kang, E.-A. Kim, S.-Y. Heo, S.-J. Heo and J. S. Lee, *ACS Omega*, 2024, **9**, 27592–27609, DOI: [10.1021/acsomega.4c03241](https://doi.org/10.1021/acsomega.4c03241).



- 76 F. Han, J. Li, S. Li, Z. Wang, Y. Guo and T. Ye, *Angew. Chem., Int. Ed.*, 2024, **63**, e202317636, DOI: [10.1002/anie.202317636](https://doi.org/10.1002/anie.202317636).
- 77 P. Karamanis, J. Muldoon, C. D. Murphy and M. Rubini, *J. Pept. Sci.*, 2024, **30**, e3569, DOI: [10.1002/psc.3569](https://doi.org/10.1002/psc.3569).
- 78 S. Kreeger, B. Sarrels and C. Lucero, *Tetrahedron*, 2024, **163**, 134139, DOI: [10.1016/j.tet.2024.134139](https://doi.org/10.1016/j.tet.2024.134139).
- 79 A. F. Anwar, Y. M. Elbatrawi, D. Degen, R. H. Ebright and J. R. Del Valle, *Org. Lett.*, 2024, **26**, 10056–10060, DOI: [10.1021/acs.orglett.4c04013](https://doi.org/10.1021/acs.orglett.4c04013).
- 80 C. Prudel and U. Kazmaier, *Synlett*, 2024, **35**, 2133–2137, DOI: [10.1055/s-0042-1751578](https://doi.org/10.1055/s-0042-1751578).
- 81 J. Wu and M. J. Krische, *Org. Lett.*, 2024, **26**, 4830–4834, DOI: [10.1021/acs.orglett.4c01644](https://doi.org/10.1021/acs.orglett.4c01644).
- 82 M. J. Bedding, B. C. Forster, A. M. Giltrap, M. T. Stevens, L. Corcilius, W. J. Britton and R. J. Payne, *Org. Lett.*, 2024, **26**, 10993–10998, DOI: [10.1021/acs.orglett.4c04163](https://doi.org/10.1021/acs.orglett.4c04163).
- 83 K. Gan, Y. Zhu, G. Shi, C. Wu, F. Ni, L. Zhao, X. Dou, Z. Zhang and H. Lu, *Chin. J. Chem.*, 2024, **43**, 79–89, DOI: [10.1002/cjoc.202400751](https://doi.org/10.1002/cjoc.202400751).
- 84 S. M. Astle, S. Guggiari, J. R. Frost, H. B. Hepburn, D. J. Klauber, K. E. Christensen and J. W. Burton, *J. Am. Chem. Soc.*, 2024, **146**, 17757–17764, DOI: [10.1021/jacs.4c02969](https://doi.org/10.1021/jacs.4c02969).
- 85 S. Wang, X. Li, W. Yang and R. Huang, *Microb. Biotechnol.*, 2024, **17**, e14533, DOI: [10.1111/1751-7915.14533](https://doi.org/10.1111/1751-7915.14533).
- 86 C.-H. Ji, H.-W. Je, H. Kim and H.-S. Kang, *Nat. Prod. Rep.*, 2024, **41**, 672–699, DOI: [10.1039/D3NP00049D](https://doi.org/10.1039/D3NP00049D).
- 87 H. Put, H. Gerstmans, H. V. Capelle, M. Fauvart, J. Michiels and J. Masschelein, *Nat. Prod. Rep.*, 2024, **41**, 1113–1151, DOI: [10.1039/D3NP00065F](https://doi.org/10.1039/D3NP00065F).
- 88 M. Gonzales, P. Jacquet, F. Gaucher, É. Chabrière, L. Plener and D. Daudé, *J. Nat. Prod.*, 2024, **87**, 1268–1284, DOI: [10.1021/acs.jnatprod.3c00672](https://doi.org/10.1021/acs.jnatprod.3c00672).
- 89 W.-C. Chin, Y.-Z. Zhou, H.-Y. Wang, Y.-T. Feng, R.-Y. Yang, Z.-F. Huang and Y.-L. Yang, *Nat. Prod. Rep.*, 2024, **41**, 977–989, DOI: [10.1039/D3NP00059A](https://doi.org/10.1039/D3NP00059A).
- 90 Z. Liu, W. Sun, Z. Hu, W. Wang and H. Zhang, *Mar. Drugs*, 2024, **22**, 51, DOI: [10.3390/md22010051](https://doi.org/10.3390/md22010051).
- 91 R. T. Magar and J. K. Sohng, *Appl. Microbiol. Biotechnol.*, 2024, **108**, 471, DOI: [10.1007/s00253-024-13296-y](https://doi.org/10.1007/s00253-024-13296-y).
- 92 D. Sweeney, A. B. Chase, A. Bogdanov and P. R. Jensen, *J. Nat. Prod.*, 2024, **87**, 439–452, DOI: [10.1021/acs.jnatprod.3c01007](https://doi.org/10.1021/acs.jnatprod.3c01007).
- 93 A. L. B. Canellas and M. S. Laport, *Lett. Appl. Microbiol.*, 2024, **77**, ovae099, DOI: [10.1093/lambio/ovae099](https://doi.org/10.1093/lambio/ovae099).
- 94 L. T. H. Diethelm, A. B. da Silva Bellini Ramos, G. B. de Lorena, B. I. Trajano, R. D. do Espírito Santo, R. P. B. de Menezes, M. T. Scotti, F. A. Colombo, M. J. Marques, C. R. D. Correia and J. Q. Reimão, *Pharmaceutics*, 2024, **16**, 262, DOI: [10.3390/pharmaceutics16020262](https://doi.org/10.3390/pharmaceutics16020262).
- 95 G. Castro-Falcón, J. Straetener, J. Bornikoel, D. Reimer, T. N. Purdy, A. Berscheid, F. M. Schempp, D. Y. Liu, R. G. Linington, H. Brötz-Oesterhelt and C. C. Hughes, *ACS Chem. Biol.*, 2024, **19**, 743–752, DOI: [10.1021/acscchembio.3c00773](https://doi.org/10.1021/acscchembio.3c00773).
- 96 T. L. de Brito, E. A. Edson, K. G. D. Florêncio, J. A. Machado-Neto, A. D. M. B. Garnique, J. P. M. Luiz, F. de Queiroz Cunha, J. C. Alves-Filho, M. Haygood and D. V. Wilke, *Chem.-Biol. Interact.*, 2024, **400**, 111177, DOI: [10.1016/j.cbi.2024.111177](https://doi.org/10.1016/j.cbi.2024.111177).
- 97 J. Lee, J.-Y. Hwang, D. Oh, D.-C. Oh, H.-G. Park, J. Shin and K.-B. Oh, *Mar. Drugs*, 2024, **22**, 293, DOI: [10.3390/md22070293](https://doi.org/10.3390/md22070293).
- 98 R. M. Johnson, K. Li, X. Chen, G. L. Morgan, J. Aubé and B. Li, *ACS Infect. Dis.*, 2024, **10**, 582–593, DOI: [10.1021/acsinfectdis.3c00504](https://doi.org/10.1021/acsinfectdis.3c00504).
- 99 A. Chatterjee, J. Mondal, S. Paul, H. Sharma, R. K. Goswami and P. Sen, *J. Med. Chem.*, 2024, **67**, 21952–21974, DOI: [10.1021/acs.jmedchem.4c01902](https://doi.org/10.1021/acs.jmedchem.4c01902).
- 100 K. Irie, G. Jeelani, T. Nozaki and A. Iwasaki, *J. Nat. Prod.*, 2024, **87**, 2292–2301, DOI: [10.1021/acs.jnatprod.4c00733](https://doi.org/10.1021/acs.jnatprod.4c00733).
- 101 N. E. Avalon, M. A. Reis, C. C. Thornburg, R. T. Williamson, D. Petras, A. T. Aron, G. F. Neuhaus, M. Al-Hindy, J. Mitrevska, L. Ferreira, J. Morais, Y. E. Abiead, E. Glukhov, K. L. Alexander, F. A. Vulpanovici, M. J. Bertin, S. Whitner, H. Choi, G. Spengler, K. Blinov, A. M. Almohammadi, L. A. Shaala, W. R. Kew, L. Paša-Tolić, D. T. A. Youssef, P. C. Dorrestein, V. Vasconcelos, L. Gerwick, K. L. McPhail and W. H. Gerwick, *J. Am. Chem. Soc.*, 2024, **146**, 18626–18638, DOI: [10.1021/jacs.4c05399](https://doi.org/10.1021/jacs.4c05399).
- 102 F. H. Al-Awadhi, S. Kokkaliari, R. Ratnayake, V. J. Paul and H. Luesch, *J. Nat. Prod.*, 2024, **87**, 2355–2365, DOI: [10.1021/acs.jnatprod.4c00420](https://doi.org/10.1021/acs.jnatprod.4c00420).
- 103 S. Kokkaliari, L. Grauso, A. Mangoni, G. Seabra, V. J. Paul and H. Luesch, *Chemistry*, 2024, **30**, e202401393, DOI: [10.1002/chem.202401393](https://doi.org/10.1002/chem.202401393).
- 104 T.-E. Ngo, A. Ecker, B. Ryu, A. Guild, A. Rimmel, P. D. Boudreau, K. L. Alexander, C. B. Naman, E. Glukhov, N. E. Avalon, V. V. Shende, L. Thomas, S. Dahesh, V. Nizet, L. Gerwick and W. H. Gerwick, *ACS Chem. Biol.*, 2024, **19**, 619–628, DOI: [10.1021/acscchembio.3c00391](https://doi.org/10.1021/acscchembio.3c00391).
- 105 B. Ryu, E. Glukhov, T. R. Teixeira, C. R. Caffrey, S. Madiyan, V. Joseph, N. E. Avalon, C. A. Leber, C. B. Naman and W. H. Gerwick, *J. Nat. Prod.*, 2024, **87**, 1601–1610, DOI: [10.1021/acs.jnatprod.4c00242](https://doi.org/10.1021/acs.jnatprod.4c00242).
- 106 N. Kurisawa, G. Jeelani, T. Nozaki, A. L. Agusta, K. Suenaga and A. Iwasaki, *J. Nat. Prod.*, 2024, **87**, 1838–1843, DOI: [10.1021/acs.jnatprod.4c00499](https://doi.org/10.1021/acs.jnatprod.4c00499).
- 107 R. Fernández, M. Pérez, A. Losada, S. Reboredo, A. G.-S. Juan, M. J. Martín, A. Francesch, S. Munt and C. Cuevas, *Mar. Drugs*, 2024, **22**, 303, DOI: [10.3390/md22070303](https://doi.org/10.3390/md22070303).
- 108 J. J. Mehjabin, C.-S. Phan and T. Okino, *J. Nat. Prod.*, 2024, **87**, 984–993, DOI: [10.1021/acs.jnatprod.3c01272](https://doi.org/10.1021/acs.jnatprod.3c01272).
- 109 K. Umeda, N. Kurisawa, G. Jeelani, T. Nozaki, K. Suenaga and A. Iwasaki, *Beilstein J. Org. Chem.*, 2024, **20**, 645–652, DOI: [10.3762/bjoc.20.57](https://doi.org/10.3762/bjoc.20.57).
- 110 M. Yotsu-Yamashita, R. L. Haddock and T. Yasumoto, *J. Am. Chem. Soc.*, 2002, **115**, 1147–1148, DOI: [10.1021/ja00056a048](https://doi.org/10.1021/ja00056a048).



- 111 M. Yotsu-Yamashita, T. Seki, V. J. Paul, H. Naoki and T. Yasumoto, *Tetrahedron Lett.*, 1995, **36**, 5563–5566, DOI: [10.1016/0040-4039\(95\)01052-J](https://doi.org/10.1016/0040-4039(95)01052-J).
- 112 M. Yotsu-Yamashita, K. Abe, T. Seki, K. Fujiwara and T. Yasumoto, *Tetrahedron Lett.*, 2007, **48**, 2255–2259, DOI: [10.1016/j.tetlet.2007.02.003](https://doi.org/10.1016/j.tetlet.2007.02.003).
- 113 G. Navarro, M. E. Cummings, J. Lee, N. Moss, E. Glukhov, F. A. Valeriote, L. Gerwick and W. H. Gerwick, *Environ. Sci. Technol. Lett.*, 2015, **2**, 166–170, DOI: [10.1021/acs.estlett.5b00116](https://doi.org/10.1021/acs.estlett.5b00116).
- 114 M. Yotsu-Yamashita, K. Umeda, A. Iwasaki and K. Suenaga, *Toxicon*, 2024, **250**, 108122, DOI: [10.1016/j.toxicon.2024.108122](https://doi.org/10.1016/j.toxicon.2024.108122).
- 115 B.-T. Zhang, H. Nishino, R. Kawabe, M. Kamio, R. Watanabe, H. Uchida, M. Satake and H. Nagai, *Biosci. Biotechnol. Biochem.*, 2024, **88**, 517–521, DOI: [10.1093/bbb/zbac015](https://doi.org/10.1093/bbb/zbac015).
- 116 B. Zhang, R. Kawabe, M. Kamio, H. Uchida, N. Matsuura, K. Mori-Yasumoto, M. Satake and H. Nagai, *Tetrahedron Lett.*, 2024, **150**, 155284, DOI: [10.1016/j.tetlet.2024.155284](https://doi.org/10.1016/j.tetlet.2024.155284).
- 117 N. Watanabe, G. Jeelani, T. Nozaki and A. Iwasaki, *ACS Omega*, 2024, **9**, 36795–36801, DOI: [10.1021/acsomega.4c05909](https://doi.org/10.1021/acsomega.4c05909).
- 118 A. Ebihara, R. Taguchi, G. Jeelani, T. Nozaki, K. Suenaga and A. Iwasaki, *J. Nat. Prod.*, 2024, **87**, 1116–1123, DOI: [10.1021/acs.jnatprod.4c00056](https://doi.org/10.1021/acs.jnatprod.4c00056).
- 119 L. A. Elsadek, E. K. Ellis, G. Seabra, V. J. Paul and H. Luesch, *Mar. Drugs*, 2023, **22**, 28, DOI: [10.3390/md22010028](https://doi.org/10.3390/md22010028).
- 120 J. P. A. Reis, S. Freitas, T. Prochazkova and P. N. Leão, *J. Nat. Prod.*, 2024, **87**, 2709–2715, DOI: [10.1021/acs.jnatprod.4c00832](https://doi.org/10.1021/acs.jnatprod.4c00832).
- 121 M. L. Matos-Hernández, R. Samples, G. Dyer, V. M. Casimir Montán, C. A. Morales-Colón, J. M. Salvino, L. J. Montaner, J. A. Cassel, T. E. Messick, I. Tietjen and E. J. E. Caro-Díaz, *J. Nat. Prod.*, 2024, **87**, 1513–1520, DOI: [10.1021/acs.jnatprod.3c01071](https://doi.org/10.1021/acs.jnatprod.3c01071).
- 122 K. Hagimoto, S. Tojo, T. Teruya, M. Yoshida and H. Kigoshi, *Tetrahedron*, 2024, **154**, 133871, DOI: [10.1016/j.tet.2024.133871](https://doi.org/10.1016/j.tet.2024.133871).
- 123 A. Tripathi, J. Puddick, M. R. Prinsep, M. Rottmann, K. P. Chan, D. Y. Chen and L. T. Tan, *Phytochemistry*, 2011, **72**, 2369–2375, DOI: [10.1016/j.phytochem.2011.08.019](https://doi.org/10.1016/j.phytochem.2011.08.019).
- 124 K. Sueyoshi, M. Kaneda, S. Sumimoto, S. Oishi, N. Fujii, K. Suenaga and T. Teruya, *Tetrahedron*, 2016, **72**, 5472–5478, DOI: [10.1016/j.tet.2016.07.031](https://doi.org/10.1016/j.tet.2016.07.031).
- 125 B. P. Smith, N. J. Truax, A. S. Pollatos, M. Meanwell, P. Bedekar, A. F. Garrido-Castro and P. S. Baran, *Angew. Chem., Int. Ed.*, 2024, **63**, e202401107, DOI: [10.1002/anie.202401107](https://doi.org/10.1002/anie.202401107).
- 126 T. Suwa, M. Sasaki and A. Umehara, *Org. Lett.*, 2024, **26**, 4377–4382, DOI: [10.1021/acs.orglett.4c01367](https://doi.org/10.1021/acs.orglett.4c01367).
- 127 R. Shigeta, T. Suzuki, K. Kaneko, H. Tanaka, I. Haishima, K. Norio, A. Tanaka-Yanuma and T. Usuki, *Org. Biomol. Chem.*, 2024, **22**, 4637–4640, DOI: [10.1039/d4ob00580e](https://doi.org/10.1039/d4ob00580e).
- 128 X. Zhao, M. Lv, X. Xi, Y. Lu, L. Wang and Y. Chen, *Chin. J. Chem.*, 2023, **42**, 605–610, DOI: [10.1002/cjoc.202300526](https://doi.org/10.1002/cjoc.202300526).
- 129 K. Tokushige and T. Abe, *Chem.–Eur. J.*, 2024, **30**, e202302963, DOI: [10.1002/chem.202302963](https://doi.org/10.1002/chem.202302963).
- 130 S. B. Waghmode and A. R. Jadhao, *Synthesis*, 2023, **56**, 1407–1414, DOI: [10.1055/a-2222-3822](https://doi.org/10.1055/a-2222-3822).
- 131 K. Annapurna and A. Venkat Narsaiah, *Tetrahedron Lett.*, 2024, **142**, 155081, DOI: [10.1016/j.tetlet.2024.155081](https://doi.org/10.1016/j.tetlet.2024.155081).
- 132 L. T. Tan and N. F. Salleh, *Molecules*, 2024, **29**, 5307, DOI: [10.3390/molecules29225307](https://doi.org/10.3390/molecules29225307).
- 133 Â. P. Matos, F. M. P. Saldanha-Corrêa, R. D. S. Gomes and G. R. Hurtado, *Acta Trop.*, 2024, **251**, 107116, DOI: [10.1016/j.actatropica.2023.107116](https://doi.org/10.1016/j.actatropica.2023.107116).
- 134 A. Bouyahya, S. Bakrim, I. Chamkhi, D. Taha, N. El Omari, N. El Mneyiy, N. El Hachlafi, M. El-Shazly, A. Khalid, A. N. Abdalla, K. W. Goh, L. C. Ming, B. H. Goh and T. Aanniz, *Biomed. Pharmacother.*, 2024, **170**, 115989, DOI: [10.1016/j.biopha.2023.115989](https://doi.org/10.1016/j.biopha.2023.115989).
- 135 A. K. Bishoyi, C. P. Mandhata, C. R. Sahoo, S. K. Paidesetty and R. N. Padhy, *Naumyn-Schmiedeberg's Arch. Pharmacol.*, 2024, **397**, 1347–1375, DOI: [10.1007/s00210-023-02719-8](https://doi.org/10.1007/s00210-023-02719-8).
- 136 D. Kallifidas, D. Dhakal, M. Chen, Q. Y. Chen, S. Kokkaliari, N. A. Colon Rosa, R. Ratnayake, S. D. Bruner, V. J. Paul, Y. Ding and H. Luesch, *Org. Lett.*, 2024, **26**, 1321–1325, DOI: [10.1021/acs.orglett.3c04083](https://doi.org/10.1021/acs.orglett.3c04083).
- 137 P. Mandal, D. Paul, H. Sharma, S. Saha, P. Chakrabarti and R. K. Goswami, *ACS Med. Chem. Lett.*, 2024, **15**, 696–705, DOI: [10.1021/acsmchemlett.4c00094](https://doi.org/10.1021/acsmchemlett.4c00094).
- 138 L. J. Liu, K. R. Francisco, Y. U. Sun, M. S. M. Serafim, D. K. Amarasinghe, T. R. Teixeira, B. Lucero, T. Kronenberger, W. Elsayed, H. Elwakeel, M. Al-Hindy, J. Almaliti, W. H. Gerwick, A. J. O'Donoghue and C. R. Caffrey, *ACS Infect. Dis.*, 2024, **10**, 4182–4193, DOI: [10.1021/acsinfectdis.4c00441](https://doi.org/10.1021/acsinfectdis.4c00441).
- 139 N. Kurisawa, K. Teranuma, A. Noto, A. Iwasaki, Y. Kabashima, R. Nakajima, C. Toyoshima and K. Suenaga, *Bull. Chem. Soc. Jpn.*, 2024, **97**, uoae070, DOI: [10.1093/bulcsj/uoae070](https://doi.org/10.1093/bulcsj/uoae070).
- 140 X. Zhao, X. Xi, M. Zhang, M. Lv, X. Zhang, Y. Lu, L. Wang and Y. Chen, *Mar. Drugs*, 2024, **22**, 537, DOI: [10.3390/md22120537](https://doi.org/10.3390/md22120537).
- 141 É. C. de Almeida, L. S. Passos, P. N. N. de Freitas, A. O. de Souza and E. Pinto, *Aquac. Res.*, 2024, **16**, 1186–1211, DOI: [10.1111/raq.12891](https://doi.org/10.1111/raq.12891).
- 142 L. Biessy, J. Puddick, S. A. Wood, A. I. Selwood, M. Carbines and K. F. Smith, *Toxins*, 2024, **16**, 522, DOI: [10.3390/toxins16120522](https://doi.org/10.3390/toxins16120522).
- 143 X. Hao, S. Li, J. Li, G. Wang, J. Li, Z. Peng and M. Gan, *J. Nat. Prod.*, 2024, **87**, 1059–1066, DOI: [10.1021/acs.jnatprod.4c00015](https://doi.org/10.1021/acs.jnatprod.4c00015).
- 144 Y. Zhang, J.-B. Zhou, S.-T. Yang, X. Liu, W. Cao, P.-H. Li, H. Chen and Y.-Q. Fan, *Mar. Drugs*, 2024, **22**, 25, DOI: [10.3390/md22010025](https://doi.org/10.3390/md22010025).
- 145 C.-L. An, Q.-Y. Ma, Q.-Y. Xie, L. Yang, J.-Z. Yuan, H.-F. Dai and Y.-X. Zhao, *J. Asian Nat. Prod. Res.*, 2024, **26**, 489–496, DOI: [10.1080/10286020.2023.2251114](https://doi.org/10.1080/10286020.2023.2251114).
- 146 Z. Luo, K. Liu, Z. Tang, L. Peng, C. Gao, C. Xia, Y. Liu and X. Chen, *Rec. Nat. Prod.*, 2024, **18**, 136–142, DOI: [10.25135/rnp.438.12310.2940](https://doi.org/10.25135/rnp.438.12310.2940).



- 147 Y.-H. Kim, D.-C. Choi, D.-W. Ki, Y.-S. Won, S.-J. Lee, J.-Y. Kim, I.-K. Lee and B.-S. Yun, *J. Antibiot.*, 2024, 77, 466–470, DOI: [10.1038/s41429-024-00735-1](https://doi.org/10.1038/s41429-024-00735-1).
- 148 S.-T. Fang, Y.-P. Song, Z.-Z. Shi and N.-Y. Ji, *Phytochem. Lett.*, 2024, 61, 218–221, DOI: [10.1016/j.phytol.2024.05.006](https://doi.org/10.1016/j.phytol.2024.05.006).
- 149 J.-J. Xue, F.-P. Miao and S.-T. Fang, *Nat. Prod. Res.*, 2024, 38, 2480–2485, DOI: [10.1080/14786419.2023.2183500](https://doi.org/10.1080/14786419.2023.2183500).
- 150 Y.-S. Lee, H.-C. Wu, S.-J. Huang, G. Hsiao, W.-C. Chi and T.-H. Lee, *Phytochemistry*, 2024, 219, 113998, DOI: [10.1016/j.phytochem.2024.113998](https://doi.org/10.1016/j.phytochem.2024.113998).
- 151 Y. Hu, X. Zhao, Y. Song, J. Jiang, T. Long, M. Cong, Y. Miao, Y. Liu, Z. Yang, Y. Zhu and J. Wang, *J. Nat. Prod.*, 2024, 87, 1975–1982, DOI: [10.1021/acs.jnatprod.4c00393](https://doi.org/10.1021/acs.jnatprod.4c00393).
- 152 P. P. Edjah, M.-F. Lu, W.-C. Chen, J. Wu, Y.-T. Zhong, M. Li, Q. Chen, K.-K. Zhu, R.-Y. Yuan, W.-X. Tao, K.-J. Wu and Y.-S. Cai, *Fitoterapia*, 2024, 179, 106251, DOI: [10.1016/j.fitote.2024.106251](https://doi.org/10.1016/j.fitote.2024.106251).
- 153 P. E. Mandelare, S. S. Yee, D. G. Icenhour, E. N. Kaweesa, M. McCauley, A. L. Risinger, J. D. Rudolf and S. Loesgen, *ChemBioChem*, 2024, 25, e202400398, DOI: [10.1002/cbic.202400398](https://doi.org/10.1002/cbic.202400398).
- 154 Y. Zeng, S. Wang, H. Peng, W. Zhao, W. Chang, H. Wang, H. Chen and H. Dai, *Mar. Drugs*, 2024, 22, 13, DOI: [10.3390/md22010013](https://doi.org/10.3390/md22010013).
- 155 C.-J. Lu, L.-F. Liang, G.-S. Zhang, H.-Y. Li, C.-Q. Fu, Q. Yu, D.-M. Zhou, Z.-W. Su, K. Liu, C.-H. Gao, X.-Y. Xu and Y.-H. Liu, *Mar. Drugs*, 2024, 22, 63, DOI: [10.3390/md22020063](https://doi.org/10.3390/md22020063).
- 156 D. H. El-Kashef, D. D. Obidake, K. Schiedlauske, A. Deipenbrock, S. Scharf, H. Wang, D. Naumann, D. Friedrich, S. Miljanovic, T. Haj Hassani Sohi, C. Janiak, K. Pfeffer and N. Teusch, *Mar. Drugs*, 2024, 22, 5, DOI: [10.3390/md22010005](https://doi.org/10.3390/md22010005).
- 157 H. R. A. Cardona, B. C. da Silva, F. O. de Lima, F. H. A. Leite, B. C. de Souza, H. N. Brandão, J. M. David, C. Q. Alves and A. Kijjoa, *Mar. Drugs*, 2024, 22, 97, DOI: [10.3390/md22030097](https://doi.org/10.3390/md22030097).
- 158 E. Doro-Goldsmith, Q. Song, X.-L. Li, X.-M. Li, X.-Y. Hu, H.-L. Li, H.-R. Liu, B.-G. Wang and H. Sun, *J. Nat. Prod.*, 2024, 87, 381–387, DOI: [10.1021/acs.jnatprod.3c01157](https://doi.org/10.1021/acs.jnatprod.3c01157).
- 159 Q. Li, S. Li, S. Li, X. Hao, A. Wang, S. Si, Y. Xu, J. Shu and M. Gan, *J. Nat. Prod.*, 2024, 87, 365–370, DOI: [10.1021/acs.jnatprod.3c01115](https://doi.org/10.1021/acs.jnatprod.3c01115).
- 160 S. Kankanamge, P. V. Bernhardt, Z. G. Khalil and R. J. Capon, *Mar. Drugs*, 2024, 22, 243, DOI: [10.3390/md22060243](https://doi.org/10.3390/md22060243).
- 161 Y. He, T. Yang, J. Li, K. Li, C. Zhuang, M. Zhang, R. Li, Y. Zhao, Q. Song, M. Jiang, S. Mao, X. Song, Y. Guo, X. Li, F. Tan, S. Jitkaew, W. Zhang and Z. Cai, *Br. J. Pharmacol.*, 2024, 181, 2443–2458, DOI: [10.1111/bph.16364](https://doi.org/10.1111/bph.16364).
- 162 Z.-B. Zou, Y. Li, Y. Wang, C.-L. Xie, Z.-Q. Li, S.-S. Nie, Y. Li, S.-Y. Fang, T.-H. Zhong, L.-S. Li and X.-W. Yang, *Org. Lett.*, 2024, 26, 5695–5699, DOI: [10.1021/acs.orglett.4c01745](https://doi.org/10.1021/acs.orglett.4c01745).
- 163 S. Niu, S. Huang, J. Wang, J. He, M. Chen, G. Zhang and B. Hong, *Nat. Prod. Res.*, 2024, 38, 1362–1368, DOI: [10.1080/14786419.2022.2144299](https://doi.org/10.1080/14786419.2022.2144299).
- 164 Y.-H. Che, W.-P. Ding, Z.-H. Xiao, J.-M. Wu, H. Yin, F.-Z. Wang and S. Zhang, *Mar. Drugs*, 2024, 22, 117, DOI: [10.3390/md22030117](https://doi.org/10.3390/md22030117).
- 165 X. Zhang, Y. Dong, X. Liu, R. Wang, J. Lu and F. Song, *Nat. Prod. Res.*, 2024, 38, 2792–2799, DOI: [10.1080/14786419.2023.2236764](https://doi.org/10.1080/14786419.2023.2236764).
- 166 X. Shi, X. Li, X. He, D. Zhang, C. Quan, Z. Xiu and Y. Dong, *Molecules*, 2024, 29, 670, DOI: [10.3390/molecules29030670](https://doi.org/10.3390/molecules29030670).
- 167 R. Ge, X. Guo, H. Jia, J. Zhang, A. Fan, D. Liu, J. Huang and W. Lin, *J. Org. Chem.*, 2024, 89, 7692–7704, DOI: [10.1021/acs.joc.4c00367](https://doi.org/10.1021/acs.joc.4c00367).
- 168 Y. Chen, S.-P. Wang, L.-C. Xu, C. Liang, G.-D. Liu, X. Ji, W.-H. Luo, S. Liu, Z.-X. Zhang and G.-Y. Cao, *Fitoterapia*, 2024, 172, 105763, DOI: [10.1016/j.fitote.2023.105763](https://doi.org/10.1016/j.fitote.2023.105763).
- 169 J. Zhang, L. Gao, H. Lin, Y. Liang, M. You, L. Ding, F. Feng, B. Yang and Y. Liu, *J. Agric. Food Chem.*, 2024, 72, 12596–12606, DOI: [10.1021/acs.jafc.4c02769](https://doi.org/10.1021/acs.jafc.4c02769).
- 170 Z. Zhang, Y. Li, H. Wang, W. Xu, C. Wang, H. Ma, F. Zhong, J. Ou, Z. Luo, H.-B. Luo and Z. Cheng, *J. Nat. Prod.*, 2024, 87, 1563–1573, DOI: [10.1021/acs.jnatprod.4c00154](https://doi.org/10.1021/acs.jnatprod.4c00154).
- 171 G.-Y. Zhang, B.-J. Liu, H.-L. Pan, H.-J. Li, Z.-F. Huang, T. Mahmud, W.-Z. Ma and W.-J. Lan, *Fitoterapia*, 2024, 175, 105967, DOI: [10.1016/j.fitote.2024.105967](https://doi.org/10.1016/j.fitote.2024.105967).
- 172 H. Fan, X.-H. Shao, P. Wu, A.-L. Hao, Z.-W. Luo, M.-D. Zhang, J. Xie, B. Peng and C.-X. Zhang, *J. Agric. Food Chem.*, 2024, 72, 19869–19882, DOI: [10.1021/acs.jafc.4c04728](https://doi.org/10.1021/acs.jafc.4c04728).
- 173 H. Cui, Y. Tang, C. Yang, H. Deng, L. Chen, X. Fan, L. Zhu, Y. Liu, Z. Zhao and T. Su, *Phytochemistry*, 2024, 219, 113983, DOI: [10.1016/j.phytochem.2024.113983](https://doi.org/10.1016/j.phytochem.2024.113983).
- 174 Y. Zhou, X. Chen, Y. Tang, Y. Liu, Z. Zhao and H. Cui, *Nat. Prod. Res.*, 2024, 38, 1334–1340, DOI: [10.1080/14786419.2022.2140338](https://doi.org/10.1080/14786419.2022.2140338).
- 175 Y.-Y. Zheng, J.-Q. Mao, Y. Liu, N. Han, L. Lv, Y.-H. Zhang, M. Chen, Z.-Q. Liu, C.-L. Shao, G.-S. Yao and C.-Y. Wang, *Bioorg. Chem.*, 2024, 153, 107832, DOI: [10.1016/j.bioorg.2024.107832](https://doi.org/10.1016/j.bioorg.2024.107832).
- 176 Y.-P. He, Z.-K. Zhang, Z.-J. Li, P.-P. Wu, J.-S. Hu, H. Fan and C.-X. Zhang, *Fitoterapia*, 2024, 177, 106136, DOI: [10.1016/j.fitote.2024.106136](https://doi.org/10.1016/j.fitote.2024.106136).
- 177 H.-M. Wen, Y.-W. Zhang, F.-J. Feng, G.-B. Huang, Y.-H. Lv, Z.-Y. Zhang and L.-J. Ding, *J. Asian Nat. Prod. Res.*, 2024, 26, 548–554, DOI: [10.1080/10286020.2023.2259317](https://doi.org/10.1080/10286020.2023.2259317).
- 178 N. T. Ngoc, T. H. Quang, T. T. H. Hanh, N. X. Cuong, V. T. Quyen, N. T. T. Ngan, D. V. Ha, N. H. Nam and C. V. Minh, *Phytochem. Lett.*, 2024, 61, 29–34, DOI: [10.1016/j.phytol.2024.03.010](https://doi.org/10.1016/j.phytol.2024.03.010).
- 179 K. Anjum, X. Huang, L. Zhou, T. Zhu, Q. Che, G. Zhang and D. Li, *Nat. Prod. Res.*, 2024, 38, 3231–3236, DOI: [10.1080/14786419.2023.2227754](https://doi.org/10.1080/14786419.2023.2227754).
- 180 C. Sun, Y. Ha, X. Liu, N. Wang, X.-Y. Lian and Z. Zhang, *Molecules*, 2024, 29, 459, DOI: [10.3390/molecules29020459](https://doi.org/10.3390/molecules29020459).
- 181 Y. Ha, Y. Zhou, M. Ma, N. Wang, P. Wang and Z. Zhang, *Phytochemistry*, 2024, 224, 114164, DOI: [10.1016/j.phytochem.2024.114164](https://doi.org/10.1016/j.phytochem.2024.114164).



- 182 Y.-H. Chen, Q. Zhu, J. Li, R. Yang, J. Zhang, M. You, L. Luo and B. Yang, *Metabolites*, 2024, **14**, 99, DOI: [10.3390/metabo14020099](https://doi.org/10.3390/metabo14020099).
- 183 A. Abulaizi, R. Wang, Z. Xiong, S. Zhang, Y. Li, H. Ge and Z. Guo, *Mar. Drugs*, 2024, **22**, 270, DOI: [10.3390/md22060270](https://doi.org/10.3390/md22060270).
- 184 C. Li, X. Lin, S. Wang, D. Guan, X. Wang, B. Yang, X. Zhou, J. Li, B. Xiong, Y. Liu and Y. Sun, *Chem. Biodiversity*, 2024, **21**, e202301706, DOI: [10.1002/cbdv.202301706](https://doi.org/10.1002/cbdv.202301706).
- 185 X. Xu, J. Han, X. Zhang, W. Xu, J. Yang and F. Song, *Nat. Prod. Res.*, 2024, **38**, 1369–1374, DOI: [10.1080/14786419.2022.2144300](https://doi.org/10.1080/14786419.2022.2144300).
- 186 P. Lu, Y. Shi, J. Zhang, K. Hong, Y. Xue and L. Liu, *Int. J. Biol. Macromol.*, 2024, **257**, 128808, DOI: [10.1016/j.ijbiomac.2023.128808](https://doi.org/10.1016/j.ijbiomac.2023.128808).
- 187 D. Tian, M. Chen, W. Ding, J. Li, S. Wu, K. Hong and J. Tang, *Nat. Prod. J.*, 2024, **14**, e031023221602, DOI: [10.2174/0122103155274328230926160728](https://doi.org/10.2174/0122103155274328230926160728).
- 188 J. Xue, X. Guo, G. Xu, X. Chen, L. Jiao and X. Tang, *J. Agric. Food Chem.*, 2024, **72**, 2989–2996, DOI: [10.1021/acs.jafc.3c07826](https://doi.org/10.1021/acs.jafc.3c07826).
- 189 H.-Y. Dong, Y. Wang, X.-Y. Zhang, M. Zhang, L.-H. Yang, Z.-B. Zou, Y. Li, M.-M. Xie, X.-W. Yang and B. Wang, *Chem. Biodiversity*, 2024, **21**, e202400507, DOI: [10.1002/cbdv.202400507](https://doi.org/10.1002/cbdv.202400507).
- 190 G.-J. Ran, P. Wang, H.-K. Wang, T.-T. Wei, Y.-Y. Liu, H.-X. Jin, T.-T. Wang and S. He, *Chem. Biodiversity*, 2024, **21**, e202401689, DOI: [10.1002/cbdv.202401689](https://doi.org/10.1002/cbdv.202401689).
- 191 S.-T. Fang, F.-P. Miao, X.-L. Yin and N.-Y. Ji, *Nat. Prod. Res.*, 2024, **38**, 1510–1516, DOI: [10.1080/14786419.2022.2154345](https://doi.org/10.1080/14786419.2022.2154345).
- 192 X.-D. Li, X.-M. Li, B.-G. Wang and X. Li, *Org. Biomol. Chem.*, 2024, **22**, 3979–3985, DOI: [10.1039/D4OB00449C](https://doi.org/10.1039/D4OB00449C).
- 193 X.-X. Miao, L.-L. Hong, H.-Y. Liu, R.-Y. Shang, W.-H. Jiao, S.-H. Xu and H.-W. Lin, *Chem. Biodiversity*, 2024, **21**, e202400832, DOI: [10.1002/cbdv.202400832](https://doi.org/10.1002/cbdv.202400832).
- 194 X. Miao, L. Hong, Z. Ju, H. Liu, R. Shang, P. Li, K. Liu, B. Cheng, W. Jiao, S. Xu and H. Lin, *ACS Omega*, 2024, **9**, 22450–22458, DOI: [10.1021/acsomega.4c02488](https://doi.org/10.1021/acsomega.4c02488).
- 195 Q. Gao, Y. Shi, H. Wen, D. Zhang, X. Yan, S. He and L. Ding, *J. Mol. Struct.*, 2024, **1296**, 136723, DOI: [10.1016/j.molstruc.2023.136723](https://doi.org/10.1016/j.molstruc.2023.136723).
- 196 C.-P. Li, Y.-P. Song, X.-R. Liang and N.-Y. Ji, *Nat. Prod. Res.*, 2024, **38**, 3051–3056, DOI: [10.1080/14786419.2023.2214291](https://doi.org/10.1080/14786419.2023.2214291).
- 197 X. Li, Y. Chen, Z. Liu, S. Li, H. Liu, Y. Wang, W. Zhang and H. Yan, *Nat. Prod. Res.*, 2024, **38**, 594–600, DOI: [10.1080/14786419.2023.2187794](https://doi.org/10.1080/14786419.2023.2187794).
- 198 W. Zhang, Y. Yuan, X. Pang, Y. Liu, X. Zhang, Q. Liu and J. Wang, *Chem. Biodiversity*, 2024, **21**, e202401751, DOI: [10.1002/cbdv.202401751](https://doi.org/10.1002/cbdv.202401751).
- 199 L. Dai, Q. Xie, J. Guo, Q. Ma, L. Yang, J. Yuan, H. Dai, Z. Yu and Y. Zhao, *J. Oceanol. Limnol.*, 2024, **42**, 905–914, DOI: [10.1007/s00343-023-3084-7](https://doi.org/10.1007/s00343-023-3084-7).
- 200 H. Kamauchi, M. Tanaka, M. Suzuki, M. Furukawa, A. Ikeda, C. Sasho, Y. Kiba, M. Kitamura, K. Takao and Y. Sugita, *Chem. Pharm. Bull.*, 2024, **72**, 98–101, DOI: [10.1248/cpb.c23-00723](https://doi.org/10.1248/cpb.c23-00723).
- 201 J. Wang, H. Lu, W. Fang, M. Lin, Y. Feng, X. Qi, C. Gao, Y. Liu, X. Wang and X. Luo, *RSC Adv.*, 2024, **14**, 38697–38705, DOI: [10.1039/D4RA06292B](https://doi.org/10.1039/D4RA06292B).
- 202 S. Xu, Z. Li, X. Xin and F. An, *Mar. Drugs*, 2024, **22**, 227, DOI: [10.3390/md22050227](https://doi.org/10.3390/md22050227).
- 203 J. R. Virués-Segovia, C. Pinedo, D. Zorrilla, J. Sánchez-Márquez, P. Sánchez, M. C. Ramos, M. de la Cruz, J. Aleu and R. Durán-Patrón, *Front. Mar. Sci.*, 2024, **11**, 1386175, DOI: [10.3389/fmars.2024.1386175](https://doi.org/10.3389/fmars.2024.1386175).
- 204 J. He, Q. Zou, H. Deng, S. He, D. Yan, K. Pan, Y. Zhou, Z. Zhao, H. Cui and Y. Liu, *Fitoterapia*, 2024, **173**, 105804, DOI: [10.1016/j.fitote.2023.105804](https://doi.org/10.1016/j.fitote.2023.105804).
- 205 Z.-P. Jiang, R. Su, M.-T. Chen, J.-Y. Li, H.-Y. Chen, L. Yang, F.-F. Liu, J. Liu, C.-J. Xu, W.-S. Li, Y. Rao and L. Huang, *Phytochemistry*, 2024, **223**, 114121, DOI: [10.1016/j.phytochem.2024.114121](https://doi.org/10.1016/j.phytochem.2024.114121).
- 206 X.-D. Chen, X. Li, X.-M. Li, S.-Q. Yang and B.-G. Wang, *Mar. Drugs*, 2024, **22**, 574, DOI: [10.3390/md22120574](https://doi.org/10.3390/md22120574).
- 207 G. N. Pham, B. Josselin, A. Cousseau, B. Baratte, M. Dayras, C. Le Meur, S. Debaets, A. Weill, T. Robert, G. Burgaud, I. Probert, F. M. Abdoul-Latif, L. Boyer, S. Bach and M. Mehiri, *Mar. Drugs*, 2024, **22**, 444, DOI: [10.3390/md22100444](https://doi.org/10.3390/md22100444).
- 208 Z. Zhao, F. Cen, T. Mo, W. Li and G. Y. Chen, *Chem. Biodiversity*, 2024, **21**, e202400118, DOI: [10.1002/cbdv.202400118](https://doi.org/10.1002/cbdv.202400118).
- 209 Y. Zhang, W. Liu, G. Li, C. Wu, J. Yan, D. Feng, S. Yuan, R. Zhang, H. Lou and X. Peng, *Nat. Prod. Res.*, 2024, **38**, 2957–2963, DOI: [10.1080/14786419.2023.2201885](https://doi.org/10.1080/14786419.2023.2201885).
- 210 H. Li, Y. Chen, B. Tang, Z. Liu, B. Peng, J. Li, H. Gao, S. Wang and Z. Li, *J. Nat. Prod.*, 2024, **87**, 2695–2708, DOI: [10.1021/acs.jnatprod.4c00740](https://doi.org/10.1021/acs.jnatprod.4c00740).
- 211 T. Wang, J. Tian, J. Huang, Y. Yuan, C. B. Naman, S. Wu, H. Wang, W. Lin, Z. Tong, L. Ding, W. Wang and S. He, *J. Nat. Prod.*, 2024, **87**, 1203–1208, DOI: [10.1021/acs.jnatprod.3c01078](https://doi.org/10.1021/acs.jnatprod.3c01078).
- 212 M. A. Lee, J. S. Kang, J.-W. Yang, H.-S. Lee, C.-S. Heo, S. J. Park and H. J. Shin, *Mar. Drugs*, 2024, **22**, 87, DOI: [10.3390/md22020087](https://doi.org/10.3390/md22020087).
- 213 F.-H. Yao, X. Liang, W.-B. Shen, X.-H. Lu, G.-C. Li and S.-H. Qi, *J. Nat. Prod.*, 2024, **87**, 810–819, DOI: [10.1021/acs.jnatprod.3c00984](https://doi.org/10.1021/acs.jnatprod.3c00984).
- 214 T. Feng, R. Wu, Y. Wang, P. Wang, L. Zhou, C. Wang and F. Kong, *Mar. Drugs*, 2024, **22**, 241, DOI: [10.3390/md22060241](https://doi.org/10.3390/md22060241).
- 215 X. Chen, S. Chen, H. Guo, X. Lu, H. Shen, L. Liu, L. Wang, B. Chen, Y. Zhang and Y. Liu, *Mar. Drugs*, 2024, **22**, 214, DOI: [10.3390/md22050214](https://doi.org/10.3390/md22050214).
- 216 M. T. Ngo, J. W. Han, M. V. Nguyen, Y. Choi, B. Kim, E. S. Gho, H. Kim and G. J. Choi, *J. Agric. Food Chem.*, 2024, **72**, 16359–16367, DOI: [10.1021/acs.jafc.4c04706](https://doi.org/10.1021/acs.jafc.4c04706).
- 217 X. Li, W. Shen, G. Li, Y. Song, X. Lu, N.-K. Wong and Y. Yan, *Nat. Prod. Res.*, 2024, **38**, 3917–3923, DOI: [10.1080/14786419.2023.2269462](https://doi.org/10.1080/14786419.2023.2269462).
- 218 J. Yang, X. Zhang, W. Xu, X. Xu and F. Song, *Nat. Prod. Res.*, 2024, **38**, 2231–2236, DOI: [10.1080/14786419.2023.2169917](https://doi.org/10.1080/14786419.2023.2169917).



- 219 Y.-H. Zhang, H.-F. Du, Y.-F. Liu, F. Cao, D.-Q. Luo and C.-Y. Wang, *Appl. Microbiol. Biotechnol.*, 2024, **108**, 194, DOI: [10.1007/s00253-024-13026-4](https://doi.org/10.1007/s00253-024-13026-4).
- 220 J. Yang, Y. Zong, C. Wang, K. Li, Y. Zhang and P. Li, *Mar. Drugs*, 2024, **22**, 517, DOI: [10.3390/md22110517](https://doi.org/10.3390/md22110517).
- 221 N. M. Anh, L. T. H. Minh, N. T. Linh, P. T. Dao, D. T. Quynh, D. T. M. Huong, P. V. Cuong, V. T. T. Huyen and T. T. H. Dat, *Biosci., Biotechnol., Biochem.*, 2024, **88**, 1254–1260, DOI: [10.1093/bbb/zbae113](https://doi.org/10.1093/bbb/zbae113).
- 222 J. Wei, X. Chen and B. Wu, *J. Org. Chem.*, 2024, **89**, 15264–15270, DOI: [10.1021/acs.joc.4c02067](https://doi.org/10.1021/acs.joc.4c02067).
- 223 C.-S. Zhu, X.-M. Li, S.-Q. Yang, Y.-W. Liu, B.-G. Wang and X. Li, *Chem. Biodiversity*, 2024, **21**, e202400584, DOI: [10.1002/cbdv.202400584](https://doi.org/10.1002/cbdv.202400584).
- 224 G. Yu, L. Zhou, G. Wu and D. Li, *J. Ocean Univ. China*, 2024, **23**, 216–220, DOI: [10.1007/s11802-023-5676-4](https://doi.org/10.1007/s11802-023-5676-4).
- 225 B.-C. Hao, Y.-Y. Zheng, Z.-H. Li, C.-J. Zheng, C.-Y. Wang and M. Chen, *Nat. Prod. Res.*, 2024, **38**, 2252–2257, DOI: [10.1080/14786419.2023.2171401](https://doi.org/10.1080/14786419.2023.2171401).
- 226 W. Song, L. Ji, Y. Zhang and L. Cao, *Front. Microbiol.*, 2024, **15**, 1400803, DOI: [10.3389/fmicb.2024.1400803](https://doi.org/10.3389/fmicb.2024.1400803).
- 227 F. Lu, W. Song, H. Li and L. Cao, *Rec. Nat. Prod.*, 2024, **18**, 699–704, DOI: [10.25135/rnp.486.2410.3339](https://doi.org/10.25135/rnp.486.2410.3339).
- 228 Z. Chen, Y. Lv, Z. Luo, B. Hong and S. Niu, *Fitoterapia*, 2024, **179**, 106248, DOI: [10.1016/j.fitote.2024.106248](https://doi.org/10.1016/j.fitote.2024.106248).
- 229 X. Xu, Y. Dong, J. Yang, L. Wang, L. Ma, F. Song and X. Ma, *J. Fungi*, 2024, **10**, 424, DOI: [10.3390/jof10060424](https://doi.org/10.3390/jof10060424).
- 230 J. Bao, Y.-F. Zhao, X.-X. Wang, K. Zhu, R. Ao, H. Liu, X.-X. Li, J.-S. Zhang and H. Zhang, *Bioorg. Chem.*, 2024, **148**, 107434, DOI: [10.1016/j.bioorg.2024.107434](https://doi.org/10.1016/j.bioorg.2024.107434).
- 231 Y. Yang, X. Han, C. Jiang, Y. Chen, X. Tang and G. Li, *Bioorg. Chem.*, 2024, **153**, 107897, DOI: [10.1016/j.bioorg.2024.107897](https://doi.org/10.1016/j.bioorg.2024.107897).
- 232 F.-Z. Zhang, X.-M. Li, L.-H. Meng and B.-G. Wang, *J. Antibiot.*, 2024, **77**, 13–20, DOI: [10.1038/s41429-023-00666-3](https://doi.org/10.1038/s41429-023-00666-3).
- 233 G. V. Borkunov, E. V. Leshchenko, D. V. Berdyshev, R. S. Popov, E. A. Chingizova, N. P. Shlyk, A. V. Gerasimenko, N. N. Kirichuk, Y. V. Khudyakova, V. E. Chausova, A. S. Antonov, A. I. Kalinovsky, A. R. Chingizov, E. A. Yurchenko, M. P. Isaeva and A. N. Yurchenko, *Nat. Prod. Bioprospect.*, 2024, **14**, 32, DOI: [10.1007/s13659-024-00449-9](https://doi.org/10.1007/s13659-024-00449-9).
- 234 E. V. Leshchenko, E. A. Chingizova, A. S. Antonov, N. P. Shlyk, G. V. Borkunov, D. V. Berdyshev, V. E. Chausova, N. N. Kirichuk, Y. V. Khudyakova, A. R. Chingizov, A. I. Kalinovsky, R. S. Popov, N. Y. Kim, K. A. Chadova, E. A. Yurchenko, M. P. Isaeva and A. N. Yurchenko, *Mar. Drugs*, 2024, **22**, 317, DOI: [10.3390/md22070317](https://doi.org/10.3390/md22070317).
- 235 Y.-Q. Tang, X. Liang, Q.-H. Zou, H. Cui, L.-X. Luo and S.-H. Qi, *J. Nat. Prod.*, 2024, **87**, 1209–1216, DOI: [10.1021/acs.jnatprod.3c01121](https://doi.org/10.1021/acs.jnatprod.3c01121).
- 236 F.-P. Miao, S.-T. Fang, Z.-Z. Shi, Y.-P. Song and N.-Y. Ji, *Fitoterapia*, 2024, **177**, 106110, DOI: [10.1016/j.fitote.2024.106110](https://doi.org/10.1016/j.fitote.2024.106110).
- 237 L.-T. Dai, L. Yang, J.-C. Guo, Q.-Y. Ma, Q.-Y. Xie, L. Jiang, Z.-F. Yu, H.-F. Dai and Y.-X. Zhao, *Bioorg. Chem.*, 2024, **145**, 107205, DOI: [10.1016/j.bioorg.2024.107205](https://doi.org/10.1016/j.bioorg.2024.107205).
- 238 L. Yang, M.-M. Quan, Q.-Y. Ma, Q.-Y. Xie, C.-J. Gai, H.-F. Dai, Y.-E. Hao and Y.-X. Zhao, *Chem. Biodiversity*, 2024, **21**, e202401871, DOI: [10.1002/cbdv.202401871](https://doi.org/10.1002/cbdv.202401871).
- 239 C. Wang, Y. Zhou, L. Yang, H. Hu, J. Chen, Y. Ying and H. Wang, *Nat. Prod. Res.*, 2024, **38**, 3605–3612, DOI: [10.1080/14786419.2023.2258441](https://doi.org/10.1080/14786419.2023.2258441).
- 240 W. Chen, J. Jiang, X. Pang, Y. Song, Z. Yang, J. Wang and Y. Liu, *Org. Biomol. Chem.*, 2024, **22**, 4521–4527, DOI: [10.1039/D4OB00586D](https://doi.org/10.1039/D4OB00586D).
- 241 Y.-Y. Sun, B. Hu, H.-B. Yu, J. Zhou, X.-C. Meng, Z. Ning, J.-F. Ding, M.-H. Cui and X.-Y. Liu, *Mar. Drugs*, 2024, **22**, 236, DOI: [10.3390/md22060236](https://doi.org/10.3390/md22060236).
- 242 C. Wang, C. Wang, Y. Liu, Y. Yue, X. Lu, H. Wang, Y. Ying and J. Chen, *Appl. Environ. Microbiol.*, 2024, **90**, e01603–e01624, DOI: [10.1128/aem.01603-24](https://doi.org/10.1128/aem.01603-24).
- 243 C. Wang, Y. Fan, C. Wang, J. Tang, Y. Qiu, K. Xu, Y. Ding, Y. Liu, Y. Ying and H. Wang, *Mar. Drugs*, 2024, **22**, 296, DOI: [10.3390/md22070296](https://doi.org/10.3390/md22070296).
- 244 N. J. Morehouse, K. M. Graham, S. L. Cox, J. A. Johnson and C. A. Gray, *Can. J. Chem.*, 2024, **102**, 86–90, DOI: [10.1139/cjc-2023-0100](https://doi.org/10.1139/cjc-2023-0100).
- 245 H. Fan, X.-H. Shao, Z.-K. Zhang, C. Ni, C. Feng, X. Wei, J.-Q. Zhu, X.-H. Li and C.-X. Zhang, *J. Nat. Prod.*, 2024, **87**, 705–712, DOI: [10.1021/acs.jnatprod.3c00765](https://doi.org/10.1021/acs.jnatprod.3c00765).
- 246 H. Li, J. Long, X. Wang, J. She, Y. Liu, Y. Li and B. Yang, *Nat. Prod. Res.*, 2024, **38**, 2996–3003, DOI: [10.1080/14786419.2023.2207133](https://doi.org/10.1080/14786419.2023.2207133).
- 247 J.-W. Wang, Y.-Y. Liu, G.-Y. Chen, C.-J. Zheng and X.-H. Nong, *Fitoterapia*, 2024, **175**, 105906, DOI: [10.1016/j.fitote.2024.105906](https://doi.org/10.1016/j.fitote.2024.105906).
- 248 Y. Chen, J. Cai, Z. Xia, C. Chen, Y. Liu, L. Jayasinghe, X. Wang and X. Zhou, *Mar. Drugs*, 2024, **22**, 384, DOI: [10.3390/md22090384](https://doi.org/10.3390/md22090384).
- 249 L. Huang, C. Chen, J. Cai, Y. Chen, Y. Zhu, B. Yang, X. Zhou, Y. Liu and H. Tao, *Mar. Drugs*, 2024, **22**, 393, DOI: [10.3390/md22090393](https://doi.org/10.3390/md22090393).
- 250 J. Cai, M. Li, C. Chen, B. Yang, C. Gao, Y. Liu, X. Luo, Y. Tan and X. Zhou, *J. Nat. Prod.*, 2024, **87**, 1401–1406, DOI: [10.1021/acs.jnatprod.4c00116](https://doi.org/10.1021/acs.jnatprod.4c00116).
- 251 J. Cai, L. Gao, Y. Wang, Y. Zheng, X. Lin, P. Zhou, C. Chen, K. Liu, L. Tang, Y. Liu, Y. Tan, M. Jin and X. Zhou, *Eur. J. Med. Chem.*, 2024, **265**, 116068, DOI: [10.1016/j.ejmech.2023.116068](https://doi.org/10.1016/j.ejmech.2023.116068).
- 252 C. Chen, L. Xiao, X. Luo, J. Cai, L. Huang, H. Tao, X. Zhou, Y. Tan and Y. Liu, *J. Med. Chem.*, 2024, **67**, 2602–2618, DOI: [10.1021/acs.jmedchem.3c01748](https://doi.org/10.1021/acs.jmedchem.3c01748).
- 253 Y.-R. Wang, Y.-L. Dong, X.-M. Li, X.-S. Shi, H.-L. Li, L.-H. Meng, R. Xu and B.-G. Wang, *Phytochemistry*, 2024, **220**, 114000, DOI: [10.1016/j.phytochem.2024.114000](https://doi.org/10.1016/j.phytochem.2024.114000).
- 254 B. Yang, S. Shao, M. Nie, Q. Tie, X. Pang, X. Lin, X. Zhou, Y. Liu, X. Wang and Y. Li, *Mar. Drugs*, 2024, **22**, 218, DOI: [10.3390/md22050218](https://doi.org/10.3390/md22050218).
- 255 M. R. de Amorim, S. M. Schoellhorn, C. d. S. Barbosa, G. R. Mendes, K. de L. Macedo, A. G. Ferreira,



- T. Venâncio, R. V. C. Guido, A. N. L. Batista, J. M. Batista Jr, E. Skellam and R. G. S. Berlinck, *J. Nat. Prod.*, 2024, **87**, 2204–2215, DOI: [10.1021/acs.jnatprod.4c00516](https://doi.org/10.1021/acs.jnatprod.4c00516).
- 256 T. Wang, Y. Feng, J. Huang, S. Wu, K. Hu, J. Wu, C. B. Naman, H. Wang, W. Lin and S. He, *J. Nat. Prod.*, 2024, **87**, 160–165, DOI: [10.1021/acs.jnatprod.3c00892](https://doi.org/10.1021/acs.jnatprod.3c00892).
- 257 L. Jiang, B. Teng, M. Zhang, S. Chen, D. Zhang, L. Zhai, J. Lin and H. Lei, *Mar. Drugs*, 2024, **22**, 15, DOI: [10.3390/md22010015](https://doi.org/10.3390/md22010015).
- 258 Z.-L. Tan, Y.-C. Chen, J.-P. Zhang, H.-X. Liu, W.-M. Zhang and H.-J. Yan, *J. Asian Nat. Prod. Res.*, 2024, **26**, 534–540, DOI: [10.1080/10286020.2023.2249832](https://doi.org/10.1080/10286020.2023.2249832).
- 259 X. Li, Y. Chen, S. Li, W. Zhang, H. Yan, H. Liu and W. Zhang, *Fitoterapia*, 2024, **172**, 105772, DOI: [10.1016/j.fitote.2023.105772](https://doi.org/10.1016/j.fitote.2023.105772).
- 260 J. Wu, J. Ye, J. Cen, Y. Chen and J. Xu, *Mar. Drugs*, 2024, **22**, 332, DOI: [10.3390/md22080332](https://doi.org/10.3390/md22080332).
- 261 J. Wu, D. Chen, Q. Li, T. Feng and J. Xu, *Mar. Drugs*, 2024, **22**, 102, DOI: [10.3390/md22030102](https://doi.org/10.3390/md22030102).
- 262 Z. Ying, X.-M. Li, S.-Q. Yang, H.-L. Li, X. Li, B.-G. Wang and L.-H. Meng, *J. Nat. Prod.*, 2024, **87**, 1347–1357, DOI: [10.1021/acs.jnatprod.3c01175](https://doi.org/10.1021/acs.jnatprod.3c01175).
- 263 C. Sun, Y. Jiang, C. Li, S. Sun, J. Lin, W. Wang, L. Zhou, L. Li, M. Shah, Q. Che, G. Zhang, D. Wang, T. Zhu and D. Li, *Angew. Chem., Int. Ed.*, 2024, **63**, e202405860, DOI: [10.1002/anie.202405860](https://doi.org/10.1002/anie.202405860).
- 264 L. Yang, Q. Zhang, D. Yu, W. Zhu and Y. Wang, *Chem. Biodiversity*, 2024, **21**, e202400578, DOI: [10.1002/cbdv.202400578](https://doi.org/10.1002/cbdv.202400578).
- 265 J.-W. Wang, D.-Y. Liu, H.-Z. Zhang, Z. Tan, C.-J. Zheng, G.-Y. Chen and X.-H. Nong, *Nat. Prod. Res.*, 2024, **38**, 1711–1718, DOI: [10.1080/14786419.2023.2218008](https://doi.org/10.1080/14786419.2023.2218008).
- 266 X. Liu, Y. Lv, J. He, B. Hong, Z. Shao, M. Yu and S. Niu, *Chem. Biodiversity*, 2024, **21**, e202400805, DOI: [10.1002/cbdv.202400805](https://doi.org/10.1002/cbdv.202400805).
- 267 Y. Xie, Y. Wang, K. Zhang, Q.-M. Liu, Q.-X. Yan, Z.-B. Zou, Y.-Q. Xu, G.-M. Liu, X.-F. Huang and X.-W. Yang, *Eur. J. Org. Chem.*, 2024, **27**, e202301250, DOI: [10.1002/ejoc.202301250](https://doi.org/10.1002/ejoc.202301250).
- 268 H. Azami, Y. Watanabe, K. Sakai, H. Nakahara, H. Kojima, T. Tokiwa, K. Nonaka, Y. Noguchi, Y. Nagano, T. Hirose, T. Sunazuka, H. Matsui, N. Arima, K. Abe, H. Hanaki and M. Iwatsuki, *J. Antibiot.*, 2024, **77**, 156–162, DOI: [10.1038/s41429-023-00696-x](https://doi.org/10.1038/s41429-023-00696-x).
- 269 Z. Xiao, J. Cai, T. Chen, Y. Wang, Y. Chen, Y. Zhu, C. Chen, B. Yang, X. Zhou and H. Tao, *Mar. Drugs*, 2024, **22**, 103, DOI: [10.3390/md22030103](https://doi.org/10.3390/md22030103).
- 270 X. Pang, B. Yang, X. Zhou, J. Wang, J. Yang and Y. Liu, *Chem. Biodiversity*, 2024, **21**, e202302069, DOI: [10.1002/cbdv.202302069](https://doi.org/10.1002/cbdv.202302069).
- 271 H. Kwon, B.-R. Jin, S. Yoo, H.-J. Kim, B. Y. Hwang, Y. Guo, J. H. Yim, I.-C. Kim, S. H. Shim, H. J. An and D. Lee, *Bioorg. Chem.*, 2024, **143**, 107070, DOI: [10.1016/j.bioorg.2023.107070](https://doi.org/10.1016/j.bioorg.2023.107070).
- 272 X. Liang, J.-F. Yang, Z.-H. Huang, X. Ma, Y. Yan and S.-H. Qi, *J. Agric. Food Chem.*, 2024, **72**, 6402–6413, DOI: [10.1021/acs.jafc.4c00493](https://doi.org/10.1021/acs.jafc.4c00493).
- 273 Y.-C. He, M.-Q. Wang, Q.-Q. Tie, X.-W. Huang, Y.-H. Liu, Y.-Q. Li and B. Yang, *J. Antibiot.*, 2024, **78**, 64–67, DOI: [10.1038/s41429-024-00780-w](https://doi.org/10.1038/s41429-024-00780-w).
- 274 Z. Wu, W. Wang, J. Li, C. Ma, L. Chen, Q. Che, G. Zhang, T. Zhu and D. Li, *J. Nat. Prod.*, 2024, **87**, 1222–1229, DOI: [10.1021/acs.jnatprod.3c01202](https://doi.org/10.1021/acs.jnatprod.3c01202).
- 275 Z. Huang, D. Liu, S. Chen, J. Ren, C. Gao, Z. Li, A. Fan and W. Lin, *Mar. Drugs*, 2024, **22**, 78, DOI: [10.3390/md22020078](https://doi.org/10.3390/md22020078).
- 276 S. Lin, Z. Chai, H. Zeng, B. Yang, J. Chi, Y. Zhang and Z. Hu, *Phytochemistry*, 2024, **225**, 114199, DOI: [10.1016/j.phytochem.2024.114199](https://doi.org/10.1016/j.phytochem.2024.114199).
- 277 X. Ma, M. Wu, Z. Chen, F. Cao, T. Zhong, Z. Luo, Z. Shao, Y. Zhang, L. Chen and Z. Zhang, *Molecules*, 2024, **29**, 1685, DOI: [10.3390/molecules29071685](https://doi.org/10.3390/molecules29071685).
- 278 T. Berger, J. Alenfelder, S. Steinmüller, D. Heimann, N. Gohain, D. Petras, M. Wang, R. Berger, E. Kostenis and R. Reher, *J. Nat. Prod.*, 2024, **87**, 692–704, DOI: [10.1021/acs.jnatprod.3c00750](https://doi.org/10.1021/acs.jnatprod.3c00750).
- 279 M. Zheng, Q. Li, H. Liao, Y. Li, C. Zhou, X. Zhao, C. Chen, W. Sun, Y. Zhang and H. Zhu, *J. Nat. Prod.*, 2024, **87**, 1921–1929, DOI: [10.1021/acs.jnatprod.4c00330](https://doi.org/10.1021/acs.jnatprod.4c00330).
- 280 J. Wu, W. Wang, Y. Yang, M. Shah, J. Peng, L. Zhou, G. Zhang, Q. Che, J. Li, T. Zhu and D. Li, *J. Nat. Prod.*, 2024, **87**, 1407–1415, DOI: [10.1021/acs.jnatprod.4c00132](https://doi.org/10.1021/acs.jnatprod.4c00132).
- 281 Z.-H. Peng, H. Jia, Y.-L. Luo, L.-J. Zhang, J.-T. Zhou, Y.-H. Xie, L.-J. Wang, J.-K. Qin, J. Li, G.-H. Zhang, R.-Y. Yang and W.-F. Xu, *Mar. Drugs*, 2024, **22**, 475, DOI: [10.3390/md22100475](https://doi.org/10.3390/md22100475).
- 282 K. Zhang, Y. Wang, T. Long, Y.-P. Xie, Z.-B. Zou, Y. Zhang, Y. Li, M.-M. Xie, G.-M. Liu and X.-W. Yang, *Biochem. Syst. Ecol.*, 2024, **114**, 104829, DOI: [10.1016/j.bse.2024.104829](https://doi.org/10.1016/j.bse.2024.104829).
- 283 W. Wang, J. Wang, F. Song, R. Jia, L. Wang, X. Xu and N. Yang, *Mar. Drugs*, 2024, **22**, 237, DOI: [10.3390/md22060237](https://doi.org/10.3390/md22060237).
- 284 X. Liang, Z.-H. Huang, W.-B. Shen, X.-H. Lu, X.-X. Zhang, X. Ma and S.-H. Qi, *Phytochemistry*, 2024, **223**, 114119, DOI: [10.1016/j.phytochem.2024.114119](https://doi.org/10.1016/j.phytochem.2024.114119).
- 285 Y. Wang, X.-M. Li, N. Song, B.-G. Wang, H.-L. Li and L.-H. Meng, *Bioorg. Chem.*, 2024, **147**, 107417, DOI: [10.1016/j.bioorg.2024.107417](https://doi.org/10.1016/j.bioorg.2024.107417).
- 286 Y. Cho, K.-H. Park, E. Kim, S. Kim, W. Wang, H. Choi and H. Kang, *J. Nat. Prod.*, 2024, **87**, 1230–1234, DOI: [10.1021/acs.jnatprod.3c01227](https://doi.org/10.1021/acs.jnatprod.3c01227).
- 287 L. Tang, J. Xia, Z. Chen, X. Wu, G. Li, Q. Lai, Z. Shao, W. Wang and X. Hong, *Mar. Drugs*, 2024, **22**, 274, DOI: [10.3390/md22060274](https://doi.org/10.3390/md22060274).
- 288 L. Tang, J. Xia, Z. Chen, F. Lin, Z. Shao, W. Wang and X. Hong, *Mar. Drugs*, 2024, **22**, 186, DOI: [10.3390/md22040186](https://doi.org/10.3390/md22040186).
- 289 X. Hong, L. Tang, Z. Chen, Q. Lai, B. Zhang, Y. Jiang, X. Wang, R. He, J. Lin, Z. Shao, S. Lin and W. Wang, *Nat. Prod. Res.*, 2024, **38**, 320–326, DOI: [10.1080/14786419.2022.2121830](https://doi.org/10.1080/14786419.2022.2121830).
- 290 T. Y. Kim, C. Lee, J. W. Lee and S. H. Shim, *Nat. Prod. Sci.*, 2024, **30**, 59–63, DOI: [10.20307/nps.2024.30.1.59](https://doi.org/10.20307/nps.2024.30.1.59).



- 291 Z. Wu, X.-M. Li, S.-Q. Yang, B.-G. Wang and X. Li, *Mar. Drugs*, 2024, **22**, 204, DOI: [10.3390/md22050204](https://doi.org/10.3390/md22050204).
- 292 H. Zeng, Y. Wang, C. Wang, Y. Huang, S. Lin, J. Wei, W. Sun, F. Cao, Y. Zhang and Z. Hu, *J. Nat. Prod.*, 2024, **87**, 2592–2603, DOI: [10.1021/acs.jnatprod.4c00709](https://doi.org/10.1021/acs.jnatprod.4c00709).
- 293 Y. Shi, X.-Q. Sun, J.-X. Zhang, R.-H. Zhang, K. Hong, Y.-X. Xue, H. Qiu and L. Liu, *J. Agric. Food Chem.*, 2024, **72**, 17431–17443, DOI: [10.1021/acs.jafc.4c03959](https://doi.org/10.1021/acs.jafc.4c03959).
- 294 T. Gu, J. Cai, D. Xie, J. She, Y. Liu, X. Zhou and L. Tang, *Mar. Drugs*, 2024, **22**, 403, DOI: [10.3390/md22090403](https://doi.org/10.3390/md22090403).
- 295 J. Cai, Q. Zhou, X. Qi, F. Zhang, J. Yang, C. Chen, K. Zhang, Z. Chen, H.-B. Luo, Y. Liu, Y.-Y. Huang and X. Zhou, *J. Nat. Prod.*, 2024, **87**, 1808–1816, DOI: [10.1021/acs.jnatprod.4c00422](https://doi.org/10.1021/acs.jnatprod.4c00422).
- 296 Z. Zeng, J. Cai, Y. Chen, X. Li, C. Chen, Y. Liu, L. Jayasinghe and X. Zhou, *Mar. Drugs*, 2024, **22**, 559, DOI: [10.3390/md22120559](https://doi.org/10.3390/md22120559).
- 297 C. Chen, X. Wang, W. Fang, J. Liang, J. Cai, D. Yang, X. Luo, C. Gao, X. Yi, Y. Liu and X. Zhou, *Chin. J. Nat. Med.*, 2024, **22**, 1047–1056, DOI: [10.1016/S1875-5364\(24\)60659-X](https://doi.org/10.1016/S1875-5364(24)60659-X).
- 298 Y. Jung, C. Kwon, T. Kim, J. W. Lee, M.-K. Shin and S. H. Shim, *Mar. Life Sci. & Technol.*, 2024, **6**, 84–92, DOI: [10.1007/s42995-023-00198-7](https://doi.org/10.1007/s42995-023-00198-7).
- 299 Y. Liu, L. Qi, M. Xu, W. Li, N. Liu, X. He and Y. Zhang, *Front. Microbiol.*, 2024, **15**, 1446283, DOI: [10.3389/fmicb.2024.1446283](https://doi.org/10.3389/fmicb.2024.1446283).
- 300 W. Yang, S. Tian, Y.-F. Du, X.-L. Zeng, J.-J. Liang, W.-J. Lan and H. Li, *J. Nat. Prod.*, 2024, **87**, 2746–2756, DOI: [10.1021/acs.jnatprod.4c00905](https://doi.org/10.1021/acs.jnatprod.4c00905).
- 301 X. Dong, L.-C. Jia, H.-Q. Chai, X.-X. Liu and L. Yang, *Rec. Nat. Prod.*, 2024, **18**, 352–356, DOI: [10.25135/rnp.456.2403.3175](https://doi.org/10.25135/rnp.456.2403.3175).
- 302 C. Zhou, Y. Ge, D. Lan, M. Zhao and B. Wu, *Fitoterapia*, 2024, **176**, 106039, DOI: [10.1016/j.fitote.2024.106039](https://doi.org/10.1016/j.fitote.2024.106039).
- 303 Y. Zhang, Y. Zhang, G. Li, K. Dong, J. Wang, S. Xiao, H. Lou and X. Peng, *Fitoterapia*, 2024, **175**, 105963, DOI: [10.1016/j.fitote.2024.105963](https://doi.org/10.1016/j.fitote.2024.105963).
- 304 C. Chen, L. Liu, S. Ye, J. Li, L. Wu, J. Li, H. Jia and Y. Long, *Steroids*, 2024, **208**, 109449, DOI: [10.1016/j.steroids.2024.109449](https://doi.org/10.1016/j.steroids.2024.109449).
- 305 L. Huang, C. Chen, J. Cai, Y. Chen, Y. Zhu, B. Yang, X. Zhou, Y. Liu and H. Tao, *Chem. Biodiversity*, 2024, **21**, e202400070, DOI: [10.1002/cbdv.202400070](https://doi.org/10.1002/cbdv.202400070).
- 306 H.-Y. Hsi, S.-W. Wang, G. Hsiao, L.-K. Chang, Y.-C. Cheng, S.-J. Huang, Y.-S. Lu and T.-H. Lee, *J. Food Drug Anal.*, 2024, **32**, 155–167, DOI: [10.38212/2224-6614.3501](https://doi.org/10.38212/2224-6614.3501).
- 307 Y. Xiao, J. Liu, Y. Jiang, Y. Guo and T. Ye, *Molecules*, 2024, **29**, 3599, DOI: [10.3390/molecules29153599](https://doi.org/10.3390/molecules29153599).
- 308 S. B. Khandekar and R. A. Fernandes, *Org. Biomol. Chem.*, 2024, **22**, 4508–4515, DOI: [10.1039/D4OB00549J](https://doi.org/10.1039/D4OB00549J).
- 309 Z. Xu, X.-T. Liang, L. V. White, M. G. Banwell and S. Tan, *J. Org. Chem.*, 2024, **89**, 12639–12650, DOI: [10.1021/acs.joc.4c01559](https://doi.org/10.1021/acs.joc.4c01559).
- 310 Z. Xu, X.-T. Liang, P. Lan and M. G. Banwell, *Org. Chem. Front.*, 2024, **11**, 2433–2441, DOI: [10.1039/D4OQ00131A](https://doi.org/10.1039/D4OQ00131A).
- 311 R. Ghosh and S. K. Chattopadhyay, *Eur. J. Org. Chem.*, 2024, **27**, e202400228, DOI: [10.1002/ejoc.202400228](https://doi.org/10.1002/ejoc.202400228).
- 312 G. Han, W. Zhang, E. Acs, A. Paquin, Q. Ronzon, N. Casaretto and B. Nay, *Org. Lett.*, 2024, **26**, 2629–2634, DOI: [10.1021/acs.orglett.4c00658](https://doi.org/10.1021/acs.orglett.4c00658).
- 313 E. Fukuda, I. Fujiwara, S. Maruno, K. Motomura, S. Endo, A. Iwasaki, T. Fukuta, A. Nakayama and T. Shinada, *J. Nat. Prod.*, 2024, **87**, 1394–1400, DOI: [10.1021/acs.jnatprod.4c00114](https://doi.org/10.1021/acs.jnatprod.4c00114).
- 314 M. Wang, Y. Jia, F. Wu, Z. Li, L. Shao and X. Chen, *Org. Biomol. Chem.*, 2024, **22**, 7649–7658, DOI: [10.1039/D4OB01186D](https://doi.org/10.1039/D4OB01186D).
- 315 T. Li, S. Jiang, Y. Dai, X. Wu, H. Guo, L. Shi, X. Sang, L. Ren, J. Wang, L. Shi, W. Zhou, H. Li and H.-D. Hao, *Nat. Commun.*, 2024, **15**, 10581, DOI: [10.1038/s41467-024-55189-8](https://doi.org/10.1038/s41467-024-55189-8).
- 316 W. M. Hussein, Y. Zhu, A. A. Salim and R. J. Capon, *Mar. Drugs*, 2024, **22**, 454, DOI: [10.3390/md22100454](https://doi.org/10.3390/md22100454).
- 317 Y. Kawamoto, A. Nishitani, Y. Yoshimura, T. Kobayashi and H. Ito, *Chem. Commun.*, 2024, **60**, 95–97, DOI: [10.1039/D3CC05130G](https://doi.org/10.1039/D3CC05130G).
- 318 G. Rajendar and M. Rehman, *Tetrahedron*, 2024, **168**, 134343, DOI: [10.1016/j.tet.2024.134343](https://doi.org/10.1016/j.tet.2024.134343).
- 319 Y. Liu, Z.-B. Zou, F.-D. Gu, Y.-F. Lin, Y. Li, H.-Y. Chen, H. Liu, X.-W. Yang, G.-M. Liu and Q.-M. Liu, *J. Agric. Food Chem.*, 2024, **72**, 5463–5476, DOI: [10.1021/acs.jafc.3c08126](https://doi.org/10.1021/acs.jafc.3c08126).
- 320 T. Jiang, K. P. Godinez-Macias, J. E. Collins, J. W. Lee, K. L. Wendt, K. Carolino, D. Chakrabarti, R. H. Cichewicz and E. A. Winzeler, *Microbiol. Spectr.*, 2024, **12**, e04142–23, DOI: [10.1128/spectrum.04142-23](https://doi.org/10.1128/spectrum.04142-23).
- 321 H. Zhang, C. Sun, Q. Xia, P. Li, K. Liu and Y. Zhang, *Mar. Drugs*, 2024, **22**, 439, DOI: [10.3390/md22100439](https://doi.org/10.3390/md22100439).
- 322 A. C. Jordão, G. S. dos Santos, T. R. Teixeira, A. J. P. Gluzezak, C. B. de Souza Azevedo, K. de Castro Pereira, L. Tonani, L. R. Gaspar, M. R. von Zeska Kress, P. Colepicolo and H. M. Debonisi, *Arch. Microbiol.*, 2024, **206**, 35, DOI: [10.1007/s00203-023-03756-w](https://doi.org/10.1007/s00203-023-03756-w).
- 323 Y. Fang, J. She, X. Zhang, T. Gu, D. Xie, X. Luo, X. Yi, C. Gao, Y. Liu, C. Zhang, L. Tang and X. Zhou, *J. Nat. Prod.*, 2024, **87**, 322–331, DOI: [10.1021/acs.jnatprod.3c01029](https://doi.org/10.1021/acs.jnatprod.3c01029).
- 324 Y. Rao, R. Su, C. Wu, G. Yang, R. Fu, J. Wu, J. Liang, J. Liu, Z. Jiang, C. Xu and L. Huang, *Front. Pharmacol.*, 2024, **15**, 1320040, DOI: [10.3389/fphar.2024.1320040](https://doi.org/10.3389/fphar.2024.1320040).
- 325 Y. Rao, R. Su, C. Wu, X. Chai, J. Li, G. Yang, J. Wu, T. Fu, Z. Jiang, Z. Guo, C. Xu and L. Huang, *Acta Pharm. Sin. B*, 2024, **14**, 304–318, DOI: [10.1016/j.apsb.2023.08.032](https://doi.org/10.1016/j.apsb.2023.08.032).
- 326 C. Wu, Y. Chen, M. Chen, T. Fu, J. Liu, F. Liu, C. Xu, W. Li, B. Li, Z. Jiang, Y. Rao and L. Huang, *Free Radical Biol. Med.*, 2024, **224**, 630–643, DOI: [10.1016/j.freeradbiomed.2024.09.019](https://doi.org/10.1016/j.freeradbiomed.2024.09.019).
- 327 Y.-S. Liu, W.-L. Chen, Y.-W. Zeng, Z.-H. Li, H.-L. Zheng, N. Pan, L.-Y. Zhao, S. Wang, S.-H. Chen, M.-H. Jiang, C.-C. Jin, Y.-C. Mi, Z.-H. Cai, X.-Z. Fang, Y.-J. Liu, L. Liu and G.-L. Wang, *Mar. Drugs*, 2024, **22**, 283, DOI: [10.3390/md22060283](https://doi.org/10.3390/md22060283).
- 328 X. Niu, S. Chen, X. Wang, J. Wen, X. Liu, Y. Yong, Z. Yu, X. Ma, A. M. Abd El-Aty and X. Ju, *Mar. Drugs*, 2024, **22**, 564, DOI: [10.3390/md22120564](https://doi.org/10.3390/md22120564).



- 329 J. M. Oh, Q. Gao, W.-H. Shin, E.-Y. Lee, D. Chung, G. Choi, S.-J. Nam and H. Kim, *Appl. Biol. Chem.*, 2024, **67**, 26, DOI: [10.1186/s13765-024-00878-7](https://doi.org/10.1186/s13765-024-00878-7).
- 330 Y. Zeng, T. Lu, S. Ren, Z. Hu, J. Fang, Z. Guan, J. Li, L. Liu and Z. Gao, *Org. Lett.*, 2024, **26**, 6692–6697, DOI: [10.1021/acs.orglett.4c02372](https://doi.org/10.1021/acs.orglett.4c02372).
- 331 Z.-F. Zhang, F. Liu, L.-R. Liu, M. Li, L. Cai, S. Liu and J. Mao, *Fungal Biol. Rev.*, 2024, **48**, 100353, DOI: [10.1016/j.fbr.2023.100353](https://doi.org/10.1016/j.fbr.2023.100353).
- 332 C. Han, A. Song, Y. He, L. Yang, L. Chen, W. Dai, Q. Wu and S. Yuan, *Front. Microbiol.*, 2024, **15**, 1520446, DOI: [10.3389/fmicb.2024.1520446](https://doi.org/10.3389/fmicb.2024.1520446).
- 333 Y. Choi, E. J. Jeong, Y. D. Yoo, J. Park and J.-R. Rho, *J. Nat. Prod.*, 2024, **87**, 2537–2542, DOI: [10.1021/acs.jnatprod.4c00851](https://doi.org/10.1021/acs.jnatprod.4c00851).
- 334 S. Moon, D. H. Choi, Y. Lee and J.-R. Rho, *J. Korean Magn. Reson.*, 2024, **28**, 25–31, DOI: [10.6564/JKMRS.2024.28.3.025](https://doi.org/10.6564/JKMRS.2024.28.3.025).
- 335 N. T. T. Thuy, H. T. M. Hien, N. C. Ha, L. T. Thom, D. D. Hong, N. V. Thinh, N. T. Dan, N. D. Hoi, V. T. Loan, H. D. Quang, D. T. T. Hang, P. V. Kiem, B. H. Tai, N. T. M. Hang, N. T. Dat and N. X. Nhiem, *Nat. Prod. Res.*, 2024, **38**, 789–795, DOI: [10.1080/14786419.2023.2201882](https://doi.org/10.1080/14786419.2023.2201882).
- 336 M. Azizah and G. Pohnert, *J. Nat. Prod.*, 2024, **87**, 50–57, DOI: [10.1021/acs.jnatprod.3c00766](https://doi.org/10.1021/acs.jnatprod.3c00766).
- 337 M. Hakamada, C. Tokairin, H. Ishizuka, K. Adachi, T. Osawa, S. Aonuma, R. Hirozumi, S. Tsuchiya, Y. Cho, Y. Kudo, K. Konoki, Y. Oshima, K. Nagasawa and M. Yotsu-Yamashita, *Chem.–Eur. J.*, 2024, **30**, e202304238, DOI: [10.1002/chem.202304238](https://doi.org/10.1002/chem.202304238).
- 338 K. Zhang, X. Yin, Y. Huang, C. Liu, Q. Zhang, Q. Liu, S. Wang, W. Fei, Q. Shi and L. Qiu, *Int. J. Mol. Sci.*, 2024, **25**, 13736–13751, DOI: [10.3390/ijms252413736](https://doi.org/10.3390/ijms252413736).
- 339 A. M. White, B. D. Schwartz, M. G. Gardiner and L. R. Malins, *Org. Lett.*, 2024, **26**, 6803–6808, DOI: [10.1021/acs.orglett.4c02004](https://doi.org/10.1021/acs.orglett.4c02004).
- 340 Y. Ning, Y. Wang and J. Gui, *JACS Au*, 2024, **4**, 635–641, DOI: [10.1021/jacsau.3c00698](https://doi.org/10.1021/jacsau.3c00698).
- 341 T. Obana, M. Nakajima, K. Nakazato, H. Nakagawa, K. Murata, M. Tsuda and H. Fuwa, *J. Am. Chem. Soc.*, 2024, **146**, 29836–29846, DOI: [10.1021/jacs.4c11714](https://doi.org/10.1021/jacs.4c11714).
- 342 E. Montuori, D. De Luca, A. Penna, D. Stalberga and C. Lauritano, *Mar. Drugs*, 2024, **22**, 31–50, DOI: [10.3390/md22010031](https://doi.org/10.3390/md22010031).
- 343 I. Ciss, M. Seck, B. Figadère and L. Ferrié, *Chem.–Eur. J.*, 2024, **30**, e202400471, DOI: [10.1002/chem.202400471](https://doi.org/10.1002/chem.202400471).
- 344 J. Yang, W. Sun, M. Sun, Y. Cui and L. Wang, *Mar. Drugs*, 2024, **22**, 79–95, DOI: [10.3390/md22020079](https://doi.org/10.3390/md22020079).
- 345 J. C. Perkins, K. R. Zenger, Y. Liu and J. M. Strugnell, *Harmful Algae*, 2024, **139**, 102735–102754, DOI: [10.1016/j.hal.2024.102735](https://doi.org/10.1016/j.hal.2024.102735).
- 346 C. O. Miles, I. W. Burton, N. I. Lewis, A. Robertson, S. D. Giddings, P. McCarron and E. M. Mudge, *Tetrahedron*, 2024, **162**, 134115–134124, DOI: [10.1016/j.tet.2024.134115](https://doi.org/10.1016/j.tet.2024.134115).
- 347 J. S. Wood, J. Tang, W. K. Strangman and R. T. Williamson, *J. Nat. Prod.*, 2024, **87**, 2120–2125, DOI: [10.1021/acs.jnatprod.4c00525](https://doi.org/10.1021/acs.jnatprod.4c00525).
- 348 E. M. Mudge, A. L. Wilkins, J. S. Murray, F. Rise and C. O. Miles, *Toxicon*, 2024, **251**, 108154–108165, DOI: [10.1016/j.toxicon.2024.108154](https://doi.org/10.1016/j.toxicon.2024.108154).
- 349 J. L. García-Corona, H. Hegaret, M. Lassudrie, A. Derrien, A. Terre-Terrillon, T. Delaire and C. Fabioux, *Aquat. Toxicol.*, 2024, **266**, 106793, DOI: [10.1016/j.aquatox.2023.106793](https://doi.org/10.1016/j.aquatox.2023.106793).
- 350 T. R. Fallon, V. V. Shende, I. H. Wierzbicki, A. L. Pendleton, N. F. Watervoort, R. P. Auber, D. J. Gonzalez, J. H. Wisecaver and B. S. Moore, *Science*, 2024, **385**, 671–678, DOI: [10.3390/md22050213](https://doi.org/10.3390/md22050213).
- 351 D. Li, X. Liao, S. Xu and B. Zhao, *Nat. Prod. Res.*, 2024, **38**, 1320–1325, DOI: [10.1080/14786419.2022.2140152](https://doi.org/10.1080/14786419.2022.2140152).
- 352 S. Komori, S. Yamabe, R. Matsuta, Y. Yamazaki, M. Fukuoka, S. Sato and K. Takada, *Biosci. Biotechnol. Biochem.*, 2024, **88**, 399–404, DOI: [10.1093/bbb/zbac007](https://doi.org/10.1093/bbb/zbac007).
- 353 Y. Zeng, X. Yang, Z. Xia, R. Chen, F. He, J. Zhang and P. He, *Biology*, 2024, **13**, 456–471, DOI: [10.3390/biology13060456](https://doi.org/10.3390/biology13060456).
- 354 A. R. Mendes, M. P. Spinola, M. Lordelo and J. A. M. Prates, *Appl. Sci.*, 2024, **14**, 10810–10835, DOI: [10.3390/app142310810](https://doi.org/10.3390/app142310810).
- 355 Y. Sun, H. Li, X. Ma, M. Pu, Y. Zhang, Z. Dong, P. He and S. Zheng, *Mar. Drugs*, 2024, **22**, 279–294, DOI: [10.3390/md22060279](https://doi.org/10.3390/md22060279).
- 356 H. S. Kim, J. W. Ahn, N. R. Ha, K. Damodar, S. K. Jang, Y.-M. Yoo, Y. S. Gyoung and S. S. Joo, *Mar. Drugs*, 2024, **22**, 354–374, DOI: [10.3390/md22080354](https://doi.org/10.3390/md22080354).
- 357 J.-Y. Kim, G.-W. Oh, D.-C. Choi, D.-W. Ki, S.-C. Ko, K. W. Kim, D. Yang, M.-J. Yim, J. M. Lee, K. Lee, D.-M. Jo, C. Grace and D.-S. Lee, *J. Appl. Biol. Chem.*, 2024, **67**, 459–463, DOI: [10.3839/jabc.2024.063](https://doi.org/10.3839/jabc.2024.063).
- 358 J. Pan, H. Liu, J. Xu, X. Xu, Y. Qi, C. Wang, C. Wu, Z. Sun, Y. Lv and P. Yan, *Phytochem. Lett.*, 2024, **64**, 133–137, DOI: [10.1016/j.phytol.2024.11.001](https://doi.org/10.1016/j.phytol.2024.11.001).
- 359 K. Ashwini, B. Siva, P. Poornima, S. D. Reddy, V. G. Sastry and K. S. Babu, *Fitoterapia*, 2024, **177**, 106071–110677, DOI: [10.1016/j.fitote.2024.106071](https://doi.org/10.1016/j.fitote.2024.106071).
- 360 K. Wang, C.-H. Xu, L. Tong, B. Wang, A.-H. Liu, S.-W. Li and S.-C. Mao, *J. Nat. Prod.*, 2024, **87**, 121–131, DOI: [10.1021/acs.jnatprod.3c00898](https://doi.org/10.1021/acs.jnatprod.3c00898).
- 361 A. Guy, S. Delly, J. Galano, T. Durand and C. Oger, *Chem.–Eur. J.*, 2024, **30**, e202401632, DOI: [10.1002/chem.202401632](https://doi.org/10.1002/chem.202401632).
- 362 Y. Aalilou, H. Moussa, L. H. Lee, A. Bouyahya, G. Zengin and M. El Abbes Faouzi, *Fitoterapia*, 2024, **176**, 106016–106030, DOI: [10.1016/j.fitote.2024.106016](https://doi.org/10.1016/j.fitote.2024.106016).
- 363 Y. Peng, X. Yang, R. Huang, B. Ren, B. Chen, Y. Liu and H. Zhang, *Mar. Drugs*, 2024, **22**, 59–78, DOI: [10.3390/md22020059](https://doi.org/10.3390/md22020059).
- 364 P. Anand, A. Kaur and S. Singh, *Eur. J. Pharmacol.*, 2024, **967**, 176335–176348, DOI: [10.1016/j.ejphar.2024.176335](https://doi.org/10.1016/j.ejphar.2024.176335).
- 365 S. Ishigami, R. Fukada, G. Nagasaka, T. Tsuruta, K. Nishikawa, Y. Sasaki, K. Nimura, I. Oshima, Y. Yamagishi, Y. Morimoto, T. Kamada and T. Ishii,



- Chem. Biodiversity*, 2024, **21**, e202400436, DOI: [10.1002/cbdv.202400436](https://doi.org/10.1002/cbdv.202400436).
- 366 V. T. Doan, V. T. Pham, C. H. D. Le, T. T. N. Luu, T. U. Dam, H. K. H. Le, Q. T. That, K. P. P. Nguyen and T. T. D. Ngo, *Nat. Prod. Res.*, 2024, **38**, 2258–2263, DOI: [10.1080/14786419.2023.2171407](https://doi.org/10.1080/14786419.2023.2171407).
- 367 K. Kirita, H. Matsumoto, G. Endo and S. Hosokawa, *Angew. Chem., Int. Ed.*, 2024, **63**, e202400586, DOI: [10.1002/anie.202400586](https://doi.org/10.1002/anie.202400586).
- 368 K. Kirita, H. Matsumoto, G. Endo, K. Inoue and S. Hosokawa, *Biosci., Biotechnol., Biochem.*, 2024, **88**, 1144–1154, DOI: [10.1093/bbb/zbac104](https://doi.org/10.1093/bbb/zbac104).
- 369 N. Castejón, A. Adrien, L. Spitzer and S. C. M. Fernandes, *J. Appl. Phycol.*, 2024, **36**, 291–311, DOI: [10.1007/s10811-023-03115-8](https://doi.org/10.1007/s10811-023-03115-8).
- 370 Y. Yamagishi, T. Kamada, T. Ishii, H. Matsuura, N. Kikuchi, T. Abe and M. Suzuki, *Chem. Biodiversity*, 2024, **21**, e202400833, DOI: [10.1002/cbdv.202400833](https://doi.org/10.1002/cbdv.202400833).
- 371 K.-L. Cheong, G. Li, M. Wang, Y. Yang and S. Zhong, *Front. Mar. Sci.*, 2024, **11**, 1360425–1360435, DOI: [10.3389/fmars.2024.1360425](https://doi.org/10.3389/fmars.2024.1360425).
- 372 K. Kaneko, M. Shinke, T. Kamada, T. Okino and A. Yamamoto, *J. Appl. Phycol.*, 2024, **36**, 3795–3808, DOI: [10.1007/s10811-024-03327-6](https://doi.org/10.1007/s10811-024-03327-6).
- 373 T. F. Molinski, *J. Org. Chem.*, 2024, **89**, 10027–10036, DOI: [10.1021/acs.joc.4c00905](https://doi.org/10.1021/acs.joc.4c00905).
- 374 Y. Oyadomari, Y. Goto, K. Suganuma, S.-I. Kawazu, L. E. Becking, N. Fusetani and Y. Nakao, *Mar. Drugs*, 2024, **22**, 171, DOI: [10.3390/md22040171](https://doi.org/10.3390/md22040171).
- 375 A.-Y. Shin and J. Lee, *J. Nat. Prod.*, 2024, **87**, 1872–1880, DOI: [10.1021/acs.jnatprod.4c00692](https://doi.org/10.1021/acs.jnatprod.4c00692).
- 376 R. Kumar, R. K. Akee, L. Martínez-Fructuoso, V. F. Freire, C. C. Thornburg, J. R. Evans, B. D. Peyser, S. Ensel, B. R. O'Keefe and T. Grkovic, *Bioorg. Med. Chem. Lett.*, 2024, **113**, 129963–129967, DOI: [10.1016/j.bmcl.2024.129963](https://doi.org/10.1016/j.bmcl.2024.129963).
- 377 L. A. Amador, A. D. Rodríguez, L. Carmona-Sarabia, E. E. Colón-Lorenzo and A. E. Serrano, *Appl. Sci.*, 2024, **14**, 281–294, DOI: [10.3390/app14010281](https://doi.org/10.3390/app14010281).
- 378 W. Zhong, J. O. Olugbami, P. Rathakrishnan, I. Mohanty, S. G. Moore, N. Garg, A. K. Oyelere, T. L. Turner, A. C. McShan and V. Agarwal, *J. Org. Chem.*, 2024, **89**, 12748–12752, DOI: [10.1021/acs.joc.4c01104](https://doi.org/10.1021/acs.joc.4c01104).
- 379 Y. Hitora, A. H. El-Desoky, Y. Sadahiro, A. Sejiyama, A. Kinoshita, Y. Ise, E. D. Angkouw, R. E. P. Mangindaan, T. Higaki and S. Tsukamoto, *J. Nat. Prod.*, 2024, **87**, 1197–1202, DOI: [10.1021/acs.jnatprod.4c00158](https://doi.org/10.1021/acs.jnatprod.4c00158).
- 380 P. G. Cruz, R. Fernández, R. Rodríguez-Acebes, M. Martínez-Díez, G. Santamaría-Núñez, M. Pérez and C. Cuevas, *Mar. Drugs*, 2024, **22**, 339–349, DOI: [10.3390/md22080339](https://doi.org/10.3390/md22080339).
- 381 D. Lucena-Agell, M. J. Guillén, R. Matesanz, B. Álvarez-Bernad, R. Hortigüela, P. Avilés, M. Martínez-Díez, G. Santamaría-Núñez, J. Contreras, I. Plaza-Menacho, J. F. Giménez-Abián, M. A. Oliva, C. Cuevas and J. F. Díaz, *J. Med. Chem.*, 2024, **67**, 2619–2630, DOI: [10.1021/acs.jmedchem.3c01775](https://doi.org/10.1021/acs.jmedchem.3c01775).
- 382 J. A. Gerke, S. F. Odrón, J. Kim, N. Dutta, J. G. Clarke, J. Media, D. A. Coppage, M. Oorloff, A. Alcalá, G. García, M. E. F. Kang, C. L. Gerke, J. C. Peterson, J. D. Morris, R. Higuchi-Sanabria, F. A. Valeriote, P. Crews and T. A. Johnson, *J. Nat. Prod.*, 2024, **87**, 2523–2529, DOI: [10.1021/acs.jnatprod.4c00691](https://doi.org/10.1021/acs.jnatprod.4c00691).
- 383 C. S. Hassane, F. Tintillier, P.-E. Campos, G. Herbet, N. J. de Voogd, J. Ouazzani, M. Fouillaud, L. Dufossé and A. Gauvin-Bialecki, *Nat. Prod. Res.*, 2024, **38**, 2973–2982, DOI: [10.1080/14786419.2023.2204431](https://doi.org/10.1080/14786419.2023.2204431).
- 384 A. A. E. Said, E. Z. Attia, A. S. Wanas, M. N. Samy, U. R. Abdelmohsen, K. Matsunami and M. A. Fouad, *Nat. Prod. Res.*, 2024, **38**, 3581–3587, DOI: [10.1080/14786419.2023.2256448](https://doi.org/10.1080/14786419.2023.2256448).
- 385 Z. Wang, G.-Q. Li and H.-T. Zhang, *Chem. Biodiversity*, 2024, **21**, e202400939, DOI: [10.1002/cbdv.202400939](https://doi.org/10.1002/cbdv.202400939).
- 386 Z.-J. Guo, H.-X. Liang, X.-Y. Lian, X.-J. Liao, X.-W. Xing, S.-H. Xu and B.-X. Zhao, *J. Asian Nat. Prod. Res.*, 2024, **26**, 328–333, DOI: [10.1080/10286020.2023.2244432](https://doi.org/10.1080/10286020.2023.2244432).
- 387 D. Wang, W. Jiang, M. Churiwal, K. Jia, S. P. D. Senadeera, H. R. Bokesch, G. M. Woldemichael, Y. Kim, R. G. Hawley, J. S. Wei, J. Khan, B. R. O'Keefe, J. A. Beutler and K. R. Gustafson, *J. Nat. Prod.*, 2024, **87**, 332–339, DOI: [10.1021/acs.jnatprod.3c01041](https://doi.org/10.1021/acs.jnatprod.3c01041).
- 388 K. M. Tabakmakher, T. N. Makarieva, Y. E. Sabutski, M. S. Kokoulin, A. S. Menshov, R. S. Popov, A. G. Guzii, L. K. Shubina, E. A. Chingizova, A. R. Chingizov, E. A. Yurchenko, S. N. Fedorov, B. B. Grebnev, G. v. Amsberg, S. A. Dyshlovoy, N. V. Ivanchina and P. S. Dmitrenok, *Mar. Drugs*, 2024, **22**, 396–413, DOI: [10.3390/md22090396](https://doi.org/10.3390/md22090396).
- 389 S. Sala, M. Shimomura, L. Tham, J. Sakata, A. N. Sobolev, S. A. Moggach, J. Fromont, O. Gomez, M. J. Piggott, H. Tokuyama, S. G. Stewart and G. R. Flematti, *J. Nat. Prod.*, 2024, **87**, 2507–2514, DOI: [10.1021/acs.jnatprod.4c00549](https://doi.org/10.1021/acs.jnatprod.4c00549).
- 390 M. Orfanoudaki, M. Dalilian, L. Du, C. H. Chau, W. D. Figg, B. R. O'Keefe and T. Grkovic, *J. Nat. Prod.*, 2024, **87**, 2640–2648, DOI: [10.1021/acs.jnatprod.4c01036](https://doi.org/10.1021/acs.jnatprod.4c01036).
- 391 L.-P. He, X.-C. Luo, C.-X. Zhang and H.-W. Lin, *Phytochemistry*, 2024, **228**, 114250–114256, DOI: [10.1016/j.phytochem.2024.114250](https://doi.org/10.1016/j.phytochem.2024.114250).
- 392 D. Peng, X. Luo, R. Zhu, W. Tong, Y. Yang, G. Li and Q. Wang, *Org. Lett.*, 2024, **26**, 5794–5798, DOI: [10.1021/acs.orglett.4c01976](https://doi.org/10.1021/acs.orglett.4c01976).
- 393 X. Liu, Q. Wang, Y. Zhang and H. Zhang, *Mar. Drugs*, 2024, **22**, 477–486, DOI: [10.3390/md22100477](https://doi.org/10.3390/md22100477).
- 394 L. Gris, M. R. Prinsep, L. M. Peters and C. N. Battershill, *Mar. Drugs*, 2024, **22**, 231–243, DOI: [10.3390/md22050231](https://doi.org/10.3390/md22050231).
- 395 C. J. de Muizon, C. Moriou, M. Levasseur, D. Touboul, B. I. Iorga, H. Nedev, E. V. Elslande, P. Retailleau, S. Petek, E. Folcher, A. Bianchi, M. Thomas, S. Viallon, S. Peyroche, S. Nahle, M. Rousseau and A. Al-Mourabit, *Mar. Drugs*, 2024, **22**, 196–215, DOI: [10.3390/md22050196](https://doi.org/10.3390/md22050196).
- 396 S. Hayes, Y. Lu, B. H. A. Rehm and R. A. Davis, *Beilstein J. Org. Chem.*, 2024, **20**, 3205–3214, DOI: [10.3762/bjoc.20.266](https://doi.org/10.3762/bjoc.20.266).



- 397 A. M. G. Ang, M. M. Uy, E. Ohta, H. Ômura and S. Ohta, *Chem. Biodiversity*, 2024, **21**, e202400962, DOI: [10.1002/cbdv.202400962](https://doi.org/10.1002/cbdv.202400962).
- 398 S.-I. Kurimoto, K. Inoue, T. Ohno and T. Kubota, *Beilstein J. Org. Chem.*, 2024, **20**, 3215–3220, DOI: [10.3762/bjoc.20.267](https://doi.org/10.3762/bjoc.20.267).
- 399 X. Yu, X. Han, Y. Mi, Y. Cui, A. Fu, K. Liu, X. Li, X. Tang and G. Li, *Phytochemistry*, 2024, **226**, 114220–114230, DOI: [10.1016/j.phytochem.2024.114220](https://doi.org/10.1016/j.phytochem.2024.114220).
- 400 J.-X. Li, R.-Y. Shang, D.-D. Xie, X.-C. Luo, T.-Y. Hu, B.-H. Cheng, H.-W. Lin and W.-H. Jiao, *J. Nat. Prod.*, 2024, **87**, 396–403, DOI: [10.1021/acs.jnatprod.3c01239](https://doi.org/10.1021/acs.jnatprod.3c01239).
- 401 J. Wang, Y.-L. Yan, X.-Y. Yu, J.-Y. Pan, X.-L. Liu, L.-L. Hong and B. Wang, *Mar. Drugs*, 2024, **22**, 183–193, DOI: [10.3390/md22040183](https://doi.org/10.3390/md22040183).
- 402 D.-X. Fan, X.-C. Luo, Y.-F. Ding, L.-Y. Liu, X. Wang, J.-Y. Pan, Y.-Y. Ji, J. Wang, C. Li, L.-L. Hong and H.-W. Lin, *Phytochemistry*, 2024, **220**, 114017–114025, DOI: [10.1016/j.phytochem.2024.114017](https://doi.org/10.1016/j.phytochem.2024.114017).
- 403 Y. Cho, C. Bawkar, J. M. Hyun, M. J. Song, K. Jeong and Y.-J. Lee, *J. Nat. Prod.*, 2024, **87**, 358–364, DOI: [10.1021/acs.jnatprod.3c01104](https://doi.org/10.1021/acs.jnatprod.3c01104).
- 404 A. A. E. Said, I. M. Abdel-Rahman, Y. A. Mostafa, E. Z. Attia, M. N. Samy, U. R. Abdelmohsen, K. Matsunami, M. A. Fouad and Y. G. Gouda, *BMC Chem.*, 2024, **18**, 213–222, DOI: [10.1186/s13065-024-01325-w](https://doi.org/10.1186/s13065-024-01325-w).
- 405 A. Ruiz-Molina, D. Pech-Puch, R. E. Millán, L. Ageitos, H. Villegas-Hernández, J. Pachón, J. P. Sestelo, J. Sánchez-Céspedes, J. Rodríguez and C. Jiménez, *Chem.–Eur. J.*, 2024, **30**, e202401844, DOI: [10.1002/chem.202401844](https://doi.org/10.1002/chem.202401844).
- 406 S. S. H. Olsen, S. K. Morrow, J. L. Szabo, M. N. Teng, K. C. Tran, C. D. Amsler, J. B. McClintock and B. J. Baker, *Mar. Drugs*, 2024, **22**, 551–561, DOI: [10.3390/md22120551](https://doi.org/10.3390/md22120551).
- 407 L.-G. Zheng, Y.-Y. Chen, C.-C. Liaw, Y.-C. Lin, S.-Y. Chien, Y.-H. Lo, Y.-C. Tsai, H.-C. Hu, Y.-Y. Chen, T.-L. Hwang, T.-M. Ou and P.-J. Sung, *Phytochem. Lett.*, 2024, **61**, 177–181, DOI: [10.1016/j.phytol.2024.04.013](https://doi.org/10.1016/j.phytol.2024.04.013).
- 408 R. L. Mama, C. D. Gelani, J. M. T. Daluz, M. M. Uy, E. Ohta and S. Ohta, *Nat. Prod. Res.*, 2024, **38**, 2395–2403, DOI: [10.1080/14786419.2023.2175359](https://doi.org/10.1080/14786419.2023.2175359).
- 409 M.-C. Liu, X.-J. Liao, X.-W. Xing, S.-H. Xu and B.-X. Zhao, *J. Asian Nat. Prod. Res.*, 2024, **26**, 1254–1260, DOI: [10.1080/10286020.2024.2360114](https://doi.org/10.1080/10286020.2024.2360114).
- 410 Y.-F. Ding, L.-Y. Liu, J. Tang, D.-X. Fan, Y.-Y. Ji, H.-W. Lin, J. Wang and L.-L. Hong, *Nat. Prod. Res.*, 2024, **38**, 2562–2568, DOI: [10.1080/14786419.2023.2188588](https://doi.org/10.1080/14786419.2023.2188588).
- 411 J. Liu, Z.-Y. Liu, Y. Fu, Y.-C. Gu, S.-W. Li, H.-Y. Zhang and Y.-W. Guo, *Chem. Biodiversity*, 2024, **21**, e202400519, DOI: [10.1002/cbdv.202400519](https://doi.org/10.1002/cbdv.202400519).
- 412 X. Leng, H. He, J. E. H. Lazaro, X. Chen, H. Ouyang, T. Li, X. Yan and S. He, *Phytochemistry*, 2024, **223**, 114097, DOI: [10.1016/j.phytochem.2024.114097](https://doi.org/10.1016/j.phytochem.2024.114097).
- 413 P. Vlachou, N. Tsafantakis, N. Milic, A. Polyzois, E. Baira, A. Termentzi, G. L. Goff, J. Ouazzani and N. Fokialakis, *Mar. Drugs*, 2024, **22**, 522–540, DOI: [10.3390/md22110522](https://doi.org/10.3390/md22110522).
- 414 M. Sauer, A. M. Kany, S. Götze, R. Müller and C. Beemelmans, *Angew. Chem., Int. Ed.*, 2024, **63**, e202416036, DOI: [10.1002/anie.202416036](https://doi.org/10.1002/anie.202416036).
- 415 N. Jamey and L. Ferrié, *Org. Lett.*, 2024, **26**, 5741–5745, DOI: [10.1021/acs.orglett.4c01893](https://doi.org/10.1021/acs.orglett.4c01893).
- 416 H. Ogawa, Y. Nagata and H. Nakamura, *Chin. J. Chem.*, 2024, **42**, 3023–3028, DOI: [10.1002/cjoc.202400637](https://doi.org/10.1002/cjoc.202400637).
- 417 D. M. Wilson and R. Britton, *J. Am. Chem. Soc.*, 2024, **146**, 8456–8463, DOI: [10.1021/jacs.3c14553](https://doi.org/10.1021/jacs.3c14553).
- 418 A. Leblond, A. Nguyen, C. Alcover, K. Leblanc, J.-F. Gallard, D. Joseph, E. Poupon and M. A. Beniddir, *Org. Lett.*, 2024, **26**, 2163–2168, DOI: [10.1021/acs.orglett.4c00194](https://doi.org/10.1021/acs.orglett.4c00194).
- 419 K. Yamazaki, Y. Okuda, A. Takaya and T. Nemoto, *Org. Lett.*, 2024, **26**, 670–675, DOI: [10.1021/acs.orglett.3c04039](https://doi.org/10.1021/acs.orglett.3c04039).
- 420 S. Tsuruda, T. Sato, H. Aoyama, N. Sirimangkalakitti, S. Fujimura, M. Arisawa and K. Murai, *J. Nat. Prod.*, 2024, **87**, 1556–1562, DOI: [10.1021/acs.jnatprod.4c00129](https://doi.org/10.1021/acs.jnatprod.4c00129).
- 421 L. Tham, B. W. Bulcock, S. Sala, G. L. Nealon, G. R. Flematti, S. A. Moggach and M. J. Piggott, *J. Nat. Prod.*, 2024, **87**, 2499–2506, DOI: [10.1021/acs.jnatprod.4c00973](https://doi.org/10.1021/acs.jnatprod.4c00973).
- 422 L. Yi, S. Tan, L. V. White, M.-Y. Liang and M. G. Banwell, *J. Nat. Prod.*, 2024, **87**, 1521–1531, DOI: [10.1021/acs.jnatprod.3c01177](https://doi.org/10.1021/acs.jnatprod.3c01177).
- 423 T. Tsukamoto, K. Takahashi, N. Murase, K. Someya, F. Sakata, T. Yue, T. Kusakabe and K. Kato, *Org. Lett.*, 2024, **26**, 9011–9016, DOI: [10.1021/acs.orglett.4c03158](https://doi.org/10.1021/acs.orglett.4c03158).
- 424 G. L. Barnes, N. L. Magann, D. Perrotta, F. M. Hörmann, S. Fernandez, P. Vydyam, J.-Y. Choi, J. Prudhomme, A. Neal, K. G. Le Roch, C. B. Mamoun and C. D. Vanderwal, *J. Am. Chem. Soc.*, 2024, **146**, 29883–29894, DOI: [10.1021/jacs.4c11897](https://doi.org/10.1021/jacs.4c11897).
- 425 S. P. Rezgui, J. Farhi, H. Yu, Z. P. Sercel, S. C. Virgil and B. M. Stoltz, *Chem. Sci.*, 2024, **15**, 12284–12290, DOI: [10.1039/D4SC02981J](https://doi.org/10.1039/D4SC02981J).
- 426 C. Tolle, M. Fresia, P. G. Jones and T. Lindel, *ChemPhotoChem*, 2024, **8**, e202400065, DOI: [10.1002/cptc.202400065](https://doi.org/10.1002/cptc.202400065).
- 427 A. P. Morrow and M. W. Smith, *J. Am. Chem. Soc.*, 2024, **146**, 2913–2918, DOI: [10.1021/jacs.3c14120](https://doi.org/10.1021/jacs.3c14120).
- 428 R. Fujita, K. Ooka, H. Wasada, Y. Ichikawa and S. Hosokawa, *Org. Biomol. Chem.*, 2024, **22**, 1901–1906, DOI: [10.1039/D4OB00088A](https://doi.org/10.1039/D4OB00088A).
- 429 A. Ortigosa-Palomo, F. Quiñonero, R. Ortiz, F. Sarabia, J. Prados and C. Melguizo, *Mar. Drugs*, 2024, **22**, 101–122, DOI: [10.3390/md22030101](https://doi.org/10.3390/md22030101).
- 430 Q.-B. Yang and L.-F. Liang, *Mar. Drugs*, 2024, **22**, 213–234, DOI: [10.3390/md22050213](https://doi.org/10.3390/md22050213).
- 431 N. Barzkar, S. Sukhikh and O. Babich, *Int. J. Biol. Macromol.*, 2024, **280**, 135823–135840, DOI: [10.1016/j.ijbiomac.2024.135823](https://doi.org/10.1016/j.ijbiomac.2024.135823).
- 432 J. M. J. Nolsøe, J. Underhaug, Å. M. Sørskar, S. G. Antonsen, K. E. Malterud, O. Gani, Q. Fan, M. Hjorth, T. Sæther, T. V. Hansen and Y. H. Stenstrøm, *Molecules*, 2024, **29**, 994–1008, DOI: [10.3390/molecules29050994](https://doi.org/10.3390/molecules29050994).
- 433 D. S. Dissanayake, D. P. Nagahawatta, J.-S. Lee and Y.-J. Jeon, *Mar. Drugs*, 2024, **22**, 426–447, DOI: [10.3390/md22090426](https://doi.org/10.3390/md22090426).
- 434 N. K. Tailor, G. Deswal and A. S. Grewal, *Chemistry*, 2024, **6**, 677–694, DOI: [10.3390/chemistry6040040](https://doi.org/10.3390/chemistry6040040).



- 435 M. Xu, Z. Bai, B. Xie, R. Peng, Z. Du, Y. Liu, G. Zhang, S. Yan, X. Xiao and S. Qin, *Molecules*, 2024, **29**, 933–956, DOI: [10.3390/molecules29050933](https://doi.org/10.3390/molecules29050933).
- 436 P. F. Montenegro, G. N. Pham, F. M. Abdoul-Latif, E. Taffin-de-Givenchy and M. Mehiri, *Mar. Drugs*, 2024, **22**, 132–166, DOI: [10.3390/md22030132](https://doi.org/10.3390/md22030132).
- 437 A. Khatun, K. Hazra, B. Mahato, R. Koley, R. Ghorai, A. Adhikary, B. Debnath and S. Das, *Eur. J. Med. Chem. Rep.*, 2024, **11**, 100158–100196, DOI: [10.1016/j.ejmcr.2024.100158](https://doi.org/10.1016/j.ejmcr.2024.100158).
- 438 J. Xia, X. Chen, G. Li, P. Qiu, W. Wang and Z. Shao, *Mar. Drugs*, 2024, **22**, 447–482, DOI: [10.3390/md22100447](https://doi.org/10.3390/md22100447).
- 439 L. A. Amador, E. E. Colón-Lorenzo, A. D. Rodríguez and A. E. Serrano, *Life*, 2024, **14**, 684–696, DOI: [10.3390/life14060684](https://doi.org/10.3390/life14060684).
- 440 A. Capuano, G. D'Urso, M. Aliberti, D. Ruggiero, S. Terracciano, C. Festa, A. Tosco, M. G. Chini, G. Lauro, G. Bifulco and A. Casapullo, *Mar. Drugs*, 2024, **22**, 41–51, DOI: [10.3390/md22010041](https://doi.org/10.3390/md22010041).
- 441 Y. Sun, A. Dakiiwa, M. Zhang, T. Shibata and M. Kita, *Chem.–Eur. J.*, 2024, **30**, e202402049, DOI: [10.1002/chem.202402049](https://doi.org/10.1002/chem.202402049).
- 442 C.-L. Gao, J.-Q. Song, Z.-N. Yang, H. Wang, X.-Y. Wu, C. Shao, H.-X. Dai, K. Chen, Y.-W. Guo, T. Pang and X.-W. Li, *J. Am. Chem. Soc.*, 2024, **146**, 28384–28397, DOI: [10.1021/jacs.4c09695](https://doi.org/10.1021/jacs.4c09695).
- 443 E. J. Strong, L. Tan, S. Hayes, H. Whyte, R. A. Davis and N. P. West, *Mar. Drugs*, 2024, **22**, 298–306, DOI: [10.3390/md22070298](https://doi.org/10.3390/md22070298).
- 444 M. Sinane, C. Grunberger, L. Gentile, C. Moriou, V. Chaker, P. Coutrot, A. Guenneguez, M.-A. Poullaouec, S. Connan, V. Stiger-Pouvreau, M. Zubia, Y. Fleury, S. Cérantola, N. Kervarec, A. Al-Mourabit, S. Petek and C. Voisset, *Mar. Drugs*, 2024, **22**, 456–473, DOI: [10.3390/md22100456](https://doi.org/10.3390/md22100456).
- 445 G. Risi, M. Liu, F. Vairoletti, R. J. Quinn and G. Salinas, *J. Nat. Prod.*, 2024, **87**, 1532–1539, DOI: [10.1021/acs.jnatprod.3c01212](https://doi.org/10.1021/acs.jnatprod.3c01212).
- 446 X. Wang, W. Xu, Z. Wang, Q. Yu, L. Yuan, Y. Liu, J. Sang, W. Li, S. Zhu, W. Jiang, Z. Li, W. Zhang and Y. Dang, *J. Nat. Prod.*, 2024, **87**, 713–721, DOI: [10.1021/acs.jnatprod.3c00811](https://doi.org/10.1021/acs.jnatprod.3c00811).
- 447 T. Tian, Y. Ogura, Y. Ise, H. Takikawa, S. Okada, S. Matsunaga and G.-L. Tang, *J. Org. Chem.*, 2024, **89**, 9135–9138, DOI: [10.1021/acs.joc.4c00913](https://doi.org/10.1021/acs.joc.4c00913).
- 448 M. Orfanoudaki, R. K. Akee, L. Martínez-Fructuoso, D. Wang, J. A. Kelley, E. A. Smith, C. J. Henrich, M. J. Schnermann, B. R. O'Keefe and T. Grkovic, *J. Nat. Prod.*, 2024, **87**, 415–423, DOI: [10.1021/acs.jnatprod.3c01113](https://doi.org/10.1021/acs.jnatprod.3c01113).
- 449 A. Galitz, S. Vargas, O. P. Thomas, M. M. Reddy, G. Wörheide and D. Erpenbeck, *Chem. Biodiversity*, 2024, **21**, e202400549, DOI: [10.1002/cbdv.202400549](https://doi.org/10.1002/cbdv.202400549).
- 450 A. G. Silva, J. V. C. Rostirola, F. D. Speri, J. G. Pina, M. V. Kitahara, G. B. Longato and J. M. Sciani, *J. Cell. Mol. Med.*, 2024, **28**, e70165, DOI: [10.1111/jcmm.70165](https://doi.org/10.1111/jcmm.70165).
- 451 S. P. D. Senadeera, D. Wang, B. A. P. Wilson, E. A. Smith, A. Wamiru, J. A. Martinez Fiesco, L. Du, P. Zhang, B. R. O'Keefe and J. A. Beutler, *J. Nat. Prod.*, 2024, **87**, 2014–2020, DOI: [10.1021/acs.jnatprod.4c00477](https://doi.org/10.1021/acs.jnatprod.4c00477).
- 452 Y. Sun, X. Zhang, F. Jiang, M. Zhang, W. Wu and Y. Sun, *J. Am. Chem. Soc.*, 2024, **146**, 32305–32310, DOI: [10.1021/jacs.4c14081](https://doi.org/10.1021/jacs.4c14081).
- 453 M.-Y. Cheng, I.-C. Hsu, S.-Y. Huang, Y.-T. Chuang, T.-Y. Ke, H.-W. Chang, T.-H. Chu, C.-Y. Chen and Y.-B. Cheng, *Mar. Drugs*, 2024, **22**, 219, DOI: [10.3390/md22050219](https://doi.org/10.3390/md22050219).
- 454 J. Lackner, C. Alberti, T. Bock, K. Nešmerak, E. Urban, S. Khom and N. Schützenmeister, *Chem.–Eur. J.*, 2024, **30**, e202401921, DOI: [10.1002/chem.202401921](https://doi.org/10.1002/chem.202401921).
- 455 A. Koutsaviti, M. Kvasnicová, G. Gonzalez, T. Štenclová, S. Agusti, C. M. Duarte, L. Rarová, M. Strnad, V. Roussis and E. Ioannou, *Chem. Biodiversity*, 2024, **21**, e202400235, DOI: [10.1002/cbdv.202400235](https://doi.org/10.1002/cbdv.202400235).
- 456 X. Yu, X. Han, Y. Mi, Y. Cui, A. Fu, K. Liu, X. Tang and G. Li, *J. Nat. Prod.*, 2024, **87**, 1150–1158, DOI: [10.1021/acs.jnatprod.4c00112](https://doi.org/10.1021/acs.jnatprod.4c00112).
- 457 Y. Zong, T.-Y. Jin, J.-J. Yang, K.-Y. Wang, X. Shi, Y. Zhang and P.-L. Li, *Mar. Drugs*, 2024, **22**, 145, DOI: [10.3390/md22040145](https://doi.org/10.3390/md22040145).
- 458 V. Kurnianda, H.-C. Hu, P.-J. Sung, Y. Hayashi, K. Mori-Yasumoto, A. Suetake, H. Nakayama, M. Yasumoto-Hirose, Y. Tsutsumi, G. Koja, Y.-C. Tsai, T. Jomori and J. Tanaka, *Tetrahedron Lett.*, 2024, **151**, 155323, DOI: [10.1016/j.tetlet.2024.155323](https://doi.org/10.1016/j.tetlet.2024.155323).
- 459 S. S. H. Olsen, S. Afoullouss, R. M. Young, M. Johnson, A. L. Allcock, M. N. Teng, K. C. Tran and B. J. Baker, *Org. Lett.*, 2024, **26**, 9419–9424, DOI: [10.1021/acs.orglett.4c02549](https://doi.org/10.1021/acs.orglett.4c02549).
- 460 T. Kobayashi, R. Sugitate, K. Uchida, Y. Kawamoto and H. Ito, *Org. Lett.*, 2024, **26**, 1803–1806, DOI: [10.1021/acs.orglett.3c04314](https://doi.org/10.1021/acs.orglett.3c04314).
- 461 T. Kobayashi, F. Hayano, A. Kamiya, Y. Kawamoto and H. Ito, *Org. Lett.*, 2024, **26**, 5105–5109, DOI: [10.1021/acs.orglett.4c01384](https://doi.org/10.1021/acs.orglett.4c01384).
- 462 H. Abe, N. Inaba, M. Nomura, Y. Ogura, N. Fukazawa, R. Saito-Moriya, Y. Kawamoto, T. Kobayashi and H. Ito, *Tetrahedron Lett.*, 2024, **139**, 154991, DOI: [10.1016/j.tetlet.2024.154991](https://doi.org/10.1016/j.tetlet.2024.154991).
- 463 G. Stakanovs, A. Blazevica, D. Rasina, S. Belyakov and A. Jirgensons, *Org. Lett.*, 2024, **26**, 8074–8078, DOI: [10.1021/acs.orglett.4c02942](https://doi.org/10.1021/acs.orglett.4c02942).
- 464 C. Wang, J. Zhang, K. Li, J. Yang, L. Li, S. Wang, H. Hou and P. Li, *Mar. Drugs*, 2024, **22**, 442, DOI: [10.3390/md22100442](https://doi.org/10.3390/md22100442).
- 465 X. Chen, J. Zhang, J. Yang, B. Li, T. Li, H. Ouyang, W. Lin, H. Hu, X. Yan and S. He, *Mar. Drugs*, 2024, **22**, 392, DOI: [10.3390/md22090392](https://doi.org/10.3390/md22090392).
- 466 D. H. Dethe, S. Juyal, N. Sharma and U. Kundu, *Org. Lett.*, 2024, **26**, 3010–3013, DOI: [10.1021/acs.orglett.4c00605](https://doi.org/10.1021/acs.orglett.4c00605).
- 467 X. Han, H. Wang, B. Li, X. Chen, T. Li, X. Yan, H. Ouyang, W. Lin and S. He, *Mar. Drugs*, 2024, **22**, 157, DOI: [10.3390/md22040157](https://doi.org/10.3390/md22040157).
- 468 J. Marrero, L. A. Amador, I. M. Novitskiy, A. G. Kutateladze and A. D. Rodríguez, *Molecules*, 2024, **29**, 2493, DOI: [10.3390/molecules29112493](https://doi.org/10.3390/molecules29112493).



- 469 J. Marrero, L. A. Amador, I. M. Novitskiy, A. G. Kutateladze and A. D. Rodriguez, *Molecules*, 2025, **30**, 2929, DOI: [10.3390/molecules30142929](https://doi.org/10.3390/molecules30142929).
- 470 S.-W. Li, C. Cuadrado, J. Liu, H. Li, W. Tang, A. H. Daranas and Y.-W. Guo, *J. Chromatogr., A*, 2024, **1736**, 465416, DOI: [10.1016/j.chroma.2024.465416](https://doi.org/10.1016/j.chroma.2024.465416).
- 471 D. Zhang, Y. Li, X. Li, X. Han, Z. Wang and G. Li, *Nat. Prod. Res.*, 2024, **38**, 97–102, DOI: [10.1080/14786419.2022.2106568](https://doi.org/10.1080/14786419.2022.2106568).
- 472 Z.-Y. Xia, M.-M. Sun, Y. Jin, M.-Z. Su, S.-W. Li, H. Wang and Y.-W. Guo, *Phytochemistry*, 2024, **219**, 113960, DOI: [10.1016/j.phytochem.2023.113960](https://doi.org/10.1016/j.phytochem.2023.113960).
- 473 N. B. M. Janzing, C. H. R. Senges, P. Dietze, B. Haltli, D. H. Marchbank, R. G. Kerr and J. E. Bandow, *Proteomics*, 2024, **24**, 2300390, DOI: [10.1002/pmic.202300390](https://doi.org/10.1002/pmic.202300390).
- 474 Y. Zong, J.-J. Yang, K. Li, Y.-F. Pei, H.-Y. Hou, Y. Zhang, C.-L. Wang and P.-L. Li, *Phytochemistry*, 2024, **223**, 114113, DOI: [10.1016/j.phytochem.2024.114113](https://doi.org/10.1016/j.phytochem.2024.114113).
- 475 S.-H. Zhu, Y.-M. Chang, M.-Z. Su, L.-G. Yao, S.-W. Li, H. Wang and Y.-W. Guo, *Mar. Drugs*, 2024, **22**, 50, DOI: [10.3390/md22010050](https://doi.org/10.3390/md22010050).
- 476 J.-H. Sheu, L.-G. Zheng, Y.-Y. Chen, S.-Y. Chien and P.-J. Sung, *Phytochem. Lett.*, 2024, **63**, 14–17, DOI: [10.1016/j.phytol.2024.07.012](https://doi.org/10.1016/j.phytol.2024.07.012).
- 477 M. Shaaban and M. A. Ghani, *Z. Naturforsch., C: J. Biosci.*, 2024, **79**, 81–87, DOI: [10.1515/znc-2024-0004](https://doi.org/10.1515/znc-2024-0004).
- 478 C. Wang, J. Zhang, T. Jin, X. Shi, M. Wang, W. Zhang, Y. Zong, G. Li and P. Li, *Nat. Prod. Res.*, 2024, **38**, 3597–3604, DOI: [10.1080/14786419.2023.2258436](https://doi.org/10.1080/14786419.2023.2258436).
- 479 H. Peng, Y. Zeng, R. Zhang, L. Yang, F. Wu, C. Gai, J. Yuan, W. Chang, H. Dai and X. Wang, *Mar. Drugs*, 2024, **22**, 536, DOI: [10.3390/md22120536](https://doi.org/10.3390/md22120536).
- 480 Y. Mi, Y. Yang, X. Tang, X. Han and G. Li, *Chin. J. Chem.*, 2024, **42**, 815–822, DOI: [10.1002/cjoc.202300592](https://doi.org/10.1002/cjoc.202300592).
- 481 Y. Mi, X. Han, X. Yu, L. Li, X. Tang and G. Li, *Phytochemistry*, 2024, **223**, 114109, DOI: [10.1016/j.phytochem.2024.114109](https://doi.org/10.1016/j.phytochem.2024.114109).
- 482 T. A. Mohamed, S. M. Abdelmawgoud, A. A. Hamdy, M. A. A. Ibrahim, A. I. Elshamy, M. A. M. Atia, H. A. Kassem, M.-E. F. Hegazy and N. M. Selim, *Nat. Prod. Res.*, 2024, **38**, 512–522, DOI: [10.1080/14786419.2022.2130915](https://doi.org/10.1080/14786419.2022.2130915).
- 483 Y.-Y. Cui, Y. Jin, R.-N. Sun, X. Wang, C.-L. Gao, X.-Y. Cui, K.-X. Chen, Y.-L. Sun, Y.-W. Guo, J. Li and X.-W. Li, *J. Med. Chem.*, 2024, **67**, 12248–12260, DOI: [10.1021/acs.jmedchem.4c00950](https://doi.org/10.1021/acs.jmedchem.4c00950).
- 484 C. Huang, Y. Jin, R. Sun, K. Hu, L. Yao, Y. Guo, Z. Yuan and X. Li, *Chem. Biodiversity*, 2024, **21**, e202401146, DOI: [10.1002/cbdv.202401146](https://doi.org/10.1002/cbdv.202401146).
- 485 M. Sun, S. Li, J. Zeng, Y. Guo, C. Wang and M. Su, *Mar. Drugs*, 2024, **22**, 553, DOI: [10.3390/md22120553](https://doi.org/10.3390/md22120553).
- 486 P. D. Scesa, S. P. Roche and L. West, *Org. Lett.*, 2024, **26**, 1123–1127, DOI: [10.1021/acs.orglett.3c03604](https://doi.org/10.1021/acs.orglett.3c03604).
- 487 D. S. Lin, G. Späth, Z. Meng, L. H. E. Wieske, C. Farès and A. Fürstner, *J. Am. Chem. Soc.*, 2024, **146**, 24250–24256, DOI: [10.1021/jacs.4c09467](https://doi.org/10.1021/jacs.4c09467).
- 488 E. J. Simmons, D. B. Ryffel, D. A. Lopez, Y. D. Boyko and D. Sarlah, *J. Am. Chem. Soc.*, 2024, **147**, 130–135, DOI: [10.1021/jacs.4c16629](https://doi.org/10.1021/jacs.4c16629).
- 489 H. Takamura, Y. Sugitani, R. Morishita, T. Yorisue and I. Kadota, *Org. Biomol. Chem.*, 2024, **22**, 5739, DOI: [10.1039/D4OB00698D](https://doi.org/10.1039/D4OB00698D).
- 490 E. M. Elkady, H. R. Tadros, Y. A. Soliman, M. Raafat and A. M. Abdel-Tawab, *Egypt. J. Aquat. Res.*, 2024, **50**, 234–240, DOI: [10.1016/j.ejar.2024.06.004](https://doi.org/10.1016/j.ejar.2024.06.004).
- 491 S.-W. Tsai, Y.-C. Cheng, Y.-H. Chao and D.-H. Yang, *J. Inflammation Res.*, 2024, **17**, 8299–8311, DOI: [10.2147/JIR.S476847](https://doi.org/10.2147/JIR.S476847).
- 492 L.-M. Ke, D.-D. Yu, M.-Z. Su, L. Cui and Y.-W. Guo, *Mar. Drugs*, 2024, **22**, 95, DOI: [10.3390/md22020095](https://doi.org/10.3390/md22020095).
- 493 X.-Y. Meng, K.-J. Wang, S.-Z. Ye, J.-F. Chen, Z.-Y. Chen, Z.-Y. Zhang, W.-Q. Yin, X.-L. Jia, Y. Li, R. Yu and Q. Ma, *Biochem. Pharmacol.*, 2024, **220**, 116011, DOI: [10.1016/j.bcp.2023.116011](https://doi.org/10.1016/j.bcp.2023.116011).
- 494 R.-Y. Fang, Y.-W. Liu, Y.-G. Goan, J.-J. Lin, J.-H. Su, W.-T. Wu, Y.-J. Wu and N. Ismail, *Adv. Pharmacol. Pharm. Sci.*, 2024, **2024**, 6695837, DOI: [10.1155/2024/6695837](https://doi.org/10.1155/2024/6695837).
- 495 M. A. Bashar, A. Hamed, M. A. M. El-Tabakh, A. H. Hashem, A. A. Zaki, A. A. Al-Askar, E. S. Abou-Amra, M. E. El-Beeh, A. B. M. Mehany, M. Shaaban, A. M. Abdelaziz, K. A. Shaaban and A. I. Hasaballah, *Sci. Rep.*, 2024, **14**, 28028, DOI: [10.1038/s41598-024-75446-6](https://doi.org/10.1038/s41598-024-75446-6).
- 496 Y.-W. Chuang, W.-C. Ting, S.-Y. Chien, L.-G. Zheng, C.-Y. Huang, Z.-H. Wen, Y.-J. Wu, J. Tanaka, Y.-Y. Chen, J.-H. Su and P.-J. Sung, *Phytochem. Lett.*, 2024, **64**, 117–122, DOI: [10.1016/j.phytol.2024.10.008](https://doi.org/10.1016/j.phytol.2024.10.008).
- 497 H. N. Do, Y.-T. Chen, S.-Y. Chien, Y.-Y. Chen, M. M. Zhang, L. K. Tsou, J.-J. Chen, Z.-H. Wen, Y.-H. Lo, L.-G. Zheng and P.-J. Sung, *RSC Adv.*, 2024, **14**, 17195–17201, DOI: [10.1039/D4RA03062A](https://doi.org/10.1039/D4RA03062A).
- 498 H. N. Do, C.-Y. Huang, S.-Y. Chien, L.-G. Zheng, Y.-Y. Chen, Y.-T. Zeng, J.-H. Su, Z.-H. Wen, Y.-C. Chang and P.-J. Sung, *Chem. Pharm. Bull.*, 2024, **72**, 1110–1115, DOI: [10.1248/cpb.c24-00574](https://doi.org/10.1248/cpb.c24-00574).
- 499 H.-W. Chen, F.-C. Liu, H.-M. Kuo, S.-H. Tang, G.-H. Niu, M. M. Zhang, L. K. Tsou, P.-J. Sung and Z.-H. Wen, *Biomed. Pharmacother.*, 2024, **172**, 116279, DOI: [10.1016/j.biopha.2024.116279](https://doi.org/10.1016/j.biopha.2024.116279).
- 500 G. Yao, M. R. Parris, W. C. Kuo, P. Pörzgen, B. Castillo, E. S. Mason, A. Chinchilla, J. Huang, S. Suzuki, R. Ross, E. Akana, S. V. Schuit, S. P. Miller, R. Penner, H.-S. Sun, Z.-P. Feng, K. G. Hull, D. Romo, A. Fleig and F. D. Horgen, *J. Nat. Prod.*, 2024, **87**, 783–797, DOI: [10.1021/acs.jnatprod.3c00942](https://doi.org/10.1021/acs.jnatprod.3c00942).
- 501 S. Afoullouss, R. M. Young, L. K. Jennings, J. Doyle, K. Croke, D. Livorsi, J. H. Adams, M. P. Johnson, O. P. Thomas and A. L. Allcock, *ACS Omega*, 2024, **9**, 41914–41922, DOI: [10.1021/acsomega.4c06361](https://doi.org/10.1021/acsomega.4c06361).
- 502 L. Betschart and K. Altmann, *Angew. Chem., Int. Ed.*, 2024, **63**, e202315423, DOI: [10.1002/anie.202315423](https://doi.org/10.1002/anie.202315423).
- 503 A. H. Eissa, A. M. Abdel-Tawab, E. S. A. E. Hamed, F. Z. El-Ablack and S.-E. N. Ayyad, *Nat. Prod. Res.*, 2024, **38**, 4390–4398, DOI: [10.1080/14786419.2023.2284259](https://doi.org/10.1080/14786419.2023.2284259).



- 504 T.-Y. Cheng, C.-J. Yang, P.-J. Chen, Y.-L. Chen, B.-R. Peng, T.-A. Kung, Z.-H. Wen, K.-H. Lai and H.-M. Chung, *Phytochem. Lett.*, 2024, **61**, 115–119, DOI: [10.1016/j.phytol.2024.03.015](https://doi.org/10.1016/j.phytol.2024.03.015).
- 505 C.-L. Hung, L.-C. Huang, K.-Y. Shih, K.-H. Lai, B.-R. Peng, T.-L. Hwang, S.-Y. Chien, C.-Y. Li and H.-M. Chung, *J. Mol. Struct.*, 2024, **1296**, 136790, DOI: [10.1016/j.molstruc.2023.136790](https://doi.org/10.1016/j.molstruc.2023.136790).
- 506 H.-M. Chung, Z.-Z. Tsai, C.-K. Hsu, P.-J. Chen, Y.-L. Chen, S.-Y. Chien, Z.-H. Wen, P.-J. Sung and K.-H. Lai, *Rec. Nat. Prod.*, 2024, **18**, 357, DOI: [10.25135/rnp.455.2402.3141](https://doi.org/10.25135/rnp.455.2402.3141).
- 507 T.-H. Lee, J.-L. Chen, C.-H. Chang, M.-M. Tsai, H.-C. Tseng, Y.-C. Chang, V. Shanmugam and H.-L. Hsieh, *Biomedicines*, 2024, **12**, 226, DOI: [10.3390/biomedicines12010226](https://doi.org/10.3390/biomedicines12010226).
- 508 C.-Q. Liu, Q.-B. Yang, L. Zhang and L.-F. Liang, *Mar. Drugs*, 2024, **22**, 402, DOI: [10.3390/md22090402](https://doi.org/10.3390/md22090402).
- 509 H. L. Hammond and C. J. Roy, *Toxins*, 2024, **16**, 417, DOI: [10.3390/toxins16100417](https://doi.org/10.3390/toxins16100417).
- 510 V. M. Dembitsky, *Microbiol. Res.*, 2024, **15**, 1516–1575, DOI: [10.3390/microbiolres15030103](https://doi.org/10.3390/microbiolres15030103).
- 511 K. A. Elnahriry, D. C. Wai, L. M. Ashwood, M. U. Naseem, T. G. Szanto, S. Guo, G. Panyi, P. J. Prentis and R. S. Norton, *Biochim. Biophys. Acta, Proteins Proteomics*, 2024, **1872**, 140952, DOI: [10.1016/j.bbapap.2023.140952](https://doi.org/10.1016/j.bbapap.2023.140952).
- 512 A. N. Kvetkina, S. D. Oreshkov, P. A. Mironov, M. M. Zagraev, A. A. Klimovich, Y. V. Deriavko, A. S. Menshov, D. S. Kulbatskii, Y. A. Logashina, Y. A. Andreev, A. O. Chugunov, M. P. Kirpichnikov, E. N. Lyukmanova, E. V. Leychenko and Z. O. Shenkarev, *Mar. Drugs*, 2024, **22**, 542, DOI: [10.3390/md22120542](https://doi.org/10.3390/md22120542).
- 513 T. Homma, M. Ishida, Y. Nagashima and K. Shiomi, *J. Venom. Anim. Toxins Incl. Trop. Dis.*, 2024, **30**, e20240019, DOI: [10.1590/1678-9199-jvatitd-2024-0019](https://doi.org/10.1590/1678-9199-jvatitd-2024-0019).
- 514 B.-Y. Wu and C.-K. Lin, *Asian J. Org. Chem.*, 2024, **13**, e202400074, DOI: [10.1002/ajoc.202400074](https://doi.org/10.1002/ajoc.202400074).
- 515 Y.-Y. Kou, J. Liu, Y.-T. Chang, L.-Y. Liu, F. Sun, Y.-L. Li, J.-R. Leng, H.-W. Lin and F. Yang, *Cell. Oncol.*, 2024, **47**, 1863–1878, DOI: [10.1007/s13402-024-00968-0](https://doi.org/10.1007/s13402-024-00968-0).
- 516 X. Zhang, M. Su, M. Zhu, S. Chen, Z. Gao, Y. Guo and X. Li, *Chin. J. Nat. Med.*, 2024, **22**, 1030–1039, DOI: [10.1016/S1875-5364\(24\)60611-4](https://doi.org/10.1016/S1875-5364(24)60611-4).
- 517 Y. A. T. Ngandjui, T. T. Kereeditse, I. Kamika, L. M. Madikizela and T. A. M. Msagati, *Mar. Drugs*, 2024, **22**, 201, DOI: [10.3390/md22050201](https://doi.org/10.3390/md22050201).
- 518 S. Nandi, S. Khatua, A. Nag, S. Sen, N. Chakraborty, A. Naskar, K. Acharya, R. H. Mekky, M. del Mar Contreras, D. Calina, I. Dini and J. Sharifi-Rad, *Curr. Res. Biotechnol.*, 2024, **7**, 100167, DOI: [10.1016/j.crbiot.2023.100167](https://doi.org/10.1016/j.crbiot.2023.100167).
- 519 M. Tost and U. Kazmaier, *Mar. Drugs*, 2024, **22**, 165, DOI: [10.3390/md22040165](https://doi.org/10.3390/md22040165).
- 520 I. W. Mudianta, J. E. Siregar, A. F. M. Rizki, W. A. Azmi, N. E. Pravitasari, G. M. Gholam, F. R. Putri, R. Kristiana, N. K. D. Cahyani and I. M. Artika, *Biochem. Biophys. Res. Commun.*, 2024, **737**, 150921, DOI: [10.1016/j.bbrc.2024.150921](https://doi.org/10.1016/j.bbrc.2024.150921).
- 521 P. Kancharla, D. Ortiz, C. M. Fargo, X. Zhang, Y. Li, M. Sanchez, A. Kumar, M. Yeluguri, R. A. Dodean, D. Caridha, M. S. Madejczyk, M. Martin, X. Jin, C. Blount, R. Chetree, K. Pannone, H. T. Dinh, J. DeLuca, M. Evans, R. Nadeau, C. Vuong, S. Leed, W. E. Dennis, N. Roncal, B. S. Pybus, P. J. Lee, A. Roth, K. A. Reynolds, J. X. Kelly and S. M. Landfear, *J. Med. Chem.*, 2024, **67**, 8323–8345, DOI: [10.1021/acs.jmedchem.4c00517](https://doi.org/10.1021/acs.jmedchem.4c00517).
- 522 M. Wei, J. Chen, Y. Song, J.-P. Monserrat, Y. Zhang and L. Shen, *Eur. J. Med. Chem.*, 2024, **269**, 116294, DOI: [10.1016/j.ejmech.2024.116294](https://doi.org/10.1016/j.ejmech.2024.116294).
- 523 P. Sivaganesan, S. VL, A. Sahoo, C. Elanchezhian, G. Nataraj and S. Chaudhuri, *ChemistrySelect*, 2024, **9**, e202403112, DOI: [10.1002/slct.202403112](https://doi.org/10.1002/slct.202403112).
- 524 V.-P. M. Mitsiou, A.-M. N. Antonaki, M. D. Douka and K. E. Litinas, *Molecules*, 2024, **29**, 4032, DOI: [10.3390/molecules29174032](https://doi.org/10.3390/molecules29174032).
- 525 S. C. Finch, D. T. Harwood, M. J. Boundy and A. I. Selwood, *Mar. Drugs*, 2024, **22**, 129, DOI: [10.3390/md22030129](https://doi.org/10.3390/md22030129).
- 526 Z.-R. Lin, H.-X. Geng and R.-C. Yu, *J. Hazard. Mater.*, 2024, **469**, 133896, DOI: [10.1016/j.jhazmat.2024.133896](https://doi.org/10.1016/j.jhazmat.2024.133896).
- 527 J. L. García-Corona, C. Fabioux, J. Vanmaldergem, S. Petek, A. Derrien, A. Terre-Terrillon, L. Bressolier, F. Breton and H. Hegaret, *Harmful Algae*, 2024, **133**, 102607, DOI: [10.1016/j.hal.2024.102607](https://doi.org/10.1016/j.hal.2024.102607).
- 528 T. Teng, Z. Zheng, W. Jiao, N. Liu, A. Wang, M. Liu, L. Xie, Z. Yang, J. Hu and Z. Bao, *Genes*, 2024, **15**, 365, DOI: [10.3390/genes15030365](https://doi.org/10.3390/genes15030365).
- 529 G. Chiappa, G. Fassio, M. V. Modica and M. Oliverio, *Toxins*, 2024, **16**, 348, DOI: [10.3390/toxins16080348](https://doi.org/10.3390/toxins16080348).
- 530 Z. Ratibou, N. Inguibert and S. Dutertre, *Toxins*, 2024, **16**, 94, DOI: [10.3390/toxins16020094](https://doi.org/10.3390/toxins16020094).
- 531 L. Martínez-Hernández, E. López-Vera and M. B. Aguilar, *Toxins*, 2024, **16**, 504, DOI: [10.3390/toxins16120504](https://doi.org/10.3390/toxins16120504).
- 532 S. Shekh, S. Dhannura, P. Dhurjad, C. Ravali, S. M. M. S. Kakkat, Vishwajyothi, M. Vijayasathy, R. Sonti and K. H. Gowd, *Toxicon*, 2024, **250**, 108087, DOI: [10.1016/j.toxicon.2024.108087](https://doi.org/10.1016/j.toxicon.2024.108087).
- 533 M. Wang, Z. Liao, D. Zhangsun, Y. Wu and S. Luo, *Molecules*, 2024, **29**, 1181, DOI: [10.3390/molecules29051181](https://doi.org/10.3390/molecules29051181).
- 534 C. Lin, H. Qin, Y. Liao, J. Chen and B. Gao, *Molecules*, 2024, **29**, 2846, DOI: [10.3390/molecules29122846](https://doi.org/10.3390/molecules29122846).
- 535 H. Wang, X. Li, Y. Qiao, M. Wang, W. Wang, J. M. McIntosh, D. Zhangsun and S. Luo, *Mar. Drugs*, 2024, **22**, 49, DOI: [10.3390/md22010049](https://doi.org/10.3390/md22010049).
- 536 X. Guo, L. He, W. Xu, W. Wang, X. Feng, Y. Fu, X. Zhang, R.-B. Ding, X. Qi, J. Bao and S. Luo, *Mar. Drugs*, 2024, **22**, 252, DOI: [10.3390/md22060252](https://doi.org/10.3390/md22060252).
- 537 X. Li, H.-S. Tae, S. Chen, A. Yousuf, L. Huang, J. Zhang, T. Jiang, D. J. Adams and R. Yu, *J. Med. Chem.*, 2024, **67**, 971–987, DOI: [10.1021/acs.jmedchem.3c00979](https://doi.org/10.1021/acs.jmedchem.3c00979).
- 538 S. Raghuraman, J. Carter, M. Walter, M. Karthikeyan, K. Chase, M. L. Giglio, M. Giacobassi, R. W. Teichert, H. Terlau and B. M. Olivera, *Sci. Rep.*, 2024, **14**, 31001, DOI: [10.1038/s41598-024-82165-5](https://doi.org/10.1038/s41598-024-82165-5).



- 539 R. Sakai, K. Matsumura, H. Uchimasu, K. Miyako, T. Taniguchi, V. Raghavendra Rao Kovvuri, A. D. Acharige, K. G. Hull, D. Romo, L. Thaveepornkul, S. Chinnaronk, H. Miyamoto, A. Takada, H. Watari, M. J. Fujita and J. Sakaue, *J. Org. Chem.*, 2024, **89**, 5977–5987, DOI: [10.1021/acs.joc.3c02765](https://doi.org/10.1021/acs.joc.3c02765).
- 540 Y. Li, K. Takamatsu, T. Aota and H. Konno, *Org. Lett.*, 2024, **26**, 3375–3379, DOI: [10.1021/acs.orglett.4c00808](https://doi.org/10.1021/acs.orglett.4c00808).
- 541 D. M. Dukes, V. K. Atanassov and J. M. Smith, *Chem. Sci.*, 2024, **15**, 16554–16558, DOI: [10.1039/D4SC04910A](https://doi.org/10.1039/D4SC04910A).
- 542 L. A. Segura-Quezada, E. D. Hernández-Velázquez, A. R. Corrales-Escobosa, C. de León-Solis and C. R. Solorio-Alvarado, *Chem. Biodiversity*, 2024, **21**, e202300883, DOI: [10.1002/cbdv.202300883](https://doi.org/10.1002/cbdv.202300883).
- 543 M. A. Tammam, M. I. Gamal El-Din, A. Aouidate and A. El-Demerdash, *Bioorg. Chem.*, 2024, **151**, 107654, DOI: [10.1016/j.bioorg.2024.107654](https://doi.org/10.1016/j.bioorg.2024.107654).
- 544 P. J. Batista, G. Nuzzo, C. Gallo, D. Carbone, M. dell'Isola, M. Affuso, G. Barra, F. Albiani, F. Crocetta, R. Virgili, V. Mazzella, D. Castiglia, G. d'Ippolito, E. Manzo and A. Fontana, *Mar. Drugs*, 2024, **22**, 75, DOI: [10.3390/md22020075](https://doi.org/10.3390/md22020075).
- 545 A. L. Agognon, M. Casertano, A. Vito, S. Orso, S. Cabaro, F. Mormone, C. Morelli, G. Perruolo, P. Formisano, M. Menna, C. Imperatore and F. Oriente, *Int. J. Mol. Sci.*, 2024, **25**, 6039, DOI: [10.3390/ijms25116039](https://doi.org/10.3390/ijms25116039).
- 546 N. Spacho, M. Casertano, C. Imperatore, C. Papadopoulos, M. Menna and N. Eleftheriadis, *Chem.–Eur. J.*, 2024, **30**, e202402279, DOI: [10.1002/chem.202402279](https://doi.org/10.1002/chem.202402279).
- 547 H. Wu, G. Lv, L. Liu, R. Hu, F. Zhao, M. Song, S. Zhang, H. Fan, S. Dai, S. u. Rehman, H. Wang and X. Mou, *J. Nat. Prod.*, 2024, **87**, 2779–2789, DOI: [10.1021/acs.jnatprod.4c00946](https://doi.org/10.1021/acs.jnatprod.4c00946).
- 548 T. Kalnins, V. Vitkovska, M. Kazak, D. Zelencova-Gopejenko, M. Ozola, N. Narvaiss, M. Makrečka-Kuka, I. Domračeva, A. Kinens, B. Gukalova, N. Konrad, R. Aav, F. Bonato, D. Lucena-Agell, J. F. Díaz, E. Liepinsh and E. Suna, *J. Med. Chem.*, 2024, **67**, 9227–9259, DOI: [10.1021/acs.jmedchem.4c00388](https://doi.org/10.1021/acs.jmedchem.4c00388).
- 549 H. Zhang, X. Li, Z. Hui, S. Huang, M. Cai, W. Shi, Y. Lin, J. Shen, M. Sui, Q. Lai, Z. Shao, J. Dou, X. Luo, Y. Ge and X. Tang, *Angew. Chem., Int. Ed.*, 2024, **63**, e202318784, DOI: [10.1002/anie.202318784](https://doi.org/10.1002/anie.202318784).
- 550 P. Landete, O.-A. Caliman-Sturdza, J. A. Lopez-Martin, L. Preotescu, M.-C. Luca, A. Kotanidou, P. Villares, S.-P. Iglesias, P. Guisado-Vasco, E.-M. Saiz-Lou, M. del Carmen Farinas-Alvarez, E. M. de Lucas, E. Perez-Alba, J.-M. Cisneros, V. Estrada, C. Hidalgo-Tenorio, G. Poulakou, M. Torralba, J. Fortun, P. Garcia-Ocana, A. Lemaigen, M. Marcos-Martin, M. Molina, R. Paredes, M. T. Perez-Rodriguez, D. Raev, P. Ryan, F. Meira, J. Gomez, N. Torres, D. Lopez-Mendoza, J. Jimeno and J.-F. Varona, *Clin. Infect. Dis.*, 2024, **79**, 910–919, DOI: [10.1093/cid/ciae227](https://doi.org/10.1093/cid/ciae227).
- 551 J. Zhang, L. Zhao, Y. Bai, S. Li, M. Zhang, B. Wei, X. Wang, Y. Xue, L. Li, G. Ma, Y. Tang and X. Wang, *Bioorg. Med. Chem. Lett.*, 2024, **103**, 129706, DOI: [10.1016/j.bmcl.2024.129706](https://doi.org/10.1016/j.bmcl.2024.129706).
- 552 A. Povo-Retana, R. Landauro-Vera, C. Alvarez-Lucena, M. Cascante and L. Boscá, *Molecules*, 2024, **29**, 331, DOI: [10.3390/molecules29020331](https://doi.org/10.3390/molecules29020331).
- 553 R. Matsuta, S. Shigeno, T. Ohshiro, M. Ueda and K. Takada, *Fish. Sci.*, 2024, **90**, 787–793, DOI: [10.1007/s12562-024-01803-z](https://doi.org/10.1007/s12562-024-01803-z).
- 554 A. A. Kicha, T. V. Malyarenko, A. S. Kuzmich, O. S. Malyarenko, A. I. Kalinovsky, R. S. Popov, D. K. Tolkanov and N. V. Ivanchina, *Mar. Drugs*, 2024, **22**, 43, DOI: [10.3390/md22010043](https://doi.org/10.3390/md22010043).
- 555 S. J. Smith, T. Wang and S. F. Cummins, *Mar. Drugs*, 2024, **22**, 552, DOI: [10.3390/md22120552](https://doi.org/10.3390/md22120552).
- 556 R. Zhang, Z. Lu, D. Wang, Z. Yan, X. Sun, X. Li, X. Yin, S. Wang and K. Li, *Mar. Drugs*, 2024, **22**, 545, DOI: [10.3390/md22120545](https://doi.org/10.3390/md22120545).
- 557 A. A. Kicha, D. K. Tolkanov, T. V. Malyarenko, O. S. Malyarenko, A. S. Kuzmich, A. I. Kalinovsky, R. S. Popov, V. A. Stonik, N. V. Ivanchina and P. S. Dmitrenok, *Mar. Drugs*, 2024, **22**, 294, DOI: [10.3390/md22070294](https://doi.org/10.3390/md22070294).
- 558 T. V. Malyarenko, V. M. Zakharenko, A. A. Kicha, A. I. Ponomarenko, I. V. Manzhulo, A. I. Kalinovsky, R. S. Popov, P. S. Dmitrenok and N. V. Ivanchina, *Mar. Drugs*, 2024, **22**, 508, DOI: [10.3390/md22110508](https://doi.org/10.3390/md22110508).
- 559 T. V. Malyarenko, A. A. Kicha, A. S. Kuzmich, O. S. Malyarenko, A. I. Kalinovsky, R. S. Popov, P. S. Dmitrenok, N. V. Ivanchina and V. A. Stonik, *Mar. Drugs*, 2024, **22**, 19, DOI: [10.3390/md22010019](https://doi.org/10.3390/md22010019).
- 560 A. S. Silchenko, A. I. Kalinovsky, S. A. Avilov, R. S. Popov, E. A. Chingizova, E. S. Menchinskaya, E. A. Zelepuga, K. M. Tabakmakher, V. G. Stepanov and V. I. Kalinin, *Mar. Drugs*, 2024, **22**, 292, DOI: [10.3390/md22070292](https://doi.org/10.3390/md22070292).
- 561 A. S. Silchenko, E. A. Chingizova, E. S. Menchinskaya, E. A. Zelepuga, A. I. Kalinovsky, S. A. Avilov, K. M. Tabakmakher, R. S. Popov, P. S. Dmitrenok, S. S. Dautov and V. I. Kalinin, *Mar. Drugs*, 2024, **22**, 560, DOI: [10.3390/md22120560](https://doi.org/10.3390/md22120560).
- 562 Y. Liu, Z. Lu, Z. Yan, X. Li, X. Yin, R. Zhang, Y. Li, S. Wang, R. Xie and K. Li, *Chem. Biodiversity*, 2024, **21**, e202400335, DOI: [10.1002/cbdv.202400335](https://doi.org/10.1002/cbdv.202400335).
- 563 Y. Liu, Z. Lu, Z. Yan, A. Lin, S. Han, Y. Li, X. Yang, X. Li, X. Yin, R. Zhang and K. Li, *J. Chem. Ecol.*, 2024, **50**, 185–196, DOI: [10.1007/s10886-024-01483-0](https://doi.org/10.1007/s10886-024-01483-0).
- 564 F. M. Abdelkarem, H. K. Assaf, Y. A. Mostafa, A. Mahdy, M. F. Hussein, S. A. Ross and N. M. Mohamed, *RSC Adv.*, 2024, **14**, 14185–14193, DOI: [10.1039/D4RA01966K](https://doi.org/10.1039/D4RA01966K).
- 565 E. A. Vasileva, D. V. Berdyshev, N. P. Mishchenko, A. V. Gerasimenko, E. S. Menchinskaya, E. A. Pislyagin, E. A. Chingizova, L. A. Kaluzhskiy, S. S. Dautov and S. A. Fedoreyev, *Biomolecules*, 2024, **14**, 151, DOI: [10.3390/biom14020151](https://doi.org/10.3390/biom14020151).
- 566 I. A. Abdelmawgood, N. A. Mahana, A. M. Badr and A. S. Mohamed, *Naunyn-Schmiedeberg's Arch. Pharmacol.*, 2024, **397**, 1803, DOI: [10.1007/s00210-023-02678-0](https://doi.org/10.1007/s00210-023-02678-0).



- 567 D.-Y. Eum, C. Lee, C. S. Tran, J. Lee, S. Y. Park, M.-S. Jeong, Y. Jin, J. W. Shim, S. R. Lee, M. Koh, E. A. Vasileva, N. P. Mishchenko, S.-J. Park, S. H. Choi, Y. J. Choi, H. Yun and K. Heo, *Toxicol. Res.*, 2024, **40**, 409–419, DOI: [10.1007/s43188-024-00232-5](https://doi.org/10.1007/s43188-024-00232-5).
- 568 S. Promtang, T. Sanguanphun, P. Chalorak, L. S. Pe, N. Niamnont, P. Sobhon and K. Meemon, *ACS Chem. Neurosci.*, 2024, **15**, 2182–2197, DOI: [10.1021/acscchemneuro.4c00008](https://doi.org/10.1021/acscchemneuro.4c00008).
- 569 L. Herr, J. R. Ye, S. W. Kang, S. T. Ro, Y. K. Chae, K. E. Lee, M. S. Kim, M. K. Jih, C. Lee, S. C. Choi and O. H. Nam, *Dent. Traumatol.*, 2024, **40**, 178–186, DOI: [10.1111/edt.12891](https://doi.org/10.1111/edt.12891).
- 570 A. Elkattan, M. Matsumoto, M. Nagata, Y. Mittraphab, G. A. Wahab, A. Ashour, A. A. Zaki, A. E. H. El-Sayed and K. Shimizu, *Cytotechnology*, 2024, **76**, 721–734, DOI: [10.1007/s10616-024-00649-8](https://doi.org/10.1007/s10616-024-00649-8).
- 571 C. Lu, X. Wang, J. Ma, M. Wang, W. Liu, G. Wang, Y. Ding, Z. Lin and Y. Li, *Arch. Pharm.*, 2024, **357**, 2300427, DOI: [10.1002/ardp.202300427](https://doi.org/10.1002/ardp.202300427).
- 572 M. Liao, X. Xia, Q. Meng, C. Zhu, B. Liao, J. Wang, L. Gou, X. Zhou, W. Yuan, L. Cheng and B. Ren, *Br. J. Pharmacol.*, 2024, **181**, 1857–1873, DOI: [10.1111/bph.16333](https://doi.org/10.1111/bph.16333).
- 573 E. S. Menchinskaya, S. A. Dyshlovoy, S. Venz, C. Jacobsen, J. Hauschild, T. Rohlfing, A. S. Silchenko, S. A. Avilov, S. Balabanov, C. Bokemeyer, D. L. Aminin, G. von Amsberg and F. Honecker, *Mar. Drugs*, 2024, **22**, 20, DOI: [10.3390/md22010020](https://doi.org/10.3390/md22010020).
- 574 E. S. Menchinskaya, E. A. Chingizova, E. A. Pisyagin, E. A. Yurchenko, A. A. Klimovich, E. A. Zelepuga, D. L. Aminin, S. A. Avilov and A. S. Silchenko, *Mar. Drugs*, 2024, **22**, 474, DOI: [10.3390/md22100474](https://doi.org/10.3390/md22100474).
- 575 S. Wang, C. Zhong, Y. Huang and P. Lu, *Angew. Chem., Int. Ed.*, 2024, **63**, e202400515, DOI: [10.1002/anie.202400515](https://doi.org/10.1002/anie.202400515).
- 576 H. M. Ameen, A. Jayadev, G. Prasad and D. I. Nair, *Molecules*, 2024, **29**, 4596, DOI: [10.3390/molecules29194596](https://doi.org/10.3390/molecules29194596).
- 577 Z. Chebaro, J. E. Mesmar, A. Badran, A. Al-Sawalmih, M. Maresca and E. Baydoun, *Biomolecules*, 2024, **14**, 991, DOI: [10.3390/biom14080991](https://doi.org/10.3390/biom14080991).
- 578 N. Wehbe, M. Bechelany, A. Badran, A. Al-Sawalmih, J. E. Mesmar and E. Baydoun, *Pharmaceuticals*, 2024, **17**, 993, DOI: [10.3390/ph17080993](https://doi.org/10.3390/ph17080993).
- 579 N. A. Cacciola, P. De Cicco, R. Amico, F. Sepe, Y. Li, L. Grauso, M. F. Nani, S. Scarpatto, C. Zidorn, A. Mangoni and F. Borrelli, *Phytother. Res.*, 2024, **38**, 4168–4176, DOI: [10.1002/ptr.8269](https://doi.org/10.1002/ptr.8269).
- 580 W. R. Kem, F. Soti, J. R. Rocca and J. V. Johnson, *Mar. Drugs*, 2024, **22**, 141, DOI: [10.3390/md22040141](https://doi.org/10.3390/md22040141).
- 581 L. K. Jennings, N. Kaur, M. C. Ramos, F. Reyes, M. M. Reddy and O. P. Thomas, *J. Nat. Prod.*, 2024, **87**, 906–913, DOI: [10.1021/acs.jnatprod.3c01186](https://doi.org/10.1021/acs.jnatprod.3c01186).
- 582 O. A. Shepperson, P. W. R. Harris, M. A. Brimble and A. J. Cameron, *Antibiotics*, 2024, **13**, 615, DOI: [10.3390/antibiotics13070615](https://doi.org/10.3390/antibiotics13070615).
- 583 V. N. Safronova, P. V. Pantelev, R. N. Kruglikov, I. A. Bolosov, E. I. Finkina and T. V. Ovchinnikova, *Russ. J. Bioorg. Chem.*, 2024, **50**, 629–643, DOI: [10.1134/S1068162024030087](https://doi.org/10.1134/S1068162024030087).
- 584 M. Dhanji-Rapkova, R. G. Hatfield, D. I. Walker, C. Hooper, S. Alewijnse, C. Baker-Austin, A. D. Turner and J. M. Ritchie, *Mar. Drugs*, 2024, **22**, 458, DOI: [10.3390/md22100458](https://doi.org/10.3390/md22100458).
- 585 Y. Bai, W. Zhang, W. Zheng, X.-Z. Meng, Y. Duan, C. Zhang, F. Chen and K.-J. Wang, *Biochem. Pharmacol.*, 2024, **226**, 116344, DOI: [10.1016/j.bcp.2024.116344](https://doi.org/10.1016/j.bcp.2024.116344).
- 586 H.-H. Yu, L.-Y. Wu, P.-L. Hsu, C.-W. Lee and B.-C. Su, *Probiotics Antimicrob. Proteins*, 2024, **16**, 1724, DOI: [10.1007/s12602-023-10130-1](https://doi.org/10.1007/s12602-023-10130-1).
- 587 F.-C. Chiu, H.-M. Kuo, C.-L. Yu, P. Selvam, I.-L. Su, C.-C. Tseng, C.-H. Yuan and Z.-H. Wen, *Free Radical Biol. Med.*, 2024, **220**, 28–42, DOI: [10.1016/j.freeradbiomed.2024.04.235](https://doi.org/10.1016/j.freeradbiomed.2024.04.235).
- 588 Q. Wu, C. Munschy, N. Bodin and W. Vetter, *J. Agric. Food Chem.*, 2024, **72**, 15643–15652, DOI: [10.1021/acs.jafc.4c03503](https://doi.org/10.1021/acs.jafc.4c03503).
- 589 A. Moores and V. G. Zuin Zeidler, *Nat. Rev. Chem.*, 2025, **9**, 649–650, DOI: [10.1038/s41570-025-00757-9](https://doi.org/10.1038/s41570-025-00757-9).
- 590 M. Varadi, D. Bertoni, P. Magana, U. Paramval, I. Pidruchna, M. Radhakrishnan, M. Tsenkov, S. Nair, M. Mirdita, J. Yeo, O. Kovalevskiy, K. Tunyasuvunakool, A. Laydon, A. Zidek, H. Tomlinson, D. Hariharan, J. Abrahamson, T. Green, J. Jumper, E. Birney, M. Steinegger, D. Hassabis and S. Velankar, *Nucleic Acids Res.*, 2024, **52**, D368–D375, DOI: [10.1093/nar/gkad1011](https://doi.org/10.1093/nar/gkad1011).
- 591 F. I. Saldivar-Gonzalez, V. D. Aldas-Bulos, J. L. Medina-Franco and F. Plisson, *Chem. Sci.*, 2022, **13**, 1526–1546, DOI: [10.1039/d1sc04471k](https://doi.org/10.1039/d1sc04471k).
- 592 G. Hu and M. Qiu, *Nat. Prod. Rep.*, 2023, **40**, 1735–1753, DOI: [10.1039/d3np00025g](https://doi.org/10.1039/d3np00025g).
- 593 D. Meijer, M. A. Beniddir, C. W. Coley, Y. M. Mejri, M. Ozturk, J. J. J. van der Hooft, M. H. Medema and A. Skiredj, *Nat. Prod. Rep.*, 2025, **42**, 654–662, DOI: [10.1039/d4np00008k](https://doi.org/10.1039/d4np00008k).
- 594 A. Gangwal and A. Lavecchia, *J. Med. Chem.*, 2025, **68**, 3948–3969, DOI: [10.1021/acs.jmedchem.4c01257](https://doi.org/10.1021/acs.jmedchem.4c01257).
- 595 A. Rutz, W. Bittremieux, R. Schmid, O. Cailloux, J. J. J. van der Hooft and M. A. Beniddir, *Nat. Prod. Rep.*, 2025, DOI: [10.1039/d5np00034c](https://doi.org/10.1039/d5np00034c).
- 596 I. M. Novitskiy and A. G. Kutateladze, *J. Org. Chem.*, 2022, **87**, 4818–4828, DOI: [10.1021/acs.joc.2c00169](https://doi.org/10.1021/acs.joc.2c00169).
- 597 A. Howarth, K. Ermanis and J. M. Goodman, *Chem. Sci.*, 2020, **11**, 4351–4359, DOI: [10.1039/d0sc00442a](https://doi.org/10.1039/d0sc00442a).
- 598 G. D. Hannigan, D. Prihoda, A. Palicka, J. Soukup, O. Klempir, L. Rampula, J. Durcak, M. Wurst, J. Kotowski, D. Chang, R. Wang, G. Piizzi, G. Temesi, D. J. Hazuda, C. H. Woelk and D. A. Bitton, *Nucleic Acids Res.*, 2019, **47**, e110, DOI: [10.1093/nar/gkz654](https://doi.org/10.1093/nar/gkz654).
- 599 Y. Y. Lee, M. Guler, D. N. Chigumba, S. Wang, N. Mittal, C. Miller, B. Krummenacher, H. Liu, L. Cao, A. Kannan, K. Narayan, S. T. Slocum, B. L. Roth, A. Gurevich, B. Behsaz, R. D. Kersten and H. Mohimani, *Nat. Commun.*, 2023, **14**, 4219, DOI: [10.1038/s41467-023-39905-4](https://doi.org/10.1038/s41467-023-39905-4).



- 600 N. J. van Eck and L. Waltman, *Scientometrics*, 2010, **84**, 523–538, DOI: [10.1007/s11192-009-0146-3](https://doi.org/10.1007/s11192-009-0146-3).
- 601 A. R. Carroll, B. R. Copp, R. A. Davis, R. A. Keyzers and M. R. Prinsep, *Nat. Prod. Rep.*, 2022, **39**, 1122–1171, DOI: [10.1039/d1np00076d](https://doi.org/10.1039/d1np00076d).
- 602 R. Reher, H. W. Kim, C. Zhang, H. H. Mao, M. Wang, L. F. Nothias, A. M. Caraballo-Rodriguez, E. Glukhov, B. Teke, T. Leao, K. L. Alexander, B. M. Duggan, E. L. Van Everbroeck, P. C. Dorrestein, G. W. Cottrell and W. H. Gerwick, *J. Am. Chem. Soc.*, 2020, **142**, 4114–4120, DOI: [10.1021/jacs.9b13786](https://doi.org/10.1021/jacs.9b13786).
- 603 H. W. Kim, C. Zhang, R. Reher, M. Wang, K. L. Alexander, L. F. Nothias, Y. K. Han, H. Shin, K. Y. Lee, K. H. Lee, M. J. Kim, P. C. Dorrestein, W. H. Gerwick and G. W. Cottrell, *J. Cheminf.*, 2023, **15**, 71, DOI: [10.1186/s13321-023-00738-4](https://doi.org/10.1186/s13321-023-00738-4).
- 604 H. W. Kim, M. Wang, C. A. Leber, L. F. Nothias, R. Reher, K. B. Kang, J. J. J. van der Hooft, P. C. Dorrestein, W. H. Gerwick and G. W. Cottrell, *J. Nat. Prod.*, 2021, **84**, 2795–2807, DOI: [10.1021/acs.jnatprod.1c00399](https://doi.org/10.1021/acs.jnatprod.1c00399).
- 605 J. R. Pardos-Blas, M. J. Tenorio, J. C. G. Galindo and R. Zardoya, *Mar. Drugs*, 2022, **20**, 149, DOI: [10.3390/md20020149](https://doi.org/10.3390/md20020149).
- 606 C. R. Pye, M. J. Bertin, R. S. Lokey, W. H. Gerwick and R. G. Linington, *Proc. Natl. Acad. Sci. U. S. A.*, 2017, **114**, 5601–5606, DOI: [10.1073/pnas.1614680114](https://doi.org/10.1073/pnas.1614680114).
- 607 C. R. McClain, T. J. Webb, N. A. Heim, M. L. Knope, P. M. Monarrez and J. L. Payne, *Oikos*, 2025, e10828, DOI: [10.1111/oik.10828](https://doi.org/10.1111/oik.10828).
- 608 P. G. Albano, B. Sabelli and P. Bouchet, *Biodivers. Conserv.*, 2011, **20**, 3223–3237, DOI: [10.1007/s10531-011-0117-x](https://doi.org/10.1007/s10531-011-0117-x).
- 609 D. Levine, M. Shuaibi, E. C. Spotte-Smith, M. G. Taylor, M. R. Hasyim, K. Michel, I. Batatia, G. Csányi, M. Dzamba, P. Eastman, N. C. Frey, X. Fu, V. Gharakhanyan, A. S. Krishnapriyan, J. A. Rackers, S. Raja, A. Rizvi, A. S. Rosen, Z. Ulissi, S. Vargas, C. L. Zitnick, S. M. Blau and B. M. Wood, *arXiv*, 2025, preprint, arXiv:2505.08762, DOI: [10.48550/arXiv.2505.08762](https://doi.org/10.48550/arXiv.2505.08762).
- 610 X. Chang, *Adv. Econ. Manag. Political Sci.*, 2023, **23**, 134–140, DOI: [10.54254/2754-1169/23/20230367](https://doi.org/10.54254/2754-1169/23/20230367).
- 611 M. Wang, J. J. Carver, V. V. Phelan, L. M. Sanchez, N. Garg, Y. Peng, D. D. Nguyen, J. Watrous, C. A. Kapon, T. Luzzatto-Knaan, C. Porto, A. Bouslimani, A. V. Melnik, M. J. Meehan, W. T. Liu, M. Crusemann, P. D. Boudreau, E. Esquenazi, M. Sandoval-Calderon, R. D. Kersten, L. A. Pace, R. A. Quinn, K. R. Duncan, C. C. Hsu, D. J. Floros, R. G. Gavilan, K. Kleigrewe, T. Northen, R. J. Dutton, D. Parrot, E. E. Carlson, B. Aigle, C. F. Michelsen, L. Jelsbak, C. Sohlenkamp, P. Pevzner, A. Edlund, J. McLean, J. Piel, B. T. Murphy, L. Gerwick, C. C. Liaw, Y. L. Yang, H. U. Humpf, M. Maansson, R. A. Keyzers, A. C. Sims, A. R. Johnson, A. M. Sidebottom, B. E. Sedio, A. Klitgaard, C. B. Larson, C. A. B. P, D. Torres-Mendoza, D. J. Gonzalez, D. B. Silva, L. M. Marques, D. P. Demarque, E. Pociute, E. C. O'Neill, E. Briand, E. J. N. Helfrich, E. A. Granatosky, E. Glukhov, F. Ryffel, H. Houson, H. Mohimani, J. J. Kharbush, Y. Zeng, J. A. Vorholt, K. L. Kurita, P. Charusanti, K. L. McPhail, K. F. Nielsen, L. Vuong, M. Elfeki, M. F. Traxler, N. Engene, N. Koyama, O. B. Vining, R. Baric, R. R. Silva, S. J. Mascuch, S. Tomasi, S. Jenkins, V. Macherla, T. Hoffman, V. Agarwal, P. G. Williams, J. Dai, R. Neupane, J. Gurr, A. M. C. Rodriguez, A. Lamsa, C. Zhang, K. Dorrestein, B. M. Duggan, J. Almaliti, P. M. Allard, P. Phapale, L. F. Nothias, T. Alexandrov, M. Litaudon, J. L. Wolfender, J. E. Kyle, T. O. Metz, T. Peryea, D. T. Nguyen, D. VanLeer, P. Shinn, A. Jadhav, R. Muller, K. M. Waters, W. Shi, X. Liu, L. Zhang, R. Knight, P. R. Jensen, B. O. Palsson, K. Pogliano, R. G. Linington, M. Gutierrez, N. P. Lopes, W. H. Gerwick, B. S. Moore, P. C. Dorrestein and N. Bandeira, *Nat. Biotechnol.*, 2016, **34**, 828–837, DOI: [10.1038/nbt.3597](https://doi.org/10.1038/nbt.3597).
- 612 R. Schmid, S. Heuckeroth, A. Korf, A. Smirnov, O. Myers, T. S. Dyrland, R. Bushuiev, K. J. Murray, N. Hoffmann, M. Lu, A. Sarvepalli, Z. Zhang, M. Fleischauer, K. Duhrop, M. Wesner, S. J. Hoogstra, E. Rudt, O. Mokshyna, C. Brungs, K. Ponomarov, L. Mutabdzija, T. Damiani, C. J. Pudney, M. Earll, P. O. Helmer, T. R. Fallon, T. Schulze, A. Rivas-Ubach, A. Bilbao, H. Richter, L. F. Nothias, M. Wang, M. Oresic, J. K. Weng, S. Bocker, A. Jeibmann, H. Hayen, U. Karst, P. C. Dorrestein, D. Petras, X. Du and T. Pluskal, *Nat. Biotechnol.*, 2023, **41**, 447–449, DOI: [10.1038/s41587-023-01690-2](https://doi.org/10.1038/s41587-023-01690-2).
- 613 WoRMS Editorial Board, World Register of Marine Species (WoRMS), <https://www.marinespecies.org>, accessed 2025-11-06, DOI: [10.14284/170](https://doi.org/10.14284/170).
- 614 Intergovernmental Oceanographic Commission of UNESCO, Ocean Biodiversity Information System (OBIS), obis.org, accessed 2025-11-06.
- 615 iNaturalist, <https://www.inaturalist.org>, accessed 2025-11-06.
- 616 A. Rutz, M. Sorokina, J. Galgonek, D. Mietchen, E. Willighagen, A. Gaudry, J. G. Graham, R. Stephan, R. Page, J. Vondrasek, C. Steinbeck, G. F. Pauli, J. L. Wolfender, J. Bisson and P. M. Allard, *eLife*, 2022, **11**, e70780, DOI: [10.7554/eLife.70780](https://doi.org/10.7554/eLife.70780).
- 617 A. Rutz, D. Probst, C. Aguilar, D. Y. Akiyama, F. Alberti, H. E. Augustijn, N. E. Avalon, C. Beemelmans, H. B. Barbieri, F. Biermann, A. J. Bridge, E. C. Giron, R. Cox, M. Crusemann, P. M. D'Agostino, M. Feuermann, J. Gerke, K. G. Garcia, J. E. Holme, J. Y. Hwang, R. Iacovelli, J. C. Jeronimo Barbosa, N. Kaur, M. Klapper, A. M. Kohler, A. Korenskaia, N. Kubach, B. T. Lee, C. Loureiro, S. Mantri, S. Narula, D. Meijer, J. C. Navarro-Munoz, G. S. Nguyen, S. Paliyal, M. Panghal, L. Rao, S. Sieber, N. Sokolova, S. T. Sowa, J. Szenei, B. R. Terlouw, H. G. Weddeling, J. Yu, N. Ziemert, T. Weber, K. Blin, J. J. J. van der Hooft, M. H. Medema and M. M. Zdouc, *Nucleic Acids Res.*, 2025, gkaf969, DOI: [10.1093/nar/gkaf969](https://doi.org/10.1093/nar/gkaf969).

