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ARTICLE

Structure–activity relationship for calantheside, a potential hair-growth stimulant with an indole 2-*S*,3-*O*-bis-glucoside structure. Part 1: Role of two glucoside moieties in promoting HFDPc proliferationReceived 00th January 20xx,
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The development of new and effective drugs for treating androgenetic alopecia (AGA) remains challenging. Minoxidil and finasteride, two drugs currently used to treat AGA, were originally developed for treating hypertension and prostate disease, respectively. Therefore, the pharmacological effects of these drugs may be double-edged in patients with AGA. Calantheside (**1**) is an indole 2-*S*,3-*O*-bis-glucoside isolated from the *Calanthe* genus. Compared to minoxidil, **1** induces significantly higher proliferation of human hair follicle dermal papilla cells (HFDPcs). This study aimed to identify the key structures responsible for the activity of **1**. To this end, calantheside derivatives were synthesised and evaluated. Analogues (**9b**, **9d–9f**), in which the 3-*O*-glucoside moiety of **1** was replaced with different glycosides, exhibited minimal activity. In contrast, analogues (**8c–8f**), in which the 2-*S*-thioglycoside unit of **1** was substituted with various thioglycosides, demonstrated potent activity comparable to that of **1**. These results suggest that 3-*O*-glucoside is an essential structural feature for activity. The 2-*S*-ethylated derivative (**8g**) exhibited a complete loss of activity. Similarly, compounds **10a** and **10b**, in which all the hydroxyl groups of the sugar residue of **1** were ester-protected, also exhibited a complete loss of activity. Therefore, highly polar sugar structures are required at the 2- and 3-positions. Collectively, the findings of this initial evaluation of the structure–activity relationship (SAR) provide valuable insights for expanding the chemical space for the future development of AGA treatments.

Introduction

Androgenetic alopecia (AGA), also known as male pattern hair loss (MPHL) or female pattern hair loss (FPHL), is the most common form of hair loss worldwide. AGA affects up to 80% of men and 50% of women during their lifetime, and its prevalence increases with age.¹ Hair loss can significantly impair patients' quality of life, reduce self-esteem, and elevate daily stress levels, potentially leading to psychological disorders.^{2–4} Currently, only two drugs, minoxidil and finasteride, are approved by the US Food and Drug Administration (FDA) for the treatment of AGA. However, both were originally developed for unrelated indications: minoxidil for hypertension and finasteride for prostate disorders.^{5,6} Consequently, these drugs can exert double-edged pharmacological actions in patients with AGA.^{5,6} Topical minoxidil is associated with localised side-effects such as scalp itching, rash, irritation, dryness, and dandruff.⁷ In some cases, its vasodilatory properties may also lead to systemic adverse effects, including headaches, dizziness, palpitations, and peripheral oedema.⁸ Studies indicate that finasteride, a 5 α -reductase inhibitor, can cause sexual dysfunction in men, including reduced libido and erectile

dysfunction.⁹ Moreover, its use in women is limited owing to potential teratogenicity.¹⁰ Thus, the development of new therapeutic agents for AGA remains an ongoing challenge.^{11–13} Identifying alternative lead compounds from natural sources has emerged as a promising strategy for addressing these issues.^{14–17}

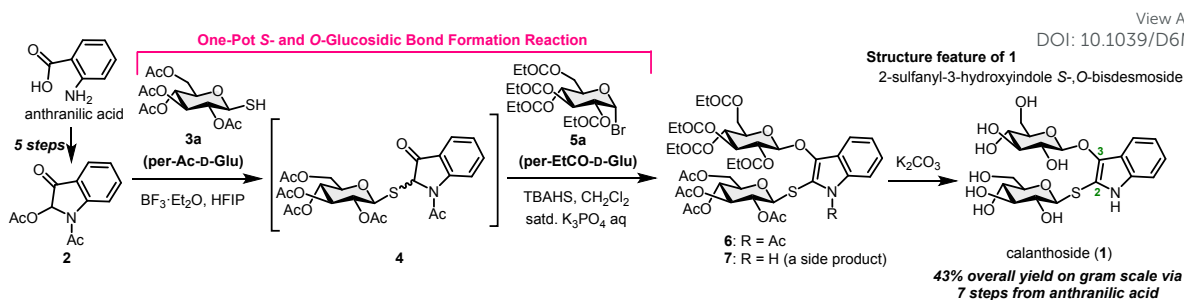
In 1998, Yoshikawa et al. isolated calantheside (**1**), an indole alkaloid, from the plants *Calanthe discolor* LINDL. (known as "ebine" in Japanese) and *C. liukiensis* SCHLTR.^{18–19} Compound **1** induces potent proliferation of human hair follicle dermal papilla cells (HFDPcs), exhibiting a level of activity superior to that of minoxidil.²⁰ Moreover, studies have shown that its mechanism of action does not involve 5 α -reductase inhibition,²⁰ suggesting that **1** can prospectively serve as an alternative to finasteride for treating androgenetic alopecia (AGA) in both sexes. Structurally, compound (**1**) features a unique indole 2-*S*,3-*O*-bis-glucoside in which two β -glucose units are attached to the 2-sulfanyl-3-hydroxyindole framework *via* *S*- and *O*-glycosidic bonds. The novel architecture and attractive biological activity of **1** make it particularly intriguing. However, despite its promising profile, no synthetic route to **1** has been established, and no structure–activity relationship (SAR) studies have been conducted to date. Recently, we accomplished the first total synthesis of **1** on a preparative scale.²¹ Building on this achievement, a straightforward synthetic strategy (Scheme 1)

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Scheme 1 Overview of the synthetic route to calanthiside (**1**) using one-pot *S*,*O*-glycosidic bond-formation reaction. TBAHS = tetrabutylammonium hydrogen sulfate.

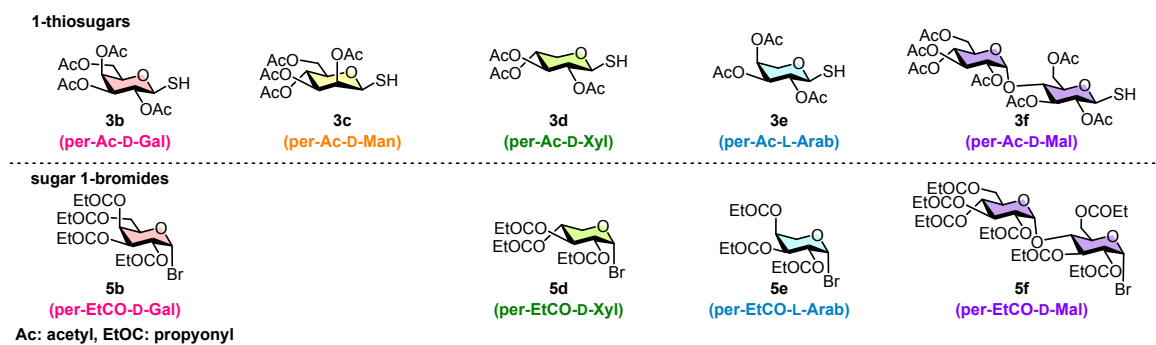


Fig 1 Structure of synthons for one-pot *S*,*O*-glycosidic bond-formation reaction of **2**.

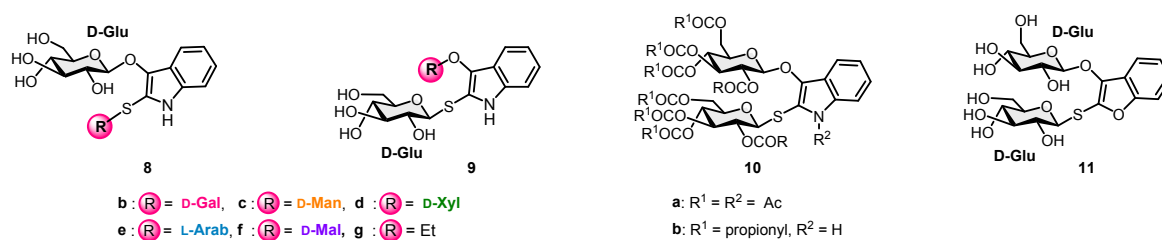


Fig 2 Designed calanthiside derivatives (**8a–8e**, **9a–9e**, **10a**, **10b** and **11**).

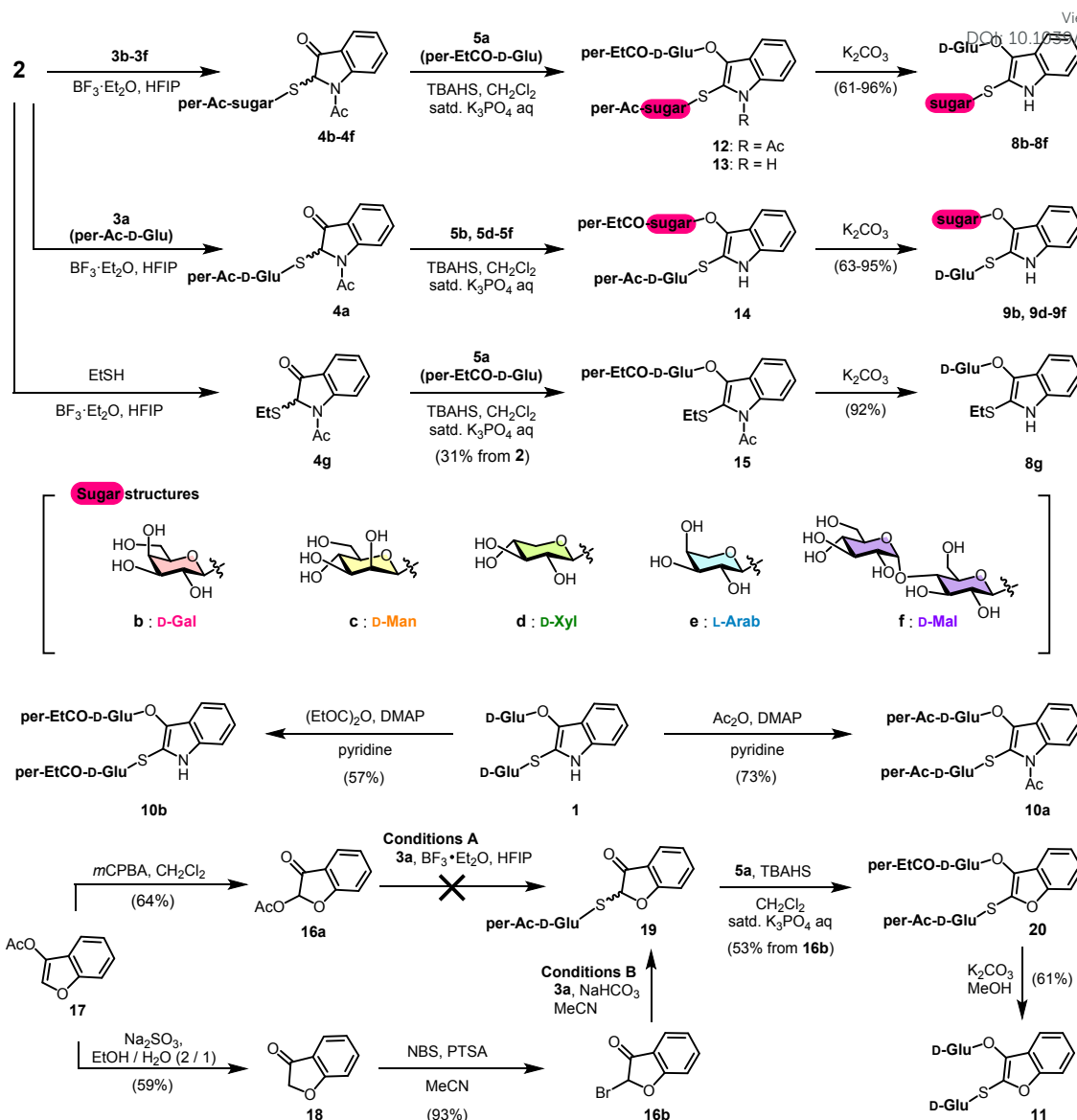
was established for accessing structurally related analogues, either through replacement of the sugar-type intermediates (**3a** and **5a**) used in the one-pot *S*,*O*-glycosidic bond-forming sequence (**2** → **4** → **6**) with alternative sugar derivatives (**3b–3f**, **5b**, and **5d–5f**; see Figure 1), or through the use of isosteres of **2** in which an oxygen atom replaces the nitrogen atom. The structural modification of **1** represents a promising approach for discovering novel lead compounds for the treatment of AGA. In this study, we report the first evaluation of the SAR of **1**. To examine the influence of structural variations of the sugar moiety on the biological activity, a series of analogues (**8b–8f**, **9b**, and **9d–9f**; see Figure 2) was synthesised, in which the *D*-glucose moiety attached to either the sulfur or oxygen atom of **1** was replaced with different sugar units. In addition, analogue **8g**, bearing an ethyl group instead of the *D*-glucose unit on the oxygen atom of **1**, was prepared. Per-*O*-acetylated derivatives (**10a** and **10b**) were also synthesised to evaluate the role of the hydroxyl groups in the sugar moiety. The effects of the synthesised compounds on HFDPCs proliferation were evaluated. In addition to these biological evaluations, benzofuran (**11**) was synthesised and evaluated. Although **11** was virtually inactive, during its synthesis, a distinctly different

mechanism of bond formation at the 2-position of intermediate **16b** was revealed, compared to the *S*_N1-type mechanism²¹ observed for intermediate **2** in the synthesis of compounds **1**, **8b–8f**, **9b**, and **9d–9f**, which is also discussed herein.

Results and discussion

Following our previously reported method,²¹ indolinone **2**, prepared from anthranilic acid, was treated sequentially with peracylated 1-thiosugars **3b–3f** and peracylated glucose 1-bromide **5a** to afford a mixture of *S*,*O*-bisdesmosides **12b–12f** and their de-*N*-deacylated products **13b–13f** in good yields. Without separating these two components, the acyl groups were subjected to methanolysis in the presence of K_2CO_3 to furnish 3-*O*-glucoside-type analogues **8b–8f** in good yields. Conversely, when compound **2** was first reacted with peracylated 1-thioglucose **3a**, followed by 3-*O*-glycosylation of the resulting 2-*S*-glucoside **4a** with **5b**, **5d**, **5e**, and **5f**, the corresponding *S*,*O*-bisdesmosides **14b**, **14d**, **14e**, and **14f** were obtained in good yields. These compounds were then converted to the target analogues **9b**, **9d**, **9e**, and **9f** under methanolytic conditions similar to those used in the preparation of **8b–8f**. In





Scheme 2 Synthesis of derivatives (**8–11**).

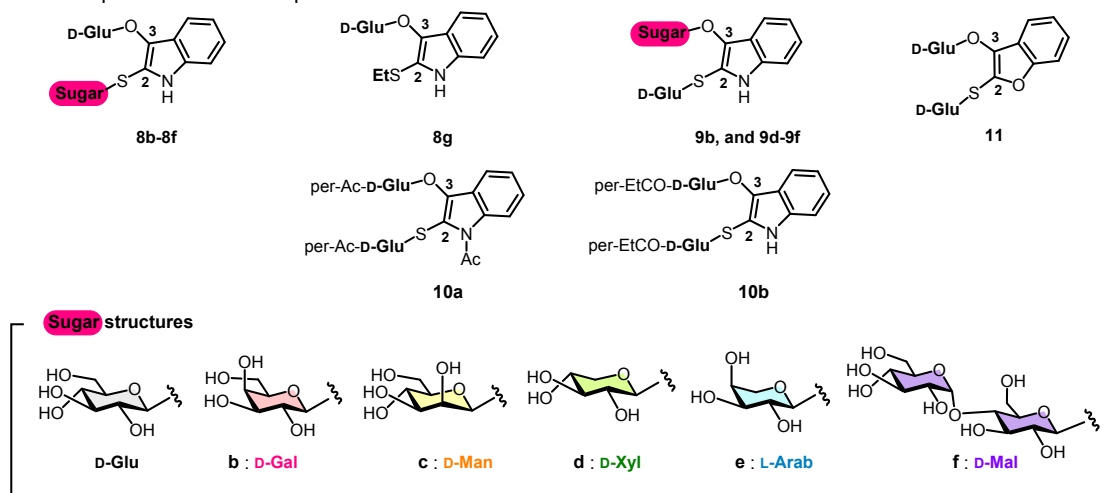
the positive-mode electrospray ionization (ESI) mass spectra, the quasimolecular ion peaks $[M+Na]^+$ at m/z 512, 512, 482, 482, 674, 512, 482, 482, and 674 correspond to products **8b**, **8c**, **8d**, **8e**, **8f**, **9b**, **9d**, **9e**, and **9f**, respectively, consistent with the proposed molecular structures. The 1H and ^{13}C NMR spectra of these analogues were generally similar, except for differences in the signals arising from the hydrogen and carbon atoms of the sugar residues. For instance, the ^{13}C NMR spectra of series **8** and **9** showed eight signals assignable to the carbons of the 2-sulfanyl-2-hydroxyindole ring in the range of δ_c 110–143 ppm. Additionally, signals arising from the anomeric carbons, supporting their *S*- and *O*-glycoside structures, appeared at δ_c 87 ppm and δ_c 105 ppm, respectively. Furthermore, the vicinal coupling constants ($J_{1,2} = 9.0$ – 9.7 Hz for the *S*-glycoside moiety and $J_{1,2} = 7.2$ – 7.9 Hz for the *O*-glycoside moiety) for the anomeric proton H-1 and the H-2 methine protons in these analogues (except for the signal of H-1 in the 2-*S*-mannoside structure of analogue **8c**) indicated that both sugar units are

linked via β -glycosidic bonds. The spectrum of compound **8c** exhibited a signal of the anomeric proton H-1 at δ_H 4.98, with a small coupling constant ($J_{1,2} \approx 0$ Hz), characteristic of the β -configuration of the anomeric carbon in the mannose unit. The β -stereochemistry at the anomeric centre was confirmed by nuclear Overhauser effect (NOE) correlations among the three axial protons (H-1, H-3, and H-5) of the mannose residue. Additionally, the related analogue **8g** was synthesised *via* a three-step sequence comprising *S*-glucosylation with ethyl mercaptan, *O*-glucosidation of the resulting intermediate **4g**, and deprotection of compound **15** under basic conditions. The ^{13}C NMR spectrum of **8g** was highly similar to those of **8b–8f**, except for the observation of signals at δ_c 15.5 and 31.0 ppm, arising from the ethyl group carbons in **8g**, which appeared in place of the sugar residues in **8b–8f**. Calanthoside (**1**), pre-prepared in large quantities,²¹ was acylated with either acetic or propionic anhydride in pyridine in the presence of a catalytic amount of 4-dimethylaminopyridine (DMAP) to produce **10a**



Table 1. Effects of compounds **8–11** on the proliferation of HFDPCs

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entry	compound	Relative activity level (% of control) ^{a)}				
		0 μ M	12.5 μ M	25 μ M	50 μ M	100 μ M
1	8b (2S-Gal;3O-Glu)	100.0 \pm 6.9	89.1 \pm 2.6	94.9 \pm 6.4	95.9 \pm 4.0	99.5 \pm 1.9
2	8c (2S-Man;3O-Glu)	100.0 \pm 1.3	103.6 \pm 5.2	110.5 \pm 1.2	119.0 \pm 0.7**	115.5 \pm 5.4*
3	8d (2S-Xyl;3O-Glu)	100.0 \pm 6.3	105.6 \pm 5.4	106.2 \pm 3.9	105.3 \pm 4.3	120.8 \pm 2.7*
4	8e (2S-Arab;3O-Glu)	100.0 \pm 4.6	104.4 \pm 3.4	108.1 \pm 0.8	134.9 \pm 8.8**	138.0 \pm 2.7**
5	8f (2S-Mal;3O-Glu)	100.0 \pm 1.8	101.8 \pm 0.3	103.1 \pm 2.7	109.6 \pm 6.1	122.4 \pm 3.5**
6	8g (2S-Et;3O-Glu)	100.0 \pm 4.5	87.7 \pm 6.6	99.4 \pm 6.7	100.6 \pm 6.5	106.1 \pm 2.3
7	9b (2S-Glu;3O-Gal)	100.0 \pm 4.4	96.2 \pm 6.4	97.5 \pm 5.2	96.6 \pm 4.5	98.5 \pm 4.3
8	9d (2S-Glu;3O-Xyl)	100.0 \pm 10.4	100.4 \pm 9.5	106.1 \pm 9.9	100.4 \pm 8.6	101.1 \pm 0.5
9	9e (2S-Glu;3O-Arab)	100.0 \pm 16.3	108.0 \pm 19.7	117.7 \pm 15.3	111.7 \pm 14.3	105.5 \pm 2.8
10	9f (2S-Glu;3O-Mal)	100.0 \pm 4.8	101.4 \pm 8.7	94.3 \pm 3.8	95.0 \pm 3.3	79.9 \pm 1.8
11	10a (2S-Ac-Glu;3O-Ac-Glu)	100.0 \pm 7.2	76.6 \pm 4.8**	50.6 \pm 3.3**	50.9 \pm 0.3**	66.4 \pm 1.5**
12	10b (2S-ProGlu;3O-Pro-Glu)	100.0 \pm 5.5	98.1 \pm 5.4	97.3 \pm 4.7	119.0 \pm 4.5	108.6 \pm 3.4
13	calanthoside (1 , synthetic) ^{b)}	100.0 \pm 2.9	134.0 \pm 5.2**	138.9 \pm 4.1**	139.6 \pm 8.6**	125.2 \pm 2.8
14	11	100.0 \pm 7.4	89.0 \pm 5.7	87.5 \pm 4.6	99.1 \pm 17.3	90.4 \pm 3.3**
15	minoxidil sulfate ^{c)}	100.0 \pm 1.7	108.3 \pm 5.5	101.5 \pm 4.4	107.5 \pm 2.7	112.4 \pm 3.3

a) Each value represents the mean \pm S.E.M. ($N = 3$). Significantly different from the control, * $p < 0.05$, ** $p < 0.01$. b) The activity of natural calanthoside (**1**) had been previously reported.²⁰ However, to compare the activity of candidates **8–11**, we re-examined the activity of the synthetic calanthoside (**1**). c) Minoxidil sulfate was purchased from FUJIFILM Wako Pure Chemical Corporation (Osaka, Japan).

and **10b**, with yields of 73 and 57%, respectively (Scheme 3). In the ¹³C NMR spectra of **10a** and **10b**, the signals of the sugar carbons (C2, C3, C4, C6) shifted to a lower field compared to those of **1**, providing strong evidence that the hydroxyl groups were fully acylated. The quasimolecular [M+Na]⁺ peaks observed at m/z 868 and 938 in the ESI mass spectra of **10a** and **10b**, respectively, are consistent with the molecular weights of the corresponding acylated compounds. Subsequently, benzofuran-type analogue **11** was synthesised as follows: Using a literature procedure,²² key intermediate 3-acetoxybenzofuran (**17**) was prepared from salicylic acid. After treatment of **17** with *m*-chloroperoxybenzoic acid (*m*CPBA), the resulting oxidation product **16a** was subjected to the *S*-glycosidic bond-formation

reaction under the same conditions used in the synthesis of **4a**. Interestingly, the desired intermediate **19** was not observed, suggesting that the S_N1 -type mechanism involving electron donation from nitrogen, which proceeds with compound **4a**,^{23–24} does not occur in the case of **16a**. To overcome this limitation, compound **16b**, in which the leaving group was changed from an acetate moiety to a bromine moiety, was synthesised from **17** in two steps and subsequently reacted with the thiolate anion generated from **3a** under basic conditions to afford **19**. The successful synthesis of this compound supports the conclusion that the reaction proceeds via an S_N2 -type mechanism. Compound **19** was found to be rather unstable and was, therefore, immediately *O*-glucosylated with **5a** to afford



the fully acylated analogue **20** in 53% yield *via* a two-step reaction from **16b**. Finally, methanolysis of **20** furnished the desired product **11** in 61% yield. The spectra of compounds **11** exhibited quasimolecular ion $[M+Na]^+$ peaks at m/z 905; the molecular formula of these compounds, $C_{20}H_{50}O_{20}SNa$, was confirmed by high-resolution ESI mass spectrometry (HRESIMS). The ^{13}C NMR spectra of **11** closely match those of calanthoside (**1**), strongly suggesting that **11** is an isostere of **1** (Scheme 2).

With all thirteen target compounds (**8b–8g**, **9b**, **9d–9f**, **10a**, **10b**, and **11**) in hand, their potency as activators of HFDPs proliferation was examined and compared with that of **1** and minoxidil sulfate, as shown in Table 1. Although the biological activity of natural calanthoside (**1**) has been previously reported,²⁰ its synthetic counterpart was reevaluated in the present study as a reference standard. Compound **1** exhibited potent activity in promoting the proliferation of HFDPs at concentrations ranging from 12.5 μ M to 50 μ M, with activity levels of $134.0\% \pm 5.2\%$ to $139.6\% \pm 8.6\%$ (entry 13) relative to that of the control. Although the proliferation-promoting effect of **1** was somewhat reduced at 100 μ M compared with that at 50 μ M, the compound still showed a favourable effect, with a proliferation rate of $125.0 \pm 2.8\%$ relative to that of the control. Furthermore, the efficacy of **1** was confirmed to be significantly higher than that of minoxidil sulfate (entry 15). Subsequently, proliferation activity tests were conducted on the series of 3-*O*-glucoside-type analogues **8b–8g**. Unlike **8b** and **8g**, four analogues (**8c**, **8d**, **8e**, and **8f**) were effective activators, exhibiting proliferation rates of $115.5\% \pm 5.4\%$, $120.8\% \pm 2.7\%$, $138.0\% \pm 2.7\%$, and $122.4\% \pm 3.5\%$ versus that of the control at 100 μ M, respectively (entries 2–5). Therefore, based on the results for compound **8d** (entry 3), the hydroxymethyl group in the thioglucoside moiety of **1** is plausibly not essential for activity. Consistent with this structural insight, removal of this group from analogue **8b** (entry 1) markedly increased its activity, yielding analogue **8e**, which exhibited a proliferation-promoting effect of $138.0\% \pm 2.7\%$ versus that of the control, surpassing that of **1** ($125.0\% \pm 2.8\%$ relative to the control). These findings suggest that in the absence of a hydroxymethyl group at the 5-position, the stereochemistry of the hydroxyl groups at the 2- or 4-position of the sugar moiety (axial or equatorial) is not a determining factor for proliferative activity. However, the activity of **8g** was almost completely lost, suggesting that highly polar pyranoside moieties at the 2-position of the indole core are essential for maintaining the activity (entry 6). In contrast, among the 2-thioglucoside-type analogues (**9b**, **9d**, **9e**, and **9f**), only **9e** retained slight activity. The remaining compounds (**9b**, **9d**, and **9f**) showed no significant proliferative effect, even at a concentration of 100 μ M (entries 7–10, $79.9\% \pm 1.8\%$ to $101.1\% \pm 0.5\%$ versus the control). These results indicate that the 3-*O*-glucoside moiety is requisite for the activity of this series. The per-acylated derivatives **10a** and **10b** exhibited lower inhibitory activities than the parent compound **1** (entries 10 and 11). Notably, **10a** exhibited markedly reduced activity and clear toxicity towards HFDPs (entry 11). These findings indicate that the polarity of the hydroxyl groups of the sugar moieties is essential for the biological activity of the compounds. Finally, compound **11**

exhibited a markedly diminished effect on cell proliferation compared to **1** and also showed mild toxicity towards HFDPs. This result suggests that substituting the nitrogen atom with an oxygen atom is an unfavourable structural modification.

Conclusions

Calanthoside (**1**) is an indole 2-*S*,3-*O*-bis-glucoside found in traditional medicinal plants. Because of its potential to promote hair growth, it is considered a promising lead compound for the development of novel pharmaceuticals to treat androgenetic alopecia (AGA). This study investigated the relationship between the two glucoside structures and the proliferative activity of HFDPs as an initial exploration of the structure–activity relationship. Compounds **8c**, **8d**, **8e**, and **8f** (excluding **8b**) exhibit strong proliferation-promoting effects. In contrast, the latter group (**9b**, **9d**, **9e**, and **9f**) shows almost no activity. These findings indicate that the 3-*O*-glucoside structure is essential for promoting proliferation. Furthermore, the inactivity of the 2-*S*-ethylated derivative **8g**, which is related to **8b–8f**, suggests that introducing a hydrophobic substituent at the 2-position of the sulfur atom is undesirable. Thus, it was determined that a highly polar pyranoside unit at the 2-*S* position of the indole core is required for achieving activity. Additionally, no activity was detected for the peracylated analogues **10a** and **10b**, indicating that the polarity of the sugar moiety is crucial for the manifestation of activity. Taken together, these findings provide a foundation for the further development of calanthoside derivatives. In our laboratory, structure–activity relationship studies of calanthoside (**1**) are currently being advanced, with particular emphasis on substitutions on the indole ring. In addition, further studies including toxicity profiling, transcriptomic analysis, and combination studies with reference compounds such as minoxidil are planned in order to further elucidate the pharmacological properties of these derivatives.

Experimental section

Chemistry

Optical rotations were determined with a digital polarimeter. IR spectra were recorded on a FT-IR spectrophotometer. NMR spectra were recorded on FT-NMR spectrometers (1H , 800 MHz; ^{13}C , 200 MHz). Tetramethylsilane (TMS) was used as an internal standard for 1H NMR measurements in $CDCl_3$, whereas ^{13}C NMR measurements utilized the solvent signal (77.0 ppm) of $CDCl_3$ for this purpose. When CD_3OD or $DMSO-d_6$ was used for the measurement of 1H and ^{13}C NMR spectra, solvent signals [CD_3OD (δ_H 3.30 ppm and δ_C 49.0 ppm); $DMSO-d_6$ (δ_H 2.49 ppm and δ_C 39.7 ppm)] were used as standards for 1H and ^{13}C NMR measurements. Sodium 2,2-dimethyl-2-silapentane-5-sulfonate (DSS) was used as an external standard for the measurement of 1H and ^{13}C NMR spectra in D_2O . 1D NMR peak assignments were confirmed by COSY, HSQC, and HMBC spectra. High-resolution mass spectra were recorded on an Orbitrap mass spectrometer (ESI). All organic extracts were dried over anhydrous Na_2SO_4 prior to evaporation. Column chromatography was performed over silica gel (45–106 μ m).



Mixtures of each **12b** and **13b**, **12c** and **13c**, **12d** and **13d**, **12e** and **13e**, and **12f** and **13f**, as well as compounds **14b**, **14d**, **14e**, and **14f**, were prepared according to the literature procedures²¹ prior to methanolysis, which was then carried out as described below.

3-(β-D-Glucopyranosyloxy)-1H-indol-2-yl 1-thio-β-D-galactopyranoside (8b). The mixture of **12b** (207 mg, 0.22 mmol), and **13b** (198 mg, 0.22 mmol) was treated with K₂CO₃ (6 mg, 0.044 mmol) in MeOH (5 mL) at room temperature (rt) for 4 h. After the reaction mixture was condensed *in vacuo*, the residue was purified by column chromatography (CHCl₃/MeOH = 20/1) to give the title compound (**8b**, 179 mg, 0.37 mmol, 83%) as a pale yellow microcrystalline solid. Mp 138–139 °C. [α]_D²⁵ –10.0 (*c* = 1.0, MeOH). IR (KBr): 3366, 2920, 2853, 1449, 1344, 1240, 1134, 1074, 1011, 856, 824 cm⁻¹. ¹H NMR (800 MHz, D₂O) δ: 3.28 (1H, ddd, *J* = 9.6, 5.1, 2.4, OGU H-5), 3.44 (1H, dd, *J* = 9.3, 9.3, OGU H-4), 3.46 (1H, dd, *J* = 9.7, 9.7, SGal H-2), 3.48 (1H, dd, *J* = 9.6, 9.6, OGU H-3), 3.51–3.54 (2H, m, SGal H-3 and SGal H-5), 3.52 (1H, dd, *J* = 7.9, 9.6, OGU H-2), 3.60 (1H, dd, *J* = 11.7, 4.3, SGal H-6a), 3.66 (1H, dd, *J* = 12.5, 5.1, OGU H-6a), 3.67 (1H, dd, *J* = 11.7, 7.9, SGal H-6b), 3.74 (1H, dd, *J* = 12.5, 2.4, OGU H-6b), 3.82 (1H, dd, *J* = 3.3, 1.0, SGal H-4), 4.58 (1H, d, *J* = 9.7, SGal H-1), 5.02 (1H, d, *J* = 7.9, OGU H-1), 7.03 (1H, ddd, *J* = 8.0, 6.9, 1.0, H-5), 7.14 (1H, ddd, *J* = 8.2, 6.9, 1.2, H-6), 7.28 (1H, br d-like, *J* = *ca.* 8.2, 1.0, H-7), 7.62 (1H, dd, *J* = 8.0, 1.2, H-4). ¹³C NMR (200 MHz, D₂O) δ: 60.5 (OGU C-6), 61.2 (SGal C-6), 68.8 (SGal C-4), 69.3 (OGU C-4), 69.6 (SGal C-2), 73.4 (OGU C-2), 73.9 (SGal C-3), 75.6 (OGU C-3), 76.2 (SGal C-5), 79.3 (OGU C-5), 88.3 (SGal C-1), 104.7 (OGU C-1), 111.9 (C-7), 113.4 (C-2), 117.9 (C-4), 120.0 (C-3a), 120.1 (C-5), 123.9 (C-6), 134.5 (C-7a), 139.0 (C-3). HRMS (ESI) *m/z*: [M+Na]⁺ Calcd for C₂₀H₂₇NNaO₁₁S 512.1197; Found 512.1197.

3-(β-D-Glucopyranosyloxy)-1H-indol-2-yl 1-thio-β-D-mannopyranoside (8c). Following the method similar to that used for the preparation of **8b**, the mixture of **12c** (143 mg, 0.16 mmol) and **13c** (137 mg, 0.16 mmol) was treated with K₂CO₃ in MeOH to give the title compound (**8c**, 139 mg, 0.28 mmol, 88%) as a pale yellow microcrystalline solid. Mp 154–155 °C. [α]_D²⁵ –12.0 (*c* = 1.0, MeOH). IR (KBr): 3632, 2920, 2881, 1449, 1343, 1240, 1097, 1068 cm⁻¹. ¹H NMR (800 MHz, D₂O) δ: 3.27 (1H, dd, *J* = 9.4, 6.5, 2.4, SMan H-5), 3.36 (1H, ddd, *J* = 9.4, 5.1, 2.3, OGU H-5), 3.54 (1H, dd, *J* = 9.4, 9.4, OGU H-4), 3.59 (1H, dd, *J* = 9.2, 9.2, OGU H-3), 3.60–3.66 (3H, m, OGU H-2, SMan H-3 and H-4), 3.73 (1H, dd, *J* = 12.4, 6.5, SMan H-6a), 3.75 (1H, dd, *J* = 12.4, 5.1, OGU H-6a), 3.82 (1H, dd, *J* = 12.4, 2.3, OGU H-6b), 3.84 (1H, dd, *J* = 12.4, 2.4, SMan H-6b), 4.22 (1H, d, *J* = 2.4, SMan H-2), 4.98 (br s-like, man H-1), 5.00 (1H, d, *J* = 7.9, OGU H-1), 7.13 (1H, ddd, *J* = 7.9, 6.9, 1.0, H-5), 7.25 (1H, ddd, *J* = 8.2, 6.9, 1.2, H-6), 7.39 (1H, dd, *J* = 8.2, 1.0, H-7), 7.72 (1H, dd, *J* = 6.9, 1.0, H-4). ¹³C-NMR (200 MHz, D₂O) δ: 60.4 (SMan C-6), 61.0 (OGU C-6), 66.5 (SMan C-4), 69.2 (OGU C-4), 71.6 (SMan C-2), 73.3 (OGU C-2), 73.6 (SMan C-3), 75.6 (OGU C-3), 76.1 (SMan C-5), 80.3 (OGU C-5), 87.5 (C-1 SMan), 104.7 (OGU C-1), 111.7 (C-7), 114.9 (C-2), 117.7 (C-4), 119.7 (C-3a), 119.9 (C-5), 123.7 (C-6), 134.3

(C-7a), 138.2 (C-3). HRMS (ESI) *m/z*: [M+Na]⁺ Calcd for C₂₀H₂₇NNaO₁₁S 512.1195; Found 512.1197. DOI: 10.1039/D6MD00273K

3-(β-D-Glucopyranosyloxy)-1H-indol-2-yl 1-thio-β-D-xylopyranoside (8d). Following the method similar to that used for the preparation of **8b**, the mixture of **12d** (51 mg, 60 μmol) and **13d** (49 mg, 60 μmol) was treated with K₂CO₃ in MeOH to give the title compound (**8d**, 53 mg, 0.115 mmol, 96%) as a pale yellow microcrystalline solid. Mp 170–171 °C. [α]_D²⁵ –6.6 (*c* = 1.0, MeOH). IR (KBr): 3385, 2922, 1740, 1653, 1341, 1238, 1098, 1051, 1009, 897 cm⁻¹. ¹H NMR (800 MHz, D₂O) δ: 3.13–3.17 (2H, m, SXyl H-2 and H-5a), 3.27 (1H, ddd, *J* = 9.5, 5.0, 2.3, OGU H-5), 3.34 (1H, dd, *J* = 9.0, 9.0, SXyl H-3), 3.41–3.50 (3H, m, OGU H-3, H-4 and SXyl H-4), 3.54 (1H, dd, *J* = 9.2, 7.9, OGU H-2), 3.66 (1H, dd, *J* = 12.4, 5.0, OGU H-6a), 3.74 (1H, dd, *J* = 12.4, 2.3, OGU H-6b), 3.90 (1H, dd, *J* = 11.5, 5.3, SXyl H-5b), 4.47 (1H, d, *J* = 9.5, SXyl H-1), 4.90 (1H, d, *J* = 7.9, OGU H-1), 7.00 (1H, ddd, *J* = 8.1, 7.0, 0.9, H-5), 7.27 (1H, ddd, *J* = 8.3, 7.0, 1.2, H-6), 7.35 (1H, br d-like, *J* = *ca.* 8.3, H-7), 7.69 (1H, br d-like, *J* = *ca.* 8.1, H-4). ¹³C-NMR (200 MHz, D₂O) δ: 60.5 (OGU C-6), 69.0(2C)/69.3 (OGU C-4, SGLU C-4 and C-5), 72.0 (SXyl H-2), 73.4 (OGU H-2), 75.7 (OGU H-3), 76.3 (OGU H-5), 77.1 (SXyl C-3), 87.9 (SXyl C-1), 104.6 (OGU C-1), 112.0 (C-7), 112.6 (C-2), 117.9 (C-4), 119.6 (C-3a), 120.1 (C-5), 124.0 (C-6), 134.6 (C-7a), 139.5 (C-3). HRMS (ESI) *m/z*: [M+Na]⁺ Calcd for C₁₉H₂₅NNaO₁₀S 482.1091; Found 482.1091.

3-(β-D-Glucopyranosyloxy)-1H-indol-2-yl 1-thio-β-D-arabinopyranoside (8e). Following the method similar to that used for the preparation of **8b**, the mixture of **12e** (94 mg, 0.110 mmol) and **13e** (87 mg, 0.108 mmol) was treated with K₂CO₃ in MeOH to give the title compound (**8e**, 84 mg, 0.183 mmol, 84%) as a pale yellow microcrystalline solid. Mp 150–151 °C. [α]_D²⁵ –23.2 (*c* = 1.0, MeOH). IR (KBr): 3524, 2922, 2855, 1701, 1458, 1240, 1076 cm⁻¹. ¹H NMR (800 MHz, D₂O) δ: 3.34 (1H, ddd, *J* = 7.2, 4.9, 2.2, OGU H-5), 3.36–3.58 (3H, m, OGU H-4, SARab H-3 and H-5a), 3.60–3.65 (3H, m, OGU H-2 and H-3, SARab H-2), 3.75 (1H, dd, *J* = 12.5, 5.0, OGU H-6a), 3.82 (1H, dd, *J* = 12.5, 2.2, OGU H-6b), 3.91–3.99 (2H, m, SARab H-4 and H-5b), 4.56 (1H, d, *J* = 9.0, SARab H-1), 4.98 (1H, d, *J* = 7.9, OGU H-1), 7.12 (1H, dd, *J* = 7.8, 7.8, H-5), 7.23 (1H, dd, *J* = 7.8, 7.8, H-6), 7.37 (1H, d, *J* = 7.8, H-7), 7.71 (1H, d, *J* = 7.8, H-4). ¹³C NMR (200 MHz, D₂O) δ: 60.3 (OGU C-6), 68.2 (SARab C-4), 69.2 (OGU C-4), 69.5 (SARab C-5), 69.6 (SARab C-2), 73.0 (SARab C-3), 73.3 (OGU C-2), 75.5 (OGU C-3), 76.1 (OGU C-5), 88.3 (SARab C-1), 104.6 (OGU C-1), 111.8 (C-7), 113.3 (C-2), 117.7 (C-4), 119.5 (C-3a), 119.9 (C-6), 123.8 (C-5), 134.4 (C-7a), 139.0 (C-3). HRMS (ESI) *m/z*: [M+Na]⁺ Calcd for C₁₉H₂₅NNaO₁₀S 482.1091; Found 482.1092.

3-(β-D-Glucopyranosyloxy)-1H-indol-2-yl 1-thio-β-D-maltopyranoside (8f). Following the method similar to that used for the preparation of **8b**, the mixture of **12f** (51 mg, 0.42 μmol) and **13f** (49 mg, 0.41 μmol) was treated with K₂CO₃ in MeOH to give the title compound (**8f**, 34 mg, 0.51 mmol, 61%) as a pale yellow microcrystalline solid. Mp 150–151 °C. [α]_D¹⁹ +3.6 (*c* = 1.0, MeOH). IR (KBr): 3395, 2920, 2888, 1616, 1449, 1342, 1240, 1148, 1059, 1032, 916 cm⁻¹. ¹H NMR (800 MHz, D₂O)



δ : 3.28 (1H, dd, $J = 9.4, 9.4$), 3.40–3.43 (2H, m), 3.48 (1H, ddd, $J = 9.9, 5.0, 2.3$), 3.55–3.58 (3H, m), 3.61 (1H, dd, $J = 9.2, 9.2$), 3.65–3.71 (3H, m), 3.74–3.81 (4H, m), 3.83–3.90 (3H, m), 4.60 (1H, d, $J = 9.7$, SMal H-1), 5.03 (1H, d, $J = 7.8$, OGlu H-1), 5.33 (1H, d, $J = 4.0$, SMal H-1'), 7.14 (1H, br dd-like, $J = ca. 8.2, 6.9$, H-5), 7.26 (1H, dd, $J = 8.4, 6.9, 1.2$, H-6), 7.41 (1H, br d-like, $J = ca. 8.4, H-7$), 7.76 (1H, dd, $J = 8.2, 1.2$, H-4). ^{13}C NMR (200 MHz, D_2O) δ : 60.5, 60.6, 60.7, 69.3, 69.4, 71.8, 71.9, 72.8, 72.9, 73.4, 75.6, 76.2, 76.6, 77.5, 78.6, 87.1 (SMal C-1), 99.8 (OGlu C-1), 104.6 (OMal C-1'), 111.9 (C-7), 112.5 (C-2), 117.9 (C-4), 119.5 (C-3a), 120.1 (C-5), 124.0 (C-6), 134.6 (C-7a), 139.5 (C-3). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{26}\text{H}_{37}\text{NNaO}_{16}\text{S}$ 674.1725; Found 674.1726.

3-(β -D-Galactopyranosyloxy)-1H-indol-2-yl 1-thio- β -D-glucopyranoside (9b). Following the method similar to that used for the preparation of **8b**, **14b** (150 mg, 0.170 mmol) was treated with K_2CO_3 in MeOH to give the title compound (**9b**, 83 mg, 0.155 mmol, 95%) as a pale red microcrystalline solid. Mp 159–160 °C. $[\alpha]_{\text{D}}^{25} +83.1$ ($c = 1.0$, MeOH). IR (KBr): 3337, 2930, 2859, 1748, 1715, 1636, 1506, 1429, 1340, 1240, 1067, 1048, 1026 cm^{-1} . ^1H NMR (800 MHz, CD_3OD) δ : 3.10 (1H, dd, $J = 9.5, 8.9$, SGlu H-2), 3.18 (1H, dd, $J = 8.9, 8.9$, SGlu H-4), 3.29 (1H, ddd, $J = 8.9, 5.7, 2.4$, SGlu H-5), 3.37 (1H, dd, $J = 8.9, 8.9$, SGlu H-3), 3.56 (1H, ddd, $J = 6.2, 6.2, 1.2$, OGal H-5), 3.58 (1H, dd, $J = 7.8, 3.4$, OGal H-3), 3.66 (1H, dd, $J = 12.0, 5.7$, SGlu H-6a), 3.76–3.77 (2H, br m, OGal H-6), 3.86 (1H, dd, $J = 12.0, 2.4$, SGlu H-6b), 3.87 (1H, dd, $J = 8.9, 7.8$, OGal H-2), 3.89 (1H, dd, $J = 3.4, 1.2$, OGal H-4), 4.41 (1H, d, $J = 9.5$, SGlu H-1), 4.86 (1H, d, $J = 7.8$, OGal H-1), 6.98 (1H, ddd, $J = 8.0, 7.0, 0.9$, H-5), 7.11 (1H, ddd, $J = 8.2, 7.0, 1.2$, H-6), 7.24 (1H, dd, $J = 8.2, 0.9$, H-7), 7.78 (1H, dd, $J = 8.0, 1.2$, H-4). ^{13}C NMR (200 MHz, CD_3OD) δ : 62.2 (OGal C-6), 62.6 (SGlu C-6), 70.2 (OGal C-4), 71.1 (SGlu C-4), 72.9 (OGal C-2), 73.6 (SGlu C-2), 74.8 (OGal C-3), 76.8 (OGal C-5), 79.1 (SGlu C-3), 81.9 (SGlu C-5), 89.0 (SGlu C-1), 107.2 (OGal C-1), 112.0 (C-7), 112.9 (C-2), 119.7 (C-4), 120.2 (C-5), 121.7 (C-3a), 124.2 (C-6), 136.3 (C-7a), 142.3 (C-3). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{20}\text{H}_{27}\text{NNaO}_{11}\text{S}$ 512.1197; Found 512.1191.

3-(β -D-Xylopyranosyloxy)-1H-indol-2-yl 1-thio- β -D-glucopyranoside (9d). Following the method similar to that used for the preparation of **8b**, **14d** (180 mg, 0.23 mmol) was treated with K_2CO_3 in MeOH to give the title compound (**9d**, 87 mg, 0.19 mmol, 83%) as a pale yellow microcrystalline solid. Mp 154–155 °C. $[\alpha]_{\text{D}}^{25} -6.7$ ($c = 1.0$, MeOH). IR (KBr): 3337, 2930, 2859, 1748, 1715, 1636, 1506, 1429, 1340, 1240, 1067 cm^{-1} . ^1H NMR (800 MHz, CD_3OD) δ : 3.05 (1H, dd, $J = 9.5, 8.9$, SGlu H-2), 3.15 (1H, dd, $J = 9.8, 8.9$, SGlu H-4), 3.22 (1H, dd, $J = 11.6, 9.8$, OXyl H-5b), 3.26 (1H, ddd, $J = 9.8, 5.6, 2.4$, SGlu H-5), 3.34 (1H, dd, $J = 8.9, 8.9$, SGlu H-3), 3.42 (1H, dd, $J = 8.7, 8.7$, OXyl H-3), 3.52 (1H, dd, $J = 8.7, 7.2$, OXyl H-2), 3.58 (1H, ddd, $J = 9.8, 8.7, 5.2$, OXyl H-4), 3.61 (1H, dd, $J = 12.0, 5.6$, SGlu H-6a), 3.81 (1H, dd, $J = 12.0, 2.4$, SGlu H-6b), 3.96 (1H, dd, $J = 11.6, 5.2$, OXyl H-5b), 4.36 (1H, d, $J = 9.5$, SGlu H-1), 4.88 (1H, d, $J = 7.2$, OXyl H-1), 6.96 (1H, ddd, $J = 8.1, 7.0, 0.9$, H-5), 7.09 (1H, ddd, $J = 8.3, 7.0, 1.1$, H-6), 7.22 (1H, d, $J = 8.3, 0.9$, H-7), 7.63 (1H, dd, $J = 8.1, 1.1, H-4$). ^{13}C NMR (200 MHz, CD_3OD) δ : 62.6 (SGlu C-6), 66.9 (SXyl

C-5), 71.06 (OXyl C-4), 71.08 (SGlu C-4), 73.6 (SGlu C-2), 75.0 (OXyl C-2), 77.2 (OXyl C-3), 79.1 (SGlu C-3), 82.0 (SGlu C-5), 88.9 (SGlu C-1), 107.0 (OXyl C-1), 112.2 (C-7), 112.9 (C-2), 119.2 (C-4), 120.2 (C-5), 121.7 (C-3a), 124.2 (C-6), 136.3 (C-7a), 141.9 (C-3). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{19}\text{H}_{25}\text{NNaO}_{10}\text{S}$ 482.1091; Found 482.1085.

3-(β -D-Arabinopyranosyloxy)-1H-indol-2-yl 1-thio- β -D-glucopyranoside (9e). Following the method similar to that used for the preparation of **8b**, **14e** (168 mg, 0.21 mmol) was treated with K_2CO_3 in MeOH to give the title compound (**9e**, 73 mg, 0.16 mmol, 79%) as a pale yellow microcrystalline solid. Mp 144–145 °C. $[\alpha]_{\text{D}}^{26} -7.6$ ($c = 1.0$, MeOH). IR (KBr): 3397, 2922, 2881, 2444, 1456, 1341, 1240, 1068, 1022, 947, 912, 873 cm^{-1} . ^1H NMR (800 MHz, D_2O) δ : 3.11 (1H, dd, $J = 9.7, 9.0$, SGlu H-2), 3.21 (1H, dd, $J = 9.9, 9.0$, SGlu H-4), 3.29 (1H, ddd, $J = 9.9, 5.6, 2.3$, SGlu H-5), 3.39 (1H, dd, $J = 9.0, 9.0$, SGlu H-3), 3.45 (1H, dd, $J = 13.5, 2.3$, SGlu H-6a), 3.60 (1H, dd, $J = 12.3, 5.6$, OArab H-5a), 3.64 (1H, dd, $J = 9.4, 3.4$, OArab H-3), 3.76 (1H, dd, $J = 12.3, 2.3$, OArab H-5b), 3.83 (1H, dd, $J = 9.4, 7.4$, OArab H-2), 3.85–3.87 (2H, m, SGlu H-6b and OArab H-4), 4.50 (1H, d, $J = 9.7$, SGlu H-1), 4.87 (1H, d, $J = 7.4$, OArab H-1), 7.05 (1H, ddd, $J = 8.1, 8.1, 0.9$, H-5), 7.17 (1H, ddd, $J = 8.1, 8.1, 1.0$, H-6), 7.31 (1H, dd, $J = 8.1, 0.9$, H-7), 7.60 (1H, dd, $J = 8.1, 1.0$, H-4). ^{13}C NMR (200 MHz, D_2O) δ : 60.8 (SGlu C-6), 66.2 (OArab C-5), 68.0 (OArab C-4), 69.3 (SGlu C-4), 71.1 (OArab C-2), 72.0 (SGlu C-2), 72.1 (OArab C-3), 77.0 (SGlu C-3), 80.1 (SGlu C-5), 87.1 (SGlu C-1), 104.9 (OArab C-1), 111.9 (C-7), 112.4 (C-2), 117.7 (C-4), 119.5 (C-3a), 120.1 (C-5), 124.0 (C-6), 134.6 (C-7a), 139.3 (C-3). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{19}\text{H}_{25}\text{NNaO}_{10}\text{S}$ 482.1091; Found 482.1085.

3-(β -D-Maltopyranosyloxy)-1H-indol-2-yl 1-thio- β -D-glucopyranoside (9f). Following the method similar to that used for the preparation of **8b**, **14f** (100 mg, 82 μmol) was treated with K_2CO_3 in MeOH to give the title compound (**9f**, 35 mg, 52 mmol, 63%) as a pale red microcrystalline solid. Mp 169–170 °C. $[\alpha]_{\text{D}}^{26} +8.7$ ($c = 1.0$, MeOH). IR (KBr): 3345, 3277, 2868, 1683, 1340, 1240, 1146, 1103, 1071, 1045, 914, 841 cm^{-1} . ^1H NMR (800 MHz, D_2O) δ : 3.25 (1H, dd, $J = 9.7, 9.0$, SGlu H-2), 3.35 (1H, dd, $J = 9.8, 9.0$, SGlu H-4), 3.42 (1H, ddd, $J = 9.8, 5.6, 2.3$, SGlu H-5), 3.44 (1H, dd, $J = 10.1, 9.2$, OMal H-4), 3.49 (1H, br m, OMal H-5), 3.51 (1H, dd, $J = 9.1, 9.0$, SGlu H-3), 3.61 (1H, dd, $J = 9.7, 4.0$, OMal H-2'), 3.68 (1H, dd, $J = 9.6, 7.9$, OMal H-2), 3.71 (1H, dd, $J = 9.7, 9.7$, OMal H-3'), 3.72–3.76 (2H, m, OMal H-5' and SGlu H-6a), 3.77–3.83 (3H, m, OMal H-4', OMal H-6a and H-6'a), 3.84–3.95 (4H, m, OMal H-3, H-6b, H-6'b, SGlu H-6b), 4.62 (1H, d, $J = 9.7$, SGlu H-1), 5.02 (1H, d, $J = 7.9$, OMal H-1), 5.44 (1H, d, $J = 4.0$, OMal H-1'), 7.17 (1H, ddd, $J = 8.0, 7.2, 0.9$, H-5), 7.29 (1H, ddd, $J = 8.3, 7.2, 1.0$, H-6), 7.43 (1H, dd, $J = 8.3, 0.9$, H-7), 7.75 (1H, dd, $J = 8.0, 1.0$, H-4). ^{13}C NMR (200 MHz, D_2O) δ : 60.4/60.6/60.8 (OMal C-6, C-6' and SGlu C-6), 69.3 (SGlu C-4), 69.4 (OMal C-4), 71.8 (OMal C-2'), 72.0 (SGlu C-2), 72.8 (OMal C-5'), 72.9 (OMal C-3'), 73.3 (OMal C-2), 74.8 (OMal C-5), 76.0 (OMal C-3), 76.6 (OMal C-4'), 77.1 (SGlu C-3), 80.0 (SGlu C-5), 87.2 (SGlu C-1), 99.8 (OMal C-1'), 104.4 (OMal C-1), 111.9 (C-7), 112.7 (C-2), 117.9 (C-4), 119.5 (C-3a), 120.1 (C-5), 124.0 (C-6),



134.6 (C-7a), 139.4 (C-3). HRMS (ESI) m/z : $[M+Na]^+$ Calcd for $C_{26}H_{37}NNaO_{16}S$ 674.1725; Found 674.1724.

N-Acetyl-2-(ethylthio)-1H-indol-3-yl 2,3,4,6-tetra-O-propionyl-1-thio-β-D-glucopyranoside (15). A mixture of **2** (300 mg, 1.29 mmol), ethanethiol (98 μL, 1.35 mmol), $BF_3 \cdot Et_2O$ (0.49 mL, 3.86 mmol), and 1,1,1,3,3,3-hexafluoropropan-2-ol (HFIP, 1.5 mL) was stirred at rt for 2 h. After removal of HFIP *in vacuo*, the residue was treated with **5a** (3.0g, 6.43 mmol) in a mixture of saturated aqueous K_3PO_4 (30 mL) and CH_2Cl_2 (10 mL) containing tetrabutylammonium hydrogen sulfate (TBAHS, 218 mg, 0.64 mmol) at rt for 30 min. The reaction mixture was poured into cold water (30 mL) and extracted with EtOAc (20 mL × 3). The extract was washed with brine (30 mL) and concentrated *in vacuo*. The residue was purified by column chromatography (*n*-hexane/EtOAc = 7/1) to give the title compound (**15**, 251 mg, 0.40 mmol, 31%) as a colourless viscous oil. $[\alpha]_D^{26} -9.5$ ($c = 1.0$, CH_2Cl_2). IR (neat): 1738, 1697, 1360, 1296, 1163, 1064, 1016, 887, 806, 752 cm^{-1} . 1H NMR (800 MHz, $CDCl_3$) δ : 1.09 [9H, t, $J = 7.8$, $C(O)CH_2CH_3$], 1.13 [3H, t, $J = 7.8$, $C(O)CH_2CH_3$], 1.16 (each 3H, t, $J = 7.8$, SCH_2CH_3), 2.26–2.44 [8H, m, $C(O)CH_2CH_3$], 2.76–2.85 (2H, m, SCH_2CH_3), 2.89 [3H, s, $C(O)CH_3$], 3.75–3.77 (1H, m, Glu H-5), 4.16 (1H, d, $J = 12.0$, Glu H-6a), 4.26 (1H, dd, $J = 12.0$, 5.2, H-6b), 5.16 (1H, d, $J = 7.8$, Glu H-1), 5.22 (1H, dd, $J = 9.6$, 9.6 H-4), 5.31 (1H, dd, $J = 9.6$, 9.6 H-3), 5.42 (1H, dd, $J = 9.6$, 7.8, H-2), 7.27 (1H, dd, $J = 7.8$, 7.8, H-5), 7.39 (1H, dd, $J = 7.8$, 7.8, H-6), 7.72 (1H, d, $J = 7.8$, H-4), 8.35 (1H, d, $J = 7.8$, H-7). ^{13}C NMR (200 MHz, $CDCl_3$) δ : 8.89/8.95/8.98/9.1 [$C(O)CH_2CH_3$], 14.5 (SCH_2CH_3), 27.26/27.34/27.4/27.6 [$C(O)CH_2CH_3$], 28.2 [$C(O)CH_3$], 31.4 (SCH_2CH_3), 61.9 (Glu C-6), 68.0 (Glu C-4), 71.3 (Glu C-2), 72.3 (Glu C-5), 72.6 (Glu C-3), 102.2 (Glu C-1), 116.2 (C-7), 118.2 (C-4), 118.3 (C-2), 122.9 (C-3a), 123.5 (C-5), 126.7 (C-6), 135.7 (C-7a), 146.9 (C-3), 171.1/172.8/172.9/173.6/174.0 (CO). HRMS (ESI) m/z : $[M+Na]^+$ Calcd for $C_{30}H_{39}NNaO_{15}S$ 644.2136; Found 644.2133.

2-(Ethylthio)-1H-indol-3-yl β-D-glucopyranoside (8g). Following the method similar to that used for the preparation of **8b**, **15** (101 mg, 0.16 mmol) was treated with K_2CO_3 in MeOH to give the title compound (**8g**, 53 mg, 0.15 mmol, 92%) as a pale green microcrystalline solid. Mp 95–96 °C. $[\alpha]_D^{26} +167.8$ ($c = 1.0$, MeOH). IR (KBr): 3395, 1622, 1533, 1449, 1341, 1238, 1071, 1011, 897 cm^{-1} . 1H NMR (800 MHz, CD_3OD) δ : 1.24 (3H, t, $J = 7.4$, SCH_2CH_3), 2.86/2.94 (each 1H, dq, $J = 14.7$, 7.4, SCH_2CH_3), 3.30–3.32 (1H, m, OGlu H-5), 3.46 (2H, br m, OGlu H-2 and H-4), 3.53 (1H, dd, $J = 8.6$, 8.6, OGlu H-3), 3.73 (1H, dd, $J = 11.8$, 5.1, OGlu H-6a), 3.84 (1H, dd, $J = 11.8$, 2.5, H-6b), 4.91 (1H, d, $J = 7.8$, OGlu H-1), 6.99 (1H, ddd, $J = 8.0$, 7.0, 1.0, H-5), 7.10 (1H, ddd, $J = 8.2$, 7.0, 1.2, H-6), 7.24 (1H, d, $J = 8.2$, H-7), 7.75 (1H, d, $J = 8.0$, H-4). ^{13}C NMR (200 MHz, CD_3OD) δ : 15.5 (SCH_2CH_3), 31.0 (SCH_2CH_3), 62.6 (OGlu C-6), 71.4 (OGlu C-4), 75.4 (OGlu C-2), 77.9 (OGlu C-3), 78.1 (OGlu C-5), 106.4 (OGlu C-1), 112.0 (C-7), 117.3 (C-2), 119.1 (C-4), 120.1 (C-5), 122.2 (C-6), 123.6 (C-3a), 135.8 (C-7a), 140.5 (C-3). HRMS (ESI) m/z : $[M+Na]^+$ Calcd for $C_{16}H_{21}NNaO_{16}S$ 378.0982; Found 378.0977.

N-acetyl-3-(2,3,4,6-tetra-O-acetyl-β-D-glucopyranosyloxy)-1H-indol-2-yl 2,3,4,6-tetra-O-acetyl-1-thio-β-D-glucopyranoside (10a). A mixture of **1** (70 mg, 0.143mmol), acetic anhydride (Ac_2O , 0.16 mL, 1.72 mmol), DMAP (1.0 mg, 7 μmol), and pyridine (3.0 mL) was stirred at rt for 16 h. The reaction mixture was poured into cold water (10 mL) and extracted with EtOAc (7 mL × 3). The extract was successively washed with 10% H_2SO_4 (7 mL × 4) and brine (7 mL) and concentrated *in vacuo*. The residue was purified by column chromatography (*n*-hexane/EtOAc = 5/1) to give the title compound (**10a**, 91 mg, 0.104 mmol, 73%) as a white amorphous. Mp 84–85 °C. IR (KBr): 2986, 2943, 2887, 1755, 1748, 1713, 1464, 1445, 1371, 1294, 1225, 1163, 1084, 925, 887 cm^{-1} . 1H NMR (800 MHz, $CDCl_3$) δ : 1.96/1.98/1.99/2.04/2.05/2.06/2.14/2.18 [each 3H, s, $OC(O)CH_3$], 2.87 [each 3H, s, $NC(O)CH_3$], 3.53 (1H, ddd, $J = 10.2$, 5.6, 2.2, SGlu H-5), 3.75 (1H, ddd, $J = 10.2$, 5.6, 2.5, OGlu H-5), 3.97 (1H, dd, $J = 12.3$, 2.2, SGlu H-6a), 4.12 (1H, dd, $J = 12.3$, 5.6, SGlu H-6b), 4.25 (1H, dd, $J = 12.4$, 2.5, OGlu H-6a), 4.27 (1H, dd, $J = 12.4$, 5.6, OGlu H-6b), 4.70 (1H, d, $J = 10.1$, SGlu H-1), 5.00 (1H, dd, $J = 10.2$, 9.4, SGlu H-2), 5.02 (1H, dd, $J = 10.2$, 9.4, SGlu H-4), 5.14 (1H, dd, $J = 10.1$, 9.4, OGlu H-4), 5.18 (1H, dd, $J = 9.4$, 9.4, SGlu H-3), 5.26 (1H, d, $J = 7.8$, OGlu H-1), 5.28 (1H, dd, $J = 9.4$, 9.4, OGlu H-3), 5.32 (1H, dd, $J = 9.4$, 7.8, OGlu H-2), 7.28 (1H, ddd, $J = 8.0$, 7.1, 0.9, H-5), 7.28 (1H, ddd, $J = 8.0$, 7.1, 0.9, H-5), 7.41 (1H, ddd, $J = 8.5$, 7.1, 1.3, H-6), 7.71 (1H, dd, $J = 8.0$, 1.3, H-4), 8.34 (1H, dd, $J = 8.5$, 0.9, H-7). ^{13}C NMR data were in good agreement with the literature values.¹⁸ HRMS (ESI) m/z : $[M+Na]^+$ Calcd for $C_{38}H_{46}NNaO_{20}S$ 868.2318; Found 868.2318.

3-(2,3,4,6-Tetra-O-propionyl-β-D-glucopyranosyloxy)-1H-5-indol-2-yl 2,3,4,6-tetra-O-propionyl-1-thio-β-D-glucopyranoside (10b). A mixture of **1** (70 mg, 0.143mmol), propionic anhydride (0.22 mL, 1.72 mmol), DMAP (1.0 mg, 7 μmol), and pyridine (3.0 mL) was stirred at rt for 24 h. The reaction mixture was poured into cold water (10 mL) and extracted with EtOAc (7 mL × 3). The extract was successively washed with 10% H_2SO_4 (7 mL × 4) and brine (7 mL) and concentrated *in vacuo*. The residue was purified by column chromatography (*n*-hexane/EtOAc =10/1) to give the title compound (**10b**, 77 mg, 82 μmol, 57%) as a pale yellow amorphous. Mp 81–82 °C. $[\alpha]_D^{20} -6.4$ ($c = 1.0$, $CHCl_3$). IR (KBr): 2982, 2943, 1751, 1352, 1275, 1179, 1084, 1067 cm^{-1} . 1H NMR (800 MHz, $CDCl_3$) δ : 1.04/1.07/1.08/1.09/1.12/1.13/1.15/1.18 [each 3H, t, $J = 7.6$, $C(O)CH_2CH_3$], 2.20–2.43 [16H, m, $C(O)CH_2CH_3$], 3.71 (1H, ddd, $J = 9.8$, 4.4, 2.0, SGlu H-5), 3.74 (1H, ddd, $J = 9.7$, 5.5, 2.3, OGlu H-5), 4.18 (1H, dd, $J = 12.3$, 2.0, OGlu H-6a), 4.24 (1H, dd, $J = 12.4$, 4.4, SGlu H-6a), 4.27 (1H, dd, $J = 12.3$, 5.5, OGlu H-6b), 4.33 (1H, dd, $J = 12.4$, 2.0, SGlu H-6b), 4.56 (1H, d, $J = 9.8$, SGlu H-1), 4.98 (1H, dd, $J = 9.8$, 9.8, SGlu H-2), 5.01 (1H, dd, $J = 9.8$, 9.8, SGlu H-4), 5.03 (1H, d, $J = 8.0$, OGlu H-1), 5.18 (1H, dd, $J = 9.7$, 9.7, OGlu H-4), 5.22 (1H, dd, $J = 9.8$, 9.8, SGlu H-3), 5.26 (1H, dd, $J = 9.7$, 9.7, OGlu H-3), 5.31 (1H, dd, $J = 9.7$, 8.0, OGlu H-2), 7.07 (1H, ddd, $J = 8.0$, 6.9, 0.9, H-5), 7.22 (1H, ddd, $J = 8.2$, 6.9, 1.2, H-6), 7.34 (1H, br d-like, $J = ca. 8.2$, H-7), 7.67 (1H, dd, $J = 8.0$, 0.9, H-4), 8.72 (1H, br s, NH). ^{13}C NMR (200 MHz, $CDCl_3$) δ : 8.89/8.96(3C)/8.03/9.0/9.1 [$C(O)CH_2CH_3$],



27.30/27.31/27.34(2C)/27.42/27.45/27.5/27.6 [C(O)CH₂CH₃], 61.4 (SGLu C-6), 62.1 (OGLu C-6), 67.6 (SGLu C-4), 68.3 (OGLu C-4), 69.9 (SGLu C-2), 71.3 (OGLu C-2), 72.1 (OGLu C-5), 72.7 (OGLu C-3), 73.7 (SGLu C-3), 76.5 (SGLu C-5), 74.0 (SGLu C-1), 102.2 (OGLu C-1), 110.8 (C-2), 111.2 (C-7), 118.6 (C-4), 119.8 (C-5), 120.5 (C-3a), 123.7 (C-6), 135.0 (C-7a), 140.6 (C-3), 172.7/172.8/172.9/173.3/173.4/173.6/174.1/174.6 [C(O)CH₂CH₃]. HRMS (ESI) *m/z*: [M+Na]⁺ Calcd for C₄₄H₆₀NNaO₁₉S 938.3475; Found 938.3475.

2-Acetyloxy-2,3-dihydro-1-benzofuran-3-one (16a). A mixture of **17**²² (600 mg, 3.41 mmol), *m*CPBA (70%, 1.26 mg, 5.11 mmol), and CH₂Cl₂ (15 mL) was stirred at rt for 3 h. The reaction was quenched by addition of a mixture of saturated aqueous NaHCO₃ solution containing 10% NaHSO₃ (15 mL), and the resulting mixture was extracted with CH₂Cl₂ (10 mL × 3). The extract was washed with brine (20 mL) and concentrated *in vacuo*. The residue was purified by column chromatography (*n*-hexane/EtOAc = 15/1) to give the title compound (**16a**,²⁵ 417 mg, 2.17 mmol, 64%) as a colourless oil. ¹H NMR (800 MHz, CDCl₃) δ: 2.20 [3H, s, C(O)CH₃], 6.21 (1H, s), 7.11 (1H, d, *J* = 8.3, H-7), 7.14 (1H, dd, *J* = 7.5, 7.5, H-5), 7.67 (1H, dd, *J* = 8.3, 7.5, H-6), 7.69 (1H, d, *J* = 7.5, H-4). ¹³C NMR (200 MHz, CDCl₃) δ: 20.5 [C(O)CH₃], 91.8 (C-2), 113.4 (C-7), 119.2 (C-3a), 123.1 (C-5), 124.8 (C-4), 139.0 (C-6), 169 [C(O)CH₃], 171.4 (C-7a), 194.9 (C-3). HRMS (ESI) *m/z*: [M+Na]⁺ Calcd for C₁₀H₈NaO₄ 215.0315; Found 215.0307.

2,3-Dihydro-1-benzofuran-3-one (18). A mixture of **17** (1.77 g, 10.1 mmol), Na₂SO₃ (1.89 g, 15.1 mmol), H₂O (25 mL), and MeOH (50 mL) was stirred at rt for 3 h. After removal of MeOH *in vacuo*, the aqueous residue was extracted with EtOAc (30 mL × 3). The extract was washed with brine (30 mL) and concentrated *in vacuo*. The residue was recrystallized from a mixture of *n*-hexane and EtOAc to give the title compound (**18**, 789 mg, 5.88 mmol, 59%) as a pale yellow microcrystalline solid. Mp 98–99 (lit.²⁶ 100–102 °C, lit.²⁷ 101–103 °C). ¹H NMR (800 MHz, CDCl₃) δ: 4.61 (2H, s, H-2), 7.08 (1H, dd, *J* = 7.7, 7.7, H-5), 7.13 (1H, d, *J* = 8.4, H-7), 7.60 (1H, dd, *J* = 8.4, 7.7, H-6), 7.66 (1H, d, *J* = 7.7, H-4). ¹³C NMR (200 MHz, CDCl₃) δ: 74.6 (C-2), 113.6 (C-7), 121.1 (C-3a), 121.9 (C-5), 124.0 (C-4), 137.8 (C-6), 173.9 (C-7a), 199.8 (C-3). HRMS (ESI) *m/z*: [M+H]⁺ Calcd for C₈H₇O₂ 9135.0441; Found 135.0439.

2-Bromo-2,3-dihydro-1-benzofuran-3-one (16b). A mixture of **18** (771 mg, 5.75 mmol), NBS (1.13 g, 6.33 mmol), PTSA (109 mg, 0.58 mmol), and MeCN (30.0 mL) was stirred at rt for 6 h. The reaction mixture was poured into cold water (30 mL) and extracted with EtOAc (20 mL × 3). The extract was washed with brine (30 mL) and concentrated *in vacuo*. The residue was purified by column chromatography (*n*-hexane/EtOAc = 7/1) to give the title compound (**16b**,²⁸ 1.14 g, 5.36 mmol, 93%) as a pale yellow microcrystalline solid. Mp 74–75 °C. IR (KBr): 1735, 1614, 1463, 1325, 1182, 1011, 762 cm⁻¹. ¹H NMR (800 MHz, CDCl₃) δ: 6.50 (1H, s, H-2), 7.18 (1H, d, *J* = 8.4, H-7), 7.23 (1H, dd, *J* = 7.4, 7.4, H-5), 7.70 (1H, ddd, *J* = 8.4, 7.4, 1.5, H-6), 7.77 (1H, br d-like, *J* = ca. 7.4, 1.5, H-4). ¹³C NMR (200 MHz, CDCl₃) δ: 75.7

(C-2), 113.9 (C-7), 117.9 (C-3a), 124.0 (C-5), 125.5 (C-4), 138.8 (C-6), 170.2 (C-7a), 194.5 (C-3). HRMS (ESI) *m/z*: [M+H]⁺ Calcd for BrC₈H₆O₂ 212.9546; Found 212.9541.

3-(2,3,4,6-Tetra-O-propionyl-β-D-glucopyranosyloxy)benzofuran-2-yl 2,3,4,6-tetra-O-acetyl-1-thio-β-D-glucopyranoside (20). A mixture of **16b** (300 mg, 1.41 mmol), **3a** (550 mg, 1.41 mmol), NaHCO₃ (237 mg, 2.82 mmol), and CH₃CN (20 mL) was stirred at rt for 1 h. After removal of CH₃CN *in vacuo*, the residue, consisting primarily of **19**, was treated with **5a** (3.2 g, 7.04 mmol) in a mixture of saturated aqueous K₃PO₄ (43 mL) and CH₂Cl₂ (14 mL) containing TBAHS (239 mg, 0.70 mmol) at rt for 30 min. The reaction mixture was poured into cold water (100 mL) and extracted with EtOAc (100 mL × 4). The extract was washed with brine (100 mL) and concentrated *in vacuo*. The residue was purified by column chromatography (*n*-hexane/EtOAc = 7/1) to give the title compound (**20**, 649 mg, 0.73 mmol, 53%) as pale red microcrystalline solid. Mp 86–87 °C. [α]_D²² –18.3 (*c* = 1.0, CHCl₃). IR (KBr): 2984, 2945, 2882, 1763, 1379, 1364, 1240, 1169, 1086, 1059 cm⁻¹. ¹H NMR (800 MHz, CDCl₃) δ: 1.08/1.09/1.11/1.15 [each 3H, t, *J* = 7.6, C(O)CH₂CH₃], 1.98/2.011/2.013/2.15 [each 3H, s, OC(O)CH₃], 2.24–2.44 [8H, m, C(O)CH₂CH₃], 3.69 (1H, ddd, *J* = 10.2, 5.3, 2.2, SGLu H-5), 3.87 (1H, ddd, *J* = 10.2, 5.3, 2.3, OGLu H-5), 4.16 (1H, dd, *J* = 12.5, 5.3, SGLu H-6a), 4.18 (1H, dd, *J* = 12.5, 2.2, OGLu H-6a), 4.22 (1H, dd, *J* = 12.5, 2.2, SGLu H-6b), 4.27 (1H, dd, *J* = 12.3, OGLu H-6b), 4.78 (1H, d, *J* = 10.0, SGLu H-1), 4.91 (1H, dd, *J* = 10.1, 9.6, SGLu H-2), 5.02 (1H, dd, *J* = 9.6, 9.6, SGLu H-4), 5.33 (1H, dd, *J* = 9.6, 9.6, SGLu H-3), 5.34 (1H, dd, *J* = 9.6, 9.6, OGLu H-4), 5.34 (1H, dd, *J* = 9.6, 9.6, OGLu H-3), 5.34 (1H, dd, *J* = 7.7, 9.6, OGLu H-2), 5.40 (1H, d, *J* = 7.7, OGLu H-1), 7.22 (1H, ddd, *J* = 8.0, 6.9, 1.2, H-6), 7.34 (1H, ddd, *J* = 8.0, 6.9, 1.3, H-5), 7.36 (1H, dd, *J* = 8.0, 1.2, H-4), 7.61 (1H, dd, *J* = 8.0, 1.3, H-7). ¹³C NMR (200 MHz, CDCl₃) δ: 9.0/9.1/9.16/9.19 [C(O)CH₂CH₃], 20.61/20.63(2C)/20.8 [C(O)CH₃], 27.38/27.42/27.51/27.59 [C(O)CH₂CH₃], 61.8 (SGLu C-6), 62.0 (OGLu C-6), 67.9 (SGLu C-4), 68.3 (OGLu C-4), 70.6 (SGLu C-2), 71.1 (OGLu C-2), 72.0 (OGLu C-5), 72.6 (OGLu C-3), 74.1 (SGLu C-3), 76.6 (SGLu C-5), 84.1 (SGLu C-1), 100.6 (OGLu C-1), 111.8 (C-4), 119.7 (C-7), 122.9 (C-3a), 123.1 (C-6), 126.6 (C-5), 130.0 (C-2), 145.7 (C-3), 154.4 (C-7a), 169.5(2C)/170.2/170.7 [C(O)CH₃], 172.9/173.0/173.6/174.2(CO). HRMS (ESI) *m/z*: [M+Na]⁺ Calcd for C₂₀H₅₀NaO₂₀S 905.2508; Found 905.2509.

3-(β-D-Glucopyranosyloxy)benzofuran-2-yl 1-thio-β-D-glucopyranoside (11). A mixture of **20** (160 mg, 0.18 mmol), K₂CO₃ (8 mg, 0.06 mmol), and MeOH (6.0 mL) was stirred at rt for 4 h. After removal of MeOH *in vacuo*, the residue was purified by column chromatography (CHCl₃/MeOH = 5/1) to give the title compound (**11**, 53 mg, 0.11 mmol, 61%) as a colourless microcrystalline solid. Mp 198–199 °C. [α]_D²³ –4.1 (*c* = 1.0, DMSO). IR (KBr): 3385, 2934, 2886, 1447, 1354, 1273, 1117, 1026 cm⁻¹. ¹H NMR (800 MHz, DMSO-*d*₆) δ: 3.08–3.12 (2H, m, SGLu H-2 and H-4), 3.17 (1H, ddd, *J* = 9.7, 5.4, 2.4, SGLu H-5), 3.23–3.28 (2H, m, SGLu H-3 and OGLu H-4), 3.31–3.35 (3H, m, OGLu H-2, H-3, and H-5), 3.47 (1H, ddd, *J* = 11.8, 5.7, 5.7, SGLu H-6a), 3.57 (1H, ddd, *J* = 11.5, 5.7, 5.7, OGLu H-6a), 3.65 (1H, ddd,



$J = 11.8, 5.3, 2.4$, SGLu H-6b), 3.73 (1H, ddd, $J = 11.5, 5.1, 2.4$, OGLu H-6b), 4.07/4.18/4.63/4.70/4.73/4.75/4.97/5.04 (each 1H, br s-like, OH), 4.59 (1H, d, $J = 9.6$, SGLu H-1), 5.20 (1H, d, $J = 7.2$, OGLu H-1), 7.24 (1H, ddd, $J = 8.1, 7.2, 0.9$, H-6), 7.36 (1H, ddd, $J = 7.2, 7.2, 1.3$, H-5), 7.47 (1H, dd, $J = 7.2, 0.9$, H-4), 7.78 (1H, dd, $J = 8.1, 1.3$, H-7). ^{13}C NMR (200 MHz, DMSO- d_6) δ : 60.7 (OGLu C-6), 60.8 (SGLu C-6), 69.6 (OGLu C-4), 69.7 (SGLu C-4), 73.1 (SGLu C-2), 73.4 (OGLu C-2), 76.2 (OGLu C-5), 76.6 (OGLu C-3), 77.6 (SGLu C-3), 80.7 (SGLu C-5), 87.1 (SGLu C-1), 102.4 (OGLu C-1), 111.0 (C-4), 119.3 (C-7), 122.2 (C-6), 122.3 (C-3a), 125.3 (C-5), 132.8 (C-3), 144.0 (C-2), 153.3 (C-7a). HRMS (ESI) m/z : $[\text{M}+\text{Na}]^+$ Calcd for $\text{C}_{20}\text{H}_{50}\text{NaO}_{20}\text{S}$ 905.2508; Found 905.2509.

Bioassays

Cell Culture

The HFDPCs were originally purchased from TaKaRa Bio Inc. (Shiga, Japan), and the cells were grown in Dulbecco's modified eagle medium (DMEM) supplemented with 10% fetal bovine serum (FBS), penicillin (100 units/mL), and streptomycin (100 $\mu\text{g}/\text{mL}$) at 37 °C in 5% $\text{CO}_2/95\%$ air. The cells were harvested by incubation in phosphate-buffered saline (PBS) containing 1 mM ethylenediaminetetraacetic acid (EDTA) and 0.25% trypsin for approximately 5 min at 37 °C and were used for the subsequent bioassays.

Measurement of Cell Proliferation

The HFDPCs (1.0×10^4 cells/mL) were seeded in 96-well plates in serum-free DMEM (100 $\mu\text{L}/\text{well}$) and cultured for 24 h. Afterward, 100 μL of the test compounds at various concentrations dissolved in serum-free DMEM were added to each well, followed by incubation for 4 d. After the incubation, 20 μL of Cell Counting Kit-8 (CCK-8, Dojindo Laboratories, Kumamoto, Japan) reagent was added to each well, followed by incubation for 1 h at 37 °C. The absorbance was recorded at 450 nm (absorbance at 650 nm as reference) on a microplate reader (MTP-900Lab; HITACHI, Japan). The results are expressed as mean percentages of the control \pm standard error of the mean (S.E.M.).

Cell proliferation was calculated using the following equation:

$$\text{Cell proliferation (\%)} = A/B \times 100$$

where A is the absorbance at 450 nm (650 nm for the test sample) and B is the absorbance at 450 nm (650 nm for the control).

Author contributions

RS, KT, HN and SM performed chemistry and analytical experiments. YM, FI and TM performed the biological experiments and analysed data. GT supervised the study, obtained funding and wrote the paper. The manuscript was reviewed and approved by all authors.

Conflicts of interest

There are no conflicts to declare.

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Data Availability Statement

- All data associated with this study have been included in either the manuscript or in the supplementary file associated with the manuscript.

