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Tradescantia pallida extract incorporated chitosan/pullulan intelligent biodegradable films: an eco-friendly packaging to preserve the freshness of chicken

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Herein, biodegradable active films were developed using a polysaccharide based blend of chitosan (CS) and pullulan (PU), incorporated with the ethanolic extract of *Tradescantia pallida* (TP), to function as intelligent food packaging materials. FTIR confirmed intermolecular hydrogen bonding between the polyphenolic constituents of TP extract and the CS/PU matrix. The resulting CPT intelligent biodegradable films exhibited an impressive tensile strength of 45.84 MPa, surpassing that of conventional plastic films. SEM micrographs revealed a uniform and compact surface morphology, demonstrating successful blending and structural integrity. The incorporation of polyphenolic compounds not only enhanced the antimicrobial efficacy of the films against foodborne pathogens but also imparted remarkable antioxidant potential, as evidenced by DPPH (89.81%) and ABTS (85.14%) radical scavenging activities. A distinct color shift from purple to yellow upon spoilage enabled real-time monitoring of freshness. Chicken samples packaged in polyethylene (PE) and CPT-2 intelligent biodegradable films were stored under ambient conditions. Analyses of microbial growth, ammonia release, free fatty acid degradation, and the effects of UV exposure revealed that CPT-2 intelligent biodegradable films offered superior protection against microbial contamination and oxidative deterioration, thereby extending the chicken's shelf life compared to PE packaging. The CPT intelligent biodegradable films also showed biodegradation potential, addressing the persistence, landfill accumulation, and greenhouse gas emissions associated with single-use plastics. Thereby mitigating the adverse impacts of SUPs on human health and the ecosystem. Overall, the physicochemically enhanced intelligent films offer a sustainable and effective alternative to plastic packaging, improving food safety and environmental health.

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Sustainability spotlight

Sustainability has become an imperative across all sectors due to the growing environmental burden caused by SUPs and their persistent pollution. In the food sector, the challenges of resource scarcity and rapid spoilage demand innovative and eco-friendly packaging solutions. Herein, a chitosan/pullulan blend was developed, both polymers being biodegradable and currently at a Technology Readiness Level (TRL) of 5–6, indicating strong potential for commercial advancement. The incorporation of anthocyanin-rich *Tradescantia pallida* extract introduces both active and intelligent functionalities to the film, making it a sustainable alternative to conventional synthetic packaging materials. The prepared intelligent film demonstrated the ability to extend the shelf life of chicken by 48 hours at room temperature, effectively doubling that of polyethylene (PE) packaging under similar conditions. Beyond its superior preservation capacity, the film's complete biodegradability ensures that, after disposal, it does not contribute to landfill accumulation or microplastic pollution. Thus, the developed film not only addresses food preservation challenges but also aligns with circular economy principles by minimizing environmental impact as a promising step toward a sustainable, pollution-free packaging ecosystem and a greener future.

1. Introduction

Excessive plastic consumption has become a significant environmental threat, with waste disposal and management presenting substantial global challenges. Plastics in landfills contribute to global warming and microplastic pollution. Across its life cycle, from extraction to manufacturing, plastic production emits large amounts of greenhouse gases,

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generating approximately 1.96 gigatons in 2015, according to the Minderoo-Monaco Commission on Plastics and Human Health (2023). The food sector significantly contributes to this problem through the use of single-use, non-biodegradable packaging, driven by modern lifestyles. Valued at \$338.34 billion in 2023, the global food packaging industry is projected to reach \$478.18 billion by 2028. Although recycled plastic is considered a better packaging alternative, it often contains harmful substances such as BPA, phthalates, flame retardants, and heavy metals, which limit its suitability for food-related uses.¹⁻³ The rise in demand for food packaging, combined with the detrimental effects of single-use and recycled plastics, has prompted the development of environmentally acceptable and sustainable alternatives. Bioplastics degrade in the environment with minimal hazardous environmental impact and are receiving greater attention from researchers. There are three primary categories in bioplastics: partially bio-based, such as Bio-PET, which is non-biodegradable, and another group generated from conventional fossil resources that are biodegradable, such as CPL and PBHT. Conversely, biopolymers are naturally occurring polysaccharides, lipids, and proteins that are biodegradable and eco-friendly.⁴⁻⁶

Polysaccharide based films are gaining popularity due to their natural origin. The deacetylation of β -(1-4)-poly-*N*-acetyl-*D*-glucosamine (chitin) leads to the formation of the second most abundant polysaccharide, *i.e.*, chitosan (CS). Chitosan consists of β -1,4-linked 2-amino-2-deoxy- β -*D*-glucose (deacetylated *D*-glucosamine) and *N*-acetyl-*D*-glucosamine units, a copolymer that has received FDA certification for food-related studies. The biodegradable, biocompatible, non-toxic, and antimicrobial properties, along with the film-forming capacity of chitosan, have received considerable attention from a commercial perspective in food packaging and biomedical applications.⁷⁻⁹ Emulsion formation capability, good tensile strength, and antimicrobial activity suggest that CS can be considered for food packaging applications. However, the brittleness and high water vapour permeability of pure CS films restrict their applicability as a food packaging material.¹⁰

The extracellular polysaccharide obtained from *Aureobasidium* spp. Fungi are known to produce pullulan (PU). Pullulan is a biopolymer that received GRAS status from the FDA in 2004 and was registered by the European Union (EU) as a food additive (E1204) in the same year.¹¹ The characteristic structural flexibility in pullulan is due to α -1,6-glycosidic linkages present between the linearly polymerized maltotriose units.^{12,13} Pullulan films are transparent, possess thermal capacity, and exhibit resistance to oil and grease, while also showing good oxygen barrier properties.¹⁴ Despite all these excellent tensile strength and physicochemical qualities, pullulan film's lack of water resistance properties limits its usage as a food packaging material.

In the present study, a polymer blend of CS/PU is considered to overcome the limitations of chitosan and pullulan neat films. Glycerol was added to the CS/PU blend as the plasticizing agent to enhance the elastic nature of the polymer blend. The glycerol's acceptor and donor hydrogen bonding ability suggests it as a better plasticizing agent for chitosan.¹⁵ Adding glycerol as

a plasticizer increases the free volume in the polymer matrix, thereby enhancing the flexibility of the brittle CS/PU blend. Food freshness is crucial, as every product has a defined shelf life. To monitor this, intelligent packaging materials have been developed that can sense spoilage through various detection methods. Among these, pH-based sensing is effective in identifying food deterioration. Anthocyanins, naturally occurring polyphenols in fruits and vegetables, are pH-sensitive pigments responsible for their vivid colors. They appear red in acidic conditions due to the flavylium cation and yellow in basic conditions due to the chalcone structure.¹⁶

Hence, the anthocyanin content in plant and fruit extracts was utilized intelligently in packaging to identify pH variations by changing the color of the films. Yan Qin *et al.* reported that betalains from red pitaya (*Hylocereus polyrhizus*) peel extract are extracted into starch/PVA to fabricate biodegradable films for intelligent packaging applications.¹⁷ Tilak Gasti *et al.* fabricated chitosan/methylcellulose films to monitor fish fillet freshness, incorporating *Phyllanthus reticulatus* as an anthocyanin source.¹⁸ Thus, *Tradescantia pallida* (also known as purple heart or wandering jew) is an extremely popular, inexpensive, and readily available ornamental groundcover worldwide. This accessibility, stemming from its ease of growth, high nursery availability, drought tolerance, and simple propagation *via* cuttings, makes it a cost-effective alternative for medicinal cultivation. Historically, its leaves have been used as an anti-inflammatory, antitoxic, and circulatory tonic. Modern pharmacological interest, particularly in the leaf extract, is driven by its rich phytochemical profile of phenolics, flavonoids, and anthocyanins, making it suitable for active and intelligent packaging applications.¹⁹ Traditionally, TP treated venereal diseases, mucosal infections, and gastrointestinal disorders.²⁰ The polyphenolic constituents of the *Tradescantia pallida* (TP) extract include phenolic acids such as *p*-coumaric acid, syringic acid, and chlorogenic acid; flavonoids such as naringenin, morin, catechin, epicatechin, and quercetin; and anthocyanins such as cyanidin-3,7,3'-triglucoside and petunidin, which are primarily responsible for the colouration of TP leaves. These phenolic acids, flavonoids, and anthocyanins collectively contribute to the film's antimicrobial and antioxidant properties, while also enabling real-time color variation in response to pH changes, making them suitable for intelligent food packaging applications.²¹ Due to all these considerations, TP extract is considered the active agent in the present study because polyphenolic extract consists of anthocyanin content capable of identifying pH variation on spoilage.

According to the UN's Food and Agriculture Organization (FAO), worldwide chicken meat production reached 137 million tons in 2020.²² Chicken meat is susceptible to microbial assault, perishable at room temperature, and challenging to preserve. Plastic is the primary packaging material used in the meat industry. In India, meat packaging is dependent on 10 μ PET/45 μ HDPE – LDPE, LDPE – BA – Nylon – BA – LLDPE, HDPE – LDPE – HDPE, HDPE – LLDPE, and HDPE + LDPE (blended) plastics, according to the Indian Centre for Plastic in the Environment.²³ These plastics, derived from fossil fuels, are the primary source of environmental contamination because they



persist in the environment for extended periods.^{24,25} Therefore, the current study hypothesized that polysaccharides based CS/PU films incorporated with TP extract should be employed instead of using single-use plastic for chicken meat packaging because CPT intelligent biodegradable films enhanced physicochemical, superior antibacterial and antioxidant properties, colour changing capabilities due to pH variation and biodegradable nature made CPT intelligent biodegradable films an appealing alternative to microplastic generating tradition plastic packaging and serve as active and intelligent packaging materials for eco-friendly and sustainable future.

2. Materials and methods

2.1. Materials

Chitosan (viscosity: 200–600 mPa s, 0.5% in 0.5% acetic acid, deacetylation value: min. 80% after drying, product no.: C0831, CAS: 9012-76-4, MW = 19–31 kDa) and pullulan (product no.: P0978, CAS: 9057-02-7, MW ≈ 200 kDa) procured from TCI, Tokyo. Mueller–Hinton agar (MHA), yeast extract, peptone, sodium chloride L-ascorbic acid Hi-Media, methanol SD Fines, 2,2-diphenyl-1-picrylhydrazyl (DPPH), 2-[(2,6-dichlorophenyl)amino]benzene acetic acid sodium salt (diclofenac sodium salt) from SRL.

2.2. Extraction of anthocyanin content from *Tradescantia pallida* (TP)

The leaves of the *Tradescantia pallida* plant, also called purple heart, were collected from the Karnatak Science College botanical garden, Dharwad. The anthocyanin content was extracted according to the procedure suggested by Valdir Aniceto Pereira *et al.*, with some modifications.²⁶ Distilled water was used to wash the collected leaves. A 100 g sample of finely chopped TP leaves was placed in a 150 ml ethanolic solution (ethanol : water = 2 : 1), and 1 mol l⁻¹ HCl was added dropwise to adjust the pH of the solution to 2. The sample was protected from the sunlight and stored at 4 °C for 48 h. After filtering the solution, the extract was centrifuged for 10 min at 7000 rpm. After filtering, the resultant supernatant pH was adjusted to 7 using 2.5 ml l⁻¹ of NaOH and dried at 40 °C. The dried powder extract was then stored at 4 °C. The total phenolic content of ethanolic leaf extract was 134.44 ± 0.29 mg g⁻¹ of leaf, and the total flavonoid content in ethanolic leaf extract was 117.78 ± 0.57 mg g⁻¹ of leaf.

2.3. Fabrication of CPT intelligent films

Chitosan (CS) and pullulan (PU) polysaccharides are the polymers considered in the study. 1.5 g of CS and 0.5 g of PU were dissolved in 1% acetic acid and distilled water, respectively. After achieving a homogenous blend of CS and PU, 10 ml of 1% glycerol was added as a plasticizer. The blend solution was poured into a Petri dish and dried at room temperature for 6 days to obtain the control CP film. To the control blend solution, 100 and 200 mg of TP extract dissolved in ethanol were added and stirred on a magnetic stirrer for 4–5 h at 350 rpm. The CPT-1 and CPT-2 intelligent biodegradable films fabricated

from the solvent casting method poured in Petri plates were peeled after 6 d and stored in a desiccator.

3. Characterizations

3.1. Fourier transform infrared (FTIR) spectroscopy

Fundamental alterations induced by the CS/PU blend and the incorporation of TP extract were characterized using attenuated total reflection (ATR) mode with a Shimadzu IR spectrometer (Japan). Samples measuring 4 × 1 cm² were placed directly in the spectrometer, and spectral analysis was performed across the 400–4000 cm⁻¹ range at a resolution of 4 cm⁻¹.²⁷

3.2. Mechanical properties

The mechanical properties were assessed using the Universal Testing Machine (series 7200-1 kN) manufactured by Dak System Inc., as per the ASTM-D882 standard test (ASTM, 1992). The films consider a grip separation of 50 mm, a crosshead speed of 0.1 mm min⁻¹, and film sample parameters of 10 cm in length and 2.5 cm in width, carried out at RH of 50–55% at RT.^{28,29}

3.3. Morphological analysis

To examine the surface morphology, 2 cm × 2 cm control CP and CPT intelligent films were considered. The 10 kV acceleration voltage JEOL JSM-6360 (Japan) was used for SEM imaging. A thin layer of gold is sputtered onto films to prevent charging.³⁰

3.4. Water contact angle (WCA)

A contact angle analyzer (model DMs-401, Kyowa Interface Science Co., Ltd, Tokyo) was used to study hydrophilic or hydrophobic behavior. Each time, a liquid drop volume of 1 µl was injected onto 1 × 5 cm² films to capture the nature of contact in 5 different positions.³¹

3.5. Water solubility (WS)

The water solubility of CP and CPT intelligent films is determined according to the ASTM standard D570-98. After pre-drying 2 × 2 cm² film samples for 24 h at 60 °C in a hot air oven, the initial weight (W_1) was recorded. After immersing the films in 30 milliliters of double-distilled water for a day at room temperature, the excess water from the immersed films was removed using filter paper. Following another day of drying at 100 °C, the films were weighed, and the final weight was recorded (W_2). The percentage of water solubility was determined using the equation below.³²

$$\text{Water solubility (\%)} = \frac{W_1 - W_2}{W_1} \times 100$$

3.6. Moisture adsorption (MA)

The intelligent film samples, measuring 2 × 2 cm², were subjected to a drying process in an oven at a temperature of 100 °C for 24 h, and the initial weight (W_1) was recorded. The films were reweighed after exposure to atmospheres with relative



humidity levels of 19%, 50%, and 75% for 24 hours, yielding the final weight (W_2). Each sample underwent three separate measurements, and MA was calculated as follows,³³

$$\text{Moisture adsorption (\%)} = \frac{W_2 - W_1}{W_1} \times 100$$

3.7. Water vapour transmission rate (WVTR)

The cylindrical vials, with an inner diameter of 1.7 cm, were filled with 10 ml of distilled water. The objective was to assess the films' water vapor transmission rate (WVTR). The measurement was done gravimetrically using the ASTM E00996-00 method. A $3 \times 3 \text{ cm}^2$ film was used to cover the opening of the vial, and Teflon tape was wrapped around it to ensure airtightness. The vials were subjected to a temperature of 40 °C in a hot air oven for 24 hours. The WVTR capacity of each film was determined by calculating the weight fluctuations of vials at 24 h using the equations given below,³⁴

$$\text{WVTR (g per m}^2 \text{ per h)} = \frac{W_1 - W_2}{A \times T} \times 100$$

3.8. Soil burial test

The investigation measured the degradation ability of a $2 \times 2 \text{ cm}^2$ control CP and CPT intelligent films, initially weighing as W_1 . The films were buried 5 cm deep in the soil on mesh and watered daily. After 10, 20, and 30 days, the films were removed, cleaned with double-distilled water, dried at 45 °C, and weighed to obtain a decreased final weight of W_2 . The rate of degradation of active films was calculated using the following equation.³⁵

$$\text{Rate of degradation (\%)} = \frac{W_1 - W_2}{W_1} \times 100$$

3.9. Anti-microbial sensitivity

The antimicrobial property of CPT-intelligent films was determined against *Escherichia coli* (ATCC 10799), *Staphylococcus aureus* (ATCC 6538), *Bacillus subtilis*, and *Candida albicans* (ATCC 24433). Pure bacterial cultures were subcultured on Luria–Bertani (LB) medium until the optical density (OD) at 600 nm reached 0.5. A swab was used to smear the microorganisms onto the MHA (4 mm thickness) plates. 100 μl of the solution was gently dropped into the wells created using a gel puncher (8 mm diameter). The plates were then incubated overnight at 37 °C in the incubator. The inhibition zone around the drop was measured using a vernier scale.

3.10. Anti-oxidant activity

3.10.1. DPPH free radical scavenging assay. The procedure was performed in accordance with Blois (1958), with minor

modifications.³⁶ Briefly, 100–500 μl of standard ascorbic acid and film solutions (1 mg ml^{-1}) were made up to 1000 μl using methanol and treated with 500 μl methanolic DPPH (0.5 mM) and incubated for 30 min in the dark at room temperature. OD was measured in a UV-visible spectrophotometer (Labman, LMSP UV-1200) with 1.5 ml glass cuvettes of 1 cm pathlength at 517 nm. As a control, DPPH diluted in methanol was used, and methanol alone served as the blank solution.³⁷

3.10.2. ABTS free radical scavenging assay. The assay was performed in accordance with Re *et al.* (1999), with slight modifications.³⁸ Briefly, an ABTS⁺ stock solution was prepared by reacting equal quantities of aqueous ABTS (7 mM) and aqueous ammonium persulfate (2.45 mM) in solution; the mixture was allowed to stand in the dark at room temperature for 12–16 hours. The working solution of ABTS⁺ was obtained by diluting the stock solution in methanol to give an absorbance of 0.70 ± 0.05 at 734 nm. Then, 1000 μl of ABTS⁺ solution was mixed with 100–500 μl of film solutions and standard ascorbic acid (1 mg ml^{-1}). The mixture was then incubated in the dark at RT for 10 min. OD was measured using a UV-visible spectrophotometer (Labman, LMSP UV-1200) with 1 ml glass cuvettes of 1 cm pathlength at 734 nm. The ABTS⁺ solution with water was used as the control, and water was used to set the blank.

The free radical scavenging activity was expressed in terms of percent inhibition, which was calculated using the formula shown below,

$$\% \text{ scavenging} = \frac{Ab_c - Ab_s}{Ab_c} \times 100.$$

where Ab_c denotes the absorbance of the control and Ab_s denotes the absorption of the sample.

3.11. Microbiological analysis of chicken

3.11.1. Total bacterial count (TBC). The enumeration of bacterial CFU determined TBC after the incubation period. Chicken meat (CM) slices (1 g) were packed in control CP and CPT-2 intelligent biodegradable films and PE plastic before incubating at room temperature for 24, 48, and 72 h, respectively. After incubation, the chicken meat was dissolved in 0.9% saline (0.1 g ml^{-1}) and vortexed for a minute, followed by 15 minutes in a shaker incubator at 37 °C and 180 rpm. The solution was serially diluted, and 100 μl of each dilution was distributed on Luria–Bertani (LB) agar plates and incubated for 36 h at 37 °C. After incubation, the plates were examined for colonies, and the imaged plate colonies were counted using Open-CFU colony counter software. The CFU per g of CM values of all the samples were calculated by using the following equation:

$$\text{CFU per g of CM} = \frac{\text{number of colonies} \times 100 \text{ (initial dilution factor)} \times \text{serial dilution factor}}{\text{volume plated in ml (0.1)}}$$



3.11.2. Inoculation of CM with common foodborne pathogenic bacteria. The CM slices (1 g) were sprayed with 90% ethanol and wiped thoroughly with tissue paper. The sterile CM was dipped in LB broth containing $\sim 1.5 \times 10^6$ CFU per ml of *S. aureus* and *E. coli* and allowed to stand in a shaker incubator for 30 min at 37 °C and 180 rpm. The infected CM slices were packed with control CP, CPT-2 intelligent biodegradable films, and PE plastic before incubating at room temperature (RT) for 24, 48, and 72 h. After incubation, the slices were dissolved in 0.9% saline (0.1 g ml^{-1}) and vortexed for 0.5 min, followed by 15 min in a shaker incubator at 37 °C at 180 rpm. The solution was serially diluted, and 100 μl from each dilution was spread on LB agar plates and incubated for 36 h at 37 °C. The CFU per g of CM was then calculated.

3.12. Evaluation of chicken meat spoilage activities after incubation time

Soon after the treatment, chicken meat was blended with 18 ml of 0.9% saline using a tissue homogenizer (Labico, LT-451, Labtech Instruments, India) for 0.5 min and then vortexed for 0.5 min. The blend was filtered through an individual four-layered muslin cloth. The chicken meat broth solution (CMBS) was further subjected to biochemical characterizations to evaluate nutritional levels.

3.12.1. DPPH free radical scavenging assay. The procedure was performed in accordance with Blois (1958), with minor modifications.³⁶ Briefly, 200 μl of CMBS was diluted to 1000 μl with methanol, treated with 500 μl of methanolic DPPH (0.5 mM), and incubated for 30 min in the dark at room temperature. Absorbance was measured in a UV-visible spectrophotometer (Labman, LMSP UV-1200) with 1.5 ml glass cuvettes of 1 cm path length at 517 nm. DPPH diluted with methanol was used as a control, and pure methanol was used as a blank solution. The free radical scavenging activity was expressed in terms of percent inhibition, calculated using the formula below.³⁹

$$\% \text{ scavenging} = \frac{Ab_c - Ab_t}{Ab_c} \times 100.$$

Ab_c denotes the absorbance of the control, and Ab_t represents the absorbance of CMBS.

3.12.2. Estimation of ammonia and ammonium salts. The ammonia and ammonium salts were estimated using Nessler's reagent with minor modifications.^{40,41} Briefly, 0.25 ml Rochelle salt solution (25%) was mixed with 1.5 ml of CMBS and 0.25 ml of Nessler's reagent (4% KI, 1.6% HgCl_2 , 10% KOH). The solution was allowed to stand in the dark for 15 min at RT. After incubation, absorbance was measured in a UV-visible spectrophotometer (Labman, LMSP UV-1200) with 1.5 ml glass cuvettes of 1 cm path length at 420 nm. A solution of 1 ml 0.9% saline was used as a blank. The concentration of ammonia was estimated using the standard calibration curve of ammonium chloride.

3.12.3. Estimation of total protein content. Initially, to minimize ammonia interference in protein quantification, CMBS was five-fold diluted with 50 mM Tris–borate buffer at pH

8.5 (50 mM) and incubated for 30 min at 37 °C to allow precipitation of ammonium ions and evaporation of ammonia gas.^{42,43} After incubation, the mixture was centrifuged at 4000 rpm for 5 min, and the supernatant was subjected to total protein estimation.

The total protein content of the buffered CMBS was quantified using the FCR method with slight modifications.⁴⁴ Briefly, 40 μl of buffered CMBS was diluted to 400 μl with water, followed by the addition of 1 ml of alkaline–copper sulfate solution. The mixture was incubated for 10 min, 100 μl of 50% FCR reagent was added, and incubated for 30 min in the dark at RT. After incubation, absorbance was measured in a UV-visible spectrophotometer (Labman, LMSP UV-1200) with 1.5 ml glass cuvettes of 1 cm path length at 750 nm. A solution with 0.4 ml buffer was used as a set blank. The total protein content was estimated using the standard calibration curve of bovine serum albumin. The relative decrease in % of protein content compared to the fresh chicken was expressed using the following formula.

$$\text{Decrease in \% of total protein content} = 100 \times \left[1 - \left(\frac{Ab_t}{Ab_f} \right) \right]$$

where Ab_f denotes the absorbance of fresh buffered CMBS and Ab_t denotes the absorption of treatment.

3.12.4. Estimation of acidity index of total free soluble fat. Initially, CMBS was added with an equal volume of ethanol (95%) and vortex to dissolve the fat content for 1 min. The NaOH titration method was used to estimate the total acid value of the soluble fat content, employing a phenolphthalein indicator with slight modifications.^{45,46} Briefly, 10 ml of ethanol CMBS mixture was added to 0.1 ml of phenolphthalein indicator (1% in ethanol) and titrated against a 10 mM NaOH solution. The results are expressed in grams of oleic acid equivalents per 100 g of meat.

3.13. Applications of UV treatment for packaged meat

A meat packaging experiment was conducted to assess the feasibility of implementing UV-blocking properties in prepared active film in comparison to traditional PE plastic and unpacked chicken, with a focus on its performance in the context of preserving meat. 2 g of sliced fresh chicken meat was kept in a sterile Petri plate (unpacked meat and meat packed with CPT-2, PEP). The experiment was conducted in a laminar airflow chamber with UV light. A broad-spectrum UV lamp was used as the UV source, with a distance of 30 cm between the source and the meat treatment. The samples were exposed to UV light for 20, 40, and 60 min at RT. CMBS, after UV treatment, was prepared as described above and was subjected to microbiological and biochemical analysis. The biochemical analysis involved the assay of antioxidants by DPPH free radical scavenging activity (as described above) and the estimation of free thiol groups using Ellman's test.

3.13.1. Estimation of free thiol group by Ellman's test. This method is based on spectrophotometric quantification of the free sulfhydryl (–SH) group. Ellman's reagent (DTNB) reacts with the free-SH group by forming a stable yellow colored



complex, which can be measured spectrometrically at 412 nm.⁴⁷ Briefly, 1 ml of UV-treated CMBS was mixed with 0.5 ml DTNB (0.5 mM in 0.1 mM PBS of pH 7) reagent and allowed to stand for 10 min at RT, and absorbance was measured spectrophotometrically at 412 nm. The relative % decrease in the concentration of free thiol groups was expressed using the formula below,

$$\% \text{ decrease free thiol group} = 100 \times \left[1 - \left(\frac{Ab_t}{Ab_f} \right) \right]$$

where Ab_f denotes the absorbance of fresh CMBS and Ab_t denotes the absorbance of UV-treated samples.

3.14. Statistical analysis

Using the Origin 9.0 program, statistical analysis was performed using ANOVA (One-Way Analysis of Variance). The results of each experiment were obtained in triplicate and are presented as the mean \pm SD. The Tukey test ($p \leq 0.05$) was used to assess the difference between the mean values.

4. Results and discussions

4.1. FTIR analysis

FTIR spectroscopy, using the Attenuated Total Reflectance (ATR) method, is a primary tool for analyzing the interactions between the functional groups of chitosan, pullulan, and TP extract in control CP and CPT intelligent biodegradable films. The control PC film exhibits the characteristic -OH stretching band at 3284 cm^{-1} , as well as C-H symmetric and asymmetric stretching at 2932 and 2889 cm^{-1} , respectively.⁴⁸ Due to chitosan and pullulan polysaccharides, peaks observed at 1642 and 1533 cm^{-1} are due to C-O (amide-I) stretching and N-H (amide-II) bending. 1376 and 1152 cm^{-1} because of -OH bending and C-O-C bridge by asymmetric stretching, respectively.^{49,50} The characteristic peak observed at 850 cm^{-1} is due to the α -glucopyranose units of pullulan,⁵¹ as represented in Fig. 1(a).

Incorporating TD extract into the CS/PL matrix results in the CPT intelligent biodegradable films. The change in -OH stretching frequency to a higher wavelength of 3289–3294 cm^{-1} and the variation in the wavelength of polysaccharide peaks

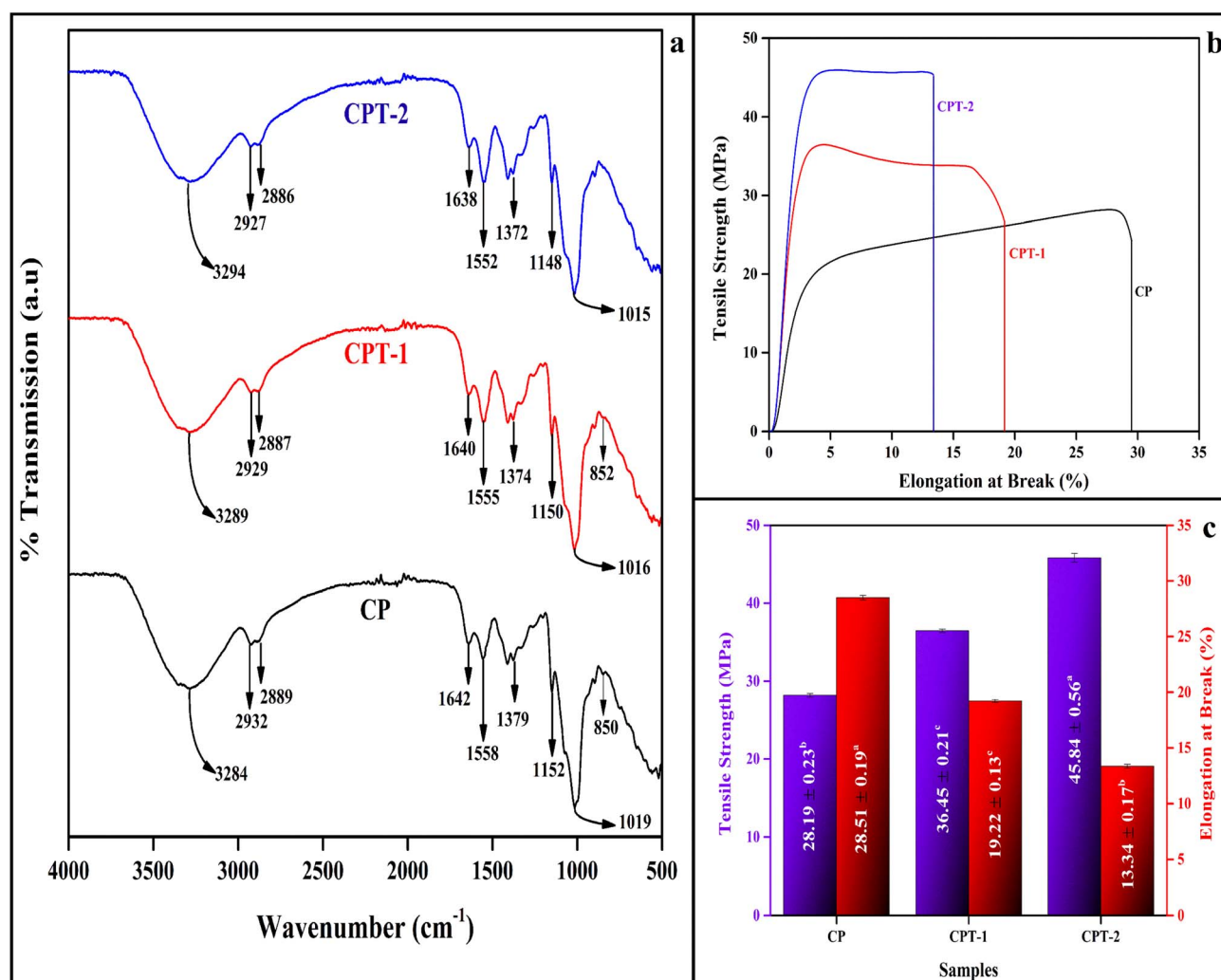


Fig. 1 (a) FTIR analysis, mechanical properties (b) stress vs. strain curve and (c) TS and EB of control CP and CPT intelligent biodegradable films.



suggested intermolecular hydrogen bond formation, attributed to the interaction between the polymer and the active agent.⁵² The disappearance of characteristic α -glucopyranose units at 850 cm^{-1} in the case of CPT-2 intelligent biodegradable films indicated homogeneity between the polymer blend and the active agent.⁵³ Further, the homogenous nature was confirmed by the SEM analysis of CPT intelligent biodegradable films. By forming intermolecular hydrogen bonds, this interaction between the CS/PL and TD confirms the compatibility between the active agent and the polymer matrix.

4.2. Mechanical properties

The CPT intelligent films are fabricated by incorporating *Tradescantia pallida* (TP) anthocyanin extract as an active and intelligent agent in the CS/PU polymer blend. The tensile strength (TS) and elongation at break (EB) of CP and CPT intelligent films were evaluated. The mechanical properties of films used for food packaging become particularly significant as they must withstand the stress of various environmental factors during transportation while preserving the food from mechanical damage. The TS and EB of the control CP film are $28.19 \pm 0.23\text{ MPa}$ and $28.51 \pm 0.19\%$, respectively. Adding TP anthocyanin extract as an active agent increases the tensile strength of CPT-1 and CPT-2 intelligent biodegradable films by 36.45 ± 0.13 and $45.84 \pm 0.56\text{ MPa}$, respectively. However, the elongation at break decreased to $19.22 \pm 0.13\%$ and $13.35 \pm 0.17\%$, respectively, as demonstrated in Fig. 1(c).

The incorporation of anthocyanin-rich polyphenols resulted in a 79% increase in tensile strength (TS) of the CPT-2 intelligent biodegradable film compared to the control CP film. This improvement can be attributed to the interactions between the phenolic groups of the TP extract and the $-\text{NH}_2$ and $-\text{OH}$

functional groups of chitosan (CS), as well as the $-\text{OH}$ groups of pullulan (PU).¹⁸ However, a 47% reduction in elongation at break was observed, likely due to the reduced flexibility of the CPT biodegradable films resulting from these intermolecular interactions.⁵⁴ The CPT intelligent biodegradable films exhibit the TS in the range of 36–45 MPa, superior to the microplastic generating conventional polyethylene (22–31 MPa) and polypropylene (31–38 MPa) plastics, which are widely used commercially in everyday life.³⁹

The mechanical robustness and flexibility of CPT-2 biodegradable films, with a tensile strength (TS) of $45.84 \pm 0.56\text{ MPa}$ and an elongation at break (EB) of $13.35 \pm 0.17\%$, indicate a balanced performance suitable for food packaging applications. The TS value suggests the film can withstand considerable stress without breaking, ensuring durability during storage and handling. Meanwhile, the EB reflects moderate flexibility. Together, these properties enable the film to maintain structural integrity and adapt to the mechanical stresses encountered during packaging, transportation, and consumer use, making it a promising alternative to conventional plastics in sustainable packaging solutions.

4.3. Scanning electron microscopy (SEM) and water contact angle (WCA)

Scanning electron microscopy is the most adaptable and prominent method for determining a biopolymer's homogeneous nature, surface morphology, and phase separation, which is essential for fabricating polymer films.⁵⁵ The micrographs from the SEM analysis were correlated with the mechanical characteristics of the produced films and used to evaluate how active additives cause agglomeration, which further affects the physicochemical characteristics of the

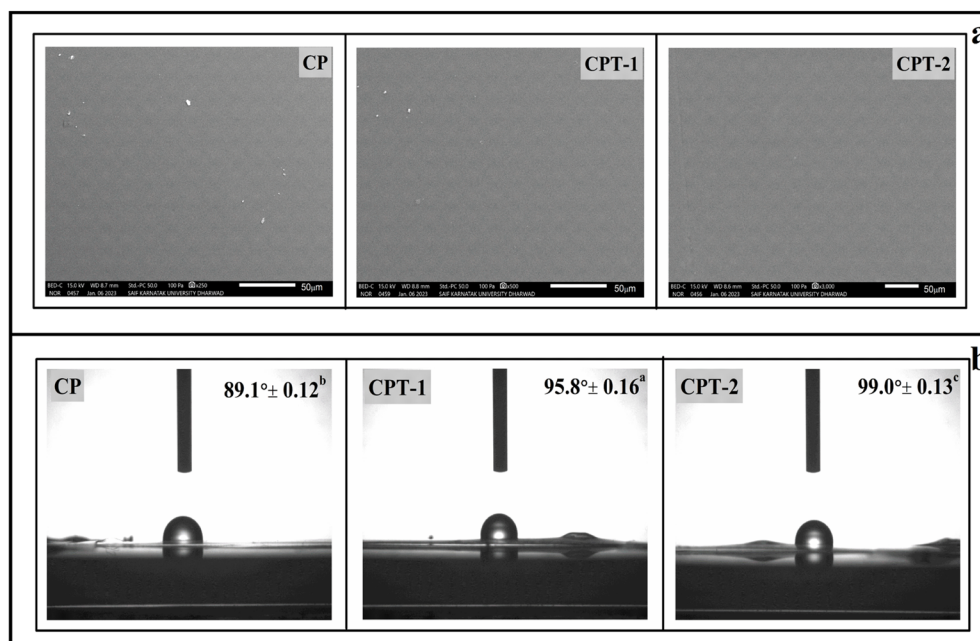


Fig. 2 (a) SEM micrographs and (b) water contact images of control CP and CPT intelligent biodegradable films.



polymer films.^{56,57} In the present study (Fig. 2(a)), the control CP films had uniform surfaces with a few white spots resulting from the plasticizing effect of glycerol. As the TP extract's anthocyanin content increased in CPT intelligent biodegradable films, the white spots on the film surface became less noticeable. This may be due to the formation of intermolecular hydrogen bonds between the polyphenols in the TP extract and the CS, PU, and glycerol components of the polymer blend. Similar results were obtained in the vanillic acid-incorporated CS/PVA blend.³⁹ These findings indicate that homogenous dense surface morphology was relevant in enhancing the tensile strength of CPT intelligent biodegradable films.

The hydrophobic properties of the CPT intelligent biodegradable film were assessed using water contact angle analysis. The films must exhibit a hydrophobic nature to be suitable for use in the food packaging industry. Food deterioration is frequently caused by increased water activity in packaging because it promotes the growth of microorganisms. The control CP films exhibited a contact angle of $89.1^\circ \pm 0.12^\circ$. It may be due to the availability of excess hydroxyl groups through unbonded glycerol molecules. Generally, hydrophilic films exhibit a water contact angle of less than 90° , while hydrophobic films display a water contact angle of more than 90° .⁴⁸ The WCA values of CPT-1 and CPT-2 intelligent biodegradable films rise by $95.8^\circ \pm 0.16^a$ and $99.0^\circ \pm 0.13^c$, respectively, as the concentration of TP extract increases, as represented in Fig. 2(b). The lesser availability of $-OH$ groups to form the hydrogen bond with water molecules caused a decrease in the adhesive nature of intelligent biodegradable films. A similar enhancement in the contact angle was reported by Li-Ting Wu *et al.* by incorporating *Clitoria ternatea* (CT) anthocyanin extracts into gellan gum (G) film.⁵⁸

4.4. Water solubility (WS) and moisture adsorption (MA)

The prepared films were water-resistant, which influenced the physico-mechanical characteristics in food packaging

applications. The fabricated intelligent biodegradable film's water solubility values are presented in Table 1; the control CP films demonstrated a solubility of $22.29 \pm 0.15\%$ during 24 h. With the addition of the TP anthocyanin extract, the solubility of CPT-1 and CPT-2 intelligent biodegradable films decreased by $17.61 \pm 0.13\%$ and $15.31 \pm 0.12\%$, respectively. The reduced solubility of films involving CS and PU may be due to the enhanced surface area of the CS upon contact with PU, which results in intermolecular hydrogen bonding.⁵⁹ However, free unbonded glycerol may lead to greater solubility in the CP film. In contrast, a 7% decrement in the CPT-2 intelligent biodegradable films' solubility compared to the control CP film interaction of polyphenolic groups of TP extract with $-NH_2$ and $-OH$ groups of chitosan, pullulan, and unbonded glycerol.

Packaged food must resist deterioration from moisture adsorption. Therefore, the packing material is crucial for protecting food from microbial harm caused by excessive moisture absorption, which is conducive to microbial development.⁶⁰ Relative humidity is essential for moisture absorption. The fabricated films were evaluated at three relative humidity levels: 19%, 50%, and 75%. According to the data in Table 1, compared to CPT intelligent biodegradable films, the control CP has a significant moisture uptake under all relative humidity levels. The reduction in moisture absorption may be attributed to the addition of TP extract, which is due to the dense surface morphology and reduced availability of $-OH$ groups in CPT intelligent biodegradable films. The findings on water solubility and moisture adsorption suggest that CPT intelligent biodegradable films could serve as a suitable food packaging material.

4.5. Water vapour transmission rate (WVTR) and soil burial test

Water vapour transmission significantly depends on the interaction between polymers and active agents. The homogeneous and dense surface of the fabricated film ensures the entrapment of moisture in packed food and prevents water vapor from the

Table 1 Thickness, water solubility, moisture adsorption, water vapour permeability, and biodegradability control CP and CPT intelligent biodegradable films

	CP	CPT-1	CPT-2
Thickness (μm)	80 ± 4^a	80 ± 3.5^b	90 ± 2^c
Water solubility (%)	22.29 ± 0.16^a	17.602 ± 0.13^b	15.313 ± 0.12^c
Moisture adsorption (%)			
RH 19%	3.16 ± 0.12^b	2.76 ± 0.13^a	2.24 ± 0.11^c
RH 50%	4.53 ± 0.11^c	3.86 ± 0.14^a	3.13 ± 0.13^b
RH 75%	5.51 ± 0.14^c	5.09 ± 0.17^a	4.61 ± 0.15^b
WVTR ($\text{g per m}^2 \text{ per 24 h}$)	34.61 ± 0.25^c	30.02 ± 0.31^a	26.55 ± 0.28^b
Degradation in soil (%)			
Day-10	5.32 ± 0.13^a	4.79 ± 0.11^b	3.23 ± 0.12^c
Day-20	16.34 ± 0.21^a	10.02 ± 0.18^b	8.91 ± 0.15^c
Day-30	25.34 ± 0.24^a	19.64 ± 0.20^c	16.82 ± 0.21^b
Antioxidant activity (%)			
DPPH	58.26 ± 1.59^c	75.16 ± 1.55^b	89.81 ± 1.47^a
ABTS	54.97 ± 1.52^a	69.73 ± 1.48^c	85.14 ± 1.46^b



environment from entering the package. In the present study, the control CP film showed the highest transmission rate of water, 34.61 ± 0.25 g per m² per 24 h, due to the presence of polar groups in CS (–NH₂ and –OH), PU (–OH), and glycerol (–OH). As the anthocyanin content of TP extract increased in CPT-1 and CPT-2 intelligent biodegradable films, intermolecular hydrogen bonding reduced the availability of free polar groups, thereby restricting water molecules from passing through the film layer. Hence, CPT-1 and CPT-2 intelligent biodegradable films exhibit lower WVTR values of 30.05 ± 0.31^a and 26.55 ± 0.28^b g per m² per 24 h is due to filling the structural void between polymers with the TP anthocyanin extract as the active agent. A similar decline in the WVTR value resulted in chitosan films with the addition of starchy powder from white turmeric rhizomes.⁶¹

The packaging materials in the market are primarily made of plastic, and after their usefulness, they end up in landfills, generating microplastics that have profound health implications. Hence, the fabricated intelligent biodegradable film should be degradable and eco-friendly. Soil burial tests were conducted for 10, 20, and 30 d to evaluate the biodegradability of the CP and CPT intelligent biodegradable films. The soil microbes initially colonize the film after burial, generating film fragments. The enzymatic function of extracellular hydrolases depolymerized the polymer content, resulting in the creation of CO₂ and water. These benefits of microbial activity in soil promote degradation and improve the nutrient cycle.⁶² After 30 d, control CP films exhibited a $25.34 \pm 0.24\%$ degradation. It is higher than the degradation rates of CPT-1 ($19.64 \pm 0.2\%$) and CPT-2 ($16.82 \pm 0.21\%$) in intelligent biodegradable films, as mentioned in Table 1. CPT intelligent biodegradable films express enhanced antibacterial activity due to the incorporated anthocyanin content of TP extract, restrict the colonization of microorganisms, and slow down the degradation rate compared to control CP films. The rate of degradation of fabricated CP and CPT intelligent films is indicative of the environmental sustainability, considering CPT intelligent biodegradable films as an alternative to conventional PE packaging.

4.6. Antimicrobial activity against foodborne pathogens

The antimicrobial ability of packaging materials is essential from a consumer's health perspective. According to the World Health Organisation (WHO) report, over 200 diseases are caused by unsafe food, *i.e.*, contaminated by harmful foodborne bacteria and fungi.⁶³ In the present study, *S. aureus* (ATCC 6538), *B. subtilis* (MTCC 13343) (Gram +ve), *E. coli* (ATCC 10799) (Gram –ve), and *C. albicans* (ATCC 24433) (fungi) are considered for analysis of the antimicrobial activity of the CP and CPT intelligent biodegradable films. Literature studies show evidence that *S. aureus* causes foodborne illness mainly due to consuming ready-to-eat foods contaminated from the nasopharynx, which produces enterotoxins leading to vomiting and diarrhea. *Bacillus* pathogens often cause food poisoning by contaminating food.^{64,65}

Similarly, toxins produced by spoilage due to *E. coli* can affect the human central nervous system and kidneys upon

consumption.⁶⁵ For all these reasons, food contaminated by foodborne microorganisms is dangerous for consumption. Intelligent biodegradable packaging films that exhibit activity against foodborne pathogens are necessary to prevent spoilage and ensure consumer health. The control CP films demonstrated antimicrobial activity against all tested pathogens, attributed to the inherent antimicrobial nature of chitosan. The inhibition zones measured 18 ± 0.17 mm and 14 ± 0.11 mm for Gram-positive *Bacillus subtilis* and *Staphylococcus aureus*, 14 ± 0.17 mm for Gram-negative *Escherichia coli*, and 13 ± 0.11 mm for the fungal strain *Candida albicans*.^{39,48} As shown in Fig. 3, the CPT-2 intelligent biodegradable films exhibited enhanced antimicrobial efficacy, with inhibition zones of 26 ± 0.24 mm and 17.5 ± 0.18 mm for *B. subtilis* and *S. aureus*, 18.5 ± 0.15 mm for *E. coli*, and 15 ± 0.12 mm for *C. albicans*, respectively.

The extensive zone of inhibition resulting from the CPT-2 intelligent biodegradable films compared to the control CP film is due to the polyphenolic proportion present in the CPT-2, specifically the anthocyanin content of the TP extract. Gram-positive and Gram-negative bacteria have well-protected cell walls composed of peptidoglycan, which protects against osmotic pressure. The anthocyanin (polyphenol) content of TP extract in CPT-2 intelligent biodegradable active films inhibits bacterial growth by disrupting phospholipid or lipid bilayers in the cell wall, thereby enhancing membrane permeability and altering ion transport.^{66,67} Hence, the growth suppression of pathogenic cells was ensured by an excellent zone of inhibition exhibited by the CPT-2.

These findings demonstrate that the fabricated CPT-2 intelligent biodegradable films may be employed in food packaging to extend shelf life without the risk of food deterioration and foodborne pathogen-related illnesses.

4.7. Antioxidant activity

The imbalance between the antioxidant defense and free radical generation is the cause of oxidative stress. Oxidative stress initiates damage to molecular species, such as lipids and proteins.⁶⁸ Free radicals are highly reactive species capable of causing cell damage in biologically relevant carbohydrates, lipids, and proteins, and are also a contributing factor to homeostatic disruption.^{69,70} During fresh meat processing, antioxidants are added to counteract oxidation, which can cause spoilage.⁷¹ Hence, antioxidant activity is necessary for food packaging materials to avoid directly incorporating antioxidants into food. DPPH and ABTS scavenging assays were used to evaluate the antioxidant activity of CP and CPT intelligent biodegradable active films.

In Table 1, control CP films exhibit $58.26 \pm 1.59\%$ and $54.9 \pm 1.55\%$ of antioxidant activity in DPPH and ABTS assays, respectively. Due to the anthocyanin content present in the TP extract, CPT-2 intelligent biodegradable films exhibit activity of $89.81 \pm 1.47\%$ and $85.14 \pm 1.52\%$. The polyphenolic content in CPT-2 intelligent film may initiate the chain-breaking mechanism, fulfilling the additional requirement of electrons for free radicals as they act as a primary antioxidant donor.⁷² Through this phenomenon, polyphenols help maintain a balanced



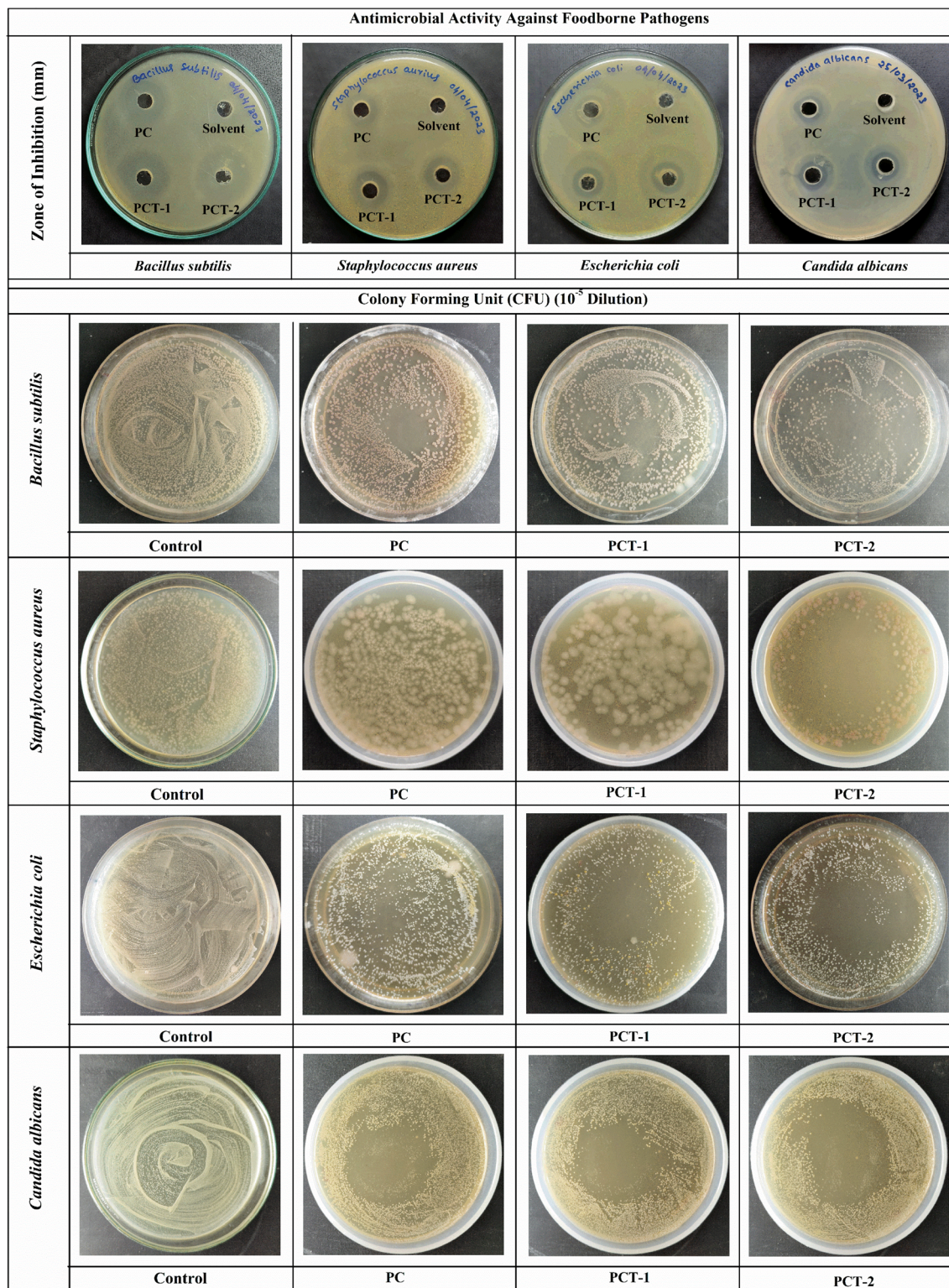


Fig. 3 Zone of inhibition and CFU against *S. aureus*, *B. subtilis*, *E. coli*, and *C. albicans* for control CP and CPT intelligent biodegradable films.

oxidative stress, and DPPH and ABTS assay analysis confirmed that the fabricated CPT intelligent biodegradable films effectively preserve packed food from oxidative deterioration. Similar enhancements in antioxidant activity have been reported by

Amina Sadi *et al.*⁷³ for incorporating anthocyanin content from red cabbage into a CMC/gelatin polymer blend⁵¹ and by Gabriel da Silva Filipini *et al.* for methylcellulose films with *Syzygium cumini* skin extract.⁷⁴



4.8. Preservation of chicken meat and biochemical assessment of its nutrients

Around the world, most people regularly include eggs and chicken in their diet due to their rich source of protein. According to a United Nations report by the Food and Agriculture Organization, 73 million tons of eggs and 100 million tons of chicken were produced worldwide in 2016.⁷⁵ Due to human lifestyle changes, urbanization, and growing populations, demand increases rapidly each year.^{76,77} Poultry is the industry's second-largest source of meat, meaning that poultry products are a significant contributor to foodborne illnesses. Meeting the market's demand for well-preserved chicken meat in the face of microbial attacks that cause spoilage is a primary concern nowadays. Therefore, the current study focuses on using sustainable CPT-2 intelligent biodegradable film packaging as an alternative to plastic packaging to protect chicken meat from foodborne microbe spoilage.

The chicken was packed in plastic (PE), control-CP film, and CPT intelligent biodegradable film to evaluate the effectiveness of packaging. The chicken had been packaged and stored at room temperature, including with *S. aureus* and *E. coli* foodborne pathogens for 24, 48, and 72 h. Following the incubation period, 2 g of chicken flesh was blended for 0.5 min using a tissue homogenizer (Labico, LT-451, Labtech Instruments, India) with 18 ml of 0.9% saline, and then for an additional 0.5 min using a vortex. The mixture was passed through a single, four-layered muslin fabric filter. The CMBS was further subjected to biochemical characterizations to evaluate nutritional levels.

4.8.1. Microbiological analysis

4.8.1.1. Total bacterial count (TBC). Since chicken meat is very perishable and readily contaminated by microorganisms, it undergoes decontamination by modifying its nutritional value. Plastics are mainly used for packaging meat products, and packaging waste accounts for a significant portion of municipal waste. Single-use plastic packaging degrades meat products since it lacks antibacterial and antioxidant properties. This investigation employed plastic, control CP, and CPT-2 intelligent biodegradable packaging film to package chicken meat. Instead of refrigeration to prolong its shelf life, the packaged chicken flesh was kept at room temperature for 24, 48, and 72 h to evaluate the CPT intelligent biodegradable film's preservation capacity. The overall bacterial growth was expressed as log CFU per g of chicken, with a fresh chicken bacterial count of 4.956 log CFU per g. According to the International Commission on Microbiological Specifications for Foods (ICMSF), the permissible microbiological limit for edible meat is 7 log CFU per g.⁷⁸

Microbial growth data for three packaging types are presented in Fig. 4 and Table S3. After 24 hours of incubation at room temperature, chicken packed in conventional plastic showed a bacterial count of 6.55 log CFU per g. This elevated count, attributed to plastic's inability to inhibit microbial activity, led to spoilage within a day. In contrast, CP films, leveraging the natural antibacterial properties of chitosan, preserved chicken freshness for a longer duration. Notably, chicken packed with CPT-2 intelligent biodegradable films, which incorporate polyphenolic TP extract and anthocyanins,

Presevation Studies Chicken Samples with PE (Polyethylene), CP, and CPT-2 Smart Biodegradable film for 24 h, 48 h, and 72 h at Room Temperature												
	Total Bacterial Count (TBC)			Chicken Samples Included with Foodborne Pathogens	Total Bacterial Count (TBC) on inclusion of <i>S. Aureus</i>			Total Bacterial Count (TBC) on inclusion of <i>E. Coli</i>				
	PE	CP	CPT-2		PE	CP	CPT-2	PE	CP	CPT-2		
24 hours Dilutions 10 ⁻¹				24 hours Dilutions 10 ⁻²								
48 hours Dilutions 10 ⁻²					48 hours Dilutions 10 ⁻³							
72 hours Dilutions 10 ⁻³						72 hours Dilutions 10 ⁻⁴						
					72 hours Dilutions 10 ⁻⁴							

Fig. 4 Total bacterial count (TBC) of chicken meat preservation carried out in PE, control CP, and CPT-2 intelligent biodegradable films.



achieved an extended shelf life of up to two days, with microbial growth reaching 6.99 log CFU per g. The enhanced preservation effect is likely due to altered ion transport and lipid bilayer disruption in microbial cells. Comparable results have been reported by S. R. Kanatt *et al.*, demonstrating that the inclusion of amaranthus leaf extract (ALE) anthocyanins in PVA packaging films extends the shelf life of chicken meat.⁷⁹

4.8.1.2. Total bacterial count by inclusion of foodborne pathogens. To analyze packaging effectiveness against contamination involving aerobic mesophilic bacteria, such as *S. aureus* and *E. coli*, in the present study. Chicken packaging was preserved at room temperature, and the aerobic mesophilic bacteria grew and thrived in the moderate temperature range of 20–45 °C.⁸⁰ When contaminated food is consumed, these microorganisms can cause severe health issues.⁸¹ When chicken, seafood, and red meat are contaminated with *S. aureus*, it frequently leads to food poisoning.⁸² The plastic packaging has a log CFU per g of chicken count ranging from 7.36 to 10.89. However, the CPT-2 intelligent biodegradable films packaging has a log CFU per g of chicken ranging from 5.67 to 8.84 for 24 to 72 h. Similarly, *E. coli* also negatively affects human health and is used as a hygiene indicator in poultry-related food manufacturers.⁸³ After 72 hours of incubation, CPT-2 intelligent biodegradable packaging showed 8.97 log CFU per g compared to plastic packaging, which had 10.87 log CFU per g.

Both pathogenic results suggested that the CPT-2 intelligent biodegradable packaging is superior to single-use plastic packaging. The anthocyanin content in the active agent successfully inhibits microbial growth. The active agent in the polymer network effectively acted against foodborne pathogens by penetrating the cell wall, resulting in suppression of pathogenic growth. Intelligent biodegradable packaging successfully prolongs the shelf life by ensuring the quality of the meat.⁸⁴

4.8.2. Freshness monitoring of chicken and estimation of ammonia and ammonium salts. The anthocyanin content of TP extract serves as an active ingredient in the current investigation of CPT-2 intelligent biodegradable packaging film. According to the anthocyanin concentration, the films exhibit colour changes in response to alterations in pH. The anthocyanin content in an acidic pH medium causes CPT-2 intelligent biodegradable films to turn pink, whereas it turns yellow in a basic pH medium. Food should be maintained at a proper pH level for the health benefit of the consumer. In fact, due to transportation errors and improper packaging, food comes into contact with microbial attacks that are invisible to the naked eye. The growth of microorganisms on food materials and changes in food quality, consistency, and composition due to chemical reactions may all influence pH variations.⁸⁵ Because of this, the fabricated biodegradable films serve as both a freshness indicator and a means of food preservation. Freshly sliced chicken with a pH of 6.17 was evaluated by a freshness monitor in Fig. 5. Following 48 h, a gradually turning yellow colouration was observed, and the pH changed to a slightly basic 7.46. At the 72-hour assessment of the freshness of the chicken meat, the CPT-2 intelligent biodegradable packaging film turns yellow, signifying that the meat has spoiled and is no longer suitable to be consumed as a source of protein. These visual indicators of food freshness help consumers identify suitable food items that will not harm their health upon consumption.

The primary sources impacted by the microbial attack that causes spoilage during storage include the availability of lactic acid, glucose, amino acids, and other nitrogenous substances used as energy sources in meat.^{86,87} Ammonia (NH₃) and methylamines are the amine sources obtained by biomolecules due to microbial and enzyme decomposition of meat products.⁸⁸ As a result, as meat spoilage is initiated, its total volatile basic nitrogen (TVB-N) value increases.⁸⁹ In the current

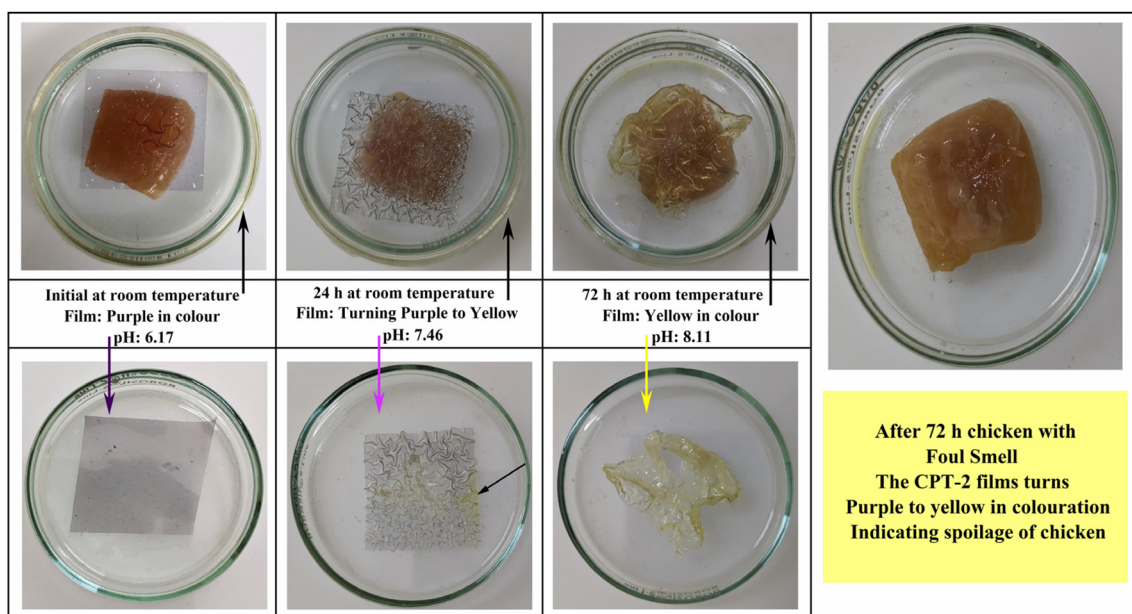


Fig. 5 Visual identification of spoilage of chicken meat using CPT-2 intelligent biodegradable film as an intelligent indicator of pH variation.



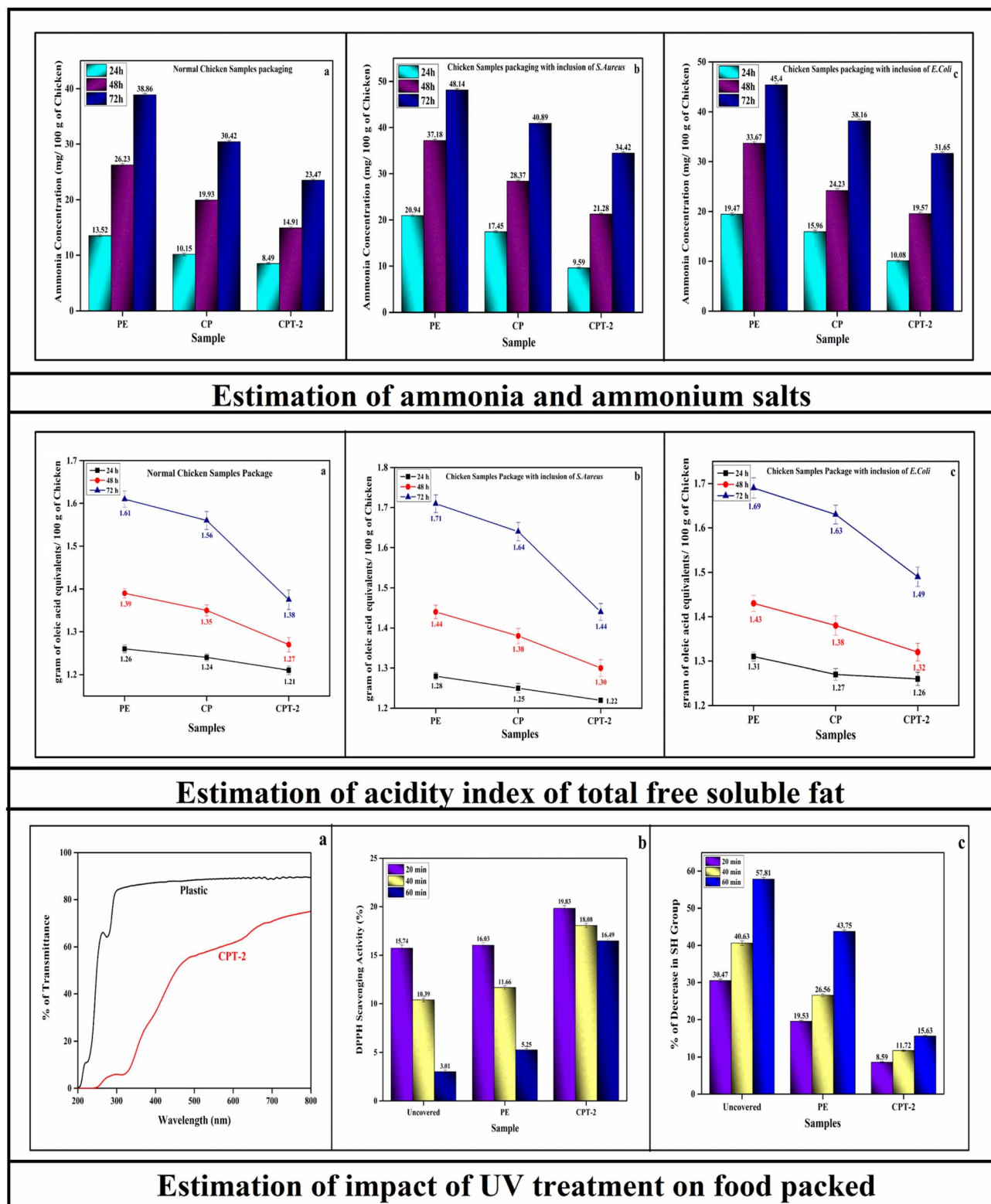


Fig. 6 Spoilage analysis (TVB-N, acidity index, and impact of UV light) of chicken by packaging in PE, control CP, and CPT-2 intelligent biodegradable films.

investigation, the TVB-N variation in chicken meat packed in plastic, control CP, and CPT-2 intelligent biodegradable films is presented in Fig. 6. During a 24 h packing period, chicken

packaged in plastic has a TVB-N value of 13.52 mg per 100 g. However, chicken packaged in CPT-2 intelligent biodegradable film has a value of 8.49 mg per 100 g. It may be due to the



anthocyanin content in CPT-2 intelligent biodegradable film's significant antibacterial and barrier qualities, which prevent microbial spoiling and protein degradation, attributed to a reduced TVB-N value.⁸⁷ Senapati and Sahu (2020)⁹⁰ state that the acceptable TVB-N levels for dietary chicken meat fall between 25 to 28 mg per 100 g. After 72 h of preservation, the CPT-2 intelligent packaging chicken meat has a TVB-N value of 23.47 mg per 100 g. At the same time, plastic packaging contains 38.86 mg per 100 g, while control CP film contains 30.42 mg per 100 g. Considering these findings, it can be concluded that CPT-2 intelligent biodegradable packaging effectively extends the shelf life of chicken meat by preventing spoilage due to the production of ammonia and ammonium salts over 72 h at room temperature.

The impact of foodborne microorganisms, specifically *Staphylococcus aureus* and *Escherichia coli*, on total volatile basic nitrogen (TVB-N) levels, an indicator of spoilage, was investigated in chicken meat packaged using plastic, control CP, and CPT-2 intelligent biodegradable films. *E. coli* is a Gram-negative bacteria well known for food spoilage on contamination; these pathogens utilize mixed acids and trimethylamine oxide (TMAO) to produce slime production with an off odour by producing formic acid, ethanol, and trimethylamine (TMA) as spoilage products.^{86,91} Hence, the plastic packaging lacked resistance against microbial growth, and chicken meat spoiled within a day at room temperature (19.47 mg per 100 g). Due to their inherent antimicrobial properties, CPT-2 intelligent biodegradable films restrict microbial growth and extend the shelf life to 2 days at room temperature (19.57 mg per 100 g). In the case of *S. aureus* contamination, these pathogens utilize glucose, amino acids, and TMAO to produce souring and off-odour in meat by producing acetate, pyruvate, succinate, and TMA as spoilage products.^{87,92} Similar TVB-N values are observed with plastic packaging, acceptable freshness up to a day (20.94 mg per 100 g), and extension up to 48 h with CPT-2 intelligent biodegradable packaging (21.28 mg per 100 g). The slight enhancement in the TVB-N values with *S. aureus* compared to *E. coli* may be due to the thick outer peptidoglycan layers of Gram +ve bacteria, which make it difficult to penetrate the cell of bacteria by an active agent to neutralize the spoilage action.³⁹

The rapid increase in chicken meat's TVB-N value reduces the meat's glucose content.⁹³ The microbial deterioration of meat products is associated with the conversion of TMAO, which results in the formation of spoilage-initiating byproducts such as TMA, DMA, and formaldehyde. Hence, packaging material should have antimicrobial properties to avoid spoilage. Foodborne pathogens (*S. aureus* and *E. coli*) adapt to facultative anaerobic growth conditions. In the absence of oxygen, it switches to fermentation and proliferates in the presence of oxygen. Hence, the packaging material has good barrier properties. Incorporated anthocyanin content creates a compact structure with a polymer blend and enhances the barrier properties.⁹⁴ Therefore, due to the improved antimicrobial ability and barrier nature of CPT-2 intelligent biodegradable films with the incorporation of anthocyanin content, TP extract is an active agent, extending the shelf life of packed chicken

meat compared to traditional plastic packaging at room temperature.

4.8.3. Estimation of acidity index of total free soluble fat.

Meat spoilage is initiated by microbial attack on nutrient substances, such as fatty acids, free amino acids, and sugars, which release undesirable volatile compounds (VOCs).⁹⁵ Meat spoilage not only depends on microbial spoilage but also on non-microbial spoilage, such as the generation of toxic metabolites from lipid peroxidation and a reduction in the availability of essential fatty acids, caused by variations in flavor and coloration of meat.⁹⁶ When lipid peroxidation begins, hydroperoxides are the primary product. Estimating hydroperoxides during spoilage gives essential information on the packaging capacity of the packaging material. The foul odour and flavour of spoilage are due to the breakdown of hydroperoxides into several organic compounds.⁷⁸ The present study estimated the acidity index of total free soluble fat by considering the oleic acid (hydroperoxide) over time at room temperature, as shown in Fig. 6.

Oleic acid is a monounsaturated omega-9 fatty acid (C18:1) commonly found in animal fats and vegetable oils.⁹⁷ The plastic packaging, which is similar to normal chicken packaging and includes foodborne pathogens, exhibits a comparatively higher acidity index of total free soluble fat than chicken packed in CP and CPT-2 intelligent biodegradable films, respectively. Lipid oxidation primarily occurs due to photo- and enzyme-catalyzed oxidation.^{97,98} Photooxidation mainly occurs in the presence of light and oxygen. Enzymatically catalyzed oxidation is caused by the action of lipases produced by microorganisms, such as bacteria and molds, which arises due to spoilage. Lipolysis forms glycerol and fatty acids by the hydrolysis of triglycerides.⁸⁷ Hence, the lack of antimicrobial properties of plastic packaging attributed to an increase in the acidity index of total free soluble fat by 1.61 g of oleic acid equivalents per 100 g of meat in normal chicken packaging, and by including foodborne pathogens, it increased by 1.71 and 1.69 g of oleic acid equivalents per 100 g of meat, respectively. In the case of CP and CPT-2 intelligent biodegradable packaging, the inherent antimicrobial properties, due to the anthocyanin content in the packaging environment, adversely impact pathogen growth. Hence, the acidity index is restricted by 1.56 and 1.38 g of oleic acid equivalents per 100 g of meat in normal packaging. On inclusion of *S. aureus* 1.64 and 1.44 g of oleic acid equivalents per 100 g of meat, while with *E. coli* 1.63 and 1.49 g of oleic acid equivalents per 100 g of meat, respectively. Therefore, reducing lipid oxidation in CPT-2 intelligent packaging prolongs the shelf life of packed chicken compared to control CP as well as plastic packaging.

4.9. Impact of UV treatment on packaged chicken meat

The UV radiation present in food is detrimental to consumption because it activates free radicals through the initiation of photochemical reactions. These undesirable photochemical reactions damage food products by digesting protein, oxidizing lipids, causing color changes, reducing vitamin content, and producing foul odors.^{99,100} The packaging films should have



better UV shielding properties to protect the food from these unwanted photochemical reactions caused by UV impact. The impact of UV radiation on chicken meat was analyzed by determining the free thiol (–SH) groups present after 20, 40, and 60 min of irradiation.

Sulfur containing amino acids, cysteine, and glutathione are the compounds that act as antioxidants in meat products, consisting of the thiol groups (–SH). These protect meat products from oxidative stress and prevent lipid and protein oxidation.¹⁰¹ Hence, by assessing the impact of UV radiation on thiol groups present in the meat as uncovered, covered with plastic, and CPT-2 intelligent packaging film. According to Fig. 6, which illustrates the percentage of UV radiation transmitted through plastic and CPT-2 intelligent biodegradable films, UV-C penetration at 280 nm was 65% in plastic and 5% in CPT-2 intelligent biodegradable films. Similarly, the CPT-2 intelligent biodegradable film's UV-B (at 320 nm) and UV-A (at 400 nm) area penetration of around 6% and 33%, respectively, was noticeably lower than that of the plastic film, which was 85% and 88%, respectively. UV-B and UV-A radiation frequently cause lipid oxidation, vitamin breakdown, and decolorization of meat products.¹⁰² According to this finding, the CPT-2 intelligent biodegradable films prevented the meat items from spoiling by effectively acting as a UV barrier.

UV irradiation affected the free thiol groups, and Ellman's test was used to determine the thiol groups in the chicken meat after 20, 40, and 60 minutes of irradiation. As a result of successive thiol groups oxidizing when exposed to UV radiation, thiol groups oxidized in proteins led to the formation of –S–S– (disulfide) bonds and also caused the protein crosslinking.¹⁰³ The protein network is disrupted by the polymerization of proteins, which significantly affects softness, and by the uncontrolled oxidation of thiol caused by UV light, which causes food to spoil.¹⁰⁴ The percentage of decrease in the thiol groups after UV irradiation was presented in Fig. 6. After 60 minutes of UV irradiation, the uncovered chicken meat declined by 50% in free thiol groups; in contrast, the chicken enclosed in PE packaging was almost as close to the uncovered chicken, with 43.75% of the drop indicating impotent behaviour of PE packaging against UV radiation. After 60 minutes of radiation, the CPT-2 intelligent biodegradable film packaging effectively blocks UV penetration, reducing the oxidation of thiol groups by only 15.63%. These findings confirm the optical properties of CPT-2 intelligent biodegradable films, demonstrating their suitability as a packaging material with UV barrier properties.

In the biological system, UV light produces reactive oxygen species (ROS), which interact with free thiol groups to initiate oxidative damage. The depletion of thiol groups correlates with the antioxidant capacity of meat products. In the current investigation, the DPPH scavenging assay revealed that fresh chicken meat has an antioxidant activity of 20.42%. After 60 minutes of UV irradiation, the antioxidant capacity of uncovered meat remained at only 3%, while PE-covered meat reached 5.25%. In the case of CPT-2 intelligent biodegradable films covering chicken meat, it retained its antioxidant capacity at 16.49%. Antioxidants, including phenol components, shield

proteins and lipids from oxidation, which might disrupt the disulfide network.^{105,106} The polyphenolic content present in CPT-2 intelligent biodegradable film, as well as the anthocyanin content of TP extract, is also capable of protecting proteins and lipids from oxidation. Therefore, chicken meat covered with CPT-2 intelligent biodegradable films showed a slight decrease in antioxidant capacity and a percentage decline in thiol content compared to PE packaging. According to Sisse Jongberg *et al.*, a phenolic extract from green tea can prevent thiol oxidation in chicken meat.¹⁰⁴

The enhancement in physicochemical properties and the superior capability of CPT-2 intelligent biodegradable films against microbial attacks to preserve the nutritional value of packed meat at room temperature confirm that the fabricated biodegradable film of a chitosan and pullulan blend incorporating TP extract as an active agent is a suitable alternative to PE packaging and holds promise for a sustainable future.

5. Conclusions

Poultry represents one of the significant sources of meat production, where effective preservation is essential to satisfy consumer demand. However, the meat packaging industry continues to rely predominantly on single-use plastic materials, contributing substantially to environmental pollution. In this context, the present study developed chitosan/pullulan-based CPT-2 intelligent biodegradable films incorporating 200 mg of *Tradescantia pallida* (TP) extract as an active and intelligent packaging alternative to conventional polyethylene (PE) films. The CPT-2 intelligent biodegradable films demonstrated enhanced preservation performance by suppressing bacterial growth, the release of ammonia and ammonium salts, and lipid oxidation regarding free fatty acid content at room temperature, thereby extending the shelf life of chicken up to 2 days at room temperature, which is an environment favourable for foodborne pathogen activity compared to refrigeration temperature. In contrast, PE packaging maintained quality for only 1 day under similar conditions.

Despite their promising functionality and complete biodegradability, cost optimization remains a significant challenge for the commercialization of CPT-2 intelligent biodegradable films. The current development corresponds to a Technology Readiness Level (TRL) of 4–5, indicating successful laboratory validation but requiring further investigation to advance toward TRL-9 for industrial-scale application as a cost-effective substitute for single-use plastics (SUP). Considering that all polymers used are food-grade and that *Tradescantia pallida* (TP) is a readily available, drought-tolerant, widely cultivated, and simple-to-propagate plant through cuttings, it offers a sustainable and economical source of active agents. Therefore, future research should focus on optimizing fabrication techniques and production costs to enhance scalability and economic feasibility. Overall, this study confirms that *T. pallida* extract serves as a viable and cost-effective intelligent component in biodegradable packaging films, highlighting the need for continued innovation to replace SUPs and ensure a sustainable future for the upcoming generation.



Consent for publication

All authors agreed to publish the paper.

Author contributions

Manjunath P. Eelager: conceptualization; data curation; formal analysis; methodology; visualization; resources; funding; software; roles: writing – original draft; review & editing. Saraswati P. Masti: the corresponding author; investigation; supervision; validation; writing – review & editing, visualization. Suhasini Madihalli: investigation; formal analysis. Ravindra B. Chougale: formal analysis. Nagarjuna Prakash Dalbanjan: data curation, resources, formal analysis. S. K. Praveen Kumar: formal analysis.

Conflicts of interest

The authors declare that they have no competing interests.

Data availability

The data supporting this study's findings are available from the corresponding author upon reasonable request.

Supplementary information: tables presenting the optimization parameters for the base matrix (CS/PU), composition details of the control (CP) and CPT intelligent biodegradable films, zone of inhibition and colony-forming unit (CFU) analysis of CP and CPT films against various foodborne pathogens, total bacterial count (TBC) of chicken meat (normal and inoculated with *S. aureus* and *E. coli*) packaged in PE, CP, and CPT-2 films, and a comparative assessment of tensile strength, antimicrobial, and antioxidant properties among different intelligent compounds incorporated into the polymer blend films. Additionally, it also consists of the results and discussion related to the total protein content, antioxidant capability of chicken meat after the incubation period, and the tentative cost analysis and life cycle assessment (LCA) of the CPT-2 intelligent packaging film. See DOI: <https://doi.org/10.1039/d5fb00349k>.

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References

- 1 P. J. Landrigan, H. Raps, M. Cropper, C. Bald, M. Brunner, E. M. Canonizado, D. Charles, T. C. Chiles, M. J. Donohue and J. Enck, *Ann. Glob. Health*, 2023, **89**, 23.
- 2 S. *Food Packaging Market Size, Trends & Growth [2030], Market Research Report*, <https://www.fortunebusinessinsights.com/industry-reports/food-packaging-market-101941>.
- 3 B. Dewangan, M. Mittal and M. P. Eelager, *Sustainable Chem. Pharm.*, 2025, **46**, 102092.
- 4 S. Ayu Rafiqah, A. Khalina, K. Zaman, I. Tawakkal, A. Harmaen and N. M. Nurrazi, *Bio-Based Packaging: Material, Environmental and Economic Aspects*, 2021, pp. 335–351, DOI: [10.1002/9781119381228.ch19](https://doi.org/10.1002/9781119381228.ch19).
- 5 S. Sid, R. S. Mor, A. Kishore and V. S. Sharanagat, *Trends Food Sci. Technol.*, 2021, **115**, 87–104.
- 6 M. P. Eelager, N. P. Dalbanjan, S. Madihalli, M. Madar, N. P. Agadi, K. Korganokar and B. K. Kiran, *Sustainable Futures*, 2025, **10**, 101208.
- 7 S. Madihalli, S. P. Masti, M. P. Eelager, R. B. Chougale, L. K. Kurabetta, A. A. Hunashyal, N. P. Dalbanjan and S. K. Praveen Kumar, *Food Biosci.*, 2024, **62**, 105492.
- 8 S. Madihalli, S. P. Masti, M. P. Eelager, R. B. Chougale, B. M. Anilkumar and A. N. Priyadarshini, *Int. J. Biol. Macromol.*, 2025, **303**, 140611.
- 9 N. P. Dalbanjan, K. Korgaonkar, M. P. Eelager, B. N. Gonal, A. J. Kadapure, S. B. Arakera and S. K. Praveen Kumar, *Nano TransMed*, 2025, **4**, 100091.
- 10 V. D. Hiremani, N. Goudar, S. Khanapure, T. Gasti, M. P. Eelager, S. S. Narasagoudr, S. P. Masti and R. B. Chougale, *J. Food Meas. Charact.*, 2023, **17**, 1548–1561.
- 11 K. Kraśniewska, K. Pobiega and M. Gniewosz, *Int. J. Food Eng.*, 2019, **15**, 20190030.
- 12 G. Mugnaini, C. Resta, G. Poggi and M. Bonini, *J. Colloid Interface Sci.*, 2021, **592**, 430–439.
- 13 N. P. Dalbanjan, M. P. Eelager and S. S. Narasagoudr, *Food Humanity*, 2024, **3**, 100366.
- 14 P. R. Vuddanda, M. Montenegro-Nicolini, J. O. Morales and S. Velaga, *Eur. J. Pharm. Sci.*, 2017, **96**, 290–298.
- 15 M. Chen, T. Runge, L. Wang, R. Li, J. Feng, X.-L. Shu and Q.-S. Shi, *Carbohydr. Polym.*, 2018, **200**, 115–121.
- 16 V. Shukla, G. Kandeepan, M. R. Vishnuraj and A. Soni, *Agric. Res.*, 2016, **5**, 205–209.
- 17 Y. Qin, Y. Liu, X. Zhang and J. Liu, *Food Hydrocolloids*, 2020, **100**, 105410.
- 18 T. Gasti, S. Dixit, O. J. D'souza, V. D. Hiremani, S. K. Vootla, S. P. Masti, R. B. Chougale and R. B. Malabadi, *Int. J. Biol. Macromol.*, 2021, **187**, 451–461.
- 19 N. Hidayah, M. S. Hawa, B. Purwono and S. N. Rahmadhia, *Trends Sci.*, 2025, **22**, 10077.
- 20 M. Butnariu, Á. Fernández Ochoa, A. Segura Carretero and M. d. I. L. Cádiz Gurrea, *Front. Biosci.*, 2022, **27**, 197, DOI: [10.31083/j.fbl2706197](https://doi.org/10.31083/j.fbl2706197).



- 21 F. Imtiaz, M. Islam, H. Saeed, A. Ahmed, F. K. Hashmi, K. M. Khan, U. I. Dar, K. Ullah, S. M. Rana and B. Saleem, *Pharmaceutics*, 2022, **14**, 2578.
- 22 D. Yıldız, *Global Poultry Industry and Trends*, <https://www.feedandadditive.com/global-poultry-industry-and-trends/>.
- 23 ICPE, *Packaging of Meat and Poultry Products*, https://www.icpe.in/icpefoodnpackaging/pdfs/19_meat.pdf.
- 24 L. Cabernard, S. Pfister, C. Oberschelp and S. Hellweg, *Nat. Sustainability*, 2022, **5**, 139–148.
- 25 M. Madar, V. Srinivasan, M. P. Eelager, N. P. Dalbanjan, K. N. Kummur, V. Nayak, M. M. Basanagouda and A. Sidarai, *Tetrahedron*, 2025, **185**, 134796.
- 26 V. A. Pereira Jr, I. N. Q. de Arruda and R. Stefani, *Food Hydrocolloids*, 2015, **43**, 180–188.
- 27 M. P. Eelager, S. P. Masti, S. Madihalli, N. Gouda, L. K. Kurabetta, M. N. Gunaki, A. A. Hunashyal and R. B. Chougale, *J. Environ. Chem. Eng.*, 2025, **13**, 116029.
- 28 S. Madihalli, S. P. Masti, M. P. Eelager, R. B. Chougale, N. P. Dalbanjan and S. K. Praveen Kumar, *Int. J. Biol. Macromol.*, 2025, **302**, 140926.
- 29 R. Mirji, B. Lobo, D. Dutta, S. P. Masti and M. P. Eelager, *Appl. Radiat. Isot.*, 2023, **196**, 110773.
- 30 S. Madihalli, S. P. Masti, M. P. Eelager, M. N. Gunaki, R. B. Chougale, N. P. Dalbanjan and S. K. Praveen Kumar, *Sustainable Food Technol.*, 2025, 1035–1052.
- 31 M. P. Eelager, S. P. Masti, N. P. Dalbanjan, S. Madihalli, M. N. Gunaki, L. K. Kurabetta, S. K. Praveen Kumar and R. B. Chougale, *Prog. Org. Coat.*, 2024, **192**, 108510.
- 32 M. P. Eelager, S. P. Masti, R. B. Chougale, N. P. Dalbanjan and S. K. Praveen Kumar, *Int. J. Biol. Macromol.*, 2024, **269**, 132270.
- 33 L. K. Kurabetta, S. P. Masti, M. P. Eelager, M. N. Gunaki, S. Madihalli, A. A. Hunashyal, R. B. Chougale, S. K. Praveen Kumar and A. J. Kadapure, *Int. J. Biol. Macromol.*, 2023, **253**, 127552.
- 34 L. K. Kurabetta, S. P. Masti, M. N. Gunaki, A. A. Hunashyal, M. P. Eelager, R. B. Chougale, N. P. Dalbanjan and S. K. Praveen Kumar, *Int. J. Biol. Macromol.*, 2024, **277**, 134191.
- 35 M. N. Gunaki, S. P. Masti, O. J. D'Souza, M. P. Eelager, L. K. Kurabetta, R. B. Chougale, A. J. Kadapure and S. K. Praveen Kumar, *Food Hydrocolloids*, 2024, **152**, 109937.
- 36 M. S. Blois, *Nature*, 1958, **181**, 1199–1200.
- 37 N. P. Dalbanjan, L. Bheemayya, A. J. Kadapure, M. P. Eelager, T. M. C. Swamy, R. R. Kamble and S. K. Praveen Kumar, *ASPET Discovery*, 2025, **1**, 100004.
- 38 R. Re, N. Pellegrini, A. Proteggente, A. Pannala, M. Yang and C. Rice-Evans, *Free Radicals Biol. Med.*, 1999, **26**, 1231–1237.
- 39 M. P. Eelager, S. P. Masti, R. B. Chougale, V. D. Hiremani, S. S. Narasgoudar, N. P. Dalbanjan and P. K. SK, *Int. J. Biol. Macromol.*, 2023, **232**, 123499.
- 40 I. Borovuk and N. Zazharska, *J. Adv. Vet. Anim. Res.*, 2022, **9**, 155.
- 41 L. Zhou and C. E. Boyd, *Aquaculture*, 2016, **450**, 187–193.
- 42 S. Guštin and R. Marinšek-Logar, *Process Saf. Environ. Prot.*, 2011, **89**, 61–66.
- 43 B. Jiang, R. Tsao, Y. Li and M. Miao, Food Safety: Food Analysis Technologies/Techniques, in *Encyclopedia of Agriculture and Food Systems*, Elsevier, 2014, pp. 273–288.
- 44 O. Lowry, N. Rosebrough, A. L. Farr and R. Randall, *J. Biol. Chem.*, 1951, **193**, 265–275.
- 45 P. Cunniff, *Official Methods of Analysis of AOAC International*, AOAC International, Arlington, 1995, vol. 1.
- 46 V. Feddern, L. Kupski, E. P. Cipolatti, G. Giacobbo, G. L. Mendes, E. Badiale-Furlong and L. A. de Souza-Soares, *Eur. J. Lipid Sci. Technol.*, 2010, **112**, 1277–1284.
- 47 N. P. Dalbanjan, A. J. Kadapure, P. Huded, V. B. Chachadi, S. Nayaka and S. K. Kumar, *Ukr. Biochem. J.*, 2022, **94**, 61–66.
- 48 S. S. Narasagoudr, V. G. Hegde, V. N. Vanjeri, R. B. Chougale and S. P. Masti, *Carbohydr. Polym.*, 2020, **236**, 116049.
- 49 T. Gasti, S. Dixit, V. D. Hiremani, R. B. Chougale, S. P. Masti, S. K. Vootla and B. S. Mudigoudra, *Carbohydr. Polym.*, 2022, **277**, 118866.
- 50 M. F. Queiroz, K. R. Teodosio Melo, D. A. Sabry, G. L. Sasaki and H. A. O. Rocha, *Mar. Drugs*, 2014, **13**, 141–158.
- 51 S. Roy and J.-W. Rhim, *Polymers*, 2020, **12**, 2665.
- 52 J. Ghaderi, S. F. Hosseini, N. Keyvani and M. C. Gómez-Guillén, *Food Hydrocolloids*, 2019, **95**, 122–132.
- 53 L. H. Gaabour, *AIP Adv.*, 2021, **11**(10), 105120.
- 54 C. M. Yoshida, V. B. V. Maciel, M. E. D. Mendonça and T. T. Franco, *LWT – Food Sci. Technol.*, 2014, **55**, 83–89.
- 55 H. Saari, C. Fuentes, M. Sjöo, M. Rayner and M. Wahlgren, *Carbohydr. Polym.*, 2017, **157**, 558–566.
- 56 S. Mafirad, M. R. Mehrnia, P. Zahedi and S. N. Hosseini, *Polym. Compos.*, 2018, **39**, 4452–4466.
- 57 Z.-H. Zhang, Z. Han, X.-A. Zeng, X.-Y. Xiong and Y.-J. Liu, *Int. J. Biol. Macromol.*, 2015, **81**, 638–643.
- 58 L.-T. Wu, I.-L. Tsai, Y.-C. Ho, Y.-H. Hang, C. Lin, M.-L. Tsai and F.-L. Mi, *Carbohydr. Polym.*, 2021, **254**, 117410.
- 59 J. Wu, F. Zhong, Y. Li, C. Shoemaker and W. Xia, *Food Hydrocolloids*, 2013, **30**, 82–91.
- 60 L. Vermeiren, F. Devlieghere, M. van Beest, N. de Kruijf and J. Debevere, *Trends Food Sci. Technol.*, 1999, **10**, 77–86.
- 61 V. D. Hiremani, S. Khanapure, T. Gasti, N. Goudar, S. K. Vootla, S. P. Masti, R. B. Malabadi, B. S. Mudigoudra and R. B. Chougale, *Int. J. Biol. Macromol.*, 2021, **193**, 2192–2201.
- 62 Y. Sun, W. Yang, H. Shi, S. K. Tanveer and J. Hai, *Front. Bioeng. Biotechnol.*, 2022, **10**, 1006388.
- 63 WHO, *Food Safety*, <https://www.who.int/news-room/fact-sheets/detail/food-safety>.
- 64 P. C. B. Turnbull, *Medical Microbiology*, 4th edn, 1996.
- 65 L. H. ALManseeqanaa and R. H. Ogaili, *Acad. Int. J. Med. Sci.*, 2023, **1**(2), 7–10.
- 66 H. Kumar, K. Bhardwaj, N. Cruz-Martins, E. Nepovimova, P. Oleksak, D. S. Dhanjal, S. Bhardwaj, R. Singh, C. Chopra and R. Verma, *Molecules*, 2021, **26**, 3447.
- 67 F. Nazzaro, F. Fratianni, L. De Martino, R. Coppola and V. De Feo, *Pharmaceutics*, 2013, **6**, 1451–1474.
- 68 A. B. Falowo, P. O. Fayemi and V. Muchenje, *Food Res. Int.*, 2014, **64**, 171–181.



- 69 V. Lobo, A. Patil, A. Phatak and N. Chandra, *Pharmacogn. Rev.*, 2010, **4**, 118.
- 70 C. Kh, *Br. Med. Bull.*, 1993, **49**, 481–493.
- 71 M. A. Shah, S. J. D. Bosco and S. A. Mir, *Meat Sci.*, 2014, **98**, 21–33.
- 72 A. M. Pisoschi, A. Pop, F. Iordache, L. Stanca, G. Predoi and A. I. Serban, *Eur. J. Med. Chem.*, 2021, **209**, 112891.
- 73 A. Sadi and H. Ferfera-Harrar, *Int. J. Biol. Macromol.*, 2023, **242**, 124964.
- 74 G. da Silva Filipini, V. P. Romani and V. G. Martins, *Food Hydrocolloids*, 2020, **109**, 106139.
- 75 K. Wessels, D. Rip and P. Gouws, *Foods*, 2021, **10**, 1742.
- 76 P. Magdelaine, M. Spiess and E. Valceschini, *World's Poult. Sci. J.*, 2008, **64**, 53–64.
- 77 N. Alexandratos and J. Bruinsma, *World Agriculture towards 2030/2050: The 2012 Revision*, 2012, DOI: [10.22004/ag.econ.288998](https://doi.org/10.22004/ag.econ.288998).
- 78 R. Priyadarshi, S.-M. Kim and J.-W. Rhim, *Sustainable Mater. Technol.*, 2021, **29**, e00325.
- 79 S. R. Kanatt, *Food Packag. Shelf Life*, 2020, **24**, 100506.
- 80 A. Wisal, A. Ullah, W. Anwar, C. M. Morel and S. S. Hassan, *Genomics Inf.*, 2023, **21**(3), e34.
- 81 C. E. Manyi-Loh and R. Lues, *Microorganisms*, 2023, **11**, 2484.
- 82 W. Yuan and H.-G. Yuk, *Food Microbiol.*, 2018, **72**, 176–184.
- 83 Ç. Soysal, H. Bozkurt, E. Dirican, M. Güçlü, E. D. Bozhüyük, A. E. Uslu and S. Kaya, *Food Control*, 2015, **54**, 294–299.
- 84 J. H. Han, *Antimicrobial Food Packaging*, 2003.
- 85 E. Balbinot-Alfaro, D. V. Craveiro, K. O. Lima, H. L. G. Costa, D. R. Lopes and C. Prentice, *Food Eng. Rev.*, 2019, **11**, 235–244.
- 86 R. K. Robinson, *Encyclopedia of Food Microbiology*, Academic Press, 2014.
- 87 A. E.-D. A. Bekhit, B. W. Holman, S. G. Giteru and D. L. Hopkins, *Trends Food Sci. Technol.*, 2021, **109**, 280–302.
- 88 K. Dong, X. Luo, L. Liu, F. An, D. Tang, L. Fu, H. Teng and Q. Huang, *Int. J. Food Sci. Technol.*, 2021, **56**, 1597–1607.
- 89 X. Lu, Y. Zhang, L. Zhu, X. Luo and D. L. Hopkins, *Meat Sci.*, 2019, **149**, 79–84.
- 90 M. Senapati and P. P. Sahu, *Food Chem.*, 2020, **324**, 126893.
- 91 F. Li and L. L. Guan, *Appl. Environ. Microbiol.*, 2017, **83**, e00061.
- 92 A. E.-D. A. Bekhit, B. W. B. Holman, S. G. Giteru and D. L. Hopkins, *Trends Food Sci. Technol.*, 2021, **109**, 280–302.
- 93 L. C. Umuhumuza and X. Sun, *Eur. Food Res. Technol.*, 2011, **232**, 425–431.
- 94 Q. Ma, L. Du and L. Wang, *Sens. Actuators, B*, 2017, **244**, 759–766.
- 95 A. J. Pellissery, P. G. Vinayamohan, M. A. R. Amalaradjou and K. Venkitanarayanan, in *Meat Quality Analysis*, Elsevier, 2020, pp. 307–334.
- 96 C. Papuc, G. V. Goran, C. N. Predescu and V. Nicorescu, *Compr. Rev. Food Sci. Food Saf.*, 2017, **16**, 96–123.
- 97 Y. Yu, G. Wang, X. Yin, C. Ge and G. Liao, *Food Res. Int.*, 2021, **149**, 110696.
- 98 F. S. Fedorov, A. Yaqin, D. V. Krasnikov, V. A. Kondrashov, G. Ovchinnikov, Y. Kostyukevich, S. Osipenko and A. G. Nasibulin, *Food Chem.*, 2021, **345**, 128747.
- 99 J. Csapó, J. Prokisch, C. Albert and P. Sipos, *Acta Univ. Sapientiae, Aliment.*, 2019, **12**, 21–41.
- 100 S. E. Duncan and H.-H. Chang, *Adv. Food Nutr. Res.*, 2012, **67**, 25–73.
- 101 M. Parcheta, R. Świsłocka, S. Orzechowska, M. Akimowicz, R. Chojińska and W. Lewandowski, *Materials*, 2021, **14**, 1984.
- 102 A. Khan, R. Priyadarshi, T. Bhattacharya and J.-W. Rhim, *Food Bioprocess Technol.*, 2023, 1–15, DOI: [10.1007/s11947-023-03048-7](https://doi.org/10.1007/s11947-023-03048-7).
- 103 M. N. Lund, R. Lametsch, M. S. Hviid, O. N. Jensen and L. H. Skibsted, *Meat Sci.*, 2007, **77**, 295–303.
- 104 S. Jongberg, A. M. Racanicci and L. H. Skibsted, *Food Chem.*, 2019, **300**, 125134.
- 105 Y. Li, S. Jongberg, M. L. Andersen, M. J. Davies and M. N. Lund, *Free Radicals Biol. Med.*, 2016, **97**, 148–157.
- 106 S. Jongberg, L. d. S. Terkelsen, R. Miklos and M. N. Lund, *Meat Sci.*, 2015, **100**, 2–9.

