



Cite this: *Chem. Soc. Rev.*, 2026, 55, 336

Electric fields at hydrophobic water interfaces: spectroscopic evidence, physical origin, and implications on reactivity

Kwing Yeung Chan, Chenjie Zhuang, Vinh Gia Vuong, Naixin Qian, Xin Gao  and Wei Min *

Water at interfaces exhibits unique properties that differ markedly from those of bulk water. In particular, a myriad of water-interface-related enhanced reactivities including on-water catalysis and microdroplet chemistry have been documented since the 1980s but remain mechanistically unclear. This review focuses on recent advances in optical spectroscopy and imaging techniques—including fluorescence imaging, vibrational Stark spectroscopy, electrochromism, sum-frequency generation, and high-resolution Raman micro-spectroscopy—that have successfully enabled the detection of interfacial electric fields at different hydrophobic water interfaces (air, liquid and solid). We summarize how both probe-based and label-free optical spectroscopic techniques can consistently quantify the on-water electric field strengths to be on the order of tens of MV cm^{-1} , corroborated by independent non-spectroscopic techniques, such as electrokinetic and surface charge measurements. The surprisingly close agreement among these different measurements and across broad experimental systems strongly hints at the existence of strong electric fields being a general feature of water–hydrophobe interfaces. We further discuss the physical origins of the interfacial electric field with a particular emphasis on the mechanism of preferential hydroxide accumulation at hydrophobic interfaces. Finally, we examine the implications of strong interfacial electric fields for chemical kinetics, radical generation and thermodynamics, thereby making important connections to interfacial water reactivity. These insights not only contribute to our fundamental understanding of water at interfaces but also point toward new strategies for harnessing interfacial water electrostatics in biomedicine, catalysis, green chemistry, and environmental science.

Received 4th July 2025

DOI: 10.1039/d5cs00778j

rsc.li/chem-soc-rev

Department of Chemistry, Columbia University, New York, NY 10027, USA. E-mail: wm2256@columbia.edu



Kwing Yeung Chan

(Hong Kong). His research focuses on interfacial chemistry and using spectroscopy and microscopy to study physical and chemical processes.

Kwing Yeung Chan received his BBiomedSc degree (major in biomedical sciences and minor in chemistry) from Li Ka Shing Faculty of Medicine, the University of Hong Kong, in 2021. He is currently a PhD candidate in chemistry at Columbia University under the supervision of Prof. Wei Min. His PhD studies are supported by the Croucher Scholarship (Hong Kong) and the Joseph Needham Merit Scholarship



Chenjie Zhuang

Chenjie Zhuang received her BS degree (major in theoretical and applied mechanics and minor in biomedical engineering) from Peking University in 2024. She is currently a PhD student in chemistry at Columbia University under the supervision of Prof. Wei Min. Her research focuses on interfacial phenomena and applying spectroscopy and microscopy in various biomedical systems.



Interfacial water reactivity: from on-water catalysis to microdroplet chemistry and beyond

Water is likely the most mysterious, yet universal, substance on Earth. Although its physical and chemical properties have been well studied as a bulk solvent, much evidence suggests that water at interfaces (interfacial water) has intriguing behaviors and properties that are vastly distinct from those of bulk water. In the realm of chemistry, water playing a role in organic reactions was perhaps first recognized by Breslow more than 40 years ago.^{1,2} It was shown in this pioneering study that Diels–Alder reactions could proceed at a much higher rate in water than in common organic solvents. Breslow intuitively attributed this observation to “hydrophobic effects” with the remarkable conclusion that the reaction acceleration occurs

because of, rather than in spite of, the poor solubility of the organic substances in water (Fig. 1a).³

In the 2000s, Sharpless and co-workers revisited this phenomenon and systematically studied the unique reactivity of organic compounds in aqueous suspension, coining the term “on-water” catalysis.⁴ Their approach involved vigorous stirring of organic reactants in water to create suspensions with extensive interfaces between water and organic molecules (Fig. 1b). Importantly, this “on-water” catalysis was demonstrated to be a general mechanism beyond Diels–Alder reactions. They suggested that the phase heterogeneity—the coexistence of organic (oil) and aqueous (water) phases—has a crucial role in the drastic acceleration of reaction kinetics. Though without direct evidence at the time, they hypothesized that the mesoscopic phase boundary itself might be responsible for the observed effect. Jung and Marcus later proposed a molecular theory for this, in which “dangling” OH groups arising from the water’s



Vinh Gia Vuong

Vinh Gia Vuong obtained his BS degree in chemistry with honors from New York University in 2024. He is currently pursuing his PhD at Columbia University under the supervision of Professor Wei Min. His research focuses on understanding chemical reactivity at hydrophobic interfaces and developing microfluidic platforms for biomolecular analysis.



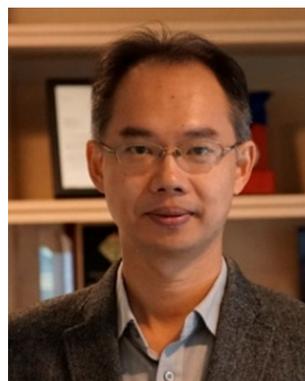
Naixin Qian

Naixin Qian received her BS degree in chemistry from Li Yun Honors School, Beijing Normal University, China, in 2019 and PhD with the Pegram Award from Columbia University in 2025. Her research focuses on developing novel vibrational spectroscopy and microscopy methods for biomedical and environmental applications.



Xin Gao

Xin Gao received his BS degree in chemistry from Kuang Yaming Honors School, Nanjing University, China, in 2019 and PhD with the Pegram Award from Columbia University in 2025. His research focuses on applying chemical spectroscopy and microscopy in various biomedical systems.



Wei Min

Wei Min received his BS from Peking University in 2003 and PhD from Harvard University in 2008 studying single-molecule biophysics with Prof. Sunney Xie. After continuing his post-doctoral work in the Xie group, Dr Min joined the faculty at Columbia University in 2010 and was promoted to a Full Professor there in 2017. Dr Min’s contribution has been recognized by a number of honors, including the Biophotonics Technology Innovator Award from SPIE (2023), the Craver Award of Vibrational Spectroscopy (2022), the Scientific Achievement Award from Royal Microscopical Society (2021), the Pittsburgh Conference Achievement Award (2019) and the ACS Early Career Award in Experimental Physical Chemistry (2017).



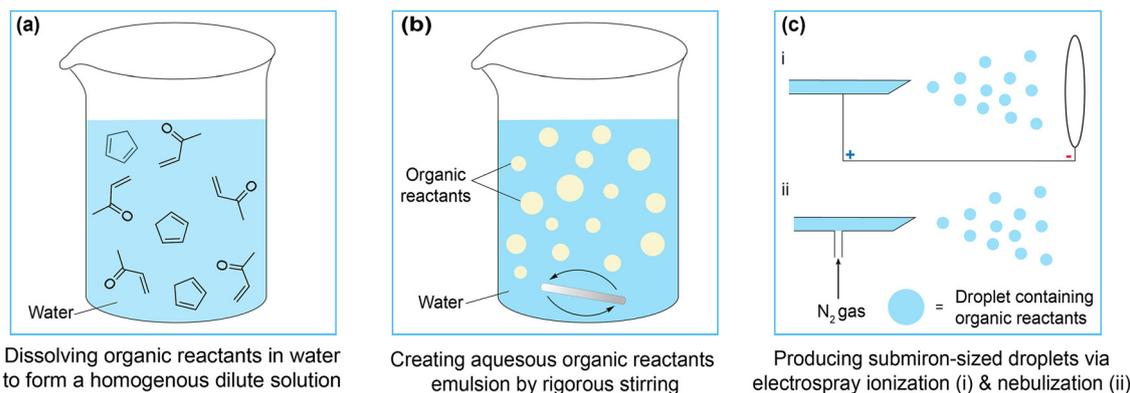


Fig. 1 Experimental setups of on-water catalysis and microdroplet chemistry. (a) Dissolving organic reactants (such as the diene and dienophile in a Diels–Alder reaction) in water at low concentrations (μM to mM range) to form a homogeneous dilute solution demonstrated by Breslow in the 1980s. (b) Mixing organic reactants at much higher concentration (several M) in water by rigorous stirring to form an aqueous organic emulsion demonstrated by Sharpless in the 2000s. (c) In microdroplet chemistry, there are two common methods to prepare a more confined aqueous reaction environment (i) preparing microdroplets using electrospray ionization and (ii) creating aerosol microdroplets using high-pressure nitrogen gas (nebulization). Reagents inside these droplets are allowed to react before entering a mass spectrometer for analyzing the outcome. The distance between the emitter and the mass spectrometer inlet is known as the reaction distance for microdroplets.

broken hydrogen bonding network at the interface could stabilize the transition state of nearby organic reactions.⁵ Since then, on-water catalysis has continued to attract attention in the field of organic chemistry.^{6,7}

While on-water catalysis highlights the role of water–oil interfaces in accelerating reactions, the emergence of microdroplet chemistry is a parallel and perhaps more remarkable development.^{8,9} In microdroplet chemistry, these micron-sized droplets containing reactants are generated through methods such as electrospray ionization and inert gas nebulization, to name but a few (Fig. 1c). These droplets form extensive interfaces with air, and the reaction outcome is analyzed with a mass spectrometer. In the 2010s, several groups, particularly Cooks and Zare, reported substantial (up to a million folds) rate acceleration of reactions in microdroplets compared to their counterparts in bulk water.^{10–20} These reactions occur entirely in the aqueous phase, hence complementing “on-water” catalysis in the water–oil systems. One striking demonstration is that the on-water chemistry reaction $[(2\sigma + 2\sigma + 2\pi)]$ cycloaddition studied earlier by Sharpless and co-workers⁴ was reproduced in microdroplets with another 100-fold higher rate.²¹ Thus, these examples underscore a broader principle: the acceleration of chemical reactions is not exclusive to water–oil mixtures but also applicable to water–air interfaces.

Among a plethora of microdroplet reactions, the spontaneous generation of H_2O_2 from sprayed water microdroplets, initially reported by Zare and co-workers, took the field by surprise, although this discovery is surrounded by ongoing debates.^{22–33} Its production did not require any obvious oxidants/reductants or an applied potential. It seemingly contradicts one’s intuition: the standard potential for oxidizing water to hydroxyl radicals is -2.8 V (against the standard hydrogen electrode),³⁴ which is thermodynamically highly unfavorable. This suggests that water interfaces not only accelerate the reaction kinetics but also reshape thermodynamic landscapes

of reactions. Since then, an expanding list of reactions (mostly redox by nature) has been explored and realized in microdroplets, for instance, $\text{C}(\text{sp}^3)\text{–N}$ bond formation, Menshutkin reactions, C–H/N–H cross-coupling, reduction of transition metal ions, even simultaneous oxidation and reduction of phosphonates, *etc.*^{35–54} These unique chemical reactivities have implications across broad fields including chemical synthesis, green chemistry, atmospheric chemistry and environmental science. For example, nitrogen fixation,⁵⁵ CO_2 capture,^{56–59} greenhouse gas formation,⁶⁰ sulfate formation in aerosol,⁶¹ degradation of PFAS,⁶² high-throughput synthesis,⁶³ photochemical synthesis,^{64,65} and prebiotic chemistry^{66–71} have all recently been demonstrated in microdroplets.

In addition to water–oil interfaces as in “on-water” catalysis and water–air interfaces as in microdroplet chemistry, similar reactivity has also been found at water–solid interfaces. For example, water droplets have been demonstrated to spontaneously degrade organic dye molecules when sliding over a hydrophobic polymer surface.⁷² Redox reactivity like that of microdroplets has also been discovered at glass–water interfaces.⁷³ Even biological systems have been reported to exhibit similar behaviors.^{74,75} Together, decades of experimental evidence accumulated from a wide range of different water interfaces (oil, air and solid) seem to all suggest that interfacial water on hydrophobic surfaces has a chemical reactivity distinct from that of bulk water.

Electrostatic properties of interfacial water

Mechanistic studies over the years have attributed the unique chemistry in microdroplets to several intrinsic properties of water interfaces, such as partial solvation of reactants,⁷⁶ kinetic confinement,¹⁷ stabilization of transition states,⁵ entropic



factor,^{8,67} extreme pH values (super-acid or super-base),^{77–79} *etc.* These effects can be amplified in micron-sized droplets due to their high surface-to-volume ratios. Among various explanations, one particularly intriguing hypothesis is the existence of a strong electric field across the aqueous interface.^{9,80–85}

While the underlying mechanism may not appear obvious (which will be discussed later), several pieces of evidence and considerations imply the unique interfacial electrostatics and the potential existence of a localized electric field.

(1) Firstly, unlike in a bulk solution, the symmetry is broken at the interface at the molecular level. The ordered orientation of water molecules, which have a large dipole moment at the interface, can generate a local electric field. This picture is adopted in some theoretical calculations of the surface potential.^{86–89}

(2) Secondly, although there are debates on the direction of pH deviation, nearly all studies agree that the pH of the water interface is not neutral, with more evidence supporting that it is basic in nature.^{90–95} Since only pure water is present in the system, a non-neutral interface means either anions or cations, presumably hydroxide (OH^-) or hydronium (H_3O^+), are preferentially enriched at water interfaces. This ionic imbalance can also generate an interfacial electric field.

(3) Thirdly, micron-sized objects such as oil droplets, air bubbles and solid particles have been repeatedly and robustly shown to be electrokinetically active in electrophoresis experiments (Fig. 2a).^{91,96–98} Zeta (ζ) potential is typically around -60 to -90 mV at pH 7, supporting excellent long-term colloidal

stability. The electrokinetic response of micron-sized objects to an external electric field unescapably implies that they carry net charges, likely due to the structured distribution of cations and anions, which gives rise to an interfacial electrical double layer (Fig. 2b).

(4) Fourthly, there is an accumulation of negative charges on the hydrophobic polymer surface when in contact with water.⁹³ When water droplets move across a hydrophobic surface, such as polytetrafluoroethylene (PTFE), two phases (water and polymer, respectively) become oppositely charged—a phenomenon called slide electrification. While this had been observed nearly a century ago in a largely curious manner, it has been recently harnessed to generate electrical energy to power light-emitting diodes (LEDs) (Fig. 2c and d).⁹⁹ Further experiments suggested that slide electrification could yield an electric potential exceeding 1 kilovolt.¹⁰⁰ In addition, electrostatic breakdown has also been observed at liquid–solid–gas triple-phase interfaces most recently,¹⁰¹ further suggesting the involvement of strong electric fields.

These pieces of evidence, albeit from different communities, are not isolated. For example, the surface pH measurement can be viewed as the static (only contact) version of slide electrification between water and air; electrokinetic measurement of micron-size objects can be viewed as the miniaturized version of slide electrification. Taken together, non-neutral surface pH, negative ζ potential and water electrification all point to the electrically active nature of aqueous interfaces and may be indicative of a local electric field. Such a convergence of

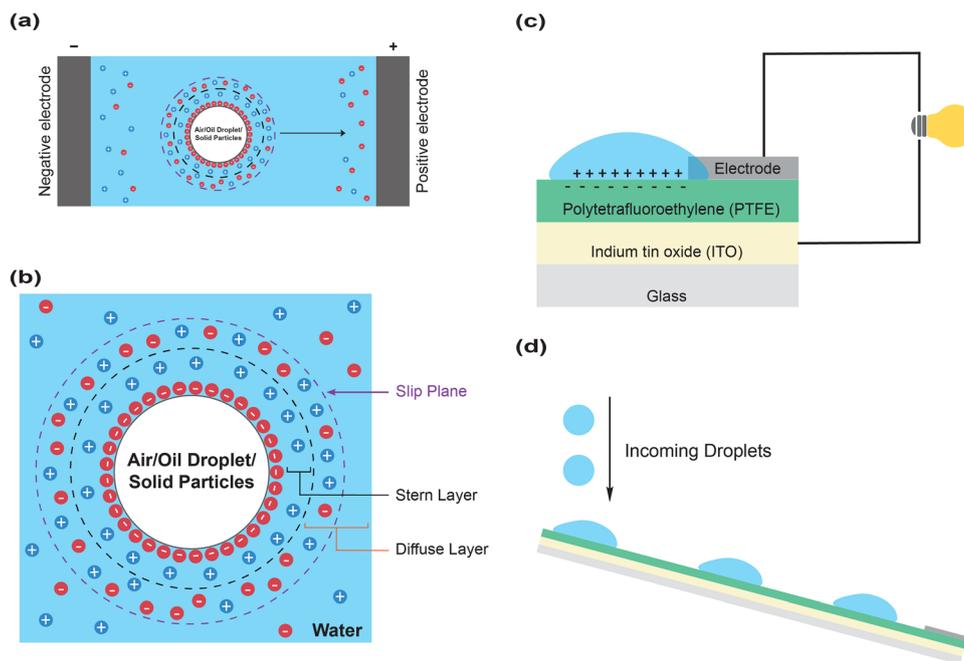


Fig. 2 Electrostatic properties of interfacial water. (a) A typical electrokinetic measurement setup showing how a negatively charged object (air bubbles, oil droplets or solid particles dispersed in water) moves towards the positive electrode under an external electric field. (b) The structure of the electrical double layer surrounding bubbles/droplets/particles in water. Note that the length scale does not represent the actual thickness of the ion layers. (c) A schematic diagram showing the layout of the droplet-based electricity generator, which harvests energy through sliding electrification as water droplets move across the PTFE surface. The electrical energy of four droplets ($100\ \mu\text{L}$ each) is enough to power 400 LEDs. (d) The setup is tilted at an angle to ensure water can slide down from the PTFE surface.



findings raises two fundamental questions for physical chemistry: is there direct experimental evidence for the existence of such an electric field? And if so, how can their strength be quantitatively determined?

Evidence of on-water electric fields by optical imaging

Investigating the interfacial electric field poses both a valuable opportunity and a formidable challenge for experimentalists. Due to water's high dielectric constant and the associated

strong screening effect, this interfacial electric field is expected to decay very quickly into the bulk phase, making its detection inherently difficult. Moreover, one cannot prepare only interfacial water without its bulk counterpart in an experiment. Therefore, either strict interfacial selectivity or rigorous separation is required to isolate the interfacial component from the bulk counterpart, which creates another hurdle.

However, if an interfacial electric field does exist, it will exert a directional force on polar probe molecules, attracting them to the interface and aligning their dipoles (Fig. 3a). This physical consequence provides an indirect route to test the presence of the interfacial electric field. Optical imaging offers a powerful

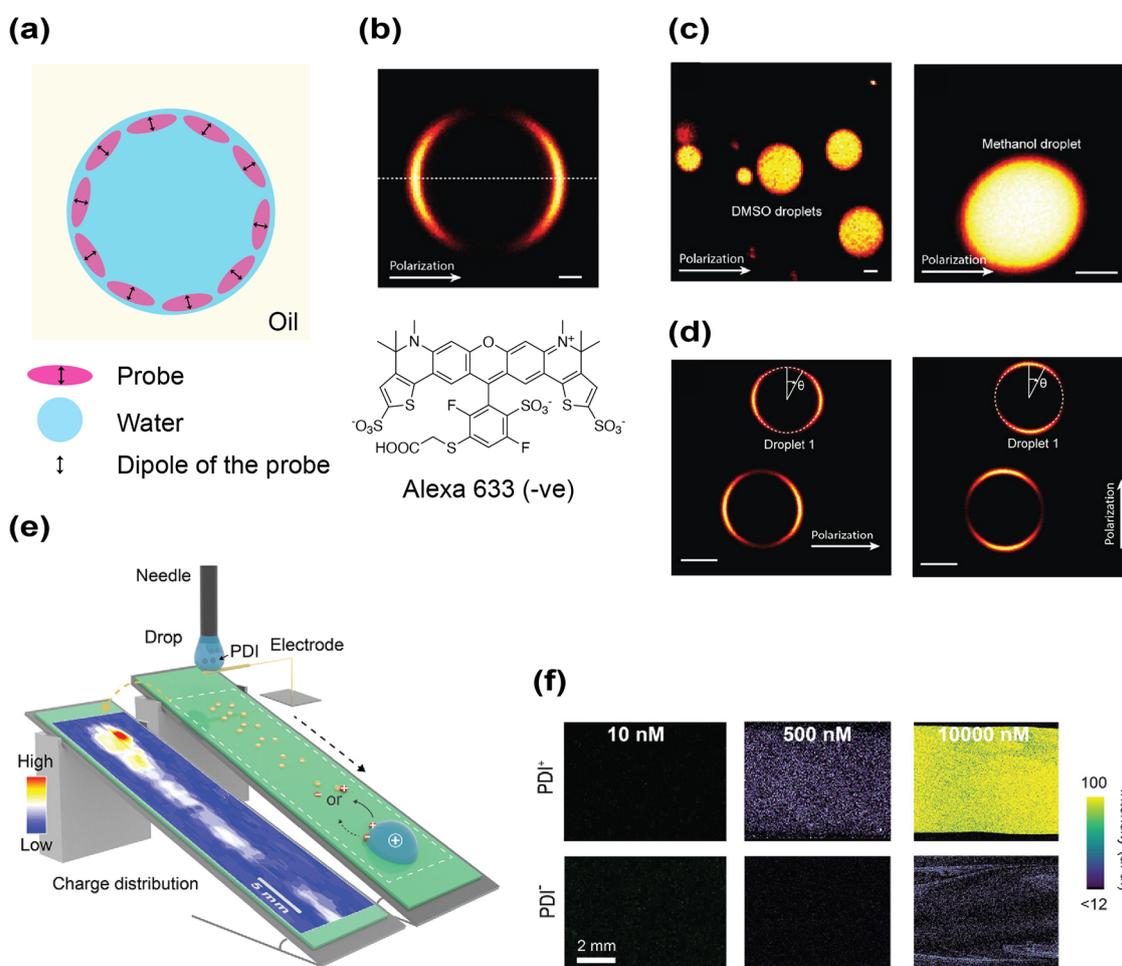


Fig. 3 Probing the interfacial electric field using optical imaging of fluorophores. (a) A Schematic diagram showing how a probe can enrich and orient at the water–oil interface (b) Two-photon fluorescence imaging showing the spatial distribution of Alexa 633 (negatively charged) for aqueous droplets emulsified in hexadecane. A similar ring pattern was observed for rhodamine 101 (zwitterionic) and rhodamine 6G (positively charged). Scale bar: 1 μm . Reproduced with permission from ref. 81. Copyright 2020, the American Chemical Society. (c) The probe showed no interfacial enrichment or orientational preference in microdroplets prepared from DMSO/MeOH in hexadecane. Scale bar: 2 μm . Reproduced with permission from ref. 81. Copyright 2020, the American Chemical Society. (d) Strong anisotropy of rhodamine 800 can be observed by rotating the polarization direction of the excitation laser. The fluorescence signal is most intense along the excitation polarization vector where the coupling between the transition dipole and the incident laser's electric field is the strongest. Scale bar: 5 μm . Reproduced with permission from ref. 82. Copyright 2020, the American Chemical Society. (e) An experimental setup for demonstrating slide electrification between water and a hydrophobic material, PFOTS. Water droplets containing either a positively charged fluorophore (PDI⁺) or a negatively charged fluorophore (PDI⁻) slide down PFOTS and their fluorescence distribution pattern is recorded using a confocal microscope. Reproduced with permission from ref. 104. Copyright 2025, Wiley-VCH. (f) PDI⁺ is deposited on the surface when its concentration reaches 0.5 μM , but PDI⁻ only begins to deposit on the surface when its concentration reaches 10 μM . Reproduced with permission from ref. 104. Copyright 2025, Wiley-VCH.



means to examine this hypothesis by directly visualizing the spatial and orientational distribution of fluorescent probes in microdroplets. Studies aiming to examine the electrostatics of the interface were performed on water–oil emulsions using either single-photon confocal fluorescence or two-photon excited fluorescence microscopy.^{80–82,102} They investigated the spatial (translational degree of freedom) and orientational (rotational degree of freedom) distributions of the probe at interfaces to evaluate whether such interfacial electric fields exist. These studies showed two consistent observations:

Firstly, probe localization at the interface with a clear ring pattern was observed in water microdroplets suspended in hexadecane oil (Fig. 3b).^{80–82,102} Although Fig. 3b only shows the localization image of a negatively charged dye (Alexa 633), the zwitterionic and positively charged probe exhibited the same ring pattern. As mentioned before, preferential adsorption of certain ions to interfaces leads to an ionic imbalance, which can produce an electrical double layer and attract probes to the interface. Interestingly, when water was replaced by other organic solvents immiscible with the oil such as methanol (MeOH) and dimethyl sulfoxide (DMSO), this characteristic ring pattern disappeared, and the probes distributed uniformly (Fig. 3c). Remarkably, the hexadecane oil does not form stable emulsions in MeOH, DMSO or acetonitrile (MeCN) environments, which implies that their resulting ζ potentials were negligible (*i.e.*, high tendency towards aggregation). This would be accompanied by a much weaker interfacial electric field, which agrees with the disappearance of the ring pattern (Fig. 3c). Therefore, these two observations convey a consistent conclusion—water–oil interfaces possess uniquely strong electric fields.

Secondly, the detected fluorescence signal was not evenly distributed around the ring (Fig. 3d).^{80–82,102} The angular dependence of fluorescence intensity is known as excitation anisotropy. Fluorescence intensity is the highest when the transition dipole of the probe aligns with the polarization vector of the excitation beam and it is the weakest when perpendicular to it.¹⁰³ In their studies, by rotating the laser polarization direction, the fluorescence signal distribution rotated accordingly, but the distribution pattern remained nearly the same. This phenomenon can be readily rationalized by the presence of an interfacial electric field, which is shown to be perpendicular to the water–oil interface in simulations,⁸⁴ recruiting and strictly aligning the dipole of fluorescent probes on the interface through electrostatic interactions. Therefore, this intensity modulation is a strong manifestation for the rigid alignment of the probe's dipoles perpendicular to the water–oil interface. Again, this anisotropic effect was absent in droplets formed between oil and MeOH, MeCN or DMSO.

Similar surface enrichment of fluorescent probes has also been imaged on the water–solid planar interface.¹⁰⁴ In this study, water droplets containing either positively or negatively charged fluorescent dyes were allowed to slide down an inclined hydrophobic surface coated with perfluorodecyltrichlorosilane (PFOTS). Fluorophore deposition on the surface was monitored using a confocal fluorescence microscope

(Fig. 3e). Both dyes, PDI⁺ and PDI[−], are perylene derivatives and share very similar chemical structures. At a concentration of 0.5 μM , PDI⁺ could readily deposit on the surface, whereas the deposition of PDI[−] required at least a 20-fold higher concentration (10 μM) (Fig. 3f). Such a disparity between two dyes with nearly identical molecular structures would hardly be explained by common hydrophobic interactions towards the interface. The positively charged dye having more pronounced surface partition than its negative counterpart is consistent with previous literature that hydrophobic water–solid interfaces are negatively charged,⁹³ thereby recruiting the dye of opposite charge through the electric field.

Quantifying on-water electric fields with probe-based optical spectroscopy

Optical imaging experiments on water–oil/solid interfaces provide strong evidence to support the presence of the on-water electric field. However, quantitative measurement of the field strength is more challenging, as the exact relation between the electric field and the probe distribution (either translational or orientational) at interfaces could be non-trivial and depend on multiple parameters. One approach that has proven fruitful is to examine the spectroscopic shift of vibrational or fluorescence emission frequency of molecules on the interface. It is well established in physical chemistry that the energy eigenvalues are sensitive to the local electrostatic environment, and the amount of energy shift can be used to calculate the strength of the local electric field after proper calibration. Both probe-based and label-free spectroscopic experiments have been implemented to this end, and we will first introduce the former.

The first reported field strength was measured by Xiong *et al.* when studying the vibrational frequency shift of a nitrile (CN) group in a probe molecule on the water–oil interface.⁸² The influence of an external electric field on a bond's vibrational frequency is known as the vibrational Stark effect (VSE) (Fig. 4a). VSE typically occurs in chemical bonds with anharmonic molecular potential. The first vibrational excited state usually has a larger dipole moment than the ground state. Hence, when the dipole aligns with the external electric field, the first excited state will be energetically more stabilized than the ground state does, leading to a slightly lower energy (red-shift) of the vibrational transition between the ground state to the excited state. Mathematically, linear VSE is described by the following equation:

$$\Delta\bar{\nu} = -\Delta\bar{\mu}\cdot\vec{F} \quad (1)$$

$\Delta\bar{\nu}$, $\Delta\bar{\mu}$, and \vec{F} are the shift in vibrational frequency, the Stark tuning rate of the vibrational mode, and the strength of the external electric field, respectively.¹⁰⁵ To probe the electric field strength at the water–oil interface, a fluorescent probe, rhodamine 800 (Rh800), was employed. The bond axis of its CN group is almost parallel to the dipole moment of the molecule,



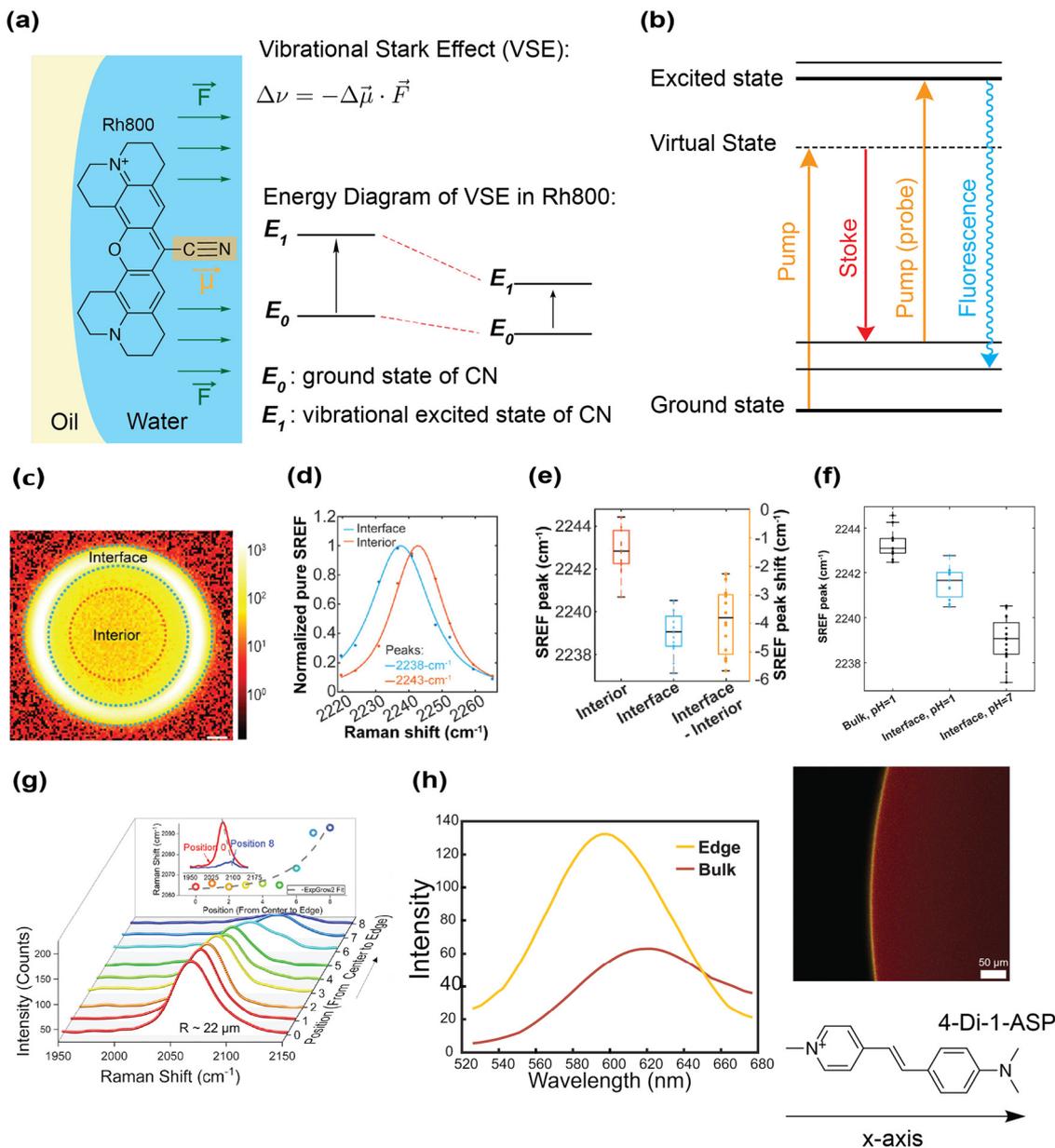


Fig. 4 Quantifying the interfacial electric field's strength by probe-based spectroscopy. (a) A schematic diagram showing how VSE decreases the vibrational frequency of the CN bond of rhodamine 800 at the water–oil interface. The dipole moment vector on rhodamine 800 aligns almost parallel to the CN bond axis, allowing good projection of the electric field onto the vibrational bond axis. $\Delta\nu$: shift in vibrational frequency, $\Delta\vec{\mu}$: Stark tuning rate, and \vec{F} : the external electric field that exerts on the CN bond. The vibrational excited state of the CN bond is more stabilized than its ground state, thus causing a redshift of the CN vibrational frequency. (b) Energy diagram of stimulated Raman excited fluorescence (SREF) spectroscopy. (c) First frame of SREF of the Rh 800 CN mode. The signal from a droplet is divided into three regions: interface, interface-interior and interior. Scale bar: 2 μm . Reproduced with permission from ref. 82. Copyright 2020, the American Chemical Society. (d) Background-subtracted SREF spectrum. A redshift of $4 \pm 1 \text{ cm}^{-1}$ is recorded for the CN vibrational frequency on the interface. Reproduced with permission from ref. 82. Copyright 2020, the American Chemical Society. (e) 16 independent measurements of the CN vibrational frequency on different droplets. Reproduced with permission from ref. 82. Copyright 2020, the American Chemical Society. (f) SREF measurement of the CN group on Rh800 under different pH. At pH = 1, the Stark effect on the interface is less prominent. The CN vibrational frequency for Rh800 in the droplet interior remains unaffected at pH = 1. Reproduced with permission from ref. 82. Copyright 2020, the American Chemical Society. (g) Probing the electric field at water–air interfaces using SERS spectroscopy. The VSE is studied by comparing the vibrational frequency of CN on gold nanoparticles (dissolved in the droplet) functionalized with SCN between the droplet interface and center. A shift in the CN vibrational frequency is observed on the interface compared with the center. Reproduced with permission from ref. 108. Copyright 2024, the Cell Press. (h) Probing the electric field at the water–air interface using a voltage-sensitive fluorophore (4-Di-1-ASP). The interfacial electric field can affect its fluorescence emission maximum (electrochromism). The emission maximum on the interface is shifted by 18 nm, suggesting that the electrostatic property on the interface and the bulk phase differs. Reproduced with permission from ref. 109. Copyright 2025, ChemRxiv.



allowing a good projection of the local electric field onto the vibrational CN bond axis.⁸²

To study the shift in the vibrational frequency of the CN bond, the vibrational Stark measurement was implemented through stimulated Raman excited fluorescence (SREF) microscopy (Fig. 4b). In SREF, the vibrational spectrum of a chemical bond is mapped into the fluorescence excitation spectrum, allowing superb detection sensitivity down to single molecule level.¹⁰⁶ Thus, the SREF signal can serve as a basis for direct comparison of the CN group's vibrational frequency in different environments (*i.e.*, interface *vs.* bulk) (Fig. 4c). By scanning the SREF images across the line shape of the CN vibrational mode, SREF spectra at different regions (interface and interior) were obtained. Direct comparisons of the SREF spectra from 16 independent measurements between the interface and the interior on different water droplets showed a clear redshift of the CN vibrational frequency ($\sim 4 \pm 1 \text{ cm}^{-1}$) at the interface (Fig. 4d and e). The same SREF experiment was repeated with a lower Rh800 concentration to circumvent the possible interference from high probe concentration. Although the SREF signal was no longer detectable in the droplet interior with a low probe concentration, a similar redshift of CN vibrational frequency ($\sim 3 \pm 1 \text{ cm}^{-1}$) was observed on the interface. Applying the eqn (1) with $\Delta\bar{\nu}$ of $\sim 0.53 \pm 0.09 \text{ cm}^{-1} \text{ MV}^{-1} \text{ cm}^{-1}$) for the CN group (estimated based on vibrational solvatochromic measurements and Onsager reaction field theory),¹⁰⁷ this redshift corresponds to a local interfacial electric field of approximately 8 MV cm^{-1} . The redshift's magnitude is also found to be pH dependent (Fig. 4f) but is much less sensitive to the concentration of salt ions such as Cl^- , suggesting there is a chemical selectivity of the water-hydrophobe interface in attracting anionic species. The observed pH dependent redshift is also consistent with the pH-dependent ζ -potential of oil droplets.^{91,94}

Another VSE-based measurement was later performed at the water–air interface of aerosol particles.¹⁰⁸ In this study, gold nanoparticles self-assembled with the vibrational thiocyanate (SCN) probes were utilized. The vibrational frequency of the CN mode inside aerosol particles was measured using surface-enhanced Raman scattering (SERS) spectroscopy. By spatially resolving the Raman signal from the droplet center to the surface, a Stark shift of 28.89 cm^{-1} of the CN mode was detected (droplet radius $\sim 22 \mu\text{m}$) (Fig. 4g). A negative correlation between the Stark shift and droplet size was also found: Stark shift in droplets with a radius of $39 \mu\text{m}$ was 3.31 cm^{-1} . Based on their calculation, the electric field on the surface of droplets was around 10 to 100 MV cm^{-1} , which appears to be in line with the 8 MV cm^{-1} field strength obtained above by SREF microscopy on the water–oil interface, although the use of SERS nanoparticles, which are much larger than fluorophores, might suffer from a higher chance of perturbing the local air–water interface.

In addition to VSE, the shift in the fluorescence emission frequency, a phenomenon known as electrochromism, is another method to probe the strength of the on-water electric field. In their experiment studying the water–air interface in a

water droplet (diameter $\sim 4 \text{ mm}$), a voltage-sensitive dye, 4-Di-1-ASP, was applied and imaged using spectral confocal fluorescence microscopy.¹⁰⁹ On the water–air interface, the emission maximum of the dye was shifted by 18 nm (Fig. 4h). Their computational results further showed that only an electric field applied along the x -axis (from pyridinium to the amine group) of the molecule could lead to such a shift. Therefore, this shift suggested that the dye is oriented perpendicularly to the water–air interface. Moreover, since the transition dipole of 4-Di-1-ASP is also parallel to the x -axis of the molecule,¹¹⁰ its perpendicular alignment at the interface allows efficient perturbation of its transition dipole.¹¹¹ Hence, the Stark effect experienced by the dye can probe the strength of the local electric field at the interface. By comparing the emission maximum on the interface and that in the bulk phase, the strength of the interfacial electric field can be estimated. Depending on the exact solvent polarity model employed, this shift corresponds to an electric field of 7.2 to 13 MV cm^{-1} , which again is comparable to the 8 MV cm^{-1} field strength obtained above by SREF microscopy.⁸² When water was replaced by either anhydrous DMSO or dimethylformamide (DMF) under a nitrogen atmosphere, no shift in the emission maximum was observed, indicating vanishing electrostatics. This is consistent with solvent-dependent probe localization discussed above.^{80–82} This result confirmed that water is essential for generating the electrochromic shift of dye on the water–air interface.

Probing on-water electric fields with label-free optical spectroscopy

While the probe-based spectroscopic techniques, including SREF, SERS and electrochromism, successfully measured the interfacial electric field strength with largely comparable magnitudes of about 10 MV cm^{-1} , interference or perturbation from the probe (either organic dyes or SERS nanoparticles) to the interfacial water layer is inevitable. Moreover, the size of the probe could be comparable or even larger than the length (presumably on the order of nanometers) of the interfacial water layer, which may lead to systematic deviation. Therefore, a label-free technique that can directly detect the interfacial electric field would be an important complement to existing probe-based techniques. To this end, the intrinsic water OH stretching mode offers a valuable opportunity to investigate the on-water electrostatics in a label-free manner.

Due to the sensitivity of the OH stretching frequency to the local electrostatic environment, vibrational spectroscopy has long been a desirable tool to investigate the electrostatics of interfacial water.^{112–114} One approach to achieve the required surface selectivity is to decompose the Raman/IR spectrum of a mixture containing water and a hydrophobe using the multivariate curve resolution (MCR) algorithm. While this approach was initially developed to study the hydration shell of small solutes for homogenous solutions such as water–methane and water–alcohol mixtures,^{115–118} Shi *et al.* reported the first Raman-MCR spectroscopic study of



mesoscale water–oil emulsions in 2025.⁸⁰ Although Raman spectroscopy is inherently not a surface selective technique, the measured Raman spectrum of emulsions can be modeled and decomposed into a pure water spectrum and a solute-correlated (SC) spectrum with high interface selectivity for an in-solution sample (Fig. 5a).¹¹⁷ The decomposition algorithm is described by the following equation:

$$D = C_B S_B + C_{SC} S_{SC} + E \quad (2)$$

D is the measured Raman spectrum of the emulsion, C_B and C_{SC} are the fractions of the solvent (water) and the oil, respectively, S_B and S_{SC} are the solvent and solute-corrected (contributed from both the oil and the OH stretching mode perturbed by the oil) spectrum, respectively, and E is the residual error matrix.⁸⁰ Its application to water–oil emulsions successfully achieved a mesoscale-resolution measurement of the interfacial water. Furthermore, unlike sum frequency generation spectroscopy (which does not produce a signal on bulk materials), the interfacial water spectrum obtained from Raman-MCR can be readily compared to the bulk water Raman spectrum, as well as hydration shells of other small hydrophobic solutes such as alcohols.^{115,116,118}

Raman-MCR spectroscopy revealed several noteworthy features. Firstly, the characteristic shoulder peak of OH stretching at 3250 cm^{-1} that typically appears in the bulk water's spectrum almost disappeared (Fig. 5b; see the red box).⁸⁰ In bulk water, this shoulder peak is associated with strong hydrogen bonding and structural order.^{119,120} Hence, the vanishing of this spectral feature at 3250 cm^{-1} suggests that the hydrogen bonding network is weakened and more disordered on the mesoscopic water–oil interface. This observation was absent in previously reported SC spectra of small solutes such as *tert*-butyl alcohol.^{80,115} This insight was further supported by molecular-dynamics (MD) simulations. Near the water–oil interface, the Errington–Debenedetti order parameter was substantially reduced, indicating a significantly more disordered and weakened (nearly truncated) hydrogen bonding network at the interface. Secondly, there was a substantial water population with free OH groups on the mesoscopic water–oil interface.⁸⁰ Both the experimental and simulated SC spectrum revealed that the amount of free OH groups made up about 13% of all OH groups within 2 \AA of the Willard–Chandler Interface (WCI).¹²¹ The emergence of these free OH groups is also in line with the weakened hydrogen bonding network at the interface. Thirdly, the stretching frequency of free OH groups in the experimental SC spectrum of water–oil emulsions appeared at $\sim 3575\text{ cm}^{-1}$ (Fig. 5b and c; see the green box), which is redshifted by $\sim 95\text{ cm}^{-1}$ compared to the free OH peak of the planar water–oil (hexane and tetrachloromethane) interface ($\sim 3670\text{ cm}^{-1}$) and water–vapor interface ($\sim 3700\text{ cm}^{-1}$) (Fig. 5d).^{122,123} One likely explanation for this redshift is the VSE, in which a strong electric field at the water–oil interface modifies the free OH vibrational frequency. Using the previously reported Stark tuning rate $\Delta\bar{\nu}$ of $1.6\text{ cm}^{-1}\text{ MV}^{-1}\text{ cm}^{-1}$,¹²⁴ the observed 95 cm^{-1} redshift corresponds to an electric field

of $\sim 60\text{ MV cm}^{-1}$. Note that the previously reported interfacial electric field was $\sim 8.5\text{ MV cm}^{-1}$ in the probe-based method (CN group in Rh800),⁸² which is likely to be an underestimation since the probe's size is comparable to or even larger than the length of the interfacial water layer.

Computational Raman spectrum (both interface and bulk) using a monomer-field Raman model of water reproduced similar spectral features (Fig. 5e).⁸⁰ The shoulder peak at 3250 cm^{-1} was greatly reduced, which is consistent with the more disordered and weakened hydrogen bonds at the interface. However, this model did not consider the presence of interfacial charges, thereby showing a free OH peak between 3650 and 3700 cm^{-1} . This resembles the reported value in the planar water–oil/air interface. By applying an extra interfacial electric field of 67.5 (SPC/E model) or 92.5 MV cm^{-1} (AMOEBa model) to free OH groups at the interface, the stretching frequency of free OH in the simulated SC spectrum would be redshifted to $\sim 3575\text{ cm}^{-1}$ and become nearly identical to the experimental free OH stretching frequency (Fig. 5f). This computational resemblance supports the experimental spectral redshift being a result of the influence of interfacial electric field on free OH groups.

Another important evidence to support the electrostatic origin of the redshift came from surfactant modulation of the interface.⁸⁰ The ζ potential of pristine oil droplets in water was -60 mV . Addition of 0.01% Triton X-100, an amphiphilic, non-ionic surfactant that inserts into the water–oil interface, reduced the oil droplets' ζ potential to -22 mV (Fig. 5g). Correspondingly, the free OH peak then appeared at $\sim 3626\text{ cm}^{-1}$, and the extent of redshift was much lower compared to the surfactant-free emulsion. A smaller ζ potential reflects a weaker interfacial electric field, hence a correspondingly smaller redshift of the free OH group.

Besides spectral decomposition in Raman-MCR, vibrational sum frequency generation (VSFG) spectroscopy, thanks to the inherent surface selectivity of the second-order nonlinear susceptibility $\chi^{(2)}$, has been a widely adopted technique to study interfaces.¹²⁵ In VSFG experiments, an IR and visible beam are spatially and temporally focused on the interface-of-interest. The sum frequency signal is resonantly enhanced when the frequency of the IR beam matches the water vibrational mode on the interface.¹²⁶ VSFG implemented with heterodyne detection also reveals the orientations of interfacial water molecules.^{127,128} Strazdaite *et al.* used heterodyne-detected VSFG to study a wide range of D_2O -hydrophobe interfaces, including air and various oils.¹²⁹ Interestingly, the structure of interfacial water differs markedly between D_2O -air and D_2O -oil interfaces. Compared to D_2O -air interfaces, D_2O -oil interfaces exhibited enhanced hydrogen-bond ordering, as evidenced by their much stronger OD stretch signals in the lower frequency region (2390 to 2500 cm^{-1}) (Fig. 6a). Moreover, $\text{Im}(\chi^{(2)})$ of this OD stretch band in the D_2O -air interface was negative, while that in the D_2O -oil interface was positive. This indicated that hydrogen-bonded OD groups point toward bulk water at the air interface but toward the hydrophobic phase at the oil interface (Fig. 6b). The only similarity was that, in both



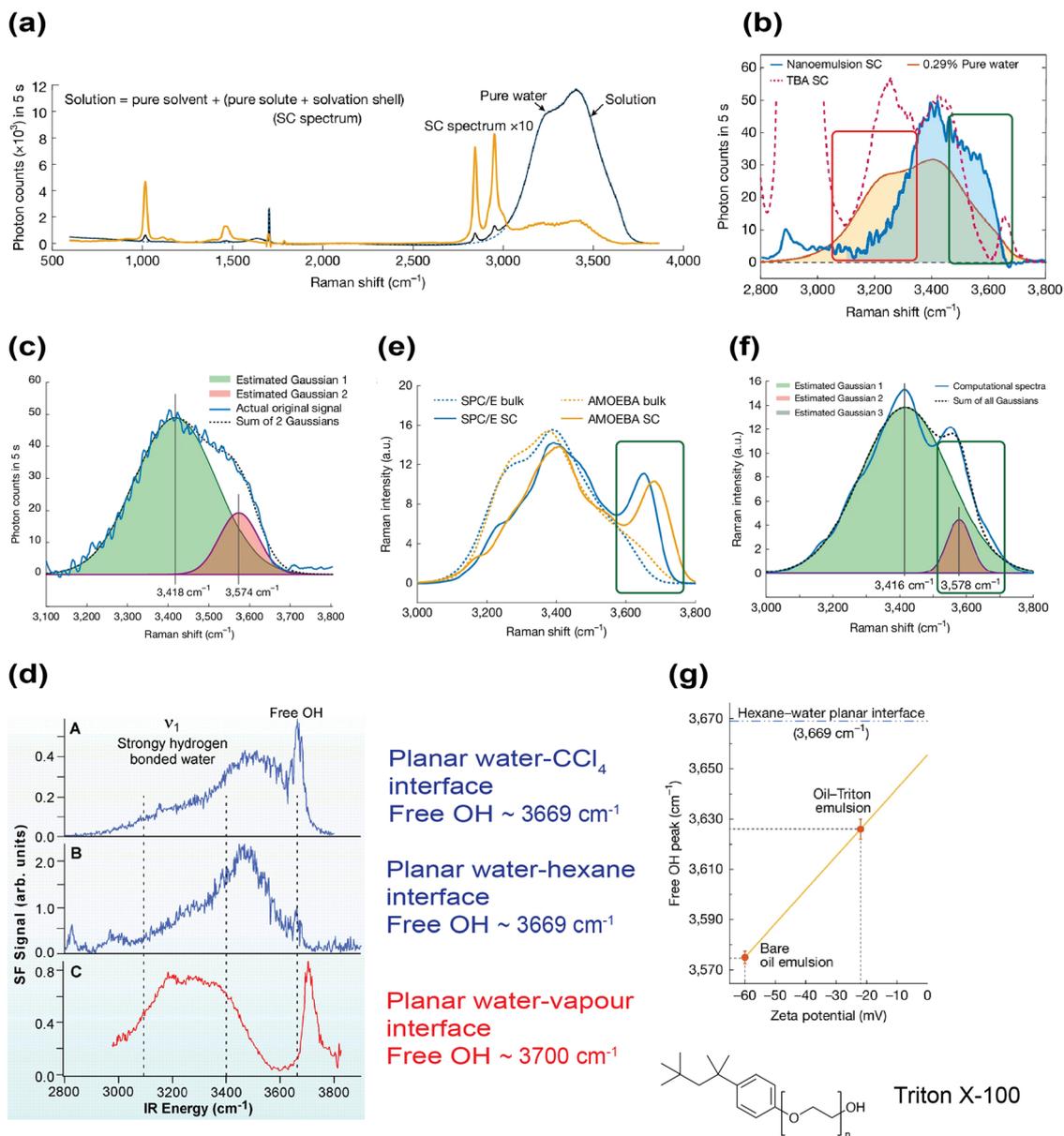


Fig. 5 Quantifying the interfacial electric field's strength using Raman-MCR spectroscopy. (a) A proof-of-concept experiment (0.5 M MeOH in water) showing that the high-resolution Raman spectrum of the solution can be decomposed into the solvent (water) and the SC spectrum (contribution from the solute and the solvation shell) using the MCR algorithm. Reproduced with permission from ref. 80. Copyright 2025, Nature Portfolio. (b) Area-scaled Raman spectrum of pure water (orange line), SC spectrum of water-hexadecane (deuterated) emulsion (blue line) and SC spectrum of 0.5 M TBA solution (red dotted line), respectively. Two characteristic features are highlighted in the SC spectrum of water-hexadecane emulsions, namely reduction of the 3250 cm^{-1} peak intensity (comparing with the pure water spectrum; highlighted in the red box) and redshift of the free OH stretching frequency to 3575 cm^{-1} (comparing with the SC spectrum of TBA; highlighted in the green box). Reproduced with permission from ref. 80. Copyright 2025, Nature Portfolio. (c) Dual Gaussian fit of the water-hexadecane SC spectrum. Note the subpeak at 3574 cm^{-1} . Reproduced with permission from ref. 80. Copyright 2025, Nature Portfolio. (d) The vibrational sum frequency spectrum of the planar water-oil (tetrachloromethane and hexane) and water-vapor interface. Note the free OH stretching frequency, which is $\sim 110 \text{ cm}^{-1}$ higher than that in water-hexadecane emulsions. Reproduced with permission from ref. 123. Copyright 2001, the American Association for the Advancement of Science. (e) Computational Raman spectrum using the monomer-field model, which is parametrized by either the SPC/E or AMOEBA model. Before applying the interfacial electric field, the free OH group stretching frequency (ranging from 3650 to 3700 cm^{-1}) on the simulated SC spectra resembles the case in small solute/planar water-oil/air interface. Reproduced with permission from ref. 80. Copyright 2025, Nature Portfolio. (f) By applying an external electric field (92.5 MV cm^{-1}) on free OH group for the AMOEBA model, the free OH stretching frequency on the simulated SC spectrum is redshifted to 3578 cm^{-1} , similar to the experimental observation. Reproduced with permission from ref. 80. Copyright 2025, Nature Portfolio. (g) Correlation between free OH stretching frequency and the ζ potential. Addition of Triton X-100 reduces the ζ potential of hexadecane droplets in water and the magnitude of the free OH stretching frequency's redshift. Reproduced with permission from ref. 80. Copyright 2025, Nature Portfolio.



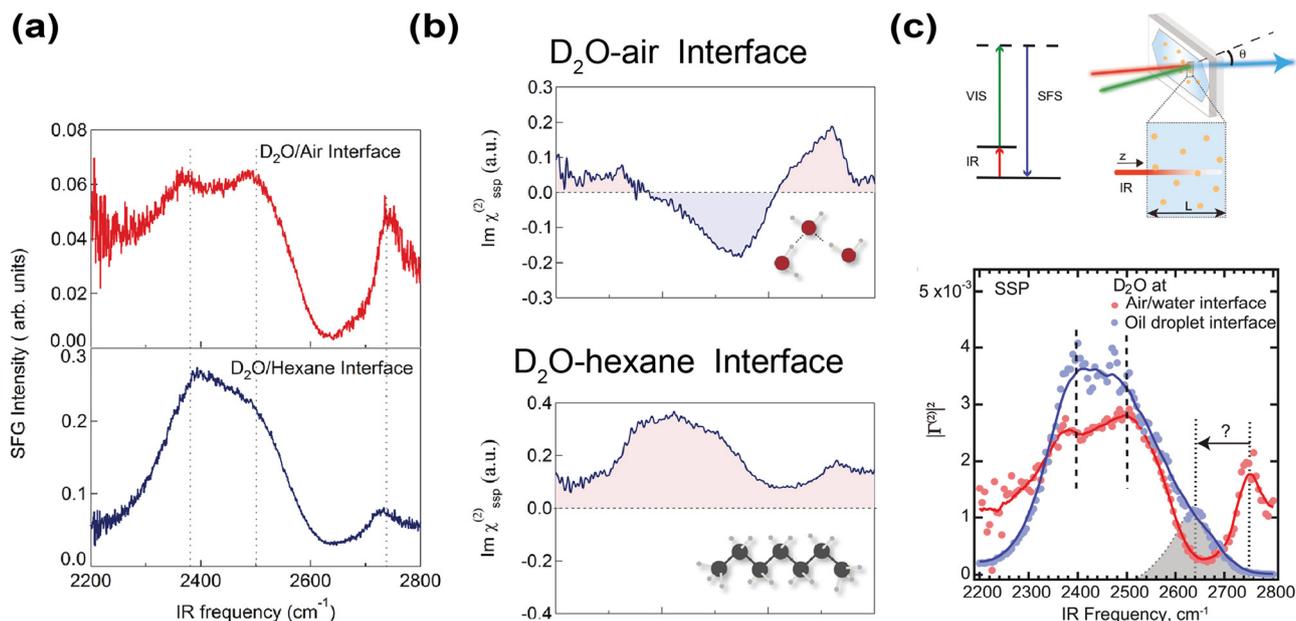


Fig. 6 Probing hydrophobic water interface using sum frequency-based spectroscopy. (a) VSGF spectrum of D₂O–hexane and D₂O–air interface, respectively. Reproduced with permission from ref. 129. Copyright 2015, the American Institute of Physics. (b) VSGF spectrum showing Im(χ⁽²⁾) of D₂O–hexane and D₂O–air interfaces, respectively. Reproduced with permission from ref. 129. Copyright 2015, the American Institute of Physics. (c) Energy level diagram of VSFS and a schematic diagram of the experimental setup. Sum frequency scattering spectrum of D₂O–hexadecane (blue) and water–air interface (red), respectively. Similar to the Raman-MCR spectrum, the stretching frequency of the free OD on the D₂O–hexadecane interface is redshifted compared to the D₂O–air interface. Reproduced with permission from ref. 132. Copyright 2021, the American Association for the Advancement of Science.

D₂O–air/oil interfaces, their free OD groups at higher stretching frequency ($\sim 2750\text{ cm}^{-1}$) oriented toward the hydrophobic phase (Fig. 6b). Their observed spectra were consistent with what Tian *et al.* measured for the water–silane/hexane interface, where Im(χ⁽²⁾) likewise exhibited an enhanced positive band for the OH stretch band ($\sim 3200\text{ cm}^{-1}$) relative to water–air interface, suggesting that the OH groups point towards the hydrophobic phase.¹³⁰ They reasoned that such OH orientation was likely due to the negative surface field arising from OH[−] adsorption on the interface.

While these VSFG studies on planar interfaces yielded valuable information about the interfacial water structure, planar interfaces may not accurately capture chemistry in microdroplets. Moreover, planar interfaces are also more prone to surface contaminations because of a much smaller surface-to-volume ratio. Despite these shortcomings, most VSFG studies have focused on planar interfaces, as probing water–oil emulsions was long considered impossible due to strong IR attenuation and spectral distortion in water.¹³¹ Nevertheless, in 2020, Pullanchery *et al.* demonstrated that vibrational sum frequency scattering (VSFS) could overcome this challenge by correcting the signal with a frequency-dependent normalization factor accounting for IR absorption and efficiency of optical collection, thereby enabling reliable measurements from water–oil emulsions (Fig. 6c).¹³² Compared to planar D₂O–oil/air interfaces, the VSFS spectrum of D₂O–oil emulsions did not show the high-frequency free OD stretching peak ($\sim 2750\text{ cm}^{-1}$) (Fig. 6c). Instead, a new shoulder peak at around 2640 cm^{-1}

was observed, so there was a redshift of around 110 cm^{-1} compared to the planar D₂O–air interface (Fig. 6c). Note the magnitude of this redshift is comparable to that of the free OH peak in Raman-MCR.⁸⁰ Thus, Raman-MCR and VSFS are largely consistent regarding the appearance and stretching frequency of the free OH(D) groups. The VSFS study attributed this redshift to a charge-transfer mechanism, in which electron density was transferred from water to oil molecules, as an explanation for the redshift of the free OD in their VSFS experiments and for the negative ζ potential of oil droplets dispersed in water.^{132,133} However, a recent energy decomposition analysis argued that the forward and backward charge transfer between water and oil nearly cancel so that the net flow charge is negligible.¹³⁴ On the other hand, if one were to interpret the redshift of free OH in VSFS using VSE, nearly identical conclusions about the electric field strength would have been reached.

Apart from the interfacial water structure, VSFG can also provide insight about the adsorption of ions on the hydrophobic water interface, which has been a highly debated topic. For instance, by monitoring the change in Im(χ⁽²⁾) at 3200 cm^{-1} upon ion titration, Tian *et al.* showed that OH[−] had the highest adsorption energy ($\Delta G = -45\text{ kJ mol}^{-1}$) compared to H₃O⁺ ($\Delta G = -39.5\text{ kJ mol}^{-1}$) and Cl[−] ($\Delta G = -28\text{ kJ mol}^{-1}$) on a water–silane interface.¹³⁰ Their findings were later substantiated by Gan *et al.* in the water–hexadecane interface.¹³⁵ Using second harmonic generation spectroscopy and a Langmuir adsorption model, they concluded that OH[−] has an adsorption free energy of $-8.3\text{ kcal mol}^{-1}$, while H₃O⁺ subsequently localizes to the



Table 1 Summary of electric field strengths reported across different methods and systems

Methods	Interfaces	Measured field strengths
SREF-VSE	Water-hexadecane interface (emulsion)	$\sim 4 \text{ MV cm}^{-1}$
SERS	water-air interface (aerosol)	$\sim 10\text{--}100 \text{ MV cm}^{-1}$ (size dependent)
Spectral confocal fluorescence microscopy	water-air interface	$\sim 10 \text{ MV cm}^{-1}$
Raman-MCR	Water-hexadecane interface (emulsion)	$\sim 60 \text{ MV cm}^{-1}$
Electroacoustic Measurement	Water-hexadecane interface (emulsion)	$\sim 55 \text{ MV cm}^{-1}$; deduced from a charge density ($-4.9 \mu\text{C cm}^{-2}$ at pH = 9)
Potentiometric mass titration	Water-Teflon interface (nanoparticles in water)	$\sim 50 \text{ MV cm}^{-1}$; deduced from a charge density ($-4.5 \mu\text{C cm}^{-2}$ at pH = 5)
pH Measurement	water-air interface (microbubbles in water)	$\sim 45 \text{ MV cm}^{-1}$; deduced from surface OH^- excess ($4.2 \times 10^{-11} \text{ mol cm}^{-2}$ at pH = 7)

interface due to electrostatic attraction with OH^- . On the other hand, claims about preferential adsorption of H_3O^+ on the water surface do exist from VSFG experiments. For instance, the sum frequency response of the water-air interface collected by Tarbuck *et al.* at different pH levels revealed that the addition of H_3O^+ caused profound perturbation to the interfacial water structure, including orientation, coordination and strength of hydrogen bond, whereas the perturbation is much less significant when OH^- was added.¹³⁶ Therefore, they concluded that H_3O^+ is enriched on the interface. Their finding is also supported by some subsequent studies.^{137–139} Again, it is worth emphasizing that VSFG experiments employed a planar water interface, not a curved interface where ζ potential is present or where microdroplet chemistry happens. The planar model can potentially neglect the effect of local curvature on interfacial water structure and charge distribution. Although the precise origin and nature of interfacial ions remain debated, extensive evidence supports the presence of excess charges at water interfaces (Table 1), leaving the detailed physical picture an open question.

Connection and consistency with other experimental studies

Is the interfacial electric field of $\sim 60 \text{ MV cm}^{-1}$, as concluded by Raman-MCR and VSE, consistent with other studies performed on similar samples? We would like to point out that there exists a striking consistency between this value and those deduced from non-spectroscopic approaches. In 2004, Beattie and Djerdjev designed an ingenious experiment to estimate the surface charge density of pristine oil-in-water emulsions.⁹⁴ They conducted pH-stat experiments combined with an electroacoustic technique to measure the size and surface charge of oil droplets as new water-oil interfaces were being formed. In their method, an emulsion of hexadecane in water was homogenized in a nitrogen atmosphere. The pH was kept constant by titrating with a proper amount of NaOH. As homogenization proceeded, new water-oil interfaces were generated and the amount of OH^- added to maintain a constant pH was recorded. Meanwhile, electroacoustic measurements allowed real-time monitoring of droplet size and ζ potential during the entire homogenization process. It was revealed that there was a linear

relationship between the amount of added OH^- and the increase in surface area (Fig. 7a). Thus, this suggested an intrinsic and constant surface charge density on the oil-water interface, for which hexadecane oil droplets in water were fitted to be $-4.9 \mu\text{C cm}^{-2}$. Using a similar pH-stat approach, Gan *et al.* estimated a charge density of -0.14 e nm^{-2} for hexadecane oil droplets in water, corresponding to $-2.3 \mu\text{C cm}^{-2}$,¹³⁵ comparable to the value obtained by Beattie and Djerdjev. The difference might be from the higher pH (~ 13) used in Gan *et al.* and/or the error associated with the estimation of the surface area. They also emphasized that if the observed ζ potential arose from carboxylic acid contaminants in hexadecane, such species would have been spectroscopically detectable, as a charge density of -0.14 e nm^{-2} corresponds to a concentration of mM, far above the detection limit of second harmonic generation spectroscopy.¹³⁵

The mathematical relationship between the surface charge density and the electric field strength is described by the Gauss's law (in a vacuum):

$$E = \frac{\rho}{\epsilon_0} \quad (3)$$

E is the electric field, ρ is the surface charge density, and ϵ_0 is the vacuum permittivity (absolute dielectric constant in vacuum). When using eqn (3) to water interfaces, it should be recognized that the dielectric response of interfacial water differs significantly from the bulk water (whose dielectric constant is ~ 78). The effective permittivity of interfacial water, however, has been shown to rapidly approach the vacuum value.^{140,141} Thus, applying eqn (3) and ϵ_0 , the surface charge density ($-4.9 \mu\text{C cm}^{-2}$) estimated by Beattie and Djerdjev can generate an interfacial electric field of $\sim 55 \text{ MV cm}^{-1}$, remarkably consistent with the Raman-MCR inferred value of $\sim 60 \text{ MV cm}^{-1}$.⁸⁰

In addition to the water-oil emulsions above, similar densities (~ 4 to $5 \mu\text{C cm}^{-2}$) of negative surface charge have been observed in solid Teflon nanoparticles (Fig. 7b) and nitrogen gas microbubbles dispersed in water (Fig. 7c).^{142,143} It would be hardly coincidental that all three mesoscopic (*i.e.*, nanodroplets, microbubbles and nanoparticles) hydrophobic phases (lipid, air and solid) of aqueous interfaces exhibit a fairly similar level of surface charge density. Even two-dimensional materials such as hexagonal boron nitride, when in contact



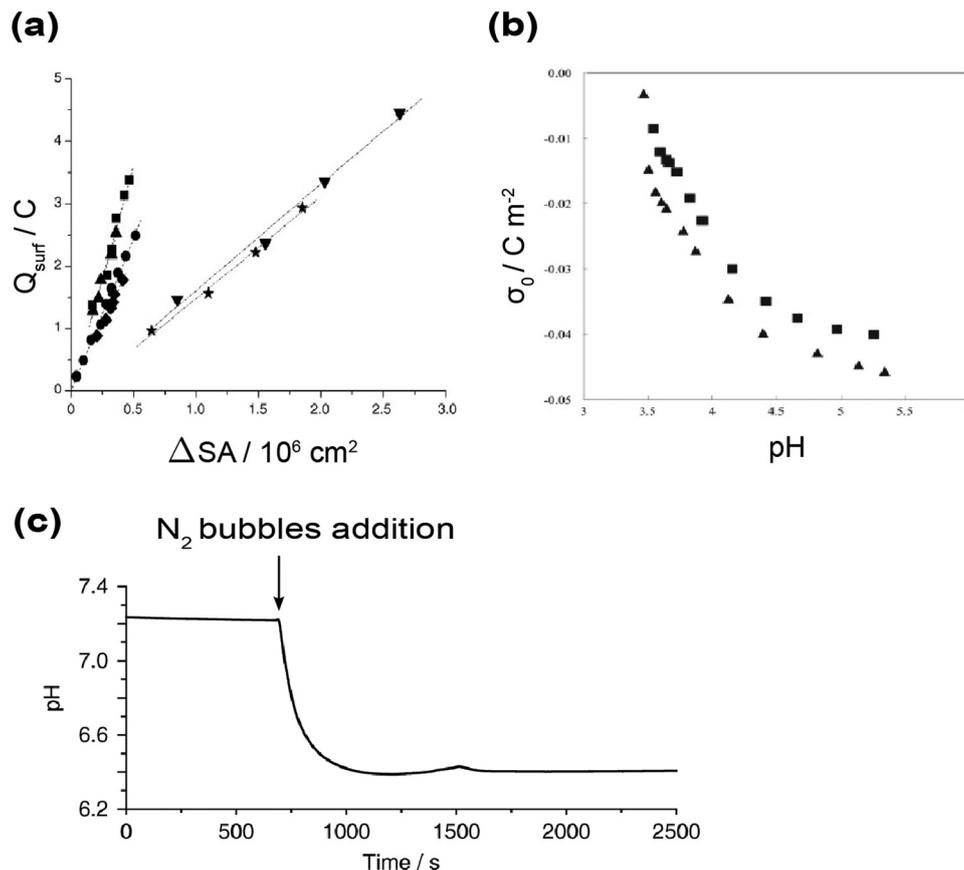


Fig. 7 Non-spectroscopic studies on the electrostatics of hydrophobic water interfaces. (a) Correlation between the change in surface area (ΔSA) and surface charge (Q_{surf}) in different water–oil emulsions: perfluoromethyldecalin (\blacksquare) at pH = 9: $-7.3 \mu\text{C cm}^{-2}$, squalene (\blacktriangle) at pH = 9: $-6.7 \mu\text{C cm}^{-2}$, hexadecane (\bullet) at pH = 9: $-4.9 \mu\text{C cm}^{-2}$, hexadecane (\blacklozenge) at pH = 7: $-4.6 \mu\text{C cm}^{-2}$, hexadecane with 2 mM SDS (\blacktriangledown) at pH = 9: $-1.7 \mu\text{C cm}^{-2}$, and squalene with 2 mM SDS (\blackstar) at pH = 9: $-1.6 \mu\text{C cm}^{-2}$ respectively. Reproduced with permission from ref. 94. Copyright 2004, Wiley-VCH. (b) Teflon surface charge density as a function of pH. At pH = 5, the surface charge density on Teflon is around $-4.5 \mu\text{C cm}^{-2}$. Reproduced with permission from ref. 142. Copyright 2012, Elsevier. (c) Reported pH drop when nitrogen bubbles are added to ultrapure water. Based on the pH drop, the calculated surface OH^- excess is calculated to be $4.2 \times 10^{-11} \text{ mol cm}^{-2}$ in nearly neutral pH. This corresponds to a surface charge density of around $-4 \mu\text{C cm}^{-2}$. Reproduced with permission from ref. 143. Copyright 2020, Nature Portfolio.

with water, were found to acquire a surface charge density of approximately $-2.3 \mu\text{C cm}^{-2}$ at pH = 7, which was assigned to OH^- chemisorption and physisorption.¹⁴⁴ Applying the Gauss's law again to these systems would result in an interfacial electric field with comparable strength to the one determined for the water–oil interface. Considering the wide range of experimental techniques employed and interfacial water samples examined, the close agreement among these electric field strengths is notable and strongly suggests a common underlying principle for all water–hydrophobe interfaces.

Possible mechanism of hydroxide accumulation on hydrophobic interfaces

Despite still being a controversial topic with ongoing debates, mounting experimental evidence has indicated the preferential accumulation of OH^- ions on water–hydrophobe

interfaces,^{91,94,96} regardless of the specific hydrophobic material involved (*e.g.*, air and oil). Unfortunately, the underlying driving force of this phenomenon remains sparsely discussed in the literature. Here, we highlight an insightful work of Gray-Weale and Beattie, who proposed a thermodynamically grounded framework that coherently accounts for the available experimental observations.⁷⁸ They attribute the interfacial adsorption of OH^- to a suppression in the collective dipole-moment fluctuations of nearby water, which leads to a free energy reduction near interfaces. The depth of the potential well ($\sim 20 k_{\text{B}}T$) quantitatively underscores the energetic favorability and strength of this adsorption preference. More broadly, this mechanism unifies seemingly disparate systems, including water–oil, water–air, and water–solid interfaces.

Central to this so-called fluctuation-suppression model is the ion's ability to lower the relative permittivity of an aqueous solution, and this can be quantitatively described by the following equation:

$$\varepsilon(c) = \varepsilon_{\text{w}} - \delta \cdot c \quad (4)$$



c is the molar concentration of the electrolyte, ϵ_w is the relative permittivity of pure water (~ 78), and δ is the dielectric decrement. Experimental studies have shown that OH^- exhibits almost the largest dielectric decrement among all monovalent anions, such as Cl^- .^{145,146} This is plausibly related to the unique solvation structure proposed in other studies, where water molecules are tightly coordinated to the oxygen atom of OH^- .^{147–150} This property uniquely positions OH^- to accumulate at the interface, a behavior made even more remarkable by the fact that OH^- is a natural product of water's autoionization and is inherently present in all aqueous systems.

As illustrated in Fig. 8a, an anion in a homogeneous polar solvent imposes constraints on its solvation shell, suppressing local collective dipole-moment fluctuations. The surrounding region, which would normally exhibit correlated fluctuations with the constrained shell, is disrupted—resulting in a free energy penalty. Although the correlation strength decays smoothly with distance, the two-region model (inner constrained zone and outer correlation zone) serves as a useful conceptual simplification. However, when the ion approaches a hydrophobic interface, as shown in Fig. 8b, this energetic penalty is reduced. This is because part of the fluctuation-affected region overlaps with the low-permittivity phase (air, oil or solid, compared to that of water), where water molecules are absent. As a result, fewer “broken” correlations occur within range, and the corresponding free energy cost is diminished. This translates into a net force that attracts ions towards the interface with a vacuum or another medium of low permittivity,

which is analogous in form to Hamaker-type dispersion interactions.¹⁵¹

The full derivation of the fluctuation–suppression force can be found in the original work. Here we briefly summarize the key result. Total free energy is modeled as the sum of two components: a fluctuation-suppression term $v(z)$ and a self-image repulsion term $u(z)$, where z is the distance from the ion. The former arises from the loss of solvent dipole correlations as discussed above. The latter accounts for electrostatic image forces that act to repel the ion from the low-permittivity phase, which dominates at very short distances.¹⁵¹ The total potential of mean force, $v(z) + u(z)$, is calculated and plotted as shown in Fig. 8c. In the case of OH^- , a conservatively estimated dielectric decrement ($\delta_- = 10.7 \pm 3.1 \text{ M}^{-1}$) is employed. The model predicts a deep potential well with a minimum around 6.7 \AA and a depth of approximately $20 k_B T$.

This magnitude has been validated by a very recent experimental study, which reported an even higher adsorption free energy of $28.4 k_B T$ per OH^- at the water–oil interface.¹⁵² This value was determined by analyzing the heat released during acid-induced breaking of pristine oil-in-water emulsions, after subtracting the estimated contributions from neutralization and interfacial tension. Consistent results for the desorption enthalpy were observed across three systems: two emulsions with isopropyl myristate and one with hexadecane.

According to Boltzmann statistics, surface ion concentration is expected to scale exponentially with the depth of this potential well. Thus, the $\sim 20 k_B T$ well for OH^- leads to a

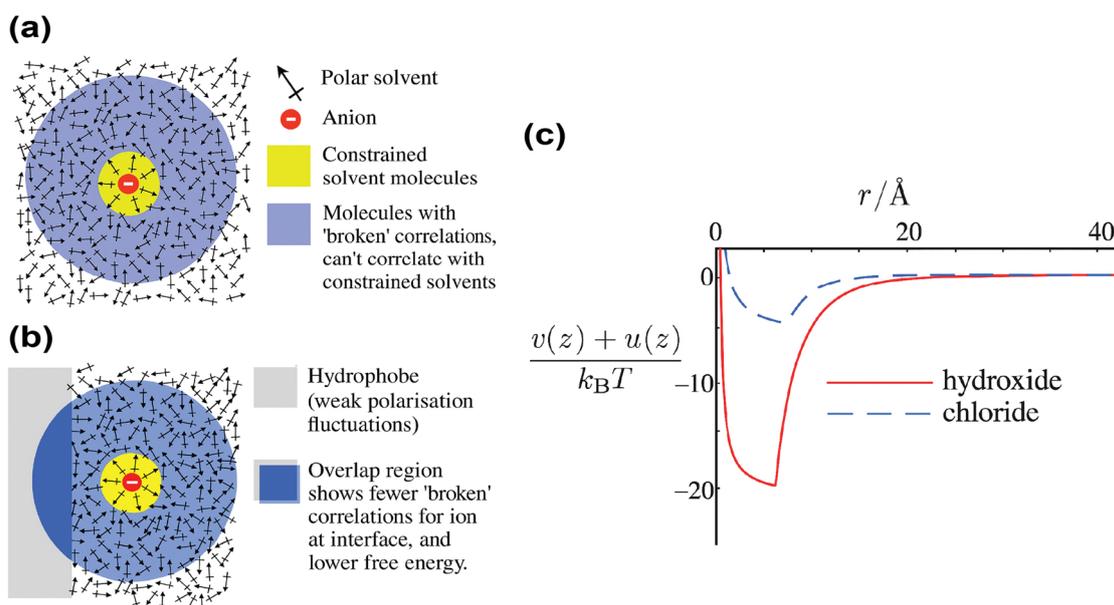


Fig. 8 The fluctuation–suppression model and the potential profiles for OH^- and Cl^- . (a) An ion in a polar solvent: molecules in the constrained solvation shell (yellow) are not as free as the others to fluctuate, breaking correlations in the surrounding region (larger blue circle). (b) When the same ion is near a hydrophobic interface (gray), part of the correlation zone overlaps with the low-permittivity phase, reducing the number of broken correlations and thus the free energy cost. (c) Calculated potentials of mean force for hydroxide and chloride ions. OH^- (red) exhibits a deep potential well ($\sim 20 k_B T$) at $\sim 6.7 \text{ \AA}$, consistent with strong interfacial adsorption; Cl^- (blue) shows a much shallower well. Reproduced with permission from ref. 78. Copyright 2009, the Royal Society of Chemistry.



significant interfacial enrichment, while other anions such as Cl^- , with much shallower wells, show lower surface affinity. The study calculated that each OH^- within the shear plane occupies an area of approximately 4.5 nm^2 . This theoretical estimate closely matches the experimental value of $\sim 3.2 \text{ nm}^2$ reported earlier by Beattie and Djerdjev.⁹⁴ Such close agreement between theory and experiment reinforces the credibility of the model and confirms the high interfacial density of OH^- .

Notably, this model successfully reproduces a wide range of experimental observations, including low isoelectric points of hexadecane emulsions,¹⁵³ the pH dependence of ζ potentials of hexadecane droplets,⁹⁴ and the Jones-Ray effect as reflected in surface tension measurements.^{154–156} To predict ζ potential, by numerically solving a modified Poisson–Boltzmann equation that incorporates the fluctuation–suppression potential, the electrostatic potential profile near the interface can be obtained. The slip plane was assumed to lie 2.5 nm away from the interface, and the potential at that position was taken as the ζ potential. By varying the fluctuation-suppression radius a , the simulated ζ potential curves show excellent agreement with experimental data across a wide pH range, with the best fit achieved at $a = 6.7 \text{ \AA}$. This value corresponds to a potential well depth of $20 k_{\text{B}}T$, as discussed above. In addition, the predicted isoelectric point was shown to be highly sensitive to both the dielectric decrement δ and the value of a . A decrement as small as that of chloride (5 M^{-1}), or a slightly larger a , would shift the isoelectric point upward toward neutral pH, in conflict with experimental results. The model thus naturally explains the low isoelectric point near pH 3 to 4. Importantly, these predictions arise directly from the computed potential well, without reliance on empirical fitting parameters. Moreover, it accounts for previous contradictions in surface-sensitive spectroscopic data, such as VSFG,^{137,139} by suggesting that OH^- mostly resides below the outermost molecular layers, beyond the typical probing depth of those interface selective techniques.

Implications of on-water electric fields on chemical reactivity

What are the consequences of a strong electric field on chemistry? One elegant and well-established example to demonstrate the effect of electric field on chemistry is enzyme catalysis. As demonstrated by Boxer and co-workers, the active site of an enzyme can exert a large and homogenous electric field on the substrate through pre-organizing its residues to stabilize the transition state, thereby reducing the activation barrier.¹⁵⁷ Different from reactions in bulk solution, where solvent molecules mostly interact and stabilize the ground state of the reactant molecule, the active site of an enzyme has evolved to orient the dipoles and charges of amino acid residues so that the resulting electric field is strong and precisely focused on specific bonds of the substrate, especially those undergoing charge rearrangement in the transition state. An enzyme to showcase this principle is ketosteroid isomerase (KSI).¹⁵⁸ KSI catalyzes the isomerization of 3-oxo- Δ^5 -steroid to 3-oxo- Δ^4 -steroid,

which involves the conversion of a ketone ($\text{C}=\text{O}$) to a charged enolate ($\text{C}-\text{O}^-$). Using the vibrational frequency of the ketone group as the Stark probe, KSI was shown to exert a strong electric field ($\sim 150 \text{ MV cm}^{-1}$) via the OH and COOH groups in Tyr16 and Asp103, respectively. Moreover, by mutating the wild-type residues, the kinetic barrier for forming the enolate intermediate was found to increase from ~ 11 to 16 kcal mol^{-1} while the electric field was reduced from ~ 150 to 60 MV cm^{-1} . Taken together, enzyme catalysis highlights how an electric field can be a robust regulator of reactivity.

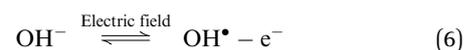
Just as enzymes exploit localized electric fields in their active sites to stabilize transition states, aqueous microdroplets have also been experimentally shown to possess a strong electric field at their interfaces, to which their unusual reactivity is often attributed. Herein, we would like to focus on the chemical reactivity of microdroplets. If such an electric field truly exists on microdroplet interfaces, it can potentially account for the accelerated kinetics and altered thermodynamics in microdroplet chemistry (Fig. 9a). We will illustrate by using simple chemical principles.

A strong interfacial electric field will greatly impact the chemical kinetics observed in microdroplets. The effect of electric field on the activation energy can be captured by the following equation:

$$\Delta\Delta G^\ddagger = -\Delta\vec{\mu}_{\text{rxn}} \cdot \vec{F} \quad (5)$$

$\Delta\Delta G^\ddagger$, $\Delta\vec{\mu}_{\text{rxn}}$ and \vec{F} are the change in activation free energy, the change in dipole moment between ground state and transition state, and the external electric field, respectively.^{157,159} Assuming a dipole moment difference of 2 Debye between the transition state and the reactant ground state, which is a reasonable magnitude for many reactions (for instance the dipole moment difference for *tert*-butylchloride in a $\text{S}_{\text{N}}1$ reaction is ~ 5.5 Debye),¹⁵⁷ a lower bound of the interfacial electric field ($\sim 50 \text{ MV cm}^{-1}$) can lower the free energy barrier by $4.8 \text{ kcal mol}^{-1}$. According to simple transition state theory, this will accelerate the rate constant by 3 to 4 orders of magnitude and thus it is a key factor for explaining the rate acceleration of many reactions in aqueous microdroplets relative to the bulk solvent.

Take the conversion of OH^- to hydroxyl radicals as a concrete example with an immediate implication to redox chemistry. While examining the energetics of water back in 1988, Kloss suggested that a minor fraction of OH^- exists as a radical-electron pair ($\text{OH}^\bullet - \text{e}^-$), which may reversibly dissociate into a hydroxyl radical and an electron when it is subject to an external perturbation such as an electric field in pure water.¹⁶⁰



Along this line, a sophisticated calculation using MD simulations from Head-Gordon and co-workers suggested that OH^- are under-coordinated on the droplet surface relative to the bulk, exhibiting much lower vertical ionization energy.¹⁶¹ Such partial solvation is likely related to the significantly weakened (nearly-truncated) hydrogen bonding network of the interfacial



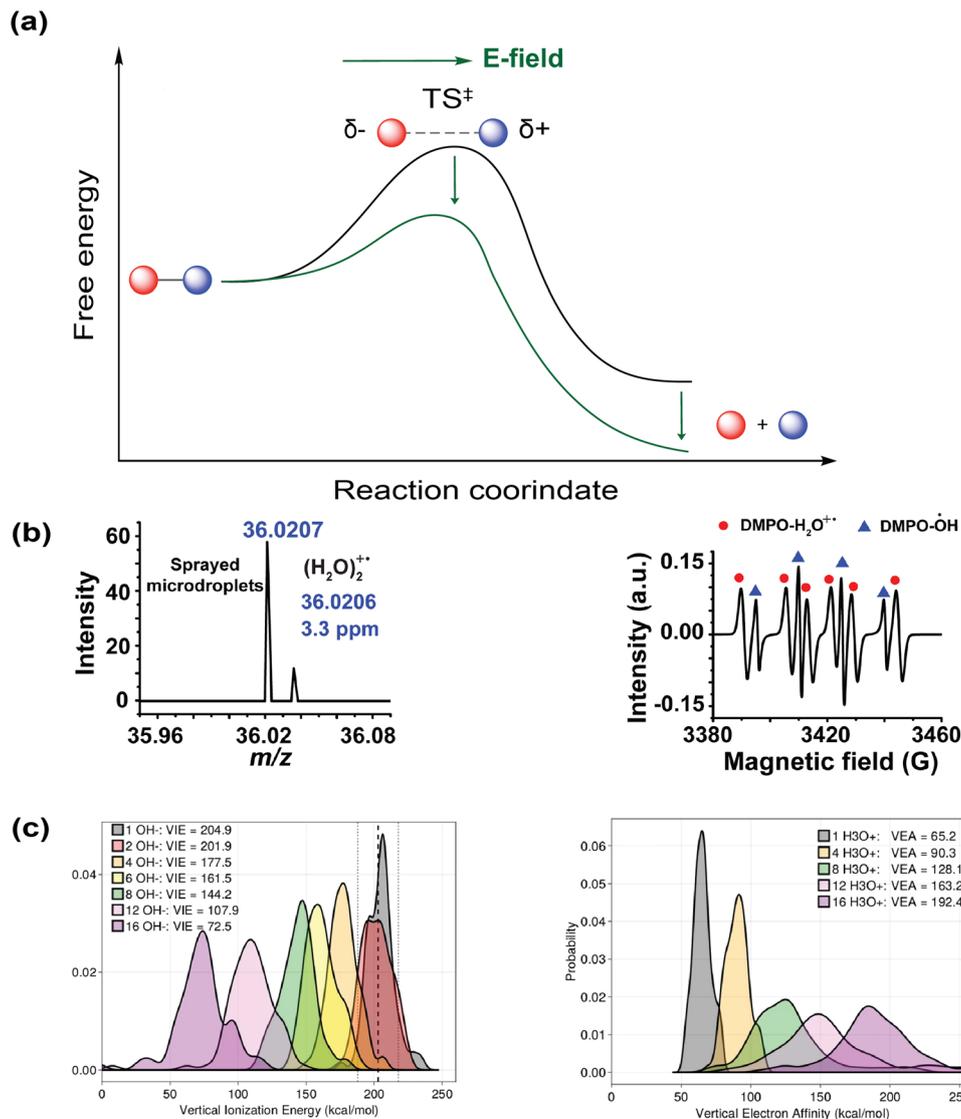


Fig. 9 Implications of the interfacial electric field on chemical kinetics and thermodynamics. (a) A schematic diagram showing the altered reaction kinetics and thermodynamics under the influence of an external electric field. (b) Generation of water dimer radical cation is detected using high-resolution mass spectrometry (note the peak with *m/z* = 36.0207) and electron paramagnetic resonance spectroscopy. Reproduced with permission from ref. 33. Copyright 2025, the American Chemical Society. (c) MD simulations showing that in the presence of excess charges in a microdroplet, the vertical ionization energy of OH⁻ is greatly reduced, whereas the vertical electron affinity of H⁺ is significantly increased. Reproduced with permission from ref. 169. Copyright 2024, Nature Portfolio.

water itself, as unveiled by the recent Raman-MCR spectroscopic study.⁸⁰ Consistent with Kloss's vision, this simulation further revealed that a field strength of 80 MV cm⁻¹ exerting ~5 Å is sufficient to convert the interfacial OH⁻ to a hydroxyl radical.¹⁶¹ Note that the Raman-MCR inferred value of ~60 MV cm⁻¹ is an average over space and time. Thus, its fluctuations around the average are inevitable, resulting in a rather large probability of reaching the required 80 MV cm⁻¹,^{84,162} thereby catalyzing the radical generation. The interfacial electric field is also believed to be connected to the curvature of microdroplets.¹⁶³ Different from the above mechanism of hydroxyl radical oxidizing from OH⁻, it has been recently hypothesized that the strong electric field can also

promote the natural dissociation of water into radical cation/anion pairs (H₂O^{•+}/H₂O^{•-}) and separate the free radical ions into different layers.^{164,165} While water is more abundant than OH⁻ at the interface, the loss of an electron from OH⁻ is expected to be much lower in energy. Experimentally, Zare, Zhang and co-workers reported the capture of hydroxyl radicals in the form of OH⁻-H₃O⁺ (hydrogen bonding with a hydronium cation) using a mass spectrometer.²⁹ Moreover, its isomeric configuration, water dimer radical cation (H₂O)₂^{•+}, has been recently detected using both high-resolution mass spectrometry (observed peak at *m/z* 36.0207 vs. the calculated exact mass of 36.0206 with a mass accuracy error of 3.3 ppm) and electron paramagnetic resonance spectroscopy (Fig. 9b).³³ The hydroxyl



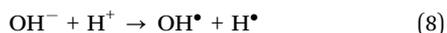
radicals and hydrated free electrons from interfacial OH⁻ can trigger a plethora of downstream redox reactions.

Besides, the electric field and the associated electrostatic charge can also alter the thermodynamics of reactions profoundly. Intuitively, an electric field can align and enrich charged or polar molecules near the interface and consequently minimize the entropic barrier of chemical reactions. However, chemical thermodynamics textbooks virtually neglect the effect of matter's electrification. This had promoted McCarthy and Whitesides to ask "One of the basic assumptions of chemical thermodynamics is that bulk matter is electrically neutral. ... What is the chemistry of materials that bear a net electrostatic charge?"¹⁶⁶ Quantitatively, the electrochemical potential μ_i of an ionic species i in an aqueous solution is governed by the following equation:

$$\mu_i = \mu_i^\circ + RT \ln a_i + z_i FV \quad (7)$$

μ_i° is the standard chemical potential of species i , a_i is the thermodynamic activity of species i , R is the ideal gas constant, T is the temperature, z_i is the charge of species i , F is the Faraday constant, and V is the local potential.^{167,168} The first part of the equation $\mu_i = \mu_i^\circ + RT \ln a_i$ is the textbook knowledge when describing chemical thermodynamics. However, the $z_i FV$ term is often neglected ($V = 0$) unless the reaction involves a charged interface (*i.e.*, an electrode), an external potential, or the material's electrification.

If one puts the above equation in the context of redox microdroplet chemistry, it can provide an explanation of why some thermodynamically unfavorable reactions (*i.e.*, H₂O₂ generation) can occur spontaneously in a microdroplet environment. Head-Gordon and co-workers elucidated the role of charge in microdroplet chemistry and explicitly emphasized that the reaction thermodynamics in charged microdroplets, especially for redox reactions, are not the same as those occurring in bulk solution.¹⁶⁹ In their simulations, they examined the following reaction:



This reaction is normally highly endothermic (+107 kcal mol⁻¹), yet in a water microdroplet with excess charge (~20 to 50% of the Rayleigh limit), it is shown that the reaction thermodynamics can be altered dramatically. The strong Coulombic repulsion within the droplet destabilizes solvated ions. Increasing the net charge of a 4-nm water microdroplet (from ±1 to ±16) substantially reduces the hydration enthalpy, shifting from -126 to +10 kcal mol⁻¹ for OH⁻ and from -263 to -146 kcal mol⁻¹ for H⁺. Thus, the thermodynamic penalty in bulk solution for removing charged species is relieved due to the presence of excess charges in microdroplets, *i.e.*, eqn (8) becomes favorable. At the same time, the vertical ionization energy of OH⁻ decreases whereas the vertical electron affinity of the H⁺ increases in the presence of charges, indicating that electron transfer between these species becomes more electronically favored (Fig. 9c). Moreover, these results are not confined to nanoscale droplets but can be directly extended to the

micro-sized ones, which are more relevant to real experimental conditions. Altogether, in charged microdroplets, the combined shifts in hydration and ionization energy levels act together to make radical formation thermodynamically spontaneous. This provides a molecular-level explanation to rationalize why simultaneous reduction and oxidation become possible in a microdroplet environment,³⁶ despite the thermodynamic barrier.

Conclusions and outlook

Building upon and inspired by pioneering work in "on-water" catalysis and the emerging microdroplet chemistry, this review systematically summarizes the presence, characteristics, and implications of strong electric fields on different water-hydrophobe interfaces. A wide variety of experimental methods, including fluorescence imaging, SREF-vibrational Stark spectroscopy, electrochromism, Raman-MCR, and VSFG spectroscopy, among others, have collectively provided extensive evidence substantiating the existence of an electric field at water interfaces. Remarkably, the inferred strengths of these fields all fall within the range of tens of MV cm⁻¹, in close agreement with values independently estimated from electrokinetic and surface charge measurements. This strongly hints at the existence of strong electric fields being a general feature of water-hydrophobe interfaces. Moreover, this review highlights an intriguing thermodynamic mechanism proposed by Gray-Weale and Beattie that is able to explain the accumulation of OH⁻ at interfaces and reproduce a wide range of experimental observations such as high interfacial density of OH⁻, isoelectric points of hexadecane emulsions, and the pH dependence of ζ potentials of hexadecane droplets. Central to this mechanism is the suppression of dielectric fluctuations and the associated reduction in free energy near the interface.

Our current understanding of hydrophobic water interfaces is still not complete.¹⁷⁰⁻¹⁷³ Firstly, all aforementioned spectroscopic/microscopic studies of water interfaces capture steady-state response. Important dynamic features, including hydrogen bond rearrangements, vibrational relaxation timescales, transient fluctuations of ions on interfaces, *etc.* are still unclear. To this end, time-resolved spectroscopy, such as time-resolved VSFG, can offer a powerful method moving forward. It can directly probe how the vibrational frequency of water fluctuates in time (spectral diffusion) and water reorientation kinetics.¹⁷⁴⁻¹⁷⁶ Beyond the temporal aspect, another incomplete puzzle lies in the spatial resolution, as the reported electric field strength was derived from ensemble-averaged measurements. Future progress will benefit from methods that can directly probe the spatially resolved water structure and electric fields. Moreover, it should also be recognized that the role of the interfacial electric field may be coupled to, or confounded with, other effects like solvation. For instance, a recent simulation has indicated that the solvation structure of a molecule on the interface can largely determine the electric field it experiences.¹⁷⁷ Further studies



on disentangling the relative contributions to the observed reactivity are highly desirable. Finally, current studies have not conclusively determined the identity of the charged species on the interface. While ζ potential measurements for solids, gas bubbles and oil droplets dispersed in water almost unequivocally suggest that hydrophobic water interfaces are negatively charged,^{93,96,98} spectroscopic evidence remains less conclusive. On the theoretical side, some earlier simulations predicted preferential accumulation of H_3O^+ on the interface,^{90,178,179} and recent ones suggest the presence of both H_3O^+ and OH^- .^{180,181} Thus, experiments that can reconcile the measured ζ potential with the identity of the charged species will be incredibly constructive.

The presence of these intrinsic electric fields would reshape our understanding of how interfacial environments influence chemical reaction kinetics and thermodynamics. As shown by simple chemical principles, strong electric fields can potentially account for the accelerated kinetics, altered thermodynamics and radical-based redox reactions. Harnessing interfacial electrostatics thus offers a promising engineering strategy for modulating chemical reactivity and aqueous-phase synthesis, as being implemented most recently.¹⁸² Continued advances in spectroscopy and imaging are anticipated to illuminate the intricate relationships between interfacial water structure, electrostatic environment, and chemical reactivity. Such advances will be critical to developing predictive models of interfacial chemistry. This will also pave the way for future innovations across different areas, ranging from catalysis and materials synthesis to atmospheric chemistry and environmental remediation, where interfacial processes play a central role.

Conflicts of interest

There are no conflicts to declare.

Data availability

No primary research results, software or code have been included and no new data were generated or analyzed as part of this review.

Acknowledgements

W. M. acknowledges support from the Multi-University Research Initiative (MURI) of the Air Force Office of Scientific Research (FA9550-21-1-0170), the National Institute of Health (R35 GM149256), and Chan Zuckerberg Initiative (Dynamic Imaging 2023-321166). K. Y. C. acknowledges Croucher Scholarships for Doctoral Study (Croucher Foundation, Hong Kong).

References

- D. C. Rideout and R. Breslow, *J. Am. Chem. Soc.*, 1980, **102**, 7816–7817.
- R. Breslow, U. Maitra and D. Rideout, *Tetrahedron Lett.*, 1983, **24**, 1901–1904.
- R. Breslow, *Acc. Chem. Res.*, 1991, **24**, 159–164.
- S. Narayan, J. Muldoon, M. G. Finn, V. V. Fokin, H. C. Kolb and K. B. Sharpless, *Angew. Chem., Int. Ed.*, 2005, **117**, 3339–3343.
- Y. Jung and R. A. Marcus, *J. Am. Chem. Soc.*, 2007, **129**, 5492–5502.
- A. Chanda and V. V. Fokin, *Chem. Rev.*, 2009, **109**, 725–748.
- M. F. Ruiz-Lopez, J. S. Francisco, M. T. Martins-Costa and J. M. Anglada, *Nat. Rev. Chem.*, 2020, **4**, 459–475.
- Z. Wei, Y. Li, R. G. Cooks and X. Yan, *Annu. Rev. Phys. Chem.*, 2020, **71**, 31–51.
- R. A. LaCour, J. P. Heindel, R. Zhao and T. Head-Gordon, *J. Am. Chem. Soc.*, 2025, **147**, 6299–6317.
- X. Jia, J. Wu and F. Wang, *JACS Au*, 2024, **4**, 4141–4147.
- H. Chen, X. Li, B. Li, Y. Chen, H. Ouyang, Y. Li and X. Zhang, *J. Am. Chem. Soc.*, 2025, **147**, 11399–11406.
- M. F. Ruiz-López and M. T. Martins-Costa, *Phys. Chem. Chem. Phys.*, 2022, **24**, 29700–29704.
- Y. Wang, Y. Ju, J. He, J. Zhao, Z. Zhou, J. Jiang and H. Zhang, *J. Phys. Chem. Lett.*, 2025, **16**, 2621–2626.
- Y.-W. Zhou, M.-Y. Jia, J.-L. Yang, Q. Liu and Z.-F. Cai, *Chem. Sci.*, 2025, **16**, 8470–8477.
- X. Yan, R. M. Bain and R. G. Cooks, *Angew. Chem., Int. Ed.*, 2016, **55**, 12960–12972.
- G. Rovelli, M. I. Jacobs, M. D. Willis, R. J. Rapf, A. M. Prophet and K. R. Wilson, *Chem. Sci.*, 2020, **11**, 13026–13043.
- K. R. Wilson and A. M. Prophet, *Annu. Rev. Phys. Chem.*, 2024, **75**, 185–208.
- S. Banerjee, E. Gnanamani, X. Yan and R. N. Zare, *Analyst*, 2017, **142**, 1399–1402.
- M. Girod, E. Moyano, D. I. Campbell and R. G. Cooks, *Chem. Sci.*, 2011, **2**, 501–510.
- J. K. Lee, S. Banerjee, H. G. Nam and R. N. Zare, *Q. Rev. Biophys.*, 2015, **48**, 437–444.
- R. M. Bain, S. Sathyamoorthi and R. N. Zare, *Angew. Chem., Int. Ed.*, 2017, **56**, 15083–15087.
- J. K. Lee, K. L. Walker, H. S. Han, J. Kang, F. B. Prinz, R. M. Waymouth, H. G. Nam and R. N. Zare, *Proc. Natl. Acad. Sci. U. S. A.*, 2019, **116**, 19294–19298.
- M. A. Mehrgardi, M. Mofidfar and R. N. Zare, *J. Am. Chem. Soc.*, 2022, **144**, 7606–7609.
- D. Nguyen and S. C. Nguyen, *J. Phys. Chem. B*, 2022, **126**, 3180–3185.
- A. Gallo Jr, N. H. Musskopf, X. Liu, Z. Yang, J. Petry, P. Zhang, S. Thoroddsen, H. Im and H. Mishra, *Chem. Sci.*, 2022, **13**, 2574–2583.
- M. Angelaki, Y. Carreira Mendes Da Silva, S. Perrier and C. George, *J. Am. Chem. Soc.*, 2024, **146**, 8327–8334.
- K. Li, Y. Guo, S. A. Nizkorodov, Y. Rudich, M. Angelaki, X. Wang, T. An, S. Perrier and C. George, *Proc. Natl. Acad. Sci. U. S. A.*, 2023, **120**, e2220228120.
- M. A. Eatoo and H. Mishra, *Chem. Sci.*, 2024, **15**, 3093–3103.



- 29 D. Xing, Y. Meng, X. Yuan, S. Jin, X. Song, R. N. Zare and X. Zhang, *Angew. Chem., Int. Ed.*, 2022, **134**, e202207587.
- 30 A. J. Colussi, *J. Am. Chem. Soc.*, 2023, **145**, 16315–16317.
- 31 C. J. Chen and E. R. Williams, *Angew. Chem., Int. Ed.*, 2024, **136**, e202407433.
- 32 D. Xing, X. Gao, H. Chen, J. Zhang, M. E. Edwards, C. Liang, C. Zhu, Y. Meng, R. N. Zare, Y. Xia and X. Zhang, *Anal. Chem.*, 2025, **97**, 7995–8000.
- 33 X. Zhang, M. Duan, K. Chingjin, R. Balabin, L. Liu, X. Zhang and H. Chen, *Anal. Chem.*, 2025, **97**, 16004–16010.
- 34 Y. Tian, L. Kong, H. Zou, M. Liu and G. Liu, *Sustain. Horiz.*, 2024, **11**, 100103.
- 35 S. Jin, H. Chen, X. Yuan, D. Xing, R. Wang, L. Zhao, D. Zhang, C. Gong, C. Zhu, X. Gao, Y. Chen and X. Zhang, *JACS Au*, 2023, **3**, 1563–1571.
- 36 L. Qiu and R. G. Cooks, *Angew. Chem., Int. Ed.*, 2022, **134**, e202210765.
- 37 C. Zhu, L. N. Pham, X. Yuan, H. Ouyang, M. L. Coote and X. Zhang, *J. Am. Chem. Soc.*, 2023, **145**, 21207–21212.
- 38 Y. Meng, E. Gnanamani and R. N. Zare, *J. Am. Chem. Soc.*, 2022, **144**, 19709–19713.
- 39 Z. Song, C. Liang, K. Gong, S. Zhao, X. Yuan, X. Zhang and J. Xie, *J. Am. Chem. Soc.*, 2023, **145**, 26003–26008.
- 40 D. Zhang, X. Yuan, C. Gong and X. Zhang, *J. Am. Chem. Soc.*, 2022, **144**, 16184–16190.
- 41 X. Yuan, D. Zhang, C. Liang and X. Zhang, *J. Am. Chem. Soc.*, 2023, **145**, 2800–2805.
- 42 A. Nandy, A. Kumar, S. Mondal, D. Koner and S. Banerjee, *J. Am. Chem. Soc.*, 2023, **145**, 15674–15679.
- 43 H. Chen, R. Wang, J. Xu, X. Yuan, D. Zhang, Z. Zhu, M. Marshall, K. Bowen and X. Zhang, *J. Am. Chem. Soc.*, 2023, **145**, 2647–2652.
- 44 L. Zhao, X. Song, C. Gong, D. Zhang, R. Wang, R. N. Zare and X. Zhang, *Proc. Natl. Acad. Sci. U. S. A.*, 2022, **119**, e2200991119.
- 45 K. Gong, Y. Meng, R. N. Zare and J. Xie, *J. Am. Chem. Soc.*, 2024, **146**, 8576–8584.
- 46 X. Chen, Y. Xia, Y. Wu, Y. Xu, X. Jia, R. N. Zare and F. Wang, *J. Am. Chem. Soc.*, 2024, **146**, 10868–10874.
- 47 A. Kumar, V. S. Avadhani, A. Nandy, S. Mondal, B. Pathak, V. K. N. Pavuluri, M. M. Avulapati and S. Banerjee, *Anal. Chem.*, 2024, **96**, 10515–10523.
- 48 C. J. Chen, V. S. Avadhani and E. R. Williams, *Angew. Chem., Int. Ed.*, 2025, **64**, e202424662.
- 49 Y.-G. Fang, X. Li, C. Yuan, X. Li, X. Yuan, D. Zhang, X. Zhang, C. Zhu and W.-H. Fang, *Angew. Chem., Int. Ed.*, 2025, **137**, e202417920.
- 50 J. Dong, J. Xu, Z.-D. Meng, Z.-A. Nan, W. Li, R. N. Zare, Z.-Q. Tian and F. R. Fan, *J. Am. Chem. Soc.*, 2025, **147**, 16060–16069.
- 51 J. Zhou, Q. Wang, G. Cheng, W. Shen, R. N. Zare and X. Sun, *J. Am. Chem. Soc.*, 2025, **147**, 10916–10924.
- 52 S. Wei, Q. Wan, S. Zhou, W. Nie and S. Chen, *J. Am. Chem. Soc.*, 2024, **146**, 32777–32784.
- 53 K. Li, W. You, Y. Zhu, W. Wang, L. Wang, Q. Ge, Y. Liu, X. Ruan, H. Cheng, M. A. Tahir and L. Zhang, *J. Hazard. Mater.*, 2025, **493**, 138311.
- 54 R. Zhang, Z. Zhang, X. Chen, J. Jiang, L. Hua, X. Jia, R. Bao and F. Wang, *J. Am. Chem. Soc.*, 2024, **146**, 8528–8535.
- 55 Y. Wang, J. Luo, Y.-G. Fang, Z.-A. Nan, X. Cui, T. Chen, X. Zeng, X. Wang, X. Song, J. Zhao, W. Li, C. Zeng, D. Chen, C. Zhu, Z. Wei, Z.-Q. Tian and F. R. Fan, *J. Am. Chem. Soc.*, 2025, **147**, 2756–2765.
- 56 M. A. Mehrgardi, M. Mofidfar, J. Li, C. F. Chamberlayne, S. R. Lynch and R. N. Zare, *Adv. Sci.*, 2024, **11**, 2406785.
- 57 J. Dong, J. Chen, W. Wang, Z. Wei, Z.-Q. Tian and F. R. Fan, *J. Am. Chem. Soc.*, 2024, **146**, 2227–2236.
- 58 X. Liu, X. Li, X. Wang, Y. Li, X. Hu, Z. Zhou, F. Zhang, Z. Zhang and Y. Chen, *J. Am. Chem. Soc.*, 2025, **147**, 3529–3538.
- 59 Q. Ge, Y. Liu, K. Li, L. Xie, X. Ruan, W. Wang, L. Wang, T. Wang, W. You and L. Zhang, *Angew. Chem., Int. Ed.*, 2023, **62**, e202304189.
- 60 Q. Jiang, D. Xia, X. Li, H. Zhang, R. Yin, H. Xie, H. Xie, J. Jiang, J. Chen and J. S. Francisco, *Angew. Chem., Int. Ed.*, 2025, **64**, e202421002.
- 61 W. Wang, Y. Liu, T. Wang, Q. Ge, K. Li, J. Liu, W. You, L. Wang, L. Xie, H. Fu, J. Chen and L. Zhang, *J. Am. Chem. Soc.*, 2024, **146**, 6580–6590.
- 62 D. Xia, H. Zhang, Y. Ju, H. Xie, L. Su, F. Ma, J. Jiang, J. Chen and J. S. Francisco, *J. Am. Chem. Soc.*, 2024, **146**, 11266–11271.
- 63 K.-H. Huang, K. Chen, N. M. Morato, T. C. Sams, E. T. Dziekonski and R. G. Cooks, *Chem. Sci.*, 2025, **16**, 7544–7550.
- 64 K. Li, K. Gong, J. Liu, L. Ohnoutek, J. Ao, Y. Liu, X. Chen, G. Xu, X. Ruan, H. Cheng, J. Han, G. Sui, M. Ji, V. K. Valev and L. Zhang, *Cell Rep. Phys. Sci.*, 2022, **3**, 100917.
- 65 K. Li, Q. Ge, Y. Liu, L. Wang, K. Gong, J. Liu, L. Xie, W. Wang, X. Ruan and L. Zhang, *Energy Environ. Sci.*, 2023, **16**, 1135–1145.
- 66 I. Nam, J. K. Lee, H. G. Nam and R. N. Zare, *Proc. Natl. Acad. Sci. U. S. A.*, 2017, **114**, 12396–12400.
- 67 I. Nam, H. G. Nam and R. N. Zare, *Proc. Natl. Acad. Sci. U. S. A.*, 2017, **115**, 36–40.
- 68 D. Xia, F. Zeng, W. Chen, H. Zhao, H. Xie, J. Chen and J. S. Francisco, *Proc. Natl. Acad. Sci. U. S. A.*, 2025, **122**, e2501323122.
- 69 D. T. Holden, N. M. Morato and R. G. Cooks, *Proc. Natl. Acad. Sci. U. S. A.*, 2022, **119**, e2212642119.
- 70 Y. Meng, Y. Xia, J. Xu and R. N. Zare, *Sci. Adv.*, 2025, **11**, eadt8979.
- 71 M. Piejko, J. E. Alfonso-Ramos, J. Moran and T. Stuyver, *ChemistryEurope*, 2025, **3**, e202400093.
- 72 W.-Z. Song, M. Zhang, H.-J. Qiu, C.-L. Li, T. Chen, L.-L. Jiang, M. Yu, S. Ramakrishna, Z.-L. Wang and Y.-Z. Long, *Water Res.*, 2022, **226**, 119242.
- 73 B. Chen, Y. Xia, R. He, H. Sang, W. Zhang, J. Li, L. Chen, P. Wang, S. Guo, Y. Yin, L. Hu, M. Song, Y. Liang, Y. Wang,



- G. Jiang and R. N. Zare, *Proc. Natl. Acad. Sci. U. S. A.*, 2022, **119**, e2209056119.
- 74 H. Gu, C. Gu, A. Du Toit, W. Yu, M. W. Chen, H. L. Struckman, J. R. Silva, Y. Dai and A. G. Ewing, *J. Am. Chem. Soc.*, 2025, **147**, 27020–27029.
- 75 M. W. Chen, X. Ren, X. Song, N. Qian, Y. Ma, W. Yu, L. Yang, W. Min, R. N. Zare and Y. Dai, *J. Am. Chem. Soc.*, 2025, **147**, 8267–8279.
- 76 L. Qiu, Z. Wei, H. Nie and R. G. Cooks, *ChemPlusChem*, 2021, **86**, 1362–1365.
- 77 H. Wei, E. P. Vejerano, W. Leng, Q. Huang, M. R. Willner, L. C. Marr and P. J. Vikesland, *Proc. Natl. Acad. Sci. U. S. A.*, 2018, **115**, 7272–7277.
- 78 A. Gray-Weale and J. K. Beattie, *Phys. Chem. Chem. Phys.*, 2009, **11**, 10994–11005.
- 79 K. Gong, J. Ao, K. Li, L. Liu, Y. Liu, G. Xu, T. Wang, H. Cheng, Z. Wang, X. Zhang, H. Wei, C. George, A. Mellouki, H. Herrmann, L. Wang, J. Chen, M. Ji, L. Zhang and J. S. Francisco, *Proc. Natl. Acad. Sci. U. S. A.*, 2023, **120**, e2219588120.
- 80 L. Shi, R. A. LaCour, N. Qian, J. P. Heindel, X. Lang, R. Zhao, T. Head-Gordon and W. Min, *Nature*, 2025, **640**, 87–93.
- 81 H. Xiong, J. K. Lee, R. N. Zare and W. Min, *J. Phys. Chem. B*, 2020, **124**, 9938–9944.
- 82 H. Xiong, J. K. Lee, R. N. Zare and W. Min, *J. Phys. Chem. Lett.*, 2020, **11**, 7423–7428.
- 83 C. F. Chamberlayne and R. N. Zare, *J. Chem. Phys.*, 2020, **152**, 184702.
- 84 H. Hao, I. Leven and T. Head-Gordon, *Nat. Commun.*, 2022, **13**, 280.
- 85 C. F. Chamberlayne and R. N. Zare, *J. Chem. Phys.*, 2022, **156**, 054705.
- 86 J. R. Cendagorta and T. Ichiye, *J. Phys. Chem. B*, 2015, **119**, 9114–9122.
- 87 M. Matsumoto and Y. Kataoka, *J. Chem. Phys.*, 1988, **88**, 3233–3245.
- 88 M. A. Wilson, A. Pohorille and L. R. Pratt, *J. Chem. Phys.*, 1988, **88**, 3281–3285.
- 89 S. M. Kathmann, I.-F. W. Kuo and C. J. Mundy, *J. Am. Chem. Soc.*, 2008, **130**, 16556–16561.
- 90 V. Buch, A. Milet, R. Vácha, P. Jungwirth and J. P. Devlin, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 7342–7347.
- 91 J. K. Beattie, A. M. Djerdjev and G. G. Warr, *Faraday Discuss.*, 2009, **141**, 31–39.
- 92 M. Takahashi, *J. Phys. Chem. B*, 2005, **109**, 21858–21864.
- 93 R. Zimmermann, S. Dukhin and C. Werner, *J. Phys. Chem. B*, 2001, **105**, 8544–8549.
- 94 J. K. Beattie and A. M. Djerdjev, *Angew. Chem., Int. Ed.*, 2004, **43**, 3568–3571.
- 95 H. Mishra, S. Enami, R. J. Nielsen, L. A. Stewart, M. R. Hoffmann, W. A. Goddard III and A. J. Colussi, *Proc. Natl. Acad. Sci. U. S. A.*, 2012, **109**, 18679–18683.
- 96 K. G. Marinova, R. G. Alargova, N. D. Denkov, O. D. Velev, D. N. Petsev, I. B. Ivanov and R. P. Borwankar, *Langmuir*, 1996, **12**, 2045–2051.
- 97 R. Vácha, S. W. Rick, P. Jungwirth, A. G. de Beer, H. B. de Aguiar, J.-S. Samson and S. Roke, *J. Am. Chem. Soc.*, 2011, **133**, 10204–10210.
- 98 N. Agmon, H. J. Bakker, R. K. Campen, R. H. Henchman, P. Pohl, S. Roke, M. Thämer and A. Hassanali, *Chem. Rev.*, 2016, **116**, 7642–7672.
- 99 W. Xu, H. Zheng, Y. Liu, X. Zhou, C. Zhang, Y. Song, X. Deng, M. Leung, Z. Yang, R. X. Xu, Z. L. Wang, X. C. Zeng and Z. Wang, *Nature*, 2020, **578**, 392–396.
- 100 P. Bista, A. D. Ratschow, H.-J. Butt and S. A. Weber, *J. Phys. Chem. Lett.*, 2023, **14**, 11110–11116.
- 101 C. Ye, D. Liu, Y. Gao, F. Liu, H. Xu, T. Jiang and Z. L. Wang, *Matter*, 2025, **8**, 102007.
- 102 Z. Zhou, X. Yan, Y.-H. Lai and R. N. Zare, *J. Phys. Chem. Lett.*, 2018, **9**, 2928–2932.
- 103 R. F. Steiner, *Topics in fluorescence spectroscopy: Principles*, Springer, 1991, pp. 1–52.
- 104 X. Zhou, Y. Ji, Z. Ni, J. G. Lopez, K. Peneva, S. Jiang, N. Knorr, R. Berger, K. Koynov and H.-J. Butt, *Adv. Mater.*, 2025, **37**, 2420263.
- 105 S. D. Fried and S. G. Boxer, *Acc. Chem. Res.*, 2015, **48**, 998–1006.
- 106 N. Qian, H. Xiong, L. Wei, L. Shi and W. Min, *Annu. Rev. Phys. Chem.*, 2025, **76**, 279–301.
- 107 L. Shi, F. Hu and W. Min, *Nat. Commun.*, 2019, **10**, 4764.
- 108 Y. Liu, Q. Ge, T. Wang, R. Zhang, K. Li, K. Gong, L. Xie, W. Wang, L. Wang, W. You, X. Ruan, Z. Shi, J. Han, R. Wang, J. Chen, C. K. Chan and L. Zhang, *Chem*, 2024, **10**, 330–351.
- 109 A. M. Kol, T. Herriman, W. Salmon, G. DiLabio and M. Jackson, *ChemRxiv*, 2025, preprint, DOI: [10.26434/chemrxiv-2025-dh30c-v2](https://doi.org/10.26434/chemrxiv-2025-dh30c-v2).
- 110 L. M. Loew and L. L. Simpson, *Biophys. J.*, 1981, **34**, 353–365.
- 111 S. G. Boxer, *J. Phys. Chem. B*, 2009, **113**, 2972–2983.
- 112 A. Montenegro, C. Dutta, M. Mammetskuliev, H. Shi, B. Hou, D. Bhattacharyya, B. Zhao, S. B. Cronin and A. V. Benderskii, *Nature*, 2021, **594**, 62–65.
- 113 Q. Du, E. Freysz and Y. R. Shen, *Phys. Rev. Lett.*, 1994, **72**, 238–241.
- 114 Y. Zhang, H. B. de Aguiar, J. T. Hynes and D. Laage, *J. Phys. Chem. Lett.*, 2020, **11**, 624–631.
- 115 J. G. Davis, K. P. Gierszal, P. Wang and D. Ben-Amotz, *Nature*, 2012, **491**, 582–585.
- 116 X. Wu, W. Lu, L. M. Streacker, H. S. Ashbaugh and D. Ben-Amotz, *Angew. Chem., Int. Ed.*, 2018, **57**, 15133–15137.
- 117 D. Ben-Amotz, *J. Am. Chem. Soc.*, 2019, **141**, 10569–10580.
- 118 P. N. Perera, K. R. Fega, C. Lawrence, E. J. Sundstrom, J. Tomlinson-Phillips and D. Ben-Amotz, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 12230–12234.
- 119 K. M. Hunter, F. A. Shakib and F. Paesani, *J. Phys. Chem. B*, 2018, **122**, 10754–10761.
- 120 R. A. LaCour, J. P. Heindel and T. Head-Gordon, *J. Phys. Chem. Lett.*, 2023, **14**, 11742–11749.
- 121 A. P. Willard and D. Chandler, *J. Phys. Chem. B*, 2010, **114**, 1954–1958.



- 122 Q. Du, E. Freysz and Y. R. Shen, *Science*, 1994, **264**, 826–828.
- 123 L. F. Scatena, M. G. Brown and G. L. Richmond, *Science*, 2001, **292**, 908–912.
- 124 J. D. Smith, R. J. Saykally and P. L. Geissler, *J. Am. Chem. Soc.*, 2007, **129**, 13847–13856.
- 125 Y. R. Shen, *Nature*, 1989, **337**, 519–525.
- 126 A. G. Lambert, P. B. Davies and D. J. Neivandt, *Appl. Spectrosc. Rev.*, 2005, **40**, 103–145.
- 127 S. Nihonyanagi, J. A. Mondal, S. Yamaguchi and T. Tahara, *Annu. Rev. Phys. Chem.*, 2013, **64**, 579–603.
- 128 Y. R. Shen, *Annu. Rev. Phys. Chem.*, 2013, **64**, 129–150.
- 129 S. Strazdaite, J. Versluis and H. J. Bakker, *J. Chem. Phys.*, 2015, **143**, 084708.
- 130 C. S. Tian and Y. R. Shen, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 15148–15153.
- 131 S. Kulik, S. Pullanchery and S. Roke, *J. Phys. Chem. C*, 2020, **124**, 23078–23085.
- 132 S. Pullanchery, S. Kulik, B. Rehl, A. Hassanali and S. Roke, *Science*, 2021, **374**, 1366–1370.
- 133 E. Poli, K. H. Jong and A. Hassanali, *Nat. Commun.*, 2020, **11**, 901.
- 134 R. Zhao, H. Shen, R. A. LaCour, J. P. Heindel, M. Head-Gordon and T. Head-Gordon, *Angew. Chem., Int. Ed.*, 2025, **64**, e202508145.
- 135 W. Gan, W. Wu, F. Yang, D. Hu, H. Fang, Z. Lan and Q. Yuan, *Soft Matter*, 2017, **13**, 7962–7968.
- 136 T. L. Tarbuck, S. T. Ota and G. L. Richmond, *J. Am. Chem. Soc.*, 2006, **128**, 14519–14527.
- 137 C. Tian, N. Ji, G. A. Waychunas and Y. R. Shen, *J. Am. Chem. Soc.*, 2008, **130**, 13033–13039.
- 138 L. M. Levering, M. R. Sierra-Hernández and H. C. Allen, *J. Phys. Chem. C*, 2007, **111**, 8814–8826.
- 139 P. B. Petersen and R. J. Saykally, *Chem. Phys. Lett.*, 2008, **458**, 255–261.
- 140 K.-Y. Chiang, T. Seki, C.-C. Yu, T. Ohto, J. Hunger, M. Bonn and Y. Nagata, *Proc. Natl. Acad. Sci. U. S. A.*, 2022, **119**, e2204156119.
- 141 M. Dinpajoo and D. V. Matyushov, *J. Chem. Phys.*, 2016, **145**, 014504.
- 142 T. Preočanin, A. Selmani, P. Lindqvist-Reis, F. Heberling, N. Kallay and J. Lützenkirchen, *Colloids Surf., A*, 2012, **412**, 120–128.
- 143 Y. B. Vogel, C. W. Evans, M. Belotti, L. Xu, I. C. Russell, L.-J. Yu, A. K. Fung, N. S. Hill, N. Darwish, V. R. Gonçalves, M. L. Coote, K. S. Iyer and S. Ciampi, *Nat. Commun.*, 2020, **11**, 6323.
- 144 Y. Wang, H. Luo, X. R. Advincula, Z. Zhao, A. Esfandiari, D. Wu, K. D. Fong, L. Gao, A. S. Hazrah, T. Taniguchi, C. Schran, Y. Nagata, L. Bocquet, M.-L. Bocquet, Y. Jiang, A. Michaelides and M. Bonn, *J. Am. Chem. Soc.*, 2025, **147**, 30107–30116.
- 145 R. Buchner, G. T. Hefter and P. M. May, *J. Phys. Chem. A*, 1999, **103**, 1–9.
- 146 R. Buchner, G. Hefter, P. M. May and P. Sipos, *J. Phys. Chem. B*, 1999, **103**, 11186–11190.
- 147 T. Megyes, S. Bálint, T. Grósz, T. Radnai, I. Bakó and P. Sipos, *J. Chem. Phys.*, 2008, **128**, 044501.
- 148 M. E. Tuckerman, D. Marx and M. Parrinello, *Nature*, 2002, **417**, 925–929.
- 149 B. Chen, J. M. Park, I. Ivanov, G. Tabacchi, M. L. Klein and M. Parrinello, *J. Am. Chem. Soc.*, 2002, **124**, 8534–8535.
- 150 C. D. Cappa, J. D. Smith, B. M. Messer, R. C. Cohen and R. J. Saykally, *J. Phys. Chem. A*, 2007, **111**, 4776–4785.
- 151 H. C. Hamaker, *Physica*, 1937, **4**, 1058–1072.
- 152 A. M. Djerdjev and J. K. Beattie, *Molecules*, 2025, **30**, 785.
- 153 P. Creux, J. Lachaise, A. Graciaa, J. K. Beattie and A. M. Djerdjev, *J. Phys. Chem. B*, 2009, **113**, 14146–14150.
- 154 P. B. Petersen, J. C. Johnson, K. P. Knutsen and R. J. Saykally, *Chem. Phys. Lett.*, 2004, **397**, 46–50.
- 155 G. Jones and W. A. Ray, *J. Am. Chem. Soc.*, 1935, **57**, 957–958.
- 156 P. B. Petersen and R. J. Saykally, *J. Am. Chem. Soc.*, 2005, **127**, 15446–15452.
- 157 S. D. Fried and S. G. Boxer, *Annu. Rev. Biochem.*, 2017, **86**, 387–415.
- 158 S. D. Fried, S. Bagchi and S. G. Boxer, *Science*, 2014, **346**, 1510–1514.
- 159 S. H. Schneider and S. G. Boxer, *J. Phys. Chem. B*, 2016, **120**, 9672–9684.
- 160 A. I. Kloss, *Dokl. Akad. Nauk SSSR*, 1988, **303**, 1403–1407.
- 161 J. P. Heindel, H. Hao, R. A. LaCour and T. Head-Gordon, *J. Phys. Chem. Lett.*, 2022, **13**, 10035–10041.
- 162 M. T. Martins-Costa and M. F. Ruiz-López, *J. Am. Chem. Soc.*, 2023, **145**, 1400–1406.
- 163 K. Lee, M. A. Mehrgardi and R. N. Zare, *J. Am. Chem. Soc.*, 2025, **147**, 33240–33247.
- 164 D. Ben-Amotz, *Science*, 2022, **376**, 800–801.
- 165 L. Qiu and R. G. Cooks, *Angew. Chem., Int. Ed.*, 2024, **136**, e202400118.
- 166 L. S. McCarty and G. M. Whitesides, *Angew. Chem., Int. Ed.*, 2008, **47**, 2188–2207.
- 167 F. Galembeck, L. P. Santos, T. A. Burgo and A. Galembeck, *Chem. Soc. Rev.*, 2024, **53**, 2578–2602.
- 168 Y. Dai, Z.-G. Wang and R. N. Zare, *Nat. Chem. Biol.*, 2024, **20**, 1420–1433.
- 169 J. P. Heindel, R. A. LaCour and T. Head-Gordon, *Nat. Commun.*, 2024, **15**, 3670.
- 170 K. D. Judd, S. W. Parsons, T. Majumder and J. M. Dawlaty, *Chem. Rev.*, 2025, **125**, 2440–2473.
- 171 S. W. Devlin, F. Bernal, E. J. Riffe, K. R. Wilson and R. J. Saykally, *Faraday Discuss.*, 2024, **249**, 9–37.
- 172 G. Gonella, E. H. G. Backus, Y. Nagata, D. J. Bonthuis, P. Loche, A. Schlaich, R. R. Netz, A. Kühnle, I. T. McCrum, M. T. M. Koper, M. Wolf, B. Winter, G. Meijer, R. K. Campen and M. Bonn, *Nat. Rev. Chem.*, 2021, **5**, 466–485.
- 173 Z. L. Wang and A. C. Wang, *Mater. Today*, 2019, **30**, 34–51.
- 174 W. Sung, K. Inoue, S. Nihonyanagi and T. Tahara, *Nat. Commun.*, 2024, **15**, 1258.
- 175 D. Ojha, N. K. Kaliannan and T. D. Kühne, *Commun. Chem.*, 2019, **2**, 116.
- 176 D. Ojha and T. D. Kühne, *Sci. Rep.*, 2021, **11**, 2456.



- 177 S. D. Pino, D. Banerjee, M. Monti, G. D. Miron, G. Cassone and A. Hassanali, *arXiv*, 2025, preprint, DOI: [10.48550/arXiv.2506.23988](https://doi.org/10.48550/arXiv.2506.23988).
- 178 R. Vácha, D. Horinek, M. L. Berkowitz and P. Jungwirth, *Phys. Chem. Chem. Phys.*, 2008, **10**, 4975–4980.
- 179 Y.-L. S. Tse, C. Chen, G. E. Lindberg, R. Kumar and G. A. Voth, *J. Am. Chem. Soc.*, 2015, **137**, 12610–12616.
- 180 P. Zhang and X. Xu, *Langmuir*, 2025, **41**, 3675–3683.
- 181 P. Zhang, M. Feng and X. Xu, *ACS Phys. Chem. Au*, 2024, **4**, 336–346.
- 182 Z. Chen, Y. Zhang, P. Lv, T. Wu, J. He, J. Du, Q. Sun, Q. Wu, J. Yang, Y. Zhang, Y. Zhang, F. He, C. Cui, G. Hong, H. Zhu, Y. Li, J. Guo and X. Deng, *Nat. Nanotechnol.*, 2025, 1–9.

