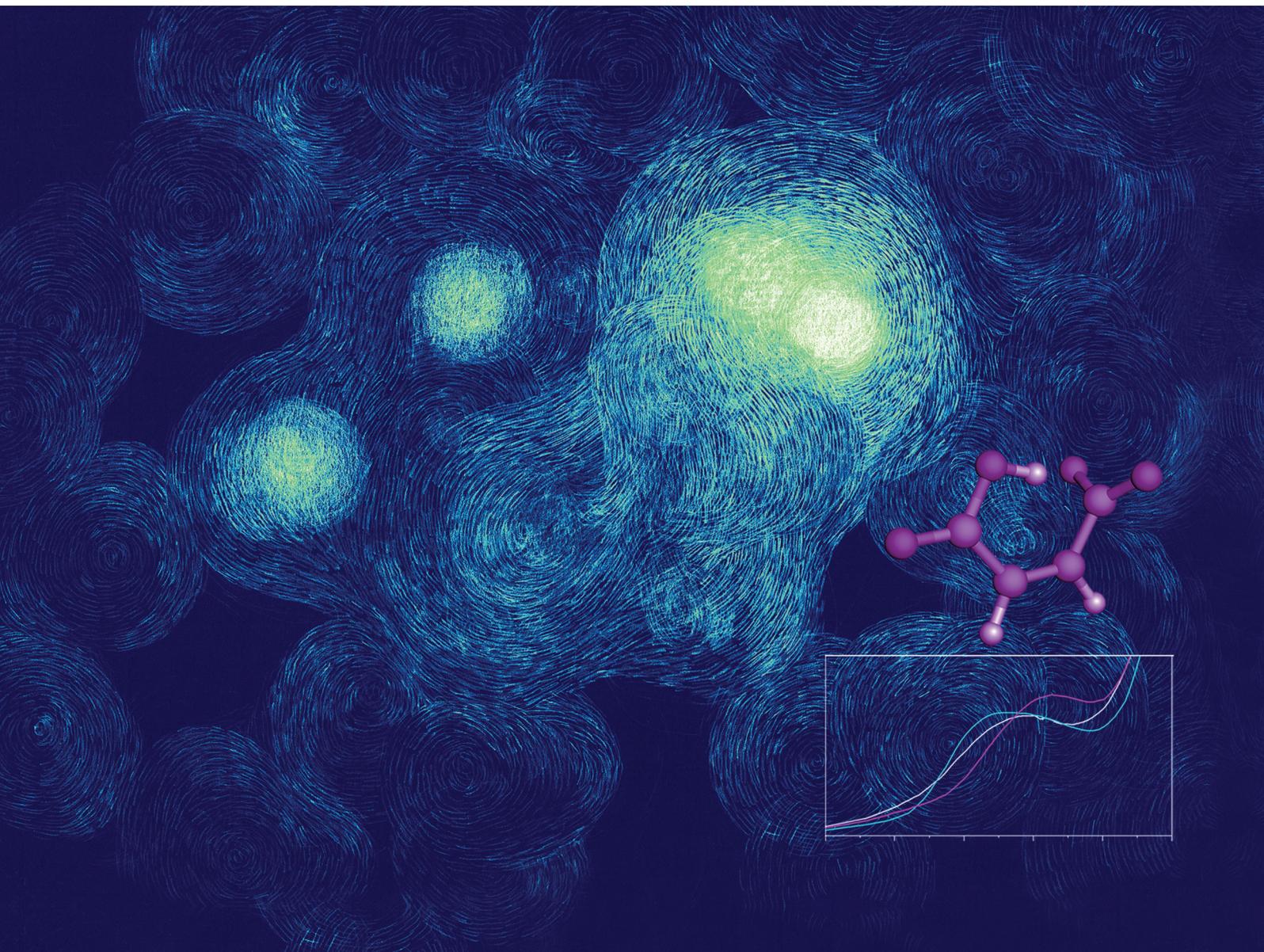


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**PAPER**

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Controlling the pH of aqueous succinic and maleic acids  
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# Controlling the pH of aqueous succinic and maleic acids analyzed by X-ray absorption spectroscopy

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X-ray absorption spectroscopy (XAS) measurements were performed at the oxygen K-edge using aqueous solutions of succinic and maleic acids. Molecular dynamics simulations were employed to perform structural sampling and calculate the theoretical spectra using the framework of the density functional theory. When the pH was varied, the first peak in the XAS profiles exhibited different behaviors for succinic and maleic acids. This peak behavior was reproduced using theoretical calculations. Maleic acid, which has a double bond between its central carbon atoms, exhibits resonance effects between its two carboxyl groups. Because this resonance effect was not observed for succinic acid, it was considered to account for the different peak shifts. Hydrogen bonding analyses of the water molecules surrounding succinic and maleic acids were performed to elucidate the structure of the water network.

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## Introduction

Since the beginning of the 21st century, structural research based on the molecular theory of liquids under ambient conditions has progressed considerably.<sup>1</sup> The development of technologies such as liquid jets<sup>2,3</sup> and liquid flow cells<sup>4–7</sup> has made it possible to quantify the X-ray absorption spectroscopy (XAS) profiles of liquid samples. To date, XAS studies have been conducted on liquid samples such as water,<sup>2,8</sup> alcohols,<sup>9</sup> and aqueous solutions of organic acids.<sup>10</sup>

A typical example of a study in aqueous solutions using soft X-rays is acetic acid. This molecule has two oxygen atoms in the carboxyl group, C=O and OH, which have different chemical environments and are observed in a site-selective manner.<sup>10</sup> In addition, Horikawa *et al.*<sup>11,12</sup> quantified the XAS and X-ray emission spectroscopy (XES) profiles of aqueous acetic acid solutions at various pH levels and reported their pH dependence.

The molar fractions of the neutral and anionic species quantitatively analyzed from the XES profiles were consistent with the Henderson–Hasselbalch equation. For oxalic acid, which is a dicarboxylic acid, both XAS and site-selective XES of its aqueous solution were conducted, and the respective profiles were quantified. By combining experimental findings with theoretical calculations, a resonance effect between the two carboxyl groups, which was not observed in monocarboxylic acids, was demonstrated.<sup>13</sup> The pH control of aqueous solutions of organic molecules has also been studied for amino acids; for example, Alías-Rodríguez *et al.*<sup>14</sup> confirmed that three protonation states dependent on the pH could be distinguished for proline. Using XPS and XAS, Greenspoon *et al.*<sup>15</sup> also showed that changes in the pH affect the surface composition of ammonium sulfate aerosols. Horikawa *et al.*<sup>16</sup> applied XAS to aqueous glycine as a biomolecular prototype.

Theoretical calculations based on the electronic structure theory are also powerful tools for studying solutions. Reinholdt *et al.*<sup>17</sup> investigated the coupled cluster modeling of nitrogen K-edge XAS for aqueous ammonia and ammonium based on quantum mechanics/molecular dynamics (MD) and compared their results with published experimental data and transition potential density functional theory (TP-DFT)-based simulations. Odellius *et al.*<sup>18</sup> conducted a theoretical and experimental study on the same system and showed symmetry breaking around the ammonia molecules in an aqueous solution. In addition, they simulated spectra using both  $\Delta$ SCF (where orbitals were optimized for the final state) and second-order algebraic diagrammatic

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construction [ADC(2)] (which uses molecular orbitals optimized for the ground state), and interpreted the observed solvatochromism by comparing the XAS profiles in the gas phase and in aqueous solutions.<sup>19</sup>

In this study, we focused on succinic and maleic acids as dicarboxylic acids. Dicarboxylic acids with two carboxyl groups play important roles in the metabolic pathways of living organisms. Succinic acid is among the compounds that makes up the citric acid cycle and is oxidized by succinate dehydrogenase to form fumarate. Maleic acid has the same empirical formula as fumaric acid, but refers to the *cis*-isomer. Maleic acid ( $pK_a = 1.92, 6.23$ ) and succinic acid ( $pK_a = 4.21, 5.64$ )<sup>20</sup> are four-carbon dicarboxylic acids. Maleic acid contains a carbon-carbon double bond, whereas succinic acid does not. Maleic acid has several conformational isomers owing to the rotation of its carboxyl groups and the orientation of the hydrogen atoms on its OH groups, whereas succinic acid has additional conformational isomers owing to the rotation around the C2–C3 axis. It would be interesting to observe the spectral features upon changing the pH and bond order. Furthermore, owing to intra- and intermolecular hydrogen bonding, succinic and maleic acids adopt various conformations. Theoretical analysis is also important to elucidate the features of these compounds at the molecular level.

Previous studies using IR spectroscopy<sup>21,22</sup> and NMR<sup>23,24</sup> have provided insights into the solvation structures and hydrogen bonding environments of these molecules. Conformational studies of dicarboxylic acids have been performed using IR and Raman spectroscopy by Suzuki and coworkers<sup>25</sup> and theoretically.<sup>26,27</sup> However, direct experimental information on how differences in hydrogen bonding motifs are reflected in the electronic states of oxygen atoms remains limited. In the present study, we employ soft XAS at the oxygen K-edge to investigate aqueous solutions of succinic and maleic acids, with the aim of elucidating how intra- and intermolecular hydrogen bonding influence their electronic structures.

## Methods

### Experimental methods

Aqueous 0.3 M succinic acid solutions at pH levels of 2.4, 5.0, and 12.5 was prepared by mixing two reagents [namely succinic acid (99.5% purity) and sodium hydroxide (97% purity), provided by Nacalai Tesque] with water. A 0.2 M maleic acid solution at pH levels of 1.4, 4.2, and 11.7 were prepared by mixing two reagents provided by Nacalai Tesque, maleic anhydride (99% purity) and sodium hydroxide (97% purity), with water.

XAS profiles were recorded using the soft X-ray beamline BL17SU at the SPring-8 facility.<sup>28,29</sup> All spectra were quantified using a liquid flow cell with a 150 nm-thick Au-coated thin Si<sub>3</sub>N<sub>4</sub> window to separate the liquid flowing at atmospheric pressure from the high-vacuum region.<sup>5,30</sup> The energy resolution of the XAS profiles was 0.1 eV. The background consisted of oxygen impurities in the Si<sub>3</sub>N<sub>4</sub> window.

### Theoretical methods

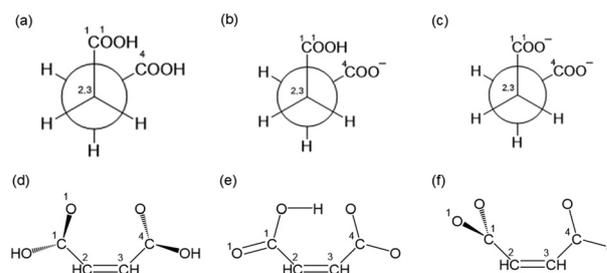
Maleic acid has several conformational isomers owing to the rotation of the carboxyl group and the orientation of the H atom of the OH group. The chemical structures of succinic and maleic acids are shown in Scheme 1. Succinic acid and its rotation around the C2–C3 axis should be considered. Conformational searches for succinic and maleic acids were performed using the CONFLEX<sup>31</sup> program for neutral, anionic, and dianionic acids. Each conformation was structurally optimized using the molecular orbital program package Gaussian 09<sup>32</sup> based on the Møller–Plesset second-order perturbation theory and cc-pVTZ basis sets.<sup>33</sup> The polarizable continuum model (PCM)<sup>34–36</sup> was also used to account for solvent effects.

MD simulations were performed using the simulation program GROMACS.<sup>37</sup> The aqueous solution model of maleic acid was created by randomly placing nine maleic acid molecules and 2500 water molecules in a box (0.2 M). The aqueous solution model of succinic acid was prepared by randomly adding 14 molecules of succinate and 2500 molecules of water to a box (0.3 M). Sodium ions were added to the anionic and dianionic acid solutions to achieve a total charge of zero. The water model used was the TIP4P force field.<sup>38</sup>

The bond and angular force constants of succinic and maleic acids were determined for the most stable succinic and maleic acid conformations. The lists of force constants are compiled in the SI.

We first performed energy minimization, followed by 20 ps simulations in the NVT ensemble (*i.e.*, the number of particles  $N$ , volume  $V$ , and temperature  $T$  of the system were kept constant). Short-range interactions were corrected using the Verlet algorithm<sup>39</sup> with a cutoff distance of 0.9 nm. Velocity-rescaled temperature coupling was used.<sup>40</sup> Long-range electrostatics were corrected using the particle mesh Ewald (PME) algorithm<sup>41,42</sup> during equilibration. The subsequent simulations were followed by 10 ns simulations in the NPT ensemble (*i.e.*, the number of particles  $N$ , pressure  $P$ , and temperature  $T$  of the system were kept constant) with a time step of 0.1 fs, pressure of 1 bar, and a temperature of 300 K.

From several snapshots of the MD simulations, 90 clusters consisting of one succinic and one maleic acid molecule and surrounding 30 water molecules were sampled. XAS calculations were performed using the density functional theory implemented with the deMon2k code.<sup>43</sup> The standard Perdew–Burke–Ernzerhof



**Scheme 1** Chemical structures of succinic (a)–(c) and maleic (d)–(f) acids.



gradient-corrected exchange–correlation functional<sup>14,45</sup> was used. In addition, the IGRO-III basis set<sup>46</sup> was used to describe core-excited oxygen atoms. Triple-zeta valence plus polarization basis sets were used for hydrogen atoms. The effective core potential was used for the carbon and non-core-excited oxygen atoms. The obtained XAS line spectra were convolved using a Gaussian function (full width at half maximum, FWHM: 1.2 eV at 534 eV and 5.0 eV at 547 eV for maleic acid, 1.0 eV at 535 eV and 5.0 eV at 547 eV for succinic acid) and averaged.

## Results and discussion

### XAS profiles of succinic and maleic acids

Fig. 1 shows the XAS profile of succinic acid. The insets show a broad energy range (528–548 eV). The first peaks assigned as O(1s) to  $\pi^*(\text{C}=\text{O})$  of succinic acid are observed at 532.4, 532.6, and 532.6 eV corresponding to pH levels of 2.4, 5.0, and 12.5. Note that there is no water absorption in this energy region, as also previously observed for acetic acid<sup>10</sup> and oxalic acid.<sup>13</sup> The positions and intensities of the first peaks increase with increasing pH. The theoretical X-ray absorption spectra are shown in Fig. 1(b). The theoretical spectra reproduce the experimental trend well; the first peaks are at 532.3, 532.3, and 533.4 eV for the neutral, anionic, and dianionic forms, respectively.

Fig. 2 shows XAS profile of maleic acid. The insets show a broad energy range (528–548 eV). The assignment of the first peak is the same as that of succinic acid. In contrast to succinic acid, the order of the first peaks for different pH levels is different, *i.e.*, the peaks are at 532.8, 532.6, and 533.3 eV corresponding to pH levels of 1.4, 4.2, and 11.7. The neutral, anionic, and dianionic forms, respectively, are theoretically reproduced at 532.9, 532.1, and 533.3 eV.

### Conformation analysis of succinic and maleic acids

To elucidate the spectral features of succinic and maleic acids, conformational analyses were performed. Succinic and maleic acids have multiple conformations. To systematically explore

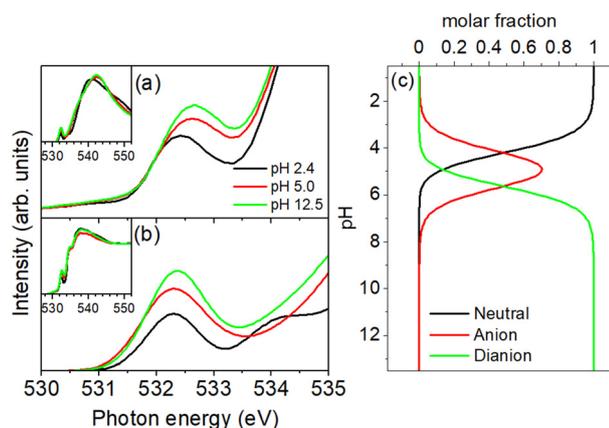


Fig. 1 (a) Experimental and (b) theoretical spectra of succinic acid obtained using XAS (black: pH = 2.4, red: pH = 5.0, and green: pH = 12.5). (c) Molar fractions for neutral, anionic, and dianionic species.

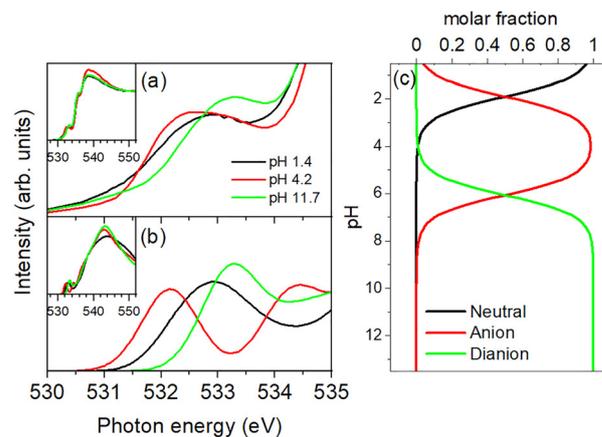


Fig. 2 (a) Experimental and (b) theoretical spectra of maleic acid obtained using XAS (black: pH = 1.4, red: pH = 4.2, green: pH = 11.7). (c) Molar fractions for neutral, anionic, and dianionic species.

all the conformers, a conformational search for a single molecule was performed using CONFLEX. Eight neutral maleic acids, six anionic maleic acids, three dianionic maleic acids, 14 neutral succinic acids, 11 anionic succinic acids, and three dianionic succinic acids were obtained. The complete results are presented in the SI. Geometry optimization was performed for the isolated molecule using the PCM. The most stable conformations are shown in Fig. 3 and are the same as those for the isolated molecule and PCM, except for dianionic succinic acid. For neutral and anionic maleic acids, the most stable conformer was planar owing to intramolecular hydrogen bonding. In dianionic maleic acid, the most stable conformation is nonplanar owing to the repulsion between the negative charges of the oxygen atoms, and the two carboxylic groups are orthogonal to each other. Succinic acid has both *gauche* and anti-conformations; however, in the PCM, the *gauche* conformation is the most stable.

Fig. 3 shows the conformations of the succinic and maleic acids used in the MD simulations. The most stable conformational isomers of dianionic succinic acid differ between a single molecular model and PCM, with the linear (unfolded) conformation being more stable in the single molecular model and the folded conformation being more stable in the PCM. Wanko *et al.*<sup>47</sup> showed that the dianion of succinic acid adopts a folded

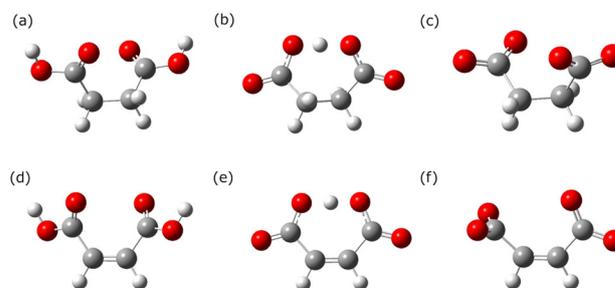


Fig. 3 Most stable molecular structures of succinic and maleic acids. The following text was used for atom labeling. (a) and (d) Neutral, (b), (e) anionic, and (c) and (f) dianionic forms.



conformation in aqueous solutions. Neutral maleic acid was the most stable in the gas phase and the PCM, with intramolecular hydrogen-bonded cyclic structures. However, Hyttinen *et al.*<sup>48</sup> suggested that neutral maleic acid in water forms hydrogen bonds with the surrounding water rather than intramolecular hydrogen bonds. The single molecule and PCM calculations do not fully account for interactions with the surrounding water molecules. Therefore, for neutral maleic acid, we adopted a conformational isomer without intramolecular hydrogen bonds.

In the XAS profile of oxalic acid, the resonance effect between the two COOH functional groups was observed as a high-energy shift in the peak.<sup>13</sup> This resonance effect was observed when the two COOH groups of oxalic acid were parallel; when the COOH groups were twisted, the  $\pi$  resonance was destroyed, and this did not occur. To elucidate this effect explicitly, the XAS profile of a single molecule was obtained by rotating the carboxyl group of neutral maleic acid by  $10^\circ$  with respect to the molecular plane. The spectral changes observed in this case are shown in Fig. 4(a). The angle shown in the figure corresponds to the C3–C2–C1–O1 dihedral angle. The first peak, which is assigned as  $O(1s) \rightarrow \pi^*(C=O)$ , blue-shifts, and its intensity increases as the dihedral angle increases. The peak position is at 531.1 eV when the dihedral angle is  $0^\circ$ , but it is at 531.9 eV at  $90^\circ$ . The  $\pi^*$  orbitals of the excited destination of neutral maleic acid when the dihedral angles of the carboxylic groups are  $\phi = 0^\circ, 30^\circ, 60^\circ,$  and  $90^\circ$  are shown in Fig. 4(b). When the dihedral angle is  $0^\circ$ , all atoms are on the same plane, and  $\pi$  resonance between the two carboxylic groups is observed. As the dihedral angle increases, the  $\pi$  resonance begins to break, and completely disappears at  $\phi = 90^\circ$ . Yamamura *et al.*<sup>13</sup> showed that it is possible to adjust the  $\pi$  resonance of oxalic acid. The same mechanism was observed in the case of maleic acid.

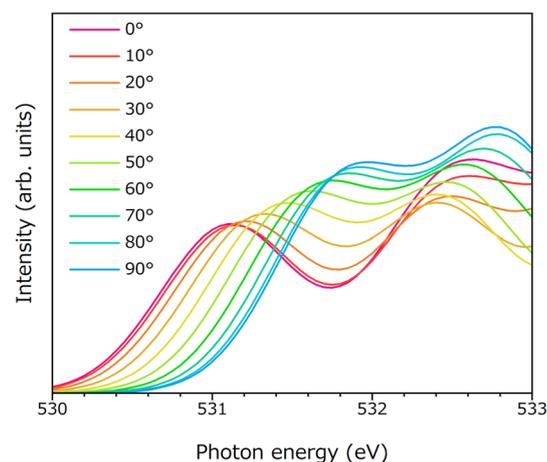


Fig. 4 (upper) Dihedral angle C3–C2–C1–O1 dependence of XAS profiles for neutral maleic acid from  $0^\circ$  to  $90^\circ$ . (lower) HOMOs at  $0^\circ, 30^\circ, 60^\circ,$  and  $90^\circ$ .

The  $\pi^*$  orbitals of succinic and maleic acids are shown in Fig. 5. Succinic acid has four C atoms that are not in the same plane, and its dihedral angle is approximately  $60^\circ$ . Therefore,  $\pi$ -orbital delocalization through the four C atoms does not occur. In contrast, maleic acid has a dihedral angle of  $0^\circ$  and consists of four C atoms owing to its double bond. In anionic maleic acid, the two carboxyl groups are parallel to a plane consisting of four C atoms, thus preserving resonance, whereas in neutral and dianionic maleic acids, the carboxyl groups are not parallel to the molecular plane and have a twisted structure. This twisting breaks the  $\pi$  resonance and blue-shifts the first peak in the XAS profiles. The peculiar peak behavior of the aqueous maleic acid solutions in the XAS profile was caused by the C atoms in the same plane and the different dihedral angles of the carboxylic groups for each valence.

To understand the behavior at the interface between the organic substances and water, it is interesting to investigate the hydrogen bonding behavior of the water surrounding the organic substances. Based on the MD simulation results of the aqueous solutions of succinic and maleic acids, hydrogen bond analysis was conducted. The results are shown in Fig. 6.

The difference between the structures of succinic and maleic acids lies in the bond distance between the C2–C3 atoms and the number of hydrogen atoms attached to the carbons. These differences had only a minor effect on the hydrogen-bonded network of the surrounding water molecules, with the effect being more prominent for maleic acid. This was due to the difference in the C2–C3 bond length. In the first hydration shell, tetra-coordinated water decreased, and bi-coordinated and tri-coordinated water increased, indicating the disruption of the hydrogen-bond network of water caused by succinic and maleic acids. In the first and second hydration shells, which were closer to the solute, the structural differences were greater compared with those in liquid water, suggesting that the surrounding water structure was disturbed by the presence of these acids. In the third hydration shell, the effect of the solute decreased, and the number of hydrogen bonds became closer to that in liquid water. As the valence increased from neutral to monovalent to divalent, the difference from liquid water increased because the water molecules were more strongly attracted to the solute.

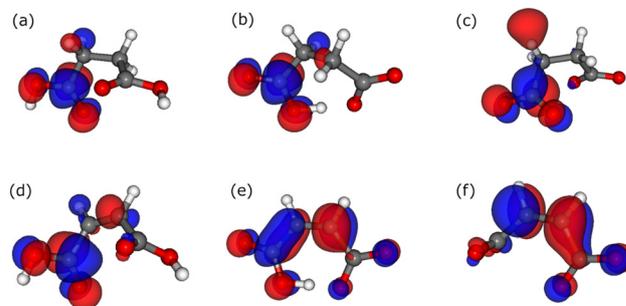


Fig. 5 HOMOs for succinic acid in the (a) neutral, (b) anionic, and (c) dianionic forms, and HOMOs for maleic acid in the (d) neutral, (e) anionic, and (f) dianionic forms.



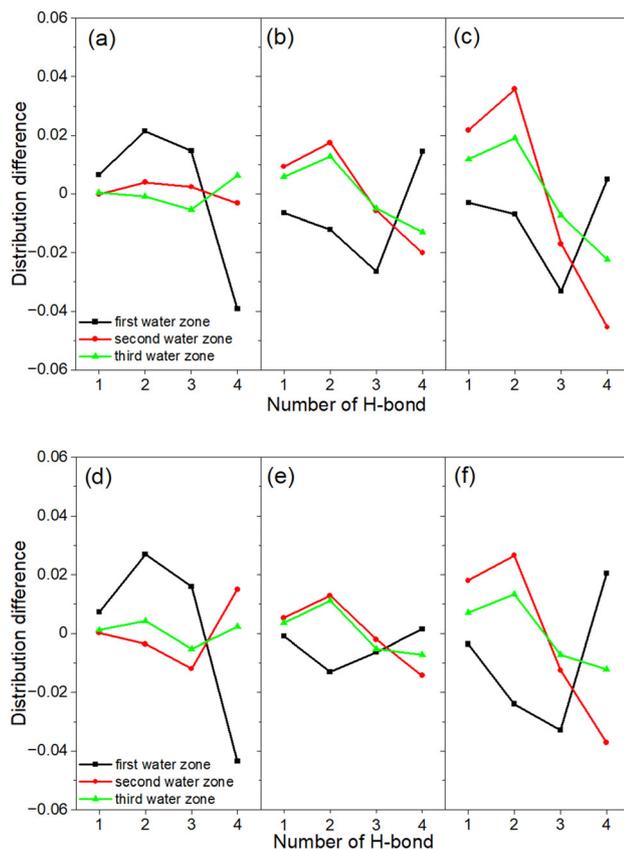


Fig. 6 Distribution difference from bulk water as a function of the number of hydrogen bonds. The black, red, and blue lines indicate the first, second, and third hydration zones, respectively. (a) Neutral, (b) anionic, and (c) dianionic succinic acid forms. (d) Neutral, (e) anionic, and (f) dianionic maleic acid forms.

### Conformations of succinic and maleic acids controlling pH

Dicarboxylic acids can exist in three different states depending on the pH adjustment. In the case of succinic acid, there is no resonance between the carboxyl groups at either end, and each functional group acts independently because the central C2–C3 bond can rotate freely. In such case, succinic acid behaves as if there were two acetic acid molecules. Horikawa *et al.*<sup>11,13</sup> showed that, in acetic acid, as the pH increased, the first peak gradually increased. The behaviors of dicarboxylic acids with longer carbon chains, such as glutaric and adipic acids, can also be predicted.

Although maleic acid has the same number of carbon atoms as succinic acid, the behavior of its peaks differs from that of succinic acid. The first peak of neutral maleic acid was larger and shifted to a higher energy compared with the others. This is because the two carboxyl groups cannot lie in the same plane owing to the electrostatic repulsion between the functional groups, even though resonance exists *via* the double bond between the two carboxyl groups at either end. When the pH is increased and maleic acid becomes a monovalent anion, intramolecular hydrogen bonding allows the two carboxyl groups to remain in the same plane. Therefore, the first peak shifted to a lower energy compared with that of the neutral

case. When the pH is further increased and the carboxyl group becomes a divalent anion, the two carboxyl groups twist considerably, as in the case of oxalic acid, shifting the peak to a higher energy.

## Summary

In this study, we investigated the structures of succinic and maleic acids in aqueous solutions using XAS. Although succinic and maleic acids have the same number of carbon atoms, their XAS behaviors differ in response to pH. This can be explained based on the electrostatic repulsion between the two carboxyl groups and the presence or absence of resonant structures mediated by double bonds. By combining XAS measurements with theoretical calculations, we were able to discuss the rich chemical information present in the aqueous solutions in detail.

While NMR and IR provide averaged information, XAS/XES captures the local electronic state obtained by core-excitation from oxygen 1s electron site-selectively as an extremely fast process occurring on the femtosecond scale. Observing how the concept of resonance actually behaves in the structurally fluctuating environment of an aqueous solution is not straightforward, but soft X-ray spectroscopy has made it possible for the first time. The methodology used in this study can be applied to other organic compounds in aqueous solutions that undergo structural changes depending on the pH, and is expected to be useful for future research on the solution chemistry of aqueous systems.

## Author contributions

OT devised the project and prepared the beamtime proposal. TT, YH, and MO designed the experimental setup. RY, TS, TT, and OT performed the experiments and analyzed the data under guidance of TT and YH. RY, RO, RA and OT performed theoretical analysis. RO and OT wrote the original draft. OT is a corresponding author.

## Conflicts of interest

There are no conflicts to declare.

## Data availability

The authors confirm that the data supporting the findings of this study are available within the article and/or its supplementary information (SI). Supplementary information: Tables S1–S20. See DOI: <https://doi.org/10.1039/d6cp00520a>.

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## Notes and references

- J. W. Smith and R. J. Saykally, *Chem. Rev.*, 2017, **117**, 13909–13934.
- K. R. Wilson, B. S. Rude, T. Catalano, R. D. Schaller, J. G. Tobin, D. T. Co and R. J. Saykally, *J. Phys. Chem. B*, 2001, **105**, 3346–3349.
- M. Ekimova, W. Quevedo, M. Faubel, P. Wernet and E. T. J. Nibbering, *Struct. Dyn.*, 2015, **2**, 054301.
- M. Freiwald, S. Cramm, W. Eberhardt and S. Eisebitt, *J. Electron Spectrosc. Relat. Phenom.*, 2004, **137–140**, 413–416.
- T. Tokushima, Y. Harada, H. Ohashi, Y. Senba and S. Shin, *Rev. Sci. Instrum.*, 2006, **77**, 063107.
- M. Nagasaka, T. Hatsui, T. Horigome, Y. Hamamura and N. Kosugi, *J. Electron Spectrosc. Relat. Phenom.*, 2010, **177**, 130–134.
- Y. Harada, M. Kobayashi, H. Niwa, Y. Senba, H. Ohashi, T. Tokushima, Y. Horikawa, S. Shin and M. Oshima, *Rev. Sci. Instrum.*, 2012, **83**, 013116.
- P. Wernet, D. Nordlund, U. Bergmann, M. Cavalleri, M. Odelius, H. Ogasawara, L. A. Naslund, T. K. Hirsch, L. Ojamae, P. Glatzel, L. G. M. Pettersson and A. Nilsson, *Science*, 2004, **304**, 995–999.
- K. R. Wilson, M. Cavalleri, B. S. Rude, R. D. Schaller, T. Catalano, A. Nilsson, R. J. Saykally and L. G. M. Pettersson, *J. Phys. Chem. B*, 2005, **109**, 10194–10203.
- T. Tokushima, Y. Horikawa, Y. Harada, O. Takahashi, A. Hiraya and S. Shin, *Phys. Chem. Chem. Phys.*, 2009, **11**, 1679–1682.
- Y. Horikawa, T. Tokushima, Y. Harada, O. Takahashi, A. Chainani, Y. Senba, H. Ohashi, A. Hiraya and S. Shin, *Phys. Chem. Chem. Phys.*, 2009, **11**, 8676–8679.
- Y. Horikawa, T. Tokushima, A. Hiraya and S. Shin, *Phys. Chem. Chem. Phys.*, 2010, **12**, 9165–9168.
- R. Yamamura, T. Suenaga, M. Oura, T. Tokushima and O. Takahashi, *Chem. Phys. Lett.*, 2020, **738**, 136895.
- M. Alias-Rodríguez, S. Bonfrate, W. Park, N. Ferré, C. H. Choi and M. Huix-Rotllant, *J. Phys. Chem. A*, 2023, **127**, 10382–10392.
- E. H. Greenspoon, S. Warkander, P. Kim, A. Jana, J. Qian, K. R. Wilson, M. Ahmed and J. B. Bergner, *J. Chem. Phys.*, 2025, **162**, 234307.
- Y. Horikawa, T. Tokushima, O. Takahashi, Y. Harada, A. Hiraya and S. Shin, *Phys. Chem. Chem. Phys.*, 2018, **20**, 23214–23221.
- P. Reinholdt, M. L. Vidal, J. Kongsted, M. Iannuzzi, S. Coriani and M. Odelius, *J. Phys. Chem. Lett.*, 2021, **12**, 8865–8871.
- M. Odelius, S. D. Folkestad, T. Saisopa, Y. Rattanachai, W. Sailuam, H. Yuzawa, N. Kosugi, A. C. Paul, H. Koch and D. Céolin, *J. Phys. Chem. Lett.*, 2025, **16**, 3411–3419.
- S. Tsuru and M. Nagasaka, *J. Phys. Chem. A*, 2025, **129**, 3020–3031.
- R. D. Lide, *CRC Handbook of Chemistry and Physics*, CRC, Taylor and Francis, Boca Raton, FL, 87th edn, 2007.
- M. a J. Ayora-Cañada and B. Lendl, *Vib. Spectrosc.*, 2000, **24**, 297–306.
- J. T. Hołaj-Krzak, N. Rekik, N. A. M. Alsaif and G. Lakshminarayana, *J. Phys. Chem. A*, 2022, **126**, 5604–5620.
- J. D. Roberts, *Acc. Chem. Res.*, 2006, **39**, 889–896.
- J. Guo, P. M. Tolstoy, B. Koeppe, G. S. Denisov and H.-H. Limbach, *J. Phys. Chem. A*, 2011, **115**, 9828–9836.
- M. Suzuki and T. Shimanouchi, *J. Mol. Spectrosc.*, 1968, **28**, 394–410.
- P. Tarakeshwar and S. Manogaran, *THEOCHEM*, 1996, **362**, 77–99.
- T. H. Nguyen, D. E. Hibbs and S. T. Howard, *J. Comput. Chem.*, 2005, **26**, 1233–1241.
- H. Ohashi, Y. Senba, H. Kishimoto, T. Miura, E. Ishiguro, T. Takeuchi, M. Oura, K. Shirasawa, T. Tanaka, M. Takeuchi, K. Takeshita, S. Goto, S. Takahashi, H. Aoyagi, M. Sano, Y. Furukawa, T. Ohata, T. Matsushita, Y. Ishizawa, S. Taniguchi, Y. Asano, Y. Harada, T. Tokushima, K. Horiba, H. Kitamura, T. Ishikawa and S. Shin, *AIP Conf. Proc.*, 2007, **879**, 523–526.
- Y. Senba, H. Ohashi, H. Kishimoto, T. Miura, S. Goto, S. Shin, T. Shintake and T. Ishikawa, *AIP Conf. Proc.*, 2007, **879**, 718–721.
- T. Tokushima, Y. Harada, O. Takahashi, Y. Senba, H. Ohashi, L. G. M. Pettersson, A. Nilsson and S. Shin, *Chem. Phys. Lett.*, 2008, **460**, 387–400.
- H. Goto, S. Obata, N. Nakayama and K. Ohta, CONFLEX Corporation, Tokyo, Japan, 2012.
- M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. J. A. Montgomery, J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, O. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski and D. J. Fox, Gaussian, Inc., Wallingford CT, 2010.
- J. T. H. Dunning, *J. Chem. Phys.*, 1989, **90**, 1007–1023.



- 34 B. Mennucci, E. Cancès and J. Tomasi, *J. Phys. Chem. B*, 1997, **101**, 10506–10517.
- 35 M. Cossi, V. Barone, B. Mennucci and J. Tomasi, *Chem. Phys. Lett.*, 1998, **286**, 253–260.
- 36 E. Cancès, B. Mennucci and J. Tomasi, *J. Chem. Phys.*, 1997, **107**, 3032–3041.
- 37 M. J. Abraham, T. Murtola, R. Schulz, S. Páll, J. C. Smith, B. Hess and E. Lindahl, *SoftwareX*, 2015, **1–2**, 19–25.
- 38 H. W. Horn, W. C. Swope, J. W. Pitera, J. D. Madura, T. J. Dick, G. L. Hura and T. Head-Gordon, *J. Chem. Phys.*, 2004, **120**, 9665–9678.
- 39 L. Verlet, *Phys. Rev.*, 1967, **159**, 98–103.
- 40 G. Bussi, D. Donadio and M. Parrinello, *J. Chem. Phys.*, 2007, **126**, 614101.
- 41 T. Darden, D. York and L. Pedersen, *J. Chem. Phys.*, 1993, **98**, 10089–10092.
- 42 U. Essmann, L. Perera, M. L. Berkowitz, T. Darden, H. Lee and L. G. Pedersen, *J. Chem. Phys.*, 1995, **103**, 8577–8593.
- 43 A. M. Köster, G. Geudtner, A. Alvarez-Ibarra, P. Calaminici, M. E. Casida, J. Carmona-Espindola, V. D. Domínguez-Soria, R. Flores-Moreno, G. U. Gamboa, A. Goursot, T. Heine, A. Ipatov, A. de la Lande, F. Janetzko, J. M. del Campo, D. Mejia-Rodriguez, J. U. Reveles, J. Vasquez-Perez, A. Vela, B. Zúñiga-Gutierrez and D. R. Salahub, *deMon2k, version 5*, The deMon developers, 2018.
- 44 J. P. Perdew, K. Burke and M. Ernzerhof, *Phys. Rev. Lett.*, 1996, **77**, 3865–3868.
- 45 J. P. Perdew, K. Burke and M. Ernzerhof, *Phys. Rev. Lett.*, 1997, **78**, 1396.
- 46 W. Kutzelnigg, U. Fleischer and M. Schindler, *NMR-Basic Principles and Progress*, Springer-Verlag, Heidelberg, 1990.
- 47 M. Wanko, T. Wende, M. M. Saralegui, L. Jiang, A. Rubio and K. R. Asmis, *Phys. Chem. Chem. Phys.*, 2013, **15**, 20463–20472.
- 48 N. Hyttinen and N. L. Prisle, *J. Phys. Chem. A*, 2020, **124**, 4801–4812.

