







Characterization of strongly hyperfine-split protons by DNP

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Dynamic nuclear polarization experiments use microwave irradiation to transfer the larger electron polarization to nuclear spins of interest, and thus enhance the NMR transitions above thermal equilibrium. How the polarization transfer from the electron spin to the nuclear spins in such experiments proceeds and which nuclear spins close to an unpaired electron get polarized and contribute through spin diffusion to the observable bulk nuclear magnetization are not fully understood. We address these questions by combining reverse DNP and band-selective inversion pulses on nuclear spins. We report the electron-detected NMR spectrum of proton spins involved in the direct DNP process in Ox063 trityl samples with protonated and deuterated solvents and variable radical concentrations. We also determine the spin-diffusion barrier surrounding the trityl radical and find that proton spin diffusion is quenched for hyperfine couplings exceeding ~ 250 kHz. This corresponds to a radius of the spin diffusion barrier in the range from 5.4 to 6.8 Å. Burning a hole into the NMR spectrum of proton spins involved in the direct DNP step reveals an electron–electron spin diffusion process imprinted on the proton spectrum. We explain this diffusion process using a three-spin system consisting of two electron spins and one proton, and quantify the electron spin diffusion rate constant.

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1 Introduction

Dynamic nuclear polarization (DNP) is a widely applied method to increase the sensitivity of nuclear magnetic resonance (NMR) spectroscopy by transferring the higher electron polarization to the nuclei of interest.^{1–3} The experiments are typically performed at cryogenic temperatures with a radical embedded in a frozen glassy matrix.^{4–9} Currently, two different DNP implementations are used in practice: (i) magic-angle spinning (MAS) DNP operating typically at around 100 K with the aim to investigate materials, surfaces, and biomolecules.^{2,10–12} (ii) Dissolution DNP typically carried out around 1 K to generate highly polarized small molecules for metabolic imaging or spectroscopic applications.^{13–18} Both DNP techniques rely on an efficient spin diffusion process from nuclei (usually protons) near the unpaired electron spin to the bulk nuclear spin bath.^{19–21} The nearby nuclei are not directly observable *via* conventional NMR due to fast relaxation and/or the strong hyperfine coupling to the electron spin. Therefore, it is not well understood how the polarization is transferred from the nearby nuclei to the bulk and where these nuclei are located

spatially and spectrally.²² Recent studies have demonstrated that the spin diffusion barrier lies within less than 0.6 nm for a trityl radical in a glassy water/glycerol mixture.²³ Spins within this barrier do not participate in spin diffusion to the bulk due to the large hyperfine splitting.²⁴ The spectrum and spin diffusion of proton spins nearby an unpaired electron to bulk protons in TEMPOL embedded in a frozen glycerol–water mixture was demonstrated at 1.4 K using a dissolution DNP setup and conventional NMR bulk spin detection using a modified version of the CEST experiment.^{22,25} Another example is the nuclear–nuclear double resonance (NUDOR) experiment, which detects strongly coupled protons in different nitroxide biradicals using the cross effect under MAS at 14 T at around 100 K.²⁶ Although those are impressive results, the direct NMR detection of nearby spins requires a lot of signal averaging and long measurement times. In this work, we demonstrate the measurement of an electron-detected spectrum of protons near a trityl radical Ox063 dissolved in protonated and deuterated solvents at 80 K. Electron detection has much higher sensitivity and is thus much faster than a direct detection of the proton spectrum. In particular, the Ox063 trityl radical in a $\text{gly-d}_3\text{:D}_2\text{O:H}_2\text{O}$ (6:3:1 by volume) gives high enhancement factors in conventional DNP experiments as first described in ref. 9 and knowledge about this system is of high value. We also report a spectral and spatial diffusion barrier with spin diffusion of protons within the barrier to the bulk protons being

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quenched. Furthermore, we were able to quantify electron spin diffusion rate constants by burning a hole into the electron-detected proton spectrum and observing its time evolution.

2 Results and discussion

All pulse schemes (Fig. 1) used in this work are based on the concept of a reverse DNP contact as first described in ref. 27. We extend the concept by introducing pulses on the proton channel. For both DNP steps the NOVEL sequence²⁸ is used. One of the advantages of using NOVEL as a DNP contact compared to, e.g., the solid effect (SE)²⁹ or pulsed DNP sequences^{30–33} is that the resonance condition in the NOVEL sequence is satisfied at the center of the EPR line ($\Omega_{0,S} = 0$ MHz) and thus at the same spectral position as the two pulse Hahn echo detection. This reduces the risk of potential artifacts due to spectral diffusion. Moreover, NOVEL is one of the best performing DNP sequences with respect to the enhancement and available mw power, especially below 3 T. The NOVEL matching condition $|\nu_{1,S}| = |\nu_{0,I}|$ was determined by performing a single NOVEL DNP contact followed by a spin echo as described in ref. 27 and outlined in more detail Sections A.1.

and A.2. of the SI. All pulse sequences have in common that after the initial NOVEL DNP contact and a variable delay t_{del} the electron spins are saturated to facilitate the back transfer from the polarized proton spins to the electron spin. Only electron polarization that is initially transferred to the proton spin in the first DNP step is transferred back to the electron spin with the reverse DNP step. This is shown in Fig. S9 Section A.2. of the SI. Depending on the experimental scheme, the proton spin polarization during the delay t_{del} is manipulated using one or two band-selective Gauss pulses (red and blue pulses in Fig. 1). Using pulse parameters that correspond to a π -pulse for the blue Gauss pulse (detection pulse) and sweeping the carrier frequency of that pulse, an electron-detected proton spectrum can be recorded. After the reverse DNP step, the detection of the electron spin is done by a two-pulse Hahn echo sequence.³⁴

2.1 Electron-detected proton spectrum

To record the electron-detected proton spectrum, the pulse sequence shown in Fig. 1(A) is used. The band-selective variable-amplitude Gauss pulse on the proton channel was placed between the initial DNP contact and the reverse DNP block, i.e., in the middle of the delay t_{del} . The relative amplitude of the Gauss pulse with a pulse length of 10 μs was changed from 0 to 40, where a maximum output power of 300 W corresponds to a relative amplitude of 100. The amplitude nutation traces were recorded for carrier frequencies $\nu_{0,I}$ ranging from 14.14 MHz to 15.10 MHz. The Gauss function used for the Gauss pulse has a standard deviation $\sigma = 1.96 \mu\text{s}$. In Fig. 2 we show the result for 5 mM trityl in gly- d_8 : $D_2O:H_2O$ (6:3:1) for $t_{\text{del}} = 15 \mu\text{s}$ without electron decoupling (A) and with electron decoupling (B) for different amplitudes of the variable-amplitude Gauss pulse. We see from the amplitude nutation traces for different carrier frequencies $\nu_{0,I}$ that the first minimum is found at an amplitude $a = 25$ corresponding to ~ 100 kHz rf-field amplitude which agrees with the theoretical pulse profile as shown in Fig. S28 in Section B. of the SI. We clearly see in Fig. 2(A) that we are not able to invert the signal nor do we achieve a negative electron echo. The reason for this is that the proton spectrum is rather broad with a FWHM of ~ 650 kHz. This effect is also seen in numerical simulations as shown in Fig. S29 in Section B. of the SI.

The inversion efficiency increases by narrowing the proton spectrum through electron decoupling during the nutation pulse. With electron decoupling an almost complete inversion of the Hahn echo reaching a minimum signal intensity of about -0.8 in the center of the proton spectrum at $\nu_{0,I} = 14.89$ MHz. The center frequency is shifted by ~ 40 kHz compared to the thermal equilibrium proton signal from the bulk. The electron-detected proton spectrum is obtained by extracting the first minima of the amplitude nutation traces. This is shown in Fig. 2(A) and (B) on the bottom. Without electron-decoupling during the nutation pulse, we obtain an electron-detected proton spectrum with a full-width-half-maximum (FWHM) of ~ 650 kHz. Note that due to the limited tuning range of the proton circuit, only part of the undecoupled line could be detected. The proton line is by a factor of ~ 20 wider than for

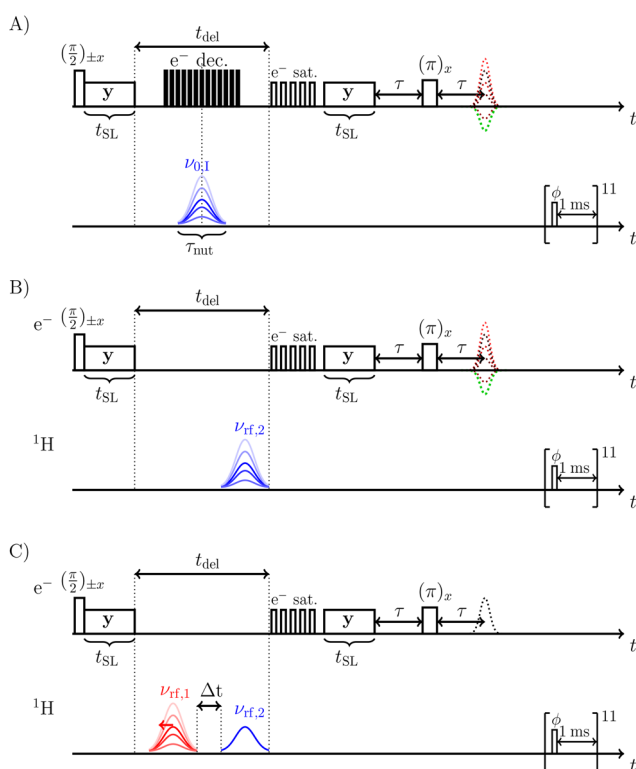


Fig. 1 Pulse sequences used in the realm of this work. (A) Is the pulse sequence to measure the electron-detected proton spectra, (B) the pulse sequence to monitor the spin diffusion towards the bulk and (C) to burn a hole into the proton spectrum. For all experiments a saturation train on the proton spins at the end of each sequence was applied with a carrier frequency equal to the blue Gauss pulses and $\phi = 100^\circ$. The carrier frequencies of the Gaussian pulses on the proton channel are indicated above the corresponding pulses. The dashed lines indicate the Hahn echo signal, which appear after the second τ delay.



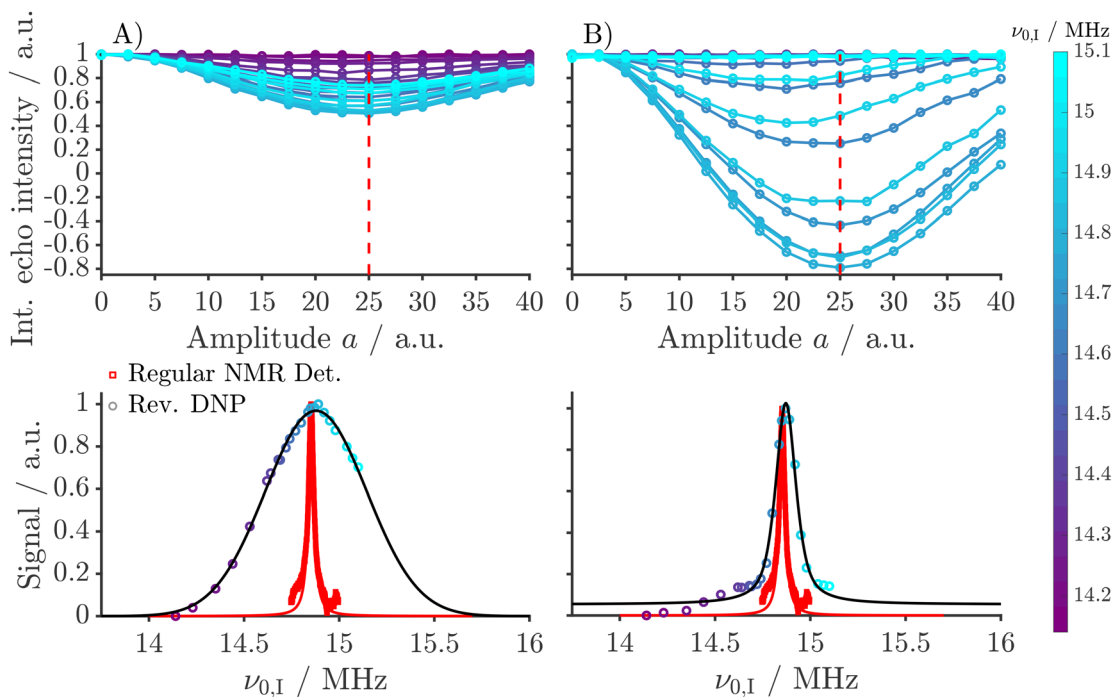


Fig. 2 Electron-detected proton spectrum for 5 mM trityl in gly- d_8 : D_2O : H_2O (6 : 3 : 1) using the pulse sequence of Fig. 1(A) with $t_{\text{del}} = 15 \mu\text{s}$ and $t_{\text{SL}} = 4000 \text{ ns}$. The variable-amplitude Gauss pulse was placed in the middle of the delay t_{del} . (A) Amplitude nutation traces upon application of a band-selective Gauss pulse of $10 \mu\text{s}$ length. The relative amplitude a of this Gauss pulse is varied from 0 to 40. The first minimum of the amplitude nutation trace is at a relative amplitude of 25 corresponding to a Rabi frequency of $\sim 100 \text{ kHz}$. The carrier frequency of the amplitude nutation trace was varied from 14.14 to 15.10 MHz throughout different experiments. The minima of the amplitude nutation traces were extracted and plotted against the frequency to obtain the electron-detected proton spectrum. Comparison with the thermal equilibrium signal of the bulk protons using regular NMR detection is shown in red. (B) Amplitude nutation traces upon application of the same band-selective Gauss pulse as in (A) with simultaneous electron decoupling during that pulse.

the thermal equilibrium proton signal from the bulk (see Fig. S38 in Section C.1. of the SI). Under electron decoupling during the nutation pulse, the line width of the proton spectrum is reduced by a factor of ~ 5 compared to the line width without electron decoupling, resulting in a FWHM of $\sim 130 \text{ kHz}$. This is still a factor of ~ 4 larger than the line width of the thermal equilibrium proton signal from the bulk. However, the FWHM of the electron-detected proton spectrum recorded with simultaneous electron decoupling during the pulse is equal to the FWHM of the theoretical inversion bandwidth of the variable-amplitude Gauss pulse, as shown in Fig. S28 in Section B. of the SI. The larger linewidth with electron decoupling as compared to direct NMR observation of bulk protons can thus be attributed to power broadening.

This motivated us to perform numerical simulations for a more quantitative understanding. For this, we assumed proton lines with FWHM that are equal or smaller than the FWHM of the pulse inversion band. If the proton line width is on the same order as the inversion pulse or even less, a slightly broader proton spectrum is obtained by extracting the minima of the amplitude nutation traces compared to the “true” proton spectrum (see Fig. S30 for a proton FWHM of 30 kHz and S31 for a proton FWHM of 130 kHz in the SI). In both cases, the FWHM of the proton spectrum obtained by the minima of the amplitude nutation traces has a FWHM of $\sim 130 \text{ kHz}$.

That such an approach with extracting the minima of the amplitude nutation traces is suitable to record a proton spectrum that is significantly broader than the bandwidth of the inversion pulse ($\sim 130 \text{ kHz}$) is shown in Fig. S32 in the SI. This shows that our method of indirect measurement of the proton line width works best for spectra that are significantly broader than the theoretical pulse inversion profile. Different types of band-selective rf pulses were tested (I-BURP,^{35,36} optimal control designed pulses^{37,38}). Those band-selective pulses have a narrower inversion profile than the $10 \mu\text{s}$ Gauss pulse, but are much longer (up to $100 \mu\text{s}$ or even more). We found that the long duration of such pulses induces substantial loss of signal.

Another reason for the relatively broad proton line observed under electron decoupling could be that the electron decoupling is not perfect as can be seen from the shoulder on the left hand side of the main peak in Fig. 2(B) and that the Hahn echo is only inverted up to 80%. The electron-decoupling consisted of 99 π -pulses of 12 ns length spaced by 80.2 ns with an FWHM of $\sim 66.4 \text{ MHz}$ of the theoretical inversion profile (see Fig. S36 in the SI). The EPR spectrum of trityl has an FWHM of $\sim 6.23 \text{ MHz}$. Hence, our pulses should in principle be sufficient to invert the entire EPR spectrum uniformly. Further improvements in the decoupling efficiency can be envisioned by using broadband chirp pulses²⁷ or broadband decoupling sequences as used in solution-state NMR.³⁹



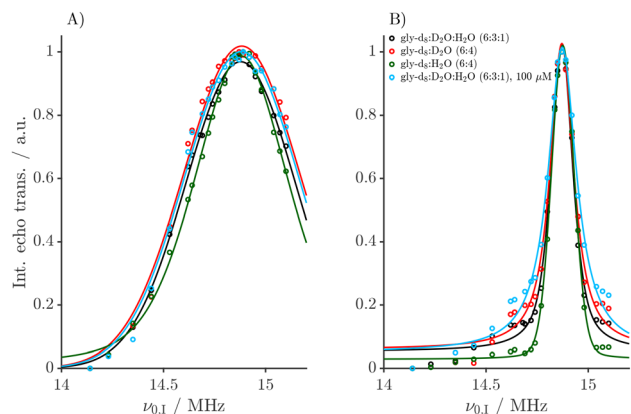


Fig. 3 Electron-detected proton spectrum for 5 mM trityl Ox063 in gly- d_8 : D_2O : H_2O (6:3:1) black, gly- d_8 : D_2O (6:4) red, gly- d_8 : H_2O (6:4) green and for 100 μ M trityl Ox063 in gly- d_8 : D_2O : H_2O (6:3:1) light-blue using the pulse sequence of Fig. 1(A). The variable-amplitude Gauss pulse was placed in the middle of the delay $t_{\text{del}} = 15 \mu\text{s}$ and $t_{\text{SL}} = 4000 \text{ ns}$. Panels (A) and (B) show the electron-detected proton spectra without and with electron decoupling, respectively. Within experimental error between the four different measurement sessions the electron-detected proton spectra are identical. Please note that the data in black were already shown in Fig. 2.

In Fig. 3 we compare the electron-detected proton spectra for samples with different protonation levels of the water, *i.e.*, 5 mM trityl in gly- d_8 : D_2O : H_2O (6:3:1), gly- d_8 : D_2O (6:4) and gly- d_8 : H_2O (6:4) and of different trityl concentration (100 μ M and 5 mM both in gly- d_8 : D_2O : H_2O (6:3:1)). In Fig. 3(A) we show the electron-detected proton spectra without electron decoupling and in Fig. 3(B) with electron decoupling for the four different sample compositions for a value of $t_{\text{del}} = 15 \mu\text{s}$. We can see that except for deviations within experimental uncertainty between the four samples, the electron-detected proton spectra are almost identical. For $t_{\text{del}} = 800 \mu\text{s}$ as shown in Fig. S37 in the SI the agreement within the four different data sets is even better. Here, we discuss the experiments for a short time $t_{\text{del}} = 15 \mu\text{s}$ to ensure that spin diffusion to the bulk is negligible (see Section 2.2). The observation that the electron-detected proton spectra of the four different samples are almost identical in shape and intensity lets us conclude that we detect protons that are located on the trityl molecule or on close-by solvent molecules rather than the bulk protons. The bulk proton concentration in the four samples is quite different, and we compared the intensity of the thermal equilibrium proton spectra with the theoretical number of protons in the samples (see Table S6 in the SI). The integrated intensities and the number of protons per sample are in good agreement. The calculation for the number of protons in the samples includes the purity of the deuterated molecules and the protons of the trityl molecule. This indicates that there was no contamination during preparation of the samples. Except for the unavoidable proton impurities in gly- d_8 ($\sim 2\%$ protons) and D_2O ($\sim 0.15\%$ protons) as used for the sample preparation of the gly- d_8 : D_2O (6:4) mixture, the amount of protons in the bulk of the matrix of gly- d_8 : H_2O (6:4) as compared to gly- d_8 : D_2O (6:4) is by a factor $\frac{100}{2.15} \sim 47$ larger. For such a large difference of numbers

in the bulk protons, we would expect a large difference in the electron-detected proton spectra if our method would detect a significant amount of bulk protons (see also thermal equilibrium spectra shown in Fig. S38 and S39 in the SI).

Another indication that our experimental scheme in Fig. 1(A) detects proton spins that are located on the trityl molecule itself or on close-by molecules is by comparing it to the ^1H MIMS ENDOR spectrum reported in Fig. 3 of ref. 23. The ^1H MIMS ENDOR spectrum of the d_{36} -Finland trityl that contains no protons directly bound to itself in gly- d_8 : D_2O : H_2O (6:3:1 by volume) shows a FWHM of $\sim 200 \text{ kHz}$. This is a factor of 3.25 smaller than our observed linewidth. Note that MIMS ENDOR has a blind spot behavior, which also influences the hyperfine couplings that are observable.⁴⁰ Our experimental scheme is not hampered by such a blind spot behavior.

2.2 Detection of proton spin diffusion towards the bulk

In a second step, we investigated the decay of proton polarization during the delay t_{del} (see Fig. 1(B)). This kind of experiment was already reported in ref. 27 for a different matrix composition and different trityl concentrations at Q-band (1.2 T field) without pulses on the proton channel. However, our goal here is to measure the decay of proton polarization as a function of the proton frequency $\nu_{\text{rf},2}$. Therefore, the Gauss pulse to record the frequency-selective proton spectrum was placed at the end of the delay t_{del} as shown in Fig. 1(B). To capture the full proton spectrum and monitor spin diffusion from those protons nearby an electron spin to the bulk protons, we omitted electron decoupling during the Gauss pulse. The delay t_{del} was increased from 15 μs to 3000 μs using a NOVEL contact time of $t_{\text{SL}} = 4000 \text{ ns}$ for both DNP contacts. The amplitude nutation traces were recorded for a range of frequencies from $\nu_{\text{rf},2} = 14.14$ to 15.1 MHz for 5 mM trityl in gly- d_8 : H_2O (6:4). Fig. 4(A) shows the measurement for $\nu_{\text{rf},2} = 14.77 \text{ MHz}$ and (B) for $\nu_{\text{rf},2} = 14.89 \text{ MHz}$. The reconstructed proton spectra for different delays t_{del} are shown in Fig. 4(C). They were obtained by extracting the minima of the amplitude nutation traces at an amplitude $a_2 = 25$, which is indicated by the green dashed lines in Fig. 4(A) and (B). One can clearly see that the proton polarization decays and that this decay is more pronounced in the center of the spectra, *i.e.*, in the range of $14.64 \text{ MHz} \leq \nu_{\text{rf},2} \leq 15.07 \text{ MHz}$. Outside this range no significant decay of the proton polarization can be observed on this time scale. Comparing this observation with the thermal bulk proton spectrum as shown in Fig. S38 in the SI with a center frequency $\sim \nu_{0,1} = 14.853 \text{ MHz}$ and a FWHM = 0.054 MHz, we attribute this loss of magnetization to spin diffusion to the bulk protons rather than a decay originating from longitudinal relaxation of the protons (bulk proton relaxation times $T_{1,\text{H}}$ are on the order of seconds for all samples). Outside this range spin diffusion to the bulk protons is much slower due to the large difference in the proton frequency between the bulk and the protons nearby the electron spin. For a shift in frequency of $14.853 \text{ MHz} - 14.6 \text{ MHz} = 0.253 \text{ MHz}$ the electron-proton distance can be calculated to define a spatial diffusion barrier. For an angle θ between the external magnetic field vector \vec{B} and the electron-proton



distance vector \vec{r}_{SI} we find for $\theta = 0^\circ$ $|\vec{r}_{\text{SI}}| = 6.8 \text{ \AA}$ and for $\theta = 90^\circ$ $|\vec{r}_{\text{SI}}| = 5.4 \text{ \AA}$. This is in very good agreement with the diffusion barrier of $< 6 \text{ \AA}$ as reported by Tan *et al.*²³ The spin diffusion to the bulk protons is faster at the center of the proton line ($\nu_{\text{rf},2} = 14.89 \text{ MHz}$) than at the edge of the active range ($\nu_{\text{rf},2} = 14.64 \text{ MHz}$ or $\nu_{\text{rf},2} = 15.07 \text{ MHz}$) as can be seen in Fig. 4(D). In the absence of any nutation pulses ($a_2 = 0$), an average decay is measured due to the missing proton frequency selectivity.

A simple fit to a mono-exponential or stretched exponential decay as shown in Fig. 4(D) did not give satisfactory or stable results. A longer time scale would be required to unambiguously characterize the decay constant of the spin diffusion process. In reality a sum of mono-exponential or stretched exponential functions might describe the spin diffusion accurately, similar to ref. 20. Also the electron spin polarization plays a role in the nuclear spin diffusion.²¹ It was found experimentally and explained using Lindblads Master equation in ref. 21 that the lower the electron spin polarization the faster the nuclear spin diffusion. A detailed investigation of the spin diffusion for different matrix compositions and electron concentrations is beyond the scope of this article, but would be an interesting research topic. In the context of this work the aim is to quantify the spectral range in the proton spectrum where spin diffusion to the bulk takes place. In Fig. S48 in the SI we show the experimental data for a NOVEL contact time t_{SL} of 800 ns. The protons nearby the electron spin are thus less

polarized, and the observed spin diffusion towards the bulk is slower. This indicates that a significant amount of the observed loss in polarization for increasing t_{del} can be attributed to spin diffusion towards the bulk rather than to spin relaxation of the nuclei. The latter is expected to be independent of the polarization level of the nearby protons. With our experimental data as shown in Fig. 4 we observe a spectral region ranging from $\sim 14.64 \text{ MHz}$ to 15.07 MHz in which spin diffusion to bulk protons takes place. Outside this spectral region spin diffusion to the bulk is quenched due to the strong HFI.

2.3 Electron-spin diffusion imprinted on the proton spectrum

In a final experiment (Fig. 1(C)) the pulse sequence has an additional band-selective Gauss pulse to burn a hole into the electron-detected proton spectrum. The first pulse drawn in red is the hole burning pulse with a variable amplitude a_1 to ensure optimal hole-burning efficiency at a fixed carrier frequency $\nu_{\text{rf},1}$. The blue pulse is the detection pulse to record the proton spectrum and has an amplitude a_2 and a carrier frequency $\nu_{\text{rf},2}$. To maintain optimal detection excitation and sensitivity, the NMR coil of the resonator is tuned and matched with respect to the carrier frequency of the detection pulse. The amplitude a_2 of this pulse is kept constant during the experiment. The carrier frequency $\nu_{\text{rf},2}$ of the detection pulse is systematically varied across the proton spectrum from 14.14 MHz to 15.10 MHz . Tuning and matching was always on resonance with respect to

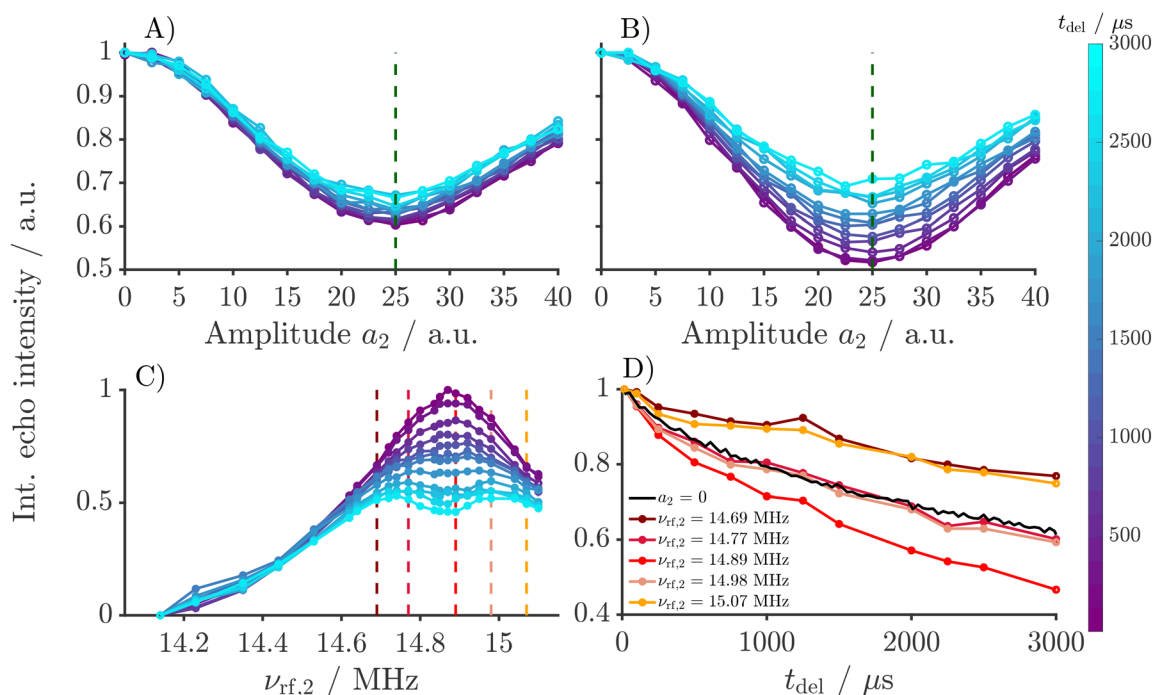


Fig. 4 Measurement of spin diffusion from protons nearby an electron spin to bulk protons using the pulse sequence of Fig. 1(B). (A) and (B) Recorded amplitude nutation traces for t_{del} ranging from $15 \mu\text{s}$ to $3000 \mu\text{s}$ by varying the amplitude a_2 of the Gauss pulse. The sample was 5 mM trityl in gly- d_8 : H_2O (6 : 4) and $t_{\text{SL}} = 4000 \text{ ns}$. In (A) the carrier frequency of the Gauss pulse was $\nu_{\text{rf},2} = 14.77 \text{ MHz}$ and in (B) 14.89 MHz , with the latter corresponding to the center of the proton spectrum. The data points to plot the proton spectra were extracted along the green dashed line. (C) Proton spectra for different delays t_{del} . The spin diffusion towards the bulk is more pronounced in the center of the spectrum. The reddish dashed lines indicate the spectral positions where the data points for the traces in (D) were extracted. (D) Diffusion traces extracted from (C) for different frequencies $\nu_{\text{rf},2}$. The diffusion to the bulk is fastest for $\nu_{\text{rf},2} = 14.89 \text{ MHz}$ corresponding to the center of the spectrum and gets slower moving away from the center. The black solid line indicates a measurement in the absence of any Gauss pulse *i.e.* $a_2 = 0$.



the detection pulse to maintain the same detection sensitivity across the proton spectrum. Here we discuss the experiments where the hole was burned at $\nu_{\text{rf},1} = 14.69$ MHz and $\nu_{\text{rf},1} = 14.89$ MHz. Experiments with a hole burned at $\nu_{\text{rf},1} = 15.04$ MHz can be found in the SI, Fig. S50 Section E.1. At large differences between $\nu_{\text{rf},1}$ and $\nu_{\text{rf},2}$, the amplitude of the hole burning pulse would no longer be the same as the one of the second pulse. To account for that, the amplitude a_1 of the hole-burning pulse is incremented during the experiments. In Fig. S53 Section E.2. in the SI we show such an experimental amplitude trace for variation of a_1 for different frequencies $\nu_{\text{rf},2}$ and Δt for the sample 5 mM trityl in gly-d₈:D₂O:H₂O (6:3:1) and $\nu_{\text{rf},1} = 14.69$ MHz. The amplitude traces of all other samples and frequencies $\nu_{\text{rf},1}$ can be found in Sections E.2.–E.6. of the SI. To observe time-dependent changes in the spectra, the delay between the two Gauss pulses Δt was incremented independently. All details about the data analysis and processing of the experimental data can be found in Section 4.

Fig. 5 shows the proton spectra as a function of Δt for a hole burning frequency of $\nu_{\text{rf},1} = 14.69$ MHz for four different samples. In all spectra, it can be observed that the hole burning was successful and that the hole is slowly filled for larger values of Δt . We observe an intensity loss symmetrically shifted around the center of the proton spectrum at $\nu_{\text{rf},2} \sim 14.89$ MHz. The hole in the proton spectrum has a width of ~ 320 kHz which matches the width of the theoretical inversion profile of ~ 360 kHz as shown in Fig. S1.

This diffusion process is observed in samples with 5 mM trityl concentration with different degrees of protonation as shown in Fig. 5(A)–(C). It is strongly reduced in the sample with a trityl concentration of 100 μM . This constitutes first evidence that the observed process includes a spin system consisting of two or more electron spins. The proton polarization transport process occurs on a time scale of ~ 100 μs (see Fig. 8 and Table 1), while the relaxation time of the electrons is $T_{1,e} \sim 2.5$ ms for all samples regardless of the trityl or proton concentration. The bulk proton

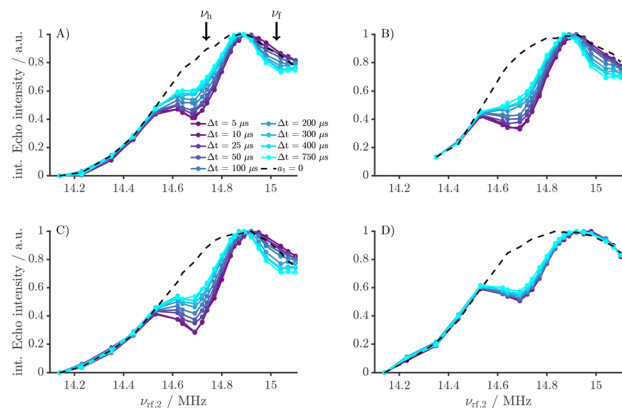


Fig. 5 Hole burning spectra for delays Δt between the two Gauss pulses varying from 5 to 750 μs , $t_{\text{del}} = 800$ μs and $t_{\text{SL}} = 4000$ ns. The hole was burned at $\nu_{\text{rf},1} = 14.69$ MHz. The black dashed lines are obtained by using the data points with $a_1 = 0$ i.e. in the absence of any hole burning pulse. The sample in subplot (A) is 5 mM trityl in gly-d₈:D₂O:H₂O (6:3:1), in subplot (B) 5 mM trityl in gly-d₈:D₂O (6:4), in subplot (C) 5 mM trityl in gly-d₈:H₂O (6:4) and in subplot (D) 100 μM trityl in gly-d₈:D₂O:H₂O (6:3:1).

Table 1 Comparison of the fit parameters from a mono-exponential decay as given in eqn (S.6) in the SI and shown in Fig. 8 for difference magnetization I_{Δ} for all four samples. The hole was burned at $\nu_{\text{rf},1} = 14.69$ MHz and $\nu_{\text{h}} = 14.69$ MHz, $\nu_{\text{r}} = 15.04$ MHz

Sample	$\Delta I/\text{a.u.}$	$\tau/\mu\text{s}$	$I_{\infty}/\text{a.u.}$
gly-d ₈ :D ₂ O:H ₂ O (6:3:1) 5 mM	0.361	168.499	0.132
gly-d ₈ :D ₂ O (6:4) 5 mM	0.356	172.961	0.142
gly-d ₈ :H ₂ O (6:4) 5 mM/4000 ns	0.347	155.591	0.148
gly-d ₈ :H ₂ O (6:4) 5 mM/800 ns	0.367	140.626	0.142
gly-d ₈ :D ₂ O:H ₂ O (6:3:1) 100 μM	0.129	530.830	0.367

relaxation times $T_{1,\text{H}}$ are on the order of seconds for all samples, and even protons near the radicals are unlikely to relax so fast.¹ The hole forming at $\nu_{\text{rf},2} \sim 15.04$ MHz broadens over time, which is another indication that the observed spin transport process is not caused by T_1 relaxation. The latter would only reduce the depth of the hole over time, but not influence its width. The process is symmetric for holes burned at $\nu_{\text{rf},1} = 15.04$ MHz and $\nu_{\text{rf},1} = 14.69$ MHz (see Fig. S50 Section E.1. in SI). For a hole burned at the center of the spectrum at $\sim \nu_{\text{rf},1} = 14.89$ MHz we do not observe any spectral transport process at all, as shown in Fig. 6.

These observations can be explained by a simple three-spin model consisting of two coupled electron spins and one proton spin. The proton spin needs to be hyperfine coupled to one of the electron spins. The Hamilton operator for such a spin system can be written in the rotating frame of the electron spins (S_1 and S_2) and the laboratory frame of the proton spin I as

$$\begin{aligned} \hat{\mathcal{H}} = & \Omega_{0,S_1} \hat{S}_{1z} + \Omega_{0,S_2} \hat{S}_{2z} + \omega_{0,I} \hat{I}_z \\ & + D^{(1,2)} \left(2\hat{S}_{1z} \hat{S}_{2z} - \frac{1}{2} (\hat{S}_1^+ \hat{S}_2^- + \hat{S}_1^- \hat{S}_2^+) \right) \quad (1) \\ & + A_{zz}^{(1,I)} \hat{S}_{1z} \hat{I}_z + B^{(1,I)} \hat{S}_{1z} \hat{I}_x, \end{aligned}$$

where $\Omega_{0,S_1} = \omega_{0,S_1} - \omega_{\text{mw}}$ and $\Omega_{0,S_2} = \omega_{0,S_2} - \omega_{\text{mw}}$ are the offset of electron 1 and 2 with respect to the microwave frequency ω_{mw} ,

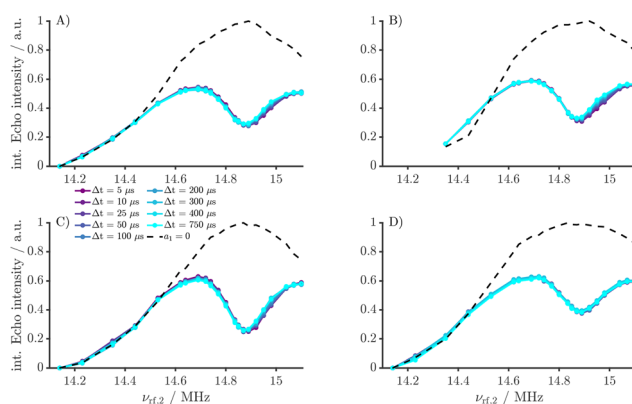


Fig. 6 Hole burning spectra for delays Δt between the two Gauss pulses varying from 5 to 750 μs , $t_{\text{del}} = 800$ μs and $t_{\text{SL}} = 4000$ ns. The hole was burned at $\nu_{\text{rf},1} = 14.89$ MHz. The black dashed lines are obtained by using the data points with $a_1 = 0$ i.e. in the absence of any hole-burning pulse. The sample in subplot (A) is 5 mM trityl in gly-d₈:D₂O:H₂O (6:3:1), in subplot (B) 5 mM trityl in gly-d₈:D₂O (6:4), in subplot (C) 5 mM trityl in gly-d₈:H₂O (6:4) and in subplot (D) 100 μM trityl in gly-d₈:D₂O:H₂O (6:3:1).



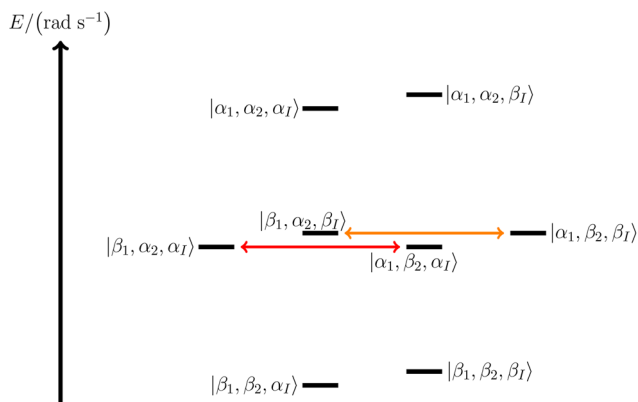


Fig. 7 Energy level diagram for a three-spin system consisting of two electron spins 1 and 2 and a proton spin I , where the difference in the Larmor frequencies of the electron spins 1 and 2 is roughly equal to half of the secular hyperfine coupling constant, *i.e.*, $|\omega_{0,S_1} - \omega_{0,S_2}| \approx \left| \frac{A_{zz}^{(1,I)}}{2} \right|$. The two transitions that are responsible for the observed electron spin diffusion process are shown in red with the proton spin in the α_I -state and in orange with the proton spin in the β_I -state, respectively. The energy levels and populations in this diagram are drawn only schematically and are not to be taken as exact.

$A_{zz}^{(1,I)} = \frac{\delta_{1,I}}{2}(3\cos^2(\beta_{1,I}) - 1)$ the secular and $B^{(1,I)} = \frac{3}{4}\delta_{1,I}\sin(2\beta_{1,I})$ the pseudo-secular part of the hyperfine coupling between electron 1 and the proton. The electron resonance frequencies are given by $\omega_{0,S_1} = g_{zz}^{(1)}\frac{\mu_B}{\hbar}B_0$ and $\omega_{0,S_2} = g_{zz}^{(2)}\frac{\mu_B}{\hbar}B_0$, while B_0 is the external static magnetic field applied along the laboratory z -axis. Thus, the proton Larmor frequency is given by $\omega_{0,I} = -\gamma_I B_0$. $\delta_{1,I} = \frac{2\mu_0\mu_B g_{zz}^{(1)}\gamma_I}{4\pi r_{1,I}^3}$ is the hyperfine coupling constant and $D^{(1,2)} = -\frac{2\mu_0 g_{zz}^{(1)}g_{zz}^{(2)}\mu_B^2}{4\pi\hbar r_{1,2}^3} \frac{3\cos^2(\theta_{1,2}) - 1}{2}$ is the dipole-dipole coupling constant between electron spin 1 and 2. If the two electrons undergo a flip-flop transition ($\alpha_1 \rightarrow \beta_1$ and $\beta_2 \rightarrow \alpha_2$, *i.e.*, electron spin diffusion), the hyperfine coupling to the proton changes sign, and the frequency of the proton changes. If the difference in the Larmor frequencies between the two electron spins is roughly equal to half of the secular hyperfine coupling constant, *i.e.*, $|\omega_{0,S_1} - \omega_{0,S_2}| \approx \left| \frac{A_{zz}^{(1,I)}}{2} \right|$, the process in the three-spin system would be energy conserving and, therefore, fast. The matrix elements that drive this process are $\langle \beta_1, \alpha_2, \alpha_I | \hat{\mathcal{H}} | \alpha_1, \beta_2, \alpha_I \rangle = \langle \alpha_1, \beta_2, \alpha_I | \hat{\mathcal{H}} | \beta_1, \alpha_2, \alpha_I \rangle = -\frac{D^{(1,2)}}{2}$ and originate from the flip-flop operators $\hat{S}_1^+\hat{S}_2^-$ and $\hat{S}_1^-\hat{S}_2^+$ in eqn (1). Note that for the proton spin in the β_I -state the description is equivalent. This electron spin diffusion process is schematically shown in an energy level diagram in Fig. 7 and is what we observe experimentally when burning a hole at $\nu_{rf,1} = 14.69$ MHz or $\nu_{rf,1} = 15.04$ MHz. The hole initially burned at $\nu_{rf,1}$ is slowly filled with magnetization originating from the spectral region symmetric around the center of the proton spectrum.

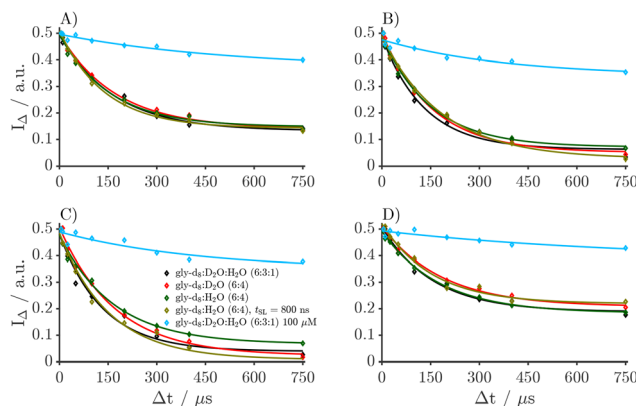


Fig. 8 Comparison of the difference magnetization according to eqn (3) for different combinations of ν_h and ν_f across the four different samples. The hole is burned at $\nu_{rf,1} = 14.69$ MHz. (A) Corresponds to $\nu_h = 14.69$ MHz, $\nu_f = 15.04$ MHz, (B) to $\nu_h = 14.77$ MHz, $\nu_f = 14.98$ MHz, (C) to $\nu_h = 14.80$ MHz, $\nu_f = 14.95$ MHz and (D) to $\nu_h = 14.64$ MHz, $\nu_f = 15.10$ MHz.

In the center of the proton spectrum at $\sim \nu_{rf,1} = 14.89$ MHz the electron spin diffusion is not visible on the proton spectra because the hyperfine coupling is too small to generate a visible splitting of the resonances. Thus, the exchange of magnetization between the two spectral positions in the proton spectrum is most likely caused by an electron spin-diffusion process. Similar phenomena were observed in MAS NMR in the ^{13}C spectra of adamantane⁴¹ or in ^{13}C spectra of a small organic molecule with trifluoromethyl group.⁴² In a three-spin system analogous to ours case, *i.e.*, two coupled electron spins and one proton coupled to one of them, a $T_{1,e}$ -driven mechanism was observed in Davis ENDOR for a frozen solution of copper(*l*-histidine)₂.⁴³ The three-spin mechanism in ENDOR is observed by matching $|\omega_{0,S_1} - \omega_{0,S_2}| = |\omega_{0,I}|$ analogous to the cross effect in DNP.⁴⁴⁻⁴⁷

To check that magnetization is indeed transported symmetrically across the center of the proton line, one can analyze the behavior of the sum $I_\Sigma(\Delta t)$ and difference magnetization $I_A(\Delta t)$ for two spectral positions ν_h and ν_f . The frequency ν_h denotes the spectral position near the hole and ν_f the symmetric position where magnetization is lost. The two frequencies are symmetric around the center of the spectrum, *i.e.*, $\nu_h = \nu_{0,I} + \Delta\nu$ and $\nu_f = \nu_{0,I} - \Delta\nu$, and are shown in Fig. 5(A). Then the sum magnetization I_Σ and the difference magnetization I_A can be defined as

$$I_\Sigma(\Delta t) = \frac{1}{2}(I_h(\Delta t) + I_f(\Delta t))/(I_h(0) + I_f(0)), \quad (2)$$

$$I_A(\Delta t) = \frac{1}{2}(I_f(\Delta t) - I_h(\Delta t))/(I_f(0) - I_h(0)), \quad (3)$$

where $I_h(\Delta t)$ and $I_f(\Delta t)$ represent the intensities of the proton spectra at position ν_h and ν_f as shown in Fig. 5. If the magnetization is transferred symmetrically around the center of the spectrum, the sum magnetization I_Σ is constant over Δt and the difference magnetization I_A decays towards a plateau I_∞ . Please note that the intensity $I_h(\Delta t = 0)$ was set to the intensity at $\Delta t = 5 \mu\text{s}$. In Sections E.2.–E.6. of the SI we show the



comparison of I_{Σ} and I_A for all four different samples and combinations of ν_h and ν_r . The sum magnetization I_{Σ} is indeed constant over the time scale of Δt for almost all cases. An exception is the case for the sample with 100 μM trityl in gly- d_8 : D_2O : H_2O (6:3:1), where almost no diffusion process is visible. In Fig. 8 we only show the difference magnetization for the hole burned at $\nu_{\text{rf},1} = 14.69$ MHz for all four different samples. We fitted the difference magnetization $I_A(\Delta t)$ to an exponential decaying function including a plateau I_{∞} as described in eqn (S.6) in the SI. The extracted fitting parameters are given in Table 1. From the data in Fig. 8 and Table 1 we can see that the three samples with 5 mM trityl concentration show almost identical decays. A small variation in the decay-rate constant and the final value with respect to the proton concentration can be observed especially at the center of the line. Among the samples with 5 mM trityl concentrations, the deuterated sample shows the slowest decay and the sample with only H_2O the fastest decay. Here we used a simple mono-exponential function to fit the data. A sum of exponentials or stretched exponentials might be a better fitting model. We refrain from optimizing fit quality by making the model more complex, as we aim to describe the origin of the diffusion process only qualitatively here.

When we compare the 5 mM samples to the 100 μM sample, we see a drastic slowdown of the spectral diffusion process by a factor of 4–5 in the decay rate constant τ upon reducing electron concentration by a factor of 50. This is consistent with our initial assessment that the observed proton polarization transport is the result of an electron spin diffusion process within a three-spin system consisting of two coupled electron spins and on proton spin. A decrease in the electron–electron coupling strength between the 5 mM samples and the 100 μM sample is also visible in phase memory time T_M measured using a two pulse Hahn echo and shown in Fig. S25 Section A.7. in the SI.

3 Conclusions

We have demonstrated that it is possible to indirectly detect the protons close to electron spins using the reverse DNP scheme and frequency-band selective proton pulses. Substantial spectral splittings caused by the large hyperfine couplings make direct detection *via* regular NMR experiments difficult. We show experimentally that the observed proton line originates from protons very close to the electron spins most likely to a large extent from the protons on the trityl molecule itself. This can be inferred from the fact that the intensity of the electron-detected spectrum is largely independent of the proton concentration of the solvent. The electron-detected proton spectrum is also independent of the trityl concentration.

The center of the electron-detected proton spectrum is shifted by ~ 40 kHz and the FWHM increases by a factor ~ 20 compared to the bulk proton signal. We validated our experimental results by simultaneous application of electron decoupling during the band-selective inversion pulse, which

reduces the proton line width significantly. Under optimized π -pulse decoupling on the electrons, the FWHM is reduced by a factor of 5. The observed proton line width under electron decoupling is still a factor ~ 4 larger than the line width of the bulk spectrum. This is most likely due to the large radio-frequency field amplitude of the band-selective proton pulses. In addition, imperfect electron decoupling might also contribute to the larger line width, and we expect that the decoupling efficiency could be increased by using broadband chirp pulses.

By measuring the spin-diffusion rate constant to the bulk as a function of the proton spectral frequency, we were able to characterize the spectral diffusion barrier of the proton spins close to the electrons. Thus, we observe a spectral region ranging from ~ 14.64 MHz to 15.07 MHz in which spin diffusion to bulk protons centered at $\nu_{0,1} \sim 14.85$ MHz takes place. This results in a radius for the diffusion barrier ranging from ~ 5.4 to 6.8 Å. For proton spins with an electron–proton distance shorter than ~ 5.4 Å, diffusion to the bulk protons is quenched. A straightforward analysis of the diffusion time constant is complicated by different effects that influence the diffusion of polarization from protons close to an electron spin to the bulk protons.

Using hole-burning experiments on the proton line, we were able to characterize electron spin diffusion through a change in the proton resonance frequency. The electron spin diffusion mechanism was confirmed by comparing samples with two different concentrations of trityl. The electron spin diffusion process is absent in a sample of 100 μM trityl in gly- d_8 : D_2O : H_2O (6:3:1) and is almost independent of the concentration of protons in the matrix.

The observations in this publication have implications for the future design of new DNP experiments and probably also ENDOR experiments. We were able to detect the proton spectrum of protons spins nearby an unpaired electron spin in trityl Ox063 and to reduce the line width of that spectrum by applying electron-decoupling. Furthermore, we showed that a hole can be burned into the spectrum and an electron–electron spin diffusion process within an electron–electron–proton spin system can be observed. More important, we showed that not all protons nearby an electron spin take part in the spin diffusion process towards the bulk proton spin bath. This has important consequences for the design of more efficient DNP radicals. In this work, we measured a narrow-line trityl radical. We expect that the experiments in this work can be extended by pulsed broadband DNP sequences^{48,49} to also investigate radicals with broader EPR spectra such as nitroxides. Extending the pulse sequences in this work to large magnetic fields would also be an interesting direction for further research. However, the NOVEL matching condition is difficult to fulfill at magnetic fields larger than 3 T. In principle other DNP sequences like the solid effect and its frequency-swept versions (integrated SE,^{50–54} stretched SE^{53,54} and adiabatic SE^{54,55}) or pulsed DNP sequences like TOP,³⁰ XiX^{31,32} or TPPM³³ can be used. Those sequences show a decreased DNP efficiency compared to NOVEL at the same magnetic field⁵⁶ and use off-resonance mw irradiation on the electron spin. While a decrease in DNP efficiency can be overcome by slightly increasing the number of



Table 2 List of molecular weight (MW) and density (ρ_0) used for sample preparation. Ox063 trityl is a solid and therefore no density is listed

Compound	MW/(g mol ⁻¹)	ρ_0 /(g mL ⁻¹)
Ox063 trityl	1359	—
H ₂ O	18.02	0.997
D ₂ O	20.03	1.11
Glycerol-d ₈	100.14	1.371

DNP contacts, off-resonance mw irradiation could potentially lead to artifacts due to spectral diffusion *i.e.* for a Hahn echo detection applied on-resonance on the electron spin. However, we expect that the measurement of the spin diffusion barrier is dominated by the hyperfine coupling strength, and thus independent of the external magnetic field.

4 Materials and methods

4.1 Sample preparation

The molecular weights and densities used to prepare the four different samples are listed in Table 2. The four different samples were prepared by the following procedures:

4.1.1 5 mM trityl in gly-d₈:D₂O:H₂O (6:3:1). 1.69 mg of the trityl were dissolved in 24.15 mg water (deionized water from the lab) and 80.93 mg D₂O (Sigma Aldrich, ≥ 99.85 atom% D). This gives a ~ 1.2 μ M solution of trityl dissolved in a D₂O:H₂O matrix of 3:1 by volume. From that resulting solution, 48.6 μ L were added to 99.92 mg glycerol-d₈ (Sigma Aldrich, ≥ 98 atom% D by Chromatography Purity) to give the desired 5 mM solution of trityl in DNP juice. 40 μ L of that final solution were added to a 3 mM OD quartz capillary. All masses were weighed in with a AT261 delta range scale with a precision of 0.01 mg. A pipetman from GILSON (P100) with volumes adjustable to 0.1 μ L precision was used to transfer and measure the volumes.

4.1.2 5 mM trityl in gly-d₈:D₂O (6:4). The sample with a fully deuterated matrix was prepared inside a glove box. All chemicals and equipment were placed in the glove box 24 hours prior to sample preparation to prevent contamination from residual surface moisture.

1.7 mg of the trityl were dissolved in 112.5 mg D₂O (Cambridge Isotope Laboratory, ≥ 99.85 atom% D, sealed glass ampule). This gives a ~ 1.2 μ M solution of trityl dissolved in a D₂O. From that resulting solution, 48.6 μ L were added to 100.2 mg glycerol-d₈ (Sigma Aldrich, ≥ 98 atom% D by CP, freshly opened in the glove box) to give the desired 5 mM solution of trityl in a gly-d₈:D₂O (6:4 by volume) matrix. 40 μ L of that final solution were added to a 3 mM OD quartz capillary inside the glove box. The quartz capillary was then transferred under inert condition and sealed using a Schlenk line. All masses were weighed in with a Mettler Toledo scale with a precision of 0.1 mg. As GILSON PIPETMAN (P100 and P20) with volumes adjustable to 0.1 μ L precision were used to transfer and measure the volumes.

4.1.3 5 mM trityl in gly-d₈:H₂O (6:4). 1.68 mg of the trityl were dissolved in 101.34 mg H₂O (deionized water from the

lab). This gives a ~ 1.2 μ M solution of trityl dissolved in a H₂O. From that resulting solution, 48.6 μ L were added to 108.36 mg glycerol-d₈ (Sigma Aldrich, ≥ 98 atom% D by CP) to give the desired 5 mM solution of trityl in a gly-d₈:H₂O (6:4 by volume) matrix. 40 μ L of that final solution were added to a 3 mM OD quartz capillary. All masses were weighed in with a Mettler Toledo AX205 scale with a precision of 0.01 mg. A GILSON PIPETMAN (P100) and GILSON (P20) with volumes adjustable to 0.1 μ L precision were used to transfer and measure the volumes.

4.1.4 100 μ M trityl in gly-d₈:D₂O:H₂O (6:3:1). 0.33 mg of the trityl were dissolved in 242.94 mg water (deionized water from the lab) and 810.24 mg D₂O (Sigma Aldrich, ≥ 99.85 atom% D). This gives a ~ 0.25 μ M solution of trityl dissolved in a D₂O:H₂O matrix of 3:1 by volume. From that resulting solution 486 μ L were added to 1000.2 mg glycerol-d₈ (Sigma Aldrich, ≥ 98 atom% D by CP) to give the desired 100 μ M solution of trityl in DNP juice. 40 μ L of that final solution were added to a 3 mM OD quartz capillary. All masses were weighed in with a Mettler Toledo AX205 scale with a precision of 0.01 mg. A GILSON PIPETMAN (P100) with volumes adjustable to 0.1 μ L precision was used to transfer and measure the volumes.

All experiments were conducted at 80 K and an external magnetic field of around 0.35 T (X-band, $\nu_{0,e} \sim 9.8$ GHz, $\nu_{0,H} \sim 14.83$ MHz). The sample was flash frozen in liquid nitrogen before placed in the resonator.

4.2 EPR and NMR measurements

All experiments were acquired on a home-built X-band spectrometer similar to the spectrometer described in ref. 57. A constant temperature of 80 K was achieved with a closed-cycle cryogen-free cryostat from Cryogenic Limited. An arbitrary waveform generator (Keysight model M8190A) was used to generate mw pulses and the pulses were amplified using a 1 kW traveling wave tube (TWT) amplifier. The Hahn echoes were recorded using a digitizer running at 1.8 GSa s⁻¹ (SP devices ADQ412) for the measurements of the samples 5 mM trityl in gly-d₈:D₂O:H₂O (6:3:1), 5 mM trityl in gly-d₈:D₂O (6:4) and 5 mM trityl in gly-d₈:H₂O (6:4). The sample 100 μ M trityl in gly-d₈:D₂O:H₂O (6:3:1) was measured using a digitizer running at 10 GSa s⁻¹ (ADQ7DC). A Bruker EN4118X-MD4 resonator was used with a home-built external rf tuning and matching box. NMR pulses were generated with an OpenCore spectrometer.^{58,59} A schematic representation of a general DNP experiment used in this work can be seen in Fig. 1. The general sequence starts with a NOVEL DNP contact consisting of a $\frac{\pi}{2}$ pulse of 6 ns length and a digital amplitude 1 (max. amplitude) and a spin lock of length t_{SL} . In most of the experiments t_{SL} was set to 4000 ns. The optimal amplitude and length of the spin lock was determined by a single DNP contact followed by a Hahn echo detection as described in Section A.1. of the SI. After the first DNP contact the AWG was delayed for t_{del} . During this delay pulses on the proton channel were applied using a BLAX300RS amplifier, where the maximum output power is 300 W corresponding to a relative amplitude of 100. A detailed description of the pulse scheme on the proton channel is given below. After the delay t_{del}



the electron spin was saturated by a train consisting of five pulses of 10 ns length spaced by 1800 ns. This step is necessary to ensure that the proton spins are more strongly polarized than the electron spin and thus to facilitate the reverse DNP transfer (second spin lock). The reverse DNP was started 1800 ns after the last saturation pulse. The electron signal was then read out by a Hahn echo with $\tau = 600$ ns and a π of 12 ns length and a digital amplitude of 1. Each single-shot experiment was followed by a ^1H saturation pulse train consisting of eleven 100° pulses spaced by 1 ms to destroy any polarization left on the protons. The frequency of the saturation pulses was always on resonance with respect to tuning and matching. The shot repetition time was set to 15 ms $\sim 5T_{1,e}$.

4.2.1 Electron-detected proton spectra. To record the electron-detected proton spectra as described in Section 2.1 the nutation pulse (Gauss or rectangular pulse) on the proton channel was placed in the middle of the delay as shown in Fig. 1(A) for a variable-amplitude Gauss pulse. The carrier frequency of the nutation pulse $\nu_{0,I}$ was swept from 14.14 MHz to 15.10 MHz. For each frequency the rf coil in the resonator was tuned and matched accordingly. An overview of the minimum of the dip after tuning and matching is shown in Fig. S46 and S47 in the SI. In case of a rectangular nutation pulse the relative amplitude of the pulse was kept constant at 30 corresponding to 100 kHz and the length of the pulse was varied from 0 to 9 μs in 0.5 μs steps. In case of the Gauss pulse, the pulse length was kept constant at 10 μs and the relative amplitude was varied from 0 to 40 in 2.5 steps. The minimum of the amplitude nutation trace was read out for $\nu_{0,I} = 14.89$ MHz corresponding to the center of the electron-detected proton spectra *i.e.* amplitude nutation trace with the largest inversion. For the variable-amplitude Gauss pulse the minimum was at a relative amplitude of 25 with and without electron decoupling. For the rectangular nutation pulse the minimum of the amplitude nutation trace was at $\tau_{\text{nut}} = 3.5$ μs without electron decoupling and at $\tau_{\text{nut}} = 4$ μs with electron decoupling applied during the nutation pulse. The electron-detected proton spectrum was then obtained by reading out the data points at the minimum position determined for $\nu_{0,I} = 14.89$ MHz. The spectra were baseline corrected to the signal corresponding to $\nu_{0,I} = 14.14$ MHz which did not show any nutation. They were afterwards multiplied by -1 to obtain a positive signal. Electron decoupling sequence consisted of 99 π pulses (12 ns pulses at digital scale of 1) spaced by 80.2 ns and was applied during the nutation pulse (see Fig. S35 Section B. in the SI). Experiments were recorded for $t_{\text{del}} = 15$ μs and 800 μs .

4.2.2 Detection of proton spin diffusion towards the bulk. To observe the proton spin diffusion into the bulk for different carrier $\nu_{\text{rf},2}$ the Gauss pulse was placed at the end of the delay (see blue Gauss pulse in Fig. 1(B)). The length of the Gauss pulse at the end was kept constant at 10 μs and the relative amplitude of this pulse was swept from 0 to 40 in 2.5 steps. The delay t_{del} was varied from 15 μs to 3000 μs . The spin diffusion spectra were then obtained by the minimum of the amplitude nutation traces at a relative amplitude of 25 (equal to the electron detected proton spectra). The NMR coil of the resonator was tuned and impedance matched following the same procedure as outlined for the electron-detected proton spectra.

4.2.3 Electron spin diffusion imprinted on the proton spectrum. The pulse sequence containing two Gauss pulses on the proton channel is schematically shown in Fig. 1(C). For all experiments $t_{\text{del}} = 800$ μs was fixed. The blue pulse was the detection pulse (inversion pulse). The length of this pulse was set to 10 μs and the amplitude was set to the minimum of the amplitude nutation trace recorded at $t_{\text{del}} = 800$ μs in the experiment with only one Gauss pulse at the end. The NMR coil of the resonator was tuned and matched on resonance with respect to the carrier frequency $\nu_{\text{rf},2}$ of the detection pulse. The frequency $\nu_{\text{rf},2}$ was scanned through the proton spectrum from 14.14 MHz to 15.10 MHz. The red Gauss pulse served as an hole-burning pulse. The frequency $\nu_{\text{rf},1}$ was fixed to 14.69, 14.89 or 15.04 MHz and the pulse length was set to 10 μs . The space Δt between the two Gauss pulses was varied from 5 μs to 750 μs . The amplitude of red Gauss pulse a_1 was varied for every single Δt to ensure optimal hole burning performance also for large frequency differences between the carrier frequencies of the two Gauss pulses *i.e.* $\nu_{\text{rf},1}$ and $\nu_{\text{rf},2}$. The minimum or maximum of the amplitude traces for a fixed Δt were extracted and examples for all four different samples can be found in Sections E.2.–E.6. of the SI. The obtained spectra were zero-corrected to the data point at 14.14 MHz, inverted and compared to the data with $a_1 = 0$, *i.e.* in the absence of an hole burning pulse.

The NMR spectra without any DNP involved were recorded with only pulses applied to the proton channel. The proton spectra were recorded by a solid echo consisting of two 90° pulses of 2.5 μs length separated by $t_{\text{SE}} = 20$ μs . For the solid echo a eight step phase cycle was used with $\{x, x, y, y, -x, -x, -y, -y\}$ for the first pulse and detection and $\{y, -y, x, -x, y, -y, x, -x\}$ for the second pulse. A dwell time of 4 μs and a number of 1024 sampling points was used to sample the solid-echo signal. A saturation recovery pulse sequence with adjustable delay τ_{delay} was used to record the $T_{1,H}$ of the proton spin. The reference experiment was recorded with $\tau_{\text{delay}} = 180$ s $\approx 5 \cdot T_{1,H}$. To process the NMR data, a cosine-squared apodization function was used and time-domain data was zero-filled to twice the number of recorded data points. After Fourier transformation of the resulting time-domain data, the peak was fitted by a Lorentzian–Gauss blend.

Author contributions

N. W., M. E. and G. J. designed the research. G. M. C. performed all experimental measurements, simulations, data processing, and data evaluation. G. M. C. wrote the first draft and all authors were involved in editing the final manuscript.

Conflicts of interest

There are no conflicts to declare.

Data availability

Supplementary information is (SI) available. See DOI: <https://doi.org/10.1039/d6cp00086j>.



The data that support the findings of this study are available at <https://doi.org/10.3929/ethz-c-000791347>.

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