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Designing soft and tough multiple-network elastomers: impact of reversible radical deactivation on filler network architecture and fracture toughness†

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Polymer networks are crucial for engineering and biomedical applications; however, their excessive brittleness in the absence of viscoelastic dissipation limits their use in applications requiring high temperatures and water concentrations. Multiple-networks, consisting of a stiff and pre-stretched “filler” network phase embedded within a soft and extensible “matrix” network, offer a promising route to overcome this limitation; yet, the relationship between “filler” network architecture and fracture toughness remains unknown. Here, we synthesized three poly(ethyl acrylate) “filler” networks via free radical polymerization (FRP), RAFT, and ATRP, and assessed the interplay between “filler” network architecture, irreversible chain breakage, and fracture toughness by labelling their crosslinks with fluorogenic mechanophores. These networks had similar elastic chain densities but distinct chain length distributions and mesoscopic structures. They were used to prepare multiple-networks, whose structure and mechanical properties were evaluated using mechanical tests, confocal microscopy, reactive Monte Carlo, and coarse-grained molecular dynamics simulations. Our results reveal that “filler” networks synthesized by RAFT and ATRP yield more brittle multiple-networks than those made by FRP, primarily due to the reduced average extensibility of their “filler” network chains. Their narrower chain length distributions in the load-bearing phase promote strain hardening but compromise energy dissipation through molecular friction and chain breakage, as well as fracture toughness. Overall, these findings underscore the need for advanced gelation methods that provide control over elastic chain distributions within networks, as, without such improvements, the use of RAFT and ATRP may result in long curing times, opacity, and a greater tendency to fracture.

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Introduction

Polymer networks are irreplaceable in engineering and biomedical applications, such as elastomers in rubber tires, dampers, and seals;¹ hydrogels in contact lenses and superabsorbent diapers;² and pressure-sensitive-adhesives (PSAs) in labels, band-aids, and double-sided tape.³ These materials feature elastic moduli E ranging from 1 kPa to 1 MPa and can sustain large deformations near pre-existing defects or cracks.^{4–6} However, at high temperatures or solvent concentrations, they readily break, precluding their use in applications such as water purification membranes,⁷ hydrogels for artificial prosthetics and drug delivery,⁸ solid polymer electrolytes for energy conversion and storage devices,⁹ and electro-adhesives for advanced manufacturing.¹⁰ In these

conditions, the polymer segmental dynamics facilitate ion or small-molecule transport, but the networks exhibit limited viscoelastic dissipation, which leads to localized stresses, crack nucleation, and fracture. Designing polymer networks that are both soft and tough under these conditions is critical for developing advanced materials in emerging energy, healthcare, and manufacturing applications.

A common practice to design soft and tough polymer networks is to embed stiff carbon black or silica particles within a rubber-like polymer matrix.¹¹ This approach leads to an uneven load distribution between the stiff and soft phases,¹² which helps mitigate the nucleation and growth of cracks. However, probing this load distribution under increasing strain is challenging because of load transfer from the stiff particles to the soft matrix, molecular-scale damage, and large-strain cavitation.^{13–15} Gaining insight into this load distribution, along with its evolution and impact on energy dissipation and fracture, is essential for establishing structure–property relationships and designing next-generation polymer networks.

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We aim to understand and control the load distribution, energy dissipation, and fracture toughness of soft and tough polymer networks, leveraging three recent advances in polymer science. First, the introduction of double-network hydrogels and multiple-network elastomers by Gong *et al.* and Ducrot *et al.*,^{16,17} which consist of a stiff and brittle “filler” network phase embedded within a soft and extensible “matrix” network. These materials result from sequential polymerization and swelling steps, which isotropically pre-stretch the “filler” network chains to a stretch ratio λ_0 ; and have mechanical properties governed by an uneven load distribution between the stiff and soft phases, just like conventional filled networks.^{18–22}

Specifically, at small strains, the “filler” network carries the load within its chains; while at large strains, these chains elongate and break, transferring the load to the “matrix” network. Therefore, the mechanical properties of these materials closely depend on the pre-stretch of the “filler” network chains, λ_0 , as well as the architecture (*i.e.*, topology) and interconnectivity between the “filler” and “matrix” networks. For instance, when the “filler” network chains elongate to their contour lengths, the multiple-networks undergo strain-hardening; and when they break, the multiple-networks dissipate significant energy and exhibit enhanced toughness.^{17,21–23} Thus, we consider multiple-networks as ideal systems for designing soft and tough polymer networks by focusing on the impact of polymerization methods on gelation.

The second important development is the use of reversible deactivation radical polymerizations (RDRPs) such as reversible addition–fragmentation transfer (RAFT), atom transfer radical polymerization (ATRP), and nitroxide-mediated polymerization (NMP) to structurally tailor and engineer polymer networks.^{24,25} These techniques provide control over the molecular weight, dispersity, and microstructure of linear polymers (*i.e.*, comonomer sequence) and can influence the topology of polymer networks. Fukuda,^{26,27} Billingham and Armes,^{28,29} Matyjaszewski,^{30–33} Allonas and Croutxé-Barghorn,^{34,35} and their coworkers have demonstrated that RDRPs prevent the formation of “dead” gels or clusters at the early stages of monomer-crosslinker copolymerization, shifting the percolation threshold to higher vinyl group conversions and narrowing the chain length distribution within the networks. These RDRPs were conducted at either high crosslinker, ≈ 85 mol%,^{34,35} or high solvent concentrations, ≈ 50 wt%,^{36,37} resulting in polymer networks with mechanical properties ranging from stiff and brittle to soft and tacky.

More recently, one of our investigations showed that conducting RAFT and ATRP copolymerizations at lower crosslinker concentrations and under bulk conditions (*i.e.*, without solvent) also induces sol–gel demixing at high vinyl group conversions (*i.e.*, microsyneresis), resulting in polymer networks with crosslinker-rich and crosslinker-poor mesophases that form at the late stages of gelation, an opaque appearance, and elastomer-like mechanical properties (*i.e.*, Young’s modulus, $E \sim 1$ MPa, and fracture toughness, $G_c \sim 100$ J m^{−2}).³⁸ Therefore, we consider RDRPs to be promising techniques for controlling the architecture of both single- and multiple-networks, and understanding their impact on load distribution, energy dissipation, and mechanical properties.

The third important development is the use of mechanochemistry as a tool to map and quantify damage in polymers.^{39–44} This approach involves incorporating force-responsive probes, known as mechanophores, at load-bearing points within polymer networks, such as the crosslinks. These probes selectively activate at forces similar to those required to break a covalent bond, inducing changes in the photophysical properties of the bulk in response to force. For example, force-induced chemical reactions can turn these probes into fluorophores, with fluorescence detectable under a confocal microscope.^{45–48} This fluorescence provides a direct damage measure and is particularly useful for studying fracture.^{23,47,49–52} Hence, we consider this fluorogenic mechanochemistry technique a powerful tool for unraveling the impact of “filler” network architecture, as controlled by RDRPs, on load distribution, damage, energy dissipation, and fracture toughness of multiple-networks.

Before proceeding, we deem it important to highlight two key bodies of work on the structure–property relationships of these multiple-networks. The first is by Kawauchi *et al.*, which examined the effect of “filler” network heterogeneities on the fracture toughness of double-network hydrogels.⁵³ This study revealed that “filler” networks with no or large-scale heterogeneities, $\xi > 300$ nm, synthesized by monomer-crosslinker copolymerization in either good or poor solvents, offered more brittle double-networks than those with moderately sized heterogeneities, $\xi = 10$ –300 nm, where ξ represents the correlation length of the spatial heterogeneities within the “filler” network.

The second study, by Slootman *et al.*, investigated the impact of “filler” network pre-stretch, λ_0 , on the fracture toughness of multiple-network elastomers.²³ Their results demonstrated that pre-stretched “filler” network chains improve the high-temperature (*i.e.*, low-rate) fracture toughness of multiple-networks by facilitating damage delocalization ahead of cracks and improving the load transfer efficiency between the “filler” and “matrix” networks. Namely, this work illustrated that when the “filler” network pre-stretch exceeds $\lambda_0 > 1.6$, the “matrix” network can more effectively carry the load transferred upon “filler” network breakage, delaying the transition from random (*i.e.*, mean-field) to localized (*i.e.*, correlated) “filler” network scission, crack nucleation, and fracture.

In addition, Slootman *et al.* demonstrated that rate and temperature strongly influence this damage delocalization mechanism, highlighting the critical role played by viscoelasticity in energy dissipation, bulk deformation, and fracture.²³ Their findings showed that multiple-networks dissipate energy through both viscoelasticity and damage, with the number of broken “filler” network chains per unit area of propagated crack, Σ_x , scaling with the effective stretch of the “filler” network chains at the fracture point, $\lambda_c \lambda_0$, $\Sigma_x \sim \exp(\alpha \cdot \lambda_c \lambda_0)$, where $\alpha = 1.75$. This effective stretch, $\lambda_c \lambda_0$, depends on both the loading rate and temperature, indicating that viscoelasticity and damage are inherently coupled in dissipating energy ahead of cracks.

With these considerations in mind, we hypothesized that RDRPs such as RAFT and ATRP could offer control over the “filler” network architecture, as well as the resulting load transfer mechanism, energy dissipation, and fracture toughness of



multiple-networks. To validate this hypothesis, we synthesized a series of polymer networks *via* free radical copolymerization (FRP), RAFT, and ATRP, using ethyl acrylate (EA) as the monomer and 1,4-butanediol diacrylate (BDA) as the crosslinker. These networks were similar to those reported in our previous work, where we showed that RAFT and ATRP lead to a different interplay between elasticity (*i.e.*, Young's modulus) and strain-hardening than FRPs.³⁸ Here, we used these networks as “filler” phases in multiple-network systems, focusing specifically on how their architecture impacts energy dissipation and fracture toughness.

Through mechanical testing, mechanochemistry, reactive Monte-Carlo (RMC) and coarse-grained molecular dynamics (CGMD) simulations, we show that, at the same crosslink density, ν_x , (*i.e.*, modulus, E , or pre-stretch ratio λ_0), “filler” networks synthesized *via* RAFT and ATRP result in more brittle multiple-networks than those synthesized by FRP. The reversible deactivation of radical chain ends during gelation leads to the formation of crosslinker-rich and crosslinker-poor mesophases, along with narrower chain length distributions within the crosslinker-rich, load-bearing phase (*i.e.*, lower average chain extensibility, λ_h). This reduced chain extensibility limits the load transfer efficiency between the “filler” and “matrix” networks, and decreases the energy dissipated per broken “filler” network bond ahead of cracks, thereby reducing overall toughness. These results inform the molecular design of soft and tough polymer networks, underscoring the need for advanced polymerization methods that offer control over elastic chain distribution, mesoscopic architecture, and load transfer efficiency of multiple-networks.

Experimental

Materials

Unless otherwise specified, all chemicals were used as received. Ethyl acrylate (EA) and 1,4-butanediol diacrylate (BDA) were sourced from TCI America; 2-hydroxy-2-methylpropiophenone (HMP), cyanomethyl dodecyl trithiocarbonate (CTA), ethyl α -bromoisobutyrate (EBiB) copper(i) bromide (CuBr), tris[2-(dimethylamino)ethyl]amine (Me₆TREN), anhydrous THF, triethylamine, acryloyl chloride, hydroquinone, magnesium sulfate and anhydrous ethyl acetate from Millipore Sigma; basic aluminum oxide, toluene, dichloromethane (DCM), sodium chloride, sodium bicarbonate and anhydrous DMSO from VWR; and the π -extended anthracene-maleimide adduct, diol precursor of the fluorogenic mechanophore (DACL), from Concept Life Sciences.

EA (100 mL) and BDA (25 mL) were purified by elution over basic aluminum oxide. Purified EA, BDA and as-received HMP, CTA, EBiB, and Me₆TREN were placed in dry septum-sealed bottles, sparged with N₂ for 45 min, and transferred into a N₂-filled glovebox.

Synthesis of fluorogenic mechanophore

The fluorogenic mechanophore (DACL) was synthesized following the procedure reported by Göstl *et al.*⁴⁵ Briefly, the diol precursor (50 mg, 0.11 mmol) was dissolved in anhydrous THF (2 mL) and

activated with excess triethylamine (160 μ L, 1.1 mmol). Separately, acryloyl chloride (90 μ L, 1.1 mmol) was dissolved in anhydrous THF (3 mL) in a round-bottom flask pre-cooled in an ice bath. The diol solution was then added dropwise to the acryloyl chloride solution, and the reaction mixture was allowed to proceed overnight at room temperature.

The following day, DI water (25 mL) was added to the reaction mixture, and the product was extracted with DCM (25 mL). The organic layer was washed with saturated sodium bicarbonate (3 $\times$ 25 mL) and saturated sodium chloride (3 $\times$ 25 mL), dried with magnesium sulfate, filtered, and used to dissolve hydroquinone (1 mg, 0.01 mmol). Finally, DCM was evaporated under vacuum, yielding the mechanophore crosslinker, DACL, as a pale-yellow solid. This mechanophore was dried overnight in a vacuum oven at room temperature, resulting in a reaction yield of 85% (see ¹H NMR in Fig. S1, ESI†).

Synthesis of filler networks

Polymer networks were synthesized in a N₂-filled glovebox, following a procedure introduced by Ducrot *et al.*^{17,54,55} and detailed in our previous work.³⁸ The concentration of propagating clusters was fixed at approximately 0.1 mol% through the concentration of initiator (FRP and ATRP) or chain transfer agent (RAFT).

For FRP, monomer EA, crosslinker BDA (0.5 mol%), and initiator HMP (0.1 mol%) were mixed in a 20 mL scintillation vial and transferred to a mold composed of two PET-covered glass plates sealed with a silicone rubber spacer approximately 0.1 cm thick. Instead, for RAFT and ATRP, the mixture consisted of monomer EA, crosslinker BDA (1.0 mol%), initiator HMP (0.02 mol%), CTA agent (0.1 mol%), and toluene (10%v/v); and monomer EA, crosslinker BDA (1.0 mol%), initiator EBiB (0.07 mol%), catalyst CuBr (0.007 mol%), ligand Me₆TREN (0.028 mol%), and DMSO (10% v/v). Importantly, ATRP polymerizations were conducted directly on glass, as their kinetics were too slow to prevent diffusion of the EA monomer within the PET layer and excessive adhesion of the networks (see a detailed summary of the polymerization conditions in Table S1, ESI†).

For networks labelled with fluorogenic mechanophores, a small fraction of crosslinker BDA, 0.02 mol%, was substituted with the mechanophore crosslinker (see a detailed summary of the polymerization conditions in Table S3, ESI†), just like in previous work.⁴⁷ This substitution resulted in networks with the same mechanical properties as those synthesized without mechanophore, consistent with the findings of Slootman *et al.*⁴⁷ (see representative stress–stretch curves in Fig. S10 and S11, ESI†).

The copolymerizations were initiated with UV light (365 nm, 0.30 mW cm^{−2}) and carried out until approximately 100% conversion: 2 h, 24 h, and 48 h for networks synthesized by FRP, RAFT, and ATRP, respectively. These times were determined from the kinetics of EA polymerization reported in Dookhith *et al.*³⁸

The resulting polymer networks were removed from the glovebox and dried under vacuum oven overnight at 30 °C. These networks had dimensions of approximately 8 $\times$ 4 $\times$ 0.1 cm³ and gel fractions above 99%.



Synthesis of multiple networks

Multiple-networks were synthesized in a N₂-filled glovebox according to a procedure introduced by Ducrot *et al.*^{17,54,55} and detailed in our previous work.³⁸ Briefly, the filler networks were swollen to equilibrium with a solution of monomer EA (99.98 mol%), crosslinker BDA (0.01 mol%) and initiator HMP (0.01 mol%) for 2 h, and transferred to a mold composed of two PET-covered glass plates sealed with a silicone spacer (≈ 0.1 cm thick). The polymerizations were initiated by irradiation with UV light (365 nm, 0.30 mW cm⁻²) and conducted for 2 h.

The resulting multiple-networks were transferred outside of the glovebox and dried overnight under vacuum at 30 °C. These networks had dimensions dictated by the filler network pre-stretch, λ_0 , and gel fractions above 99%.

Following Millereau *et al.*, this procedure was adjusted to fine tune the pre-stretch the “filler” network, leveraging the composition of the EA solution with ethyl acetate solvent to control the swelling, Q , and pre-stretch ratios, $\lambda_0 = Q^{-1/3}$.²¹

Visualization of phase separation in filler networks

Polymer networks were punch-cut into cylindrical specimens of 8 mm diameter, cleaned with ethanol and optical paper, and placed on a glass slide. Then, a Zeiss Axio Scope A1 microscope was used to examine their structure, acquiring brightfield images with a 20 $\times$ LD Plan – Neofluar objective.

Rheology

Polymer networks were punch-cut into cylindrical specimens of 8 mm diameter, and their rheological properties were evaluated in a discovery HR-2 rheometer equipped with stainless steel flat plates of the same diameter. Frequency sweeps from 0.1 to 100 rad s⁻¹ at temperatures from 30 to 75 °C were performed within the linear viscoelastic regime at a strain of 0.10%. Master curves for the storage, G' , and loss, G'' , moduli were constructed by time-temperature-superposition (TTS), using a reference temperature of 30 °C and horizontal a_T , and vertical, b_T , shift factors.

Uniaxial tension

Polymer networks were punch-cut into dog-bone shaped specimens of 20 mm gauge-length, 4 mm width, and approximately 1.3 mm thickness. These specimens were marked with two dots of white paint and then deformed with an Instron 34TM5 equipped with a 100 N load cell and a video extensometer. The initial stretch rate was varied between 3×10^{-4} and 3×10^{-2} s⁻¹, and the temperature between 23 and 75 °C.

The resulting force–displacement curves were used to compute the stress, σ , and stretch, λ , according to:

$$\sigma = \frac{F}{A_0} \quad (1)$$

and

$$\lambda = \frac{L}{L_0} \quad (2)$$

where F is the measured force, A_0 is the specimen cross-sectional area in the undeformed state, and L and L_0 are the

specimen lengths in the deformed and undeformed configurations, respectively.

The stress–stretch curves were used to estimate the Young's modulus, E , according to:

$$E = \left. \frac{d\sigma}{d\lambda} \right|_{1.05} \quad (3)$$

And the stress–stretch curves of the multiple-networks were fitted to Gent's hyperelastic model to estimate the strain hardening, λ_m , according to:

$$\sigma = \frac{E \left(\lambda - \frac{1}{\lambda^2} \right)}{3 \left(1 - \frac{J_1}{J_m} \right)} \quad (4)$$

where $J_1 = \lambda^2 + 2/\lambda - 3$ is the first stress invariant, and $J_m = \lambda_m^2 + 2/\lambda_m - 3$ is the maximum of the first stress invariant at the limiting extensibility, λ_m . Using this value, the limiting extensibility of the “filler” network chains, λ_h , was calculated as $\lambda_h = \lambda_0 \lambda_m$.

Uniaxial tensile tests were conducted by triplicate at each stretch rate and temperature to determine the average and 95% confidence interval of the modulus, E , and strain hardening, λ_m .

Single-edge notch crack propagation tests

Polymer networks were punch-cut into dog-bone shaped specimens of 20 mm gauge length, 4 mm width, and approximately 1.3 mm thickness. These specimens were cut with a fresh razor blade to introduce a crack of ≈ 1 mm length, marked with two dots of white paint, and deformed with an Instron 34TM5 equipped with a 100 N load cell and a video extensometer. The initial stretch rate was varied between 3×10^{-4} and 3×10^{-2} s⁻¹ and the temperature between 23 and 75 °C.

The resulting force–displacement curves were used to compute the engineering stress, σ , and stretch, λ , according to eqn (1) and (2).

The fracture toughness or critical energy release rate, G_c , was calculated using the Greensmith model:⁵⁶

$$G_c = \frac{6W(\lambda)c_0}{\sqrt{\lambda_c}} \quad (5)$$

where c_0 is the crack length in the initial state, λ_c is the critical stretch at which the crack starts to propagate, and $W(\lambda_c)$ is the strain energy density stored in the bulk at the fracture point,

$$W(\lambda_c) = \int_1^{\lambda_c} \sigma d\lambda.$$

Single-edge notch fracture tests were conducted by triplicate at each stretch rate and temperature to determine the average and 95% confidence interval of the fracture toughness, G_c .

Damage quantification through mechanochemistry

Fractured specimens were cleaned with ethanol and optical paper, and then taped to a Petri dish using double-sided tape. Fluorescent images were acquired on a Nikon AXR (Galvano and Resonant Scanner) confocal system equipped with a CFI60



Plan Apochromat Lambda D 20× N.A. 0.80, W.D. 0.80 mm, F.O.V. 25 mm, DIC, spring-loaded objective lens. π -Extended anthracene fluorophores were excited with a 405-nm laser, and their fluorescent emission recorded in the 424–524 nm range. The optical magnification was set to 20×, resulting in 2048 × 2048 raw images (0.43 $\mu\text{m pixel}^{-1}$). 3D scans were collected from the specimen surface (*i.e.*, plane of maximum intensity) through a 300 μm depth using a step size of 20 μm .

The fluorescence intensity was used to estimate the number of broken monolayers of “filler” network chains following a previously established procedure.^{47,49} Briefly, a calibration curve was used to determine the number density of activated mechanophores, $\phi(x,y,z)$, which was assumed to represent the number density of broken “filler” network bonds. This assumption was thoroughly discussed by Slootman *et al.*,⁴⁷ who evaluated the force-induced fluorescence of pol(methyl acrylate) networks labeled with different mechanophore fractions and subjected to uniaxial deformation or single-edge notch crack propagation at various rates and temperatures. Their results indicated that, within the range of mechanophore concentrations, strain rates, and temperatures considered in this work, the force-induced fluorescence serves as a representative measure of chain breakage and damage (see Appendices 6 and 7 in Slootman *et al.*, ¹H NMR spectrum for calibration molecule in Fig. S2 and representative calibration curve in Fig. S12, ESI†). The calculation was done according to:

$$\phi(x,y,z) = \frac{I(x,y,z) - I_0}{\alpha c_0 \phi_0} \times 100 \quad (6)$$

where $I(x,y,z)$ is the fluorescence intensity within a voxel, I_0 is the fluorescence intensity of the undamaged bulk, recorded far from the fracture surface, α is a linear coefficient relating the fluorescence intensity to the fluorophore concentration, obtained from the calibration curve, c_0 is the mechanophore concentration in the “filler” network, and ϕ_0 is the volume fraction of “filler” network within the multiple-network, as estimated from the pre-stretch ratio, $\phi_0 = \lambda_0^{-3}$.

This density of broken “filler” network bonds was averaged across the imaged plane, xz , to calculate the distribution of broken “filler” network chains within the multiple-networks, $\phi(y,z) = \frac{\int_0^{L_x} \phi(x,y,z) dx}{L_x}$, where $L_x = 880 \mu\text{m}$ is the dimension of the imaged volume along the crack propagation direction, x .

This damage profile, $\phi(y,z)$, was integrated along the penetration depth, $L_y = 300 \mu\text{m}$, to estimate the number of broken “filler” network bonds per unit area of propagated crack, $\Sigma_x(z)$:

$$\Sigma_x(z) = 2\nu_x \int_0^{L_y} \phi(y,z) dy \quad (7)$$

where ν_x is the density of elastically active chains in the “filler” network, calculated from Young’s modulus as $\nu_x = E/3k_B T$, and 2 is a symmetry factor.

Averaging $\Sigma_x(z)$ along the imaged thickness, $L_z = 300 \mu\text{m}$, offers a measure of the number of broken “filler” network bonds per unit area of propagated crack, $\Sigma_x = \frac{\int_0^{L_z} \Sigma_x(z) dz}{L_z}$.

This Σ_x was normalized by the areal density of “filler” network chains, Σ_0 , to determine the number of Lake–Thomas monolayers that break upon fracture, $\bar{\Sigma} = \Sigma_x/\Sigma_0$ (see image processing details in Fig. S13, ESI†). Here, Σ_0 was calculated according to:

$$\Sigma_0 = \frac{1}{2} \nu_x \langle R_0^2 \rangle^{1/2} = l_0 \phi_0^{2/3} \left(\frac{E \rho C_\infty N_A}{6 M_0 k_B T} \right)^{1/2} \quad (8)$$

Where $\langle R_0^2 \rangle^{1/2}$ is the average distance between crosslinks, E is the Young’s modulus of the filler network, l_0 is the length of a C–C bond ($l_0 \approx 0.154 \text{ nm}$), ρ is the density of poly(ethyl acrylate) ($\rho \approx 1.12 \text{ g cm}^{-3}$), C_∞ is the characteristic ratio ($C_\infty \approx 9.3$), N_A is Avogadro’s number, and M_0 is the molecular weight of EA ($M_0 \approx 100.12 \text{ g mol}^{-1}$).

Reactive Monte-Carlo and coarse-grained molecular dynamics simulations

Reactive Monte-Carlo (RMC) and coarse-grained molecular dynamics (CGMD) simulations were conducted following procedures from our recent work and are described in detail in the ESI.†^{38,57}

The polymerization scheme used in the RMC for FRP, RAFT, and ATRP utilized an initiator concentration of 0.2%. This scheme differentiated RAFT and ATRP by varying the termination rate constant (see details in ESI†). The crosslinker concentrations were set to 0.5% (FRP) and 1.0% (RAFT and ATRP). The simulation box contained 106 250 monomers, and the reaction probabilities for initiation, propagation, and termination reactions were based on previously reported values.^{38,57}

The RMC simulations were conducted until 98% vinyl group conversion. The resulting polymer networks were then mapped onto a coarse-grained bead-spring representation, purging unreacted monomers and initiators. The polymer network number density was then maintained at $0.85\sigma^{-3}$.

In the coarse-grained simulations, pairwise interactions between beads were modelled by the repulsive, truncated-shifted Lennard-Jones (LJ) potential:

$$U_{\text{LJ}}(r) = \begin{cases} 4\varepsilon_{\text{LJ}} \left[(\sigma/r)^{12} - (\sigma/r)^6 - (\sigma/r_{\text{cut}})^{12} + (\sigma/r_{\text{cut}})^6 \right] & r \leq r_{\text{cut}} \\ 0 & r > r_{\text{cut}} \end{cases} \quad (9)$$

where $\varepsilon_{\text{LJ}} = 1.0 k_B T$ is the Lennard-Jones interaction strength, k_B is Boltzmann’s constant, $T = 1.0$ is the temperature, $\sigma = 1.0$ is the bead diameter, $r_{\text{cut}} = 2^{1/6} \sigma$ is the cutoff radius, and r is the distance between beads.

The connectivity between beads was modelled using the finite extensible nonlinear elastic (FENE) potential:

$$U_{\text{FENE}}(r) = -\frac{1}{2} k_{\text{spring}} R_{\text{max}}^2 \ln \left(1 - \frac{r^2}{R_{\text{max}}^2} \right) \quad (10)$$

where $k_{\text{spring}} = 30.0 k_B T / \sigma^2$ is the spring constant, and $R_{\text{max}} = 1.5 \sigma^2$ is the maximum bond length.

The rigidity of the strands was described using a bending potential:



$$U_{i,i+1}^{\text{bend}} = k_{\text{B}}TK_{\text{bend}}(1 - (\mathbf{n}_i \cdot \mathbf{n}_{i+1})) \quad (11)$$

where $K_{\text{bend}} = 1.5$ is the bending constant, and $\mathbf{n}_i$ and $\mathbf{n}_{i+1}$ are the bond vectors.

The equilibration of the coarse-grained polymer network was conducted under the canonical ensemble (NVT). Specifically, Langevin dynamics were integrated using the velocity Verlet algorithm⁵⁸ with a timestep of $\Delta t = 0.005\tau$ and a duration of $50\,000\tau$, where τ is the timescale in the coarse-grained simulation described as $\sqrt{m\sigma^2/\epsilon_{\text{LJ}}}$, where $m = 1.0$ is the mass of the coarse-grained bead. After equilibration, a topological analysis was conducted to determine the elastically active chains, explicitly accounting for the Scanlan-Case criterion.⁵⁹

After equilibration, the bending potential was removed, and the FENE potential was replaced with a breakable quartic bond potential, which allows for chain scission during uniaxial deformation:

$$U_{\text{quartic}} = K_{\text{quartic}}(r - R_{\text{c}})^2(r - R_{\text{c}} - B_1)(r - R_{\text{c}} - B_2) + U_0 \quad (12)$$

Where $K_{\text{quartic}} = 2351\epsilon_{\text{LJ}}/\sigma^2$, $R_{\text{c}} = 1.5\sigma$, $B_1 = 0$, $B_2 = -0.7425\sigma$, and $U_0 = 92.74467\epsilon_{\text{LJ}}$. This setup provides the same equilibrium bond length as the FENE potential.⁶⁰ The pairwise potential is the same as in eqn (9), but the cut-off radius is now set to $r_{\text{cut}} = 2.5\sigma$.

After switching the FENE bond potential to the breakable quartic potential, the polymer network was re-equilibrated for 2500τ under the NPT ensemble. The temperature and pressure were set to $T = 1.5$ and $P = 1.0$, using a Nosé–Hoover barostat.⁶¹ Such a choice ensures that the number density of all three systems is maintained around $0.85\sigma^{-3}$ (see Fig. S31, ESI†). Subsequently, uniaxial deformation was performed under the NPT ensemble until the stretch ratio reached at least $\lambda = 21$, with an elongation rate of $5 \times 10^{-5}\tau^{-1}$. Simulation results were averaged over five ensembles with completely different initial configurations for statistical purposes. All coarse-grained molecular dynamics simulations were conducted using LAMMPS (Update 1, 2nd Aug 2023).⁶²

Results and discussion

“Filler” networks synthesized by RDRPs have different molecular and mesoscopic structures than analogues synthesized by FRP

We synthesized three polymer networks by UV-initiated radical copolymerization of ethyl acrylate (EA) monomer and 1,4-butanediol diacrylate crosslinker (BDA), using FRP, RAFT, or ATRP. Detailed synthetic conditions are located in the Materials and methods section and ESI,† but their primary difference lies in the crosslinker concentration: 0.5 mol% for FRP and 1.0 mol% for RAFT and ATRP. These differences reflect variations in the initiation, propagation, and termination rates inherent to each polymerization method but ultimately yield networks with similar small-strain mechanical properties (see detailed discussion in Zhang *et al.*⁵⁷).

These networks were rubbery at room temperature, with a glass transition temperature, $T_{\text{g}} \approx -18^\circ\text{C}$. Moreover, based on

rubber elasticity and Flory–Rehner theories,⁶³ they were indistinguishable, featuring the same elastic modulus, $E \approx 0.89 \pm 0.02$ MPa, and swelling ratio $Q \approx 4.0$, in an athermal solvent like ethyl acrylate. Thus, they had the same volumetric and areal densities of elastically active chains: $\nu_{\text{x}} \approx 7.1 \times 10^{25} \text{ m}^{-3}$ and $\Sigma_0 \approx 2.3 \times 10^{17} \text{ m}^{-2}$.

However, both the structure and optical properties of these networks differed. RAFT- and ATRP-synthesized networks were opaque due to sol–gel demixing during gelation, consisting of crosslinker-rich and crosslinker-poor phases. In addition, the early-stage kinetics of RAFT and ATRP resulted in networks with narrower chain length distributions and higher load-bearing capacities in the crosslinker-rich phase, which formed under higher local crosslinker concentrations and percolated throughout the network (see properties in Table 1, images in Fig. 1A, stress–strain curves in Fig. 1B, and elastic chain distributions estimated from reactive Monte-Carlo simulations in Fig. 1C).

These structural differences were reported in Dookhith *et al.*,³⁸ and, here, we hypothesize that they should impact the fracture properties of multiple-networks.

“Filler” networks synthesized by RAFT and ATRP offer more brittle multiple-networks than analogues synthesized by FRP

Inspired by the seminal work of Gong *et al.* on double-network hydrogels and Ducrot *et al.* on multiple-network elastomers,^{16,17} we interpenetrated these “filler” networks within a soft and loosely crosslinked “matrix” network, synthesized by FRP of EA and BDA (0.01 mol%) using HMP (0.01 mol%) as a photo-initiator (see scheme in Fig. 2A). These multiple-networks were also rubbery at room temperature, with a glass transition temperature, $T_{\text{g}} \approx -18^\circ\text{C}$. They had the same pre-stretch ratio, $\lambda_0 = Q^{-1/3} = 1.6$ (see swelling ratio in Fig. S3, ESI†), and Young’s modulus, $E = 1.30 \pm 0.03$ MPa, but exhibited distinct strain hardening. Specifically, the multiple-networks containing “filler” networks synthesized by RAFT and ATRP strain hardened at $\lambda_{\text{m,RAFT}} \approx \lambda_{\text{m,ATRP}} \approx 2.4 \pm 0.2$, whereas that containing a “filler” network synthesized by FRP did so at $\lambda_{\text{m,FRP}} = 3.1 \pm 0.1$ (see rheological master curves in Fig. 2B, stress–stretch curves in Fig. 2C, and summary of multiple-network properties in Table 2).

In Dookhith *et al.*,³⁸ we noted that these differences in large-strain mechanical properties likely arise from variations in the average extensibility of the “filler” network chains, λ_{h} , in line with the analyses reported by Millereau *et al.* and Kong *et al.* (see Table 1),^{21,64} and molecular and mesoscopic structures. Here, we examine their impact on fracture toughness, as measured by the critical energy release rate, G_{c} .

Table 1 Key properties of “filler” networks synthesized by FRP, RAFT, and ATRP. Young’s modulus, E , swelling ratio in ethyl acrylate, Q , volumetric, ν_{x} , and areal densities of elastic chains, Σ_0 , and average extensibility of the elastic chains, λ_{h}

Mechanism	E [MPa]	Q	ν_{x} [10^{25} m^{-3}]	Σ_0 [10^{17} m^{-2}]	λ_{h}
FRP	0.9	4.0	7.1	2.3	4.9
RAFT	0.9	4.0	7.1	2.3	3.9
ATRP	0.9	4.0	7.1	2.3	3.7



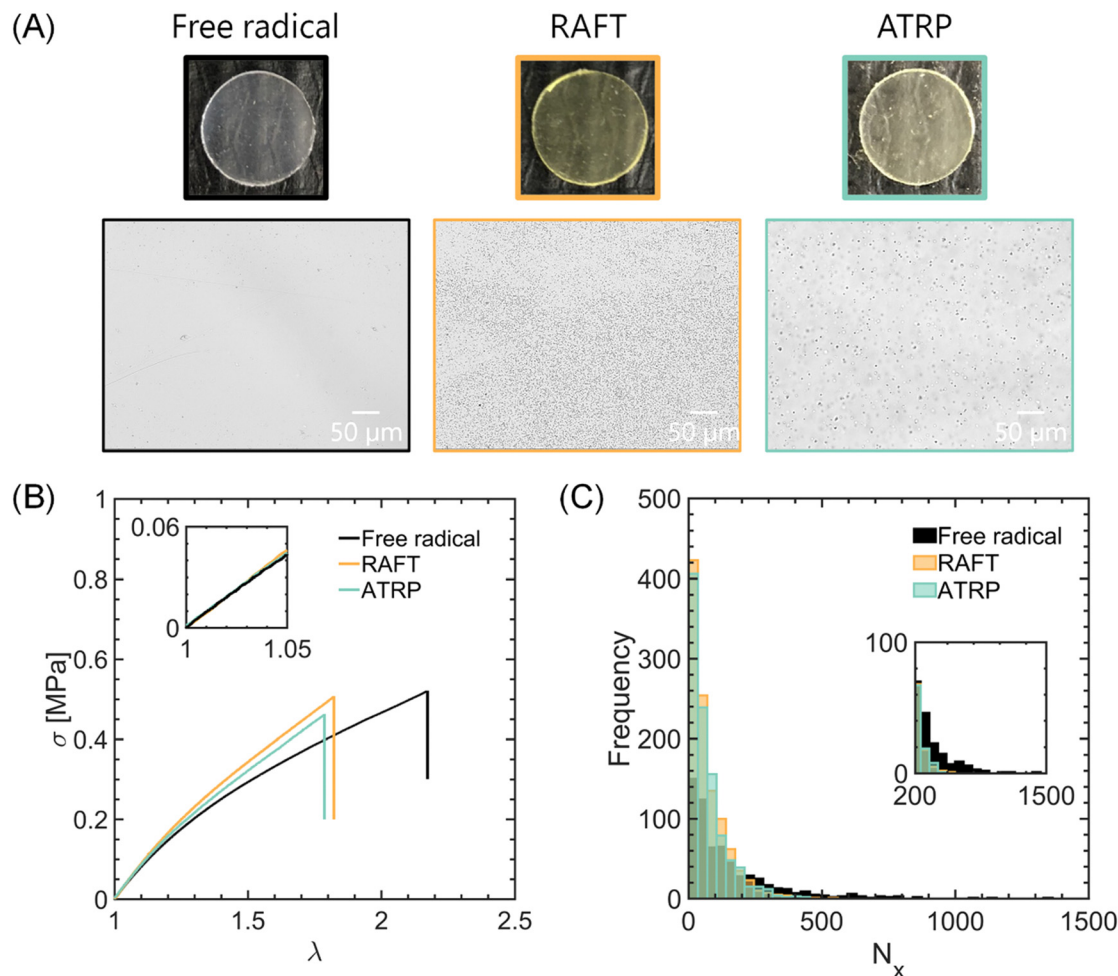


Fig. 1 Properties of “filler” networks: optical, mechanical and molecular (A) filler networks synthesized by RAFT and ATRP appear opaque, suggesting phase-separation into crosslinker-rich and crosslinker-poor phases. (B) These networks, as well as that synthesized by FRP, exhibit similar mechanical properties in the low-deformation regime. Specifically, they have comparable elastic modulus $E \approx 0.9$ MPa, indicating equivalent densities of elastically active chains. (C) However, at the molecular scale, these three networks likely differ in their chain length distributions, with RAFT- and ATRP- synthesized networks having narrower chain length distributions than that synthesized via FRP, based on reactive Monte Carlo simulations (see distributions in a log-log scale in Fig. S29, ESI†). Reprinted (adapted) from Dookhith *et al.*³⁴ Copyright 2024 American Chemical Society.

Remarkably, at a temperature of 23 °C and an initial loading rate of $3 \times 10^{-3} \text{ s}^{-1}$, the multiple-networks based on “filler” networks synthesized by RAFT and ATRP were more brittle than that based on a “filler” network synthesized by FRP, with $G_{\text{c,RAFT}} = 740 \pm 40 \text{ J m}^{-2}$, $G_{\text{c,ATRP}} = 760 \pm 50 \text{ J m}^{-2}$ and $G_{\text{c,FRP}} = 1200 \pm 40 \text{ J m}^{-2}$. At other rates and temperatures, similar differences in fracture behavior were observed despite variations in G_{c} due to viscoelastic dissipation (see Fig. 3, representative stress-stretch curves in Fig. S4 and S7, S8, critical stretch ahead of the crack in Fig. S9, and statistics in Table S2, ESI†). Thus, multiple-networks composed of “filler” networks synthesized by RAFT and ATRP appeared to be less effective at dissipating energy than that with a “filler” network synthesized by FRP, presumably due to less efficient load transfer from the “filler” to the “matrix” network.

To us, these observations were interesting given the similar viscoelastic behavior of the multiple-networks in both linear amplitude oscillatory shear rheology and step-cyclic loading. The storage and loss moduli, G' and G'' , were indistinguishable

over a wide range of reduced frequencies, $a_T\omega$, and the stress-stretch curves showed negligible hysteresis under step-cyclic loads (see rheological master curves in Fig. 2B, stress-stretch curves in Fig. S5 and shift factors in Fig. S6, ESI†). Thus, we consider this family of materials to be both identically and nearly perfectly elastic in bulk, with all differences in dissipation mechanisms (*i.e.*, viscoelasticity and damage) confined to the large-strain region ahead of a crack – similar linear rheology but different nonlinear response.

“Filler” networks synthesized by RAFT and ATRP lead to a different interplay between viscoelasticity and damage ahead of cracks

Having examined the impact of “filler” network architecture on the fracture toughness, G_{c} , we now focus on its effect on the load transfer efficiency and energy dissipation mechanisms ahead of cracks. This efficiency was evaluated by labelling the “filler” networks with fluorogenic mechanophores and



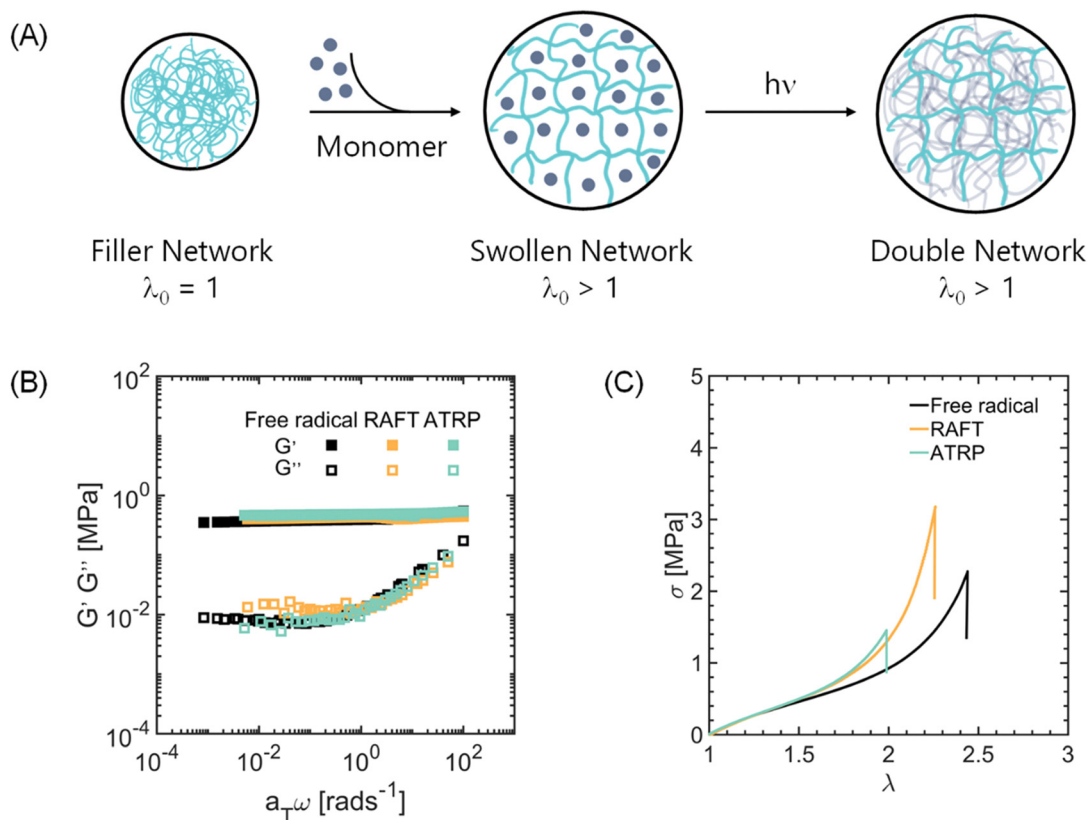


Fig. 2 Small and large-strain mechanical properties of multiple-networks. (A) Filler networks swollen in a solution of monomer, crosslinker, and initiator consist of chains pre-stretched to a stretch ratio, λ_0 . Subsequent free radical polymerization kinetically arrests these networks in their pre-stretched state, resulting in multiple-networks. (B) The linear rheology of these networks is indistinguishable, featuring similar storage, G' , and loss moduli, G'' , over a wide frequency range. (C) However, the tensile properties differ at large strains, with the multiple-networks derived from filler networks synthesized via RAFT and ATRP strain-hardening at a lower stretch ratio than that derived from the filler network synthesized via FRP.

Table 2 Key properties of multiple networks with “filler” networks synthesized by FRP, RAFT, and ATRP. Young’s modulus, E , pre-stretch ratio, λ_0 , onset of strain hardening, λ_m , and fracture toughness, G_c , at 23 °C and $3 \times 10^{-3} \text{ s}^{-1}$

“Filler” network	E [MPa]	λ_0	λ_m	G_c [J m $^{-2}$]
FRP	1.3	1.6	3.1	1200
RAFT	1.3	1.6	2.3	740
ATRP	1.3	1.6	2.4	760

imaging the regions next to the fracture surfaces with a confocal microscope. Details on the mechanophore synthesis and quantification are provided in the Materials and methods section, as well as in Göstl *et al.*,⁴⁵ Sloodman *et al.*,^{23,47} and Sanoja *et al.*,⁴⁹ but, briefly, we estimated the (i) number of activated mechanophores per unit area of propagated crack, Σ_x , from a calibration curve and the Young’s modulus, E , of the “filler” network; and (ii) the number of broken “filler” network monolayers per unit area of propagated crack, $\bar{\Sigma}$, from the prediction of the Lake and Thomas model, Σ_0 .⁶⁵ This number of activated mechanophores was considered representative of the number of broken “filler” network bonds, in line with Sloodman *et al.* (see a detailed discussion of this assumption in

Appendices 6 and 7 of Sloodman *et al.*, as well as in the recent theoretical work of Dubach *et al.*).^{47,66}

Consistent with the measured G_c , the multiple-networks composed of “filler” networks synthesized by RAFT, ATRP, and FRP experienced different extents of damage during fracture, $\bar{\Sigma}$. For example, at a loading rate of $3 \times 10^{-3} \text{ s}^{-1}$ and temperature of 23 °C, the multiple-networks with “filler” networks synthesized by RAFT and ATRP evolved larger, more delocalized damage zones than that with a “filler” network synthesized by FRP, with $\bar{\Sigma}_{\text{RAFT}} = 280 \pm 10$, $\bar{\Sigma}_{\text{ATRP}} = 320 \pm 20$, and $\bar{\Sigma}_{\text{FRP}} = 250 \pm 10$. Additionally, at lower loading rates and higher temperatures, all multiple-networks exhibited less, more localized damage likely due to reduced, large-strain viscoelastic dissipation ahead of cracks (see Fig. 4, representative confocal images and damage profiles in Fig. S14–S17 (ESI[†]), and estimates of damage delocalization length scale in Fig. S18 and S19, ESI[†]). Therefore, the multiple-networks composed of “filler” networks synthesized by RAFT and ATRP incurred more damage than that composed of a “filler” network synthesized by FRP, despite being more brittle.

To us, this observation was particularly interesting because the fracture toughness, G_c , often increases with the damage-zone size, $\bar{\Sigma}$, in fracture mechanics. Specifically, the fracture toughness is typically described by the equation $G_c = G_0$



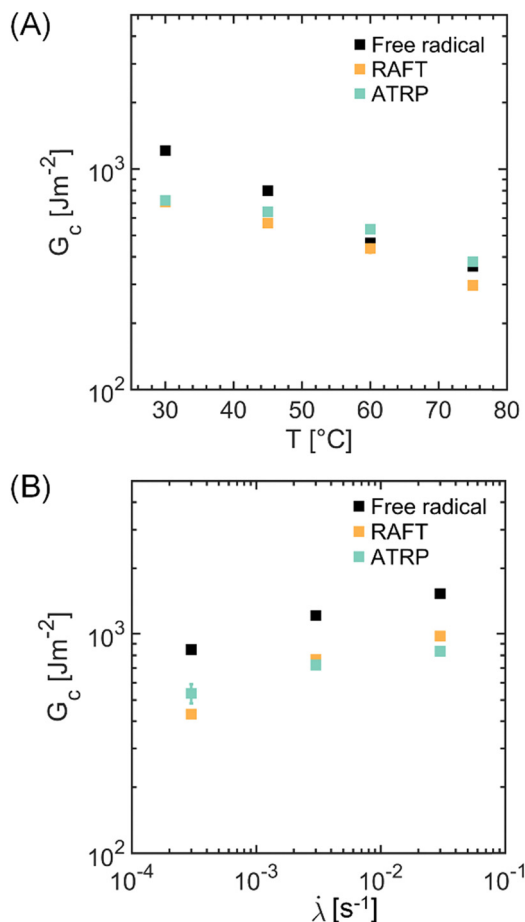


Fig. 3 Rate- and temperature-dependent fracture of multiple-networks. The fracture toughness, G_c , (A) decreases with temperature and (B) increases with loading rate due to viscoelastic dissipation ahead of the crack. Under most loading conditions, filler networks synthesized by RAFT and ATRP offer more brittle multiple-networks than that produced through free radical polymerization (FRP).

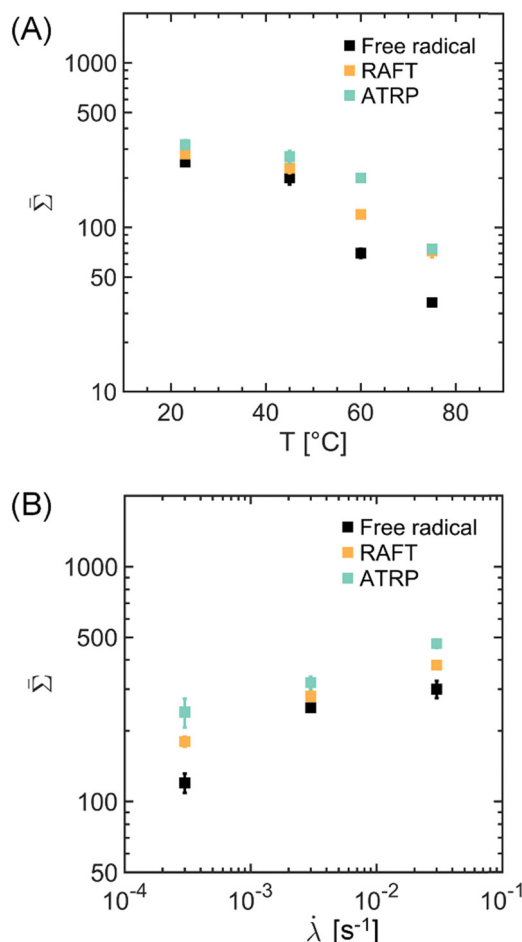


Fig. 4 Rate- and temperature-dependent damage of multiple-networks. The damage due to filler networks scission, Σ (A) decreases with temperature and (B) increases with rate due to viscoelastic dissipation ahead of the crack. This observation indicates that dissipation mechanisms – viscoelasticity and damage – synergistically operate ahead of the crack. Under most loading conditions, filler networks synthesized by RAFT and ATRP suffer more damage compared to that made by free radical polymerization (FRP).

$(1 + f(a_T v))$, where G_0 is the intrinsic fracture toughness and a function of damage, and $f(a_T v)$ is a viscoelastic toughness enhancement that depends on the reduced crack propagation velocity, $a_T v$.^{67–71} Within this framework, the intrinsic fracture energy, G_0 , is defined as the energy required to break a monolayer of elastically active chains across the crack plane. According to Lake and Thomas,⁶⁵ this energy is given by $G_0 = N_x U_b \Sigma_0$, where N_x is the average elastic chain length, U_b is the energy of a C–C bond, and Σ_0 is the areal density of elastically active chains along the fracture plane (see crack velocities in Table S3 (ESI†) and related analyses of fracture toughness and damage in Fig. S20 and S21, ESI†).

However, our results challenge this description, revealing a larger damage zone, in terms of broken “filler” network chains, for the more brittle multiple-networks, as well as a dependence of the damage zone on both rate and temperature. They indicate that the total energy dissipated per broken “filler” network bond, G_c/Σ_x , depends on the method used to synthesize the “filler” network, irrespective of the loading rate and temperature. For instance, at a temperature of 23 °C and a rate of

$3 \times 10^{-3} \text{ s}^{-1}$, $G_c/\Sigma_{x,\text{RAFT}} = 1.2 \pm 0.3 \times 10^{-17} \text{ J per chain}$, $G_c/\Sigma_{x,\text{ATRP}} = 0.9 \pm 0.5 \times 10^{-17} \text{ J per chain}$, and $G_c/\Sigma_{x,\text{FRP}} = 2.4 \pm 0.6 \times 10^{-17} \text{ J per chain}$. In comparison, at 75 °C and the same loading rate of $3 \times 10^{-3} \text{ s}^{-1}$, $G_c/\Sigma_{x,\text{RAFT}} = 0.9 \pm 0.6 \times 10^{-17} \text{ J per chain}$, $G_c/\Sigma_{x,\text{ATRP}} = 0.8 \pm 0.5 \times 10^{-17} \text{ J per chain}$, and $G_c/\Sigma_{x,\text{FRP}} = 2.6 \pm 1.0 \times 10^{-17} \text{ J per chain}$ (see energy dissipated per filler chain under different fracture conditions in Fig. S27, ESI†).

From “filler” network synthesis to multiple-network fracture: a molecular picture

Based on these results, we pose a molecular picture of fracture, which closely aligns with that of Slootman *et al.*²³ and is inspired by the damage models of Brown and Tanaka.^{18,19} In this picture, fracture results from (i) random and (ii) correlated scission of “filler” network bonds, followed by (iii) breakage of the “matrix” network; that is, from load transfer from the “filler” to the “matrix” network and the associated energy dissipation. In their seminal work, Slootman *et al.*



demonstrated that the “filler” network pre-stretch, λ_0 , is critical in this load transfer efficiency, controlling the formation of a stable damage zone, energy dissipation, and fracture.²³ Here, we focus on the influence of “filler” network architecture, as dictated by the gelation method, on the fracture toughness.

Let us first examine the impact of “filler” network architecture on the size of the damage zone, Σ_x , which, in damage models, is typically governed by the effective stretch within the “filler” network, $\lambda\lambda_0$, relative to the maximum extensibility of the “filler” network chains, λ_h .^{72–74} This ratio measures the probability of breaking a chain within the “filler” network and was evaluated at the fracture point, $\lambda_c\lambda_0/\lambda_h$, which depends on both rate and temperature because of viscoelastic dissipation (see variation in λ_c with rate and temperature in Fig. S9 (ESI†) and changes in toughness and damage with λ_c in Fig. S21, ESI†).

In line with damage models, our results indicate that the probability of breaking a “filler” network bond controls the size of the damage zone, Σ_x , revealing a master curve between Σ_x and $\lambda_c\lambda_0/\lambda_h$, which scales as $\Sigma_x \sim \exp(\beta\lambda_c\lambda_0/\lambda_h)$, with $\beta = 6.9 \pm 1.5$ (see Fig. 5). This scaling exponent is similar to that reported by Sloodman *et al.* for another family of multiple-networks composed of a “filler” network synthesized by FRP and having a modulus of $E = 1$ MPa, $\beta = 6.3 \pm 1.2$ (see Fig. S26, ESI†).²³

This result highlights two key points. First, the coupling between viscoelasticity and damage, as measured by β , is independent of the “filler” network architecture. All multiple-networks consist of poly(ethyl acrylate) chains, presumably featuring the same friction coefficient and intermolecular interactions when elongated from their undeformed configuration to their contour length. Second, “filler” networks synthesized by RAFT and ATRP experience more damage than those synthesized by FRP when fractured at the same effective stretch,

$\lambda_c\lambda_0$; likely due to the reduced average extensibility of the “filler” network chains, λ_h , resulting from the controlled gelation methods and their impact on the architecture.

With this picture in mind, we can now discuss the impact of “filler” network structure on the fracture toughness of multiple-networks, G_c . Unlike the size of the damage zone, Σ_x , which depends on the fracture stretch of the “filler” network relative to its average contour length, this fracture toughness, G_c , depends only on the fracture stretch of the “filler” network, $\lambda_c\lambda_0$, and collapses onto a master curve described by $G_c \sim \exp(\gamma\lambda_c\lambda_0)$, with $\gamma = 1.5 \pm 0.4$ (see Fig. 6). This curve suggests that viscoelasticity ultimately governs energy dissipation, dictating the fracture behavior of multiple-networks. Namely, multiple-networks with distinct “filler” network architectures exhibit similar fracture toughness as long as their filler network chains experience a similar fracture stretch, $\lambda_c\lambda_0$. Reducing the average extensibility of the “filler” network chains through an RDRP method, such as RAFT and ATRP, thus compromises both (i) the energy dissipated as the “filler” network chains extend from their relaxed configuration to their fracture length (*i.e.*, from λ_0 to $\lambda_0\lambda_c$, which dictates viscoelasticity), as well as (ii) the energy released as the “filler” network chains progressively break as they approach their contour length (*i.e.*, from λ_0/λ_h to $\lambda_0\lambda_c/\lambda_h$, which governs damage).

The impact of RAFT and ATRP on “filler” network architecture and multiple-network fracture is then two-fold. First, RAFT and ATRP lower the average extensibility of the “filler” network chains, resulting in higher chain breakage probabilities at the crack front and in multiple-networks with larger damage zones. Second, RAFT and ATRP lead to “filler” networks with limited large-strain viscoelastic dissipation ahead of the crack, ultimately promoting multiple-network fracture. Thus, RAFT- and

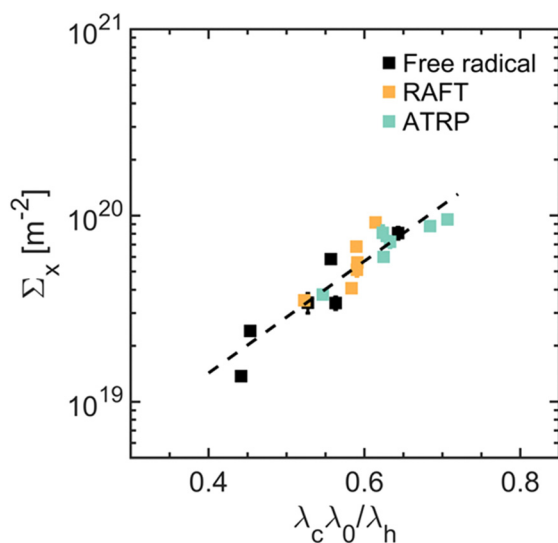


Fig. 5 Interplay between damage and filler network stretch during fracture. The filler network stretch, $\lambda_c\lambda_0$, relative to the limiting extensibility of the filler network chains, λ_h , governs the probability of incurring damage, Σ_x . As such, the data collapses onto a log-lin master curve, with slope $\beta \approx 6.9 \pm 1.5$.

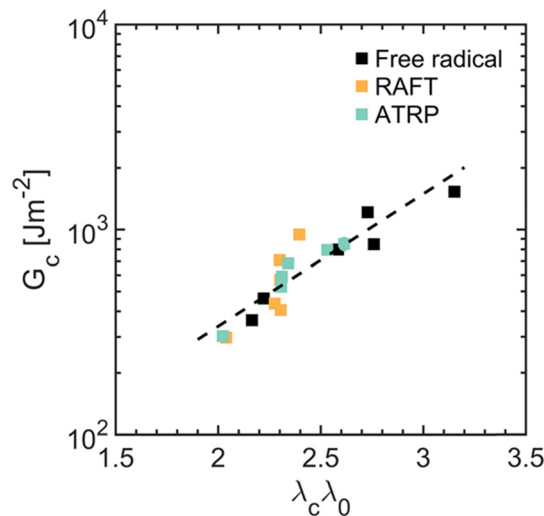


Fig. 6 Relationship between toughness and filler network stretch during fracture. The filler network stretch, $\lambda_c\lambda_0$, dictates the energy dissipated through molecular friction as the chains elongate from their Gaussian configuration to their contour length; that is, the fracture toughness. Therefore, the data collapses onto a log-lin master curve with slope, $\gamma \approx 1.5 \pm 0.4$.



ATRP-synthesized “filler” networks localize stress and transition from random to correlated chain breakage at lower deformations than that synthesized by FRP, compromising energy dissipation and fracture toughness.

Coarse-grained molecular dynamics simulations

Having painted this molecular picture of fracture, we now address whether variations in the “filler” network structure at the molecular or mesoscopic length scales have a greater impact on the toughness. Ideally, to isolate the impact of these variations, we would synthesize networks with uniform mesoscopic structures (*i.e.*, without crosslinker-rich and crosslinker-poor mesophases) using FRP, ATRP, and RAFT. However, synthesizing such networks proved challenging; all networks synthesized by RAFT and ATRP turned opaque due to sol-gel demixing at the late stages of gelation.

Nevertheless, our observations suggest that variations in chain length distributions within the load-bearing phase of the “filler” network play a more significant role than the mere presence of crosslinker-rich and crosslinker-poor mesophases. After all, RAFT- and ATRP-synthesized networks differ significantly in light transmittance ($T \approx 67\%$ for “filler” network synthesized by RAFT and $T \approx 19\%$ for the “filler” network synthesized by ATRP) but exhibit a similar relationship between viscoelasticity, damage, and fracture toughness (see images in Fig. 1A, master curves in Fig. 5 and 6, and visualization of phase separation in filler and multiple networks in Fig. S28, ESI†). Furthermore, the crosslinker-rich phase within the “filler” network, which forms during the early, reaction-controlled stages of gelation, should carry most of the load; both because it percolates and constitutes the majority phase, and because it consists, on average, of shorter chains than the crosslinker-poor phase.

To support this picture, we generated three networks by reactive Monte-Carlo simulations and deformed them in uniaxial tension through coarse-grained molecular dynamics simulations. As in experiments, these networks had similar elastic chain densities but different chain length distributions due to reversible radical deactivation during gelation (see Fig. 1C). Simulation details are provided in the Materials and methods section, as well as in Dookhith *et al.* and Zhang *et al.*,^{38,57} but briefly, the reactive Monte-Carlo simulations generated networks through a three-dimensional bond fluctuation model (3DBFM), while the coarse-grained molecular dynamics offered stress-strain curves and damage evolution estimates under uniaxial deformation, leveraging a quartic bond potential to quantify chain breakage. Importantly, these simulations focused exclusively on the crosslinker-rich phase of the “filler” networks, neglecting both the matrix network and any coexisting phases within the architecture. As discussed earlier, the crosslinker-rich phase forms during the early, reaction-controlled stages of gelation and should carry most of the load within the “filler” network.

Interestingly, our simulation results show tensile curves consistent with networks having similar elastic chain densities but distinct chain length distributions. These networks exhibited indistinguishable Young's moduli but different strain hardening and failure (*i.e.*, maximum stress) points (see Fig. 7A and

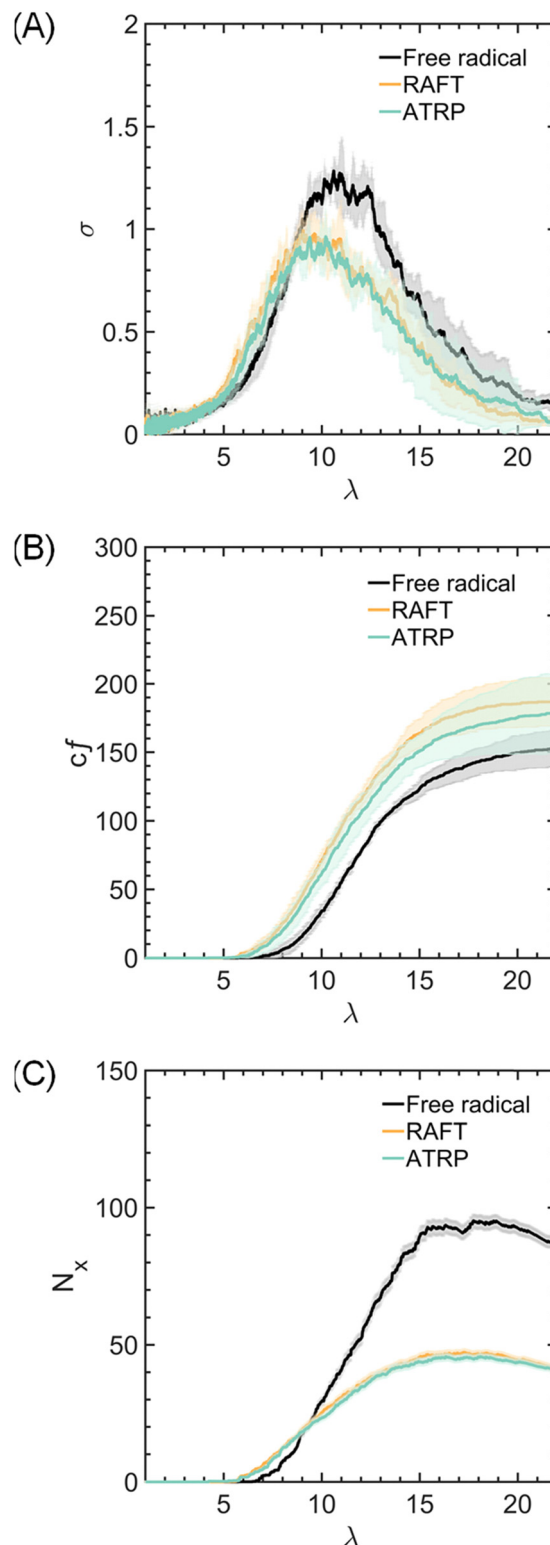
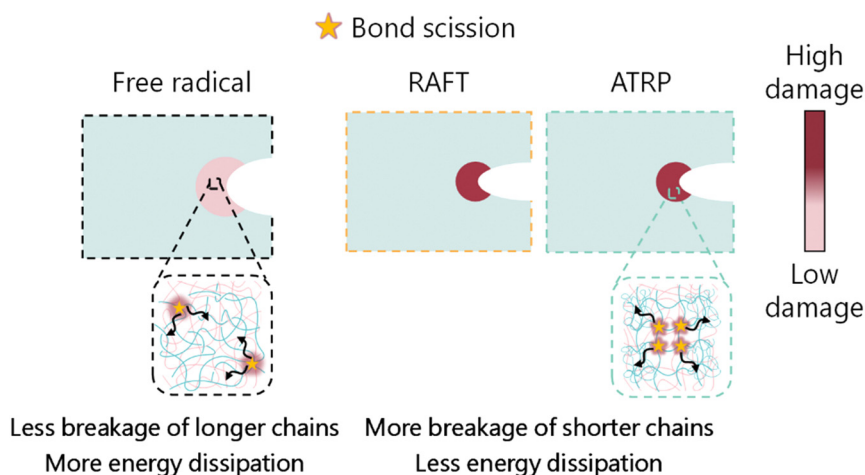


Fig. 7 Coarse-grained molecular dynamics simulations comparing the damage evolution of networks synthesized by RAFT, ATRP, and FRP in tension. (A) Stress–stretch curves show that, while all families of networks have similar moduli, the networks synthesized by RAFT and ATRP break at a lower stretch than that synthesized by FRP. Thus, (B) these networks suffer (*i.e.*, accumulate) more chain scission events prior to fracture, (C) which, on average, involve shorter chains.





Scheme 1 Networks synthesized by RAFT and ATRP have shorter chain lengths, fracture at a lower stretch, and suffer more bond scission (damage) than those synthesized by FRP. Thus, they offer multiple-networks that dissipate less energy during fracture and are therefore more brittle.

Fig. S32, ESI†). Additionally, networks synthesized by RAFT and ATRP start to damage at lower strains than those synthesized by FRP (see Fig. 7B), suffering more chain breakage events, particularly among shorter chains, when subjected to a similar stretch. Lastly, the simulations unveil a mismatch between the number of chain breakage events and the associated energy dissipation, with networks synthesized by RAFT and ATRP dissipating less energy per chain elongation and breakage event than that synthesized by FRP, rendering them more brittle. This mismatch likely underpins the differences in fracture toughness between the multiple-networks. RAFT- and ATRP-synthesized “filler” networks may experience more chain breakage events ahead of cracks but do not necessarily dissipate more energy. Since their broken chains are, on average, shorter, they are unable to store and dissipate as much energy as that synthesized by FRP, in line with the Lake and Thomas model (see the cumulative average length of the broken chains in Fig. 7C).⁶⁵

The picture that emerges from these coarse-grained MD simulations is thus qualitatively similar to that posed through mechanochemistry experiments. RAFT and ATRP offer networks with narrower chain length distributions within the crosslinker-rich, load-bearing phase, which, on average, have a limited ability to dissipate and store energy. These networks suffer chain breakage events at lower strains than those synthesized by FRP, experiencing an accelerated transition from random to correlated chain scission, rapidly localizing stress (*i.e.*, inefficiently transferring it from the “filler” to the “matrix” network), and readily undergoing crack nucleation and fracture (see estimates of this transition in Fig. S33 and S34, ESI†). Therefore, at the same elastic chain density, ν_x , RAFT and ATRP seemingly afford more brittle networks than conventional FRP (see Scheme 1).

Concluding remarks

We investigated the impact of “filler” network architecture on the fracture toughness of multiple-networks, leveraging recent

advances in RDRPs and mechanochemistry to synthesize three families of polymer networks (i) labelled with fluorogenic mechanophores and (ii) having similar elastic chain densities, ν_x , but different chain length distributions and mesoscopic structures.

Using mechanical testing, confocal microscopy, reactive Monte-Carlo, and coarse-grained molecular dynamics simulations, we showed that these networks offered multiple-networks with distinct load transfer efficiency, energy dissipation, and fracture toughness. Specifically, “filler” networks synthesized by RAFT and ATRP resulted in more brittle multiple-networks than that made by FRP due to the reduced average extensibility of their elastic chains, which lowers both the energy stored per chain in its fully elongated state and the energy dissipated through viscoelastic chain friction as the chains stretch from their relaxed, Gaussian configuration to their contour length.

This result has important implications for both polymer science and fracture mechanics, bridging the gelation kinetics with the network architecture and fracture toughness. First, it reveals an inherent trade-off between strain hardening and fracture toughness, which arises from the gelation method and its impact on the elastic chains density and chain length distribution. Networks with narrower chain lengths, particularly within the load-bearing phase, may strain harden at lower strains but are more susceptible to damage and fracture. Second, it highlights differences in the damage zone in terms of the number of chain breakage events and the energy dissipated upon such breakage. Ahead of a crack, numerous short chains may break, but these events do not necessarily dissipate much energy and afford toughness. Finally, it highlights challenges raised by RAFT, ATRP, and other living polymerization methods (*e.g.*, ring-opening polymerizations^{75–77}) in manufacturing soft and tough elastomers (*i.e.*, with elastic modulus and fracture toughness on the order of 1 MPa and 1000 J m^{−2}).^{78,79} Relative to FRP, these techniques offer slower gelation rates and opaque and embrittled networks, particularly at high temperatures.

Overall, this result underscores the need to develop new techniques that offer high gelation rates and controlled



molecular architectures, both to advance understanding of the interplay between gelation, structure, and mechanical properties and to guide the molecular design of advanced networks for emerging applications.

Further, it highlights two aspects that require future work. First, the generality of our results, since we focused on three “filler” networks synthesized *via* FRP, ATRP, and RAFT under very restricted crosslinker and initiator concentrations. Second, their validity under other conditions of minimal viscoelastic dissipation, such as the high-water concentrations typical of hydrogels. Addressing these points could deepen our understanding of the fundamental synthesis–structure–property relationships governing the mechanical properties of polymer networks.

Author contributions

Conceptualization (A. Z. D. and G. E. S.), data curation (A. Z. D. and Z. Z.), data analysis (A. Z. D., Z. Z., V. G., and G. E. S.), funding acquisitions (V. G., and G. E. S.), investigation (A. Z. D., and Z. Z.), methodology (A. Z. D., Z. Z., V. G., and G. E. S.), resources (V. G., and G. E. S.), supervision (V. G., and G. E. S.), validation (A. Z. D., Z. Z., and G. E. S.), visualization (A. Z. D. and G. E. S.), writing – original draft (A. Z. D.), writing – review and editing (A. Z. D., Z. Z., V. G. and G. E. S.).

Data availability

Data for this article, including experimental data, reactive Monte Carlo simulations, and coarse-grained molecular dynamics simulations are available at the Texas Data Repository at <https://doi.org/10.18738/T8/OVB8P1>.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

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Notes and references

- 1 A. N. Gent, in *Engineering with Rubber*, ed. A. N. B. T.-E. Gent and R. Gent, Carl Hanser Verlag GmbH & Co. KG, München, 3rd edn, 2012, pp. I–XVIII.
- 2 *Hydrogels in Medicine and Pharmacy*, ed. N. A. Peppas, CRC Press, 2019.
- 3 C. Creton, *MRS Bull.*, 2003, **28**, 434–439.
- 4 C. Creton and M. Ciccotti, *Rep. Prog. Phys.*, 2016, **79**, 046601.
- 5 C. Creton, *Macromolecules*, 2017, **50**, 8297–8316.
- 6 R. Long, C.-Y. Hui, J. P. Gong and E. Bouchbinder, *Annu. Rev. Condens. Matter Phys.*, 2021, **12**, 71–94.
- 7 G. M. Geise, D. R. Paul and B. D. Freeman, *Prog. Polym. Sci.*, 2014, **39**, 1–42.
- 8 X. Zhao, X. Chen, H. Yuk, S. Lin, X. Liu and G. Parada, *Chem. Rev.*, 2021, **121**, 4309–4372.
- 9 D. T. Hallinan and N. P. Balsara, *Annu. Rev. Mater. Res.*, 2013, **43**, 503–525.
- 10 H. J. Kim, L. Paquin, C. W. Barney, S. So, B. Chen, Z. Suo, A. J. Crosby and R. C. Hayward, *Adv. Mater.*, 2020, **32**, 1–7.
- 11 S. K. Kumar, B. C. Benicewicz, R. A. Vaia and K. I. Winey, *Macromolecules*, 2017, **50**, 714–731.
- 12 Y. Chen, A. Kovalenko, A. Brûlet, B. Bresson, A. Lantheaume, L. Olanier and C. Creton, *Macromolecules*, 2023, **56**, 5336–5345.
- 13 H. Zhang, A. K. Scholz, J. de Crevoisier, F. Vion-Loisel, G. Besnard, A. Hexemer, H. R. Brown, E. J. Kramer and C. Creton, *Macromolecules*, 2012, **45**, 1529–1543.
- 14 H. Zhang, A. K. Scholz, F. Vion-Loisel, Y. Merckel, M. Brieu, H. Brown, S. Roux, E. J. Kramer and C. Creton, *Macromolecules*, 2013, **46**, 900–913.
- 15 J. A. Gohl, T. J. Wiley, H.-C. Chang, C.-C. Chang and C. S. Davis, *Front. Soft Matter*, 2023, **3**, 1–10.
- 16 J. P. Gong, Y. Katsuyama, T. Kurokawa and Y. Osada, *Adv. Mater.*, 2003, **15**, 1155–1158.
- 17 E. Ducrot, Y. Chen, M. Bulters, R. P. Sijbesma and C. Creton, *Science*, 2014, **344**, 186–189.
- 18 H. R. Brown, *Macromolecules*, 2007, **40**, 3815–3818.
- 19 Y. Tanaka, *Europhys. Lett.*, 2007, **78**, 56005.
- 20 J. P. Gong, *Soft Matter*, 2010, **6**, 2583.
- 21 P. Millereau, E. Ducrot, J. M. Clough, M. E. Wiseman, H. R. Brown, R. P. Sijbesma and C. Creton, *Proc. Natl. Acad. Sci. U. S. A.*, 2018, **115**, 9110–9115.
- 22 Y. Chen, G. Sanoja and C. Creton, *Chem. Sci.*, 2021, **12**, 11098–11108.
- 23 J. Sloodman, C. J. Yeh, P. Millereau, J. Comtet and C. Creton, *Proc. Natl. Acad. Sci. U. S. A.*, 2022, **119**, 1–11.
- 24 H. Gao and K. Matyjaszewski, *Prog. Polym. Sci.*, 2009, **34**, 317–350.
- 25 J. Cuthbert, A. C. Balazs, T. Kowalewski and K. Matyjaszewski, *Trends Chem.*, 2020, **2**, 341–353.
- 26 N. Ide and T. Fukuda, *Macromolecules*, 1997, **30**, 4268–4271.
- 27 N. Ide and T. Fukuda, *Macromolecules*, 1999, **32**, 95–99.
- 28 I. Bannister, N. C. Billingham, S. P. Armes, S. P. Rannard and P. Findlay, *Macromolecules*, 2006, **39**, 7483–7492.
- 29 C.-D. Vo, J. Rosselgong, S. P. Armes and N. C. Billingham, *Macromolecules*, 2007, **40**, 7119–7125.
- 30 H. Gao, K. Min and K. Matyjaszewski, *Macromolecules*, 2007, **40**, 7763–7770.
- 31 H. Gao, W. Li and K. Matyjaszewski, *Macromolecules*, 2008, **41**, 2335–2340.
- 32 H. Gao, A. Miasnikova and K. Matyjaszewski, *Macromolecules*, 2008, **41**, 7843–7849.
- 33 H. Gao, P. Polanowski and K. Matyjaszewski, *Macromolecules*, 2009, **42**, 5925–5932.
- 34 E. Goldbach, X. Allonas, L. Halbardier, C. Ley and C. Croutxé-Barghorn, *Eur. Polym. J.*, 2023, **197**, 112335.



- 35 E. Goldbach, X. Allonas, C. Croutxé-Barghorn, C. Ley, L. Halbardier and G. L'Hostis, *Eur. Polym. J.*, 2023, **188**, 111947.
- 36 A. Beziau, A. Fortney, L. Fu, C. Nishiura, H. Wang, J. Cuthbert, E. Gottlieb, A. C. Balazs, T. Kowalewski and K. Matyjaszewski, *Polymer*, 2017, **126**, 224–230.
- 37 J. Cuthbert, S. V. Wanasinghe, K. Matyjaszewski and D. Konkolewicz, *Macromolecules*, 2021, **54**, 8331–8340.
- 38 A. Z. Dookhith, Z. Zhang, V. Ganesan and G. E. Sanoja, *Macromolecules*, 2024, **57**, 8698–8711.
- 39 A. L. Black, J. M. Lenhardt and S. L. Craig, *J. Mater. Chem.*, 2011, **21**, 1655–1663.
- 40 J. Li, C. Nagamani and J. S. Moore, *Acc. Chem. Res.*, 2015, **48**, 2181–2190.
- 41 R. Göstl, J. M. Clough and R. P. Sijbesma, in *Mechanochemistry in Materials*, ed. Y. C. Simon and S. L. Craig, The Royal Society of Chemistry, 2018, pp. 53–75.
- 42 N. Deneke, M. L. Rencheck and C. S. Davis, *Soft Matter*, 2020, **16**, 6230–6252.
- 43 Y. Chen, G. Mellot, D. van Luijk, C. Creton and R. P. Sijbesma, *Chem. Soc. Rev.*, 2021, **50**, 4100–4140.
- 44 G. E. Sanoja and C. Creton, *Annu. Rev. Chem. Biomol. Eng.*, 2025, DOI: [10.1146/annurev-chembioeng-092220-113154](https://doi.org/10.1146/annurev-chembioeng-092220-113154).
- 45 R. Göstl and R. P. Sijbesma, *Chem. Sci.*, 2016, **7**, 370–375.
- 46 M. Stratigaki, C. Baumann, L. C. A. van Breemen, J. P. A. Heuts, R. P. Sijbesma and R. Göstl, *Polym. Chem.*, 2020, **11**, 358–366.
- 47 J. Sloodman, V. Waltz, C. J. Yeh, C. Baumann, R. Göstl, J. Comtet and C. Creton, *Phys. Rev. X*, 2020, **10**, 041045.
- 48 S. He, M. Stratigaki, S. P. Centeno, A. Dreuw and R. Göstl, *Chem. – Eur. J.*, 2021, **27**, 15889–15897.
- 49 G. E. Sanoja, X. P. Morelle, J. Comtet, C. J. Yeh, M. Ciccotti and C. Creton, *Sci. Adv.*, 2021, **7**, 1–23.
- 50 X. P. Morelle, G. E. Sanoja, S. Castagnet and C. Creton, *Soft Matter*, 2021, **17**, 4266–4274.
- 51 J. Ju, G. E. Sanoja, M. Y. Nagazi, L. Cipelletti, Z. Liu, C. Y. Hui, M. Ciccotti, T. Narita and C. Creton, *Phys. Rev. X*, 2023, **13**, 021030.
- 52 J. Ju, G. E. Sanoja, L. Cipelletti, M. Ciccotti, B. Zhu, T. Narita, C. Yuen Hui and C. Creton, *Proc. Natl. Acad. Sci. U. S. A.*, 2024, **121**, 2017.
- 53 Y. Kawauchi, Y. Tanaka, H. Furukawa, T. Kurokawa, T. Nakajima, Y. Osada and J. P. Gong, *J. Phys.:Conf. Ser.*, 2009, **184**, 012016.
- 54 E. Ducrot, H. Montes and C. Creton, *Macromolecules*, 2015, **48**, 7945–7952.
- 55 E. Ducrot and C. Creton, *Adv. Funct. Mater.*, 2016, **26**, 2482–2492.
- 56 H. W. Greensmith, *J. Appl. Polym. Sci.*, 1963, **7**, 993–1002.
- 57 Z. Zhang, J. Krajniak, A. Z. Dookhith, Y. Tian, H. S. Sachar, N. Marion, T. J. Duncan, J. Liu, G. E. Sanoja and V. Ganesan, *Macromolecules*, 2025, **58**, 3168–3187.
- 58 W. C. Swope, H. C. Andersen, P. H. Berens and K. R. Wilson, *J. Chem. Phys.*, 1982, **76**, 637–649.
- 59 J. Scanlan, *J. Polym. Sci.*, 1960, **43**, 501–508.
- 60 T. Ge, F. Pierce, D. Perahia, G. S. Grest and M. O. Robbins, *Phys. Rev. Lett.*, 2013, **110**, 098301.
- 61 D. J. Evans and B. L. Holian, *J. Chem. Phys.*, 1985, **83**, 4069–4074.
- 62 A. P. Thompson, H. M. Aktulga, R. Berger, D. S. Bolintineanu, W. M. Brown, P. S. Crozier, P. J. in't Veld, A. Kohlmeyer, S. G. Moore, T. D. Nguyen, R. Shan, M. J. Stevens, J. Tranchida, C. Trott and S. J. Plimpton, *Comput. Phys. Commun.*, 2022, **271**, 108171.
- 63 P. J. Flory, *Principles of Polymer Chemistry*, Cornell University Press, New York, 1953.
- 64 V. A. Kong, T. A. Staunton and J. E. Laaser, *Macromolecules*, 2024, **57**, 4670–4679.
- 65 G. J. Lake and A. G. Thomas, *Proc. R. Soc. A*, 1967, **300**, 108–119.
- 66 F. F. C. Dubach, W. G. Ellenbroek and C. Storm, *J. Polym. Sci.*, 2021, **59**, 1188–1199.
- 67 A. N. Gent, *Langmuir*, 1996, **12**, 4492–4496.
- 68 B. N. J. Persson and E. A. Brener, *Phys. Rev. E:Stat., Non-linear, Soft Matter Phys.*, 2005, **71**, 036123.
- 69 C.-Y. Hui, B. Zhu and R. Long, *J. Mech. Phys. Solids*, 2022, **159**, 104748.
- 70 B. N. J. Persson, G. Carbone, C. Creton, G. Heinrich and T. Tada, *Extreme Mech. Lett.*, 2024, **68**, 102143.
- 71 S.-Q. Wang, Z. Fan, A. Siavoshani, M. Wang and J. Wang, *Extreme Mech. Lett.*, 2025, **74**, 102277.
- 72 M. Bacca, C. Creton and R. M. McMeeking, *J. Appl. Mech.*, 2017, **84**, 1–7.
- 73 F. J. Vernerey, R. Brighenti, R. Long and T. Shen, *Macromolecules*, 2018, **51**, 6609–6622.
- 74 S. R. Lavoie, P. Millereau, C. Creton, R. Long and T. Tang, *J. Mech. Phys. Solids*, 2019, **125**, 523–549.
- 75 A. Z. Dookhith, N. A. Lynd, C. Creton and G. E. Sanoja, *Macromolecules*, 2022, **55**, 5601–5609.
- 76 A. Z. Dookhith, N. A. Lynd and G. E. Sanoja, *Macromolecules*, 2023, **56**, 40–48.
- 77 M. Yasir, B. Hu, T.-C. Lin and K. Matyjaszewski, *Langmuir*, 2025, **41**, 378–382.
- 78 A. Bagheri, C. M. Fellows and C. Boyer, *Adv. Sci.*, 2021, **8**, 2003701.
- 79 V. A. Bobrin, J. Zhang, N. Corrigan and C. Boyer, *Adv. Mater. Technol.*, 2023, **8**, 2201054.

