

Cite this: *Chem. Sci.*, 2025, 16, 19777

All publication charges for this article have been paid for by the Royal Society of Chemistry

Aryl azopyrroles as visible light photoswitchable TRPA1 ligands

Lisa C. Dollhopf,^{†a} Jordan A. Munos,^{†b} Kai Y. Zheng,^c Rui Xin Tao,^a Peter R. Haycock,^a Philip J. Parsons,^{†a} Randall T. Peterson,^d Pui-Ying Lam^{*b} and Matthew J. Fuchter^{†ac}

Access to high quality optochemogenetic tools will enable precise interrogation of the complex biology of diverse cell types, allowing temporally and spatially controlled manipulation of cell autonomous functions. This level of control is expected to yield deeper mechanistic insights and foster the development of targeted strategies and molecular interventions for translational photopharmacology. Recent years have demonstrated the exciting potential of heteroaromatic azoarene photoswitches, particularly to tune azo photoswitch performance. However, strategies to integrate such motifs into photopharmacological ligands remain underdeveloped. Leveraging a previously developed screening assay, we report the discovery of a robust aryl azopyrrole ligand for the TRPA1 channel, TRPswitch-C. TRPswitch-C provides a red-shifted chemotype to address this channel, allowing optical control in live zebrafish larvae in a controllable fashion. Not only does this study expand the tool kit for optochemogenetic experiments using TRPA1, but also advocates the further exploration of heteroaromatic azoarenes in photopharmacology.

Received 8th July 2025
Accepted 15th September 2025

DOI: 10.1039/d5sc05070g

rsc.li/chemical-science

Introduction

Photopharmacology using molecular photoswitches represents a powerful way to endow bioactive ligands with reversible light addressability. By using light to interconvert the structural states of the photoswitch, the bioactivity of the corresponding photopharmacological ligand can be modulated as a function of the wavelength.^{1–5} The most common photoswitch for photopharmacology, by far, is azobenzene. However, recent studies have shown that superior photoswitch performance can be achieved using heteroaromatic azo switches.^{6,7} Despite this advance, heteroaromatic azo photoswitches remain relatively underexplored in photopharmacology.⁸

The discovery of photopharmacological ligands mostly relies on modification of a known non-photoswitchable ligand, using one of two complementary approaches called “azologization” and “azo extension”.^{1,9} The former seeks to replace elements of the ligand core structure with an isosteric azo photoswitch,⁹ while the latter adds an azo photoswitch to the periphery of the

ligand. While these are powerful and effective ways to develop photopharmaceuticals, they are reliant on preexisting non-switchable ligands. *De novo* discovery of photopharmacological ligands remains poorly developed. In a rare example, we previously screened a curated library of photoswitches in a phenotypic zebrafish assay and discovered the first photoswitchable ligands of the TRPA1 channel, TRPswitch-A and B.¹⁰ A more recent example of *de novo* discovery of a different photoswitch scaffold, Plinabulin and analogues as tubulin-binding hemipiperazine switches, has been reported by Kirchner *et al.*¹¹ Beyond validation of screening as an effective means for photopharmacological ligand discovery, this work resulted in the identification of light-addressable tool molecules that have further enabled interrogation of TRPA1 biology¹² and hold potential in other chemo-optogenetic experiments in large neurons, such as those in the nucleus gigantocellularis,¹³ where a large depolarization current is needed.

A notable omission from our initial screening library was heteroaromatic azo photoswitches; our reported azylazopyrazole ligand TRPswitch-B was a subsequently optimized version of our initial azobenzene screening hit, TRPswitch-A.¹⁰ Here we use our assay to identify an aryl azopyrrole ligand of TRPA1, which represents a distinct chemotype over TRPswitch-A/B. This ligand can be robustly addressed in live zebrafish larvae using visible light and represents the first azopyrrole used in photopharmacology. We believe this work further advocates new discovery methodologies for photopharmacology and the strengthens the case for heteroaromatic azo photoswitches in this area.

^aMolecular Sciences Research Hub, Department of Chemistry, Imperial College London, Wood Lane, London W12 0BZ, UK

^bDepartment of Cell Biology, Neurobiology and Anatomy, Medical College of Wisconsin, 53226 Milwaukee, WI, USA. E-mail: plam@mcw.edu

^cDepartment of Chemistry, University of Oxford, Chemical Research Laboratory, 12 Mansfield Road, Oxford, OX1 3TA, UK. E-mail: matthew.fuchter@chem.ox.ac.uk

^dDepartment of Pharmacology and Toxicology, University of Utah, Salt Lake City, UT, USA

† L. D. and J. M. contributed equally.



Results and discussion

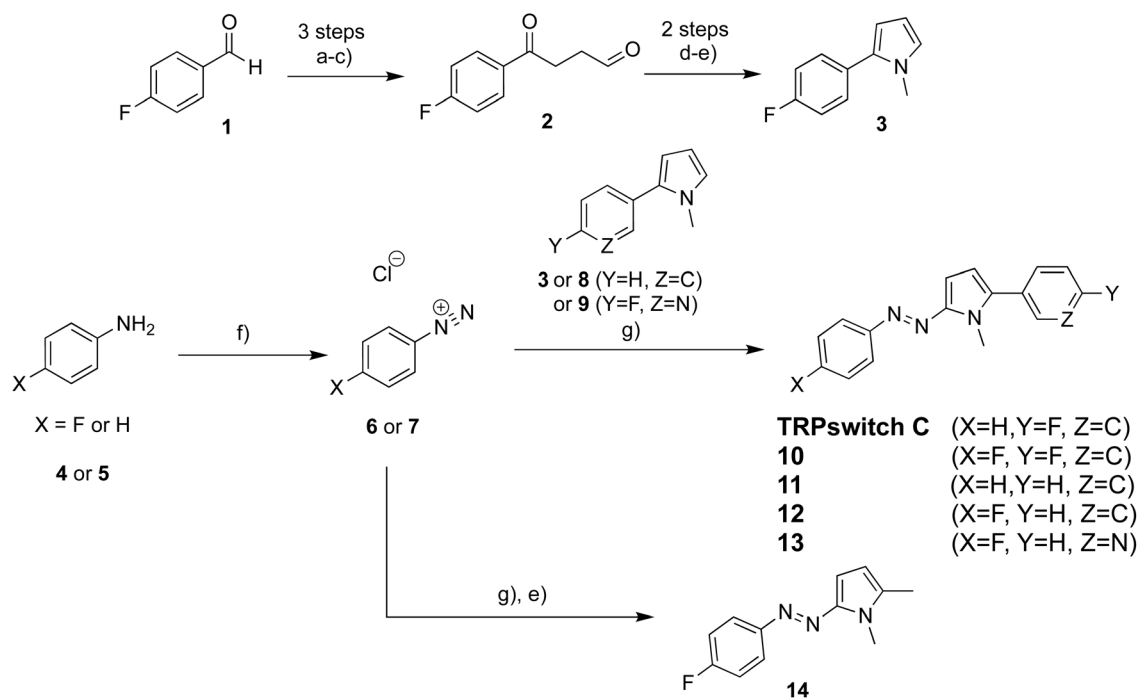
By screening small libraries of heteroaromatic switches, accessed through our previous structure–property studies,^{14–16} and using our previously reported light induced motion response assay,¹⁰ we identified aryl azopyrroles as a promising chemotype to control movement in zebrafish larvae. The electron-rich pyrrole ring results in red-shifting of the absorption bands for such switches, which in principle allows for visible light addressability.^{14,15} However, aryl azopyrroles can be prone to rapid thermal *Z*–*E* isomerization, especially in the presence of water,¹⁵ which has limited their wider use in photoswitch applications.¹⁷ Nonetheless, **TRPswitch-C**, which is an effective modulator of TRPA1 (see below), was also found to be a robust aryl azopyrrole photoswitch, with an extended thermal half-life for the *Z* isomer.

All azopyrrole photoswitches were synthesized *via* diazonium coupling between an aryl diazonium salt and a corresponding pyrrole (Scheme 1). Intermediate **2** was synthesized according to a known literature procedure.¹⁸ Briefly, 4-fluorobenzaldehyde **1** was reacted with allyl magnesium chloride, followed by a subsequent hydroboration-oxidation and Swern oxidation. This resulted in the desired aldehyde **2** in moderate yield (33% over 3 steps). Key pyrrole intermediate **3** was obtained through condensation of **2** with ammonium acetate, followed by *N*-alkylation of the generated pyrrole with NaH and iodomethane in THF (61% yield). Anilines **4** and **5** were first diazotized, followed by coupling with the respective pyrroles under basic conditions in an EtOH/water mixture. The final

products **TRPswitch-C** and **10–13** were obtained in moderate yields (8–28% over 2 steps). Final compound **14** was obtained after subsequent *N*-methylation with NaH and iodomethane in THF.

The UV/vis spectrum of **TRPswitch-C** is shown in Fig. 1A. **TRPswitch-C** showed a strong π – π^* absorption band with a characteristic shoulder that was observed for all studied azopyrrole photoswitches. Irradiation with UV (340 and 365 nm) and visible light (405 and 450 nm) resulted in good *E* to *Z* isomerization of up to 82% in DMSO at room temperature. The absorption of these aryl azopyrrole switches are significantly red-shifted (*Z* isomer $\lambda_{\text{max}} = 414$ nm) in comparison to previously reported arylazopyrazole based TRPswitches.¹⁰ Due to the broad absorption bands, relatively poor *Z* to *E* photoswitching was observed (39% *E* isomer after irradiation with 525 nm light). A half-life of 70 minutes in DMSO was obtained from thermal isomerization kinetics experiments that measured the absorption at 400 nm of a previously irradiated *Z*-isomer rich sample (Fig. 1B).

We assessed our aryl azopyrrole compounds in our previously reported motion response assay. In short, three 3 day post fertilization (dpf) wildtype (WT) zebrafish (*zf*) larvae were placed in a well of a 96 well plate and exposed to test compounds (20 μM) at 28 °C for 1 h in the dark prior to the experiment. At 3 dpf, *zf* larvae do not respond to a light stimulus, thus any light-induced motion response observed would be due to compound treatment. DMSO (1%) was used as a negative control for all experiments. Each well was individually stimulated with three 1 s pulses of stimulation light (Fig. 2A).



Scheme 1 Reagents and conditions. (a) Allyl magnesium chloride, THF, 0 °C to room temperature, 3 h. (b) 2 M $\text{BH}_3 \cdot \text{DMS}$ in THF, then aq. NaOH and H_2O_2 , 0 °C to room temperature, 5 h. (c) $(\text{COCl})_2$, DMSO, NEt_3 , DCM, –78 °C to room temperature, 1.5 h, 33% over 3 steps. (d) NH_4OAc , EtOH, reflux, 1.5 h, 85%. (e) Iodomethane, NaH, THF, 0 °C to 60 °C, 1.5 h, 40–61%. (f) NaNO_2 , 1 M HCl, MeOH, 0 °C, 30 min; then (g) pyrroles, NaOAc, EtOH, water, 0 °C, 1 h, 8–28%.



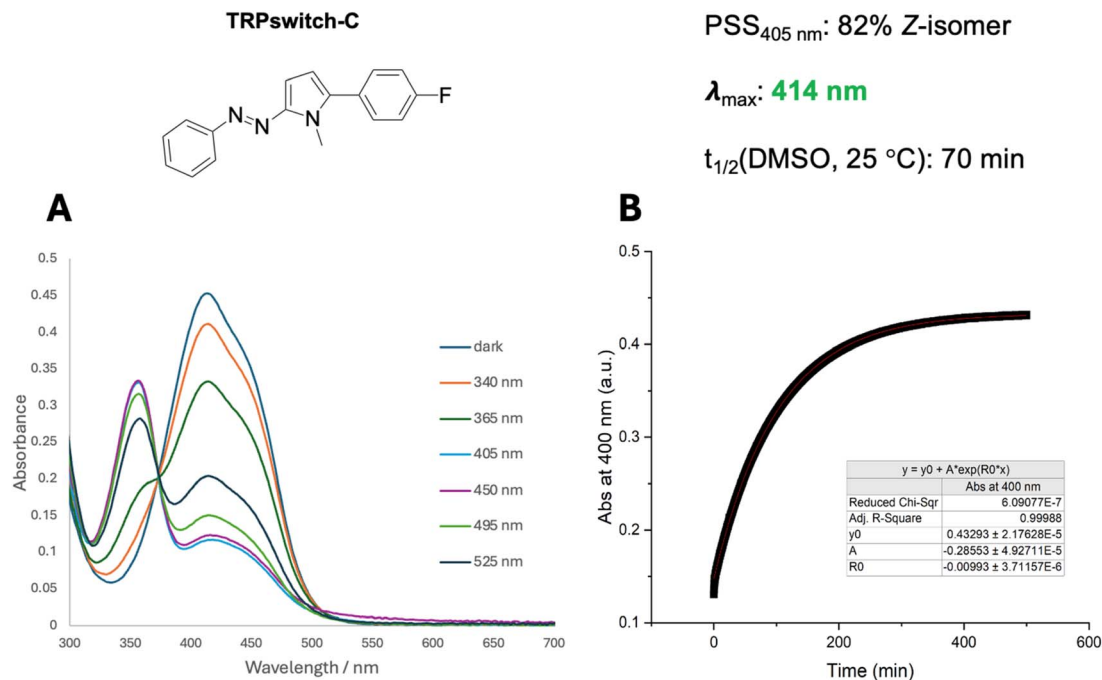


Fig. 1 Photochemical characterization of TRPswitch-C. (A) UV-vis spectra of TRPswitch-C in DMSO (50 μM) at 25 $^{\circ}\text{C}$ were recorded before and after irradiation with LEDs of different wavelengths for 3 minutes. PSS ratios were determined according to the method described by Fischer.¹⁹ (B) Thermal isomerization kinetics of TRPswitch-C in DMSO (50 μM) at 25 $^{\circ}\text{C}$. The absorbance at 400 nm was fitted to an exponential fit to determine the rate constant and thermal half-life of Z-E isomerization.

Throughout the light stimulation sequence, images were acquired without delay. The readout of the assay is a light induced motion response. We quantified the motion before and after light illumination with a custom script developed using NIS Elements GA3 software, thus allowing us to compare DMSO controls to compound treated wells (Fig. 2B). TRPswitch-C caused a robust motion response produced upon each light illumination when stimulated with violet light (Fig. 2C and D and SI Movie 1). The light induced motion response was not observed in larvae incubated in DMSO.

We sought to briefly survey the structure property relationships that were important for the activity of TRPswitch-C, particularly the importance of fluorine substitution and the need for an aromatic ring attach to the pyrrole unit. As such, we generated a series of derivatives (compounds 10–14) and characterized them using our motion response assay. Compound 10 contains a fluorine group in the para position on either benzene ring (Fig. 3) and undergoes E/Z isomerization most efficiently with 440 nm light. We observed reduced photoswitching and biological activity when 390 nm light was used for stimulation with compound 10. When 3 dpf WT zf were exposed to compound 10 there was an increase in motion after 440 nm light stimulation (Fig. 3) compared to DMSO controls, but less than that observed for TRPswitch-C. When both fluorines were removed from the compound (compound 11) the optimal wavelength of photoactivation becomes 390 nm. The motion response of 11 is similar to 10. When the F is removed from the para position on the right side of the compound (compound 12), the motion response is, again, similar to that of compounds

10 and 11. As such, it seems this scaffold is modest in its sensitivity to fluorine substitution, with TRPswitch-C having the optimum combination of those analogues we tested. To examine if different substitutions of the ring were tolerated, compounds 13 and 14 were synthesized. Compound 13 replaced the benzene ring with a pyridine ring while compound 14 removed the ring altogether. A lack of motion was observed with both 13 and 14 (Fig. 3) suggesting the benzene ring attached to the pyrrole to be critical for motion response in our assay.

Two types of photoactivatable molecules have been previously reported that activate Trpa1b in zebrafish, Optovin,²⁰ which is irreversible, and TRPswitch-A/B,¹⁰ which are reversible. In 3 dpf larval zebrafish, Trpa1b is normally expressed in Rohon-Beard neurons²¹ and its activation causes a characteristic motion response that is highly reproducible.^{10,20,22} A similar motion response was observed in zf larvae exposed to TRPswitch-C and light stimulation in comparison to the response of these prior compounds. To test whether Trpa1b was the target of TRPswitch-C, 3 dpf WT, *trpa1b*^{-/-}, or *trpv1*^{-/-} mutant larvae were exposed to DMSO (1%) or TRPswitch-C (20 μM) and stimulated with 390 nm light. The motion response to TRPswitch-C was completely abolished in *trpa1b*^{-/-} mutant larvae (Fig. 4A and SI Movie 2). However, when *trpv1*^{-/-} mutants were exposed to TRPswitch-C, these larvae exhibited a similar motion response as WT larvae (Fig. 4A). These results suggest that the zTrpa1b channel is required for TRPswitch-C light-induced motion response.

To test the cross-reactivity of TRPswitch-C with other Trpa1 orthologs, we transiently expressed zebrafish Trpa1b (zTrpa1b)



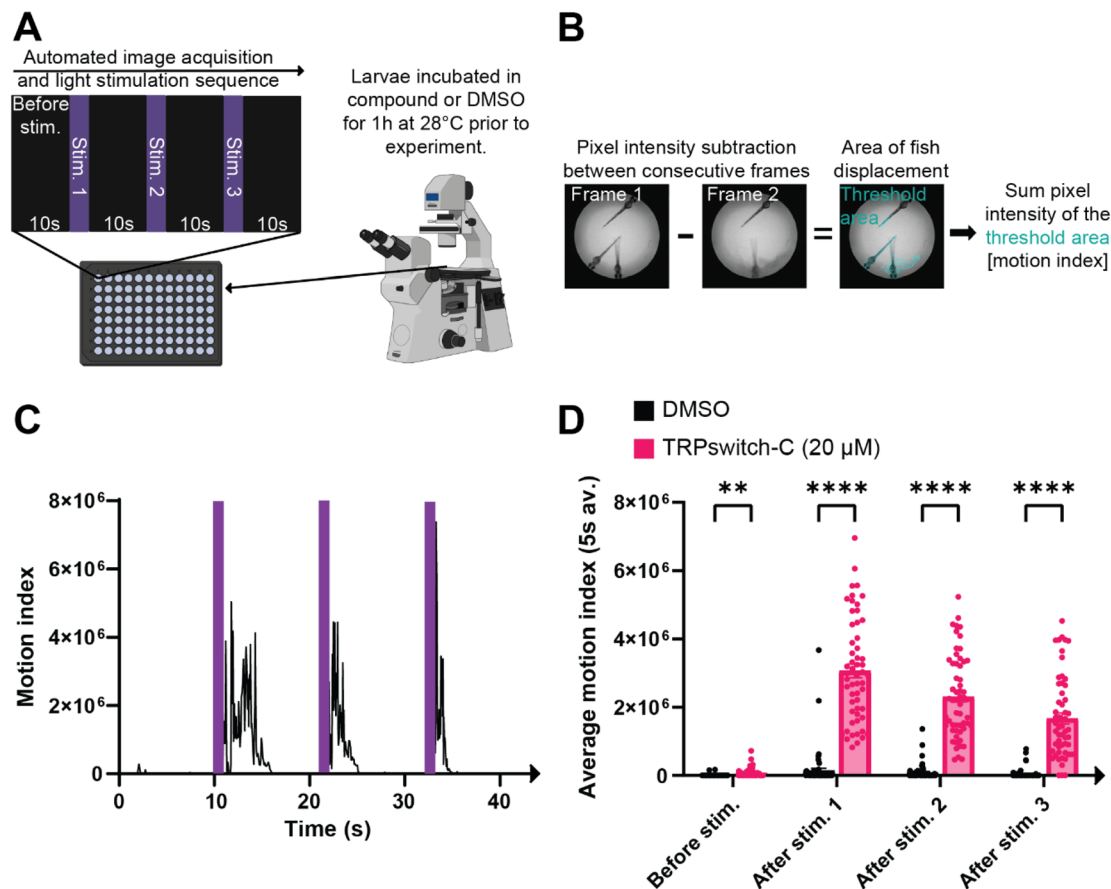


Fig. 2 Properties of TRPswitch-C. (A) Schematic of the experimental setup for the light induced motion response assay. Compounds were screened in a 96-well plate using 3 day post fertilization (dpf) wild-type zebrafish (zf) larvae. Three 3dpf zf were placed into 1 well and compounds were added. A motorized inverted compound microscope and an automated light sequence were used. Hit compounds have a motion response after light stimulation when compared to DMSO controls. (B) Quantification of the motion response (see methods section for details). (C) Representative line graph of the zf behavioral response when exposed to 20 μM TRPswitch-C. Violet bars indicate when stimulation light is on (duration 1 s). (D) Quantification of the behavioral response of fish exposed to TRPswitch-C or DMSO before and after light stimulation. Each data point is a single well. Values are mean \pm SEM. (p -value ** < 0.0021; **** < 0.0001) see SI Movie 1.

or human TRPA1 (hTRPA1) in Rohon Beard neurons of *trpa1b*^{-/-} mutant fish. Expression of zTrpa1b in *trpa1b*^{-/-} mutant fish restored the light induced motion response in fish exposed to TRPswitch-C upon 390 nm light activation (Fig. 4B). Conversely, expression of hTRPA1 in *trpa1b*^{-/-} mutant fish did not restore the light induced motion response in the presence of TRPswitch-C (Fig. 4B). These results suggest that the activity of TRPswitch-C is specific to the zebrafish Trpa1b and does not react with the human TRPA1 ortholog. This is also consistent with our previously reported compounds TRPswitch-A/B, despite the significantly different chemotype of TRPswitch-C. Zebrafish Trpa1b and human TRPA1 share 46% amino acid sequence identity. The differences in amino acid sequences, which are distributed across various domains of TRPA1, may account for the specificity of TRPswitches towards zTrpa1b.

Given the visible light addressability of aryl azopyrrole switches, we surveyed the light-induced motion response of TRPswitch-C at longer wavelengths. We found that TRPswitch-C can produce a significant light-induced motion response when stimulated with 440 nm and 475 nm light (Fig. 5A and SI Movie

3). No motion response was observed for 510 nm stimulation (data not shown), which likely relates to the lower absorbance and lower conversion to the active Z isomer at this wavelength (see Fig. 1). At longer wavelengths such as 475 nm the activity of TRPswitch-C increases with longer stimulation times (Fig. 5B), likely due to the lower absorbance at this wavelength. Together, this suggests that a range of wavelengths and irradiation times can be used for controlling the function of zTrpa1b when using TRPswitch-C.

To demonstrate the reversible and repeatable activity of the Trpa1b/TRPswitch-C system, we utilized a previously established heartbeat interruption assay.¹⁰ This assay involves the use of 3 dpf zebrafish larvae from the transgenic line *Tg(cmlc2:Trpa1b-2A-EGFP)* in *trpa1b*^{-/-} mutants, in which Trpa1b is expressed exclusively in the cardiomyocyte of the larvae. Zebrafish larvae incubated in either DMSO or TRPswitch-C were then subjected to repeated cycles of activation and deactivation light. Since calcium is essential for heart contractions, the heartbeat serves as a biological readout of this assay—and by proxy, for the activation and deactivation of the



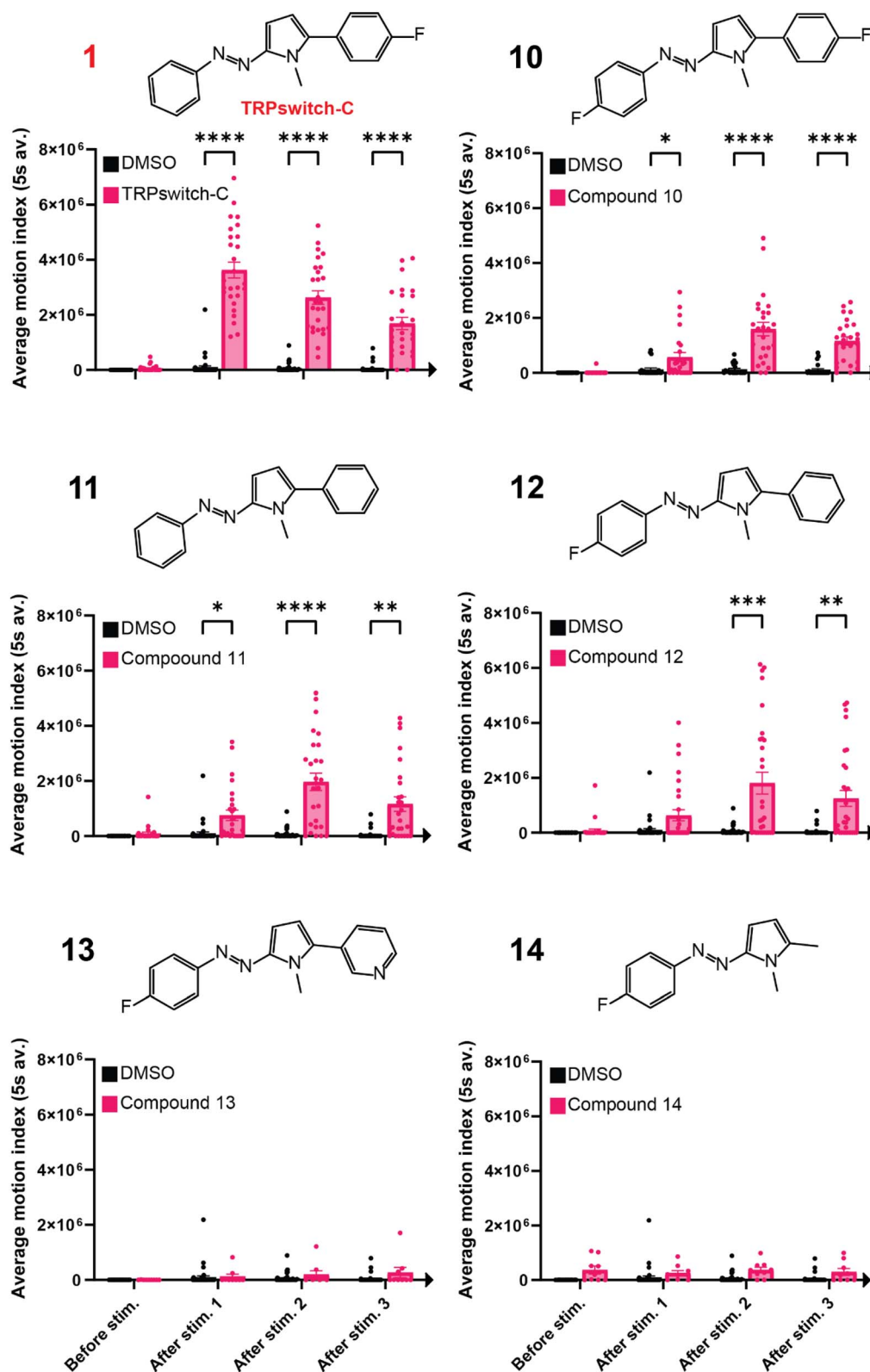


Fig. 3 TRPswitch-C structure-activity relationship analysis. Structures for each compound are on top of the corresponding graph of the light induced motion assay result. 390 nm light was used for stimulation for all compounds except compound 10, which used 440 nm for stimulation. Data points are individual wells. Values are mean \pm SEM. (p -value * < 0.0332; ** < 0.0021; *** < 0.0002; **** < 0.0001).



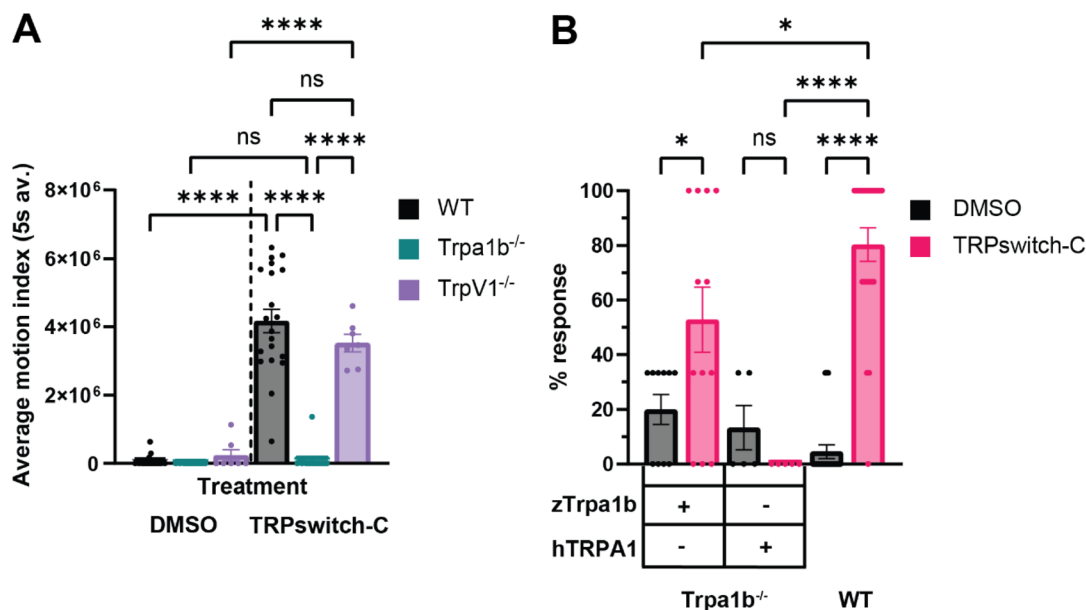


Fig. 4 The activity of TRPswitch-C is specific for zebrafish *Trpa1b*. (A) Light induced motion response assay was performed with TRPswitch-C on WT, *trpa1b*^{-/-} mutant or *trpV1*^{-/-} mutant larvae. Values are mean \pm SEM. (p -value **** $<$ 0.0001) see SI Movie 2. (B) Cross reactivity testing of TRPswitch-C on zebrafish *Trpa1b* ohnolog. Light induced motion response assays were performed on 2 dpf WT, *trpa1b*^{-/-} mutants or *trpa1b*^{-/-} mutants with transient mosaic expression of either zTrpa1b or hTRPA1 in Rohon Beard neurons. The percentage of larvae showing motion response upon light activation with either DMSO or 20 μ M TRPswitch-C treatment was quantified. Each data point is a single larva. Values are mean \pm SEM. (p -value ns \geq 0.1234, * $<$ 0.0332 or **** $<$ 0.0001).

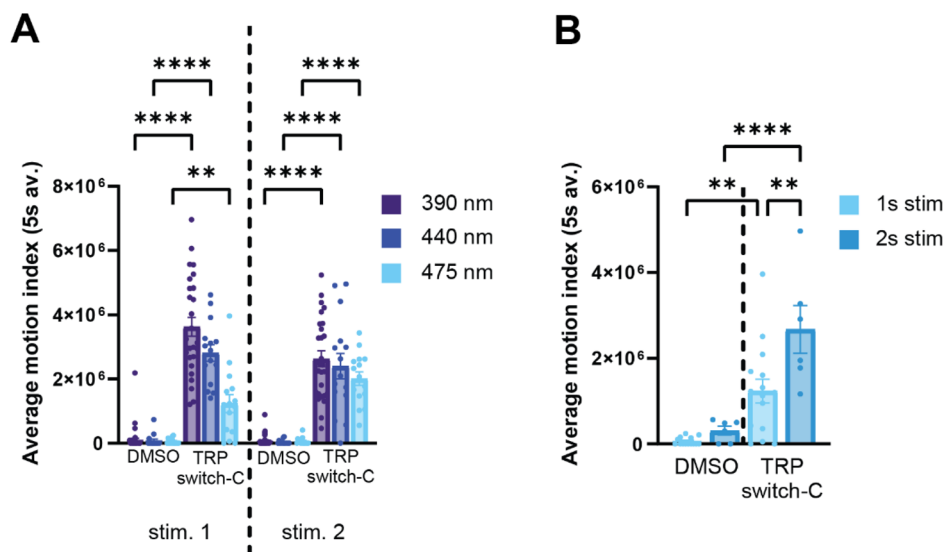


Fig. 5 Comparing the light-induced activities of TRPswitch-C at different wavelengths. (A) Light-induced motion response assay was performed with TRPswitch-C on WT larvae at 390 nm, 440 nm or 475 nm. Values are mean \pm SEM. (p -value ** $<$ 0.0021; **** $<$ 0.0001) see SI Movie 3. (B) Light-induced motion response assay was performed with TRPswitch-C on WT larvae at 475 nm for 1 s or 2 s stimulation. Values are mean \pm SEM. (p -value ** $<$ 0.0021; **** $<$ 0.0001) see SI Movie 4.

Trpa1b channel (Fig. 6A). We found that when incubated in TRPswitch-C, activation with 390 nm light resulted in a stopping of the heartbeat, a response not observed in the DMSO controls (Fig. 6B and C). Subsequent illumination with deactivating 555 nm light was sufficient to restart the heartbeat (Fig. 6C and D). Importantly, repeated activation and

deactivation cycles led to consistent stopping and restarting of the heartbeat (Fig. 6C and D), although with a progressive decrease in heart rate compared to the original baseline (Fig. 6D). Our results demonstrate that the *Trpa1b* channel can be repeatedly activated and deactivated with 390 nm and 555 nm light, respectively, in the presence of TRPswitch-C.



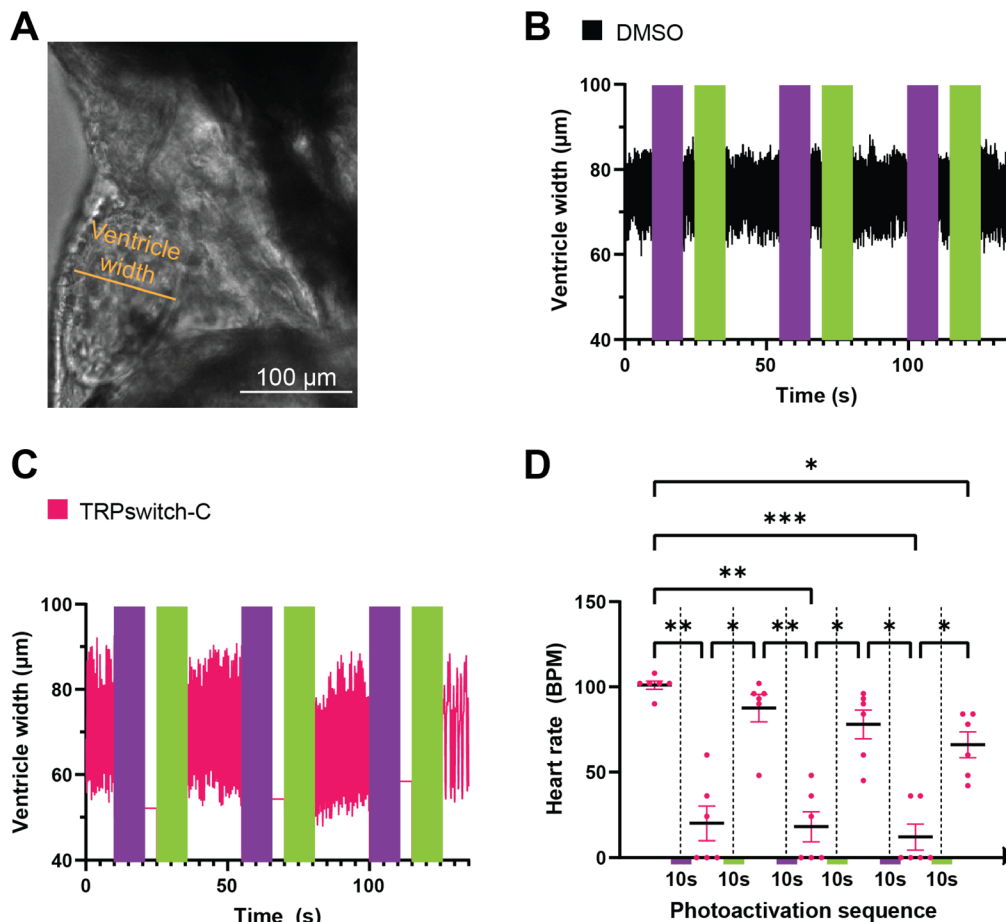


Fig. 6 Photocontrol of larval zebrafish heartbeats using *Trpa1b*/TRPswitch-C. The heartbeat interruption assay was performed on *Tg(cmlc2:Trpa1b-2A-EGFP)* larvae in a *trpa1b*^{-/-} background. (A) Image of the zebrafish heart showing the ventricle chamber and the location of ventricle width measurements. (B and C) Representative traces of the ventricle width measured every 100 ms during the photoactivation sequence from larvae incubated in DMSO (B) or TRPswitch-C (C). Violet and green vertical lines indicate photoactivation with 10 s of 390 nm or 555 nm light, respectively. (D) Quantification of the heart rate (beat per minute, BPM) in larvae incubated in TRPswitch-C during the photoactivation sequence. Values are mean \pm SEM. (*p*-value ns \geq 0.1234; * < 0.0332; ** < 0.0021; *** < 0.0002). see SI Movie 5.

Conclusions

In summary, we report the discovery of an aryl azopyrrole ligand, **TRPswitch-C**, for the optical control of TRPA1, including in live zebrafish. The discovery of this ligand expands the tool molecules available to address TRPA1, which we expect to be particularly useful in optochemogenetic experiments studying excitable cells where a large depolarization current is needed. It seems that using the large absorption tuneability of hetero-aromatic azos, together with suitable targets has large potential in future photopharmacology.

Author contributions

Conceptualization: LCD, JAM, RTP, PYL, MJF; Funding acquisition: PJP, RTP, PYL, MJF; Investigation: LCD, JAM, KYZ, RXT, PRH; Methodology: LCD, JAM, RTP, PYL, MJF; Supervision: PJP, RTP, PYL, MJF; Writing – original draft: LCD, JAM, PYL, MJF; Writing – review & editing: all authors.

Conflicts of interest

There are no conflicts to declare.

Data availability

The data supporting this article have been included as part of the supplementary information (SI). Further raw data can be obtained from the authors upon request.

Further experimental data is included in the SI and supplementary movie files. Supplementary information: SI Movie 1: Corresponding to Fig. 2C and D. Motion response of wild type zebrafish larvae in a 96 well plate treated with 1% DMSO (left) or 20 μ M TRPswitch-C (right) in response to 1 s stimulation of 390 nm light illumination. Light illumination is indicated by the purple box in the corner of each well. SI Movie 2: Corresponding to Fig. 4A. Motion response of wild type (top row) or *trpa1b* mutant (bottom row) zebrafish larvae in a 96 well plate treated with 1% DMSO (left) or 20 μ M TRPswitch-C (right) in



response to 1 s stimulation of 390 nm light illumination. Light illumination is indicated by the purple box in the corner of each well. SI Movie 3: Corresponding to Fig. 5A. Motion response of wild type zebrafish larvae in a 96 well plate treated with 1% DMSO (top row) or 20 μ M TRPswitch-C (bottom row) in response to 1 s stimulation of 390 nm (left), 440 nm (center), or 475 nm (right) light illumination. Light illumination is indicated by the corresponding-colored box between wells of the same stimulation condition. SI Movie 4: Corresponding to Fig. 5B. Motion response of wild type zebrafish larvae in a 96 well plate treated with 1% DMSO (top row) 20 μ M TRPswitch-C (bottom row) in response to 1 s stimulation (left) or 2 s stimulation of 475 nm light illumination. Light illumination is indicated by the blue box in the corner of each well. SI Movie 5: Corresponding to Fig. 6. Heartbeat of *Tg(cmlc2:Trpa1b-2A-EGFP)* in a *trpa1b^{-/-}* background, shown in DMSO (left) or 20 μ M TRPswitch-C (right) before and after the first stimulation cycle of the heartbeat interruption assay. Violet and green squares below the scale bar in each video indicate when 390 nm and 555 nm stimulations occur, respectively. The red circle indicates the area of stimulation. See DOI: <https://doi.org/10.1039/d5sc05070g>.

Acknowledgements

This work was supported by the National Institute Of Neurological Disorders And Stroke of the National Institutes of Health under Award Number R00NS112599 (P. Y. L.), the Advancing a Healthier Wisconsin Endowment (P. Y. L.) and the EPSRC (EP/R00188X/1 to M. J. F.) The authors also acknowledge the funding support from “Laboratory for Synthetic Chemistry and Chemical Biology” under the Health@InnoHK Program launched by Innovation and Technology Commission, The Government of Hong Kong Special Administrative Region of the People’s Republic of China. The authors would like to acknowledge Thomas Grimes of the Centre for Medicines Discovery at the University of Oxford for expertise and assistance with LCMS data acquisition and analysis. The graphical abstract was created with BioRender. Munos, J. (2025) <https://BioRender.com/q4hh9t6>.

References

- M. J. Fuchter, On the Promise of Photopharmacology Using Photoswitches: A Medicinal Chemist’s Perspective, *J. Med. Chem.*, 2020, **63**, 11436–11447.
- K. Hull, J. Morstein and D. Trauner, In Vivo Photopharmacology, *Chem. Rev.*, 2018, **118**, 10710–10747.
- W. A. Velema, W. Szymanski and B. L. Feringa, Photopharmacology: beyond proof of principle, *J. Am. Chem. Soc.*, 2014, **136**, 2178–2191.
- A. A. Beharry and G. A. Woolley, Azobenzene photoswitches for biomolecules, *Chem. Soc. Rev.*, 2011, **40**, 4422–4437.
- J. Morstein and D. Trauner, New players in phototherapy: photopharmacology and bio-integrated optoelectronics, *Curr. Opin. Chem. Biol.*, 2019, **50**, 145–151.
- S. Crespi, N. A. Simeth and B. König, Heteroaryl azo dyes as molecular photoswitches, *Nat. Rev. Chem.*, 2019, **3**, 133–146.
- J. L. Greenfield, A. R. Thawani, M. Odaybat, R. S. L. Gibson, T. B. Jackson, and M. J. Fuchter, in *Molecular Photoswitches*, ed. Z. L. Pianowski, Wiley VCH, 2022, pp. 84–113.
- Select examples include: (a) Q. Zhang, C. S. Kounde, M. Mondal, J. L. Greenfield, J. R. Baker, S. Kotelnikov, M. Ignatov, C. P. Tinworth, L. Zhang, D. Conole, E. De Vita, D. Kozakov, A. McCluskey, J. D. Harling, M. J. Fuchter and E. W. Tate, Light-mediated multi-target protein degradation using arylazopyrazole photoswitchable PROTACs (AP-PROTACs), *Chem. Commun.*, 2022, **58**, 10933–10936; (b) T. Aguirre, E. Teichmann, F. Q. Rompp, R. Vivier, C. Bryant, M. A. Hulverson, W. C. Van Voorhis, K. K. Ojo, J. S. Doggett, D. Fiedler and S. Hecht, Photoswitchable Inhibitors to Optically Control Specific Kinase Activity, *ACS Chem. Biol.*, 2023, **18**, 1378–1387; (c) A. Sink, H. Gerwe, H. Hubner, V. Boivin-Jahns, J. Fender, K. Lorenz, P. Gmeiner and M. Decker, Photo-Adrenalines: Photoswitchable beta(2)-Adrenergic Receptor Agonists as Molecular Probes for the Study of Spatiotemporal Adrenergic Signaling, *Chem.–Eur. J.*, 2024, **30**, e202303506; (d) S. A. M. Steinmuller, J. Fender, M. H. Deventer, A. Tutov, K. Lorenz, C. P. Stove, J. N. Hislop and M. Decker, Visible-Light Photoswitchable Benzimidazole Azo-Arenes as beta-Arrestin2-Biased Selective Cannabinoid 2 Receptor Agonists, *Angew Chem. Int. Ed. Engl.*, 2023, **62**, e202306176; (e) N. N. Mafy, K. Matsuo, S. Hiruma, R. Uehara and N. Tamaoki, Photoswitchable CENP-E Inhibitor Enabling the Dynamic Control of Chromosome Movement and Mitotic Progression, *J. Am. Chem. Soc.*, 2020, **142**, 1763–1767.
- J. Morstein, M. Awale, J. Reymond and D. Trauner, Mapping the Azolog Space Enables the Optical Control of New Biological Targets, *ACS Cent. Sci.*, 2019, **5**, 607–618.
- P. Lam, A. R. Thawani, E. Balderas, A. J. P. White, D. Chaudhuri, M. J. Fuchter and R. T. Peterson, TRPswitch-A Step-Function Chemo-optogenetic Ligand for the Vertebrate TRPA1 Channel, *J. Am. Chem. Soc.*, 2020, **142**, 17457–17468.
- S. Kirchner, A. Leistner, P. Godtel, A. Seliwjorstow, S. Weber, J. Karcher, M. Nieger and Z. Pianowski, Hemipiperazines as peptide-derived molecular photoswitches with low-nanomolar cytotoxicity, *Nat. Commun.*, 2022, **13**, 6066.
- A. Sampieri, T. Padilla-Flores, A. R. Thawani, P. Lam, M. J. Fuchter, R. Peterson and L. Vaca, The conducting state of TRPA1 modulates channel lateral mobility, *Cell Calcium*, 2023, **116**, 102800.
- I. Tabansky, Y. Liang, M. Frankfurt, M. A. Daniels, M. Harrigan, S. Stern, T. A. Milner, R. Leshan, R. Rama, T. Moll, J. M. Friedman, J. N. H. Stern and D. W. Pfaff, Molecular profiling of reticular gigantocellularis neurons indicates that eNOS modulates environmentally dependent levels of arousal, *Proc. Natl. Acad. Sci. U. S. A.*, 2018, **115**, E6900–E6909.
- C. E. Weston, R. D. Richardson, P. R. Haycock, A. J. P. White and M. J. Fuchter, Arylazopyrazoles: azoheteroarene



- photoswitches offering quantitative isomerization and long thermal half-lives, *J. Am. Chem. Soc.*, 2014, **136**, 11878–11881.
- 15 J. Calbo, C. E. Weston, A. J. P. White, H. S. Rzepa, J. Contreras-Garcia and M. J. Fuchter, Tuning Azoheteroarene Photoswitch Performance through Heteroaryl Design, *J. Am. Chem. Soc.*, 2017, **139**, 1261–1274.
- 16 C. E. Weston, R. D. Richardson and M. J. Fuchter, Photoswitchable basicity through the use of azoheteroarenes, *Chem. Commun.*, 2016, **52**, 4521–4524.
- 17 V. Jelinkova, A. Dellai, L. Verwaerde, V. Rodriguez, M. Feckova, M. Vachtlova, J. Podlesny, M. Klikar, B. Sempe, M. Hugget, P. Parik, O. Pytela, F. Castet, Z. Ruzickova and F. Bures, Property Tuning in N-Methylpyrrole Azo-Photoswitches *via* Modification of the Peripheral Substituents, *Chem.–Eur. J.*, 2025, **31**, e202404221.
- 18 K. W. Shimkin, P. G. Gildner and D. A. Watson, Copper-Catalyzed Alkylation of Nitroalkanes with alpha-Bromonitriles: Synthesis of beta-Cyanonitroalkanes, *Org. Lett.*, 2016, **18**, 988–991.
- 19 E. Fischer, Calculation of photostationary states in systems $A \rightleftharpoons B$ when only A is known, *J. Phys. Chem.*, 1967, **71**, 3704–3706.
- 20 P. Lam, S. K. Mendu, R. W. Mills, B. Zheng, H. Padilla, D. J. Milan, B. N. Desai and R. T. Peterson, A high-conductance chemo-optogenetic system based on the vertebrate channel Trpa1b, *Sci. Rep.*, 2017, **7**, 11839.
- 21 D. A. Prober, S. Zimmerman, B. R. Myers, B. M. J. McDermott, S. Kim, S. Caron, J. Rihel, L. Solnica-Krezel, D. Julius, A. J. Hudspeth and A. F. Schier, Zebrafish TRPA1 channels are required for chemosensation but not for thermosensation or mechanosensory hair cell function, *J. Neurosci.*, 2008, **28**, 10102–10110.
- 22 A. D. Douglass, S. Kraves, K. Deisseroth, A. F. Schier and F. Engert, Escape behavior elicited by single, channelrhodopsin-2-evoked spikes in zebrafish somatosensory neurons, *Curr. Biol.*, 2008, **18**, 1133–1137.

