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Mapping photoisomerization dynamics on a three-state model potential energy surface in bacteriorhodopsin using femtosecond stimulated Raman spectroscopy†

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The process of proton translocation in *Halobacterium salinarum*, triggered by light, is powered by the photoisomerization of all-*trans*-retinal in bacteriorhodopsin (bR). The primary events in bR involving rapid structural changes upon light absorption occur within subpicoseconds to picoseconds. While the three-state model has received extensive support in describing the primary events between the H and K states, precise characterization of each excited state in the three-state model during photoisomerization remains elusive. In this study, we investigate the ultrafast structural dynamics of all-*trans*-retinal in bR using femtosecond stimulated Raman spectroscopy. We report Raman modes at 1820 cm⁻¹ which arise from C=C stretch vibronic coupling and provide direct experimental evidence for the involvement of the I and J states with 2A_g⁻ symmetric character in the three-state model. The detection of the C=C vibronic coupling mode, C=N stretching mode (1700 cm⁻¹), and hydrogen out-of-plane (HOOP) mode (954 cm⁻¹) further supports the three-state model that elucidates the initial charge translocation along the conjugated chain accompanied by *trans*-to-*cis* photoisomerization dynamics through H(1B_u⁺) → I(2A_g⁻) → J(2A_g⁻) → K(13-*cis* ground state) transitions in all-*trans*-retinal in bR.

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Introduction

The photophysical and photochemical properties of retinal proteins have been extensively investigated to comprehend the initial capture of visible light and subsequent structural modifications in the retinal chromophore.^{1–6} Bacteriorhodopsin (bR) is a photoreceptor protein discovered in the purple membrane of *Halobacterium salinarum*, which functions as a light-activated proton pump. Upon photon excitation, a complex photocycle process is initiated in the all-*trans*-retinal in bR. This intricate process involves multiple intermediates, including H → I → J → K → L → M → N → O states.⁷

The ultrafast primary events involving temporal and structural changes upon light absorption in bR occur within subpicoseconds to a few picoseconds.^{8–10} Previous experimental findings and calculations strongly support the three-state model (see Fig. 1a) instead of the two-state model for describing the ultrafast primary events in bR.^{11–16} The all-*trans*-retinal molecule is formally classified under the C_{2h} point group; the three-state model for all *trans*-retinal comprises one

ground state and two excited states with B_u and A_g symmetries.¹⁷ It has been widely acknowledged that the absorption band observed in all *trans*-retinal in bR within the green spectral range is attributed to one photo-allowed transition from the S₀ (1A_g⁻) ground state to the S₁ excited state with the 1B_u⁺ symmetry; while the S₂ excited state, which is about 3500 cm⁻¹ higher than the S₁ (1B_u⁺) state, shares the same symmetry (2A_g⁻) as the ground state S₀, resulting in a forbidden one-photon transition between them.¹⁷ The calculations suggest that after light absorption by the first excited state S₁ (1B_u⁺), retinal carries out a rapid torsion around its C₁₃=C₁₄ bond, subsequently undergoing two nonadiabatic transitions between S₁ (1B_u⁺) and S₂ (2A_g⁻) as well as S₂ (2A_g⁻) and the *cis* ground state (see Fig. 1a).¹⁴ Despite conducting numerous time-resolved spectroscopic investigations with a high temporal resolution, there has been no direct experimental evidence to support the involvement of the S₂ state with 2A_g⁻ symmetric character in the excited-state dynamics of bR.

For short polyenes with a conjugation length of N ≤ 5, previous studies have identified a C=C stretching Raman mode at 1750–1800 cm⁻¹, attributed to an upshift in the frequency of the 2A_g⁻ state C=C stretching Raman mode, resulting from the adiabatic vibronic coupling between the 2A_g⁻ and 1A_g⁻ states.^{18–21} Governed by the symmetry rules of the C_{2h} point group in all-*trans* polyenes, this coupling results in a well-known frequency upshift of 100–150 cm⁻¹ for the C=C stretching mode in the

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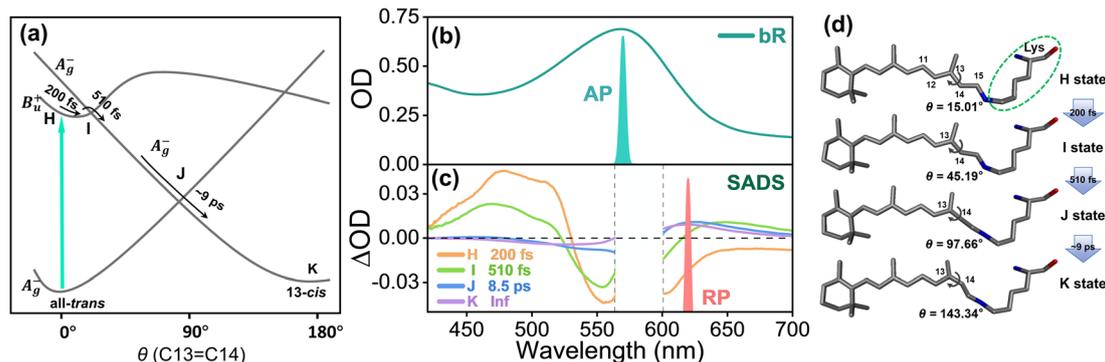


Fig. 1 (a) Schematic potential energy curves of the three-state model along the torsional angle of the C13=C14 bond of all-*trans*-retinal in bR. (b) The absorption spectrum of bR under light-adapted conditions with the actinic pump (AP) wavelength in FSRs depicted by a color spike. (c) The SADS from global analysis of the TA spectra of bR with the Raman pump (RP) wavelength in FSRs depicted by a color spike. (d) The chemical structures of the retinal molecule in bR in different intermediate states obtained by TR-SFX; θ represents the dihedral angle of C12–C13=C14–C15. The PDB IDs for various intermediate states are as follows: H state (6G7H), I state (6G7I), J state (6G7J), and K state (6G7K).¹⁰

$2A_g^-$ state, while simultaneously lowering the frequency of the C=C stretching mode in the $1A_g^-$ state.^{19,22,23} More recently, femtosecond stimulated Raman spectroscopy (FSRS) has revealed a high-frequency Raman mode at 1712 cm^{-1} in UV-absorbing rhodopsin HKR1 (*histidine kinase rhodopsin 1*), attributed to the ethylenic C=C stretch vibronic coupling mode of the 13-*cis* retinal molecule.⁵ These findings have motivated us to reexamine the ultrafast structural dynamics of all-*trans*-retinal in bR using FSRs, with a primary focus on high-frequency Raman signals ($>1600\text{ cm}^{-1}$) associated with C=C stretch vibronic coupling between the two A_g^- states.

In this study, the utilization of a Raman pump passing through a grating filter system in FSRs significantly enhances both spectral resolution and sensitivity, thereby facilitating the acquisition of more intricate insights into the excited state vibrational dynamics of all-*trans*-retinal in bR (see Fig. S1b and c†).^{24–28} The observation of the highest frequency Raman modes at 1820 cm^{-1} , exhibiting biexponential decay dynamics, can be attributed to C=C stretch vibronic coupling between I ($2A_g^-$)/J ($2A_g^-$) states and the $1A_g^-$ ground state. Furthermore, the identification of the vibronic coupling C=C stretching mode, C=N stretching mode (1700 cm^{-1}), and the HOOP mode (954 cm^{-1}) provides support for the three-state model that describes the initial charge transfer accompanied by transition of H ($1B_u^+$) \rightarrow I ($2A_g^-$) \rightarrow J ($2A_g^-$) \rightarrow K (13-*cis* ground state) \rightarrow L states in all-*trans*-retinal in bR.

Results and discussion

The absorption spectrum of all-*trans* retinal in bR under light-adapted conditions is illustrated in Fig. 1b. As shown in Fig. S3b and S4b,† the two-dimensional transient absorption (TA) spectra with an excitation energy of 200 nJ per pulse exhibit intricate excited state features between 400 nm and 750 nm. Global analysis with a sequential model was employed to obtain the species-associated difference spectra (SADS) (see Fig. 1c), yielding four characteristic lifetimes with time constants of $\tau_1 = 200\text{ fs}$ (H \rightarrow I state), $\tau_2 = 510\text{ fs}$ (I \rightarrow J state), $\tau_3 = 8.5\text{ ps}$ (J \rightarrow K

state), and $\tau_4 =$ an infinitely long lifetime (K \rightarrow L state).^{9,29–31} Notably, the lifetime of the J state ($\tau_3 = 8.5\text{ ps}$) observed in our measurement exceeds the commonly recognized value of $\sim 3\text{--}4\text{ ps}$.^{9,10,13,15} To explore this discrepancy, we collected TA spectra using a lower excitation energy of 50 nJ per pulse (see Fig. S3a and S4a†), where the SADS for the J-state lifetime ($\tau_3 = 4.5\text{ ps}$) were consistent with previous studies. It has been demonstrated that at sufficiently high light intensity, two-photon absorption occurs in bR, resulting in the excitation of the tryptophan residue (Trp86) close to the retinal chromophore.^{31,32} This excitation induces strong excitonic coupling between Trp86 and retinal, which significantly influences the structural dynamics of the retinal chromophore. To further validate our findings, we left the bR sample in the dark overnight following the TA measurement at 200 nJ per pulse. TA data were then recollected at low excitation energy (50 nJ per pulse), and the results were consistent with those from a fresh bR sample under identical excitation conditions (see Fig. S3c†).

To gain insights into the structural dynamics beyond electronic features, we used FSRs to track the excited-state vibrational marker bands of bR. Fig. S5† shows the time-resolved FSRs spectra of bR across a frequency range of $900\text{--}2000\text{ cm}^{-1}$ at an excitation energy of 50 nJ per pulse. We strategically selected a 620 nm Raman pump to pre-resonantly enhance the signal on the Stokes side (Fig. 1b) by matching the transition of the ESA band of I,¹² J, and K states (see Fig. S4†). The decay lifetimes for the three states are consistent with those observed in TA spectra (see Fig. S6†). However, due to the limited signal-to-noise ratio at this low excitation power, capturing more detailed Raman signal dynamics was challenging. In contrast, Fig. 2 presents time-resolved FSRs spectra of bR at a higher excitation energy of 200 nJ per pulse (with the baseline and raw data shown in Fig. S7†), revealing more pronounced excited-state Raman signals with intricate dynamics. The ground state spectrum (bottom of Fig. 2) reveals six major Raman peaks: the methyl rocking mode at 1006 cm^{-1} , the C–C stretching doublet mode at 1168 and 1200 cm^{-1} , and the ethylenic C=C stretching mode at 1529 cm^{-1} .^{9,33–37}



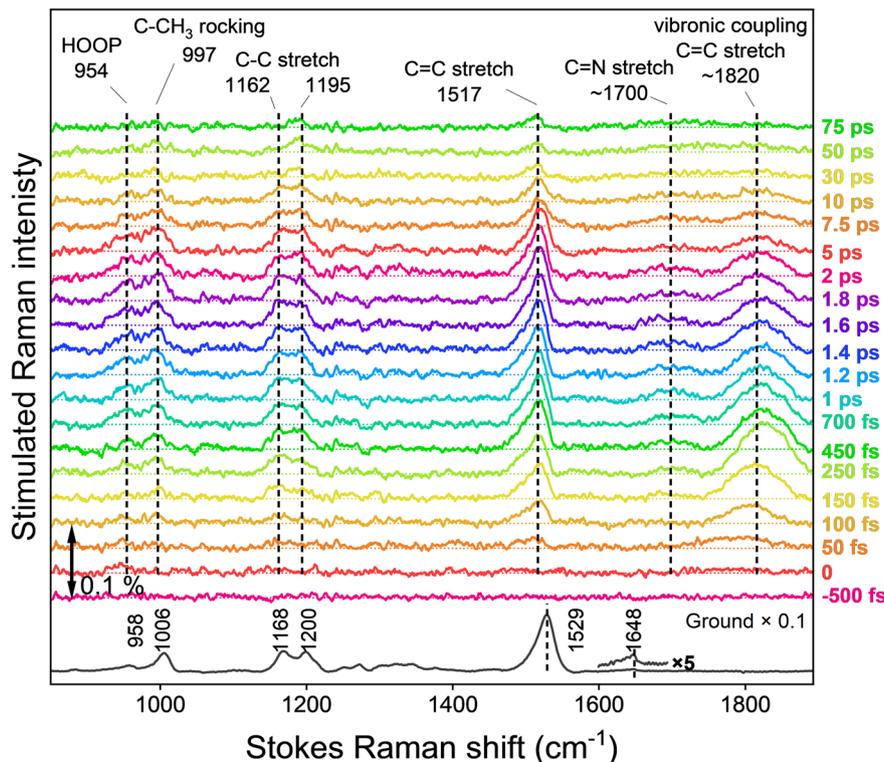


Fig. 2 Excited-state FSRS spectra of all-*trans*-retinal in bR from 900–1850 cm^{-1} at an excitation energy of 200 nJ per pulse. The ground state FSRS spectrum is shown at the bottom for comparison.

Additionally, the HOOP mode at 958 cm^{-1} and the C=N stretch mode at 1648 cm^{-1} are also discernible.^{33,37,38}

As depicted in Fig. 2, upon photoexcitation at 570 nm, the aforementioned five major Raman peaks in the excited state exhibit a significant redshift in frequency to 1162 and 1195 cm^{-1} (C–C stretch), 997 cm^{-1} (C–CH₃ rocking), 954 cm^{-1} (HOOP), and to 1517 cm^{-1} (C=C stretch) compared to their counterparts in the ground state.⁹ In contrast, the C=N stretching mode associated with the Schiff base experiences a significant frequency blueshift to ~1700 cm^{-1} in the excited state.³⁹ The presence of a prominent Raman mode at approximately 1820 cm^{-1} has not been previously reported for all-*trans*-retinal in bR using FSRS,^{9,40,41} and its unusually high frequency suggests a specific origin. Notably, such a Raman mode was commonly observed in carotenoid molecules possessing linear conjugated polyene chains with conjugation lengths (*N*) ranging from 9 to 15.^{42–45} A similar excited state Raman mode at 1712 cm^{-1} has been reported for unprotonated 13-*cis* retinal in UV-absorbing rhodopsin,⁵ which is attributed to the C=C stretching mode of the optically forbidden $2A_g^-$ state resulting from strong adiabatic vibronic coupling between $2A_g^-$ and $1A_g^-$ states. The Raman mode at 1820 cm^{-1} in our case is thereby assigned to a significant blueshift of the $2A_g^-$ state C=C ethylenic stretch, arising from the vibronic coupling between the two A_g^- states in the all-*trans* retinal in bR.¹⁸ The discrepancy in frequencies between the vibronic-coupled C=C stretching mode at 1820 cm^{-1} and the all-*trans* C=C stretching mode (S_0) at 1529 cm^{-1} , *i.e.*, 1820–1529 = 291 cm^{-1} , exhibits

a significantly higher magnitude compared to that observed for UV-absorbing rhodopsin (~200 cm^{-1}). This observation suggests a very strong vibronic coupling within the all-*trans* retinal of bR.⁴⁶

We obtained SADS extracted from global analysis in a sequential mode using a kinetic scheme H_{invisible} state → I state → J state → K state → of FSRS spectra,^{47,48} as shown in Fig. 3. Importantly, the H state exhibiting negligible FSRS signal intensity was designated as an invisible component in SADS analysis (*i.e.*, a spectrally silent species^{48,49}), primarily due to the

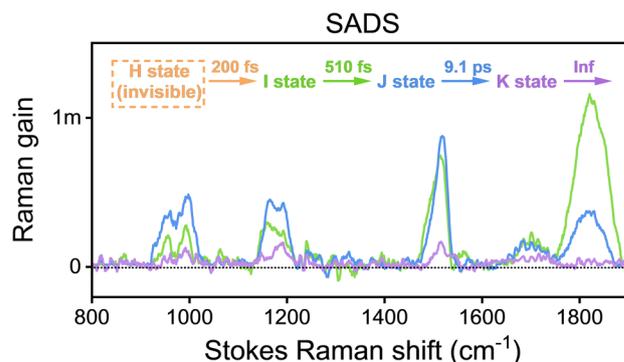


Fig. 3 The SADS from global analysis of the excited state FSRS spectra of bR. Due to the experimental resonance Raman conditions, the H state with a negligible FSRS signal was set as an invisible component (not contributing to the spectra) in the global analysis, denoted by the dashed-line box.



experimental resonance Raman conditions where the H species is off-resonant with a 620 Raman pump. The FSRS spectra of bR are satisfactorily fitted in SADS using a model that includes three species of $\tau_2 = 500$ fs (I state), $\tau_3 = 9.1$ ps (J state), and $\tau_4 =$ an infinitely long lifetime (K state), which reveals significant dynamics of formation and relaxation in the fingerprint Raman modes within the intermediate states I, J, and K.

To quantitatively examine the temporal variation of stimulated Raman signals, we analyzed the transient amplitude observed in each Raman band. Fig. 4a, b, and S8† present the transient kinetic trace of the C–C and C=C stretching bands at 1162, 1195, and 1517 cm^{-1} .⁹ The area of these peaks was determined by fitting to a Gaussian line shape function. The time evolution of the peak area was fitted using least-squares fits with four-time components retrieved from SADS global fitting in FSRS spectra. All three Raman peaks exhibit a double exponential rise with time constants of $\tau_1 = 200$ fs and $\tau_2 = 500$ fs followed by two decay components of $\tau_3 = 9.1$ ps and $\tau_4 =$ an infinitely long lifetime, indicating observation of the I state (green dashed line), J state (blue dashed line), and K state (purple dashed line) *via* the relaxation scheme $H_{\text{invisible}} \rightarrow \tau_1 I \rightarrow \tau_2 J \rightarrow \tau_3 K \rightarrow \tau_4 L$ states.

Notably, different from the C–C and C=C stretching modes, the transient amplitude of the C=C vibronic coupling Raman mode at 1820 cm^{-1} exhibits a rise time of 200 fs accompanied by two exponential decay components of 500 fs and 9.1 ps, as

depicted in Fig. 4c. Based on previous studies, this behavior suggests that the excited states I and J can be assigned to the optically forbidden $2A_g^-$ state, which undergoes strong vibronic coupling with the *trans*- S_0 ground state ($1A_g^-$). This coupling with the $2A_g^- - 1A_g^-$ states, shifts the C=C stretching in the $2A_g^-$ (I and J) state upward to 1820 cm^{-1} . Simultaneously, the vibronic interaction lowers the C=C stretching of the $1A_g^-$ state to 1529 cm^{-1} .^{19,22,23} The previous experimental and computational findings provide support for the three-state model,^{13–15} in which the optically allowed state, termed the single excited state $\pi-\pi^* 1B_u$ state, is positioned lower than an optically forbidden state (the $2A_g^-$ state with a double $\pi-\pi^*$ excitation) in the Frank-Condon geometry, based on the C_{2h} symmetry. The primary determinant of the reaction coordinate is considered to be the torsional motion revolving around the $C_{13}=C_{14}$ double bond involved in the reaction process.^{1,50} The vertical photoexcitation of the all-*trans*-retinal in bR initially leads to the formation of the intermediate excited state H ($1B_u^+$), which corresponds to a charge transfer state involving hole-pair (ionic) excitation.^{51–53} Consequently, the interaction between the H ($1B_u^+$) state and the upper $2A_g^-$ state surface results in a slight energy barrier on the H ($1B_u^+$) state surface caused by a small torsion process,¹⁴ after which an inversion of energy levels occurs between $1B_u^+$ and $2A_g^-$ at the first crossing point to form the intermediate state I ($2A_g^-$) after 200 fs. Importantly, the observed C=C vibronic coupling in the I ($2A_g^-$) state suggests that the minor torsion

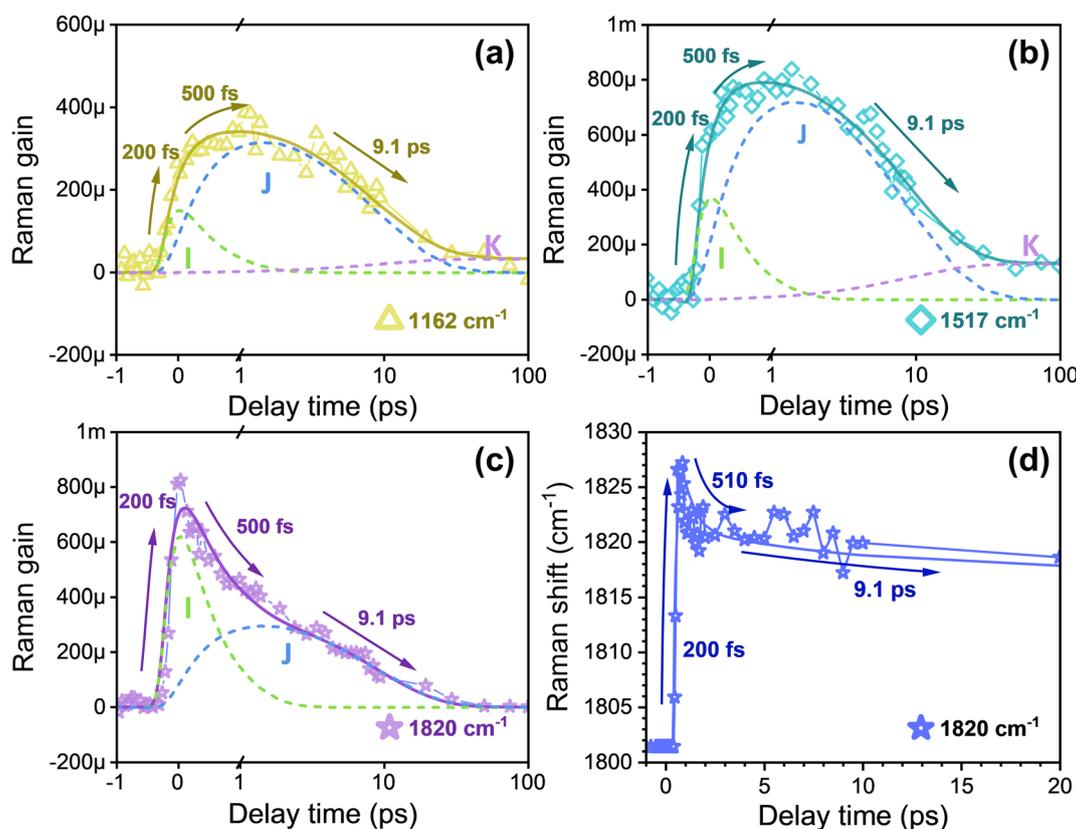


Fig. 4 The transient Raman intensities of the (a) C–C stretching mode at 1162 cm^{-1} and (b) C=C stretching mode at 1520 cm^{-1} . The transient Raman intensities (c) and frequencies (d) of the C=C vibronic coupling mode at 1820 cm^{-1} . The dashed lines represent different intermediate states: the I state (green), J state (blue), and K state (purple).



following photoexcitation is unlikely to break the C_{2h} symmetry of retinal in bR. This observation is consistent with the findings from time-resolved serial femtosecond crystallography (TR-SFX), which determined a change in torsion angle of about 30° via the $C_{12}-C_{13}=C_{14}-C_{15}$ bonds (shifting from 15° in the H state to 45° in the I state), as shown in Fig. 1d.¹⁰ Having established the I ($2A_g^-$) state in 200 fs, further structure distortion is induced by twisting the $C_{12}-C_{13}=C_{14}-C_{15}$ dihedral angle, and the observed $\tau_2 = 500$ fs ultrafast decay dynamics in Fig. 4c can be attributed to the torsion-induced weakening of the C_{2h} symmetry in the $2A_g^-$ state along the I \rightarrow J transition. However, this interpretation is challenged by TR-SFX data, which reveal a dihedral angle of approximately 97° in the J ($2A_g^-$) state (Fig. 1d), suggesting a potential loss of C_{2h} symmetry.¹⁹ A possible explanation for this discrepancy may lie in the difference in experimental conditions. The TR-SFX experiments were conducted on bR in a crystalline state, where confinement and lower hydration levels may affect the structural dynamics differently than in physiological environments (in solution), potentially altering the torsional behavior.³⁴ Our FSRS results for bR suggest that the dihedral angle in the J state is likely less than 90° under physiological conditions. Following the formation of the J ($2A_g^-$) state, the potential energy surfaces intersect with the K (*cis*-ground) state surface along the reaction coordinate at a second weakly crossing point.¹⁴ This reveals a J \rightarrow K state transition by further twisting the $C_{12}-C_{13}=C_{14}-C_{15}$ dihedral angle (to approximately 143° , see Fig. 1d), resulting in decreased vibronic coupling strength to zero due to the complete breaking of the C_{2h} symmetry. Eventually, this leads to the formation of *cis*-retinal. As depicted in Fig. 4d, the transient blueshift of the vibronic coupling mode reveals an ultrafast blueshift with a time constant of $\tau_1 = 200$ fs followed by two components of redshift with lifetimes of $\tau_2 = 510$ fs and $\tau_3 = 9.1$ ps. The frequency blueshift indicates an enhancement in the strength of vibronic coupling during the formation of the I ($2A_g^-$) state, while the biexponential redshift can be attributed to a weakening effect for vibronic coupling caused by torsion.⁵⁵

The vibrational frequency at 954 cm^{-1} corresponds to the HOOP vibrational mode, which is highly sensitive to local torsion in the $C_{13}=C_{14}$ region during retinal

isomerization.^{3,9,25,37,38,56,57} As shown in Fig. 5a, akin to the transient dynamics observed in C–C and C=C bonds, the C_{14} -H HOOP mode exhibits two rising components at $\tau_1 = 200$ fs and $\tau_2 = 500$ fs followed by two decay components of $\tau_3 = 9.1$ ps and $\tau_4 =$ an infinitely long lifetime, suggesting a direct probe of the isomerization in retinal during the I \rightarrow J \rightarrow K \rightarrow L transition. Our findings are consistent with the previous reports using TR-SFX, as structural changes were already observed at the I state.¹⁰ Due to the sensitivity of the HOOP modes towards the tilting angle around the C=C bond,¹⁵ the observed weak signal intensity in the I state indicates a twisting of the polyene backbone via $C_{12}-C_{13}=C_{14}-C_{15}$ torsion at a small angle (see Fig. 1d).^{10,15} The increase in FSRS signal intensity observed during the development of the J state indicates a subsequent enhancement in the torsion angle of $C_{12}-C_{13}=C_{14}-C_{15}$.¹⁰ During this period, there is an alteration in the torsion angles of the $C_{13}=C_{14}$ and adjacent $C_{11}=C_{12}$ and $C_{15}=N$ bonds in opposing orientations, suggesting that the reaction follows a space-saving bicycle-pedal model of retinal isomerization.⁵⁸ Subsequently, the decrease in transient intensity of the HOOP signal with a time constant of 9.1 ps arises due to a further twist in the $C_{12}-C_{13}=C_{14}-C_{15}$ bond (to approximately 143°), indicating complete isomerization into its 13-*cis* configuration.⁵⁸ The infinitely long decay lifetime is assigned to the decay from the K to the L state; observing this decay dynamics in the transient HOOP mode provides evidence for further conformation change during the K \rightarrow L transition.^{59,60} This observation aligns with the TR-SFX diffraction data, which reveal that, during the K \rightarrow L transition, the 13-*cis* retinal progressively adopts a planar configuration, ultimately achieving the essentially *cis*-planar configuration within approximately 290 ns.⁷

As depicted in Fig. 5b, a vibration mode observed in the high-frequency region at $\sim 1700\text{ cm}^{-1}$ by FSRS corresponds to the C=N stretching mode.³⁹ The previous TA results have demonstrated excitonic coupling between the dipole moment of retinal and nearby tryptophan residues (Trp86); the experimental findings reflect a translocation of the positive charge from the protonated Schiff base toward the β -ionone end of retinal within 200 fs after photoexcitation.^{51,52} This leads to an enhancement of the dipole moment and a redistribution of

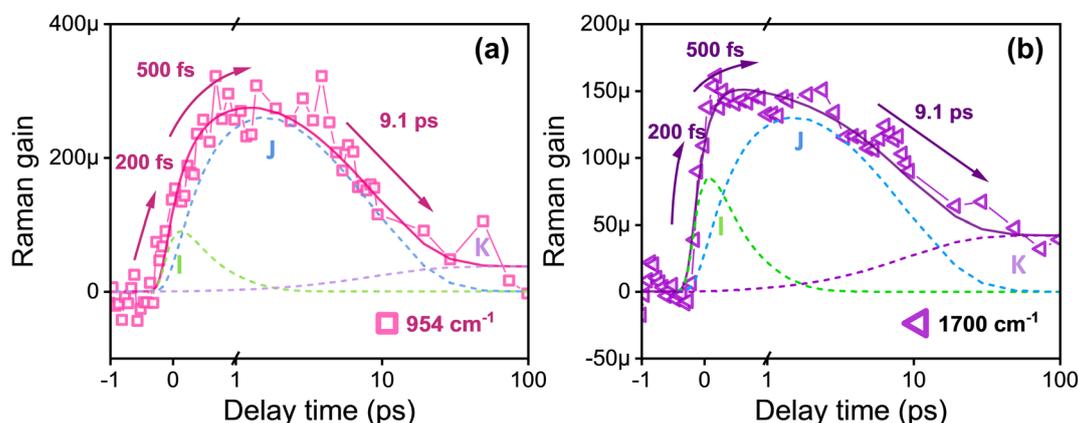


Fig. 5 The transient Raman intensities of the (a) HOOP vibrational mode at 954 cm^{-1} and (b) C=N stretching vibrational mode at $\sim 1700\text{ cm}^{-1}$.



charge within the retinal. Furthermore, considering that the occurrences at 200 fs involve a change in the configuration of the retinal framework, the increase in dipole moment is related to structural changes and possibly serves as the driving force for subsequent *trans-to-cis* isomerization.^{51,52,61,62} The FSRS spectra of the C=N stretching mode are satisfactorily fitted including $\tau_1 = 200$ fs rising followed by three decay components of $\tau_2 = 500$ fs (I \rightarrow J state), $\tau_3 = 9.1$ ps (J \rightarrow K state), and $\tau_4 =$ an infinitely long lifetime (K \rightarrow L state), similar to the transient dynamics observed in the HOOP mode. The findings suggest a strong correlation between the gradual decrease in dipole moment and simultaneous modifications observed in the retinal structure.^{15,51,52}

Conclusion

In this study, we investigate the ultrafast structural dynamics of all-*trans*-retinal in bR using FSRS and report the previously undiscovered Raman modes at 1820 cm^{-1} , which arise from vibronic coupling between the two-photon allowed $2A_g^-$ states and $1A_g^-$ ground state. The observation of the vibronic coupling C=C stretching mode and C=N stretching mode (at 1700 cm^{-1}) as well as the HOOP mode (at 954 cm^{-1}) provides support for the three-state model in elucidating the initial photoisomerization dynamics of retinal in bR (schematic potential energy curves of the three-state model are depicted in Fig. 1a). In our experiment, the I and J states are identified as higher-energy two-photon allowed $2A_g^-$ states, in comparison to the $2B_u^+$ -like H state. The transition from H ($2B_u^+$) to the I ($2A_g^-$) state occurs within 200 fs and involves an enhancement in dipole moment resulting from the charge transfer and a small change of the $C_{12}-C_{13}=C_{14}-C_{15}$ dihedral angle in the all-*trans* retinal structure. The vibronic coupling is subsequently attenuated by twisting the $C_{12}-C_{13}=C_{14}-C_{15}$ dihedral angle, accompanied by a reduction in dipole moment within ~ 500 fs, resulting in a transition from I ($2A_g^-$) to J ($2A_g^-$) states. Afterward, a transition from J ($2A_g^-$) to K 13-*cis* ground states occurs by further twisting the dihedral angle of $C_{12}-C_{13}=C_{14}-C_{15}$, resulting in reduced vibronic coupling strength towards zero due to complete disruption of the C_{2h} symmetry. Eventually, during the K \rightarrow L transition, there is a further conformation change that results in the formation of a *cis*-planar configuration. Importantly, our experimental findings are significantly supported by the TR-SFX experimental results,¹⁰ providing novel insights into the previously unexplored optical properties of bR and emphasizing the unique utility of FSRS in advancing our comprehension of photoreceptor protein dynamics.

Data availability

The data supporting this article have been included as part of the ESI.†

Author contributions

Z. W., Y. C. and W. L. conceived, designed and performed the experiments. Y. C., Z. W. and J. J. analysed the experimental

results. Z. W., Y. C., and W. L. co-wrote the manuscript. X. Z. provided the bR samples. All authors participated in discussions and edited the manuscript. W. L. directed the project.

Conflicts of interest

There are no conflicts of interest to declare.

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