Chemical Science

ROYAL SOCIETY OF CHEMISTRY

EDGE ARTICLE

View Article Online
View Journal | View Issue

Cite this: *Chem. Sci.*, 2025, **16**, 318

All publication charges for this article have been paid for by the Royal Society of Chemistry

Received 23rd August 2024
Accepted 12th November 2024

DOI: 10.1039/d4sc05679e

rsc.li/chemical-science

A flexible and scalable synthesis of 4′-thionucleosides†

Callum Lucas,‡[a] Ethan Fung, ‡[a] Matthew Nodwell,[a] Steven Silverman,[b] Bara Singh,[a] Louis-Charles Campeau[b] and Robert Britton *[a]

4′-Thionucleosides (thNAs) are synthetic nucleoside analogues that have attracted attention as leads for drug discovery in oncology and virology. Here we report a *de novo* thNA synthesis that relies on a scalable α-fluorination and aldol reaction of α-heteroaryl acetaldehydes followed by a streamlined process involving carbonyl reduction, mesylate formation and a double displacement reaction using NaSH. We demonstrate the multigram preparation of 4′-thio-5-methyluridine and highlight the production of purine and pyrimidine thNAs as well as C2′-modified thNAs.

Introduction

For several decades nucleoside analogues (NAs) have served as a prolific source of antiviral and anticancer therapeutics, and account for more than half of all approved antiviral drugs.[1] However, the pressure of new pathogens and emergence of drug resistance has highlighted the need for continued exploration of NA-relevant chemical space to identify compounds with novel mechanisms of action or enhanced resistance-combating properties.[2] One promising class of NAs that have been studied since the 1960s[3] are 4′-thio NAs (thNAs), where the ring oxygen is replaced with a sulfur atom. This single modification can have a profound impact on biological activity, including pharmacokinetic and pharmacodynamic properties.[1,4] Further, due to the increased stability of the C–N anomeric bond, thNAs are generally more resistant to hydrolysis.[5] For example, thiarabine (4′-thioaraC (**1**), Fig. 1),[4] a thNA of the sponge metabolite cytarabine, was developed to treat hematological malignancies and solid tumors. Here, replacement of the endocyclic oxygen with sulfur resulted in an improved once daily oral dosing regimen compared to cytarabine, which requires twice-daily intravenous administration.[6] The structurally related 2′-deoxy-fluoro thNA FF-10502 (**2**)[7] is an anticancer agent with improved potency over the related NA gemcitabine. Additionally, 4′thio-DMDC (**3**)[8] and the C4′ alkyne thNA **4**[9] have demonstrated potent anticancer and anti-HIV activities, respectively. In particular, the C4′-alkyne containing thNA **4** is a nucleoside/nucleotide reverse transcriptase inhibitor (NRTI) that also demonstrated an excellent selectivity index.[9] The use of thNAs in oligonucleotide sequences is also of importance, and processes to access 4′-thio locked nucleic acids (LNAs)[10] or to carry out nucleobase diversification using biocatalysis[11] have advanced efforts in this area.

A common approach to thNAs involves production of a protected 4-thioribose (*e.g.*, **6**), which can be achieved in as little as 6 steps.[12] For example, Miller has shown that **6** can be accessed from the ribose-derived bromo aldehyde **5** on multigram scale by bromide displacement using NaSH.[12] This synthesis supported production of 4′-thiouridine (**7**),[12] a precursor to thiarabine (**1**). An important contribution by Guindon[13] demonstrated that thNAs can also be constructed using an acyclic approach where the nucleobase is attached prior to cyclization. For example, the ribose-derived ^{t}Bu thioether **9** was cyclized under basic conditions to form the 2′-fluoro thNA **10**.[13]

Our groups have previously reported[14] a straightforward synthesis of NAs **14** that relies on two key steps: (i) a one-pot α-fluorination and aldol reaction (αFAR), and (ii) an annulative fluoride displacement (AFD) reaction (Fig. 1C). Considering the ease of access to ketofluorohydrins of general structure **13**, and precedent for the formation of thioribose analogues *via* displacement strategies (*e.g.*, Fig. 1B), we sought to extend our NA synthesis platform to the preparation of thNAs. Importantly, this approach would afford orthogonally protected thNAs and should support the synthesis of C2′-modified thNAs (*e.g.*, **1** and **2**). Here, we report the development of this process, its application to the synthesis of purine and pyrimidine thNAs and a 4′-seleno NA, and the multigram-scale synthesis of 5-methyl 4′-thiouridine.

Results and discussion

Our initial efforts focused on the use of thymine derivative **17**, which was prepared on 100 g scale and is a stable solid that can

[a]*Department of Chemistry, Simon Fraser University, Burnaby, British Columbia, V5A 1S6, Canada. E-mail: rbritton@sfu.ca*

[b]*Department of Process Research & Development, Merck & Co., Inc., Rahway, NJ 07065, USA*

† Electronic supplementary information (ESI) available: The experimental procedures, characterization data and ^{1}H and ^{13}C NMR spectroscopic data. See DOI: https://doi.org/10.1039/d4sc05679e

‡ These authors contributed equally.

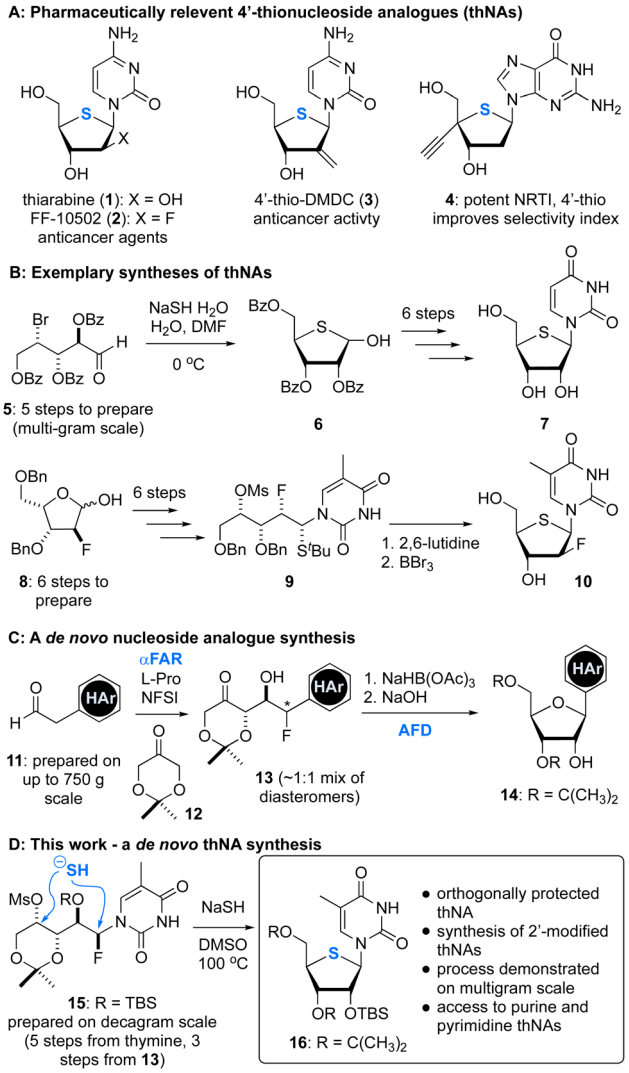
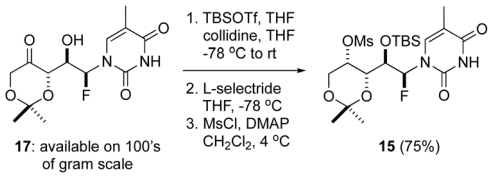


Fig. 1 4′-Thionucleoside analogues (thNAs) and strategies used to prepare these compounds. (A) Examples of pharmaceutically relevnt 4′-thionucleosides. (B) Syntheses of 4′-thionucleosides. (C) A one-pot organocatalytic α-fluorination and aldol reaction (αFAR) and its application to nucleoside synthesis. (D) Synthesis of 4′-thionucleosides from αFAR products. Blue colouring is used to emphasize the endocyclic sulfur atom and two key reactions: (i) αFAR, and (ii) annulative fluoride displacement (AFD).

be stored for months without notable degradation.[14] We have shown that the direct reduction of the ketone function in **17** using $Me_4N{\cdot}BH(OAc)_3$ affords 1,3-*syn* diols with high levels of diastereoselectivity.[14] Application to thNA synthesis would require 1,3-*anti* selectivity in the reduction step owing to the planned invertive cyclization process (*i.e.*, S_N2 reaction at C4′). We found this could be readily achieved by first protecting the secondary alcohol function as a TBS ether and subsequently reducing the carbonyl function with L-selectride.[15] With the mono-TBS protected 1,3-*anti* diol in hand, several activation strategies were examined and ultimately mesylation proved to be optimal. Thus, the fluoro mesylate **15** could be reliably



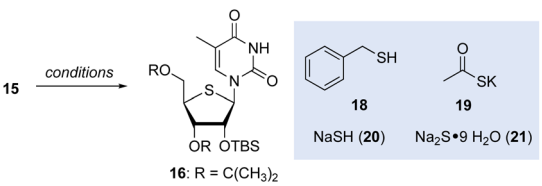
Scheme 1 Synthesis of the fluoromesylate **15**.

prepared in 3 steps from **17** in 75% overall yield following this straightforward process (Scheme 1).

We next explored the reaction of fluoromesylate **15** with various sulfur nucleophiles[15] with an aim to effect a one-pot double displacement and gain direct access to thNA **16** or generate a masked thiolate group and intercept intermediates related to those described by Guindon[13] (*e.g.*, **9**, Fig. 1). Surprisingly, common thiol nucleophiles, including benzyl mercaptan (**18**), potassium thioacetate (**19**), NaSH (**20**) and $Na_2S{\cdot}9H_2O$ (**21**), did not react with mesylate **15** in DMF, even at 90 °C (Table 1, entries 1–4). Further heating of these reactions led to hydrolysis of the nucleobase and degradation. However, we were pleased to find that using freshly recrystallized $Na_2S{\cdot}9H_2O$, the desired double displacement occurred readily at 90 °C, giving the thNA **16** in 50% yield (entry 5). Further optimization ultimately identified DMSO as the optimal solvent for this reaction and we found additionally that in DMSO, NaSH was an efficient sulfur nucleophile that reproducibly gave the thNA **16** in ~60% yield. Thus, following this straightforward sequence, the αFAR product **17** could be converted into thNA **16** in four steps with an overall yield of ~50%. Importantly, owing to the orthogonal protection of the secondary alcohol functions in **16**, this route should support the synthesis of thNA functionalized at C2′ (see below).

Having established a route to the thNA **16**, we next evaluated the scope of this thNA synthesis starting with the readily available TBS-protected fluorohydrins **22a–d** (Fig. 2). The fluorohydrins can be prepared in 2 or 3 steps from the commercial

Table 1 A double displacement reaction to access thNA **16**



Entry	Thiol	Solvent	Temp.	Yield (%)
1	**18**	DMF	90 °C	0
2	**19**	DMF	90 °C	0
3	**20**	DMF	90 °C	0
4	**21**	DMF	90 °C	0
5	**21**[a]	DMF	90 °C	50
6	**20**	DMSO	100 °C	61

[a] Freshly recrystalized **21** (Na_2S).

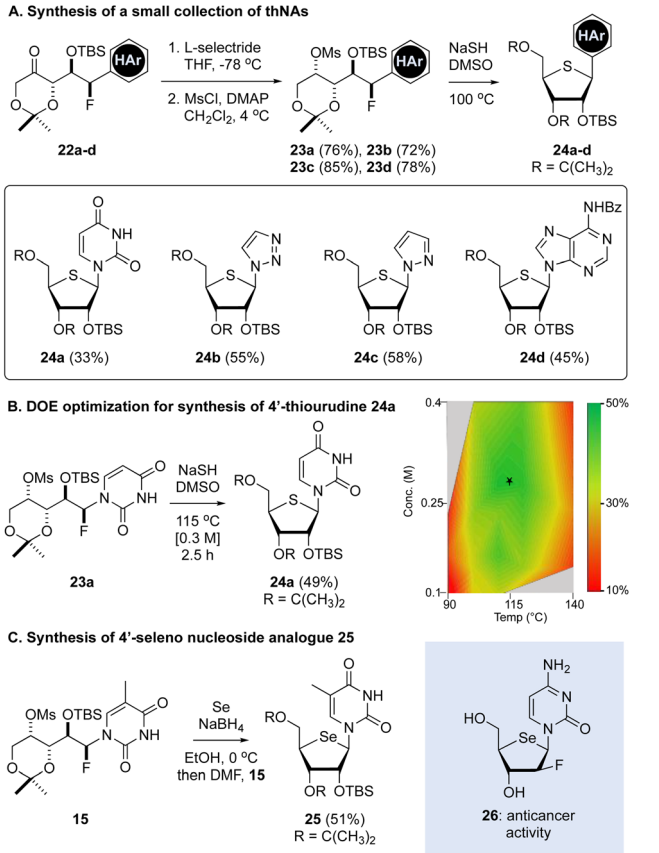


Fig. 2 Synthesis of a purine, pyrimidine and other thNAs. (A) Examples of pyrimidine, purine and other C4′-thNAs produced from αFAR products. (B) Optimization of reaction temperature and concentration for the production of **24a**. (C) Synthesis of the 4′-seleno nucleoside analogue **25**.

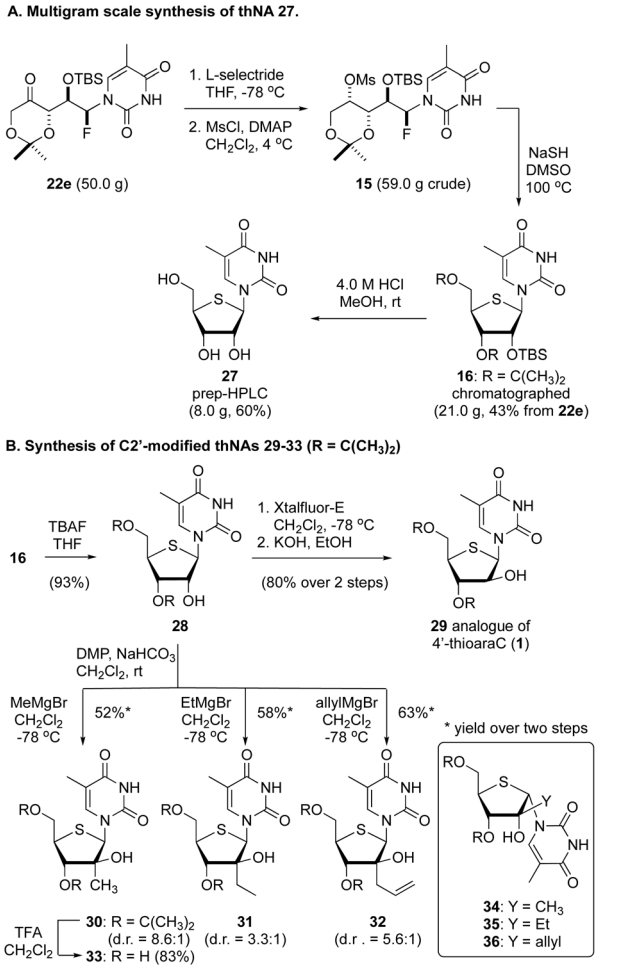


Fig. 3 Large scale synthesis of thNA **27** and synthesis of C2′-modified thNAs **29–36**. (A) A multigram scale synthesis of the thNAs **16** and **27**. (B) Synthesis of C2′-modified thNAs.

nucleobase/heterocycle,[14,16] though in the case of uracil- and triazole-containing fluorohydrins these were produced as inseparable mixtures of *syn*- and *anti*-fluorhydrins as described.[12] These diastereomers were separable following TBS protection and mesylate formation (see ESI†). We further demonstrated that this process was compatible with pyrazole and benzoyl-protected adenosine, each of which gave the corresponding thNAs **24a–d** in good overall yield. This reaction sequence was further optimized for execution with minimal chromatography and this sequence of steps could be executed as a through process with little impact on the overall yield. Due to challenges in accessing the corresponding cytosine and guanine aldol products (*e.g.*, **22**, HAr = cytosine or guanine), synthesis of the corresponding thNAs was not explored.

Unfortunately, reaction of the uracil containing fluoro mesylate **23a** with NaSH led to substantial cleavage of the uracil function and degradation, with uracil being released at a similar rate as thNA **24a** formation. Thus, using the standard reaction conditions (Table 1, entry 6), the 4′-thiouridine **24a** was produced in 33% yield. In an effort to improve on this result, we conducted Design of Experiment (DOE) optimization,[17] focusing on the relationship between temperature, concentration, and time (Fig. 2B). Here, we found a correlation between concentration and time, with a maximum yield of 47% at a concentration of ~0.3 M after 3 hours at 115 °C. Using these optimized conditions, the 4′-thiouridine **24a** could be produced in 4 steps and ~40% overall yield from the TBS-protected keto fluoride **22a**. To demonstrate the versatility of this approach, we also reacted the thymine derivative **15** with NaSeH, generated in EtOH by the reduction of Se with $NaBH_4$. As highlighted in Fig. 2C, this reaction gave the 4′-selenonucleoside analogue **25**, which is an analogue of the known 4′-selenouridine.[18] Notably, selenonucleosides (*e.g.*, **26**[19]) have also attracted attention as anticancer agents.

To assess the scalability of this thNA synthesis, we additionally executed the process starting with 50.0 g of the protected αFAR product **22e** (Fig. 3a). Without additional optimization we found that the sequence of reduction and mesylation proceeded in good overall yield, affording 59.0 g of the mesylate **15**. From here, reaction with NaSH in DMSO at 100 °C gave 21.0 g of the thNA **16**, which was purified by flash column chromatography. Removal of the silyl and acetonide

protecting groups by treatment with 4 M HCl in MeOH then afforded 8.0 g of 4′-thio-5-methyluridine **27**.[20]

Finally, considering that this process affords orthogonally protected thNAs (*e.g.*, **16**), we investigated the selective C2′-functionalization of thymine thNA **16**. As highlighted in Fig. 3B, removal of the TBS protecting group afforded the 2′-OH thNA **28** in excellent yield. From here, a 2-step sequence involving formation of the anhydro thNA and hydrolysis gave the arabino-configured thNA **29** in good overall yield. Notably, the corresponding triol (*i.e.*, deprotected) has demonstrated activity as low as 0.77 μg mL^{-1} against HSV-1.[21] Additionally, despite concerns regarding anomerization of 2′-keto thNAs,[22] we found that oxidation of compound **28** using Dess Martin periodinane buffered with $NaHCO_3$ in CH_2Cl_2 gave clean conversion to the corresponding 2′-keto derivative. The use of $NaHCO_3$ in this reaction proved critical, and several other standard oxidation conditions failed to provide the 2′-ketone in any reasonable yield. This latter material proved to be unstable on all stationary phases used for chromatographic purifications and thus the crude material was reacted directly with Grignard reagents to afford a small collection of previously unreported C2′-modified thNAs **30–36**. In all cases, the arabino-configured stereoisomer was the major product, and the minor product was that derived from epimerization at C1′ prior to reaction with the Grignard reagent (*e.g.*, **34–36**, see inset). Similar results have been reported by Matsuda.[22] The use of more hindered Grignard reagents (*e.g.*, ^{c}PrMgBr or ^{i}PrMgBr) resulted in larger amounts of C1′-epimerization. Removal of the acetonide protecting group from **30** using TFA gave the corresponding triol **33** in excellent yield. Notably, Liotta has recently reported related, ribose-configured C2′-modified thNAs.[23]

Conclusions

In summary, we report a streamlined process for the synthesis of various thNAs that exploits the ready availability of keto-fluorohydrin aldol products. Importantly, the resulting thNAs are orthogonally protected, which enables synthesis of C2′-modified thNAs. This overall 7-step sequence was also demonstrated on multi-gram scale in the preparation of 5-methyl-4′-thiouridine (**27**) suggesting potential utility for larger scale, process research efforts. Importantly, the demonstration that purine and pyrimidine thNAs, 4′-seleno NAs, and C2′-modified thNAs can be readily prepared following straightforward strategies suggests that this new approach should inspire and support medicinal chemistry efforts in this area.

Data availability

The experimental procedures, characterization data and ^{1}H and ^{13}C NMR spectroscopic data generated in this study are provided in the ESI.†

Author contributions

R. B., C. L. and M. N. designed the study, R. B., C. L. and M. N. developed the synthetic plans and C. L., E. F., M. N. and B. S. optimized and executed the synthesis of all new compounds. S. S. and L.-C. C. supervised the large (multi-gram) scale reactions. R. B. and C. L. prepared the manuscript text. All authors contributed to the ESI.†

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

Financial support from the Natural Sciences and Engineering Research Council (NSERC) of Canada was received by R. B. (Discovery Grant: 2019-06368). R. B. acknowledges support from the Canadian Glycomics Network (Strategic Initiatives Grant CD-81), the Consortium de Recherche Biopharmaceutique (CQDM Quantum Leap Grant), Merck & Co., Inc., Rahway, NJ, USA.

Notes and references

1 (*a*) L. P. Jordheim, D. Durantel, F. Zoulim and C. Dumontet, Advances in the Development of Nucleoside and Nucleotide Analogues for Cancer and Viral Diseases, *Nat. Rev. Drug Discovery*, 2013, **12**(6), 447–464; (*b*) M. K. Yates and K. L. Seley-Radtke, The Evolution of Antiviral Nucleoside Analogues: A Review for Chemists and Non-Chemists. Part II: Complex Modifications to the Nucleoside Scaffold, *Antiviral Res.*, 2019, **162**, 5–21; (*c*) K. L. Seley-Radtke and M. K. Yates, The Evolution of Nucleoside Analogue Antivirals: A Review for Chemists and Non-Chemists. Part 1: Early Structural Modifications to the Nucleoside Scaffold, *Antiviral Res.*, 2018, **154**, 66–86.

2 (*a*) N. Tsesmetzis, C. B. J. Paulin, S. G. Rudd and N. Herold, Nucleobase and Nucleoside Analogues: Resistance and Re-Sensitisation at the Level of Pharmacokinetics, Pharmacodynamics and Metabolism, *Cancers*, 2018, **10**(7), 240; (*b*) C. M. Galmarini, J. R. Mackey and C. Dumontet, Nucleoside analogues: mechanisms of drug resistance and reversal strategies, *Leukemia*, 2001, **15**(6), 875–890.

3 E. J. Reist, D. E. Gueffroy and L. Goodman, Synthesis of 4-Thio-D- and -L-ribofuranose and the Corresponding Adenine Nucleosides, *J. Am. Chem. Soc.*, 1964, **86**(24), 5658–5663.

4 (*a*) W. R. Waud, K. S. Gilbert and J. A. Secrist, Preclinical Antitumor Activity of Thiarabine in Human Leukemia and Lymphoma Xenograft Models, *Nucleosides, Nucleotides Nucleic Acids*, 2012, **31**(9), 647–660; (*b*) P. Gunaga, H. R. Moon, W. J. Choi, D. H. Shin, J. G. Park and L. S. Jeong, Recent advances in 4′-thionucleosides as potential antiviral and antitumor agents, *Curr. Med. Chem.*, 2004, **11**(19), 2585–2637; (*c*) Y. Yoshimura, Y. Saito, Y. Natori and H. Wakamatsu, Synthesis of 4′-Thionucleosides as Antitumor and Antiviral Agents, *Chem. Pharm. Bull.*, 2018, **66**(2), 139–146; (*d*) Y. Yoshimura, M. Endo, S. Miura and S. Sakata, An Alternative Synthesis of the Antineoplastic Nucleoside 4′-ThioFAC and Its

Application to the Synthesis of 4′-ThioFAG and 4′-Thiocytarazid, *J. Org. Chem.*, 1999, **64**(21), 7912–7920; (*e*) M. Yokoyama, Synthesis and Biological Activity of Thionucleosides, *Synthesis*, 2000, **2000**(12), 1637–1655.

5 M. I. Elzagheid, M. Oivanen, R. T. Walker and J. A. Secrist, Kinetics for the Acid-Catalyzed Hydrolysis of Purine and Cytosine 2′-Deoxy-4′-thionucleosides, *Nucleosides*, 1999, **18**(2), 181–186.

6 W. B. Parker, W. R. Waud and J. A. Secrist, 3rd, Thiarabine, 1-(4-Thio-β-D-arabinofuranosyl)cytosine. A Deoxycytidine Analog With Excellent Anticancer Activity, *Curr. Med. Chem.*, 2015, **22**(34), 3881–3896.

7 M. Shinji, K. Chihaya, H. Tamami, S. Kazunori, Y. Takayuki, U. Rena, I. Miki, K. Nobuko, N. Hiroki, K. Hidenobu, M. Takuya, F. Hideyasu, J. P. Linda, S. Yasuhiro and I. Hiroyuki, FF-10502, an Antimetabolite with Novel Activity on Dormant Cells, Is Superior to Gemcitabine for Targeting Pancreatic Cancer Cells, *J. Pharmacol. Exp. Ther.*, 2018, **366**(1), 125.

8 S. Miura, M. Tanaka, Y. Yoshimura, H. Satoh, S. Sakata, H. Machida, A. Matsuda and T. Sasaki, In Vitro and in Vivo Antitumor Activity of a Novel Nucleoside, 4′-Thio-2′-deoxy-2′-methylidenecytidine, *Biol. Pharm. Bull.*, 1996, **19**(10), 1311–1315.

9 (*a*) K. Haraguchi, H. Shimada, K. Kimura, G. Akutsu, H. Tanaka, H. Abe, T. Hamasaki, M. Baba, E. A. Gullen, G. E. Dutschman, Y.-C. Cheng and J. Balzarini, Synthesis of 4′-Ethynyl-2′-deoxy-4′-thioribonucleosides and Discovery of a Highly Potent and Less Toxic NRTI, *ACS Med. Chem. Lett.*, 2011, **2**(9), 692–697; (*b*) K. Haraguchi, H. Shimada, H. Tanaka, T. Hamasaki, M. Baba, E. A. Gullen, G. E. Dutschman and Y.-C. Cheng, Synthesis and Anti-HIV Activity of 4′-Substituted 4′-Thiothymidines: A New Entry Based on Nucleophilic Substitution of the 4′-Acetoxy Group, *J. Med. Chem.*, 2008, **51**(6), 1885–1893.

10 R. Maeda, N. Saito-Tarashima, H. Wakamatsu, Y. Natori, N. Minakawa and Y. Yoshimura, Synthesis and Properties of 4′-ThioLNA/BNA, *Org. Lett.*, 2021, **23**(10), 4062–4066.

11 S. Westarp, C. M. M. Benckendorff, J. Motter, V. Röhrs, Y. S. Sanghvi, P. Neubauer, J. Kurreck, A. Kurreck and G. J. Miller, Biocatalytic Nucleobase Diversification of 4′-Thionucleosides and Application of Derived 5-Ethynyl-4′-thiouridine for RNA Synthesis Detection, *Angew. Chem., Int. Ed.*, 2024, **63**(33), e202405040.

12 M. Guinan, N. Huang, C. S. Hawes, M. A. Lima, M. Smith and G. J. Miller, Chemical synthesis of 4′-thio and 4′-sulfinyl pyrimidine nucleoside analogues, *Org. Biomol. Chem.*, 2022, **20**(7), 1401–1406.

13 (*a*) D. Chapdelaine, B. Cardinal-David, M. Prevost, M. Gagnon, I. Thumin and Y. Guindon, A stereoselective approach to nucleosides and 4′-thioanalogues from acyclic precursors, *J. Am. Chem. Soc.*, 2009, **131**(47), 17242–17245; (*b*) S. Dostie, M. Prévost, P. Mochirian, K. Tanveer, N. Andrella, A. Rostami, G. Tambutet and Y. Guindon, Diastereoselective Synthesis of C2′-Fluorinated Nucleoside Analogues Using an Acyclic Approach, *J. Org. Chem.*, 2016, **81**(22), 10769–10790.

14 (*a*) M. Meanwell, S. M. Silverman, J. Lehmann, B. Adluri, Y. Wang, R. Cohen, L.-C. Campeau and R. Britton, A short de novo synthesis of nucleoside analogs, *Science*, 2020, **369**(6504), 725; (*b*) E. K. Davison, D. A. Petrone, M. Meanwell, M. B. Nodwell, S. M. Silverman, L.-C. Campeau and R. Britton, Practical and concise synthesis of nucleoside analogs, *Nat. Protoc.*, 2022, **17**(9), 2008–2024.

15 C. Grondal and D. Enders, A Direct Organocatalytic Entry to Selectively Protected Aldopentoses and Derivatives, *Adv. Synth. Catal.*, 2007, **349**(4-5), 694–702.

16 C. Huxley, C. Lucas, J. M. M. Bruno, M. J. Anketell, E. K. Davison, G. Muir, M. B. Nodwell, M. Meanwell, S. M. Silverman, R. Britton and L.-C. Campeau, Efficient protocol for the preparation of α-heteroaryl acetaldehydes, *Can. J. Chem.*, 2023, **101**(7), 487–490.

17 S. A. Weissman and N. G. Anderson, Design of Experiments (DoE) and Process Optimization. A Review of Recent Publications, *Org. Process Res. Dev.*, 2015, **19**(11), 1605–1633.

18 P. K. Sahu, G. Kim, J. Yu, J. Y. Ahn, J. Song, Y. Choi, X. Jin, J.-H. Kim, S. K. Lee, S. Park and L. S. Jeong, Stereoselective Synthesis of 4′-Selenonucleosides via Seleno-Michael Reaction as Potent Antiviral Agents, *Org. Lett.*, 2014, **16**(21), 5796–5799.

19 L. S. Jeong, D. K. Tosh, W. J. Choi, S. K. Lee, Y.-J. Kang, S. Choi, J. H. Lee, H. Lee, H. W. Lee and H. O. Kim, Discovery of a New Template for Anticancer Agents: 2′-deoxy-2′-fluoro-4′-selenoarabinofuranosyl-cytosine (2′-F-4′-Seleno-ara-C), *J. Med. Chem.*, 2009, **52**(17), 5303–5306.

20 V. Pejanović, Z. Stokić, B. Stojanović, V. Piperski, M. Popsavin and V. Popsavin, Synthesis and biological evaluation of some novel 4′-Thio-l-ribonucleosides with modified nucleobase moieties, *Bioorg. Med. Chem. Lett.*, 2003, **13**(11), 1849–1852.

21 Y. Yoshimura, M. Watanabe, H. Satoh, N. Ashida, K. Ijichi, S. Sakata, H. Machida and A. Matsuda, A Facile, Alternative Synthesis of 4′-Thioarabinonucleosides and Their Biological Activities, *J. Med. Chem.*, 1997, **40**(14), 2177–2183.

22 D. Kaga, N. Minakawa and A. Matsuda, Nucleosides and Nucleotides. 232. Synthesis of 2′-C-Methyl-4′-thiocytidine: Unexpected Anomerization of the 2′-Keto-4′-thionucleoside Precursor, *Nucleosides, Nucleotides Nucleic Acids*, 2005, **24**(10–12), 1789–1800.

23 Z. W. Dentmon, T. M. Kaiser and D. C. Liotta, Synthesis and Antiviral Activity of a Series of 2′-C-Methyl-4′-thionucleoside Monophosphate Prodrugs, *Molecules*, 2020, **25**, 5165.