


 Cite this: *RSC Adv.*, 2025, 15, 45387

Biodegradable graphene nanocomposites as functional biomaterials: a review of their role in controlled drug delivery and tissue engineering

 Md. Mohiuddin,^a Md. Mahbubur Rahman,^{id} *^a Md. Nizam Uddin,^{*b} Rakib Hasan^c and Ismail Rahman^{id} *^d

Biodegradable graphene nanocomposites (BGNs) have emerged as highly versatile platforms at the intersection of nanotechnology, materials science, and biomedicine. By combining the exceptional physicochemical properties of graphene-based materials with the biocompatibility and environmental sustainability of biodegradable polymers, BGNs constitute a unique class of materials for advanced biomedical applications. Key features of BGNs, such as high surface area, tunable surface chemistry, excellent mechanical strength, and the ability to interface effectively with biological systems, make them promising candidates for controlled drug delivery and tissue engineering. In drug delivery, BGNs facilitate high drug loading and enable spatially and temporally controlled release, which can be triggered by internal or external stimuli, thereby improving therapeutic efficiency while minimizing side effects. In tissue engineering, the mechanical robustness and customizable structure of BGNs support cellular attachment, proliferation, and differentiation, rendering them suitable as scaffolds for regenerating bone, cartilage, skin, and neural tissues. This review explores recent advancements in the fabrication techniques and biomedical applications of BGNs, emphasizing their role in achieving precise drug delivery and effective tissue regeneration.

 Received 23rd August 2025
 Accepted 23rd October 2025

DOI: 10.1039/d5ra06280b

rsc.li/rsc-advances

1 Introduction

1.1 Background and rationale

Conventional drug administration systems present several limitations, including poor bioavailability, uncontrolled release kinetics, off-target effects, and systemic toxicity.^{1,2} Similarly, traditional materials used in tissue repair often lack the requisite biocompatibility and mechanical properties, leading to failed integration with native tissue, implant failure, and adverse immune responses.³ These significant clinical challenges have motivated researchers to develop advanced materials capable of enhancing therapeutic efficacy, promoting predictable tissue regeneration, and minimizing adverse effects.

In recent decades, biomaterials science has progressed significantly, leading to the introduction of biodegradable polymers for biomedical applications. These polymers offer

favorable properties, including minimal toxicity and high biocompatibility.⁴ In drug delivery, they enable stimuli-responsive (*e.g.*, to temperature, pH, light) controlled release of therapeutic agents,⁵ while in tissue engineering, they provide a temporary structural support that degrades in coordination with new tissue formation.⁶ However, the utility of many biodegradable polymers is constrained by inherent limitations in mechanical strength, electrical conductivity, or targeted functionality, properties which are essential for more demanding applications like load-bearing tissue repair or electro-active tissue stimulation.^{4,7}

To overcome these shortcomings, research has shifted toward nanotechnology, which enables the engineering of materials at the molecular scale. Among various nanomaterials, graphene and its derivatives, graphene oxide (GO) and reduced graphene oxide (rGO), have gained substantial attention. Their exceptional characteristics, including an extremely high surface-area-to-volume ratio, low weight, excellent thermal and mechanical properties, and the capacity to adsorb drugs or bioactive molecules *via* non-covalent interactions, make them ideal reinforcing and functional agents.⁸ However, the standalone use of pristine graphene-based materials faces its own challenges, including potential cytotoxicity,⁹ poor physiological stability, and a tendency to aggregate in aqueous biological environments, which can limit their practical application.¹⁰

^aDepartment of Mechanical Engineering, Khulna University of Engineering & Technology, Khulna 9203, Bangladesh. E-mail: mahbub_rahman@me.kuet.ac.bd

^bJames C. Morriss Division of Engineering, Texas A&M University-Texarkana, 7101 University Ave, Texarkana, TX 75503, USA. E-mail: muddin@tamut.edu

^cDepartment of Mechanical, Aerospace, and Industrial Engineering, The University of Texas at San Antonio, 1 UTSA Circle, San Antonio, TX 78249, USA

^dInstitute of Environmental Radioactivity, Fukushima University, 1 Kanayagawa, Fukushima City, Fukushima 960-1296, Japan. E-mail: immrahman@ipc.fukushima-u.ac.jp



Consequently, a synergistic approach has emerged: incorporating these nanomaterials into biocompatible polymeric matrices to improve their biostability, biocompatibility, and overall functionality. This has led to the development of BGNs, a class of materials that strategically combines the functional advantages of graphene with the safety and biodegradability of polymers. In these composites, graphene provides significant mechanical reinforcement, high drug-loading capacity, and responsiveness to external stimuli. Simultaneously, the polymer matrix ensures biocompatibility and produces safe, absorbable degradation byproducts. Furthermore, BGNs are valued for their sustainable nature, especially when fabricated with natural polymers, addressing the growing need for environmentally conscious medical technologies.

1.2 Scope and methodology

Numerous reviews have addressed the biomedical applications of biodegradable polymers and graphene-based materials separately. Comprehensive analyses exist for chitosan-functionalized GO in cancer therapy,¹¹ GO-based hydrogels for drug delivery,¹² natural polymeric nanobiocomposites for anti-cancer therapeutics,¹³ and polysaccharide-based nanomedicines for cancer immunotherapy.¹⁴ Other reviews have covered topics such as plasma modification of drug delivery systems,¹⁵ bacterial cellulose for wound dressings,¹⁶ 3D bioprinting with chitosan,¹⁷ and nanostructured composites for bone regeneration.¹⁸

Although these articles provide valuable insights, most focus on either drug delivery or tissue engineering, and many concentrate solely on either biodegradable polymers or graphene. Reviews that explore the synergy of polymer–graphene nanocomposites in both drug delivery and tissue engineering are limited. This highlights a gap in the literature: a comprehensive review that simultaneously addresses both applications using BGNs and explores advancements in their fabrication is lacking. As shown in Fig. 1, research on BGNs for drug delivery and tissue engineering has gained significant attention since 2014, indicating a rapidly emerging field that warrants an in-depth, integrated review.

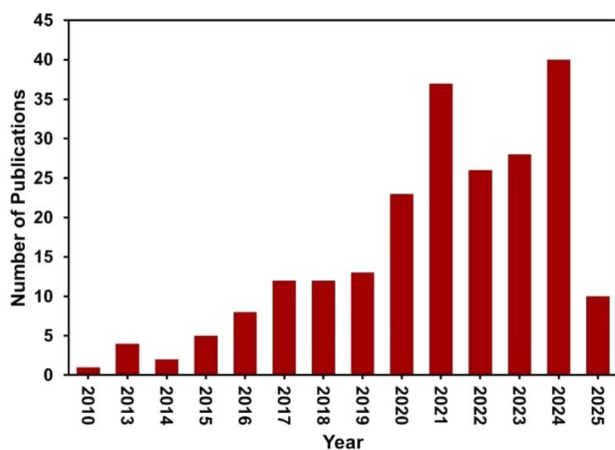


Fig. 1 Annual number of publications on BGNs since 2010. Data sourced from <https://scopus.com>.

This review bridges this gap by systematically examining both the fabrication techniques and the dual applications of BGNs in controlled drug delivery and tissue engineering. A comprehensive literature search was conducted using the Scopus database with the keywords: (“biopolymer” OR “natural polymer” OR “biodegradable polymer”) AND (“graphene” OR “graphene oxide” OR “rGO”) AND (“nanocomposite” OR “nanomaterial”) AND (“drug delivery” OR “controlled release”) OR (“tissue engineering” OR “regenerative medicine”). This query yielded 221 records published between 2010 and June 2025. Following the PRISMA guidelines (Fig. 2), records were screened, and 66 primary research articles meeting all inclusion criteria were selected for synthesis. These core studies, supplemented by additional references for background context, form the basis of this review.

2 Foundational materials

2.1 Graphene and its oxidized derivatives

Graphene is a single layer of carbon atoms arranged in a two-dimensional honeycomb lattice (Fig. 3a). Each carbon atom is sp^2 hybridized, forming strong covalent bonds approximately 0.142 nm in length.¹⁹ This unique structure imparts exceptionally high electrical conductivity, mechanical strength (with a Young's modulus of ~ 1 TPa), thermal conductivity, and a large theoretical specific surface area (~ 2630 m² g⁻¹). Each carbon atom in graphene is covalently bonded to three neighboring atoms, forming a robust and stable hexagonal framework. This sp^2 bonding provides delocalized π -electrons across the basal plane, which are responsible for graphene's high conductivity and chemical inertness.²⁰ While chemically inert, its surface can be functionalized through covalent or non-covalent methods to attach biomolecules or drugs, though its intrinsic hydrophobicity can be a challenge for biological dispersion.^{21,22} Such functionalization exploits graphene's π -system for noncovalent π - π stacking with aromatic drug molecules, while covalent grafting (e.g., carbodiimide-mediated amidation) allows linkage of $-COOH$ groups on GO with $-NH_2$ groups of polymers like chitosan. Similarly, hydroxyl and epoxy groups on GO readily form hydrogen bonds with hydrophilic polymers (e.g., PVA, cellulose), enabling stable polymer–graphene nanocomposites. These interfacial interactions dictate dispersion, mechanical reinforcement, and drug release behavior. Such functionalization strategies are crucial for biomedical applications because they allow the attachment of drugs, peptides, or targeting ligands, improving dispersibility and specificity.²³ Common synthesis methods include mechanical exfoliation, which yields high-quality flakes but lacks scalability, and chemical vapor deposition, which allows for large-area film production.²⁴ Other routes include epitaxial growth on SiC substrates,²⁵ unzipping of carbon nanotubes,²⁶ and chemical reduction of GO.²⁷ Each method produces graphene with different levels of purity, layer control, and defect density, which in turn affect its chemical reactivity and biomedical performance.

GO is derived from graphite *via* aggressive oxidation and subsequent exfoliation. Its structure contains a high density of oxygen-containing functional groups (hydroxyl, epoxy, and



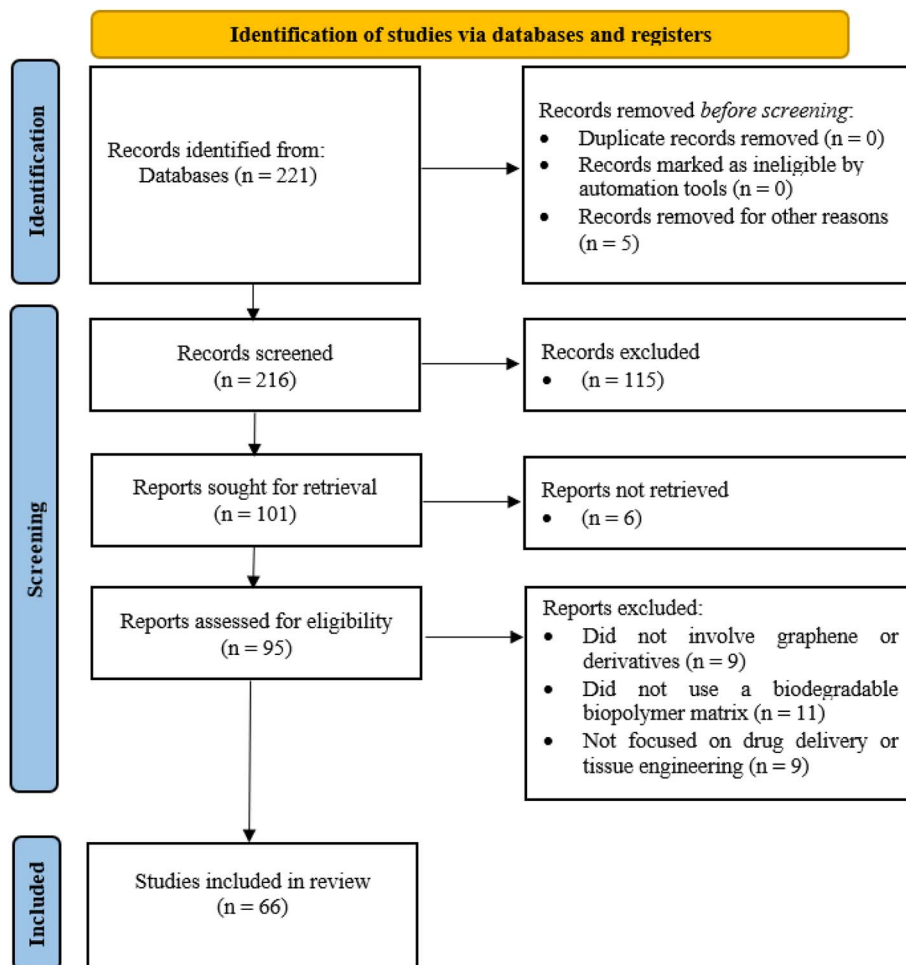


Fig. 2 PRISMA flow diagram illustrating the study selection process for the systematic review.

carboxyl groups) that disrupt the planar sp^2 network, introducing sp^3 -hybridized carbon atoms and rendering it electrically insulating but highly hydrophilic and easily dispersible in water (Fig. 3b).²² This aqueous dispersibility and the abundance of functional groups for further chemical modification make GO particularly attractive for biomedical applications such as drug delivery, biosensing, and tissue engineering.²⁸ The type and density of oxygen functionalities depend on the oxidation method used (Hummers', Brodie's, or Staudenmaier's), which not only influence dispersibility but also toxicity and stability.^{29,30} Hummers' method involves strong acids (typically sulfuric acid) and oxidants like potassium permanganate to rapidly oxidize graphite, producing GO with various oxygen functional groups.³¹ Brodie's method, one of the earliest, uses fuming nitric acid and potassium chlorate, resulting in slower oxidation and more defects.³² Staudenmaier's method improves on Brodie's by combining concentrated sulfuric and nitric acids with potassium chlorate for faster oxidation and higher oxygen content, but it produces hazardous chlorine dioxide gas.³³ These oxygen functionalities ($-OH$, $-COOH$, $-O-$) provide chemical handles for bonding with polymers: $-COOH$ groups undergo esterification or amidation with polymeric hydroxyl/

amine groups, while hydroxyl and epoxy moieties participate in hydrogen bonding and ionic crosslinking. Such interactions govern composite stability, swelling, and enzymatic degradation in physiological environments.

rGO is produced by removing a significant portion of the oxygen-containing groups from GO through chemical, thermal, or electrochemical reduction (Fig. 3c). This process partially restores the sp^2 carbon network, improving electrical conductivity to a level between that of pristine graphene and GO, although residual defects remain.^{19,34} Reduction routes include chemical reductants (*e.g.*, hydrazine, sodium borohydride, or green alternatives such as ascorbic acid), high-temperature annealing, and electrochemical reduction. Each pathway affects the residual oxygen content and defect density, which determine conductivity, dispersibility, and biocompatibility.³⁵ Chemical reduction of GO typically involves the use of strong reducing agents like hydrazine or sodium borohydride, which react with oxygen-containing functional groups (such as epoxides, hydroxyls, and carboxyls) to remove them from the GO surface. This significantly restores the conjugated sp^2 carbon network but can introduce defects and leave toxic residues, negatively affecting electrical and structural properties.³⁵



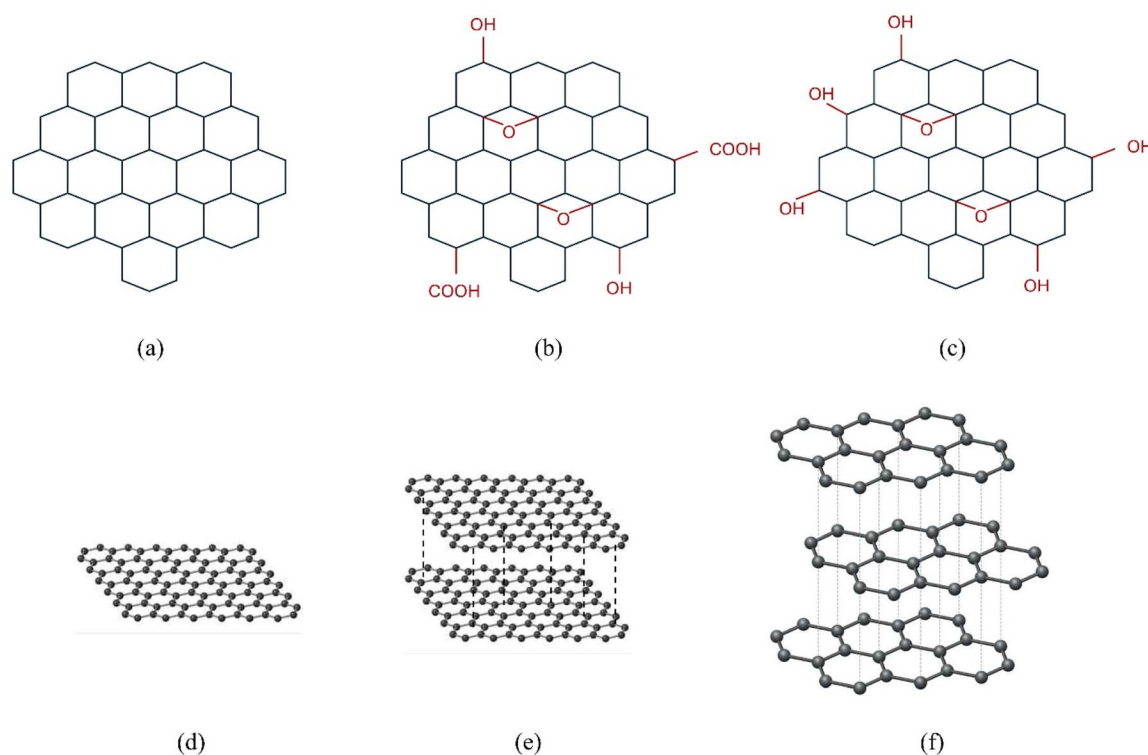


Fig. 3 Schematic representations of (a) pristine graphene, (b) graphene oxide, (c) reduced graphene oxide, and 3D models of (d) single-layer graphene, (e) bi-layer graphene, and (f) tri-layer graphene stack.

Green reductants like ascorbic acid operate through similar mechanisms but involve milder redox reactions, offering moderate deoxygenation while maintaining better biocompatibility and aqueous dispersibility.³⁶ Thermal annealing, on the other hand, works by heating GO to high temperatures (typically 500–1100 °C) under inert or reducing atmospheres (*e.g.*, argon or hydrogen), which causes decomposition of oxygen functional groups and reconstructs sp^2 domains. This method yields highly conductive rGO but often at the expense of scalability and may cause layer restacking.³⁷ Finally, electrochemical reduction involves applying a potential across GO films in an electrolyte solution, triggering electron transfer that selectively removes oxygen groups. This technique avoids harsh chemicals and allows fine-tuning of reduction levels, producing rGO with controlled surface chemistry and good dispersibility.³⁸ The residual oxygen groups on rGO still permit limited hydrogen bonding or ionic interactions with polymers, while defect sites act as nucleation points for covalent grafting. Thus, tuning the reduction level enables control over degradation rate, electrical conductivity, and mechanical reinforcement of the resulting nanocomposites. The resulting material offers a tunable balance of conductivity and dispersibility, making rGO a cost-effective and versatile option for similar biomedical applications.^{19,24}

Graphene materials can exist as monolayer, bilayer, trilayer, or multilayer sheets (Fig. 3d–f). As the number of layers increases, the specific surface area and some unique quantum electronic properties decrease, but the potential for

functionalization for biomedical use remains a key feature.^{22,24} Specifically, one, two, and three layers are termed monolayer, bilayer, and trilayer graphene, respectively, while 5–10 layers are considered few-layer graphene and 20–30 layers as multilayer graphene (nanocrystalline thin graphite).

2.2 Common biodegradable polymers

Biodegradable polymers, derived from both natural and synthetic sources, are central to the development of BGNs. They provide essential biocompatibility and degrade into non-toxic byproducts that can be safely metabolized or excreted by the body, aligning with the need for sustainable and eco-friendly biomedical solutions.³⁹ When combined with graphene, these polymers form composites with enhanced mechanical strength, electrical conductivity, and tailored functionality, making them ideal for drug delivery and tissue engineering.⁴⁰

Table 1 provides an overview of common biodegradable polymers used in BGNs, detailing their origin, formulation possibilities, and key functional properties.

2.3 Polymer–graphene interfacial interactions

The interactions at the polymer–graphene interface are a key determinant of the performance of BGNs, and they have a significant impact on important parameters like dispersion, mechanical augmentation, degradation, and drug release behaviors. Usually, these interfacial interactions are divided into two categories: covalent bonds which involve the sharing of



Table 1 Overview of biodegradable polymers for drug delivery and tissue engineering applications

| Polymer | Origin/type | Formulation possibilities | Key functional properties | References |
|--|--|---|---|------------|
| (A) Natural proteins | | | | |
| Silk fibroin (SF) | Silkworm-derived (<i>Bombyx mori</i>) | Fabricated as hydrogels, porous sponges/scaffolds, films, fibers, and nanoparticles | High mechanical strength; tunable degradation; minimal immunogenicity | 41 and 42 |
| Gelatin | Denatured collagen from animal bone/skin | Used in hydrogels, porous scaffolds, films, microspheres, and electrospun nanofibers | Non-toxic and non-immunogenic; promotes cell adhesion (collagen-mimetic RGD sequences) | 43 and 44 |
| Collagen (COL) | Major ECM protein from connective tissues | Sponges/fibrous scaffolds, hydrogels, sheets (<i>via</i> molding, electrospinning, 3D bioprinting) | Low immunogenicity; contains natural cell-binding sites (RGD) and directs cell behavior | 45 |
| Zein | Corn protein (GRAS excipient) | Films/coatings, nanoparticles, electrospun fibers, scaffolds | Biodegradable, biocompatible protein; hydrophobic (alcohol-soluble) enabling controlled drug release; good film-former with moderate mechanical strength; supports drug-loaded membranes and tissue scaffolds | 46 |
| (B) Natural polysaccharides and derivatives | | | | |
| Chitosan (CS) | Derived from chitin in crustacean shells | Processed into hydrogels, nanoparticles, nanofibers, membranes, films, and 3D scaffolds | Inherently mucoadhesive and hemostatic; antimicrobial; low immune rejection | 47–49 |
| Alginate | From brown seaweed | Formed into ionically crosslinked hydrogels/gels, porous foams/sponges, microcapsules/microspheres, fibers, and films | Low-toxicity; undergoes mild Ca ²⁺ -induced gelation; yields high-porosity, tunable-stiffness networks | 50 |
| Cellulose nanofibers | Plant-derived nanoscale fibers | Fabricated into ionically crosslinked hydrogels, freeze-dried porous scaffolds/aerogels and composite matrices | Extremely high surface area and tensile strength; forms interconnected porous networks that support cell growth and may confer antibacterial effects | 51 |
| Starch | Plant polysaccharide (amylose/amylopectin) | Hydrogels (drug-loaded matrices), electrospun nanofibrous scaffolds, films, microparticles | Highly hydrophilic (water-absorbing); low-cost; promotes cell proliferation and wound healing | 52 and 53 |
| Agarose | Red algae-derived galactose polymer | Thermoresponsive hydrogels (injectable gels, bioinks), cryogels, sponges/scaffolds (<i>e.g.</i> 3D-printed) | Reversible gelation (thermo-sensitive sol-gel transition); good mechanical strength and high water retention; inert | 54 and 55 |
| Carboxymethyl cellulose (CMC) | Cellulose-derived anionic polysaccharide | Hydrogels, films, 3D porous scaffolds (<i>e.g.</i> for bone or soft tissue), wound dressings | Very hydrophilic (forms swellable gels); thixotropic (viscoelastic) with high viscosity | 56 |
| Hyaluronic acid | ECM glycosaminoglycan | Hydrogels (injectable gels, bioinks, cryogels), 3D-printed scaffolds, composite matrices; also viscous solutions/films (<i>e.g.</i> eye drops, dermal fillers) | Mucoadhesive (interacts with tissues like cartilage, skin); highly hydrophilic (excellent water retention); tunable viscoelasticity and porosity | 57 |
| Arabinoxylan (ARX) | Plant hemicellulose | Hydrogel matrices (films, injectable gels, tablets, capsules) and particulate systems (micro/nanogels) | Polysaccharide; forms gel networks for controlled release; shown to promote wound healing | 58 and 59 |
| β-Glucan (BG) | Found in cereal grains, fungi, yeast cell walls | Porous scaffolds (<i>e.g.</i> freeze-dried foams, nanocomposites with hydroxyapatite) and hydrogels | Highly hydrophilic (water-adsorbing) polymer; supports cell attachment and proliferation | 60 |
| Guar gum (GG) | From guar bean (<i>Cyamopsis tetragonolobus</i>) | Injectable hydrogels, films/membranes, and freeze-dried scaffolds (often blended with other polymers) | Mucoadhesive polymer; exhibits gel-forming ability, high swellability and controlled-release characteristics | 61 and 62 |
| Chondroitin sulfate (CS-MA) | Cartilage ECM-derived sulfated polysaccharide | Photocrosslinked hydrogels (often blended with HA) for cartilage TE | Cartilage-mimetic biopolymer; provides hydration and growth-factor binding; tunable mechanics, swelling, and enzymatic degradability | 63 |
| Carboxymethyl arabinoxylan (CMARX) | Psyllium-derived hemicellulose | Hydrogels, polyelectrolyte nanoparticles (<i>e.g.</i> with CS), films, composite scaffolds | Biocompatible, biodegradable polysaccharide; CM modification increases crystallinity and thermal | 64 |



Table 1 (Contd.)

| Polymer | Origin/type | Formulation possibilities | Key functional properties | References |
|---|---|---|---|------------|
| Kappa-carrageenan (K-CG) | Red algae-derived sulfated polysaccharide | Ionic hydrogels (beads, gels, membranes), composite scaffolds, electrospun mats | stability; supports pH-responsive swelling and sustained drug release Forms strong ionically crosslinked gels (with K^+/Ca^{2+}); highly hydrophilic (swelling); biodegradable; excellent encapsulation and sustained release of growth factors/drugs | 65 |
| 2-Hydroxyethyl cellulose (2-HEC) | Cellulose ether | Cryogels, hydrogels, films, microparticles | Biocompatible and biodegradable; highly hydrophilic with large swelling; forms viscous, stable gels; demonstrated use in sustained-release cryogel systems | 66 |
| Konjac glucomannan (KGM) | From konjac tuber | Hydrogels (thermal or alkali-induced gelation), sponges/aerogels, films | Biocompatible and biodegradable; extremely high viscosity and water uptake; strong gelation (acetyl-dependent); forms robust matrices for sustained drug release and wound dressings | 67 |
| (C) Synthetic polyesters and copolymers | | | | |
| Poly(glycerol sebacate) (PGS) | Synthetic thermoset elastomer | Made into porous elastomeric scaffolds (foams), fibrous meshes, and composite grafts | Elastomer; rubber-like elasticity; tunable mechanical properties and degradation rate (matched to native soft tissues) | 68 and 69 |
| Poly(3-hydroxybutyrate) P(3HB) | Bacterial polyester | Scaffolds (<i>e.g.</i> bone, tissue engineering), cardiovascular patches, biodegradable microspheres/carriers | Intrinsically osteoinductive (promotes MSC osteogenic differentiation and bone regeneration) | 70 and 71 |
| Poly(ϵ -caprolactone) (PCL) | Aliphatic polyester | Processed into porous scaffolds (<i>e.g.</i> bone or vascular), electrospun fiber mats, and biodegradable microspheres/nanoparticles | Thermoplastic; good mechanical strength and toughness; easily fabricated (melt, 3D-printing, electrospinning) for sustained-release applications | 72 |
| Poly-L-lactic acid (PLLA) | Aliphatic polyester | 3D porous scaffolds (electrospun fibers, meshes, foams), microspheres/nanoparticles (drug carriers), composites (<i>e.g.</i> with bioceramics for bone TE) | High tensile strength; tunable mechanical stiffness and degradation rate (<i>via</i> crystallinity/porosity); FDA-approved for implants | 73 and 74 |
| Poly(lactic acid) (PLA) | Aliphatic polyester | 3D-printed scaffolds, electrospun mats, films, microparticles, sutures | Biodegradable (hydrolyzes to lactic acid); high tensile strength and stiffness (brittle); hydrophobic; tunable crystallinity; biocompatible but relatively inert (often blended or surface-modified for cell adhesion) | 75 |
| Poly(lactic-co-glycolic acid) (PLGA) | Copolymer | Microparticles, nanoparticles, 3D scaffolds, implants | Biocompatible and biodegradable with tunable degradation rate (by LA/GA ratio); relatively hydrophobic; good mechanical strength; extensively used for controlled release of drugs, proteins and growth factors | 76 |
| Poly(propylene fumarate) (PPF) | Unsaturated polyester | Injectable/crosslinked hydrogels, 3D-printed scaffolds, composites | Biodegradable (degrades to fumaric acid and propylene glycol); tunable mechanical properties and degradation rate; inherently crosslinkable (unsaturated sites) for high-strength bone/regenerative scaffolds | 77 |
| Poly(glycolic acid-co-propylene fumarate) (PGA-co-PF) | Copolymer | Electrospun nanofibrous scaffolds, composite fibers | Biodegradable copolymer; high water uptake and accelerated degradation when formulated with nanofillers; reinforced scaffolds exhibit enhanced mechanical strength and osteoconductivity (<i>e.g.</i> with GO/hydroxyapatite) | 78 |



Table 1 (Contd.)

| Polymer | Origin/type | Formulation possibilities | Key functional properties | References |
|--|--|--|---|------------|
| (D) Semi-synthetic methacrylated polymers | | | | |
| AESO (acrylated epoxidized soybean oil) | Acrylated vegetable oil | Photocurable 3D-printed resins and hydrogels (often composite with nano-hydroxyapatite for bone TE) | Thermoset resin; UV-crosslinkable (photopolymerizable) with tunable mechanics; supports cell growth/differentiation | 79 and 80 |
| Alginate-methacrylate (AlgMA) | Methacrylated alginate | Photocrosslinked hydrogels and 3D-printed scaffolds | Tunable stiffness and degradation (hydrolytic/enzymatic) | 80 |
| Gelatin-methacryloyl (GelMA) | Methacrylated gelatin | Photocurable hydrogels and bioinks for 3D bioprinting; microcarriers | Retains natural RGD and matrix metalloproteinases-cleavage sequences | 81 |
| Chondroitin sulfate-methacrylate (CS-MA) | Methacrylated cartilage ECM | Photocrosslinked hydrogels (often blended with HA) for cartilage TE | Cartilage-mimetic biopolymer; provides hydration and growth-factor binding; tunable mechanics, swelling, and enzymatic degradability | 63 |
| (E) Natural or bioactive polymers | | | | |
| Peptides | Natural or synthetic short proteins/polypeptides | Self-assembling hydrogels and nanofiber networks (3D scaffolds), injectable gels, composite coatings; can form nanoparticles for drug delivery | Often bioactive (e.g. signaling peptides); good mechanical stability and tissue-like elasticity in hydrogel form; tunable assembly (e.g. β -sheet fibers) | 82 |

electron pairs between atoms and result in strong, stable chemical linkages such as amide, ester, or radical-mediated grafts; and non-covalent interactions which are weaker and reversible forces that do not involve sharing of electrons such as hydrogen bonding, electrostatic forces, π - π stacking, and hydrophobic interactions. Different classes of biodegradable polymers tend to favor different types of bonding depending on their functional groups.

Hydrogen bonding is a specific type of non-covalent interaction that occurs when a hydrogen atom, covalently attached to an electronegative atom (such as oxygen or nitrogen) within a molecule, experiences an attractive force to another electronegative atom with lone electron pairs. In GO, the abundant oxygenated functional groups including hydroxyl (-OH), carboxyl (-COOH), and epoxy (-C-O-C) act as both hydrogen bond donors and acceptors, allowing physiochemical interactions with complementary sites on surrounding polymers.⁸³ Natural polysaccharides and their derivatives (such as alginate, cellulose, starch, hyaluronic acid, and carboxymethyl cellulose), as well as natural proteins (collagen, gelatin, silk fibroin), are especially suited for forming hydrogen bonds with GO. Their molecular structures are rich in hydroxyl, carboxyl, and amide groups, providing ample sites for hydrogen bonding interactions with the oxygenated functionalities on GO.⁸³ It enhances water dispersibility by stabilizing the nanocomposite network, boosts swelling by increasing the matrix's hydrophilicity, and enables better control of drug release by slowing burst release and supporting sustained release. A dense hydrogen-bonding network also strengthens mechanical properties and elasticity while regulating degradation, which is important for biomedical hydrogels and drug delivery systems.⁸⁴

Electrostatic interactions are fundamental to the interfacial binding between biodegradable polymers and graphene derivatives. Specifically, positively charged groups on cationic biodegradable polymers such as the amine groups found in chitosan and polycationic peptides are attracted to the negatively charged carboxyl groups on GO or other oxidized graphene surfaces. This electrostatic attraction enhances compatibility and mechanical integrity in composite materials, which is especially important in biomedical applications that require controlled interactions and stable matrices.⁸⁵ Similarly, anionic biodegradable polymers including alginate, hyaluronic acid, and CMC can interact with amine functionalized or protonated graphene derivatives through complementary charge pairing. The carboxyl and sulfate groups in these polysaccharides form strong ionic bonds with positively charged sites on modified graphene. These electrostatic interactions often lead to pH responsive behavior, as the ionization states of both polymer and graphene surfaces change with pH, enabling tunable adhesion and controlled drug release.^{86,87}

The basal plane of graphene and rGO provides an extended π -conjugated system, which facilitates π - π stacking and hydrophobic interactions with biodegradable polymers containing aromatic rings or hydrophobic backbones. Proteins such as gelatin and peptides enriched with aromatic residues such as phenylalanine, tyrosine, and tryptophan offer abundant sites for π - π interactions, while biodegradable polyesters like PLA, PLLA, PCL, and PLGA possess hydrophobic domains that participate in both π - π stacking and hydrophobic contacts with graphene surfaces. These interactions enhance compatibility and matrix integrity by stabilizing polymer chains along the graphene plane, promoting uniform dispersion and superior



Table 2 Fabrication techniques for graphene–polymer composites: a comparative overview

| Method | Process steps | Advantage/limitations | References |
|---|---|--|---------------|
| Solvent casting | Dissolve polymer and graphene (<i>e.g.</i> GO) in solvent; cast into mold; evaporate solvent to form film | Simple, cheap, and scalable yields uniform thin films; enhances mechanical, barrier, and antibacterial properties Limited 3D architecture control, potential nanoparticle aggregation, solvent residues | 92–94 |
| Solution casting with sonication | As above but apply ultrasonic waves during mixing to disperse fillers; then cast and dry | Improved nanoparticle dispersion compared to plain casting; more uniform composites Excessive sonication may degrade polymers, some aggregation and solvent residues may persist | 92, 95 and 96 |
| Emulsification and crosslinking | Create a (water-in-oil or double) emulsion of polymer solution containing graphene; solidify droplets by chemical crosslinking (<i>e.g.</i> GLA) | Enables formation of micro/nanoparticles with high surface area and uniform graphene dispersion for drug delivery; crosslinking provides stability and tunable release Use of surfactants/oil phases may affect biocompatibility; batch variability possible | 83 and 97 |
| Sonication-assisted (hybrid) synthesis | Use ultrasonication to drive <i>in situ</i> reactions or mixing (<i>e.g.</i> solvothermal or co-precipitation) of graphene and other components | Enables efficient exfoliation and uniform dispersion of graphene, scalable and simple, can be combined with hybrid synthesis for multifunctionality Excessive sonication can cause defects and reduce flake size; risk of agglomeration or residual surfactants if not optimized | 98 and 99 |
| <i>In situ</i> polymerization | Polymerize monomers in presence of graphene fillers (<i>e.g.</i> Michael addition, free-radical) to form network | Enables uniform graphene dispersion and strong interfacial bonding, leading to enhanced mechanical, thermal, and drug release properties; allows covalent grafting and functionalization Complex chemistry, need for initiators/catalysts, risk of aggregation at high graphene content, and potential cytotoxicity from residual chemicals | 99 and 100 |
| Free-radical polymerization (lyophilization) | Initiate polymerization (<i>e.g.</i> acrylic monomers) in solution; freeze-dry the gel to form porous scaffold | Creates porous 3D scaffolds after freeze-drying; controllable pore structure Requires initiators and freeze-drying, which may affect drug stability and loading | 96 and 101 |
| Magnetic functionalization (co-precipitation) | Synthesize magnetic nanoparticles (<i>e.g.</i> Fe ₃ O ₄) onto graphene (GO) by co-precipitation; incorporate into polymer (<i>e.g.</i> <i>via</i> emulsification or casting) | Enables magnetically targeted drug delivery and imaging; stable integration of Fe ₃ O ₄ enhances multifunctionality Potential agglomeration, requires precise chemical control | 102 and 103 |
| Electrospinning | Apply high-voltage to polymer/graphene solution to draw nanofibers onto a collector; then (optionally) crosslink fibers (<i>e.g.</i> heat or ionic) | Produces nanofibrous mats that mimic ECM, supporting cell growth and sustained drug release Limited to thin mats and challenges in loading cells or drugs uniformly in 3D structures | 104 and 105 |
| UV-assisted crosslinking | Mix photopolymerizable polymers with graphene and a photoinitiator; | Rapid solidification; spatial control of crosslinking; GO reinforcement enhances stiffness | 106 |



Table 2 (Contd.)

| Method | Process steps | Advantage/limitations | References |
|--------------------------------------|---|---|-------------|
| 3D printing (additive manufacturing) | expose to UV light to induce covalent crosslinking into a network | UV penetration depth limits thickness; photoinitiators may introduce toxicity | 107 and 108 |
| | Extrude thermoplastic melt or hydrogel ink layer-by-layer to build 3D scaffold; may include post-print coatings | Enables precise, patient-specific scaffold geometry. Surface coatings (e.g. polydopamine) can aid GO deposition High processing temperatures may limit cell/drug loading and can affect sensitive biomolecules | |

reinforcement.^{40,88} π - π stacking is particularly efficient at adsorbing aromatic drug molecules, increasing entrapment efficiency and allowing controlled release profiles due to strong molecular interactions at the interface. Furthermore, hydrophobic contacts between graphene and biodegradable polymer chains reduce water uptake and permeability, improving barrier properties and retarding degradation rates under aqueous conditions, an effect desirable for prolonged drug delivery and enhanced packaging performance.⁸⁸

Amide bond formation typically utilizes the carboxyl (-COOH) groups abundant on GO surfaces. These can be activated using carbodiimide chemistry, most commonly with EDC (*N*-ethyl-*N'*-(3-dimethylaminopropyl)carbodiimide) and NHS (*N*-hydroxysuccinimide), to form amide bonds with amine containing biopolymers such as chitosan, collagen, or gelatin. This reaction proceeds by forming an active ester intermediate on GO that readily reacts with amino groups on the polymer, resulting in a stable amide linkage that covalently anchors the polymer chain to the graphene surface.⁸⁹

Esterification reactions exploit the reaction between GO's carboxyl groups and hydroxyl (-OH) functional groups present in hydroxyl-rich biopolymers, such as polysaccharides and polyesters. This process, often catalyzed by acid or activation agents, leads to the formation of ester bonds, chemically grafting the polymer backbone onto the graphene architecture and yielding a network with improved mechanical and degradation resistance.⁸⁹

Radical grafting is widely used for methacrylated biopolymers like GelMA, AlgMA, or CS-MA. Under UV irradiation, these methacryloyl groups form free radicals that can react with activated double bonds on the GO surface or pre-functionalized graphene, leading to the formation of robust covalent cross-links. This method is especially effective in hydrogel synthesis, resulting in nanocomposite networks with high crosslinking density and mechanical stability suitable for scaffolds.^{90,91}

3 Fabrication of BGNs

The fabrication of BGNs involves a variety of techniques aimed at achieving optimal dispersion of graphene, mechanical integrity, and functional performance. The choice of methods depends heavily on the desired final form (e.g., film, hydrogel,

fiber, 3D scaffold) and application. Many studies employ hybrid approaches, combining multiple methods to leverage their respective advantages. For instance, sonication is often integrated with solvent casting or crosslinking to enhance nanoparticle dispersion. A comparative summary of the techniques discussed is provided in Table 2.

3.1 Solution-based and casting methods

3.1.1 Solvent casting. This widely used and straightforward method involves dissolving a polymer and dispersing graphene in a suitable solvent, casting the mixture onto a flat surface, and slowly evaporating the solvent to form a solid film (Fig. 4).¹⁰⁹⁻¹¹¹ During this process, solvent molecules act as mediators that control both polymer chain mobility and graphene dispersion within the solution. The viscosity of the casting solution and the polarity of the solvent influence the degree of mixing, with well-matched solvents promoting uniform dispersion and poor solvents increasing the risk of aggregation.¹¹² As the solvent evaporates, the polymer chains reorganize and gradually entrap graphene sheets, locking in the noncovalent interactions (hydrogen bonding, electrostatic forces, or π - π stacking) formed in solution. The evaporation rate is a critical factor: slow solvent removal facilitates homogeneous film formation, whereas rapid evaporation can lead to nanoparticle migration or surface aggregation. In some systems, residual oxygen-containing groups on graphene derivatives may undergo limited condensation reactions with polymer hydroxyl or amine

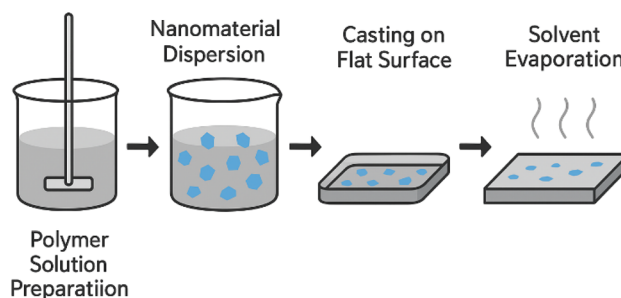


Fig. 4 Schematic representation of the solvent casting process for nanocomposite thin films.



groups during drying, forming ester or amide linkages that further stabilize the composite structure.¹¹³ Solvent casting remains the simplest fabrication route, but its lack of 3D control makes it unsuitable for regenerative scaffolds compared to electrospinning or 3D printing. For instance, SF/GO nanocomposite films were fabricated by blending the components in solution, where the presence of GO induced a conformational transition in the SF matrix from a random coil to a more stable β -sheet structure, thereby enhancing mechanical integrity.¹¹⁴ This technique has also been used to create 2-HEC/graphene films with improved thermal stability¹¹⁵ and SA/hydroxyapatite (HA)/graphene nanoplatelets (GnP) bionanocomposite films for bone tissue engineering.¹¹⁶ A hybrid approach combining solvent casting with porogen leaching, using a sacrificial material like salt to create pores, has been used to create porous PPF/GO nanocomposites with a hierarchical architecture suitable for cell infiltration.¹¹⁷

3.1.2 Casting with sonication. To overcome the natural tendency of nanoparticles to agglomerate, high-frequency ultrasonic waves are applied to the polymer-graphene solution before casting (Fig. 5). The process, known as sonication, generates acoustic cavitation, which involves the formation and violent collapse of microscopic bubbles. This creates intense localized shear forces that effectively break apart particle aggregates, ensuring a uniform and stable dispersion.^{118–120} This method has been successfully used to fabricate PCL/PGS/GO tubular scaffolds with a uniform porous morphology,¹²¹ P(3HB)/GnP scaffolds for neuronal studies,¹²² and KGM/GO films with significantly improved mechanical strength (tensile

strength of 183.3 MPa) and a well-organized, bioinspired brick-and-mortar structure.¹²³

3.1.3 Emulsification. This process creates a stable dispersion of two immiscible liquids, such as oil and water, stabilized by a surfactant (Fig. 6). The emulsification step typically involves vigorous stirring or ultrasonication to generate fine droplets of one liquid phase dispersed within the other. Surfactant molecules orient at the interface, reducing interfacial tension and preventing droplet coalescence.¹²⁴ In the context of polymer-graphene systems, the choice of surfactant and solvent pair strongly influences droplet size, stability, and the final morphology of the nanocomposite. Hydrophilic-lipophilic balance governs whether a water-in-oil or oil-in-water emulsion is formed, while the concentration of emulsifier controls droplet size distribution.¹²⁵ Once the emulsion is stabilized, crosslinking or solvent evaporation solidifies the polymer phase, entrapping graphene sheets within spherical domains. The interaction between surfactant functional groups and graphene surfaces can also contribute to dispersion stability, as electrostatic repulsion or steric hindrance suppresses aggregation during processing.¹²⁶ In nanocomposite fabrication, emulsification enables the uniform incorporation of graphene into a polymer matrix, often to form spherical micro- or nanoparticles.^{127,128} For example, a water-in-oil emulsification was used to fabricate starch/agarose/GO nanoparticles for 5-fluorouracil (5FU) delivery, achieving good colloidal stability and drug encapsulation.¹²⁹ A more complex double emulsion (water-in-oil-in-water) method was employed to create a pH-sensitive CS/CMC/GQD/ZnO nanocomposite for quercetin delivery.¹³⁰

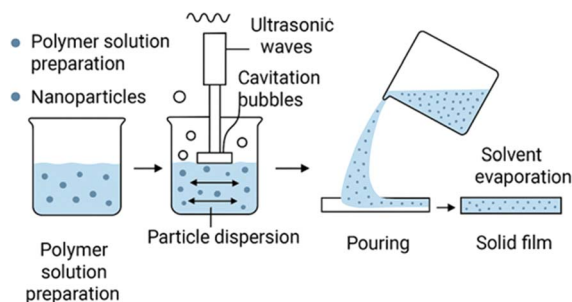


Fig. 5 Schematic illustration of the casting process enhanced by ultrasonic sonication.

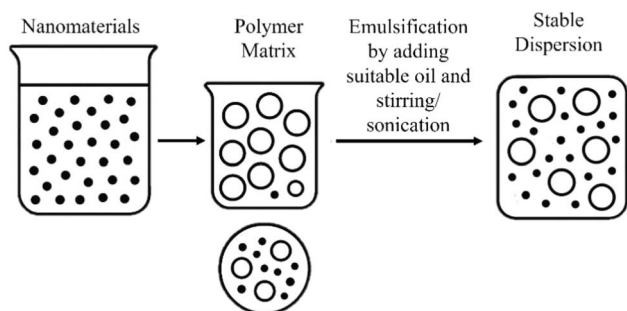


Fig. 6 Schematic illustration of emulsification.

3.2 Network formation and crosslinking

3.2.1 Crosslinking. This process forms covalent or non-covalent bonds between polymer chains or between polymers and nanofillers, creating a stable three-dimensional network. This network structure is fundamental to hydrogels and enhances the strength, thermal stability, and chemical resistance of the composite (Fig. 7). One common method is chemical crosslinking, which involves the formation of permanent covalent bonds using a chemical agent. For example, hydroxyl-functionalized polymers and polyurethane prepolymers can be used to form robust, crosslinked rGO composite films.⁹⁷ Similarly, a folic acid (FA)-CS-GO quantum dot (GOQD) nanocomposite was synthesized by conjugating FA with CS *via* carbodiimide chemistry using 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide and *N*-hydroxysuccinimide, followed by mixing with GO quantum dots.¹³¹ In such carbodiimide-mediated reactions, the $-\text{COOH}$ groups on GO or FA are activated to form *O*-acylisourea intermediates, which then react with $-\text{NH}_2$ groups of chitosan, creating stable amide linkages. This covalent grafting not only improves dispersion and interfacial bonding but also alters degradation behavior, since the hydrolytic cleavage of amide bonds is slower compared to ester linkages. In contrast, physical crosslinking relies on weaker, reversible non-covalent interactions like hydrogen bonding, ionic interactions, or hydrophobic



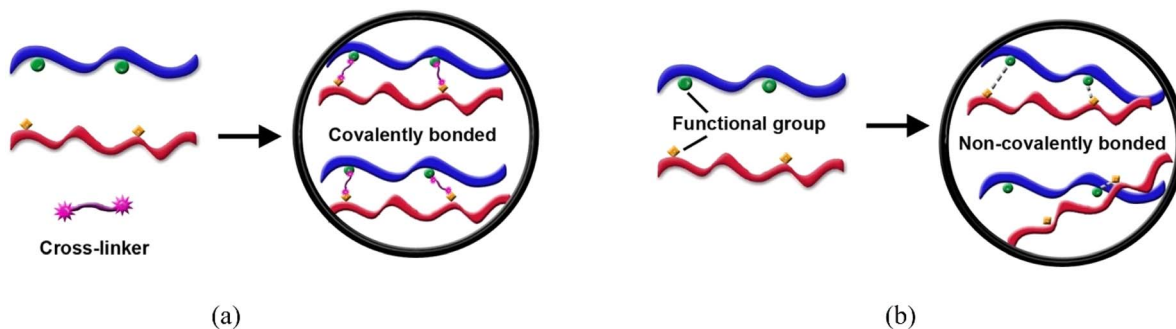


Fig. 7 Schematic illustration of polymer crosslinking mechanisms: (a) covalent bonding *via* cross-linkers and (b) non-covalent interactions *via* functional groups.¹³⁵ Ramachandran *et al.*, Cross-linking dots on metal oxides, *NPG Asia Mater.*, 2019, **11**, 19, under the terms of the Creative Commons Attribution 4.0 International Licensee (<https://creativecommons.org/licenses/by/4.0/>).

associations. GO/ γ -poly(glutamic acid) films achieve a strong, nacre-like structure through a combination of hydrogen and ionic bonds (with Ca^{2+} ions).¹³² Similarly, GO-COL scaffolds can be formed *via* pH-dependent electrostatic self-assembly between the negatively charged GO and positively charged COL.¹³³ These noncovalent systems are particularly relevant for drug delivery because protonation/deprotonation of functional groups (*e.g.*, $-\text{COOH}/-\text{COO}^-$ or $-\text{NH}_3^+/-\text{NH}_2$) under physiological pH changes can reversibly weaken or strengthen the

interactions, thereby triggering controlled drug release. Thermal crosslinking is another approach, achieved by heating the material to induce bond formation, often through carbon-carbon networks. In graphene-based biodegradable polymer composites, this method has been shown to significantly improve thermal conductivity and stability.¹³⁴

Another approach is UV-assisted crosslinking (photo-crosslinking), which uses ultraviolet light to activate a photo-initiator, which then generates free radicals that initiate

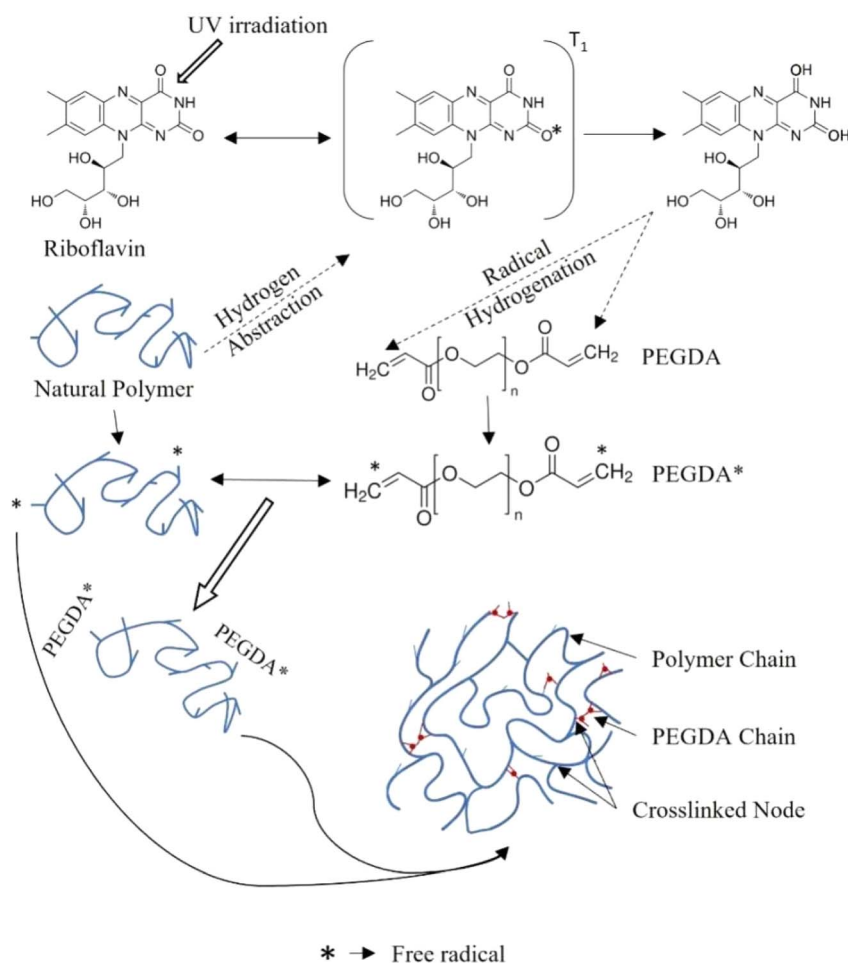


Fig. 8 Schematic representation of UV-assisted free radical crosslinking.



polymerization and crosslinking (Fig. 8). Upon exposure to UV light, photoinitiator molecules absorb photons and are promoted from their ground state to an excited triplet state. In this high-energy state, the photoinitiator can abstract a hydrogen atom from a neighboring polymer chain, generating a polymer radical and a hydrogenated photoinitiator radical. If an auxiliary crosslinking agent is present, the hydrogenated photoinitiator can also induce cleavage of carbon–carbon double bonds within the crosslinker, producing additional radicals. These transient radicals then recombine to form new covalent bonds between polymer chains and/or between polymer and crosslinker segments, resulting in a three-dimensional network.^{136,137} While photoinitiators and crosslinkers are commonly used, some modified polymers, such as methacrylated derivatives (*e.g.*, GelMA, AlgMA), can undergo self-crosslinking under UV light with only a photoinitiator, eliminating the need for an external crosslinking agent.

This technique offers rapid curing and spatial control and has been used to create AESO/PVDF/GO nanocomposites¹³⁸ and to fabricate 3D bioprinted hydrogels from bioinks containing methacrylated polymers (AlgMA, GelMA) and GO for cartilage engineering.¹³⁹ Finally, radiation-induced crosslinking using high-energy sources like γ -rays can generate free radicals on polymer chains and water molecules, initiating polymerization and network formation without the need for chemical initiators, as was done to fabricate sterile GO/(AAc-co-SA) interpenetrating network hydrogels.¹⁴⁰

3.2.2 *In situ* polymerization. This elegant method involves synthesizing the polymer matrix directly in a solution containing pre-dispersed nanoparticles (Fig. 9). The process begins by uniformly dispersing nanoparticles in a liquid monomer or low-molecular-weight precursor. Polymerization is then initiated through heat, radiation, or chemical initiators, allowing polymer chains to grow around and chemically bond with the nanoparticles. This facilitates the integration of nanoparticles into the polymer network by enabling interactions during chain propagation and network formation.¹¹² For instance, when GO or rGO is incorporated, its surface functional groups can form covalent or non-covalent bonds with the polymerizing matrix. In many systems, these groups can act as anchoring sites, leading to interfacial polymerization where the growing polymer chains

either initiate from or become physically entangled with the nanoparticle surface.¹⁴¹ This results in improved filler–matrix compatibility, reduced agglomeration, and enhanced load transfer efficiency.¹⁴² Furthermore, in biodegradable polymer systems such as PLA, PCL, or chitosan, *in situ* polymerization can promote uniform dispersion and interfacial adhesion due to hydrogen bonding and polar interactions formed during the growth of the polymer chains. This approach promotes homogeneous filler distribution and allows for the formation of strong interfacial bonds between the growing polymer chains and the nanoparticle surface.¹⁴³

It has been used to create hybrid rGO–multi-walled carbon nanotubes (MWCNT) nanocarriers functionalized with CoNi₂S₄ and ZnO, coated with CS and alginate, nanoparticles for co-delivery of drugs and genes,¹⁴⁵ and to fabricate thiol–maleimide hydrogels incorporating rGO for chemo-photothermal therapy *via* a Michael addition reaction, where hyaluronic acid was functionalized with thiol groups and CS was modified with maleimide groups.¹⁴⁶ A solvent-free approach using *in situ* polycondensation has also been developed for PGS/gelatin/GO nanocomposites, avoiding the use of potentially toxic solvents.¹⁴⁷

3.2.3 Free radical polymerization. This common chain-growth process involves initiation, propagation, and termination steps to form polymer chains from monomer precursors (Fig. 10). The process begins with the thermal or photochemical decomposition of initiators such as benzoyl peroxide, AIBN, or ammonium persulfate, generating free radicals that attack the π -bonds of vinyl monomers like acrylamide or methyl methacrylate. This initiates chain growth, where monomers are sequentially added during the propagation phase. Termination occurs through radical recombination or disproportionation, yielding stable polymer chains.¹⁴⁸ When applied in the synthesis of graphene-based biodegradable nanocomposites, this method enables the polymer matrix to form in direct contact with dispersed graphene derivatives. The presence of oxygen-containing functional groups on materials like GO can influence radical polymerization kinetics, affect the local polymerization environment, and improve filler compatibility by modulating surface energy and dispersion stability.¹⁴⁹ It is a versatile method for creating hydrogels and is often followed

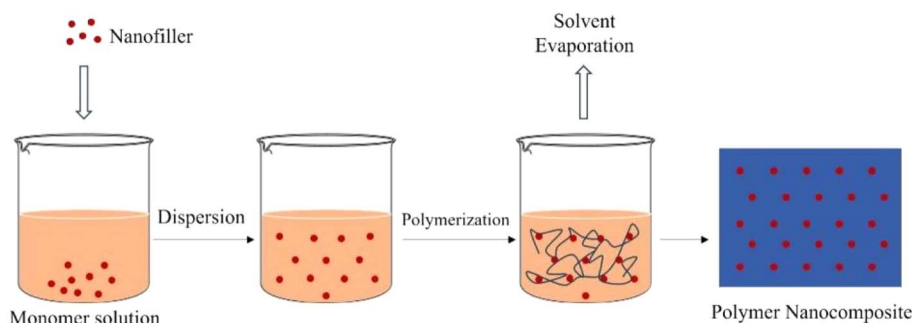


Fig. 9 Schematic representation of polymer nanocomposite fabrication *via in situ* polymerization.¹⁴⁴ Adopted from Basavegowda *et al.*, Advances in functional biopolymer-based nanocomposites for active food packaging applications, *Polymers*, 2021, 13(23), 4198, under the terms of the Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by/4.0/>).



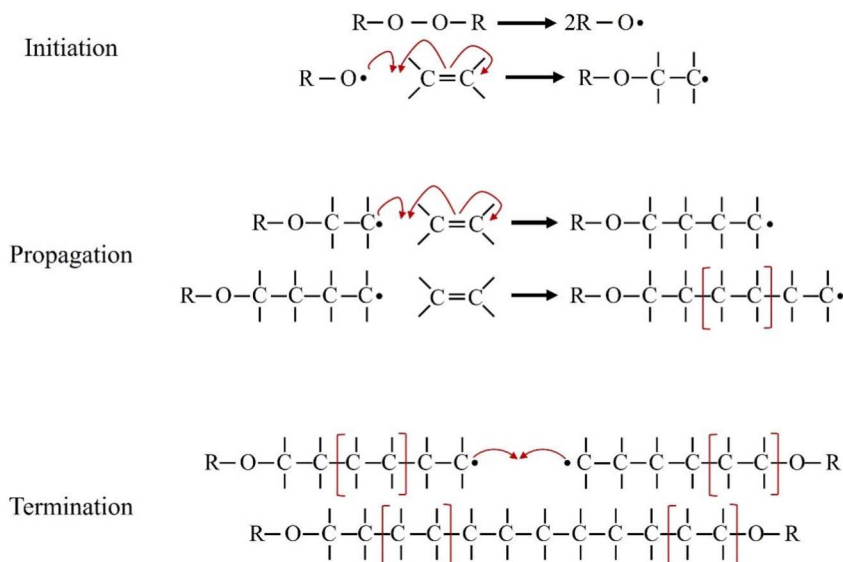


Fig. 10 Steps of free radical polymerization.¹⁵² Adopted from Ribas-Massonis et al., Free-radical photopolymerization for curing products for refinishing coatings market, *Polymers*, 2022, 14(14), 2856, under the terms of the Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by/4.0/>).

by lyophilization (freeze-drying) to produce highly porous scaffolds. This technique has been used to synthesize multifunctional porous scaffolds by grafting sodium alginate (SA) with acrylic acid (AAc) in the presence of nHA, SiO₂, and GO,¹⁵⁰ and to create hybrid scaffolds from GG, AAc, GO, and other nanoparticles for bone tissue engineering.¹⁵¹

3.3 Advanced and hybrid fabrication techniques

3.3.1 Electrospinning. This technique uses a high-voltage electric field to draw exceedingly fine nanocomposite fibers (typically nanometer to micrometer scale) from a polymer–graphene solution (Fig. 11). Under high voltage, a polymer–nanofiller solution is ejected as a fine jet from a syringe, stretching and solidifying into nanocomposite fibers with embedded nanofillers as the solvent evaporates. The resulting non-woven nanofibrous mats have a high surface-area-to-volume ratio and a structure that can effectively mimic the native extracellular matrix (ECM).^{153,154} However, the resulting thin mats limit applications to skin or vascular grafts rather than bulk bone regeneration. The success of electrospinning depends on a delicate balance of solution parameters such as viscosity, surface tension, and electrical conductivity,¹⁵⁵ all of which are strongly influenced by the presence of graphene derivatives. GO, for instance, increases the conductivity of the spinning solution due to its surface charges and polar groups, which can lead to finer fiber diameters and more uniform fiber morphology.¹⁵⁶ Additionally, uniform dispersion of GO within the polymer matrix is crucial to avoid defects during fiber formation; this often requires prior functionalization or surfactant stabilization.¹⁵⁷ The interactions between the polymer chains and the nanofillers during electrospinning can influence chain alignment and packing density, ultimately affecting mechanical properties and porosity of the final

scaffold. Moreover, residual solvent–filler interactions and rapid solvent evaporation can induce local phase separation or filler aggregation if chemical compatibility is poor.¹⁵⁵ Electrospinning has been used to fabricate SA/PVA/GnP composite wound dressings,¹⁵⁸ CS-based nanofibrous mats with GO and carbon quantum dot (CQD)-doped TiO₂ for accelerated wound healing,¹⁵⁹ and plasmonic rGO@AuNP–PCL composite scaffolds for synergistic cancer therapy and nerve regeneration.¹⁶⁰

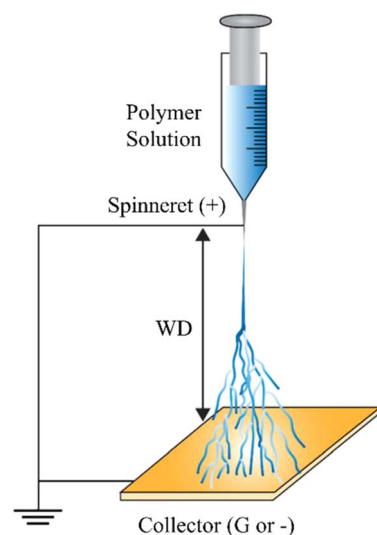


Fig. 11 Electrospinning setup for nanofiber fabrication.¹⁶¹ Adopted from Antonios Keirouz, Zhe Wang, Vundrala Sumedha Reddy, Zsombor Kristóf Nagy, Panna Vass, Matej Buzgo, Seeram Ramakrishna, and Norbert Radacsi, The History of Electrospinning: Past, Present, and Future Developments, *Adv. Mater. Technol.*, 2023, 8, 2201723, under the terms of the Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by/4.0/>).



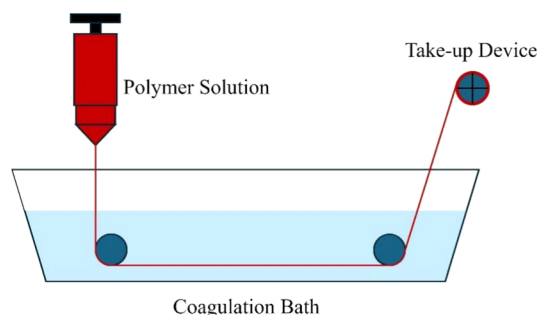


Fig. 12 Schematic representation of wet spinning procedure.

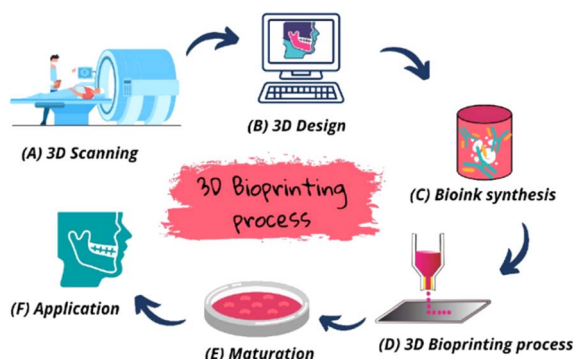


Fig. 13 Schematic workflow of the 3D bioprinting process for tissue engineering applications.¹⁷⁶ Reproduced from Lima *et al.*, 3D bioprinting technology and hydrogels used in the process, *J. Funct. Biomater.*, 2022, 13(4), 214, under the terms of the Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by/4.0/>).

3.3.2 Wet spinning. In this process, a viscous polymer-graphene solution is extruded through a spinneret directly into a liquid coagulation bath. The solvent exchange between the extruded jet and the bath causes the polymer and any embedded nanofillers to solidify into continuous filaments (Fig. 12).¹⁶² Wet spinning relies on phase inversion, where the solvent in the polymer solution diffuses into the coagulation

bath while nonsolvent molecules simultaneously diffuse into the jet. This induces polymer precipitation, driven by a decrease in solubility and thermodynamic instability of the polymer-solvent system. The presence of graphene-based nanofillers can significantly alter this process by influencing local viscosity, diffusion rates, and nucleation during solidification.¹⁶³ If well-dispersed, graphene derivatives may serve as nucleating agents that guide polymer chain alignment and promote the formation of more crystalline or oriented domains along the fiber axis.¹⁶⁴ However, poor dispersion or interfacial incompatibility can lead to phase separation or filler aggregation, resulting in heterogeneous fiber morphology.¹⁶³ Surface-functionalized graphene can mitigate these issues by improving interfacial affinity with the polymer matrix, enhancing mechanical integrity.¹⁶⁵ It has been used to fabricate flexible and electrically conductive alginate-graphene hydrogel biofibers with enhanced mechanical properties and thermal stability.¹⁶⁶

3.3.3 Freeze-drying (lyophilization). This dehydration process involves freezing a material (typically a hydrogel) and then reducing the surrounding pressure to allow the frozen water to sublimate directly from a solid to a gas phase.¹⁶⁷ This gentle removal of solvent preserves the material's delicate porous structure. It is widely used as a crucial post-processing step after hydrogel formation to create stable, highly porous scaffolds suitable for tissue engineering, as demonstrated in the fabrication of CS-GO nanocomposites.¹⁶⁸ GO-COL composite aerogels further exemplify this approach and were validated through *in vivo* studies.¹⁶⁹

3.3.4 3D printing and bioprinting. This transformative additive manufacturing technology enables the layer-by-layer fabrication of complex, patient-specific scaffolds with precise control over geometry and porosity. In 3D printing, a thermoplastic polymer-graphene composite is typically melted and extruded as a filament.^{170,171} This has been used to create PCL/GO scaffolds with mussel-inspired coatings¹⁷² and PLA/GO scaffolds with controlled porosity.^{173,174} 3D printing consistently outperforms traditional methods in shape fidelity and porosity control, though processing temperatures may denature bioactive molecules. 3D bioprinting advances this concept by

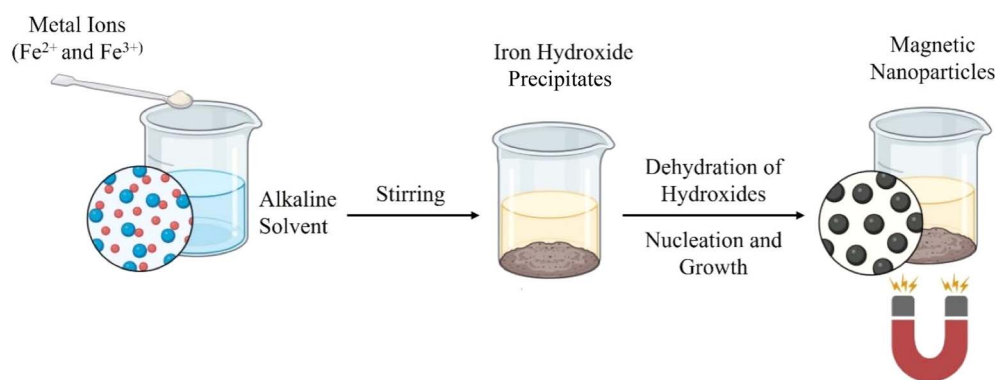


Fig. 14 Synthesis of magnetic nanoparticles.¹⁸² Adopted from Stiuftuc *et al.*, Magnetic nanoparticles: synthesis, characterization, and their use in biomedical field, *Appl. Sci.*, 2024, 14(4), 1623, under the terms of the Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by/4.0/>).



directly printing “bioinks” containing living cells, biomaterials, and growth factors to construct living tissue structures (Fig. 13).¹⁷⁵ In bioprinting, patient-specific images (CT/MRI) are converted into 3D models; cells are cultured and mixed with biomaterials to form bioink, which is then printed layer-by-layer and stabilized, followed by maturation to promote tissue formation.¹⁷⁶ This has been successfully used to create cell-

laden hydrogels for cartilage regeneration¹³⁹ and functional, spontaneously beating cardiac rings from rGO-containing bioinks.¹⁷⁷

3.3.5 Magnetic functionalization. This involves the integration of magnetic nanoparticles, typically iron oxide (Fe_3O_4), into the composite to impart magnetic responsiveness. This is commonly achieved *via* an *in situ* co-precipitation method,

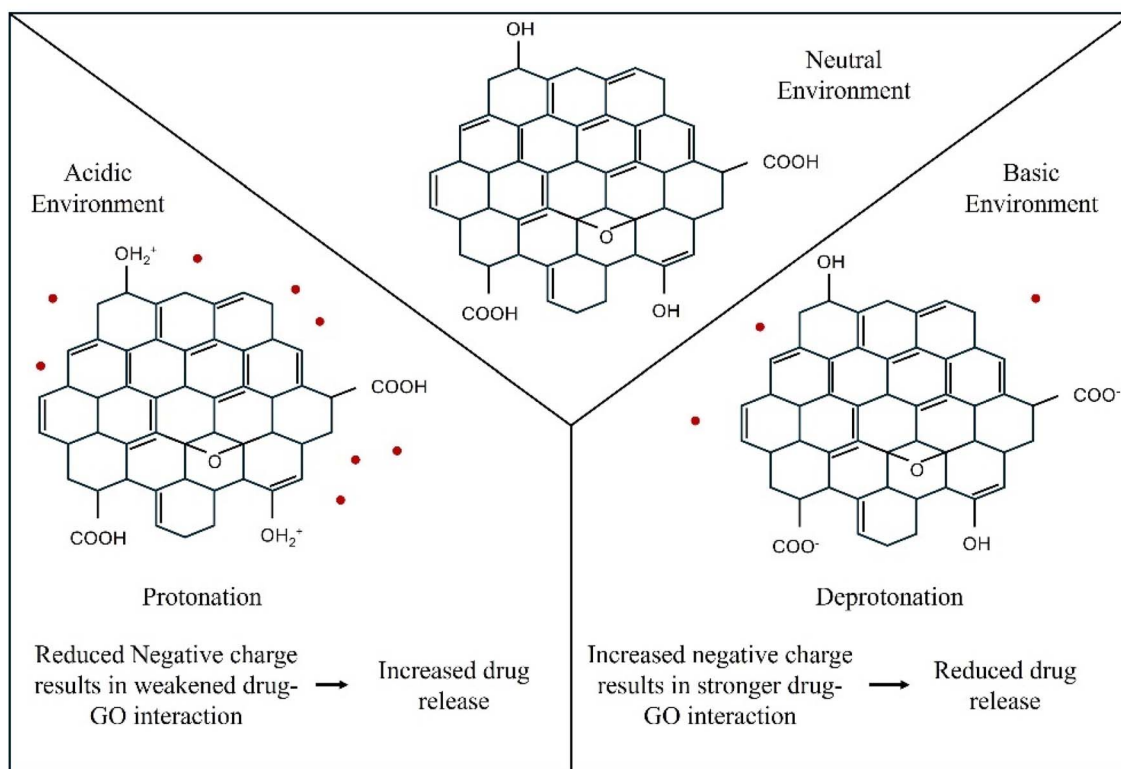
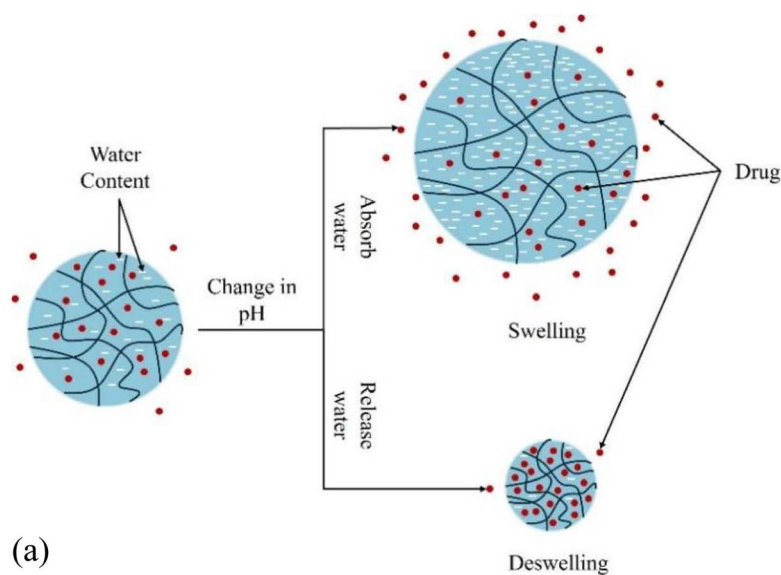


Fig. 15 pH-Responsive drug delivery system: (a) swelling and deswelling mechanism, (b) protonation and deprotonation mechanism.



where ferrous (Fe^{2+}) and ferric (Fe^{3+}) salts are mixed with GO in an aqueous medium under alkaline conditions. As the pH increases, usually above 10, iron ions react with hydroxide ions to form $\text{Fe}(\text{OH})_2$ and $\text{Fe}(\text{OH})_3$, which subsequently undergo nucleation and crystallization into magnetite (Fe_3O_4) nanoparticles. The oxygen-containing functional groups on GO act as nucleation and anchoring sites, enabling uniform deposition of nanoparticles onto the graphene surface (Fig. 14). The resulting magnetic BGNs can be guided by external magnetic fields for targeted drug delivery, manipulated for remote-controlled release, or used as contrast agents in magnetic resonance imaging.^{178,179} This method was used to create a multi-responsive SA-g-poly(2-hydroxypropyl methacrylamide) (PHPM)/mGO nanocomposite for etoposide delivery.¹⁸⁰ And a CS-based composite with magnetic GQDs for transdermal microneedle arrays.¹⁸¹

4 Applications in controlled drug delivery

BGNs are highly effective platforms for controlled drug delivery due to their high drug-loading capacity, enhanced stability, and responsiveness to a range of biological and external stimuli. The large surface area and π -electron system of graphene facilitate high drug loading *via* π - π stacking and hydrogen bonding, while the polymer matrix ensures biocompatibility and biodegradability.^{183,184}

4.1 Mechanisms of stimuli-responsive release

The “smart” behavior of BGNs stems from their ability to release therapeutic payloads in response to specific triggers. This enables on-demand, site-specific delivery, which is crucial for maximizing therapeutic efficacy and minimizing systemic toxicity.

One of the most widely exploited mechanisms is pH-sensitive release, which leverages the pH gradients that exist in the body and in pathological tissues. The mechanism typically relies on the ionization of functional groups within the polymer matrix or on the GO surface. For polymers with acidic (*e.g.*, $-\text{COOH}$) or basic (*e.g.*, $-\text{NH}_2$) groups, a change in pH alters their charge state, leading to changes in swelling behavior, as illustrated in Fig. 15a.¹⁸⁵ In an acidic environment, for instance, amine groups on CS become protonated ($-\text{NH}_3^+$), leading to electrostatic repulsion between polymer chains. This causes the hydrogel network to swell, increasing the mesh size and facilitating drug diffusion. At the same time, protonation weakens π - π stacking interactions and hydrogen bonding between chitosan chains and GO nanosheets, thereby loosening the carrier network. The protonation/deprotonation of GO's functional groups also plays a key role (Fig. 15b).¹⁸⁶

At low pH, carboxyl groups are protonated, reducing electrostatic repulsion and weakening the hydrogen bonding interactions that hold the drug, thereby promoting its release. Conversely, at neutral or basic pH, deprotonation enhances interactions (*e.g.*, hydrogen bonding or π - π stacking), stabilizing drug loading and slowing release.^{186,187} Additionally,

hydrolysis of ester linkages in polyesters such as PLA and PLGA is accelerated in acidic environments, producing lactic and glycolic acid that further decreases local pH and autocatalyzes matrix degradation.^{188,189} In some systems, pH-sensitive bonds such as Schiff bases and acetals are commonly used in drug delivery systems because they selectively cleave in acidic environments. Under low pH, Schiff bases hydrolyze due to protonation of the imine nitrogen, allowing water to attack and cleave the bond (*e.g.*, $\text{R}-\text{CH}=\text{N}-\text{R}' + \text{H}_2\text{O} \rightarrow \text{R}-\text{CHO} + \text{R}'-\text{NH}_2$), leading to controlled drug release in acidic environments.¹⁹⁰ For instance, imine ($\text{C}=\text{N}$) bonds in Schiff base linkages undergo protonation followed by nucleophilic attack by water, whereas acetals ($\text{C}-\text{O}-\text{C}$) are cleaved through acid-catalyzed oxonium ion intermediates, both providing predictable release profiles in tumor-like acidic conditions.^{191,192}

Another important mechanism is thermo-responsive release, which uses temperature as a trigger (Fig. 16). It is often designed using polymers like poly(*N*-isopropylacrylamide) (PNIPAAm) that exhibit a lower critical solution temperature (LCST). Below the LCST, the polymer is hydrophilic and swollen, retaining the drug. Above the LCST, the polymer undergoes a phase transition, this transition arises from disruption of hydrogen bonding between PNIPAAm's amide groups and water, leading to hydrophobic association within the network, which squeezes out the encapsulated drug.^{193,194} Graphene's excellent photothermal properties can be harnessed here; when incorporated into a thermosensitive polymer matrix, near-infrared (NIR) irradiation can be used to remotely heat the BGN above its LCST, triggering on-demand drug release.¹⁹⁵

Enzyme-responsive release offers high specificity by exploiting enzymes that are overexpressed in pathological tissues, such as matrix metalloproteinases (MMPs) in the tumor microenvironment. In the context of BGN, this responsiveness can be engineered by incorporating enzyme-cleavable peptide linkers either between the therapeutic agent and the nanocarrier or within the polymer backbone itself. Upon exposure to the target enzyme, specific peptide bonds ($-\text{CO}-\text{NH}-$) are hydrolyzed ($\text{R}-\text{CO}-\text{NH}-\text{R}' \rightarrow \text{R}-\text{COOH} + \text{H}_2\text{N}-\text{R}'$), resulting in cleavage of the linker or degradation of the polymer matrix. This cleavage destabilizes the nanocomposite structure, triggering the controlled release of the encapsulated or conjugated drug. In particular, hydrophilic oxygen-containing groups on GO and surface functionalities of the polymer can enhance enzyme

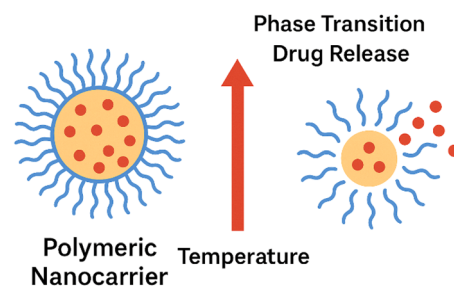


Fig. 16 Schematic illustration of thermo-responsive drug release: at temperatures below the LCST, the nanocarrier remains swollen and retains the drug; upon heating above the LCST, the polymer network collapses, triggering.



accessibility and interaction, facilitating more efficient degradation and payload release payload.^{196–198}

Photo-responsive release, illustrated in Fig. 17, can be achieved through several pathways beyond the photothermal effect. Photo-cleavable linkers can be incorporated into the BGN structure; upon irradiation with a specific wavelength of light, these bonds break, releasing the drug (Fig. 17b).¹⁹⁹ Alternatively, photoisomerization (Fig. 17a) uses molecules like azobenzene that change conformation under light, altering the carrier's structure and porosity to allow drug diffusion.²⁰⁰ Additionally, photosensitization involves a photosensitizer that, upon light exposure, produces reactive oxygen species (ROS) causing cell damage or drug release, forming the basis of photodynamic therapy (Fig. 17c).²⁰¹ Similarly, photoactivation uses light to convert inactive prodrugs into active therapeutic forms at the target site (Fig. 17d).

Finally, electro- and magnetically-triggered release offer remote control. Graphene's conductivity allows for electro-responsive systems where an applied electric field can induce conformational changes or electrophoretic movement to trigger release (Fig. 18).²⁰³ For magnetic release, incorporating magnetic nanoparticles allows an external oscillating magnetic field to induce localized hyperthermia or mechanical deformation of the scaffold, both of which can drive drug release.^{204,205}

A comparative summary of these release mechanisms is provided in Table 3.

4.2 Drug diffusion mechanisms

In addition to stimuli-responsive release, the passive diffusion of drugs from the BGN matrix is a fundamental process. The rate of release is governed by the concentration gradient and the physical properties of the matrix.²⁰⁶ In erodible matrices, the drug is released as the polymer matrix gradually degrades or dissolves from the surface inward, exposing new layers of drug

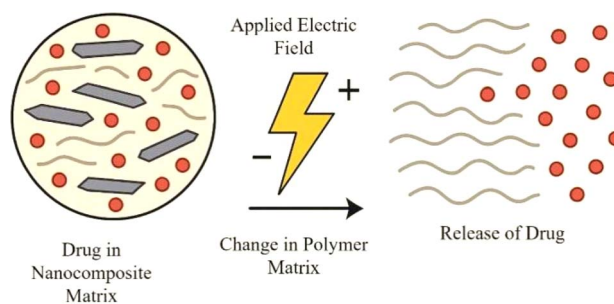


Fig. 18 Electro-responsive release mechanism.

over time (Fig. 19a). Enzymatically degradable polymers (*e.g.*, collagen, gelatin) coupled with GO generally degrade faster than synthetic polyesters, highlighting their suitability for soft tissue repair rather than long-term implants.²⁰⁷ In contrast, hydrophilic matrices swell upon contact with aqueous fluids, forming a gel layer (Fig. 19b). The drug must then diffuse through this viscous gel layer, and the release rate is controlled by the rate of swelling and the thickness of the gel.²⁰⁸ Polyester-based BGNs degrade primarily through hydrolysis, which is slowed by graphene reinforcement due to reduced water uptake and chain mobility.²⁰⁹ Interestingly, electrostatic polymer–graphene interactions accelerate degradation in acidic microenvironments by destabilizing the network, a feature exploited in tumor-targeted release systems.²¹⁰ In reservoir-type systems, the drug is contained within a core that is encapsulated by a rate-controlling polymer membrane. The drug diffuses through this membrane to the external environment (Fig. 19c).²¹¹ Comparisons show that covalently bonded systems maintain mechanical stability during degradation, while non-covalent systems show early-stage swelling and burst release.²¹²

The release kinetics in these systems are often described by mathematical models, such as Fickian diffusion, where release

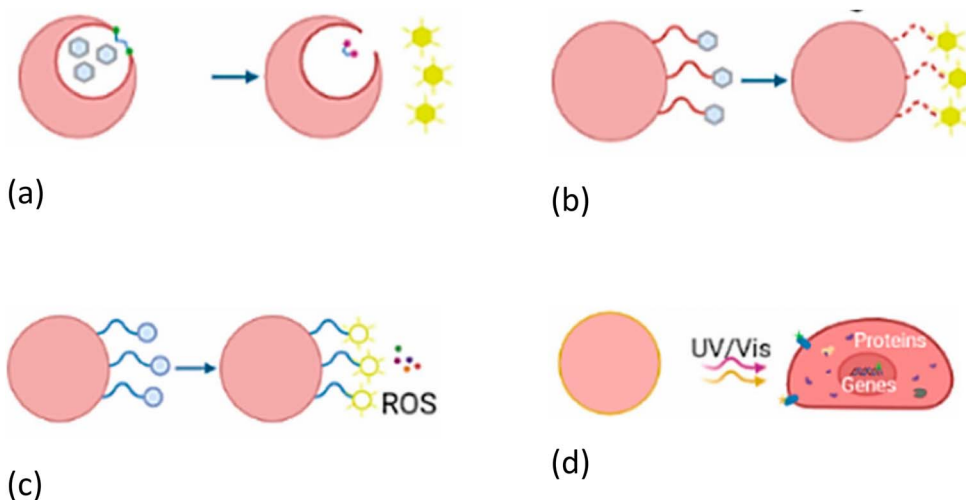


Fig. 17 Schematic representation of different photosensitive drug release mechanisms: (a) photoisomerization, (b) photocleavage, (c) photosensitization, and (d) photoactivation.²⁰² Adopted from Fernández *et al.*, *Advances in functionalized photosensitive polymeric nanocarriers*, *Polymers*, 2021, 13(15), 2464, under the terms of the Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by/4.0/>).



Table 3 Comparison of drug release mechanisms in biodegradable polymer–graphene nanocomposites

| Release mechanism | Advantages | Disadvantages | Usage frequency in BGN |
|------------------------|---|---|------------------------|
| pH-Sensitive | <ul style="list-style-type: none"> Targeted release in acidic microenvironment Minimal release at normal pH Polymer swelling at low pH enhances release | <ul style="list-style-type: none"> Off-target release in any acidic site (<i>e.g.</i> stomach) Requires precise tuning of polymer pK_a | Common |
| Thermo-responsive | <ul style="list-style-type: none"> On-demand release <i>via</i> external heating Synergistic with hyperthermia-based therapy | <ul style="list-style-type: none"> Possible drug degradation in strong acid Risk of tissue damage if overheated Requires effective heat delivery to target site (depth/penetration issues) | Moderately used |
| Enzyme-responsive | <ul style="list-style-type: none"> High specificity if target enzyme is unique | <ul style="list-style-type: none"> Complex design (requires enzyme-cleavable linkers) Variability in enzyme expression levels (patient-to-patient) | Less common |
| Photo-responsive | <ul style="list-style-type: none"> Precise spatiotemporal control | <ul style="list-style-type: none"> Phototoxicity/heating of healthy tissue is possible Limited penetration for visible/UV light | Less common |
| Electroresponsive | <ul style="list-style-type: none"> Deep NIR penetration enables efficient photothermal ablation Fine control over drug release by modulating electrical signal Can be easily integrated into implantable devices | <ul style="list-style-type: none"> Requires conductive materials | Less common |
| Magnetically triggered | <ul style="list-style-type: none"> Remote, non-invasive control Enables localized hyperthermia and mechanical actuation Magnetic targeting enhances accumulation at desired site | <ul style="list-style-type: none"> Risk of electrochemical reactions and tissue irritation Limited penetration depth in tissue Requires incorporation of magnetic nanoparticles Risk of local overheating or unintended tissue exposure | Moderately used |

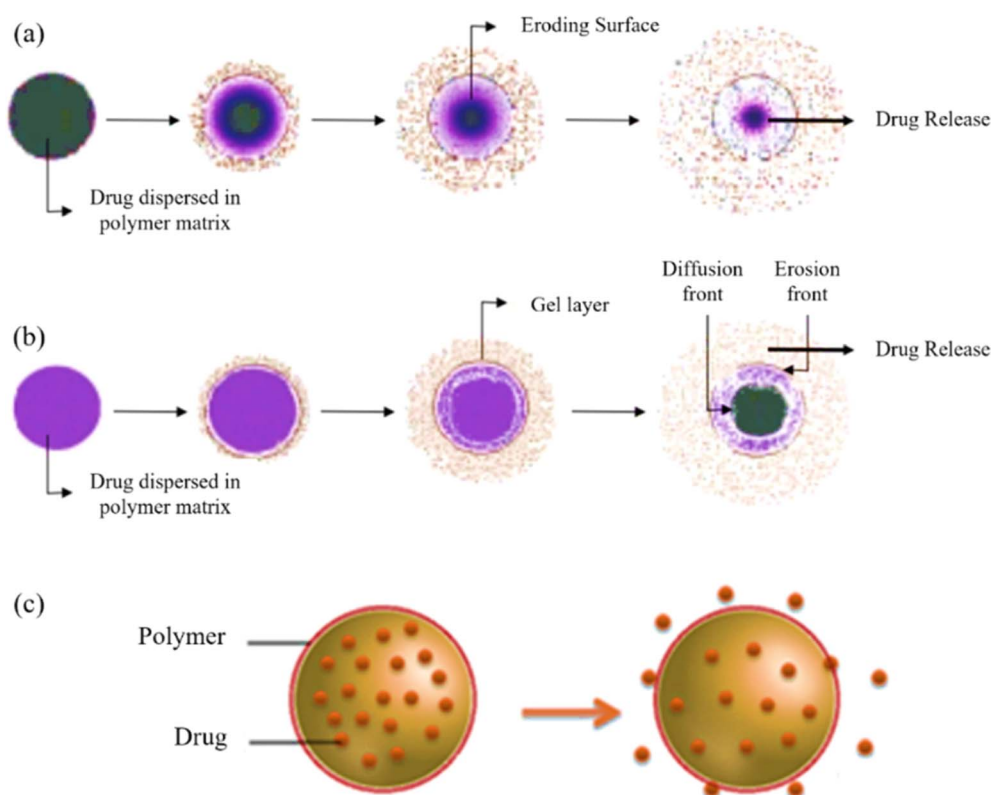


Fig. 19 Illustration of drug release from diffusion-controlled delivery systems, where the drug is uniformly distributed in: (a) an erodible polymer matrix, (b) a hydrophilic, swellable polymer matrix, and (c) a reservoir-type system.^{211,213} Adapted from Joseph *et al.*, Emerging Bio-Based Polymers from Lab to Market: Current Strategies, Market Dynamics and Research Trends, *C*, 2023, 9(1), 30, and Hossain *et al.*, Scope of bio-based nanoparticle targeted through the cancer zone to deactivate cancer affected cells, *Chem. Phys. Impact*, 2023, 6, 100180, both under the terms of the Creative Commons Attribution 4.0 International License (<https://creativecommons.org/licenses/by/4.0/>).



Table 4 Applications of BGN systems in controlled drug delivery

| Polymer matrix | Graphene type and additives | Application (target, drug, mechanism) | Observed outcomes (e.g. release%, viability, responsiveness) | References |
|----------------------------|---|---|---|------------|
| CS | rGO–5FU blended with CMARX (from <i>Plantago ovata</i>) | Melanoma therapy and wound healing; pH-responsive 5FU delivery | 93.1% release at pH 7.4; sustained release at pH 6.4; promotes skin cell proliferation; antibacterial vs. <i>S. aureus</i> , <i>P. aeruginosa</i> | 218 |
| FA-functionalized CS | GOQDs; DOX | Cancer therapy (A549, SH-SY5Y), folate-receptor targeting | 57% DOX release at pH 5.5 vs. 12% at pH 7.4; nuclear fragmentation; less than 5% hemolysis; selective cytotoxicity | 131 |
| CS/CMC | GQD beads in CMC, sodium salicylate | Oral delivery, inflammation therapy (pH-responsive) | Minimal release at pH 1.2; enhanced at pH 6.8 and 7.4; more than 70% HT29 cell viability | 221 |
| Alg/AAc | GO | Colon-specific drug delivery; pH-responsive cefadroxil release | Minimal release at pH 1 (stomach); sustained release at pH 7 (intestine); GO reduces burst release and regulates swelling | 140 |
| SA/K-CG | rGO | Amoxicillin delivery for wound treatment | 94% loading efficiency; 26% release at pH 5.5 vs. 34% at pH 7.4 over 95 h; Fickian release; strong antibacterial activity | 223 |
| SF | GO (0.5–3 wt%) | Controlled drug delivery (rhodamine B) and tissue engineering scaffolds | β -Sheet content peaked at 1.0% GO then declined; reduced burst release and sustained RhB release; faster degradation with higher β -sheet | 114 |
| 2-HEC | HCl/HNO ₃ -modified graphene | Nanocomposite films for potential biomedical applications (drug delivery, biosensors) | Enhanced thermal stability (11 °C increase in T_{max}); hydrophilic transformation of graphene | 115 |
| CS | GO–CS–FA decorated with camptothecin and diindolylmethane | Breast cancer (MCF-7); targeted co-delivery system | 95.67% inhibition (MCF-7); enhanced bioavailability (AUC \sim 33 858 ng mL ⁻¹ h ⁻¹); low renal/liver toxicity | 215 |
| Gelatin-coated GO | FA-functionalized rGO | Cervical cancer (Siha), chlorambucil delivery | 82% release at pH 1.2, 62.1% at pH 5.4, 43.7% at pH 7.4; IC ₅₀ : 125.9 μ g mL ⁻¹ (vs. 86 μ g mL ⁻¹ for free drug) | 217 |
| Starch/agarose | GO | Breast cancer (MCF-7), 5FU delivery | High 5FU encapsulation (\sim 87.3%); significant MCF-7 growth inhibition; acidic pH triggers release (targeted tumor delivery) | 129 |
| SA grafted with PHPM | mGO, Fe ₃ O ₄ -decorated GO | Lung cancer (H1299), etoposide delivery | Triple stimuli-responsive release (pH 5.5, NIR, magnetic); cell viability drops to \sim 19.1% (H1299); acid/NIR/magnet enhance release | 180 |
| CS/CMC | GQDs; ZnO nanoparticles | Brain cancer (U-87 MG), quercetin delivery | 49% inhibition of U-87 MG (cancer cells); 85% viability in L929 (normal); pH-sensitive release ($t_{1/2} \approx$ 72 h) | 130 |
| CS (folate-functionalized) | rGO; NiO nanoparticles; FA targeting ligand | Lung (A549) and breast (MCF-7), DOX delivery | DOX release \sim 98.6% at pH 5 vs. 9.6% at pH 7.4; A549 viability 12.3%, MCF-7 7.1%; low zebrafish toxicity | 224 |
| CS | rGO, Pd nanoparticles | Colon cancer (HT-29), dual-drug system (5FU + another) | \sim 95–98% encapsulation; pH-sensitive release; IC ₅₀ : 9.87 μ g mL; Fickian diffusion kinetics | 225 |
| CS (folic acid-modified) | GO nanoscrolls; FA-functionalization; DOX + caffeic acid | Lung carcinoma (A549), co-delivery of DOX and caffeic acid (CA) | DOX release \sim 83% at pH 5 vs. 71% at pH 7.4; selective apoptosis in A549, high viability in HEK293 (normal) | 226 |
| CS | GO; TiO ₂ nanoparticles; escin | Colon cancer (COLO 205) | IC ₅₀ \approx 22.7 μ g mL ⁻¹ (COLO 205); induces ROS-mediated apoptosis; minimal toxicity to normal cells | 216 |
| CS | GQDs | Ocular drug delivery; latanoprost for glaucoma; real-time tracking <i>via</i> photoluminescence | “On–Off–On” photoluminescence response with lysozyme; more than 80% cell viability; protective effect against H ₂ O ₂ -induced oxidative damage in human corneal epithelial cells | 222 |



Table 4 (Contd.)

| Polymer matrix | Graphene type and additives | Application (target, drug, mechanism) | Observed outcomes (e.g. release%, viability, responsiveness) | References |
|---|--|--|---|------------|
| CS/alginate | rGO; MWCNTs; CoNi ₂ S ₄ and ZnO nanoparticles | Co-delivery (HeLa/HEK-293), DOX and pCRISPR | pH-Responsive sustained DOX release (more in acidic environment); enhanced cellular uptake; improved therapeutic efficacy (co-delivery) | 145 |
| Thiol-maleimide hydrogel | Dopamine-rGO; DOX | Breast cancer (MCF-7), chemo-photothermal therapy | MCF-7 viability ~21% under NIR; pH-sensitive DOX release accelerated by NIR ($\Delta T \approx 22$ °C); targeted chemo/photothermal effect | 146 |
| Self-assembling peptide hydrogel | GO; (with antibiotics: isoniazid, amphotericin B, ciprofloxacin) | Sustained delivery of TB/antifungal/antibacterial drugs | Sustained, controlled release of each loaded drug (isoniazid, amphotericin B, ciprofloxacin); good biocompatibility | 219 |
| Double-network hydrogel (PAAm/agarose or PNIPAAm/agarose) | GO, CNT (photothermal agents); Fmoc-protected amino acids | Neurological (spasticity), baclofen release (NIR-triggered) | NIR-triggered baclofen release; inclusion of GO/CNT enables photothermal actuation (CNT-containing gel shows highest release efficiency) | 220 |
| CS | MrGO (Fe ₃ O ₄ @RGO) + Pluronic F127; α -mangosteen | Breast cancer (MCF-7), α -mangosteen delivery | Faster release at pH 5.5 (tumor-like) vs. 7.4; magnetic field enables targeting; inhibited MCF-7 proliferation | 227 |
| CS | mGQDs | Transdermal microneedle arrays; electrically triggered drug delivery | 96.4% drug release under electrical stimulation vs. 25.7% passive diffusion; detachable design with PEG base for rapid fluid response | 181 |
| PCL | GO, Fe ₃ O ₄ , TRAIL, DOX | Magnetically-triggered cancer therapy (scaffold system) | Magnetic stimulation enhances drug release; dual action therapy; localized delivery; <i>in vivo</i> validation not provided | 228 |
| Aminated CMC | GO; Fe ₃ O ₄ magnetic nanoparticles; curcumin | Breast cancer (MDA-MB-231), curcumin delivery | Enhanced cytotoxicity vs. free curcumin; rapid curcumin release at pH 5.5 vs. slower at 7.4; magnetic targeting improves localization | 186 |
| CMC | N-doped graphene, imatinib mesylate | pH-Responsive cancer therapy (simulated tumor microenvironment) | ~74% loading at pH 7.0 in 3 h; ~58% release at pH 4.0; reduced release at neutral/basic pH | 229 |

is proportional to the square root of time, or more complex non-Fickian (anomalous) models when polymer relaxation and drug diffusion rates are comparable.²¹⁴ This evidence suggests that degradation can be fine-tuned by adjusting graphene loading and interfacial chemistry rather than polymer choice alone.

4.3 Applications in cancer therapy

BGNs have shown significant promise in cancer therapy by enabling targeted delivery and controlled release, thereby enhancing efficacy and minimizing side effects. In the context of breast cancer, multiple studies have targeted MCF-7 and MDA-MB-231 cells. A GO-CS-FA nanobiocomposite co-loaded with camptothecin and diindolylmethane achieved 95.7% inhibition of MCF-7 cells and showed enhanced *in vivo* bioavailability.²¹⁵ A starch/agarose/GO system for 5FU delivery demonstrated accelerated release at pH 5.4.¹²⁹ Combining chemo- and photothermal therapy, a dopamine-rGO hydrogel loaded with doxorubicin (DOX) reduced MCF-7 viability to 21% under NIR irradiation.¹⁴⁶ For lung cancer, GO-based systems have been developed to target A549 and H1299 cells. An folic acid-CS-GOQD nanocomposite showed pH-responsive DOX release and folate receptor-mediated uptake, inducing

apoptosis selectively in cancer cells.¹³¹ A multi-responsive SA-G-PPHM/mGO system for etoposide delivery was triggered by pH, NIR light, and magnetic fields, enabling precise, multi-modal therapy that led to a sharp reduction in H1299 cell viability.¹⁸⁰ BGNs have also been applied to other cancers. For colon cancer, a CS/GO/TiO₂/escin nanocomposite demonstrated significant cytotoxicity against COLO 205 cells through TiO₂-induced reactive oxygen species (ROS) generation.²¹⁶ For cervical cancer, an FA-functionalized gelatin-coated GO nanocarrier delivered chlorambucil with pH-responsive release kinetics.²¹⁷ For melanoma, an rGO-5FU-CMARX hydrogel showed pH-dependent 5FU release and strong antibacterial activity.²¹⁸

4.4 Treatment of infectious and other diseases

Beyond cancer, BGNs are being developed for a range of therapeutic areas. For infectious diseases, nanoengineered self-assembling peptide-based hydrogels incorporating GO have been used for the sustained release of drugs against tuberculosis (isoniazid), fungal infections (amphotericin B), and bacterial infections (ciprofloxacin).²¹⁹ In the treatment of neurological disorders, double-network hydrogels composed of amino-acid-based networks reinforced with polyacrylamide



(PAAm), PNIPAAm, agarose, or low-gelling agarose, containing GO or carbon nanotubes (CNTs) as photothermal agents, have been used for the on-demand, NIR light-triggered release of baclofen to treat severe spasticity.²²⁰ For gastrointestinal conditions, a CS-GQD/sodium salicylate@CMC hydrogel bead system was designed for oral delivery, protecting the drug from the acidic stomach and enabling controlled release in the intestines.²²¹ Finally, in ocular drug delivery, a CS-GQD nanocomposite demonstrated enzyme-triggered release of latanoprost for glaucoma treatment, responding to lysozyme in tear fluid.²²²

A summary of selected studies on BGNs for controlled drug delivery is presented in Table 4.

5 Applications in tissue engineering

In tissue engineering, scaffolds provide a three-dimensional framework that mimics the native ECM, offering physical support and biological cues to guide cell adhesion, proliferation, and differentiation for the regeneration of new tissue.^{230,231} BGNs are excellent scaffold materials because the biopolymer component provides essential biocompatibility and biodegradability, while the graphene component significantly enhances mechanical strength, stability, and bioactivity.^{232–235}

5.1 Scaffold design principles and tissue engineering strategies

The success of a tissue engineering strategy depends on the careful design of scaffolds that can orchestrate a complex series of biological events. This can be achieved through two primary approaches: *in vitro* and *in vivo* tissue engineering.

The classic approach is *in vitro* tissue engineering, which involves creating tissues outside the body in a controlled laboratory setting (Fig. 20). The process begins with isolating cells from a patient or donor and seeding them onto a pre-fabricated

3D scaffold. This cell-seeded construct is then cultured in a bioreactor, which provides a dynamic environment with nutrients, oxygen, and mechanical or electrical stimuli to promote cell proliferation, differentiation, and ECM production. Once the engineered tissue reaches a desired level of maturity, it is implanted into the patient.^{236–238} However, implantation into the human body is not always the ultimate or mandatory step, as *in vitro* tissue constructs are also crucial for evaluating scaffold performance, studying cell-material interactions, modeling diseases, and testing drugs in a physiologically relevant environment.

In contrast, *in vivo* tissue engineering leverages the body's own regenerative capacity by using it as a natural bioreactor (Fig. 21). A biomaterial scaffold, often loaded with growth factors or other signaling molecules, is implanted directly into the site of injury. The scaffold is designed to recruit endogenous stem or progenitor cells from surrounding tissues. These recruited cells then populate the scaffold, differentiate into the appropriate cell types, and regenerate the damaged tissue *in situ*. As the new tissue forms, the scaffold gradually degrades and is replaced.^{239,240} However, studies in this area remain comparatively limited, as this approach requires significant prior research and validation to ensure safety and efficacy.

Most of the studies discussed in the following sections are thereby focused only up to various stages of the *in vitro* process. Some involve scaffold fabrication alone, others extend to cell seeding and bioreactor-based culture, while only a few report implantation into a living body. Understandably, *in vivo* studies are far less common than *in vitro* ones due to the extensive safety and regulatory requirements involved. Regardless of the approach, the scaffold's properties are paramount. It must be biocompatible, biodegradable at an appropriate rate, and possess sufficient mechanical integrity. Furthermore, its architecture must feature an interconnected porous network to allow for cell infiltration and nutrient transport.^{230,231,241} The

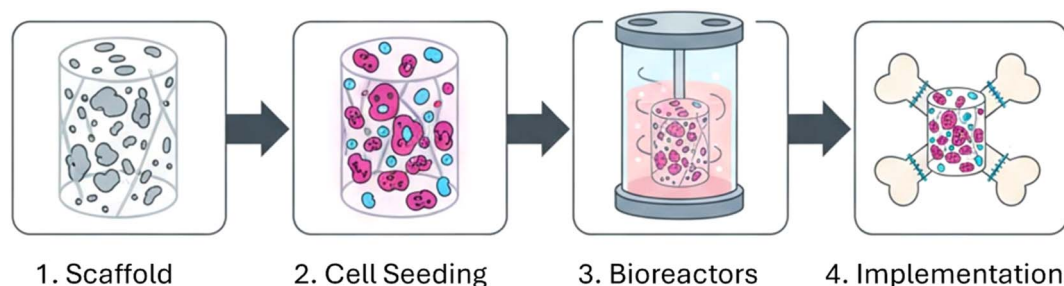


Fig. 20 Steps of *in vitro* tissue engineering.

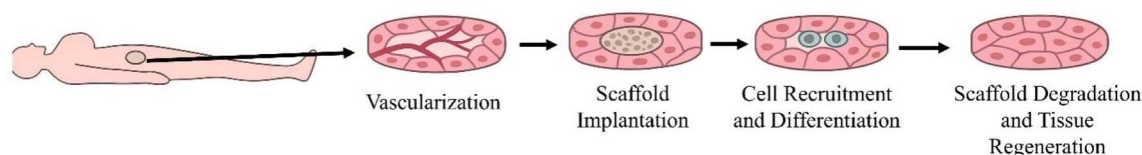


Fig. 21 Steps of *in vivo* tissue engineering.



Table 5 Applications of BGN systems in tissue engineering

| Polymer matrix | Graphene type and additives | Applications (target tissue and role) | Observed outcomes | References |
|---|--|---|--|------------|
| PGA-co-PF | GO, HA | Bone – electrospun scaffold | Increased alkaline phosphatase activity; osteoblastic differentiation; more than 5 times protein adsorption; greater than 98% metabolic activity | 78 |
| COL | GO | Neural – scaffold for nerve tissue engineering | Schwann cell adhesion/spreading; 3D porous (20–100 μm); mechanically stable; cell infiltration | 133 |
| AESO/PVDF blend (UV-crosslinked) | GO; curcumin | Bone – bone tissue scaffold | Semi-crystalline matrix with enhanced stiffness; antibacterial against common pathogens; supports osteoblast viability | 138 |
| Alginate/gelatin/chondroitin sulfate bioink | GO | Cartilage – 3D bioprinted scaffold | Promoted intrinsic chondrogenic differentiation of stem cells; high cell viability; improved ECM synthesis | 139 |
| GelMA/AlgMA bioink | rGO | Cardiac – 3D bioprinted cardiac tissue scaffold | Supported cardiomyocyte viability, alignment, and beating; enhanced electrical conductivity for synchronized activity | 177 |
| Type I COL | GO (0.05–0.2% w/v) | Bone – aerogel scaffold for cranial defects | Enhanced compressive modulus (0.20–0.51 MPa); superior biomineralization (Ca/P = 1.67); 1.5 \times increased bone volume <i>in vivo</i> ; improved rat bone marrow mesenchymal stem cell proliferation | 169 |
| SA | GnP; HA | Bone – scaffold for bone regeneration | Nearly doubled tensile strength at 0.5% GP; promoted apatite-like mineralization; excellent biocompatibility and biodegradability | 116 |
| CS (anisotropic membrane) | GO; HA | Bone – bone-mimicking membrane scaffold | Improved early osteoblast viability and long-term growth; enhanced cell spreading and adhesion; favorable microenvironment <i>via</i> GO/HA | 246 |
| PPF | GO nanoribbons/nanoplatelets | Bone – porous scaffold for bone regeneration | 26% modulus increase; cell infiltration; ECM formation; enhanced osteoblast viability | 117 |
| PCL | rGO-Ag nanoparticles | Bone – film scaffold | Increased mechanical/electrical properties; promotes stem cell differentiation; antimicrobial | 247 |
| CS beads | GO, TiO ₂ nanoparticles | Bone – injectable beads | Less than 1% resorption over 90 days; osteoconductivity; COL type I formation; enhanced crystallinity | 248 |
| PCL/PGS tubular scaffold | GO | Nervous/vascular/renal – nerve conduit | About 84% fibroblast viability; enhanced compressive modulus and thermal stability; channel-like porosity aiding regeneration | 121 |
| P(3HB) | GnP | Neural – conductive neuronal scaffold | Restored physiological neuronal firing patterns; increased responsiveness at low stimulus; dense neuronal network formation | 122 |
| CS (GLA-crosslinked beads) | GO; TiO ₂ nanoparticles; blackberry extract | Bone – injectable bead scaffold | Strong <i>in vivo</i> biocompatibility; stimulated new bone formation and mineral deposition; COL fiber development | 249 |
| KGM | GO | Scaffold through solvent casting | Young's modulus: 16.8 GPa; tensile strength: 183.3 MPa; over 90% cell viability; strong cell adherence; bioactive and biocompatible | 123 |
| PLGA | GO, MoS ₂ nanoplatelets | Bone – porous scaffold | 20–27% early bone regeneration; minimal inflammation; enhanced mechanical strength | 250 |
| CS | GO | Skin – scaffold | Around 78 μm porosity; improved vascularization and healing; mild inflammation | 168 |
| SA/PVA | GnP; curcumin | Skin – electrospun wound dressing | Controlled curcumin release (\sim 80% in 24 h); combined antimicrobial and antioxidant effects; supports tissue regeneration | 158 |
| PVA + CS nanofibers | GO; CQD-doped TiO ₂ | Skin – nanofiber wound healing scaffold | Accelerated wound closure (greater than 93% in 14 days); promoted fibroblast migration; antibacterial against gram-positive and negative bacteria | 159 |



Table 5 (Contd.)

| Polymer matrix | Graphene type and additives | Applications (target tissue and role) | Observed outcomes | References |
|---------------------------|--|--|--|-------------|
| CS/PVP nanofibers | GO | Wound healing – nanofibrous mat | <i>In vitro</i> study: more cell viability (40%); faster wound closure (33%); mimics ECM; water-permeable | 244 |
| Zein nanofibers | GO, curcumin | Wound healing – dressing | <i>In vitro</i> study: biphasic drug release; fibroblast proliferation; low swelling, controlled CUR release | 251 |
| PLGA/gelatin | GO | Bone – electrospun scaffold | Increased ALP, RUNX2, calcium deposition; ECM-like morphology; supports osteogenic differentiation | 252 |
| PCL | rGO, AuNPs | Neural – aligned nanofiber scaffold | Improved neurite outgrowth (2.5×); 90% Schwann cell viability; NIR-induced tumor cell ablation | 160 |
| Gelatin | GO | Bone – electrospun scaffold | Improved Young's modulus (70%) and tensile strength (200%); supports osteoblast proliferation | 253 |
| Alginate | GO | Muscle – biofiber scaffold | <i>In vitro</i> study: increased tensile strength/modulus; C2C12 myoblast viability and differentiation; myogenic morphology | 166 |
| PLLA | GO–HA hybrid composite | Bone – load-bearing scaffold | Increased compressive strength (~21.5 MPa) and modulus (~5 GPa); apatite layer formation in simulated body fluid; excellent osteoblast compatibility | 243 |
| CS–PPPOEMA | GO–Ag | Wound healing – scaffold | Improved wound closure (89.81%); antimicrobial activity; selective cytotoxicity | 254 |
| PGS/gelatin | GO; Clay | Scaffold through <i>in situ</i> polymerization | Enhanced thermal and mechanical properties; controlled degradation; homogeneous filler dispersion | 147 |
| SA grafted AAC | GO; nHA@SiO ₂ | Bone – bioactive porous scaffold | <i>In vitro</i> study: enhanced osteoblast proliferation and adhesion; increased mineralization; improved hydrophilicity and porosity | 150 |
| ARX and BG | GO; nHA | Bone – porous hydrogel scaffold | High compressive strength and modulus; optimal porosity (~55%) for cell migration; promoted osteogenic differentiation | 255 |
| Poly(acrylic acid) hybrid | GO; nHA; TiO ₂ ; Ag-sulfadiazine | Bone – antibacterial fracture scaffold | Sustained silver drug release; increased mechanical strength; enhanced osteoblast adhesion and proliferation | 151 |
| PPF | GO nanoplatelets, MoS ₂ nanoplatelets | Bone – porous scaffold | Enhanced compressive modulus (up to 108%); high cytocompatibility; ECM deposition | 256 |
| PPF | CNTs, GO nanoribbons/nanoplatelets | Bone – tissue scaffold | High cell viability; mild degradation cytotoxicity; enhanced spreading and attachment | 257 |
| PCL (3D-printed) | GO (mussel-inspired coating) | Bone – surface-modified scaffold | Improved osteoblast proliferation and differentiation; increased alkaline phosphatase activity and calcium deposition | 172 |
| PCL | Graphene/GO | Bone – 3D-printed scaffold | Enhanced modulus (136.74 MPa); cell proliferation; trabecular bone mimicry | 242 |
| PLA | GO | Bone – 3D printed scaffold | Improved biocompatibility; 2× mineralization; 30% increase in Young's modulus | 173 and 174 |
| GG hydrogel film | rGO; TiO ₂ nanowires | Skin – wound healing hydrogel | <i>In vitro</i> study: stimulated fibroblast migration and wound closure; bioactive composite aiding skin regeneration | 258 |

incorporation of graphene into biopolymer scaffolds can enhance all these properties, improving mechanical strength while providing a surface that promotes cell adhesion and guides differentiation.^{234,235}

5.2 Bone and cartilage regeneration

BGNs are particularly well-suited for bone and cartilage tissue engineering due to their ability to provide robust mechanical



support and promote chondro/osteogenesis. In terms of osteogenic potential, numerous studies have shown that GO-based composites enhance osteoblastic differentiation. For example, PGA-co-PF/GO/HA electrospun scaffolds significantly increased alkaline phosphatase activity in MG63 cells,⁷⁸ and SA/HA/GnP films promoted apatite-like mineral formation in simulated body fluid.¹¹⁶ For load-bearing applications, the reinforcing effect of graphene is critical. PCL scaffolds reinforced with 3% graphene achieved a compressive modulus of 136.74 MPa,²⁴² while PLLA scaffolds incorporating 12% GO-hydroxyapatite reached a compressive strength of 21.52 MPa.²⁴³ In the challenging area of cartilage regeneration, biomimetic hydrogels composed of alginate, gelatin, chondroitin sulfate, and GO have been 3D bioprinted with mesenchymal stem cells, inducing intrinsic chondrogenic differentiation without exogenous growth factors.¹³⁹

5.3 Neural and cardiac tissue engineering

For neural tissue, scaffolds must support cell viability and provide electrical cues to guide regeneration. Graphene's conductivity is a major advantage. GO-COL and rGO-COL scaffolds supported extensive Schwann cell spreading and attachment,¹³³ while P(3HB) scaffolds containing GnP restored physiological firing patterns in cultured neurons.¹²² For cardiac tissue, which relies on electrical signal propagation for synchronized contractions, a 3D bioprinted cardiac "BioRing" fabricated from a GelMA/AlgMA/rGO bioink successfully mimicked key aspects of native heart tissue, including spontaneous and synchronous beating.¹⁷⁷

5.4 Skin and wound healing

BGNs can accelerate wound healing by acting as both protective wound dressings and pro-regenerative templates. Electrospun mats of SA/PVA/GnP loaded with curcumin provided sustained antimicrobial and antioxidant effects,¹⁵⁸ while GO-CS/PVP membranes enhanced skin wound repair in rat models, showing 33% faster wound closure than sterile gauze.²⁴⁴ For skin regeneration, CS-GO scaffolds implanted subdermally in rats supported tissue encapsulation and vascularization.¹⁶⁸ Furthermore, PLLA nanofiber scaffolds coated with polydopamine and carbon nanomaterials such as GO and CNTs have been shown to exhibit piezoelectric behavior, suggesting they could generate therapeutic electrical cues to accelerate healing in response to body movement.²⁴⁵

A detailed summary of selected studies on BGNs for tissue engineering is presented in Table 5.

6 Future outlook

The integration of biodegradable polymers with graphene-based nanomaterials has produced a formidable class of BGNs with multifunctional advantages for biomedicine. This review has detailed their significant advancements in both controlled drug delivery and tissue engineering. Looking forward, the primary goal is to translate these promising materials from the laboratory to clinical settings. This transition requires comprehensive long-term *in vivo* studies focusing

on BGN biocompatibility, degradation kinetics, biodistribution, and immunogenicity. Standardized testing protocols and adherence to good manufacturing practice guidelines are critical hurdles for achieving regulatory approval and enabling scalable, reproducible production.

Future research will likely focus on several key areas. One is the development of smart, multifunctional platforms (theranostics) capable of simultaneous diagnosis and therapy. At the chemical level, this will require more precise tailoring of interfacial chemistry between graphene derivatives and polymers. Another is 4D bioprinting, which can create scaffolds that change their shape or function in response to physiological stimuli after implantation. Advances here will depend not only on printing resolution but also on chemical innovation in photo-crosslinkable or reversible-bonding polymers, which dictate scaffold adaptability and long-term stability. BGNs are also exceptionally well-suited for personalized and AI-driven medicine, where therapies could be tailored based on a patient's specific needs. Mechanistic studies on hydrolytic, enzymatic, and oxidative degradation pathways will be critical to integrate predictive modeling with clinical translation. Finally, a growing focus on bioinspired and sustainable design, using green synthesis methods and mimicking natural tissue structures, will continue to drive the field forward. In particular, greener chemical synthesis routes for graphene and biodegradable polymers can help reduce cytotoxic byproducts while maintaining functional surface chemistry, aligning with both regulatory and sustainability goals.

7 Conclusions

BGNs represent a powerful convergence of nanotechnology, materials science, and biomedicine. These materials synergistically combine the exceptional mechanical, electrical, and responsive properties of graphene with the biocompatibility and sustainability of biodegradable polymers. In controlled drug delivery, BGNs enhance drug-loading capacity and enable targeted, stimuli-triggered release, improving therapeutic efficacy while minimizing side effects. In tissue engineering, they function as robust scaffolds that promote cellular growth and guide the regeneration of diverse tissues, including bone, skin, and cartilage. The fabrication of BGNs has progressed from simple solution-based methods to advanced techniques like 3D bioprinting, where innovations in crosslinking chemistry, surface functionalization, and reversible bonding are crucial for scaffold adaptability and stability. While BGNs hold immense promise for sustainable and effective biomedical solutions, further research, particularly rigorous *in vivo* experiments and clinical trials, is essential to advance these innovative materials toward practical medical applications.

Author contributions

M. Mohiuddin: writing – original draft, writing – review and editing, methodology. M. M. Rahman: writing – original draft, supervision, project administration, formal analysis, conceptualization. M. N. Uddin: writing – review and editing, supervision, project administration, conceptualization. R.



Hasan: writing – review and editing, methodology, investigation. I. Rahman: writing – review and editing, investigation, formal analysis, visualization.

Conflicts of interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability

All data used to prepare the manuscript are included. Additional explanations will be available upon request.

Acknowledgements

While preparing this work, the author(s) used *Gemini Advanced*, *QuillBot*, and *Grammarly* to paraphrase and edit the language. After using those tools, the author(s) reviewed and revised the content as needed and take(s) full responsibility for the publication's content.

References

- S. Adepun and S. Ramakrishna, *Molecules*, 2021, **26**, 5905.
- M. S. Rao, R. Gupta, M. J. Liguori, M. Hu, X. Huang, S. R. Mantena, S. W. Mittelstadt, E. A. Blomme and T. R. Van Vleet, *Frontiers in Big Data*, 2019, **2**, 25.
- D. F. Williams, *Front. Bioeng. Biotechnol.*, 2019, **7**, 127.
- J. Kurowiak, T. Klekiel and R. Będziński, *Int. J. Mol. Sci.*, 2023, **24**, 16952.
- C. M. Wells, M. Harris, L. Choi, V. P. Murali, F. D. Guerra and J. A. Jennings, *J. Funct. Biomater.*, 2019, **10**, 34.
- M. Modrák, M. Trebuňová, A. F. Balogová, R. Hudák and J. Živčák, *J. Funct. Biomater.*, 2023, **14**, 159.
- V. Gayathri, T. Khan, M. Gowtham, R. Balan and T. A. Sebaey, *Front. Bioeng. Biotechnol.*, 2025, **13**, 1533944.
- H. Zhao, R. Ding, X. Zhao, Y. Li, L. Qu, H. Pei, L. Yildirimer, Z. Wu and W. Zhang, *Drug Discovery Today*, 2017, **22**, 1302–1317.
- J. Zhang, H.-Y. Cao, J.-Q. Wang, G.-D. Wu and L. Wang, *Front. Cell Dev. Biol.*, 2021, **9**, 616888.
- H. Tang, S. Zhang, T. Huang, J. Zhang and B. Xing, *Environ. Sci. Technol.*, 2021, **55**, 14639–14648.
- P. Rajput and S. Khanchandani, *Int. J. Biol. Macromol.*, 2025, **311**, 143999.
- R. Saharan, S. K. Paliwal, A. Tiwari, M. A. Babu, V. Tiwari, R. Singh, S. K. Beniwal, M. Kumar, A. Sharma and W. H. Almalki, *J. Drug Delivery Sci. Technol.*, 2024, **94**, 105506.
- A. Mondal, A. K. Nayak, P. Chakraborty, S. Banerjee and B. C. Nandy, *Pharmaceutics*, 2023, **15**, 2064.
- Y. Zeng, Y. Xiang, R. Sheng, H. Tomás, J. Rodrigues, Z. Gu, H. Zhang, Q. Gong and K. Luo, *Bioact. Mater.*, 2021, **6**, 3358–3382.
- P. Bhatt, V. Kumar, V. Subramaniyan, K. Nagarajan, M. Sekar, S. V. Chinni and G. Ramachawolran, *Pharmaceutics*, 2023, **15**, 2066.
- M. Horue, J. M. Silva, I. R. Berti, L. R. Brandão, H. d. S. Barud and G. R. Castro, *Pharmaceutics*, 2023, **15**, 424.
- S. Mallakpour, F. Sirous and C. M. Hussain, *New J. Chem.*, 2021, **45**, 10565–10576.
- E. V. Prathyusha, S. S. Gomte, H. Ahmed, A. Prabakaran, M. Agrawal, N. Chella and A. Alexander, *Int. J. Biol. Macromol.*, 2024, 137834.
- M. Azizi-Lalabadi and S. M. Jafari, *Adv. Colloid Interface Sci.*, 2021, **292**, 102416.
- A. D. Ghuge, A. R. Shirode and V. J. Kadam, *Curr. Drug Targets*, 2017, **18**, 724–733.
- H. Shen, L. Zhang, M. Liu and Z. Zhang, *Theranostics*, 2012, **2**, 283–294.
- Y. Xiao, Y. X. Pang, Y. Yan, P. Qian, H. Zhao, S. Manickam, T. Wu and C. H. Pang, *Adv. Sci.*, 2023, **10**, 2205292.
- A. Gostaviceanu, S. Gavrilas, L. Copolovici and D. M. Copolovici, *Int. J. Mol. Sci.*, 2024, **25**, 10174.
- M. S. A. Bhuyan, M. N. Uddin, M. M. Islam, F. A. Bipasha and S. S. Hossain, *Int. Nano Lett.*, 2016, **6**, 65–83.
- H. I. Røst, R. K. Chellappan, F. S. Strand, A. Grubišić-Čabo, B. P. Reed, M. J. Prieto, L. C. Tănase, L. de Souza Caldas, T. Wongpinij and C. Euaruksakul, *J. Phys. Chem. C*, 2021, **125**, 4243–4252.
- Y. Liu, J. Zheng, X. Zhang, K. Li, Y. Du, G. Yu, Y. Jia and Y. Zhang, *J. Appl. Polym. Sci.*, 2021, **138**, 50474.
- J. Narayan and K. Bezborah, *RSC Adv.*, 2024, **14**, 13413–13444.
- X.-M. Han, K.-W. Zheng, R.-L. Wang, S.-F. Yue, J. Chen, Z.-W. Zhao, F. Song, Y. Su and Q. Ma, *Am. J. Transl. Res.*, 2020, **12**, 1515.
- V. Marsala, Y. Gerasymchuk, M. L. Saladino, E. Paluch, M. Wawrzyńska, V. Boiko, X. Li, C. Giordano, D. Hreniak and B. Sobieszkańska, *Molecules*, 2025, **30**, 240.
- J. V. Lim, S. T. Bee, L. Tin Sin, C. T. Ratnam and Z. A. Abdul Hamid, *Polymers*, 2021, **13**, 3547.
- W. S. Hummers Jr and R. E. Offeman, *J. Am. Chem. Soc.*, 1958, **80**, 1339.
- P. Feicht, J. Biskupek, T. E. Gorelik, J. Renner, C. E. Halbig, M. Maranska, F. Puchtler, U. Kaiser and S. Eigler, *Chem.–Eur. J.*, 2019, **25**, 8955–8959.
- H. L. Poh, F. Šaněk, A. Ambrosi, G. Zhao, Z. Sofer and M. Pumera, *Nanoscale*, 2012, **4**, 3515–3522.
- G. Utkan, G. Yumusak, B. C. Tunalı, T. Ozturk and M. Turk, *ACS Omega*, 2023, **8**, 31188–31200.
- B. Lesiak, G. Trykowski, J. Tóth, S. Biniak, L. Kövér, N. Rangam, L. Stobinski and A. Malolepszy, *J. Mater. Sci.*, 2021, **56**, 3738–3754.
- K. K. H. De Silva, H.-H. Huang and M. Yoshimura, *Appl. Surf. Sci.*, 2018, **447**, 338–346.
- P. Zhang, Z. Li, S. Zhang and G. Shao, *Energy Environ. Mater.*, 2018, **1**, 5–12.
- N.-F. Chiu, C.-D. Yang, C.-C. Chen and C.-T. Kuo, *Sens. Actuators, B*, 2018, **258**, 981–990.



- 39 A. Samir, F. H. Ashour, A. A. A. Hakim and M. Bassyouni, *npj Mater. Degrad.*, 2022, **6**, 68.
- 40 E. Avcu, F. E. Bastan, M. Guney, Y. Yildiran Avcu, M. A. Ur Rehman and A. R. Boccaccini, *Acta Biomater.*, 2022, **151**, 1–44.
- 41 Z.-H. Li, S.-C. Ji, Y.-Z. Wang, X.-C. Shen and H. Liang, *Front. Mater. Sci.*, 2013, **7**, 237–247.
- 42 Y. Lyu, Y. Liu, H. He and H. Wang, *Gels*, 2023, **9**, 431.
- 43 R. Andreazza, A. Morales, S. Pieniz and J. Labidi, *Polymers*, 2023, **15**, 1026.
- 44 X. Jiang, Z. Du, X. Zhang, F. Zaman, Z. Song, Y. Guan, T. Yu and Y. Huang, *Front. Bioeng. Biotechnol.*, 2023, **11**, 1158749.
- 45 Y. Isobe, T. Kosaka, G. Kuwahara, H. Mikami, T. Saku and S. Kodama, *Materials*, 2012, **5**, 501–511.
- 46 S. Tortorella, M. Maturi, V. V. Buratti, G. Vozzolo, E. Locatelli, L. Sambri and M. C. Franchini, *RSC Adv.*, 2021, **11**, 39004–39026.
- 47 V. B. Sindhusa and J. N. Doraiswamy, *Cureus*, 2023, **15**, e48667.
- 48 Z. Arabpour, F. Abedi, M. Salehi, S. M. Baharnoori, M. Soleimani and A. R. Djalilian, *Int. J. Mol. Sci.*, 2024, **25**, 1982.
- 49 T. M. M. Ways, W. M. Lau and V. V. Khutoryanskiy, *Polymers*, 2018, **10**, 267.
- 50 N. Farshidfar, S. Iravani and R. S. Varma, *Mar. Drugs*, 2023, **21**, 189.
- 51 P. Zou, J. Yao, Y.-N. Cui, T. Zhao, J. Che, M. Yang, Z. Li and C. Gao, *Gels*, 2022, **8**, 364.
- 52 C.-S. Lee and H. S. Hwang, *Gels*, 2023, **9**, 951.
- 53 V. S. Waghmare, P. R. Wadke, S. Dyawanapelly, A. Deshpande, R. Jain and P. Dandekar, *Bioact. Mater.*, 2018, **3**, 255–266.
- 54 F. Jiang, X.-W. Xu, F.-Q. Chen, H.-F. Weng, J. Chen, Y. Ru, Q. Xiao and A.-F. Xiao, *Mar. Drugs*, 2023, **21**, 299.
- 55 Y. Huang, S. Peng, Y. Chen and B. Chu, *Gels*, 2025, **11**, 255.
- 56 M. S. Rahman, M. S. Hasan, A. S. Nitai, S. Nam, A. K. Karmakar, M. S. Ahsan, M. J. Shiddiky and M. B. Ahmed, *Polymers*, 2021, **13**, 1345.
- 57 A. Di Mola, M. R. Landi, A. Massa, U. D'Amora and V. Guarino, *Int. J. Mol. Sci.*, 2022, **23**, 14372.
- 58 J. G. Pérez-Flores, L. García-Curiel, E. Pérez-Escalante, E. Contreras-López and E. J. Olloqui, *Heliyon*, 2024, **10**, e25445.
- 59 M. U. A. Khan, M. A. Raza, S. I. A. Razak, M. R. Abdul Kadir, A. Haider, S. A. Shah, A. H. Mohd Yusof, S. Haider, I. Shakir and S. Aftab, *J. Tissue Eng. Regen. Med.*, 2020, **14**, 1488–1501.
- 60 M. U. A. Khan, M. A. Al-Thebaiti, M. U. Hashmi, S. Aftab, S. I. Abd Razak, S. Abu Hassan, M. R. Abdul Kadir and R. Amin, *Materials*, 2020, **13**, 971.
- 61 C. M. Robles-Kanafany, M. L. Del Prado-Audelo, M. González-Torres, D. M. Giraldo-Gomez, I. H. Caballero-Florán, M. González-Del Carmen, J. Sharifi-Rad, G. Figueroa-González, O. D. Reyes-Hernández and H. Cortés, *Cell. Mol. Biol.*, 2021, **67**, 89–95.
- 62 H. Poudel, A. B. RanguMagar, P. Singh, A. Oluremi, N. Ali, F. Watanabe, J. Batta-Mpouma, J. W. Kim, A. Ghosh and A. Ghosh, *Bioengineering*, 2023, **10**, 1088.
- 63 T. Hardingham, *Osteoarthr. Cartil.*, 1998, **6**, 3–5.
- 64 M. Bhatia and M. Ahuja, *Int. J. Biol. Macromol.*, 2015, **72**, 495–501.
- 65 V. E. Santo, A. M. Frias, M. Carida, R. Cancedda, M. E. Gomes, J. F. Mano and R. L. Reis, *Biomacromolecules*, 2009, **10**, 1392–1401.
- 66 D. Momekova, E. Ivanov, S. Konstantinov, F. Ublekov and P. D. Petrov, *Polymers*, 2020, **12**, 1172.
- 67 Y. Sun, X. Xu, Q. Zhang, D. Zhang, X. Xie, H. Zhou, Z. Wu, R. Liu and J. Pang, *Polymers*, 2023, **15**, 1852.
- 68 M. Rosalia, D. Rubes, M. Serra, I. Genta, R. Dorati and B. Conti, *Polymers*, 2024, **16**, 1405.
- 69 P. Piszko, M. Włodarczyk, S. Zielińska, M. Gazińska, P. Płociński, K. Rudnicka, A. Szwed, A. Krupa, M. Grzymajło, A. Sobczak-Kupiec, D. Słota, M. Kobielarz, M. Wojtków and K. Szustakiewicz, *Int. J. Mol. Sci.*, 2021, **22**, 8587.
- 70 B. Dariš and Ž. Knez, *Acta Pharm.*, 2020, **70**, 1–15.
- 71 A. P. Bonartsev, V. V. Voinova, A. V. Volkov, A. A. Muraev, E. M. Boyko, A. A. Venediktov, N. N. Didenko and A. A. Dolgalev, *Sovremennye Tehnologii v Medicine*, 2022, **14**, 78–90.
- 72 R. Dwivedi, S. Kumar, R. Pandey, A. Mahajan, D. Nandana, D. S. Katti and D. Mehrotra, *J. Oral Biol. Craniofac. Res.*, 2020, **10**, 381–388.
- 73 E. Capuana, F. Lopresti, M. Ceraulo and V. La Carrubba, *Polymers*, 2022, **14**, 1153.
- 74 Y. Dai, T. Lu, M. Shao and F. Lyu, *Front. Bioeng. Biotechnol.*, 2022, **10**, 1011783.
- 75 S. Castañeda-Rodríguez, M. González-Torres, R. M. Ribas-Aparicio, M. L. Del Prado-Audelo, G. Leyva-Gómez, E. S. Güerer and J. Sharifi-Rad, *J. Biol. Eng.*, 2023, **17**, 21.
- 76 H. K. Makadia and S. J. Siegel, *Polymers*, 2011, **3**, 1377–1397.
- 77 Z. Cai, Y. Wan, M. L. Becker, Y.-Z. Long and D. Dean, *Biomaterials*, 2019, **208**, 45–71.
- 78 A. M. Díez-Pascual and A. L. Díez-Vicente, *J. Mater. Chem. B*, 2017, **5**, 4084–4096.
- 79 D. Mondal, A. Srinivasan, P. Comeau, Y.-C. Toh and T. L. Willett, *Mater. Sci. Eng., C*, 2021, **118**, 111400.
- 80 S. C. Mauck, S. Wang, W. Ding, B. J. Rohde, C. K. Fortune, G. Yang, S.-K. Ahn and M. L. Robertson, *Macromolecules*, 2016, **49**, 1605–1615.
- 81 H. Yin, M. Zhu, Y. Wang, L. Luo, Q. Ye and B. H. Lee, *Frontiers in Soft Matter*, 2023, **2**, 1101680.
- 82 R. Binaymotlagh, L. Chronopoulou and C. Palocci, *J. Funct. Biomater.*, 2023, **14**, 233.
- 83 A. M. Díez-Pascual, *Polymers*, 2021, **13**, 2978.
- 84 H. Xue, P. Wang, L. Ji, K. Zhang, S. Ge and J. Tan, *J. Agric. Food Res.*, 2025, **24**, 102395.
- 85 W. Feng and Z. Wang, *iScience*, 2022, **25**, 103629.
- 86 S. Zhang, J. Dong, R. Pan, Z. Xu, M. Li and R. Zang, *Polymers*, 2023, **15**, 2149.
- 87 N. Sittisanguanphan, N. Paradee and A. Sirivat, *J. Pharm. Sci.*, 2022, **111**, 1633–1641.
- 88 R. Binaymotlagh, L. Chronopoulou and C. Palocci, *Materials*, 2025, **18**, 2978.



- 89 H. J. Salavagione, G. Martínez and G. Ellis, *Macromol. Rapid Commun.*, 2011, **32**, 1771–1789.
- 90 R. Muñoz, L. León-Boigues, E. López-Elvira, C. Munuera, L. Vázquez, F. Mompeán, J. Á. Martín-Gago, I. Palacio and M. García-Hernández, *ACS Appl. Mater. Interfaces*, 2023, **15**, 46171–46180.
- 91 X. Liu, A. L. Miller II, B. E. Waletzki and L. Lu, *J. Biomed. Mater. Res., Part A*, 2018, **106**, 1247–1257.
- 92 A. Kausar, I. Ahmad, M. H. Eisa, M. Maaza and H. Khan, *Nanomanufacturing*, 2023, **3**, 1–19.
- 93 S. J. Lee, S. J. Yoon and I.-Y. Jeon, *Polymers*, 2022, **14**, 4733.
- 94 A. M. Díez-Pascual, *Polymers*, 2022, **14**, 2102.
- 95 M. Dong, Y. Sun, D. J. Dunstan, R. J. Young and D. G. Papageorgiou, *Nanoscale*, 2024, **16**, 13247–13299.
- 96 M. Silva, N. M. Alves and M. C. Paiva, *Polym. Adv. Technol.*, 2018, **29**, 687–700.
- 97 D. Spasevska, G. Leal, M. Fernández, J. B. Gilev, M. Paulis and R. Tomovska, *RSC Adv.*, 2015, **5**, 16414–16421.
- 98 A. F. Ghanem and M. H. Abdel Rehim, *Biomedicines*, 2018, **6**, 63.
- 99 S. Qamar, N. Ramzan and W. Aleem, *Synth. Met.*, 2024, **307**, 117697.
- 100 L. Sisti, G. Totaro, A. Celli, L. Giorgini, S. Ligi and M. Vannini, *Polymers*, 2021, **13**, 1377.
- 101 A. T. Lawal, *Cogent Chem.*, 2020, **6**, 1833476.
- 102 S. Uten, P. Boonbanjong, Y. Prueksathaporn, K. Treerattrakoon, N. Sathirapongsasuti, N. Chanlek, S. Pinitsoontorn, P. Luksirikul and D. Japrungr, *ACS Omega*, 2024, **9**, 2263–2271.
- 103 F. Li, Y. Huang, K. Huang, J. Lin and P. Huang, *Int. J. Mol. Sci.*, 2020, **21**, 390.
- 104 R. Jiffrin, S. I. A. Razak, M. I. Jamaludin, A. S. A. Hamzah, M. A. Mazian, M. A. T. Jaya, M. Z. Nasrullah, M. Majrashi, A. Theyab, A. A. Aldarmahi, Z. Awan, M. M. Abdel-Daim and A. K. Azad, *Polymers*, 2022, **14**, 3725.
- 105 A. M. Muñoz-Gonzalez, S. Leal-Marin, D. Clavijo-Grimaldo and B. Glasmacher, *Int. J. Artif. Organs*, 2024, **47**, 633–641.
- 106 W. Xu, X. Zhang, P. Yang, O. Långvik, X. Wang, Y. Zhang, F. Cheng, M. Österberg, S. Willför and C. Xu, *ACS Appl. Mater. Interfaces*, 2019, **11**, 12389–12400.
- 107 P. H. N. Cardoso and E. S. Araújo, *Compounds*, 2024, **4**, 71–105.
- 108 R. Patil and S. Alimperti, *J. Funct. Biomater.*, 2024, **15**, 82.
- 109 R. Mohan, A. Jineesh and N. M. Prabu, *Environ. Eng. Res.*, 2022, **27**, 210161.
- 110 L. Di Muzio, P. Simonetti, V. C. Carriero, C. Brandelli, J. Trilli, C. Sergi, J. Tirillò, F. Cairone, S. Cesa, G. Radochia, S. Schippa, S. Petralito, P. Paolicelli and M. A. Casadei, *Molecules*, 2022, **27**, 2959.
- 111 N. Jouault, D. Zhao and S. K. Kumar, *Macromolecules*, 2014, **47**, 5246–5255.
- 112 S. N. Tripathi, G. S. Rao, A. B. Mathur and R. Jasra, *RSC Adv.*, 2017, **7**, 23615–23632.
- 113 S. Cheng and G. S. Grest, *ACS Macro Lett.*, 2016, **5**, 694–698.
- 114 X. Xu, Z. Ren, M. Zhang and L. Ma, *J. Appl. Polym. Sci.*, 2021, **138**, 51173.
- 115 E. Kılıç and N. P. Bayramgil, *Results Chem.*, 2022, **4**, 100544.
- 116 S. Iswarya, T. Theivasanthi, K. Chinnaiah and S. C. B. Gopinath, *Appl. Nanosci.*, 2024, **14**, 109–122.
- 117 B. Farshid, G. Lalwani, M. S. Mohammadi, J. S. Sankaran, S. Patel, S. Judex, J. Simonsen and B. Sitharaman, *J. Biomed. Mater. Res., Part A*, 2019, **107**, 1143–1153.
- 118 S. Sinha Ray and M. Okamoto, *Prog. Polym. Sci.*, 2003, **28**, 1539–1641.
- 119 M. M. Rahman, K. H. Khan, M. M. H. Parvez, N. Irizarry and M. N. Uddin, *Processes*, 2025, **13**, 994.
- 120 H. M. Shanshool, M. Yahaya, W. M. M. Yunus and I. Y. Abdullah, *J. Mater. Sci.: Mater. Electron.*, 2016, **27**, 9804–9811.
- 121 B. Davoodi, V. Goodarzi, H. Hosseini, M. Tirgar, S. Shojaei, A. Asefnejad, A. Saedi, F. Oroojalian and S. Zamanlui, *J. Polym. Res.*, 2022, **29**, 54.
- 122 M. Moschetta, M. Chiacchiaretta, F. Cesca, I. Roy, A. Athanassiou, F. Benfenati, E. L. Papadopoulou and M. Bramini, *Front. Neurosci.*, 2021, **15**, 731198.
- 123 W. K. Zhu, H. P. Cong, H. B. Yao, L. B. Mao, A. M. Asiri, K. A. Alamry, H. M. Marwani and S. H. Yu, *Small*, 2015, **11**, 4298–4302.
- 124 S. P. Ogilvie, M. J. Large, M. A. O'mara, A. C. Sehnal, A. Amorim Graf, P. J. Lynch, A. J. Cass, J. P. Salvage, M. Alfonso and P. Poulin, *ACS Nano*, 2022, **16**, 1963–1973.
- 125 T. D. Gamot, A. R. Bhattacharyya, T. Sridhar, F. Beach, R. F. Tabor and M. Majumder, *Langmuir*, 2017, **33**, 10311–10321.
- 126 J. Luo, L. Yang, D. Sun, Z. Gao, K. Jiao and J. Zhang, *Small*, 2020, **16**, 2003426.
- 127 N. M. Farokhi, J. M. Milani and Z. R. Amiri, *J. Food Eng.*, 2024, **367**, 111885.
- 128 Z. Chen, G. Zhang, Y. Luo and Z. Suo, *Proc. Natl. Acad. Sci. U. S. A.*, 2024, **121**, e2322684121.
- 129 M. Ghotbi, M. Pourmadadi, F. Yazdian and A. Hallajani, *Inorg. Chem. Commun.*, 2024, **170**, 113119.
- 130 S. Ostovar, M. Pourmadadi and M. A. Zaker, *Int. J. Biol. Macromol.*, 2023, **253**, 127091.
- 131 S. De, K. Patra, D. Ghosh, K. Dutta, A. Dey, G. Sarkar, J. Maiti, A. Basu, D. Rana and D. Chattopadhyay, *ACS Biomater. Sci. Eng.*, 2018, **4**, 514–531.
- 132 K. Liang, E. M. Spiesz, D. T. Schmieden, A.-W. Xu, A. S. Meyer and M.-E. Aubin-Tam, *ACS Nano*, 2020, **14**, 14731–14739.
- 133 A. F. Girão, G. Gonçalves, K. S. Bhangra, J. B. Phillips, J. Knowles, G. Irueta, M. K. Singh, I. Bdkin, A. Completo and P. A. Marques, *RSC Adv.*, 2016, **6**, 49039–49051.
- 134 J. Chen, B. Liu and X. Gao, *Results Phys.*, 2020, **16**, 102974.
- 135 R. Ramachandran, D. Jung and A. M. Spokoiny, *NPG Asia Mater.*, 2019, **11**, 19.
- 136 Y.-W. Fu, W.-F. Sun and X. Wang, *Polymers*, 2020, **12**, 420.
- 137 S. H. Moon, H. J. Hwang, H. R. Jeon, S. J. Park, I. S. Bae and Y. J. Yang, *Front. Bioeng. Biotechnol.*, 2023, **11**, 1127757.
- 138 M. Rahimi, V. Pirouzfard and H. Sakhaeina, *J. Polym. Environ.*, 2024, **32**, 2762–2779.



- 139 F. Olate-Moya, G. Rubí-Sans, E. Engel, M. Á. Mateos-Timoneda and H. Palza, *Biomacromolecules*, 2024, **25**, 3312–3324.
- 140 A. I. Raafat and A. E.-H. Ali, *Polym. Bull.*, 2017, **74**, 2045–2062.
- 141 K. Mylvaganam and L. Zhang, *J. Phys. Chem. C*, 2013, **117**, 2817–2823.
- 142 S. Das, A. S. Wajid, J. L. Shelburne, Y.-C. Liao and M. J. Green, *ACS Appl. Mater. Interfaces*, 2011, **3**, 1844–1851.
- 143 J. Fawaz and V. Mittal, in *Synthesis Techniques for Polymer Nanocomposites*, ed. V. Mittal, Wiley-VCH GmbH, Weinheim, Germany, 2014, pp. 1–30.
- 144 N. Basavegowda and K.-H. Baek, *Polymers*, 2021, **13**, 4198.
- 145 N. Rabiee, M. Bagherzadeh, A. M. Ghadiri, Y. Fatahi, A. Aldhafer, P. Makvandi, R. Dinarvand, M. Jouyandeh, M. R. Saeb, M. Mozafari, M. Shokouhimehr, M. R. Hamblin and R. S. Varma, *ACS Appl. Bio Mater.*, 2021, **4**, 5336–5351.
- 146 F. J. P. Costa, M. Nave, R. Lima-Sousa, C. G. Alves, B. L. Melo, I. J. Correia and D. de Melo-Diogo, *Int. J. Pharm.*, 2023, **635**, 122713.
- 147 M. H. Aghajan, M. Panahi-Sarmad, N. Alikarami, S. Shojaei, A. Saeidi, H. A. Khonakdar, M. Shahrousvan and V. Goodarzi, *Eur. Polym. J.*, 2020, **131**, 109720.
- 148 Z. Wang, F. Cui, Y. Sui and J. Yan, *Beilstein J. Org. Chem.*, 2023, **19**, 1580–1603.
- 149 I. S. Tsagkalias, T. K. Manios and D. S. Achilias, *Polymers*, 2017, **9**, 432.
- 150 M. U. A. Khan, S. I. A. Razak, S. Rehman, A. Hasan, S. Qureshi and G. M. Stojanović, *Int. J. Biol. Macromol.*, 2022, **222**, 462–472.
- 151 M. U. Aslam Khan, W. S. Al-Arjan, M. S. Binkadem, H. Mehboob, A. Haider, M. A. Raza, S. I. Abd Razak, A. Hasan and R. Amin, *Nanomaterials*, 2021, **11**, 1319.
- 152 A. Ribas-Massonis, M. Cicujano, J. Duran, E. Besalú and A. Poater, *Polymers*, 2022, **14**, 2856.
- 153 A. Kausar and I. Ahmad, *J. Compos. Sci.*, 2023, **7**, 290.
- 154 M. Toriello, M. Afsari, H. K. Shon and L. D. Tijing, *Membranes*, 2020, **10**, 204.
- 155 A. Al-Abduljabbar and I. Farooq, *Polymers*, 2023, **15**, 65.
- 156 R. Nayak, R. Padhye, I. L. Kyratzis, Y. B. Truong and L. Arnold, *Text. Res. J.*, 2013, **83**, 606–617.
- 157 Y. Sun, S. Cheng, W. Lu, Y. Wang, P. Zhang and Q. Yao, *RSC Adv.*, 2019, **9**, 25712–25729.
- 158 M. Rezaei, M. Nikkhah, S. Mohammadi, S. H. Bahrami and M. Sadeghizadeh, *J. Appl. Polym. Sci.*, 2021, **138**, 50884.
- 159 F. Norouzi, M. Pourmadadi, F. Yazdian, K. Khoshmaram, J. Mohammadnejad, M. H. Sanati, F. Chogan, A. Rahdar and F. Baino, *J. Funct. Biomater.*, 2022, **13**, 300.
- 160 R. Jaswal, D. Kumar, A. I. Rezk, V. K. Kaliannagounder, C. H. Park and K. H. Min, *Colloids Surf., B*, 2024, **237**, 113820.
- 161 A. Keirouz, Z. Wang, V. S. Reddy, Z. K. Nagy, P. Vass, M. Buzgo, S. Ramakrishna and N. Radacsi, *Adv. Mater. Technol.*, 2023, **8**, 2201723.
- 162 N. Ren, A. Qiao, M. Cui, R. Huang, W. Qi and R. Su, *Chem. Eng. Sci.*, 2023, **282**, 119320.
- 163 M. A. Al Faruque, R. Remadevi, A. Guirguis, A. Kiziltas, D. Mielewski and M. Naebe, *Sci. Rep.*, 2021, **11**, 12068.
- 164 H. He, L. Guan and H. Le Ferrand, *J. Mater. Chem. A*, 2022, **10**, 19129–19168.
- 165 M. Oksuz and H. Y. Erbil, *RSC Adv.*, 2018, **8**, 17443–17452.
- 166 S. Talebian, M. Mehrali, R. Raad, F. Safaei, J. Xi, Z. Liu and J. Foroughi, *Front. Chem.*, 2020, **8**, 88.
- 167 C. Katrilaka, N. Karipidou, N. Petrou, C. Manglaris, G. Katrilakas, A. N. Tzavellas, M. Pitou, E. E. Tsiridis, T. Choli-Papadopoulou and A. Aggeli, *Materials*, 2023, **16**, 4425.
- 168 C. Valencia, C. H. Valencia, F. Zuluaga, M. E. Valencia, J. H. Mina and C. D. Grande-Tovar, *Molecules*, 2018, **23**, 2651.
- 169 S. Liu, C. Zhou, S. Mou, J. Li, M. Zhou, Y. Zeng, C. Luo, J. Sun, Z. Wang and W. Xu, *Mater. Sci. Eng., C*, 2019, **105**, 110137.
- 170 A.-V. Do, B. Khorsand, S. M. Geary and A. K. Salem, *Adv. Healthcare Mater.*, 2015, **4**, 1742–1762.
- 171 A. Seyed-salehi, L. Daneshmandi, M. Barajaa, J. Riordan and C. T. Laurencin, *Sci. Rep.*, 2020, **10**, 22210.
- 172 J. M. Seok, G. Choe, S. J. Lee, M.-A. Yoon, K.-S. Kim, J. H. Lee, W. D. Kim, J. Y. Lee, K. Lee and S. A. Park, *Mater. Des.*, 2021, **209**, 109941.
- 173 M. Mashhadi Keshtiban, H. Taghvaei, R. Noroozi, V. Eskandari, Z. U. Arif, M. Bodaghi, H. Bardania and A. Hadi, *Adv. Eng. Mater.*, 2024, **26**, 2301260.
- 174 H. Belaid, S. Nagarajan, C. Teyssier, C. Barou, J. Barés, S. Balme, H. Garay, V. Huon, D. Cornu, V. Cavallès and M. Bechelany, *Mater. Sci. Eng., C*, 2020, **110**, 110595.
- 175 S. Agarwal, S. Saha, V. K. Balla, A. Pal, A. Barui and S. Bodhak, *Frontiers in Mechanical Engineering*, 2020, **6**, 589171.
- 176 T. d. P. L. Lima, C. A. d. A. Canelas, V. O. C. Concha, F. A. M. d. Costa and M. F. Passos, *J. Funct. Biomater.*, 2022, **13**, 214.
- 177 A. Mousavi, A. Hedayatnia, P. P. van Vliet, D. R. Dartora, N. Wong, N. Rafatian, A. M. Nuyt, C. Moraes, A. Ajji, G. Andelfinger and H. Savoji, *Appl. Mater. Today*, 2024, **36**, 102035.
- 178 S. Sharma, H. Sharma and R. Sharma, *Chemistry of Inorganic Materials*, 2024, **2**, 100035.
- 179 C. Comanescu, *Coatings*, 2023, **13**, 1772.
- 180 N. İşiklan, G. Geyik and E. Güncüm, *Mater. Today Chem.*, 2024, **41**, 102323.
- 181 R. Justin and B. Chen, *Interface Focus*, 2018, **8**, 20170055.
- 182 G. F. Stiufluc and R. I. Stiufluc, *Appl. Sci.*, 2024, **14**, 1623.
- 183 Q. Zhang, Z. Wu, N. Li, Y. Pu, B. Wang, T. Zhang and J. Tao, *Mater. Sci. Eng., C*, 2017, **77**, 1363–1375.
- 184 E. Khakpour, S. Salehi, S. M. Naghib, S. Ghorbanzadeh and W. Zhang, *Front. Bioeng. Biotechnol.*, 2023, **11**, 1129768.
- 185 J. Singh and P. Nayak, *J. Polym. Sci.*, 2023, **61**, 2828–2850.
- 186 E. Daneshmoghlanlou, M. Miralinaghi, E. Moniri and S. K. Sadjady, *J. Polym. Environ.*, 2022, **30**, 3718–3736.
- 187 M. U. A. Khan, Z. Yaqoob, M. N. M. Ansari, S. I. A. Razak, M. A. Raza, A. Sajjad, S. Haider and F. M. Busra, *Polymers*, 2021, **13**, 3124.



- 188 J. Siepmann, K. Elkharraz, F. Siepmann and D. Klose, *Biomacromolecules*, 2005, **6**, 2312–2319.
- 189 S. Li and S. McCarthy, *Biomaterials*, 1999, **20**, 35–44.
- 190 Y. Liu, S. Longqing, J. Yuxuan, J. Shaojing, Z. Xiaokang, L. Song, C. Jing and J. Hu, *Int. J. Nanomed.*, 2025, **20**, 705–721.
- 191 P. Misra, B. K. Mishra and G. B. Behera, *Int. J. Chem. Kinet.*, 1991, **23**, 639–654.
- 192 A. C. Dash, B. Dash and S. Praharaj, *J. Chem. Soc., Dalton Trans.*, 1981, (10), 2063–2069.
- 193 M. Karimi, P. Sahandi Zangabad, A. Ghasemi, M. Amiri, M. Bahrami, H. Malekzad, H. Ghahramanzadeh Asl, Z. Mahdih, M. Bozorgomid, A. Ghasemi, M. R. Rahmani Taji Boyuk and M. R. Hamblin, *ACS Appl. Mater. Interfaces*, 2016, **8**, 21107–21133.
- 194 M. Bikram and J. L. West, *Expet Opin. Drug Deliv.*, 2008, **5**, 1077–1091.
- 195 D. N. Céspedes-Valenzuela, S. Sánchez-Rentería, J. Cifuentes, S. C. Gómez, J. A. Serna, L. Rueda-Gensini, C. Ostos, C. Muñoz-Camargo and J. C. Cruz, *Front. Bioeng. Biotechnol.*, 2022, **10**, 947616.
- 196 Q. Hu, P. S. Katti and Z. Gu, *Nanoscale*, 2014, **6**, 12273–12286.
- 197 M. Li, G. Zhao, W.-K. Su and Q. Shuai, *Front. Chem.*, 2020, **8**, 647.
- 198 R. L. Minehan and M. P. Del Borgo, *Frontiers in Biomaterials Science*, 2022, **1**, 916985.
- 199 J. Liu, W. Kang and W. Wang, *Photochem. Photobiol.*, 2022, **98**, 288–302.
- 200 Y. Xing, B. Zeng and W. Yang, *Front. Bioeng. Biotechnol.*, 2022, **10**, 1075670.
- 201 R. J. Mitchell, D. Havrylyuk, A. C. Hachey, D. K. Heidary and E. C. Glazer, *Chem. Sci.*, 2025, **16**, 721–734.
- 202 M. Fernández and J. Orozco, *Polymers*, 2021, **13**, 2464.
- 203 C. L. Weaver, J. M. LaRosa, X. Luo and X. T. Cui, *ACS Nano*, 2014, **8**, 1834–1843.
- 204 D. Zahn, A. Weidner, K. Saatchi, U. O. Häfeli and S. Dutz, *Current Directions in Biomedical Engineering*, 2019, **5**, 161–164.
- 205 Z. Wang, C. Liu, B. Chen and Y. Luo, *Int. J. Biol. Macromol.*, 2021, **168**, 38–45.
- 206 F. V. Lavrentev, V. V. Shilovskikh, V. S. Alabusheva, V. Y. Yurova, A. A. Nikitina, S. A. Ulasevich and E. V. Skorb, *Molecules*, 2023, **28**, 5931.
- 207 M. S. B. Reddy, D. Ponnamma, R. Choudhary and K. K. Sadasivuni, *Polymers*, 2021, **13**, 1105.
- 208 M. V. S. Varma, A. M. Kaushal, A. Garg and S. Garg, *Am. J. Drug Delivery*, 2004, **2**, 43–57.
- 209 D. Hazarika, K. Gupta, M. Mandal and N. Karak, *ACS Omega*, 2018, **3**, 2292–2303.
- 210 V. J. Sawant, B. V. Tawade, V. M. Desai, B. B. Dongare and S. V. Nipane, *Prog. Biomater.*, 2022, **11**, 193–205.
- 211 T. M. Joseph, A. B. Unni, K. S. Joshy, D. Kar Mahapatra, J. Haponiuk and S. Thomas, *C-J. Carbon Res.*, 2023, **9**, 30.
- 212 L. Moradi, L. Witek, V. Vivekanand Nayak, A. Cabrera Pereira, E. Kim, J. Good and C. J. Liu, *Biomaterials*, 2023, **301**, 122289.
- 213 N. Hossain, M. A. Chowdhury, M. Kchaou, A. Alam and M. M. Rahman, *Chem. Phys. Impact*, 2023, **6**, 100180.
- 214 E. Gudiño, C. M. Oishi and A. Sequeira, *Int. J. Numer. Methods Eng.*, 2018, **114**, 292–320.
- 215 A. Deb, N. G. Andrews and V. Raghavan, *Int. J. Biol. Macromol.*, 2018, **113**, 515–525.
- 216 I. A. A. Ibrahim, A. R. Alzahrani, I. M. Alanazi, N. Shahzad, I. Shahid, A. H. Falemban, M. F. N. Azlina and P. Arulsevan, *Int. J. Biol. Macromol.*, 2023, **253**, 127334.
- 217 G. Singh, B. P. Nenavathu, K. Imtiyaz and M. Moshahid A Rizvi, *Biomed. Pharmacother.*, 2020, **129**, 110443.
- 218 S. Nazir, M. Umar Aslam Khan, W. Shamsan Al-Arjan, S. Izwan Abd Razak, A. Javed and M. Rafiq Abdul Kadir, *Arab. J. Chem.*, 2021, **14**, 103120.
- 219 I. Sahu and P. Chakraborty, *Colloids Surf., B*, 2024, **233**, 113654.
- 220 S. Xiang, C. Guilbaud-Chéreau, P. Hoschtettler, L. Stefan, A. Bianco and C. Ménard-Moyon, *Int. J. Biol. Macromol.*, 2024, **255**, 127919.
- 221 S. Javanbakht and A. Shaabani, *Int. J. Biol. Macromol.*, 2019, **123**, 389–397.
- 222 B. N. Kumara, R. Shambhu, A. Prabhu and K. S. Prasad, *Carbohydr. Polym.*, 2022, **289**, 119426.
- 223 Z. Karimzadeh and H. Namazi, *J. Polym. Res.*, 2022, **29**, 37.
- 224 S. R. Obireddy, A. Ayyakannu, G. Kasi, B. Sridharan, W.-F. Lai and K. Viswanathan, *Int. J. Nanomed.*, 2025, **20**, 1965–1981.
- 225 S. Dhanavel, P. Praveena, V. Narayanan and A. Stephen, *Polym. Bull.*, 2020, **77**, 5681–5696.
- 226 A. D. Sontakke, P. Gupta, S. K. Banerjee and M. K. Purkait, *Int. J. Biol. Macromol.*, 2024, **271**, 132621.
- 227 A. Hardiansyah, A. Randy, R. T. Dewi, M. Angelina, N. Yudasari, S. Rahayu, I. M. Ulfah, F. Maryani, Y.-W. Cheng and T.-Y. Liu, *Polymers*, 2022, **14**, 3163.
- 228 S. Agila and J. Poornima, in *2015 IEEE 15th International Conference on Nanotechnology (IEEE-NANO)*, IEEE, 2015, pp. 1058–1061.
- 229 N. S. Tehrani, M. Masoumi, F. Chekin and M. S. Baei, *Russ. J. Appl. Chem.*, 2020, **93**, 1221–1228.
- 230 M. Krishani, W. Y. Shin, H. Suhaimi and N. S. Sambudi, *Gels*, 2023, **9**, 100.
- 231 M. I. Echeverria Molina, K. G. Malollari and K. Komvopoulos, *Front. Bioeng. Biotechnol.*, 2021, **9**, 617141.
- 232 P. Zarrintaj, F. Seidi, M. Y. Azarfam, M. K. Yazdi, A. Erfani, M. Barani, N. P. S. Chauhan, N. Rabiee, T. Kuang and J. Kucinska-Lipka, *Composites, Part B*, 2023, **258**, 110701.
- 233 M. Z. A. Mahmud, M. D. Islam and M. H. Mobarak, *J. Nanomater.*, 2023, **2023**, 9270064.
- 234 S. D. Purohit, R. Bhaskar, H. Singh, I. Yadav, M. K. Gupta and N. C. Mishra, *Int. J. Biol. Macromol.*, 2019, **133**, 592–602.
- 235 S. D. Purohit, H. Singh, R. Bhaskar, I. Yadav, S. Bhushan, M. K. Gupta, A. Kumar and N. C. Mishra, *Front. Mater.*, 2020, **7**, 250.
- 236 W. T. Godbey and A. Atala, *Ann. N. Y. Acad. Sci.*, 2002, **961**, 10–26.
- 237 N. A. Kurniawan, *Curr. Opin. Organ Transplant.*, 2019, **24**, 590–597.



- 238 S. Caddeo, M. Boffito and S. Sartori, *Front. Bioeng. Biotechnol.*, 2017, **5**, 40.
- 239 C. I. Chamorro, S. Zeiai, N. Juul, O. Willacy, J. Huo, J. Hilborn and M. Fossum, *Int. J. Mol. Sci.*, 2022, **23**, 12703.
- 240 W. R. Dong, Y. Q. Xiao, Y. J. Piao and Y. H. Chen, *Acad. J. First Med. Coll. PLA*, 2004, **24**, 969–974.
- 241 F. J. O'Brien, *Mater. Today*, 2011, **14**, 88–95.
- 242 Y. Hou, W. Wang and P. Bártolo, *Int. J. Bioprint.*, 2020, **6**, 266.
- 243 C. Shuai, B. Peng, P. Feng, L. Yu, R. Lai and A. Min, *J. Adv. Res.*, 2022, **35**, 13–24.
- 244 N. Mahmoudi and A. Simchi, *Mater. Sci. Eng., C*, 2017, **70**, 121–131.
- 245 M. S. Ramasamy, R. Bhaskar, K. B. Narayanan, S. D. Purohit, S. S. Park, A. Manikkavel, B. Kim and S. S. Han, *Mater. Today Commun.*, 2022, **33**, 104659.
- 246 F. Puza, S. Rostami, B. Özçolak-Aslan, S. Odabaş, K. D. Jandt and B. Garipcan, *Adv. Eng. Mater.*, 2023, **25**, 2200777.
- 247 S. Kumar, S. Raj, S. Jain and K. Chatterjee, *Mater. Des.*, 2016, **108**, 319–332.
- 248 C. D. Grande Tovar, J. I. Castro, C. H. Valencia, P. A. Zapata, M. A. Solano, E. Florez López, M. N. Chaur, M. E. Valencia Zapata and J. H. Mina Hernandez, *Molecules*, 2020, **25**, 2308.
- 249 C. H. Valencia-Llano, M. A. Solano and C. D. Grande-Tovar, *Polymers*, 2021, **13**, 3877.
- 250 J. T. Rashkow, Y. Talukdar, G. Lalwani and B. Sitharaman, *ACS Biomater. Sci. Eng.*, 2017, **3**, 2533–2541.
- 251 G. L. Breitenbach, B. S. Caldas, M. C. G. Pellá, E. C. Muniz and D. C. Dragunski, *Colloids Surf., A*, 2024, **701**, 134872.
- 252 C. Fu, H. Bai, Q. Hu, T. Gao and Y. Bai, *RSC Adv.*, 2017, **7**, 8886–8897.
- 253 S. Nagarajan, C. Pochat-Bohatier, C. Teyssier, S. Balme, P. Miele, N. Kalkura, V. Cavallès and M. Bechelany, *RSC Adv.*, 2016, **6**, 109150–109156.
- 254 G. Khamidov, Ö. Hazman, M. N. Hasanovich and I. Erol, *J. Drug Delivery Sci. Technol.*, 2024, **91**, 105258.
- 255 M. U. A. Khan, S. I. A. Razak, M. N. M. Ansari, R. M. Zulkifli, N. Ahmad Zawawi and M. Arshad, *Polymers*, 2021, **13**, 3611.
- 256 G. Lalwani, A. M. Henslee, B. Farshid, L. Lin, F. K. Kasper, Y.-X. Qin, A. G. Mikos and B. Sitharaman, *Biomacromolecules*, 2013, **14**, 900–909.
- 257 B. Farshid, G. Lalwani and B. Sitharaman, *J. Biomed. Mater. Res., Part A*, 2015, **103**, 2309–2321.
- 258 J. Yang, J. Zhang, M. Yusoff, N. A. Roslan and M. H. Razali, *Dig. J. Nanomater. Biostruct.*, 2025, **20**, 301–314.

