


Cite this: *RSC Adv.*, 2025, 15, 36300

# Influence of C3'- and C4'-substitutions on fluorescence, crystal packing, and physicochemical properties of flavonol

Oleksii O. Demidov,<sup>a</sup> Ludmila V. Chepeleva,<sup>a</sup> Svitlana V. Shishkina,<sup>bc</sup> Eugene S. Gladkov,<sup>ab</sup> Alexander Kyrychenko,<sup>\*ab</sup> Rostyslav P. Linnik<sup>d</sup> and Alexander D. Roshal<sup>a</sup>

Natural flavonols exhibit a wide range of pharmacological activities and possess unique dual ESIPT (Excited-State Intramolecular Proton Transfer) fluorescence, making them sensitive to microenvironments. This sensitivity allows for the detection of metal ions, anions, small ligands, and biomacromolecules. However, the diversity in their structure, including the number and position of hydroxyl groups and potential chemical modifications, complicates the relationship between structure and fluorescence, posing challenges for their practical use as fluorescent probes. In this study, we focus on fine-tuning the ESIPT fluorescence, crystal packing, physicochemical properties, and ADMET (Absorption, Distribution, Metabolism, Excretion, and Toxicity) characteristics of a series of flavonols. We achieve this by introducing hydroxy, methoxy and benzyl groups at the C3' and C4' positions of the 2-phenyl side ring. The photophysical properties of the synthesized flavonols were systematically examined by UV-vis and fluorescence measurements in terms of their structure–property relationship. Our findings indicate that the nature and position of the substituent groups in flavonols can significantly influence their crystal packing in the solid state, tuning contributions of intra- and intermolecular hydrogen bonding and the ESIPT behavior. Lastly, through fluorescence titration and molecular docking calculations, we explored how the introduction of a bulky benzyl moiety and its alteration between C3' and C4' positions can influence the binding interactions of flavonols with  $\beta$ -glucosidases. We believe our findings shed light on the structure–fluorescence relationship in flavonols and open up new opportunities for the design of innovative flavonol-based probes.

Received 7th August 2025  
Accepted 21st September 2025

DOI: 10.1039/d5ra05790f

rsc.li/rsc-advances

## Introduction

Flavonols are widely distributed compounds found in nature, primarily as secondary metabolites of plants and fungi.<sup>1</sup> These substances possess a variety of biological effects, making them of great interest to pharmaceutical researchers.<sup>2</sup> They are known for their strong antioxidant and cancerostatic properties, and their P-vitamin activity.<sup>1,2</sup> Additionally, flavonols serve as effective models for studying intramolecular excited-state proton transfer (ESIPT) processes.<sup>3–8</sup> The phototautomerization of flavonols leads to the formation of two distinct emission bands in fluorescence spectra. The intensity ratio of these bands—corresponding to the initial excited form

and the phototautomer — depends not only on the structure of the flavonol compounds but also on external factors like the polarity of the surrounding environment and specific interactions with solvent molecules.<sup>3,9–14</sup> Due to their properties, flavonols can serve as sensors in biochemical and medicobiological studies.<sup>9,15,16</sup> The 7-hydroxy group in flavones can undergo photodissociation, resulting in fluorescent anions when the corresponding flavones are in an excited state.<sup>17,18</sup> In the case of 7-hydroxyflavonols, photodissociation occurs alongside excited-state intramolecular proton transfer (ESIPT), leading to the formation of more complex protolytic forms known as anion-tautomers.<sup>19</sup> The fluorescence of mentioned flavonol molecules is utilized to study micelles and cell membranes.<sup>20–22</sup> Additionally, flavonols possess one to four complexation centers, enabling them to function as fluorescent indicators for detecting metal ions.<sup>21,23,24</sup> They also may act as ligands that enhance the efficient extraction of metal ions into the organic phase.<sup>25</sup>

All the manifestations of flavonols' reactivity share common characteristics in terms of their spectral and pharmacological effects. These changes in flavonol structures, such as photodissociation, photoisomerization, complex formation, and

<sup>a</sup>Institute of Chemistry, V.N. Karazin Kharkiv National University, 4 Svobody Sq., Kharkiv 61022, Ukraine. E-mail: a.v.kyrychenko@karazin.ua

<sup>b</sup>Institute of Functional Materials Chemistry, State Scientific Institution "Institute for Single Crystals" of National Academy of Sciences of Ukraine, 60 Nauky Ave, 61072 Kharkiv, Ukraine

<sup>c</sup>Institute of Organic Chemistry, NAS of Ukraine, 5 Akademik Kukhar Str., Kyiv 02066, Ukraine

<sup>d</sup>Taras Shevchenko National University of Kyiv, Analytical Chemistry Department, 64/13 Volodymyrska Str., Kyiv 01601, Ukraine



interactions with surrounding molecules, typically involve the chromone fragment of the molecule.<sup>24</sup> The effects of the side phenyl ring and its substituents on the reactivity, spectral, and pharmacological properties have not been thoroughly studied.

The introduction of strong electron-donating and electron-accepting groups, such as alkylamino substituents or nitro groups, into the side ring can result in significant interfragment charge transfer (ICT) between the phenyl and chromone fragments. This transfer leads to the appearance of new long-wavelength absorption bands in the spectra, attributed to the charge-transfer transition, along with changes in the fluorescence characteristics. Natural and many synthetic flavonols typically contain less effective electron-donating substituents, such as hydroxy or various alkoxy groups.<sup>8</sup> Their impact on the spectral properties of flavonols, as previously noted, has not been thoroughly studied. Additionally, our recent studies indicate that flavonols with lipophilic alkoxy substituents on the side phenyl ring can effectively bind to hydrolase enzymes, such as glucosidases, and inhibit their catalytic activity.<sup>21,26</sup>

Physicochemical and pharmacological properties of flavonols have complex interplay on the position and number of hydroxyl groups, as well as their chemical modifications.<sup>2</sup> Therefore, in this study, we focus on some flavonols with various combinations of substituents in the C3' and C4' positions of the side benzene ring. The introduction of methoxy and benzyl groups and altering their positions aimed to establish the relationship between the structure and fluorescent properties of flavonols. Our study demonstrates how the nature and position of substituents in the side ring affect the structure of flavonol molecules in both gas and crystalline phases. It also discusses how these factors influence the position of bands in absorption spectra, as well as the position and intensity ratio of emission bands for both the original and photo-tautomeric forms in fluorescence spectra. Finally, we consider how peripheral phenyl ring substitutions can tune the binding interactions with proteins.

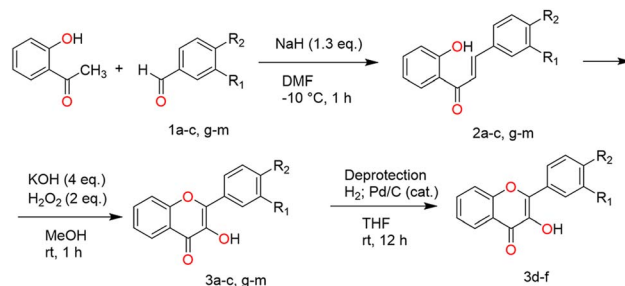
## Experimental section

### Chemistry

To study the effect of the C3' and C4' substitutions in the flavonol's side benzene ring on fluorescence, crystal packing,

Flavonol	3'-R <sub>1</sub>	4'-R <sub>2</sub>
<b>3</b>	H	H
<b>3a</b>	H	OH
<b>3b</b>	H	OCH <sub>3</sub>
<b>3c</b>	H	OBn
<b>3d</b>	OH	H
<b>3e</b>	OH	OH
<b>3f</b>	OH	OCH <sub>3</sub>
<b>3g</b>	OCH <sub>3</sub>	H
<b>3h</b>	OCH <sub>3</sub>	OH
<b>3i</b>	OCH <sub>3</sub>	OCH <sub>3</sub>
<b>3j</b>	OCH <sub>3</sub>	OBn
<b>3k</b>	OBn	H
<b>3l</b>	OBn	OCH <sub>3</sub>
<b>3m</b>	OBn	OBn

Scheme 1 Structure of model flavonol **3** and abbreviations for its derivatives **3a–m**.



Scheme 2 Synthesis of flavonols (3-hydroxy-4H-chromen-4-ones) **3a–m**.

physicochemical and ADMET properties the following flavonols were considered (Scheme 1).

Structure of unsubstituted flavonol **3** and synthesized flavonol derivatives **3a–m** are summarized in Scheme 1. Unsaturated ketones (**2a–c**, **g–m**), used as precursors, were obtained by a modified known methods (Scheme 2).<sup>27,28</sup> Preliminary spectral analysis was provided by Enamine Ltd (Ukraine). All solvents and reagents were commercial grade and, if required, purified in accordance with the standard procedures.

### General procedure of synthesis of chalcones (**2a–c**, **g–m**)

To a suspension of NaH (5.2 mmol, 60% dispersion in mineral oil) in DMF (20 mL), under a nitrogen atmosphere at 0 °C added drop wisely 2'-hydroxyacetophenone (4 mmol). The resulting mixture stirred at 0 °C for 10 min. A solution of corresponding benzaldehyde (4 mmol) in DMF (10 mL) was added drop wisely in the mixture and stirred at room temperature for 2 hours. Therefore, the mixture was acidified drop wisely with glacial acetic acid until pH 4. After filtration the precipitate washed with MeOH (10 mL). The resulting solid was concentrated under vacuum to obtain a crud product with uses in next stage without purification.

### General procedure of synthesis of flavonols (**3a–c**, **g–m**)

To a suspension of chalcone **2a–c**, **g–m** (6 mmol) in MeOH (30 mL) was added KOH (15 mmol), the reaction mixture stirred at –15 °C for 10 min. Then 30% H<sub>2</sub>O<sub>2</sub> (18 mmol) was drop wisely added in the mixture and stirred at rt for 2.5 hours. Therefore, the mixture was acidified drop wisely with glacial acetic acid until pH 2. After filtration the precipitate washed with MeOH (10 mL). The resulting solid was concentrated under vacuum to obtain a pure product. The synthesis scheme is summarized in Scheme 2 and Table 1.

### General procedure of synthesis of flavonols (**3d–f**)

The flavonols **3k–m** (3 mmol) was dissolved in THF (40 mL) and 300 mg of Pd/C 10% was added. The mixture was stirred for 12 h at room temperature under hydrogen atmosphere. The Pd/C was filtered off. After the solvent was evaporated, the precipitate washed with MeOH (10 mL) and filtered. The resulting solid was concentrated under vacuum to obtain a pure product.

**Table 1** Synthesized flavonols (3-hydroxy-4H-chromen-4-ones) **3a–m**

Flavonol	3' (R <sub>1</sub> )	4' (R <sub>2</sub> )	Starting material	Solvent	Yield, %
<b>3a</b>	H	OH	<b>2a</b>	MeOH	80
<b>3b</b>	H	OCH <sub>3</sub>	<b>2b</b> <sup>28</sup>	MeOH	78
<b>3c</b>	H	OBn	<b>2c</b>	MeOH	73
<b>3d</b>	OH	H	<b>3k</b>	THF	85
<b>3e</b>	OH	OH	<b>3l</b>	THF	76
<b>3f</b>	OH	OCH <sub>3</sub>	<b>3m</b>	THF	90
<b>3g</b>	OCH <sub>3</sub>	H	<b>2g</b> <sup>28</sup>	MeOH	69
<b>3h</b>	OCH <sub>3</sub>	OH	<b>2h</b>	MeOH	90
<b>3i</b>	OCH <sub>3</sub>	OCH <sub>3</sub>	<b>2i</b>	MeOH	73
<b>3j</b>	OCH <sub>3</sub>	OBn	<b>2j</b>	MeOH	67
<b>3k</b>	OBn	H	<b>2k</b> <sup>27</sup>	MeOH	85
<b>3l</b>	OBn	OCH <sub>3</sub>	<b>2l</b> <sup>27</sup>	MeOH	68
<b>3m</b>	OBn	OBn	<b>2m</b> <sup>27</sup>	MeOH	74

**4'-Hydroxyflavonol (3-hydroxy-2-(4-hydroxyphenyl)-4H-chromen-4-one) (3a).**<sup>29</sup> <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 10.08 (s, 1H), 9.33 (s, 1H), 8.10 (t, *J* = 7.7 Hz, 3H), 7.79–7.72 (m, 2H), 7.44 (s, 1H), 6.93 (d, *J* = 8.3 Hz, 2H). <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 172.5, 159.1, 154.4, 146.1, 137.8, 133.3, 129.5, 124.7, 124.4, 122.0, 121.3, 118.2, 115.4 (SI Fig. S1). Mass spectrum, *m/z* (*I*<sub>rel</sub>, %): 253.0 [*M* – H]–(100) (SI Fig. S14).

**4'-Methoxyflavonol (3-hydroxy-2-(4-methoxyphenyl)-4H-chromen-4-one) (3b).**<sup>29</sup> <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 9.46 (s, 1H), 8.20 (d, *J* = 8.6 Hz, 2H), 8.10 (d, *J* = 8.0 Hz, 1H), 7.82–7.72 (m, 2H), 7.45 (t, *J* = 7.4 Hz, 1H), 7.13 (d, *J* = 8.6 Hz, 2H), 3.84 (s, 3H). <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 172.6, 160.4, 154.4, 145.5, 138.1, 133.4, 129.4, 124.7, 124.4, 123.5, 121.3, 118.3, 114.0, 55.3 (Fig. S2). Mass spectrum, *m/z* (*I*<sub>rel</sub>, %): 157.0 (100), 269.0 [*M* + 1]<sup>+</sup> (55) (Fig. S15).

**4'-Benzyloxyflavonol (2-(4-(benzyloxy)phenyl)-3-hydroxy-4H-chromen-4-one) (3c).**<sup>26</sup> <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 9.31 (s, 1H), 8.18 (d, 2H), 8.09 (d, *J* = 8.2 Hz, 1H), 7.79–7.68 (m, 2H), 7.49–7.29 (m, 6H), 7.18 (d, *J* = 8.0 Hz, 2H), 5.18 (d, *J* = 2.5 Hz, 2H). <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 172.6, 159.5, 154.4, 145.5, 138.2, 136.6, 133.4, 129.3, 128.4, 127.9, 127.7, 124.7, 124.4, 123.8, 121.3, 118.3, 114.8, 69.4 (Fig. S3). Mass spectrum, *m/z* (*I*<sub>rel</sub>, %): 345.2 [*M* + 1]<sup>+</sup> (92), 367.2(8) (Fig. S16).

**3'-Hydroxyflavonol (3-hydroxy-2-(3-hydroxy-4-methoxyphenyl)-4H-chromen-4-one) (3d).**<sup>29</sup> <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 9.29–9.21 (m, 2H), 8.11 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.82–7.67 (m, 4H), 7.45 (t, *J* = 7.4 Hz, 1H), 7.10 (d, *J* = 8.6 Hz, 1H), 3.86 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 172.5, 154.4, 149.3, 146.2, 145.6, 138.2, 133.4, 124.7, 124.4, 123.7, 121.3, 119.7, 118.2, 114.8, 111.8, 55.6 (Fig. S4). Mass spectrum, *m/z* (*I*<sub>rel</sub>, %): 157.0 (100), 239.2 (23), 255.0 [*M* + 1]<sup>+</sup> (55) (Fig. S17).

**3',4'-Dihydroxyflavonol (2-(3,4-dihydroxyphenyl)-3-hydroxy-4H-chromen-4-one) (3e).**<sup>30</sup> <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 9.39 (s, 3H), 8.09 (dd, *J* = 7.9, 1.7 Hz, 1H), 7.80–7.66 (m, 3H), 7.61 (dd, *J* = 8.4, 2.2 Hz, 1H), 7.43 (t, *J* = 7.4 Hz, 1H), 6.91 (d, *J* = 8.4 Hz, 1H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 172.9, 154.8, 148.1, 146.6, 145.6, 138.4, 133.8, 125.2, 124.8,

122.8, 121.8, 120.5, 118.6, 116.1, 115.8 (Fig. S5). Mass spectrum, *m/z* (*I*<sub>rel</sub>, %): 271.0 [*M* + 1]<sup>+</sup> (88), 272.0 (8) (Fig. S18).

**3'-Hydroxy-4'-methoxyflavonol (3-hydroxy-2-(3-hydroxy-4-methoxyphenyl)-4H-chromen-4-one) (3f).**<sup>31</sup> <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 9.29–9.21 (m, 2H), 8.11 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.82–7.67 (m, 4H), 7.45 (t, *J* = 7.4 Hz, 1H), 7.10 (d, *J* = 8.6 Hz, 1H), 3.86 (s, 3H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 172.5, 154.4, 149.3, 146.2, 145.6, 138.2, 133.4, 124.6, 124.4, 123.7, 121.3, 119.7, 118.2, 114.8, 111.8, 55.6 (Fig. S6). Mass spectrum, *m/z* (*I*<sub>rel</sub>, %): 285.2 [*M* + 1]<sup>+</sup> (94), 286.2 (6) (Fig. S19).

**3'-Methoxyflavonol (3-hydroxy-2-(3-methoxyphenyl)-4H-chromen-4-one) (3g).**<sup>29</sup> <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 9.66 (s, 1H), 8.12 (d, *J* = 7.6 Hz, 1H), 7.86–7.75 (m, 5H), 7.53–7.43 (m, 3H), 7.10 (dd, *J* = 8.2, 2.6 Hz, 1H), 3.83 (s, 4H). <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 173.5, 159.7, 155.0, 145.3, 139.7, 134.3, 133.0, 130.2, 125.3, 125.1, 121.7, 120.5, 119.0, 115.7, 113.9, 55.7 (Fig. S7). Mass spectrum, *m/z* (*I*<sub>rel</sub>, %): 269.0 [*M* + 1]<sup>+</sup> (93), 270.0 (7) (Fig. S20).

**4'-Hydroxy-3'-methoxyflavonol (3-hydroxy-2-(4-hydroxy-3-methoxyphenyl)-4H-chromen-4-one) (3h).**<sup>32</sup> <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 9.55 (s, 2H), 8.09 (d, *J* = 7.9 Hz, 1H), 7.83 (s, 1H), 7.80–7.73 (m, 3H), 7.50–7.40 (m, 1H), 6.96 (dd, *J* = 8.5, 3.7 Hz, 1H), 3.86 (s, 3H). <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 172.5, 154.3, 148.7, 147.4, 145.9, 138.0, 133.3, 124.6, 124.4, 122.2, 121.8, 121.2, 118.3, 115.5, 111.7, 55.7 (Fig. S8). Mass spectrum, *m/z* (*I*<sub>rel</sub>, %): 285.0 [*M* + 1]<sup>+</sup> (96), 285.0(3) (Fig. S21).

**3',4'-Dimethoxyflavonol (2-(3,4-dimethoxyphenyl)-3-hydroxy-4H-chromen-4-one) (3i).**<sup>33</sup> <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 9.47 (s, 1H), 8.09 (d, *J* = 7.9 Hz, 1H), 7.86 (d, *J* = 8.6 Hz, 1H), 7.83–7.75 (m, 3H), 7.48–7.42 (m, 1H), 7.14 (d, *J* = 8.6 Hz, 1H), 3.84 (s, 6H). <sup>13</sup>C NMR (151 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 173.1, 154.9, 150.8, 148.9, 145.9, 138.7, 133.9, 125.2, 124.9, 124.1, 122.0, 121.7, 118.9, 112.0, 111.5, 56.1, 56.1 (Fig. S9). Mass spectrum, *m/z* (*I*<sub>rel</sub>, %): 299.2 [*M* + 1]<sup>+</sup> (91), 300.2 (15) (Fig. S22).

**3'-Methoxy-4'-benzyloxyflavonol (2-(4-(benzyloxy)-3-methoxyphenyl)-3-hydroxy-4H-chromen-4-one) (3j).**<sup>34</sup> <sup>1</sup>H NMR (500 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 9.48 (s, 1H), 8.09 (d, *J* = 7.3 Hz, 1H), 7.86–7.72 (m, 4H), 7.50–7.30 (m, 6H), 7.22 (d, *J* = 8.8 Hz, 1H), 5.17 (s, 2H), 3.85 (s, 3H). <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 172.6, 154.4, 149.3, 148.7, 145.3, 138.3, 136.7, 133.4, 128.4, 127.9, 127.9, 124.7, 124.4, 123.9, 121.3, 121.2, 118.3, 113.0, 111.3, 69.8, 55.7 (Fig. S10). Mass spectrum, *m/z* (*I*<sub>rel</sub>, %): 375.2 [*M* + 1]<sup>+</sup> (97), 397.2 (3) (Fig. S23).

**3'-Benzyloxyflavonol (2-(3-(benzyloxy)phenyl)-3-hydroxy-4H-chromen-4-one) (3k).** <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 9.69 (s, 1H), 8.11 (d, *J* = 8.0 Hz, 1H), 7.89–7.71 (m, 4H), 7.53–7.29 (m, 7H), 7.15 (dd, *J* = 8.4, 2.7 Hz, 1H), 5.17 (s, 2H). <sup>13</sup>C NMR (126 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 173.0, 158.2, 154.5, 144.7, 139.2, 136.8, 133.7, 132.5, 129.6, 128.4, 127.9, 127.8, 124.7, 124.5, 121.2, 120.2, 118.4, 115.9, 114.3, 69.4 (Fig. S11). White solid with a yellowish tint, mp 137–138 °C. Mass spectrum, *m/z* (*I*<sub>rel</sub>, %): [*M* + H]<sup>+</sup> = 345.0 [*M* + 1]<sup>+</sup> (97), 346 (20) (Fig. S24).

**3'-Benzyloxy-4'-methoxyflavonol (2-(3-(benzyloxy)-4-methoxyphenyl)-3-hydroxy-4H-chromen-4-one) (3l).**<sup>31</sup> <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>),  $\delta$ , ppm: 9.50 (s, 1H), 8.10 (dd, *J* = 8.0, 1.6 Hz, 1H), 7.94–7.87 (m, 2H), 7.84–7.72 (m, 2H), 7.55–7.30 (m, 6H), 7.21–7.14 (m, 1H), 5.18 (s, 2H), 3.86 (s, 3H). <sup>13</sup>C NMR (126 MHz,



DMSO- $d_6$ ),  $\delta$ , ppm: 172.6, 154.3, 150.6, 147.4, 145.3, 138.3, 136.9, 133.4, 128.4, 128.1, 127.9, 124.7, 124.5, 123.5, 121.9, 121.2, 118.3, 112.7, 111.7, 70.2, 55.6 (Fig. S12). Mass spectrum,  $m/z$  ( $I_{\text{rel}}$ , %): 375.0  $[M + 1]^+$  (88), 376.0 (12) (Fig. S25).

**3',4'-Dibenzoylflavonol (2-(3,4-bis(benzoyloxy)phenyl)-3-hydroxy-4H-chromen-4-one) (3m).**<sup>26</sup>  $^1\text{H}$  NMR (500 MHz, DMSO- $d_6$ ),  $\delta$ , ppm: 9.52 (s, 1H), 8.09 (d,  $J = 8.0$  Hz, 1H), 7.94 (s, 1H), 7.87 (d,  $J = 8.6$  Hz, 1H), 7.83–7.70 (m, 2H), 7.54–7.28 (m, 11H), 7.25 (d,  $J = 8.7$  Hz, 1H), 5.22 (d,  $J = 7.3$  Hz, 4H).  $^{13}\text{C}$  NMR (126 MHz, DMSO- $d_6$ ),  $\delta$ , ppm: 172.6, 154.3, 149.8, 147.7, 145.2, 138.4, 137.0, 136.8, 133.4, 128.4, 127.9, 127.8, 127.6, 124.7, 124.4, 123.9, 121.9, 121.2, 118.3, 113.7, 70.4, 69.9 (Fig. S13). Mass spectrum,  $m/z$  ( $I_{\text{rel}}$ , %): 451.0  $[M + 1]^+$  (74), 452.0 (16) (Fig. S26).

### Quantum-chemical calculations

Unconstrained and constrained geometry optimizations of isolated molecules of flavonols in the ground electronic state and analysis of rotation barriers were carried out at the DFT levels of theory using the B3LYP functional<sup>35</sup> and cc-pVDZ basis set<sup>36</sup> implemented in the GAUSSIAN 16 program package.<sup>37</sup> The solvent (dichloromethane and acetonitrile) effect at 298.14 K and standard pressure was estimated at the level of the Polarized Continuum Model (PCM).<sup>38</sup>

### Spectroscopic measurements

**NMR spectra.**  $^1\text{H}$  NMR spectra were recorded using Unity Inova 400, Bruker Avance DRX 500, and Bruker Avance III 400 MHz spectrometers in DMSO- $d_6$ .  $^{13}\text{C}$  NMR spectra were recorded on Bruker Avance DRX 500 and Agilent ProPulse 500 MHz spectrometers at a resonance frequency of 126 MHz in DMSO- $d_6$ . Chemical shifts are reported in the  $\delta$  scale (ppm). Mass spectra were recorded on an Agilent 1100 high-performance liquid chromatograph (HPLC) equipped with a diode matrix and an Agilent LC/MSD SL mass-selective detector, a SUPELCO Ascentis Express C18 chromatographic column 2.7  $\mu\text{m}$  4.6 mm  $\times$  15 cm. Control over the course of the reaction and the individuality of the obtained substances was carried out by TLC method on silica gel-coated "Polychrom SI F254" plates with a fluorescent detector in the hexane–ethyl acetate 2 : 1 system, the developer was an ultraviolet lamp. If necessary, additional purification of the obtained compounds was carried out using flash chromatography (UPLC) on a PuriFlash XS520 Plus device using gradient elution. The melting points of all synthesized compounds were determined using a Hanon Instruments MP450 open capillary tube automatic melting point apparatus.

**UV-vis and fluorescence spectra.** Absorption and fluorescence spectra of flavonols in liquid media were registered in dichloromethane and acetonitrile solutions. Commercial solvents were additionally rectified and dried before use. The flavonols' concentrations in investigated solutions ranged from  $1.0 \times 10^{-5}$  to  $5.0 \times 10^{-5} \text{ M l}^{-1}$ . Absorption and fluorescence spectra were recorded on a PerkinElmer LS55 spectrofluorimeter and a Shimadzu UV-2401PC spectrophotometer. The latter was also used to record the excitation and fluorescence spectra of flavonols in the solid state.

**Fluorescence titrations.** The interaction between flavonols and protein molecules was studied using  $\beta$ -glucosidase ( $\beta$ -D-glucoside glucohydrolase) from Almonds purchased from Sigma-Aldrich as a lyophilized powder with > 98% purity. Stock solutions of flavonols were prepared by dissolving their samples in DMSO. A stock solution of  $\beta$ -glucosidase was prepared by dissolving a sample of the enzyme in an aqueous phosphate buffer with pH 6.86. The fluorescence was excited at 380 nm and the emission spectra were recorded in the range of 400–700 nm using a Hitachi 850 spectrofluorimeter. The experimental details are described elsewhere.<sup>39</sup>

**X-ray diffraction study.** X-Ray diffraction studies were performed on an automatic "Bruker APEX II" diffractometer (graphite monochromated MoK $\alpha$  radiation, CCD-detector,  $\phi$ - and  $\omega$ -scanning). The structures were solved by direct method using OLEX2 ref. 40 package with SHELXT<sup>41</sup> and SHELXL modules.<sup>42</sup> The phenyl ring of the benzyl group in structure 3k is disordered over two positions (A and B) due to rotation around the C<sub>sp<sup>3</sup></sub>–C<sub>ar</sub> bond. The restrictions on the bond lengths in the disordered fragment were applied (C<sub>ar</sub>–C<sub>ar</sub> = 1.39 Å). The hydrogen atoms were placed in their ideal positions and refined by "riding" model with  $U_{\text{iso}} = nU_{\text{eq}}$  of the carrier atom ( $n = 1.5$  for methyl groups and  $n = 1.2$  for other hydrogen atoms). The crystallographic data and experimental parameters are listed in Table S1 in Supplementary section. Deposition Numbers CCDC 2416411 for structure 3b and CCDC 2416347 for structure 3k contain the supplementary crystallographic data for this paper. These data can be obtained free of charge by the joint Cambridge Crystallographic Data Centre and Fachinformationszentrum Karlsruhe Access Structures service <http://www.ccdc.cam.ac.uk/structures>.

### Molecular docking setup

The molecular docking setting up was carried out with the AutoDock Tools (ADT) software, version 1.5.7.<sup>43</sup> The addition of hydrogen, the calculation of the Gasteiger charges of the receptor, and ligands were also performed using the ADT software. Molecular docking calculations were performed with the AutoDock Vina 1.1.2 software.<sup>43,44</sup> The 3D X-ray structure of *Paenibacillus polymyxa*  $\beta$ -glucosidase B (BglB, PDB 2O9R),<sup>45</sup> *Raucaffricine*  $\beta$ -glucosidase (rBG, PDB 4A3Y),<sup>46</sup> *Thermotoga maritima*  $\beta$ -glucosidase (TmGH1, PDB 1OD0),<sup>47</sup> and human cytosolic  $\beta$ -glucosidase (hCBG, PDB 2JFE)<sup>48</sup> were downloaded from the RCSB Protein Data Bank. The calculations details were described in details elsewhere.<sup>26,39</sup>

## Results and discussion

### Absorption and fluorescence spectra in solutions

Research into the electronic transitions of flavonol molecules, conducted using both semi-empirical methods and density functional theory (DFT), has revealed that broad absorption bands in the 340–380 nm and 240–270 nm ranges result from intense electronic transitions occurring primarily between molecular orbitals located on the chromone fragment of these molecules.<sup>49</sup> In this case, the side benzene ring can affect the





Table 2 Calculated absorption and fluorescence parameters of flavonols 3a–f in water<sup>a</sup>

Transitions	I ( $S_0 \rightarrow S_1$ )					II ( $S_0 \rightarrow S_2$ )				
Flavonol	$\lambda_{\text{abs}}$ , nm ( $\nu_{\text{abs}}$ , $\text{cm}^{-1}$ )	$f$	C.I.	Type	ICT	$\lambda_{\text{abs}}$ , nm ( $\nu_{\text{abs}}$ , $\text{cm}^{-1}$ )	$f$	C.I.	Type	
H	349 (28 625)	0.354	0.69 $\chi_{1 \rightarrow 1'}$	CHR	0.102	308 (32 520)	0.000	0.70 $\chi_{4 \rightarrow 1'}$	$n \rightarrow \pi^*$	
3'-OH	356 (28 110)	0.308	0.68 $\chi_{1 \rightarrow 1'}$	CHR	0.133	332 (30 120)	0.041	0.68 $\chi_{2 \rightarrow 1'}$	CT	
4'-OH	361 (27 695)	0.435	0.70 $\chi_{1 \rightarrow 1'}$	CHR	0.152	306 (32 730)	0.000	0.70 $\chi_{4 \rightarrow 1'}$	$n \rightarrow \pi^*$	
3',4'-OH	372 (26 855)	0.421	0.70 $\chi_{1 \rightarrow 1'}$	CHR	0.232	315 (31 750)	0.068	0.70 $\chi_{2 \rightarrow 1'}$	FL	
Transitions	III ( $S_0 \rightarrow S_3$ )					IV ( $S_0 \rightarrow S_4$ )				
	$\lambda_{\text{abs}}$ , nm ( $\nu_{\text{abs}}$ , $\text{cm}^{-1}$ )	$f$	C.I.	Type		$\lambda_{\text{abs}}$ , nm ( $\nu_{\text{abs}}$ , $\text{cm}^{-1}$ )	$f$	C.I.	Type	
H	292 (34 190)	0.116	0.70 $\chi_{2 \rightarrow 1'}$	FL		284 (35 195)	0.137	0.64 $\chi_{3 \rightarrow 1'}$	CT	
3'-OH	306 (32 680)	0.000	0.70 $\chi_{4 \rightarrow 1'}$	$n \rightarrow \pi^*$		289 (34 600)	0.137	0.64 $\chi_{3 \rightarrow 1'}$	FL	
4'-OH	298 (33 595)	0.089	0.67 $\chi_{2 \rightarrow 1'}$	FL		282 (35 385)	0.005	0.41 $\chi_{1 \rightarrow 2'} + 0.41\chi_{1 \rightarrow 3'}$	FL	
3',4'-OH	305 (32 840)	0.000	0.70 $\chi_{4 \rightarrow 1'}$	$n \rightarrow \pi^*$		290 (34 455)	0.058	0.65 $\chi_{3 \rightarrow 1'}$	CT	

<sup>a</sup>  $\lambda_{\text{abs}}$ ,  $\nu_{\text{abs}}$  – wavelength and wave number of the electronic transition,  $f$  – transition oscillator strength, C.I. – configuration composition of the transition, types of transitions – CHR – local, located predominantly on the chromone fragment, CT – transition with interfragment charge transfer, FL – transition localized on the molecule as a whole,  $n \rightarrow \pi^*$  – forbidden  $n\pi^*$  transition. ICT – value of interfragment charge transfer during the  $S_0 \rightarrow S_1$  transition.

electron density distribution within the chromone fragment, which may result in a shift in the absorption bands. Therefore, according to chromophore theory, the side benzene ring operates as an auxochrome of a complex structure. Electronic transitions involving inter-fragmental charge transfer between the side benzene ring and the chromone part of the molecule typically occur in the short-wavelength region of the spectrum. However, when a strong electron donor, such as  $-\text{N}(\text{CH}_3)_2$ , or a strong electron acceptor, such as  $-\text{NO}_2$ , is present in the side ring, the electronic transition shifts to the long-wavelength region. This transition can occur either from the molecular orbitals localized on the benzene ring to the orbitals of the chromone fragment, or *vice versa*. The presence of a charge transfer transition is manifested by the appearance of a new long-wavelength absorption band in the range of 400–430 nm.<sup>12</sup> Table 2 summarizes the calculated absorption and fluorescence parameters of flavonols 3a–f in water.

The molecular orbitals of unsubstituted flavonol, as well as its monosubstituted derivatives that have hydroxyl groups in the C3' and C4' positions, are illustrated in Fig. 1. Meanwhile, the disubstituted derivative, 3',4'-dihydroxyflavonol, is also included in this analysis. Fig. 2 presents a diagram showing the relative energies of these molecular orbitals. Additionally, Table 2 provides details on the theoretical parameters of long-wavelength transitions observed in the spectra of these compounds. The analysis of configurational interaction reveals that the long-wavelength intense transitions of all flavonols can be characterized by a transition between the highest occupied molecular orbital (MO)  $\phi_1$  and the lowest vacant molecular orbital  $\phi_1'$ . The MO  $\phi_1$  is delocalized between the chromone and the side phenyl fragment. However, as the torsion angle between these fragments increases, the localization of MO  $\phi_1$  increasingly favors the chromone fragment. In contrast, the vacant MO  $\phi_1'$  is primarily localized on the chromone fragment.

Therefore, it can be concluded that the long-wavelength electronic transition generally exhibits a “chromone” nature (CHR).

This conclusion is supported by the relatively low values of interfragment charge transfer during the transition from the ground state to the Frank–Condon excited state  $S_1$ , which are approximately 0.1–0.2 electronic charges ( $e^-$ ) only. These low values suggest that the side phenyl ring does not significantly participate in the excitation process.

The Table 2 illustrates that increasing the number of substituents in the phenyl side ring results in a decrease in  $S_0 \rightarrow S_1$  transition energy, *i.e.* to a bathochromic shift of the corresponding absorption band from 350 nm to 370 nm approximately. Adding a substituent at position C4' enhances the transition oscillator strength  $f$ , which in turn increases the intensity of the associated absorption band. Conversely, introducing a substituent at position C3' has the opposite effect, reducing the intensity of the absorption band.

DFT calculations of the electronic spectrum for the compound featuring an OH group at the C3' position reveal an additional long-wavelength band in the  $\sim 330$  nm range. This band results from an electronic transition between the  $\phi_2$  orbital, which is localized on the side phenyl ring, and the  $\phi_1'$  orbital, which is localized on the chromone fragment. This transition is associated with significant inter-fragment charge transfer (ICT). In the spectra of other compounds, similar transitions are observed in the shorter wavelength region of 285–290 nm (see Table 2). This phenomenon occurs due to an increase in the energy of the corresponding occupied molecular orbital when a substituent is introduced at the C3' position. In Fig. 2, the energies of the molecular orbitals localized on the side phenyl ring are highlighted in red. In the 305–310 nm spectrum range, all compounds have a forbidden  $n\pi^*$  transition from the  $n$ -orbital  $\phi_4$  to the orbital  $\phi_1'$ . The short-wave intense band ranging from 280 to 315 nm in all spectra is



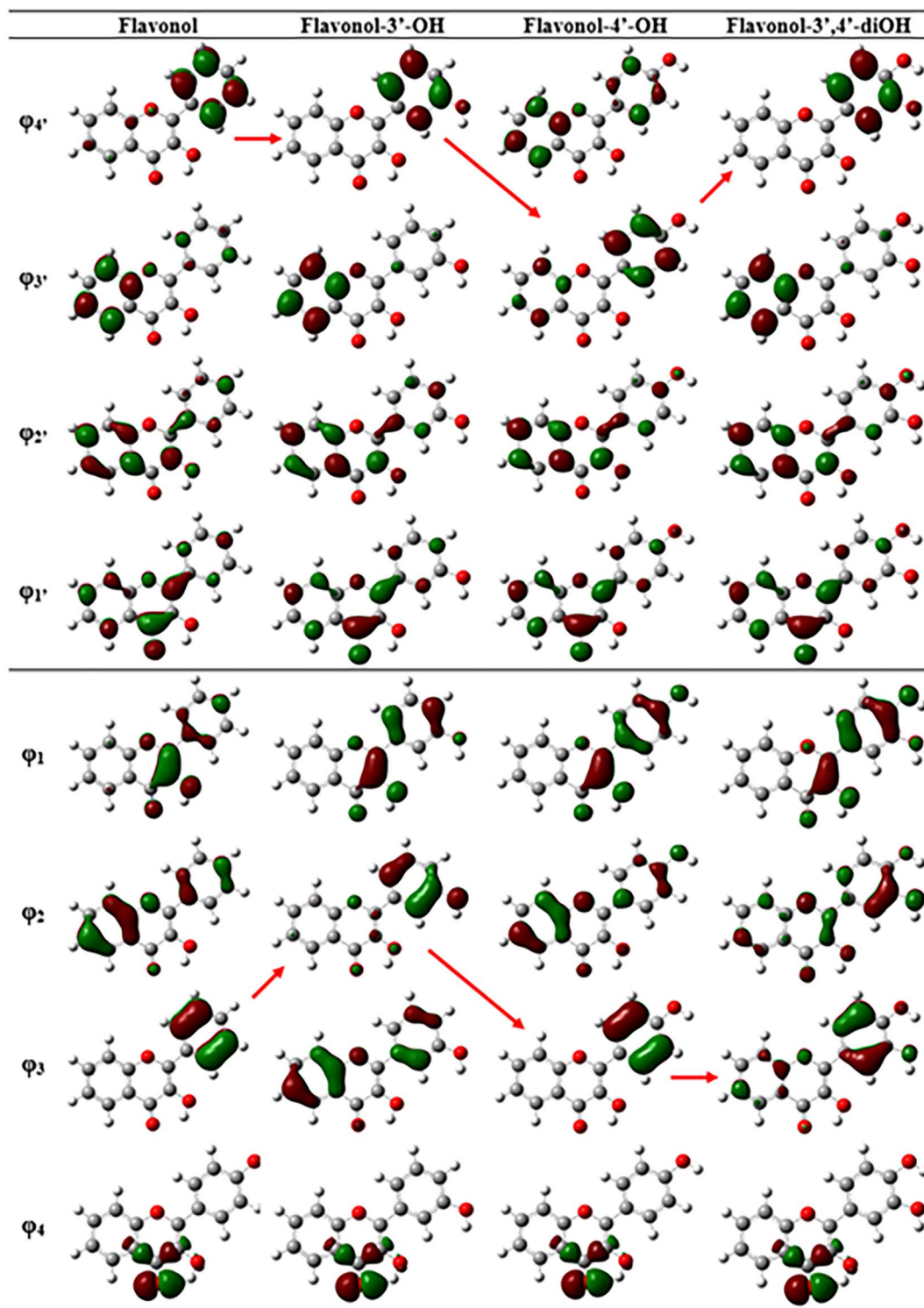


Fig. 1 Frontier molecular orbitals of flavonol, and substituted flavonol with 3'-OH, 4'-OH, and 3',4'-OH.

attributed to the electronic transition  $\phi_2 \rightarrow \phi_1'$  (with  $\phi_3 \rightarrow \phi_1'$  occurring in the compound substituted at the C3' position). As illustrated in Fig. 1, the  $\phi_2$  orbital is delocalized across both fragments of the flavonol molecule. Therefore, the transition from  $\phi_2$  to  $\phi_1'$  is not localized and can be understood as the excitation of the molecule as a whole. It should be noted that

the results from the DFT method correlate well with semi-empirical methods.<sup>49</sup>

The impact of the side phenyl ring on the chromone fragment arises from both the electronic effects of its substituents and the extent of their electronic conjugation governed by the torsion angle between the planes of the both fragments. The

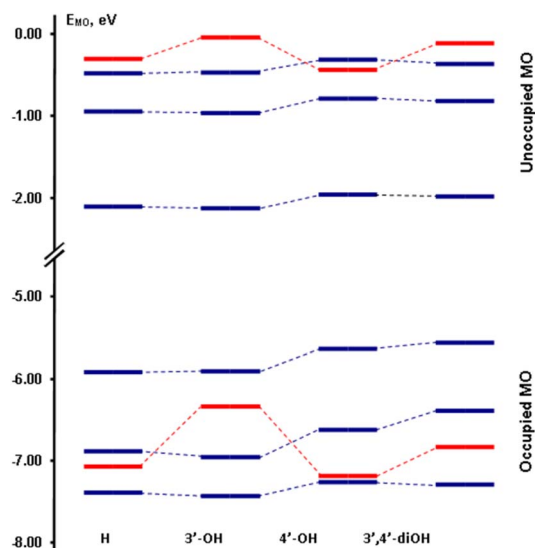


Fig. 2 Influence of substituents' positions on MO energies in flavonols obtained using the b3lyp/cc-pVDZ level of theory (see Table 2 for more details).

conformations of flavonol molecules in the gas phase, as well as in dichloromethane and acetonitrile, were investigated by modeling the potential energy curves associated with the rotation of the side ring relative to the plane of the chromone fragment. This rotation was analyzed over an angle range from  $-30^\circ$  to  $150^\circ$  for 3'-hydroxy-, 4'-hydroxy-, and 3',4'-dihydroxyflavonols. As illustrated in Fig. 3, regardless of the substituent's position, the minimum potential energy occurs at angles between  $-10^\circ$  and  $10^\circ$ . This observation indicates that over 97% of flavonol molecules are planar in both solutions and the gas phase. Furthermore, the calculations reveal that an increase in the polarity of the medium results in a decrease in the rotation barrier by approximately 0.6 to 0.7 kcal mol $^{-1}$ .

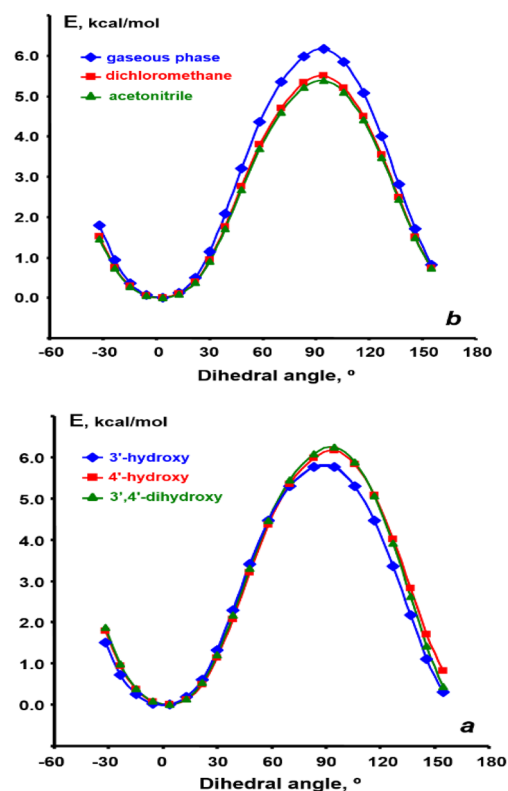


Fig. 3 DFT-calculated rotation barriers between the chromone moiety and phenyl side ring calculated using b3lyp/cc-pVDZ level of theory. (a) – if substituents are in different positions of the benzene ring, (b) – for the same flavonol **3a** in different media.

The influence of electronic effects from substituents is quantified by the Hammett constants  $\sigma_{para}$  and  $\sigma_{meta}$ . However, since multiple substituents can simultaneously occupy the para and meta positions in the side ring, and considering the

Table 3 Spectral characteristics of long-wavelength absorption bands of the flavonols in dichloromethane and acetonitrile<sup>a</sup>

Flavonol	3'-R	4'-R	$\sigma_{para}$	Dichloromethane		Acetonitrile		$\Delta\nu_{abs}$
				$\lambda_{abs}$	$\nu_{abs}$	$\lambda_{abs}$	$\nu_{abs}$	
<b>3<sup>b</sup></b>	H	H	0.000	344	29 070	340	29 400	330
<b>3a</b>	H	OH	-0.047	351	28 490	350	28 570	80
<b>3b</b>	H	OCH <sub>3</sub>	-0.037	352	28 410	350	28 570	160
<b>3c</b>	H	OBn	-0.007	355	28 170	349	28 655	485
<b>3d</b>	OH	H	+0.007	346	28 900	343	29 155	255
<b>3e</b>	OH	OH	-0.046	353	28 330	355	28 170	-160
<b>3f</b>	OH	OCH <sub>3</sub>	-0.040	356	28 090	354	28 250	160
<b>3g</b>	OCH <sub>3</sub>	H	+0.017	345	28 984	342	29 240	256
<b>3h</b>	OCH <sub>3</sub>	OH	-0.041	357	28 010	355	28 170	160
<b>3i</b>	OCH <sub>3</sub>	OCH <sub>3</sub>	-0.037	361	27 700	356	28 090	390
<b>3j</b>	OCH <sub>3</sub>	OBn	-0.003	361	27 700	356	28 090	390
<b>3k</b>	OBn	H	+0.032	346	28 900	343	29 155	255
<b>3l</b>	OBn	OCH <sub>3</sub>	-0.005	360	27 780	356	28 090	310
<b>3m</b>	OBn	OBn	-0.001	361	27 700	355	28 170	470

<sup>a</sup>  $\sigma_{para}$  – Hammett constants for R-oxyphenyl fragments,  $\lambda_{abs}$  – absorption band maxima in nm scale,  $\nu_{abs}$  – absorption band maxima in cm $^{-1}$  scale,  $\Delta\nu_{abs}$  – shifts of absorption band maxima on going from dichloromethane to acetonitrile. <sup>b</sup> Spectral data for unsubstituted flavonol **3** were taken from ref. 50. The data for nonpolar medium are shown for chloroform.

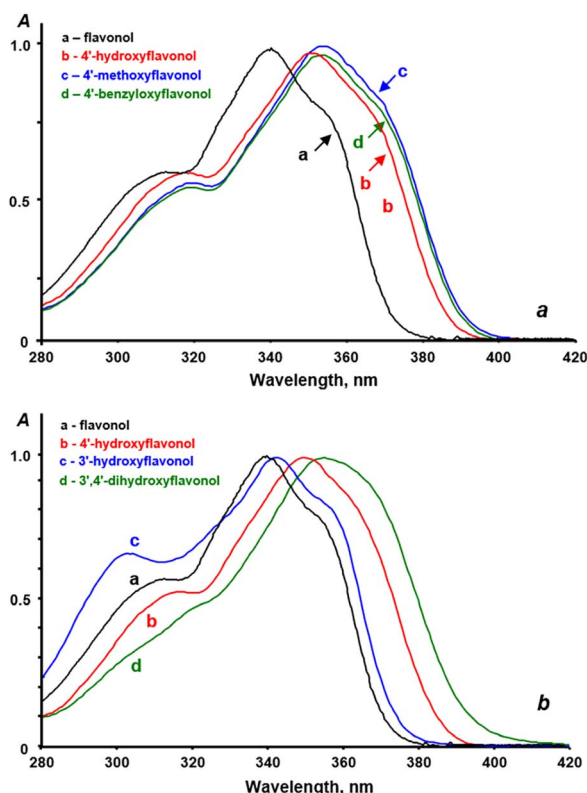


Fig. 4 Normalized absorption spectra of the studied flavonols in dichloromethane. a – different substituents in the same position of the side benzene ring, b – the same substituents in different positions of the side benzene ring. Concentrations of flavonols were in a range of  $1\text{--}5 \cdot 10^{-5}$  M.

dampening effect of the aromatic  $\pi$ -system, we calculated the overall  $\sigma_{\text{para}}$  constants for various RO-phenyl substituents using the web tool outlined in the article.<sup>51</sup> The calculated  $\sigma_{\text{para}}$  values are presented in Table 3. It is important to note that these  $\sigma_{\text{para}}$  values are notably low, indicating a very weak influence of the substituents in the side benzene ring on the electronic structure of the chromone moiety of the molecules.

A comparison of the  $\sigma_{\text{para}}$  constants with the positions of the long-wavelength absorption bands in the experimental spectra of flavonols revealed no correlation between them.

This indicates that substituents do not have a significant impact on the spectral parameters of flavonols. For example, substituting a hydroxy group with either a methoxy group or a benzyloxy group in the same position on the aromatic ring does not cause any shift in the absorption band (see Table 3 and Fig. 4a). Additionally, no shifts in the band position were observed with any combinations of substituents in 3',4'-disubstituted flavonols.

More significant differences in the absorption band parameters are observed depending on the positions of the substituents in the benzene ring (see Fig. 4b). For 3'-R-hydroxyflavonols in dichloromethane, regardless of the type of substituents, the absorption maximum falls within the range of 345–346 nm ( $28\,900\text{--}28\,985\text{ cm}^{-1}$ ). The  $\sigma_{\text{para}}$  constants for these compounds have positive values of +0.01 to +0.03, indicating a very weak

electron-withdrawing effect of the benzene ring on the chromone fragment. In contrast, 4'-R-oxyflavonols and 3',4'-di-R-hydroxyflavonols exhibit long-wavelength band maxima in approximately similar range: at 351–355 nm ( $28\,170\text{--}28\,490\text{ cm}^{-1}$ ) and 353–361 nm ( $27\,700\text{--}28\,330\text{ cm}^{-1}$ ), respectively. The corresponding  $\sigma_{\text{para}}$  constants for these compounds range from  $-0.01$  to  $-0.05$ , suggesting that the benzene ring demonstrates weakly expressed electron-releasing properties. When going to a more polar environment, such as acetonitrile, a slight hypsochromic shift of the absorption bands from 80 to  $500\text{ cm}^{-1}$  is observed (see Table 3). Additionally, the effect of substituent position is even less pronounced in acetonitrile.

The  $\sigma_{\text{para}}$  values indicate that substituents on the side benzene ring have a very weak effect on the spectral properties of flavonols. In contrast, the introduction of 4'-diethylamino- and 4'-nitro substituents into the side ring results in significant changes to the absorption spectra. In these latter cases, the  $\sigma_{\text{para}}$  values are tens of times greater, measuring  $-0.146$  and  $+0.372$ , respectively. Natural flavonols commonly contain hydroxy and various alkoxy groups, which typically have low  $\sigma_{\text{para}}$  values. Therefore, it can be assumed that natural flavonols with a similar structure in the chromone moiety of their molecules will exhibit approximately the same absorption spectra, regardless of the type or location of substituents in the side phenolic ring.

Due to the ESIPT process, the fluorescence spectra of flavonols can exhibit two distinct fluorescence bands. The first is a short-wavelength emission band corresponding to the excited form of the original flavonol ( $N^*$ ), and the second is a long-wavelength emission band from the phototautomer ( $T^*$ ), which is formed by the transfer of a proton from the 3-hydroxy group to the carbonyl group of the chromone fragment. Proton transfer is typically a very rapid and irreversible process, resulting in the fluorescence spectrum usually displaying only

Table 4 Spectral characteristics of long-wavelength emission bands of the flavonols in dichloromethane and acetonitrile<sup>a</sup>

Flavonol	3'-R	4'-R	Dichloromethane			Acetonitrile		
			$\lambda_{\text{fl}}$	$\nu_{\text{fl}}$	$\Delta\nu_{\text{St}}$	$\lambda_{\text{fl}}$	$\nu_{\text{fl}}$	$\Delta\nu_{\text{St}}$
3 <sup>b</sup>	H	H	530	18 865	10 200	526	18 860	10 500
3a	H	OH	528	18 940	9550	547	18 280	10 290
3b	H	OCH <sub>3</sub>	530	18 870	9540	532	18 795	9775
3c	H	OBn	529	18 905	9265	539	18 550	10 105
3d	OH	H	527	18 975	9925	527	18 960	10 195
3e	OH	OH	529	18 905	9425	536	18 655	9515
3f	OH	OCH <sub>3</sub>	534	18 725	9365	538	18 585	9665
3g	OCH <sub>3</sub>	H	524	19 085	9900	528	18 940	10 300
3h	OCH <sub>3</sub>	OH	531	18 830	9180	534	18 730	9440
3i	OCH <sub>3</sub>	OCH <sub>3</sub>	533	18 760	8940	528	18 940	9150
3j	OCH <sub>3</sub>	OBn	534	18 725	8975	542	18 450	9640
3k	OBn	H	525	19 050	9850	536	18 655	10 500
3l	OBn	OCH <sub>3</sub>	534	18 725	9055	540	18 520	9570
3m	OBn	OBn	534	18 725	8975	542	18 450	9720

<sup>a</sup>  $\lambda_{\text{fl}}$ ,  $\nu_{\text{fl}}$  – these designations are given in Table 2,  $\Delta\nu_{\text{St}}$  – Stokes shifts of fluorescence,  $\text{cm}^{-1}$ . <sup>b</sup> Spectral data for unsubstituted flavonol were taken from ref. 50 data for nonpolar medium are shown for chloroform.



one long-wavelength band of the phototautomer ( $T^*$ ). However, as the polarity of the medium increases, the activation barrier for the proton transfer reaction also increases. Consequently, a band corresponding to the  $N^*$  form may appear in the fluorescence spectra. In proton-donating solvents or when proton-donating impurities are present, flavonols can form intermolecular hydrogen bonds. This leads to suppressing the excited-state intramolecular proton transfer (ESIPT) and results in the fluorescence of the  $N^*$  form. Additionally, substituents on the side ring of flavonols can influence the dynamics of phototautomerization. Specifically, introducing an electron-releasing group at the C4' position may decrease the acidity of the 3-hydroxy group in the chromone ring and slow down ESIPT, which can lead to the emergence of two-band fluorescence.

The experimental fluorescence spectra of all the studied flavonols exhibited a single emission band. This observed emission band is attributed to the  $T^*$  phototautomer, as indicated by the high Stokes shift values in the range of 8900–10500  $\text{cm}^{-1}$  (see Table 4). A trace amount of fluorescence from the  $N^*$  form can be detected in the short-wavelength region of the spectra. However, this may be attributed to water impurities that were adsorbed by acetonitrile during the experiment, as well as the formation of trace amounts of flavonol hydrates, which may not undergo proton phototransfer. Considering the low  $\sigma_{\text{para}}$  values and the spectral parameters outlined in Table 3, we can conclude that the fluorescence spectra of natural flavonols in aprotic solvents will be single-band, regardless of the nature and position of the substituents in the side benzene ring.

The effects of the nature and position of these substituents are illustrated in Fig. 5a and b, while the quantitative parameters of the emission bands are provided in Table 3.

The analysis of the data in Table 3 indicates that the nature and position of the substituents in the side benzene ring have a lesser impact on the position of the  $T^*$  emission band compared to the position of the absorption bands. Specifically, the average emission band maxima for 3'-R-oxy-, 4'-R-oxy-, and 3',4'-R-dioxyflavonols are  $525 \pm 2$  nm,  $529 \pm 1$  nm, and  $533 \pm 4$  nm, respectively. The shifts of the bands when changing the position of the substituent range from 135 to 145  $\text{cm}^{-1}$  on the energy scale, which indicates that these shifts are relatively insignificant.

The energy costs involved in rearranging the geometry of molecules and their solvate shell is characterized by the Stokes shifts of fluorescence ( $\Delta\nu_{\text{St}}$ ). For the flavonols studied in dichloromethane, the values of  $\Delta\nu_{\text{St}}$  range from 9850 to 9925  $\text{cm}^{-1}$  for 3'-R-hydroxyflavonols, 9265 to 9550  $\text{cm}^{-1}$  for 4'-R-hydroxyflavonols, and 8940 to 9425  $\text{cm}^{-1}$  for 3',4'-R-dihydroxyflavonols. Notably, the lowest Stokes shift values are observed in flavonols that lack hydroxyl groups. Considering the similar mechanism and kinetics of excited-state intramolecular proton transfer (ESIPT), as well as the larger  $\Delta\nu_{\text{St}}$  values for hydroxyl-containing flavonols and the planar geometry of molecules in both the initial and phototautomeric forms ( $T^*$ ), we can conclude that the differences in Stokes shifts arise from the structural characteristics of the solvation shell in the ground state and its subsequent rearrangement during phototautomerization.

The changes in solvent polarity on going from dichloromethane to acetonitrile do not significantly affect the position of the phototautomer emission bands. The observed shifts in the bands are multidirectional, do not exceed a few nanometers, and are statistically insignificant. The Stokes shifts of fluorescence for 3'-R-hydroxyflavonols, 4'-R-hydroxyflavonols, and 3',4'-R-dihydroxyflavonols in acetonitrile are within the ranges of 10 190–10500  $\text{cm}^{-1}$ , 9775–10290  $\text{cm}^{-1}$ , and 9150–9665  $\text{cm}^{-1}$ , respectively. Thus,  $\Delta\nu_{\text{St}}$  values of flavonols in more polar acetonitrile are correspondingly 460, 625, and 225  $\text{cm}^{-1}$  higher than those in less polar dichloromethane. A comparison of the spectral characteristics of the absorption and fluorescence bands indicates that the increase in  $\Delta\nu_{\text{St}}$  is due to a hypsochromic shift of the absorption bands when going from dichloromethane to acetonitrile solutions. This suggests additional energy costs are associated with the relaxation of the solvate shell of flavonols upon excitation in polar media.

### Fluorescence spectra in the solid state

To investigate the spectral properties of flavonols in the solid phase, we grew crystals of the compounds we were studying and recorded their fluorescence spectra. Since recording the absorption spectra of the crystals was not possible, we obtained fluorescence excitation spectra for each compound instead. The shape of these fluorescence excitation spectra closely resembles that of the corresponding absorption spectra, enabling us to

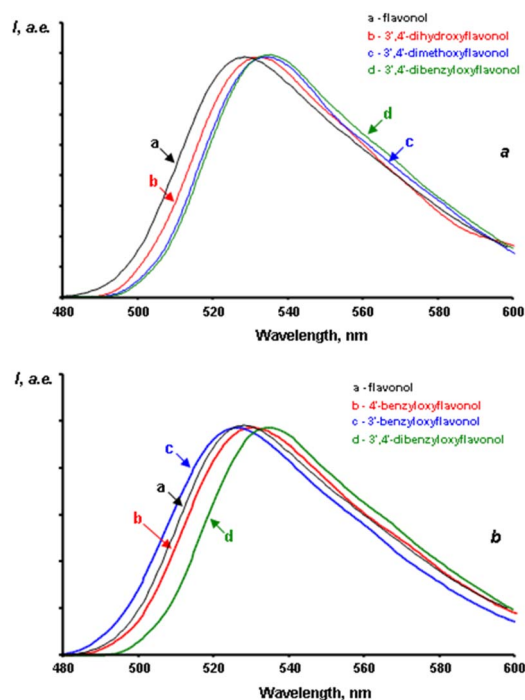


Fig. 5 Fluorescence spectra of studied flavonols in dichloromethane. a – different substituents in the same position of the side benzene ring, b – the same substituents in different positions of the side benzene ring. Concentrations of flavonols were in a range of  $1\text{--}5 \cdot 10^{-5}$  M.



Table 5 Spectral characteristics of studied flavonols in the solid state<sup>a</sup>

Flavonol	3'-R	4'-R	Excitation spectra		Fluorescence spectra						
					Form N*			Form T*			
			$\lambda_{\text{ex}}$	$\nu_{\text{ex}}$	$\lambda_{\text{N}}$	$\nu_{\text{N}}$	$\Delta\nu_{\text{St,N}}$	$\lambda_{\text{T}}$	$\nu_{\text{T}}$	$\Delta\nu_{\text{St,T}}$	$I_{\text{N}}/I_{\text{T}}$
3 <sup>b</sup>	H	H	357	28 020	461	21 690	6330	547	18 290	9730	0.85
3a	H	OH	363	27 610	453	22 080	5530	553	18 080	9530	1.14
3b	H	OCH <sub>3</sub>	364	27 530	452	22 130	5400	545	18 350	9180	1.73
3c	H	OBn	363	27 580	467	21 394	6186	557	17 946	9634	1.26
3d	OH	H	363	27 590	475	21 060	6530	539	18 560	9030	1.13
3e	OH	OH	365	27 400	453	22 080	5320	539	18 560	8840	1.58
3f	OH	OCH <sub>3</sub>	363	27 570	460	21 720	5850	561	17 840	9730	1.11
3g	OCH <sub>3</sub>	H	362	27 660	450	22 210	5450	537	18 630	9030	1.76
3h	OCH <sub>3</sub>	OH	362	27 640	464	21 540	6100	550	18 180	9460	1.32
3i	OCH <sub>3</sub>	OCH <sub>3</sub>	364	27 460	486	20 550	6910	536	18 660	8800	1.57
3j	OCH <sub>3</sub>	OBn	365	24 625	456	21 950	5475	545	18 342	9083	1.62
3k	OBn	H	375	26 670	457	21 865	4805	555	18 030	8640	2.57
3l	OBn	OCH <sub>3</sub>	362	27 670	454	22 040	5630	542	18 450	9220	1.63
3m	OBn	OBn	357	28 020	461	21 690	6330	547	18 290	9730	0.85

<sup>a</sup>  $\lambda_{\text{ex}}$ ,  $\nu_{\text{ex}}$  – positions of long-wavelength excitation bands in nm and  $\text{cm}^{-1}$ ;  $\lambda_{\text{N}}$ ,  $\nu_{\text{N}}$ ,  $\lambda_{\text{T}}$ ,  $\nu_{\text{T}}$  – positions of emission bands of N\* and T\* forms in nm and  $\text{cm}^{-1}$ ;  $\Delta\nu_{\text{St,N}}$  and  $\Delta\nu_{\text{St,T}}$  – Stokes shifts of fluorescence of N\* and T\* forms in  $\text{cm}^{-1}$  calculated relatively long-wavelength excitation bands:  $\Delta\nu_{\text{St,N}} = \nu_{\text{exc}} - \nu_{\text{N}}$  and  $\Delta\nu_{\text{St,T}} = \nu_{\text{exc}} - \nu_{\text{T}}$ ;  $I_{\text{N}}/I_{\text{T}}$  – ratio of intensities of forms N\* and T\*. <sup>b</sup> Spectral data for unsubstituted flavonol were taken from ref. 50 data for nonpolar medium are shown for chloroform.

assess the light absorption characteristics of flavonols in their solid state.

Fluorescence excitation spectra were measured by assessing the luminescence intensity at 450 nm and 550 nm, corresponding to the emission regions of the N\* and T\* forms, respectively. The refined positions of the maxima of the long-wavelength band in the excitation spectra ( $\lambda_{\text{max}}^{\text{ex}}/\nu_{\text{max}}^{\text{ex}}$ ) were determined through double differentiation of the spectral curves. The differences in maxima positions in the spectra measured at 450 nm and 550 nm did not exceed 3 nm; therefore, averaged values of  $\lambda_{\text{max}}^{\text{ex}}/\nu_{\text{max}}^{\text{ex}}$  were utilized for further

analysis of the obtained data. The spectral characteristics of flavonols in the solid state are presented in Table 5.

The data presented in the Table 5 indicate that the positions of the fluorescence excitation bands in the solid phase do not correspond with the positions of the absorption bands in flavonol solutions, nor do they align with the Hammett constants of the side ring. When going from the less polar solvent dichloromethane to the more polar solvent acetonitrile, the positions of absorption band maxima shift hypsochromically by an average of  $415 \text{ cm}^{-1}$ . In contrast, moving to the crystalline phase results in a bathochromic shift of the bands by

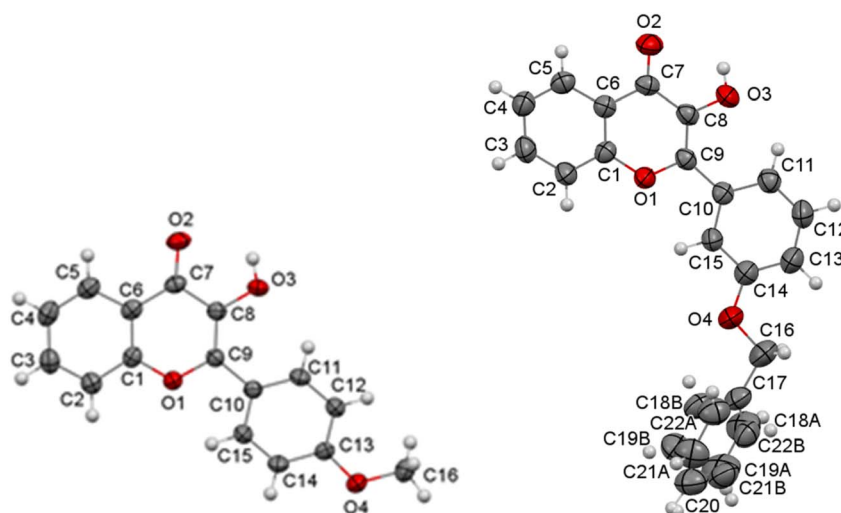


Fig. 6 Molecular structure of compounds 3b (on the left) and 3k (on the right) according to the X-ray diffraction data. Thermal ellipsoids are shown at 50% probability level.



an average of  $730\text{ cm}^{-1}$ , which suggests a decrease in the environmental polarity. Additionally, flavonols in the crystalline state exhibit a smaller variation in the bands' positions.

To understand how substituents on the side benzene ring of flavonols influence their spectral properties, X-ray structural analysis was conducted on compounds **3b** and **3k**. The structures and geometries of the substances studied, as obtained from X-ray diffraction, are illustrated in Fig. 6. It is important to note that the "crystallographic" numbering of atoms, assigned during structure resolution using the OLEX2 package, does not align with the atom numbering typically used for flavonols. Therefore, when referencing the "crystallographic" atom numbers below, an asterisk will be used to indicate them.

The presence of a phenyl substituent at the C9\* atom in molecules **3b** and **3k** (Fig. 6) suggests conjugation between the  $\pi$ -systems of the chromone fragment and the side benzene ring. The X-ray analysis revealed that, in contrast to their geometry in solution or the gas phase, flavonols in a crystalline state are non-planar molecules. Thus, in molecules **3b** and **3k**, these fragments are rotated relative to each other (torsion angle C8\*-C9\*-C10\*-C11\* in Fig. 6) by  $12.3(3)^\circ$  in molecule **3b** and by  $-17.3(4)^\circ$  in molecule **3k**, and the length of the C9\*-C10\* bond ( $1.468(2)\text{ \AA}$  in molecule **3b** and  $1.463(3)\text{ \AA}$  in molecule **3k**) is comparable to the average length of the  $C_{sp^2}-C_{ar}$  bond in conjugated systems  $1.470\text{ \AA}$ .<sup>52</sup>

The non-planarity of flavonol molecules in the solid state, *i.e.* the presence of some non-zero dihedral angles between the molecular fragments additionally diminishes the influence of substituents on the side ring regarding the electron density distribution in the chromone part of the molecule. Thus, the effect of substituents on the spectral properties of flavonols occurs only indirectly, mediated by conformational effects and the polarity of the surrounding environment. The latter is influenced by the parameters of the crystal lattice, which can in turn be affected by the volumetric characteristics and positions of the substituents.

Unlike the fluorescence spectra of flavonol solutions, which show only one emission band from the T\* phototautomer, the spectra of flavonols in the solid state exhibit two emission bands (Fig. 7).

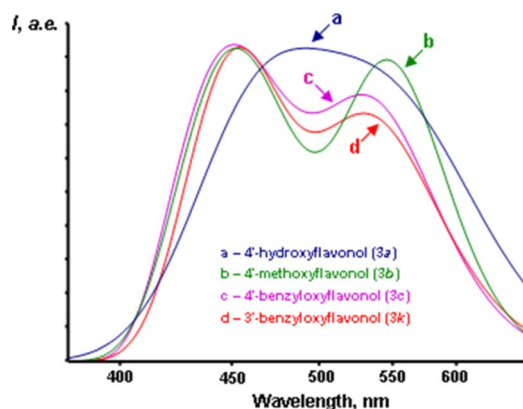


Fig. 7 Fluorescence spectra of flavonols in the solid state.

The short-wavelength fluorescence bands of the N\* form in most flavonols typically range from 450 to 460 nm (approximately  $22\,200\text{--}21\,700\text{ cm}^{-1}$ ). However, in compounds **3d**, **3e**, and **3j**, these bands are slightly shifted toward the long-wavelength region. It is important to note the large Stokes shifts of fluorescence, which range from  $4800$  to  $6900\text{ cm}^{-1}$ . This range is approximately  $1500$  to  $2500\text{ cm}^{-1}$  greater than that observed in most fluorophores. This discrepancy likely results from significant energy costs associated with the structural relaxation of the excited molecule within a rigid crystal lattice.

The fluorescence bands of the T\* form are centered between 540 and 560 nm (corresponding to  $18\,550\text{--}17\,850\text{ cm}^{-1}$ ). These bands are shifted by 20 to 30 nm toward the long-wavelength region of the spectrum compared to the fluorescence bands of the same form in the solutions. In the solid state, the Stokes shifts of the fluorescence of the flavonol phototautomer are smaller than those in the solutions, measuring between  $8800$  and  $9730\text{ cm}^{-1}$ . This suggests the absence of a significant structural relaxation of the phototautomer in the excited state.

No dependence was found between the positions of the N\* and T\* bands in the fluorescence spectra of crystalline flavonols and the nature or position of the substituents in the side ring.

The appearance of the short-wavelength N\* emission band evidences that the intramolecular proton transfer is partially suppressed. The side ring substituents have weak electronic effects, and the non-planar geometry of the molecules suggests that the side rings have a minimal impact on proton transfer kinetics. Another factor contributing to the reduced efficiency of the excited-state intramolecular proton transfer is the weakening of the intramolecular hydrogen bond necessary for the proton transfer between the carbonyl group and the 3-hydroxy group of chromone.

As illustrated in Fig. 8, the hydrogen atoms of the 3-hydroxy group form bifurcated hydrogen bonds. According to Etter's rules,<sup>53,54</sup> the formation of intramolecular hydrogen bonds is preferred over intermolecular hydrogen bonds. However, the hydrogen bond that closes the five-membered ring is not sufficiently effective. This likely leads to the hydroxyl group in all three structures acting as a proton donor in two hydrogen bonds simultaneously: the intramolecular  $O_3^*-\text{H}\cdots O_2^*$  and the intermolecular  $O_3^*-\text{H}\cdots O_2^*$ . The  $O_2^*$  atom, in turn, serves as a proton acceptor in both hydrogen bonds.

The nature of the primary structural motif formed by the intermolecular hydrogen bonds varies. In structure **3b**, hydrogen-bonded chains are formed, while in structures **3c** and **3k**, centrosymmetric hydrogen-bonded dimers are established (see Fig. 8). It is important to note that the formation of bifurcated hydrogen bonds weakens each bond, as demonstrated by their characteristics (refer to Table 6).

Since the formation of multi-center hydrogen bonds weakens the intramolecular hydrogen-bond component, it leads to a decrease in the rate of phototautomerization. Consequently, the rate of the proton transfer becomes comparable to the rate of emission of the original form N\*, which results in the appearance of the corresponding short-wavelength band N\* in the spectra.



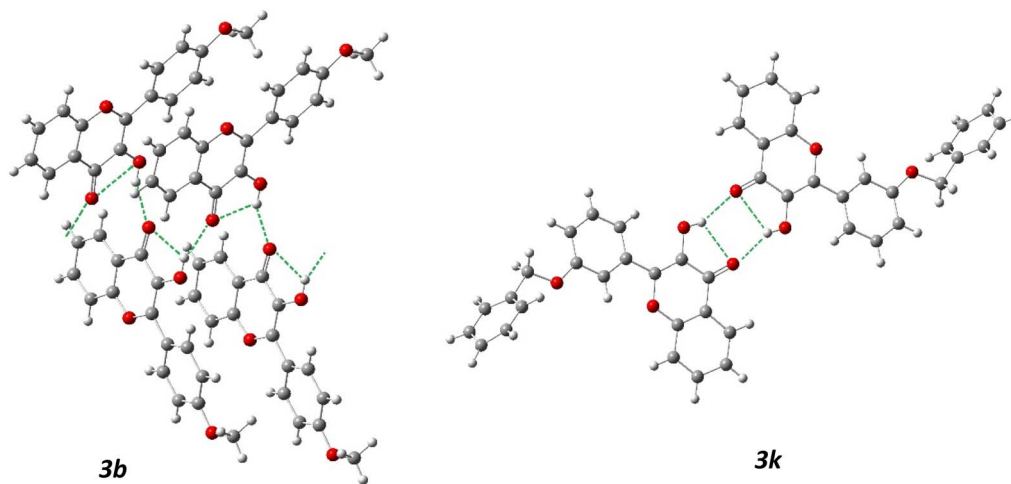


Fig. 8 Crystalline structure of 4'-methoxy (**3b**) and 3'-benzyloxy (**3k**) flavonols. Intramolecular and intermolecular hydrogen bonds are shown with green dashed lines.

Table 6 Geometric characteristics of hydrogen bonds in structures **3b** and **3k**

		Geometric characteristics		
Hydrogen bond	Symmetry operation	H···O, Å	O···O, Å	O–H···O, deg
<b>Structure 3b</b>				
O3*–H···O2*		2.23	2.677(2)	113.30
O3*–H···O2’*	1 – x,0.5 + y,0.5 – z	1.98	2.753(2)	153.6
<b>Structure 3k</b>				
O3*–H···O2*		2.27	2.712(3)	113.5
O3*–H···O2’*	–x, –y, 1 – z	2.01	2.751(3)	146.0

The fluorescence intensity ratio of the initial form ( $N^*$ ) to the phototautomer ( $T^*$ ), represented as  $I_N/I_T$ , indicates that for most compounds, the emission intensity of form  $N^*$  is greater than that of form  $T^*$ . In several flavonols, the intensity  $I_N$  can exceed  $I_T$  by 10–20%. However, most compounds show low ESIPT efficiency with the luminescence intensity of form  $N^*$  surpassing that of form  $T^*$  by 1.5 to 2.5 times. An analysis of the  $I_N/I_T$  values reveals that the ratio of forms  $N^*$  and  $T^*$  in the excited state does not depend on the position and nature of the substituents. Nonetheless, a significant slowdown in ESIPT is primarily observed in flavonols with bulky benzyl substituents. This suggests that the spectral properties of these compounds must be determined by the characteristics of their crystal.

### Binding interactions of flavonols with $\beta$ -glucosidase

Fluorescent flavonols serve as promising molecular probes for investigating protein-ligand interactions, with the goal of understanding the mechanisms of enzyme action.<sup>55,56</sup> They also provide a theoretical foundation for the design and discovery of new regulators of enzymatic activity.<sup>55,57,58</sup> Recently, we demonstrated that the ESIPT fluorescence of flavonols is highly sensitive to binding interactions with proteins, making it an

effective tool for probing protein structure and their hydrophobic pockets.<sup>59,60</sup> Furthermore, incorporating bulky alkyl and benzoyl groups into the 2-phenyl ring of flavonols increases their hydrophobicity and enhances the binding affinity to  $\beta$ -glucosidase, allowing for the inhibition of the enzyme's activity.<sup>26</sup>

In this study, we aimed to investigate the influence of C3'- and C4'-substitutions in flavonols on their protein binding affinity and selectivity. We examined the fluorescence properties of flavonols when interacting with  $\beta$ -glucosidase. Fig. 9 displays the results of fluorescence titration of flavonols **3k** and **3l** by varying concentrations of  $\beta$ -glucosidase in a phosphate buffer at pH 6.86. Despite their common use in biomedical applications, flavonols have low solubility in water.<sup>61</sup> To address this issue, we dissolved them in the buffer by adding small aliquots of their DMSO stock solution. To explore the interaction between flavonols and the enzyme, we followed a fluorescence titration protocol similar to the one used in our recent study.<sup>26</sup> In summary, during the titration, we maintained the concentration of flavonols at approximately  $1\text{--}2 \times 10^{-5}$  M, while gradually varying the concentration of glucosidase from 0 to  $9.5 \times 10^{-4}$  M.

As shown in Fig. 9, the fluorescence intensity of both flavonols **3k** and **3l** increases with higher concentrations of  $\beta$ -glucosidase. This phenomenon, which has also been observed with other flavonols, is attributed to a protein-induced “turn-on” effect.<sup>26,39,58,59,62</sup> The increase in fluorescence can be explained by the reduction of water-induced fluorescence quenching, which occurs as the probes bind to and penetrate deeper into the water-free hydrophobic regions of biomacromolecules.<sup>6,63</sup> In addition to the significant “turn-on” effect observed, there is also a noticeable redistribution of the normal-to-phototautomer emission intensity. Specifically, the contribution of the normal form, represented by the short-wavelength band around 450 nm, gradually increases. This dual-band redistribution is more prominent in the case of





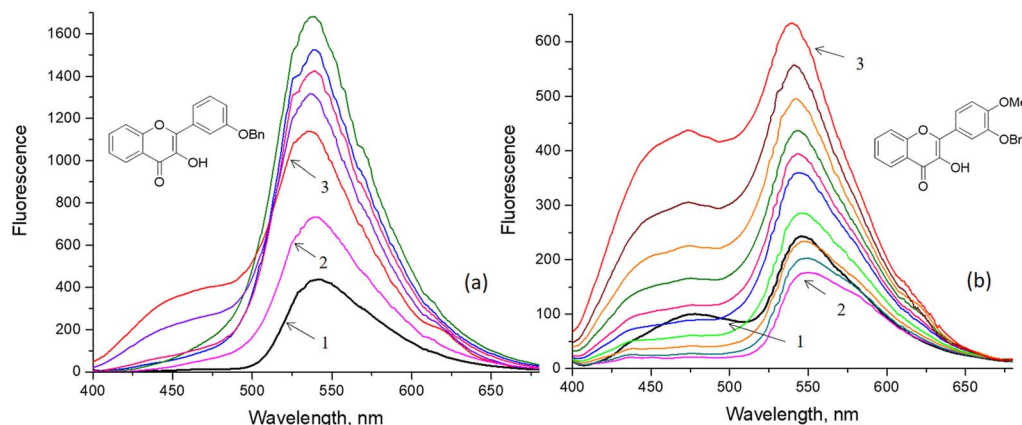


Fig. 9 Fluorescence titration of flavonols **3k** (a) and **3l** (b) with  $\beta$ -glucosidase. The flavonol fluorescence was measured in the phosphate buffer pH 6.86 at 298 K after excitation at 380 nm. The arrows indicate the flavonol spectra: (1) in the absence of  $\beta$ -glucosidase, (2) after adding the first  $\beta$ -glucosidase aliquot, (3) in the presence of maximal concentration of  $\beta$ -glucosidase. The flavonol concentration of  $2.3 \cdot 10^{-5}$  M (a, b) was constant, whereas the  $\beta$ -glucosidase concentration was varied in a range from  $1.1 \cdot 10^{-5}$  up to  $9.5 \cdot 10^{-4}$  M (a) and from  $2.2 \cdot 10^{-5}$  up to  $8.8 \cdot 10^{-4}$  M (b).

flavonol **3l**, indicating a possible alteration in its binding mode to the  $\beta$ -glucosidase protein (see Fig. 9b).

The strong fluorescence of the normal form suggests that the ESIPT process is somewhat inhibited in the protein-bound state of flavonols **3k** and **3l**. Several factors could explain this, including: (i) the formation of intermolecular hydrogen bonds with protein residues, or (ii) the disruption of the intramolecular hydrogen bond between the 3-OH group and the 4O carbonyl oxygen due to steric constraints within the protein-binding pocket. Additionally, it is possible that both mechanisms contribute to the ESIPT-sensitive behavior of the flavonols. This implies that even small modifications in the peripheral side ring of the chromone moiety could significantly influence ligand-protein interactions, including their strength and selectivity.

### ADMET properties

The physicochemical properties of molecular probes influence their water solubility, lipophilicity, and binding affinity to organized hydrophobic environments, such as micelles, lipid membranes, plasma proteins and other biomacromolecules. In the context of *in vitro* and in-cell studies, a proper balance of these properties is crucial for governing the adsorption, distribution, metabolism, and pharmacokinetics of flavonols used as ESIPT probes. Therefore, a thorough understanding of these properties, referred to as ADMET (Absorption, Distribution, Metabolism, Excretion, and Toxicity), along with their measurement and prediction, is essential for successful probe design and development.<sup>64</sup>

Probe's lipophilicity affects its permeability across cell membranes, the partitioning and distribution of a drug-like molecule, as well as its metabolism and renal excretion.<sup>65</sup> Our fluorescence titration experiments shown in Fig. 9 demonstrated that flavonol binding to the protein was primarily driven by the hydrophobic effect because of their high lipophilicity. Therefore, we first tried to see if there is some correlations

between spectral changes seen in Fig. 9 and lipophilicity of the studied flavonols.

Lipophilicity is defined as the logarithm of the octanol–water partition coefficient ( $\log P_{o/w}$ ) that reflects the ability of a molecule to dissolve in the lipophilic environment, such as a lipid membrane.

Table 7 summarizes calculated consensus  $\log P$  for model flavonol **3** and studied flavonols **3a–m**. Table 7 illustrates that the gradual introduction of methoxy and benzoyl groups into the 2-phenyl ring increases the  $\log P_{o/w}$  value from 2.84 for the unsubstituted flavonol to as high as 5.87 for the compound **3m**. It is also noteworthy that empirical *in silico* predictions of  $\log P_{o/w}$  parameters are not sensitive to the specific positions of substitutions. For instance, both **3j** and **3l** were predicted to have the same  $\log P_{o/w}$  value of 4.33. To thoroughly investigate the molecular-level details of how variations in the C3' and C4'-substituents fine-tune protein-ligand binding interactions, computational tools, such as molecular docking, that account for the implicit structure of proteins should be utilized.

### Molecular docking calculations

Due to the unavailability of the high-resolution 3D structure of the  $\beta$ -glucosidase from commercial almonds used in our experimental studies,<sup>66</sup> we turned to X-ray structures of  $\beta$ -glucosidases from other sources. The  $\beta$ -glucosidase family includes a variety of enzymes that exhibit different activities and are widely distributed across many living organisms.<sup>67</sup> These enzymes share the common ability to hydrolyze  $\beta$ -glucosidic linkages found in disaccharides, oligosaccharides, and conjugated saccharides.

Fig. 10 illustrates the X-ray structure of the  $\beta$ -glucosidase from *Thermotoga maritima* (TmGH1). Although there are some variations in the primary structure at the peripheral regions of the protein, all enzymes within the glucosidase family exhibit a high sequence identity of over 90%. The main common features of these enzymes include: (i) the presence of a deep



Table 7 Calculated ADMET properties of the studied flavonols<sup>a</sup>

Flavonol	3'-R <sub>1</sub>	4'-R <sub>2</sub>	Mw (g mol <sup>-1</sup> )	TPSA (Å <sup>2</sup> )	Number of H-bond donors	Number of H-bond acceptors	Consensus log P <sub>o/w</sub> <sup>*</sup>
<b>3</b>	H	H	238.2	50.4	1	3	2.84
<b>3a</b>	H	OH	254.2	70.7	2	4	2.77
<b>3b</b>	H	OCH <sub>3</sub>	268.3	59.7	1	4	3.02
<b>3c</b>	H	OBn	330.3	59.7	1	4	4.57
<b>3d</b>	OH	H	254.2	70.7	2	4	2.77
<b>3e</b>	OH	OH	270.2	90.9	3	5	2.34
<b>3f</b>	OH	OCH <sub>3</sub>	284.3	79.9	2	5	2.62
<b>3g</b>	OCH <sub>3</sub>	H	268.3	59.7	1	4	3.02
<b>3h</b>	OCH <sub>3</sub>	OH	284.3	79.9	2	5	2.62
<b>3i</b>	OCH <sub>3</sub>	OCH <sub>3</sub>	298.3	68.9	1	5	2.85
<b>3j</b>	OCH <sub>3</sub>	OBn	360.4	68.9	1	5	4.33
<b>3k</b>	OBn	H	330.3	59.7	1	4	4.57
<b>3l</b>	OBn	OCH <sub>3</sub>	360.4	68.9	1	5	4.33
<b>3m</b>	OBn	OBn	422.4	68.9	1	5	5.87

<sup>a</sup> Consensus log P<sub>o/w</sub> was estimated based on set of *in silico* predicting tools, such as PreADME, Molinspiration, XLOGP3, ALOGPS 2.1, ChemDoodle, pkCSM, and Osiris Property Explorer, respectively.

hydrophobic pocket that can accommodate substrate molecules during cellulolytic hydrolysis (Fig. 10a), and (ii) a catalytic active site composed of two glutamate (Glu) residues situated in close proximity to each other (see Fig. 10a and b).

For our molecular docking analysis of flavonol–glucosidase interactions, we selected  $\beta$ -glucosidases from four different sources: *Paenibacillus polymyxa*  $\beta$ -glucosidase B (BglB),<sup>45</sup> *Raucaffricine*  $\beta$ -glucosidase (rBG),<sup>46</sup> *Thermotoga maritima*  $\beta$ -glucosidase (TmGH1),<sup>47</sup> and human cytosolic  $\beta$ -glucosidase (hCBG).<sup>48</sup> The X-ray 3D structures of these proteins have been well resolved, so that some of them already used as receptor models for molecular docking calculations.<sup>26,39,62,68–70</sup>

The structural and energetic characteristics of the interactions between flavonols **3k** and **3l**, and  $\beta$ -glucosidases were

analyzed using molecular docking calculations, which are summarized in Table 8.

The results indicate that all examined flavonols exhibit strong binding affinities, with values exceeding  $-8.5$  kcal mol<sup>-1</sup>. Furthermore, flavonols can penetrate deeply into the central cavity of  $\beta$ -glucosidase and bind closely to the catalytic glutamic acid (Glu) residues.

Examples of binding modes for flavonols are illustrated in Fig. 11, specifically for  $\beta$ -glucosidase TmGH1. Molecular docking studies indicate that the binding affinity of compounds **3k** and **3l** to  $\beta$ -glucosidase depends on the specific characteristics of the peripheral substituents located at the C3'- and C4'-positions of the flavonol's phenyl ring.

In the case of flavonol **3k**, it binds deeper into the enzyme pocket, with the 3OH-chromone moiety favoring a position

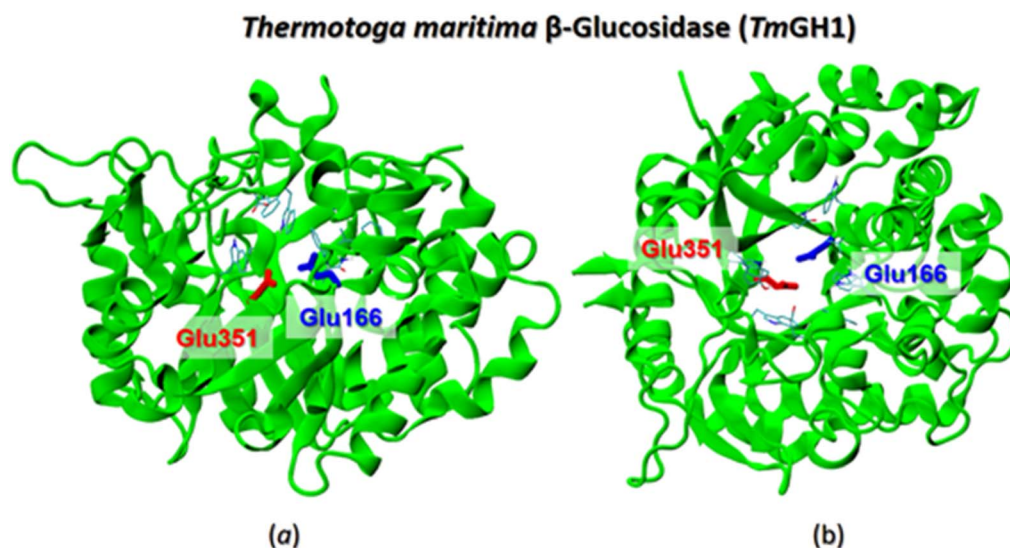


Fig. 10 The X-ray structure of *Thermotoga maritima*  $\beta$ -glucosidase (PDB 1OD0)<sup>47</sup> shown in a side (a) and top (b) views, respectively. The catalytic Glu166 and Glu351 residues for are shown by color-coded sticks.



Table 8 The binding affinity of flavonols with  $\beta$ -glucosidases from various sources estimated by molecular docking calculations

Flavonol	3'-R <sub>1</sub>	4'-R <sub>2</sub>	Binding affinity, kcal mol <sup>-1</sup>			
			Human cytosolic $\beta$ -glucosidase (PDB 2JFE)	<i>Paenibacillus polymyxa</i> $\beta$ -glucosidase (PDB 2O9R)	<i>Thermotoga maritima</i> $\beta$ -glucosidase (PDB 1OD0)	<i>Raucaffricine</i> $\beta$ -glucosidase (PDB 4A3Y)
<b>3</b>	H	H	-8.8	-8.3	-8.5	-8.7
<b>3a</b>	H	OH	-9.2	-8.5	-8.9	-8.5
<b>3b</b>	H	OCH <sub>3</sub>	-9.3	-8.9	-8.6	-8.8
<b>3c</b>	H	OBn	-10.4	-9.1	-9.3	-9.3
<b>3d</b>	OH	H	-9.0	-8.6	-8.9	-9.5
<b>3e</b>	OH	OH	-9.6	-9.0	-8.8	-9.0
<b>3f</b>	OH	OCH <sub>3</sub>	-8.6	-8.4	-8.2	-9.0
<b>3g</b>	OCH <sub>3</sub>	H	-8.9	-8.9	-8.7	-9.4
<b>3h</b>	OCH <sub>3</sub>	OH	-9.0	-8.7	-8.9	-9.3
<b>3i</b>	OCH <sub>3</sub>	OCH <sub>3</sub>	-8.8	-8.7	-8.6	-8.0
<b>3j</b>	OCH <sub>3</sub>	OBn	-10.4	-9.4	-9.3	-9.3
<b>3k</b>	OBn	H	-10.6	-10.0	-9.9	-9.8
<b>3l</b>	OBn	OCH <sub>3</sub>	-10.6	-9.7	-9.4	-9.2
<b>3m</b>	OBn	OBn	-11.9	-11.1	-11.0	-10.2

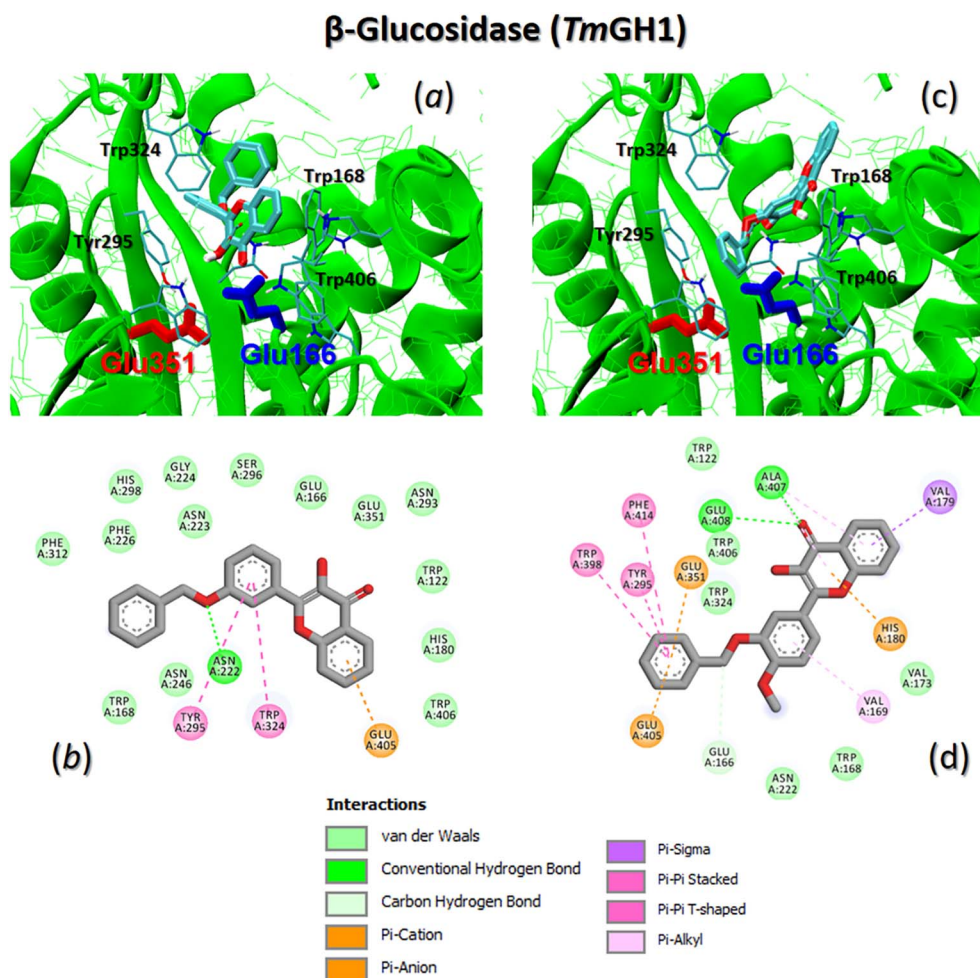


Fig. 11 The best binding mode of flavonols **3k** (a and b) and **3l** (c and d) to  $\beta$ -glucosidase TmGH1 (PDB 1OD0). The catalytic residues Glu166 and Glu351 are shown as color-coded sticks. Some key enzyme interacting residues are also shown. (b and d) 2D interaction maps of selected flavonols with key protein residues.



close to the catalytic residue Glu166. The bulky 3'-benzyl ring protrudes outward from the enzyme pocket (see Fig. 11a). In contrast, the introduction of an additional 4'-methoxy group results in a reversal of the binding mode. For flavonol 3l, the 2-phenyl ring, featuring the bulky 3'-benzyl and 4'-methoxy groups, occupies the catalytic pocket, displacing the 3OH-chromone moiety outwards (see Fig. 11c).

Fig. 11b and d illustrate 2D interaction maps of flavonols with  $\beta$ -glucosidase TmGH1. Flavonol **3k** binds to TmGH1 through non-covalent interactions, including  $\pi$ - $\pi$  stacking of the 2-phenyl ring with TYR295 and TRP324. It also involves short-range  $\pi$ -anion interactions between the aromatic ring of the chromone and GLU405, as well as hydrogen bonding between the 3'-benzyl oxygen atom and ASN222 (Fig. 11b). In contrast, flavonol 3l exhibits much more complex interaction patterns. The major contributions to its binding include classical  $\pi$ - $\pi$  stacking or  $\pi$ - $\pi$  T-shaped stacking of the 3'-benzyl ring with TYR295, TRP298, and PHE414. Additionally, there are  $\pi$ -anion interactions of the 3'-benzyl ring with GLU405 and catalytic GLU351, along with hydrogen bonding of the 2O carbonyl oxygen atom with ALA407 and GLU408 (Fig. 11d).

### Summary and perspectives

Natural flavonols, including compounds like quercetin, morin, fisetin, and galangin, are among the most abundant classes of metabolic polyphenols. They exhibit a wide range of pharmacological activities beneficial to human health.<sup>1,2</sup> Many flavonols also display unique dual ESIPT-emission fluorescence, which makes them sensitive to changes in their microenvironment.<sup>6,7</sup> This property offers exciting opportunities for sensing and detecting metal ions, anions, small ligands, and biomacromolecules.<sup>3,24,58,71</sup> However, the diverse structures of flavonols—characterized by variations in the number and position of hydroxyl groups, as well as various chemical modifications—complicate the relationship between their structure and fluorescence.<sup>23,72</sup> As a result, using them as fluorescent probes can often be challenging.<sup>58</sup>

This study aims to clarify the influence of hydroxy, methoxy, and benzyl groups at the C3' and C4' positions of the 2-phenyl side ring of flavonols. We focus on examining their ESIPT fluorescence, crystal packing, physicochemical properties, and ADMET characteristics. A series of flavonols were synthesized, and their structures were characterized using NMR and MS analysis. We systematically investigated the fluorescence properties of the synthesized flavonols in aprotic solvents, analyzing the relationships between their structures and properties. Our main findings suggest that the nature and position of substituent groups in flavonols significantly influence their crystal packing in the solid state. We discovered that the molecular arrangement in the crystal lattice could be affected by intra- and intermolecular hydrogen bonding ratio, which in turn affects the ESIPT dual-band ratio. Furthermore, by employing fluorescence titration and molecular docking calculations, we explored how the introduction of a bulky benzyl moiety and the alteration of its position between C3'- and C4'- can impact the binding interactions of flavonols with  $\beta$ -glucosidases. We

believe our findings shed light on the structure–fluorescence relationship in flavonols and open up new possibilities for the design of innovative flavonol-based probes.

### Author contributions

Oleksii O. Demidov: investigation, writing – original draft, writing – review and editing. Liudmyla V. Chepeleva, Svitlana V. Shishkina: investigation, writing – original draft, writing – review and editing. Eugene S. Gladkov: investigation, writing – original draft. Alexander V. Kyrchenko: investigation, conceptualization, writing – original draft, writing – review and editing. Rostyslav P. Linnik: conceptualization, writing – original draft. Alexander D. Roshal: supervision, conceptualization, writing – original draft, writing – review and editing.

### Conflicts of interest

There are no conflicts to declare.

### Data availability

The additional data for this study are available from the corresponding author upon request.

CCDC 2416411 (**3b**), 2416412 (**3c**) and 2416347 (**3k**) contain the supplementary crystallographic data for this paper.<sup>73a–c</sup>

Supplementary information: the crystallographic data in Table S1, <sup>1</sup>H and <sup>13</sup>C NMR spectra of **3a–m** in Fig. S1–S13 and mass spectra of **3a–m** in Fig. S14–26. See DOI: <https://doi.org/10.1039/d5ra05790f>.

### Acknowledgements

O.O.D., L.V.C., E.S.G., A.D.R. acknowledge Grant 2023.03/0083 of the National Research Foundation of Ukraine.

### References

- 1 S. Tang, B. Wang, X. Liu, W. Xi, Y. Yue, X. Tan, J. Bai and L. Huang, Structural insights and biological activities of flavonoids: Implications for novel applications, *Food Front.*, 2025, **6**, 218–247.
- 2 Y. Liu, J. Qian, J. Li, M. Xing, D. Grierson, C. Sun, C. Xu, X. Li and K. Chen, Hydroxylation decoration patterns of flavonoids in horticultural crops: chemistry, bioactivity, and biosynthesis, *Hortic. Res.*, 2022, **9**, uhab068.
- 3 T. Qin, B. Liu, Z. Xu, G. Yao, H. Xu and C. Zhao, Flavonol-based small-molecule fluorescent probes, *Sens. Actuators, B*, 2021, **336**, 129718.
- 4 A. D. Roshal, J. A. Organero and A. Douhal, Tuning the mechanism of proton-transfer in a hydroxyflavone derivative, *Chem. Phys. Lett.*, 2003, **379**, 53–59.
- 5 I. E. Serdiuk and A. D. Roshal, Single and double intramolecular proton transfers in the electronically excited state of flavone derivatives, *RSC Adv.*, 2015, **5**, 102191–102203.





- 6 V. G. Pivovarenko and A. S. Klymchenko, Fluorescent Probes Based on Charge and Proton Transfer for Probing Biomolecular Environment, *Chem. Rec.*, 2024, **24**(2), e202300321.
- 7 V. G. Pivovarenko, Multi-parametric sensing by multi-channel molecular fluorescent probes based on excited state intramolecular proton transfer and charge transfer processes, *BBA Adv.*, 2023, **3**, 100094.
- 8 P. K. Sengupta, Excited state proton transfer based two color fluorescence: Perspectives and some biophysical applications, *J. Ind. Chem. Soc.*, 2021, **98**, 100059.
- 9 I. E. Serdiuk and A. D. Roshal, Exploring double proton transfer: A review on photochemical features of compounds with two proton-transfer sites, *Dyes Pigm.*, 2017, **138**, 223–244.
- 10 A. S. Klymchenko, Solvatochromic and Fluorogenic Dyes as Environment-Sensitive Probes: Design and Biological Applications, *Acc. Chem. Res.*, 2017, **50**, 366–375.
- 11 S. Höfener, P. C. Kooijman, J. Groen, F. Ariese and L. Visscher, Fluorescence behavior of (selected) flavonols: a combined experimental and computational study, *Phys. Chem. Chem. Phys.*, 2013, **15**, 12572–12581.
- 12 A. Y. Chumak, V. O. Mudrak, V. M. Kotlyar and A. O. Doroshenko, 4'-Nitroflavonol fluorescence: Excited state intramolecular proton transfer reaction from the non-emissive excited state, *J. Photochem. Photobiol., A*, 2021, **406**, 112978.
- 13 Z. Xu, X. Zhao, M. Zhou, Z. Zhang, T. Qin, D. Wang, L. Wang, X. Peng and B. Liu, Donor engineering on flavonoid-based probes to enhance the fluorescence brightness in water: Design, characterization, photophysical properties, and application for cysteine detection, *Sens. Actuators, B*, 2021, **345**, 130367.
- 14 T. Qin, B. Liu, Y. Huang, K. Yang, K. Zhu, Z. Luo, C. Pan and L. Wang, Ratiometric fluorescent monitoring of methanol in biodiesel by using an ESIPT-based flavonoid probe, *Sens. Actuators, B*, 2018, **277**, 484–491.
- 15 A. O. Doroshenko, A. V. Kyrychenko, O. M. Valyashko, V. N. Kotlyar and D. A. Svechkarev, 4'-Methoxy-3-hydroxyflavone excited state intramolecular proton transfer reaction in alcoholic solutions: Intermolecular versus intramolecular hydrogen bonding effect, *J. Photochem. Photobiol., A*, 2019, **383**, 111964.
- 16 A. P. Demchenko, S. Ercelen, A. D. Roshal and A. S. Klymchenko, Excited-State Proton Transfer Reaction in a New Benzofuryl 3-Hydroxychromone Derivative: The Influence of Low-Polar Solvents, *Pol. J. Chem.*, 2002, **76**, 1287–1299.
- 17 I. E. Serdiuk and A. D. Roshal, 7-Hydroxyflavone Revisited. 2. Substitution Effect on Spectral and Acid–Base Properties in the Ground and Excited States, *J. Phys. Chem. A*, 2015, **119**, 12672–12685.
- 18 I. E. Serdiuk, A. S. Varenikov and A. D. Roshal, 7-Hydroxyflavone Revisited: Spectral, Acid–Base Properties, and Interplay of the Protolytic Forms in the Ground and Excited States, *J. Phys. Chem. A*, 2014, **118**, 3068–3080.
- 19 I. E. Serdiuk, A. D. Roshal and J. Błażejowski, Origin of Spectral Features and Acid–Base Properties of 3,7-Dihydroxyflavone and Its Monofunctional Derivatives in the Ground and Excited States, *J. Phys. Chem. A*, 2016, **120**, 4325–4337.
- 20 S. Concilio, M. Di Martino, A. M. Nardiello, B. Panunzi, L. Sessa, Y. Miele, F. Rossi and S. Piotta, A Flavone-Based Solvatochromic Probe with A Low Expected Perturbation Impact on the Membrane Physical State, *Molecules*, 2020, **25**, 3458.
- 21 R. De, K. W. Jo and K.-T. Kim, Influence of Molecular Structures on Fluorescence of Flavonoids and Their Detection in Mammalian Cells, *Biomedicines*, 2022, **10**, 1265.
- 22 S. Saha, E. Panieri, S. Suzen and L. Saso, The Interaction of Flavonols with Membrane Components: Potential Effect on Antioxidant Activity, *J. Membr. Biol.*, 2020, **253**, 57–71.
- 23 A. D. Roshal, Complexation of Flavonoids: Spectral Phenomena, Regioselectivity, Interplay with Charge and Proton Transfer, *Chem. Rec.*, 2024, **24**, e202300249.
- 24 M. D. Yilmaz, S. Altves and S. Erbas-Cakmak, Naturally Occurring Flavonol, Quercetagenin 5,6,7,3',4'-Pentamethyl Ether (Marionol), as a Nontoxic Plant-Based Fluorescent Probe for Rapid, Sensitive, and Selective Detection of Cu<sup>2+</sup> in Water, *ACS Omega*, 2024, **9**, 47342–47349.
- 25 O. O. Demidov, A. P. Krasnopyorova, G. D. Yukhno, N. V. Efimova and A. D. Roshal, Flavonol assisted extraction of divalent and trivalent metal ions, *Funct. Mater.*, 2024, **31**, 601–608.
- 26 L. V. Chepeleva, O. O. Demidov, A. D. Snizhko, D. O. Tarasenko, A. Y. Chumak, O. O. Kolomoitsev, V. M. Kotliar, E. S. Gladkov, A. Kyrychenko and A. D. Roshal, Binding interactions of hydrophobically-modified flavonols with  $\beta$ -glucosidase: fluorescence spectroscopy and molecular modelling study, *RSC Adv.*, 2023, **13**, 34107–34121.
- 27 J. Lee, Y. Jeong, H. Jin Jung, S. Ullah, J. Ko, G. Young Kim, D. Yoon, S. Hong, D. Kang, Y. Park, P. Chun, H. Young Chung and H. Ryong Moon, Anti-tyrosinase flavone derivatives and their anti-melanogenic activities: Importance of the  $\beta$ -phenyl- $\alpha,\beta$ -unsaturated carbonyl scaffold, *Bioorg. Chem.*, 2023, **135**, 106504.
- 28 V. A. Chebanov, S. M. Desenko and T. W. Gurley, *Azaheterocycles Based on  $\alpha,\beta$ -Unsaturated Carbonyls*, Springer-Verlag, Berlin Heidelberg, 2008.
- 29 W. Xiong, X. Wang, X. Shen, C. Hu, X. Wang, F. Wang, G. Zhang and C. Wang, Synthesis of Flavonols via Pyrrolidine Catalysis: Origins of the Selectivity for Flavonol versus Aurone, *J. Org. Chem.*, 2020, **85**, 13160–13176.
- 30 T. Kan, A. Hiza, Y. Tsukaguchi, T. Ogawa, M. Inai, T. Asakawa and Y. Hamashima, Synthetic Studies of Fisetin, Myricetin and Nobiletin Analogs and Related Probe Molecules, *Heterocycles*, 2014, **88**, 1371–1396.
- 31 F. Estévez-Sarmiento, M. Said, I. Brouard, F. León, C. García, J. Quintana and F. Estévez, 3'-Hydroxy-3,4'-dimethoxyflavone blocks tubulin polymerization and is a potent apoptotic inducer in human SK-MEL-1 melanoma cells, *Bioorg. Med. Chem.*, 2017, **25**, 6060–6070.



- 32 E. Venkateswararao, M.-J. Son, N. Sharma, M. Manickam, P. Boggu, Y. H. Kim, S.-H. Woo and S.-H. Jung, Exploration of Pharmacophore in Chrysosplenol C as Activator in Ventricular Myocyte Contraction, *ACS Med. Chem. Lett.*, 2015, **6**, 758–763.
- 33 A. W. Weig, P. M. O'Conner, J. M. Kwiecinski, O. M. Marciano, A. Nunag, A. T. Gutierrez, R. J. Melander, A. R. Horswill and C. Melander, A structure activity relationship study of 3,4'-dimethoxyflavone for ArlRS inhibition in *Staphylococcus aureus*, *Org. Biomol. Chem.*, 2023, **21**, 3373–3380.
- 34 C. X. Qin, X. Chen, R. A. Hughes, S. J. Williams and O. L. Woodman, Understanding the Cardioprotective Effects of Flavonols: Discovery of Relaxant Flavonols without Antioxidant Activity, *J. Med. Chem.*, 2008, **51**, 1874–1884.
- 35 A. D. Becke, Density-functional thermochemistry. III. The role of exact exchange, *J. Chem. Phys.*, 1993, **98**, 5648–5652.
- 36 T. H. Dunning, Gaussian basis sets for use in correlated molecular calculations. I. The atoms boron through neon and hydrogen, *J. Chem. Phys.*, 1989, **90**, 1007–1023.
- 37 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. V. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, Williams, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery Jr, J. E. Peralta, F. Ogliaro, M. J. Bearpark, J. J. Heyd, E. N. Brothers, K. N. Kudin, V. N. Staroverov, T. A. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. P. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman and D. J. Fox, *Gaussian 16 rev. C.01*, Wallingford, CT, 2016.
- 38 J. Tomasi, B. Mennucci and R. Cammi, Quantum Mechanical Continuum Solvation Models, *Chem. Rev.*, 2005, **105**, 2999–3094.
- 39 L. V. Chepeleva, D. O. Tarasenko, A. Y. Chumak, O. O. Demidov, A. D. Snizhko, O. O. Kolomoitsev, V. M. Kotliar, E. S. Gladkov, A. L. Tatarets, A. V. Kyrchenko and A. D. Roshal, 4'-Benzoyloxyflavonol glucoside as fluorescent indicator for  $\beta$ -glucosidase activity, *Funct. Mater.*, 2023, **30**, 494–505.
- 40 O. V. Dolomanov, L. J. Bourhis, R. J. Gildea, J. A. K. Howard and H. Puschmann, OLEX2: a complete structure solution, refinement and analysis program, *J. Appl. Crystallogr.*, 2009, **42**, 339–341.
- 41 G. Sheldrick, SHELXT - Integrated space-group and crystal-structure determination, *Acta Crystallogr. A*, 2015, **71**, 3–8.
- 42 G. Sheldrick, Crystal structure refinement with SHELXL, *Acta Crystallogr. C*, 2015, **71**, 3–8.
- 43 D. S. Goodsell, G. M. Morris and A. J. Olson, Automated docking of flexible ligands: Applications of AutoDock, *J. Mol. Recognit.*, 1996, **9**, 1–5.
- 44 O. Trott and A. J. Olson, AutoDock Vina: Improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading, *J. Comput. Chem.*, 2010, **31**, 455–461.
- 45 P. Isorna, J. Polaina, L. Latorre-García, F. J. Cañada, B. González and J. Sanz-Aparicio, Crystal Structures of *Paenibacillus polymyxa*  $\beta$ -Glucosidase B Complexes Reveal the Molecular Basis of Substrate Specificity and Give New Insights into the Catalytic Machinery of Family I Glycosidases, *J. Mol. Biol.*, 2007, **371**, 1204–1218.
- 46 L. Xia, M. Ruppert, M. Wang, S. Panjikar, H. Lin, C. Rajendran, L. Barleben and J. Stöckigt, Structures of Alkaloid Biosynthetic Glucosidases Decode Substrate Specificity, *ACS Chem. Biol.*, 2012, **7**, 226–234.
- 47 D. L. Zechel, A. B. Boraston, T. Gloster, C. M. Boraston, J. M. Macdonald, D. M. G. Tilbrook, R. V. Stick and G. J. Davies, Iminosugar Glycosidase Inhibitors: Structural and Thermodynamic Dissection of the Binding of Isofagomine and 1-Deoxynojirimycin to  $\beta$ -Glucosidases, *J. Am. Chem. Soc.*, 2003, **125**, 14313–14323.
- 48 S. Tribolo, J.-G. Berrin, P. A. Kroon, M. Czjzek and N. Juge, The Crystal Structure of Human Cytosolic  $\beta$ -Glucosidase Unravels the Substrate Aglycone Specificity of a Family 1 Glycoside Hydrolase, *J. Mol. Biol.*, 2007, **370**, 964–975.
- 49 A. D. Roshal, V. G. Mitina, V. D. Orlov, O. A. Ponomarev, A. A. Sukhorukov and S. F. Fialkova, Interpretation of electron transitions in absorption spectra of phenylchromones and their hydroxy derivatives, *Funct. Mater.*, 1997, **4**, 121–127.
- 50 A. D. Roshal, V. I. Moroz, V. G. Pivovarenko, A. Wróblewska and J. Błażejowski, Spectral and Acid–Base Features of 3,7-Dihydroxy-2,8-diphenyl-4H,6H-pyrano[3,2-g]chromene-4,6-dione (Diflavonol) A Potential Probe for Monitoring the Properties of Liquid Phases, *J. Org. Chem.*, 2003, **68**, 5860–5869.
- 51 P. Ertl, A Web Tool for Calculating Substituent Descriptors Compatible with Hammett Sigma Constants, *Chem.: Methods*, 2022, **2**, e202200041.
- 52 H.-B. Burgi and J. D. Dunitz, *Structure Correlation*, VCH, Weinheim, 1994.
- 53 M. C. Etter, Encoding and decoding hydrogen-bond patterns of organic compounds, *Acc. Chem. Res.*, 1990, **23**, 120–126.
- 54 M. C. Etter, Hydrogen bonds as design elements in organic chemistry, *J. Phys. Chem.*, 1991, **95**, 4601–4610.
- 55 S. Pal and C. Saha, A review on structure–affinity relationship of dietary flavonoids with serum albumins, *J. Biomol. Struct. Dyn.*, 2014, **32**, 1132–1147.
- 56 J. Xiao and G. Kai, A Review of Dietary Polyphenol-Plasma Protein Interactions: Characterization, Influence on the Bioactivity, and Structure-Affinity Relationship, *Crit. Rev. Food Sci. Nutr.*, 2012, **52**, 85–101.
- 57 S. Sengupta, M. Datta and S. Datta, in *Glycoside Hydrolases*, ed. A. Goyal and K. Sharma, Academic Press, 2023, vol. 5, pp. 97–120.



- 58 G. N. Sagandykova, P. P. Pomastowski and B. Buszewski, Multi-instrumental approach to unravel molecular mechanisms of natural bioactive compounds: Case studies for flavonoids, *Trac. Trends Anal. Chem.*, 2020, **126**, 115865.
- 59 O. O. Demidov, E. S. Gladkov, A. V. Kyrychenko and A. D. Roshal, Synthetic and Natural Flavonols as Promising Fluorescence Probes for  $\beta$ -Glucosidase Activity Screening, *Funct. Mater.*, 2022, **29**, 252–262.
- 60 I. E. Serdiuk, M. Reszka, H. Myska, K. Krzyński, B. Liberek and A. D. Roshal, Flavonol-based fluorescent indicator for determination of  $\beta$ -glucosidase activity, *RSC Adv.*, 2016, **6**, 42532–42536.
- 61 L. Chebil, C. Humeau, J. Anthoni, F. Dehez, J.-M. Engasser and M. Ghoul, Solubility of Flavonoids in Organic Solvents, *J. Chem. Eng. Data*, 2007, **52**, 1552–1556.
- 62 S. Khan, T. Pozzo, M. Megyeri, S. Lindahl, A. Sundin, C. Turner and E. N. Karlsson, Aglycone specificity of *Thermotoga neapolitana*  $\beta$ -glucosidase 1A modified by mutagenesis, leading to increased catalytic efficiency in quercetin-3-glucoside hydrolysis, *BMC Biochem.*, 2011, **12**, 11.
- 63 P. Zhou and K. Han, ESIPT-based AIE luminogens: Design strategies, applications, and mechanisms, *Aggregate*, 2022, **3**, e160.
- 64 S. Tian, J. Wang, Y. Li, D. Li, L. Xu and T. Hou, The application of in silico drug-likeness predictions in pharmaceutical research, *Adv. Drug Delivery Rev.*, 2015, **86**, 2–10.
- 65 D. E. V. Pires, T. L. Blundell and D. B. Ascher, pkCSM: Predicting Small-Molecule Pharmacokinetic and Toxicity Properties Using Graph-Based Signatures, *J. Med. Chem.*, 2015, **58**, 4066–4072.
- 66 S. He and S. G. Withers, Assignment of Sweet Almond  $\beta$ -Glucosidase as a Family 1 Glycosidase and Identification of Its Active Site Nucleophile, *J. Biol. Chem.*, 1997, **272**, 24864–24867.
- 67 J. R. Ketudat Cairns, B. Mahong, S. Baiya and J.-S. Jeon,  $\beta$ -Glucosidases: Multitasking, moonlighting or simply misunderstood?, *Plant Sci.*, 2015, **241**, 246–259.
- 68 A. D. Snizhko, A. V. Kyrychenko and E. S. Gladkov, Synthesis of Novel Derivatives of 5,6,7,8-Tetrahydro-quinazolines using of  $\alpha$ -Aminoamidines and In Silico Screening of Their Biological Activity, *Int. J. Mol. Sci.*, 2022, **23**, 3781.
- 69 G.-Y. Chen, H. Zhang and F.-Q. Yang, A simple and portable method for  $\beta$ -Glucosidase activity assay and its inhibitor screening based on a personal glucose meter, *Anal. Chim. Acta*, 2021, **1142**, 19–27.
- 70 S. Riaz, I. U. Khan, M. Yar, M. Ashraf, T. U. Rehman, A. Shaukat, S. B. Jamal, V. C. M. Duarte and M. J. Alves, Novel pyridine-2,4,6-tricarbohydrazide derivatives: Design, synthesis, characterization and in vitro biological evaluation as  $\alpha$ - and  $\beta$ -glucosidase inhibitors, *Bioorg. Chem.*, 2014, **57**, 148–154.
- 71 A. Kyrychenko and A. S. Ladokhin, Fluorescent Probes and Quenchers in Studies of Protein Folding and Protein-Lipid Interactions, *Chem. Rec.*, 2024, **24**(2), e202300232.
- 72 L. Giordano, V. V. Shvadchak, N. Arrupe, L. J. Falomir Lockhart, V. Sanchez and T. M. Jovin, Tuning of environment-sensitive 3-hydroxychromone fluorophores based on strong donor substituents in positions 2 or 7, *Dyes Pigm.*, 2023, **218**, 111479.
- 73 (a) CCDC 2416411: Experimental Crystal Structure Determination, 2025, DOI: [10.5517/ccdc.csd.cc2m3grz](https://doi.org/10.5517/ccdc.csd.cc2m3grz); (b) CCDC 2416412: Experimental Crystal Structure Determination, 2025, DOI: [10.5517/ccdc.csd.cc2m3gs0](https://doi.org/10.5517/ccdc.csd.cc2m3gs0); (c) CCDC 2416347: Experimental Crystal Structure Determination, 2025, DOI: [10.5517/ccdc.csd.cc2m3dpv](https://doi.org/10.5517/ccdc.csd.cc2m3dpv).

