


Cite this: *RSC Adv.*, 2025, 15, 28481

# Tuning metal ion affinity in acyclic phenanthrene schiff bases: comparative study of ethylene and phenylene linkers

Haritha C, Swathi M  and Chinna Ayya Swamy P \*

We have successfully designed and synthesized two structurally simple salen-type Schiff base probes, designated as **SB-1** and **SB-2**, for the selective detection of biologically and environmentally relevant metal ions. Fluorescence studies revealed that **SB-1** exhibits a distinct "turn-on" fluorescence response in the presence of  $\text{Zn}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Na}^+$ , and  $\text{K}^+$  ions, while **SB-2** demonstrated a selective fluorescence enhancement exclusively for  $\text{Zn}^{2+}$  ions. In addition to its fluorescence response, **SB-1** displayed distinct colorimetric changes upon interaction with  $\text{Zn}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Na}^+$ , and  $\text{K}^+$  ions, highlighting its broad-spectrum sensing capability. In contrast, **SB-2** exhibited selective colorimetric responses only toward  $\text{Zn}^{2+}$  and  $\text{Cu}^{2+}$  ions. These results underscore the dual-mode sensing potential of the probes, with **SB-1** offering broader ion recognition and **SB-2** demonstrating higher selectivity. To gain insights into the interaction mechanism and validate the spectral changes observed experimentally, density functional theory (DFT) calculations were performed. The computational results supported the experimental findings, confirming significant electronic transitions associated with metal ion binding and providing a detailed understanding of the coordination environment and binding modes of **SB-1** and **SB-2** with various metal ions.  $^1\text{H}$  NMR titration studies further substantiated these results by revealing that the metal ions coordinate with the imine ( $\text{C}=\text{N}$ ) moiety at the core of the Schiff base structure, consistent with the UV-visible absorption and fluorescence spectroscopy data. The binding stoichiometry between the probes and metal ions ( $\text{Zn}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Mg}^{2+}$ ,  $\text{Na}^+$ , and  $\text{K}^+$ ) was determined to be 1 : 1, as confirmed through Job's plot analysis and supported by UV-Vis, fluorescence, DFT, and  $^1\text{H}$  NMR (for  $\text{Zn}^{2+}$ ) studies. The binding affinities were quantified using the Benesi-Hildebrand method, with association constants ( $K_a$ ) found to be in the range of  $0.88\text{--}2.28 \times 10^3 \text{ M}^{-1}$ . The limit of detection (LOD) for each metal ion was calculated to be in the micromolar ( $\mu\text{M}$ ) range (0.1 to 0.05  $\mu\text{M}$ ), demonstrating the high sensitivity of the probes for practical sensing applications.

Received 23rd May 2025  
Accepted 4th August 2025

DOI: 10.1039/d5ra03617h

rsc.li/rsc-advances

## 1. Introduction

Metal ion sensing is a critical area of research with far-reaching implications in both biological and environmental sciences.<sup>1</sup> Within this broad field, multi-metal ion sensing is especially valuable, as biological systems are inherently complex and often involve the simultaneous presence and interaction of multiple metal ions.<sup>2</sup> These ions play vital roles in cellular processes such as enzymatic catalysis, ion transport, gene regulation, and signal transduction.<sup>3</sup> Accurately detecting and distinguishing between these ions, particularly in dynamic or physiologically relevant environments, remains a significant challenge due to their similar charge-to-radius ratios and overlapping coordination preferences.<sup>4</sup> Among the biologically essential metal ions, zinc ( $\text{Zn}^{2+}$ ) occupies a central position due to its abundance and

multifunctionality in the human body. It is involved in DNA synthesis, transcriptional regulation, immune function, and neurotransmission, functioning either as a catalytic or structural cofactor.<sup>5</sup> Unlike redox-active metals like iron or copper,  $\text{Zn}^{2+}$  is redox-inert but exhibits a wide range of coordination geometries and ligand affinities, making it an ideal target for selective fluorescence sensing.<sup>6</sup> Dysregulation of zinc homeostasis has been implicated in numerous pathological conditions, including Alzheimer's disease, epilepsy, diabetes, and prostate cancer.<sup>7</sup> Environmentally, zinc contamination can disrupt microbial ecology and plant growth, underscoring the importance of its detection in soil and water samples.<sup>8</sup> Other metal ions such as sodium ( $\text{Na}^+$ ), potassium ( $\text{K}^+$ ), magnesium ( $\text{Mg}^{2+}$ ), and copper ( $\text{Cu}^{2+}$ ) are also indispensable for proper physiological functioning.  $\text{Na}^+$  and  $\text{K}^+$  play key roles in maintaining osmotic balance, nerve impulse transmission, and muscle contraction.<sup>9</sup>  $\text{Mg}^{2+}$  acts as a cofactor in over several enzymatic reactions, including those involved in ATP metabolism and nucleic acid synthesis.<sup>10</sup>  $\text{Cu}^{2+}$ , while essential in trace

Main Group Organometallics Optoelectronic Materials and Catalysis Lab, Department of Chemistry, National Institute of Technology, Calicut-673601, India. E-mail: swamy@nitc.ac.in



amounts for redox enzymes and iron metabolism, can be highly toxic when accumulated in excess, contributing to oxidative stress and neurodegenerative diseases.<sup>11</sup> The environmental and physiological significance of these ions, coupled with the narrow window between their beneficial and toxic levels, highlights the urgent need for accurate and efficient detection methods.<sup>12</sup>

To address this need, traditionally metal ion detection has relied on techniques such as atomic absorption spectroscopy (AAS) and inductively coupled plasma mass spectrometry (ICP-MS), which offer high sensitivity but require expensive instrumentation, extensive sample preparation, and are not amenable to real-time or *in situ* analysis.<sup>13</sup> In contrast, fluorescence-based sensors have emerged as powerful alternatives owing to their high sensitivity, operational simplicity, cost-effectiveness, and compatibility with live-cell imaging and real-time monitoring.<sup>14</sup> A wide variety of photophysical mechanisms – such as chelation-enhanced fluorescence (CHEF), photoinduced electron transfer (PET), internal charge transfer (ICT), fluorescence resonance energy transfer (FRET), and excimer/excimer formation – have been employed in sensor design.<sup>15</sup> Among these, PET-based sensors are particularly attractive for their “turn-on” response mechanism, in which fluorescence is quenched in the free state and restored upon metal ion coordination.<sup>16,20</sup> This mechanism ensures high signal-to-noise ratios and makes PET sensors especially useful for detecting trace levels of metal ions.<sup>17</sup> In recent years, phenanthrene has gained attention as a promising but underutilized fluorophore in sensor development.<sup>18</sup> Its rigid fused-ring structure, high fluorescence quantum yield, and excellent photostability make it ideal for robust sensing applications. Despite these advantages, phenanthrene-based systems have been less explored compared to more commonly used fluorophores like coumarin, anthracene, or triphenylamine.<sup>16c</sup>

In the present study, we report the development of two novel Schiff base fluorescent probes **SB-1** and **SB-2** anchored on a phenanthrene core. These ligands were synthesized *via* simple and high-yielding condensation reactions involving 5-(4a,4b-dihydrophenanthren-9-yl)-2-hydroxybenzaldehyde and either benzene-1,2-diamine (**SB-1**) or ethane-1,2-diamine (**SB-2**). The photophysical behavior and metal ion sensing properties of both receptors were extensively evaluated using UV-Vis absorption and fluorescence spectroscopy in DMSO. One of the most remarkable and unexpected findings of this work is the multi-metal ion sensing capability of **SB-1**, which exhibits fluorescence enhancement in the presence of a suite of biologically relevant metal ions, including Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Cu<sup>2+</sup>, and Zn<sup>2+</sup>. This broad yet selective responsiveness is particularly noteworthy given that the detection of monovalent ions such as Na<sup>+</sup> and K<sup>+</sup>, and even divalent Mg<sup>2+</sup> and Zn<sup>2+</sup>, through fluorescence “turn-on” sensors is rarely reported in the literature.<sup>3,8</sup> These ions typically exhibit weak binding affinities due to their high hydration energies and lack of strong coordination tendencies, making them challenging targets for optical sensing.<sup>3,9</sup> The ability of **SB-1** to detect these metal ions suggests a finely tuned binding cavity and a favorable spatial arrangement around the phenanthrene-based Schiff base, possibly facilitating interactions that enhance fluorescence upon ion

binding. In contrast, **SB-2** demonstrates high selectivity, exhibiting a significant fluorescence *turn-on* response exclusively in the presence of Zn<sup>2+</sup>, making it an excellent candidate for selective zinc sensing in complex biological environments where other metal ions may be present. This study not only underscores the potential of phenanthrene as a robust and versatile fluorophore scaffold but also highlights the importance of structural design in achieving selective *versus* broad-spectrum metal ion detection. The unique sensing behavior of **SB-1**, capable of responding to a range of biologically essential cations, and the specificity of **SB-2** for Zn<sup>2+</sup>, opens new avenues for the development of smart, adaptable fluorescent probes for real-world applications in diagnostics, environmental monitoring, and cellular imaging.

## 2. Experimental section

### 2.1. Materials and instruments

All reagents and solvents used in this study were of analytical grade. The compounds 4-bromophenol, benzene-1,2-diamine, ethane-1,2-diamine, (4a,4b-dihydrophenanthren-9-yl)boronic acid, trifluoroacetic acid (TFA, Spectrochem), hexamethylenetetramine, and various metal salts (purchased from Merck India) were utilized directly without further purification. Solvents such as acetonitrile, ethanol, and *N,N*-dimethylformamide (DMF), all procured from Spectrochem, were purified according to standard laboratory procedures prior to use. Deionized water, further purified by double distillation, was used for preparing all aqueous solutions. Stock solutions of metal ions including Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, Al<sup>3+</sup>, Fe<sup>2+</sup>, Co<sup>2+</sup>, Hg<sup>2+</sup>, Ni<sup>2+</sup>, Mn<sup>2+</sup>, Sn<sup>2+</sup>, Pb<sup>2+</sup>, Cd<sup>2+</sup>, Cu<sup>2+</sup>, and Zn<sup>2+</sup> were freshly prepared in deionized water. Column chromatography was performed using silica gel (60–120 mesh) as the stationary phase. NMR spectroscopic data were recorded on a JEOL JNM-ECZ-500R/M1 instrument (500 MHz for <sup>1</sup>H and 125 MHz for <sup>13</sup>C) using CDCl<sub>3</sub> as the solvent, with tetramethylsilane (TMS) as the internal standard. High-resolution mass spectra (HRMS) were acquired on a Thermo Scientific Exactive Orbitrap mass spectrometer, and results are reported in mass-to-charge (*m/z*) ratios. UV-visible absorption measurements were carried out on a SHIMADZU UV-2600 spectrophotometer with a slit width of 2 nm. Fluorescence spectra were recorded using a PerkinElmer LS 6500 fluorescence spectrometer. Theoretical calculations were performed using density functional theory (DFT) at the B3LYP/6-31G\* level with the Gaussian 09W software package.

### 2.2. UV-visible and fluorescence titration

UV-visible and fluorescence titration experiments for the compounds **SB-1** and **SB-2** were conducted using 1 mL of a 1 × 10<sup>−5</sup> M solution prepared in DMSO (stock concentration: 1 × 10<sup>−4</sup> M). Stock solutions of metal salts were prepared in water at a concentration of 1 × 10<sup>−3</sup> M. For the titration studies, incremental additions of a 1 × 10<sup>−4</sup> M aqueous solution of the respective metal ions were added to the probe solution. The changes in absorbance and fluorescence intensity were



monitored to evaluate the binding interactions between the receptors and metal ions.

### 2.3. Synthesis and characterization

**2.3.1. Synthesis of 5-bromo-2-hydroxybenzaldehyde.** 4-Bromophenol (10 g, 58 mmol) was dissolved in trifluoroacetic acid (100 mL), followed by the addition of hexamethylenetetramine (9.39 g, 67 mmol). The reaction mixture was refluxed at 90–100 °C for 5 hours. After completion, the mixture was cooled and stirred with 6 N HCl (100 mL) for 20 minutes. The product was then extracted using DCM and purified by column chromatography, yielding a yellow powder with 88% yield (Scheme 1).

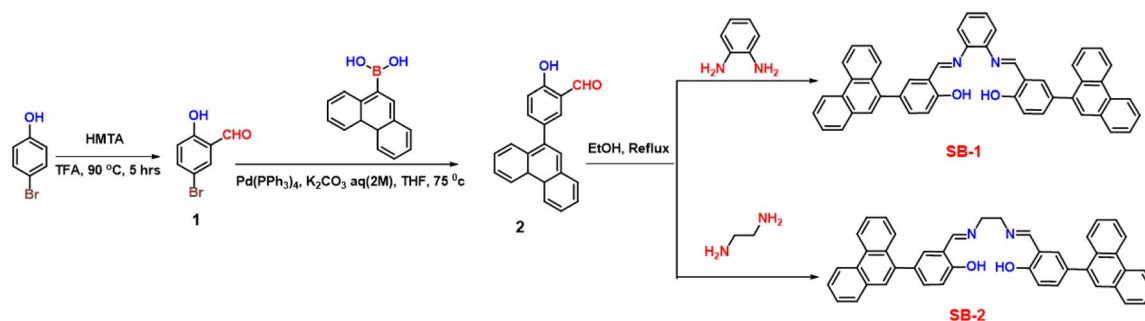
**2.3.2. Synthesis of 5-(4a,4b-dihydrophenanthren-9-yl)-2-hydroxybenzaldehyde.** In an oven-dried Schlenk flask, 5-bromo-2-hydroxybenzaldehyde (700 mg, 3.48 mmol) and phenanthren-9-ylboronic acid (850 mg, 3.83 mmol) were combined and dissolved in 50 mL of tetrahydrofuran (THF). A freshly prepared aqueous solution of potassium carbonate (1.4 g in 5 mL H<sub>2</sub>O) was added to the mixture. The resulting suspension was thoroughly degassed by bubbling nitrogen gas for 30 minutes. Following degassing, Pd(PPh<sub>3</sub>)<sub>4</sub> (40 mg) was added as the catalyst under nitrogen atmosphere, and the reaction mixture was stirred under reflux at 75 °C for 24 hours. Reaction progress was monitored by TLC until complete consumption of the starting material was confirmed. After completion, the reaction mixture was cooled to room temperature and concentrated under reduced pressure. The residue was extracted using ethyl acetate and water, and the organic phase was subsequently washed with brine. The organic layer was dried over anhydrous sodium sulfate, and concentrated. Purification of the crude product was carried out by column chromatography on silica gel using a gradient of hexane and ethyl acetate as eluent. The desired product was obtained as an off-white solid in an isolated yield of 80.6% (837.9 mg). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 11.14 (s, 1H), 9.95 (s, 1H), 8.79 (d, *J* = 8.2 Hz, 1H), 8.73 (d, *J* = 8.3 Hz, 1H), 7.90 (d, *J* = 7.8 Hz, 1H), 7.85 (d, *J* = 8.2 Hz, 1H), 7.71 (dd, *J* = 7.4, 1.6 Hz, 3H), 7.68 (m, 2H), 7.64 (t, *J* = 6.9 Hz, 1H), 7.57 (t, *J* = 7.1 Hz, 1H), 7.15 (d, *J* = 9.2 Hz, 1H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 197.1, 161.5, 139.2, 137.2, 135.2, 133.0, 131.9, 131.4, 131.2, 130.5, 129.1, 128.3, 127.5, 127.4, 127.2, 127.2, 126.8, 123.6, 123.1, 121.0, 118.1.

**2.3.3. Synthesis of SB-1.** An oven-dried round-bottom flask was charged with 2-hydroxy-5-(phenanthren-9-yl)benzaldehyde (250 mg, 0.838 mmol), which was dissolved in ethanol. To this solution, benzene-1,2-diamine (50 mg, 0.461 mmol) was added, and the mixture was refluxed at 90 °C for 12 hours. As the reaction progressed, a solid precipitate gradually formed. Upon completion, the reaction mixture was allowed to cool and was further stored at low temperature for 30 minutes to enhance precipitation. The resulting orange solid was collected by vacuum filtration, washed thoroughly with cold ethanol, and dried under reduced pressure. Yield = 82%. <sup>1</sup>H NMR (500 MHz) δ 13.19 (s, 2H), 9.07 (s), 8.94 (d, *J* = 8.2 Hz, 2H), 8.87 (d, *J* = 8.0 Hz, 2H), 8.03 (d, *J* = 7.5 Hz, 2H), 7.93 (d, *J* = 8.4 Hz, 2H), 7.90 (d, *J* = 6.9 Hz, 2H), 7.81 (s, 2H), 7.75–7.71 (m, 4H), 7.68 (d, *J* = 7.5 Hz, 2H), 7.62 (dd, *J* = 14.3, 5.7 Hz, 4H), 7.50 (d, *J* = 3.4 Hz, 2H), 7.42 (d, *J* = 2.5 Hz, 2H), 7.17 (d, *J* = 8.6 Hz, 2H). <sup>13</sup>C NMR (500 MHz, DMSO) δ 164.6, 160.5, 142.8, 137.6, 135.4, 134.8, 133.4, 131.6, 131.3, 130.9, 130.8, 129.1, 128.4, 127.8, 127.6, 127.3, 126.7, 126.7, 123.9, 123.3, 120.3, 120.0, 117.4, 117.1. HRMS (EI): calculated for C<sub>48</sub>H<sub>32</sub>N<sub>2</sub>O<sub>2</sub> [M–H]<sup>+</sup>, *m/z* 669.2542, found *m/z* 669.2540.

**2.3.4. Synthesis of SB-2.** Compound SB-2 was prepared following a procedure similar to that used for compound SB-1 replacing the benzene-1,2-diamine with ethylenediamine. The quantities involved and characterization data are as follows. 2-hydroxy-5-(phenanthren-9-yl)benzaldehyde (250 mg, 0.838 mmol), ethane-1,2-diamine (27.69 mg, 0.461 mmol) and ethanol (50 mL) yield = 89%. <sup>1</sup>H NMR (400 MHz, DMSO) δ 13.61 (s, 2H), 8.93 (d, *J* = 8.3 Hz, 2H), 8.85 (s, 2H), 8.73 (s, 2H), 8.00 (d, *J* = 7.5 Hz, 2H), 7.86 (d, *J* = 8.2 Hz, 2H), 7.76 (s, 2H), 7.74–7.67 (m, 5H), 7.65 (d, *J* = 2.1 Hz, 3H), 7.59 (t, *J* = 7.6 Hz, 2H), 7.51 (d, *J* = 8.4 Hz, 2H), 7.08 (s, 2H), 4.00 (s, 4H). <sup>13</sup>C NMR (101 MHz, DMSO) δ 167.5, 160.8, 137.7, 134.3, 133.2, 131.6, 130.9, 130.7, 130.6, 129.8, 129.1, 127.7, 127.6, 127.4, 127.3, 126.6, 123.9, 123.2, 118.9, 117.2, 59.2. HRMS (EI): calculated for C<sub>44</sub>H<sub>32</sub>N<sub>2</sub>O<sub>2</sub> [M–H]<sup>+</sup>, *m/z* 621.2542, found *m/z* 620.2547.

## 3. Results and discussions

SB-1 and SB-2, two Schiff base-based chemosensors, were synthesized *via* a simple condensation reaction between 5-(4a,4b-dihydrophenanthren-9-yl)-2-hydroxybenzaldehyde and either benzene-1,2-diamine (for SB-1) or ethane-1,2-diamine



Scheme 1 Synthesis of SB-1 and SB-2.

(for **SB-2**) in ethanol under reflux conditions, as depicted in Scheme 1. Following the reaction, the pure products were isolated as red solids by washing with cold ethanol and diethyl ether. The compounds were subsequently purified and characterized using a combination of  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectroscopy, along with high-resolution ESI-HRMS to confirm their molecular structures. In the  $^1\text{H}$  NMR spectrum of **SB-1**, a prominent signal at  $\delta = 9.06$  ppm was attributed to the proton of the extended phenyl conjugation, consistent with the expected structure. For **SB-2**, the imine proton ( $\text{HC}=\text{N}$ ) appeared at  $\delta = 8.73$  ppm, showing a slight upfield shift compared to **SB-1**, indicative of structural variations in the imine linkage. The  $^{13}\text{C}$  NMR spectrum revealed the imine carbon in **SB-1** at  $\delta = 160.4$  ppm, whereas **SB-2** showed a corresponding signal at  $\delta = 160.8$  ppm, further confirming the subtle differences in their molecular environments. The NMR spectra for both compounds displayed well-defined chemical shifts and integration values, supporting the successful formation of the intended Schiff base structures. The identities of **SB-1** and **SB-2** were further validated by ESI-HRMS analysis. The molecular ion for **SB-1** was observed at  $m/z$  669.2540, while **SB-2** displayed a peak at  $m/z$  621.2547, corresponding to their respective molecular weights. These results, in combination with the NMR data, explicitly confirm the successful synthesis of the Schiff base chemosensors and their expected molecular compositions.

### 3.1. Photophysical properties

After successfully isolating pure form of **SB-1** and **SB-2**, their optical properties were thoroughly investigated using UV-Vis absorption and fluorescence spectroscopy in DMSO solutions. The absorption spectrum of **SB-1** exhibited three distinct peaks at 260, 300, and 365 nm, which can be attributed to the electronic transitions within the aromatic system, the azomethine ( $-\text{CH}=\text{N}-$ ) bond, and the hydroxyl ( $-\text{OH}$ ) group. These absorption features are indicative of  $\pi-\pi$  and  $n-\pi$  transitions that are typical for Schiff base compounds containing conjugated structures. On the other hand, **SB-2** displayed two major absorption peaks at 260 and 300 nm, suggesting a lack of additional conjugation from the phenyl group present in **SB-1**. The absence of the 365 nm absorption peak in **SB-2** supports the idea that the phenanthrene unit in **SB-1** contributes significantly to its enhanced absorption characteristics, likely due to the extended conjugation that facilitates additional  $\pi-\pi^*$  transitions. These absorption profiles are summarized in Fig. S9. In terms of fluorescence properties, both **SB-1** and **SB-2** demonstrated relatively weak emission in DMSO solution. This low fluorescence intensity can be attributed to the well-known photon-induced electron transfer (PET) process commonly observed in Schiff base derivatives, thus quenching the fluorescence. In addition to our photophysical investigations, we measured the fluorescence quantum yields of **SB-1** and **SB-2** to further validate their emission characteristics. **SB-1** exhibited a moderate quantum yield of 0.04, whereas **SB-2** showed a significantly lower value of 0.01. These results are in good agreement with previously reported values for similar Schiff base systems and reinforce the conclusion that **SB-2** possesses

inherently weaker emissive properties compared to **SB-1**, which can be explained by the presence of the phenyl unit, which makes the system rigid. Overall, these optical studies underscore the significant role of molecular structure in governing the photophysical properties of Schiff base-based sensors. The differences observed in the absorption and fluorescence behaviour of **SB-1** and **SB-2** highlight the potential for tuning optical properties by modifying the ligand environment, which could be crucial for improving their performance in sensing applications.

### 3.2. UV-Vis absorption and fluorescence titration studies

UV-visible and fluorescence spectroscopy were employed to investigate the metal ion sensing capabilities of **SB-1** and **SB-2**. The UV-visible absorption spectra were monitored in DMSO upon the addition of various metal ions, including  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Al}^{3+}$ ,  $\text{Fe}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Hg}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Sn}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Cu}^{2+}$ , and  $\text{Zn}^{2+}$ . Significant spectral changes were observed for **SB-1** in the presence of  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Cu}^{2+}$ , and  $\text{Zn}^{2+}$  ions, as illustrated in Fig. 1, and S10–S12. In contrast, **SB-2** exhibited noticeable UV-spectral changes primarily only towards  $\text{Cu}^{2+}$  and  $\text{Zn}^{2+}$  ions. For **SB-1**, the addition of  $\text{Zn}^{2+}$  ions resulted in a gradual increase in absorption bands at 415 and 315 nm, while absorption intensities at 365 and 295 nm simultaneously decreased. Similarly,  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ , and  $\text{Cu}^{2+}$  ions influenced the absorption spectrum in a comparable manner. Upon addition of  $\text{Zn}^{2+}$  to **SB-1**, a distinct increase in absorption bands at 415 and 315 nm was observed, accompanied by a decrease in intensities at 365 and 295 nm. Comparable spectral alterations were noted in the presence of  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ , and  $\text{Cu}^{2+}$ , suggesting similar binding interactions. **SB-2**, on the other hand, showed a subtle increase at 325 nm and a decrease at 300 nm upon  $\text{Zn}^{2+}$  addition, although the spectral response was significantly weaker than that of **SB-1**. A notable visual colour change from colourless to yellow was observed upon adding  $\text{Zn}^{2+}$  ions to the **SB-1** solution, attributed to the formation of a new absorption band in the bathochromic region ( $\sim 415$  nm) (Fig. 1 and S10–S12). This result demonstrates that **SB-1** can detect  $\text{Zn}^{2+}$  ions in aqueous solutions with the naked eye, even in the presence of other competing metal ions. The emergence of this bathochromic band is likely due to intra-ligand charge transfer, redshifted upon strong metal chelation. In the case of  $\text{Cu}^{2+}$ , **SB-1** displayed enhanced absorption at 315 and 415 nm, while **SB-2** exhibited a peak shift centered around 320 nm. Competitive experiments, involving an excess of other metal ions, confirmed the selective sensitivity of **SB-1** toward  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Cu}^{2+}$ ,  $\text{Mg}^{2+}$  and  $\text{Zn}^{2+}$ , which induced new bands at 400 and 300 nm and a concurrent decrease at 365 and 290 nm. Meanwhile, **SB-2** remained largely inert to these metal ions, underscoring its comparatively narrower sensing profile (Scheme 2).

The metal-binding behaviour of **SB-1** and **SB-2** was further elucidated through the identification of isosbestic points in their UV-visible absorption spectra. **SB-1** displayed three well-defined isosbestic points upon interaction with selected metal ions, suggesting a clean interconversion between the free ligand and metal-bound forms. Specifically,  $\text{Zn}^{2+}$  and  $\text{Cu}^{2+}$  induced





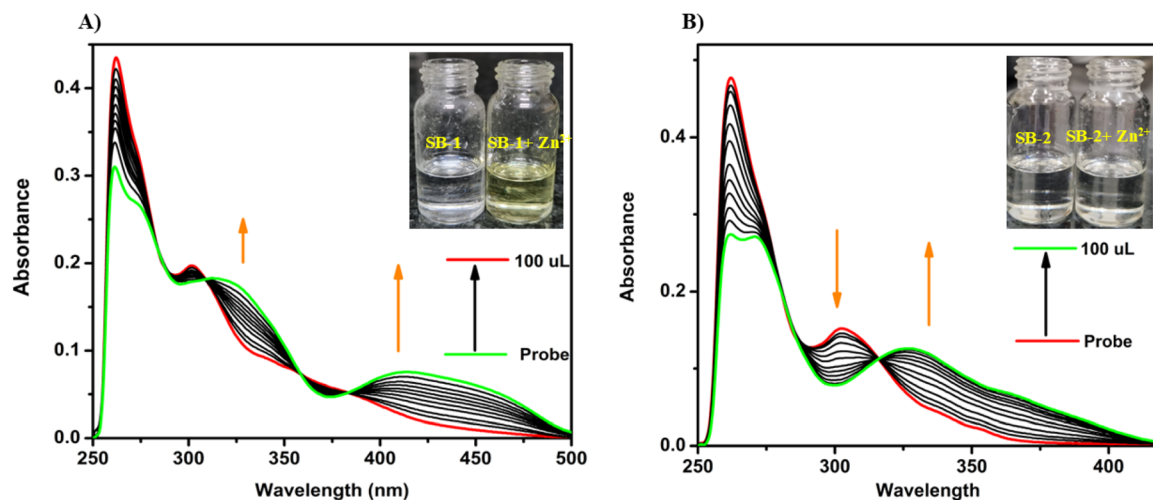
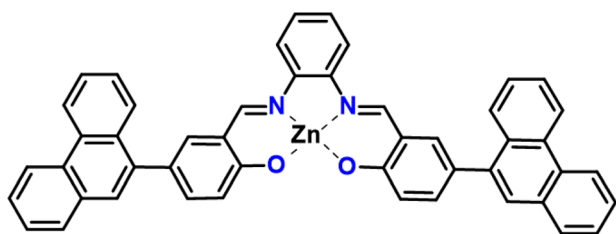


Fig. 1 UV-visible absorbance spectra of (A) SB-1 (left) and (B) SB-2 (right) in DMSO before and after the addition of  $\text{Zn}^{2+}$  ions. (Inset: images of the solutions under ambient light).



Scheme 2 Nature of metal binding of Schiff bases.

isosbestic transitions at 380, 360, and 310 nm, while  $\text{Na}^+$ ,  $\text{K}^+$ , and  $\text{Mg}^{2+}$  triggered similar transitions at 380, 350, and 320 nm, respectively. In contrast, **SB-2** showed only two isosbestic points, with  $\text{Cu}^{2+}$  producing transitions at 315 and 290 nm, and  $\text{Zn}^{2+}$  at 315 and 285 nm. These observations indicate fast and discrete complexation events, reflecting the formation of well-defined metal-ligand species. The clear visual color change observed in **SB-2**, particularly upon binding  $\text{Zn}^{2+}$  and  $\text{Cu}^{2+}$ , highlights its potential as a practical colorimetric sensor for these ions, with detectable changes even in competitive environments (Fig. S13). To quantify these interactions, association constants were determined using the Benesi-Hildebrand method. Among all tested ions,  $\text{Zn}^{2+}$  showed the highest binding affinity for **SB-1**, while  $\text{Cu}^{2+}$  displayed the strongest interaction with **SB-2**. In contrast,  $\text{Mg}^{2+}$  exhibited the weakest binding to **SB-1**, and  $\text{Zn}^{2+}$  showed the lowest affinity for **SB-2** (Table S1). The overall binding strength followed the order  $\text{Zn}^{2+} > \text{K}^+ > \text{Na}^+ > \text{Cu}^{2+} > \text{Mg}^{2+}$  for **SB-1** and  $\text{Cu}^{2+} > \text{Zn}^{2+}$  for **SB-2**. The detection limits (LOD) for each sensor-ion pair were calculated using the formula  $\text{LOD} = 3\sigma/S$ , where  $\sigma$  represents the standard deviation of the blank and  $S$  denotes the slope of the calibration curve derived from titration data. Remarkably, **SB-2** exhibited the highest sensitivity toward  $\text{Cu}^{2+}$ , with an LOD of 0.05  $\mu\text{M}$ , while **SB-1** also demonstrated strong detection capabilities with a  $\text{Cu}^{2+}$  LOD of 0.10  $\mu\text{M}$  (Table S1). In addition, other metal ions

such as  $\text{Zn}^{2+}$ ,  $\text{K}^+$ ,  $\text{Na}^+$ , and  $\text{Mg}^{2+}$  also demonstrated detection limits in  $\mu\text{M}$  with both **SB-1** and **SB-2**, highlighting their broad applicability. Absorbance values for both chemosensors plateaued after the addition of 1.0 equivalent of their respective target ions, confirming the formation of 1 : 1 ratio of metal and Schiff base complexes. Further insights into the binding stoichiometry were obtained from Job's plot analyses, which supported the proposed 1 : 1 ligand-to-metal complexation ratio for the most responsive ions. Upon adding metal ions to the receptors **SB-1** and **SB-2**, the absorption maxima at 420 nm were evaluated, to determine the binding stoichiometry, varying mole ratios of metal ions to receptors (0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, and 0.9). The Job's plot analysis, as illustrated in Fig. 2, and S15, revealed that the maximum absorption occurred at a 0.5 mole fraction for both **SB-1** and **SB-2** with  $\text{Zn}^{2+}$  ions, indicating a 1 : 1 binding stoichiometry. A similar trend was observed for other metal ions, including  $\text{Cu}^{2+}$ ,  $\text{Na}^+$ ,  $\text{K}^+$ , and  $\text{Mg}^{2+}$ , as shown in Fig. S14, and S16. These results underscore the effectiveness of **SB-1** and **SB-2** as selective and sensitive probes for transition and alkali/alkaline earth metal ions.

To gain deeper insights into the fluorescence behavior and metal ion sensing potential of **SB-1** and **SB-2**, detailed fluorescence titration experiments were conducted under standardized conditions. The selectivity and sensitivity of both chemosensors were evaluated using a comprehensive set of metal ions, including  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Al}^{3+}$ ,  $\text{Fe}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Hg}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Sn}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Cu}^{2+}$ , and  $\text{Zn}^{2+}$ . Initially, both **SB-1** and **SB-2** exhibited a very weak fluorescence emission band centered at  $\sim 490$  nm when excited at 350 nm in DMSO, indicating their non-emissive nature in the free state due to possible non-radiative decay pathways, such as  $-\text{C}=\text{N}-$  isomerization and PET (photoinduced electron transfer). Upon gradual addition of  $\text{Zn}^{2+}$  ions to a  $10^{-5}$  M solution of **SB-1** and **SB-2**, a significant "turn-on" fluorescence response was observed, with emission intensity progressively increasing with gradual addition of  $\text{Zn}^{2+}$  ions (Fig. 3). The observed fluorescence enhancement is



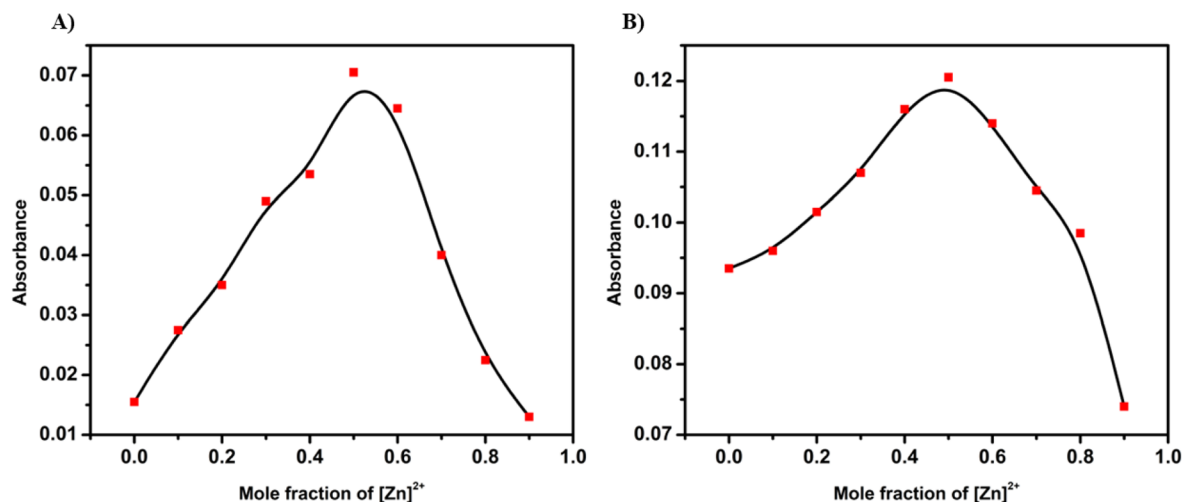


Fig. 2 Job's plot according to the method of continuous variations, indicating the 1 : 1 stoichiometry for compounds (A) SB-1 (A) and (B) SB-2 (B) with  $Zn^{2+}$  ions.

attributed to the suppression of non-radiative pathways such as C=N isomerization, PET, and ESIPT (excited-state intra-molecular proton transfer) processes, which are otherwise active in the free ligands. Coordination of metal ions with the imine nitrogen and phenolic oxygen moieties stabilizes the excited state and restricts conformational flexibility, thereby allowing efficient radiative decay. These mechanisms are well-supported by literature precedent for Schiff-base type fluorophores. Similarly, **SB-1** also showed fluorescence enhancement in the presence of  $Na^+$ ,  $K^+$ , and  $Mg^{2+}$  ions, although the intensity was lower compared to the  $Zn^{2+}$  induced emission, indicating a broader sensing profile for **SB-1** (Fig. S24). On the other hand, **SB-2** exhibited a vivid yellow fluorescence exclusively in the presence of  $Zn^{2+}$  ions, while no significant change in emission spectra was observed upon the addition of  $Na^+$ ,  $K^+$ , or  $Mg^{2+}$  ions, thereby confirming its high selectivity toward  $Zn^{2+}$ .

Furthermore, we evaluated the fluorescence quantum yields of **SB-1** and **SB-2** in the presence of various metal ions to better understand their sensing efficiencies. **SB-1** exhibited the highest quantum yield upon coordination with  $Mg^{2+}$  (0.32), followed by  $Zn^{2+}$  (0.28),  $Na^+$  (0.12), and  $K^+$  (0.10), indicating a broad yet differential fluorescence enhancement across multiple biologically relevant metal ions. In contrast, **SB-2** demonstrated a pronounced *turn-on* response specifically with  $Zn^{2+}$ , showing a significantly higher quantum yield of 0.54. Interestingly, no notable fluorescence enhancement or spectral shift was observed for either **SB-1** or **SB-2** upon exposure to other tested metal ions, even at higher concentrations. This selective response underscores the high specificity of **SB-1** and **SB-2** toward certain metal ions, particularly  $Zn^{2+}$ . Visual confirmation of this fluorescence enhancement was evident under UV light (365 nm), where the initially non-fluorescent **SB-1** solution

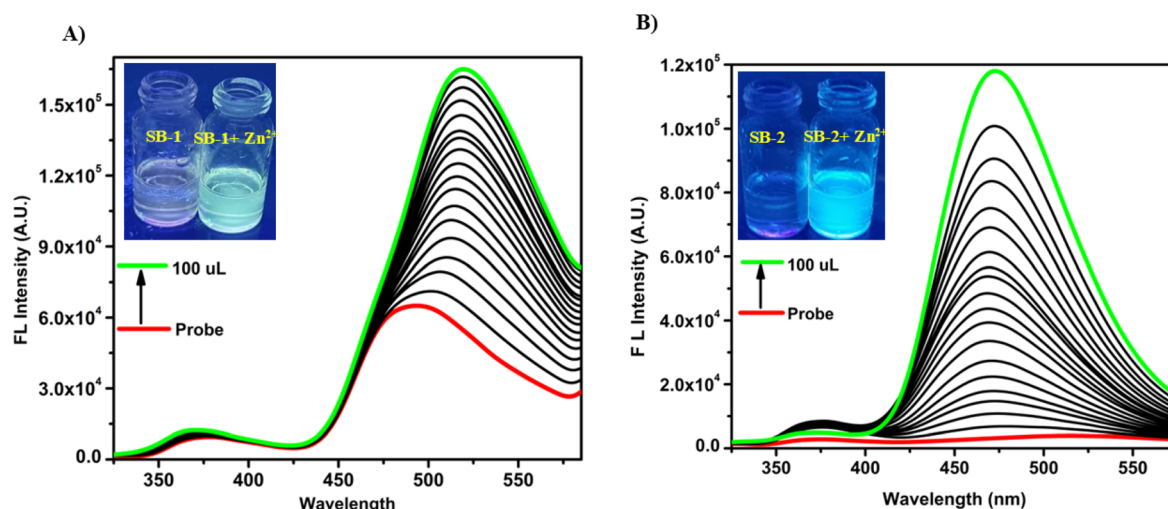


Fig. 3 Emission spectral changes of (A) **SB-1** (left,  $10^{-5}$  M) and (B) **SB-2** (right,  $10^{-5}$  M) upon the addition of  $Zn^{2+}$  ions in DMSO. (Inset: photographs captured in DMSO under UV light).



emitted a bright yellow fluorescence upon the addition of  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ , or  $\text{Zn}^{2+}$  ions. Among the two receptors, **SB-1** emerged as the more efficient fluorescent sensor for  $\text{Zn}^{2+}$ , as evidenced by its higher fluorescence enhancement and broader reactivity towards additional biologically and environmentally relevant ions like  $\text{Na}^+$ ,  $\text{K}^+$ , and  $\text{Mg}^{2+}$  (Fig. S24). This highlights its potential utility for multianalyte detection. We have systematically investigated the influence of different counter anions (such as  $\text{Cl}^-$ ,  $\text{NO}_3^-$ ,  $\text{SO}_4^{2-}$ , and  $\text{AcO}^-$ ) associated with  $\text{Zn}^{2+}$  on the fluorescence response of **SB-2**. The results revealed that the nature of the anion had negligible impact on the fluorescence behavior of **SB-2**. This suggests that the observed fluorescence enhancement is primarily driven by the coordination of the  $\text{Zn}^{2+}$  cation with the probe, and not significantly affected by its accompanying counter anion. Such stability toward varying anionic environments further demonstrates the robustness and reliability of **SB-2** as a selective  $\text{Zn}^{2+}$  sensor, even in complex media containing different zinc salts.

Achieving high selectivity and sensitivity in the presence of multiple metal ions is a key requirement for the practical application of fluorescent chemosensors. To explore this, **SB-1** and **SB-2** were systematically examined for their fluorescence responses toward a panel of biologically and environmentally relevant metal ions, including  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Al}^{3+}$ ,  $\text{Fe}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Hg}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Sn}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Cd}^{2+}$ ,  $\text{Cu}^{2+}$ , and  $\text{Zn}^{2+}$ . Fluorescence titration experiments were conducted using  $10^{-5}$  M solutions of **SB-1** and **SB-2** in DMSO to monitor changes in emission behaviour upon the addition of these metal ions (Fig. 4). The results showed that **SB-1** exhibited a clear and selective fluorescence enhancement in the presence of  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ , and  $\text{Zn}^{2+}$  ions. Notably, the fluorescence remained unaltered or only marginally affected when other metal ions were introduced, indicating strong specificity for these four analytes. This response was both rapid and stable, suggesting efficient coordination between **SB-1** and these target ions. In contrast, **SB-2**

displayed highly selective fluorescence activation exclusively with  $\text{Zn}^{2+}$  ions. No observable enhancement was detected with  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ , or any other tested metal ions, underscoring **SB-2**'s strong preference and exceptional selectivity for  $\text{Zn}^{2+}$ . This behavior was maintained even in the presence of excess amounts of potentially interfering ions, confirming the sensor's discrimination capability under competitive conditions (Fig. 5). Further, competitive studies of **SB-1** and **SB-2** with  $\text{Zn}^{2+}$  in the presence of other metal ions and results strongly suggested that the emission intensities remained stable, indicating saturation of binding interactions and does not influence the other metal ions. Together, these results highlight **SB-1** as a versatile sensor capable of detecting multiple metal ions ( $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ , and  $\text{Zn}^{2+}$ ), while **SB-2** stands out as a highly selective and sensitive probe for  $\text{Zn}^{2+}$ . These findings clearly indicate that, even in the presence of other interfering metal ions, the **SB-1** has high selectivity for  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Zn}^{2+}$ , and **SB-2** for  $\text{Zn}^{2+}$  ions. Such selective recognition profiles make these chemosensors promising tools for real-time metal ion detection in complex matrices.

### 3.3. $^1\text{H}$ NMR titration studies

To further investigate the coordination behavior of **SB-1** and **SB-2** with  $\text{Zn}^{2+}$  ions,  $^1\text{H}$  NMR titration studies were carried out in  $\text{DMSO}-d_6$ . Upon gradual addition of  $\text{Zn}^{2+}$  to **SB-2**, noticeable changes were observed in the chemical shifts of key proton signals (Fig. 6), and S25 indicating complex formation. The most prominent shift was seen in the imine proton ( $-\text{CH}=\text{N}-$ ), which moved downfield, reflecting its direct involvement in metal coordination. Concurrently, the disappearance of the phenolic  $-\text{OH}$  proton resonance at  $\delta = 13.60$  ppm strongly suggested deprotonation of the hydroxyl group upon binding to  $\text{Zn}^{2+}$ . These spectral alterations confirm that  $\text{Zn}^{2+}$  coordinates with **SB-2** through both the imine nitrogen and phenolic oxygen

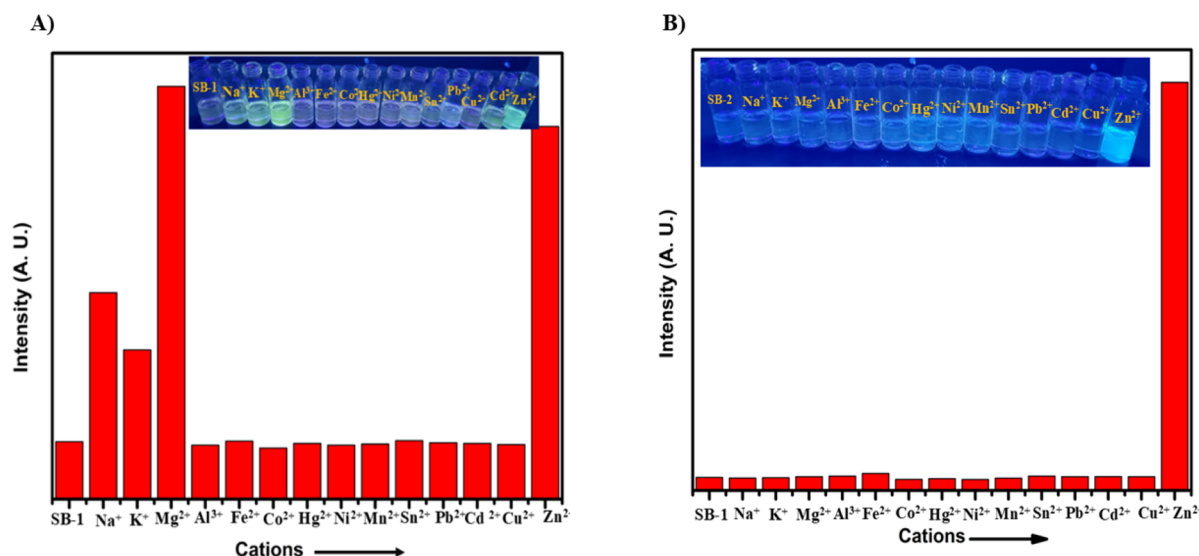


Fig. 4 Selectivity analysis of **SB-1** (A) and **SB-2** (B) ( $1 \times 10^{-5}$  M) toward various metal ions. (Inset: photographs captured in DMSO under UV light).

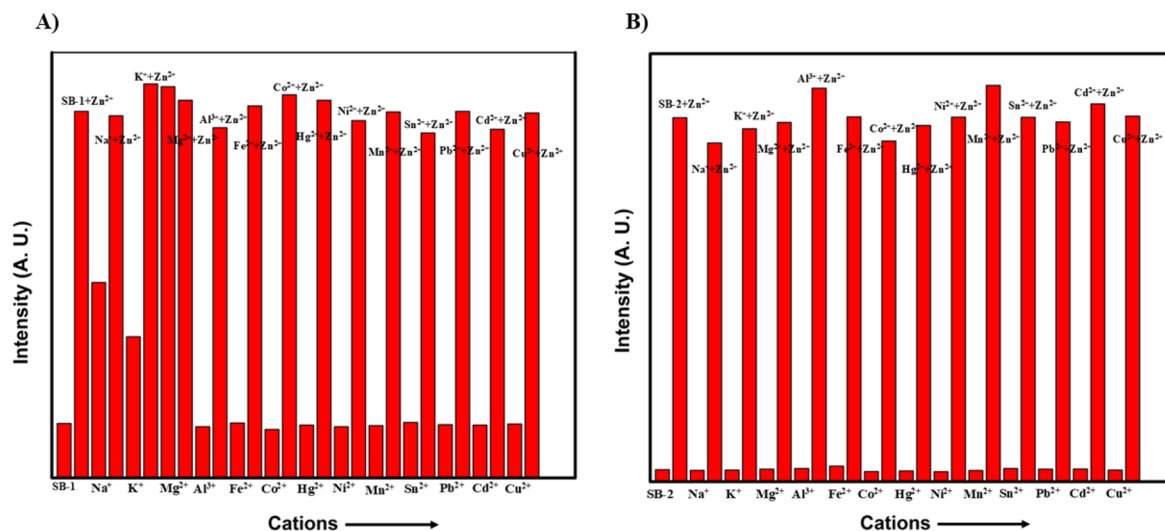


Fig. 5 Examination of competitive studies of SB-1 (A) and SB-2 (B) ( $1 \times 10^{-5}$  M) towards various metal ions. The competitive binding ability of CS-1 and CS-2 ( $1 \times 10^{-5}$  M) in the presence of other interfering metal ions and emission band was monitored at 520 nm.

atoms. The overall downfield migration of aromatic proton signals further supports the formation of a stable metal-ligand complex, likely accompanied by electronic reorganization within the aromatic system. Similar spectral behavior was observed during the titration of **SB-1** with  $\text{Zn}^{2+}$ , reinforcing a comparable binding mode for both receptors. These findings, derived from  $^1\text{H}$  NMR evidence, clearly demonstrate the strong and selective interaction between the ligands and  $\text{Zn}^{2+}$  ions.

### 3.4. DFT theoretical studies

Molecular orbital analysis provides useful information about the electronic structure and can be used to determine how receptors interact with the metal ions. Density functional theory (DFT) calculations for all compounds were performed using the Gaussian 09W program package. These calculations were carried out at the B3LYP level<sup>19</sup> for C, H, N, and O atoms,

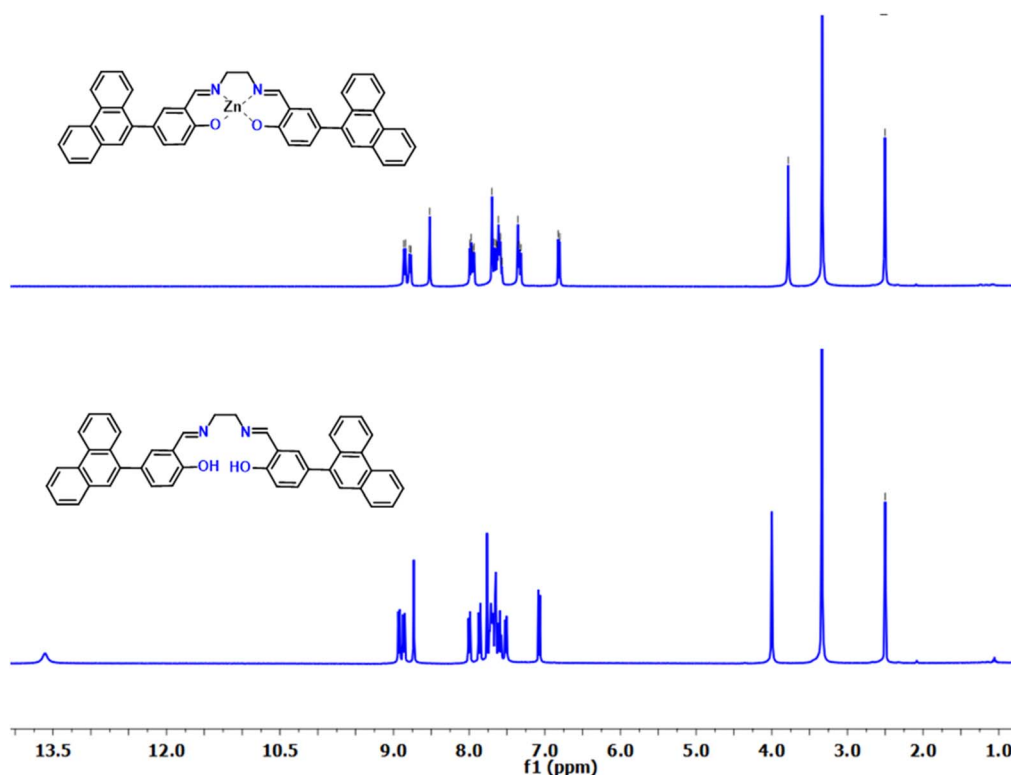


Fig. 6 Compared  $^1\text{H}$ NMR of SB-2 and SB-2 +  $\text{Zn}^{2+}$  (NMR titration in  $\text{DMSO}-d_6$ ).





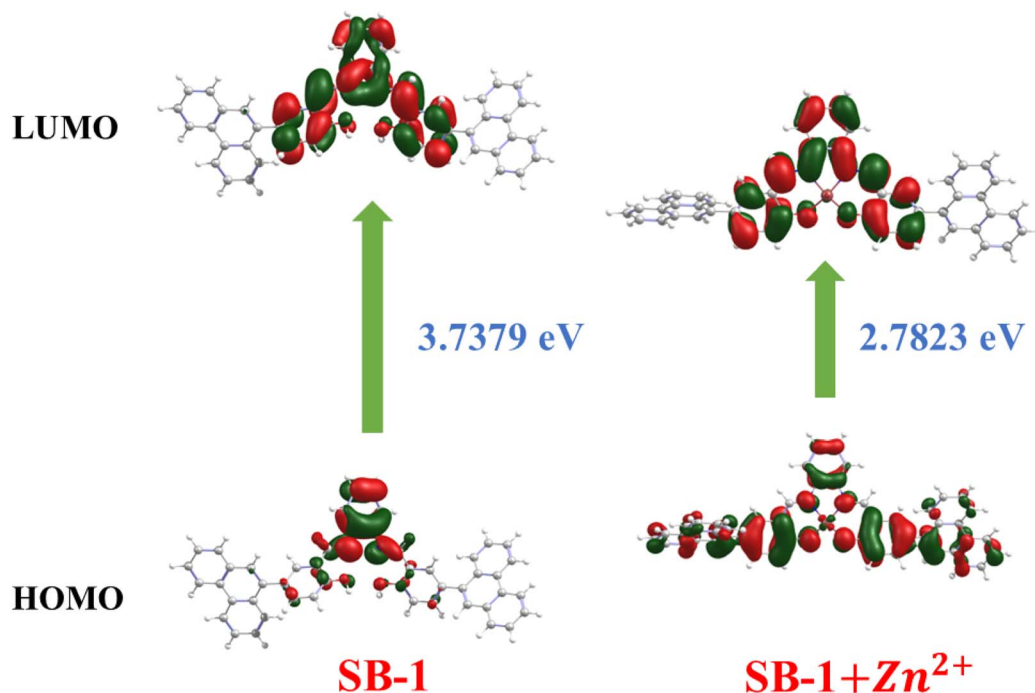


Fig. 7 Selected MOs of SB-1 and SB-1 +  $\text{Zn}^{2+}$  (not to scale; isocontour value = 0.02).

allowing for a comprehensive understanding of the electronic structure and metal-binding properties of the probes. The lowest unoccupied molecular orbital (LUMO), which shows the charge transfer from the aromatic unit to the imine moiety, is highly located on the entire  $\pi$ -moiety except the phenanthrene ring in SB-1 and while in SB-2, it extends over the entire molecule except for the ethane segment. The highest occupied molecular orbital (HOMO) is concentrated on the  $\pi$ -moiety of the diamino benzene unit in SB-1 and on one side of the molecule in SB-2 (Fig. 7 and S26). According to DFT calculations, the HOMO–LUMO energy gap of SB-1 and SB-2 was 3.74 and 4.38 eV, respectively which correlates with the experimental value. Upon the introduction of  $\text{Zn}^{2+}$  ions to the receptors, the

HOMO in SB-1 becomes localized over the entire  $\pi$ -moiety, whereas in SB-2, it is concentrated on one side of the molecule. The LUMO, on the other hand, is distributed across the  $\pi$ -system, excluding the phenanthrene ring, with minimal contribution from  $\text{Zn}^{2+}$  ions. This interaction leads to a significant reduction in the HOMO–LUMO energy gap, decreasing to 2.78 eV for SB-1 and 3.56 eV for SB-2. This reduction correlates with the emergence of a new absorption band ( $\sim 440$  nm) in the UV-visible spectrum of SB-1 +  $\text{Zn}^{2+}$  and SB-2 +  $\text{Zn}^{2+}$ . Fig. 7 illustrates the molecular structure and energy gap ( $E_g$ ) of SB-1 and its  $\text{Zn}^{2+}$  complex, while Fig. S26 presents the same for SB-2 and SB-2 +  $\text{Zn}^{2+}$ . These findings strongly support the idea that  $\text{Zn}^{2+}$  coordination with the receptors modulates intraligand

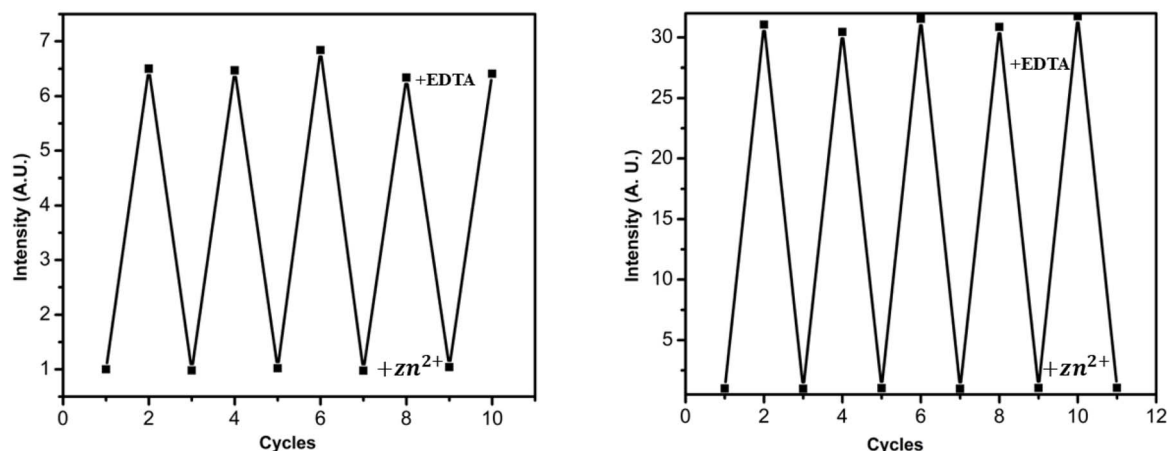


Fig. 8 Reversibility studies of SB-1 (left) and SB-2 (right) were conducted by monitoring fluorescence intensity at 520 nm over five regeneration cycles, with alternating additions of  $\text{Zn}^{2+}$  followed by EDTA.

charge transfer, leading to distinct photophysical changes. The colorimetric response of **SB-1** and **SB-2** upon zinc ion binding is well substantiated by absorption spectral shifts, with the observed bathochromic shift serving as direct evidence of the narrowing HOMO–LUMO energy gap.

### 3.5. Reversibility studies

For practical applications, a chemosensor must exhibit not only high sensitivity and selectivity but also excellent reversibility. To assess the regenerative capability of **SB-1** and **SB-2**, reversibility experiments were conducted using EDTA as a strong competing ligand. Upon the addition of 10 equivalents of EDTA to the **SB-1**·**Zn**<sup>2+</sup> and **SB-2**·**Zn**<sup>2+</sup> complexes in DMSO, a rapid quenching of fluorescence was observed, restoring the emission profile of the free receptors (Fig. 8). This clearly indicates that EDTA effectively sequesters **Zn**<sup>2+</sup> from the sensor–metal complex due to its stronger binding affinity, thereby regenerating the original receptor. These results confirm that both **SB-1** and **SB-2** exhibit excellent reversibility and can be efficiently recycled through simple EDTA treatment. Notably, the receptors maintained their structural and optical integrity over multiple cycles, with minimal variation in absorption and emission intensity observed across at least five regeneration cycles (Fig. 8). This high stability and reversibility highlight the practical utility of **SB-1** and **SB-2** as reusable fluorescent sensors for **Zn**<sup>2+</sup> detection (Fig. 8).

## 4. Conclusion

We developed and characterized two Schiff base-based chemosensors, **SB-1** and **SB-2**, using NMR and ESI-HRMS. Their sensing properties in DMSO were studied *via* UV-visible and fluorescence spectroscopy. **SB-1** selectively responded to Na<sup>+</sup>, K<sup>+</sup>, Mg<sup>2+</sup>, and **Zn**<sup>2+</sup> ions with a distinct colour change and fluorescence enhancement, while **SB-2** showed high fluorescence selectivity for **Zn**<sup>2+</sup> alone. These responses are attributed to intraligand charge transfer and suppression of –C=N– isomerization, PET, and ESIPT processes upon metal coordination. Job's plot confirmed 1 : 1 binding stoichiometry, and <sup>1</sup>H NMR titrations supported the proposed coordination *via* imine nitrogen and phenolic oxygen. Both sensors demonstrated excellent reversibility with EDTA and maintained stability over multiple cycles. **SB-1** showed superior sensitivity for **Zn**<sup>2+</sup>, with lower detection limits and higher binding constants than **SB-2**. Overall, **SB-1** and **SB-2** are effective, selective, and reversible chemosensors for biologically relevant metal ions.

## Conflicts of interest

The authors declare no competing financial interest.

## Data availability

The data underlying this study are available in the published article and its SI.

<sup>1</sup>H NMR, <sup>13</sup>C NMR, HRMS, and characterization data for all molecules, photophysical data, DFT and TD-DFT results (PDF). See DOI: <https://doi.org/10.1039/d5ra03617h>.

## Acknowledgements

CAS P thanks, SERB/EEQ/2021/000180 for funding and support. The authors acknowledge the Centre for Materials Characterisation NIT Calicut for NMR, HRMS (DST-FIST) and Centre for Computational Modelling and Simulations NIT Calicut. The authors acknowledge NMR Facility (Supported by DST-FIST) by Gandhigram rural institute.

## References

- (a) K. P. Carter, A. M. Young and A. E. Palmer, *Chem. Rev.*, 2014, **114**, 4564–4601; (b) C. Suksai and T. Tuntulani, *Chem. Soc. Rev.*, 2003, **32**, 192–202; (c) R. Martinez-Manez and F. Sancenon, *Chem. Rev.*, 2003, **103**, 4419–4476; (d) P. D. Beer and J. Cadman, *Coord. Chem. Rev.*, 2000, **205**, 131–155.
- (a) M. Li, Q. Shi, N. Song, Y. Xiao, L. Wang, Z. Chen and T. D. James, *Chem. Soc. Rev.*, 2023, **52**, 5827–5860; (b) A. Afrin and P. C. A. Swamy, *Coord. Chem. Rev.*, 2023, **494**, 215327.
- (a) R. Rodriguez, S. Müller, L. Colombeau, S. Solier, F. Sindikubwabo and T. Cañeque, *Chem. Rev.*, 2025, **125**, 660–744; (b) J. W. Erdman Jr, I. A. Macdonald, and S. H. Zeisel, *Present Knowledge in Nutrition*, John Wiley & Sons, 2012.
- (a) I. S. Joung and T. E. Cheatham, *J. Phys. Chem. B*, 2008, **112**, 9020–9041; (b) P. Mukhopadhyay, L. Monticelli and D. P. Tieleman, *Biophys. J.*, 2004, **86**, 1601.
- (a) V. Gerke, C. E. Creutz and S. E. Moss, *Nat. Rev. Mol. Cell Biol.*, 2005, **6**, 449–461; (b) J. M. Berg and Y. Shi, *Science*, 1996, **271**, 1081–1085; (c) Y. Chen, Y. Bai, Z. Han, W. He and Z. Guo, *Chem. Soc. Rev.*, 2015, **44**, 4517–4546.
- (a) M. S. Kumar, S. Pakrashi, S. Manna, S. M. Choudhury, B. Das, A. Ghosh, A. H. Seikh, M. Dolai and A. K. Das, *Anal. Methods*, 2025, **17**, 2125–2133; (b) M. V. Karmegam, S. Karuppannan, D. B. C. Leslee, S. Subramanian and S. Gandhi, *Luminescence*, 2019, **35**, 1–8; (c) H. Kim, D. Gil and C. Kim, *J. Chin. Chem. Soc.*, 2022, **69**, 856–863; (d) N. Li, W. Tang, Y. Xiang, A. Tong, P. Jin and Y. Ju, *Luminescence*, 2010, **25**, 445–451.
- (a) S. C. Burdette and S. J. Lippard, *Coord. Chem. Rev.*, 2001, **216**, 333–361; (b) C. J. Frederickson, *Int. Rev. Neurobiol.*, 1989, **31**, 145–238; (c) J.-Y. Lee, T. B. Cole, R. D. Palmiter, S. W. Suh and J.-Y. Koh, *Proc. Natl. Acad. Sci. U. S. A.*, 2002, **99**, 7705–7710; (d) D. W. Choi and J. Y. Koh, *Annu. Rev. Neurosci.*, 1998, **21**, 347–375.
- (a) P. Jiang and Z. Guo, *Coord. Chem. Rev.*, 2004, **248**, 205–229; (b) X. Li, X. Gao, W. Shi and H. Ma, *Chem. Rev.*, 2014, **114**, 590–659; (c) R. Mondal, A. Shanmughan, A. Murugeswari and S. Shanmugaraju, *Chem. Commun.*, 2023, **59**, 11456–11468.



- 9 (a) X. Zheng, W. Cheng, C. Ji and M. Yin, *Rev. Anal. Chem.*, 2020, **39**, 231–246; (b) K. P. Carter, A. M. Young and A. E. Palmer, *Chem. Rev.*, 2014, **114**, 4564–4601.
- 10 (a) D. Fiorentini, C. Cappadone, G. Farruggia and C. Prata, *Nutrients*, 2021, **13**, 1136; (b) *Present Knowledge in Nutrition*, ed. J. W. Erdman Jr, I. A. Macdonald and Zeisel, S. H., John Wiley & Sons, 2012; (c) *Encyclopedia of dietary supplements*, ed. P. M. Coates, J. M. Betz, M. R. Blackman, G. M. Cragg, M. Levine, J. Moss, J. D. White, CRC Press, 2012.
- 11 (a) H. Tapiero, D. M. Townsend and K. D. Tew, *Biomed. Pharmacother.*, 2003, **57**, 386–398; (b) T. Chopra, S. Sasan, L. Devi, R. Parkesh and K. K. Kapoor, *Coord. Chem. Rev.*, 2022, **470**, 214704; (c) M. Kaykhaili, M. Khajeh and S. H. J. Hashemi, *Anal. Chem.*, 2015, **70**, 1325–1329; (d) X. Chen, M. J. Jou, H. Lee, S. Kou, J. Lim, S. W. Nam, S. Park, K. M. Kim and J. Yoon, *Sens. Actuators, B*, 2009, **137**, 597–602; (e) T. T. T. Vo, T. Y. Peng, T. H. Nguyen, T. N. H. Bui, C. S. Wang, W. J. Lee, Y. L. Chen, Y. C. Wu and I. T. Lee, *Cell Commun. Signaling*, 2024, **22**, 353.
- 12 K. Jomova, M. Makova, S. Y. Alomar, S. H. Alwasel, E. Nepovimova, K. Kuca, C. J. Rhodes and M. Valko, *Chem.-Biol. Interact.*, 2022, **1**, 110–173.
- 13 (a) P. Pohl, *Trends Anal. Chem.*, 2009, **28**, 117–128; (b) A. T. Townsend, K. A. Miller, S. McLean and S. Aldous, *J. Anal. At. Spectrom.*, 1998, **13**, 1213–1219; (c) P. Verma, N. Kalra and S. Verma, *Microchem. J.*, 2024, **205**, 111293.
- 14 R. Govindaraju, R. Govindaraju, S. Govindaraju, K. Yun and J. Kim, *Biosensors*, 2023, **13**, 1008.
- 15 (a) B. K. Ran and S. A. John, *J. Photochem. Photobiol., A*, 2021, **418**, 113372; (b) J. Wu, W. Liu, J. Ge, H. Zhang and P. Wang, *Chem. Soc. Rev.*, 2011, **40**, 3483–3495; (c) R. A. Bissell, A. P. de Silva, H. Q. N. Gunaratne, P. L. M. Lynch, G. E. M. Maguire and K. R. A. S. Sandanayake, *Chem. Soc. Rev.*, 1992, **21**, 187–195; (d) A. P. De Silva, H. Q. N. Gunaratne, T. Gunnlaugsson, A. J. M. Huxley, C. P. McCoy, J. T. Rademacher and T. E. Rice, *Chem. Rev.*, 1997, **97**, 1515–1566.
- 16 (a) A. V. Verma, A. P. Singh and R. Shrivastava, *J. Mol. Struct.*, 2024, **1295**, 136549; (b) I. Chaturvedi, S. Vyas and R. Mishra, *Dyes Pigm.*, 2025, **239**, 112748; (c) A. Afrin, J. Anjitha, M. S. Gayathria and P. C. A. Swamy, *Sens. Diagn.*, 2023, **2**, 988–1076; (d) M. K. Goshisht, G. K. Patra and N. Tripathi, *Mater. Adv.*, 2022, **3**, 2612–2669; (e) B. Mohan, J. Kapoor, A. Shanmughan, M. K. Noushija, M. Sumithradevi and S. Shanmugaraju, *Res. Chem.*, 2024, **7**, 101402.
- 17 S. Aslam, I. Kousar, S. Rani, W. Altaf, S. Bristy and R. Skouta, *Molecules*, 2025, **30**, 1450.
- 18 D. Udhayakumari, *Talanta*, 2024, **278**, 126536.
- 19 M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. V. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery Jr, J. E. Peralta, F. Ogliaro, M. J. Bearpark, J. J. Heyd, E. N. Brothers, K. N. Kudin, V. N. Staroverov, T. A. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. P. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman and D. J. Fox, *Gaussian 09, Revision B.01*, Gaussian, Inc., Wallingford, CT, 2016.
- 20 A. Afrin, P. Chinna Ayya Swamy and R. Angel, *Anal. Methods*, 2024, (16), 6323–6336.

