RSC Advances



PAPER

View Article Online
View Journal | View Issue



Cite this: RSC Adv., 2025, 15, 13214

investigations of new pyrazolone chalcones†

Synthesis, molecular docking, and biological

Ahmed A. Noser, ** Maha M. Salem, ** Esraa M. ElSafty, ** Mohamed H. Baren, ** Adel I. Selim* and Hamada S. A. Mandour**

Heterocyclic compounds are essential to the drug development and discovery processes. Herein, we synthesized new pyrazolone chalcones (3a-g) through the reaction of azopyrazolone (2) with different aromatic aldehydes in a basic medium. Numerous techniques including elemental analysis, 1H -NMR, ^{13}C -NMR, and FT-IR spectroscopies, were used to characterize pyrazolone chalcone derivatives. Compound 3b exhibited the highest binding energy towards YAP/TEAD protein with a value of -8.45 kcal mol $^{-1}$ in *in silico* studies. This observation suggested that compound 3b inhibits the YAP/TEAD Hippo signaling pathway. In addition, compound 3b offered a prospective anticancer effect against various cancer cell lines, such as HepG-2, MCF-7, and HCT-116, among the other synthesized compounds, with IC $_{50}$ values equal to 5.03 ± 0.4 , 3.92 ± 0.2 , and $6.34 \pm 0.5~\mu$ M, respectively. These results validated our findings regarding the *in silico* suppression of the YAP/TEAD protein. Its pharmacokinetic properties were theoretically observed using ADMET. Additionally, compound 3b demonstrated a potent antioxidant scavenging action (*in vitro*) against DPPH free radicals. Thus, based on our findings, compound 3b may act as a potential anticancer scaffold owing to its inhibitory impact towards the YAP/TEAD-mediated Hippo signaling pathway with a safe toxic profile on normal cells.

Received 20th February 2025 Accepted 10th April 2025

DOI: 10.1039/d5ra01233c

rsc.li/rsc-advances

Introduction

Cancer is a worldwide civic health concern, and its diagnosis in early stage is of great importance.1 Currently, the primary treatment strategies for cancer are chemotherapy, surgery, and targeted therapy. Targeted therapy has become a standard treatment for cancer patients and has significantly increased the efficacy of cancer therapy.2 According to a previous report, regulating organ growth and inhibiting tumors depend on the Hippo signaling target pathway's control over cell division and proliferation.3 Because of its significant influence on patient prognosis and chemotherapeutic drug resistance, the Hippo pathway is a desirable therapeutic target.4 Pyrazoles have attracted significant attention owing to their effective pharmaceutical properties such as antibiotic, antifungal, antibacterial, pesticidal, antioxidant, anti-inflammatory, antiviral, and antitumor effects.⁵⁻¹¹ Chalcones are α,β-unsaturated ketones with a broad range of pharmacological actions, making them

interesting synthons and bioactive scaffolds with great therapeutic potential. It is commonly known that they have significant biological effects such as antibacterial, antioxidant, and anticancer effects. ¹²⁻¹⁵ Chalcones facilitate the synthesis of novel heterocyclic compounds including pyrimidinones, azepines, pyridine, and pyran, which have strong biological activities. ^{16,17} There have been numerous reports on marketed anticancer drugs containing pyrazolone and a chalcone core, such as metochalcone, sofalcone, butein, metamizole, and propyphenazone, ¹⁸⁻²² as shown in Fig. 1.

The goal of our study was to develop a new class of pyrazolone chalcone derivatives, examine their radical scavenging activities and investigate their anticancer effects by inhibiting the YAP/TEAD Hippo signaling pathway (*in silico*). our findings confirmed their impact on cancer cell lines (*in vitro*). Additionally, absorption–distribution–metabolism–excretion–toxicity (ADMET) pharmacokinetic features were used to assess their drug-likeness and bioavailability.

Results and discussion

Chemistry of the novel compounds

Scheme 1 describes the reaction of ethyl cyanoacetate with phenyl hydrazine, which leads to the formation of compound 1, as described previously by Weissberger.²³ Further modification of compound 1 with *p*-acetyl phenyl diazonium chloride offered azo pyrazolone 2 with 74% yield (Scheme 1). The chemical

^{*}Organic Chemistry, Chemistry Department, Faculty of Science, Tanta University, Tanta 31527, Egypt. E-mail: ahmed.nosir@science.tanta.edu.eg; esraa_elsafty@science.tanta.edu.eg; chemistnoser2010@yahoo.com; adel.saleem@science.tanta.edu.eg; hamada.mandour@science.tanta.edu.eg

^bBiochemistry Division, Chemistry Department, Faculty of Science, Tanta University, Tanta 31527, Egypt. E-mail: maha_salem@science.tanta.edu.eg

[†] Electronic supplementary information (ESI) available: Materials and instruments used during the synthesis and the spectral data of the new compounds. See DOI: https://doi.org/10.1039/d5ra01233c

Fig. 1 Marketed pyrazolone- and chalcone-based drugs.

Scheme 1 Synthesis route for compound 2.

structures of compounds 3a-g were established according to their elemental analysis and different spectral data. Elemental analysis was used to indicate the exact ratio of C, H, and N. Fourier transform infrared (FT-IR) spectroscopy was used to

illustrate the presence of the main functional groups. Nuclear magnetic resonance (NMR) was used to indicate the different types of hydrogen and carbon atoms in the synthesized compounds. The FT-IR analysis of compound 2 showed

Scheme 2 Synthesis route for compounds 3a-g.

different absorption bands at 1600 cm⁻¹ and 1540 cm⁻¹ due to C=O and N=N groups, respectively. Its ¹H NMR spectrum showed a new singlet signal at δ 2.20 ppm attributed to the CH₃ group.

Modifying azo compound (2) with a different aromatic aldehyde in a basic medium led to the formation of pyrazolone chalcone 3a-g (Scheme 2).24 The FT-IR analysis of chalcones (3a-g) showed absorption bands at 1610–1670 cm⁻¹, 1520– 1600 cm⁻¹, and 1500-1560 cm⁻¹, for C=O, C=N, and N=N groups. Compound 3b-d showed absorption bands at 3100-3310 cm⁻¹ for the OH group. The ¹H NMR spectrum of compound 3a revealed two new signals resonated at δ 7.54, and 7.68 ppm attributed to two olefinic protons. Compound 3b showed four new signals at δ 7.00, 7.40, 3.86, and 2.08 ppm attributed to two olefinic protons, OCH3 protons, and OH proton, respectively. Additionally, compounds 3c and 3d showed three new signals at δ 6.92–7.45, 7.55–7.56, and 10.12– 10.26 ppm attributed to two olefinic protons and OH proton, respectively. Finally, the ¹H NMR spectrum of compound 3g revealed four new signals resonated at δ 7.62, 7.68, 8.02, and 8.07 ppm attributed to olefinic protons.

The suggested mechanism for the synthesis of compounds 3a-g is illustrated in Scheme 3.

Antioxidant capacity

Fig. 2 and 3 show that the antioxidant activity enhanced with the increase in chalcone concentration using the stable DPPH radical. Compared to normal L-ascorbic acid (IC₅₀ = 16.81 \pm 0.10 µM), compound 3b exhibited the greatest DPPH scavenging activity with an IC₅₀ value of 19.95 \pm 0.14 μ M. Compounds 3a and 3f exhibited a strong antioxidant activity with IC₅₀ values of 21.40 \pm 0.17 and 27.25 \pm 0.19 μ M. Moreover, compounds 3e and 3d had moderate effects on scavenging with the activity of 32.79 \pm 0.21 and 48.59 \pm 0.28 μ M, respectively, while compounds 3g and 3c showed a limited ability to quench free radicals with IC $_{50}$ values of 73.63 \pm 0.39 and 94.18 \pm 0.45 μM, respectively.

Docking studies

For creating anticancer drugs, the novel chalcone ligands 3a-g were docked into YAP/TEAD, a well-known and alluring therapeutic target protein in the Hippo signaling pathway. Numerous studies in the fields of oncology and regenerative medicine have examined the Hippo pathway, and these studies have also suggested that the route may be important as a regulatory variable in human biology or as an inhibitory target for drug

Scheme 3 Suggested mechanism for compounds 3a-g.

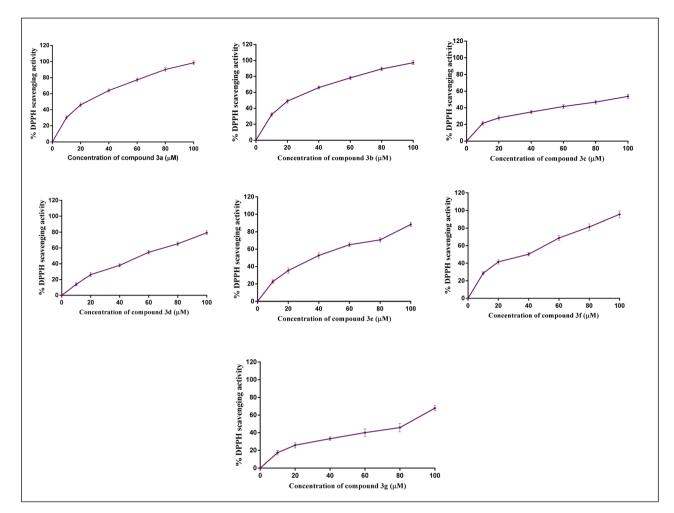


Fig. 2 Antioxidant scavenging activity of chalcone derivatives (3a-g). Results are reported as mean \pm SE; where n=3.

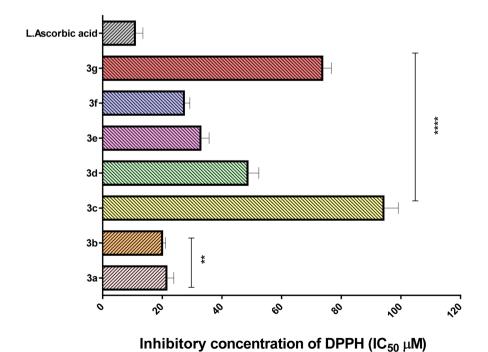


Fig. 3 Inhibitory concentrations (IC₅₀) of chalcones 3a-g via DPPH. Results are reported as mean \pm SE, where n=3.

Table 1 Docking scores of compounds 3a-g with the target protein

	YAP/TEAD target	Hippo signaling protein
Compounds	Docking score $(\Delta G_{ m bind})$	Docked complex (amino acid-ligand) interactions
3a	−7. 86	H-acceptor
		LYS344
		CYS367
		Electrostatic interactions
		PHE229
		LLE395
		MET370
_		LEU366
3b	-8.45	H-donor
		PRO365
		π-Hydrogen
		LYS344
		VAL345
		Electrostatic interactions
		LEU366
		MET370
		CYS367
		LLE395
		GLU343 CYS367
3c	-6.11	Electrostatic interactions
30	-0.11	CYS367
		LEU366
		MET370
		LYS344
		LLE395
3d	-7.11	H-acceptor
o u	7.11	LYS344
		Electrostatic interactions
		CYS367
		THR332
		PHE229
		LLE395
		PHE415
		VAL342
3e	-7.13	H-acceptor
		LYS344
		Electrostatic interactions
		CYS367
		THR332
		PHE229
		LLE395
		PHE415
- 6		VAL342
3f	-7.22	H-acceptor
		LYS344
		Electrostatic interactions
		CYS367 VAL342
		VAL342 LLE395
		MET370
		LEU366
3g	-6.09	Electrostatic interactions
95	-0.03	LEU366
		GLU368
		VAL342
		CYS367
		LYS344
		LLE395

development. Generally, the Hippo pathway consists of two constitutive modules: the upstream serine/threonine kinase cascade, which comprises MST1/2 and LATS1/2, and the downstream transcriptional modules, which mainly include the YAP/TEAD complex.²⁵

The YAP/TEAD genetic and pharmacological interaction significantly decreased carcinogenesis in YAP-dependent cancer models.26 As a result, the YAP/TEAD complex has become a desirable target for the creation of anti-cancer medications.²⁷ Table 1 and Fig. 4 summarize all new chalcone interactions with the target YAP/TEAD protein. According to our findings, compound 3b had the highest binding $(-8.45 \text{ kcal mol}^{-1})$ against the target YAP/TEAD protein. Compound 3a had high binding energy $(-7.86 \text{ kcal mol}^{-1})$ with the target protein. With binding energies of -7.11, -7.13, and -7.7.22 kcal mol⁻¹, respectively, compounds 3d, 3e, and 3f also demonstrated a mild inhibitory effect. Conversely, compounds 3c and 3g showed low binding energies of −6.11 and -6.09 kcal mol⁻¹, respectively. Compound 3b was therefore highly recommended for use as an anticancer agent because of its potential to inhibit the YAP/TEAD mediated Hippo signaling pathway.

ADMET features

The bioavailability and drug-like characteristics of the studied chalcone derivatives (3a-g) were determined, and are shown in Fig. 5. With a total polar surface area (TPSA) ranging from 102.25 to 149.35 Å^2 , the compounds 3a-g met all the requirements for exceptional permeability and had sufficient oral bioavailability. To further demonstrate flexibility, they also used rotatable bonds between 0 and 10. They were more soluble in cellular membranes because their hydrogen-bond accepted (HBA) and hydrogen-bond donated (HBD) values fell within the fulfilled range. Table 2 demonstrates that octanol/water partition coefficient $(\log p)$ values less than 5 were indicative of good lipophilicity features. The chalcone derivatives also showed a higher percentage of human intestinal absorption (HIA) ratings in accordance with the ADMET criteria, indicating that the human intestine could absorb them more effectively. The studied chalcones have a great safety profile for the central nervous system (CNS) since they do not penetrate the blood-brain barrier. They are safe because every AMES toxicity and carcinogenicity test result was negative.

In vitro anticancer study

The antitumor effect of chalcone derivatives (3a–g) on HCT-116, HepG2, MCF-7, and WI-38 normal cell lines was assessed in this study using the MTT assay (Fig. 6). Compound 3b demonstrated noteworthy antitumor effects on the cancer cell lines HepG2, MCF-7, and HCT-116, with IC $_{50}$ values of 5.03 \pm 0.4, 3.92 \pm 0.2, and 6.34 \pm 0.5 μ M, respectively. Compounds 3a and 3f demonstrated impressive antitumor effects with HCT-116, HepG2, and MCF-7 cell lines. Compounds 3e and 3d demonstrated a moderate effect on all cancer cell lines. Conversely, Compounds 3g and 3c demonstrated a weak effect

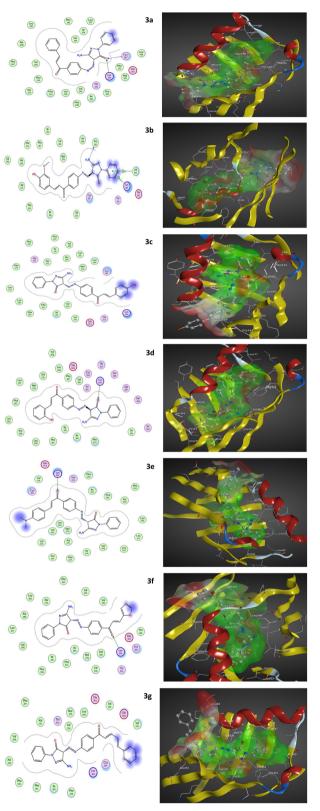


Fig. 4 Two- and three-dimensional molecular interaction networks for all chalcone derivatives (3a-g) with the target YAP/TEAD-mediated Hippo signaling pathway.

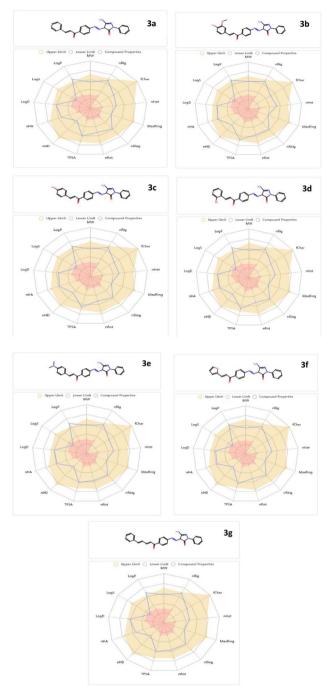


Fig. 5 Bioavailability radar plot for chalcones (3a-g).

on all cancer cell lines when compared to the reference chemotherapeutic drug (DOX). Moreover, all new derivatives (3a-g) showed lower cytotoxic effects on WI-38 normal cells except 3c. Our finding was in line with the previous study.²⁴ This suggests that compound 3b worked against proliferative cancer by blocking the Hippo signaling pathway mediated by YAP/TEAD.

Table 2 ADMET assets

	Molecular weight (g mol ⁻¹)	Blood-brain barrier (BBB)	% human intestinal absorption (HIA+)	TPSA A^2	$\log p$	nHA	nHD	N rotatable	AMES toxicity	Carcinogenicity
3a	409.15	No	89.6	100.48	3.39	7	2	6	Nontoxic	Noncarcinogenic
3b	455.16	No	92.1	129.94	2.98	9	3	7	Nontoxic	Noncarcinogenic
3 c	425.15	No	84.3	120.71	3.08	8	3	6	Nontoxic	Noncarcinogenic
3d	425.15	No	75.9	120.71	3.43	8	3	6	Nontoxic	Noncarcinogenic
3e	452.20	No	83.4	103.72	3.61	8	2	7	Nontoxic	Noncarcinogenic
3f	399.13	No	79.1	113.62	2.90	8	2	6	Nontoxic	Noncarcinogenic
3g	435.17	No	77.3	100.48	3.47	7	2	7	Nontoxic	Noncarcinogenic

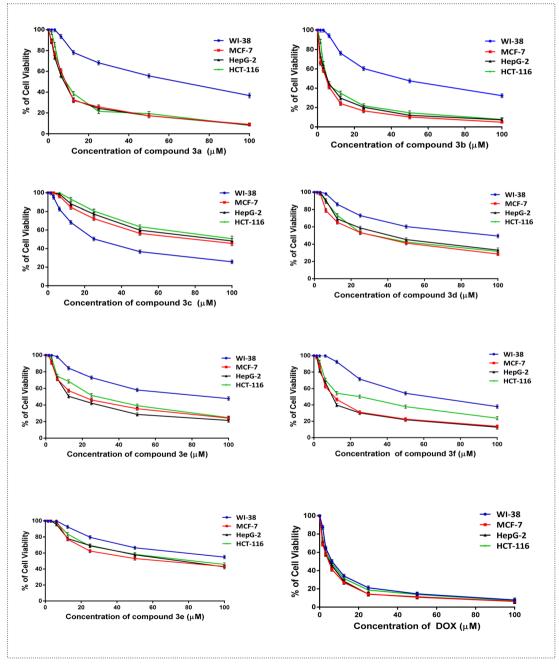


Fig. 6 Antitumor and cytotoxic impacts of chalcones 3a-g. Results are reported as mean \pm SE of three independent triplicates.

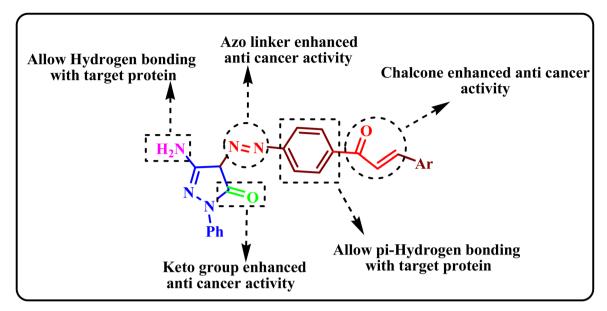


Fig. 7 SAR of the most active compound.

Structure-anti-tumor activity relationship (SAR)

The synthesized chalcones demonstrated their anticancer effect for the following reasons (Fig. 7):

- The presence of different substituents in the aryl group enhanced cytotoxic activity against different cancer cell lines (HepG-2, MCF-7, and HCT-116).
- The presence of hetero atoms such as nitrogen as well as carbonyl groups in the chemical structure of the pyrazolone chalcones enhanced the value of binding energy through the interaction between the pyrazolone chalcone and the target protein *via* hydrogen bonding.²⁸
- The azo linker enhanced the hydrogen bonding with the target protein 29.
- Compound 3b was considered the most active compound due to the presence of the phenyl group and olefinic protons, which enhanced the hydrogen bonding and π - π interaction with the target protein and, hence, increased the value of the binding energy.³⁰

Conclusion

We synthesized and characterized new pyrazolone chalcones (3a-g) using various spectral data. Among all the synthesized compounds, chalcone 3b demonstrated the strongest antioxidant activity. Furthermore, it has a high docking score because of hydrogen bonds, hydrophobic interactions, and electrostatic interactions with important residues in the binding pocket of the YAP/TEAD target protein. For that, molecular docking suggested that it may be a targeted anticancer agent. Additionally, the *in vitro* anti-cancer effect against different cancer cell lines was explained by the suppressive effect of chalcone 3b, which inhibited the YAP/TEAD-mediated Hippo signaling pathway to cause apoptosis and obstruct cell growth and survival. According to these results, chalcone 3b may be a newly targeted anticancer scaffold that targets a variety of cancer cells.

Experimental section

Materials and instrumentation

Section S1 in the ESI† includes the data of chemicals and instrumentation.

Synthesis of 3-amino-1-phenyl-1*H*-pyrazol-5(4*H*)-one (1)

Weissberger lab was previously preparing compound 1 (ref. 23)

Synthesis of 4-((4-acetylphenyl)diazenyl)-3-amino-1-phenyl-1*H*-pyrazol-5(4*H*)-one (2)

A sodium nitrite solution (12.7 mmol) was added slowly to *p*-amino acetophenone (13.7 mmol) in concentrated HCl. The resulting diazonium chloride was added to a solution of compound **1** (8.5 mmol) in pyridine. Then, the product was filtered and dried.

Orange powder; yield 74%; mp 89–91 °C; ¹H NMR (400 MHz, DMSO-d₆) δ (ppm): 9.14 (s, 2H, NH₂), 7.08–8.11 (m, 9H, Ar–H), 2.59 (s, 1H, CH–pyrazolone), 2.20 (s, 3H, CH₃); IR (KBr) ν : 1600 (CO), 1540 (N=N); anal. calcd for $C_{17}H_{15}N_5O_2$ (321.33): C, 63.54%; H, 4.71%; N, 21.79%. Found: C, 63.34%; H, 4.57%; N, 21.67%.

General procedure for the synthesis of pyrazolone chalcones (3a-g)

An ethanolic solution of compound 2 (10.0 mmol), aromatic aldehyde (10.0 mmol), and sodium hydroxide (20.0 mmol) was stirred at 25 $^{\circ}$ C for 10 h, put into freezing water, filtered, and then dried.

5-Amino-4-((*E*)-(4-cinnamoylphenyl)diazenyl)-2-phenyl-2,4-dihydro-3*H*-pyrazol-3-one (3a). Orange powder; yield 76%; mp 228–230 °C; 1 H NMR (400 MHz, DMSO-d₆) δ (ppm): 10.55 (s, 2H, NH₂), 7.68 (d, 1H, =CH), 7.54 (d, 1H, COCH=), 6.54–8.40 (m, 14H, Ar-H), 2.60 (s, 1H, CH-pyrazolone); 13 C NMR (101 MHz, DMSO-d₆) δ (ppm): 171.72 (C=O chalcone), 168.04 (C=O

Paper

pyrazolone), 146.95 (C=N pyrazolone), 113.20–133.22 (Ar–C), 127.60, 135.25 (C–olefinic), 74.43 (CH–pyrazolone); IR (KBr) ν : 3405 (NH₂), 1650 (C=O), 1597 (C=N), 1550 (N=N); anal. calcd for C₂₄H₁₉N₅O₂ (409.15): C, 70.40%; H, 4.68%; N, 17.10%. Found: C, 70.25%; H, 4.46%; N, 17.02%.

5-Amino-4-((*E*)-(4-((*E*)-3-(4-hydroxy-3-methoxyphenyl) acryloyl)phenyl)diazenyl)-2-phenyl-2,4-dihydro-3*H*-pyrazol-3-one (3b). Orange powder; yield 84%; mp 103–105 °C; ¹H NMR (400 MHz, DMSO-d₆) δ (ppm): 9.76 (s, 2H, NH₂), 7.40 (d, 1H, = CH), 7.00 (d, 1H, COCH=), 6.89–8.17 (m, 12H, Ar–H), 3.86 (s, 3H, OCH₃), 2.61 (s, 1H, CH–pyrazolone), 2.08 (s, 1H, OH); ¹³C NMR (101 MHz, DMSO-d₆) δ (ppm):153.80 (C=O chalcone), 150.94 (C=O pyrazolone), 149.03 (C=N pyrazolone), 145.49 (C-OH), 111.54–130.62 (Ar–C), 122.71, 130.93 (C–olefinic), 70.29 (CH–pyrazolone), 55.98 (OCH₃); IR (KBr) ν : 3400 (NH₂), 3310 (OH), 1610 (C=O), 1600 (C=N), 1520(N=N); anal. calcd for C₂₅H₂₁N₅O₄ (455.16): C, 65.93%; H, 4.65%; N, 15.38%. Found: C, 65.77%; H, 4.47%; N, 15.26%.

5-Amino-4-((*E*)-(4-((*E*)-3-(4-hydroxyphenyl)acryloyl)phenyl) diazenyl)-2-phenyl-2,4-dihydro-3*H*-pyrazol-3-one (3c). Brownish orange powder; yield 83%; mp 148–150 °C; 1 H NMR (400 MHz, DMSO-d₆) δ (ppm): 10.12 (s, 1H, OH), 9.79 (s, 2H, NH₂), 7.56 (d, 1H, =CH), 6.92 (d, 1H, COCH=), 6.84–8.21 (m, 13H, Ar–H), 2.59 (s, 1H, CH–pyrazolone); 13 C NMR (101 MHz, DMSO-d₆) δ (ppm):163.65 (C=O chalcone), 160.64 (C=O pyrazolone), 159.35 (C=N pyrazolone), 135.57 (C-OH), 98.54–131.58 (Ar–C), 123.23, 132.56 (C–olefinic), 74.42 (CH–pyrazolone); IR (KBr) ν : 3400 (NH₂), 3200 (OH), 1620 (C=O), 1590 (C=N), 1520(N=N); anal. calcd for $C_{24}H_{19}N_5O_3$ (425.15): C, 67.76%; H, 4.50%; N, 16.46%. Found: C, 67.66%; H, 4.36%; N, 16.38%.

5-Amino-4-((*E*)-(4-((*E*)-3-(2-hydroxyphenyl)acryloyl)phenyl) diazenyl)-2-phenyl-2,4-dihydro-3*H*-pyrazol-3-one (3d). Orange powder; yield 84%; mp 143–145 °C; ¹H NMR (400 MHz, DMSO-d₆) δ (ppm): 10.26 (s, 1H, OH), 9.79 (s, 2H, NH₂), 7.55 (d, 1H, = CH), 7.45 (d, 1H, COCH=), 6.79–8.22 (m, 13H, Ar–H), 2.60 (s, 1H, CH–pyrazolone); ¹³C NMR (101 MHz, DMSO-d₆) δ (ppm): 167.74 (C=O chalcone), 156.32 (C=O pyrazolone), 146.51 (C=N pyrazolone), 143.65 (C–OH), 116.98–131.27 (Ar–C), 122.70, 133.51 (C–olefinic), 63.92 (CH–pyrazolone); IR (KBr) ν : 3200 (NH₂), 3100 (OH), 1670 (C=O), 1600 (C=N), 1540 (N=N); anal. calcd for C₂₄H₁₉N₅O₃ (425.15): C, 67.76%; H, 4.50%; N, 16.46%. Found: C, 67.66%; H, 4.42%; N, 16.38%.

5-Amino-4-((*Z*)-(4-((*E*)-3-(4-(dimethylamino)phenyl)acryloyl) phenyl)diazenyl)-2-phenyl-2,4-dihydro-3*H*-pyrazol-3-one (3e). Dark brown powder; yield 81%; mp 112–114 °C; 1 H NMR (400 MHz, DMSO-d₆) δ (ppm): 9.67 (s, 2H, NH₂), 7.55 (d, 1H, =CH), 6.74 (d, 1H, COCH=), 7.09–8.35 (m, 13H, Ar–H), 3.04 (s, 6H, N(CH₃)₂), 2.84 (s, 1H, CH-pyrazolone); 13 C NMR (101 MHz, DMSO-d₆) δ (ppm):154.73 (C=O chalcone), 152.51 (C=O pyrazolone), 145.51 (C=N pyrazolone), 111.52–137.28 (Ar–C), 122.98, 138.22 (C-olefinic), 79.78 (CH-pyrazolone), 55.34 (N(CH₃)₂); IR (KBr) ν : 3400 (NH₂), 1660 (C=O), 1520 (C=N), 1500 (N=N); anal. calcd for $C_{26}H_{24}N_6O_2$ (452.20): C, 69.01%; H, 5.35%; N, 18.57%. Found: C, 68.91%; H, 5.55%; N, 18.39%.

(*E*)-3-Amino-4-((4-(3-(furan-2-yl)acryloyl)phenyl)diazenyl)-1-phenyl-1*H*-pyrazol-5(4*H*)-one (3*f*). Black powder; yield 79%; mp 165–167 °C; 1 H NMR (400 MHz, DMSO-d₆) δ (ppm): 9.91 (s, 2H,

NH₂), 7.92 (d, 1H, =CH), 7.13 (d, 1H, COCH=), 6.71–8.17 (m, 12H, Ar–H), 2.17 (s, 1H, CH–pyrazolone); 13 C NMR (101 MHz, DMSO-d₆) δ (ppm): 151.58 (C=O chalcone), 147.45 (C=O pyrazolone), 146.80 (C=N pyrazolone), 112.83–136.01 (Ar–C), 124.28, 136.37 (C–olefinic), 75.72 (CH–pyrazolone); IR (KBr) ν : 3420 (NH₂), 1630 (C=O), 1600 (C=N), 1530 (N=N); anal. calcd for $\rm C_{22}H_{17}N_5O_3$ (399.13): C, 66.16%; H, 4.29%; N, 17.53%. Found: C, 66.06%; H, 4.21%; N, 17.35%.

5-Amino-2-phenyl-4-((*E*)-(4-((2*E*,4*E*)-5-phenylpenta-2,4-dienoyl)phenyl)diazenyl)-2,4-dihydro-3*H*-pyrazol-3-one (3g). Brown powder; yield 83%; mp 152–154 °C; ¹H NMR (400 MHz, DMSO-d₆) δ (ppm): 6.39 (s, 2H, NH₂), 8.07 (t, 1H, =<u>CH</u>-CH), 8.02 (d, 1H, COCH=), 7.68 (t, 1H, <u>CH</u>=CH-Ar), 7.62 (d, 1H, CH=<u>CH</u>-Ar), 6.74–8.60 (m, 14H, Ar-H), 2.08 (s, 1H, CH-pyrazolone); IR (KBr) ν : 3400 (NH₂), 1620 (C=O), 1590 (C=N), 1560 (N=N); anal. calcd for C₂₆H₂₁N₅O₂ (435.17): C, 71.71%; H, 4.86%; N, 16.08%. Found: C, 71.55%; H, 4.68%; N, 15.92%.

Biological investigations

Antioxidant activity using DPPH. Using a modified Zheleva-Dimitrova approach, the synthesized chalcone derivatives' ability to scavenge free radicals [DPPH] was evaluated. 31,32

Docking and ADMET *in silico* **studies.** The pyrazolone chalcones' binding processes to the target protein YAP/TEAD were assessed by the molecular docking experiment.^{33,34} To estimate the ADMET features, the web tool Swiss ADME was used. The ESI, Section S2,† has comprehensive details.

Anticancer assessments (in vitro)

MTT assay. A panel of cell lines, namely MCF-7, HCT-116, HepG-2, and WI-38 (human lung fibroblast), was used to identify the anticancer activity of pyrazolone chalcones (3a-g). El-Nahass described the experimental details.³⁵

Statistical analysis. The statistical analysis was performed using GraphPad Prism 6 (San Diego, CA) (GraphPad Prism 6, https://www.graphpad.com/scientific-software/prism/).

Abbreviations

FT-IR Fourier transform infrared

NMR nuclear magnetic resonance

DPPH 2,2-diphenyl-1-picrylhydrazyl

ADMET absorption-distribution-metabolism-excretion-toxicity

HBA hydrogen bond accepted

HBD hydrogen bond donated HIA human intestinal absorption CNS central nervous system

CNS Central hervous system

log P octanol/water partition coefficient

Data availability

The cell lines were provided by the American Type Culture Collection (ATCC) *via* VACSERA, Cairo, Egypt, and all accession codes were added: mammary gland (MCF-7; #ATCC HTB-22 (https://www.atcc.org/products/htb-22)), colorectal

adenocarcinoma (HCT-116; #ATCC CCL-247 (https://www.atcc.org/products/htb-37)), hepatocellular carcinoma (HepG-2; #ATCC HB-8065 (https://www.atcc.org/products/hb-8065)), and human lung fibroblast (WI-38; #ATCC CCL-75 (https://www.atcc.org/products/ccl-75)). The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request. The macromolecule protein structure is deposited in the world-wide protein data bank repository (https://www.rcsb.org/structure/3KYS).

Author contributions

Ahmed A. Noser, Maha M. Salem, and Esraa M. ElSafty: writing – review & editing, writing – original draft, and methodology. Maha M. Salem: formal analysis. Adel I. Selim, Ahmed A. Noser, Mohamed H. Baren, and Hamada S. A. Mandour: supervision.

Conflicts of interest

The authors have no relevant financial or non-financial interests to disclose.

References

- 1 P. Das, G. Sayan, A. Tarun, M. Pritiprasanna, G. Sabyasachi, C. Sumita and G. Subhashis, *Mater. Chem. Phys.*, 2019, 237, 121860.
- 2 W. Cao, H. D. Chen, Y. W. Yu, N. Li and W. Q. Chen, *Chin. Med.*, 2020, **134**, 783e791.
- 3 L. Zhang, F. Ren, Q. Zhang, Y. Chen, B. Wang and J. Jiang, *Dev. Cell*, 2008, **14**, 377–387.
- 4 Y. Zhao and X. Yang, Int. J. Cancer, 2015, 137, 2767–2773.
- 5 J. V. Patil, S. S. Shubhangi, U. Shweta, G. Pankaj and B. Suresh, *ChemistrySelect*, 2024, **9**, e202303391.
- 6 S. M. Emam, B. Samir and A. M. Ahmed, *Results Chem.*, 2023, 5, 100725.
- 7 A. N. Ahmed, H. A. Aboubakr and M. S. Maha, *Bioorg. Chem.*, 2023, **131**, 106299.
- 8 A. N. Ahmed, A. S. Ihsan, H. A. Aboubakr and M. S. Maha, *ACS Omega*, 2022, 7, 25265–25277.
- M. Islam, M. Abdullah, N. Essam, Y. Sammer,
 A. Muhammad, N. Asif and W. Abdul, *J. Mol. Struct.*, 2022,
 1269, 133843.
- 10 S. Dabhade, P. Manjushri, S. Lala, A. Sachin, A. Shweta, Y. Somdatta and N. Santosh, *Chem. Biodiversity*, 2024, 21, e202400015.
- 11 F. Rizk, N. Ahmed, A. Seham and K. Amira, *J. Heterocycl. Chem.*, 2022, **59**, 2190–2206.
- 12 H. Şenol, G. Mansour, Ö. Gülbahar, T. Alim and G. Uğur, J. Mol. Struct., 2024, 1295, 136804.
- 13 A. N. Ahmed, A. A. El-Barbary, M. S. Maha, A. Hayam and S. Mohamed, *Sci. Rep.*, 2024, 14, 3530.

- 14 K. J. Setshedi, M. B. Richard, I. Kayhan, M. Dorien, C. Guy and J. L. Lesetja, *Med. Chem. Res.*, 2024, 3, 977–988.
- 15 A. N. Ahmed, A. I. Saham, A. Hayam, M. A. Nora and S. A. Hamada, *J. Iran. Chem. Soc.*, 2023, **20**, 2963–2976.
- 16 S. S. Shafi, Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem., 2021, 60, 1132–1136.
- 17 S. K. Ramadan and A. R. Sameh, *J. Iran. Chem. Soc.*, 2022, **19**, 187–201.
- 18 J. Zhou, W. Feng, X. Bin, L. Xin, P. Cheng and F. U. Peng, *Oncol. Res.*, 2024, 32, 943.
- 19 H. Tanaka, N. Seito, O. Kenji, T. Tomoya and H. Toshihiko, *Biochem. Biophys. Res. Commun.*, 2009, **381**, 566–571.
- 20 G. Padmavathi, K. R. Nand, B. Devivasha, A. Frank, M. Srishti, S. Gautam, B. Anupam and B. K. Ajaikumar, *Phytomedicine*, 2017, 25, 118–127.
- 21 A. Jasiecka, T. Maslanka and J. J. Jaroszewski, *Pol. J. Vet. Sci.*, 2014, 17, 207–214.
- 22 H. G. Kraetsch, T. Hummel, J. Lötsch, R. Kussat and G. Kobal, Eur. J. Clin. Pharmacol., 1996, 49, 377–382.
- 23 A. Weissberger, H. D. Porter and W. A. Gregory, *J. Am. Chem. Soc.*, 1944, **66**, 1851–1855.
- 24 J. Dong, H. Guang, Z. Qijing, W. Zengtao, C. Jiahua, W. Yan, M. Qingqing and L. Shaoshun, *MedChemComm*, 2019, 10, 1606–1614.
- 25 B. Zhao, X. Ye, J. Yu, L. Li, W. Li, S. Li, J. Yu, J. D. Lin, C. Y. Wang, A. M. Chinnaiyan, Z. C. Lai and K. L. Guan, *Genes Dev.*, 2008, 22, 1962–1971.
- 26 Y. Liu-Chittenden, B. Huang, J. S. Shim, Q. Chen, S. J. Lee, R. A. Anders, J. O. Liu and D. Pan, *Genes Dev.*, 2012, 26, 1300–1305.
- 27 H. Zhang, S. K. Ramakrishnan, D. Triner, B. Centofanti, D. Maitra, B. Gyorffy, J. S. Sebolt-leopold, M. K. Dame, J. Varani, D. E. Brenner, E. R. Fearon, M. B. Omary and Y. M. Shah, *Sci. Signaling*, 2015, 8, 98.
- 28 A. N. Ahmed, E. Mohamed, D. Thoria and H. A. Aboubakr, *Molecules*, 2020, 25, 4780.
- 29 S. K. Liew, M. Sharan, M. A. Norhafiza and H. N. Noor, *Biomolecules*, 2020, **10**, 138.
- 30 Y. Ali, S. A. Hamid and R. Umer, *Mini-Rev. Med. Chem.*, 2018, **18**, 1548–1558.
- 31 W. M. Hamada, M. N. El-Nahass, A. A. Noser, T. A. Fayed, M. El-Kemary, M. M. Salem and E. Bakr, *Sci. Rep.*, 2023, 13, 15420.
- 32 A. Noser, H. Mohamed, A. Saham, M. Rekaby and M. Maha, *ChemistrySelect*, 2023, **8**, e202204670.
- 33 Y. Son, J. Kim, Y. Kim, S. G. Chi, T. Kim and J. Yu, *Bioorg. Chem.*, 2023, **131**, 106274.
- 34 H. A. Hekal, O. M. Hammad, N. R. El-Brollosy, M. M. Salem and A. K. Allayeh, *Bioorg. Chem.*, 2024, 147, 107353.
- 35 M. N. El-Nahass, E. A. Bakr, T. A. Fayed, W. M. Hamada, M. M. Salem and A. M. Radwan, *J. Iran. Chem. Soc.*, 2024, 21, 699–718.