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Applicability of NMR spectroscopy to quantify microplastics across varying concentrations in polymer mixtures†

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Quantitative nuclear magnetic resonance (qNMR) spectroscopy could potentially be used for environmental microplastic analyses, provided the challenges posed by mixed polymer samples with varying concentrations and overlapping signals are understood. This study investigates the feasibility of qNMR as a reliable and cost-efficient method for quantifying synthetic polymers in mixtures of low and varying concentrations, addressing key challenges and limitations. Polymer mixtures were analysed using deuterated chloroform (CDCl₃) and deuterated tetrahydrofuran (THF-d₈) as solvents, with polystyrene (PS), polybutadiene-cis (PB), polyisoprene-cis (PI), polyvinyl chloride (PVC), polyurethane (PU), and polylactic acid (PLA) as selected polymers. Mixtures contained either low and high concentrations of each polymer or equal concentrations of all six polymers. Polymer concentrations were measured using the internal standard method. The method showed low relative errors for low concentrations of PS in CDCl₃ and PVC in THF-d₈, with values of −5% and 0%, respectively, while PB and PI in CDCl₃ show relative errors of +5% and −3%, respectively. We observe significant linearity between nominal and measured concentrations with R^2 values ranging from 0.9655 to 0.9981, except for PU, which had high relative errors and poor linearity ($R^2 = 0.9548$). Moreover, simultaneous quantification of six polymers in THF-d₈ proves effective at intermediate concentrations. However, overlapping proton signals are observed, causing high-concentration polymers to mask low-concentration ones. While this study demonstrates low limit of quantification (LOQ) and advances in simultaneous polymer quantification, further research is needed to improve qNMR accuracy for mixed polymer samples and environmentally relevant concentrations.

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Introduction

Plastic products are ubiquitous in modern society,^{1–3} which has led to increasing global concern. Correspondingly, microplastics (MP, 1 μm–5 mm)^{2,4} and nanoplastics (NP, <1 μm)^{5–8} have emerged as major environmental issues due to their potential risks to human health and the environment.^{3,9} The widespread presence, small size, and ability of MPs to be taken up and translocated into biological tissues make them a critical environmental concern,^{10–12} necessitating further research on their toxicity.^{4,13,14}

Despite growing awareness, there are no ISO standards for MP quantification, and current methods remain inadequate.

Existing approaches include optical methods such as scanning electron microscopy (SEM), Fourier transform infrared spectroscopy (FTIR), and Raman spectroscopy,^{15,16} as well as thermoanalytical techniques such as thermal desorption gas chromatography-mass spectrometry (TDS-GC-MS) and pyrolysis GC-MS.^{17–19} Novel techniques like laser-induced breakdown spectroscopy (LIBS) and laser ablation inductively coupled plasma mass spectrometry (LA-ICP-MS) also show promise in identifying aged microplastics.²⁰ However, these methods are often costly, time-consuming, and typically involve single replicate analysis due to high sample costs.²¹ Therefore, fast, reliable, and cost-effective polymer quantification is essential for environmental risk assessments.

In recent years, proton nuclear magnetic resonance (¹H NMR) and quantitative NMR (qNMR) spectroscopy has become a popular method for detecting and analysing MP particles^{22–27} due to its rapid, cost-effective, and precise nature.^{28,29} However, challenges exist, including the need for suitable solvents to dissolve polymers, the loss of particle size, colour, and shape information, and the requirement for high temperatures to dissolve resistant polymers like polypropylene (PP) and polyethylene (PE).^{30–32} Recent research has focused on analysing

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† Electronic supplementary information (ESI) available: ¹H NMR spectra of pure MPs: PS, PB and PI in CDCl₃ (Fig. S1) and PS, PVC and PU in THF-d₈ (Fig. S2) with relevant proton signals; linearity data of the calibration curves (Table S1). See DOI: <https://doi.org/10.1039/d5ra01174d>



various polymers, such as low-density polyethylene (LDPE), polystyrene (PS), polyethylene terephthalate (PET), acrylonitrile-butadiene-styrene (ABS), polyamide (PA), polyvinyl chloride (PVC), polyurethane (PU), polylactic acid (PLA), polybutadiene (PB), polyisoprene (PI), polymethylmethacrylate (PMMA), and polyacrylonitrile (PAN), offering insights into the proton signals of pure polymer.^{22–24,27,33} This first step towards differentiating various proton signals of polymers provides a basis for the quantification of polymers in mixtures.

While some previous research has explored the application of qNMR spectroscopy to polymer mixtures at high or uniform concentrations,^{33,34} the challenge of quantifying low and varying concentrations, reflecting the complexity of environmental samples, remains unaddressed. This study therefore investigates and describes the challenges and potential pitfalls in quantifying mixed polymer samples at low and varying concentrations. We report uncertainties and relative errors when quantifying polymer mixtures, using concentrations close to the previously determined limit of quantification (LOQ) for single polymers.²⁷ Additionally, we assess whether the precision in the analyses changes in polymer mixtures with differing concentrations due to overlapping or interfering proton signals. This is a highly necessary step towards understanding the reliability and limitations of qNMR spectroscopy for real environmental sample analysis (Schmidt *et al.*, in prep).

Experimental

Materials

Commercially available polymer particles of polystyrene (PS), polylactic acid (PLA), and polyurethane (PU) beads (3–5 mm) from GoodFellow Cambridge Ltd, England; polyvinyl chloride (PVC) powder (<50 μm , > 99.7% purity) from Werth-Metall, Germany, natural rubber polyisoprene-cis (PI), and synthetic polybutadiene-cis (PB) from Sigma-Aldrich®, were utilized as model particles. As solvents, deuterated chloroform (CDCl_3 ,

99.8 atom % D) from Sigma-Aldrich® with its residual proton signal at 7.26 ppm and deuterated tetrahydrofuran (THF-d_8 , ≥ 99.5 atom % D) purchased by Sigma-Aldrich® and VWR International, LLC with its residual proton signals at 3.58 ppm and 1.73 ppm, were used. Dimethyl sulfone (DMSO_2) purchased by TraceCERT®, Sigma-Aldrich® with its proton signal at 3.00 ppm was utilized as an internal standard.

Sample preparation

This study consists of three parts. The first part involves acquiring a single ^1H NMR spectrum of the polymers PS, PB and PI in CDCl_3 and PS, PVC and PU in THF-d_8 . A high concentration (1 mg mL^{-1}) was chosen and served as quality control and reference for the polymer mixtures in the setups described below. For the second part, all polymer mixtures were prepared from a stock solution with a high concentration. The first setup (Table 1) consists of a mixture of three polymers of equally high concentrations (333 $\mu\text{g mL}^{-1}$). PS, PB and PI were dissolved in CDCl_3 (setup 1A) and PVC, PU, PS in THF-d_8 (setup 1B). Setup 2A consists of mixtures of three polymers in CDCl_3 , always including one high and two low concentrations (Table 1, Mixture 1–3). Setup 2B used mixtures of three polymers in THF-d_8 , always including one low and two high concentrations (Table 1, Mixture 4–6). The lower concentrations used in setups 2A and 2B, were the LOQ for that polymer.²⁷ The third setup consisted of sets of six mixtures in CDCl_3 or THF-d_8 with concentration gradients that made up a calibration curve (Table 1, setup 3A and 3B). These mixtures served as calibration samples. The calibration concentration levels were 750, 500, 333, 250 and 125 $\mu\text{g mL}^{-1}$. The concentrations 125 and 250 $\mu\text{g mL}^{-1}$ of each polymer were used twice in different constellations with other polymers, while the concentrations 500 and 750 $\mu\text{g mL}^{-1}$ were represented once per polymer. The third part involved preparing a polymer mixture in the fourth setup, consisting of equal parts of PS, PVC, PU, PI, PB and PLA in THF-d_8 . Each polymer had a nominal concentration of 166 $\mu\text{g mL}^{-1}$.

Table 1 Overview of the four experimental setups of polymer mixtures in CDCl_3 or THF-d_8 ^a

Setup	PS	PB	PI	PS	PVC	PU
Nominal concentration [$\mu\text{g mL}^{-1}$]						
Part 2	Solvent		CDCl_3(A)	THF-d_8(B)		
Setup 1A and 1B	333	333	333	333	333	333
Setup 2A and 2B	1000	0.4	0.6	4	100	100
	4	1000	0.6	200	8	100
	4	0.4	1000	200	100	6
Setup 3A and 3B	500	250	250	500	250	250
	250	500	250	250	500	250
	250	250	500	250	250	500
	750	125	125	750	125	125
	125	750	125	125	750	125
	125	125	750	125	125	750
Part 3	Solvent		THF-d_8(B)			
Setup 4	166	166	166	166	166	166

^a PS = polystyrene, PB = polybutadiene-cis, PI = polyisoprene-cis, PVC = polyvinyl chloride, PU = polyurethane.



All polymers in their corresponding solvents were dissolved at room temperature. To quantify the polymers, present in the sample, the internal standard of DMSO₂ was prepared in both solvents and added to all setups with a known concentration. For the NMR measurements, each sample solution (600 μL) was transferred into 5 mm NMR tubes (Bruker BioSpin, 4" NMR tubes) and subsequently analysed.

qNMR

All NMR experiments were conducted using a Bruker Ascend 600 MHz spectrometer equipped with an AVANCE NEO console and a QCI-P CryoProbe™. All measurements were performed at a temperature of 298 K. For the qNMR measurements, the acquisition parameters were standardized across all polymer mixtures, with the pulse width and receiver gain automatically calibrated for each sample. The spectral width was set to 29.76 ppm, the number of scans was 8, the spectral size was 262 144 points, the acquisition time was 3.67 s, and the delay was 60 s for each sample.

For visualisation, the acquired ¹H NMR spectra were imported into the NMR software program MestReNova (v14.2.0), while the obtained qNMR spectra for quantification were imported into Bruker's TopSpin NMR software (Version 4.3.0). For all qNMR data, manual phase and baseline correction were conducted and the line broadening applied to all data was 0.1 Hz. For each polymer type, a consistent ppm range was manually integrated across all samples. The integration of the signal areas corresponds to the proton atoms and, consequently, to the concentration of the analyte in the solution. When performing quantitative analysis, care must be exercised in the integration of the signal regions of interest. To minimize potential measurement or integration errors, the internal standard method was employed for all quantitative analyses, as previously published.²⁸ The proton signal of DMSO₂ served as the internal standard across all polymer samples. When utilizing DMSO₂ as the internal standard, it is essential to ensure that the same concentration is added to each polymer sample and that the concentration is selected to be within a similar intensity range as the deuterated solvents. The concentration of the polymers in their respective solvents was measured as follows (eqn (1)):

$$C_u = C_r \frac{A_u n_r}{A_r n_u} \quad (1)$$

where C_u is the unknown polymer concentration, C_r is the known concentration of the internal standard, A_r is the integral of the proton of the internal standard with known concentration, A_u is the integral of the proton of the polymer sample with unknown concentration, n_r is the number of atoms of the proton of the internal standard with known concentration and n_u is the number of atoms of the proton of the polymer sample with unknown concentration.

Calculations and statistical analysis

Calculations and statistical analyses were performed, and graphics were made using RStudio (Version 4.3.0). In all setups,

each proton signal of each polymer in the different polymer mixtures was measured according to eqn (1) as one replicate sample. For the calibration curve (setup 3A and 3B), polymers with more than one proton signal the total polymer concentration was calculated as the average of the measured concentrations per proton signal. Additionally, for all polymers the concentrations of 125 μg mL⁻¹ and 250 μg mL⁻¹ were included twice in the calibration curve setup (Table 1, setup 3A and 3B), and were thus used as two data points each. For determining a linear regression, the measured concentrations of the different polymers in the calibration curve setup were plotted against their respective nominal concentrations in micrograms per millilitre (eqn (1)), and a linear regression analysis was run, finding the coefficient of determination (R^2) for each regression line. If R^2 is larger than 0.99, the regression can be considered linear.³⁵ The significance level was set to $p < 0.05$ and F-tests were analysed. The upper and lower confidence intervals of 95% were included in the plot.

For setup 1A and 1B, setup 2A and 2B and setup 4 the relative error reports the percent deviation from the nominal concentration. A relative error of 0% means an exact match between the nominal and the measured concentration. The relative error was measured using the formula as follows (eqn (2)):

$$\text{Relative Error}[\%] = \left(\frac{\text{measured concentration}}{\text{nominal concentration}} \times 100 \right) - 100 \quad (2)$$

Prevention of contamination and quality control

To reduce the risk of microplastic contamination from airborne particles, all glassware and other plastic-free equipment were cleaned with water, acetone, and distilled water. All glass flasks were dried for 24 h at 60 °C prior to use and then sealed with a lid once cooled. The NMR tubes were dried for 30 minutes at 60 °C in a closed heating cabinet that had been wiped clean with 95% ethanol before use. Furthermore, laboratory coats composed of pure cotton were used, and samples were covered and kept closed during sampling handling to prevent contamination of polymer fibers. Additionally, nitrile gloves were worn and routinely replaced to avoid cross-contamination. Procedural blanks, consisting of ¹H NMR and qNMR spectra of pure CDCl₃ and THF-d₈ without polymer added, were acquired.

Results

Identification – pure polymer ¹H NMR spectra

To identify the different polymers in the mixtures, pure polymer spectra in their corresponding solvents were prepared and assigned. For clarity, only the regions containing relevant polymer proton signals are considered. The proton signal ranges used for polymer concentration calculations in mixtures are as follows: PS exhibited proton signals in the range of 7.20 to 6.20 ppm (PS-H_a,H_b), PB observed proton signals at 5.38 ppm (PB-H_a) and 2.09 ppm (PB-H_b), and PI showed proton signals at 5.12 ppm (PI-H_a) and 2.04 ppm (PI-H_b). PVC presented a proton signal in the range of 4.70 to 4.25 ppm (PVC-H_a). For PU, proton signals were identified in the ranges of 8.67 to 8.47 ppm (PU-



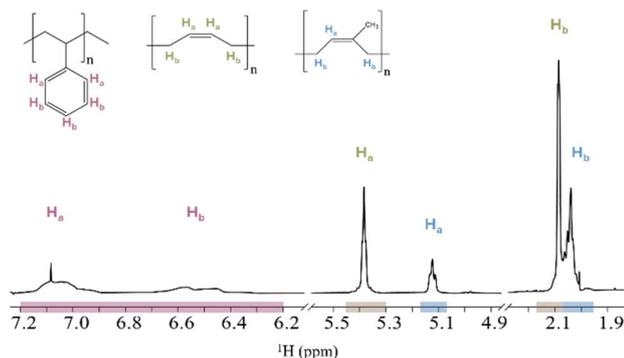


Fig. 1 ^1H NMR spectrum and their structural formula of setup 1A (PS, PB and PI in CDCl_3), showing overlap between signals of PB- H_b and PI- H_b . Integrated areas are highlighted with coloured sections: PS (red), PB (brown) and PI (blue). Proton assignments for each polymer are shown above the corresponding signal areas.

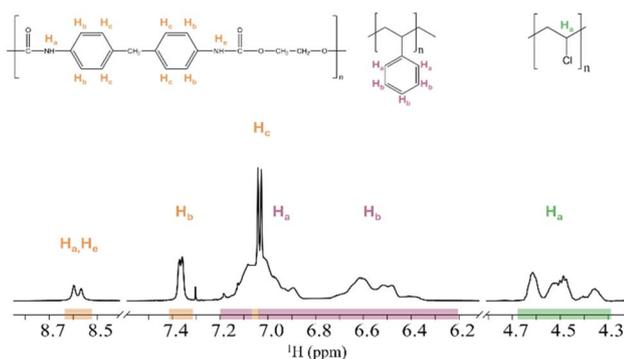


Fig. 2 ^1H NMR spectrum and their structural formula of setup 1B (PU, PS and PVC in THF-d_8), showing overlap between signals of PU- H_c and PS- H_a, H_b . Integrated areas are highlighted with coloured sections: PU (orange), PS (red), and PVC (green). Proton assignments for each polymer are shown above the corresponding signal areas.

H_a, H_c), 7.43 to 7.29 ppm (PU- H_b) and 7.07 to 6.98 ppm (PU- H_c). The spectra and signal assignments are consistent with previously published work²⁷ and are provided in Fig. S1 and S2 in the ESI† for reference.

Quantitative analysis by qNMR spectroscopy of setup 1A and 1B

For an overview of the experimental setups, see Table 1. Setup 1A and 1B were prepared with nominal concentrations of $333 \mu\text{g mL}^{-1}$ for each polymer. The ^1H NMR spectrum of PS, PB and PI in CDCl_3 (setup 1A) shows an overlap between the proton signals corresponding to PB- H_b and PI- H_b (Fig. 1). Similarly, the ^1H NMR spectrum of PS, PU and PVC in THF-d_8 (setup 1B) shows an overlap between the proton signals corresponding to PS- H_a, H_b and PU- H_c (Fig. 2). Table 2 presents setup 1A and 1B, including the nominal concentrations, the measured concentrations, and their relative error for the respective proton signals of the different polymer types.

To enable a deeper comprehension of the overlapping signals, the spectrum of the polymer mixture was superimposed with the pure spectra of overlapping polymers. This approach facilitated the identification of the start and end point of the overlapping signals, as well as the degree of their interaction. The overlapping proton signal of PB- H_b and PI- H_b (setup 1A, Fig. 1) was separated at 2.07 ppm, integrated individually and the concentration measured. The relative errors of PB- H_b and PI- H_b were +2% and -2%, respectively, using this method. In the overlapping proton signals of PS- H_a, H_b and PU- H_c (setup 1B, Fig. 2), PU- H_c appears as a sharper signal than PS. Its signal range of 7.07 to 6.98 ppm was used to measure the concentration. The concentration of PS- H_a, H_b was measured based on its proton signals range of 7.20 to 6.20 ppm. The measured concentration of PU- H_c was close to its nominal concentration with a relative error of -12%. The measured concentration of PS- H_a, H_b deviated more from its nominal concentration, with relative errors of +24% and +14% in CDCl_3 and THF-d_8 , respectively. Furthermore, the measured concentrations of the proton signals of PU- H_a, H_c and PU- H_b deviated -60% of their nominal concentrations.

Overall, with observed relative error ranges from +24% to -60% for the setups 1A and 1B, the results are not sufficiently consistent for all polymers. This inconsistency depends on solvents and the proton signals used, highlighting the importance of understanding potential interactions and proton signal interferences between polymers in mixtures. It indicates that numbers and integrations directly from the NMR should not be

Table 2 Quantification of the polymer concentrations in mixtures of PS, PB and PI in CDCl_3 (setup 1A) and PS, PVC and PU in THF-d_8 (setup 1B). The nominal and measured concentrations [$\mu\text{g mL}^{-1}$], determined using the internal standard method with DMSO_2 , along with the relative error [%] are given for each polymer and setup

Solvent	Polymer type and proton signal	Nominal concentration [$\mu\text{g mL}^{-1}$]	Measured concentration [$\mu\text{g mL}^{-1}$]	Relative error [%]
Setup 1A CDCl_3	PS- H_a, H_b	333	412	24
	PB- H_a	333	306	-8
	PB- H_b	333	340	2
	PI- H_a	333	333	0
	PI- H_b	333	325	-2
Setup 1B THF-d_8	PS- H_a, H_b	333	381	14
	PVC- H_a	333	333	0
	PU- H_a, H_c	333	134	-60
	PU- H_b	333	134	-60
	PU- H_c	333	294	-12



considered accurate and reliable for quantifications when working with mixtures of moderate to high polymer concentrations. Additionally, selecting the appropriate proton signal for polymer measurements is crucial in mixtures.

Quantitative analysis by qNMR spectroscopy of mixed polymers of different concentrations

Polymer mixtures in CDCl₃ (setup 2A). The ¹H NMR spectrum of PS, PB and PI in CDCl₃ (setup 2A) are presented in Fig. 3 and the corresponding quantification data of each proton signal in the different polymer mixtures in Table 3. Mixture 1 using low concentrations of PB and PI show that for the H_a proton signal the relative errors of the measured polymer concentrations are very low with +5% and -3%, respectively. Furthermore, the analysis of PS-H_a,H_b gives a relative error of the measured polymer concentration of +20%.

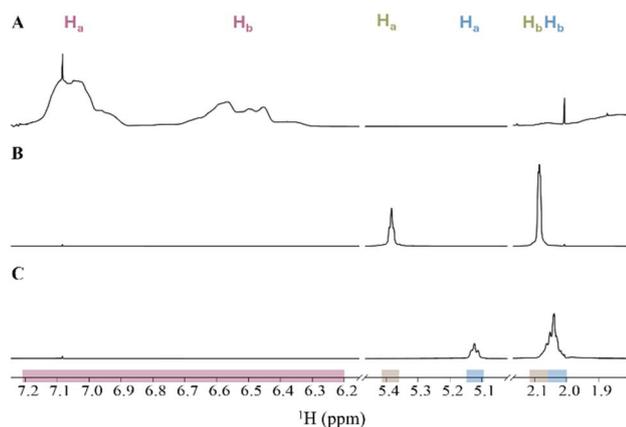


Fig. 3 ¹H NMR spectrum of setup 2A with signal ranges for PS (red), PB (brown) and PI (blue) in CDCl₃: (A) mixture 1: high PS concentration and low PB and PI concentration, (B) mixture 2: high PB concentration and low PS and PI concentration and (C) mixture 3: high PI concentrations and low PS and PB concentrations. Proton assignments for each polymer are shown above the corresponding signal areas.

In contrast, in the same mixture we observe an overlap of the proton signals PB-H_b and PI-H_b (Fig. 3A) which give large deviations from nominal values of +2100% and +2967%, respectively (Table 3). Similarly, we observe that also in mixtures 2 and 3 with high concentration of either PB or PI (Fig. 3B,C) the relative error of measured concentrations for the low-concentration polymer is unacceptable. The short distance between PB-H_a and PI-H_a affects the proton signal with the lowest concentration the most (Fig. 3B and C). However, the low concentrations of PS in mixtures 2 and 3 gave relative errors of +43% and -5%, respectively.

Polymer mixtures in THF-d₈ (setup 2B). The ¹H NMR spectrum of PS, PVC and PU in THF-d₈ (setup 2B) are presented in Fig. 4 and the corresponding quantification data of each proton signal in mixtures 4–6 in Table 4. In mixtures 4 and 6 (PVC high) the proton signal of PVC-H_a has relative errors of -2% and 4%,

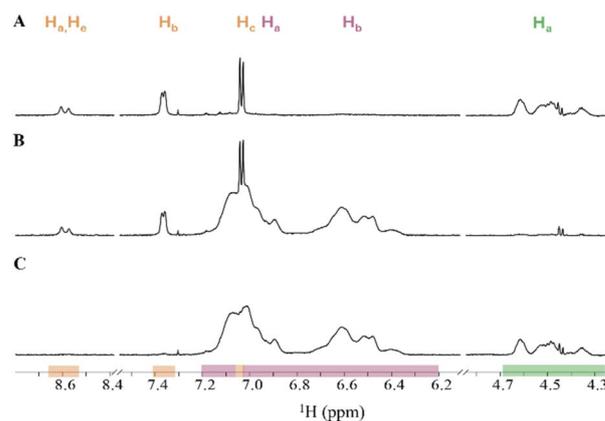


Fig. 4 ¹H NMR spectrum of setup 2B with signal ranges for PU (orange), PS (red) and PVC (green) in THF-d₈: (A) mixture 4: high PU and PVC concentrations and low PS concentration, (B) mixture 5: high PU and PS concentrations and low PVC concentration and (C) mixture 6: high PS and PVC concentrations and low PU concentration. Proton assignments for each polymer are shown above the corresponding signal areas.

Table 3 Quantification of the polymer concentrations in mixtures of PS, PB and PI in CDCl₃ (setup 2A). The nominal and measured concentrations [$\mu\text{g mL}^{-1}$], determined using the internal standard method with DMSO₂, along with the relative error [%] are given for each polymer and setup

Solvent CDCl ₃ setup 2A	Polymer type and proton signal	Nominal concentration [$\mu\text{g mL}^{-1}$]	Measured concentration [$\mu\text{g mL}^{-1}$]	Relative error [%]
Mixture 1 (Fig. 3A)	PS-H _a ,H _b	1000	1196	20
	PB-H _a	0.4	0.42	5
	PB-H _b	0.4	8.8	2100
	PI-H _a	0.6	0.58	-3
	PI-H _b	0.6	18.4	2967
Mixture 2 (Fig. 3B)	PS-H _a ,H _b	4	5.7	43
	PB-H _a	1000	1083	8
	PB-H _b	1000	1080	8
	PI-H _a	0.6	4.5	650
Mixture 3 (Fig. 3C)	PI-H _b	0.6	95.6	15 833
	PS-H _a ,H _b	4	3.8	-5
	PB-H _a	0.4	5.3	1225
	PB-H _b	0.4	187	46 650
	PI-H _a	1000	1141	14
	PI-H _b	1000	1007	1



Table 4 Quantification of the polymer concentrations in mixtures of PS, PVC and PU in THF- d_8 (setup 2B). The nominal and measured concentrations [$\mu\text{g mL}^{-1}$], determined using the internal standard method with DMSO_2 , along with the relative error [%] are given for each polymer and setup

Solvent THF- d_8 setup 2B	Polymer type and proton signal	Nominal concentration [$\mu\text{g mL}^{-1}$]	Measured concentration [$\mu\text{g mL}^{-1}$]	Relative error [%]
Mixture 4 (Fig. 4A)	PS- H_a, H_b	4	21.0	425
	PVC- H_a	100	98.0	-2
	PU- H_a, H_c	100	35.7	-64
	PU- H_b	100	35.7	-64
	PU- H_c	100	40.9	-59
Mixture 5 (Fig. 4B)	PS- H_a, H_b	200	221	11
	PVC- H_a	8	8.03	0
	PU- H_a, H_c	100	37.0	-63
	PU- H_b	100	37.2	-63
	PU- H_c	100	134	34
Mixture 6 (Fig. 4C)	PS- H_a, H_b	200	217	9
	PVC- H_a	100	104	4
	PU- H_a, H_c	6	2.5	-58
	PU- H_b	6	2.6	-57
	PU- H_c	6	101	1583

respectively (Table 4). In mixture 5 (PVC low) the proton signal of PVC- H_a has a relative error of 0%. The results imply that PVC can be safely quantified in both high and low concentrations in mixtures.

In contrast, we observe that a high concentration of PU and low concentration of PS (Mixture 4) interferes with the proton signals of PS- H_a, H_b (Fig. 4A), and gives a high relative error for both. In the same way as for setup 2A (Fig. 3), the proton signals of PS- H_a, H_b and PU- H_c overlap in mixtures 4 and 5 (Fig. 4A and B). In mixtures 5 and 6, the high concentrations of PS- H_a, H_b were measured with acceptable relative errors of 11% and 9%. The slight deviation from nominal values can be due to the overlap with the proton signals of PU- H_c . However, it is striking that all measured concentrations of PU are below the nominal concentrations. PU- H_a, H_c and PU- H_b , have unacceptably high relative errors of -64% to -57%. The relative error for the proton signal of PU- H_c varies between -59% (Mixture 4) and +1583% (Mixture 6).

Calibration curves in CDCl_3 (setup 3A) and THF- d_8 (setup 3B)

In the calibration curve (setup 3A and 3B) concentrations of the different polymers in mixtures were analysed and plotted against the nominal concentrations. To find the measured concentration of polymers with multiple proton signals, the concentrations measured from each proton signal were averaged. When there were two data points for the same concentration ($125 \mu\text{g mL}^{-1}$ and $250 \mu\text{g mL}^{-1}$ of each polymer) in mixtures, the two data points were used as two replicates for the same concentration (Fig. 5 and 6).

For setup 3A, polymer concentration mixtures in CDCl_3 of PS in mixture with PB and PI give an R^2 of 0.9655 (Fig. 5A), PB in mixture with PS and PI give an R^2 of 0.9833 (Fig. 5B), and PI in mixture with PS and PB give an R^2 of 0.9857 (Fig. 5C). Although the R^2 values are below 0.99, it still implies a good linear relationship between measured concentrations and nominal concentrations in the range of concentrations that were tested.

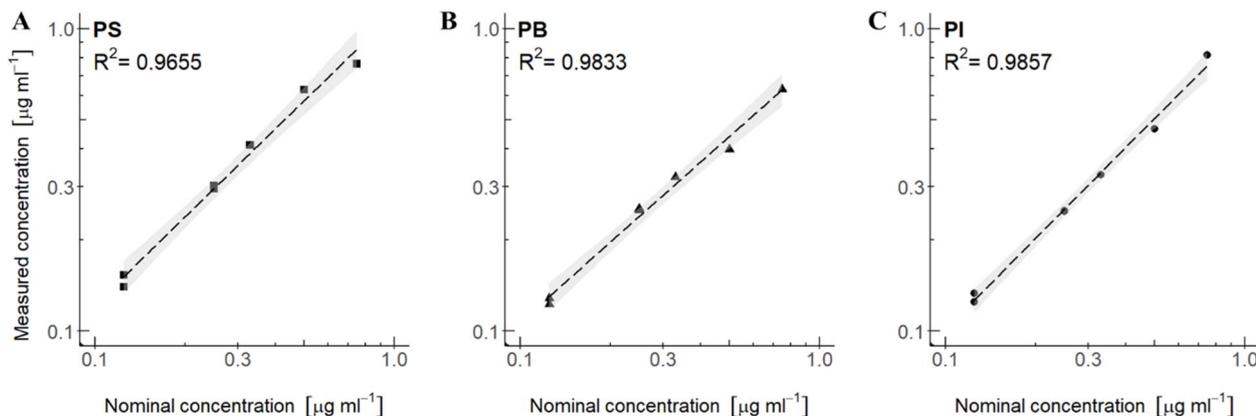


Fig. 5 Linear regression of polymer mixtures in CDCl_3 : (A) PS in mixture with PI and PB, (B) PB in mixture with PS and PI and (C) PI in mixture with PS and PB. All polymer mixtures include an internal standard of DMSO_2 and are represented with a confidence interval (95%). The nominal concentration is plotted against the measured concentration (measured by eqn (1)).



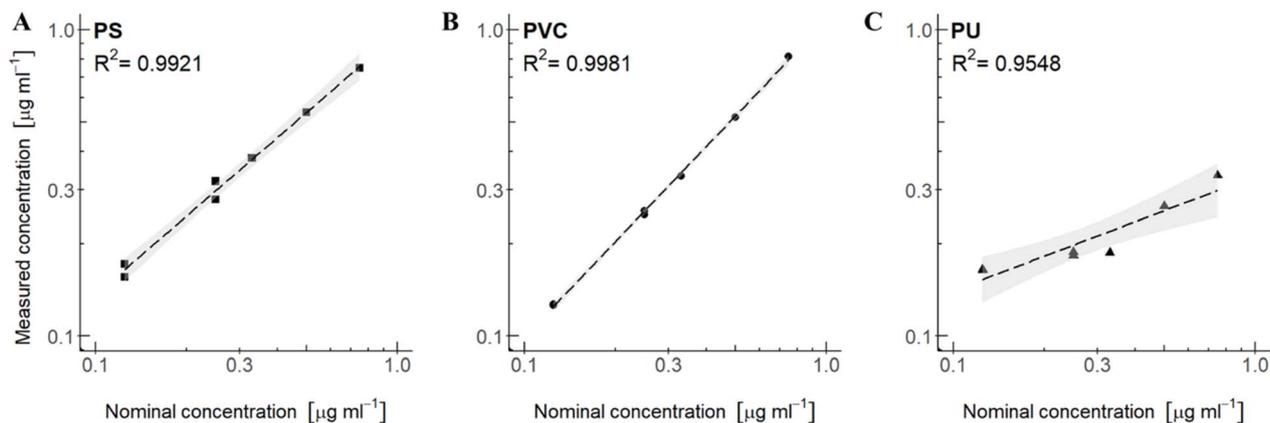


Fig. 6 Linear regression of polymer mixtures in THF- d_8 : (A) PS in mixture with PVC and PU (B) PVC in mixture with PS and PU and (C) PU in mixture with PS and PVC. All polymer mixtures include an internal standard of DMSO $_2$ and are represented with a confidence interval (95%). The nominal concentration is plotted against the measured concentration (measured by eqn (1)).

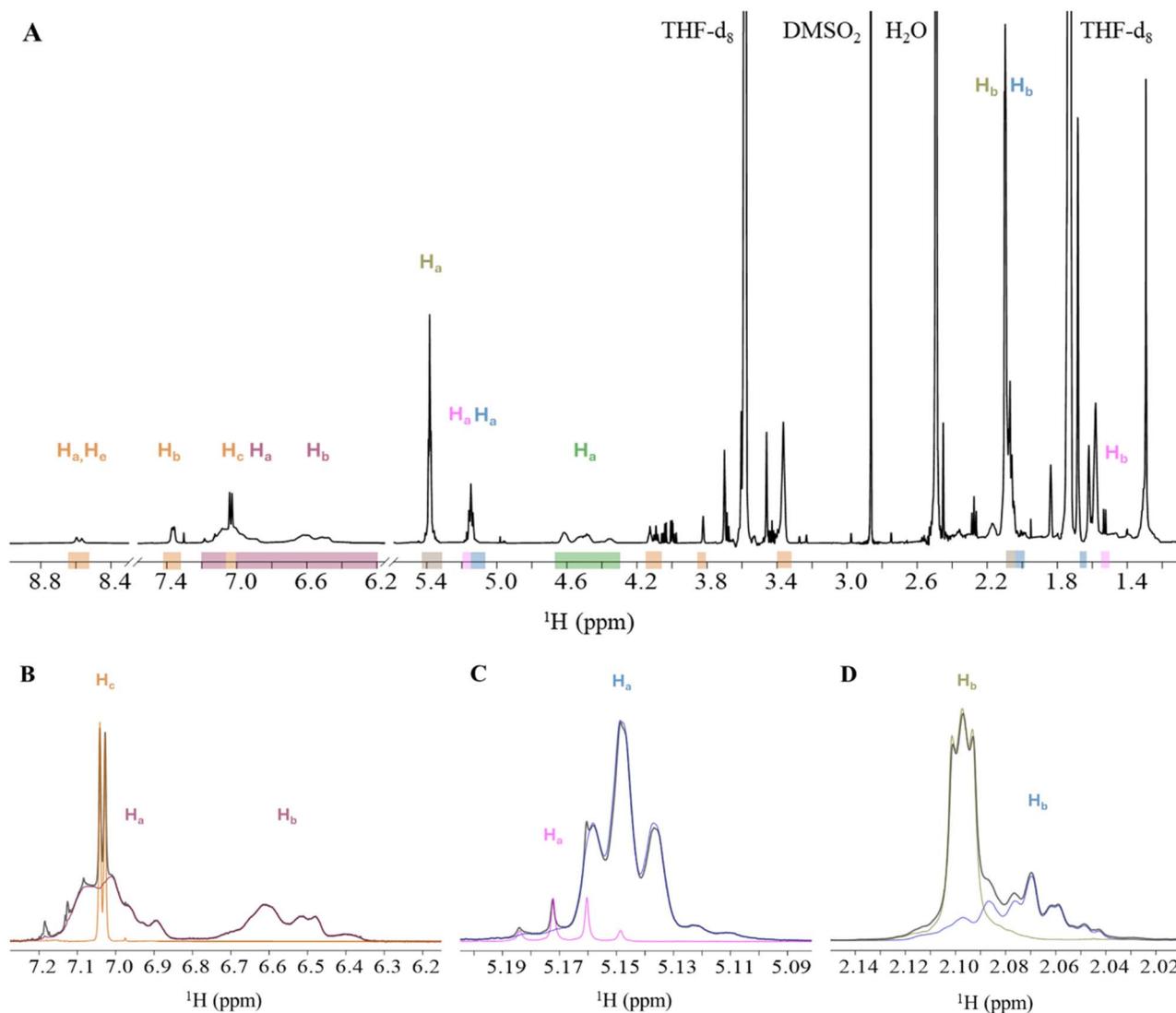


Fig. 7 (A) Full ^1H NMR spectrum of setup 4 with signal ranges for PU (orange), PS (red), PB (brown), PLA (pink), PI (blue), and PVC (green) in THF- d_8 . Panels B–D: detail showing three areas with overlapping polymer signals (black spectral line) of (B) PS- H_a, H_b and PU- H_c , (C) PLA- H_a and PI- H_a and (D) PB- H_b and PI- H_b with the same colour coding as in (A). Proton assignments for each polymer are shown above the corresponding signal areas.



For setup 3B, polymer mixtures in THF- d_8 , the measured *versus* nominal concentrations give good R^2 values for the polymers PS and PVC. R^2 for PS in mixture with PVC and PU is 0.9921 (Fig. 6A) and PVC in mixture with PS and PU has an R^2 of 0.9981 (Fig. 6B), implying a good linear relationship between measured concentrations and nominal concentrations for these polymer mixtures. PU in mixture with PS and PVC, however, gives an R^2 of 0.9548 (Fig. 6C), which corresponds with the large relative error observed for PU in mixtures (Table 4). In both setups, linear regression analysis (*cf.* the ESI†) revealed significance levels of $p < 0.05$ and high F-test values of each polymer type, indicating a significant relationship between the measured and nominal concentrations.

In contrast to setup 2A and 2B with concentration spans from 0.4 to 1000 $\mu\text{g mL}^{-1}$, the concentration span among polymers in the calibration curve are less pronounced, and we also observe a higher precision in the measurements. These spans may describe realistic concentration ratios in environmental samples, and the results indicate that the method is able to quantify the polymers in these mixtures with good accuracy except for PU.

Quantitative analysis by qNMR spectroscopy of mixed polymers in THF- d_8

To facilitate a more comprehensive examination of the potential for detecting and quantifying multiple polymers in mixtures, in the third part a sample was prepared by combining equal volumes of stock solutions containing the six polymers PS, PVC, PI, PB, PU, and PLA in THF- d_8 (Fig. 7A). Each polymer had a nominal concentration of 166 $\mu\text{g mL}^{-1}$ as shown in Table 1. The ^1H NMR spectrum of the six-polymer-mixture in THF- d_8 shows additional signals beyond those previously utilized. These signals have not previously been used for concentration calculations, and were not used in this work either, as the low ppm ranges are in regions where signals from water and DMSO₂ and other impurities of the solvents are also observed.³⁶ These omitted signal ranges include further proton signals of

PU in the range of 4.15 to 4.95 ppm and a signal at 3.36 ppm, as well as a proton signal at 3.82 ppm. Additionally, a third signal of PI (PI- H_c) was observed at 1.68 ppm and two proton signals of PLA were detected in the ranges of 5.20 to 5.12 ppm (PLA- H_a) and 1.61 to 1.56 ppm (PLA- H_b), as previously assigned.²⁷ The concentrations of these three proton signals in the polymer mixture (setup 4) were measured according to eqn (1) and are shown in Table 5.

As previously observed in setups 1 and 2, the proton signals of PS- H_a, H_b and PU- H_c (Fig. 7B), as well as those of PB- H_b and PI- H_b (Fig. 7C), were observed to overlap in this six-polymer-mixture as well. Additionally, the proton signals of PLA- H_a and PI- H_a (Fig. 7D) were observed to overlap.

The quantitative analyses of individual proton signals for the various polymers indicate that the measured concentrations of PB- H_a , PI- H_a , PI- H_c , PVC- H_a , PU- H_b , PU- H_c and PLA- H_a align closely with the nominal concentration of the polymers (Table 5), with relative errors from -7% to $+1\%$. In contrast, measured concentrations from the overlapping proton signals of PS- H_a, H_b with PU- H_c , as well as the proton signals of PB- H_b , PI- H_b , PU- H_a, H_c and PU- H_b , gave polymer concentrations significantly different from their nominal concentrations (Table 5), with relative errors in the range from -58% to $+23\%$. Additionally, although not overlapping with any other proton signal, the measured concentration of PLA- H_b , in low ppm range, differs substantially from the nominal concentration, and has a relative error of -84% .

Discussion

Recent studies on the use of NMR spectroscopy in the field of micro- and nanoplastics have advanced the understanding of how the method may be applied but has also shown that the use of qNMR spectroscopy is a method that still requires further development for the quantification of certain polymers. The method is rapid, reliable and cost-efficient, but challenges arise with mixtures, which is particularly relevant for environmental samples. Previous research has focused on examining pure polymers individually, and only a few have investigated polymer mixtures.^{33,34}

Challenges with high and low concentrations in a mixture

The current study used polymer mixtures of three different polymers in two solvents in varying concentration setups, ranging from 8 to 0.4 $\mu\text{g mL}^{-1}$, based on previously published concentration limits for single polymers.²⁷ The current investigation showed that the different concentration ratios influenced the determination of the respective polymer proton signals. The polymer mixtures containing equal parts of high concentrations showed that proton signals of PB and PI in CDCl₃ and PS and PU in THF- d_8 overlapped, a problem that persisted in the quantitative determination of the polymers in mixtures containing high and low concentrations. The overlap of the proton signals as well as the concentration ratios in the mixture can complicate an exact quantitative analysis of the overlapping polymers. To our knowledge, this study is the first

Table 5 Quantification of the polymer concentration in the six-polymer-mixture of PS, PB, PI, PVC, PU and PLA in THF- d_8 (setup 4). The nominal and measured concentrations [$\mu\text{g mL}^{-1}$], determined using the internal standard method with DMSO₂, along with the relative error [%] are given for each polymer and setup

Polymer type and proton signal	Nominal concentration [$\mu\text{g mL}^{-1}$]	Measured concentration [$\mu\text{g mL}^{-1}$]	Relative error [%]
PS- H_a, H_b	166	204.6	23
PB- H_a	166	167.2	1
PB- H_b	166	198.7	20
PI- H_a	166	159.0	-4
PI- H_b	166	107.7	-35
PI- H_c	166	155.2	-7
PVC- H_a	166	174.4	5
PU- H_a, H_c	166	70.2	-58
PU- H_b	166	70.6	-57
PU- H_c	166	154.9	-7
PLA- H_a	166	167.1	1
PLA- H_b	166	26.6	-84



to investigate overlapping proton signals and their effect on quantification, using the internal standard method. The simultaneous quantification of two polymer mixtures: one containing PVC, PS and BR, and the other containing PET and PA, using various non-deuterated solvent systems along with an internal standard, was previously reported.³⁴ The study used two concentration ranges of 2.5 to 0.5 mg mL⁻¹ and 0.5 to 0.1 mg mL⁻¹, significantly higher than those in this work. The calibration curve was based on LOD and LOQ values determined using DIN 32645 and SNR and the polymers in the mixtures were measured based on this calibration method.³⁴ Results showed accurate quantification of these polymers down to 0.1 mg mL⁻¹, achieving high linearity (R^2 above 0.99) and precision in the calibration curves. The high concentration ranges utilized may not be representative of lower concentrations typically encountered in real environmental samples.³⁴

Challenges with quantifying multiple polymers in a mixture

The polymer mixture of PS, PVC, PU, PB, PI and PLA in THF-d₈ provided a good overview of the identification and quantification of the various polymer proton signals. To our knowledge, this is the first study to use qNMR quantification to identify and accurately quantify six different polymers in one solvent and in one sample at concentrations down to 166 µg mL⁻¹ using qNMR spectroscopy.

A selective separation of eight polymers from inorganic components, along with the simultaneous fractionation of different polymer types before the use of qNMR analysis, was presented in recent research.³³ This approach tested a set of eight relevant polymers (PS, BR, PVC, PET, PA, LDPE, PMMA and PAN) dissolved sequentially in four different non-deuterated solvents (THF, trifluoroacetic acid in chloroform, formic acid in chloroform and xylene) in a concentration range of 2.5 to 0.5 mg mL⁻¹, and showed that separating different polymer types is essential for qNMR analysis of polymers, as variable solubilities require a reliable fractionation procedure to thoroughly analyse the diverse range of polymers present in environmental matrices. The study enabled the quantification of the polymers PS, PVC, PET, PA and PMMA from a single sample and observed recovery rates of over 88% for all tested polymers, whereas LDPE and PAN were absent in the NMR spectra.³³ Moreover, the lower concentrations observed and expected in real environmental samples may not be accurately reflected by the high concentrations range used. The duration of the extraction process can vary depending on whether the complete set of polymers requires analysis, with the full procedure potentially being completed within a time-frame of approximately 3 to 4 hours. The use of non-deuterated solvents for the extraction process can help maintain the costs of this procedure at a lower level compared to the utilization of deuterated solvents.³³

Overlapping proton signals in qNMR spectroscopy of mixtures

The overlapping proton signals of the polymers in different concentration ratios represent a challenge in the accurate quantification of polymers in mixtures, which is highly relevant for environmental samples, where varying concentrations must

also be expected. When a polymer has a distinct, single signal, it is preferable to be use that signal for quantification. In this study, overlapping proton signals were identified using the pure proton spectra of individual polymers. If no single signals are available, deconvolution in qNMR spectroscopy for polymer mixtures could help to resolve overlapping signals, enabling quantification of individual components despite their chemical similarities, even in complex samples. However, this approach faces certain challenges, such as the presence of broad peaks, noise, baseline distortions, and the requirement for accurate peak shape assumptions and reference data, which may potentially limit the reliability and broader applicability of the technique.^{37,38} Based on experience from this study, deconvolution did not produce spectra that resembled the pure spectra and the quantified concentrations deviated from the nominal values (Table S2 and Fig. S3 and S4 in the ESI†). Consequently, we do not recommend using deconvolution in polymer mixtures with broad, overlapping signals or highly varying concentrations. Additionally, in cases of overlapping proton signals from different polymers, lower concentrations of one polymer can remain undetected due to signal interference from other polymers with higher concentrations, complicating accurate quantification in complex polymer mixtures. Overall, while the internal standard method is generally more reliable for accurate quantification, particularly for low concentrations and signal overlap, deconvolution remains less accurate due to its limitations with broad signals. Nevertheless, it may be useful in certain cases when prior knowledge of signal appearance is available.

Solvents and internal standards

The use of the two solvents showed that the use of THF-d₈ was preferable to CDCl₃. THF-d₈ allowed the dissolution of six different polymers that could be analysed simultaneously in a single sample, additionally avoiding the collapsing between the right ¹³C satellite of CDCl₃ at 7.08 ppm with the proton signal of PS (e.g. Fig. 1A).³⁹ Nevertheless, identifying suitable solvents for a wide range of polymers, or utilizing different solvents to target various polymer types and avoid signal overlap, may be essential for the successful application of qNMR spectroscopy in the analysis of polymer mixtures. Furthermore, the use of internal standards needs to be carefully chosen when analysing polymer mixtures containing several different polymers or even environmental samples. In this study, DMSO₂, with a proton signal at 3.00 ppm, was used as the internal standard. From previous studies, it is known in which ppm ranges the different proton signals of the chosen polymers appear,^{22–24,27,33} but the use of DMSO₂ in an unknown environmental sample could lead to overlapping signals and loss of information about unexpected polymers or polymers with low concentrations. Hexamethyldisiloxane (HMDSO), with a proton signal at 0.25 ppm, proved to be suitable as an internal standard.^{33,34}

Future research needs

Considering the additional use of deconvolution of proton signals, qNMR spectroscopy may serve as a cost-effective, rapid



and reliable method for identifying and quantifying various polymers in a mixture, enabling the quantification of several different polymers in a single sample.

As the environmental concern over environmental plastic and microplastic pollution increases, and in the likely event of a global plastic treaty, management and control with release of plastic pollution through wastewater, as well as overall monitoring of environmental levels may become mandatory. A rapid and reliable method for quantification of common polymers in a range of matrices may thus be in high demand. Currently, qNMR spectroscopy could be used for the quantification of plastic polymers in highly concentrated water, sediment or soil samples in environmental compartments such as highly polluted rivers or sea-floor sediments from urban areas, or in municipal or industrial wastewater.^{40,41} Hence, besides the relevant research on pristine polymer mixtures,^{33,34} studies using realistic environmental samples with polymers that have been aged and weathered are also required. Since UV-light and heat cause degradations in polymers through chain scission, oxidation, crosslinking, and loss of side groups, these changes may manifest in ¹H NMR as new peaks, altered chemical shifts, peak broadening, and decreased intensity of original signals.^{42,43} Hence, NMR spectroscopy can serve as a tool to monitor and analyse the structural changes in polymers resulting from aging and weathering (Schmidt *et al.*, in prep). Moreover, consideration must be given to ensure that environmental samples go through a thorough sample preparation (extraction, filtration and purification) prior to NMR spectroscopy to avoid further potential overlaps with inorganic and organic compounds, whose proton signals appear in the lower ppm ranges (<3 ppm).²⁴

Another consideration is the relevance of the polymers that we can quantify so far. Since the dissolution of polymers currently limits the use of NMR spectroscopy, further research is necessary to find methods and solvents that make common polyolefins such as polypropylene (PP) and polyethylene (PE) more suitable for NMR spectroscopy. These polymers, along with PS, PVC and PU, represent the largest proportion of plastic production, and thus, the most prevalent in the environment,^{1,44} hence the demand for reliable monitoring tools. The quantification of polyolefins in polymer mixtures using qNMR spectroscopy remains a challenge that has not yet been fully overcome.³³ However, rubber-based polymers such as PB and PI, which originate from tire wear production, and the bioplastic PLA, which derives from renewable biomass sources, are becoming increasingly environmentally relevant polymers.^{45–47}

Conclusions

This study highlights the potential of qNMR spectroscopy for quantifying mixed polymer samples, offering critical insights into its application for complex environmental samples. Polymer mixtures of PS, PB, and PI in CDCl₃ show that low concentrations of PS can be effectively quantified by qNMR spectroscopy, with a relative error of –5%, when PS is present in both high and low concentrations. In the polymer mixtures, PB and PI can be quantified down to their previously measured

concentration limits for single polymers, using their first proton signal (H_a) at low concentrations, with relative errors of +5% and –3%, respectively. If one of these polymers is present at a high concentration, it can interfere with the detection and quantification of the other polymer at a lower concentration, compromising measurement accuracy. Additionally, the overlapping signal ranges for the second proton signals (H_b) of PB and PI make these signals unsuitable for quantifying their concentrations.

Analysis of polymer mixtures containing PS, PVC and PU, dissolved in THF-d₈, demonstrate that PVC concentrations can be reliably quantified in low concentration mixtures with a relative error of 0% and high concentration mixtures with relative errors of –2% and 4%. In contrast, the quantification of the measured concentration limit of PS varies from the nominal concentration due to the overlap with a proton signal of PU. Therefore, the measured concentrations of PU diverged severely from the nominal concentrations, and the third proton signal (H_c) especially seems to be less reliable basis for concentration calculations in mixtures.

Linear regression analyses in CDCl₃ demonstrated strong correlation (R^2 below 0.99) between the measured and nominal concentrations within the tested concentration range, while in THF-d₈, de, PS and PVC achieved excellent correlations (R^2 above 0.99), contrasting with PU's reduced correlation ($R^2 = 0.9548$). These findings underscore the importance of signal separation and solvent choice in achieving accurate quantification, particularly in complex polymer mixtures. Further refinement of qNMR methods is necessary to overcome the limitations posed by signal overlaps and to enhance its applicability to environmental sample analysis.

Data availability

The data supporting this article have been included as part of the ESI.†

Author contributions

J. S. designed the methodology, conducted the investigation, curated and analyzed the data, and wrote the original draft. J. S., M. H., and W. W. reviewed and edited the manuscript. J. S. and M. H. contributed to visualization. W. W. administered the project and acquired funding.

Conflicts of interest

There are no conflicts to declare.

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used ChatGPT by OpenAI to rephrase and improve the language.

References

- 1 Plastics – the fast Facts 2024 Plastics Europe [Internet]. Plastics Europe, 2024, <https://plasticseurope.org/knowledge-hub/plastics-the-fast-facts-2024/>.
- 2 A. L. Andrady and M. A. Neal, Applications and societal benefits of plastics, *Philos. Trans. R. Soc., B*, 2009, **364**, 1977–1984.
- 3 A. Lusher, Microplastics in the Marine Environment: Distribution, Interactions and Effects, in *Marine Anthropogenic Litter*, ed. M. Bergmann, L. Gutow and M. Klages, Springer International Publishing, Cham, 2015, pp. 245–307, DOI: [10.1007/978-3-319-16510-3_10](https://doi.org/10.1007/978-3-319-16510-3_10).
- 4 C. Arthur, H. Bamford and J. Baker, The Occurrence, Effects and Fate of Small Plastic Debris in the Oceans, *Proc Int Res Workshop Occur Eff Fate Microplastic Mar Debris Tacoma WA USA.*, 2008, pp. 9–11.
- 5 Y. Chae and Y. J. An, Effects of micro- and nanoplastics on aquatic ecosystems: Current research trends and perspectives, *Mar. Pollut. Bull.*, 2017, **124**, 624–632.
- 6 M. Cole and T. S. Galloway, Ingestion of Nanoplastics and Microplastics by Pacific Oyster Larvae, *Environ. Sci. Technol.*, 2015, **49**, 14625–14632.
- 7 S. Rist, A. Baun and N. B. Hartmann, Ingestion of micro- and nanoplastics in *Daphnia magna* - Quantification of body burdens and assessment of feeding rates and reproduction, *Environ. Pollut.*, 2017, **228**, 398–407.
- 8 Protection JG of E on the SA of ME, P. Kershaw, B. Carney Almroth, P. Villarrubia-Gómez, A. A. Koelmans and T. Gouin, *Proceedings of the GESAMP International Workshop on assessing the risks associated with plastics and microplastics in the marine environment*, Policy Commons, 2020, <https://policycommons.net/artifacts/8220098/proceedings-of-the-gesamp-international-workshop-on-assessing-the-risks-associated-with-plastics-and-microplastics-in-the-marine-environment/9133791/>.
- 9 S. Noventa, M. S. P. Boyles, A. Seifert, S. Belluco, A. S. Jiménez, H. J. Johnston, *et al.*, Paradigms to assess the human health risks of nano- and microplastics, *Microplast. Nanoplast.*, 2021, **1**, 9.
- 10 M. A. Browne, A. Dissanayake, T. S. Galloway, D. M. Lowe and R. C. Thompson, Ingested Microscopic Plastic Translocates to the Circulatory System of the Mussel, *Mytilus edulis* (L.), *Environ. Sci. Technol.*, 2008, **42**, 5026–5031.
- 11 A. A. Koelmans, E. Besseling, A. Wegner and E. M. Foekema, Plastic as a Carrier of POPs to Aquatic Organisms: A Model Analysis, *Environ. Sci. Technol.*, 2013, **47**, 7812–7820.
- 12 L. Rubio, R. Marcos and A. Hernández, Potential adverse health effects of ingested micro- and nanoplastics on humans. Lessons learned from in vivo and in vitro mammalian models, *J. Toxicol. Environ. Health, Part B*, 2020, **23**, 51–68.
- 13 A. L. Andrady, Microplastics in the marine environment, *Mar. Pollut. Bull.*, 2011, **62**, 1596–1605.
- 14 D. K. A. Barnes, F. Galgani, R. C. Thompson and M. Barlaz, Accumulation and fragmentation of plastic debris in global environments, *Philos. Trans. R. Soc. London, Ser. B*, 2009, **364**, 1985–1998.
- 15 M. G. J. Löder and G. Gerdtts, Methodology Used for the Detection and Identification of Microplastics—A Critical Appraisal, in M. Bergmann, L. Gutow and M. Klages, *Marine Anthropogenic Litter*, Cham, Springer International Publishing, 2015, pp. 201–227, DOI: [10.1007/978-3-319-16510-3_8](https://doi.org/10.1007/978-3-319-16510-3_8).
- 16 A. Tirkey and L. S. B. Upadhyay, Microplastics: An overview on separation, identification and characterization of microplastics, *Mar. Pollut. Bull.*, 2021, **170**, 112604.
- 17 E. Dümichen, P. Eisentraut, C. G. Bannick, A. K. Barthel, R. Senz and U. Braun, Fast identification of microplastics in complex environmental samples by a thermal degradation method, *Chemosphere*, 2017, **174**, 572–584.
- 18 E. Dümichen, A. K. Barthel, U. Braun, C. G. Bannick, K. Brand, M. Jekel, *et al.*, Analysis of polyethylene microplastics in environmental samples, using a thermal decomposition method, *Water Res.*, 2015, **85**, 451–457.
- 19 M. Fischer and B. M. Scholz-Böttcher, Simultaneous Trace Identification and Quantification of Common Types of Microplastics in Environmental Samples by Pyrolysis-Gas Chromatography–Mass Spectrometry, *Environ. Sci. Technol.*, 2017, **51**, 5052–5060.
- 20 P. Pořízka, L. Brunnbauer, M. Porkert, U. Rozman, G. Marolt, D. Holub, *et al.*, Laser-based techniques: Novel tools for the identification and characterization of aged microplastics with developed biofilm, *Chemosphere*, 2023, **313**, 137373.
- 21 L. Cabernard, L. Roscher, C. Lorenz, G. Gerdtts and S. Primpke, Comparison of Raman and Fourier Transform Infrared Spectroscopy for the Quantification of Microplastics in the Aquatic Environment, *Environ. Sci. Technol.*, 2018, **52**, 13279–13288.
- 22 N. Peez and W. Imhof, Quantitative ¹H-NMR spectroscopy as an efficient method for identification and quantification of PVC, ABS and PA microparticles, *Analyst*, 2020, **145**, 5363–5371.
- 23 N. Peez, M. C. Janiska and W. Imhof, The first application of quantitative ¹H NMR spectroscopy as a simple and fast method of identification and quantification of microplastic particles (PE, PET, and PS), *Anal. Bioanal. Chem.*, 2019, **411**, 823–833.
- 24 N. Peez, J. Becker, S. M. Ehlers, M. Fritz, C. B. Fischer, J. H. E. Koop, *et al.*, Quantitative analysis of PET microplastics in environmental model samples using quantitative ¹H-NMR spectroscopy: validation of an optimized and consistent sample clean-up method, *Anal. Bioanal. Chem.*, 2019, **411**, 7409–7418.
- 25 N. Peez, T. Rinesch, J. Kolz and W. Imhof, Applicable and cost-efficient microplastic analysis by quantitative ¹H-NMR spectroscopy using benchtop NMR and NoD methods, *Magn. Reson. Chem.*, 2022, **60**, 172–183.



- 26 G. Papini, G. Petrella, D. O. Cicero, C. Boglione and A. Rakaj, Identification and quantification of polystyrene microplastics in marine sediments facing a river mouth through NMR spectroscopy, *Mar. Pollut. Bull.*, 2024, **198**, 115784.
- 27 J. Schmidt, M. Haave, J. Underhaug and W. Wang, Unlocking the potential of NMR spectroscopy for precise and efficient quantification of microplastics, *Microplast. Nanoplast.*, 2024, **4**, 17.
- 28 S. K. Bharti and R. Roy, Quantitative ^1H NMR spectroscopy, *TrAC, Trends Anal. Chem.*, 2012, **35**, 5–26.
- 29 G. F. Pauli, B. U. Jaki and D. C. Lankin, Quantitative ^1H NMR: Development and Potential of a Method for Natural Products Analysis, *J. Nat. Prod.*, 2005, **68**, 133–149.
- 30 A. J. Brandolini and D. D. Hills, *NMR Spectra of Polymers and Polymer Additives*, CRC Press, 2000, pp. 23–117.
- 31 D. S. Achilias, C. Roupakias, P. Megalokonomos, A. A. Lappas and E. V. Antonakou, Chemical recycling of plastic wastes made from polyethylene (LDPE and HDPE) and polypropylene (PP), *J. Hazard. Mater.*, 2007, **149**, 536–542.
- 32 S. L. Wong, N. Ngadi and T. a. T. Abdullah, Study on Dissolution of Low Density Polyethylene (LDPE), *Appl. Mech. Mater.*, 2015, **695**, 170–173.
- 33 M. Günther and W. Imhof, Highly selective solid–liquid extraction of microplastic mixtures as a pre-preparation tool for quantitative nuclear magnetic resonance spectroscopy studies, *Analyst*, 2024, **149**, 5800–5811.
- 34 M. Günther and W. Imhof, Simultaneous quantification of microplastic particles by non-deuterated (NoD) ^1H -qNMR from samples comprising different polymer types, *Analyst*, 2023, **148**, 1151–1161.
- 35 C. I. Cerceau, L. C. A. Barbosa, E. S. Alvarenga, A. G. Ferreira and S. S. Thomasi, A validated ^1H NMR method for quantitative analysis of α -bisabolol in essential oils of *Eremanthus erythropappus*, *Talanta*, 2016, **161**, 71–79.
- 36 G. R. Fulmer, A. J. M. Miller, N. H. Sherden, H. E. Gottlieb, A. Nudelman, B. M. Stoltz, *et al.*, NMR Chemical Shifts of Trace Impurities: Common Laboratory Solvents, Organics, and Gases in Deuterated Solvents Relevant to the Organometallic Chemist, *Organometallics*, 2010, **29**, 2176–2179.
- 37 N. Schmid, S. Bruderer, F. Paruzzo, G. Fischetti, G. Toscano, D. Graf, *et al.*, Deconvolution of 1D NMR spectra: A deep learning-based approach, *J. Magn. Reson.*, 2023, **347**, 107357.
- 38 J. M. Pérez, C. Ruiz and I. Fernández, Synthesis of a Biodegradable PLA: NMR Signal Deconvolution and End-Group Analysis, *J. Chem. Educ.*, 2022, **99**, 1000–1007.
- 39 D. S. Dalisay and T. F. Molinski, NMR Quantitation of Natural Products at the Nanomole Scale, *J. Nat. Prod.*, 2009, **72**, 739–744.
- 40 H. K. Imhof, J. Schmid, R. Niessner, N. P. Ivleva and C. Laforsch, A novel, highly efficient method for the separation and quantification of plastic particles in sediments of aquatic environments, *Limnol. Oceanogr.:Methods*, 2012, **10**, 524–537.
- 41 M. Haave, C. Lorenz, S. Primpke and G. Gerdt, Different stories told by small and large microplastics in sediment - first report of microplastic concentrations in an urban recipient in Norway, *Mar. Pollut. Bull.*, 2019, **141**, 501–513.
- 42 M. Shang, Y. Wei, H. Zhou, T. Wu, K. Wang, H. Chen, *et al.*, Study on aging behavior of polyethylene glycol under three wavelengths of ultraviolet light irradiation, *RSC Adv.*, 2023, **13**, 34576–34586.
- 43 T. M. Alam, M. Celina, R. A. Assink, R. L. Clough, K. T. Gillen and D. R. Wheeler, Investigation of Oxidative Degradation in Polymers Using ^{17}O NMR Spectroscopy, *Macromolecules*, 2000, **33**, 1181–1190.
- 44 S. M. Brander, V. C. Renick, M. M. Foley, C. Steele, M. Woo, A. Lusher, *et al.*, Sampling and Quality Assurance and Quality Control: A Guide for Scientists Investigating the Occurrence of Microplastics Across Matrices, *Appl. Spectrosc.*, 2020, **74**, 1099–1125.
- 45 J. C. Prata, J. P. Da Costa, A. C. Duarte and T. Rocha-Santos, Methods for sampling and detection of microplastics in water and sediment: A critical review, *TrAC, Trends Anal. Chem.*, 2019, **110**, 150–159.
- 46 J. E. Tamis, A. A. Koelmans, R. Dröge, N. H. B. M. Kaag, M. C. Keur, P. C. Tromp, *et al.*, Environmental risks of car tire microplastic particles and other road runoff pollutants, *Microplast. Nanoplast.*, 2021, **1**, 10.
- 47 Z. Luo, X. Zhou, Y. Su, H. Wang, R. Yu, S. Zhou, *et al.*, Environmental occurrence, fate, impact, and potential solution of tire microplastics: Similarities and differences with tire wear particles, *Sci. Total Environ.*, 2021, **795**, 148902.

