

RESEARCH ARTICLE

View Article Online

View Journal | View Issue

Cite this: *Inorg. Chem. Front.*, 2025, 12, 7107

Feed supplementation with molybdenum complexes improves honey bee health†

Arcadie Fuior, ^{a,b} Loïc Colin-Duchevet, ^c Valentina Cebotari, ^d Amélie Noël, ^c Isabelle Ribaud, ^e Isabelle Gérard, ^a Olga Garbuz, ^d Mathieu Fregnaud, ^a Xavier López, ^f Virginie Larcher, ^c Michael A. Shestopalov, ^g Anastasiya O. Solovieva, ^h Tatiana N. Pozmogova, ^{g,i} Olesea Gliga, ^d Nadejda Railean, ^d Précillia Cochard, ^j Benjamin Poirot, ^j Leonidas Charistos, ^k Fani Hatjina, ^k Andrea Somogyi, ^l Kadda Medjoubi, ^l Sébastien Gaumer, ^m Aurelian Gulea, ^b Ion Todaras, ^{‡d} Jean-Christophe Sandoz ^{‡c} and Sébastien Floquet ^{‡a}

This study investigates the effects of dietary supplementation with molybdenum-based compounds on honey bee health. Among a series of dinuclear Mo(V) complexes, the most stable and non-toxic complexes were selected and tested over an extensive eight-year field campaign involving more than 700 beehives across diverse environmental conditions in Moldova, France, Greece and the United States. In a first part, we established that the administration of a few milligrams of the compounds Na-Mo₂O₄-EDTA or Li-Mo₂O₄-EDTA in spring or autumn enhanced colony performance: queen fecundity, hygienic behaviour, and honey production increased, while *Varroa destructor* infestation rates and winter losses were substantially reduced. A second part of the work focused on understanding these effects in beehives. Hive monitoring showed that the Mo-containing syrup can be consumed over 1.5 months and is well assimilated by larvae and workers. In particular, Mo levels increased significantly in the head of the bees. X-ray fluorescence measurements demonstrated that Na-Mo₂O₄-EDTA increases Mo levels in the brain, neurolemma and hypopharyngeal glands, which play a crucial role in honey bee health. The metabolism of Mo complexes was addressed using X-ray photoelectron spectroscopy (XPS) on bee faeces, which revealed that the complexes are oxidized into Mo(VI), suggesting that Mo complexes may function as anti-oxidant agents in bees. These findings offer promising solutions for the beekeeping industry, struggling with weakening honey bee colonies.

Received 31st March 2025,
Accepted 26th June 2025

DOI: 10.1039/d5qi00878f

rsc.li/frontiers-inorganic

1. Introduction

About 80% of all cultivated plant species and 40% of our diet directly depend on pollination.¹ The economic impact of insect pollinators, especially bees, has thus been estimated at

150 billion euros in 2005.² While the honey bee *Apis mellifera* plays a major role in insect-mediated pollination, it is gravely endangered by colony losses,³ which have systematically increased in the last decades through a multifactorial process involving biological agents (*Varroa destructor*, *Vairimorpha*,

^aInstitut Lavoisier de Versailles, Université Paris-Saclay, UVSQ, CNRS, UMR 8180, 78000 Versailles, France. E-mail: sebastien.floquet@uvsq.fr

^bState University of Moldova, MD-2009 Chisinau, Republic of Moldova

^cEvolution Genomes Behaviour & Ecology, University Paris-Saclay, CNRS, IRD, 91198 Gif Sur Yvette, France. E-mail: jean-christophe.sandoz@cnrs.fr

^dInstitute of Zoology, MD-2028 Chisinau, Republic of Moldova

^eUniversité Paris Saclay, CNRS, IN2P3, IJCLab, 91403 Orsay, France

^fUniversitat Rovira i Virgili, Departament de Química Física i Inorgànica, Marcel·lí Domingo 1, 43007 Tarragona, Spain

^gNikolaev Institute of Inorganic Chemistry SB RAS, 630090 Novosibirsk, Russia

^hResearch Institute of Clinical and Experimental Lymphology – Branch of the ICG SB RAS, 630090 Novosibirsk, Russia

ⁱNovosibirsk State University, 630090 Novosibirsk, Russia

^jApinov S.A.S, 17140 Lagord, La Rochelle, France

^kDepartment of Apiculture, Institute of Animal Science, ELGO 'DIMITRA', Nea Moudania, 63200, Greece

^lNanoscopium beamline, Synchrotron SOLEIL, L'Orme des Merisiers, Saint-Aubin, 91192 Gif-sur-Yvette, France

^mGenetics and Cell Biology Laboratory, Université Paris-Saclay, UVSQ, UR 4589 78180 Montigny-le Bretonneux, France

†Electronic supplementary information (ESI) available: Synthesis, stability studies, toxicity studies, details on test on beehives, monitoring of the Mo in beehives, assimilation of Mo by honey bees, synchrotron studies, XPS and anti-oxidative property studies. See DOI: <https://doi.org/10.1039/d5qi00878f>

‡These authors jointly supervised the study.



etc.), the use of pesticides, and a lack of resources among others.⁴ In this context, the development of new solutions to protect bees from these external aggressions and strengthen their health and resilience is therefore of great interest.

One way to improve honey bees' health and resilience to stress is to optimize their nutrition.^{5–7} This is a highly complex issue as it requires a perfect balance between macronutrients (carbohydrates, proteins, and lipids) and micronutrients (vitamins, minerals) to meet all the needs of honey bees. Floral sources are honey bees' major source of macronutrients. Nectar provides them with carbohydrates, while pollen supplies bees with lipids, proteins, and minerals. Pollen quality strongly depends on the plant species, the soil, and the time of year in which it is produced⁸ and honey bee health is directly correlated with the quality of pollen.^{9–11} These sources dynamically change in the landscape in both space and time, which can cause periods of deficiencies for honey bee colonies^{5,6} and this issue can be exacerbated by climate changes, which can potentially create mismatches between plants and honeybees.^{6,12}

To overcome nutritional limitations, beekeepers rely on food supplements usually given in spring at the beginning of the beekeeping season, during dearth periods, and in autumn to prepare the colonies for the wintering.⁵ This supplementation is usually in the form of sugar syrup or fondants, which primarily deliver carbohydrates to the colony but do not cover the other needs. Other additives are therefore needed to fill these gaps and prevent bees from feeding on risky sources containing toxic, inedible substances as much as possible.¹³ Interestingly a lot of products are used by beekeepers but very few of them are proven by scientific studies. In this field some researchers explored formulations containing vitamins,^{14,15} or proteins to boost colony strength or reduce mortality^{16–19} even if some other studies have reported that colonies fed protein supplements do not perform as well when compared with colonies receiving real pollen.²⁰ In contrast, a very recent study from Bogaert *et al.* reported impressive results on hives for the development of the colonies from May to October by supplementation with a mixture of sterols, especially isofucosterol, which appears as an essential nutrient for honeybees.²¹ More recently, the supplementation with pre and post-biotics emerged with the idea that the gut microbiome plays important roles in honey bees, including a role in honey bee colony health (pathogen defense) and nutrition.²²

Metals, are also important micronutrients for animals' health as they play an essential role in numerous physiological processes. However, surprisingly, they have been poorly studied in honey bees.²³ Honey bees likely obtain minerals from two main sources: pollen and "dirty" or turbid water. Yet, very little is known about the metals that are needed for honey bees' health.²³ Several studies have focused on the physiological role and/or toxicity of elements like Fe, Mn, Cu and Zn, as well as the deleterious effects of pollution by heavy elements such as As, Cd, and Pb.^{24–28} However, the role and the importance of trace elements for honey bees remains scarce,^{29–31} leaving us with limited knowledge of their potential role or the consequence of a deficiency in these elements.

Among trace elements, molybdenum (Mo) is of essential importance for life in almost all living organisms from bacteria to humans.³² Mo is present in about fifty enzymes, namely molybdoenzymes, which are essential constituents of the global carbon, sulfur and nitrogen metabolism in plants and animals. In these enzymes, Mo atoms play a key role in the catalysis of diverse redox reactions thanks to their oxidation state which can vary from Mo(+VI) to Mo(+IV).^{32,33} Molybdoenzymes are generally classified into different families. In eukaryotes, these enzymes mainly belong to the Sulfite Oxidase (SO) and Xanthine Oxidase (XO) families.³³ The role of SO is important for sulfite detoxification. XO molybdoenzymes are very useful for the degradation of xenobiotics and possess a broad substrate spectrum. As we demonstrate here, Mo is also present in honey bee foragers, with an average content of about 0.39 ppm (see Part I, ESI†) but to our knowledge its role in this insect is utterly unknown.

The objectives of this study are to fill this gap and thus to evidence that supplementing food for honey bees with stable, non-toxic, easy to handle, and bio-available simple molybdenum-based coordination complexes can induce positive effects on the global health of the colonies. A first part of this study is thus dedicated to the evaluation of the impact of supplementing bees feed with Mo-based compounds through an extensive eight-year campaign of field trials -conducted in Moldova, in France, in Greece and in the USA to identify the best complexes among a dozen of candidates, their effects in hives for the key parameters interesting beekeeping industry (development of the colony, productivity, mortality, action against pathogens), their optimal dose and their side effects, if so, in case of an overdosage.

For this, we chose coordination complexes of general formula (cation)_x[Mo₂O₂E₂(L)_y] (*x* = 0, 2, 4; *y* = 1 or 2, E = O or S), hereafter referred to as **Cation-Core-Ligand** combining the [Mo₂O₄]²⁺ or [Mo₂O₂S₂]²⁺ clusters with organic ligands, such as EDTA or cysteine (L-Cys), and counter cations which can be alkali or organic cations. The organic ligand should facilitate their uptake by bees and rapidly involve redox processes due to the reduced oxidation state of the Mo core. These complexes have been described in the literature as biomimetic models of molybdoenzymes since the 1970s³⁴ and more recently, it has been demonstrated that they have little to no toxicity^{35,36} with some showing potential for cyanide detoxification,³⁷ cell penetration capabilities,³⁸ and stimulation of biomass production, or acting as potent anti-oxidants.³⁶ Among a dozen previously published potential candidates,³⁶ three were selected for their high chemical stability (see Part II, ESI†) and their lack of toxicity (Part III, ESI†), especially when used as feed supplement for honey bees: [Mo₂O₄(EDTA)]^{2–}, [Mo₂O₂S₂(EDTA)]^{2–}, and [Mo₂O₂S₂(L-Cys)₂]^{2–} (see Fig. 1). [Mo₂O₄(EDTA)]^{2–} was used as PPh₄⁺, Na⁺ and Li⁺ salts (**PPh₄-Mo₂O₄-EDTA**, **Na-Mo₂O₄-EDTA** and **Li-Mo₂O₄-EDTA**), the sulphurated analogue [Mo₂O₂S₂(EDTA)]^{2–} was used as the PPh₄⁺ salt (**PPh₄-Mo₂O₂S₂-EDTA**) and [Mo₂O₂S₂(L-Cys)₂]^{2–} was used as the K⁺ salt (**K-Mo₂O₂S₂-LCys**).

A second part of this study aims to bring element to understand the mode of action of the most interesting complexes in



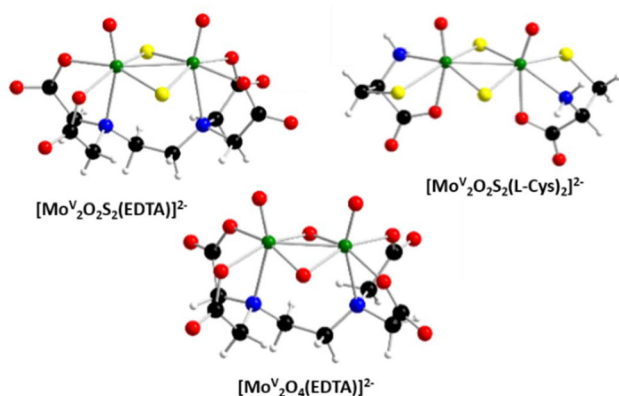


Fig. 1 Structures of the three complexes tested in beehives. Color code: Mo in green, S in yellow, O in red, N in blue, C in black, H in white.

beehives: cycle of Mo in the colonies, bio-assimilation, organs targeted by our molecules, metabolism and anti-oxidative properties.

2. Results

2.1 Tests in beehives

Field experiments aimed to assess the effects of these complexes on critical indicators of colony health, such as queen reproductive performance, honey production, levels of *Varroa destructor* infestation, and winter survival rates. The first field campaigns aimed to identify the most interesting complexes. Subsequent campaigns focused on the most promising results, particularly with regard to *Varroa* infestation, the maximum doses that can be administered without risk to the colony, the extension of tests under real operating conditions, and the study of the impact of these complexes on winter mortality in a geographical area particularly affected by these losses (California, USA).

Choice of the most efficient complex in beehives. The first two field test campaigns were performed in Moldova (forest environment 20 km outside the region of Chisinau) on 48 honey beehives of the *carpatica* ecotype of *Apis mellifera carnica*. The colonies were fed at the beginning of spring every second day over two weeks, with sugar syrup containing **PPh₄-Mo₂O₄-EDTA**, **PPh₄-Mo₂O₂S₂-EDTA** or **K-Mo₂O₂S₂-L-Cys** complexes for a total dose of 2 mg of complex per hive and compared to a control group fed only with syrup. More details and all raw data are available in Part IV of the ESI.† Among the three complexes, the best and most significant results were obtained with **PPh₄-Mo₂O₄-EDTA** (Table SIV.1, ESI†). Feeding with **PPh₄-Mo₂O₄-EDTA** induced a significant increase in queen fecundity (+10.5%, Mann-Whitney *U* test, $p = 1.5 \times 10^{-7}$), hygienic behaviour (+4.3%, *U*-test, $p = 8.8 \times 10^{-8}$), bee bread production (fermented stored pollen; +21.8%, *U*-test, $p = 8.8 \times 10^{-5}$), honey production (+19.6%, *U*-test, $p = 6.7 \times 10^{-6}$) and wax production (+39%, *U*-test, $p = 6.7 \times 10^{-6}$).³⁹ The sulphurated

complexes were further discarded, as they had poorer performance.

Choice of the most efficient salts of complex [Mo₂O₄(EDTA)]²⁻. Two more field campaigns were performed in Moldova to evaluate the influence of the counter cations PPh₄⁺, Na⁺ and Li⁺ associated with the complex [Mo₂O₄(EDTA)]²⁻, applying 2 mg of each compound per beehive. In addition to the previously monitored parameters, infestation by the mite *Varroa destructor*, a major pest for the honey bee, was also evaluated. All data are presented in Part IV of the ESI (Table SIV.2†), while the most significant results are depicted in Fig. 2. Queen fecundity increased by +13.3% and +11.7% for **Na-Mo₂O₄-EDTA** and **Li-Mo₂O₄-EDTA** but not significantly compared with the control (Kruskal Wallis test, $p = 0.574$). Conversely, the three complexes induced a decrease in the infestation of bee workers by the mite *Varroa destructor* (Kruskal Wallis test, $p < 0.001$). The lithium salt **Li-Mo₂O₄-EDTA** had the strongest effect (−42.9% on worker bees, multiple comparison Dunn's test, $p < 0.001$), while **PPh₄-Mo₂O₄-EDTA** and **Na-Mo₂O₄-EDTA** salts produced less pronounced and non-significant decreases (respectively −21.3%, $p = 0.36$ and −13.5%, $p = 1$). Finally, honey production increased by +13.0% for PPh₄-Mo₂O₄-EDTA ($p = 1$), by +42.9% for **Li-Mo₂O₄-EDTA** (Kruskal Wallis test with all data, $p = 0.41$; *U*-test compared to control, $p = 0.03$), and up to +49.6% for **Na-Mo₂O₄-EDTA** (Kruskal Wallis test with all data, $p = 0.13$; *U*-test compared to control, $p = 0.048$), which translated to an average honey production of 31.1 ± 3.0 kg per beehive vs. 20.8 ± 3.5 kg for the control group. Mo feeding did not affect honey quality. In particular, no trace of Mo was found in the produced honey, and all measured physical parameters of the honey fully complied with European Union regulations (Table SIV.3, Part IV, ESI†). In summary, these test campaigns established that the two alkali salts, **Li-Mo₂O₄-EDTA** and **Na-Mo₂O₄-EDTA**, are the most promising complexes. The most significant results of this campaign are depicted in Fig. 2A.

Effects against Varroa infestation. As lithium salts are known to have a deleterious effect on *Varroa*,⁴⁰ a further campaign in Moldova (2019) focused on the impact of the **Li-Mo₂O₄-EDTA** complex on *Varroa* infestation of worker bees and larvae. The complex was used at two dosages (2 and 6 mg per beehive), in comparison with a control group, and a group treated with lithium acetate as reference (LiOAc, 2 mg per beehive). The results are gathered in Table SIV.4 (Part IV, ESI†) and the most significant results are depicted in Fig. 2B.

The treatment had a significant effect on the worker bee infestation rate by *Varroa* (Kruskal Wallis test, $p = 0.0024$). With an overall dose of 2 mg of **Li-Mo₂O₄-EDTA** per beehive, infestation of worker bees by *Varroa* fell by −47% compared to the control group (multiple comparison Dunn's test, $p = 0.080$), similarly to the previous campaign (−42.9%). This effect increased to −61.3% with 6 mg per beehive (Dunn's test, $p = 0.002$). Despite a lithium concentration 3.7 times higher, the effect of lithium acetate was weaker and non-significant (−30.4%, $p = 1$). It could suggest that the Mo-complex in **Li-Mo₂O₄-EDTA** has a protective action against *Varroa*, in synergy with Li⁺ cations. Interestingly, the hygienic behaviour of hives



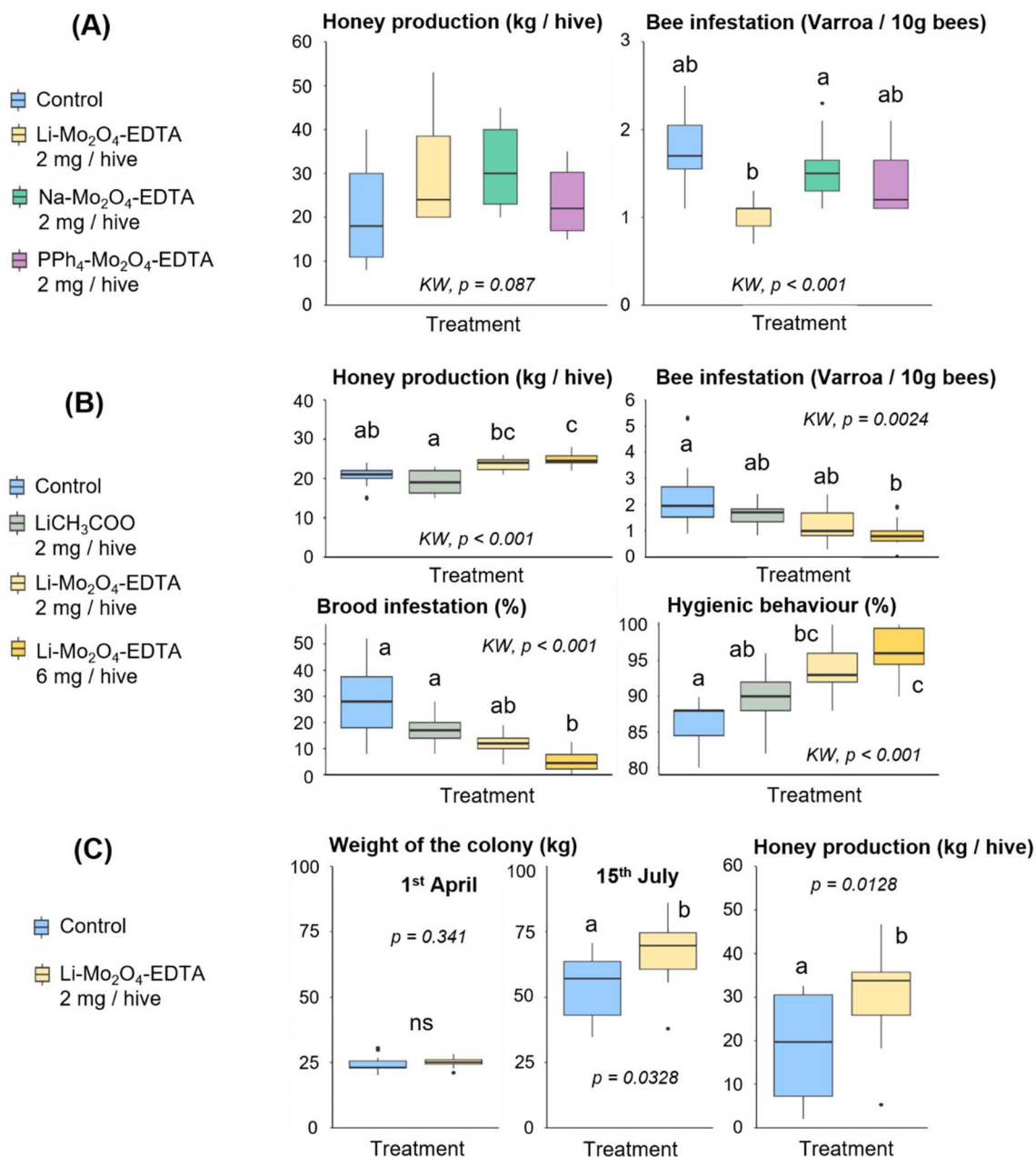


Fig. 2 Representative results obtained in the course of 3 test campaigns on honey bee colonies. (A) Effect of Na-Mo₂O₄-EDTA (2 mg per hive), Li-Mo₂O₄-EDTA (2 mg per hive) and PPh₄-Mo₂O₄-EDTA (2 mg per hive) on honey production and worker bee infestation by Varroa from the field test in Moldova (2018) (B) Effect of lithium acetate or Li-Mo₂O₄-EDTA at 2 and 6 mg per hive on worker bees and brood infestation by Varroa, hygienic behaviour and honey production from the field test in Moldova (2019); (C) Effect of Li-Mo₂O₄-EDTA at 2 mg per hive on colony weight and honey production from the field test in France (2019). For each campaign, $n = 10$ hives per treatment. Letters indicate significant differences between treatments ($p < 0.05$) based on Dunn's *post hoc* test with Bonferroni correction (A and B) or Mann-Whitney *U* test (C). Boxplots show median (horizontal crossbar) and interquartile ranges. KW: Kruskal-Wallis.

fed with 2 and 6 mg of **Li-Mo₂O₄-EDTA** concomitantly increased by +9.3% ($p = 0.007$) and +12% ($p < 0.001$), respectively. It is known that honey bee colonies with higher hygienic behaviour tend to keep *Varroa* infestation at lower levels.⁴¹

We also observed a significant effect of the treatment on the brood infestation rate by *Varroa* (Kruskal Wallis test, $p < 0.001$). Interestingly, *Varroa* infestation was very high in

control hives, *i.e.* 28.2%. After feeding with the reference compound LiOAc (2 mg per hive), the rate was lowered by –35.1% in this group (Dunn's test, $p = 1$). Again, the **Li-Mo₂O₄-EDTA** complex produced stronger effects, reducing brood infestation by –57.4% compared to the control group with 2 mg (Dunn's test, t , $p = 0.067$), and up to –81.6% with 6 mg per beehive (Dunn's test, $p < 0.001$). Beyond this result, it also suggests



that the complex is consumed not only by adult workers, but is also transferred to the brood.

Test under operating conditions. The feeding protocol employed in earlier campaigns proved impracticable under professional beekeeping conditions, as it required administration every two days.

A new test campaign was thus carried out in France, with the feeding protocol adapted to commercial beekeeping practices. The test was performed on 22 beehives of *Apis mellifera* “Buckfast”, widely used in beekeeping, in Gif-sur-Yvette (France), split into two groups: 11 control and 11 test colonies. For the test colonies, 2 mg of **Li-Mo₂O₄-EDTA** complex were introduced on 1st April in one 0.5 L syrup feeding. We monitored colony weight and the quantity of honey produced on 15th July (Table SIV.5, Part IV, ESI†). The results depicted in Fig. 2C show that a single dose of **Li-Mo₂O₄-EDTA** complex introduced at the beginning of spring resulted in a +23.6% increase in colony weight compared to the control group (*U*-test, *p* = 0.033), and in a +58.7% increase in honey production (30.3 kg on average for the test group vs. 19.1 kg for the control group, *U*-test, *p* = 0.014). This result is similar to the one from the previous campaign with 2 mg of the **Li-Mo₂O₄-EDTA** (+47%, *U*-test compared to control group, *p* = 0.03) in Moldova on *Apis mellifera carpatica* after multiple feedings, which validates the efficacy of the complex when applied in a single feeding, under professional beekeeper practices in a second geographical location and on another honey bee subspecies.

Impact on winter colony mortality. After assessing the effects of molybdenum complexes on colony development, *Varroa destructor* levels, and overall productivity, a dedicated study was initiated to investigate their potential role in mitigating one of the most pressing issues in beekeeping —winter mortality, which reaches up to 30–40% in Europe and more than 80% in other regions of the world each year.^{3,19,42–48} These losses can increase at alarming levels as measured in USA from 2024 to 2025 with 62% losses on average, mainly between autumn 2024 and spring 2025,⁴⁸ which seriously endangers the beekeeping industry and the plants and crops that depend on pollination by bees.

Two test campaigns were conducted during the winters of 2019 and 2020 in apiaries in the San Francisco Bay area (USA), a region consistently reporting high levels of winter colony losses.^{45–48} In the first campaign, 151 beehives were distributed across 6 different apiaries. In each apiary, the colonies were randomly divided into a control group (76 hives), receiving 1 US Gallon (3.78 Liters) of sugar syrup, and a test group (75 hives) receiving syrup containing 4 mg of **Li-Mo₂O₄-EDTA** complex. The feeding took place at the end of October 2019, and the surviving colonies were counted on 7th January 2020. In the control group, 47 colonies were lost out of 76, *i.e.* 61.8% winter mortality, while in the test group only 26 colonies were lost, *i.e.* 34.7% winter mortality. This 43.8% mortality rate decrease was significant (χ^2 test, *p* = 0.0008).

A second campaign (winter 2020) involved 220 beehives, distributed across 11 apiaries. The beehives were divided into

4 groups of 55 colonies. Each group was fed twice, in September and in October, with 1 US gallon of syrup each time for a total of 2 US gallons for each beehive. The control group received only syrup, the second group received 8 mg **Li-Mo₂O₄-EDTA** in September only, the third one received the same dosage in October only, and the fourth group received 4 mg of the complex in both feedings. In summary, the three test groups of colonies received 8 mg of the complex and only differed in the supplementation schedule. Lost colonies were counted on 31st December, 2020. The period of supplementation strongly and significantly impacted the results. The mortality rate in the control group was 15/55, *i.e.* 27.3% loss. The supplementation in October reduced mortality by 33% compared to the control group (10/55, *i.e.* 18.2% loss), which was not significant (χ^2 test, *p* = 0.25). The supplementation in September resulted in no winter losses (0/55, 0%, χ^2 test, *p* = 3.1 $\times 10^{-5}$). And finally, supplementation in both September and October resulted in 3 lost colonies out of 55 (5.5% loss, χ^2 test, *p* = 0.002), corresponding to 80% reduction in winter mortality compared to the control group.

Impact of an elevated dosage in beehives/tolerance studies.

After establishing that Mo complexes are effective in improving multiple health parameters in honey bee colonies by using only a few milligrams, two additional field studies were conducted to evaluate the impact of elevated dosage levels of the compounds. In a first test campaign, carried out from December 2020 to April 2021 in Nea Moudania region, Greece, we focused on the **Li-Mo₂O₄-EDTA** complex (see section IV.4 of Part IV, ESI† for more details). Thirty beehives were divided into 3 groups: a control group, a group receiving a global dose of 40 mg of **Li-Mo₂O₄-EDTA** in a candy bread (4 kg at 10 mg kg⁻¹) in December (“MoLi-B” batch) and a group receiving 80 mg of **Li-Mo₂O₄-EDTA** in a candy bread in December (4 kg at 10 mg kg⁻¹: 40 mg in total) and then in syrup in March (4 liters at 10 mg L⁻¹, 40 mg in total, “MoLi-A” batch). The size of the colonies and the number of dead bees at the entrance of the colonies (using Gary traps) were monitored throughout the whole period (see Part IV, ESI†). During the first period (December to March), we found that feeding the bee colonies with high **Li-Mo₂O₄-EDTA** dosage (colonies from both MoLi-A and MoLi-B groups) had deleterious effects. On average, about 185 dead bees were found in front of the colonies in these groups, compared to about 130 for the control group (*t*-test, *p* = 0.0001). This effect was stronger during the second period (March to April) in the “MoLi-A” group (160 dead bees per hive vs. 97 for the control group, *p* = 0.0001), while the increased mortality stopped for the MoLi-B group (57 dead bees per hive during the second period). While feeding 2–8 mg of complex **Li-Mo₂O₄-EDTA** appears to have very positive effects in hives, overdosing with 40 and 80 mg per hive increases bee mortality.

For the sake of comparison, the impact of a high **Na-Mo₂O₄-EDTA** dosage on mortality was evaluated in a field campaign performed in the west of France (see Part III.5 of the ESI† for more details) from May to the end of June 2022. 40 colonies divided into five groups of 8 beehives were each fed with 4 L of sugar syrup (2 times 2 L with 1 week between the



two feedings), and received 0, 4, 8, 16, and 80 mg of **Na-Mo₂O₄-EDTA** per beehive, respectively. Mortality was monitored in all groups over 2 months with Gary traps positioned in front of the hives. No significant difference in mortality was measured between hives supplemented with **Na-Mo₂O₄-EDTA** and control hives, even at the highest dose (80 mg, $p = 0.57$, see Fig. SIII.24, Part III, ESI†). Conversely to **Li-Mo₂O₄-EDTA**, **Na-Mo₂O₄-EDTA** demonstrated a superior safety profile, even at high concentrations, supporting its potential for safe, large-scale apicultural use.

2.2 Tracking molybdenum in beehives

Since supplementation with the complexes showed positive effects several months after administration, it was important to monitor the molybdenum cycle within the beehives to understand the underlying reasons. To this end, the control and 8 mg **Na-Mo₂O₄-EDTA** groups from the prior experiment were employed.

Impact of Mo supplementation on the food stored in brood frames. To understand the long-term effect of supplementation, we studied the Mo contents in the samples of food stored in the brood frames (which serve to produce honey for the colony, see Fig. 3A) from the feeding to two months after feeding.

At Day 0 (D0), the level of Mo in the colonies' food stores was below 0.1 ppm in all groups (Table SV.1, Part V, ESI†). At D28 and D42 the Mo content matched the levels of Mo from the syrups used for the feeding, showing that the syrup was not yet consumed. At D56, the Mo level was again below 0.1 ppm in almost all groups. The ¹H NMR studies were additionally conducted on these samples, revealing the absence of disaccharides and confirming it is pure and unadulterated honey. This study demonstrates that the syrup is stored and consumed over several weeks by the colonies (over 1.5 months in our case). This gradual consumption enables several generations of bees to be fed with **Na-Mo₂O₄-EDTA**. Once the feeding is consumed, we confirmed that the supplementation does not impact the honey produced by the colony.

Mo assimilation in worker bees and larvae. Finally, we aimed to determine Mo assimilation in worker bees and larvae. The bee samples were collected over 2 months after the feeding (May–June 2022). The Mo content was measured in all collected samples (see Fig. 3 and Part V of the ESI† for further details).

Fig. 3B and C show the variations in Mo-contents in bee larvae and in worker bees between D0 and D56 for control and 8 mg per hive batches. In bee larvae, at D0, Mo levels were similar in the control and 8 mg per beehive groups: 0.24 vs. 0.26 ppm. After two weeks (D14) a difference of +42% in Mo levels was measured in the 8 mg per hive group (U -test, $p = 0.02$). This difference increased again at D28, reaching +53% Mo (U -test, $p = 0.004$). At D56, the difference between the two groups decreased and was not significant ($p = 0.36$).

In the worker bees, Mo levels were also similar at D0 in the control and 8 mg per beehive groups: around 0.4 ppm. At D14, the Mo level increased by +68% in the 8 mg per hive group

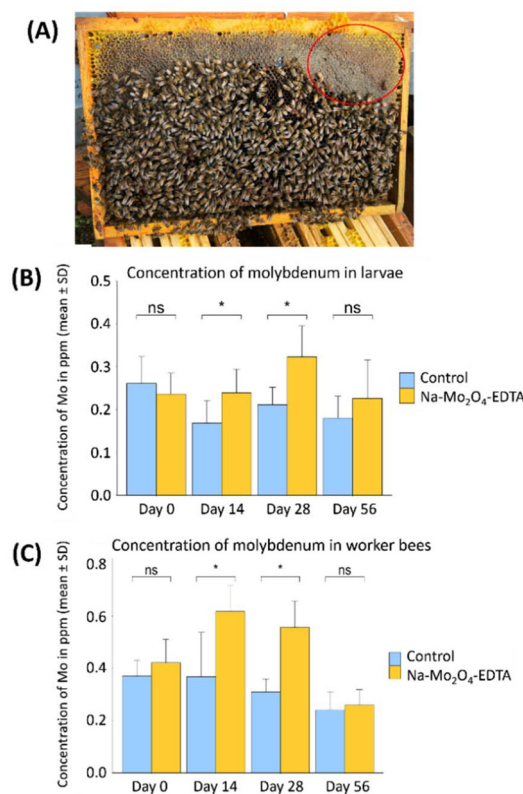


Fig. 3 (A) A typical frame found in the experimental beehives. The red circle indicates the zone where food is stored and where the samples were taken; (B) Mo content in larvae in $\mu\text{g g}^{-1}$ sampled at D0, D14, D28, D56 for the control group and the group treated with **Na-Mo₂O₄-EDTA** at 8 mg per beehive; (C) Mo content in worker bees in ppm sampled at D0, D14, D28, D56 for the control group and the group treated with **Na-Mo₂O₄-EDTA** at 8 mg per beehive. Non-parametric U -tests were performed. Differences at D0 and D56 are not significant (ns), while Mo-contents significantly differ between groups for larvae and worker bees at D14 and D28 (*: $p < 0.05$).

(0.62 ppm, U -test, $p = 0.01$), reaching +80% at D28 ($p = 0.001$). The workers consumed the syrup and assimilated the complex of molybdenum. At D56, Mo levels were again similar in both groups ($p = 0.46$). This experiment demonstrates the assimilation of Mo by larvae and worker bees over at least 1 month after the supplementation.

2.3 Effect of chronic feeding of bees in laboratory conditions

To further confirm the safety of Mo-complexes for honey bees, a chronic feeding experiment was conducted in laboratory conditions.

Three different concentrations were used in addition to a control group: 2, 20 and 400 mg L^{-1} for both **Na-Mo₂O₄-EDTA** and **Li-Mo₂O₄-EDTA** complexes. For each group, 3 cages of 50 workers collected at emergence were fed *ad libitum* with water and with Mo-enriched sugar solution. The survival was monitored throughout their lifetime (Fig. 4).

The survival curves are depicted in Fig. 4B and C (see also section III.4, Part III, ESI†). We found that the average curves



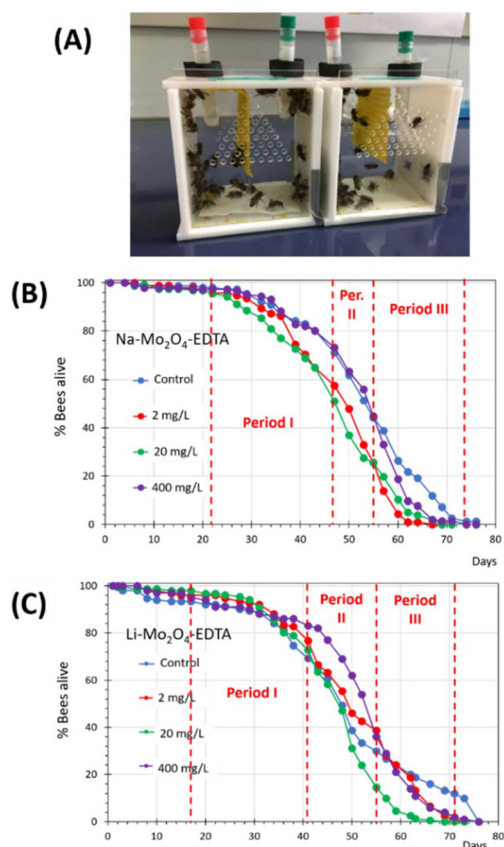


Fig. 4 Population cages used in this experiment (A); mortality curves obtained with complex $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ (B) and $\text{Li-Mo}_2\text{O}_4\text{-EDTA}$ (C) highlighting the 3 periods defined to follow Mo and Na contents in bees; controls are indicated in blue, 2 mg L⁻¹ solution in red, 20 mg L⁻¹ in green, 400 mg L⁻¹ in purple.

obtained for control batches and for bees fed with 2, 20 and 400 mg L⁻¹ solutions of both complexes are similar. The statistical analysis confirms that no difference in mortality appeared between the control group and the groups fed with both $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ and $\text{Li-Mo}_2\text{O}_4\text{-EDTA}$ during the first 10 days (Table SIII.10, Part III, ESI[†]), and for the whole period for all concentrations of $\text{Li-Mo}_2\text{O}_4\text{-EDTA}$ (Table SIII.11, Part III, ESI[†]). In contrast, the Cox model indicates a slight negative impact of $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ at 2 and 20 mg L⁻¹ but a positive impact at 400 mg L⁻¹. In summary, no notable positive or negative effect of feeding with Mo-complexes was observed on bees' longevity in laboratory conditions, even at a very high concentration.

2.4 Assimilation of Mo by the honey bees in laboratory

Dead bees collected in the previous experiment were used to evaluate the assimilation of Mo complexes within the body of the bees. The dead bees were divided into 3 batches (periods I, II, III, see Fig. 4B and C) depending on feeding duration. The Mo content was determined in heads, thoraxes, and abdomens by ICP-MS (see Table 1 and Tables SVI.2, SVI.3, Part VI, ESI[†]). First, we found low Mo levels in the control bees, with values

respectively around 0.2 ppm for $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ groups and 0.5–0.7 ppm for $\text{Li-Mo}_2\text{O}_4\text{-EDTA}$ groups in the head and in the abdomen, and lower levels in the thorax (0.12 and 0.25 ppm respectively). An increase in Mo levels was found in the head, thorax and abdomen when the concentration of $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ or $\text{Li-Mo}_2\text{O}_4\text{-EDTA}$ increased in the feeding syrup. For each concentration, the Mo level reached a different plateau which did not depend on the duration of feeding. More precisely, Mo levels in the head increased from ~0.19 ppm in the control group, to 0.58, 1.83 and 27 ppm, respectively for the groups fed with syrups at 2, 20 and 400 mg L⁻¹ of $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$, *i.e.* up to a 140-fold increase. At the same time, Mo-level variations were higher in the thorax, from 0.12 ppm to 0.73, 2.09 and 58 ppm with the same syrups. Similar observations were made with $\text{Li-Mo}_2\text{O}_4\text{-EDTA}$, with levels reaching 34.8 ppm in the head and 75.2 ppm on average in the thorax. With both complexes, as bees defecate very little in cages, strong accumulation was observed in the bees' abdomen, in particular in the faeces (up to 3600 ppm), and so these parts were further excluded from the experiment. In the case of $\text{Li-Mo}_2\text{O}_4\text{-EDTA}$, an accumulation of Lithium was also observed, in particular in the head, in which we estimate that 5.6 Li⁺ are assimilated for each Mo atom (see Table 1 and Fig. SVI.5, Part VI, ESI[†]).

2.5 X-Ray fluorescence studies

As it was confirmed that Mo levels are increased in bees after the supplementation, another study was conducted to understand in which organs it is assimilated. X-ray fluorescence spectra were recorded on the Nanoscopium beamline of Synchrotron Soleil (Gif-sur-Yvette, France) on 20–50 μm-thick slices of head, thorax, and abdomen, and on hypopharyngeal glands extracted from the heads of worker bees. The study was conducted on bees fed over 14 days after their emergence with a sugar syrup containing $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ at 400 mg L⁻¹ or pure syrup for the control group. The analysis of the slices and glands of bees not supplemented with $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ (see Fig. 5A–C) qualitatively revealed that Mo-content (i) is negligible in the brain, (ii) it is low in the neurolemma, hypopharyngeal glands and muscles (see Fig. 6A, 7A, and Fig. SVII.9, SVII.13, SVII.16, Part VII, ESI[†]) as evidenced by a shoulder on the X-ray fluorescence spectra at the energy expected for the element Mo (16.9 to 17.9 eV) and (iii) it is moderate but at least two times higher in the cuticle parts, as shown in the Fig. SVII.18 (Part VII, ESI[†]).

Fig. 5D–H show the distribution of Mo on a 20 μm thick slice of the head of a bee fed with the complex $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$. This slice contains elements of the brain, neurolemma, cuticle and oesophagus. No difference was observed in the intensity of the fluorescence of Mo in the cuticle (external cuticle and tentorial arms) and in the muscles in comparison with control bees (see Fig. SVII.18–SVII.20, Part VII, ESI[†]). More interestingly, the fluorescence of Mo is clearly seen in the brain and found especially in the neurolemma around the brain (Fig. 5F and G), in contrast with control bees (Fig. 5B). The Mo level appears slightly enhanced within the brain after feeding from an intensity of *ca.* 70–80 counts in control bees



Table 1 Mo and Na/Li contents in ppm ($\mu\text{g g}^{-1}$ of dried bee) in the head, thorax and abdomen of bees fed with the complexes Na-Mo₂O₄-EDTA and Li-Mo₂O₄-EDTA in mortality tests

Complex	Modality	Periods	Head		Thorax	
			Mo (ppm)	Na (ppm)	Mo (ppm)	Na (ppm)
Na-Mo ₂ O ₄ -EDTA	Control	I	0.20 ± 0.01	720 ± 40	0.12 ± 0.01	350 ± 5
		II	0.20 ± 0.01	650 ± 50	0.13 ± 0.01	375 ± 3
		III	0.17 ± 0.01	680 ± 30	0.11 ± 0.01	330 ± 10
	Na-Mo ₂ O ₄ -EDTA 2 mg L ⁻¹	I	0.70 ± 0.01	728 ± 9	0.6 ± 0.1	380 ± 10
		II	0.63 ± 0.01	710 ± 10	0.9 ± 0.3	400 ± 60
		III	0.42 ± 0.01	710 ± 6	0.7 ± 0.1	370 ± 30
	Na-Mo ₂ O ₄ -EDTA 20 mg L ⁻¹	I	2.02 ± 0.01	761 ± 8	2.06 ± 0.01	340 ± 20
		II	1.33 ± 0.01	661 ± 3	2.12 ± 0.04	430 ± 20
		III	2.15 ± 0.75	760 ± 10	2.08 ± 0.08	409 ± 8
	Na-Mo ₂ O ₄ -EDTA 400 mg L ⁻¹	I	30 ± 1	830 ± 5	51 ± 5	500 ± 60
		II	28 ± 1	834 ± 4	80 ± 25	530 ± 20
		III	23 ± 1	1003 ± 4	43 ± 15	540 ± 6
Complex	Modality	Periods	Head		Thorax	
			Mo (ppm)	Li (ppm)	Mo (ppm)	Li (ppm)
Li-Mo ₂ O ₄ -EDTA	Control	I	0.66 ± 0.07	0.06 ± 0.01	0.27 ± 0.01	0.03 ± 0.01
		II	0.46 ± 0.02	0.04 ± 0.01	0.26 ± 0.01	0.03 ± 0.01
		III	0.40 ± 0.01	0.04 ± 0.01	0.23 ± 0.01	0.03 ± 0.01
	Li-Mo ₂ O ₄ -EDTA 2 mg L ⁻¹	I	1.86 ± 0.01	1.06 ± 0.01	3.07 ± 0.02	1.31 ± 0.02
		II	1.48 ± 0.01	0.84 ± 0.02	2.31 ± 0.04	0.93 ± 0.01
		III	1.62 ± 0.04	1.11 ± 0.02	2.00 ± 0.04	1.23 ± 0.01
	Li-Mo ₂ O ₄ -EDTA 20 mg L ⁻¹	I	4.36 ± 0.04	1.77 ± 0.01	7.81 ± 0.04	2.27 ± 0.02
		II	3.72 ± 0.01	2.35 ± 0.01	5.50 ± 0.07	2.53 ± 0.02
		III	3.39 ± 0.02	1.58 ± 0.01	3.45 ± 0.07	1.80 ± 0.02
	Li-Mo ₂ O ₄ -EDTA 400 mg L ⁻¹	I	31.9 ± 0.1	10.89 ± 0.08	63.1 ± 0.4	16.3 ± 0.1
		II	37.6 ± 0.2	14.0 ± 0.1	58.6 ± 0.2	18.87 ± 0.05
		III	35.03 ± 0.03	16.5 ± 0.2	104.0 ± 0.3	24.9 ± 0.1

(Fig. 5C), comparable to the baseline, to *ca.* 130–140 counts in fed bees (Fig. 5H).

Concomitantly, the increase of Mo in the neurolemma is much more pronounced. From several X-ray spectra recorded in the same experimental conditions for several individuals from both groups (Fig. 6 and Fig. SVII13 and SVII14, Part VII, ESI†), Mo-content in bees fed with Na-Mo₂O₄-EDTA appeared around 10-fold higher than in the control group, showing an accumulation of Mo in the neurolemma.

Hypopharyngeal glands. The hypopharyngeal glands (HPGs) found in the head of honey bee workers are crucial for bees' nutrition and health.^{49–51} Glands were taken from control bees and bees fed with the complex Na-Mo₂O₄-EDTA at 400 mg L⁻¹ for 14 days and were analyzed by X-ray fluorescence (Fig. 7 and Fig. SVII-16, SVII-17, Part VII, ESI†).

In control bees (Fig. 7A), the fluorescence spectrum reveals a low presence of Mo in the glands. Feeding with a sugar syrup containing the Na-Mo₂O₄-EDTA complex induced a remarkable fluorescence peak, as shown in Fig. 7B. The results obtained for 7 *acini* of hypopharyngeal glands from 2 individuals are similar (see Part VII, ESI†). A 14× increase in Mo content was estimated in bees' hypopharyngeal glands after feeding bees with Na-Mo₂O₄-EDTA at 400 mg L⁻¹. These results show that the hypopharyngeal glands are a primary target for the Mo complex, which increases the Mo level naturally present in

these glands and, due to the importance of these glands, could explain its impact on the health of the bees.

2.6 Metabolism and antioxidant properties of Mo-complexes within the bee organism

The last part of this study aimed to provide elements for understanding the metabolism of the Mo complexes in the honey bee organism.

X-ray photoelectron spectroscopy (XPS) experiments were performed on the faeces of individuals fed with Na-Mo₂O₄-EDTA at 400 mg L⁻¹. The Mo 3d spectral region of Na-Mo₂O₄-EDTA and of bee faeces, as well as their associated reconstruction, are presented in Fig. 8.

The Na-Mo₂O₄-EDTA spectrum can be resolved into three doublets, with the most intense corresponding, as expected, to the Mo^{+V} oxidation state (Mo 3d_{5/2} = 231.2 eV, full width at half maximum FWHM = 1.4 eV).⁵² For the bee faeces, the Mo 3d core level spectrum displays only one broad doublet (Mo 3d_{5/2} at BE 232.7 eV, FWHM = 3.5 eV). The energy position is fully consistent with a Mo^{+VI} phase.⁵² The presence of Mo in +V or +IV oxidation states was not identified. This experiment thus unambiguously shows that the complex Na-Mo₂O₄-EDTA was oxidized in the bees, probably into the molybdate anion MoO₄²⁻. It constitutes the first element suggesting that the Na-



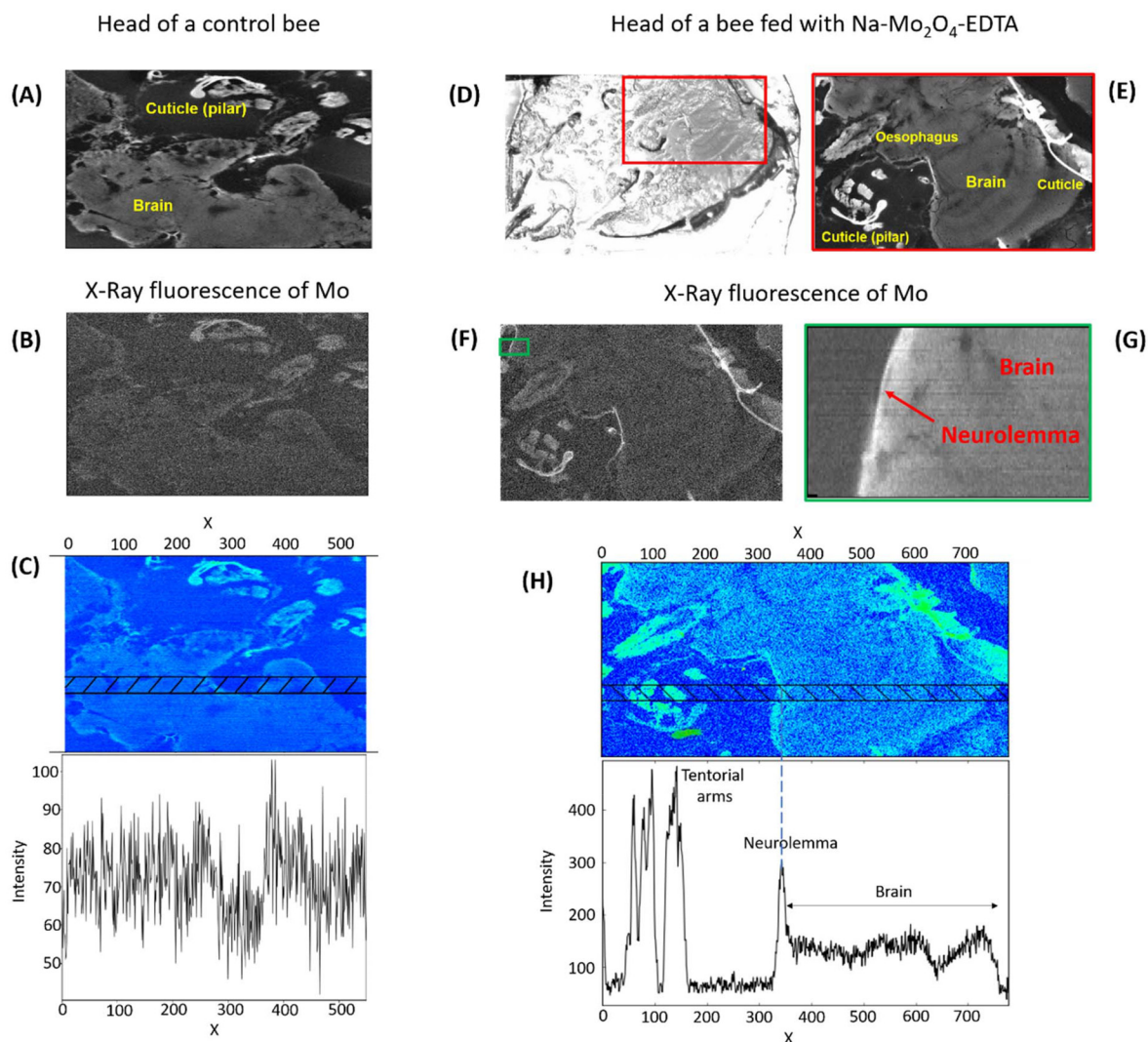


Fig. 5 Comparisons of heads of a worker bee control (left part) and a bee fed with Na-Mo₂O₄-EDTA (right part). Control bee: picture of 20 μm thick slice of the head obtained by X-ray fluorescence of all elements (A) or just by X-ray fluorescence of Mo of a portion of brain of a control bee (B); (C) Variation of intensity of the X-ray fluorescence of Mo in the hatched area (in the energy range 16.9 to 17.9 eV) along X in a 20 μm thick slice of the head of a control worker bee. Bee fed with Na-Mo₂O₄-EDTA: picture of a 20 μm thick slice of the head of a worker bee obtained by microscope (D), by X-ray fluorescence of all elements in the red rectangle (E, size 2000 $\mu\text{m} \times 1000 \mu\text{m}$, pixel size 2 \times 2 μm^2 , acquisition time per pixel 100 ms), and by X-ray fluorescence of Mo in the whole zone (F) or focused on the neurolemma (G, green rectangle of size 181 \times 64 μm^2 ; pixel size 1 \times 1 μm^2 , acquisition time per pixel 100 ms); (H) variation of intensity of the X-ray fluorescence of Mo in the hatched area (in the energy range 16.9 to 17.9 eV) along X in a 20 μm thick slice of the head of a worker bee, showing the presence of Mo in tentorial arms (cuticle), neurolemma and brain.

Mo₂O₄-EDTA complex may act as an antioxidant by oxidation of the Mo(V) atoms into the Mo(VI) oxidation state.

The antioxidant activity (AOA) of the complexes **Na-Mo₂O₄-EDTA** and **Li-Mo₂O₄-EDTA** was evaluated through field tests performed on *Apis mellifera carnica*. Experimental bee-hives were fed at the beginning of spring for 14 days with 50% sugar syrup enriched or not with **Na-Mo₂O₄-EDTA** and **Li-Mo₂O₄-EDTA** at 0.2 mg L⁻¹. Sodium molybdate Na₂MoO₄·2H₂O was also used, for comparison, with a Mo content equivalent to that of the **Na-Mo₂O₄-EDTA** and **Li-Mo₂O₄-EDTA** compounds. The AOA was evaluated by the ABTS method⁵³ on hemolymph from worker bees and larvae, wax, honey, propolis, royal jelly, and bee bread. The results are

expressed as IC₅₀ values, the half-maximal Inhibitory Concentration, defined as the concentration that causes the loss of 50% of the activity of ABTS radicals. The lower the IC₅₀ values, the higher the antioxidative activity. Some selected results are depicted in Fig. 9 while the other results are presented in the ESI (Fig. SVIII-6–SVIII-12, Part VIII, ESI†).

As shown in Fig. 9, the IC₅₀ values obtained for the hemolymph of worker bees are low, indicating a naturally good antioxidant activity (Fig. 9A). The AOA of hemolymph in worker bees increased for both **Na-Mo₂O₄-EDTA** and **Li-Mo₂O₄-EDTA** complexes compared to the control group, suggesting that these complexes act on the organisms of bees as antioxi-



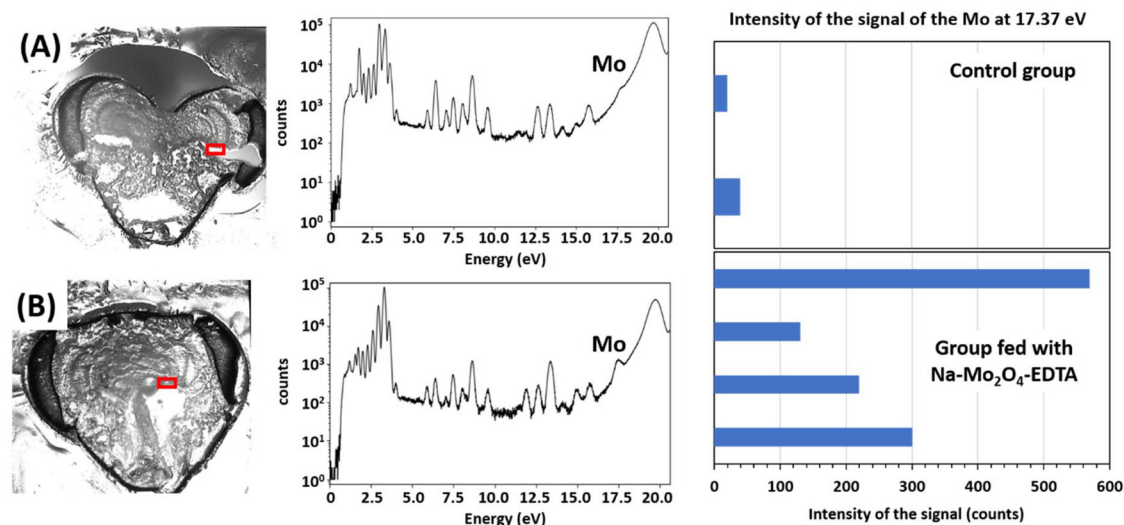


Fig. 6 Pictures of 40 μm thick slices of the head of a worker bee from the control group (A) and bees fed with $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ (B) with the corresponding fluorescence spectra of all elements in the red rectangle zones corresponding to the neurolemma ((A) Pixels $0.5 \times 0.5 \mu\text{m}^2$, size $100 \times 60 \mu\text{m}^2$, acquisition time per pixel 300 ms; (B) Pixels $0.5 \times 0.5 \mu\text{m}^2$, size $136 \times 33 \mu\text{m}^2$, acquisition time per pixel 300 ms). The energy of the spectrum corresponding to Mo is indicated.

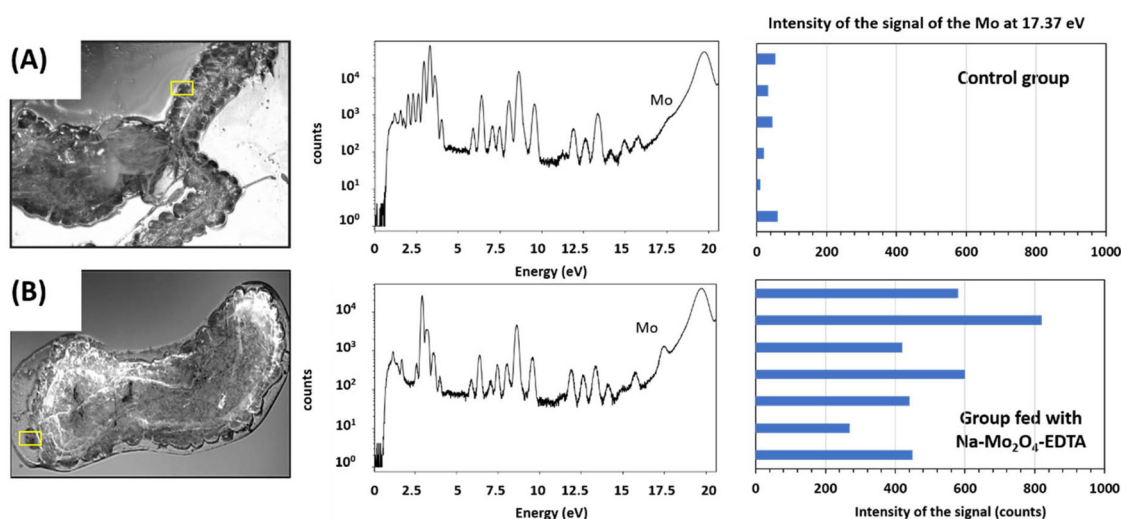


Fig. 7 Hypopharyngeal glands extracted from workers of the control group (A) and bees fed with $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ (B) with their corresponding total X-ray fluorescence spectra. The yellow rectangles indicate the part of the gland on which the fluorescence spectrum was measured ((A) pixels $0.5 \times 0.5 \mu\text{m}^2$, size $108 \times 49 \mu\text{m}^2$, acquisition time per pixel 200 ms; (B) pixels $0.5 \times 0.5 \mu\text{m}^2$, size $70 \times 80 \mu\text{m}^2$, acquisition time per pixel 200 ms).

dants or induce AOA. The AOA of both complexes appears similar, slightly better with $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$. Interestingly, even though sodium molybdate Na_2MoO_4 cannot chemically act as a direct antioxidant, the AOA of the hemolymph of bees treated with this compound increased, suggesting a more complex mechanism.

In the case of larval hemolymph (Fig. 9B), the findings are similar for the control group and sodium molybdate, showing no increase in AOA. Both $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ and $\text{Li-Mo}_2\text{O}_4\text{-EDTA}$ complexes showed an important increase in AOA in larvae with the ABTS method.

Regarding hive products (see Part VIII, ESI[†]), the AOA of honey samples was similar for control, sodium molybdate and $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ groups but strongly enhanced with $\text{Li-Mo}_2\text{O}_4\text{-EDTA}$. This result is surprising and could be either due to traces of the feeding syrup in the honey for this group or to an action of this complex on the enzymes produced in the bees' crop to produce honey. The AOA of royal jelly and bee wax taken from the 4 groups were similar in all cases. Finally, both complexes significantly amplified the antioxidant activity of bee bread (Fig. 9C), showcasing remarkable efficacy in enhancing its overall antioxidant potential⁵⁴ for the benefit of the entire colony.



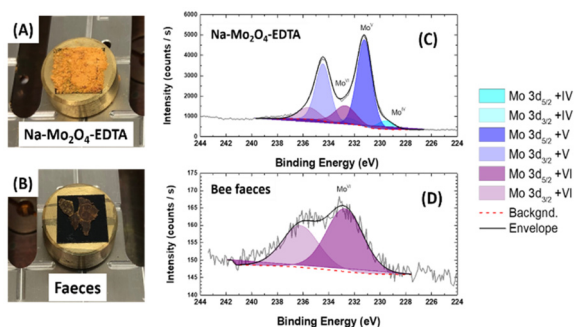


Fig. 8 Samples of $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ and bee faeces deposited on carbon tape (A and B, respectively). The diameter of the sample holder is 1 cm; XPS high-resolution spectra (Mo3d region) of $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ (C) and bee faeces (D). The different contributions and the background used for the reconstruction are plotted as well as the final envelope.

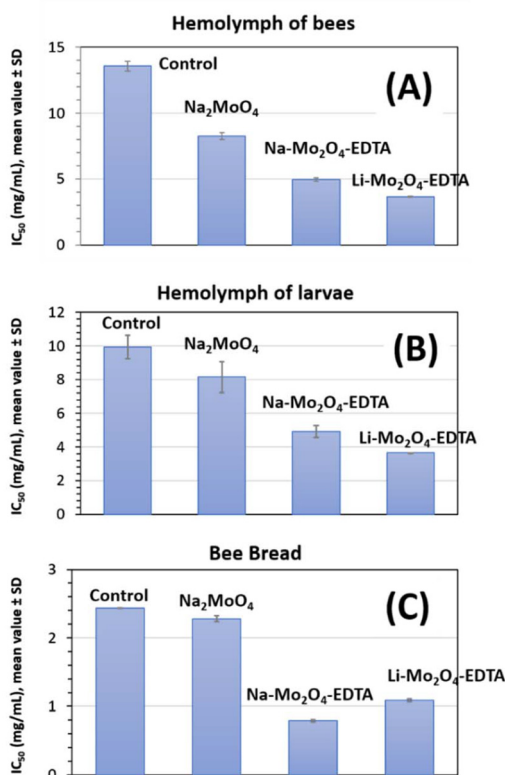


Fig. 9 Antioxidant properties of hemolymph of worker bees (A) and larvae (B) and beebread (C) obtained from beehives of the control group or the groups fed with $\text{Na}_2\text{MoO}_4\cdot 2\text{H}_2\text{O}$, $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ and $\text{Li-Mo}_2\text{O}_4\text{-EDTA}$. Each IC_{50} value corresponds to an average value between three replicates.

3. Discussion

This study provides the first large-scale demonstration of the beneficial impact of molybdenum complexes dietary supplementation on the health of honey bee colonies (*Apis mellifera*). Conducted over eight years in Moldova, in France, in

Greece, and in the USA, field trials, encompassing over 700 hives across diverse ecological settings in Europe and the United States, revealed that administering only a few milligrams of $\text{Na-Mo}_2\text{O}_4\text{-EDTA}$ or $\text{Li-Mo}_2\text{O}_4\text{-EDTA}$ complexes in spring or autumn produces consistent improvements in colony development, hygienic behaviour, honey and wax production for both complexes, whereas a strong reduction in *Varroa destructor* infestation and winter losses have been observed with the lithium salt.

All the field-tests were achieved with low doses of Mo-complexes. Such a low dosage (2–8 mg of complex, which corresponds to 0.6–2.4 mg of the Mo element) is in fact of the same order as the needs of Mo we can globally estimate for a colony across one beekeeping season. Indeed, considering an average level of about $0.4 \mu\text{g g}^{-1}$ (see Part I, ESI†), an average weight of bees of about 80 mg, a lifespan of 40–45 days for the workers bees and a population ranging from 10 000 to 50 000 individuals, we can estimate a need around 30 ng of Mo/bee and around 4 mg of Mo per colony per season. These needs must be met by pollen, but due to a growing lack of resources, the depletion of these resources, and a temporal mismatch between pollen availability and honey bees' activity,^{5,6,12} it is likely that the needs are not being totally met and that appropriate supplementation with Mo could cover the shortfall. Besides, as we demonstrated that these complexes are consumed over a long period of time, they have a prolonged action in the beehives. Therefore, they can feed several generations of bees and fill their deficiency in the Mo trace element, if so. This is a first hypothesis to explain the benefits of the colonies with our complexes, but this environmental deficiency would need to be proven in future studies and the role of Mo must be elucidated.

To date, the main part of the in-field experiments in the literature are focused on protein-based diets and very little is known about the mineral supplements and especially how transition metals cations can modulate the defence mechanisms of the bees against biotic and abiotic sources of stress and then reinforce the health of the colonies. In particular, to date, there are no studies in the literature that examine the effects of molybdenum on bees. After absorption, mineral elements enter the haemolymph and cells and then activate a large number of enzymes involved in the detoxification of xenobiotics.⁵⁵ Among transition metals, zinc has received particular attention. Zinc used as simple salt, has been shown to stimulate hypopharyngeal glands development and upregulate major royal jelly proteins.^{27c} De Almeida Longuini *et al.* evidenced an increase in the level of proteins involved in defence systems by feeding with Zinc sulfate at 50–75 ppm concentration, thus affecting nutrition and maintenance of colonies.^{27d} However, excessive zinc supplementation can result in colony-level disturbances and even abandonment.^{27b} More recently, Ghasemi *et al.* reported the effect of a mixture of transition metal complexes containing Ca, Mg, Fe, Mn, Cu, Zn, Cr and Co at various dosages.³¹ When low concentration of the mixture is used the authors observed the promotion of HPG growth, together with an increase in the level of nutrient in the



bees and a better resistance to the pesticide dimethoate at a sublethal level in laboratory conditions. In contrast, the mixture proved to be toxic when the concentration increases. The use of molybdenum complexes offers distinct advantages compared to these studies since no toxicity was evidenced even at high concentration for **Na-Mo₂O₄-EDTA** both in laboratory and in field conditions. Besides, in-depth studies in the field, in the laboratory and at the synchrotron facility have confirmed the bioavailability and assimilation of Mo in both worker bees and larvae by feeding with our complexes. In particular, we demonstrated that Mo accumulation was especially prominent in the neurolemma and hypopharyngeal glands—two critical tissues involved in nutrition, immunity, and neural function^{49–51} which already naturally contain traces of molybdenum (see Fig. 6A and 7A). These findings suggest that Mo may play a previously unrecognized physiological role in honey bee health, particularly through its incorporation into key organs. In particular, the HPGs are involved in the production of royal jelly which is the primary food of the queen bee, whereas the drone, worker bees and larvae feed upon royal jelly, honey, and pollen. An effect on the quality of the royal jelly should impact the health and the behaviour of the queen bee, what we observe in hives with an increase fecundity of 11–13%.

Honey bee gut microbiome research is an emerging field.⁵ A recent review of Smriti *et al.* evidences that many authors demonstrated that the use of probiotic-supplemented diet can promote honeybee gut health, enhance immunity, and overall well-being.⁵⁵ One of the major reasons for the loss of bee colonies is infections caused by different pathogens like *Paenibacillus larvae*, *Varroa* mites and *Nosema ceranae*. Probiotics can increase the gut health of honeybees and a better gut health favours the development of gut microbiota, which plays a crucial role to induce immune response⁵⁶ and serve various immunomodulatory functions so that they can fight bacterial infections. In particular, beneficial bacteria in the gut of bees help to reduce the impact of nosemosis and varroosis infections.⁵⁷ Thus, probiotics help to increase the survival rate of bees by preventing pathogenic infections and increase the colony strength. It also leads to more honey and royal jelly production by increasing the expression of several genes. In another study, Tejerina *et al.* demonstrated the effectiveness of feed supplementation with probiotics derived from lactic acid bacteria against mites such as *Varroa*.⁵⁸ All these elements lead to lower colony loss due to infections and increase the survival rate of bees. For instance, Kaznowski *et al.* revealed that probiotics promoted bee survival, as bee losses were 30–50% lower in colonies fed probiotics compared to control hives.⁵⁹ However, Tlak Gajger *et al.* showed that if a 5% probiotic supplementation to sugar syrup increased colony strength, a 10% concentration leads to honeybee mortality.⁶⁰

The effects observed by sugar syrup supplemented with probiotics are similar to those we observed in our field trials with Mo-complexes. It thus appears appealing to hypothesize that our Mo-complexes could have a positive impact on the gut microbiota of honey bees to explain the variety of effect

observed in hives. In a previous study, we evidenced that these complexes have no antibacterial effects and can promote the development of biomass,³⁶ which is compatible with this hypothesis.

Finally, X-ray photoelectron spectroscopy (XPS) analyses on bee's faeces revealed the oxidation of Mo(v) to Mo(vi) within bee organisms, indicating a potential antioxidant mechanism, which was confirmed by field test experiments on hemolymph of larvae and worker bees. Given the metabolic demands and oxidative stress encountered by bees, especially during periods of environmental or pathogenic stress, the antioxidant properties of these complexes could underlie many of their observed benefits in hives. Such antioxidant activity (AOA) in both bees and larvae can hold particular significance for insects with high metabolic rates, inherently generating substantial volumes of free radicals.⁶¹ Therefore, implementing proactive measures to mitigate oxidative stress effects can substantially boost the resilience of honey bees⁶² and improve their health and their productivity.

Taken together, molybdenum-based supplementation appears to offer a **multifunctional mode of action**: improving colony productivity, modulating physiological and neurological tissues, and providing redox-related protection—all without compromising honey safety. The impact on the hypopharyngeal glands and on the intestinal microbiota, in addition to compensating for molybdenum deficiency and providing an antioxidant effect, are plausible hypotheses to explain the varied effects we have observed in hives.

Of course, this does not rule out other hypotheses and paves the way towards many further investigations to elucidate the mode of action of our complexes. In particular, further studies are needed (i) to evidence an environmental deficiencies in Mo, (ii) to explore the expression and activity of Mo-dependent enzymes in bees, and to assess how these complexes might protect against combined stressors such as pesticides, pathogens, and poor nutrition, (iii) to investigate more deeply the impact on HPG glands (size, vitellogenin production), the production and quality of royal jelly and its impact on queen bees, (iv) to study the impacts of Mo-complexes on the longevity of bees in hives, on the gut microbiota, or on social behaviour, and (v) to extend this work on other pollinators.

4. Conclusion

Based on a large set of field-test campaigns unprecedented in the literature and lab experiments, this study establishes the importance of the molybdenum trace element for honey bee health. These results are promising for the beekeeping industry, which is currently suffering from numerous challenges weakening honey bee colonies, to improve winter survival and colony productivity.

In this study, we demonstrated that feeding with only a few milligrams of the coordination complexes **Li-Mo₂O₄-EDTA** or **Na-Mo₂O₄-EDTA** during spring or autumn improves key parameters of colony performance: it enhances in particular colony



development, and honey and wax production. Furthermore, it also reduces infestation by the mite *Varroa destructor* and decreases winter colony losses. Importantly, feeding with Mo-complexes had no effect on the quality of the produced honey.

In-depth studies in the field, in the laboratory and at the synchrotron facility have demonstrated that these complexes have a prolonged action in the beehives and that the Mo trace element is naturally present in bees' cuticle, muscles, hypopharyngeal glands and in the neurolemma. Interestingly, Mo levels increased in the neurolemma and in the hypopharyngeal glands after supplementation with Mo-complexes. Lastly, our experiments suggest that the complexes **Li-Mo₂O₄-EDTA** and **Na-Mo₂O₄-EDTA** act as antioxidant agents in bees.

This work repositions molybdenum—until now largely neglected in apicultural science—as a key trace element for bee health and opens many avenues for future works not only for our molybdenum-based complexes but also for coordination complexes in general, which could play a significant role in helping to protect pollinator insects.

Author contributions

I. T., J.-C. S. and S. F. initiated and coordinated the project. A. F. synthesized and characterized the complexes. X. L. performed DFT studies. M. A. S., A. O. S., and T. N. P. performed the toxicity studies on mice. O. Ga. and A. G. coordinated and performed toxicity studies on *Daphnia Magna* and Anti oxidative Properties studies. L. C-D., A. N., V. L. performed the toxicity studies on bees in laboratory conditions. V. C., N. R. and I. T. performed and followed the field tests in Moldova. A. F., S. F., P. C., and B. P. performed and followed the field tests in France and in USA. L. Ch. and F. H. performed the field tests in Greece. O. Gl. collected and provided bee samples from Moldova. I. R and L. C-D. performed the slices of bees. L. C., I. G., J.-C. S., S. F., A. S. and K. M. managed Synchrotron studies with L. C-D., I. G., S. F., and J.-C. S., M. F. performed the XPS studies. P. C. and S. G. performed statistics.

Conflicts of interest

The results of this study are partially patented to protect the complexes for their potential use in beekeeping industry. Some authors, namely Arcadie Fuior, Valentina Cebotari, Aurelian Gulea, Ion Toderas and Sébastien Floquet are co-inventors of this patent delivered in France (FR3112667B1 on 23 feb. 2024), In Europe (EP4185594B1 on 4 dec2024) and in progress in USA, in Canada, in Argentina and in China.

Data availability

All the details of the synthesis, characterizations, properties measurements, test in hives, *etc.* are given in the ESI† with the raw data.

To facilitate the reading, the ESI† is divided into 8 independent parts built as reports to cover each topic. Part I contains data about Mo level measured in honeybees by ICP-MS or ICP-OES; Part II contains all details on the synthesis of complexes, their characterization and the stability studies by DFT calculation, ¹H NMR and UV-visible spectroscopies; Part III contains the toxicity studies on mice, *Daphnia Magna*, and bees both in laboratory and in hives; Part IV gathers all the field tests in beehives and analyses on honey with the raw data obtained for each test campaign; Part V is focused on the cycle of Mo in beehives after feeding by monitoring Mo content by ICP-MS in larvae, worker bees, honey/syrup, wax, and bee bread; Part VI contains the data about the assimilation of Mo by honey bees in laboratory conditions in head, thorax and abdomen. The Mo, Na/Li contents are measured by ICP-MS; Part VII gathered all data of X-ray fluorescence acquired at synchrotron SOLEIL; Part VIII focuses on the metabolism and the antioxidant properties of Mo-complexes consumed by honey bees thank to an XPS study of bee faeces and spectroscopic determination of antioxidant activity of hemolymph of bees and larvae as well as hive products by ABTS and DPPH methods.

Acknowledgements

University of Versailles, the “Institut Universitaire de France, IUF” and the CNRS are gratefully acknowledged for financial support. AF gratefully acknowledge Campus France for Excellence Eiffel grant as well as State University of Moldova for funding his PhD thesis. This work was funded by the Labex Chammmat (project “COMPA”), the IDEX Paris-Saclay (project “COMPA2”), the “lune de miel” foundation, the UVSQ foundation (project “COMPA3”), the CNRS MITI (call Metallomix, project “MOLYBEE”), and the SATT Paris-Saclay (projects “COMPA4” and “APIMONA”). The financial support from National Agency for Research and Development (ANCD) of the Republic of Moldova (Project No. 20.80009.5007.10) is also acknowledged. Dr Clémence Riva is gratefully acknowledged for her help for the statistical analyses. The NIIC team thanks the Ministry of Science and Higher Education of the Russian Federation (No. 121031700321-3). AOS acknowledges the support of assays by state funding of RICEL—Branch of IC&G SB RAS. Mrs Aneta Ozieranska and Dr Karen Wright from Institut Lavoisier of Versailles are gratefully acknowledged for English correction.

References

- 1 A. M. Klein, B. E. Vaissiere, J. H. Cane, I. Steffan-Dewenter, S. A. Cunningham, C. Kremen and T. Tscharntke, Importance of pollinators in changing landscapes for world crops, *Proc. Biol. Sci.*, 2007, **274**, 303–313.
- 2 N. Gallai, J. M. Salles, J. Settele and B. E. Vaissiere, Economic valuation of the vulnerability of world agriculture confronted with pollinator decline, *Ecol. Econ.*, 2009, **68**, 810–821.



- 3 S. G. Potts, S. P. M. Roberts, R. Dean, G. Marris, M. A. Brown, R. Jones, P. Neumann and J. Settele, Declines of managed honey bees and beekeepers in Europe, *J. Apic. Res.*, 2010, **49**(1), 15–22.
- 4 D. Goulson, E. Nicholls, C. Botias and E. L. Rotheray, Bee declines driven by combined stress from parasites, pesticides, and lack of flowers, *Science*, 2015, **347**, 1255957.
- 5 J. M. Tsuruda, P. Chakrabarti and R. R. Sagili, Honey Bee Nutrition, *Vet. Clin. Food Anim.*, 2021, **37**, 505–519.
- 6 P. W. Lau, I. L. Esquivel, K. A. Parys, K.-L. J. Hung and P. Chakrabarti, The nutritional landscape in agroecosystems: a review on how resources and management practices can shape pollinator health in agricultural environments, *Ann. Entomol. Soc. Am.*, 2023, **116**(5), 261–275.
- 7 R. Brodschneider and K. Crailsheim, Nutrition and health in honey bees, *Apidologie*, 2010, **41**, 278–294.
- 8 E. Topal, N. Çakıcı, R. Mărgăoan, Ç. Takma, F. Güney, M. Kösoğlu, M. Cornea-Cipcigan and H. Atmaca, Annual Development Performance of Fixed Honeybee Colonies Linked with Chemical and Mineral Profile of Bee Collected Pollen, *Chem. Biodivers.*, 2022, **19**, e202200468.
- 9 C. Alaux, F. Ducloz, D. Crauser and Y. Le Conte, Diet effects on honeybee immunocompetence, *Biol. Lett.*, 2010, **6**, 562–565.
- 10 A. G. Dolezal, J. Carrillo-Tripp, T. M. Judd, W. A. Miller, B. C. Bonning and A. L. Toth, Interacting stressors matter: diet quality and virus infection in honeybee health, *R. Soc. Open Sci.*, 2019, **6**, 181803.
- 11 D. Frizzera, A. M. Ray, E. Seffin, V. Zanni, D. Annoscia, C. M. Grozinger and F. Nazzi, The Beneficial Effect of Pollen on Varroa Infested Bees Depends on Its Influence on Behavioral Maturation Genes, *Front. Insect Sci.*, 2022, **2**, 864238.
- 12 M. Gérard, M. Vanderplanck, T. Wood and D. Michez, Global warming and plant–pollinator mismatches, *Emerging Top. Life Sci.*, 2020, **4**(1), 77–86.
- 13 L. Desmedt, L. Hotier, M. Giurfa, R. Velarde and M. G. de Brito Sanchez, Absence of food alternatives promotes risk-prone feeding of unpalatable substances in honey bees, *Sci. Rep.*, 2016, **6**, 31809.
- 14 A. Brown, V. Rodriguez, J. Pfister, V. Perreten, P. Neumann and G. Retschnig, The dose makes the poison: feeding of antibiotic-treated winter honey bees, *Apis mellifera*, with probiotics and b-vitamins, *Apidologie*, 2022, **53**, 19.
- 15 N. M. Jovanovic, U. Glavinic, B. Delic, B. Vojnovic, N. Aleksic, V. Mladjan and Z. Stanimirovic, Plant-based supplement containing B-complex vitamins can improve bee health and increase colony performance, *Prev. Vet. Med.*, 2021, **190**, 105322.
- 16 R. Pavlović, B. Dojnov, M. Š. Slavić, M. Ristović, M. Vujčić, S. Stojanović and Z. Vujčić, Differential Processing of Sucrose and Invert Syrup in Honey Bees, *Arch. Insect Biochem. Physiol.*, 2025, **118**, e70052.
- 17 V. A. Ricigliano, S. T. Williams and R. Oliver, Effects of different artificial diets on commercial honey bee colony performance, health biomarkers, and gut microbiota, *BMC Vet. Res.*, 2022, **18**, 52.
- 18 A. Saffari, P. G. Kevan and J. L. Atkinson, Consumption of three dry pollen substitutes in commercial apiaries, *J. Apic. Sci.*, 2010, **54**(2), 13–20.
- 19 E. J. García-Vicente, M. Martín, I. Rey-Casero, A. Perez, J. Martín, A. García, J. M. Alonso and D. Risco, Effects of feeding with a protein liquid supplement on productivity, mortality and health of *Apis mellifera* hives in southwestern Spain, *Res. Vet. Sci.*, 2024, **169**, 105173.
- 20 G. DeGrandi-Hoffman, Y. Chen, R. Rivera, M. Carroll, M. Chambers, G. Hidalgo and E. Watkins de Jong, Honey bee colonies provided with natural forage have lower pathogen loads and higher overwinter survival than those fed protein supplements, *Apidologie*, 2016, **47**, 186–196.
- 21 T. Bogaert, T. Reams, I. Mailliet, K. Kulhanek, M. Duyck, F. Eertmans, A. M. Fauvel, B. Hopkins and J. Bogaert, A nutritionally complete pollen-replacing diet protects honeybee colonies during stressful commercial pollination — requirement for isofucosterol, *Proc. R. Soc. B*, 2025, **292**, 20243078.
- 22 E. J. García-Vicente, M. Martín, I. Rey-Casero, A. Perez, R. Martínez, M. Bravo, J. M. Alonso and D. Risco, Effect of feed supplementation with probiotics and postbiotics on strength and health status of honey bee (*Apis mellifera*) hives during late spring, *Res. Vet. Sci.*, 2023, **159**, 237–243.
- 23 (a) R. E. Bonoan, L. D. O'Connor and P. T. Starks, Seasonality of honey bee (*Apis mellifera*) micronutrient supplementation and environmental limitation, *J. Insect Physiol.*, 2018, **107**, 23–28; (b) R. E. Bonoan, T. M. Tai, M. Tagle-Rogriguez, L. Feller, S. R. Daddario, R. Czaja, L. D. O'Connor, G. Buruss and P. T. Starks, Seasonality of salt foraging in honey bees (*Apis mellifera*), *Ecol. Entomol.*, 2017, **42**, 195–201.
- 24 C. H. Liang, C. L. Chuang, J. A. Jiang and E. C. Yang, Magnetic Sensing through the Abdomen of the Honey bee, *Sci. Rep.*, 2016, **6**, 23657.
- 25 H. N. Koo, S. G. Lee, S. H. Yun, H. K. Kim, Y. S. Choi and G. H. Kim, Comparative Analyses of Cu-Zn Superoxide Dismutase (SOD1) and Thioredoxin Reductase (TrxR) at the mRNA Level between *Apis mellifera* L. and *Apis cerana* F. (Hymenoptera: Apidae) Under Stress Conditions, *J. Insect Sci.*, 2016, **16**(1), 4.
- 26 (a) Y. Ben-Shahar, The Impact of Environmental Mn Exposure on Insect Biology, *Front. Genet.*, 2018, **9**, 70; (b) E. Sovik, C. J. Perry, A. LaMora, A. B. Barron and Y. Ben-Shahar, Negative impact of manganese on honey bee foraging, *Biol. Lett.*, 2015, **11**(3), 20140989.
- 27 (a) M. P. Camilli, S. M. Kadri, M. V. N. Alvarez, P. E. M. Ribolla and R. O. Orsi, Zinc supplementation modifies brain tissue transcriptome of *Apis mellifera* honey bees, *BMC Genomics*, 2022, **23**(1), 282; (b) M. P. Carillo, A. Astolfi, E. J. D. Rodrigues, I. C. D. Lippi, L. G. R. Da Silva, M. G. Guimaraes, L. A. Justulin and R. D. Orsi, High doses of inorganic zinc modulate the hypopharyngeal glands of *Apis mellifera* but promote abandonment of colo-



- nies when offered over long periods, *Bull. Insectology*, 2022, 75(1), 27–31; (c) G. D. Ribeiro, S. M. Kadri, L. A. Justulin, P. E. M. Ribolla and R. D. Orsi, Zinc methionine or zinc sulphate supplementation modulate the development of the hypopharyngeal gland and expression of major royal jelly protein genes in *Apis mellifera* L. bees, *Physiol. Entomol.*, 2023, 48(2–3), 90–96; (d) L. A. de Almeida Longuini, G. Moreno Martineli, M. Polizel Camilli, D. Cavalcante Brambila de Barros, J. Cavalcante Souza Vieira, P. de Magalhães Padilha and R. de Oliveira Orsi, Supplementation with an Inorganic Zinc Source in the Metalloproteomic Profile of Royal Jelly in *Apis mellifera*, *Biol. Trace Elem. Res.*, 2021, 199, 4308–4318.
- 28 (a) T. V. Nikolic, D. Kojic, S. Orcic, D. Batinic, E. Vukasinovic, D. P. Blagojevic and J. Purac, The impact of sublethal concentrations of Cu, Pb and Cd on honey bee redox status, superoxide dismutase and catalase in laboratory conditions, *Chemosphere*, 2016, 164, 98–105; (b) N. Di, K. R. Hladun, K. Zhang, T. X. Liu and J. T. Trumble, Laboratory bioassays on the impact of cadmium, copper and lead on the development and survival of honey bee (*Apis mellifera* L.) larvae and foragers, *Chemosphere*, 2016, 152, 530–538; (c) C. Monchanin, E. Drujont, G. Le Roux, P. D. Lösel, A. B. Barron, J. M. Devaud, A. Elger and M. Lihoreau, Environmental exposure to metallic pollution impairs honey bee brain development and cognition, *J. Hazard. Mater.*, 2024, 465, 133218.
- 29 E. W. Herbert, A New Ash Mixture for Honeybees Maintained on a Synthetic Diet, *J. Apic. Res.*, 1979, 18, 144–147.
- 30 G. Zhang, W. Zhang, X. Cui and B. Xu, Zinc nutrition increases the antioxidant defenses of honey bees, *Entomol. Exp. Appl.*, 2015, 156, 201–210.
- 31 V. Ghasemi, G. N. Paghaleh, E. Torabi, Z. S. Arzanforoosh, R. Bideshki, S. Kalanaky, S. Fakharzadeh, M. Hafizi and M. H. Nazara, Effects of advanced chelate-based mineral supplement on adult worker honey bees' (*Apis mellifera* L.) survival, nutritional indices, hypopharyngeal glands' growth, energy reserve contents, and tolerance to dimethoate, *J. Apic. Res.*, 2025, DOI: [10.1080/00218839.2024.2446106](https://doi.org/10.1080/00218839.2024.2446106).
- 32 (a) R. Hille, Molybdenum and Tungsten in Biology, *Trends Biochem. Sci.*, 2002, 27(7), 360–367; (b) R. R. Mendel and T. Kruse, Cell biology of molybdenum in plants and humans, *Biochim. Biophys. Acta*, 2012, 1823, 1568–1579.
- 33 (a) T. Peng, Y. Xu and Y. Zhang, Comparative genomics of molybdenum utilization in prokaryotes and eukaryotes, *BMC Genomics*, 2018, 19, 691; (b) Y. Zhang, S. Rump and V. N. Gladyshev, Comparative genomics and evolution of molybdenum utilization, *Coord. Chem. Rev.*, 2011, 255(9–10), 1206–1217.
- 34 V. R. Ott, D. S. Swieter, and F. A. Schultz, Di- μ -Oxo, μ -Oxo- μ -Sulfido, and Di- μ -Sulfido Complexes of Molybdenum(v) with EDTA, Cysteine, and Cysteine Ester Ligands. Preparation and Electrochemical and Spectral Properties, *Inorg. Chem.*, 1977, 16(10), 2538–2545.
- 35 J. M. Gretarsdóttir, S. Bobersky, N. Metzler-Nolte and S. G. Suman, Cytotoxicity Studies of Water-Soluble Coordination Compounds with a $[\text{Mo}_2\text{O}_2\text{S}_2]^{2+}$ Core, *J. Inorg. Biochem.*, 2016, 160, 166–171.
- 36 A. Fuior, A. Hijazi, O. Garbuz, V. Bulimaga, L. Zosim, D. Cebotari, M. Haouas, I. Toderas, A. Gulea and S. Floquet, Screening of biological properties of $\text{MoV}_2\text{O}_2\text{S}_2$ - and MoV_2O_4 -based coordination complexes: Investigation of antibacterial, antifungal, antioxidative and antitumoral activities versus growing of *Spirulina platensis* biomass, *J. Inorg. Biochem.*, 2022, 226, 111627.
- 37 J. M. Gretarsdóttir, S. Jonsdóttir, W. Lewis, T. W. Hambley, and S. G. Suman, Water-Soluble Alpha-Amino Acid Complexes of Molybdenum as Potential Antidotes for Cyanide Poisoning: Synthesis and Catalytic Studies of Threonine, Methionine, Serine, and Leucine Complexes, *Inorg. Chem.*, 2020, 59(24), 18190–18204.
- 38 J. M. Gretarsdóttir, I. H. Lambert, S. Sturup and S. G. Suman, *In Vitro* Characterization of a Threonine-Ligated Molybdenyl-Sulfide Cluster as a Putative Cyanide Poisoning Antidote; Intracellular Distribution, Effects on Organic Osmolyte Homeostasis, and Induction of Cell Death, *ACS Pharmacol. Transl. Sci.*, 2022, 5(10), 907–918.
- 39 I. Toderas, A. Gulea, V. Cebotari, S. Floquet, E. Cadot and I. Buzu, A method for preparing a bio active food supplement for bees feed and process for feeding bee colonies *Apis mellifera*, Patent No. 4438 MD-BOPI 4, 2016 p. 16.
- 40 (a) B. Ziegelmann, E. Abele, S. Hannus, M. Beitzinger, S. Berg and P. Posenkranz, Lithium chloride effectively kills the honey bee parasite *Varroa destructor* by a systemic mode of action, *Sci. Rep.*, 2018, 8, 683; (b) S. Sevin, V. Bommuraj, Y. R. Chen, O. Afik, S. Zarchin, S. Barel, O. C. Arslan, B. Erdem, H. Tutun and J. A. Shimshoni, Lithium salts: assessment of their chronic and acute toxicities to honey bees and their anti-*Varroa* field efficacy, *Pest Manage. Sci.*, 2022, 78(11), 4507–4516.
- 41 F. Mondet, S. Blanchard, N. Barthes, D. Beslay, C. Bordier, G. Costagliola, M. R. Hervé, B. Lapeyre, S. H. Kim, B. Basso, A. R. Mercer and Y. Le Conte, Chemical detection triggers honey bee defence against a destructive parasitic threat, *Nat. Chem. Biol.*, 2021, 17, 524–530.
- 42 M.-P. Chauzat, M. Laurent, M.-P. Rivière, C. Saugeon, P. Hendrix and M. Ribiere-chabert, Report “A pan-European epidemiological study on honeybee colony losses 2012–2013”, Epilobee, 27th april 2024.
- 43 A. Gray, N. Adjlane, A. Arab, A. Ballis, V. Brusbardis, A. Bugeja Douglas, L. Cadahía, J.-D. Charrière, R. Chlebo, M. F. Coffey, B. Cornelissen, C. A. D. Costa, E. Danneels, J. Danihlík, C. Dobrescu, G. Evans, M. Fedoriak, I. Forsythe, A. Gregorc, I. I. Arakelyan, J. Johannesen, L. Kauko, P. Kristiansen, M. Martikkala, R. Martín-Hernández, E. Mazur, C. A. Medina-Flores, F. Mutinelli, E. M. Omar, S. Patalano, A. Raudmets, G. San Martin, V. Soroker, P. Stahlmann-Brown, J. Stevanovic, A. Uzunov, F. Vejsnaes, A. Williams and R. Brodschneider, Honey bee colony loss rates in 37 countries using the COLOSS survey



- for winter 2019–2020: the combined effects of operation size, migration and queen replacement, *J. Apic. Res.*, 2023, **62**, 204–210.
- 44 F. Requier, K. Antúnez, C. L. Morales, P. Aldea Sánchez, D. Castilhos, P. M. Garrido, A. Giacobino, F. J. Reynaldi, J. M. Rosso Londoño, E. Santos and L. A. Garibaldi, Trends in beekeeping and honey bee colony losses in Latin America, *J. Apic. Res.*, 2018, **57**, 657–662.
 - 45 S. Bruckner, M. Wilson, D. Aurell, K. Rennich, D. van Engelsdorp, N. Steinhauer and G. R. Williams, A national survey of managed honey bee colony losses in the USA: results from the Bee Informed Partnership for 2017–18, 2018–19, and 2019–20, *J. Apic. Res.*, 2023, **62**, 429–443.
 - 46 D. van Engelsdorp and M. D. Meixner, A historical review of managed honey bee populations in Europe and the United States and the factors that may affect them, *J. Invertebr. Pathol.*, 2023, **103**, S80–S95.
 - 47 D. van Engelsdorp, J. Hayes, R. M. Underwood and J. S. Pettis, A survey of honey bee colony losses in the United States, fall 2008 to spring 2009, *J. Apic. Res.*, 2010, **49**(1), 7–14.
 - 48 <https://www.projectapism.org/colony-loss-information>.
 - 49 S. Ahmad, S. A. Khan, K. A. Khan and J. K. Li, Novel Insight Into the Development and Function of Hypopharyngeal Glands in Honey Bees, *Front. Physiol.*, 2021, **11**, 615830.
 - 50 L. N. Standifer, A comparison of the protein quality of pollen for growth-stimulation of the hypopharyngeal glands and longevity of honey bees *Apis mellifera* L. (Hymenoptera : Apidae), *Insectes sociaux, Paris*, 1967, **14**(4), 415–426.
 - 51 G. M. Quinlan and C. M. Grozinger, Evaluating the role of social context and environmental factors in mediating overwintering physiology in honey bees (*Apis mellifera*), *J. Exp. Biol.*, 2024, **227**, jeb247314.
 - 52 (a) C. R. Clayton and Y. C. Lu, Electrochemical and XPS Evidence of the Aqueous Formation of Mo_2O_5 , *Surf. Interface Anal.*, 1989, **14**(1–2), 66–70; (b) P. A. Spevack and N. S. McIntyre, Thermal Reduction of Molybdenum Trioxide, *J. Phys. Chem.*, 1992, **96**(22), 9029–9035; (c) X. Liao, A. R. Jeong, R. G. Wilks, S. Wiesner, M. Rusu and M. Bär, X-Ray, Irradiation Induced Effects on the Chemical and Electronic Properties of MoO_3 Thin Films, *J. Electron Spectrosc. Relat. Phenom.*, 2016, **212**, 50–55; (d) J. Baltrusaitis, B. Mendoza-Sanchez, V. Fernandez, R. Veenstra, N. Dukstiene, A. Roberts and N. Fairley, Generalized Molybdenum Oxide Surface Chemical State XPS Determination via Informed Amorphous Sample Model, *Appl. Surf. Sci.*, 2015, **326**, 151–161.
 - 53 R. Re, N. Pellegrini, A. Proteggente, A. Pannala, M. Yang and C. Rice-Evans, Antioxidant activity applying an improved ABTS radical cation decolorization assay, *Free Radicals Biol. Med.*, 1999, **26**, 1231–1237.
 - 54 J. Kocot, M. Kielczykowska, D. Luchowska-Kocot, J. Kurzepa and I. Musik, Antioxidant potential of propolis, bee pollen, and royal jelly: Possible medical application, *Oxid. Med. Cell. Longevity*, 2018, 7074209.
 - 55 Smriti, A. Rana, G. Singh and G. Gupta, Prospects of probiotics in beekeeping: a review for sustainable approach to boost honeybee health, *Arch. Microbiol.*, 2024, **206**, 205.
 - 56 M. J. Molloy, N. Bouladoux and Y. Belkaid, Intestinal microbiota: shaping local and systemic immune responses, *Semin. Immunol.*, 2012, **24**(1), 58–66.
 - 57 M. C. Audisio, Gram-positive bacteria with probiotic potential for the *Apis mellifera* L. honeybee: the experience in the northwest of Argentina, *Probiotics Antimicrob. Proteins*, 2017, **9**, 22–31.
 - 58 M. R. Tejerina, M. R. Benítez-Ahrendts and M. C. Audisio, *Lactobacillus salivarius* A3iob reduces the incidence of *Varroa destructor* and *Nosema* Spp. in commercial apiaries located in the northwest of Argentina, *Probiotics Antimicrob. Proteins*, 2020, **12**, 1360–1369.
 - 59 A. Kaznowski, B. Szymas, E. Jazdzinska, M. Kazimierzczak, H. Paetz and J. Mokracka, The effects of probiotic supplementation on the content of intestinal microflora and chemical composition of worker honeybees (*Apis mellifera*), *J. Apic. Res.*, 2005, **44**(1), 10–14.
 - 60 I. Tlak Gajger, J. Vlainić, P. Šoštarić, J. Prešern, J. Bubnić and M. I. Smodiš Škerl, Effects on some therapeutical, biochemical, and immunological parameters of honeybee (*Apis mellifera*) exposed to probiotic treatments, in field and laboratory conditions, *Insects*, 2020, **11**(9), 638.
 - 61 D. J. Candy, A. Becker and B. Wegener, Coordination and integration of metabolism in insect flight, *Comp. Biochem. Physiol.*, 1997, **117B**, 497–512.
 - 62 M. Farjan, M. Dmitryjuk, Z. Lipiński, E. Biernat-Łopieńska and K. Żółtowska, Supplementation of the honey bee diet with vitamin C: The effect on the antioxidative system of *Apis mellifera carnica* brood at different stages, *J. Apic. Res.*, 2012, **51**(3), 263–270.

