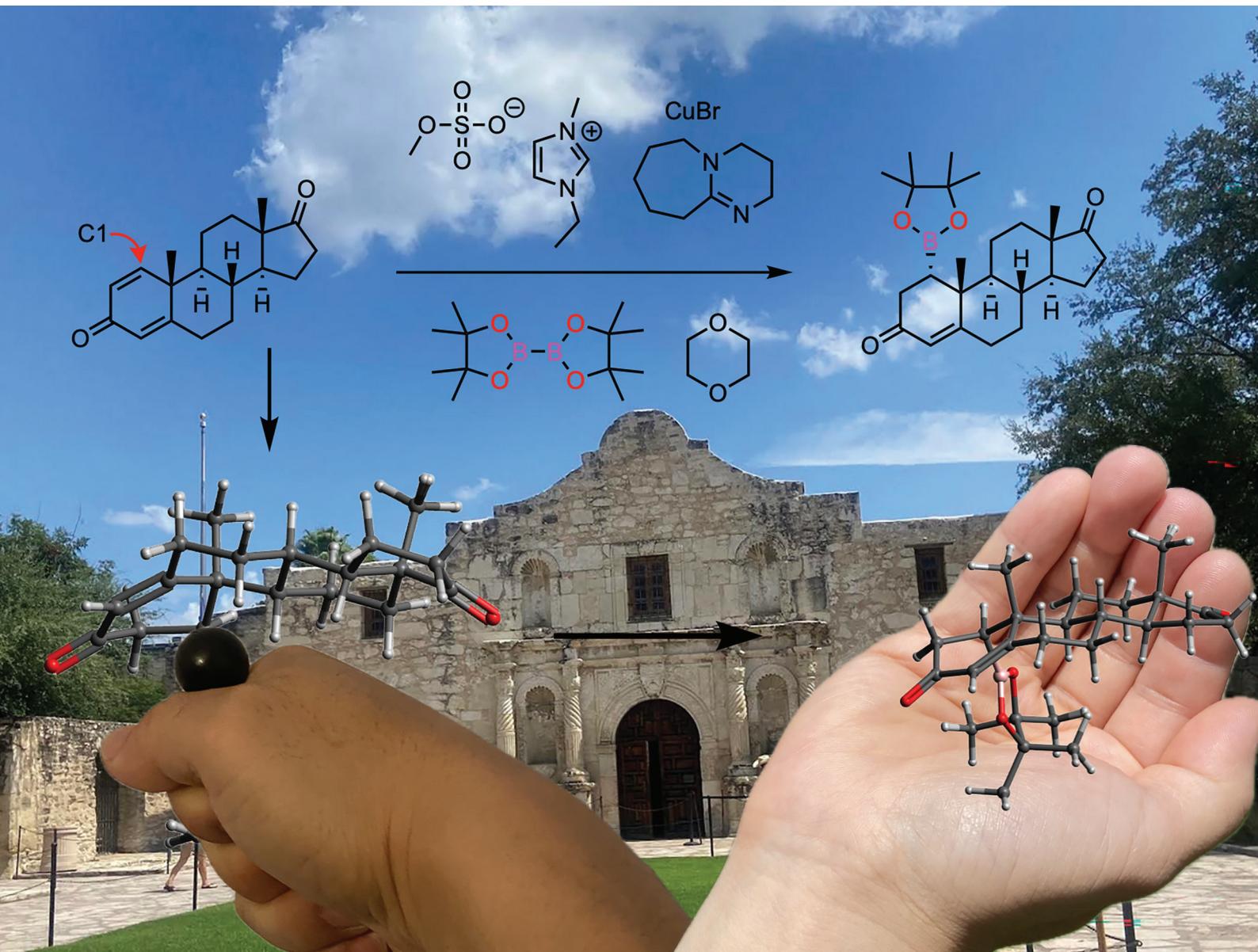


Organic & Biomolecular Chemistry

rsc.li/obc



ISSN 1477-0520

PAPER

Francis K. Yoshimoto *et al.*
A synthesis of 1 β -hydroxytestosterone, a metabolite of
xenobiotic human cytochrome P450 enzymes, beginning
with a borylation of boldione



Cite this: *Org. Biomol. Chem.*, 2025, **23**, 9618

A synthesis of 1 β -hydroxytestosterone, a metabolite of xenobiotic human cytochrome P450 enzymes, beginning with a borylation of boldione

Anna I. Elizondo, ^a Kevin D. McCarty, ^b Hadi D. Arman, ^a F. Peter Guengerich ^b and Francis K. Yoshimoto ^{*a}

Xenobiotic cytochrome P450 enzymes have been shown to hydroxylate testosterone at various positions in the steroid backbone, including C1 to produce 1 β -hydroxytestosterone. Despite the potential application to study the biochemistry of these enzymes, 1 β -hydroxytestosterone is not commercially available. A synthesis of 1 β -hydroxytestosterone from commercially available boldione (androst-1,4-dien-3,17-dione) was accomplished in eight steps. The key step to functionalize C1 was a borylation reaction catalyzed by an *in situ* generated copper carbene complex. The synthetic strategy reported will be used to access other biologically relevant C1-hydroxylated steroids to explore the biochemistry of drug metabolizing P450 enzymes.

Received 28th July 2025,
Accepted 2nd September 2025
DOI: 10.1039/d5ob01218j
rsc.li/obc

Background

Testosterone has been used as a substrate to biochemically characterize human xenobiotic P450 enzymes.^{1,2} For instance, cytochrome P450 3A4, which is a drug metabolizing liver enzyme, incorporates a hydroxy group at the 1 β -, 2 β -, 6 β -, and 15 β -positions of testosterone – the product distribution is in a ratio of 6.7 : 13 : 73 : 6.7 based on the reported k_{cat} values, respectively (Fig. 1).³ In contrast, P450 3A7, which is over-expressed in fetal liver, has been shown to monohydroxylate testosterone with a different regioselectivity.⁴ Due to its potential application to study xenobiotic drug metabolizing P450 enzymes, authentic standards of hydroxylated testosterone derivatives would be useful to study their biochemistry. However, enzymatic conversion could be low yielding⁵ and restricted to specialized laboratory equipment and plasmid strains,⁶ which directed our focus to accessing the compound through chemical synthesis.

The prior reports for the syntheses of 1 β -hydroxytestosterone⁷ and 1 α -hydroxytestosterone⁸ both involved 7 steps from 5 α -dihydrotestosterone benzoate with yields of 8.5% and 1.8%, respectively. The incorporation of the 1-oxygen was obtained from the epoxidation using *t*-butylhydroperoxide in the presence of molybdenum hexacarbonyl or NaOH to eventually yield the 1 β -hydroxy or the 1 α -hydroxy

derivatives, respectively. Our research laboratory previously reported the direct C-H hydroxylation at C1 using the Schönecker oxidation conditions,⁹ but this method would be restricted to the 5 α -reduced and 19-oxo steroid backbone.

Here, we report the synthesis of 1 β -hydroxytestosterone (2) beginning with a key 1,4-borylation at C1 onto commercially available boldione (androst-1,4-dien-3,17-dione) (Fig. 2, 6 to 7 to 2). The synthesis of 1 β -hydroxytestosterone (2) from boldione (6) was achieved in eight total steps.

Results and discussion

Optimization of the 1,4-borylation reaction onto androst-1,4-dien-3,17-dione using DBU

Although others have reported the conjugate addition of pinacolatoborane onto α,β -unsaturated carbonyls,^{10,11} our efforts to replicate the various reaction conditions onto androst-1,4-dien-3,17-dione resulted in either no reaction or low yield¹² with the copper-carbene system. We hypothesized that the low yield of the copper-catalyzed reaction was due to the base (potassium *tert*-butoxide), which made the reaction mixture viscous and cloudy with the presence of precipitate when stirring. Therefore, the base was switched to 1,8-diazabicyclo-(5.4.0)-undec-7-ene (DBU), which resulted in a homogeneous mixture. Table 1 summarizes the optimization conditions (see Fig. S2-2 for the NMR spectroscopic overlay of the 5 entries). Unlike a prior report,¹³ the presence of copper was required in our system for the reaction to occur. The use of DBU as the base optimized the yield of the C1-borylated product to 90% (Scheme 1, 6 to 7).

^aDepartment of Chemistry, the University of Texas at San Antonio (UTSA), San Antonio, Texas 78249, USA. E-mail: francis.yoshimoto@gmail.com

^bDepartment of Biochemistry, Vanderbilt University School of Medicine, Nashville, Tennessee 37232-0146, USA



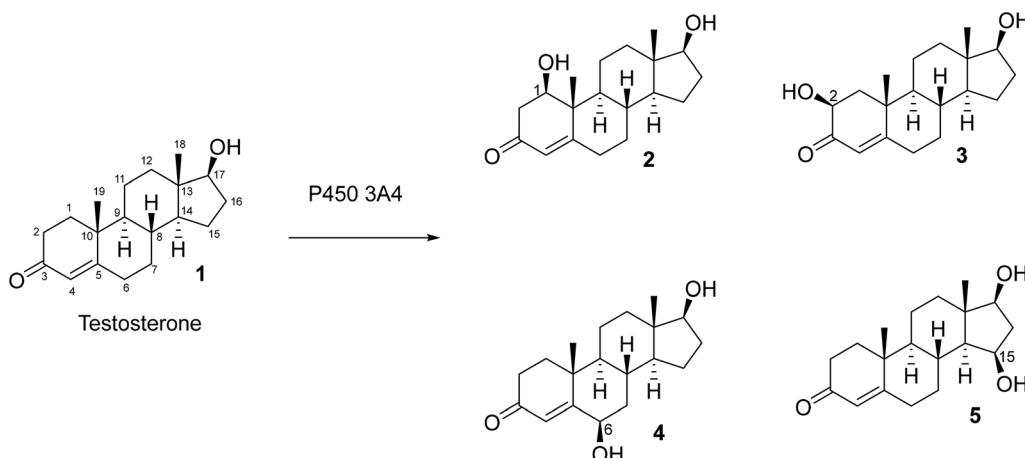


Fig. 1 Testosterone (1) hydroxylation catalyzed by cytochrome P450 3A4 yields monohydroxylated products at C1, C2, C6, and C15 (2, 3, 4, and 5).³

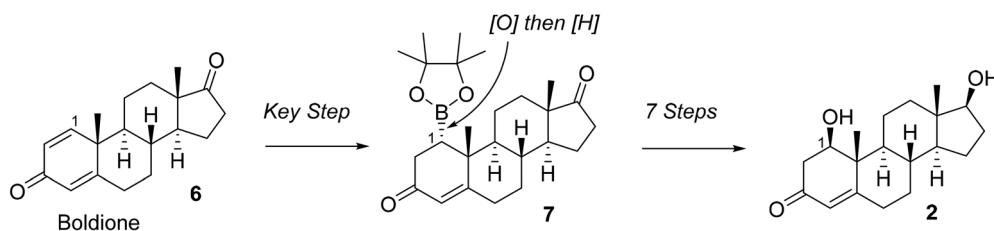


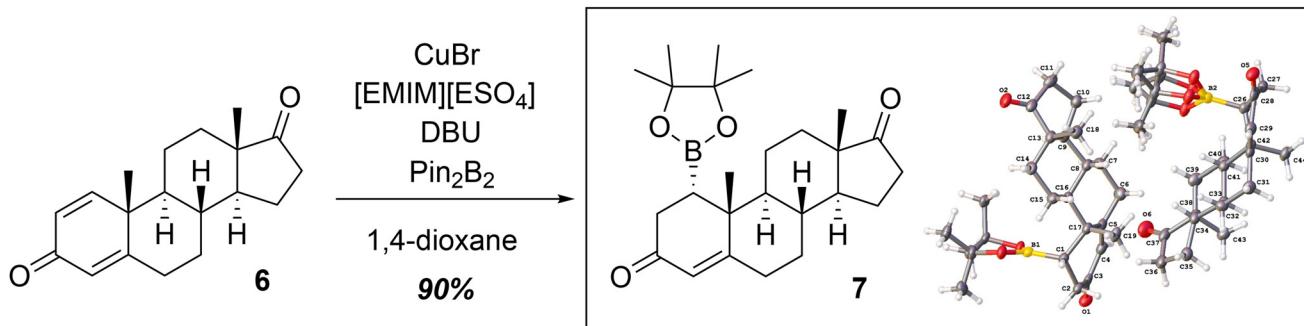
Fig. 2 Overall strategy in this report to synthesize 1 β -hydroxytestosterone (2) from boldione (androst-1,4-dien-3,17-dione, 6) through a borylated intermediate (7).

Table 1 Optimization of Step 1: C1-borylation of boldione (6) to yield 7

Entry	Conditions	Yield ^a
1	(PinB) ₂ (1.0 eq.), PPh ₃ (0.76 eq.), CH ₃ OH, Wilkinson's Catalyst (0.3 eq.)	18%
2	(PinB) ₂ (1.1 eq.), THF, DBU (45 eq.), [EMIM][ESO ₄] (45 eq.)	— ^b
3	(PinB) ₂ (3.5 eq.), THF, KOtBu (7.1 eq.), CuBr (9.3 eq.), [EMIM][ESO ₄] (4.2 eq.)	39%
4	(PinB) ₂ (1.3 eq.), THF, DBU (1.8 eq.), CuBr (0.2 eq.), [EMIM][ESO ₄] (1.8 eq.)	75%
5	(PinB) ₂ (1.5 eq.), 1,4-D, ^c DBU (1.0 eq.), CuBr (0.2 eq.), [EMIM][ESO ₄] (1.0 eq.)	88%

^aYield of 7 was calculated by integration of the C4-protons of 6 and 7 of the ¹H NMR spectra of the crude reaction mixtures (δ 6.3 and 5.8, respectively. Also see Fig. S2-2). ^bNo C4-vinyl proton corresponding to 7 in the crude reaction mixture was detected by ¹H NMR spectroscopy.

^c(1,4-D): 1,4-dioxane.



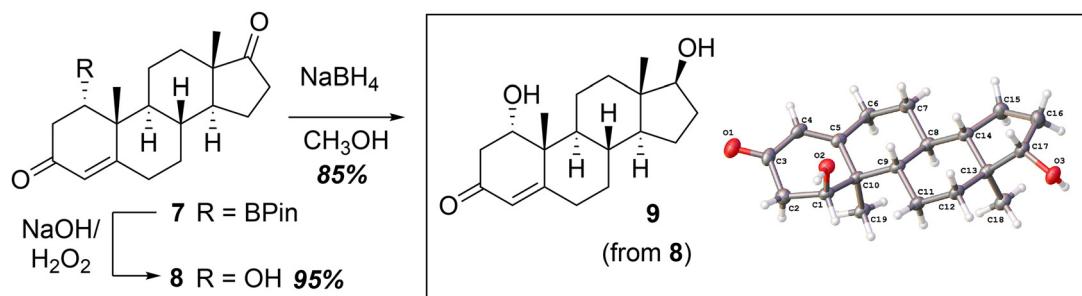
Scheme 1 Successful 1,4-borylation of androst-1,4-dien-3,17-dione (6) to yield borane adduct 7. [EMIM][ESO₄]: 1-ethyl-3-methylimidazolium ethyl sulfate (Step 1 of 8 steps).



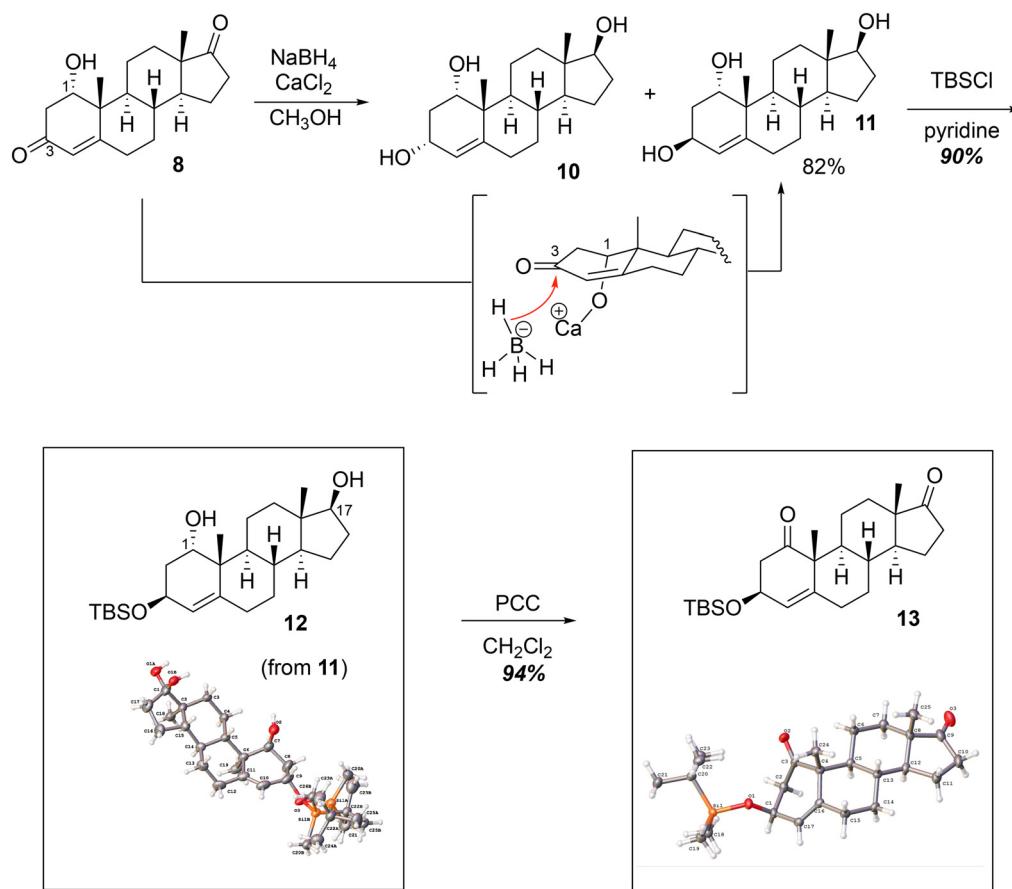
Similar to other reports of the borylation at C1 of the steroid, the boron substituent was added in the α -orientation, presumably to avoid the steric clash with the C19-axial methyl of the starting material. A crystal structure of the boron adduct is shown in Scheme 1, which confirms the stereochemistry at C1. To test the versatility of the borylation reaction, androst-2,4-dien-1-one was also used as the substrate, which underwent conjugate addition at C3 (see SI).

A 2-step sequence to 1α -hydroxytestosterone from C1-borane adduct 7

Treatment of borane 7 with stoichiometric H_2O_2 and NaOH in THF gave 1α -hydroxyandrostenedione (Scheme 2, 8, Step 2 of 8 steps). This oxidation of the C1-boryl substituent to the alcohol was stereospecific and retained the 1α -orientation of the substituent (*i.e.* the 1α -borane substituent was oxidized to



Scheme 2 Synthesis of 1α -hydroxytestosterone (9) from C1-borylated steroid intermediate (7).



Scheme 3 Synthesis of 3-tert-butyldimethylsiloxy-1,17-diketo-androst-4-ene (13) from 1 α -hydroxyandrostenedione (8) (Steps 3–5 of 8 steps). The intermediate in brackets explains the stereoselective reduction at C3 through the chelation of the calcium Lewis acid with the C1 α -hydroxy group and the borohydride to deliver the hydride at the bottom face, yielding 11.

the 1α -hydroxy substituent with H_2O_2). Reduction of the C17-ketone of 1α -hydroxyandrostenedione with NaBH_4 in CH_3OH gave 1α -hydroxytestosterone (9). This route to 1α -hydroxytestosterone from the boron intermediate contrasts with the previous strategy^{7,8} to incorporate the 1 -hydroxy group, which involved the nucleophilic epoxidation of the 3-keto- Δ^1 steroid followed by reduction with LiAlH_4 to yield the $\text{C}1\alpha$ -hydroxy steroid.

Conversion of 1α -hydroxyandrostenedione to 1β -hydroxytestosterone (8 to 2)

For the synthesis of 1β -hydroxytestosterone (2), various methods were performed to incorporate the 1β -hydroxy group (see section S12) but the ultimate strategy involved the oxidation of the 1α -hydroxy group to the C1-ketone, which was stereoselectively reduced to the 1β -hydroxy group (Scheme 3). To begin, 1α -hydroxyandrostenedione was treated with CaCl_2 and NaBH_4 in CH_3OH ¹⁴ to primarily afford the 3β -hydroxy epimer 11 in 82% yield with a minor amount of the 3α -hydroxy epimer 10 (Scheme 3, 2 to 11, Step 3 of 8 steps). The presence

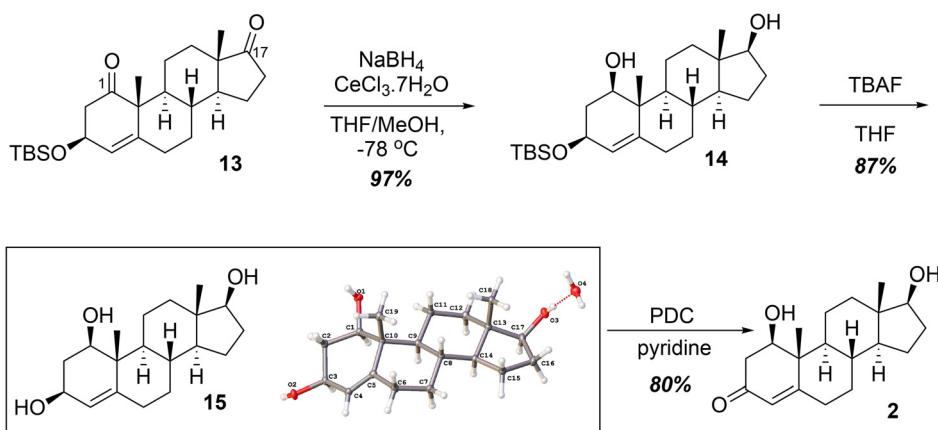
of the CaCl_2 forms $\text{Ca}(\text{BH}_4)_2$, which in turn enables calcium to chelate^{15,16} with the C1-hydroxy group of the substrate and the borohydride reducing agent. This chelation directs the hydride to attack on the bottom face giving the 3β -hydroxy epimer (11) as the major product. Alternatively, the Luche reduction conditions (NaBH_4 in the presence of $\text{CeCl}_3 \cdot 7\text{H}_2\text{O}$) yielded an epimeric mixture of the triols 10 and 11 (45 : 55 ratio, see SI for details) due to the lack of a chelation with the C1-oxygen. Triol 11 was regioselectively protected at C3 as the TBS ether 12 using TBSCl and pyridine as both the base and the solvent (11 to 12, Step 4 of 8 steps). Pyridine was required to dissolve the triol (11). The resulting 1,17-diol (12) was oxidized with 3 mol eq. of PCC in CH_2Cl_2 to yield the diketone 13 (12 to 13, Step 5 of 8 steps), which was used as the precursor to introduce the key 1β -hydroxy group in the steroid backbone.

The stereoselective reduction of the C1-ketone intermediate (13) to yield the 1β -hydroxy epimer was not trivial. Our past work in the stereoselective reduction of a C12-ketone guided us in this optimization process.¹⁷ Table 2 shows a set of reaction conditions, which led us to conclude that the Luche reduction at $-78\text{ }^\circ\text{C}$ was the optimal method to yield the desired 1β -hydroxy stereoisomer. The use of L-selectride as a sterically hindered hydride source (Table 2, entry 1), gave mostly the 1α -hydroxy epimer product (12) (74 : 26, 12 to 14). On the other hand, a smaller reducing agent such as NaBH_4 gave more of the desired 1β -hydroxy epimer product (14) relative to L-selectride (entry 2, 54 : 46, 12 to 14). In addition, three factors to optimize this reaction followed (see entry 3): (i) lowering the temperature to $-78\text{ }^\circ\text{C}$, (ii) the use of THF as a co-solvent to enhance solubility of the C1-ketone starting material (13), and (iii) the addition of $\text{CeCl}_3 \cdot 7\text{H}_2\text{O}$ to ensure reactivity of the hydride at $-78\text{ }^\circ\text{C}$. The successful stereoselective reduction of 1,17-diketone 13 using $\text{CeCl}_3 \cdot 7\text{H}_2\text{O}$ and NaBH_4 in CH_3OH and THF at $-78\text{ }^\circ\text{C}$ gave the desired $1\beta,17\beta$ -diol 14 (Table 2, entry 3, 6.0 : 94, 12 to 14, Step 6 of 8 steps). When the reaction was performed at rt, the C1-epimers ($1\alpha/1\beta$ hydroxy epimers) were obtained in a 1 to 1 ratio (entry 4). The low temperature of the reduction with a small reducing agent (Luche con-

Table 2 Optimization of Step 6: stereoselective reduction of C1-ketone (13) to yield primarily the 1β -hydroxy epimer

Entry	Reaction conditions	1 α -Hydroxy (12)	1 β -Hydroxy (14)
1 ^a	L-Selectride, THF, $-78\text{ }^\circ\text{C}$	74%	26%
2 ^a	NaBH_4 , CH_3OH , rt	54%	46%
3 ^{a,c}	NaBH_4 , $\text{CeCl}_3 \cdot 7\text{H}_2\text{O}$, $\text{CH}_3\text{OH}/\text{THF}$, $-78\text{ }^\circ\text{C}$	6.0%	94%
4 ^{b,d}	NaBH_4 , $\text{CeCl}_3 \cdot 7\text{H}_2\text{O}$, $\text{CH}_3\text{OH}/\text{THF}$, rt	47%	53%

^a The ratio of the α - and β -hydroxy epimers (12 and 14) were determined by integrating the Δ^4 proton at δ 5.32 and 5.27. ^b The ratio of the α - and β -hydroxy epimers (12 and 14) were determined by TLC analysis R_f : 0.634 and 0.846, respectively (1 to 1 ethyl acetate/hexanes, v/v). ^c 1 to 1 ratio of $\text{CH}_3\text{OH}/\text{THF}$, 2 mol eq. of NaBH_4 , 2 mol eq. of $\text{CeCl}_3 \cdot 7\text{H}_2\text{O}$. ^d 1 to 1 ratio of $\text{CH}_3\text{OH}/\text{THF}$, 3 mol eq. of NaBH_4 , 2 mol eq. of $\text{CeCl}_3 \cdot 7\text{H}_2\text{O}$.



Scheme 4 Synthesis of 1β -hydroxytestosterone (2) from 1,17-diketone 13 (Steps 6–8 of 8 steps).



ditions) avoids torsional strain between C9 and the oxygen (see Fig. S3-3).

Deprotection of the 3-TBS group with excess TBAF in THF afforded triol **15** (Step 7 of 8 steps). Triol **15** was regioselectively oxidized at C3 with PDC (1 mol equivalent) in pyridine to furnish 1 β -hydroxytestosterone (**2**) in 80% isolated yield (Step 8 of 8 steps). When pyridine was used as the solvent, the triol was completely soluble and the main product isolated was the desired oxidation product at C3 to yield 1 β -hydroxytestosterone (**2**). The C3-hydroxy group is the least sterically hindered alcohol among the three positions of triol **15** (*i.e.* C1, C3, and C17) where C1 and C17 are both adjacent to a quaternary carbon center (C10 and C13, respectively). The regioselective oxidation of the less hindered C3 alcohol over the more congested alcohols at C1 and C17 is reminiscent of a prior study, which used cholesterol oxidase¹⁸ to selectively oxidize the C3-position of 7 α -hydroxycholesterol to yield 7 α -hydroxy-cholest-4-en-3-one.

Conclusion

In conclusion, a synthesis of 1 β -hydroxytestosterone (**2**) was achieved from commercially available androst-1,4-dien-3,17-dione (**6**) through a 1,4-borylation reaction (Scheme 1). The use of DBU as the base to generate the carbene was necessary to optimize the yield for the conjugate borylation (see Fig. S2-2). Other key steps include: (i) the regioselective and stereoselective reduction of a 3-keto- Δ^4 -intermediate to yield a 3 β -hydroxy Δ^4 product using CaCl_2 and NaBH_4 in CH_3OH ¹⁴ (Scheme 3, **8** to **11**) and (ii) stereoselective reduction of a C1-ketone intermediate to yield primarily the 1 β -hydroxy epimer under Luche reduction conditions at -78°C (Scheme 4, **13** to **14**). Furthermore, 1 α -hydroxytestosterone was accessed in 3 steps from commercially available starting materials (Schemes 1 and 2, **6** to **9**), which contrasts from the previously reported synthesis of 1 α -hydroxytestosterone involving 9 steps.⁸ The C1-borylation strategy can be used to access other naturally occurring steroids,¹⁹ including those that are not commercially available and have important biological applications.^{20,21}

Conflicts of interest

There are no conflicts to declare.

Data availability

The data that support the findings of this study are available on request from the corresponding author, F. K. Y.

Supplementary information: to show the experimental details (procedures, NMR, mass spectrometry, IR) to synthesize the compounds in the main text (Section S1), optimization of the borylation of boldione (Section S2, **6** to **7**), optimization of the stereoselective reduction of the C1-ketone to the 1 β -hydroxy product (Section S3, **13** to **14**), and X-ray structures

of the synthesized compounds (Section S4, **7**, **9**, **10**, **12**, **13**, and **15**. See DOI: <https://doi.org/10.1039/d5ob01218j>.

CCDC 2448105, 2448104, 2448099, 2448100, 2448103 and 2448102 contain the supplementary crystallographic data for this paper.^{22a-f}

Acknowledgements

This research was supported by the Max and Minnie Tomerlin Voelcker Fund. Francis K. Yoshimoto, Ph.D. holds a Voelcker Fund Young Investigator Award from the MAX AND MINNIE TOMERLIN VOELCKER FUND. A. I. E. is funded by National Institutes of General Medical Sciences of the National Institutes of Health MARC under award number T34GM145507. This research was also supported by the National Institutes of Health Grant R35 GM151905 (F. P. G.) and the National Science Foundation Graduate Research Fellowship Program under Grant No. 1937963 (K. D. M.). We thank the reviewers and editors for helping us improve the quality of this manuscript. This manuscript is dedicated to Professor Richard J. Auchus (University of Michigan). F. K. Y. is grateful for his mentorship, friendship, and support.

References

- 1 S. Takeji, M. Okada, S. Hayashi, K. Kanamaru, Y. Uno, H. Imaishi and T. Uno, *Biopharm. Drug Dispos.*, 2023, **44**, 420–430.
- 2 H. Yamazaki and T. Shimada, *Arch. Biochem. Biophys.*, 1997, **346**, 161–169.
- 3 J. A. Krauser and F. P. Guengerich, *J. Biol. Chem.*, 2005, **280**, 19496–19506.
- 4 S. E. Kandel, L. W. Han, W. Mao and J. N. Lampe, *Drug Metab. Dispos.*, 2017, **45**, 1266–1275.
- 5 J. A. Krauser, M. Voehler, L.-H. Tseng, A. B. Schefer, M. Godejohann and F. P. Guengerich, *Eur. J. Biochem.*, 2004, **271**, 3962–3969.
- 6 W. Chen, M. J. Fisher, A. Leung, Y. Cao and L. L. Wong, *ACS Catal.*, 2020, **10**, 8334–8343.
- 7 P. K. Sharma and A. Akhila, *Indian J. Chem.*, 1991, **30B**, 554–556.
- 8 J. Mann and B. Pietrzak, *Tetrahedron*, 1989, **45**, 1549–1552.
- 9 S. D. Offei, H. D. Arman and F. K. Yoshimoto, *Steroids*, 2022, **186**, 109088.
- 10 A. P. Marcus and R. Sarpong, *Org. Lett.*, 2010, **12**, 4560–4563.
- 11 K.-S. Lee, A. R. Zhugralin and A. H. Hoveyda, *J. Am. Chem. Soc.*, 2009, **131**, 7253–7255.
- 12 Y. Wang, H. Tan and J. Gui, Gram-Scale Synthesis of Bufospirostenin A by a Biomimetic Skeletal Rearrangement Approach, *J. Am. Chem. Soc.*, 2021, **143**, 19576–19586.
- 13 H. Wu, S. Radomkit, J. M. O'Brien and A. H. Hoveyda, *J. Am. Chem. Soc.*, 2012, **134**, 8277–8285.



14 H. Fujii, K. Oshima and K. Utimoto, *Chem. Lett.*, 1991, 1847–1848.

15 N. V. Forkel, D. A. Henderson and M. J. Fuchter, *Tetrahedron Lett.*, 2014, **55**, 5511–5514.

16 M. Seki and Y. Takahashi, *Org. Process Res. Dev.*, 2021, **25**, 1950–1959.

17 S. D. Offei, H. D. Arman, M. O. Baig, L. S. Chavez, C. A. Paladini and F. K. Yoshimoto, *Steroids*, 2018, **140**, 185–195.

18 D. L. Alexander and J. F. Fisher, *Steroids*, 1995, **60**, 290–294.

19 R. Mohan, H. Hammers, P. Bargagna-mohan, X. Zhan, C. Herbstritt, A. Ruiz, L. Zhang, A. Hanson, B. Conner, J. Rougas and V. Pribluda, *Angiogenesis*, 2004, **7**, 115–122.

20 M. A. Hayes, X.-Q. Li, G. Gronberg, U. Diczfalusy and T. B. Andersson, *Drug Metab. Dispos.*, 2016, **44**, 1480–1489.

21 M. A. Hayes, I. Roberts, G. Gronberg, K. Lv, B. Lin, J. Bergare and C. S. Elmore, *J. Labelled Compd. Radiopharm.*, 2017, **60**, 221–229.

22 (a) A. I. Elizondo, K. D. McCarty, H. D. Arman, F. P. Guengerich and F. K. Yoshimoto, CCDC 2448105: Experimental Crystal Structure Determination, 2025, DOI: [10.5517/ccde.csd.cc2n5g4g](https://doi.org/10.5517/ccde.csd.cc2n5g4g); (b) A. I. Elizondo, K. D. McCarty, H. D. Arman, F. P. Guengerich and F. K. Yoshimoto, CCDC 2448104: Experimental Crystal Structure Determination, 2025, DOI: [10.5517/ccde.csd.cc2n5g3f](https://doi.org/10.5517/ccde.csd.cc2n5g3f); (c) A. I. Elizondo, K. D. McCarty, H. D. Arman, F. P. Guengerich and F. K. Yoshimoto, CCDC 2448099: Experimental Crystal Structure Determination, 2025, DOI: [10.5517/ccde.csd.cc2n5fy7](https://doi.org/10.5517/ccde.csd.cc2n5fy7); (d) A. I. Elizondo, K. D. McCarty, H. D. Arman, F. P. Guengerich and F. K. Yoshimoto, CCDC 2448100: Experimental Crystal Structure Determination, 2025, DOI: [10.5517/ccde.csd.cc2n5fz8](https://doi.org/10.5517/ccde.csd.cc2n5fz8); (e) A. I. Elizondo, K. D. McCarty, H. D. Arman, F. P. Guengerich and F. K. Yoshimoto, CCDC 2448103: Experimental Crystal Structure Determination, 2025, DOI: [10.5517/ccde.csd.cc2n5g2d](https://doi.org/10.5517/ccde.csd.cc2n5g2d); (f) A. I. Elizondo, K. D. McCarty, H. D. Arman, F. P. Guengerich and F. K. Yoshimoto, CCDC 2448102: Experimental Crystal Structure Determination, 2025, DOI: [10.5517/ccde.csd.cc2n5g1c](https://doi.org/10.5517/ccde.csd.cc2n5g1c).

