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Influence of magnetic convection on separation efficiency in magnetophoretic microfluidic processes: a combined simulation and experimental study†

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This work explores the complex hydrodynamics in magnetophoretic microfluidic processes, focusing on the interplay of forces and particle concentrations. The study employs a combined simulation and experimental approach to investigate the impact of magnetophoresis on magneto-responsive nanoparticles (MNPs) and their environment, including non-magneto-responsive nanoparticles (non-MNPs) in a microfluidic system. Our findings reveal that the motion of MNPs induces a hydrodynamic convective motion of non-MNPs, significantly affecting the separation efficiency and purity of the particles. The separation efficiency of MNPs increases with the Péclet number, reflecting the increase in the magneto-phoretic force, but decreases with lower concentrations. Conversely, non-MNPs exhibit high and constant separation efficiency with increasing Péclet number, independent of the magnetophoretic force. In a mixture, the separation efficiency of non-MNPs decreases, suggesting that non-MNPs drag along MNPs. The Mason number, representing the ratio between shear and magnetophoretic force, also plays a crucial role in the separation process. The results underscore the need for careful control and optimization of the Péclet and Mason numbers, as well as particle concentrations, for efficient magnetophoretic microfluidic processes. This study provides valuable information on the underlying principles of magnetophoresis in microfluidic applications, with implications for biochemistry, biomedicine, and biotechnology.

1 Introduction

In recent years, the development of lab-on-a-chip platforms has garnered significant attention in the field of biomedical research, including disease diagnosis, drug development, and

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personalized medicine. These miniaturized analytical devices offer a range of advantages over traditional laboratory techniques, including reduced sample volumes, faster analysis times, and improved sensitivity and selectivity.[1,2]

The basic principle of a lab-on-a-chip platform involves the integration of multiple laboratory functions onto a single microchip, allowing the parallel processing of multiple samples having a high degree of controllability and optimization. This integration is achieved through the use of microfluidics, which enables manipulation at the microscale level[2,3] and precise control of fluid flow with low Reynolds numbers[4] ranging from 10^{-6} to 10. Using microfluidic technology, lab-on-a-chip platforms are capable of performing a range of functions, including magnetophoresis-based cell sorting and detection,[5] DNA sequencing,[6] and protein analysis.[7] The potential impact of lab-on-a-chip platforms in biomedical research is huge. For example, using them for point-of-care diagnostics could greatly improve access to medical care in low-resource settings, where traditional laboratory techniques are often prohibitively expensive or logistically challenging.[8,9] Similarly, performing high-throughput drug screening on a microfluidic platform could greatly accel-

erate the drug development process, leading to more efficient and cost-effective drug discovery.[1]

In transport processes, the Péclet number describes the ratio of convective to diffusive forces. Controlling the Péclet number is crucial in microfluidic devices to optimize either a sorting process or to achieve efficient mixing in reaction processes.[4,10] In magnetophoresis-based processes, one key challenge is the need for a comprehensive theory for predicting and understanding the dynamic behavior of magneto-responsive nanoparticles (MNPs).[11–13] These magneto-responsive nanoparticles can be engineered with different surface coatings and functionalizations to serve as carriers for a particular substance or to specifically target cells.[14–17] Moreover, the presence of magnetic fields can lead to complex emergent behavior; such as spontaneous pattern formation and self-assembly.[18] When this magnetic field varies in space, MNPs will experience a force that causes them to migrate. This phenomenon is commonly referred to as magnetophoresis, which, similar to its analogous "electrophoresis" for charged nanoparticles,[19] has frequently been modeled *via* mean-field approximations.[20] Nonetheless, thermal fluctuations and discrete particle interactions are essential at the mesoscale, making classic mean-field theories poorly equipped to accurately simulate and predict the transport and equilibrium properties of magnetic colloidal suspensions. Particle-based simulations, such as molecular or Brownian dynamics simulations, can provide a picture of the underlying physical phenomena present in colloidal suspensions. However, due to the long-ranged and many-bodied nature of the hydrodynamic and magnetic forces present in magnetophoretic processes, previous computational work has only considered magnetic species and mainly focused on the aggregation kinetics of magnetic agglomerates.[16,21]

This study aims to explore the transport mechanisms in colloidal dispersions of MNPs and non-MNPs in the presence of a spatially varying magnetic field, particle concentration, and particle solution flow and their effect on the separation efficiency (schematically shown in Fig. 1). Notably, the influence of magnetic convection in a hydrodynamic system comprising magnetic and non-magneto-responsive entities in a microfluidic or millifluidic chip has yet to be previously investigated.[3,22,23] We compliment and support our findings with large-scale Brownian dynamics simulations coupled to the magnetic equivalent of Poisson's equation to solve for the magnetostatic field of a mixture of magneto-responsive and non-magneto-responsive nanoparticles in the presence of a magnetic field gradient. The hydrodynamic forces between colloidal particles are computed using a positively split Ewald method[24] and the magnetostatic potential is computed using a spectral Ewald method.[25,26] We present a combined simulation and experimental approach to demonstrate that convection motion is an underappreciated effect in magnetohydrodynamic systems.

2 Theory and numerical methods

2.1 Brownian dynamics simulation

We model the mixture of MNP and non-MNP as a suspension of charged, hard, spherical particles of radius a. We assume that these particles are suspended in a continuum Newtonian fluid such that the colloids interact with the solvent only *via* stochastic Brownian forces from fluctuations in the thermal energy or fluctuating hydrodynamics due to the momentum relaxation of the solvent molecules.[27] Under these assumptions we can start with Langevin's equation to derive our equations of motion:

$$\mathbf{m} \cdot \frac{\mathrm{d}\mathbf{u}_\alpha}{\mathrm{d}t} = \mathbf{F}_\alpha^H + \mathbf{F}_\alpha^I + \mathbf{F}_\alpha^M + \mathbf{F}_\alpha^B, \tag{1}$$

where $\mathbf{u}_\alpha$ is the velocity of the αth colloid, $\mathbf{F}_\alpha^H$ is the hydrodynamic force acting on the αth colloid, $\mathbf{F}_\alpha^I$ accounts for forces arising from a generic conservative potential, $\mathbf{F}_\alpha^M$ is the magnetophoretic force due to the magnetic field gradient[28] and $\mathbf{F}_\alpha^B$ is the stochastic Brownian force. The last force satisfies the fluctuation–dissipation theorem[19] with ensemble average:

$$\langle \mathbf{F}^B(t) \rangle = 0; \quad \langle \mathbf{F}^B(t)\mathbf{F}^B(t+\tau) \rangle = 2k_B T (\mathbf{M}^H)^{-1} \delta(\tau) \tag{2}$$

where $\mathbf{F}^B(t) = [\mathbf{F}_1{}^B(t), \mathbf{F}_2{}^B(t),...]$, $\langle \cdot \rangle$ indicates the expectation value, δ is the Dirac delta function, and $\mathbf{M}^H$ is the hydrodyn-
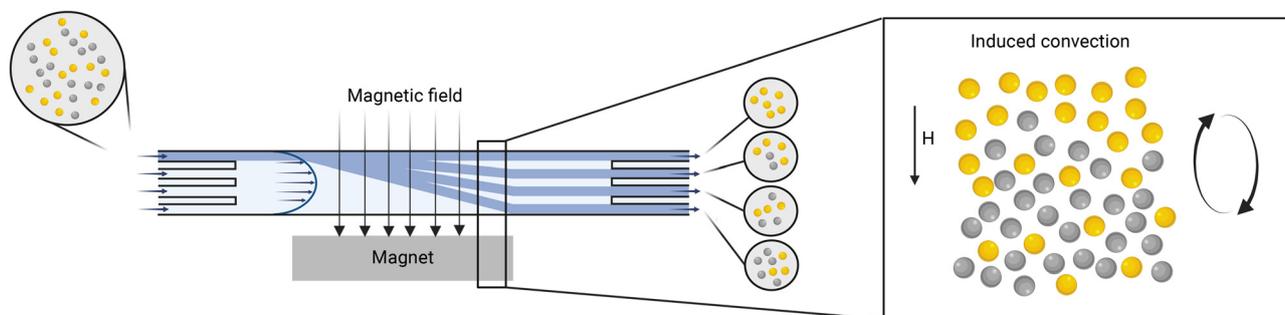

Fig. 1 Scheme of the microfluidic chip sorting a mixture of MNPs (grey) and non-MNPs (yellow) within a spatially varying magnetic field. The MNPs generate a hydrodynamic force influencing the non-MNPs, resulting in their co-migration alongside the MNPs. This magnetically induced convective motion is analyzed through both experimentally and large-scale Brownian dynamic simulation. Created with Biorender.com.

amic mobility tensor. This formulation ensures that any energy a colloid gains from a thermal fluctuation is dissipated as drag to the solvent.

A more mathematically rigorous formulation states that the Brownian force, consistent with the distribution in the second term of eqn (2), is sampled from a collection of independent Wiener processes, $\mathbf{W}(t)$, expressed as

$$\mathbf{F}^{B} = \sqrt{2k_{B}T}\left(\mathbf{M}^{H}\right)^{-1/2}\cdot\frac{d\mathbf{W}}{dt}. \quad (3)$$

Here, the square root of the inverse of the mobility matrix, $\left(\mathbf{M}^{H}\right)^{-1/2}$, satisfies the relation $\left(\mathbf{M}^{H}\right)^{-1/2} \cdot \left(\mathbf{M}^{H}\right)^{-1/2\dagger} = \left(\mathbf{M}^{H}\right)^{-1}$, where $\dagger$ denotes the adjoint (or conjugate transpose).

At the colloidal scale, inertial relaxation occurs on time scales orders of magnitude smaller than those governing colloidal motion. Consequently, any perturbation to the particle's momentum dissipates almost instantaneously. Rigorous integration of eqn (1) results in a set of stochastic differential-algebraic equations (SDAE) that faithfully capture the *overdamped* Langevin dynamics:

$$\mathbf{u}_{\alpha}(t) = \mathbf{M}^{H}_{\alpha\beta}\cdot\left(\mathbf{F}^{I}_{\beta} + \mathbf{F}^{M}_{\beta}\right) + \sqrt{2k_{B}T\mathbf{M}^{H}_{\alpha\beta}}\cdot\frac{d\mathbf{W}}{dt} \\ + k_{B}T\nabla\cdot\mathbf{M}^{H}_{\alpha\beta} \quad (4)$$

The final term in eqn (4), often referred to as the Brownian or thermal drift, emerges from integrating out the inertial degrees of freedom in eqn (1). Physically, this drift introduces an additional flux that actively displaces particles from regions of low mobility,[29] preventing their accumulation and ensuring a uniform exploration of the system's phase space.

Good agreement between experimental measurements and simulation results for a suspension of colloidal particles (including concentrated ones) can be achieved with a relatively coarse approximation for the interparticle hydrodynamic interactions. The fields generated by beads in a Newtonian fluid can be approximated as the superposition of the field generated by a point particle and a point quadrupole.[24] When all colloidal particles are of the same size, the hydrodynamic fields are given by the Rotne–Prager–Yamakawa mobility tensor:[19]

$$\mathbf{M}^{H}_{\alpha\beta} = \frac{1}{\gamma}\begin{cases}\left(\frac{3a}{4r}+\frac{a^{3}}{2r^{3}}\right)\mathbf{I} + \left(\frac{3a}{4r}-\frac{2a^{3}}{2r^{3}}\right)\hat{\mathbf{r}}\hat{\mathbf{r}}, & r > 2a \\ \left(1-\frac{9r}{32a}\right)\mathbf{I} + \left(\frac{3r}{32a}\right)\hat{\mathbf{r}}\hat{\mathbf{r}}, & r \leq 2a\end{cases} \quad (5)$$

Naive evaluation of the mobility tensor involves computational operations scaling as $O(N_{2})$, while its inversion, necessary for calculating the Brownian displacements in eqn (2), incurs a complexity of $O(N_{3})$. This computational burden often limits the feasibility of simulating large-scale systems. However, a substantial reduction in numerical complexity can be achieved by recognizing that the mobility tensor is positive-definite.[30]

This positive-definiteness allows for fast algorithms, such as those yielding $O(N\log N)$ scaling for evaluating particle vel-

ocities from applied forces.[24] Furthermore, the Positively Split Ewald (PSE) method, introduced by Fiore, Swan, and colleagues, enables linear scaling in the sampling of Brownian displacements.[31]

In periodic systems, the mobility tensor in eqn (5) exhibits translational invariance, making it divergence-free and resulting in a zero Brownian drift term in eqn (4). In this case, the trajectories over time can be numerically solved *via* a forward Euler–Maruyama integration scheme:

$$\mathbf{x}_{\alpha}(t + \Delta t) = \mathbf{x}_{\alpha}(t) + \mathbf{u}_{\alpha}(t)\Delta t \quad (6)$$

where Δt is the time step over which ion trajectories are advanced. Nonetheless, the situation becomes more intricate when simulating bounded geometries or employing more sophisticated hydrodynamic models. For example, in confined environments[32–34] or higher-order approximations,[35,36] the mobility tensor may lose its divergence-free character. In such cases, additional corrections are required. For example, when employing *constrained* Rotne–Prager–Yamakawa (RPY) hydrodynamics, it becomes essential to account for the Brownian drift to ensure the accurate resolution of particle dynamics and the generation of physically consistent trajectories.[29,36,37]

Forces arising from conservative interactions among colloidal particles are represented as the gradient of a potential energy $U(\mathcal{X})$, which is a function of the coordinates of all ions $\mathcal{X} \equiv [x_{1}, x_{2}, \ldots, x_{N}]^{T}$,

$$\mathbf{F}^{I/M}_{\alpha}(\mathcal{X}) \equiv -\nabla_{x_{\alpha}}U^{I/E}(\mathcal{X}), \quad (7)$$

where the gradient is taken with respect to the position of the αth particle. While the electrostatic interactions between the charged species can be taken into account as described by the Gouy–Champman model, the electromagnetic interactions and the influence of the magnetic field gradient are obtained by solving Maxwell's equations in the form

$$\nabla\cdot\mathbf{B} = 0, \quad \mathbf{B} = \mu\mathbf{H} \quad (8)$$

Paramagnetic particles of magnetic permeability μ_{p} in a solvent of permeability μ_{s}. By defining a magnetic scalar potential ψ, the problem reduces to Laplace's equation:

$$\nabla^{2}\psi = 0 \quad (9)$$

subject to

$$\left(\mathbf{B}_{f} - \mathbf{B}_{p}\right)\cdot\hat{n} = 0. \quad (10)$$

where $\mathbf{B}_{p}$ and $\mathbf{B}_{f}$ are the magnetic fluxes inside and outside of the particle. These equations are valid in the limit that the magnetic relaxation time inside of the particle and fluid is significantly smaller than the time scale of the study. As we are interested in timescales of particle motion $O(1s)$, the system is said to be pseudosteady. Thus, time dependence arises solely due to the time-varying boundary conditions in 10 as particles move around or the external field varies. A multipole expansion of the integral representation of Laplace's equation can be used to derive a linear system of equations that couples the magnetic flux moments on the particle to the magnetic poten-

tial and potential gradients on the fluid.[28] Considering the dipole contribution and ignoring quadrupoles and higher-order moments, we obtain:

$$\mathbf{H}_0 = \sum_{\alpha,\beta}^{N} \mathbf{M}_{\alpha\beta} \cdot \mathbf{m}_i, \tag{11}$$

where $\mathbf{M}_{\alpha\beta}$, the magnetic flux moments of particle β with particle α in a periodic box

$$\mathbf{M}_{\alpha\beta} = \frac{3\mathbf{I}\delta_{\alpha\beta}}{4\pi a^3 \left(\mu_p - \mu_f\right)} + \frac{9}{a^2 \mu_f V} \sum_{\mathbf{k} \neq 0} \frac{e^{i\mathbf{k}\cdot(x_\alpha - x_\beta)}}{k^2} j_1^2(k_a)\hat{\mathbf{k}}\hat{\mathbf{k}}, \tag{12}$$

where $j_1(x) = \sin(x)/x^2 - \cos(x)/x$ is the first order spherical Bessel function of the first kind, and $\mathbf{k} = [2\pi\kappa_x/L_x, 2\pi\kappa_y/L_y, 2\pi\kappa_z/L_z]$ for integers κ_i. The dipole–dipole interactions contribution to the potential energy can be obtained via the sum of the pairwise interactions given by

$$U^M = -\frac{1}{2}\sum_{\alpha=1}^{N} \mathbf{H}_0 \cdot \mathbf{m}_\alpha = -\frac{1}{2}\sum_{\alpha,\beta}^{N} \mathbf{M}_{\alpha\beta} : \mathbf{m}_\alpha\mathbf{m}_\beta. \tag{13}$$

Similarly, the resulting magnetic force acting on the αth particle is

$$\mathbf{F}^M = -\nabla_\alpha U^M = -\frac{1}{2}\sum_{\alpha,\beta}^{N} \frac{\partial \mathbf{M}_{\alpha\beta}}{\partial x_\alpha} : \mathbf{m}_\alpha\mathbf{m}_\beta \tag{14}$$

We simulate a mixture of MNPs and non-MNPs of the equal radius, $\alpha \approx 47$ nm, in water a room temperature. We assume that the solvent behaves as a Newtonian fluid, with a viscosity of $\mu = 1$ cp, constant magnetic permeability, $\mu_f \approx 1.25 \times 10^3$ H nm^{-1}, and constant permittivity $\varepsilon_f = 80$. Furthermore, the colloidal particles are electrostatically stabilized in a binary electrolyte, such as NaCl, which is simulated implicitly. The concentration of the implicit ions is estimated using Graham's equation[38] based on the experimental zeta potential of $\zeta_{MNP} = -24.26$ mV and $\zeta_{non-MNP} = -43.48$ mV.

2.2 Characterization of the magneto-responsive suspension using Péclet and Mason number

The displacement of the magneto-responsive particles results in a perturbation of the microstructure of the suspension which can be characterized by the Péclet number.[39] This non-dimensional number expresses the importance of the external force deforming the system, F^{ext}, with respect to the restoration of entropic spring forces from Brownian motion,[30,40,41]

$$Pe = \frac{F^{ext}}{2k_BT/a} \tag{15}$$

where F^{ext} is any externally applied force to the suspension of interest, k_BT, is the thermal energy, and a is the hydrodynamic radius of the particles. In our analysis, the external force is the magnetophoretic force, defined as the dot product of the

induced dipole on the αth colloid, $\mathbf{m}_\alpha$, and the magnetic field gradient: $\nabla\mathbf{H} = G \cdot \hat{\mathbf{H}}_0$, where $\hat{\mathbf{H}}_0 = \mathbf{H}_0/H_0$ is the unit vector of the externally applied magnetic field of magnitude $H_0 = |\mathbf{H}_0|$. In our simulations, the field is applied on the $\hat{\mathbf{z}}$ direction.

For paramagnetic particles, the magnetic dipole is *induced* by the field and assumed to be perfectly aligned with the field. Moreover, in our simulations, the magnetic field only varies in the direction of the applied field, $\hat{\mathbf{z}}$. Then, the only non-zero components of the field gradient 2-tensor $\nabla\mathbf{H}$ are those in the $\hat{\mathbf{z}}$ direction. To facilitate our analysis, we define a gradient vector containing the gradient direction and field magnitude, $G = \nabla H\hat{\mathbf{z}} = G\hat{\mathbf{z}}$. By definition, $\mathbf{m}_\alpha = \frac{4}{3}\pi a^3 \mu_f\chi\mathbf{H}_0$, where μ_f is the fluid permittivity, and $\chi = 3(\mu_p/\mu_f - 1)/(\mu_p/\mu_f + 2)$ is the susceptibility, with μ_p being the particle's permittivity. Then, this *magnetic* Péclet number is given by Pe $= (4\pi a^3 \mu_f\chi\nabla|\mathbf{H}|^2)/3k_BT \sim (a^4\mu_f\chi GH_0)/k_BT$, where G is the field gradient magnitude. This equation tells us that in the limit of very small Péclet numbers, the deformation of the particle arrangement within the suspension, and consequently the local polarization density, due to the externally imposed magnetic gradient is easily restored by the Brownian bath. On the other hand, high Péclet numbers indicate that the arrangement of particles and thus the macroscopic properties of the suspension are defined by the magnetophoretic process.

In our experimental set-up, the presence of no-slip boundary conditions in a Newtonian fluid results in an additional tangential force due to momentum transfer (a shear force), which further deforms the suspension. Assuming that the shear force is orthogonal to the magnetic gradient, as in our experimental set-up, this deformation will be opposed in the simplest of cases by the dipole–dipole interactions of the magnetic particles. The competition of both forces is given by a Mason number,[42,43] defined as

$$Ma = \frac{6\pi\eta a^2\dot{\gamma}}{F_{dip}}, \tag{16}$$

where $\dot{\gamma}$ is the shear rate and F_{dip} is obtained by taking the gradient of the potential energy with respect to the distance between a pair of magnetic interacting dipoles.[44] In the context of microfluidic devices employed for separation processes, performance evaluation revolves around two key metrics: Purity and separation efficiency. The purity, P, of the outlet streams is quantified, where the ratio of concentration to velocity serves as the defining factor for mass flux $\dot{n}$ of components A and B: $P_A = \dot{n}_{out,A}/(\dot{n}_{out,A} + \dot{n}_{out,B})$. The separation efficiency S is defined as $S_A = \dot{n}_{out,A}/\dot{n}_{in,A}$.

3 Experimental methods

3.1 Microfluidic platform

The microfluidic chip was designed in SolidWorks 2021 and printed by a FormLabs 3B + printer using clear resin (Formlabs, RS-F2-GPCL-04). The chip was placed horizontally and twisted in every direction to facilitate the later washing

process. When the print was finished, the uncured resin was flushed out with 100% 2-propanol *via* a syringe. Then, the chip was washed for 15 min in a 2-propanol bath and cured by UV light at 60 °C for 20 min. Finally, the supporting structures were removed, and the printed M5 threads were drilled into the in- and outlets, enabling the connection of the silicon tubes (1.3 mm, VWR) *via* Luer lock tubing adapters (male, 1.6 mm, Reichelt Chemie Technik GmbH & Co.).

The experiments were performed at a viscosity η of 2.2274 mPa s, increasing the colloidal stability of the nanoparticles.[45] Therefore, the nanoparticles are dissolved in 24 wt% D(+)-sucrose (Carl Roth) at pH 7. The concentration stayed the same for the mixture experiments as for the single experiments per particle species. This means the overall particle mass flux for a mixture experiment was twice the one of the single experiment ($\dot{m}_{single,SFP} = \dot{m}_{mix,SFP}$). The density for the particles is 2650 for the non-magneto-responsive and 4870 kg m^{-3} for the MNPs, respectively. For the experimental set-up, the chip and the neodymium iron boron magnet (4 × 0.5 × 1.5 cm, N40, supermagnete) were placed in the holder, and the tubes were connected. The sample was pumped at 52 μL min^{-1} (u = 0.00107 m s^{-1}) by a syringe pump (kdScientific), the buffer *via* a peristaltic pump (ISMATEC) with 37% (u = 0.00425 m s^{-1}), and the outlet samples were removed with 229 μL min^{-1} by a self-made multi-syringe adapter for a syringe pump (Krüss, Havard Apparatus) as presented in Fig. 2 and in the ESI section.† The flow of the viscous solution should be higher than the sample flow to minimize the transverse diffusional migration of the nano-

particles. For running the experiments, the chip was first slowly filled with the viscous solution, then the sample was connected, avoiding bubble formation. After 30 s of equilibration time, the outlet tubes were connected, and each experiment was run for 4 min in triplicates (n = 3, error bars presenting the standard deviation). The outlets A/B and C/D are combined to calculate the process purity and selectivity.

The nanoparticle concentration was analyzed in triplicates *via* UV absorbance at 400 nm and fluorescent measurement (excitation wavelength 488 nm, emission wavelength 520 nm) in a plate reader (Tecan Infinite M200 Microplate Reader). They were compared to standard calibration curves; if necessary, the samples were centrifugally concentrated and resuspended in an ultrasonication bath (132 kHz, Sonorex) for 15 min.

The details of nanoparticle synthesis are provided in the ESI section.†

4   Results

4.1   Exploring the dependence of magnetophoresis on magnetic field and nanoparticle concentration

The separation efficiencies and purities for separating non-MNPs from MNPs are investigated using a magnetophoretic microfluidic platform as presented in Fig. 1 and in the ESI section,† characterized by a laminar flow with a Reynold's number of 0.53. A detailed characterization of the nanoparticles used is given in the ESI section.†
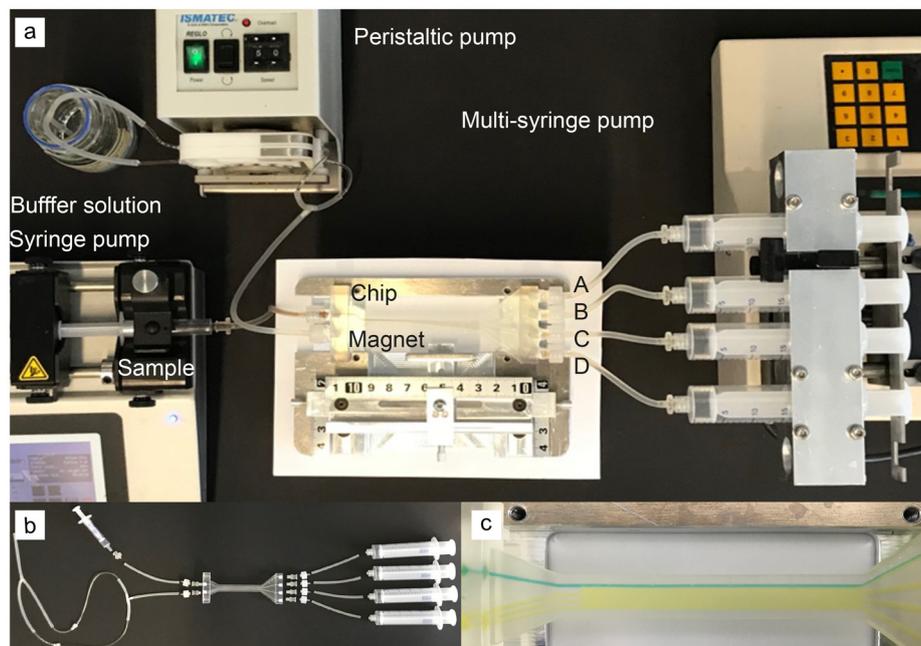

Fig. 2 (a) Experimental set-up consisting of the chip placed in the holder together with the magnet, the syringe pumps for the sample inlet, and the multi-syringe pump for the outlet. The viscous solution is pumped *via* a peristaltic pump. (b) The 3D-printed chip is connected to the tubes *via* printed threads and Luer lock adapters. (c) The flow profile of the chip is shown by food coloring (sample inlet = green, viscous solution inlet = yellow).
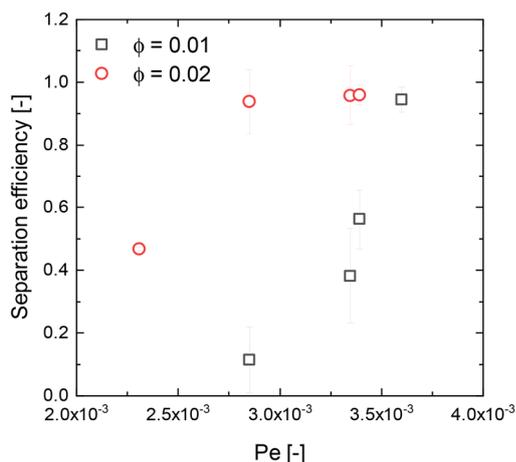
Fig. 3 The separation efficiency of soley MNPs determined with microfluidic experiments is presented as a function of the Péclet number for two different concentrations, namely $\phi = 0.02$ and $\phi = 0.01$.

Fig. 3 depicts the separation efficiency of the MNPs over varying Péclet numbers. For the lower concentration of $\phi = 0.01$, the separation efficiency is $S = 0.11 \pm 0.10$ for Pe = 2.85×10^{-3}. By increasing the Péclet number from Pe = 3.35×10^{-3} to Pe = 3.6×10^{-3}, the separation efficiency increases from $S = 0.38 \pm 0.15$ to $S = 0.94 \pm 0.04$. However, the results show that for a higher concentration, only a lower Péclet number is required to achieve higher separation efficiencies. Here, for $\phi = 0.02$, a separation efficiency of $S = 0.94 \pm 0.10$ is already reached at a Péclet number of Pe = 2.85×10^{-3}. Elevated particle concentrations promote dipole–dipole particle interactions, leading to aggregation and, therefore, an increased

magnetic moment. Consequently, these clusters experience an augmented magnetic force, which drives them towards regions of higher magnetic field intensity.

Fig. 4 compares the separation efficiencies (a and e) and purities (b and f) for varying Péclet numbers achieved by using only one particle species *versus* a mixture of both in the microfluidic chip. Fig. 4(a) refers to the separation efficiency for the non-MNPs, and Fig. 4(e) shows the separation efficiency for the magneto-responsive ones. The separation efficiency for the non-magneto-responsive particles varies little for the applied Péclet numbers but increases from $S = 0.75 \pm 0.13$ to $S = 0.86 \pm 0.05$ by halving the concentration from $\phi = 0.02$ to $\phi = 0.01$ (shaded). However, being within a mixture of both particle species, the separation efficiency decreases from $S = 0.57 \pm 0.13$ to $S = 0.24 \pm 0.01$ as the Péclet number increases. In contrast to the MNPs, it evolves for the non-MNPs that a lower concentration favors a higher separation efficiency at a constant Péclet number. For $\phi = 0.01$ (shaded), the separation efficiency reaches $S = 0.59 \pm 0.1$, compared to $S = 0.24 \pm 0.01$ for $\phi = 0.02$, both at Pe = 3.39×10^{-3}. As the separation efficiency decreases with increasing Péclet numbers for $\phi = 0.02$, the purity of the non-MNPs rises from $P = 0.51 \pm 0.13$ to 0.88 ± 0.1. For the lower concentration of $\phi = 0.01$, it decreases to a purity of $P = 0.52 \pm 0.1$.

From the perspective of the MNPs (Fig. 4e), independent of being present in a particle mixture or not, their separation efficiencies show a similar trend. As the Péclet number rises, signifying an increase in the magnetophoretic force, a notable enhancement in the separation efficiency is observed from $S = 0.44 \pm 0.07$ for Pe = 2.31×10^{-3} to $S = 0.97 \pm 0.02$ for Pe = 3.39×10^{-3} for the particles in the mixture. But again, by decreasing the concentration at a Péclet number of Pe = 3.39×10^{-3}, the
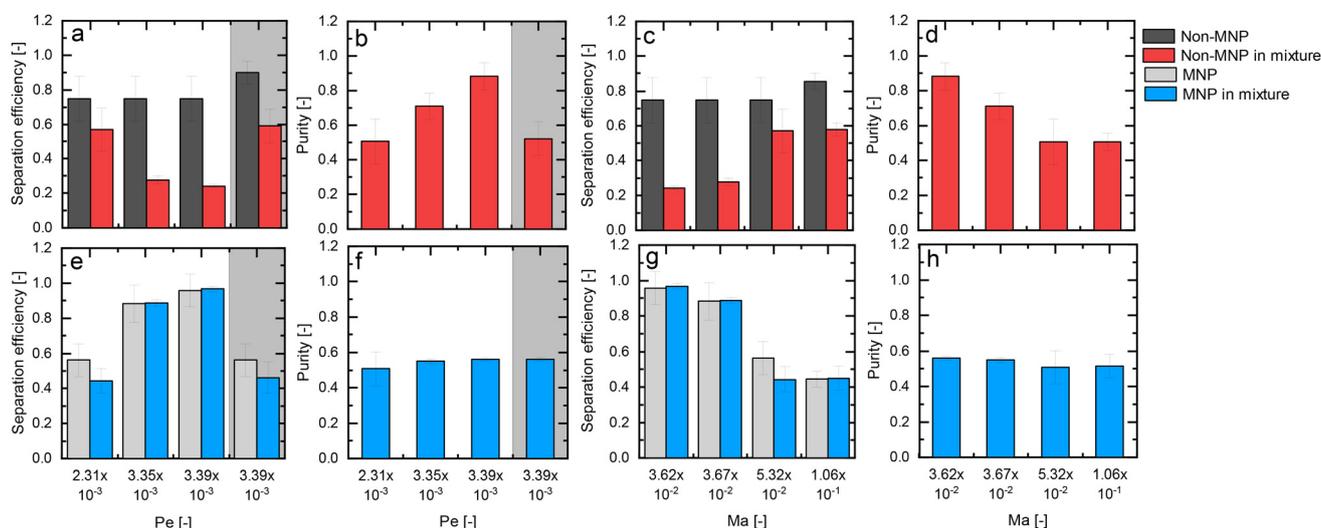

Fig. 4 Figures (a, and b) illustrate the purity and separation efficiency of the non-MNPs obtained with microfluidic separation experiments, whereas figures (e, and f) refer to the MNPs over the Péclet number. The concentration of $\phi = 0.02$ is shown in white, $\phi = 0.01$ in shaded. The mixture always has the same particle concentration as the single experiment for each component. The separation efficiency of solely non-MNPs at $\phi = 0.04$ was $S = 0.66 \pm 0.01$. Figures (c, and d) illustrate the purity and the separation efficiency of the non-MNPs, whereas figures (g, and h) refer to the MNPs over the Mason number for the concentration of $\phi = 0.02$. The mixture always has the same particle concentration as the single experiment for each component.

separation efficiency decreases from $S = 0.97 \pm 0.02$ ($\phi = 0.02$) to $S = 0.46 \pm 0.1$ ($\phi = 0.01$). For separating a mixture with both particle species, the purity (Fig. 4f) is approximately 0.5 for all applied Péclet numbers. This finding aligns with the outcomes observed for non-MNPs, as their separation efficiency diminishes with increasing Péclet numbers. Conversely, higher Péclet numbers exhibit enhanced separation capabilities for the MNPs. This implies that these non-MNPs tend to migrate alongside their magneto-responsive counterparts, consequently compromising the purity of the MNPs. As the magnetic field gradient induces the movement of the MNPs, a hydrodynamic instability appears, generating a convective flow. This flow also instigates a hydrodynamic force on non-MNPs in the direction of the moving MNPs.

4.2 The impact of hydrodynamic forces on magnetic particle interactions

The influence of varying Mason numbers on separation efficiency and purities is depicted in Fig. 4, where it serves as a metric for the ratio of shear to magnetic forces. It evolves that for the non-MNPs in Fig. 4(c), the separation efficiency is around 0.7 for all applied Mason numbers. However, when being present in a mixture with MNPs, it is $S = 0.24 \pm 0.01$ at Ma $= 3.62 \times 10^{-2}$, but increases to $S = 0.58 \pm 0.04$ at Ma $= 1.06 \times 10^{-1}$. At the same time, the purity of the non-MNPs fraction decreases from $P = 0.88 \pm 0.08$ to $P = 0.51 \pm 0.05$.

Since the shear force prevails the magnetophoretic force, the separation efficiency for the magneto-responsive nanoparticles decreases from $S = 0.97 \pm 0.02$ at Ma $= 3.62 \times 10^{-2}$ to $S = 0.45 \pm 0.07$ at Ma $= 1.06 \times 10^{-1}$, depicted in Fig. 4(g) and (h). This decline is observed irrespective of whether the particles are in a mixture with non-magneto-responsive entities or not, the same as for the variation of the Péclet number. Even if the separation efficiency for Ma $= 3.62 \times 10^{-2}$ is high, the purity is $P = 0.56 \pm 0.01$. The observed reduction in separation efficiency for non-MNPs, presented in Fig. 4(c), implies their tendency to be concurrently entrained with MNPs, which is indicative of the resultant low purity in the separation process. Under increased shear conditions, represented by the highest Mason number (Ma $= 1.06 \times 10^{-1}$), it is noteworthy that although the MNPs in the mixture exhibit a lower separation efficiency ($S = 0.45 \pm 0.07$), the non-MNPs align their movement with the MNPs again. Consequently, this leads to a purity level of only $P = 0.52 \pm 0.07$, which is comparable to the purity achieved at the lowest Mason number (Fig. 4(d)).

To further investigate the hydrodynamically induced transport of non-MNPs alongside MNPs, as suggested by the experimental observations, we employ large-scale Brownian dynamic simulations. They are designed to analyze the interplay between the nanoparticles depending on the magnetophoretic forces that contribute to the co-transport phenomenon.

4.3 Hydrodynamically induced transport of undesired species: large-scale Brownian dynamic simulations

Fig. 5(e) shows a snapshot of the simulation box in the absence of shear for a mixture of MNPs (in blue, magneto-responsive nanoparticles) and non-MNPs (in gray, non-magneto-responsive nanoparticles). Such a simple simulation, is similar to the flow present at the geometric symmetric axis along the developing convective flow ($\hat{\mathbf{z}}$). The applied magnetic field in the z direction (outward page) is $\mathbf{H}_0 = 0.4\sqrt{k_B T/a^3 \mu_f}\hat{\mathbf{z}}$, and the gradient $G = 0.02$. The gradient in units of $\sqrt{k_B T/a^5 \mu_f}$.

The motion of MNPs in the presence of a nonuniform magnetic field is known as magnetophoresis. While this topic is a meticulously studied field of study, see ref. 20 and 46 here we are interested on the importance of hydrodynamically induced transport of potentially undesired species (non-magnetic particles) in microfluidic devices. If the components of the applied field gradient $\nabla \mathbf{H}_0$ are small compared to H_0/a, the steady-state structure of the dispersion is only weakly perturbed by the magnetophoretic forces. Under these assumptions, the magnetophoretic force felt by the α_{th} colloid is equal to $\mathbf{F}_\alpha^P = (\nabla \mathbf{H}_0) \cdot \mathbf{m}_\alpha \equiv G(\hat{\mathbf{H}}_0 \cdot \mathbf{m}_\alpha)$, where $G = \nabla H_0$ and $\hat{\mathbf{H}}_0 = \mathbf{H}_0/H_0$. Similar to our magnetic Péclet number estimation, we do not need to consider the field gradient tensor, which has both the field and the gradient directions, but rather the field gradient vector, $\mathbf{G}$. This would not be the case if there is there was any permanent magnetization, for which we would need the complete tensorial expression for the force.

The velocities of the magneto-responsive particles depend on the magnetophoretic forces on the colloids, $\mathcal{F}^P = [\mathbf{F}_1, \mathbf{F}_2, \ldots, \mathbf{F}_N]^T = G(\mathcal{I} \cdot \hat{\mathbf{H}}_0)$, where $\mathcal{I}$ is a list containing the induced dipole moments, and magnetophoretic mobility, $\mathcal{U} = \mathcal{M}^H \cdot \mathcal{F}^P$. The average magnetophoretic velocity will then be given by $\langle u \rangle = \sum_\alpha^{N_M} u_\alpha/N_M$, where u_α is the velocity of the α_{th} colloid.

The first two Fig. 5(a) and (b), show the average colloid velocity of the non-MNPs and MNPs in the direction of the field as a function of the Péclet number. Concentrations vary from $\phi_{mag} = 0.01$ ($\phi_{non-mag} = 0.022$) to $\phi_{mag} = 0.08$ ($\phi_{non-mag} = 0.178$) and field gradient magnitude from $|\nabla H|/H_0 = 0.01$ to $|\nabla H|/H_0 = 0.20$. The field strength increases with the Péclet number and is varied from $O(10^{-6})$ to $O(10^{-2})$. As expected, the velocity of the magneto-responsive particles in the direction of the field (Fig. 5d) increases with the magnetophoretic force (proportional to the Péclet number). Non-MNPs do not show any clear displacement for values of Péclet numbers less than $\mathcal{O}(10^{-4})$, that is, at low-field and/or low-field gradients. The increase in the velocity of the non-MNPs, subpanel (a), with values of Péclet numbers greater than $\mathcal{O}(10^{-4})$ suggests the existence of motion induced by the hydrodynamic interactions between particles in the direction of the field. In consistency with what has been found in magnetophoretic separations,[20] Fig. 5(a) suggests low field gradients are more efficient at reducing the transport of unwanted species (represented by the non-magnetic nanoparticles). In contrast, the orthogonal component of the velocity, as shown in Fig. 5(c) and (d) for the non-MNPs and MNPs, respectively, display no dependence on the applied field or
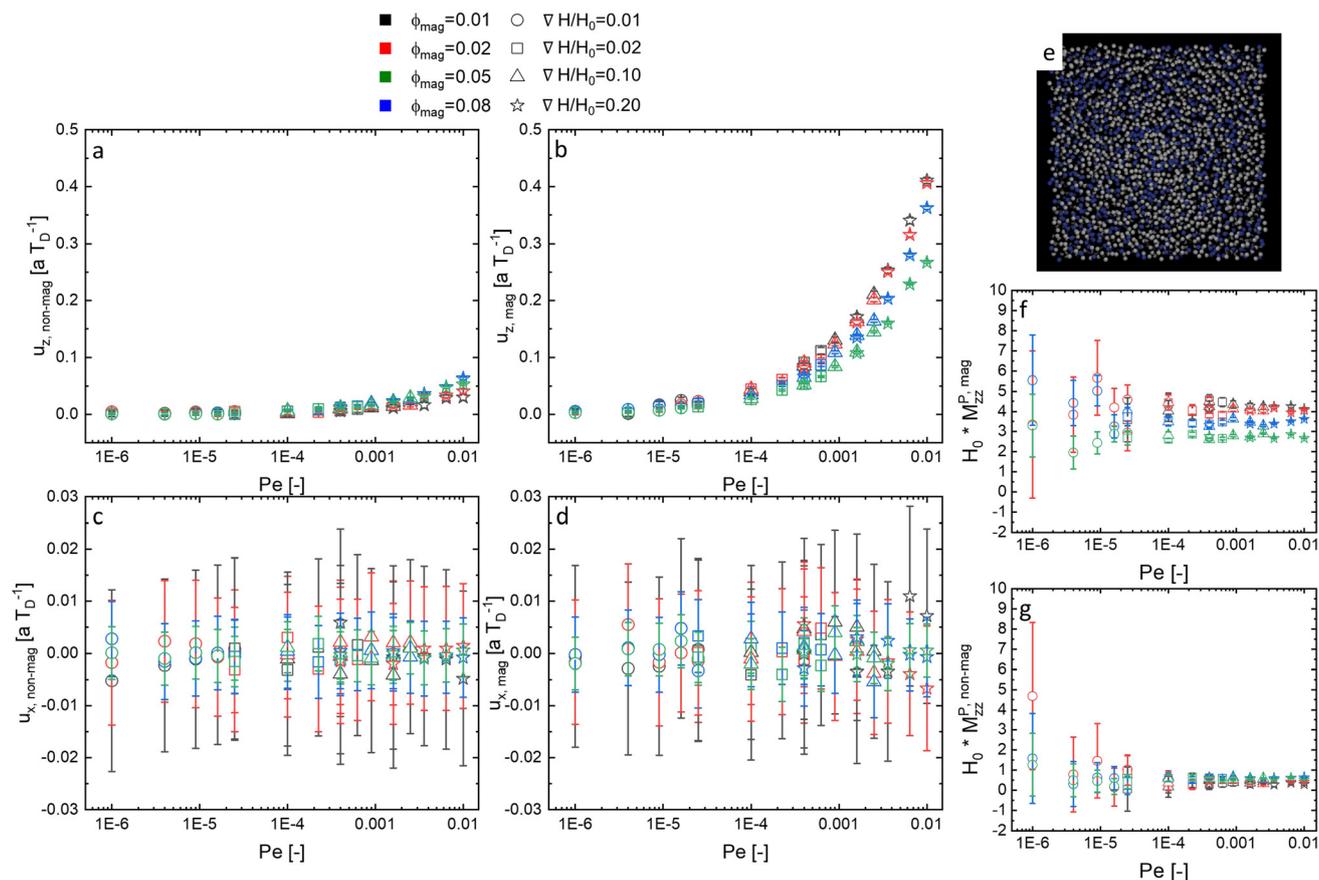
Fig. 5 (a) and (b) Simulated average particle velocity in the direction of the applied field, u_z, and (c) and (d) orthogonal, u_x, to the field as a function of the Péclet number. (a) and (c) Correspond to the average velocities of the non-magnetic particles and (b) and (d) to the magnetic particles, respectively. The concentrations of $\phi_{mag} = 0.01$ ($\phi_{non-mag} = 0.022$), $\phi_{mag} = 0.02$ ($\phi_{non-mag} = 0.044$), $\phi_{mag} = 0.05$ ($\phi_{non-mag} = 0.111$), and $\phi_{mag} = 0.08$ ($\phi_{non-mag} = 0.178$) are colored as dark gray, light gray, blue, and red respectively. The circled, squared, triangled and starred markers correspond to field gradients of $|\nabla \mathbf{H}|/H_0 = 0.01$, $|\nabla \mathbf{H}|/H_0$, $|\nabla \mathbf{H}|/H_0$, and $|\nabla \mathbf{H}|/H_0$, respectively. (e) Snapshot of a slice of the xy plane simulated mixture of MNP and non-MNPs in the absence of shear. Non- and magneto-responsive nanoparticles are shown in gray and blue, respectively. Snapshot corresponds to simulation of filling fraction $\phi_{mag} = 0.05$ ($\phi_{non-mag} = 0.111$), field $H_0 = 0.4k_BT/a_3\mu_f$ in the z direction, with constant gradient in z given by $|\nabla \mathbf{H}|/H_0 = 0.02$. (f) zz component of the mobility matrix, M_{zz}, for the magnetoresponsive particles, computed using eqn (17) (g) zz component of the mobility matrix, M_{zz}, for the non-magnetoresponsive particles, computed using eqn (17) of the MNPs as a function of the Péclet number. Concentrations of $\phi_{mag} = 0.01$ ($\phi_{non-mag} = 0.022$), $\phi_{mag} = 0.02$ ($\phi_{non-mag} = 0.044$), $\phi_{mag} = 0.05$ ($\phi_{non-mag} = 0.111$), and $\phi_{mag} = 0.08$ ($\phi_{non-mag} = 0.178$) are shown in blue, orange, green, and red, respectively. Circled, squared, triangle, and starred markers correspond to field gradients of $|\nabla \mathbf{H}|/H_0 = 0.01$, $|\nabla \mathbf{H}|/H_0 = 0.02$, $|\nabla \mathbf{H}|/H_0 = 0.10$, and $|\nabla \mathbf{H}|/H_0 = 0.20$ respectively.

the field gradients and the average velocities of both species are negligible.

From the average velocity, we can compute the approximate magnetophoretic mobility by introducing a summation tensor, Σ;[47] we find

$$\mathbf{u} = \mathbf{M}^P \cdot \mathbf{G}, \quad \mathbf{M}^P \equiv \Sigma \cdot \mathcal{M}^H \cdot \mathcal{I} \cdot \hat{\mathbf{H}}_0 \quad (17)$$

where magnetophoretic mobility $\mathbf{M}^P$ is a 2-tensor whose elements $M_{\gamma\zeta}{}^P$ couple the gradient in direction ζ to the velocity in direction γ. Considering the field and the field gradient is only nonzero in the z direction, the dominant term is the "self-term" of the magnetophoretic mobility tensor $M_{zz}{}^P$. The results for the zz component of the magnetophoretic mobility tensor for non-MNPs and MNPs are shown in Fig. 5(f) and (g), respectively. The self-term of the mobility tensor for the non-

MNPs, $M_{zz}{}^{P,non-MNP}$, exhibits little dependence on the Péclet number and field gradient. When Pe > 10^{-4}, this component appears to slightly increase in value with the concentration of *magnetic species*, underscoring the cooperative nature of the transport process. In contrast, the self-term of the magnetophoretic mobility tensor of the magneto-responsive, $M_{zz}{}^{P,MNP}$, nanoparticles slightly decreases with concentration.

5 Discussion

The hydrodynamics in magnetophoretic microfluidic separation processes are characterized by a complex interplay of forces that influence the overall process efficiency. Our experimental results show high and constant separation efficiencies

for solely non-magneto-responsive nanoparticles with increasing Péclet number, indicating that their separation is independent of the magnetic force when no magneto-responsive entities are present. But, keeping a constant Péclet number, a higher separation selectivity is achieved for lower concentrations due to the reduction of diffusive effects.[10,48] Conversely, at low concentrations of magneto-responsive nanoparticles, the separation efficiency is reduced. When a magnetic field is applied at low Péclet numbers, the separation efficiency of MNPs initially remains limited due to the predominance of thermal fluctuations over magnetic forces. However, as the Péclet number increases, enhanced dipole–dipole interactions lead to aggregation, amplifying the magnetic moment and enabling more efficient separation by the magnetic field gradient. Also, raising their concentration amplifies their total magnetic moment. These aggregates, larger in size and showing reduced Brownian motion, are more effectively moved by the magnetic field gradient towards areas of higher magnetic field intensity, resulting in improved separation.[22,49]

In a mixture of both particle species, the presence of non-magneto-responsive nanoparticles does not influence the separation efficiency of the magneto-responsive ones with rising magnetic field strength. However, the reverse is not true. The separation efficiency of non-MNPs decreases with increasing magnetic field strength (increasing Péclet number). The process purities of the MNPs are constant even though the separation efficiency of MNPs improves with increasing Péclet number, suggesting that non-MNPs drag along MNPs. That happens as the magneto-responsive nanoparticles induce a hydrodynamic force acting on the non-magneto-responsive entities, influencing the separation process significantly.[23,50] The simulation integrates and extends beyond the experimentally tested parameters of magnetic field strength and concentration, showing that this convective motion is an intrinsic aspect of the system's dynamics. Focusing on the direct interplay of the magneto-responsive entities and the non-magneto-responsive ones, the simulation confirms that the magnetic field gradient primarily induces the motion of the MNPs, as the orthogonal velocity to the field is constant for both particle species. However, the influence of magneto-responsive particles on their non-magneto-responsive counterparts is significant. Although the motion of non-MNPs is an order of magnitude smaller, it is evident that the movement of the magneto-responsive entities leads to a hydrodynamic convective motion that entrains the non-magneto-responsive particles. Since experimental measurements could only be performed offline, after the magnetophoretic process, direct experimental observation of this interaction within the magnetic field is challenging. Simulations are thus essential for capturing and analyzing the underlying dynamics comprehensively. The simulations allowed us to directly investigate the dynamic behavior within the magnetic field, providing valuable insights into parameter regimes that are costly to explore experimentally. Across all simulated particle concentrations and magnetic field gradients the hydrodynamic effect remains consistent. Exceeding the experimentally tested parameters, the average velocity of the non-magneto responsive particle in the direction of the field can be minimized by decreasing the magnetic field gradient. However, this will lead to a reduced responsiveness (velocity) of the magneto-responsive ones as well, resulting in a lower separation efficiency.[22] Experimentally, increasing the shear by increasing the volumetric buffer flow leads to an improved separation efficiency and fewer diffusive effects of the non-MNPs.[10,48] But again, in a mixture where both particle species are present, the purity of MNPs is reduced. Due to the increased shear counteracting the magnetophoretic force, the magneto-responsive nanoparticles are less attracted toward higher magnetic fields.[11]

Magnetic separation techniques are widely used in biochemistry, biomedicine, and biotechnology. Applications range from the purification of biomolecules[51] to targeted drug delivery.[52] These techniques rely on applying a magnetic field to manipulate magneto-responsive particles, most commonly superparamagnetic particles, in a non-magnetic environment. In this study, we evaluate the impact of magnetophoresis on magneto-responsive entities, which are nanoparticles in our case but can be modified for other purposes, for instance, through drug functionalization. This happens in a non-magneto-responsive environment, which is represented by non-magneto-responsive particles in a microfluidic system. The magnetic-induced convective motion is a promising technique for nanoparticle manipulation due to its simple and effective separation mechanism.[49,50,53] However, the induced hydrodynamic force acting on the non-magneto-responsive entities has to be considered when it comes to a lab-on-a-chip separation process or application in biomedicine.[25,54] For targeted processes, e.g., drug delivery, mixing motion should be avoided, allowing effective delivery and localization in the body.[55,56]

Given the complexity of the hydrodynamics in magnetophoretic microfluidic processes, it is crucial to consider strategies that could enhance the separation efficiency and purity of the particles. In the study of Solsona et al., a 3D-printed magnetophoretic chip was developed to sort catalyst particles by their magnetic moment; however, they could not see a difference in the magnetic susceptibility between the different fractions. Besides their discussion that the formation of nanoclusters agglomerates accounts for this observation, the induced convective motion reduced the purity of their fractions.[57] As investigated by Robert et al., the separation efficiency could be improved by an increased flow and a decrease in magneto-responsive entities. Their study successfully differentiated between non-magnetic monocytes and magnetic macrophages; however, they noted that during the separation process, fractions may tend to overlap due to the convective motion engendering mixing and generation of a current.[5] In another approach, the purity of a microfluidic separation process of red (non-magnetic) and white (magnetically labeled) blood cells decreased with a higher flow as the hydrodynamic drag force prevails the magnetophoretic force. Therefore, Lin et al. implemented a two-stage cell separation strategy in their chip, preventing the undesired motion of the red blood cells along with the white ones.[25]

6 Conclusion

In conclusion, our experimental and simulative results highlight that the motion of magneto-responsive nanoparticles induces a hydrodynamic motion that entrains the non-magneto-responsive ones. This interaction significantly influences the separation efficiency and purity of both nanoparticle species in the microfluidic system. The simulations directly investigated these dynamic behaviors within the magnetic field and provided insights that are difficult to obtain experimentally. Besides the careful control of the Péclet and Mason number, a practical approach to minimize the unavoidable magnetic-induced convection of non-magneto-responsive entities may include a multi-step process. Here, a prior static batch separation reduces the concentration of non-magneto-responsive nanoparticles in the system, thereby enhancing the overall separation efficiency and purity.

Author contributions

L.W., E.K., M.B., J.W.S., and S.S. conceptualization, methodology; L.W., J.S., La.W. and E.K. data curation, investigation; L. W. and E.K. formal analysis, visualization; M.B., A.A., S.S. and S.B. writing – review and editing, supervision, project administration; L.W. and E.K. writing – original draft.

Data availability

The data supporting this article have been included as part of the ESI.†

Conflicts of interest

There are no conflicts to declare. All data is available upon request.

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References

1 S. Gupta, K. Ramesh, S. Ahmed and V. Kakkar, *Int. J. Bio-Sci. Bio-Technol.*, 2016, **8**, 311–322.

2 G. Gharib, İ. Bütün, Z. Muganl, G. Kozalak, İ. Naml, S. S. Sarraf, V. E. Ahmadi, E. Toyran, A. J. van Wijnen and A. Koşar, *Biosensors*, 2022, **12**, 1–60.

3 W. H. Chong, S. S. Leong and J. K. Lim, *Electrophoresis*, 2021, **42**, 2303–2328.

4 T. M. Squires and S. R. Quake, *Rev. Mod. Phys.*, 2005, **77**, 977–1026.

5 D. Robert, N. Pamme, H. Conjeaud, F. Gazeau, A. Iles and C. Wilhelm, *Lab Chip*, 2011, **11**, 1902–1910.

6 Y. Zhao, D. Chen, H. Yue, J. B. French, J. Rufo, S. J. Benkovic and T. J. Huang, *Lab Chip*, 2013, **13**, 2183–2198.

7 A. Thiha and F. Ibrahim, *Sensors*, 2015, **15**, 11431–11441.

8 R. Augustine, A. Hasan, S. Das, R. Ahmed, Y. Mori, T. Notomi, B. D. Kevadiya and A. S. Thakor, *Biology*, 2020, **9**, 1–17.

9 E. K. Theel and S. P. Schwaminger, *Int. J. Mol. Sci.*, 2022, **23**, 1–22.

10 R. F. Ismagilov, A. D. Stroock, P. J. Kenis, G. Whitesides and H. A. Stone, *Appl. Phys. Lett.*, 2000, **76**, 2376–2378.

11 Y. W. Tan, S. S. Leong, J. K. Lim, W. M. Yeoh and P. Y. Toh, *Electrophoresis*, 2022, **43**, 2234–2249.

12 E. P. Furlani, *J. Appl. Phys.*, 2006, **99**, 1–35.

13 A. O. Ayansiji, A. V. Dighe, A. A. Linninger and M. R. Singh, *Proc. Natl. Acad. Sci. U. S. A.*, 2020, **117**, 30208–30214.

14 R. K. Singh, T. H. Kim, K. D. Patel, J. C. Knowles and H. W. Kim, *J. Biomed. Mater. Res., Part A*, 2012, **100 A**, 1734–1742.

15 J. P. Steimel, J. L. Aragones and A. Alexander-Katz, *Phys. Rev. Lett.*, 2014, **113**, 178101.

16 S. E. Favela-Camacho, E. J. Samaniego-Benítez, A. Godínez-García, L. M. Avilés-Arellano and J. F. Pérez-Robles, *Colloids Surf., A*, 2019, **574**, 29–35.

17 M. Eigenfeld, L. Wittmann, R. Kerpes, S. P. Schwaminger and T. Becker, *Biotechnol. J.*, 2023, **18**, 1–11.

18 S. A. Majetich, T. Wen and R. A. Booth, *ACS Nano*, 2011, **5**, 6081–6084.

19 D. Saville, W. Russel and R. William, *Colloidal Dispersions*, Cambridge University Press, 1989.

20 T. B. Jones, *Electromechanics of Particles*, Cambridge University Press, 1st edn, 1995.

21 R. G. Gontijo and F. R. Cunha, *Phys. Fluids*, 2017, **29**, 062004.

22 S. S. Leong, Z. Ahmad, S. C. Low, J. Camacho, J. Faraudo and J. K. Lim, *Langmuir*, 2020, **36**, 8033–8055.

23 G. Kitenbergs and A. Cēbers, *J. Magn. Magn. Mater.*, 2020, **498**, 166247.

24 J. W. Swan and G. Wang, *Phys. Fluids*, 2016, **28**, 011902.

25 S. Lin, X. Zhi, D. Chen, F. Xia, Y. Shen, J. Niu, S. Huang, J. Song, J. Miao, D. Cui and X. Ding, *Biosens. Bioelectron.*, 2019, **129**, 175–181.

26 E. S. Krucker-Velasquez, Z. Sherman and J. W. Swan, *Immersed Boundary Method for Dynamic Simulation of Polarizable Colloids of Arbitrary Shape in Explicit Ion Electrolytes*, Ssrn preprint, 2023.

27 D. Frenkel and B. Smit, *Understanding molecular simulation: from algorithms to applications*, Academic Press, 2002.

28 Z. Sherman, PhD Thesis, Massachusets Institute of Technology, 2019.

29 A. M. Fiore and J. W. Swan, *J. Fluid Mech.*, 2019, **878**, 544–597.

30 S. Kim and S. J. Karrila, *Microhydrodynamics: principles and selected applications*, Dover Publications, Mineola, N.Y, 2005.

31 A. M. Fiore, F. B. Usabiaga, A. Donev and J. W. Swan, *J. Chem. Phys.*, 2017, **146**, 124116.

32 A. Gubbiotti, M. Chinappi and C. M. Casciola, *Phys. Rev. E*, 2019, **100**, 053307.

33 S. Delong, F. B. Usabiaga, R. Delgado-Buscalioni, B. E. Griffith and A. Donev, *J. Chem. Phys.*, 2014, **140**, 134110.

34 S. Delong, F. Balboa Usabiaga and A. Donev, *J. Chem. Phys.*, 2015, **143**, 144107.

35 E. E. Keaveny, *J. Comput. Phys.*, 2014, **269**, 61–79.

36 A. M. Fiore and J. W. Swan, *J. Chem. Phys.*, 2018, **148**, 044114.

37 A. W. C. Lau and T. C. Lubensky, *Phys. Rev. E: Stat. Phys., Plasmas, Fluids, Relat. Interdiscip. Top.*, 2007, **76**, 011123.

38 J. N. Israelachvili, *Intermolecular and surface forces*, Academic Press London, San Diego, 2nd edn, 1991.

39 P. Moritz, A. Gonon, T. Blon, N. Ratel-Ramond, F. Mathieu, P. Farger, J. M. Asensio-Revert, S. Cayez, D. Bourrier, D. Saya, L. Nicu, G. Viau, T. Leïchlé and L. M. Lacroix, *ACS Nano*, 2021, **15**, 5096–5108.

40 R. Zwanzig, *Nonequilibrium statistical mechanics*, Oxford University Press, Oxford, New York, 2001.

41 J. W. Swan, R. N. Zia and J. F. Brady, *J. Rheol.*, 2014, **58**, 1–41.

42 G. Bossis, O. Volkova, S. Lacis and A. Meunier, *Magnetorheology: Fluids, Sturcutres and Rheology*, Springer-Verlag Berlin Heidelberg, 2002, vol. 594, pp. 202–233.

43 Z. Varga, V. Grenard, S. Pecorario, N. Taberlet, V. Dolique, S. Manneville, T. Divoux, G. H. McKinley and J. W. Swan, *Proc. Natl. Acad. Sci. U. S. A.*, 2019, **116**, 12193–12198.

44 J. D. Jackson, *Classical electrodynamics*, John Wiley I& Sons, 1999.

45 L. Wittmann, C. Turrina and S. P. Schwaminger, *Magnetochemistry*, 2021, **7**, 80.

46 R. Cobos and A. S. Khair, *J. Fluid Mech.*, 2023, **968**, A14.

47 E. Krucker-Velasquez and J. W. Swan, *J. Chem. Phys.*, 2021, **155**, 134903.

48 P. Agnihotri, *Microfluid. Nanofluid.*, 2023, **27**, 1–15.

49 M. Fratzl, S. Delshadi, T. Devillers, F. Bruckert, O. Cugat, N. M. Dempsey and G. Blaire, *Soft Matter*, 2018, **14**, 2671–2681.

50 J. G. Lee, V. Porter, W. A. Shelton and B. Bharti, *Langmuir*, 2018, **34**, 15416–15424.

51 A. Shiriny and M. Bayareh, *Meccanica*, 2020, **55**, 1903–1916.

52 T. Vangijzegem, D. Stanicki and S. Laurent, *Expert Opin. Drug Delivery*, 2019, **16**, 69–78.

53 S. Schuerle, A. P. Soleimany, T. Yeh, G. M. Anand, M. Häberli, H. E. Fleming, N. Mirkhani, F. Qiu, S. Hauert, X. Wang, B. J. Nelson and S. N. Bhatia, *Sci. Adv.*, 2019, **5**, 1–10.

54 S. S. Leong, S. P. Yeap and J. Lim, *Interface Focus*, 2016, **6**, 1–20.

55 X. Li, W. Li, M. Wang and Z. Liao, *J. Controlled Release*, 2021, **335**, 437–448.

56 P. M. Martins, A. C. Lima, S. Ribeiro, S. Lanceros-Mendez and P. Martins, *ACS Appl. Bio Mater.*, 2021, **4**, 5839–5870.

57 M. Solsona, A. E. Nieuwelink, F. Meirer, L. Abelmann, M. Odijk, W. Olthuis, B. M. Weckhuysen and A. van den Berg, *Angew. Chem., Int. Ed.*, 2018, **57**, 10589–10594.