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Rational experimental design and computational insights into novel heteroleptic mixed-ligand Ru(II) complexes possessing derivatized bipyridines and phendione for anti-cancer activities

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Several synthesized ruthenium(II) complexes, which are used as metallo-drug agents for cancer treatment, have fallen short of meeting expectations. In our continued efforts to search for potential anticancer drug agents for specific biological targets through rational design, this paper reports a facile, one-pot synthesis procedure, along with the photophysical and electro-redox properties, of three newly designed mixed-ligand ruthenium(II) complexes. These complexes contain triphenylphosphine (PPh₃), 4-imidazole acrylic acid (mza), and either functionalized or unfunctionalized polypyridines, specifically 2,2'-bipyridine-4,4'-dicarboxylic acid (Hbpy), 1,10-phenanthroline-5,6-dione (ptd), and 2,2'-bipyridine (bpy). The complexes are formulated as follows: **V2** = [RuCl₂(PPh₃)bpy(mza)·3H₂O], **W** = [RuCl₂(PPh₃)Hbpy(mza)·H₂O] and **X** = [RuCl₂(PPh₃)ptd(mza)·3H₂O]. The complexes were characterized by elemental analysis, Fourier-Transform infrared spectroscopy (FT-IR), ultraviolet-visible spectroscopy (UV-vis), photoluminescence (PL), ¹H, ¹³C, ³¹P nuclear magnetic resonance (NMR) spectroscopy, and mass spectrometry (MS). The UV-vis absorption spectra, showed a broad and intense metal-to-ligand charge transfer (MLCT) band with a strong emission intensity ratio near infra-red. The maximum absorbance wavelengths λ_{max} were observed at 415–530 nm ($\epsilon \approx 1.030 \times 10^3 \text{ M}^{-1} \text{ cm}^{-1}$) and λ_{em} 730–640 nm respectively. The structure–activity relationships relate to a set of voltage-current data, with ligand-based electrochemical redox processes occurring at the Ru(III/II) redox range of +1.15 and –0.82 V. Density functional theory (DFT), molecular docking, and pharmacokinetic calculations confirm that complexes **V2**, **W** and **X** exhibit superior and strong binding interactions and biological inhibition trafficking involving ER α +, WT EGFR, and ALK receptors compared to standard drugs such as Crizotinib, Doxorubicin, Gemcitabine and Lorlatinib. However, the high molecular weights and poor lipophilicity of the complexes hinder their ability to cross the blood–brain barrier (BBB), thus limiting their utility for central nervous system (CNS) targets. This study primarily focuses on the synthesis, characterization, and theoretical evaluation of complexes **V2**, **W** and **X** for their potential anticancer properties, which is useful in the discovery of new materials for drug design. No complementary *in-vitro* wet laboratory tests for anticancer or other biological methods of analysis were conducted, as they were outside the scope of the present study.

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Introduction

Cancer remains one of the deadliest diseases worldwide. It is a complex and multifaceted condition characterized by the uncontrolled growth and spread of abnormal cells. The pathogenesis of cancer can arise from various factors, including

genetic mutations, environmental factors, infections, and genetic predisposition. Carcinomas, sarcomas, leukemias, lymphomas, and colorectal cancers are the most common types worldwide. Ongoing research efforts aim to find a solution for cancer and provide a better understanding of its spread. Current treatments, such as chemotherapy, surgery, immunotherapy, and radiation therapy are used to manage the disease.^{1,2} The refinement of current therapeutic and personalized approaches is ongoing.³

In the treatment of cancer using chemotherapy, platinum-based drugs like cisplatin have been the primary agents for many years. However, due to their significant toxicity and debilitating side effects, ruthenium(II) complexes have shown

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great promise and have gained attention as potential anticancer drugs. They are being promoted as a viable alternative to platinum-based drugs because of their numerous advantages, including the ability to overcome drug resistance, stability in biological media, selectivity, lower toxicity, a wide range of biologically accessible oxidation states, and a distinct, clear mechanism of action.^{4–6} The oxidation states of most metal-based anticancer drugs play a crucial role in their bioactivity.⁷ Although no ruthenium complex has been commercialized for cancer treatment, certain derivatives such as NAMI-A, KP1019/NKP1339, and TLD1433 have advanced into various stages of clinical trials.^{8–10}

Several complexes with Ru(II) have been reported, including those with homoleptic and/or heteroleptic ligands of mono-, bi-, or polydentate architectures.^{11–21} Of great importance is the use of ruthenium(II) polypyridine complexes as molecular targets of DNA base pairs in biomolecules, which are tailored towards different structures and binding modes following sequence-specific recognition.^{22–28} This recognition involves a combination of molecular interactions such as hydrogen bonding, van der Waals interactions, electrostatic and hydrophobic contacts.^{23,29,30} Ruthenium(II) complexes containing different ligand types, including triphenylphosphine, benzoic acid, imidazole, bipyridine, and phenanthroline derivatives, are abundant.

In this study, we prepared three ruthenium(II) heteroleptic complexes, each comprising a different combination of these ligands. This included a functionalized imidazole with an α,β -unsaturated carboxylic acid. The intrinsic properties of these individual ligand types are believed to enhance the anticancer properties of the resulting complexes. It is well-established that ruthenium(II) complexes containing triphenylphosphine often show increased delivery properties due to their high hydrophobicity and strong cytotoxicity.^{31,32}

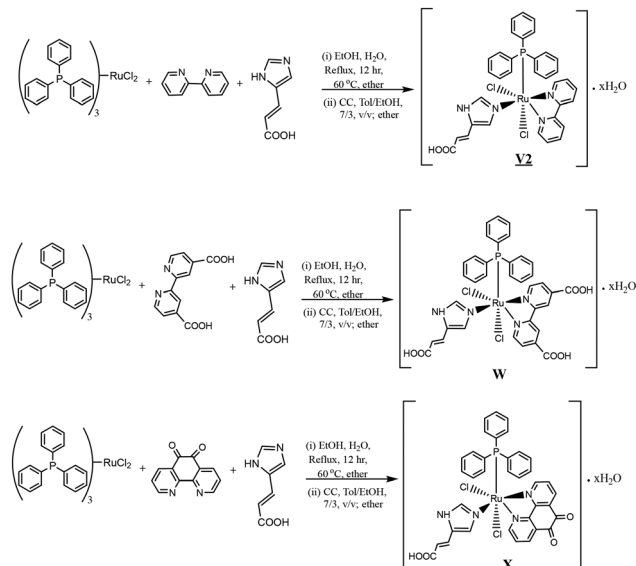
In particular, this work evaluates the photophysical, electro-redox, and pharmacokinetic properties of functionalized/non-functionalized polypyridyl ligands sandwiched by two common organic ligands: triphenylphosphine and functionalized imidazole- α,β -unsaturated carboxylic acid. Their potential anticancer properties are evaluated by density functional theory (DFT) and molecular docking techniques.

Results and discussion

General synthetic procedures for complexes **V2**, **W** and **X**

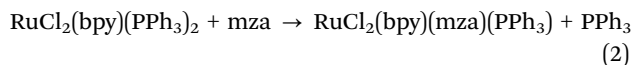
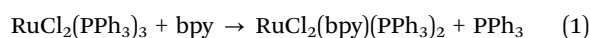
The synthetic reaction scheme for the general preparation of complexes **V2**, **W**, and **X** is shown in Scheme 1. Modifications to the general method reported by Goldman *et al.*,³³ in terms of the solvent used and synthesis procedures were utilized in the reaction conditions due to ligand solubility. The reaction was carried out under reflux using absolute ethanol with the addition of a small volume of water to aid in the total dissolution of the reagents as well as in the coordination of the ligands to the ruthenium(II)dichloro-tris(triphenylphosphine), which served as the metal precursor.

The ligands and metal precursor were added simultaneously in a one-pot reaction synthesis. All reagents and solvents used were of commercial grade with no further purification. All the



Scheme 1 Synthetic reaction procedures for complexes: **V2** = $\text{RuCl}_2(\text{PPh}_3)\text{bpy}(\text{mza}) \cdot 3\text{H}_2\text{O}$; **W** = $\text{RuCl}_2(\text{PPh}_3)\text{Hbpy}(\text{mza}) \cdot \text{H}_2\text{O}$; and **X** = $\text{RuCl}_2(\text{PPh}_3)\text{ptd}(\text{mza}) \cdot 3\text{H}_2\text{O}$.

complex products were obtained in solid form and in good percentage yield. Solubility tests carried out on the complexes showed that none of the complexes was soluble in dichloromethane, but they dissolved appreciably in DMSO and other protic solvents. Literature reports that $\text{RuCl}_2(\text{PPh}_3)_3$ or similar complexes in solution can undergo intramolecular rearrangement or dissociate triphenylphosphine to some extent. This was observed in the reported complexes where two units of the tris(triphenylphosphine) dissociated to obtain the final products, as partially represented in eqn (1) below.



where mza = imidazole-4-acrylic acid.

In fact, to obtain the six-coordinate Ru complex products, the dissociation of two triphenylphosphine units from the pentacoordinate precursor $\text{RuCl}_2(\text{PPh}_3)_3$ during the reaction would have resulted from two effects. First, the chelate effect of the polypyridine brings about increased stability of a complex when a ligand binds with more than one atom. This is followed by the coordination of a nitrogen atom from the imidazole ring with the ruthenium center. Therefore, in the product structures, the bidentate nature of bipyridine and phenanthroline allows them to form a more stable chelate ring with the ruthenium center, favoring the displacement of a single monodentate PPh_3 ligand.^{34–37}

This observation was supported by the characteristic singlet peak of triphenylphosphine at approximately 25.48 ppm in the ^{31}P NMR spectra for complexes **V2** and **W**, although the peak was unresolved in complex **X**. It has been reported that for monodentate N-donor ligands, an upfield shift in $^{31}\text{P}\{^1\text{H}\}$ NMR is observed between 30–40 ppm, whereas complexes with



bidentate N-donor ligands show their ^{31}P $\{^1\text{H}\}$ resonances shifted upfield to approximately 20 ppm.³⁸

All attempts to grow single crystals of complexes **V2**, **W**, and **X** by slow evaporation in different solvent systems were unsuccessful. The solvent combination used were *n*-hexane-toluene (1:4, v/v), *n*-hexane-ethanol (1:4, v/v), and toluene-ethanol (1:1, v/v). The prepared samples were each kept in covered WHEATON liquid scintillation vials (20 mL capacity), but no crystal formation was obtained after a period of 2–3 months.³⁹ It is believed that the hydrated complexes contain different stoichiometric ratios of water, which possibly makes it difficult to achieve the specific, repeating unit required for a well-ordered crystal.⁴⁰ Factors such as steric hindrance of the bulky triphenylphosphine ligands, ligand arrangement disorder, strong or weak ligand interactions, and the thermodynamic properties of the complexes may also play significant roles.⁴¹ The proposed chemical structures of the complexes, as reported, are in very good agreement with the individual LS-MS, proton and carbon-13 NMR spectra data.

FT-IR spectral studies of complexes **V2**, **W** and **X**

The FT-IR spectra for complexes **V2**, **W**, and **X** were recorded in the 4000–400 cm^{-1} region. These spectra were compared with the IR data of the free polypyridine ligands and 4-imidazoleacrylic acid. The IR spectra of the complexes also revealed all expected bands of interest, including the stretching vibrations of $\nu(\text{O-H})$, $\nu(\text{C-H})$, $\nu(\text{C=O})$, $\nu(\text{C=N})$, $\nu(\text{C=C})$, $\nu(\text{C-O})$, $\nu(\text{Ru-Cl})$, and $\nu(\text{Ru-P})$. The broad absorption bands at 3348 and 3406 cm^{-1} are assigned to the O–H stretching vibrations of the carboxylic acid group, as found for complexes **W** and **X**. However, this band was enhanced in complex **V2** at 3417 cm^{-1} . The vibration frequency bands in the 3066–3010 cm^{-1} region are assigned to the asymmetric carboxylate stretching bands, $\nu_{\text{asym}}(\text{COO}^-)$.⁴² The bands at 2981–2754 cm^{-1} are assigned to the C–H aromatic stretching vibrations emanating from the ligands. The C=O stretching frequencies are assigned to bands at 1712 cm^{-1} for complex **V2**, 1658 cm^{-1} for complex **W**, and 1755 cm^{-1} for complex **X**. The aromatic carbon double bonds were assigned to bands at 1612 cm^{-1} for **V2**, 1658 cm^{-1} for **W**, and 1643 cm^{-1} for **X**. Two strong vibration bands were observed at 1589 and 1597 cm^{-1} in complexes **W** and **X**. These were possibly attributed to the influence of increased contributions from carboxylate or carbonyl carbon substitutions at the 4,4'-positions of the bipyridine ring and the 5,6-dione in the phenanthroline.⁴³ The common C=N stretching frequencies are found in the 1566–1539 cm^{-1} region for all three complexes, while the C=C aromatic band frequencies were found at 1481–1411 cm^{-1} . The weak bands at 1369 and 1002 cm^{-1} , common to all complexes, are tentatively assigned to the C–O stretching. In the fingerprint regions of the spectra, the literature has shown three intense characteristic vibrations for the PPh_3 band group at 748, 698, 744, and 786 cm^{-1} for complexes **V2**, **W**, and **X**, respectively.⁴⁴ The Ru–Cl and Ru–P band frequencies for all three complexes are found in the range of 420–698 cm^{-1} .

^1H , ^{13}C and ^{31}P – NMR spectroscopic studies of Complexes **V2**, **W** and **X**

The ^1H NMR spectrum of complexes **V2**, **W**, and **X** in deuterated DMSO exhibits a very complex pattern of resonance chemical

shifts in the region of δ 7.78–6.26 ppm, which is indicative of the presence of mixed ligands comprising triphenylphosphine, 4-imidazoleacrylic acid, and polypyridine protons (see SI Sections S1, S4, and S7).

It's obvious that there are multiple non-equivalent hydrogens, as the complexes are made of two different types of monovalent ligands: 4-imidazoleacrylic acid and triphenylphosphine. For complex **V2**, a total of 27 protons were accounted for in the spectrum at various chemical shifts. The high-intensity multiplet peaks between δ 7.41–7.39 ppm (integrating for 10 protons) and at δ 7.25–7.23 ppm (integrating for 5 protons) are unequivocally assigned to the aromatic protons in the triphenylphosphine ligand. It's obvious that the separation in these hydrogen nuclei may be indicative of the non-stereochemical molecular orientation of the phenyl rings of the coordinating triphenylphosphine moiety. The other peaks are assigned to the protons of the 4-imidazole ring and the unsubstituted bipyridine. For example, the eight protons on the unsubstituted bipyridine are assigned to the multiplet peaks. These peaks include a multiplet integrating for four protons at δ 7.64–7.60 ppm, another three-proton multiplet at δ 7.57–7.53 ppm, and a single remaining proton multiplet at δ 7.22 ppm. The two *meta*-coupled protons of the imidazole ring are found as singlet peaks at δ 7.78 and δ 7.49 ppm, respectively. Also observed in the spectrum is a doublet peak at δ 6.28 ppm, which integrates for one proton with a *J*-coupling constant of 12 Hz. This peak is attributed to the H- β of the α,β -unsaturated carbonyl of the imidazole moiety. The coupled H- α is likely one of the protons with a *J*-coupling constant of 12 Hz at δ 7.46 ppm. In the upfield region, a high-intensity singlet peak was observed at δ 3.36 ppm, which integrates for 71 exchangeable protons. These hydrogens clearly form water molecules within the chemical structure of complex **V2**, and relative to the total number of hydrogens in the aromatic region, they account for at least three molecules of water in the complex.

The ^{13}C NMR spectrum showed the anticipated carbonyl carbon peak at δ 168.42 ppm which was assigned to the carboxylic acid group of the substituted 4-imidazoleacrylic acid. The other carbons in the structure were assigned as follows: δ 137.02 for C-2, δ 131.88 for C-4 (connected to acrylic side chain), δ 111.64 for C-5, δ 129.15 for the α -carbon, and δ 137.11 for the β -carbon. For the triphenylphosphine signals, the carbon directly linked to the phosphorus was assigned to the peak at δ 133.75, while the *ortho*-, *meta*- and *para*- carbons were assigned to the peaks at δ 133.60, 132.47 and 129.43 respectively. The unsubstituted bipyridine showed signals assigned to C-2/C-2' at δ 138.12; C-3/C-3' at δ 129.19; C-4/C-4' at δ 131.95; C-5/C-5' at δ 129.24; and C-6/C-6' at δ 137.95. It is believed that the steric or shielding effects in the complex may have influenced the lowering of the chemical shifts in C-2/C-2' and C-6/C-6', which are ordinarily found in the regions of δ 155–160 ppm. The ^{31}P NMR for complex **V2** was found as a singlet peak at δ 25.46 ppm (see Fig. S1–S3).

In the ^1H NMR of complex **W**, five major, distinct multiplet peaks, integrating to 25 protons, were observed in the aromatic



region. These peaks are from the various mixed-ligand compositions of the complexes. A multiplet between δ 7.64 and 7.59 ppm integrated for six protons, while another at δ 7.56 and 7.52 ppm integrated for five protons. A broad peak at δ 7.39 ppm also integrated for five protons. Three more multiplets were found: one at δ 7.25–7.21 ppm and another at δ 7.11–7.07 ppm, both integrating for four protons each, and a third at δ 6.30–6.27 ppm, which integrated for one proton. A strong, broad peak at δ 3.43 ppm integrated for 25 protons, possibly indicating at least one stoichiometric value of water molecules in the complex, this is relative to the number of protons in the aromatic region.

The ^{13}C spectrum showed peaks in the aromatic region at δ 169.36 and 165.67 ppm. These peaks were assigned to the carbonyl carbons of the 4-imidazoleacrylic acid and 2,2'-bipyridyl-4,4-dicarboxylic acid respectively. The peak at δ 159.21 and 159.17 ppm were assigned to the C-2/C-2' and C-6/C-6' of the 4,4'-disubstituted bipyridine moiety. The peak at δ 131.88 and δ 129.25 were assigned to the C-3/C-3' and C-5/C-5' signals while the C-4/C-4' was assigned to the chemical shift at δ 131.95 ppm. The ^{13}C peaks for the triphenylphosphine have the *ipso*-carbon at 137.11 ppm. The *ortho*-, *meta*-, and *para*-carbons were assigned to δ 133.75, 132.52 and 129.43 respectively. The C-2 carbon of the 4-imidazoleacrylic acid was assigned to the peak at δ 137.02 ppm, the C-4 has its peak at δ 133.60, and C-5 is at δ 129.15. The α -carbon and β -carbon were assigned to the peaks at δ 135.80 and 140.12 ppm respectively. The ^{31}P NMR for complex **W**, also showed a singlet peak at δ 25.48 ppm (see Fig. S4–S6).

Complex multiplet peak patterns were observed in the aromatic region of the ^1H NMR spectrum of **X**. The multiplet peaks were separated into eight major peaks at chemical shifts between δ 7.64–7.53 ppm, with all peaks integrating to a total of 25 protons. The complexity of the ^1H spectrum is likely due to the more mixed ligand substitutions arising from the two dione substitutions at positions 5 and 6 of the 1,10-phenanthroline and 4-imidazoleacrylic acid ligands. Similarly, as was reported for complexes **V2** and **W**, a broad singlet signal peak at δ 3.36 ppm integrating for 77 protons was attributed to the appreciable presence of water molecules in the complex. It could be inferred that the stoichiometry value for water molecules in complex **X**, relative to the total number of protons in the aromatic region is 3.

The ^{13}C NMR spectrum for complex **X** shows the carbonyl carbon peak at δ 170.15, which, due to the symmetry of the 1,10-phenadione ligand, was assigned to the carbon at position 6. The carbonyl peak at δ 166.91 was assigned to the carboxylic acid group of the 4-imidazoleacrylic acid. The other carbon peaks for the 1,10-phenadione include C-1 at δ 154.85, C-2 at δ 126.95, C-3 at δ 138.83, C-4 at δ 129.15, and C-5 at δ 148.78. The peaks for the triphenylphosphine ligand were observed for the *ipso*-carbon at 136.11 ppm. The *ortho*-, *meta*-, and *para*-carbons were assigned to δ 133.53, δ 132.72, and δ 129.25, respectively. The C-2 carbon of the 4-imidazoleacrylic acid was assigned to the peak at δ 131.95 ppm, the C-4 has its peak at δ 132.48, and C-5 is at δ 129.15. The α -carbon and β -carbon were assigned to the peaks at δ 131.88 and δ 132.50 ppm, respectively. The ^{31}P NMR spectrum of complex **X** resonates at δ 25.46 ppm (see Fig. S7–S10).

Electronic absorption studies of complexes **V2**, **W** and **X**

The electronic absorption spectra of all the newly prepared complexes **V2**, **W** and **X** are shown in Fig. 1 and summarized in Table 1. The solid-line spectrum is for **V2** (black), the dashed line is for **W** (red) and the dotted line is for **X** (blue). It has been established that at room temperature, the electronic absorption spectra of typical ruthenium(II) polypyridine complexes are often dominated by ligand-based spin-allowed $\pi \rightarrow \pi^*$ and $n \rightarrow \pi^*$ transitions in the UV region of the spectra and by $^1\text{MLCT}$ absorption bands in the visible region.^{45,46} All the spectra for complexes **V2**, **W** and **X** showed absorption bands at the UV and visible regions. Prominent peak at λ_{max} 228 nm common to all complexes but with different absorbances, is assigned to a spin-allowed ligand-centered $\pi\text{-}\pi^*$ transition, possibly originating from the 4-imidazoleacrylic acid and triphenylphosphine.

The second band was found in the region of 275–283 nm. Literature has shown that this region is assigned to the $\pi\text{-}\pi^*$ transition of phenanthroline and/or bipyridine containing ligands of typical ruthenium complexes.⁴⁷ The small absorption peak at 299 nm which is conspicuously absent in **V2** and **X**, is assumed to originate from the contribution of the bipyridine substituted carboxylic acid moieties present in complex **W**. This band has previously been assigned to intraligand $\pi\text{-}\pi^*$ transition in 2,2' bipyridyl in which the position is not affected upon the introduction of phosphine ligands to the complex inner sphere.⁴⁸ All complexes show a band ascribed to the ligand-to-ligand charge-transfer (LLCT) at λ_{max} between 344–373 nm. The bands at 492 nm, 475 nm and 494 nm were added to the metal-to-ligand charge-transfer (MLCT) for complexes **V2**, **W** and **X** respectively.

The results are supported by previously reported data for heteroleptic complexes containing both mono- and bidentate ligands.^{49–51} The presence of the extra carboxylic acid in

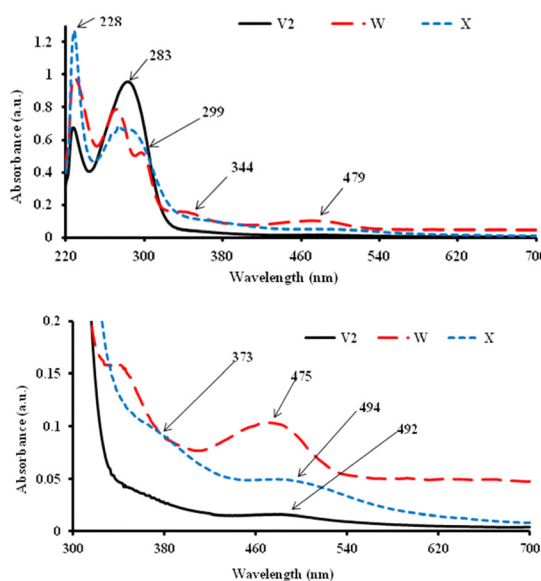


Fig. 1 Electronic absorption UV-vis spectra of Complexes **V2**, **W**, and **X** in (1×10^{-3} M) DMSO.



Table 1 UV-vis, molar extinction coefficient values, emission and CV of complexes **V2**, **W** and **X**

Complex	$\lambda_{\text{max}}/\text{nm}$, (ϵ , $\text{M}^{-1} \text{cm}^{-1}$)	$\lambda_{\text{em}}/\text{nm}$	Quantum Yield (Φ)	Stokes shift (nm)	$E_{1/2}$ (V) (Ru ^{III/II})	E_{cathodic} (V)	E_{anodic} (V)	$E_{1/2}$ (V) Ligand Reductions
V2	492 (1615), 364 (3438), 283 (9563), 227 (6760)	712	0.21	220	+0.86	+0.62	−0.81	−0.15
W	475 (1034), 336 (1603), 299 (5224), 272 (7889), 228 (9822)	643	0.31	168	+1.15	+0.94	−0.98	−0.76
X	494 (4866), 378 (9205), 287 (6599), 272 (6863), 267 (6450), 228 (12 742)	641	0.66	147	+1.08	NA	−0.82	−0.42

complex **W** enhanced the molar extinction coefficient, though with a blue-shift when compared to complexes **V2** and **X**. This has been attributed to efficient π -delocalization as a result of additional electron-donating ability, which leads to a lowering of the π -orbital energy.⁵²

Emission studies of complexes **V2**, **W** and **X**

Emission studies of metal complexes, particularly ruthenium(II) complexes, have provided a powerful tool for drug design, for investigating biochemical processes at the subcellular level, and as functional molecules for the rapid development of fluorescent probes. Complexes **V2**, **W**, and **X** are designed to possibly fit into such applications. As shown in Fig. 2, the emission spectra of complexes **V2**, **W**, and **X** were compared. Upon excitation at the ¹LC and ¹MLCT bands, ($\lambda_{\text{exc.}}$ = 400 nm), all complexes display appreciable luminescence at room temperature. An emission wavelength maximum was found at 713 nm for **V2**, which is blue-shifted to lower intensity at 643 and 641 nm for complexes **W** and **X**, respectively.

It is well known that the presence of triphenylphosphine, a strong-field ligand, often affects the strong metal-to-ligand charge transfer (MLCT) transitions of polypyridines. It also influences the excited state and the energy ordering of its low-energy excited state, in particular, the orbital nature of its lowest excited state. This is likely due to the bulky nature of the ligand, which potentially influences the geometry of the complexes, thus leading to a shift in emission wavelength as well as emission intensity.⁵³

It can be seen that the choice of ligands has a significant effect on the energy positions of the ligand center (LC), the metal center (MC), and the metal-to-ligand charge transfer (MLCT), which depends on the coordination of the donor/acceptor groups. It is obvious, when compared to **V2**, that the excess carboxylic acid group functionality at the 4,4'-positions of 2,2'-bipyridine in complex **W**, as well as the dione groups at the 5,6-positions of 1,10-phenanthroline in complex **X**, in combination with other ligands like 4-imidazole-acrylic acid and triphenylphosphine, significantly affect the energy ordering. This is such that the HOMO–LUMO energy gap and the singlet–triplet splitting energy are affected, thereby changing the overall energy splitting of the LC excited state, which has been found to depend on the intrinsic properties of each ligand.⁴²

The quantum yield was found to depend on the intrinsic properties of each ligand.⁴² The calculated quantum yield of the complexes showed that complex **X** had the highest quantum yield at 0.66, followed by complex **W** at 0.31, and **V2** at 0.21. The quantum yields seemed to increase following a pattern with a ratio of approximately 1:2:3 for complexes **V2**, **W**, and **X**, respectively. Ordinarily, it is well known that the dione functionality at positions 5 and 6 is able to introduce significant electronic perturbations into the ligand compared to unsubstituted 1,10-phenanthroline and/or bipyridine.^{54,55} This agrees with literature on the linear negative correlations between Stokes shift and emission quantum yield, which are a consequence of the reduced degrees of freedom of the intramolecular rotation, possibly due to the presence of the 1,10-phenanthroline-5,6-dione.

Accordingly, the intramolecular rotation restriction gives rise to an inverse relationship between their Stokes shift and quantum yield, whereby smaller Stokes shifts correspond to greater quantum yields, and *vice versa*⁵⁶ (see Table 1). Although, in complex **V2**, the interaction between the metal d-orbital and the ligand π -systems may possibly be responsible for the significant contributions to the excited state, which leads to the observed intense emission and extension of the wavelength to the red region, better than **W** and **X** complexes.

Electrochemical studies of complexes **V2**, **W** and **X**

The voltammograms display the Ru(III)/Ru(II) couple at positive potentials and ligand-based reduction couples at negative potentials. A typical cyclic voltammogram and square-wave voltammogram are shown in Fig. 3 and Table 1 for comparison of the structural compositions of the complexes. It is a clear indication that reduction and oxidation processes are observed in complexes **V2**, **W**, and **X**, which have unfunctionalized or functionalized polypyridines. It appeared that complexes with 2,2'-bipyridyl-4,4'-dicarboxylic acid in coordination with 4-imidazoleacrylic acid, as

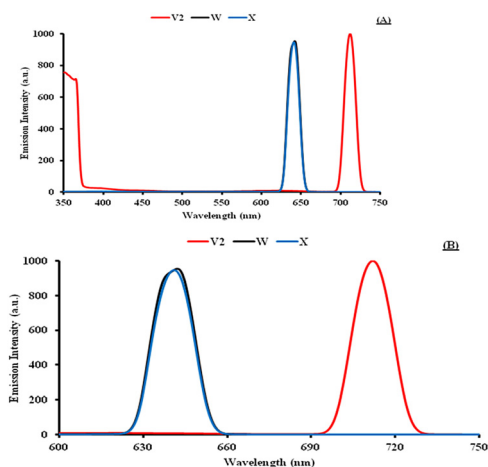


Fig. 2 Excitation spectra at emission maxima for Complexes **V2**, **W**, and **X** (1×10^{-3} M) in DMSO; room temperature. (A) Full spectra, (B) Infra-red region of emission.



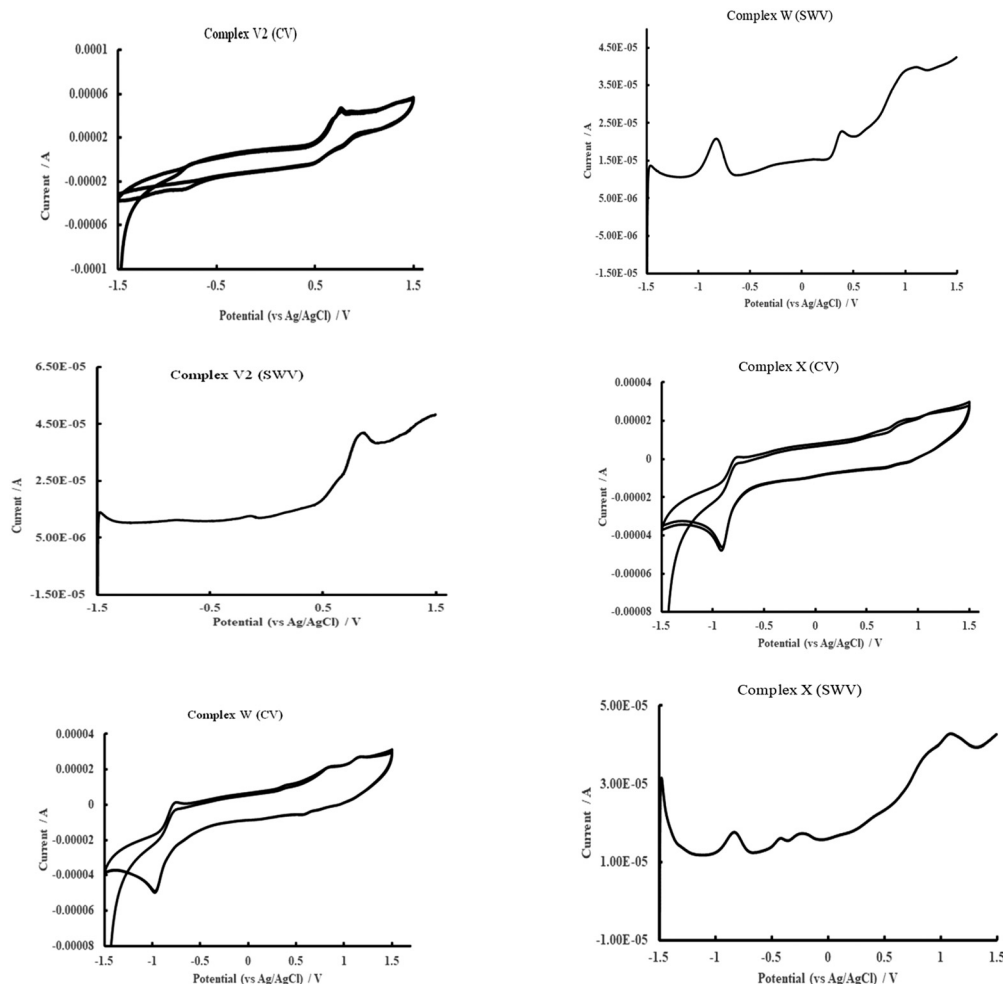
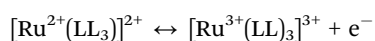


Fig. 3 Cyclic Voltammogram and square wave voltammogram of the same acetonitrile solution (degassed), in 0.2 M $[(n\text{-C}_4\text{H}_9)_4\text{N}]\text{BF}_4$ for Complexes **V2**, **W** and **X**. Scan rate: 50 mVs^{-1} , working electrode: Pt wire, counter electrode: planar Pt electrode, reference: saturated calomel electrode (SCE).

shown in complex **W** (peak oxidation +1.15 V), had a greater influence on the redox processes compared to the corresponding unsubstituted 2,2'-bipyridine and 1,10-phenanthroline-5,6-dione. These latter complexes, **V2** and **X**, had peak oxidation values of +0.86 V and +1.08 V, respectively. This was unequivocally assigned to the reversible one-electron oxidation process of the metal center, specifically the Ru(III)/Ru(II) wave couple.⁴⁶ Often, the metal-centered (parent $\pi\text{M}(\text{t}_2\text{g})$ in octahedral symmetry) orbital of polypyridine is ascribed to the formation of genuine Ru(III) complexes (low-spin 4d^5 configuration) that are inert to ligand substitution, which relates to their oxidation.⁵⁷



Other redox waves include cathodic potentials at +0.62 and +0.94 V for **V2** and **W**, respectively, neither of which is conspicuously shown for **X**. In addition, anodic potentials are found at −0.81, −0.98 V and −0.82 V for **V2**, **W**, and **X**. The half-potential for the ligand reductions was recorded at −0.15, −0.76 and −0.42 V for the complexes. The reduction corresponds to the reduction of the ligands that have the carboxyl acid and

triphenylphosphine groups.⁵⁸ In mixed-ligand complexes like these, electron transitions upon optical absorption would occur between the metal center and the ligand that is most easily reducible. The electron-withdrawing character of the carboxy group would shift the reduction potential of the ligand more positively relative to that of the unsubstituted bipyridine ligand. Thus, it was shown in this work that Ru(III)/Ru(II) redox couples are related to both donor/acceptor properties of Group 5 ligands. The effects of these properties are seen in terms of differential stabilization of Ru(III) over Ru(II) for the donor group properties, while the acceptor properties enhance the further stability of Ru(II) .⁵⁹ In general, cyclic voltammetry spectra of the complexes have shown that several reduction steps due to a strong ligand effect may be found in Ru(II) polypyridine-type complexes, which are most often caused by the presence of mixed-ligands or when the ligands are easily reduced.

Chemical reactivity descriptors and quantum chemical calculations

Density functional theory (DFT) was employed to investigate the electrostatic interactions, electronic properties, and



chemical reactivity of compounds **V2**, **W**, and **X**. Key reactivity parameters are summarized in Table 2, and the optimized molecular orbitals are depicted in Fig. 4. Frontier molecular orbitals, particularly the highest occupied molecular orbital (HOMO) and the lowest unoccupied molecular orbital (LUMO), play a critical role in determining photochemical reactions, pharmaceutical activity, and biological interaction mechanisms. The energy gap (E_g) between HOMO and LUMO is a crucial indicator of a molecule's reactivity and stability. A smaller E_g facilitates electron transfer, enhancing reactivity, which is essential for effective drug-receptor interactions, leading to stabilizing effects and improved binding affinity.⁶⁰ An optimal E_g ensures balanced binding, avoiding weak interactions associated with large gaps or excessive affinity that could result in toxicity due to very small or negative gaps.

For compound **V2**, the HOMO is primarily distributed over the 4-imidazole acrylic acid moiety, extending to the central ruthenium metal and one chlorine atom, with an energy of -5.8401 eV. The LUMO, on the other hand, is localized across the chlorine atoms and the central metal atom, with an energy of -3.3026 eV, resulting in an energy gap (ΔE) = 2.5375 eV (Fig. 4). This distribution indicates significant intramolecular charge transfer within the molecule. Similarly, for compound **W**, the HOMO is distributed over the 4-imidazole acrylic acid moiety, extending to the central ruthenium metal and both chlorine atoms, with an energy of -6.3748 eV. The LUMO is concentrated on the central metal atom bonded to both chlorine atoms, with slight spill-over onto one nitrogen atom of the 2,2'-bipyridine-4,4'-dicarboxylic acid, yielding an energy of -3.3078 eV and an energy gap (ΔE) = 3.067 eV (Fig. 4). In contrast, compound **X** exhibits a distinct electronic distribution. The HOMO is localized over the 4-imidazole acrylic acid moiety, with minor spillover onto the central ruthenium metal, and has energy of -5.7786 eV. The LUMO, however, is primarily localized on the 1,10-phenanthroline-5,6-dione moiety, with an energy of -4.5329 eV, resulting in a smaller energy gap (ΔE) of 1.2457 eV (Fig. 4). This smaller gap indicates higher reactivity and clearly defined electronegative and electron affinity sites, further confirming intramolecular charge transfer.

Additional reactivity descriptors, including chemical hardness (η) and softness (σ), are presented in Table 2. These parameters measure a molecule's resistance and susceptibility to charge transfer, respectively, and further elucidate its reactivity.⁶¹ **V2** (η = 1.2688 eV, σ = 0.7881 eV), **W** (η = 1.5335 eV, σ = 0.6521 eV), and **X** (η = 0.6229 eV, σ = 1.6054 eV) exhibit high softness, suggesting favorable interactions with biomolecules. The global electrophilicity index (ω), which quantifies a compound's tendency to attract or donate electrons, further supports their reactivity. High electrophilicity index values indicate that these

molecules are likely to act as electrophiles in biological reactions,⁶² while lower values of electrophilicity index and chemical potential signify strong nucleophilic reactivity.⁶³ Notably, a strong electrophile is characterized by $\omega > 1.5$ eV.⁶⁴ The electrophilicity index values for **V2** (ω = 8.2352 eV), **W** (ω = 7.6421 eV), and **X** (ω = 21.3375 eV) are significantly high, underscoring their potential as promising drug candidates.

To investigate the coordination environment around the Ru(II) centre, such as ligand atoms (Cl, N, and P), and assess distortions from ideal octahedral geometry, we evaluated the angular distortion parameter Σ for three complexes (**V2**, **W**, and **X**) based on DFT-optimized geometries obtained using the B3LYP functional and LANL2DZ basis set in Gaussian 16. No symmetry constraints were imposed during optimization. In addition to electronic structural analysis, bond lengths (Å) and bond angles (°) around the central metal atom are provided in the Table 3.

The Σ (Sigma) parameter, defined as the sum of absolute deviations of the 12 *cis* angles from 90° , provides a rapid and intuitive measure of angular distortion in pseudo-octahedral systems.⁶⁵ This parameter is determined by the equation:

$$\Sigma = \sum_{i=1}^{12} |\theta_i - 90^\circ|$$

where θ_i represents each *cis* bond angle around the metal centre. For an ideal octahedron, $\Sigma = 0^\circ$, and increasing values indicate greater angular distortion.^{66,67} The calculated Σ values in Table 3 reveals a progressive increase in angular distortion across the series, **V2** ($\Sigma \approx 24.73^\circ$), **W** ($\Sigma \approx 30.72^\circ$), and **X** ($\Sigma \approx 47.59^\circ$). This trend suggests increasing ligand-induced strain or asymmetry, likely arising from variations in steric bulk, electronic effects, or chelation geometry.⁶⁸ Complex **X**, with the highest Σ value, exhibits the most pronounced deviation from ideal geometry. Notably, the N6–Ru–N7 angle of 77.6° contributes significantly to this distortion, deviating markedly from the expected 90° .

These findings are consistent with reported angular distortions in low-spin Ru(II) complexes bearing asymmetric or hydrogen-bonding ligands, where Σ values typically range from 20° to 50° , depending on ligand field strength and bite angles. Although high-spin states generally exhibit larger distortions,^{66,67} the present calculations were performed at the ground-state configuration, which for Ru(II) is typically low-spin, resulting in moderate but chemically reasonable deviations. Overall, the Σ parameter effectively captures angular strain and supports the structural integrity of the DFT-optimized geometries. These quantitative insights into ligand-induced distortion provide a valuable framework for comparing coordination environments and may inform the rational

Table 2 Calculated HOMO and LUMO energies along with their corresponding quantum chemical parameters

	EH (eV)	EL (eV)	E_g (eV)	IP (eV)	EA (eV)	χ (eV)	μ (eV)	η (eV)	σ (eV)	ω (eV)
V2	-5.8401	-3.3026	2.5375	5.8401	3.3026	4.5714	-4.5714	1.2688	0.7881	8.2352
W	-6.3748	-3.3078	3.067	6.3748	3.3078	4.8413	-4.8413	1.5335	0.6521	7.6421
X	-5.7786	-4.5329	1.2457	5.7786	4.5329	5.1558	-5.1558	0.6229	1.6054	21.3375



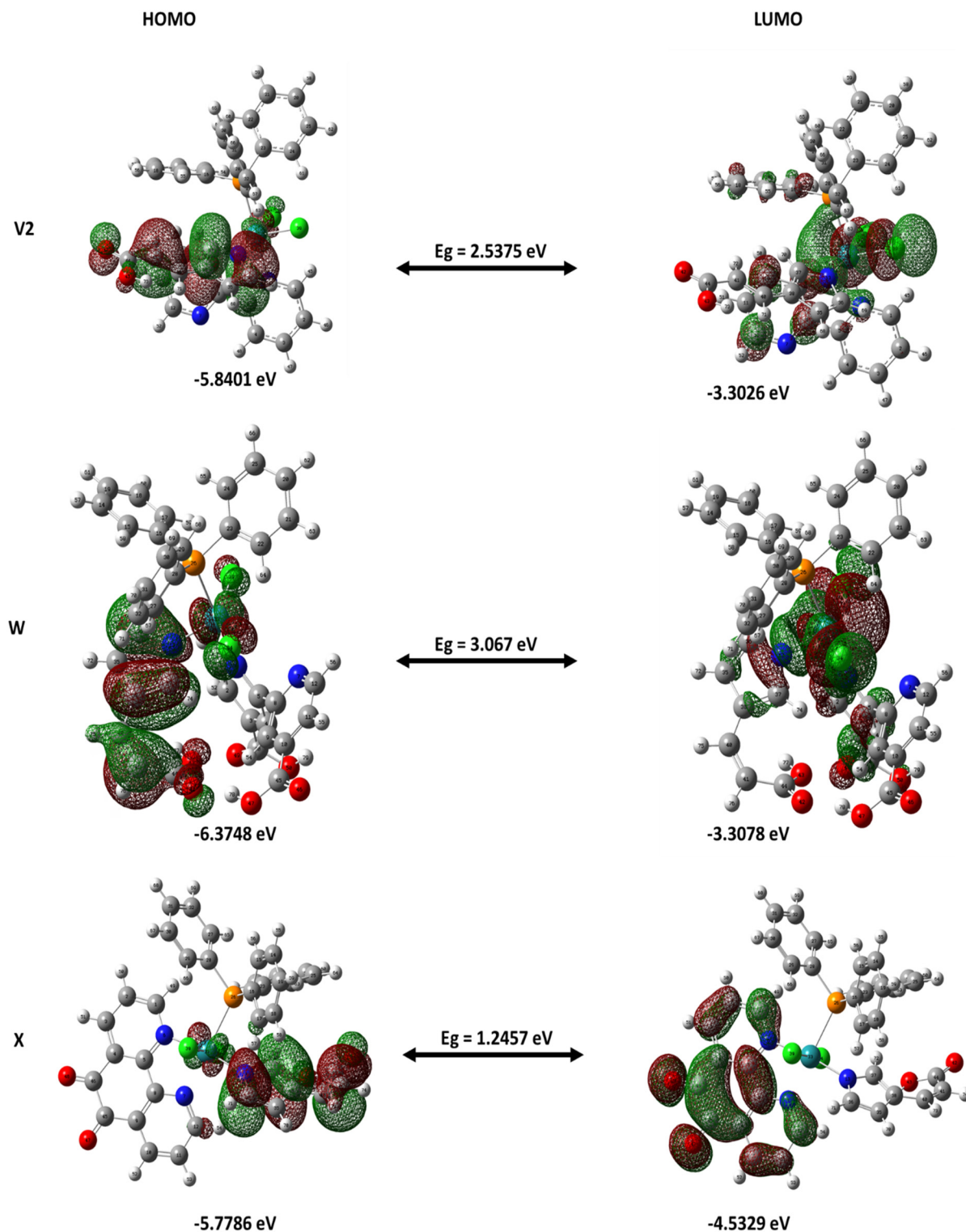


Fig. 4 Frontier molecular orbital plots of **V2**, **W**, and **X** predicted by LANL2DZ basis set with DFT/B3LYP functional.

design of Ru-based catalysts, photochemical agents, or functional materials.

Molecular electrostatic potential (MEP)

The molecular electrostatic potential (MEP) surface analysis is a powerful tool for identifying reactive sites within a molecule by

mapping the distribution of nuclear and electronic charges. This analysis highlights regions prone to nucleophilic and electrophilic attacks, providing insights into molecular reactivity. Using the B3LYP/LANL2DZ method, the MEP surfaces of compounds **V2**, **W**, and **X**, were examined. The electrostatic potentials are visualized using a red-green-blue colour gradient: blue regions



Table 3 Selected bond lengths (Å) and angles (°) for complexes **V2**, **W** and **X** from DFT calculations at B3LYP/LANL2DZ level of theory

Property	Bond length (Å)/Bond angle (°) V2	Bond length (Å)/Bond angle (°) W	Bond length (Å)/Bond angle (°) X
Ru–Cl39 (Cl1, atom 2)	2.401	2.427	2.457
Ru–Cl34 (Cl2, atom 4)	2.457	2.413	2.448
Ru–N (N13, atom 13)	2.008	1.973	2.035
Ru–N (N6, atom 6)	2.145	2.247	2.160
Ru–N (N7, atom 7)	4.390	3.813	2.101
Ru–P	2.496	2.474	2.250
Cl–Ru–Cl	98.055	151.197	170.508
N6–Ru–N	89.267 (N13), 54.355 (N7)	85.122 (N13), 55.549 (N7)	170.956 (N13), 77.626 (N7)
N7–Ru–N13	60.672	121.431	93.538
Cl–Ru–P	91.056 (Cl1), 89.209 (Cl2)	89.757 (Cl1), 87.240 (Cl2)	88.475 (Cl1), 95.000 (Cl2)
Cl–Ru–N6	92.292 (Cl1), 85.627 (Cl2)	84.997 (Cl1), 98.718 (Cl2)	85.648 (Cl1), 84.985 (Cl2)
Cl–Ru–N13	92.074 (Cl1), 168.819 (Cl2)	114.158 (Cl1), 94.639 (Cl2)	96.106 (Cl1), 92.914 (Cl2)
Cl–Ru–N7	133.913 (Cl1), 108.456 (Cl2)	104.417 (Cl1), 57.365 (Cl2)	88.196 (Cl1), 88.301 (Cl2)
P–Ru–N	174.195 (N6), 125.354 (N7), 95.352 (N13)	174.042 (N6), 128.896 (N7), 94.482 (N13)	102.194 (N6), 176.671 (N7), 86.741 (N13)
Σ (Cis-angle deviation)	24.73°	30.72°	47.59°

indicate electrophilic sites (positive electrostatic potential), red regions correspond to nucleophilic sites (negative electrostatic potential), and green areas represent neutral zones (Fig. 5).

For compound **V2**, the negative electrostatic potential regions are concentrated around the oxygen atom of the carboxylic group in the 4-imidazole acrylic acid moiety. Additionally, a yellow mapping on the chlorine atoms suggests a negative electrostatic potential, which may facilitate π - π stacking or hydrogen bond interactions with aromatic residues in receptor binding sites. This is supported by the molecular interactions observed between **V2** and the ALK receptor (Fig. 6). The remaining surface of **V2** exhibits a light blue, positive potential and a neutral electrostatic region. In the case of molecule **W**, nucleophilic centers (negative electrostatic potentials) are primarily located over one of the carboxylic groups in the 2,2'-bipyridine-4,4'-dicarboxylic acid moiety. The electron-rich regions, indicative of nucleophilic sites, are found near the carboxylic group of the 2,2'-bipyridine-4,4'-dicarboxylic acid moiety, adjacent to the carboxylic group of the 4-imidazole acrylic acid moiety. An intermediate potential, a less reactive region marked by yellow, is observed around the chlorine atom closest to the nitrogen atom on the imidazole attachment.

Compound **X**, however, displays a more scattered distribution of nucleophilic regions. These are predominantly localized on the carboxylic group of the 4-imidazole acrylic acid moiety, the chlorine atoms, and the carbonyl group of the 1,10-phenanthroline-5,6-dione moiety. The electrophilic sites, represented by light blue regions, correspond to electron-deficient areas. These patterns of electrostatic potential distribution reflect electron-rich π -interactions and suggest that the biological activity of these molecules is significantly influenced by variations in their atomic composition and electronic properties. Overall, the MEP analysis provides critical insights into the reactivity and potential binding interactions of **V2**, **W**, and **X**, underscoring the importance of electrostatic potential variations in determining their biological activity.

Molecular docking study

This study investigates the inhibition of biological trafficking involving ER α +, WT EGFR, and ALK receptors, focusing on the distinct binding interactions of compounds **V2**, **W**, and **X**. As shown in Table 4, compound **W** demonstrated the highest

binding affinity to ER α +, with a MolDock score of -202.391 . Its interaction with the ligand-binding domain of ER α is characterized by three conventional hydrogen bonds, one pi-donor hydrogen bond, one carbon-hydrogen bond, two Pi-Sigma bonds, one Pi-Pi T-shaped bond (with PHE 404), seven Pi-alkyl bonds, and one unfavorable bump with LEU 346. Notably, compound **W** forms a bifurcated hydrogen bond between ASP351 and PRO535, facilitating antagonistic docking of helix 12 (H12) in the activating function-2 (AF-2) cleft. This interaction is mediated by the carboxylic acid group on the 4-imidazole acrylic acid moiety, highlighting its role in stabilizing the binding.

Compounds **V2** and **X** also exhibited strong binding affinities, outperforming standard reference molecules. However, **V2** displayed the highest number of unfavorable bumps (5) and formed only two hydrogen bonds with active site residues. Unlike **W** and **X**, **V2** lacked interactions with the catalytic residues ASP351 and VAL533, which are critical for the activity of selective estrogen receptor modulators (SERMs) and degraders (SERDs). The presence of hydrogen bonding with ASP351, a key feature of SERMs and SERDs, was observed in both **W** and **X** but not in **V2**. This interaction enhances the potential of **W** and **X** as anti-transcriptional agents.

Fig. S17 and S18 (see SI Section) illustrate the 2D and 3D molecular interactions and the distances of these interactions within the ER α active site. The figures also provide a detailed representation of the 2D interactions of the standard reference molecules within the same active site. These findings underscore the distinct binding mechanisms of **V2**, **W**, and **X**, highlighting their potential as inhibitors of biological trafficking and their varying efficacies in targeting ER α receptors.

For wild-type EGFR (PDB ID: 8PO4), a transmembrane receptor tyrosine kinase that regulates cell proliferation, survival, differentiation, and migration, compounds **V2** (-156.575), **W** (-146.189), and **X** (-121.169) exhibited superior binding affinities compared to standard drugs. Although Crizotinib displayed a moderate binding energy (-121.169), it formed the highest number of hydrogen bonds (7), including a critical interaction with CYS 797. Compound **V2** achieved the lowest binding energy, likely due to a pi-alkyl interaction with CYS 797 via its triphenylphosphine warhead. Notably, only Crizotinib



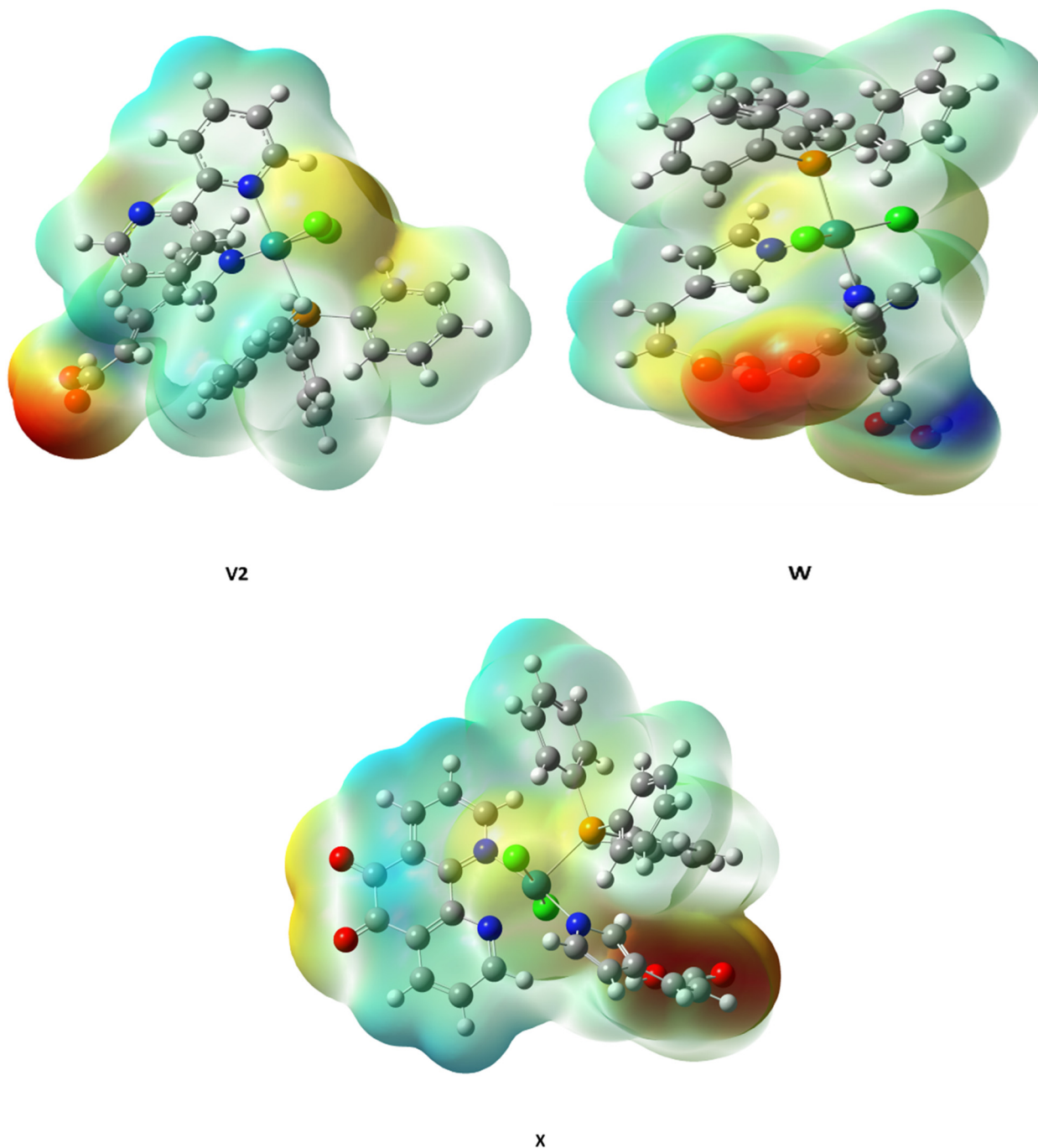


Fig. 5 Three-dimensional plot of molecular electrostatic potential (MEP) for **V2**, **W**, and **X** molecules.

and compound **V2** interacted with key binding site residues (MET 766, MET 793, and PHE 856). The binding mode was illustrated in Fig. S19 and S20.

For ALK, a receptor tyrosine kinase often activated by gene fusion, mutations, or deletions in cancers such as NSCLC, neuroblastomas, gliomas, and inflammatory myofibroblastic tumors, compounds **V2** (−170.436), **W** (−182.219), and **X** (−184.563) outperformed all standard drugs. Crizotinib, a first-generation ALK tyrosine kinase inhibitor (TKI), shows limited efficacy against acquired resistance mutations. Lorlatinib, a third-generation ALK

TKI, is brain-penetrant and effective against many resistance mutations but is limited by the emergence of compound ALK mutations, particularly those involving ARG 1202, which confer resistance to all approved ALK TKIs.³⁶

In this study, Lorlatinib and Crizotinib interacted with ARG 1202 *via* alkyl and pi-alkyl bonds, with binding scores of −148.465 and −124.025, respectively. Compound **V2** formed a pi-alkyl interaction with ARG 1202 through its bipyridine moiety, while compound **X** demonstrated the highest binding affinity (−184.563) *via* hydrogen bonding and pi-alkyl



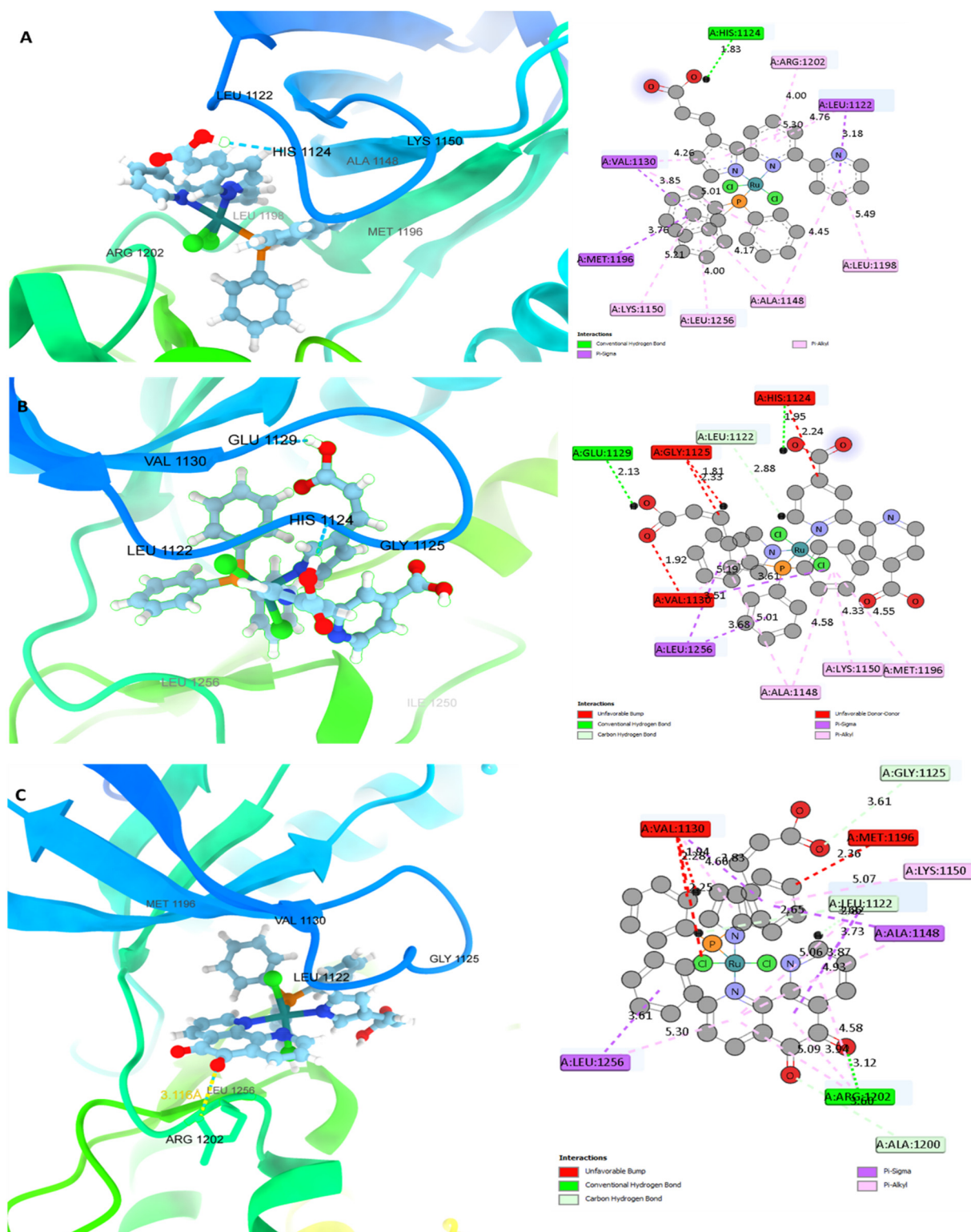


Fig. 6 3D and 2D representation of the molecular interaction between ALK (9GBE) and (A) **V2**; (B) **W**; (C) **X**.

interactions from its 1,10-phenanthroline-5,6-dione moiety. These interactions suggest that **X** is a potent inhibitor of ALK on coproteins, with minimal off-target effects on TRK in the nervous system. Most studied compounds interacted with key active site residues (LYS 1150, MET 1196, GLU 1197, LEU 1198,

MET 1199, and ARG 1202). Detailed molecular interactions and distances are shown in Fig. 6 and S21. ER α (Breast cancer receptor), WT-EGFR, and ALK are all protein targets frequently dysregulated in cancer. WT-EGFR regulates cell growth and survival and is often overexpressed in cancers like non-small



Table 4 Molecular docking results of the compounds and their corresponding inhibition constants

Ligand	ER α + (8DU6) MolDock [GRID] score	WT EGFR (8PO4) MolDock [GRID] score	ALK (9GBE) MolDock [GRID] score
V2	−157.004	−136.475	−170.436
W	−202.391	−156.575	−182.219
X	−155.301	−146.189	−184.563
Crizotinib	−138.783	−121.169	−124.025
Doxorubicin	−121.179	−94.0293	−121.057
Gemcitabine	−84.3502	−95.4603	−83.4191
Lorlatinib	−115.028	−114.656	−148.465

cell lung cancer (NSCLC). ALK drives tumorigenesis through gene rearrangements and is a common target in NSCLC and lymphoma.

Physicochemical and drug likeness properties

Optimizing lead compounds for drug development requires careful evaluation of physicochemical properties, lipophilicity, pharmacokinetics, and drug-likeness to ensure therapeutic efficacy. Here, we assess these parameters for V2, W, and X using SwissADME.⁶⁹ Classical drug-likeness rules (Lipinski, Veber, Egan, Muegge) were examined with a focus on Lipinski, Verber, Lead-likeness, and pan-assay interference (PAINS) alerts. These rules are based on the values of some molecular properties, for example, according to Lipinski rule, poor absorption and permeation are more likely when the molecular weight (MW) > 500 g mol^{−1}, lipophilicity (log *P*) and hydrogen-bond donors (HBD) are more than five, and hydrogen-bond acceptors (HBA) are greater than ten. All compounds exhibited one violation (molecular weight > 500 g mol^{−1}) of Lipinski's rule but otherwise complied (Table 5). However, compound W failed Verber's rule due to excessive rotatable bonds (RB > 10) or topological polar surface area (TPSA > 140 Å²). In drug discovery, "PAINS" (pan-assay interference compounds) are compounds that cause false-positive results in high-throughput screens, leading to the misidentification of potential drug candidates. PAINS alerts, which flag these compounds, are a common screening tool used to weed out such undesirable compounds in the early stages of drug discovery. None of the compounds triggered PAINS alerts, supporting their suitability as drug candidates.

The consensus lipophilicity partition *P*_{o/w} ranged from 3.99 (V2) – 4.05 (X), indicating moderate to high lipophilicity (Table 6). All compounds were poorly soluble (log *S* < −9.75), with X showing the lowest solubility (log *S* = −11.85). This poor solubility correlates with low gastrointestinal (GI) absorption,

necessitating alternative administration routes (e.g., intravenous) for systemic delivery. This is because low GI absorption means that a small fraction of the compound taken orally will enter the bloodstream, which is generally unfavorable for systemic cancer treatment, as it limits the sufficient concentration of the drug that reaches the tumor through oral administration, higher doses might be needed, potentially leading to increased side effects. The pharmacokinetic profiling is shown in Table 7.

None of the compounds cross the blood–brain barrier (BBB), which limits their utility for CNS targets (such as brain tumors) but also reduces off-target neurotoxicity. Most compounds showed minimal cytochrome P450 (CYP) enzymes inhibition, which means they have a lower risk of having drug–drug interactions that can alter the metabolism of other drugs metabolized by the same drug. Additionally, low log *K*_p (Skin permeation) values (−4.91 to −6.12 cm s^{−1}) rule out transdermal delivery. In conclusion, while these ruthenium complexes exhibit promising drug-like properties and lack PAINS alerts, their high molecular weight, poor solubility, and low oral bioavailability pose challenges for conventional administration. Future work should explore formulation strategies (e.g., nanoparticulate delivery) or structural modifications to improve pharmacokinetics.

Experimental

General procedure for synthesis of complexes (V2, W and X)

(V2): **synthesis of [RuCl₂(PPh₃)₃bpy(mza)]·3H₂O**. The complex was synthesized in one pot using a single-step reaction (Scheme 1) with slight modifications as described in the literature.^{70,71} The reaction was carried out in a 100 mL round-bottom flask. The flask was charged with 20 mL of absolute ethanol, and commercial dichloro-ruthenium(II)tris-(triphenylphosphine) [RuCl₂ (PPh₃)₃] (0.35 g, 0.37 mmol), 4-imidazole-acrylic acid (0.07 g, 0.37 mmol), and bipyridine (0.06 g, 0.37 mmol) were added successively. An additional 5–10 mL of water was later added to aid in the complete dissolution of the reagents. The mixture was then heated to reflux under vigorous magnetic stirring for 12 hours while monitoring the reaction with a TLC procedure. After reflux, the reaction product was allowed to cool down and was filtered. The unreacted solid was separated under gravity by the addition of a small quantity of water (20–50 mL) for washing. The filtrate was concentrated *in vacuo* to obtain a dark brown syrupy solid. The semi-crude product was purified by column chromatography using toluene-ethanol (7/3, v/v, 250 mL) to obtain a light brown solid, which was further purified by recrystallization in 30–40 mL of diethyl ether. Yield = 53%, Melting point (mp) = 212–213 °C. FT-IR

Table 5 Physicochemical properties of the studied Ruthenium complexes (V2, W and X)

Compounds	Molecular formula	M. W (g mol ^{−1})	HA	AHA	RB	HBA	HBD	MR	TPSA (Å ²)
V2	C ₃₄ H ₂₉ Cl ₂ N ₄ O ₂ PRu·3H ₂ O	805.67	44	35	7	3	1	178.62	78.57
W	C ₃₆ H ₂₉ Cl ₂ N ₄ O ₆ PRu·H ₂ O	857.20	50	35	9	7	3	192.54	153.17
X	C ₃₆ H ₂₇ Cl ₂ N ₄ O ₄ PRu·3H ₂ O	857.21	48	37	7	5	1	191.92	112.71

Molecular weight (M. W), num. heavy atoms (HA), number of aromatic heavy atoms (AHA), number of rotatable bonds (RB), number of hydrogen bond acceptors (HBA), number of hydrogen bond donors (HBD), molar refractivity (MR), and topological polar surface area (TPSA).



Table 6 Lipophilicity, solubility, and drug-likeness profiles of the studied Ruthenium complexes (V2, W and X)

Compounds	Consensus log P_{ow}	Water Solubility			Drug-likeness		
		Log S (ESOL)	Log S (Ali)	Log S (SILICOS-IT)	Lipinski	Verber	Lead likeness
V2	3.99	−9.65	−9.75	−11.29	Yes; 1 violation	Yes	No; 2 violations
W	3.32	−9.39	−10.30	−9.90	Yes; 1 violation	No; 1 violation	No; 3 violations
X	4.05	−9.44	−9.58	−11.85	Yes; 1 violation	Yes	No; 2 violations

Solubility class (log S scale): Insoluble < −10 < Poorly < −6 < moderately < −4 < soluble < −2 < very < 0 < Highly. All compounds exhibited 0 alert for PAINS.

Table 7 Pharmacokinetics profiles of the studied Ruthenium complexes (V2, W and X)

Compounds	GI absorption	BBB permeant	P-gp substrate	CYP1A2 inhibitor	CYP2C19 inhibitor	CYP2C9 inhibitor	CYP2D6 inhibitor	CYP3A4 inhibitor	Log K_p (skin permeation) cm s^{-1}
V2	Low	No	No	No	No	No	No	No	−4.91
W	Low	No	No	No	No	No	No	No	−6.12
X	Low	No	No	Yes	No	No	No	No	−5.83

(cm^{-1}): 3417, 3051 (O–H), 2981 (C–H, alkane), 1894 (COO[−]), 1712 (C=O), 1612 (C=C), 1566 (C=N), 1481, 1435, 1369, 1265 (C–O), 1145, 1118, 1087 (C–N, aryl), 995, 748, 721, 690, (Ru–P), 528 (Ru–N), 420 (Ru–Cl). UV-Vis (ϵ , $\text{M}^{-1} \text{cm}^{-1}$, nm, (DMSO)): λ_{max} = 492 (1615), 364 (3438), 283 (9563), 227 (6760). ¹H-NMR (400 MHz, DMSO- d_6 , δ): 7.78 (s, 1H), 7.64–7.60 (m, 4H), 7.57–7.53 (m, 3H), 7.49 (s, br, 1H), 7.46 (d, J = 12.0 Hz, 1H), 7.41–7.39 (m, 10H), 7.25–7.23 (m, 5H), 7.22 (m, 1H), 6.28 (d, J = 12.0 Hz, 1H). ¹³C NMR: 168.42, 138.12, 137.95, 137.11, 137.02, 133.75, 133.70, 133.60, 132.47, 131.95, 131.88, 129.43, 129.24, 129.19, 129.15, 115.29; ³¹P-NMR (162.00 MHz, DMSO- d_6 , δ): 25.46 (s, P). Anal. calcd for $\text{RuC}_{34}\text{H}_{29}\text{N}_4\text{O}_2\text{Cl}_2\text{P}\cdot 3\text{H}_2\text{O}$ (%): C, 52.17; H, 4.52; N, 7.16; found: C, 53.89; H, 4.99; N, 7.97. TOF-MS-ES +: m/z ; found = 808.35 (Anal. calcd 805.67 for $[\text{M}+\text{Na}]^+$, $3\text{H}_2\text{O}$).

(W): **synthesis of $[\text{RuCl}_2(\text{PPh}_3)_3\text{Hbpy}(\text{mza})]\cdot\text{H}_2\text{O}$** . The complex was synthesized using the same reaction procedures as reported for complex V2 (Scheme 1). The reaction was carried out in a 100 mL round-bottom flask. The flask was charged with 20 mL of absolute ethanol. Commercial dichloro-ruthenium(II) tris(triphenylphosphine) $[\text{RuCl}_2(\text{PPh}_3)_3]$ (0.35 g, 0.37 mmol), 4-imidazoleacrylic acid (0.07 g, 0.37 mmol), and 2,2'-bipyridine-4,4'-dicarboxylic acid (0.09 g, 0.37 mmol) were added successively. A 5–10 mL of water was later added to aid in the complete dissolution of the reagents. The mixture was then heated to reflux under vigorous magnetic stirring for 12 hours while monitoring the reaction with TLC (thin-layer chromatography). The reaction product was allowed to cool after reflux and then filtered. The unreacted solid was separated by gravity filtration with the addition of a small quantity of water (20–50 mL) for washing. The filtrate was concentrated *in vacuo* to obtain a dark brown solid, which was a syrup. The semi-crude product was purified by column chromatography using toluene-ethanol (7:3, v/v, 250 mL) to obtain a light brown solid, which was further purified by recrystallization in 30–40 mL of diethyl ether. The yield was 67%, with a melting point (mp) of 188–189 °C. FT-IR (cm^{-1}): 3348, 3221 (O–H), 3010 (C–H, alkane), 1658 (C=O), 1589 (C=C), 1539 (C=N), 1411, 1327, 1265, 1141, 1037 (C–N, aryl), 894, 813, 744,

698, 651, (Ru–P), 589 (Ru–N), 461 (Ru–Cl). UV-vis (ϵ , $\text{M}^{-1} \text{cm}^{-1}$, nm, (DMSO)): λ_{max} 475 (1034), 336 (1602), 299 (5224), 272 (7889), 228 (9822). ¹H-NMR (400 MHz, DMSO d_6 , δ): 7.63–7.59 (m, br, 6H), 7.56–7.52 (m, br, 5H), 7.38 (s, br, 5H), 7.25–7.21 (m, br, 4H), 7.11–7.07 (m, br, 4H), 6.30–6.27 (m, 1H). ¹³C NMR: 169.36, 165.67, 159.21, 159.17, 140.12, 137.11, 137.02, 135.80, 133.75, 133.60, 132.71, 132.50, 132.48, 131.95, 131.88, 129.43, 129.25, 129.19, 129.15. ³¹P-NMR (162.00 MHz, DMSO- d_6 , δ): 25.48 (s, P). Anal. calcd for $\text{RuC}_{36}\text{H}_{29}\text{N}_4\text{O}_6\text{Cl}_2\text{P}\cdot\text{H}_2\text{O}$ (%): C, 52.96; H, 3.59; N, 6.86; found: C, 52.71; H, 3.76; N, 6.08. TOF-MS-ES +: m/z found = 857.20 (Anal. calcd. 857.66 for $[\text{M} + \text{Na}]^+$, H_2O).

(X): **synthesis of $[\text{RuCl}_2(\text{PPh}_3)_3\text{ptd}(\text{mza})]\cdot 3\text{H}_2\text{O}$** . The complex was synthesized using the same reaction procedures as reported for complex W (Scheme 1). The reaction was carried out in a 100 mL round-bottom flask. The flask was charged with 20 mL of absolute ethanol, and commercial dichloro-ruthenium(II) tris(triphenylphosphine) $[\text{RuCl}_2(\text{PPh}_3)_3]$ (0.35 g, 0.37 mmol), 4-imidazoleacrylic acid (0.07 g, 0.37 mmol), and 1,10-phenanthroline-5,6-dione (0.08 g, 0.37 mmol) were added successively. A 5–10 mL portion of water was later added to aid in the complete dissolution of the reagents. The mixture was then heated to reflux under vigorous magnetic stirring for 12 hours while monitoring the reaction with a TLC procedure. The reaction product was allowed to cool down after reflux and filtered. The unreacted solid was separated under gravity by adding a small quantity of water (20–50 mL) to wash it. The filtrate was concentrated *in vacuo* to obtain a dark-brown syrupy solid. The semi-crude product was purified *via* column chromatography using toluene-ethanol (7/3, v/v, 250 mL) to obtain a light-brown solid, which was further purified by recrystallization in 30–40 mL of diethyl ether. Yield = 55%, melting point (mp) 150–152 °C. FT-IR (cm^{-1}): 3406, 3329, 3219 (O–H), 3066 (C–H, alkane), 2079 (C=N), 1755 (C=O), 1643 (C=C), 1597, 1566 (C=N), 1427, 1373, 1315, 1234, 1130, 1041, 1002 (C–N, aryl), 914, 786, 671, 601 (Ru–P), 516 (Ru–N), 420 (Ru–Cl). UV-Vis (ϵ , $\text{M}^{-1} \text{cm}^{-1}$, nm, (DMSO)): λ_{max} 494 (4866), 378 (9205), 287 (6599), 272 (6863), 267 (6450), 228 (12 742). ¹H-NMR (400 MHz, DMSO- d_6 , δ): 7.64–7.63 (m, 3H), 7.62 (m, 5H),



7.60 (m, 4H), 7.59 (m, 3H), 7.56 (m, 4H), 7.55 (m, 2H), 7.54 (m, 2H), 7.53 (m, 2H). ^{13}C NMR: 170.15, 166.91, 154.85, 148.78, 138.83, 136.11, 133.53, 132.72, 132.50, 132.48, 131.95, 131.88, 129.25, 129.15, 126.95. ^{31}P -NMR (162.00 MHz, DMSO- d_6 , δ): Unresolved. Anal. calcd for $\text{RuC}_{36}\text{H}_{27}\text{N}_4\text{O}_4\text{Cl}_2\text{P}\cdot 3\text{H}_2\text{O}$ (%): C, 51.69; H, 3.98; N, 6.70; found: C, 52.96; H, 4.89; N, 6.23. TOF-MS-ES $+$: m/z , found = 857.21 (Anal. calcd. 859.64 for $[\text{M}+\text{Na}]^+\cdot 3\text{H}_2\text{O}$).

Materials and methods

Experimental section

The starting materials used were $\text{RuCl}_2(\text{PPh}_3)_3$, 4-imidazoleacrylic acid, 2,2'-bipyridine, 2,2'-bipyridyl-4,4'-dicarboxylic acid, and 1,10-phenanthroline-5,6-dione, which were purchased from Sigma-Aldrich, Alfa Aesar, and Ark Pharm. All complexes, **V2**, **W**, and **X**, were synthesized according to similar reported procedures, with the exception of changes in chemical reagents, reaction time, and purification solvents.⁷¹ All required solvents were ordered from Fischer Scientific. To confirm the purity of the synthesized complexes, data were collected using a Bruker Alpha FTIR spectrometer, which was equipped with an ATR platinum Diamond 1 reflectance accessory. Microanalyses (C, H, and N) were carried out with a (Vario Elementar), University of Witwaterstrand, South Africa. A 400 MHz Bruker NMR spectrometer equipped with an auto-sampler was used for the ^1H , at 100 MHz for ^{13}C , and at 162.00 MHz for ^{31}P -NMR spectra. Chemical shift values were reported in parts per million (ppm) relative to tetramethylsilane (TMS) as an internal standard. Chemical shifts were also reported with respect to DMSO- d_6 at δ C 40.98 ppm and DMSO- d_6 at δ H 2.50 ppm. UV-vis absorption and emission data were recorded in a 1 cm path length quartz cell on a PerkinElmer Lambda 35 spectrophotometer and an LS-45 Fluorescence spectrometer, respectively. Luminescence quantum yields were calculated using eqn (1), where Φ_{D} is the quantum yield, F and A are the gradients of luminescence *versus* absorbance, and n is a solvent refractive index. Subscripts D and std refer to donor and standard, respectively. Quinine sulphate in 0.1 M H_2SO_4 was employed as the standard ($\Phi_{\text{std}} = 1.33$; $A_{\text{std}} = 0.050$; $F_{\text{std}} = 1000$); $\lambda_{\text{ex}} = 400$ nm in DMSO = 1.479; room temperature. Melting points were determined by Stuart melting point apparatus analogue SMP 11 at a temperature range between 50–350 °C. Direct injections of the respective samples into a Waters Micromass LCT Premier MS instrument equipped with electrospray ionization (ESI) source and a time-of-flight (TOF) mass analyzer afford the mass spectra. Electrochemical experiments were conducted using an Autolab PGSTAT 302 N electrochemical workstation equipped with an Electrochemical Impedance Spectroscopy (EIS) module and the Autolab NOVA 1.7 software package. During these experiments, a three-electrode voltammetric cell including a glassy carbon working electrode, Ag|AgCl wire as a pseudo reference electrode and Pt counter electrode were used. The square-wave voltammetry settings include a step potential of 4 mV, a frequency of 25 Hz and amplitude of 20 mV. The cyclic

and square wave voltammograms of the complexes **V2**, **W**, and **X** were examined in the potential range +1.5 to −1.5 V and at a scan rate 50 mV s^{−1} in acetonitrile solvent with 0.1 M tetrabutylammonium tetrafluoroborate as supporting electrolyte.

Quantum chemical calculations

Density functional theory (DFT) calculations were conducted on the structures of **V2**, **W**, and **X** using Gaussian 16 software,⁷² on the Lengau cluster at the Center for High-Performance Computing (CHPC) in Cape Town, South Africa. Initial 3D molecular structures were constructed and minimized with molecular mechanics force field by Chem 3D software, incorporating synthesized inhibitors and the obtained standard drugs (Crizotinib, Doxorubicin, Gemcitabine, and Lorlatinib) from PubChem Chemical Database. The stable structures were saved in Mol2 format and fully optimized using the exchange–correlation B3LYP hybrid functional⁷³ and ECP (effective core potential) LANL2DZ basis set was used to model the metal atoms^{74,75} in a vacuum. Meanwhile for the standard drugs to align with the molecular docking geometry, we optimized the molecules using PM6 basis set in semi-empirical method. Key quantum chemical properties, including frontier orbital energies (EHOMO and ELUMO), energy gap (E_{g}), ionization energy (IE), electron affinity (EA), electronegativity (χ), chemical hardness (η), chemical softness (σ), and global electrophilicity index (ω) were calculated.⁷⁶ These parameters provide insight into molecular reactivity and potential interactions with biological systems. The optimized structures are saved in Mol2 format for further computational study.

Molecular docking study

Molecular docking targeted three key cancer proteins: estrogen receptor alpha (ER α) (PDB: 8DU6, 2.10 Å resolution), WT EGFR (PDB: 8PO4, 1.62 Å resolution) and the receptor tyrosine kinase anaplastic lymphoma kinase (ALK) (PDB: 9GBE, 1.58 Å resolution), retrieved from the RCSB Protein Data Bank.^{77–79} Protein preparation in Molegro Virtual Docker (MVD) involved repairing and optimizing the protein structure, while removing water molecules and cofactors from crystal structure files. Optimized **V2**, **W**, and **X** structures, alongside standard drugs, were docked against these proteins using the MolDock [GRID] algorithm, which integrates differential evolution with cavity prediction.⁸⁰ MVD automatically identifies potential active sites (also referred to as cavities) by using its cavity detection algorithm. The cavity with the highest volume was selected for consideration. Ten independent docking runs generated single poses per run, ranked based on set of scoring functions, such as MolDock [GRID] and MolDock scores. However, we selected the MolDock [GRID] scores for ranking the inhibitor poses for the receptor targets. The highest-scoring poses were selected for each target receptor, and intermolecular interactions were analysed using Biovia Discovery Studio 2024 and ChimeraX.⁸¹ This methodology provided insights into binding affinities and structural interactions critical for evaluating inhibitory potential.

ADME prediction

The SwissADME web tool (<https://www.swissadme.ch/>) evaluated physicochemical properties, pharmacokinetics, lipophilicity,



and solubility of the compounds **V2**, **W**, and **X**. SMILES notations of optimized structures were submitted to predict drug-likeness and assess candidacy for further development. All computations were carried out using the computational cluster resources at the Centre for High Performance Computing (CHPC), Cape Town, South Africa.

Conclusions

In conclusion, this study reports the facile synthesis of new mixed-ligand heteroleptic Ru(II) complexes. The optical, electrical, and molecular docking properties of compounds **V2**, **W**, and **X** demonstrate the influence of polypyridyl ligand substitutions, highlighting the superior quality of using combinations of bidentate and monodentate ligands in the buildup of potential bioactive anticancer agents. Although compound **V2** shows the highest photoluminescent emission intensity and wavelength in the near-infrared region (>700 nm), it surprisingly gave the lowest quantum yield. The quantum yield ratio was found to be 1:2:3 for compounds **V2**, **W**, and **X**, respectively. The biological investigations using DFT/molecular docking methods show that the HOMO in complexes **V2** and **W** is primarily distributed over the 4-imidazole acrylic acid moiety, extending to the central ruthenium metal and one chlorine atom. The LUMO, in contrast, is localized across the chlorine atoms and the central metal atom with energy gaps (ΔE) of 2.5375 eV and 3.067 eV, respectively. In contrast, complex **X** has the HOMO localized over the 4-imidazole acrylic acid moiety, with a minor spill-over onto the central ruthenium metal. Its LUMO is primarily localized on the 1,10-phenanthroline-5,6-dione moiety and exhibits a smaller energy gap (ΔE) of 1.2457 eV. Complex **V2** achieved the lowest binding energy compared to complex **X**, which had the highest binding affinity *via* hydrogen bonding and π -alkyl interactions. This was attributed to the presence of 1,10-phenanthroline containing two carbonyl substitutions at positions 5 and 6. Overall, compounds **V2**, **W**, and **X** show superior anti-transcriptional activity compared to the standard drugs, which include crizotinib, doxorubicin, gemcitabine, and lorlatinib. Results of the physico-chemical properties, lipophilicity, pharmacokinetics, and drug-likeness to ensure therapeutic efficacy show that the complexes have one violation or another. Specifically, their high molecular weights and/or poor lipophilicity hindered them from crossing the blood-brain barrier (BBB). This limited their utility for CNS targets (such as brain tumors) but reduced off-target neurotoxicity. In conclusion, this study provides new information on the complexes, revealing the strong anticancer potential of the individual complexes, which can be used as biological electrophiles. It is envisaged that the *in vitro* and *in vivo* biological activities of the reported complexes will be investigated in the future.

Author contributions

A. O. A. conceived and designed the research idea, carried out the synthesis of the mixed-ligand Ru complexes, performed UV-vis, PL, and electrochemical analyses, and interpreted IR

and NMR data. T. K. C. was responsible for the IR and NMR acquisition and analysis. D.D.B carried out experiments on DFT, MEPs as well as the acquisition of data, analysis and interpretation of data. A. O. A. coordinated the writing of the manuscript. All authors read, edited and contributed to the manuscript and agreed to the published version of the manuscript.

Conflicts of interest

The authors declare that there are no conflicts to declare or any known competing financial interests of personal relationships that could have appeared to influence the work reported in this paper.

Data availability

All authors declare here that all the data generated or analyzed during this study are included in the manuscript and its supplementary information (SI) files and link attached below: <https://bit.ly/4c4Irsj>. Supplementary information is available. See DOI: <https://doi.org/10.1039/d5nj03704b>.

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