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Discovery of semisynthetic derivatives of (R)- and (S)-usnic acids as potential antifungal agents against *C. tropicalis* and *T. rubrum*†

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The prevalence of human fungal infections (FIs) is rapidly increasing worldwide, posing substantial challenges to public health. The underestimation of FIs risk led to a limited knowledge of the fungal pathogenicity and a concomitant paucity of antimycotic drugs that are increasingly unable to effectively address resistance liabilities. The identification of innovative antifungal drugs is therefore an urgent need. Natural products have always been under scrutiny in the drug discovery process. Of these, usnic acid (UA) represents a compelling starting point for antifungal drug development due to its natural occurrence as a secondary metabolite in various lichen species, where it serves as a natural defence mechanism against fungal invasion. This dibenzofuran derivative possesses an intrinsically rigid three-dimensional architecture with stereogenic center, providing a pre-organized chiral scaffold with potential for selective interaction with fungal targets. Despite its high therapeutic potential as antimicrobial agent, UA suffers from poor solubility and hepatotoxicity issues. The proposed research explores the modification of UA scaffold to generate the series of semisynthetic compounds 1-9 by derivatizing the (R)- and (S)-UA as enamines. Considering the inherent chirality of UA, this work aims to identify structure-activity relationships that optimize antifungal efficacy while improving the pharmacokinetic properties of UA. The resulting compounds were evaluated for their antifungal activity against three strains, showing significant differences in potency concerning their absolute configuration. This research addresses the urgent need for novel antifungal agents in an era of increasing resistance to conventional treatments, identifying (9bS,15S)-1, 3, 4, and 8 compounds as promising compounds for developing antifungal therapeutics.

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1. Introduction

Fungal infections (FIs) or mycoses have historically received less attention than bacterial and viral infections both from academia and pharmaceutical companies, due to their relatively low incidence and mortality rates in developed countries. Nonetheless, fungal infections account for 300 million cases per year, resulting in 1.5 million deaths, most of which occur in neglected populations.^{1–5}

In the last years, FIs, particularly among hospitalized and immunocompromised patients, have raised several public health concerns, prompting the World Health Organization to release a priority list of the main fungal threats in 2022.^{6,7} This formal report emphasised the need for an unprecedented research effort to thoroughly understand the mechanisms underpinning fungal virulence and, consequently, to expand the limited arsenal of available antifungal drugs.⁷ Notably, recommended therapy for the treatment of FIs has remained largely unchanged over the last decade, accounting for the use of broad-spectrum antimycotics such as amphotericin B (AmB) and azoles (e.g. fluconazole, FCZ) as mono- or combination therapy.^{8,9} Multi-drug approaches are regularly employed to counteract the increasing drug resistance acquired by several fungal strains.^{10,11}

Among pathogenic fungi, *Candida* spp. (e.g. Candida albicans, Candida tropicalis and Candida auris) represent the most common etiological agents of mucocutaneous and invasive FIs, while *Trichophyton* spp. (e.g. *Trichophyton rubrum*

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and Trichophyton mentagrophytes) is responsible for up to 50% of dermatophytosis, superficial FIs occurring on keratinous substrates. 12-14 The increasing use of antifungal drugs for cutaneous mycosis treatment, and the long-term therapeutic regimens required to eradicate the vast majority of recurrent FIs, have led to the emergence of drug-resistant fungal strains especially against AmB and azoles. 11,15

Given the magnitude of this global health issue, launching new drug discovery programs is essential for identifying new chemical entities capable of tackling, or at least limiting, the spread of FIs.16

In medicinal chemistry campaigns, natural products are extensively investigated due to their intrinsic properties and distinctive structural complexity, often leading to the discovery of biologically active metabolites and inspiring the design of new compounds. 17-19 Although still poorly studied, lichens have increasingly emerged as rich sources of secondary metabolites with multiple pharmacological activities, as evidenced by their use in traditional medicine.20-22 Among these, usnic acid (UA) is one of the best-characterized, being abundantly biosynthesised by numerous lichen species.²³ Due to the presence of a stereogenic centre, UA can occur in nature in both enantiopure and racemic forms. 24,25 Specifically, (+)-(R)-UA is the most abundant in several genera including Ramalina and Usnea. Conversely, (-)-(S)-UA is found in small amounts in a few species of Cladonia and Alectoria. This explains the low commercial availability and the high retail cost of the (S)enantiomer which has limited its investigation in a therapeutic perspective. 24,26

UA exhibits a broad spectrum of biological assets, including antineoplastic, anti-inflammatory antimicrobial activities. 23,27-29 Extensive literature examined the anti-infective properties of UA, but only a limited number of studies have investigated demonstrated its significant antimycotic activity, primarily focusing on the (R)-enantiomer. $^{25,30-34}$

Despite its pharmacological potentiality, UA has faced challenge in therapeutic significant development. Documented cases of hepatotoxicity prompted regulatory intervention, with FDA ordering the withdrawal of UAcontaining products from the market due to safety concerns. 35,36 Further, UA suffers from poor water solubility, strong binding affinity for serum proteins, and low bioavailability.37-39 These pharmaceutical limitations restrict its use for both systemic and topical treatment. 40-42 To improve the pharmacokinetic properties of UA, different chemical modifications on the scaffold have been explored, leading to different classes of biologically active compounds (e.g. enamine, benzofuran-2-one, heteroaryl, and pyrazole derivatives). 26,32-34,43,44

Building on these findings, herein we prepared a series of semisynthetic UA-based enamines (compounds 1-9, Table 1) as potential agents against FIs. We also investigated whether the configuration of the stereogenic center in UA affects antifungal potency and how systematic

Table 1 Antifungal activity of the semisynthetic UA-based enamines 1-9 against C. tropicalis, and T. rubrum

Cpds	$MIC_{99} (\mu M)$	MIC_{99} (μM)	
	C. tropicalis ^a	T. rubrum ^a	
AmB	>400	>400	
FCZ	>200	>200	
(R)-UA	17.4	580	
(S)-UA	4.54	580	
(9bR,15S)-1	6.70	450	
(9bS,15S)-1	0.22	28	
(9bR,15S)-2	12.0	400	
(9bS,15S)-2	24.0	100	
(9bR,15S)-3	1.59	405	
(9bS,15S)-3	0.40	405	
(9bR,15S)-4	11.8	394	
(9bS,15S)-4	1.54	394	
(9bR,15R)-5	223	446	
(9bS,15R)-5	>446	446	
(R)-6	214	427	
(S)-6	26.7	427	
(9b <i>R</i> ,15 <i>S</i>)-7	>246	246	
(9bS,15S)-7	15.4	7.40	
(9bR,15S)-8	7.80	>260	
(9bS,15S)-8	1.00	>260	
(9b <i>R</i> ,15 <i>S</i>)-9	>253	>253	
(9bS,15S)-9	>253	>253	

^a Experiments were performed in triplicate.

modifications of each UA enantiomer can further modulate the antifungal activity.²⁶ Indeed, the importance of chirality in the drug discovery process is well known, and the field of antifungal medicines is no exception.⁴⁵ Many antifungals are chiral, ranging from simple synthetic azoles to more intricate natural and semi-synthetic scaffolds amphotericin B, echinocandins, anidulafungin). 46-48 To evaluate the impact of the absolute configuration of UA scaffold on its antifungal activity, both the commercially available (R)-UA, and the (S)-UA isolated from Cladonia foliacea were properly functionalized, generating compounds 1-9. These latter were subsequently tested against three fungal strains, Candida albicans, Candida tropicalis, and Trichophyton rubrum to evaluate their antimycotic and fungicidal activity. Moreover, to assess the safety, the cytotoxicity on human dermal fibroblasts was assessed.

2. Results and discussion

2.1 Design and synthesis of UA-based enamines

To improve both safety and drug-like properties of UA while retaining its antifungal properties, a series of semisynthetic derivatives of (R)-UA and (S)-UA were designed (1-9). A comprehensive review of the literature has revealed that the approved antifungal drugs are medium-to-large sized molecules, characterised by intrinsic flexibility.8,49,50 Accordingly, we decided to derivatize the constrained and disk-shape UA enantiomers as enamines, since this chemical modification, unlike the others applied on the scaffold, allowed the insertion of fragments endowed with different Research Article

Fig. 1 Structure of usnic acid, with atom assignment, and of the designed enamine derivatives 1-9.

flexibility, length, and polarity (Fig. 1). Specifically, the ketone in position 13 was derivatized with amino acids (i.e. serine, arginine, phenylalanine and tyrosine), hydrophobic benzyl moieties (i.e. 1-methyl-benzylamine and 3-chloro-benzylamine). We also explored the conjugation of compounds 4 with flexible PEG chains or linear carbon chains. While the introduction of PEG moiety may offer many benefits, including the reduced renal excretion and

Scheme 1 Reagents and conditions: a) appropriate amine (1 equiv.), TEA (2 equiv.), EtOH, N2 atm, mw, 90 °C, 15 min (×3) (for 1, 3-6); b) L-arginine (1 equiv.), TEA (2 equiv.), EtOH, refl., N2 atm, 14 h (for 2).

proteolysis as well as increased water solubility, the saturated carbon chain may facilitate molecule permeability through membranes or disrupt their integrity.⁵¹

The semisynthetic derivatives 1-9 were prepared starting from homochiral UA. (S)-UA was isolated from Cladonia foliacea following our in-house well-established protocol, whereas the opposite enantiomer was purchased.²⁶

The synthesis of compounds 1-6 is described in the Scheme 1.

UA enantiomers were condensed with the appropriate amines in presence of triethylamine as base, in ethanol under heating affording the corresponding enamine derivatives 1-6, as described in the Scheme 1. To attain a green and efficient synthesis, microwaves irradiation was investigated. With the sole exception of compound 2, mw irradiation resulted an efficient procedure for the preparation of the UA-based enamines, since it allowed obtain the product in 45 minutes with yield ranging from 37 to 87%, depending on the substrate. For the synthesis of compounds 7-9, long chain amines 10-13 were prepared first, according to the Scheme 2.

Compound 10 was obtained starting from commercially available synthon 13 which was Bocdeprotected using trifluoroacetic acid in dichloromethane at 0 °C for 2 hours, to give amino acid 14 which underwent a mw-assisted Fisher esterification in methanol, using sulfuric acid as catalyst to give the corresponding methyl ester 10. For the preparation of compound 11, the N-Boc-ethylenediamine 15 was condensed with the dicarboxylic acid 16 using 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (EDC-HCl) and a catalytic amount of hydroxy-benzotriazole (HOBt) as coupling agents, and diisopropylethylamine (DIPEA) as base in anhydrous DMF under nitrogen atmosphere for 16 hours, to provide the corresponding amide 17. Treating 17 with trimethylsilyl chloride (TMSCl) in methanol allowed both the simultaneous methyl esterification and the removal of the Boc-group, thus obtaining the amine 11. For the synthesis of amine 12, 3-amino-1-propanol 18 was alkylated with hexadecyl bromide affording the intermediate 19 followed by Boc-deprotection. Lastly, compound 4 was coupled with amines 10-12 in presence of EDC-HCl, HOBt and DIPEA, affording the corresponding enamines 7-6 (Scheme 3).

All newly synthesized compounds were fully characterized by ¹H and ¹³C NMR spectroscopy. The observed chemical shifts were in accordance with previously reported values for structurally analogous compounds, confirming regioselective derivatization of usnic acid at the C-13 position. 52-54

2.2 Antifungal activity

The antifungal activity of the newly synthesized compounds 1-9, alongside the parent enantiomers (R)-UA and (S)-UA, was evaluated against the following clinical strains: Candida 1023¹⁴⁶, isolated from albicans ATCC bronchomycosis Candida tropicalis ATCC 750 isolated from bronchitis patient, and a strain of Trichophyton rubrum LM 237 isolated from patient with foot onychomycosis and maintained in the mycological collection of the University of Pavia (LM). The MIC99 values (defined as the minimum concentration required to inhibit 99% of fungal growth) was determined by means of multiwell microplates.

To assess the therapeutic potential of our compounds, we selected clinical isolates with known resistance to FCZ and AmB. Consistent with their resistant profiles, both reference drugs displayed limited effectiveness (MIC99 > 200 µM or >400 µM, Table 1). This strategic choice is in line with medical need and clinical urgency. Whilst FCZ is widely used to treat candidemia, especially against C. albicans infections,

BocHN
$$\longrightarrow$$
 O \longrightarrow OH \longrightarrow H₂N \longrightarrow OH \longrightarrow OH

Scheme 2 Reagents and conditions: a) TFA/DCM 1:1, from 0 °C to r.t., 2 h; b) H₂SO₄, MeOH, mw, 120 °C, 2 min (×3); c) appropriate amine (1 equiv.), EDC-HCI (1 equiv.), HOBt (cat.), DIPEA (2 equiv.), anh. DMF, N₂ atm, from 0 °C to r.t., 3-14 h; d) TMSCI, MeOH, from 0 °C to r.t., 5 h; e) Boc₂O; DCM, N₂ atm, from 0 °C to r.t., 6 h; f) NaH, N₂ atm, 0 °C, 1 h, followed by the alkyl bromide, N₂ atm, from 0 °C to r.t., 14 h; g) TFA, DCM, from 0 °C to r.t., 2 h.

Scheme 3 Reagents and conditions. a) Amine 10-12 (1-1.5 equiv.), TEA (2 equiv.), EDC-HCl (1.5 equiv.), HOBt (0.1 equiv.), DIPEA (4.0 equiv.), anh. DMF, N₂ atm, from 0 °C to r.t., 3-14 h.

its efficacy has started to be affected by the emergence tolerance. Persistent candidemia with C. tropicalis is showing a worrying increase in FCZ resistance, reaching rates up to 25%.55 T. rubrum, a predominant cause of chronic dermatophytosis, is increasingly resistant to azoles, especially FCZ, often exhibiting MIC₅₀ values ≥160 μM following prolonged therapeutic exposures.

The situation with amphotericin B is somewhat different. Both C. albicans and C. tropicalis exhibit low resistance to AmB, but some cases of resistant strains have started to being reported, particularly among immunocompromised patients, likely because of prior exposure to antifungal agents, as the ones used in the present study. 56,57 Although no evidence of resistance against T. rubrum has been observed, its use against dermatophytes is not common due to its nephrotoxicity and the superficial nature dermatophyte infections.⁵⁸

Regarding the newly developed series, a first screening at a concentration of 200 µM was performed. Since no effect of the UA derivatives was observed with C. albicans, this strain was not further investigated. A different behaviour was observed for C. tropicalis and T. rubrum, as summarized in Table 1.

Specifically, UA enantiomers as well as most semisynthetic derivatives (except for compounds 5, (R)-6, (9bS,15S)-7 and 9) displayed outstanding antifungal activity against C. tropicalis with MIC99 values in the low to sub-micromolar range. Among these, compounds (9bR,15S)-1, (9bS,15S)-1, (9bR,15S)-3, (9bS,15S)-3, (9bS,15S)-4, (9bR,15S)-8, and (9bS,15S)-8 stood out for their single-digit/sub-micromolar activity with MIC99 values of 6.70, 0.22, 1.59, 0.40, 1.54, 7.80 and 1.00 μM, respectively. To determine whether the observed antifungal activity was fungicidal or fungistatic, the minimum fungicidal concentrations (MFC) were determined for the most potent compounds. Compounds (9bS,15S)-3 and (9bS,15S)-4 exhibited MFC values of 25.4 μM and 49.3 μM, with an MFC/MIC ratio of 63.5 and 32, respectively, thus highlighting a prominent fungistatic rather than fungicidal mode of action.

T. rubrum exhibited modest susceptibility toward to both UA enantiomers and most semisynthetic derivatives 1-9 with MIC₉₉ values ranging from 100 to 580 μM. Notably, compounds (9bS,15S)-1 and (9bS,15S)-7 represented significative exceptions, exhibiting MIC₉₉ values of 28 and 7.4 μM, respectively, resulting in a 20-fold and 78-fold potency gains over the parent (S)-UA.

2.3 Effect of stereochemistry at position 9b on antifungal activity

Analysis of MIC99 values of compounds bearing the (R)- and (S)-UA as scaffold reveals significant insight into the role of chirality in the antifungal efficacy. Results reported in Table 1 suggest that the absolute configuration impacts antifungal activity, with structural modifications of the parent scaffold amplifying potency differences between the UA enantiomers. This relationship can be quantified by calculating the ratio between the MIC99 values of (R)-UA (or its derivative) and the MIC_{99} of (S)-UA (or its derivative). It should be noted that many compounds in our series contain an additional chiral center on the side chain at C-15, resulting in diastereomeric rather than enantiomeric pairs. However, since the absolute configuration of this additional stereocenter remains consistent across compared pairs, we can reasonably attribute the differences in the activity to the absolute configuration of configuration at position 9b of UA scaffold.

Focusing on the parent enantiomers (R)-UA and (S)-UA, they exhibited modest difference in antimycotic activity against C. tropicalis, with (S)-UA resulting 4-fold more potent (MIC₉₉ of 17.42 and 4.54 μ M, for (R)-UA and (S)-UA, respectively). However, upon derivatization, several derivatives demonstrated significantly enhanced potency and increased eudysmic ratio (ER, defined as the ratio of activity between stereoisomers at position 9b), underscoring the critical role in activity (Fig. 2). Notably, compounds with (S)configuration at position 9b demonstrated superior activity.

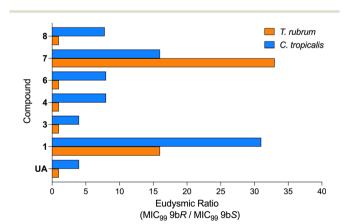


Fig. 2 Eudysmic ratios of UA and its derivatives, calculated by dividing the MIC99 of the less active stereoisomer (distomer) by the MIC99 of the more active stereoisomer (eutomer). An ER = 1 indicates equivalent activity between stereoisomers.

For example, (9bS,15S)-1 exhibited ERs of 15 and 31 in the MIC99 against T. rubrum and C. tropicalis, respectively. Similarly, compounds (9bS,15S)-3, (9bS,15S)-4, (9bS,15S)-6 and (9bS,15S)-8 showed ERs between 4 and 8 toward C. tropicalis. The most pronounced stereochemical dependence was observed with compound (9bS,15S)-7 which showed the highest eudysmic ratio in the whole series (ER = 33) in the activity against T. rubrum.

These data collectively suggest that the (S)-configuration at position 9b of UA scaffold generally confers optimal antifungal activity. This stereochemical arrangement likely facilitates more favourable interaction with fungal targets or more efficient disruption of fungal cellular processes. Our findings highlight the potential of (S)-UA and semisynthetic derivatives as promising candidates for addressing resistance mechanisms in clinically challenging fungal pathogens.26

2.4 Cytotoxicity studies

A primary requirement for in vivo application of novel antifungal agents is their biocompatibility, which includes both non-toxicity and good tissue tolerance. To determine the safety margin—the concentration range that is effective against fungi but safe for human cells-of our newly developed derivatives, we conducted cytotoxicity studies on normal human dermal fibroblasts at two different concentrations (50 µM, and 125 μM). This cell line was selected because both C. albicans and T. rubrum are responsible for mucocutaneous and cutaneous infections, making dermal fibroblast physiologically relevant for evaluation.

Results demonstrated that the enamine derivatization decreases the cytotoxicity profile of the parent UA (Fig. S1 and S2†). Specifically, most of the new derivatives display good cytocompatibility profile with cell viability percentages above 90% at both tested concentrations (Fig. 3). Exceptions include compound (9bR,15S)-7, which resulted completely

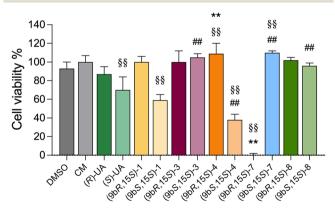


Fig. 3 Cell viability values% obtained after 24 hours of contact with the cellular substrate for the most promising samples at 50 μ M. DMSO, subjected to the same dilution of the stock solution, was used as a control. Mean values \pm sd (n = 4). ANOVA A one-way multiple range test (p < 0.01), with ** (p < 0.01) vs. (R)-UA, ## (p < 0.01) vs. (S)-UA, and §§ (p < 0.01) vs. culture medium (CM).

cytotoxic, and the pair of diastereomers (9bR,15S)-8 and (9bS,15S)-8 which exerted cytotoxicity only at the highest concentration tested.

2.5 Water thermodynamic solubility.

Given the poor water solubility associated with UA core, we envisaged a possible topical formulation for the enamine derivatives. In this context, we experimentally evaluated water solubility for the most promising compounds as this parameter is critical for developing effective topical formulations ensuring the drug delivery across skin barriers. Aqueous solubility was determined by HPLC for the bestperforming compounds (9bR,15S)-1, 3, 4, 7 and 8 as well as their corresponding diastereoisomer, applying a method already developed by us.⁵⁹ Results reported in Table 2 evidence that enamines (9bR,15S)-1, 3 and 4 incorporating amino acids motifs, exhibit a markedly enhanced water solubility ranging from 0.15 to 4.7 mM. However, no solubility improvements were observed for (9bR,15S)-7 and 8 despite the insertion of PEGylated lateral chains.

3. Conclusion

This study explored the potential of UA as a chiral scaffold for the development of semisynthetic antimycotic agents. A series of semisynthetic enamine derivatives of both (R)- and (S)-UA were rationally designed, synthesized, and evaluated for their antifungal properties against clinically relevant, drug-resistant strains of C. tropicalis and T. rubrum. Among the series, compounds (9bS,15S)-1, 3, 4, and 8 demonstrated remarkable low to sub-micromolar activity against C. tropicalis, outperforming both the parent UA and the clinical antifungals amphotericin B and fluconazole. Notably, compound (9bS,15S)-1 also exhibited activity against T. rubrum indicating a broader spectrum efficacy. Biological evaluation revealed that configuration at the 9b stereocenter of UA affects the antifungal activity. In almost all the diastereomeric pairs, derivatives bearing the outperformed configuration their 9b*R* counterparts, establishing structure activity relationship that can guide optimization. Furthermore, the systematic derivatization of UA effectively addressed some liabilities of

Table 2 Aqueous thermodynamic solubility experimentally determined by HPLC

Cmpd	Aqueous solubility (mM) at 25 °C	
UA	<0.3 (lit.) ⁶⁰	
(9bS,15S)- 1	0.15	
(9b <i>R</i> ,15 <i>S</i>)-3	4.36	
(9bS,15S)-3	4.70	
(9bR,15S)-4	3.81	
(9bS,15S)-4	1.92	
(9bS,15S)-7	<lod< td=""></lod<>	
(9bS,15S)-8	<loq< th=""></loq<>	

LOD: limit of detection (0.54 μM); LOQ: limit of quantification $(61.13 \mu M)$.

the parent compound, *i.e.* enhanced safety and improved solubility. Cytocompatibility assays on human dermal fibroblasts confirmed that enamine derivatization reduced the UA intrinsic toxicity, with most compounds maintaining >90% cell viability at therapeutically relevant concentrations. The enhanced water solubility observed for several amino acid derivatives significantly improves their drug-likeness profile, supporting further development for both topical and potentially systemic applications.

Overall, this study establishes the (S)-enantiomer of **UA** as a privileged chiral scaffold for the development of antifungal agents and highlights the derivatisation with amino acids as a valuable strategy for optimizing physicochemical and biological properties. Based on their antifungal activity, safety, and solubility profiles, (9bS,15S)-1, (9bS,15S)-3, and (9bS,15S)-4 have been identified as the most promising compounds and will be prioritized for further medicinal chemistry and pharmacological investigation.

4. Material and methods

4.1 General

Reagents and solvents for synthesis were purchased from Sigma-Aldrich (Italy) or VWR and were used as received, unless otherwise specified. Thin-layer-chromatography (TLC) was carried out on silica gel precoated aluminium-plates (Fluka Kieselgel 60 F254, Merck) and visualized by ultra-violet (UV) lamp and ninhydrin stain.

The monomodal oven Discover® SP instrument (CEM Corporation, Matthews, NC, USA) was used to perform reactions with microwave irradiations.

NMR spectra were recorded on i) a Bruker Avance 400 spectrometer with ¹H at 400.134 MHz and ¹³C at 100.62 MHz, and ii) a Bruker NMR Avance Neo 700 MHz with 13C at 176 MHz. Proton chemical shifts (δ) were reported in ppm and referenced to the solvent residual peak (CDCl₃, δ = 7.26 ppm; CD₃OD, δ = 3.31 ppm; DMSO- d_6 , δ = 2.50 ppm). Signals were abbreviated as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet) and br (broad). The coupling constant values (J) are reported in Hertz (Hz). ¹³C NMR spectra were recorded with complete proton decoupling. Carbon chemical shifts (δ) were reported in ppm and referenced to the solvent residual peak (CDCl₃, δ = 77.23 ppm; CD₃OD, δ = 49.00 ppm; DMSO d_6 , δ = 39.52 ppm). Compound purity was evaluated by HPLC-UV/vis on a Jasco (Tokyo, Japan) system consisting of a PU-1580 pump and a MD-1510 photodiode array (PDA) detector. Chromatogram acquisitions and elaborations were performed using the ChromNAV software (Tokyo, Japan). Analyses were run on a XBridgeTM Phenyl, (4.6 × 150 mm, 5 μm) column, at room temperature. The mobile phases were A: water containing 0.1% of formic acid, and B: acetonitrile containing 0.1% of formic acid. Elution was performed on a linear gradient from 50% to 100% B over 10 min, followed by an isocratic hold at 100% B for 3 min. The flow rate was 1.0 mL min⁻¹ and the injection volume 10 μ L. The chromatograms were recorded at 308 nm wavelength. All the final synthesized compounds showed a purity \geq 95%. Optical rotation values were recorded using a Jasco photoelectric polarimeter DIP 1000 with a 0.5 dm quartz cell at the sodium D line (λ = 589 nm). The IUPAC names of each compound were generated using ChemDraw Professional 16.0.

4.2 (-)-(S)-Usnic acid extraction from Cladonia foliacea

30 g of C. foliacea matrix was freshly prepared by grinding the lichen thalli. Three aliquots of 10 g were suspended in EtOH and then heated three-times under mw irradiation (100 W, 120 psi) at 80 °C for 5 minutes. The extract was filtered, and the solvent was evaporated under reduced pressure. The resulting green syrup was resuspended in MeOH to remove polar undesirable metabolites, filtered, and concentrated. The residue was partitioned between water and DCM. The organic phase was collected, dried over anhydrous Na₂SO₄, filtered, and concentrated. (-)-(S)-UA was purified from the crude by crystallisation with CHCl₃/EtOH 1:2. 250 mg (0.08% yield) of (-)-(S)-UA were obtained as a yellow solid. ¹H NMR (400 MHz, CDCl₃) δ 13.34 (s, 1H), 11.05 (s, 1H), 6.00 (s, 1H), 2.70 (s, 3H), 2.69 (s, 3H), 2.13 (s, 3H), 1.79 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 201.93, 200.48, 198.20, 191.86, 179.52, 164.03, 157.65, 155.35, 109.47, 105.37, 104.09, 101.67, 98.48, 59.22, 32.26, 31.43, 28.05, 7.69. ESI-MS (m/z): $[M-H]^-$ calcd for $C_{18}H_{15}O_7^-$, 343.1; found 343.0. HPLC k = 3.83, mp 204 °C, $[\alpha]_{\rm D}^{20}$ -477° (c = 0.2%, CHCl₃).

4.3 Chemistry

4.3.1 General procedure for the synthesis of the UA-based enamines 1, 3–6. A 30 mL mw vessel was charged with a suspension of (+)-(R)-UA or (-)-(S)-UA (1 equiv.) in absolute EtOH under nitrogen atmosphere. The suspension was stirred for 10 minutes at r.t., and the suitable amine (1 equiv.) and TEA (2 equiv.) were added dropwise. The reaction was heated under mw irradiation (200 W, 250 psi) at 90 °C for 5 minutes under vigorous stirring for three-times. The resulting suspension was filtered, and the filtrate was concentrated under reduced pressure. The residue was resuspended in water and the aqueous phase was washed three times with n-hexane to remove non-reacted UA and then extracted with DCM. The organic phase was dried over anhydrous Na₂SO₄, filtered, and concentrated. The crude was purified over silica gel.

Methyl (1-((R)-6-acetyl-7,9-dihydroxy-8,9b-dimethyl-1,3-dioxo-3,9b-dihydrodibenzo[b,d]furan-2(1H)-ylidene)ethyl)-ι-serinate ((9bR,15S)-1). The title compound was purified over silica gel, mobile phase DCM/MeOH, 95:5. White solid (50% yield). ¹H NMR (400 MHz, MeOD) δ 5.85 (s, 1H), 4.61 (s, 1H), 4.15 (dd, J = 11.5, 3.7 Hz, 2H), 3.96 (td, J = 10.9, 3.4 Hz, 2H), 3.86 (s, 3H), 2.69 (s, 3H), 2.65 (s, 3H), 2.06 (s, 5H), 1.74 (s, 3H), proton on heteroatoms exchange with the solvent. ¹³C NMR (176 MHz, CDCl₃) δ 200.77, 200.61, 198.93, 175.45, 174.90, 174.58, 168.86, 168.76, 163.94, 163.75, 158.20, 157.89, 155.90, 155.62, 108.51, 108.28, 104.50, 101.47, 101.31, 63.28, 62.95, 58.79, 58.31, 53.49, 31.33, 19.25, 7.55. ESI-MS (m/z): [M-H]⁻

calcd for $C_{22}H_{22}NO_9^-$, 444.13; found 444.08 and [M + Cl]⁻ calcd for $C_{22}H_{23}^{35}$ [Cl] NO_9^- , 480.11; found 480.78. HPLC-UV/vis: k = 2.23, $[\alpha]_D^{20}$ +264.00° (c 0.25, CHCl₃).

Methyl (1-((S)-6-acetyl-7,9-dihydroxy-8,9b-dimethyl-1,3-dioxo-3,9b-dihydrodibenzo[b,d]-furan-2(1H)-ylidene)ethyl)- ι -serinate ((9bS,15S)-1). The title compound was purified over silica gel, mobile phase DCM/MeOH, 95:5. White solid (37% yield). 1 H NMR (400 MHz, CDCl₃) δ 13.29 (s, 1H), 11.63 (s, 1H), 5.76 (s, 1H), 4.55 (dt, J = 8.1, 4.0 Hz, 2H), 4.20-4.08 (m, 1H), 4.03-3.99 (m, 1H), 3.79 (s, 3H), 2.61 (s, 3H), 2.57 (s, 3H), 2.03 (s, 3H), 1.65 (s, 3H), proton on enaminic NH and on serin hydroxyl exchange with the solvent. 13C NMR (176 MHz, CDCl₃) δ 200.81, 200.70, 199.02, 198.90, 175.41, 174.53, 168.86, 168.74, 163.85, 163.73, 158.26, 158.07, 155.94, 155.77, 108.29, 105.01, 104.75, 101.52, 63.14, 62.96, 58.53, 58.21, 53.58, 31.44, 19.17, 7.63. ESI-MS (m/z): $[M-H]^{-}$ calcd for C₂₂H₂₂NO₉, 444.13; found 444.10 and [M + Cl] calcd for C₂₂H₂₃³⁵[Cl]NO₉⁻, 480.11; found 480.51. HPLC-UV/vis: $k = 1.70, [\alpha]_D^{20} - 420.40^{\circ} (c \ 0.25, CHCl_3).$

(1-((R)-6-Acetyl-7,9-dihydroxy-8,9b-dimethyl-1,3-dioxo-3,9b-dihydrodibenzo[b,d]furan-2-(1H)-ylidene)ethyl)-ι-phenylalanine ((9bR,15S)-3). The title compound was purified over silica gel, mobile phase DCM/MeOH, 9:1. Brownish oil (87% yield). ¹H NMR (400 MHz, CDCl₃) δ, 13.57 (s, 1H), 13.36 (s, 1H), 12.16 (s, 1H), 7.47–6.84 (m, 5H), 5.76 (s, 1H), 4.53 (s, 1H), 3.42–3.40 (m, 1H), 3.12–3.05 (m, 1H), 2.66 (s, 3H), 2.16 (s, 3H), 2.07 (s, 3H), 1.66 (s, 3H), proton on enaminic NH exchanges with the solvent. ¹³C NMR (101 MHz, CDCl₃) δ 200.82, 197.94, 173.81, 171.26, 163.45, 158.52, 156.08, 136.80, 129.60, 128.71, 127.18, 107.78, 105.45, 102.90, 101.38, 60.49, 45.36, 31.36, 21.14, 18.70, 14.28, 8.61. ESI-MS (m/z): [M-H]⁻ calcd for C₂₇H₂₄NO₈⁻, 490.15; found 490.03 and [M + Cl]⁻ calcd for C₂₇H₂₄NO₈⁻, 526.13; found 526.46. HPLC-UV/vis: k = 2.84, α _D^{2C} +103.80° (c 0.25, CHCl₃).

(1-((S)-6-Acetyl-7,9-dihydroxy-8,9b-dimethyl-1,3-dioxo-3,9bdihydrodibenzo[b,d]furan-2-(1H)-ylidene)ethyl)-L-phenylalanine ((9bS,15S)-3). The title compound was purified over silica gel, mobile phase DCM/MeOH, 9:1. Brownish oil (51% yield). ¹H NMR (400 MHz, CDCl₃) δ 13.52 (s, 1H), 13.36 (s, 1H), 12.11 (s, 1H), 7.40-6.99 (m, 5H), 5.75 (s, 1H), 4.52 (td, J = 9.0, 3.8Hz, 1H), 3.42 (dd, J = 13.8, 4.0 Hz, 1H), 3.04 (d, J = 7.6 Hz, 1H), 2.67 (s, 3H), 2.18 (s, 3H), 2.08 (s, 3H), 1.67 (s, 3H), proton on enaminic NH exchanges with the solvent. ¹³C NMR (176 MHz, CDCl₃) δ 200.73, 198.35, 174.03, 163.63, 158.44, 156.01, 136.61, 129.75, 128.79, 127.36, 108.12, 105.22, 102.50, 101.46, 45.46, 40.49, 32.07, 31.38, 29.84, 29.50, 28.52, 22.83, 18.54, 14.26, 8.66, 7.62, 1.16. ESI-MS (m/z): $[M-H]^-$ calcd for C₂₇H₂₄NO₈⁻, 490.15; found 490.14 and [M + Cl]⁻ calcd for $C_{27}H_{25}^{35}$ [Cl]NO₈, 526.13; found 526.91. HPLC-UV/vis: k =2.90, $[\alpha]_D^{20}$ -267.15° (c 0.25, CHCl₃).

(1-((R)-6-Acetyl-7,9-dihydroxy-8,9b-dimethyl-1,3-dioxo-3,9b-dihydrodibenzo[b,d]furan-2-(1H)-ylidene)ethyl)-L-tyrosine ((9bR,15S)-4). The title compound was purified over silica gel, mobile phase DCM/MeOH, 9:1. White solid (48% yield). ¹H NMR (400 MHz, CDCl₃) δ 13.36 (s, 1H), 12.19 (s, 1H), 7.01 (d, J = 8.4 Hz, 2H), 6.69 (d, J = 8.4 Hz, 2H), 5.77 (s, 1H), 4.47 (td,

J = 8.2, 4.5 Hz, 1H), 3.25 (dd, J = 13.8, 4.4 Hz, 1H), 3.04 (dd, J = 13.8, 8.2 Hz, 1H), 2.66 (s, 3H), 2.28 (s, 3H), 2.08 (s, 3H), 1.67 (s, 3H), protons on enaminic NH, carboxylic acid and phenol of tyrosine exchange with the solvent. ¹³C NMR (101 MHz, DMSO) δ 200.98, 196.84, 188.18, 172.55, 162.48, 157.82, 156.05, 155.84, 130.38, 129.17, 128.23, 127.30, 115.09, 106.20, 105.27, 102.59, 101.40, 100.84, 56.03, 45.31, 31.76, 31.07, 18.61, 8.84, 7.52. ESI-MS (m/z): [M-H]⁻ calcd for $C_{27}H_{24}NO_9^-$, 506.15; found 506.64, [M + Cl]⁻ calcd for $C_{27}H_{25}^{35}$ [Cl]NO9⁻, 542.12; found 542.91 and $C_{27}H_{25}^{37}$ [Cl]NO9⁻, 544.12; found 544.05. HPLC-UV/vis: k = 1.81, [α]_D²⁰ +80.35° (c 0.2, CHCl₃).

(1-((S)-6-Acetyl-7,9-dihydroxy-8,9b-dimethyl-1,3-dioxo-3,9bdihydrodibenzo[b,d]furan-2-(1H)-ylidene)ethyl)-L-tyrosine ((9bS,15S)-4). The title compound was purified over silica gel, mobile phase DCM/MeOH, 9:1 + 0.1% NH₃ (in MeOH). Pale yellow solid (54% yield). ¹H NMR (400 MHz, MeOD) δ 7.08 (d, J = 8.4 Hz, 2H, 6.70 (d, J = 8.4 Hz, 2H), 5.76 (s, 1H), 4.57-4.54(m, 1H), 3.33-3.31 (m, 1H, under MeOD), 2.96 (dd, J = 13.9, 9.3 Hz, 1H), 2.65 (s, 3H), 2.23 (s, 3H), 2.01 (s, 3H), 1.66 (s, 3H), all the protons on heteroatoms exchange with the solvent. 13 C NMR (101 MHz, DMSO) δ 201.00, 197.43, 188.82, 173.92, 172.80, 170.81, 162.68, 156.30, 155.81, 133.15, 130.55, 126.28, 115.17, 106.29, 105.17, 102.43, 101.60, 100.89, 56.30, 45.35, 31.64, 31.08, 18.40, 8.50, 7.53. ESI-MS (m/z): $[M-H]^{-1}$ calcd for C₂₇H₂₄NO₉, 506.15; found 506.29 and [M + Cl] for C₂₇H₂₅³⁵[Cl]NO₉, 542.12; found HPLC-UV/vis: k = 1.56, $[\alpha]_D^{20} - 176.5^{\circ}$ (c 0.25, CHCl₃).

(*R*)-6-Acetyl-7,9-dihydroxy-8,9*b*-dimethyl-2-(1-(((*R*)-1-phenylethyl)amino)ethylidene) dibenzo[*b*,d]furan-1,3(2*H*,9*bH*)-dione ((9*bR*,15*R*)-5). The title compound was purified over silica gel, mobile phase DCM/MeOH, 95:5. Yellow solid (61% yield). ¹H NMR (400 MHz, MeOD) δ 7.49–7.36 (m, 5H), 5.86 (s, 1H), 5.19 (q, *J* = 6.7 Hz, 1H), 2.68 (s, 3H), 2.59 (s, 3H), 2.05 (s, 3H), 1.71 (s, 3H), 1.68 (d, *J* = 6.7 Hz, 3H), all the protons on heteroatoms exchange with the solvent. ¹³C NMR (176 MHz, CDCl₃) δ 200.78, 198.64, 191.02, 174.41, 163.60, 158.37, 155.98, 141.74, 129.44, 128.25, 125.77, 108.10, 105.17, 102.43, 102.24, 101.47, 57.49, 54.46, 31.98, 31.39, 24.26, 18.96, 7.59. ESI-MS (*m*/*z*): [M-H] calcd for C₂₆H₂₄NO₆ , 446.16; found 446.34, [M + Cl] calcd for C₂₆H₂₅³⁵[Cl]NO₆ , 482.14; found 482.28 and calcd for C₂₆H₂₅³⁷[Cl]NO₆ , 484.14; found 484.51. HPLC-UV/vis: k = 3.94, [α]²⁰ +186.3° (c 0.5, CHCl₃).

(S)-6-Acetyl-7,9-dihydroxy-8,9b-dimethyl-2-(1-(((R)-1-phenylethyl)amino)ethylidene) dibenzo[b,d]furan-1,3(2H,9bH)-dione ((9bS,15R)-5). The title compound was purified over silica gel, mobile phase DCM/MeOH, 95:5. Yellowish oil (58% yield). 1 H NMR (400 MHz, MeOD) δ 7.38–7.22 (m, 5H), 5.74 (s, 1H), 5.08 (q, J = 6.7 Hz, 1H), 2.57 (s, 3H), 2.47 (s, 3H), 1.93 (s, 3H), 1.59 (s, 3H), 1.56 (d, J = 6.8 Hz, 3H), all the protons on heteroatoms exchange with the solvent. 13 C NMR (101 MHz, CDCl₃) δ 200.85, 198.88, 192.39, 188.68, 174.92, 163.66, 159.34, 156.18, 142.42, 130.33, 128.30, 125.81, 108.19, 105.21, 102.48, 101.54, 54.50, 32.01, 31.44, 24.31, 18.99, 7.62, 1.16. ESI-MS (m/z): [M-H]⁻ calcd for $C_{26}H_{25}^{35}$ [Cl]NO₆⁻, 482.14; found 482.47 and calcd for

 $C_{26}H_{25}^{37}[Cl]NO_6^-$, 484.14; found 484.48. HPLC-UV/vis: k = 4.05, $[\alpha]_D^{20}$ –96.5° (c 0.25, CHCl₃).

Research Article

(R)-6-Acetyl-2-(1-((3-chlorobenzyl)amino)ethylidene)-7,9dihydroxy-8,9b-dimethyldibenzo [b,d]furan-1,3(2H,9bH)-dione ((R)-6). The title compound was purified over silica gel, mobile phase DCM/MeOH, 9:1 + 0.1% NH₃ (in MeOH). Yellow solid (47% yield). 1 H NMR (400 MHz, CDCl₃) δ 13.88 (s, 1H), 13.36 (s, 1H), 11.88 (s, 1H), 7.37-7.32 (m, 2H), 7.29 (s, 1H), 7.23-7.14 (m, 1H), 5.81 (s, 1H), 4.66 (d, J = 5.7 Hz, 2H), 2.68 (s, 3H), 2.65 (s, 3H), 2.10 (s, 3H), 1.72 (s, 3H). ¹³C NMR (176 MHz, CDCl₃) δ 200.82, 198.80, 191.10, 175.43, 174.69, 163.69, 158.38, 155.98, 137.14, 135.39, 130.78, 128.87, 127.60, 125.42, 108.29, 105.12, 102.61, 102.43, 101.55, 57.63, 47.31, 32.05, 29.85, 18.72, 7.64. ESI-MS (m/z): $[M-H]^-$ calcd for $C_{25}H_{21}^{35}[Cl]NO_6^-$, 466.11; found 466.03 and calcd for $C_{25}H_{21}^{37}$ [Cl]NO₆, 468.11; found 468.19. HPLC-UV/vis: k =4.07, $[\alpha]_D^{20}$ +41.30° (c 0.5, CHCl₃).

(S)-6-Acetyl-2-(1-((3-chlorobenzyl)amino)ethylidene)-7,9dihydroxy-8,9b-dimethyldibenzo [b,d]furan-1,3(2H,9bH)-dione ((S)-6). The title compound was purified over silica gel, mobile phase DCM/MeOH, 9:1 + 0.1% NH₃ (in MeOH). Yellow solid (51% yield). ¹H NMR (400 MHz, CDCl₃) δ 13.88 (s, 1H), 13.36 (s, 1H), 11.88 (s, 1H), 7.37-7.32 (m, 2H), 7.29 (s, 1H), 7.23-7.14 (m, 1H), 5.81 (s, 1H), 4.66 (d, J = 5.7 Hz, 2H), 2.68 (s, 3H), 2.65 (s, 3H), 2.10 (s, 3H), 1.72 (s, 3H). ¹³C NMR (176 MHz, CDCl₃) δ 200.82, 198.80, 191.10, 175.43, 174.69, 163.69, 158.38, 155.98, 137.14, 135.39, 130.78, 128.87, 127.60, 125.42, 108.29, 105.12, 102.61, 102.43, 101.55, 57.63, 47.31, 32.05, 29.85, 18.72, 7.64. ESI-MS (m/z): $[M-H]^-$ calcd for $C_{25}H_{21}^{35}[Cl]NO_6^-$, 466.11; found 466.08 and calcd for $C_{25}H_{21}^{37}$ [Cl]NO₆, 468.11; found 468.22. HPLC-UV/vis: k =4.07, $[\alpha]_D^{20}$ -41.10° (c 0.5, CHCl₃).

(1-((R)-6-Acetyl-7,9-dihydroxy-8,9b-dimethyl-1,3-dioxo-3,9b-dimethyl-1,3-diodihydrodibenzo[b,d]furan-2(1H)-ylidene)ethyl)-L-arginine ((9bR, 15S)-2). A suspension of (+)-(R)-UA (50 mg, 0.15 mmol), L-arginine (25 mg, 0.15 mmol), and TEA (41 μl, 0.30 mmol) in EtOH (1.5 mL, 0.1 M) was refluxed at 80 °C for 14 h under N₂. The crude was purified by crystallisation with diisopropyl ether at 0 °C, affording the title compound as pale yellow solid (35 mg, 47%). ¹H NMR (400 MHz, DMSO- d_6) δ 13.39 (s, 1H), 13.27 (s, 1H), 12.48 (s, 1H), 8.90 (s, 1H), 7.43 (s, 4H), 5.71 (s, 1H), 4.25 (s, 1H), 3.15-3.08 (m, 2H), 2.59 (s, 3H), 2.57 (s, 3H), 1.94 (s, 3H), 1.87-1.76 (m, 2H), 1.59 (s, 3H), 1.55-1.47 (m, 2H). ¹³C NMR (176 MHz, DMSO- d_6) δ 200.67, 197.02, 188.47, 172.29, 172.12, 172.05, 162.62, 157.73, 157.10, 155.67, 106.18, 105.15, 102.27, 101.59, 100.69, 58.25, 56.22, 40.33, 31.96, 30.94, 30.08, 24.85, 18.95, 7.47. ESI-MS (m/z): $[M + H]^{+}$ calcd for C₂₄H₂₉N₄O₈⁺, 501.20; found 501.27 and [M-H] calcd for $C_{24}H_{27}N_4O_8$, 499.18; found 499.31. HPLC-UV/vis: k= 0.39, $[\alpha]_D^{20}$ +255.0° (c 0.2%, DMSO).

Synthesis of (1-((S)-6-Acetyl-7,9-dihydroxy-8,9b-dimethyl-1,3-dioxo-3,9b-dihydrodibenzo[b,d]furan-2(1H)-ylidene)ethyl)-Larginine ((9bS,15S)-2). A suspension of (+)-(S)-UA (22 mg, 0.065 mmol), L-arginine (12 mg, 0.065 mmol), and TEA (18 μ l, 0.13 mmol) in EtOH (1.5 mL) was refluxed at 80 °C for 14 h under nitrogen atmosphere. The solvent was removed under

reduced pressure and the crude was purified by crystallisation with diisopropyl ether at 0 °C, affording the title compound as pale yellow solid (15 mg, 46% yield). 1 H NMR (400 MHz, DMSO- d_{6}) δ 13.06 (bs, 1H), 9.10 (bs, 1H), 7.59 (bs, 6H), 5.82 (s, 1H), 4.22 (t, J = 5.8 Hz, 1H), 3.15–3.07 (m, 2H), 2.63 (s, 3H), 2.54 (s, 3H), 1.96 (s, 3H), 1.87–1.76 (m, 2H), 1.65 (s, 3H), 1.58–1.50 (m, 2H). 13 C NMR (176 MHz, MeOD) δ 200.90, 198.13, 189.61, 174.31, 173.98, 163.07, 158.05, 157.26, 156.12, 107.09, 105.12, 102.10, 102.01, 100.92, 58.84, 56.73, 40.66, 31.01, 30.02, 29.78, 24.62, 17.90, 6.21. ESI-MS (m/z): $[M+H]^+$ calcd for $C_{24}H_{29}N_4O_8^+$, 501.20; found 501.48 and $[M-H]^-$ calcd for $C_{24}H_{27}N_4O_8^-$, 499.18; found 499.03. HPLC-UV/vis: k = 0.28, $[\alpha]_D^{20}$ –182.4° (c 0.2%, DMSO).

4.3.2 Synthesis of the lateral chains 10-12

20-Amino-3,6,9,12,15,18-hexaoxaicosanoic acid (14). To a well-stirred solution of Boc-NH-PEG₆-acid (13, 100 mg, 0.22 mmol, 1 equiv.) in DCM (1 mL), TFA was added dropwise (422 μl, 5.1 mmol, 5 equiv.) at 0 °C and the reaction was allowed to stir at r.t. for 14 h. The solvent was evaporated, and the residue was diluted with MeOH followed by the treatment with Amberlyst-15 ion-exchange acid resin for 5 h. Upon filtration, the resin was collected and swelled by alternate washing with DCM and MeOH. The resin was suspended in 50 mL of methanol in presence of 0.1% NH₃ in MeOH and the resulting solution was shaken for 14 h. The suspension was filtrated and the concentrated to give the titled compound as a brownish oil (80 mg, quantitative yield). ¹H NMR (400 MHz, CDCl₃) δ 3.62 (t, J = 10.2 Hz, 20H), 2.55 (t, J = 6.5 Hz, 4H), 1.06 (t, J = 6.5 Hz, 2H).

Methyl 20-amino-3,6,9,12,15,18-hexaoxaicosanoate (10). To a solution of 14 (80 mg, 0.22 mmol, 1 equiv.) in MeOH (2.2 mL), 1 N H₂SO₄ (4 drops) was added dropwise under stirring. The mixture was heated three times under mw irradiation (200 W, 250 psi) at 120 °C for 3 minutes. Upon completion of the reaction, the mixture was treated with Amberlyst-15 ionexchange acid resin and shaken for 5 h. After filtration, the resin was collected and swelled by alternating washing with DCM and MeOH. The resin was suspended in 50 mL of methanol in presence of 0.1% NH3 in MeOH and was shaken for 14 h. The suspension was filtrated and the concentrated to give the titled compound as a brownish oil (35 mg, 42% yield). ¹H NMR (400 MHz, CDCl₃) δ 3.75 (t, J = 6.4 Hz, 2H), 3.70-3.55 (m, 19H), 2.98 (t, J = 6.5 Hz, 2H), 2.60 (t, J = 6.5 Hz, 2H), 2.12-1.95 (br, 4H), 1.31-1.16 (m, 2H). 2.07-1.90 (br, 2H), 0.94-0.78 (m, 4H).

2,2-Dimethyl-4,9-dioxo-3,11,14,17-tetraoxa-5,8-diazanonadecan-19-oic acid (17). To a solution of 3,6,9-trioxaundecanedioic acid (16, 139 mg, 0.624 mmol, 2 equiv.) in anhydrous DMF (3.12 mL), EDC-HCl (72 mg, 0.374 mmol, 1.2 equiv.), HOBt (21 mg, 0.156 mmol, 0.5 equiv.) and DIPEA (273 μ L, 1.56 mmol, 5 equiv.) were added, and the resulting solution was stirred at r.t. under nitrogen atmosphere for 30 minutes. N-Boc-ethylenediamine (15, 49 μ L, 0.312 mmol, 1.0 equiv.) was added, and the reaction mixture was stirred for additional 6 h. n-Heptane was added to form the azeotrope with DMF, and the solvent was removed under reduced

pressure. The crude was purified on silica gel (mobile phase, DCM/MeOH + 0.1% HCOOH, from 40:1 to 20:1), to give the title compound as viscous clear oil (84 mg, 70% yield). ¹H NMR (400 MHz, CDCl₃) δ 7.61 (t, J = 6.1 Hz, 1H), 5.38 (s, 1H), 4.08 (s, 4H), 3.96 (d, J = 5.2 Hz, 2H), 3.72–3.58 (m, 6H), 3.38–3.29 (m, 2H), 3.24–3.15 (m, 2H), 1.35 (s, 9H).

Methyl 1-amino-4-oxo-6,9,12-trioxa-3-azatetradecan-14-oate hydrochloride (11). To a solution of 17 (84 mg, 0.215 mmol, 1 equiv.) in MeOH (2 mL), TMSCl (72 mg, 0.374 mmol, 1.2 equiv.) was added dropwise at 0 °C. The reaction was allowed to stir at r.t. for 6 h. The solvent was removed in vacuo and the residue was triturated with n-hexane and Et_2O , to give the title compound as light-yellow oil (quantitative yield). ¹H NMR (400 MHz, MeOD) δ 4.23–4.20 (m, 4H), 3.77–3.77 (m, 5H), 3.76–3.71 (m, 6H), 3.71–3.67 (m, 4H).

tert-Butyl (3-hydroxypropyl)carbamate (19). To a solution of 18 in anhydrous DCM, Boc₂O was added at r.t., and the resulting mixture was stirred in the same conditions for 3 h. Upon completion of the reaction, the solvent was removed *in vacuo* and the crude was filtered through a small plug of silica gel eluting with a DCM/MeOH, 20:1 to give the desired product as transparent oil (quantitative yield). ¹H NMR (400 MHz, CDCl₃) δ 4.87 (s, 1H), 3.59 (q, J = 5.4 Hz, 2H), 3.21 (q, J = 6.3 Hz, 2H), 1.60 (p, J = 6.0 Hz, 2H), 1.37 (s, 9H).

tert-Butyl (3-(hexadecyloxy)propyl)carbamate (20). To a solution of **19** (500 mg, 1.84 mmol, 1 equiv.) in anhydrous DMF (10 mL) at 0 °C, 60% NaH in mineral oil was added (66 mg, 2.75 mmol, 1.5 equiv.). After 1 h, 1-bromohexadecane (263 µL, 2.21 mmol, 1.2 equiv.) was added, and the resulting mixture was allowed to stir at r.t. for 16 h. The reaction was quenched with a saturated NH₄Cl solution (20 mL) and then extracted with EtOAc. The combined organic layers were dried over anhydrous Na₂SO₄, filtered and concentrated. The crude was purified on silica gel (petroleum ether/EtOAc, from 6:1 to 5:1) to afford the title compound as orange oil (497 mg, 75% yield). ¹H NMR (400 MHz, CDCl₃) δ 4.93–4.89 (bs, 1H), 3.40 (t, J = 5.9 Hz, 2H), 3.32 (t, J = 6.6 Hz, 2H), 3.15 (q, J = 6.3 Hz, 2H), 1.67 (p, J = 6.2 Hz, 2H), 1.49 (dt, J = 14.7, 7.1 Hz, 2H), 1.37 (s, 9H), 1.22–1.14 (m, 26H), 0.81 (t, J = 6.8 Hz, 3H).

3-(Hexadecyloxy)propan-1-aminium 2,2,2-trifluoroacetate (12). A solution of 20 (99 mg, 0.248 mmol, 1 equiv.) in anhydrous DCM (1.24 mL) was cooled to 0 °C, and TFA was added dropwise under nitrogen atmosphere. The reaction mixture was allowed to stir 0 °C for 2 h, until the consumption of the starting material. The solvent was removed *in vacuo*, affording the title compound as yellow solid (quantitative yield). ¹H NMR (400 MHz, CDCl₃) δ 10.79–10.39 (bs, 1H), 7.66–7.31 (bs, 2H), 3.56 (t, J = 5.2 Hz, 2H), 3.36 (t, J = 6.9 Hz, 2H), 3.13 (s, 2H), 1.87 (p, J = 5.2 Hz, 2H), 1.48 (p, J = 6.6 Hz, 2H), 1.28–1.15 (m, 26H), 0.80 (t, J = 6.8 Hz, 3H).

4.3.3 General procedure for the synthesis of the UA-based enamines 7–9. (9b*R*,15*S*)-4 or (9b*S*,15*S*)-4 (1 equiv.) was dissolved in anhydrous DMF under nitrogen atmosphere, and EDC-HCl (1.5 equiv.), HOBt (0.1 equiv.), DIPEA (4.0 equiv.) and the appropriate amine (1.0–1.5 equiv.) were

sequentially added at 0 °C. The mixture was stirred at r.t. for 14 h and quenched by the addiction of saturated NH₄Cl solution. The aqueous phase was extracted with EtOAc. The combined organic layers were washed with saturated NH₄Cl solution, saturated NaHCO₃ solution, and brine. The organic phase was dried over anhydrous NaSO₄, filtered, and concentrated. The crude was purified on silica gel.

Methyl (S,E)-2-((R)-6-acetyl-7,9-dihydroxy-8,9b-dimethyl-1,3dioxo-3,9b-dihydrodibenzo[b,d]furan-2(1H)-ylidene)-4-(4hydroxybenzyl)-5-oxo-9,12,15,18,21,24-hexaoxa-3,6diazahexacosan-26-oate ((9bR,15S)-7). The title compound was purified over silica gel, mobile phase DCM/MeOH, 9:1 + 0.1% NH₃ (in MeOH). Yellow solid (56% yield). ¹H NMR (400 MHz, CDCl₃) δ 13.83 (s, 1H), 13.30 (s, 1H), 11.89 (s, 1H), 7.03 (d, J = 8.1 Hz, 2H), 6.76 (d, J = 8.1 Hz, 2H), 6.55 (s, 1H), 5.75(s, 1H), 4.38 (d, J = 7.1 Hz, 1H), 3.67 (t, J = 6.4 Hz, 2H), 3.64– 3.51 (m, 25H), 3.50–3.35 (m, 2H), 3.04 (d, J = 7.0 Hz, 2H), 2.61 (s, 3H), 2.42 (s, 3H), 2.03 (s, 3H), 1.63 (s, 3H), proton on enaminic NH exchanges with the solvent. ¹³C NMR (101 MHz, MeOD) δ 205.51, 201.63, 198.54, 189.78, 172.44, 167.61, 163.90, 156.59, 155.93, 131.22, 126.16, 112.08, 109.31, 103.54, 101.85, 100.93, 99.71, 70.20, 70.14, 70.05, 69.99, 69.85, 68.89, 66.23, 59.24, 56.98, 50.73, 39.21, 34.31, 31.35, 30.87, 30.03, 29.26, 22.30, 17.61, 13.02, 6.20. ESI-MS (m/z): $[M + Na]^+$ calcd for C₄₂H₅₄N₂NaO₁₆⁺, 865.34; found 865.39 and [M-H]⁻ calcd for $C_{42}H_{53}N_2O_{16}^-$, 841.34; found 841.44. HPLC-UV/vis: k =2.11, $[\alpha]_D^{20}$ +42.30° (c 0.2, CHCl₃).

Methyl (S,E)-2-((S)-6-acetyl-7,9-dihydroxy-8,9b-dimethyl-1,3dioxo-3,9b-dihydrodibenzo[b,d]furan-2(1H)-ylidene)-4-(4hydroxybenzyl)-5-oxo-9,12,15,18,21,24-hexaoxa-3,6diazahexacosan-26-oate ((9bS,15S)-7). The title compound was purified over silica gel, mobile phase DCM/MeOH, 9:1 + 0.1% NH₃ (in MeOH). Yellow solid (49% yield). ¹H NMR (400 MHz, MeOD) δ 7.09 (d, J = 8.5 Hz, 2H), 6.73 (d, J = 8.5 Hz, 2H), 5.81 (s, 1H), 4.72 (dd, J = 8.5, 5.6 Hz, 1H), 4.01 (s, 2H), 3.71 (t, J = 6.2 Hz, 2H), 3.66 (s, 3H), 3.65-3.54 (m, 20H), 3.45-3.36 (m, 2H), 3.18 (dd, J = 13.8, 5.6 Hz, 1H), 3.00 (dd, J = 13.7, 8.6 Hz, 1H), 2.65 (s, 3H), 2.34 (s, 3H), 2.02 (s, 3H), 1.67 (s, 3H), all the protons on heteroatoms exchange with the solvent. 13 C NMR (176 MHz, MeOD) δ 200.90, 198.66, 174.59, 174.12, 173.83, 172.42, 169.94, 163.10, 157.99, 156.60, 156.09, 130.47, 129.16, 129.06, 128.09, 126.07, 115.14, 107.22, 104.98, 101.88, 100.91, 70.13, 69.84, 68.88, 66.28, 59.29, 50.73, 39.20, 38.98, 38.82, 34.31, 33.35, 30.74, 30.03, 29.36, 28.69, 24.15, 23.52, 22.62, 17.44, 10.02, 6.19. ESI-MS (m/z): $[M + Na]^+$ calcd for $C_{42}H_{54}N_2NaO_{16}^+$, 865.34; found 865.13 and $[M-H]^-$ calcd for $C_{42}H_{53}N_2O_{16}^-$, 841.34; found 841.25. HPLC-UV/vis: k =1.81, $[\alpha]_D^{20}$ -48.30° (c 0.2, CHCl₃).

Methyl (*S,E*)-2-((*R*)-6-acetyl-7,9-dihydroxy-8,9b-dimethyl-1,3-dioxo-3,9b-dihydrodibenzo[b,d]furan-2(1H)-ylidene)-4-(4-hydroxybenzyl)-5,10-dioxo-12,15,18-trioxa-3,6,9-triazaicosan-20-oate ((9bR,15S)-8). The title compound was purified over silica gel, mobile phase DCM/MeOH, 9:1 + 0.1% NH₃ (in MeOH). Yellow solid (10% yield). ¹H NMR (400 MHz, CDCl₃) δ 13.82 (s, 1H), 13.37 (s, 1H), 11.86 (s, 1H), 7.06 (d, J = 7.6 Hz, 2H), 6.81 (d, J = 7.6 Hz, 2H), 5.82 (s, 1H), 4.45 (s, 1H), 4.17 (s,

2H), 3.85–3.65 (m, 13H), 3.46–3.30 (m, 4H), 3.03–2.98 (m, 2H), 2.68 (s, 3H), 2.49 (s, 3H), 2.10 (s, 3H), 1.71 (s, 3H), proton on enaminic NH exchanges with the solvent. ¹³C NMR (101 MHz, CDCl₃) δ 200.86, 198.62, 172.24, 171.04, 171.00, 169.48169.38, 167.98, 167.54, 163.61, 158.34, 156.05, 131.02, 130.78, 128.94, 126.53, 115.93, 108.09, 105.23, 101.51, 60.23, 52.16, 52.00, 41.80, 39.04, 38.87, 38.41, 32.01, 30.50, 29.06, 23.88, 23.11, 18.86, 14.18, 11.09, 7.62. ESI-MS (m/z): [M + Na]⁺ calcd for C₃₈H₄₅N₃NaO₁₄⁺, 790.28; found 790.07 and [M–H]⁻ calcd for C₃₈H₄₄N₃O₁₄⁻, 766.28; found 766.36. HPLC-UV/vis: k = 1.65, [α]²⁰ +172.69° (c 0.3, CHCl₃).

Methyl (S,E)-2-((S)-6-acetyl-7,9-dihydroxy-8,9b-dimethyl-1,3dioxo-3,9b-dihydrodibenzo[b,d]furan-2(1H)-ylidene)-4-(4hydroxybenzyl)-5,10-dioxo-12,15,18-trioxa-3,6,9-triazaicosan-20oate ((9bS,15S)-8). The title compound was purified over silica gel, mobile phase DCM/MeOH, 9:1 + 0.1% NH₃ (in MeOH). Yellow solid (10% yield). ¹H NMR (400 MHz, MeOD) δ 7.09 (d, J = 8.5 Hz, 2H), 6.74 (d, J = 8.5 Hz, 2H), 5.79 (s, 1H), 4.70 (dd, J = 8.8, 5.3 Hz, 1H), 4.15 (s, 2H), 3.98 (s, 2H), 3.73(s, 3H), 3.71-3.63 (m, 8H), 3.41-3.37 (m, 4H), 3.21 (dd, J =13.8, 5.2 Hz, 1H), 3.00 (dd, J = 13.8, 8.8 Hz, 1H), 2.65 (s, 3H), 2.31 (s, 3H), 2.02 (s, 3H), 1.67 (s, 3H), all the protons on heteroatoms exchange with the solvent. ¹³C NMR (101 MHz, MeOD) δ 200.84, 198.68, 189.94, 174.54, 174.02, 171.94, 171.34, 170.27, 163.11, 157.92, 156.60, 156.01, 130.45, 126.13, 115.13, 107.22, 104.91, 102.19, 101.85, 100.87, 70.57, 70.46, 70.11, 69.97, 69.75, 67.69, 59.42, 57.10, 50.86, 39.01, 38.75, 38.22, 30.78, 30.03, 17.39, 6.21. ESI-MS (m/z): $[M + Na]^+$ calcd for C₃₈H₄₅N₃NaO₁₄⁺, 790.28; found 790.41 and [M-H]⁻ calcd for $C_{38}H_{44}N_3O_{14}$, 766.28; found 766.62. HPLC-UV/vis: k =1.39, $[\alpha]_{\rm D}^{20}$ -180.85° (c 0.3, CHCl).

(S)-2-(((E)-1-((R)-6-Acetyl-7,9-dihydroxy-8,9b-dimethyl-1,3dioxo-3,9b-dihydrodibenzo[b,d]furan-2(1H)-ylidene)ethyl)amino)-N-(3-(hexadecyloxy)propyl)-3-(4-hydroxyphenyl)propanamide ((9bR,15S)-9). The title compound was purified over silica gel, mobile phase DCM/MeOH, 9:1 + 0.1% NH₃ (in MeOH). Colourless oil (25% yield). 1 H NMR (400 MHz, CDCl₃) δ 13.87 (s, 1H), 13.37 (s, 1H), 11.81 (s, 1H), 7.10 (d, J = 7.8 Hz, 2H), 6.80 (d, J = 7.6 Hz, 2H), 6.50 (s, 1H), 5.84 (s, 1H), 4.38 (s, 1H),3.48 (t, J = 5.3 Hz, 2H), 3.40–3.32 (m, 4H), 3.27–3.20 (m, 1H), 3.12-3.06 (m, 1H), 2.69 (s, 3H), 2.43 (s, 3H), 2.11 (s, 3H), 1.82-1.72 (m, 5H), 1.37-1.20 (m, 28H), 0.90 (t, J = 5.3 Hz, 3H), proton on enaminic NH exchanges with the solvent. ¹³C NMR (101 MHz, CDCl₃) δ 200.83, 198.72, 168.51, 163.70, 158.29, 155.96, 155.55, 132.84, 131.04, 130.74, 128.95, 127.01, 116.01, 108.29, 104.57, 101.54, 71.58, 70.30, 68.33, 60.76, 39.40, 39.05, 38.88, 32.07, 31.43, 30.51, 29.85, 29.80, 29.64, 29.50, 29.07, 28.75, 26.28, 23.89, 23.13, 22.83, 18.88, 14.26, 14.19, 11.10, 7.63, 1.16. ESI-MS (m/z): $[M + Na]^+$ calcd for C₄₆H₆₄N₂NaO₉, 811.45; found 811.94 and [M-H] calcd for $C_{46}H_{63}N_2O_9$, 787.45; found 787.12. HPLC-UV/vis: k =5.40, $[\alpha]_{D}^{20}$ +116.93° (c 0.3, CHCl₃).

(S)-2-(((E)-1-((S)-6-Acetyl-7,9-dihydroxy-8,9b-dimethyl-1,3-dioxo-3,9b-dihydrodibenzo[b,d]furan-2(1H)-ylidene)ethyl)amino)-N-(3-(hexadecyloxy)propyl)-3-(4-hydroxyphenyl)propanamide ((9bS,15S)-9). The title compound was purified over silica gel,

mobile phase DCM/MeOH, 9:1 + 0.1% NH₃ (7 M in MeOH). White solid (11% yield). ¹H NMR (400 MHz, MeOD) δ 7.10 (d, J = 8.4 Hz, 2H, 6.75 (d, J = 8.3 Hz, 2H), 5.84 (s, 1H), 4.69 (t, J= 7.2 Hz, 1H), 3.41-3.38 (m, 6H, under MeOD), 3.22-3.13 (m, 1H), 3.01 (dd, J = 13.8, 8.3 Hz, 1H), 2.68 (s, 3H), 2.36 (s, 3H), 2.05 (s, 3H), 1.76-1.70 (m, 5H), 1.33-1.24 (m, 28H), 0.97 (t, I =7.5 Hz, 3H), all the protons on heteroatoms exchange with the solvent. 13 C NMR (100 MHz, CDCl₃) δ 200.75, 190.65, 179.76, 175.24, 168.74, 167.77, 163.75, 158.29, 156.20, 155.43, 132.61, 131.03, 130.87, 128.95, 115.98, 107.83, 104.97, 101.53, 71.64, 70.44, 68.32, 60.93, 39.54, 38.88, 32.07, 31.43, 30.51, 29.84, 29.81, 29.78, 29.66, 29.51, 29.08, 28.75, 26.24, 23.90, 23.13, 22.83, 18.99, 14.26, 14.19, 11.11, 7.63. ESI-MS (*m/z*): $[M + Na]^+$ calcd for $C_{46}H_{64}N_2NaO_9$, 811.45; found 812.03 and [M-H] calcd for C₄₆H₆₃N₂O₉, 787.45; found 787.21. HPLC-UV/vis: k = 4.69, $[\alpha]_D^{20}$ -163.32° (c 0.25, CHCl₃).

4.4 Water solubility determination

Thermodynamic aqueous solubility was determined following the previously reported procedure. ⁵⁹ Excess solid (about 10–20 mg) of each compound was suspended in 5 mL of ultrapure water for HPLC in a volumetric bottle and magnetically stirred (1500 rpm) for 24 hours at 25 \pm 1 °C.

The resulting suspension was filtered on a Nylon Syringe Filters (13 mm, 0.45 µm pore size) and the concentration of the compound in solution was quantified by HPLC-UV/PDA on a Jasco (Tokyo, Japan) system, consisting of a PU-1580 pump and a MD-1510 photodiode array (PDA) detector. Chromatogram acquisitions and elaborations were performed using the ChromNAV software (Tokyo, Japan). Analyses were carried out on a XBridgeTM Phenyl, (4.6 × 150 mm, 5 μm) column, at room temperature. The mobile phases were A: water containing 0.1% of formic acid, and B: acetonitrile containing 0.1% of formic acid. Elution was performed on a linear gradient from 50% to 100% B over 10 min, followed by an isocratic hold at 100% B for 3 min. The flow rate was 1.0 mL min⁻¹ and the injection volume 10 μL. Standard calibration curves of (+)-(R)-UA (from 0.073 mM to 2.904 mM, $R^2 = 0.998$) (Fig. S3†) was determined. Since all the derivatives share the same chromophore, the same response factor (at $\lambda = 308$ nm, the relative maximum absorption peak) was applied. Limits of detections (LOD) and quantification (LOQ) were 0.54 µM and LOQ 61.13 µM, respectively. Solubility was determined from the mean peak areas of duplicate injections.

4.5 In vitro assay

4.5.1 Microorganisms. Antifungal activity was tested against *Candida albicans* ATCC 10231, *Candida tropicalis* ATCC 750, and *Trichophyton rubrum* LM 237. The strain coded with ATCC was obtained from American Type Culture Collection (https://www.atcc.org/), while the *T. rubrum* coded LM belongs to the collection of the Mycology Laboratory of the University of Pavia and was isolated from patients

RSC Medicinal Chemistry

affected by fungal infection. All these fungi are well known to be human pathogenic agents.

4.5.2 Evaluation of minimum inhibitory concentration (MIC) and minimum fungicidal concentration (MFC). The minimum inhibitory concentration (MIC) was defined as the lowest substance concentration, which completely inhibited visible microbial growth. The minimum fungicidal concentration (MFC) corresponded to the lowest concentration resulting in a reduction of the initial inoculum by more than 99.9%.

The antifungal activity was evaluated by microdilution method using 96 microwell plates (Microtiter®), according to Clinical and Laboratory Standards Institute (CLSI; formerly NCCLS) procedures, which considers this method the best for antifungal susceptibility testing of yeasts and filamentous fungi. 61,62

Both *C. albicans*, *C. tropicalis* and *T. rubrum* were cultured on Sabouraud dextrose agar (SDA) (Merck KGaA, Darmstadt, Germany) at 27 °C for one week before microdilution tests. Dermatophytes suspension was performed by collecting actively growing mycelium in test tubes containing broken coverslips and 10 mL of sterile distillate water, while yeasts suspension was made resuspended the plate-grown cells in sterile distillate water to obtain the starting fungal inoculum of 1.0×10^7 CFU mL⁻¹.

All tested substances were dissolved in 5% dimethyl sulfoxide (DMSO) aqueous solution and tested ranging from 2.0×10^{-1} to 2.4×10^{-5} mg mL⁻¹. Twofold serial broth dilution method in SDA was performed. All microwell plates were incubated at 30 °C and visually evaluated after 24 and 48 hours for *Candida* strains or 7 days *T. rubrum*.

All experiments were conducted in triplicate, and solvent blanks were included. Amphotericin B and fluconazole reference antimycotic compound were used.

4.5.3 Cytocompatibility assay. Normal human dermal fibroblast (NHDF), obtained from Sigma Aldrich (Italy) were cultured in polystyrene flasks in Dulbecco's modified Eagle's medium (DMEM; Sigma Aldrich, Italy), supplemented with 10% v/v heat-inactivated foetal bovine serum (FBS) (VWR International S.r.l, Milan, Italy), and with 1% v/v antibioticantimycotic solution (Sigma Aldrich, Italy). Cells were incubated at 37 °C in 5% CO2 atmosphere (CO2 Incubator, PBI International, Milano, I). 200 µl of cells (p2-p8) were seeded in 96-wells plate (Corning® 96 Well TC-Treated Microplates; Biosigma, Italy) (100 000 cells per cm²) and after 24 h, medium was removed, cells were washed with phosphate buffer solution (PBS, Sigma Aldrich, Italy) and samples were added (200 µl). Test compounds were prepared as 10 mM stock solutions in DMSO and diluted with complete culture medium (CM). Aliquots of the working solutions were transferred to 96-well plates to afford final concentrations of 125 µM and 50 µM. Cells were exposed to the compounds for 24 h at 37 °C. Vehicle (0.5% DMSO) was included on every plate.

To assess cell viability, Alamar blue (ThermoFisher Scientific, Italy) assay was performed. The medium was

removed and 100 μL of a 10% v/v solution of Alamar blue in DMEM was added to each well and left in contact for 3 h. After 3 h, fluorescence was detected by means of a multimode microplate reader (FLUOstar Omega Microplate Reader, BMG LabTech, Ortenberg, G) at two different wavelengths: at 570 nm to detect the reduced form (red) of the Alamar blue, and 655 nm, to detect the oxidized one (blue). Results were expressed as cell viability% by normalizing the fluorescence measured after contact with each sample with that measured for CM, used as reference. Six replicates were performed for each sample/control.

Abbreviations

AmB Amphotericin B
DCM Dichloromethane
DIPEA Diisopropylethylamine
DMF N,N-Dimethylformamide
DMSO Dimethyl sulfoxide

EDC 1-Ethyl-3-(3-dimethylaminopropyl)carbodiimide ESI-MS Electrospray ionization-mass spectrometry

EtOAc Ethyl acetate
EtOH Ethanol
FCZ Fluconazole
FIs Fungal infections
HOBt Hydroxy-benzotriazole

ACN Acetonitrile MeOH Methanol

r.t. Room temperature

SAR Structure–activity relationship

TEA Triethylamine
TFA Trifluoroacetic acid
TMSCl Trimethylsilyl chloride

UA Usnic acid

Data availability

All relevant data are within the manuscript and its ESI† files.

Author contributions

A. F. and A. C.: synthesis of compounds; G. R.: assistance in synthesis; R. L.: solubility studies; C. V.: cell viability studies; V. C.: extraction of UA; B. V.: supervising cell viability studies; M. E. E. T.: antifungal studies; S. T.: supervising antifungal studies; all the authors: writing – original draft; G. R. and P. L.: validation, writing and editing; S. C. and E. M.: validation, writing, review and editing, supervision, funding acquisition.

Conflicts of interest

There is no conflict of interest to declare.

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