


 Cite this: *Lab Chip*, 2025, 25, 5906

CandyCollect open-to-closed (O2C) microfluidic system for rapid and user-centric detection of group A *Streptococcus*

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We created CandyCollect, a lollipop-inspired, child-friendly saliva sampling device that provides enhanced comfort compared to common throat and mouth swabs. The device has an open microchannel and a functionalized surface that effectively captures and stores the saliva and pathogens. We recently demonstrated the ability to combine the CandyCollect with rapid antigen detection tests (RADTs) for fast and accessible detection of group A *Streptococcus* (GAS). However, the current procedure necessitates manual steps to elute and transfer captured antigens from the lollipop to RADTs, which relies on the user to follow complex instructions, increasing the likelihood of error. In this work, we developed the CandyCollect open-to-closed (O2C) microfluidic system to automate antigen elution and streamline detection with RADTs. The O2C system securely seals the open microchannels of the CandyCollect device to create a continuous closed channel for seamless delivery of elution reagents to release antigens at the press of a button for subsequent lateral flow detection. Our system leverages and combines the unique benefits of both open channel and closed channel microfluidics into one platform, further enhancing the potential of both methods. We performed experiments with GAS bacteria spiked into saliva and compared the manual and O2C elution methods. Both the manual and O2C methods showed clear test lines on the RADT strips at clinically relevant GAS concentrations (between 5×10^5 and 10^9 CFU mL⁻¹). The O2C provided comparable results to the manual procedure in a more convenient form factor. The O2C platform has the potential to enable user-friendly screening for respiratory pathogens by minimally trained users in decentralized settings.

 Received 10th March 2025,
 Accepted 8th August 2025

DOI: 10.1039/d5lc00253b

rsc.li/loc

Introduction

Oral samples for the diagnosis of respiratory diseases are critical diagnostic tools for children as certain bacterial and viral diseases can lead to long-term debilitating conditions or poor prognosis when left untreated.^{1–3} However, the lack of patient-centric access to reliable, easy-to-use diagnostic testing is a barrier that limits early and patient-driven testing. Furthermore, pediatric samples are particularly challenging to

obtain as children are frequently uncooperative with procedures that cause pain, trauma, or anxiety, and parents sometimes put off procedures that induce pain and discomfort to their child, which can hinder early detection and intervention measures. Methods such as the pharyngeal swab, sputum collection, and nasal swab are unpleasant for both adults and children,⁴ and pharyngeal swabs and sputum collection are also challenging to perform in a home setting.⁵ Child-friendly diagnostics are critical for diseases like streptococcal pharyngitis, commonly known as ‘strep throat’, influenza, respiratory syncytial virus (RSV), and pneumonia that affect millions of children worldwide, occasionally resulting in severe and long-term consequences.¹

We previously developed a lollipop-inspired device termed ‘CandyCollect’ that enables user-friendly, particularly child-friendly, high-quality pathogen collection from saliva using open microfluidic channels and functionalized surfaces.^{6–8} Preliminary results show that CandyCollect captures a variety of microbes and is preferred to traditional throat swabs, oral

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swabs, and spit tubes.⁶ In a clinical study of children with group A *Streptococcus* (GAS) pharyngitis, samples were collected with both CandyCollect and traditional mouth swabs. After analysis with quantitative polymerase chain reaction (qPCR), all participants (30/30) were positive for GAS for both sampling methods demonstrating that CandyCollect can be a feasible oral swab alternative.⁸ Furthermore, we demonstrated that CandyCollect can be used in place of a traditional pharyngeal swab with a commercial GAS rapid antigen detection test (RADT) kit, expanding the testing scenarios where this device can be applied.⁹

While our initial results integrating CandyCollect with RADTs were promising, the current workflow requires manual elution steps to transfer pathogens from the open microchannels on the CandyCollect lollipop to the RADTs. These manual handling steps rely on the user to follow complex instructions, increasing the likelihood of error during testing and limiting the user-friendliness of this platform. To enable at-home use of CandyCollect, there is a need for a more streamlined device that minimizes user interaction and automates key extraction and testing steps.

Currently, there are a few sampling devices available that are similar to our device but lack some of the features that make the CandyCollect unique. Self-LolliSponge™, V-check COVID-19, and Whistling COVID-19 are all saliva collection devices that facilitate self-collection of saliva with a minimally invasive protocol,^{10,11} however these devices lack the patient-centric candy aspect of the CandyCollect and there is a need for further integration of these devices with RADTs to enable point-of-care testing.

We recently developed the prototype CandyCollect open-to-closed (O2C) microfluidic system which aims to integrate pathogen capture, inline elution, and RADT detection into a single platform. The concept of integrating closed and open microfluidic channels has been explored before^{12,13} where continuous flow is provided by the closed channels and an intermediate channel opening provides access to fluids for sampling or measurement. Our approach takes a different route by capturing saliva and pathogens in open microchannels, which are then transformed into closed microchannels to deliver and transport reagents. By integrating these complementary functions into a single system, we maximize the unique capabilities of both microfluidic designs. The CandyCollect O2C device includes a top layer with a closed microfluidic channel, an intermediate device ‘ceiling’ layer to prevent leakage, and a bottom layer with a through-hole for liquid output. The bottom and top layers enclose the CandyCollect effectively creating a tight ‘sandwich’ that converts the open pathogen collection channel into a closed channel for inline testing. Using a finger pump,¹⁴ pathogen elution reagents are flushed through the now closed CandyCollect, and out the bottom where the sample can be collected and delivered to a RADT for testing and readout. Our results show that the O2C provides comparable performance to the manual

sample elution method, while also streamlining elution steps and enabling simple manipulation of smaller sample volumes in a convenient form factor. The system combines a user-friendly sampling method with a simplified testing process which lays the foundation for future developments of this prototype into a robust patient-centric diagnostic platform for decentralized settings.

Results and discussion

Components and assembly of CandyCollect O2C device

The CandyCollect was recently validated as a more comfortable alternative to the manufacturer-provided pharyngeal swab in the Areta Strep A Swab Test™.⁹ We found that when the CandyCollect was substituted into the workflow of the RADT kit, positive test results were detected by eye and with a signal image quantification program for bacterial concentrations as low as the manufacturer’s ‘minimal detection limit’: 1.5×10^5 CFU mL⁻¹;⁹ demonstrating that the CandyCollect can be used with LFA rapid tests.

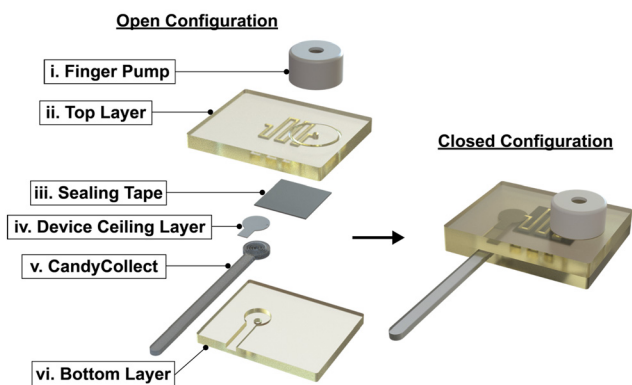
Here we engineered the O2C system to automate multiple user steps required in the manual RADT elution procedure. The open channels of CandyCollect enable capture and storage of pathogens but discourage controlled fluid delivery and automated manipulation of the sample. We transformed the open channels of CandyCollect to closed channels by placing a device ‘ceiling’ layer over the channels. A small hole in the ceiling layer at the inlet of the CandyCollect allowed reagents from the top channel to flow through the spiral of the CandyCollect. When the O2C device encloses the CandyCollect stick, the pressure of the top layer pushes down on the ceiling layer effectively and securely enclosing the open microchannels. Once the system is assembled, pressing the finger pump facilitates user-friendly addition of elution reagents to the CandyCollect to release captured antigens. The O2C device (Fig. 1A) consists of two 3D-printed layers (Fig. 1A(ii) and (vi)), a silicone finger pump retrofitted from an e-cigarette cap¹⁴ (Fig. 1A(i)), and a silicone ceiling layer (Fig. 1A(iv)) situated on top of the CandyCollect (Fig. 1A(v)). The top layer contains a 3D-printed microfluidic channel sealed with tape to retain elution reagents; upstream of the reagent channel is an inset for the finger pump attachment. The bottom layer has an inset for the CandyCollect device with a through-hole in the center for liquid elution into a reservoir. When all components are assembled and aligned, the outlet of the reagent channel aligns with the inlet of the CandyCollect microchannel (Fig. 1B), creating a closed, continuous pathway for liquid to travel through.

Simplified antigen elution workflow using CandyCollect O2C

The O2C automates the mixing of elution reagents with potentially infectious agents, enabling controlled and direct interaction of reagents with the captured pathogens in the microchannel. Mixing and elution into a reservoir occur consecutively with the single press of the finger pump. The



A CAD Schematic



B Device Photo

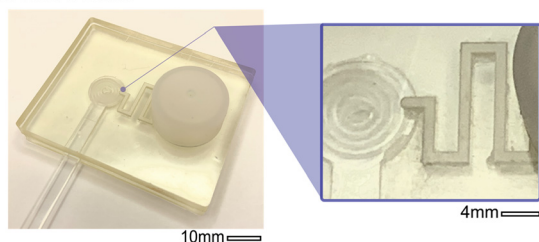


Fig. 1 Design and assembly of the O2C system for automated elution of antigens. A) Exploded CAD renderings of the O2C components. The rendering on the left contains a CandyCollect which has open channels and has yet to be sealed. The rendering on the right shows the entire system when it is assembled and the microchannels on the CandyCollect are fully enclosed. B) Image of the 3D-printed O2C device with fully assembled components. The inset on the right highlights the connection between the reagent channel on the top layer of the O2C and the CandyCollect.

complete workflow from the user's perspective is shown in Fig. S3 and Video S1. To mimic the sampling process, we first inoculated the CandyCollect with saliva and bacteria for 10 minutes (Fig. 2A(i), red food coloring used for visualization). Elution reagents are pipetted into the top channel (Fig. 2A(ii), blue food coloring used for visualization). The device components are assembled and held tightly together by binder clips to prevent leakage (Fig. 2A(iii)). Our current iteration uses binder clips for frugal prototyping; however, these can be exchanged for other clamping mechanisms in future designs to facilitate usability and manufacturability.^{15,16} The finger pump is pressed (Fig. 2A(iv)), and the liquid elutes through the CandyCollect channel and out into a reservoir (Fig. 2A(v)). Fig. 2B shows side and top views of the liquid path through the device. Elution buffer (**blue**) is flushed through the reagent channel, into the CandyCollect channel (**red**), mixing together, before being ejected into the reservoir (**dark purple droplet**) where the LFA test is performed. The current design enables automated mixing and elution; in future design iterations this workflow can be improved by including a holder for the lateral flow strip which can be directly attached to the O2C for minimal contact with potentially infectious material.

Testing the CandyCollect O2C with clinically relevant concentrations of GAS in spiked saliva

For our initial proof-of-concept experiments, a previously optimized total elution reagent volume of 200 μL (ref. 9) was used for both the manual and microfluidic elution methods. GAS concentrations between 5×10^5 and 10^9 CFU mL^{-1} showed clear test lines on the LFA strip (Fig. 3A and S1), and the quantified test line signals were above the positivity threshold (Fig. 3C). As concentrations of GAS increased, the O2C method provided comparable signal to the manual method. Both elution methods also showed faint positive test lines at 1.5×10^5 CFU mL^{-1} (the manufacturer-specified detection limit) with quantified test lines very close to the positivity threshold. At this concentration, the median values of the signal to baseline ratio (SBR) of the manual and microfluidic methods were 1.03 (interquartile range [IQR]: 1.03–1.04) and 1.03 ([IQR]: 1.02–1.05), respectively. A Wilcoxon matched pairs signed rank test to compare the median SBRs of the manual and microfluidic method showed there was no statistical difference ($p > 0.999$, $n = 4$) between the SBRs (Fig. S4).

To investigate the impact of sample pre-concentration on LFA results, we decreased the total elution volume to 100 μL . For both the manual and microfluidic methods, GAS concentrations between 5×10^5 and 10^9 CFU mL^{-1} again provided very clear test lines that were clearly above the positivity threshold (Fig. 3B and S2). At higher GAS concentrations, the test line intensities were comparable across both elution volumes, likely because the signal had already saturated the test lines. At lower GAS concentrations (10^5 – 10^6 CFU mL^{-1}), test lines from 100 μL elution volume provided larger signals than the 200 μL elution (Fig. 3C), likely because of sample pre-concentration in the smaller volume. At 1.5×10^5 CFU mL^{-1} , the median SBR values of the manual and microfluidic methods were 1.04 (IQR 1.03–1.05) and 1.05 (IQR 1.04–1.13), respectively. A Wilcoxon matched pairs signed rank test again demonstrated that there was no statistical difference ($p > 0.25$, $n = 3$) between the SBRs (Fig. S4).

Taken together, our visual, imaging, and statistical analysis suggest that the CandyCollect O2C microfluidic approach for sample elution provides comparable results to the traditional manual approach and can detect clinically relevant concentrations of GAS spiked into saliva. The O2C has the potential to enable a more integrated sample-to-answer workflow, where the user simply places a CandyCollect into the device, pushes a button, and gets LFA results. Ongoing work is focused on directly interfacing the LFA strip onto the O2C device to minimize risk with potentially infectious agents, integrating a reagent storage system for elution buffers, and conducting a usability assessment among minimally trained users prior to a clinical validation study. Our microfluidic approach also has the potential for spatial multiplexing and testing multiple LFA strips for different respiratory infections (*e.g.* COVID-19, Flu, RSV, *etc.*).



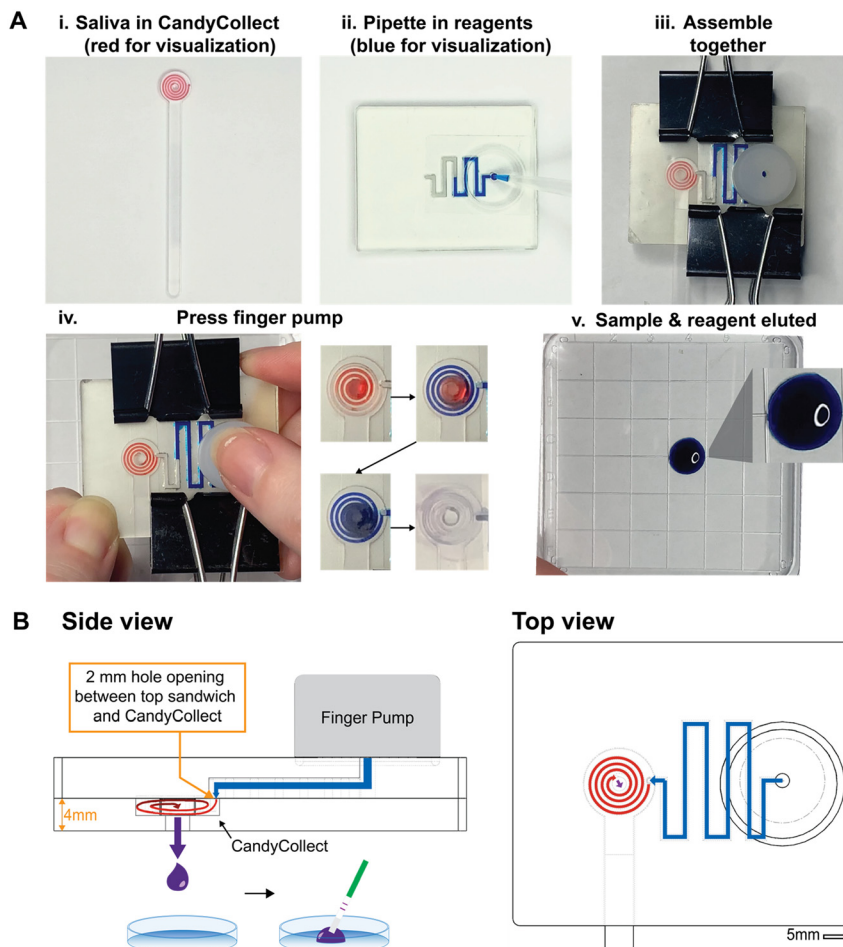


Fig. 2 Proof-of-concept operation of the CandyCollect O2C system using food dye solutions. A) Workflow for proof-of-concept testing with the CandyCollect O2C device. i) Inoculate CandyCollect with bacteria for 10 minutes (red food coloring for visualization). ii) Pipette elution buffer into reagent channel in the top layer (blue food coloring for visualization). iii) Stack components and clamp together to ensure tight seal between all layers. iv) Attach finger pump and press to elute. The time series photos depict the progression of liquid elution as the pump is pressed down. v) The sample is eluted from the CandyCollect open fluidic channel into reservoir. The inset shows a zoomed in photo of the intact droplet. B) Side view and top view of the device with fluid pathway highlighted in blue (*elution buffer*) and red (*saliva with bacteria*). When the finger pump is pressed, the two liquids mix together (*depicted in dark purple*) and output through the bottom layer outlet into a petri dish for analysis with a lateral flow strip.

Additional versions of Fig. 3 with the data reorganized for elution volume and with incorporated data from the kit-provided swabs are shown in Fig. S5 and S6, respectively.

Conclusion

We developed the CandyCollect O2C microfluidic system that uses a 3D-printed sandwich with closed channels to extract liquid from the open channel CandyCollect saliva sampling device to simplify downstream testing with RADT kits. Our modular “open-to-closed” prototype integrates the benefits of accessibility of open channels with the ability to use pressure driven flow in closed channels, presenting additional design paradigms for the lab on a chip community. Our results with GAS spiked into saliva samples show that the O2C provides comparable results to the manual elution method, while also concatenating

multiple elution steps, automating mixing, and enabling easy manipulation of smaller sample volumes. The initial prototype shows excellent promise in being able to integrate user-friendly sampling with a reliable, rapid diagnostic platform. The O2C device adds a minimal cost to the CandyCollect and elution reagents. The current cost of 3D-printing the O2C device is ~\$0.60 and we anticipate that this cost could be significantly reduced by transitioning to a manufacturing method that is more amenable to mass production like injection molding. Further work will streamline device assembly and manufacturability, incorporate stored reagents, automate the delivery of liquid to the LFA, and evaluate the usability of the device. We will also develop new prototypes that enable multiplexed detection of several respiratory pathogens within the same sample. The CandyCollect O2C microfluidic diagnostic platform has the potential to



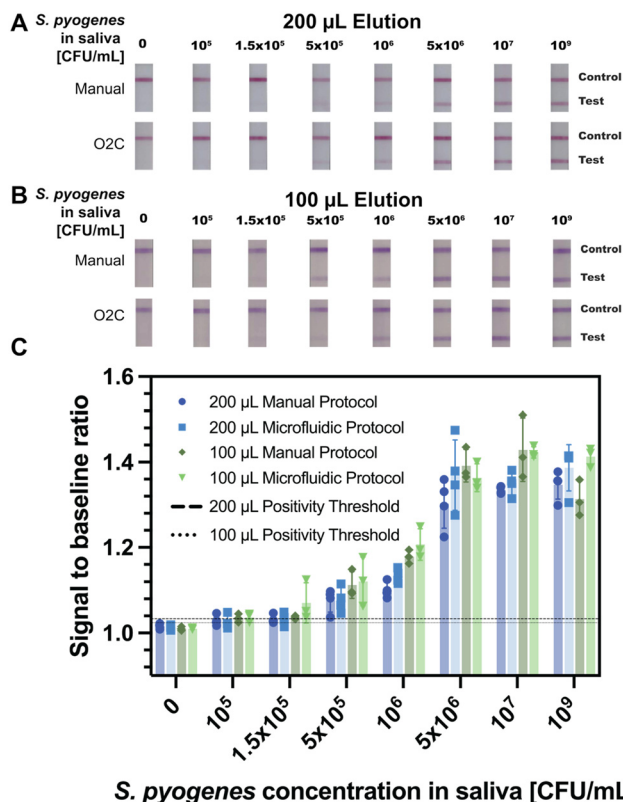


Fig. 3 The microfluidic O2C procedure provides similar performance to the manual elution protocol. A) Lateral flow strip images were taken from GAS concentrations ranging from 0 CFU mL⁻¹ to 10⁹ CFU mL⁻¹, with the manufacturer-specified limit of detection at 1.5 × 10⁵ CFU mL⁻¹. The manual protocol refers to the current RADT protocol⁹ with the CandyCollect substituted for the pharyngeal swab. O2C refers to the elution protocol performed with the microfluidic O2C platform. These results were obtained using the standard elution buffer volume of 200 µL. B) Lateral flow strips images taken from the same GAS concentrations as A with an elution buffer volume of 100 µL. C) Image analysis of the 200 µL and 100 µL dataset. Positivity threshold is the mean signal to baseline ratio +3 standard deviations of the negative controls. Generally, the 100 µL elution with the O2C device produced the highest signal to baseline ratio at the different concentration levels with a signal to baseline ratio above the positivity threshold for all concentrations tested except 0.

provide a child-friendly sampling method for consistent, hassle-free access to decentralized testing for minimally trained users.

Materials & methods

Fabrication of the CandyCollect O2C system

Fabrication of CandyCollect device. CandyCollect devices (Fig. S7) were milled from 2 mm thick sheets of polystyrene using a computer numerical control (CNC) Datron neo milling machine (GoodFellow, Cat# 235-756-86). Subsequently, the CandyCollect devices were sonicated with isopropanol (IPA) and 70% v/v ethanol (Fisher Scientific, Decon™ Labs, 07-678-004).^{6,7} Modified CandyCollect devices were created for the O2C system which had a 2 mm through-

hole cut into the center of the spiral and a 0.75 mm inlet located on the right-hand side of the lollipop face.

Fabrication of the O2C device. The layers of the microfluidic device (Fig. S8) were designed in Solidworks (Waltham, Massachusetts, USA) and 3D-printed with clear resin (Clear Microfluidics Resin V7.0a, CADworks3D, Toronto, ON, Canada) using a digital light processing printer (CADworks ProFluidics 285D, CADworks3D, Toronto, ON, Canada) which leverages a native 40 µm XY pixel resolution and an adaptive 28.5 µm XY pixel resolution. After printing, the devices were washed in an isopropanol (Sigma Aldrich, Cat#190764) bath, dried using a compressed nitrogen gun, and post-cured in a Formcure (Formlabs, Somerville, Massachusetts, USA) for 30 seconds on each side of the device.

The device ceiling layer (Fig. S8) was trimmed from a 0.3 mm thick sheet of silicone rubber (Laimeisi, Shenzhen, China) using a commercial craft cutter machine (Silhouette Cameo 4).¹⁷ A 0.8 mm circular through-hole was cut into the silicone to align with the inlet of the CandyCollect device.

A finger pump was retrofitted from an e-cigarette cap¹⁴ (ELF bar 600 disposable vape pod). A two mm hole was punched through the top of the cap to allow air to escape. After surface treating the top layer, the reagent channel was covered with hydrophobic tape (ARseal™ 90 697, Adhesives Research). A one mm outlet was left open at the end of the channel for the liquids to traverse into the CandyCollect microchannel.

Surface activation of CandyCollect device and reagent channel

The CandyCollect devices and reagent channel on the top layer of the device were activated by plasma treatment with oxygen using a Zepto LC PC Plasma Treater (Diener Electronic GmbH Plasma Treater, Ebhausen, Germany). The following protocol is consistent with previous publications from our lab.^{6,7} In short, the chamber of the plasma treater will release enclosed atmospheric gas, in order to achieve a new pressure of about 0.20 mbar. Afterwards, the chamber is then surged with oxygen gas to establish a pressure of 0.25 mbar. Lastly, the chamber is then exposed to 70 W of voltage for a duration of five minutes.

S. pyogenes culture

Liquid media preparation of Todd-Hewitt yeast (THY) broth & *S. pyogenes* maintenance. The *Streptococcus pyogenes* strain SF 370 (American Type Culture Collection, ATCC®, Cat#700294) was used for all experiments. Todd-Hewitt Broth (TH Broth) (BD Bacto™ TH broth, Fisher Scientific, Cat#DF0492-17-6), containing 0.2% yeast extract (United States Biological Corporation, Fisher Scientific, Cat# NC9796728) (THY) was used to culture *S. pyogenes*. Freeze-dried *S. pyogenes* was transferred from a frozen stock kept in a -80 °C freezer into a 15 mL conical tube (Fisher Scientific, Falcon™, Cat# 14-959-49B) containing 10 mL of THY media.



The culture was then incubated overnight at 37 °C supplied with 5% CO₂.⁹

***S. pyogenes* agar maintenance.** *S. pyogenes* was maintained by streaking bacteria onto the agar plates with a sterile disposable inoculating loops (Globe Scientific, Fisher Scientific, Cat# 22-170-201). The agar plates were incubated overnight at 37 °C supplied with 5% CO₂. Agar plates were stored at room temperature for up to 7 days. An individual colony from the agar plate was used to inoculate with 10 mL of THY liquid media confined in a 15 mL conical tube (Fisher Scientific, Falcon™, Cat# 14-959-49B) and cultured overnight as above.^{6,7}

Capture, elution, and detection of *S. pyogenes* in manual vs. microfluidic procedure

Suspension of *S. pyogenes* in filtered pooled human saliva.

To estimate the concentration of *S. pyogenes* in liquid culture media, optical density at 600 nm (OD₆₀₀) was measured using a visible 721-vis spectrometer (Vinmax). To achieve a pellet of *S. pyogenes*, the liquid culture was centrifuged for 5 minutes at 10 000 rpm. Afterwards the *S. pyogenes* pellet was resuspended with filtered pooled human saliva (purchased from Innovative Research, Cat# IRHUSL50ML; filtered using a 0.22 μm filter). Finally, serial dilutions were performed to achieve the desired *S. pyogenes* concentration.⁹

Capture of *S. pyogenes* on the open fluidic channels on the CandyCollect device. 50 μL of *S. pyogenes* in saliva were incubated on the CandyCollect device for 10 min with the following concentrations: 1.0 × 10⁷, 5.0 × 10⁶, 1.0 × 10⁶, 5.0 × 10⁵, 1.5 × 10⁵, and 1.0 × 10⁵ colony forming units (CFU) mL⁻¹; with the addition of a positive control, *S. pyogenes* at 1 × 10⁹ CFU mL⁻¹ suspended in liquid media, and a negative control, filtered pooled saliva.⁹

Extraction of surface antigen from *S. pyogenes* through manual procedure. After the 10 min incubation time, the CandyCollect device was transferred into a 15 mL test tube containing the elution reagents: 2.0 M sodium nitrite (reagent A) and 0.4 M acetic acid (reagent B). Previous studies reported that these reagents produced the highest test line intensity compared to other reagents.⁹ Afterwards, the test tube was shaken for 15 s to ensure that the liquid had adequate contact with the CandyCollect microchannel and left to sit for 45 s.

Extraction of surface antigen from *S. pyogenes* using the CandyCollect O2C system. The CandyCollect device was placed into the inset of the bottom layer. The device ceiling layer was placed on top of the CandyCollect with the inlet on the CandyCollect and the through-hole in the ceiling layer aligned. The top layer was positioned on the bottom layer and the device was clamped tightly together with binder clips. The entire assembly took around one minute, matching the timing of the manual procedure. The finger pump was placed in the inset of the top layer and the liquid was eluted into a waiting petri dish (Fisher Scientific, Corning, Cat#07-200-83) upon a single press.

Lateral flow immunoassay strip readout and data analysis

Examination by eye of lateral flow immunoassay strip. A lateral flow immunoassay (LFA) strip from the Areta kit was saturated with the eluate from either the manual elution procedure or the O2C. Following manufacturer's instructions¹⁸ to ensure enough eluted solution was added to the LFA, we held the strip in the eluted solution until wicking was observed at the start of the nitrocellulose membrane and then removed and set aside in a petri dish (Fisher Scientific, Cat# 165218) with a timer set for 18 min. After 18 min, we examined the strips in lab by eye and recorded the results which are valid from 10 min to 20 min after the test strips have been removed from the elution solution. Lastly, strips were scanned (Hewlett-Packard, HP OfficeJet Pro6978, SN# TH0AK4N0YT) at 600 dpi and darkness = 7 to a JPEG format (Fig. S9).⁹

Quantification of signal on Areta lateral flow immunoassay strip via optical analysis. We used a custom image analysis Python script to analyze the scanned LFA strips, using an algorithm published in previous work.¹⁹ The scanned images were cropped to 25 × 100 pixel images from the original full-color images obtained. The images were converted to monochrome, inverted, and profiles of digital numbers against the pixel location were obtained by averaging the rows for each strip. The code denotes the peak signal values from the test line against location on the lateral flow strip. To establish baseline signal, two regions-of-interest (ROIs) spanning between 5 to 10 pixels were manually selected, approximately 30 pixels from both sides of the peak. The average digital number from these two ROIs is calculated to compute the baseline. To obtain the signal-to-baseline ratio (SBR), the ratio of the peak signal of the test line from the profile is divided by the baseline.

The positivity threshold is a value found from the SBR, above which the test results are deemed positive. It is derived from image analysis of the three negative controls collected from each individual experiment and is calculated as such:

$$\text{Positivity Threshold} = \mu_{\text{SBR}} + 3\sigma_{\text{SBR}}$$

where μ_{SBR} is the average signal-to-baseline and σ_{SBR} is the standard deviation signal-to-baseline of the three negative controls. The python script finds the negative signal by analyzing a segment of the profile located a specific distance from the negative control line peak, a distance which is informed by the test line peak locations in positive test strips. This approach ensures that the location of the segment accurately reflects the expected position of the negative control test line.

See SI for expansion of materials and methods.

Author contributions

K. M. L., A. B. T., E. B., and A. O. O. conceptualized the initial design integrating the CandyCollect with a microfluidic device. K. M. L., J. C. S., A. B. T., and A. O. O. conceptualized



the wet lab studies to test the platform with spiked saliva. J. C. S. milled the CandyCollect devices. K. M. L. 3D printed and craft cut the microfluidic device and ceiling device. K. M. L., R. R. A., J. A. T., and C. C. designed the microfluidic device in CAD. K. M. L. and J. A. T. performed the initial device proof-of-concept tests. J. C. S. and K. M. L. designed the biological experiments, conducted the biological experiments, and performed data collection. A. B. T. and A. O. O. advised on study design and execution. J. C. S., T. R. R., M. M. C., A. O. O., E. B., and A. B. T. developed and refined data analysis methods for quantifying RADT results. All authors analyzed and interpreted data. K. M. L. and A. O. O. wrote sections of the manuscript. K. M. L. created all visual work presented in the paper. All authors revised and approved the manuscript.

Conflicts of interest

Ashleigh B. Theberge, Xiaojing Su, Erwin Berthier, and Sanitta Thongpang filed patent 63/152103 (International Publication Number: WO 2022/178291 A1) through the University of Washington on the CandyCollect oral sampling device. Kelsey M. Leong, Ayokunle O. Olanrewaju, Rene Arvizu, Erwin Berthier, Cosette Craig, Timothy Robinson, J. Carlos Sanchez, John Tatka, and Ashleigh B. Theberge filed patent 63/683571 through the University of Washington on this platform. Ashleigh B. Theberge reports filing multiple patents through the University of Washington and receiving a gift to support research outside the submitted work from Ionis Pharmaceuticals. Erwin Berthier is an inventor on multiple patents filed by Tasso, Inc., the University of Washington, and the University of Wisconsin. Sanitta Thongpang has ownership in Salus Discovery, LLC, and Tasso, Inc. Erwin Berthier has ownership in Salus Discovery, LLC, and Tasso, Inc. and is employed by Tasso, Inc. However, this research is not related to these companies. Sanitta Thongpang, Erwin Berthier, and Ashleigh B. Theberge have ownership in Seabright, LLC, which will advance new tools for diagnostics and clinical research, potentially including the CandyCollect device. The terms of this arrangement have been reviewed and approved by the University of Washington in accordance with its policies governing outside work and financial conflicts of interest in research. The other authors have no conflicts of interest to disclose.

Data availability

Supplementary information: the supplementary information (SI) includes a link to a repository containing CAD files of the O2C system and ceiling device, a link to a GitHub with the image analysis code, compiled raw images of the lateral flow strips from all experiments, an illustrated workflow of the O2C, graphs reorganized by elution volume and containing additional data from kit-provided swabs, engineering drawings of the O2C system, a description of the scanner optimization, and an expanded materials and methods section. See DOI: <https://doi.org/10.1039/D5LC00253B>.

The data supporting this article have been included as part of the SI.

Acknowledgements

We want to acknowledge the M. J. Murdock Diagnostic Foundry for Translational Research at UW for allowing us to use equipment. We would also like to thank Dr. Lucas Meza and the Meza Lab for access to instruments. We are grateful for funding from the National Institute of Health (NIH) through R35GM128648. The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health or other funding sources.

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