

# Sustainable Food Technology

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## Sustainability Spotlight

Biodegradable coatings developed from chitosan, polyvinyl alcohol, gelatin, and *Rosa rubiginosa* extract present an environmentally friendly technique for fruit preservation. These natural and synthetic biopolymers are combined to produce films with improved antioxidant, antibacterial and barrier properties, providing a sustainable alternative to traditional plastic packaging. By applying these coatings to strawberries, the study shows a significant increase in the shelf life while minimizing food waste and environmental impact. The addition of *Rosa rubiginosa*, a plant rich in bioactive compounds, enhances the value of underutilized botanical resources. This work aligns with the goals of circular bioeconomy and supports the global transition to sustainable, biodegradable materials for food packaging applications.



1 **Chitosan/PVA/Gelatin Derived Biodegradable Coatings Enriched with**  
2 ***Rosa rubiginosa* for Extended Shelf Life and Preservation of Strawberries**

3  
4 Beenish Sarwar<sup>a</sup>, Muhammad Zubair<sup>b</sup>, Asma Yaqoob<sup>c</sup>, Faiz Ahmed<sup>d</sup>, Sohail Shahzad<sup>a\*</sup>,  
5 Aman Ullah<sup>b\*</sup>

6 <sup>a</sup>*Department of Chemistry, University of Sahiwal, Sahiwal 57000, Pakistan*

7 <sup>b</sup>*Lipid Chemistry Utilization Lab, Department of Agricultural, Food & Nutritional Science, University of*  
8 *Alberta, Edmonton, AB, Canada, T6G 2P5*

9 <sup>c</sup>*Institute of Biochemistry, Biotechnology and Bioinformatics, The Islamia University of Bahawalpur*

10 <sup>d</sup>*Department of Chemistry, Government College University Faisalabad, Faisalabad 37000, Pakistan.*

11  
12 ***\*Correspondence Authors:***

13 **Sohail Shahzad:** [drsohail@uosahiwal.edu.pk](mailto:drsohail@uosahiwal.edu.pk)

14 **Aman Ullah:** [ullah2@ualberta.ca](mailto:ullah2@ualberta.ca)



## 23 Abstract

24 This study developed biodegradable composite coatings and films using chitosan (CS),  
25 polyvinyl alcohol (PVA), and gelatin enriched with *Rosa rubiginosa* extract (RRE) to enhance  
26 the shelf life of strawberries. The coatings were prepared through solution blending and applied  
27 via dip-coating, while the corresponding films were fabricated using solution casting. FTIR  
28 and SEM analyses confirmed the effective molecular interactions and uniform microstructure  
29 within the composite matrix. Increasing the proportion of RRE improved functional  
30 performance by reducing the moisture content, water solubility, water absorption, and oxygen  
31 ingress. The formulation with the highest RRE content (BS-03) exhibited the strongest  
32 antioxidant activity ( $78.8 \pm 1.01\%$  DPPH inhibition;  $IC_{50} = 0.3$  mg/mL) and enhanced  
33 antibacterial potential against *Staphylococcus aureus*, *Bacillus subtilis*, and *Escherichia coli*.  
34 Soil burial tests revealed that the films were biodegradable, with BS-02 and BS-03 showing  
35 mass losses of up to 91.29% and 86.79% after 45 days, respectively. When applied to  
36 strawberries, the coatings significantly reduced weight loss and delayed visible spoilage,  
37 extending the shelf life by up to 10 days compared to uncoated fruit. Sensory evaluation also  
38 confirmed better color, firmness, and acceptability of the coated samples. These findings  
39 demonstrate that CS/PVA/gelatin coatings enriched with RRE offer a promising, safe, and eco-  
40 friendly approach for prolonging the freshness of perishable fruits.

41 **Keywords:** Antibacterial, Antioxidant, Dip coating technique, Solution casting, Food  
42 packaging



## 43 1. Introduction

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44 Approximately 1.3 billion tons of food is wasted annually, making food waste one of the major  
45 global challenges of our time, results in resource depletion and environmental damage <sup>1</sup>.  
46 Packaging protects food from harmful ultraviolet rays and preserves its quality, aroma, taste,  
47 and physicochemical characteristics. By preserving food, we can enhance its use and protect it  
48 from microbial decay <sup>2</sup>. With the rising demand for eco-friendly packaging solutions,  
49 innovative biopolymer-based coatings play a vital role in preserving food and prolonging the  
50 shelf life of perishable items, thereby positively affecting the environment <sup>3</sup>. Films containing  
51 plasticizers such as glycerol, honey, and propylene glycol are applied to food products to  
52 improve their nutritional, antimicrobial, and antioxidant properties <sup>4</sup>. The addition of natural  
53 extracts, such as essential oils or plant extracts, to chitosan-based films improves their  
54 functional properties <sup>5</sup>. A variety of semi-synthetic and edible biopolymers are used in food  
55 packaging applications, including chitosan, agarose, alginate, agar, starch, cellulose, guar gum,  
56 polyvinyl alcohol and carboxymethyl cellulose <sup>6,7</sup>. Although plastic materials are lightweight,  
57 economical, and flexible, they contribute to waste, raising environmental concerns <sup>3</sup>.

58 Recently, there has been growing interest in utilizing biodegradable materials for food  
59 packaging and coatings<sup>8</sup>. Bio-based plastics are favoured for film coatings because they are  
60 abundantly available, biodegradable and capable of forming films <sup>9</sup>. However, these materials  
61 often lack mechanical strength and offer poor resistance <sup>10</sup>. To address these limitations,  
62 structural modifications <sup>11</sup> or blending of bio-based polymers with synthetic polymers have  
63 been performed to obtain specific properties <sup>12</sup>. Chitosan, a naturally occurring biopolymer  
64 with inherent anti-microbial <sup>13</sup> and anti-oxidant properties <sup>14-16</sup>, has been widely used in food  
65 packaging <sup>17</sup>. It is non-toxic <sup>18</sup> and has chelating ability to bind with vital trace metals  
66 selectively, bacteria and prevent toxin and bacterial growth <sup>19,20</sup>, thus making it perfect for food  
67 packaging applications <sup>21, 22</sup>. In addition, chitosan has high susceptibility to water and low



68 thermal and mechanical strength, which limits its use in food packaging materials. To address  
69 these limitations, different techniques such as cross-linking<sup>23-26</sup>, enzymatic treatment<sup>27</sup>, graft  
70 copolymerization<sup>28-30</sup>, complex formation<sup>29</sup>, surface coating<sup>31</sup>, incorporation of fillers<sup>32</sup>,  
71 high-energy radiation<sup>33</sup> and blending with other biopolymers or synthetic polymers can be  
72 used<sup>34,35</sup>.

73 PVA is a synthetic and water-soluble polymer that is an ideal biomaterial for making polymer  
74 blends. It is biocompatible, flexible, and non-toxic<sup>36</sup> and can be used in food packaging  
75 applications because of its low cost, eco-friendly nature, durability, transparency, mechanical  
76 strength, chemical stability, and film-forming properties<sup>37,38</sup>. Blending PVA with chitosan not  
77 only enhanced the mechanical strength but also improved the antibacterial activity of the  
78 fabricated films.

79 In contrast, *Rosa rugosa*, a common petal variety, has been traditionally used in biomedical  
80 applications<sup>39</sup>. *Rosa* species contain components including flavonoids, terpenes, glycosides,  
81 polyphenols, and anthocyanins. The volatile oil extracted from *Rosa* flowers possesses  
82 therapeutic properties and is primarily used for analgesic, anticonvulsant, cardiovascular, and  
83 laxative purposes<sup>40</sup>. Given the numerous bioactive components found in *Rosa rubiginosa*  
84 (RR), our objective was to explore the effects of its incorporation and proportion when  
85 combined with CS/PVA/gelatine solutions. These coatings and films are expected to exhibit  
86 enhanced mechanical strength and improved antibacterial, antioxidant, and biodegradable  
87 properties, which are essential for prolonging the shelf life of coated or packaged food  
88 products, such as strawberries. Choi *et al.* developed a ternary blend (CS/PVA/gelatin)-based  
89 composite film incorporating *Duchesnea indica* extract for the preservation of strawberries<sup>41</sup>.  
90 In another study, Islam *et al.* fabricated packaging films using a casting methodology with  
91 CS/PVA incorporated with zinc oxide nanoparticles and dragon fruit waste extract. The  
92 incorporation of ZnO nanoparticles and dragon fruit waste extract enhanced the thermal and



93 mechanical strength and provided antioxidant and antibacterial potential to the films.<sup>42</sup>  
94 Similarly, Wang *et. al.* synthesized bilayered films with excellent biodegradable, antibacterial,  
95 mechanical, and oxygen barrier properties using CS as the outer layer and PVA/gelatin as the  
96 inner layer loaded with 2.5 % curcumin nanoparticles for the preservation of fish (bigeye tuna)  
97 <sup>43</sup>. However, CS/PVA/gelatin ternary films have not yet been combined with RRE, and their  
98 application as edible coatings for strawberries has not been evaluated in terms of  
99 antioxidant/antibacterial performance, barrier properties, biodegradability, and sensory  
100 acceptance. This study aims to develop and characterizes/PVA/gelatin coatings enriched with  
101 varying concentrations of RRE and evaluate their effectiveness in extending the shelf life of  
102 strawberries.

## 104 2. Experimental

### 105 2.1 Materials

106 The chemicals and solvents utilized in this study were employed directly as obtained from  
107 manufacturers or redistilled when necessary. Macklin Chemicals (China) supplied Chitosan  
108 (CS) with a deacetylation degree exceeding 95% (molecular weight (MW) 100–200 kDa) and  
109 viscosity was 100–200 mpa.s. Polyvinyl alcohol (PVA) (MW 72,000 with 98% degree of  
110 hydrolysis) was procured from Merck (Germany). Gelatin and glacial acetic acid (C<sub>2</sub>H<sub>4</sub>O<sub>2</sub>)  
111 were bought from Sigma Aldrich–Germany. A taxonomist identified the plant sample as *Rosa*  
112 *rubiginosa* (RR), which was obtained from the University of Sahiwal, Pakistan. Strawberries  
113 were purchased from a local market in Sahiwal–Pakistan

### 114 2.2 Methodology

#### 115 2.2.1 Preparation of *Rosa rubiginosa* extract (RRE)



116 The flower petals of *Rosa rubiginosa* were thoroughly washed with deionized water to remove  
117 any debris or impurities present on the surface, followed by air drying at room temperature  
118 (20–25 °C) for 10–12 days. The petals were ground to make their powder, and then 5% w/v  
119 aqueous solution of *Rosa rubiginosa* was prepared by refluxing at 60 °C and centrifugal force  
120 of 125.8 g for a continuous 4 hours. The solution was filtered out with Whatman filter paper  
121 number 1, under vacuum conditions, and the filtrate was stored at 4 °C for further use to  
122 fabricate coatings and films.

### 123 2.2.2 Preparation of CS/PVA/Gelatin derived coatings and films loaded with RRE

124 The CS/PVA/Gelatin based composite coatings and then finally films were produced using  
125 blending of individual component solutions and then conventional solution casting  
126 methodology, as described by Rubilar *et al.*,<sup>44</sup>. A 1% chitosan (w/v) solution was prepared by  
127 dissolving 1 g of chitosan in a 1% acetic acid solution, with stirring at room temperature for a  
128 duration of 72 hours. Similarly, 5% PVA (w/v) solution was prepared by dissolving 5 g of PVA  
129 in the respective amount of deionized water by refluxing at 80 °C for 5 hours. 2% (w/v) gelatin  
130 solution was made by dissolving 2 g of gelatin in deionized water. 30% (v/v) glycerol solution  
131 was made by combining 30 mL glycerol with the required amount of deionized water. Then,  
132 different volume proportions of CS, PVA, gelatin, glycerol, and RRE (**Table 1**) were combined  
133 and stirred for 48 hours at room temperature to form a uniform solution. The resulting solutions  
134 were then used for dip coating of strawberries and were also used for evaluation of antioxidant  
135 and antibacterial potential. The other part of solution was cast in petri dishes and kept at room  
136 temperature, to fabricate composite films, for characterization and evaluation of certain other  
137 properties. The solution was evaporated within 2 to 3 days and films were formed, after which  
138 they were removed from the dishes.

139

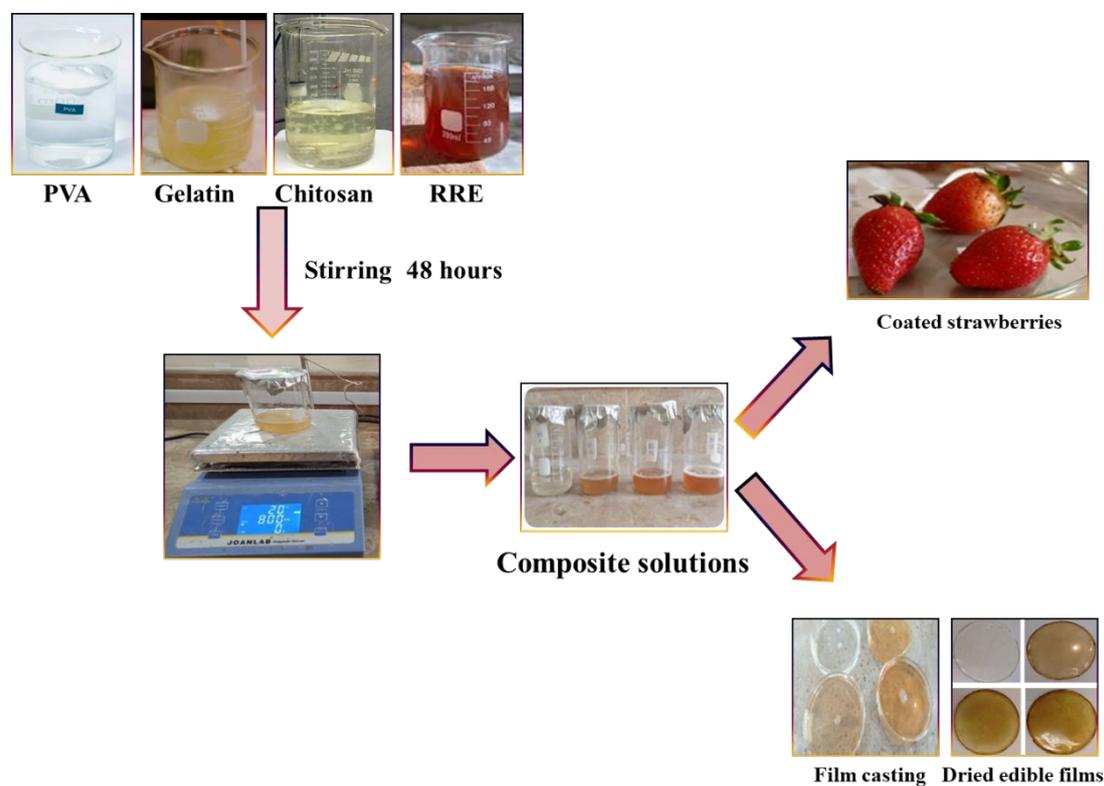
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144

145 **Figure 1.** Preparation of composite CS/PVA/Gelatin coatings and films loaded with RRE146 **Table 1.** Different volume ratios of all solutions for coatings and film formation

Samples codes	CS solution (mL)	PVA solution (mL)	Glycerol solution (mL)	Gelatin solution (mL)	RRE (mL)
BS-0	20	20	10	2	0
BS-01	20	20	10	2	2
BS-02	20	20	10	2	4
BS-03	20	20	10	2	6



147

## 148 **2.3 Characterization of synthesized film**

### 149 **2.3.1 FTIR analysis**

150 All composite fabricated films were characterized by FTIR spectrometer (Thermo Nicolet  
151 6700P, Waltham, Massachusetts, USA), to evaluate interactions among individual components  
152 and to identify peaks of prominent functional groups of films. Spectra were acquired on photo  
153 acoustic mode, with a range of 4000–400  $\text{cm}^{-1}$ , using carbon background and helium purging  
154 with 8  $\text{cm}^{-1}$  resolution and 256 scan numbers

### 155 **2.3.2 Thickness of fabricated films**

156 The thickness of fabricated films in millimetres (mm) was noted by using a digital thickness  
157 gage (manufactured by Insize company, China), with 0–10 mm range, 0.01 mm resolution and  
158  $\pm 0.02$  mm accuracy. Reported thickness values are the mean values of five random values taken  
159 at various positions, including the surroundings and centre.

### 160 **2.3.3 Camera images**

161 Camera images of synthesized films were taken to evaluate the appearance, homogeneity,  
162 colour and texture. Images were taken by the mobile camera of Infinix Hot-30 play made by  
163 Infinix Mobile China, Dual-LED flash, HDR, panorama (16 MP, f/1.8, wide, PDAF, QVGA,  
164 1080p@30fps).

### 165 **2.3.4 Scanning electron microscope (SEM) analysis**

166 To investigate the surface and cross-sectional microstructure of the fabricated films, SEM  
167 analysis was conducted using the EmCrafts Cube II tabletop model from South Korea. The  
168 samples were sputter-coated and placed on SEM holders, with images captured at different  
169 magnifications using an accelerating voltage of 10 KV.

### 170 **2.3.5 Dynamic mechanical analysis (DMA)**



171 Mechanical properties of films were evaluated by measuring tensile strength and elongation at  
172 break using a dynamic mechanical analyzer (DMA) Q800 manufactured by TA Instruments  
173 (USA) with software version of V21.3 Build 96 using controlled force module. The films were  
174 cut in 6×3 cm size and analysed by the method equilibrate at 50 °C, isothermal for 5 minutes  
175 and ramp force 3 Newton/minute to 18 Newton/minute.

### 176 **2.3.6 X-ray diffraction (XRD) analysis**

177 XRD analysis was performed to analyse the crystalline/amorphous nature of fabricated  
178 CS/PVA/gelatin enriched with RRE films using XRD powder system, D8 advance, model  
179 cube 10, EmCrafts manufactured by Bruker, South Korea. Films spectra were recorded in  
180 range of 10–70° with 20 seconds per step and 0.09 step sizes.

### 181 **2.3.7 Percent moisture content (MC %) determination**

182 The percentage moisture content was determined using the loss on drying method. Initially,  
183 small sections of the film samples were cut, and their weight was measured. These samples  
184 were then placed in an oven set at 80 °C for one hour and weighed again with precision. This  
185 process was repeated until the weight remained constant. The moisture content percentage (MC  
186 %) was calculated using equation 1<sup>45, 46</sup>.

$$187 \quad MC(\%) = \frac{M_w - M_d}{M_w} \times 100 \quad (1)$$

188  
189 Where  $M_w$  stands for initial weight of film samples before drying conditioned at 53% relative  
190 humidity to moisture equilibrium and  $M_d$  stands for dried weight. Average moisture content  
191 percentage was determined from three replications.

### 192 **2.3.8 Water solubility percent (WS %)**

193 Small pieces of films were cut and placed in oven at 80 °C for 1 hour and then their dried weight  
194 was measured. The films were then introduced into petri dishes containing 25 mL of distilled



195 water for 24 hours for their maximum solubility in water. The films were removed from petri  
196 dishes and were dried in oven at 80 °C until to attain a constant weight. The water solubility  
197 percent (WS %) was evaluated by following formula <sup>47</sup>.

$$198 \quad WS \% = \frac{W_i - W_f}{W_i} \times 100 \quad \text{Equation 2}$$

199 Where  $W_i$  stands for initially dried weight in grams and  $W_f$  stands for final/terminal dried  
200 weight (g). For each measurement, three readings were taken then WS % was evaluated by  
201 taking mean reading.

### 202 **2.3.9 Water absorption pattern of films**

203 The small pieces of films were cut and their dried weight was measured. These pieces were put  
204 into the petri dishes containing 10 mL distilled water, so that films can absorb maximum water  
205 and may swell by retaining water within their structure. The water absorption behavior of each  
206 film was observed by keeping films dipped in water for 10 minutes and then their removal and  
207 subsequently weight measurement. After weighing, the films were again immersed in water,  
208 and the process was repeated until to achieve constant weight. The percentage water absorption  
209 capacity of each film was determined by using the following formula <sup>48</sup>.

$$210 \quad \text{Water absorption (\%)} = \frac{W_s - W_d}{W_d} \times 100$$

211  
212 Where  $W_s$  stands for swollen weight and  $W_d$  stands for dried weight.

### 213 **2.3.10 Water drop test**

214 A piece of the film was placed over a support raised from the table and a water drop was poured  
215 onto the film surface. Then pass or fail judgment was given that depending on whether the drop  
216 passed through the film or not <sup>49, 50</sup>.

### 217 **2.3.11 Oxygen permeability**



218 Winkler's method, which measures dissolved oxygen in mg/mL, was employed to assess the  
219 oxygen permeability of CS/PVA/gelatin/RRE films<sup>51</sup>. In this process, distilled water was  
220 placed in glass bottles sealed with composite films using a sealant, and the quantity of oxygen  
221 that permeated through them was measured.

### 222 **2.3.12 Antibacterial potential of CS/PVA/gelatin/RRE coatings solutions**

223 The antibacterial potential of CS/PVA/gelatin coating solution and CS/PVA/gelatin infused  
224 with RRE solutions was evaluated against two gram-positive bacterial strains, *S. aureus* and *B.*  
225 *subtilis*, as well as one gram-negative strain, *E. coli*. This assessment was conducted using the  
226 standard method previously described<sup>52</sup>. A suspension of 100 µL for each of the two bacterial  
227 strains, containing approximately 10<sup>7</sup> colony-forming units (CFU) per mL, was prepared. The  
228 bacterial cells were cultured in nutrient broth (Oxoid, UK) for around 8 hours at 37 °C. Using  
229 a sterilized swab, the bacterial culture was evenly distributed on agar plates, which were then  
230 allowed to dry for 15 minutes. Filter discs soaked with CS/PVA/gelatin/RRE solutions were  
231 placed on the agar plates, with four discs, each containing a different sample, positioned  
232 equidistantly on each test plate. A separate petri dish served as a positive control, containing a  
233 filter disc impregnated with the commercially available antibiotic ciprofloxacin at a  
234 concentration of 10 µg/mL. Before incubating the plates at 37 °C for 17–24 hours, they were  
235 refrigerated at 4 °C for two hours. The inhibition zones produced by each sample were  
236 measured in millimetres using calipers, indicating the antibacterial potential of each sample  
237 and the positive control. The experiments were conducted in triplicate to ensure reliability.

### 238 **2.3.13 Antioxidant potential of CS/PVA/gelatin/RRE coatings solutions**

239 The antioxidant properties of samples containing CS/PVA/gelatin infused with RRE were  
240 assessed using the 2, 2-diphenylpicrylhydrazyl (DPPH) method. This technique, commonly  
241 employed to evaluate the antioxidant potential of samples with plant extracts, is highly  
242 regarded for its stability, practicality, and reliability. The level of antioxidant activity is gauged

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243 by the components' ability, particularly plant-based ones like flavonoids and polyphenolics, to  
 244 neutralize the DPPH radical. In this method, a sample was prepared by mixing 20  $\mu$ L of  
 245 solution with 3 mL of distilled water. Then, 1 mL of this mixture was added to 0.2 mL of a  
 246 methanolic DPPH solution (1 mM). The resulting mixture was kept in the dark at room  
 247 temperature for about 40 minutes. The absorbance of the solution was measured at 517 nm  
 248 against a blank using a UV-Visible spectrophotometer. This procedure was conducted three  
 249 times, and the percentage of DPPH radical scavenging activity was calculated using a specific  
 250 formula <sup>45, 46</sup>.

$$251 \quad \text{Scavenging activity (SA \%)} = \frac{A_{\text{control}} - A_{\text{sample}}}{A_{\text{control}}} \times 100$$

252  
 253 Where  $A_{\text{control}}$  shows the absorbance of control that is DPPH solution and  $A_{\text{sample}}$  stands for  
 254 absorbance of samples by DPPH.

#### 255 **2.3.14 Biodegradation test**

256 Film samples were cut into small strips for evaluation of their biodegradation behavior. The  
 257 experiment utilized 200 g of dry soil collected from the Sahiwal field area, which was then  
 258 moistened by spraying distilled water. Four polythene cups were filled with equal amounts of  
 259 this prepared soil. The film samples were buried at a consistent depth within the soil and  
 260 maintained at room temperature with 70% relative humidity. After 15 days, the film's condition  
 261 was evaluated based on their visual appearance and weight loss. Subsequently, the samples  
 262 were reburied in the soil and examined again after 45 days to assess their further degradation  
 263 <sup>48</sup>.

#### 264 **2.3.15 Total flavonoid contents (TFCs)**

265 TFCs of RRE with a concentration of 5 mg/mL were assessed by a procedure already declared  
 266 in literature with a few modifications <sup>53</sup>. This is the same concentration which was incorporated  
 267 into CS/PVA/gelatin films. For this assay, freshly prepared reagents including 1 M NaOH, 5%



268 NaNO<sub>2</sub> and 10 % Al<sub>2</sub>Cl<sub>3</sub> were used. The assay involved taking 1 mL of RRE in a test tube and  
269 then adding 400 µL of NaNO<sub>2</sub> and 6 mL of distilled water. The mixture was thoroughly blended  
270 to ensure maximum homogeneity. The mixture was incubated for 10 minutes, followed by  
271 addition of 700 µL of Al<sub>2</sub>Cl<sub>3</sub> (10%) and 3 mL of NaOH (1M). The absorbance was  
272 subsequently recorded at 510 nm using the STA–8100ST UV/Vis spectrophotometer from Van  
273 Nuys, Los Angeles, CA, USA, to assess TFCs as catechins equivalents per unit of dry matter.

### 274 **2.3.16 Total phenolic content (TPCs)**

275 Folin–Ciocalteu method was employed to determine the TPCs of RRE <sup>53</sup>. RRE solution was  
276 prepared at a concentration of 5 mg/mL. Then 200 µL of RRE was taken into test tubes and  
277 1000 µL of Folin-Ciocalteu reagent was added, followed by vigorous blending and mixing of  
278 solutions. Subsequently, 800 µL of Na<sub>2</sub>CO<sub>3</sub> (7.5%) was introduced to the solution, which was  
279 then left at room temperature for 2 hours. The mixture was thoroughly combined, and the  
280 absorbance of the resulting solution was recorded at 765 nm using an STA–8100ST UV/Vis  
281 spectrophotometer (Van Nuys, Los Angeles, CA, USA). Gallic acid served as the standard for  
282 quantifying the TPCs, which were expressed as GAE (Gallic Acid Equivalent) per dry matter.  
283 Each sample underwent analysis in triplicate.

### 284 **2.3.17 Application of composite coatings on strawberries**

285 The potential of synthesized solutions was evaluated as a coating application on strawberries  
286 to increase their shelf life using dip coating technique <sup>54</sup>. The synthesized composite solutions  
287 of CS/PVA/gelatin incorporating RRE encompass antibacterial and antioxidant potential,  
288 which was believed to provide better protection of strawberries against their early  
289 decomposition. The various solutions used for coatings on strawberries were coded as BS-0,  
290 BS-01, BS-02 and BS-03, with the only difference being of increasing RRE from 0 mL in BS-  
291 0 to 2 mL into BS-01, 4 mL into BS-02 and finally 6 mL into BS-03. Thus, it was believed that  
292 increasing the RRE volume ratio from 0 mL (BS-0) to 6 mL in BS-03 would also enhance their

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293 potential to protect strawberries. Initially, fresh and healthier strawberries were obtained,  
294 cleaned with tissue paper to eliminate dirt, and thoroughly dried. The prepared solutions were  
295 then applied to the dried strawberries using the dip-coating method. Each strawberry's weight  
296 was recorded, and they were stored at room temperature for 17 days with 50% relative  
297 humidity. The decay and deterioration of the strawberries, along with their weight loss, were  
298 regularly monitored. Thus, the study examined the extension of shelf life and preservation of  
299 their freshness. Strawberry samples were tested in triplicate, with their weight loss percent  
300 recorded for each. The average value was then used for calculations.

$$\text{Weight loss (\%)} = \frac{W_i - W_f}{W_i} \times 100$$

304 Where  $W_i$  stands for the initial weight of strawberries and  $W_f$  stands for the final weight of  
305 strawberries.

### 306 2.3.18 Sensory evaluation

307 The sensory assessment of coated strawberries was performed according to Nowacka et al.  
308 (2018)<sup>55</sup> for all formulations including, BS-0, BS-01, BS-02 and BS-03 and a control without  
309 any coating. The parameters such as taste, smell, color, juiciness, firmness and acceptance in  
310 general were noted on a scale of 0 to 10. A score of 8 was seen as exceptional, additional  
311 desired, juicy and acceptable whereas a score of 1 was regarded as poor, extra unwanted, dry  
312 and unsuitable.

### 313 2.3.19 Statistical analysis

314 Statistical analysis of data was run through analysis of variance (Two-way-ANOVA) using the  
315 software Origin Pro 2022, Origin Lab Corporation, Northampton, Massachusetts (USA). All  
316 measurements from the tests were taken in triplicate ( $n = 3$ ), and the data results are presented



317 as the mean  $\pm$  standard deviation (SD). The error bars on column graphs in all figures represent  
318 the standard deviation among three replicates, which is a measure of data variability and  
319 reproducibility. Differences between means were evaluated by Tukey's multiple range test ( $p$   
320  $<0.05$ ).

321

### 322 3. Result and discussion

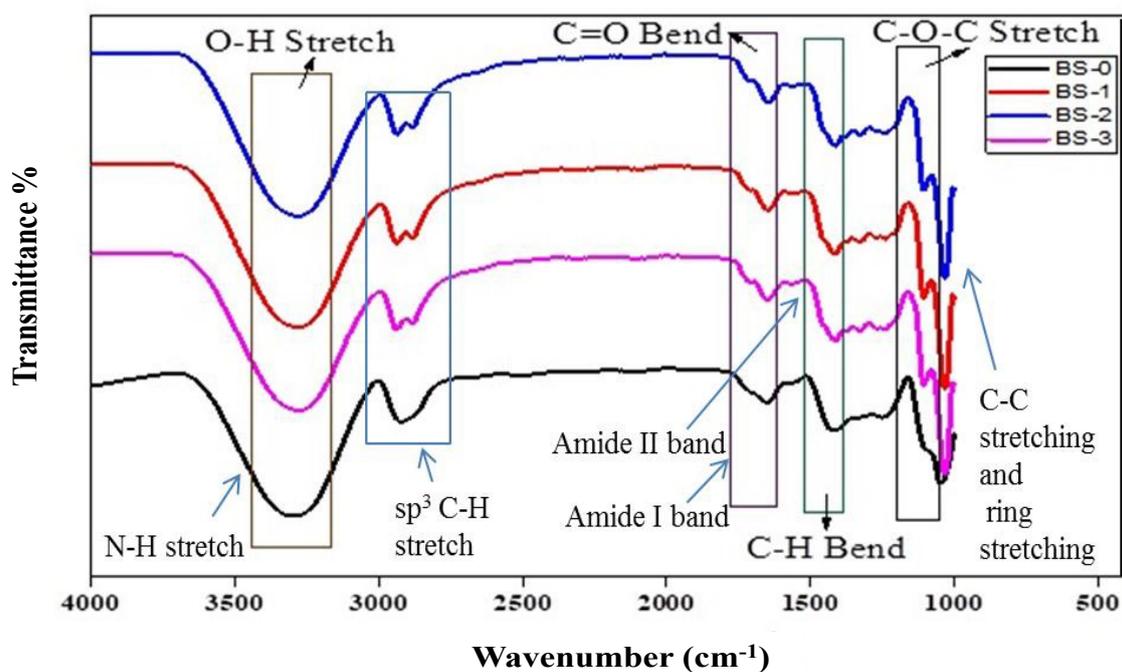
#### 323 3.1 FTIR analysis

324 FTIR analyses were performed to determine the various functional groups present in the films  
325 synthesized from CS/PVA/gelatin and loaded with RRE and to check their intermolecular  
326 interactions. **Figure 2** shows the FTIR spectra of CS/PVA/gelatin films without RRE (BS-0)  
327 and with RRE (BS-01, BS-02 and BS-03). FTIR spectra of all films in **Figure 2** showed a very  
328 broad peak at  $3300\text{ cm}^{-1}$  to  $3250\text{ cm}^{-1}$  due to the overlapping stretching frequencies of hydroxyl  
329 O-H and amino N-H groups <sup>46</sup>. Alkyl C-H group asymmetric and symmetric stretching was  
330 observed between  $2950\text{ cm}^{-1}$  and  $2916\text{ cm}^{-1}$ . The peaks for the amide I and amide II bands  
331 appeared at  $1669\text{ cm}^{-1}$  and  $1564\text{ cm}^{-1}$ , respectively <sup>56</sup>.  $\text{CH}_2$  bending peaks were seen at  $1408$   
332  $\text{cm}^{-1}$  and  $1338\text{ cm}^{-1}$ . C-O and C-O-C stretching vibrations were observed at  $1143\text{ cm}^{-1}$  and  $1099$   
333  $\text{cm}^{-1}$ . These vibration bands were also reported by Dawei Yun *et al.* <sup>57</sup>. Chitosan's ring  
334 stretching vibrations were appeared in the fingerprint region at  $895\text{ cm}^{-1}$  <sup>58</sup>.

335 The FTIR spectra of films with RRE exhibited an additional prominent peak at  $2970\text{ cm}^{-1}$  due  
336 to aromatic C-H stretch<sup>59</sup>, which was due to aromatic polyphenolic and flavonoid compounds  
337 present in RRE. Similarly, FTIR spectra of BS-01, BS-02 and BS-03 exhibited numerous small  
338 peaks from  $1800\text{ cm}^{-1}$  to  $1500\text{ cm}^{-1}$ , due to additional components of RRE <sup>60</sup>. Minor variations  
339 towards lower frequency for certain peaks were observed in FTIR spectra, possibly due to  
340 bonding interactions between CS, PVA, gelatin and RRE. Additional peaks at  $1700\text{ cm}^{-1}$ ,  
341 present in BS-01, BS-02 and BS-03 spectra and absent in BS-0 spectrum, were attributed to



342 various components like secondary metabolites and polyphenols present in RRE <sup>61</sup> The BS-0  
 343 film displayed O-H and N-H stretching at 3330  $\text{cm}^{-1}$ , the band shifted towards lower  
 344 wavenumber at 3267  $\text{cm}^{-1}$  for films incorporated with RRE (BS-01, BS-02 and BS-03), due to  
 345 the greater interactions, created by active functional groups O-H and N-H present within CS,  
 346 PVA, gelatin and RRE. As the OH group present in polyphenols develops hydrogen bonding  
 347 with the OH group present in CS and PVA, and  $\text{NH}_2$  group in chitosan, thus, greater hydrogen  
 348 bonding results in lower frequency ranges. These discussed FTIR results are also supported by  
 349 literature studies <sup>62</sup>. The hydroxyl group (-OH) and amino group (- $\text{NH}_2$ ) stretching vibrations  
 350 in the BS-0 film spectrum were appeared to be greater than those found in BS-01, BS-02, and  
 351 BS-03 spectra. This was due to presence of higher number of unreacted -OH and - $\text{NH}_2$   
 352 functional groups in BS-0 film. The reduction in stretching vibrations in spectra of BS-01, BS-  
 353 02, and BS-03 films containing RRE suggested the involvement of active -OH and - $\text{NH}_2$   
 354 groups in PVA and chitosan with the functional groups present in RRE, leading to the  
 355 formation of hydrogen bonds and other weak electrostatic interactions. Overall, FTIR analysis  
 356 confirmed the presence of the characteristic peaks for all components under consideration.

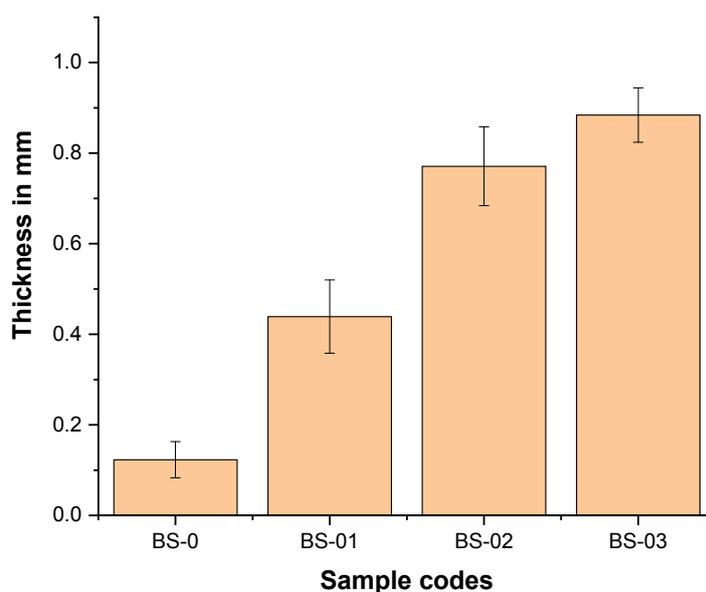


357  
 358



**Figure 2.** FTIR of films of BS-0, BS-01, BS-02 and BS-03View Article Online  
DOI: 10.1039/D5FB00934K359  
360**3.2 Film thickness**

361  
362 The thickness of the films is a vital factor in attaining desirable optical properties and optimum  
363 barrier against water and vapor transfer. An increase in film thickness may reduce the diffusion  
364 rates of oxygen and water vapors of the film <sup>63</sup>. The addition of plasticizers, antioxidants, and  
365 antimicrobial agents can increase the thickness of the films, offering better barrier properties  
366 and improved mechanical strength <sup>64</sup>. The thickness results shown in **Figure 3** indicate that the  
367 BS-0 film (without RRE) showed a minimum thickness of  $0.123\pm 0.02$  mm, while the  
368 maximum thickness was displayed by BS-03 ( $0.884\pm 0.06$  mm), followed by BS-02  
369 ( $0.771\pm 0.05$  mm), and BS-01 ( $0.439\pm 0.03$  mm). Gradually increasing the RRE from 0 mL (BS-  
370 0) to 6 mL (BS-03) also enhanced the film thickness, which is consistent with the literature <sup>65</sup>.  
371 Films with suitable or greater thicknesses offer better barrier properties, thus lower water and  
372 oxygen permeability, making them ideal for prolonged packaging of perishable fruits.



373

**Figure 3.** Film thicknesses of composite films

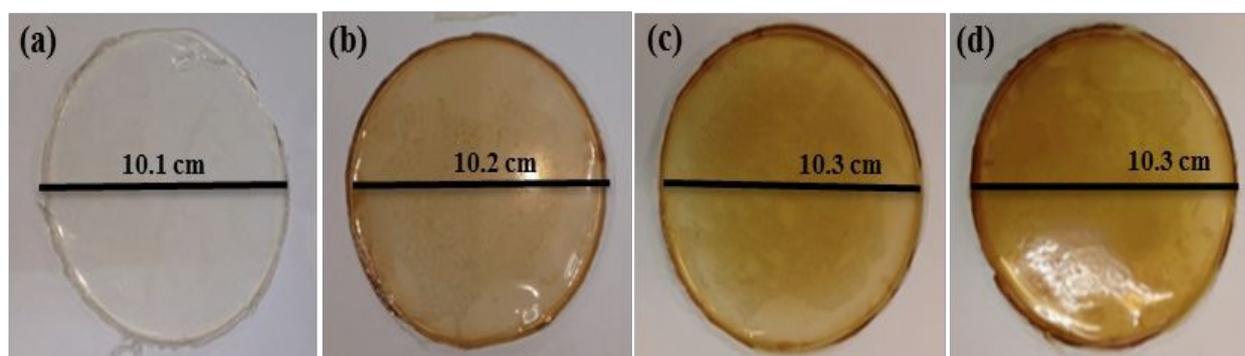
374

**3.3 Visual impact by camera images**

375



376 The visual appearance and color of films are essential parameters that affect the standard and  
 377 quality of food items. Colored films have more potential to wrap food items and preserve them  
 378 because they reduce oxidation reactions in packaged foods <sup>66</sup>. **Figure 4** shows the camera  
 379 images of the CS/PVA/gelatin films and CS/PVA/gelatin films loaded with various volumes  
 380 of RRE. The CS/PVA/gelatin films were completely transparent, whereas the other films  
 381 incorporating RRE were yellowish-brown. It was noted that, by increasing the RRE volume  
 382 from 2 mL in BS-01 to 6 mL in BS-03, the color of films became intense yellowish brown as  
 383 illustrated in **Figure 4**. Additionally, the films were seen as smooth due to better mixing,  
 384 blending and developing certain types of new interactions between film components. The dried  
 385 films were observed to be smooth, homogenous, stretchable, bubble free, mechanically strong,  
 386 flexible and were peeled off from the petri dishes swiftly without cracks.



387  
 388  
 389 **Figure 4.** Visual appearances of composite films; (a) BS-0, (b) BS-01, (c) BS-02, (d) BS-03

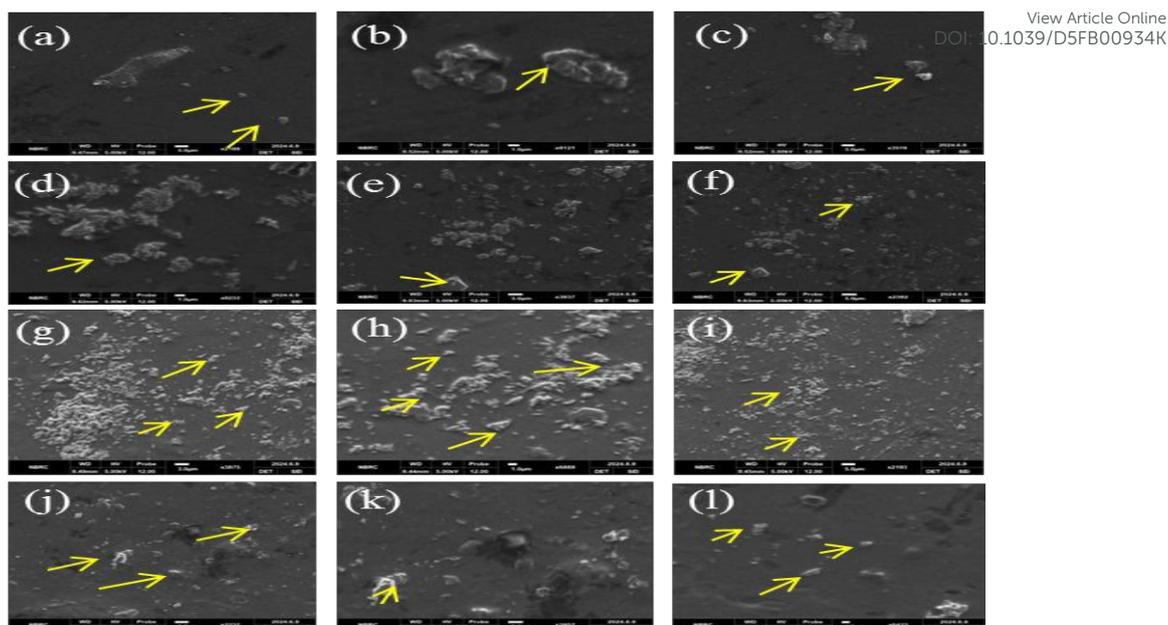
### 390 3.4 Morphological analysis using SEM

391 SEM analysis was used to examine the surface morphology of the prepared CS/PVA/gelatin  
 392 film and CS/PVA/gelatin loaded films with RRE. The SEM images of composite films are  
 393 shown **Figure 5**. The SEM images in **Figure 5 (a, b, c)** of film BS-0 (composed of only  
 394 CS/PVA/gelatin) appeared clearer and more homogenous without cracks and crystalline  
 395 particles/aggregates on their surface, indicating the amorphous nature of this film. The  
 396 micrographs in **Figure 5 (d, e, f)** of film BS-01 showed that all components were uniformly



397 distributed and appeared in one phase and homogeneous. The images showed the presence of  
398 certain aggregates/crystals within the structure, which were due to the addition of 2 mL of RRE  
399 into the CS/PVA/gelatin matrix. The addition of RRE into CS/PVA/gelatin matrix has induced  
400 certain intermolecular interactions, thus producing a few aggregates of some components. The  
401 micrographs in **Figure 5 (g, h, i)** represent BS-02 film incorporating CS/PVA/gelatin loaded  
402 with 4 mL of RRE. Here, the images also revealed the uniformity, consistency and  
403 homogeneity of all the added components. Additionally, BS-02 film also showed many white  
404 crystals, which might have been formed due to the interactions between active functional  
405 groups in CS, PVA, gelatin and RRE. Thus, it was evident from the images that increasing the  
406 RRE ratio from 2 mL (BS-01) to 4 mL (BS-02) had altered the morphology of films to some  
407 extent. The film with a higher volume ratio of RRE (4 mL in BS-02) exhibited more crystalline  
408 molecules/aggregates within its structure, as already reported in the literature <sup>67</sup>. One potential  
409 explanation is the interaction between the active components of RRE and the active groups in  
410 CS, PVA, and gelatin, which leads to the partial release of bioactive compounds such as  
411 polyphenols, alkaloids, and amino acids onto the film's surface <sup>68</sup>. A similar type of interaction  
412 was also reported by Han *et al.* in Noni (*Morinda citrifolia*) fruit polysaccharide enriched with  
413 blueberry leaf extract <sup>69</sup>. It is believed that bridging phenomena occurred inside the bio-  
414 composite films in which some molecules of RRE bonded to the CS chain while the rest of the  
415 functional groups attached to another CS chain. Thus, the overall surface morphology showed  
416 that RRE was effectively incorporated into the CS/PVA/gelatin solution matrix. In contrast to  
417 the above results, **Figure 5 (j, k, l)** represents BS-03 film (incorporating 6 mL of RRE), which  
418 showed film structure as clearer, smoother and more homogenous, without a higher number of  
419 crystalline particles/aggregates on its surface as compared to BS-02 film. Thus, it can be  
420 concluded that increasing RRE to 6 mL, into CS/PVA/gelatin matrix would again make the  
421 film texture smooth <sup>70</sup>.





422

423 **Figure 5.** SEM micrographs of BS-0 (a, b, c), BS-01 (d, e, f), BS-02 (g, h, i) and BS-03 (j, k,  
 424 l)

425

426 The cross-sectional analysis of film BS-0 is shown in **Figure 6 (a)**, which shows that the film  
 427 was homogeneous, uniform, thick, and had no cracks in its structure. Similarly cross-sectional  
 428 SEM image of BS-01 film was more homogeneous, uniform and without any pores as  
 429 compared to BS-02 film, which had certain cracks in its structure.

430 Further cross-sectional images in **Figure 6 (c, d)** revealed that BS-02 and BS-03 films  
 431 incorporating RRE addition to CS/PVA and gelatin were smoother, and their thickness  
 432 increased, respectively. The increase in thickness was primarily due to the increasing volume  
 433 ratios of RRE. The film BS-02 cross-sectional image in **Figure 6 (c)** showed some cracks and  
 434 irregular cavities, which showed its changed morphology as compared to other films. This  
 435 indicated the presence of certain types of intermolecular interactions between functional groups  
 436 of RRE active constituents and CS, PVA and gelatin.

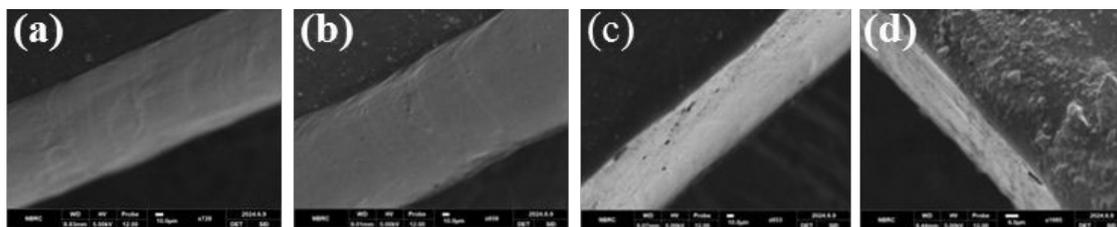
437 The BS-03 film also appeared smooth and without cracks/pores. The homogenous distribution  
 438 of bioactive components within the matrix of films ensures that their antibacterial and



439 antioxidant activities remain unaffected.

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440



441

442

443 **Figure 6.** Cross-sectional film images: BS-0 (a), BS-01 (b), BS-02 (c) and BS-03 (d).

444

### 445 3.5 Mechanical strength

446 The mechanical properties of films are crucial for their use in packaging. To assess the

447 mechanical strengths of these films, stress-strain curves were plotted and are depicted in **Figure**

448 **7.** Two sample films, including BS-0 (incorporating CS/PVA/gelatine) and BS-03

449 (incorporating CS/PVA/gelatine and RRE) were selected for the determination of mechanical

450 properties of films and to evaluate the effect of incorporation of RRE into CS/PVA/gelatin

451 matrix. The BS-0 film stress strain curve was shown by the green curve, while the BS-03 stress

452 strain curve was shown by the red colour. It was noted that the addition of RRE had slightly

453 reduced the mechanical properties of films. The stress value for BS-0 film was noted as 0.32

454 MPa, while the stress value for BS-03 film was 0.21. Similarly, the strain % for BS-01 film

455 was noted as 56 %, while for BS-03 film it was nearly 25 %. This phenomenon can be explained

456 as RRE contains many polyphenolic and flavonoid components, which have active functional

457 groups. These active functional groups had created certain interactions with polymeric chains

458 of CS, PVA and gelatin. Thus, polymeric interactions among CS, PVA and gelatin chains were

459 reduced, which ultimately altered the internal structure of films. It is well known that chitosan

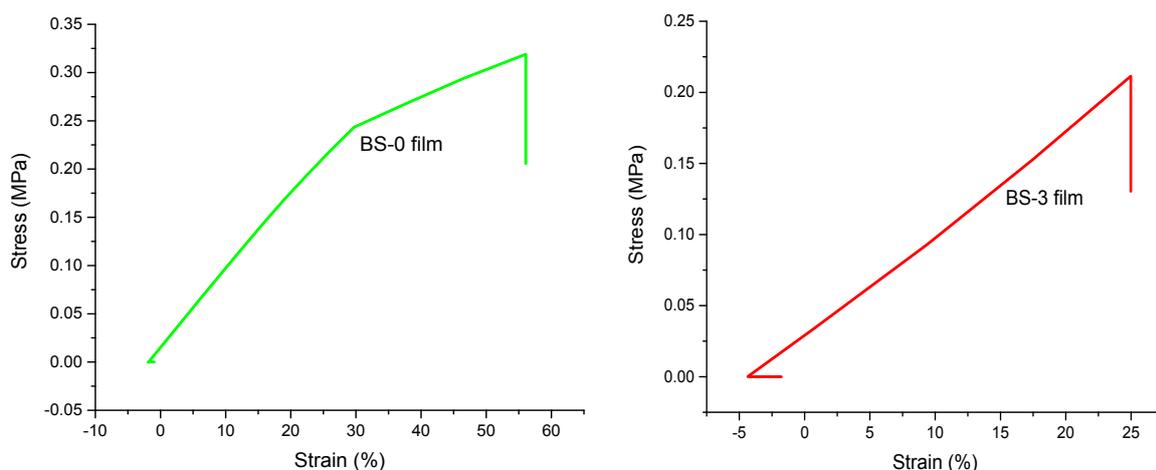
460 has low mechanical strength; thus, it is blended with synthetic polymers like PVA to enhance

461 its mechanical strength. The introduction of aqueous RRE into CS/PVA/gelatin film (BS-03)

462 has reduced its deformation, made the film stiffer and more rigid, and less flexible. The



463 Young's modulus for film BS-0 was calculated as 0.57 MPa, while Young's modulus for film  
 464 BS-03 was calculated as 0.86 MPa. Thus BS-03 film was observed to be less deforming, stiffer  
 465 and more rigid as compared to the BS-01 film. Thus BS-03 film is suitable for applications  
 466 which require more strength and dimensional stability such as packaging applications. While  
 467 the BS-01 film was deformed more under higher stress, which exhibits its more flexibility and  
 468 better suitability for those applications where flexibility and stretchability are required.



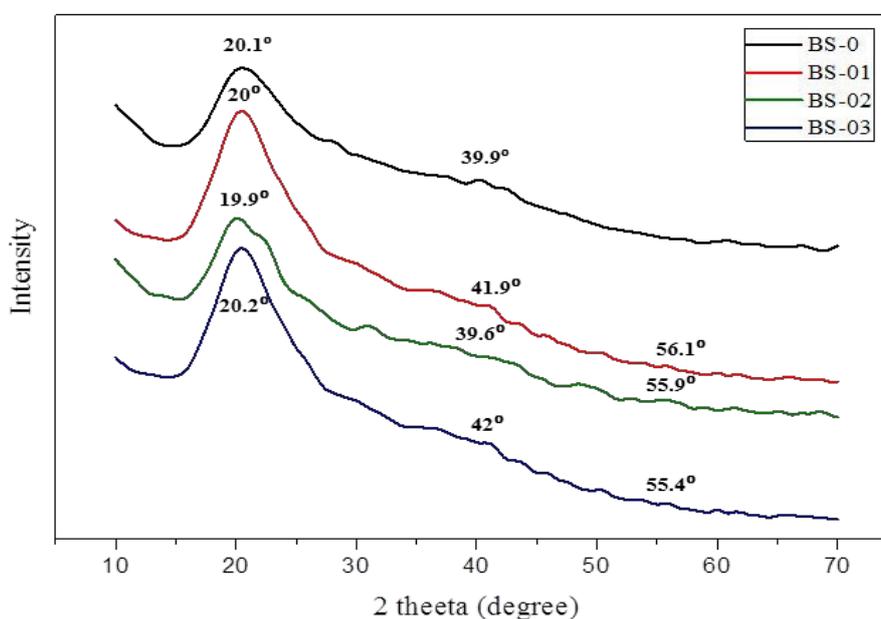
469 **Figure 7.** Mechanical properties of CS/PVA/gelatin and CS/PVA/gelatin/RRE films

### 471 3.6 XRD analysis

472 XRD analysis was performed to determine the crystalline or amorphous nature of composite  
 473 films. Literature studies of PVA and CS XRD spectra exhibited that PVA had a prominent peak  
 474 at  $2\theta = 39.9^\circ$ , while CS exhibited a sharp peak at  $2\theta = 20^\circ$ . The BS-0 coded film demonstrated  
 475 two notable peaks, one at  $2\theta = 19.59^\circ$  corresponding to the crystalline phase of chitosan, and  
 476 another at  $2\theta = 39.9^\circ$  representing the amorphous phase as a lower intensity shoulder. The  
 477 CS/PVA film BS-01 containing RRE also showed two amorphous peaks, slightly shifted to  $2\theta$   
 478  $= 20^\circ$  and  $2\theta = 41.9^\circ$ . The BS-02 film exhibited peaks at  $19.9^\circ$  and  $39.6^\circ$ , while the BS-03 film  
 479 displayed intensified peaks at  $20.2^\circ$  and  $42^\circ$ . Additional peaks were observed in BS-01, BS-  
 480 02, and BS-03 films due to the inclusion of RRE, as illustrated in **Figure 8**. The XRD values  
 481 obtained for CS/PVA/gelatin films and CS/PVA/gelatin loaded with RRE were found to be



482 aligned with previously reported values <sup>71</sup>. These XRD results confirmed the various  
 483 interactions between CS, PVA, gelatin and RRE within the film matrix.



484  
 485 **Figure 8.** XRD patterns of composite films  
 486

### 487 3.7 Moisture content percent of films

488 The moisture content of polymer-based films is the crucial parameter to determine the quality  
 489 of films, as the purpose of films is to preserve the food from microbial spoilage. Here,  
 490 CS/PVA/gelatin and CS/PVA/gelatin enriched with RRE composite films were analyzed for  
 491 their ability to absorb moisture, and the results are displayed in **Figure 9 (A)**, and the values  
 492 are mentioned in **Table 2**. The outcomes of this assay disclosed that all the films absorbed  
 493 moisture content from 15 % to 17 %. Thus, a small variation was observed in moisture content  
 494 results, which could be attributed to their different compositions. The moisture absorption  
 495 phenomenon could be explained based on interactions such as hydrogen bonds between active  
 496 amino and hydroxyl groups of CS, hydroxyl groups of PVA and water. In addition to this, the  
 497 presence of glycerol had also contributed to moisture absorption, as glycerol is a well-known  
 498 humectant. Furthermore, it was noted that the incorporation of RRE into CS/PVA/gelatin



499 matrix had slightly lessened the percent moisture content of the films. As the ratio of RRE was  
500 increased in CS/PVA/gelatin matrix, the moisture content percent was decreased to a small  
501 extent. The reported results were in accordance with literature studies, as it has been mentioned  
502 in literature that CS and PVA produced more interactions with active components of RRE and  
503 ultimately, their interaction with water was decreased, which led to hydrophobicity and a slight  
504 reduction of moisture content <sup>72</sup>. The decreased percent moisture content is most favourable  
505 for food packaging applications as it offers better water barrier properties by lowering the  
506 microbial growth and improves the texture of packaged food <sup>73</sup>.

### 507 **3.8 % Water solubility**

508 The percent solubility of composite films of CS/PVA/gelatin and CS/PVA/gelatin loaded with  
509 RRE in water is the measure of weight loss of films when fully immersed in water for 24 hours.  
510 Solubility percent determination is important to measure as it depicts the stability of films as  
511 and when they come in contact with moisture in the environment for a longer time. The  
512 solubility percent test results were presented in **Figure 9 (B)**, and their exact values were also  
513 given in **Table 2**. It was noticed that CS/PVA/gelatin film BS-0 had shown a maximum  
514 solubility % of 73%, which is mainly due to the presence of hydrophilic polymers of PVA and  
515 gelatin. However, CS is hydrophobic, and it makes interactions with both PVA and gelatin  
516 within films to decrease their solubility. The addition of RRE into CS/PVA/gelatin films had  
517 decreased their solubility, as BS-01 code film solubility was 70.2 %. Furthermore, increasing  
518 the volume ratio of RRE into CS/PVA/gelatin films further decreased the solubility of films,  
519 as observed in BS-02 (63.2 % solubility) and BS-03 films (60.2 % solubility). The intense  
520 interaction between CS and other polymers such as PVA, gelatin, and RRE led to increased  
521 intermolecular bonding, which in turn reduced their solubility <sup>74</sup>. The films with lower  
522 solubility rates are more beneficial for food packaging applications, as these films are more  
523 likely to compromise with the moisture present in the surroundings, and their dissolution rate



524 slows down. However, a balance between solubility and biodegradability is important, as  
525 excessively low solubility may slow down the degradation process post-use. A similar  
526 phenomenon of reducing the solubility of films by incorporating tea extract was also reported  
527 by Hafsa *et.al* <sup>75</sup>.

### 528 3.9 Water absorption

529 Water absorption percent of films is a vital indicator to check their quality to absorb and retain  
530 water within their structure without being dissolved or solubilized. The degree of water  
531 absorption was found to be inversely proportional to the degree of interactions or crosslinking  
532 among different constituents of films. More interactions or crosslinking between polymeric  
533 chains show lesser water absorption and vice versa. The films with lower water absorption  
534 percent are frequently opted for food packaging applications. The results of the water  
535 absorption percent test are shown in **Figure 9 (C)** and listed in **Table 2**. The results exhibited  
536 that the BS-0 film absorbed maximum water up to 225.3 % for 30 minutes. The BS-01, BS-02  
537 and BS-03 films showed 97.7 %, 71.4 % and 58.23 % of water absorption, respectively. The  
538 higher absorption by BS-0 film can be attributed to the presence of hydrophilic polymers such  
539 as PVA and gelatin, which, in the absence of RRE, had made more interactions with water and  
540 thus absorbed more. The other films showed less absorption due to the presence of RRE and  
541 more interactions/crosslinking of polymeric chains with RRE instead of water molecules. This  
542 is evident as RRE volume ratio is enhanced; % water absorption is decreased, due to apparently  
543 development of higher degree of cross linking which resulted in lower % water absorption <sup>76</sup>.

544

545

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547 **Table 2.** Film thickness, % moisture content, % solubility and % water absorption mean values  $\pm$  standard deviation were presented. View Article Online  
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Sr. No.	Sample codes	Film thickness (mm)	Moisture content (%)	Water solubility (%)	Water absorption (%)
1	BS-0	0.123 $\pm$ 0.04	17.42 $\pm$ 1.97	73 $\pm$ 3.6	225.3 $\pm$ 5.25
2	BS-01	0.439 $\pm$ 0.081	16.51 $\pm$ 2.52	70.2 $\pm$ 2.52	97.7 $\pm$ 3.44
3	BS-02	0.771 $\pm$ 0.087	16.41 $\pm$ 2.35	63.2 $\pm$ 3.07	71.4 $\pm$ 1.97
4	BS-03	0.884 $\pm$ 0.06	15.14 $\pm$ 1.46	60.2 $\pm$ 4.35	58.23 $\pm$ 2.31

549

### 550 3.10 Water drop test

551 Water drop test was performed on composite films to check their ability to pass water or not.  
552 The films that hold water appeared to be decent for their application in food packaging. The  
553 test results for all films were displayed in **Figure 9 (D)**, and the results showed that the films  
554 did not allow the water drop to pass through them. The films or other materials which show  
555 superior water barrier properties are often opted for in food packaging applications <sup>77</sup>.

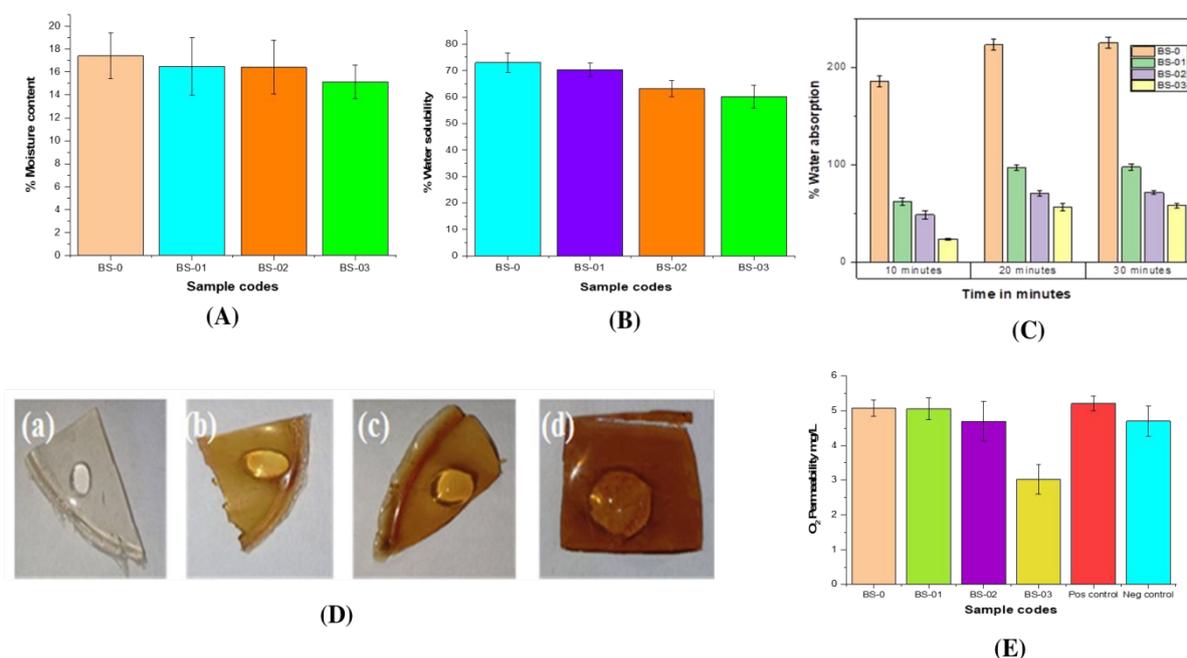
### 556 3.11 Oxygen permeability

557 Oxygen permeability of films has a noteworthy effect on the quality and shelf life of packaged  
558 food items. When oxygen is entered through films, it causes spoilage of food items by  
559 oxidation, microbial growth and colour changes. Thus, lower rates of oxygen permeability can  
560 inhibit the oxidative spoilage and growth of aerobic microbes, thus leading to extended shelf  
561 life of perishable food products like strawberries. The oxygen present in the atmosphere can  
562 also weaken the bond energy of polymers, and thus unstable peroxides are formed. These  
563 unstable species slow down the degradation rate of polymeric films and thus make them  
564 unsuitable for food packaging applications. The oxygen permeability of composite films was  
565 evaluated by the method already discussed by Wrinkler, and the results were presented in  
566 **Figure 9 (E)** <sup>78</sup>.

567 The O<sub>2</sub>-permeability test results for an airtight flask (negative control) and an open flask  
568 (positive control) were 4.7 $\pm$ 0.43 mg/L and 5.21 $\pm$ 0.22 mg/L, respectively. The oxygen



569 permeability values calculated for each film were  $5.08 \pm 0.23$  mg/L (BS-0),  $5.05 \pm 0.31$  mg/L  
 570 (BS-01),  $4.69 \pm 0.57$  mg/L (BS-02) and  $3.03 \pm 0.43$  mg/L (BS-03). The data demonstrated that  
 571 incorporating RRE reduced oxygen permeability, due to increasing RRE volume, which caused  
 572 free volumetric holes in films to contract, making it more difficult for oxygen molecules to  
 573 penetrate. Thus, lower oxygen permeability is a desirable characteristic for polymers,  
 574 particularly in food packaging applications <sup>79</sup>.



575  
 576 **Figure 9.** (A) % moisture content (B) % water solubility (C) % % water absorption (D) water  
 577 drop test on films (a) BS-0 (b) BS-01 (c) BS-02 (d) BS-04 and (E) oxygen permeability results  
 578 of the prepared films

### 579 3.12 Antibacterial potential

580 Natural plant extracts incorporating antibacterial components are more sterile, eco-friendly,  
 581 and biodegradable. They are regarded as excellent materials to be used in food packaging  
 582 against food pathogens <sup>80</sup>. The plant extracts contain bioactive phenolic and flavonoids  
 583 components, and the hydroxyl groups in these compounds provide the basis for their  
 584 antibacterial activity. The hydroxyl groups develop certain interactions, such as hydrogen

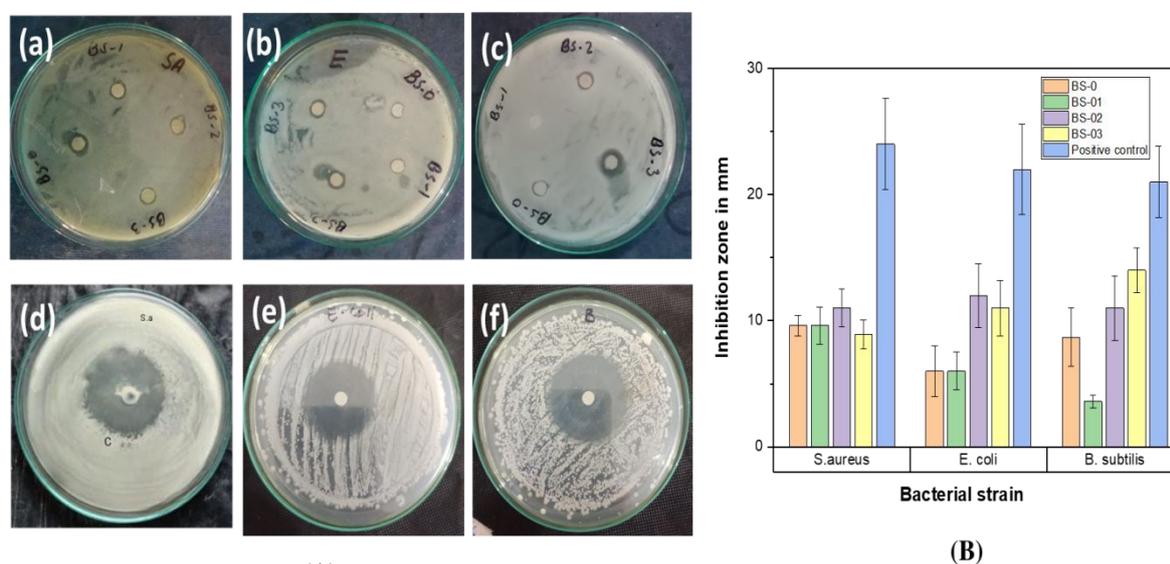


585 bonds, with the bacterial cell membrane. Based on their hydrophobic nature, these phenolic  
586 compounds may accumulate on the surface of cells or may invade the cytoplasm of microbes.  
587 After reaching the cytoplasm, these components interact with the other cellular organelles and  
588 cell's components; thus, they change the pH inside the cell membrane<sup>81</sup>. The bacterial cell wall  
589 has negative carboxylate groups ( $-\text{COO}^-$ ) and the chitosan carry positive amino groups ( $-\text{NH}_3^+$ ).  
590 The electrostatic interactions of these groups with each other provide the mechanism  
591 for antibacterial activity<sup>82</sup>. CS/PVA/gelatine and CS/PVA/gelatine incorporating RRE  
592 solutions were assessed for antibacterial potential against *S. aureus*, *B. subtilis* and *E. coli*, and  
593 results are shown in **Figures 10 (A) and 10 (B) and Table 3**. The effectiveness of each coating  
594 solution was determined by measuring the inhibition zone in mm. All the coating solutions  
595 exhibited fair to good antibacterial potential against all three bacterial strains in comparison to  
596 the control. The results demonstrated that in most tests, the BS-03 sample showed better results  
597 than other samples. This was due to the presence of a higher volume ratio of RRE in the BS-  
598 03 sample. When we compared the results of all samples, BS-03 showed higher % inhibition  
599 against *S. aureus* with the lowest  $\text{IC}_{50}$  value of  $78.65 \pm 0.13$ . Similarly, the BS-02 sample  
600 exhibited higher activity against *E. coli* with the lowest  $\text{IC}_{50}$  value of  $118.6 \pm 0.021$ ; while the  
601 BS-03 sample exhibited the highest potential against *B. subtilis* with the lowest  $\text{IC}_{50}$  value of  
602  $128.75 \pm 0.65$ . Overall, BS-02 and BS-03 samples incorporating higher volume ratios of RRE  
603 showed better antibacterial potential against all three bacterial strains as compared to BS-0 and  
604 BS-01, attributed to the presence of various active components (phenolic compounds,  
605 flavonoids, and others) in the RRE. Notably, the increasing volume of RRE correlated with  
606 improved antibacterial activity.

607 Based on these findings, the synthesized CS/PVA/gelatine/RR films, especially BS-02 and BS-  
608 03, show promise for food packaging applications. Similar antibacterial results were also  
609 reported by Dan *et al.*<sup>83</sup> for chitosan-gelatin film incorporating hop plant extract. The natural



610 compounds found in plants are considered ideal candidates for food packaging applications  
 611 due to their antibacterial properties. The two-way ANOVA demonstrated that treatment type  
 612 significantly influenced antibacterial activity ( $p < 0.001$ ), and its effect varied among bacterial  
 613 species. The positive control produced the strongest inhibition zones, BS-02 and BS-03 had  
 614 moderate effects, while BS-0 and BS-01 showed minimal inhibition. These results suggest that  
 615 specific treatments possess promising antibacterial potential depending on the bacterial species  
 616 tested.



618  
 619 **Figure 10.** (A) Anti-bacterial images of samples against (a) *S. aureus* (b) *E. coli* (c) *B. subtilis*,  
 620 and (d, e, f) representing positive controls for *S. aureus*, *E. coli* and *B. subtilis*, respectively;  
 621 (B) Anti-bacterial potential of samples in terms of inhibition zone in mm

622  
 623 **Table 3.** Antibacterial properties of all synthesized sample solutions

Sample code	Inhibition zone (mm)	IC <sub>50</sub> at 100 mg/ml	Percent inhibition
<i>S. aureus</i>			
BS-0	9.6	172.91±0.0542	17.12±0.0542
BS-01	9.6	122.91±0.047	31±0.047
BS-02	11	93.6±0.049	38.57±0.049
BS-03	8.9	78.65±0.13	38.57±0.13



Control	24	50 $\pm$ 0.9	100.714
<i>E. coli</i>			
BS-0	6	120 $\pm$ 0.014	40 $\pm$ 0.014
BS-01	6	115 $\pm$ 0.09	40 $\pm$ 0.09
BS-02	12	118.6 $\pm$ 0.021	33.6 $\pm$ 0.021
BS-03	11	120 $\pm$ 0.028	23 $\pm$ 0.028
Control	22	50 $\pm$ 0.8	100
<i>B. subtilis</i>			
BS-0	8.7	138 $\pm$ 0.604	40 $\pm$ 0.234
BS-01	3.6	225 $\pm$ 0.234	39.47368421
BS-02	11	189.64 $\pm$ 0.28	26.10526316 $\pm$ 0.2857 1
BS-03	14	128.75 $\pm$ 0.65	37.89473684 $\pm$ 0.65
Control	21	50.85 $\pm$ 0.714	100.714

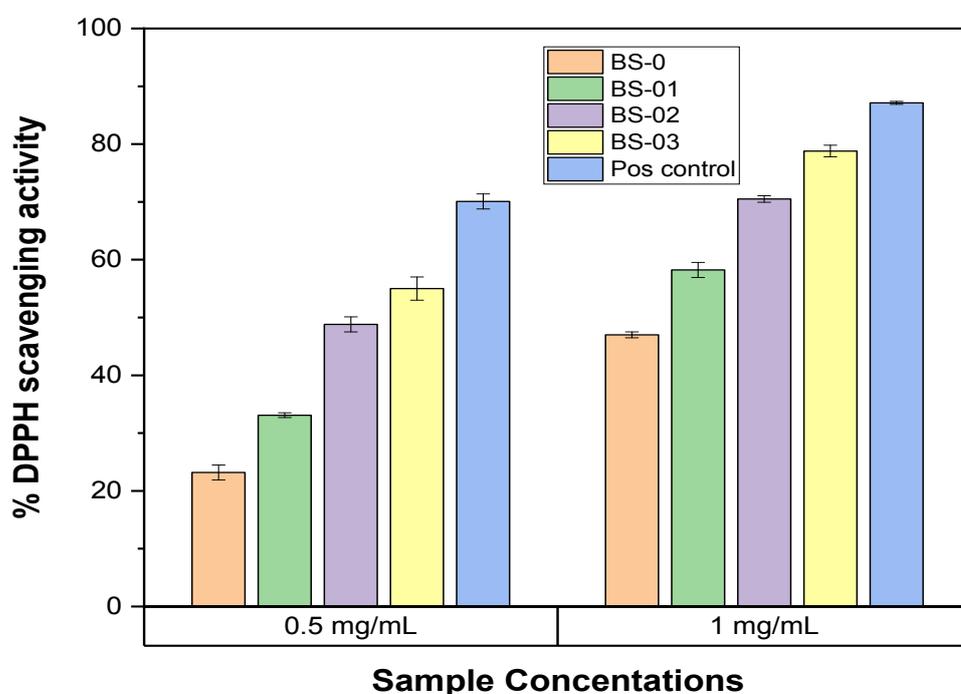
624

625 **3.13 Antioxidant potential**

626 The enhancement in food shelf life and its preservation applications depend upon the  
627 antioxidant potential of synthesized coating solutions and films. Oxidation reactions cause  
628 discoloration of food and reduce its flavor. Antioxidant materials are required to deactivate and  
629 scavenge free radicals. The antioxidant potential of solutions was determined by the 2, 2-  
630 diphenyl-1-picrylhydrazyl (DPPH) assay (**Table 4**). DPPH is a nitrogen-free stable radical of  
631 deep purple color, which turns yellow after reacting with antioxidants. Antioxidant capacity  
632 was determined using the IC<sub>50</sub> values, which represent the sample mass per mL of DPPH  
633 required to reduce the DPPH concentration by 50%. The results in **Figure 11** indicated that the  
634 BS-03 film demonstrated superior antioxidant properties (78.8 $\pm$ 1.01 with IC<sub>50</sub> value of 0.3  
635 mg/mL) attributed to the presence of plant extract (RRE) in excess. Similarly, BS-02 sample  
636 showed antioxidant potential of 70.5 $\pm$ 0.57 with IC<sub>50</sub> value of 0.5 mg/mL. The BS-01 film  
637 exhibited 58.23 $\pm$ 1.3 with IC<sub>50</sub> value of 0.7 mg/mL. The lowest antioxidant activity was  
638 exhibited by BS-0 sample having value of 47.0 $\pm$ 0.51 with IC<sub>50</sub> value of 1.1 mg/mL. The  
639 enhanced antioxidant activities of BS-03, BS-02 and BS-01 as compared to BS-0 were clearly



640 attributed to the numerous active components found in RRE, including flavonoids, phenolics,  
 641 citronellol, nerol, quercetin, catechin, epicatechin, rutin and phenethyl alcohol. These  
 642 compounds prevent oxidative degradation, thus preventing food from microbial spoilage and  
 643 resulting in prolonging freshness. The results were also in accordance to the literature analysis  
 644 <sup>84</sup>. The statistical analysis reveals that both concentration and treatment type have highly  
 645 significant effects on antioxidant activity. Furthermore, there is a significant interaction  
 646 between these factors, indicating that the effect of each treatment depends on the concentration  
 647 used. Thus, higher concentration significantly increases antioxidant activity ( $p < 0.0001$ ).



648

649

**Figure 11.** Antioxidant potential of sample films and positive control

650

**Table 4.** Antioxidant activities of all samples

Sample code	DPPH assay		IC <sub>50</sub> mg/ml
	Percent inhibition of DPPH free radicle		
	0.5mg/ml	1mg/ml	
BS-0	23.20±1.3	47.0±0.51	1.1
BS-01	33.1±0.4	58.23±1.3	0.7
BS-02	48.8±1.3	70.5±0.57	0.5



BS-03	55.0 $\pm$ 2.0	78.8 $\pm$ 1.01	0.3
Positive control (Ascorbic acid)	70.08 $\pm$ 1.3	87.14 $\pm$ 0.26	0.36

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652 **3.14 Biodegradation of composite films:**

653 A soil burial degradation test was conducted to evaluate the decomposition rate of the prepared  
654 films. The weight loss percentage with respect to the number of days for BS-0, BS-01, BS-02,  
655 and BS-03 films was measured. This method effectively indicated film biodegradability due to  
656 the presence of microbes and moisture in soil. The biodegradation process is initiated with  
657 water absorption, which disrupts hydrogen bonding between polymeric chains, followed by  
658 microbial invasion from bacteria, algae, and fungi. Biodegradation is a natural process and is  
659 also influenced by the chemical composition of composite films. Various factors affect  
660 polymeric film degradation rates, including pollutant quantity, microbial presence, and  
661 environmental conditions (oxygen, humidity, pH, and temperature). **Figure 12 (A)** displays the  
662 degraded film images after 18 and 45 days, while **Figure 12 (B)** shows the % biodegradation  
663 after 18 and 45 days. The results data showed that BS-02 film demonstrated the highest %  
664 degradation of 91.29 %, followed by BS-03 film 86.79 % and BS-01 film 85.91 %, while BS-  
665 0 film showed % degradation of 81.48%. It was observed that incorporation of RRE into  
666 CS/PVA/gelatin has improved % biodegradation of films. The biodegradation rates were  
667 higher as compared to already reported results by Riaz *et al.*,<sup>85</sup>. The increased %  
668 biodegradation, due to the incorporation of RRE, was attributed to the presence of carbon,  
669 nitrogen, and oxygen in many components of RRE, which served as food sources for  
670 microorganisms. Continuous water addition activates soil bacteria and weakens polymeric  
671 interactions between components, thus increasing degradation rates<sup>86</sup>. The incorporation of  
672 RRE into CS/PVA/gelatin films improved their biodegradability, transforming them into a  
673 rapidly degradable and eco-friendly material<sup>85</sup>. The faster degradation of these biopolymer  
674 films in soil environments provides a sustainable alternative with reduced environmental



675 impact.

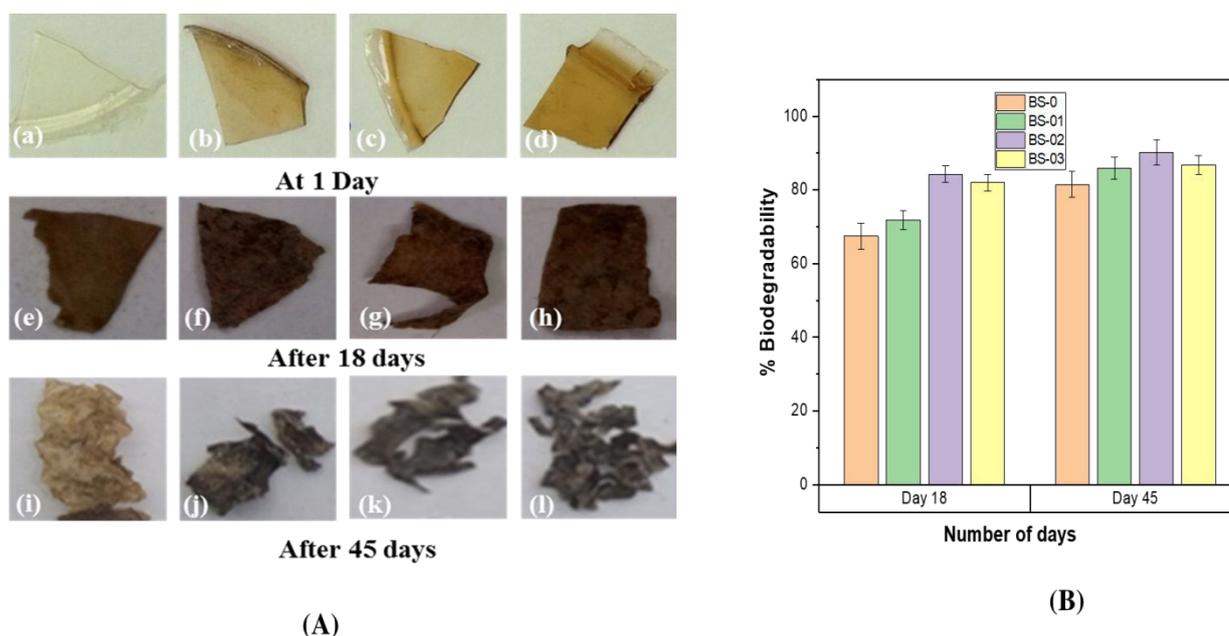
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681 **Figure 12.** (A) Images of composite films (a) BS-0; (b) BS-01 (c) BS-02 (d) BS-03 before  
682 experimentation; (e) BS-0 (f) BS-01 (g) BS-02 (h) BS-03 after day 18; (i) BS-0 (j) BS-01 (k)  
683 BS-02 (l) BS-03 after day 45; (B) % biodegradability at day 18 and day 45 of prepared films

684

### 685 3. 15 Total phenolic contents (TPCs) and total flavonoid contents (TFC)

686 Polyphenols constitute the most frequent and broadly distributed group of compounds in the  
687 plant kingdom, with more than 8000 currently known compounds<sup>87</sup>. The rose flower has high  
688 polyphenolic compounds and essential oil, and thus serves as a key source of many raw  
689 materials for cosmetics, herbal teas and rose oil<sup>88</sup>. The *Rosa rubiginosa* extracts (RRE) TPCs  
690 and TFCs were expressed in  $\mu\text{g}$  of Gallic acid equivalent (GAE) and  $\mu\text{g}$  of Quercetin equivalent



691 (QE) per mg of extract, respectively. The TPCs of RRE were found to be  $86.74 \pm 0.91$   $\mu\text{g}$   
692 GAE/mg. These values are close to as described by Murathan *et al.*<sup>89</sup> and Fascella *et al.*<sup>90</sup>  
693 Literature studies have demonstrated that among phenolic compounds, p-coumaric acid and  
694 chlorogenic acid were present abundantly in RRE extract. It was observed that the  
695 phytochemical properties of various rose species are greatly dependent upon their type of  
696 species. The RRE was found to be a promising source of natural antioxidants, which are useful  
697 in the pharmaceutical and food industries<sup>91</sup>. Similarly, TFCs of RRE were found to be  
698  $26.73 \pm 0.68$   $\mu\text{g}$  QE/mg. These values were also in accordance with already reported values by  
699 Tabaszewska *et al.*<sup>92</sup> Quercetin and Letulin are key flavonoid aglycons in Rose species.  
700 Quercetin and Lutulin belong to the flavonol and flavone groups, respectively<sup>93</sup>.

### 701 3.16 Application on strawberries

702 Coatings offer an effective solution to extend the shelf life of fruits and vegetables.  
703 Strawberries, being delicate and water-rich, have a limited shelf life and are prone to spoilage  
704 after harvesting due to pathogen attacks. Unpackaged or uncoated foods also deteriorate due to  
705 water evaporation and cellular respiration. All the composite coating solutions of BS-0, BS-01,  
706 BS-02 and BS-03 were used for application on strawberries to evaluate their potential to protect  
707 and enhance shelf life of strawberries. Uncoated strawberries were used as a negative control  
708 for comparison of results. The weight loss % was monitored after 1<sup>st</sup>, 2<sup>nd</sup>, 6<sup>th</sup>, 7<sup>th</sup>, 8<sup>th</sup> and 17<sup>th</sup>  
709 day. **Figures 13 (A)** and **13 (B)** were used to represent results in terms of images of  
710 strawberries and weight loss % over time (in days), respectively. Results indicated that coating  
711 solutions with higher RRE ratios exhibited better protection of strawberries and provided  
712 greater shelf life, as indicated by both images and weight loss % graphs. Similar results were  
713 also discussed in literature by Mukhtar *et al.*,<sup>94</sup>. The results showed that uncoated strawberries  
714 (control) started to deteriorate slowly after the 2<sup>nd</sup> day and experienced a 60 % weight loss after  
715 7<sup>th</sup> days. For coated strawberries using BS-0, BS-01, BS-02, and BS-03 coating solutions, the



716 weight loss percentages on day 6<sup>th</sup> were 39.82 %, 34.87 %, 32.12 %, and 29.28 %, respectively  
717 compared to 45.55 % for uncoated strawberries. Similarly, after 7<sup>th</sup> day, the weight loss % was  
718 45.35 % for BS-0, 39.09 % for BS-01, 35.05 % for BS-02, 32.23 % for BS-03 and 60.59 % for  
719 the negative control. It can be inferred from **Figure 13 (A)** that initial weight loss up to 6 days  
720 was mainly due to the loss of water content from strawberries, and then after 6 or 7 days, fungal  
721 attack began, which started to degrade the strawberries.

722 By day 17<sup>th</sup>, weight loss % reached 71.09 % for BS-0, 66.32 % for BS-01, 60.07 % for BS-02,  
723 57.2 % for BS-03 and 89.03 % for the uncoated negative control. It was noted that all coating  
724 solutions preserved strawberries well and enhanced their shelf life up to 8 days without being  
725 deteriorated by fungal attack. After 8 days, the strawberries coated by BS-0, BS-01 and BS-02  
726 started to deteriorate by fungal attack more rapidly as compared to BS-03 coating solution,  
727 which remained intact up to 10 days. Thus, it can be concluded that increasing the volume ratio  
728 of RRE in CS/PVA/gelatin-based coating solutions has enhanced the shelf life of strawberries  
729 up to 10 days. The BS-03 coating solution demonstrated the best performance in reducing %  
730 weight loss, suggesting its highest potential for preserving delicate food items. The results of  
731 fabricated coatings on strawberries were also compared with literature studies, and the results  
732 are presented in **Table S1** in the supplementary information. The literature studies showed the  
733 strawberries were stored for 5 days at room temperature and for 15 days at 4 °C, but our  
734 fabricated coatings protected strawberries for 10 days at room temperature.

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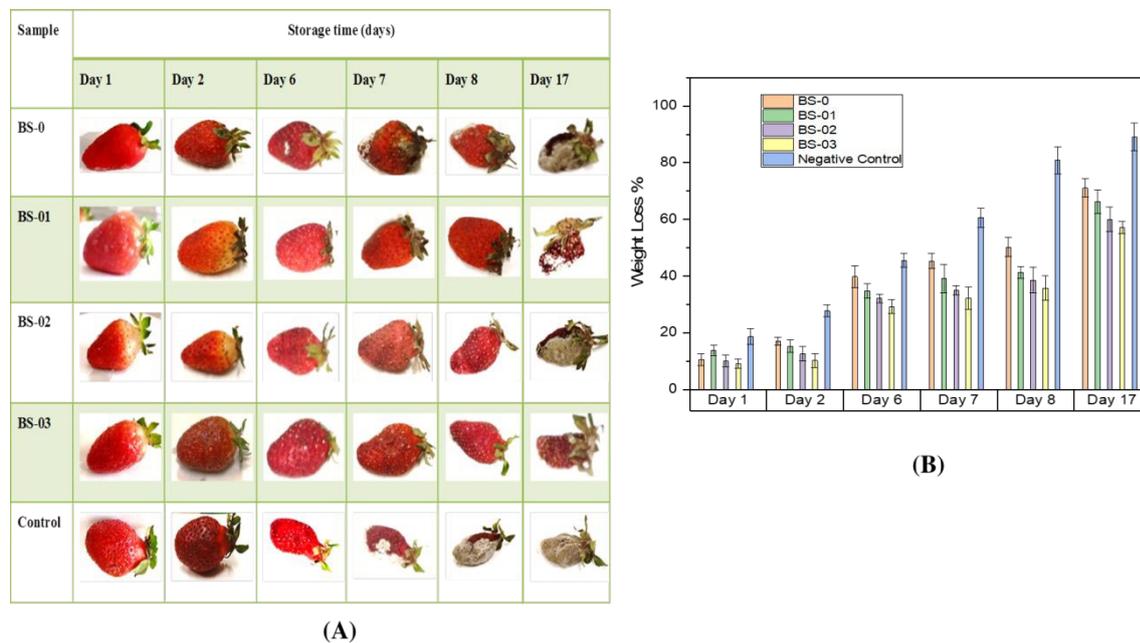
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745 **Figure 13. (A)** Images representing application of coatings on strawberries and their  
 746 deterioration with respect to the number of days; Control represents uncoated strawberries  
 747 **(B)** Weight loss % of strawberries from day 1 to day 17

### 748 3.17 Sensory evaluation

749 The results of sensory evaluation for each formulation coded as BS-0, BS-01, BS-02 and BS-  
 750 03 and a control (without any coating) are presented in **Tables S2 to S-6**. The results exhibited  
 751 that strawberries coated with BS-03 formulation showed the most favourable and better results  
 752 in terms of taste, smell, color, juiciness, firmness and acceptability in general as compared to  
 753 BS-0, BS-01 and BS-02 formulations.

754 This might be due to the presence of highest amount of RRE in this formulation, which  
 755 provided enhanced antioxidant and antibacterial properties to this formulation. The BS-0  
 756 formulation exhibited results of parameters such as taste, smell, color, juiciness, firmness and  
 757 general acceptability mostly in neither good nor bad and like somewhat categories. Similarly,



758 BS-01 showed most of the studied parameters in like somewhat category. The BS-02 coating  
759 exhibited parameters in like somewhat and moderate category. The BS-03 coating showed most  
760 attractive and convincing results of studied parameters in similar to moderate and greatly  
761 admire category. The control strawberries without any coating were subjected to the highest  
762 degradation and changes, thus their study parameters mostly fall under mildly disliked and  
763 mildly unfavourable categories.

## 764 5. Conclusion

766 This study demonstrated that CS/PVA/gelatin composite coatings enriched with RRE enhanced  
767 the preservation of strawberries. Increasing RRE volume ratio into CS/PVA/gelatin coatings  
768 (from BS-01 to BS-03) thickened the films and improved their barrier performance, while %  
769 moisture content, water solubility water absorption, and oxygen permeability were decreased.  
770 Antimicrobial tests showed that higher volume of RRE containing coatings, particularly BS-  
771 02 and BS-03, exhibited stronger activities against *S. aureus*, *B. subtilis*, and *E. coli* than the  
772 RRE free coating (BS-0). Antioxidant capacity also increased with enhancing RRE ratio, with  
773 BS-03 reaching  $78.8 \pm 1.01\%$  DPPH inhibition ( $IC_{50} = 0.3$  mg/mL). Soil burial trials confirmed  
774 that all films were biodegradable; the highest 45-day weight loss was observed for BS-02  
775 (91.29%), followed by BS-03 (86.79%). Although the incorporation of RRE reduced ductility  
776 and increased stiffness, the mechanical properties of BS-03 film would be more suitable for  
777 handling and packaging applications. When applied to strawberries, all coatings helped reduce  
778 weight loss compared to the uncoated control and prolonged their shelf life. Among them, BS-  
779 03 was the most effective, preventing visible spoilage for up to 10 days and achieving the  
780 highest sensory scores. Overall, the BS-03 formulation emerges as a promising bio-based  
781 coating for the short-term extension of berry shelf life. Future research should focus on  
782 optimizing composition through response surface methodology, conducting antifungal tests,



783 evaluating performance during refrigerated storage, and examining component migration for  
 784 applications involving direct contact with food.

### 785 **Author Contributions**

786 B.S.: Conducted experiments, investigations, data analysis and writing original draft. M.Z.:  
 787 Conceptualization, writing original draft, reviewing and editing. A.Y.: performed antibacterial  
 788 assessment. F.A.: Reviewing. S.S.: Conceptualization, supervision, editing and reviewing.  
 789 A.U.: Conceptualization, supervision, editing and reviewing. All authors have read and agreed  
 790 to the published version of the manuscript.

### 791 **Conflicts of interest**

792 There are no conflicts to declare.

### 793 **Data availability**

794 The data that support the findings of this study are available from the corresponding authors  
 795 upon reasonable request.

### 796 **References**

- 797 1. L. Kalra, R. Anushree and A. W. Sultani, *Indian Journal of Ecology*, 2024, **51**, 697-706.
- 798 2. FAO, *The state of food and agriculture 2019: Moving forward on food loss and waste*  
 799 *reduction*, UN, 2019.
- 800 3. L. Brennan, S. Langley, K. Verghese, S. Lockrey, M. Ryder, C. Francis, N. T. Phan-Le and A. Hill,  
 801 *Journal of cleaner production*, 2021, **281**, 125276.
- 802 4. S. Wardejn, S. Waclawek and G. Dudek, *International Journal of Molecular Sciences*, 2024,  
 803 **25**, 12580.
- 804 5. S. SUMNU and L. Bayindirli, *Journal of the Science of Food and Agriculture*, 1995, **67**.
- 805 6. L. Pavoni, D. R. Perinelli, G. Bonacucina, M. Cespi and G. F. Palmieri, *Nanomaterials*, 2020,  
 806 **10**, 135.
- 807 7. F. Salehi, *International Journal of Fruit Science*, 2020, **20**, S570-S589.
- 808 8. A. F. Pires, O. Díaz, A. Cobos and C. D. Pereira, *Foods*, 2024, **13**, 2638.
- 809 9. M. Mujtaba, R. E. Morsi, G. Kerch, M. Z. Elsabee, M. Kaya, J. Labidi and K. M. Khawar,  
 810 *International journal of biological macromolecules*, 2019, **121**, 889-904.
- 811 10. F. Mohanty and S. Swain, in *Nanotechnology applications in food*, Elsevier, 2017, pp. 363-  
 812 379.



- 813 11. E. Ojogbo, E. O. Ogunsona and T. H. Mekonnen, *Materials Today Sustainability*, 2020, **7**, 100028. View Article Online  
DOI: 10.1039/D5FB00934K
- 814
- 815 12. Z. Wu, J. Wu, T. Peng, Y. Li, D. Lin, B. Xing, C. Li, Y. Yang, L. Yang and L. Zhang, *Polymers*, 2017,
- 816 **9**, 102.
- 817 13. N. Ruocco, S. Costantini, S. Guariniello and M. Costantini, *Molecules*, 2016, **21**, 551.
- 818 14. M. Friedman and V. K. Juneja, *Journal of Food Protection*, 2010, **73**, 1737-1761.
- 819 15. M. Kong, X. G. Chen, K. Xing and H. J. Park, *International journal of food microbiology*, 2010,
- 820 **144**, 51-63.
- 821 16. L. A. van den Broek, R. J. Knoop, F. H. Kappen and C. G. Boeriu, *Carbohydrate polymers*,
- 822 2015, **116**, 237-242.
- 823 17. M. A. Del Nobile, N. Di Benedetto, N. Suriano, A. Conte, M. R. Corbo and M. Sinigaglia, *Food*
- 824 *Microbiology*, 2009, **26**, 587-591.
- 825 18. A. Verlee, S. Mincke and C. V. Stevens, *Carbohydrate polymers*, 2017, **164**, 268-283.
- 826 19. M. Hosseinejad and S. M. Jafari, *International journal of biological macromolecules*, 2016,
- 827 **85**, 467-475.
- 828 20. D. C. Vodnar, O. L. Pop, F. V. Dulf and C. Socaciu, *Notulae Botanicae Horti Agrobotanici Cluj-*
- 829 *Napoca*, 2015, **43**, 302-312.
- 830 21. N. A. Al-Tayyar, A. M. Youssef and R. Al-Hindi, *Food chemistry*, 2020, **310**, 125915.
- 831 22. M. Z. Elsabee and E. S. Abdou, *Materials science and engineering: C*, 2013, **33**, 1819-1841.
- 832 23. F. Jahan, R. Mathad and S. Farheen, *Materials Today: Proceedings*, 2016, **3**, 3689-3696.
- 833 24. J. Khouri, A. Penlidis and C. Moresoli, *Processes*, 2019, **7**, 157.
- 834 25. H. Haghighi, F. Licciardello, P. Fava, H. W. Siesler and A. Pulvirenti.
- 835 26. T. Yeamsuksawat and J. Liang, *Food packaging and shelf life*, 2019, **22**, 100415.
- 836 27. E. Aguila-Almanza, R. Salgado-Delgado, Z. Vargas-Galarza, E. García-Hernández and H.
- 837 Hernández-Cocoletzi, *Journal of Chemistry*, 2019, **2019**.
- 838 28. W. M. Argüelles-Monal, J. Lizardi-Mendoza, D. Fernández-Quiroz, M. T. Recillas-Mota and M.
- 839 Montiel-Herrera, *Polymers*, 2018, **10**, 342.
- 840 29. X.-Y. Wang, C.-S. Wang and M.-C. Heuzey, *International Journal of Polymeric Materials and*
- 841 *Polymeric Biomaterials*, 2016, **65**, 96-104.
- 842 30. J. Wang, L. Wang, H. Yu, Y. Chen, Q. Chen, W. Zhou, H. Zhang and X. Chen, *International*
- 843 *Journal of Biological Macromolecules*, 2016, **88**, 333-344.
- 844 31. K. Khwaldia, A. H. Basta, H. Aloui and H. El-Saied, *Carbohydrate polymers*, 2014, **99**, 508-516.
- 845 32. E. Abdelrazek, I. Elashmawi and S. Labeeb, *Physica B: Condensed Matter*, 2010, **405**, 2021-
- 846 2027.
- 847 33. M. Shahbazi, G. Rajabzadeh and S. J. Ahmadi, *Carbohydrate polymers*, 2017, **157**, 226-235.
- 848 34. A. Muxika, A. Etxabide, J. Uranga, P. Guerrero and K. De La Caba, *International journal of*
- 849 *biological macromolecules*, 2017, **105**, 1358-1368.
- 850 35. H. Wang, J. Qian and F. Ding, *Journal of agricultural and food chemistry*, 2018, **66**, 395-413.
- 851 36. E. A. Kamoun, E.-R. S. Kenawy and X. Chen, *Journal of advanced research*, 2017, **8**, 217-233.
- 852 37. S. Mohanapriya, *Polyvinyl Alcohol-Based Biocomposites and Bionanocomposites*, 2023, 59-
- 853 79.
- 854 38. J. Zanela, A. P. Bilck, M. Casagrande, M. V. E. Grossmann and F. Yamashita, *Polímeros*, 2018,
- 855 **28**, 256-265.
- 856 39. D. Gu, Y. Yang, M. Bakri, Q. Chen, X. Xin and H. A. Aisa, *Phytochemical Analysis*, 2013, **24**,
- 857 661-670.
- 858 40. B. Hamedi, A. G. Pirbalouti and F. Rajabzadeh, *Industrial Crops and Products*, 2022, **187**,
- 859 115470.
- 860 41. H.-J. Choi, S.-W. Choi, N. Lee and H.-J. Chang, *Foods*, 2022, **11**, 3963.
- 861 42. V. Jaiswal, B. Butola and A. Majumdar, *International Journal of Biological Macromolecules*,
- 862 2023, **252**, 126457.



- 863 43. Y. Wang, Y. Yu, C. Shi, Y. Ren, J. Han and R. Wu, *International Journal of Biological Macromolecules*, 2025, **308**, 142299.
- 864
- 865 44. J. F. Rubilar, R. M. Cruz, H. D. Silva, A. A. Vicente, I. Khmelinskii and M. C. Vieira, *Journal of Food Engineering*, 2013, **115**, 466-474.
- 866
- 867 45. V. G. Bhat, S. S. Narasagoudr, S. P. Masti, R. B. Chougale, A. B. Vantamuri and D. Kasai, *International Journal of Biological Macromolecules*, 2022, **200**, 50-60.
- 868
- 869 46. M. Mujahid, M. Zubair, A. Yaqoob, S. Shahzad and A. Ullah, *Bioengineering*, 2025, **12**, 439.
- 870 47. A. Hassan, M. B. K. Niazi, A. Hussain, S. Farrukh and T. Ahmad, *Journal of Polymers and the Environment*, 2018, **26**, 235-243.
- 871
- 872 48. S. Shahzad, S. Ahmed, A. Yaqoob, S. Saleem, S. Shaheen, K. Mammadova, A. K. Qureshi, M. Arshad, A. Saeed and D. H. M. Alkhalifah, *Chemical Papers*, 2025, 1-16.
- 873
- 874 49. S. Vasi, G. Ceccio, A. Cannavò, P. Pleskunov and J. Vacík, *Sustainability*, 2022, **14**, 5863.
- 875 50. D. Yun and J. Liu, *Foods*, 2024, **13**, 1174.
- 876 51. L. W. Winkler, *Berichte der deutschen chemischen Gesellschaft*, 1888, **21**, 2843-2854.
- 877 52. H. Bauer, F. Paronetto, W. A. Burns and A. Einheber, *Journal of Experimental Medicine*, 1966, **123**, 1013-1024.
- 878
- 879 53. M. Abbas, M. Arshad, M. Rafique, A. Altalhi, D. Saleh, M. Ayub, S. Sharif, M. Riaz, S. Alshawwa and N. Masood, *Arabian Journal of Chemistry*, 2022, **15**, 103766.
- 880
- 881 54. T. Wang, X. Zhai, X. Huang, Z. Li, X. Zhang, X. Zou and J. Shi, *Food Packaging and Shelf Life*, 2023, **39**, 101133.
- 882
- 883 55. M. Nowacka, K. Rybak, A. Wiktor, A. Mika, P. Boruszewski, J. Woch, K. Przybysz and D. Witrowa-Rajchert, *Food Control*, 2018, **93**, 183-190.
- 884
- 885 56. F. Nugroho, N. Nizardo and E. Saepudin, 2020.
- 886 57. D. Yun, Y. Qin, J. Zhang, M. Zhang, C. Qian and J. Liu, *International Journal of Biological Macromolecules*, 2021, **189**, 900-909.
- 887
- 888 58. M. Zubair, S. Hussain, A. Hussain, M. E. Akram, S. Shahzad, Z. Rauf, M. Mujahid and A. Ullah, *Biomaterials Science*, 2025, **13**, 130-160.
- 889
- 890 59. N. K. Howell, G. Arteaga, S. Nakai and E. C. Li-Chan, *Journal of agricultural and food chemistry*, 1999, **47**, 924-933.
- 891
- 892 60. M. Cantor, E. Buta, I. Conțiu, R. Ștefan, I. Crișan and T. Buru, *Emirates Journal of Food and Agriculture*, 2021, **33**, 899-908.
- 893
- 894 61. R. Różyło, R. Amarowicz, M. A. Janiak, M. Domin, I. Różyło, K. Rząd, A. Matwijczuk, R. Rusinek and M. Gancarz, *Molecules*, 2024, **29**, 4931.
- 895
- 896 62. D. Hu, X. Liu, Y. Qin, J. Yan, J. Li and Q. Yang, *Food Quality and Safety*, 2022, **6**, fyac028.
- 897 63. G. Qiao, Z. Xiao, W. Ding and A. Rok, *Coatings*, 2019, **9**, 828.
- 898 64. D. Lin and Y. Zhao, *Comprehensive reviews in food science and food safety*, 2007, **6**, 60-75.
- 899 65. Y. Peng, Y. Wu and Y. Li, *International journal of biological macromolecules*, 2013, **59**, 282-289.
- 900
- 901 66. U. Siripatrawan and B. R. Harte, *Food hydrocolloids*, 2010, **24**, 770-775.
- 902 67. X. Wang, H. Yong, L. Gao, L. Li, M. Jin and J. Liu, *Food hydrocolloids*, 2019, **89**, 56-66.
- 903 68. F. Bigi, H. Haghghi, H. W. Siesler, F. Licciardello and A. Pulvirenti, *Food Hydrocolloids*, 2021, **120**, 106979.
- 904
- 905 69. H.-S. Han and K. B. Song, *Food Hydrocolloids*, 2021, **112**, 106372.
- 906 70. T. A. Dinh, Y. N. Le, N. Q. Pham, P. Ton-That, T. Van-Xuan, T. G.-T. Ho, T. Nguyen and H. H. K. Phuong, *Progress in Organic Coatings*, 2023, **183**, 107772.
- 907
- 908 71. A. Ali and S. J. H. Ahmed, 2021, **7**, e06550.
- 909 72. L. Wang, Y. Dong, H. Men, J. Tong and J. Zhou, *Food hydrocolloids*, 2013, **32**, 35-41.
- 910 73. A. Ali and S. Ahmed, *Heliyon*, 2021, **7**.
- 911 74. S. B. Ghelejlou, M. Esmaili and H. Almasi, *International journal of biological macromolecules*, 2016, **86**, 613-621.
- 912



- 913 75. J. Hafsa, M. ali Smach, M. R. B. Khedher, B. Charfeddine, K. Limem, H. Majdoub and S. Rouatbi, *LWT-food Science and Technology*, 2016, **68**, 356-364. View Article Online  
DOI: 10.1039/D5FB00934K
- 914
- 915 76. S. Yadav, G. Mehrotra, P. Bhartiya, A. Singh and P. Dutta, *Carbohydrate polymers*, 2020, **227**,
- 916 115348.
- 917 77. C. Pagella, G. Spigno and D. M. De Faveri, *Food and Bioproducts Processing*, 2002, **80**, 193-
- 918 198.
- 919 78. B. Singh, J. Singh, V. Sharma, P. Sharma and R. Kumar, *Hybrid Advances*, 2023, **4**, 100096.
- 920 79. N. Nowak, W. Grzebieniarsz, A. Cholewa-Wójcik, L. Juszcak, A. Konieczna-Molenda, E. Dryzek, M. Sarnek, M. Szuwarzyński, T. Mazur and E. Jamróz, *Food and Bioprocess Technology*, 2024, **17**, 1201-1214.
- 921
- 922
- 923 80. L. K. Winkelströter, E. Bezirtzoglou and F. L. Tulini, *Journal*, 2022, **13**, 856858.
- 924 81. D. Altiok, E. Altiok and F. Tihminlioglu, *Journal of Materials Science: Materials in Medicine*,
- 925 2010, **21**, 2227-2236.
- 926 82. Y. Xing, Q. Xu, X. Li, C. Chen, L. Ma, S. Li, Z. Che and H. Lin, *International Journal of Polymer Science*, 2016, **2016**, 4851730.
- 927
- 928 83. D. Xu, T. Chen and Y. Liu, *Polymer Bulletin*, 2021, **78**, 3607-3624.
- 929 84. M. Moghadam, M. Salami, M. Mohammadian, M. Khodadadi and Z. Emam-Djomeh, *Food Hydrocolloids*, 2020, **104**, 105735.
- 930
- 931 85. A. Riaz, C. Lagnika, H. Luo, Z. Dai, M. Nie, M. M. Hashim, C. Liu, J. Song and D. Li, *International Journal of Biological Macromolecules*, 2020, **150**, 595-604.
- 932
- 933 86. T. Gasti, S. Dixit, S. P. Sataraddi, V. D. Hiremani, S. P. Masti, R. B. Chougale and R. B. Malabadi, *Journal of Polymers and the Environment*, 2020, **28**, 2918-2930.
- 934
- 935 87. J. B. Harborne, *Phytochemistry*, 1983, **22**, 1683-1684.
- 936 88. S. Kayahan, F. Gülbağ, Y. Kaya and H. Altunkanat, *Horticultural Studies*, 2024, **41**, 74-81.
- 937 89. Z. T. Murathan, M. Zarifikhosroshahi, E. Kafkas and E. Sevindik, *Italian Journal of Food Science/Rivista Italiana di Scienza degli Alimenti*, 2016, **28**.
- 938
- 939 90. G. Fascella, F. D'Angiolillo, M. M. Mammano, M. Amenta, F. V. Romeo, P. Rapisarda and G. Ballistreri, *Food Chemistry*, 2019, **289**, 56-64.
- 940
- 941 91. S. Shameh, B. Hosseini, A. Alirezalu and R. Maleki, *Journal of AOAC International*, 2018, **101**,
- 942 1788-1793.
- 943 92. M. Tabaszewska and D. Najgebauer-Lejko, *NFS Journal*, 2020, **21**, 50-56.
- 944 93. A. Y. Aldhebiani and W. A. Yaslam, *Pak. J. Bot*, 2023, **55**, 995-999.
- 945 94. N. Mukhtar, N. A. Marzuki, N. M. Zain, L. Naher, N. M. Hairin and N. M. Arsab, 2024.

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## Data Availability Statement

The data that support the findings of this study are available from the corresponding authors upon reasonable request.

