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Fruit exosomes: a sustainable green cancer therapeutic

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Extracellular vesicles, particularly their subpopulation of exosomes, have emerged as a promising avenue for cell-free anti-cancer therapies in the current decade of exosomal research, opening a new chapter in cancer precision medicine. This paradigm shift towards plant-based exosomes holds significant implications for both therapeutic efficacy and sustainability. Plant-derived exosomes offer a non-toxic source for anti-cancer agents, addressing concerns about both patient well-being and the environmental impact of treatment production. This sustainable approach has the potential to make cancer precision therapy more affordable, scalable, and accessible, while simultaneously inspiring scientific minds to explore the vast potential of this "Green Therapy". This article delves into the potential of fruit-based exosomes in cancer precision therapy, exploring their advantages, challenges, and future perspectives. We discuss the current understanding of fruit exosome biogenesis, leading and isolation techniques, cargo loading mechanisms, their interactions with recipient cellular targets and challenges associated with fruit-based exosomal research. Despite the challenges, the future of fruit-derived exosome-based cancer therapy appears promising. Continued research and interdisciplinary collaborations are crucial to fully unlock the therapeutic potential of fruit-derived exosome-based impactful natural cancer treatment.

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Sustainability spotlight

Exosomes derived from fruits present a promising avenue for sustainable and environment-friendly cancer treatment and drug delivery. These naturally sourced nanovesicles offer inherent biocompatibility, biodegradability, and low immunogenicity, positioning them as ideal carriers for therapeutic cargo. Notably, fruit-derived exosomes demonstrate significant anti-cancer properties with minimal toxicity compared to their mammalian and synthetic counterparts. Their ability to encapsulate and deliver a variety of anti-cancer agents, including anti-carcinogenic drugs, proteins, and nucleic acids, while selectively targeting cancer cells, underscores their therapeutic potential. This green nanomedicine approach, utilizing fruit-derived exosomes, minimizes the ecological footprint associated with conventional drug delivery systems, paving the way for effective, affordable, and globally accessible cancer treatments.

1. Introduction

Cancer remains a significant global health burden, necessitating the development of innovative therapeutic approaches to combat its multifaceted complexities.^{1,2} Conventional cancer treatments, such as chemotherapy, radiation therapy, and surgical interventions, have inherent limitations, including systemic toxicity, drug resistance, and a lack of tumor-targeted specificity.³⁻⁷ The rapidly evolving field of nanomedicine, particularly the use of extracellular vesicles,

including exosomes, for precise drug delivery, offers a new era for personalized treatment of chronic diseases, especially cancer.^{4,8-10} Mammalian-derived exosomes, while promising, face clinical translation hurdles due to potential immune rejection, toxicity risks, and production challenges. Fruit-derived exosomes offer a safer alternative, leveraging structural and functional similarities while mitigating significant risks.¹¹⁻¹³ Plant and fruit extracts have long been investigated for their potential anticancer properties, with numerous studies highlighting the ability of their bioactive compounds to inhibit tumor growth and progression.¹⁴⁻¹⁶ However, harnessing these beneficial effects for clinical applications has proven challenging.¹⁵ Fruit-derived exosomes, derived from abundant, edible sources, ensure scalability and biocompatibility. These exosomes represent a novel strategy for utilizing the anticancer properties of these phytochemicals. These nanovesicles offer a naturally engineered platform for encapsulating and delivering concentrated doses of these bioactive

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compounds (phytochemicals) to targeted tumor sites, enhancing their bioavailability and therapeutic efficacy.^{11,17,18} Fruit-derived exosomes simultaneously demonstrate a remarkable ability to encapsulate and deliver a variety of therapeutic cargoes, including chemotherapeutic drugs, small interfering RNAs, and microRNAs, directly to tumor sites. This targeted delivery of therapeutics using fruit-derived exosomes enhances drug efficacy by selectively transporting the cargo to tumor cells, while minimizing off-target effects on healthy tissues and associated adverse reactions. The inherent biocompatibility and tumor-homing capabilities of these



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naturally-derived nanoparticles allow for safer and more precise drug delivery, addressing a key limitation of conventional cancer treatments. Furthermore, research indicates that fruit-derived exosomes can modulate the tumor microenvironment, influencing the complex interplay between cancer cells and their surroundings to inhibit tumor growth and progression. Fruit-derived exosomes can also be engineered to enhance their targeting capabilities, drug-loading efficiency, and anti-tumor properties.^{19,20} They show immense potential for improving the clinical translatability of plant-based cancer therapies and improving cancer treatment outcomes.^{18,21}



Ketki Kalele

Dr Ketki Kalele is a distinguished oral histopathologist, has dedicated her career to advancing the fields of oral pathology, medical oncology, and cancer research. A dedicated Member of the Indian Extracellular Vesicle Society (IEVS) and an Executive Committee Member of the Malaysian Extracellular Vesicle Society (MEVS), a visionary founder of NIAR, and director of Warkas Research and Innovations Pvt. Ltd, she has also been instrumental in fostering a collaborative environment among healthcare professionals. Her commitment to social welfare and her numerous accolades, including the Women Leadership Award in 2022 and Best Research Paper by the Asia Pacific Dental Association in 2021, underscore her exceptional achievements in the field of medicine.



Vetriselvan Subramaniyan

Professor Vetriselvan Subramaniyan is a distinguished academic with over 15 years of experience in higher education. A graduate of Annamalai University, his career spans prestigious institutions globally, including Arba Minch University, MAHSA University, and currently, Sunway University where he holds a professor position. His impactful contributions to the field are evidenced by eight international patents, over 180 publications with a cumulative impact factor exceeding 500, and numerous grants from organizations like the Ministry of Higher Education, Malaysia. Professor Subramaniyan's leadership extends to his role as the Founding President of the Malaysian Extracellular Vesicle Society (<http://www.mevsociety.com>), a testament to his commitment to advancing this crucial area of research. He also holds memberships in several renowned scientific societies and serves on various editorial boards.



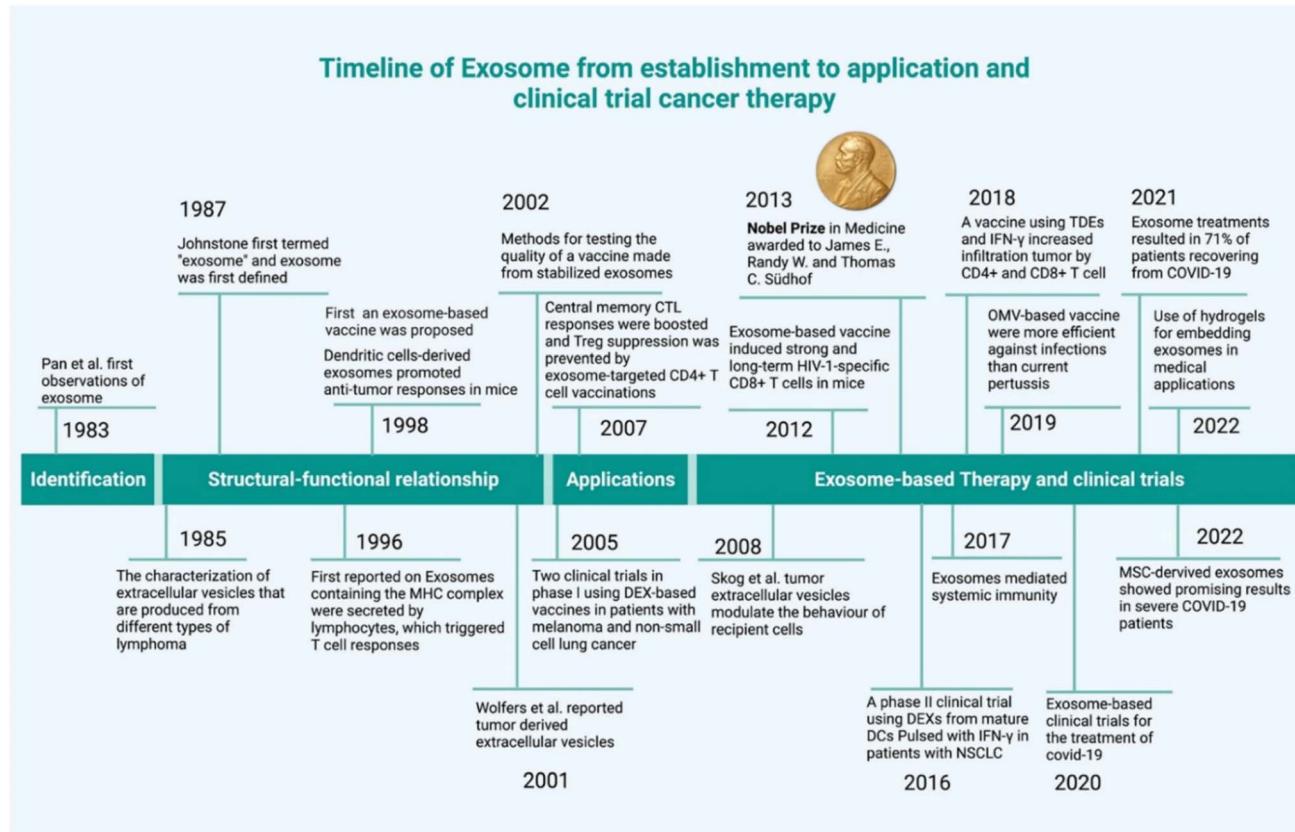


Fig. 1 Exosome-based cancer therapeutic developmental landscape (reproduced with permission under Creative Commons CC BY 4.0 license from ref. 22. Copyright © 2022 The Authors).

Despite the promising results, further research is needed to fully understand the mechanisms of action, optimize the production and purification methods, and ensure the safety and efficacy of fruit-derived exosomes in clinical settings. Ongoing clinical trials and regulatory developments will also play a crucial role in the translation of these novel therapeutics from the bench to the bedside (Fig. 1).

This review explores the burgeoning field of fruit-derived exosomes as a novel, green, and sustainable approach to cancer therapy. Fruit exosomes, naturally occurring nanovesicles, offer a unique convergence of plant-derived exosome advantages and potential edible benefits as functional foods. This dual functionality positions them as a revolutionary tool for cancer management, providing both accessible targeted therapeutic intervention and nutritional support. Their biocompatibility, inherent targeting ability, and capacity to encapsulate diverse anticancer payloads highlight their potential to enhance treatment efficacy while minimizing off-targets and systemic toxicity. Their unique ability to modulate the tumor microenvironment further positions them as a promising alternative to conventional therapies. Although the potential for edible incorporation into diets exists, further research is essential to standardize extraction and characterization methods, ensuring consistent therapeutic efficacy and safety.

2. Fruit exosome biogenesis

Plants employ various pathways for exosome biogenesis (Fig. 2), including those resembling the exosomal pathway in mammals^{23–25} (involving endosomal compartments), the Exocyst-Positive Organelle (EXPO) pathway, and a pathway involving the plant vacuole. Each pathway, with its unique mechanisms, contributes to the secretion of bioactive molecules essential for intercellular communication and various physiological processes.^{23,24,26} The EXPO pathway offers a distinct route for secreting exosomes, particularly in plants.²⁷ Originating from the ER and Golgi, specialized compartments bud off, aided by the exocyst complex and small GTPases (Rab and Ral proteins). These EXPO compartments, enriched with various biomolecules (proteins, lipids, and nucleic acids), travel along the actin cytoskeleton to the plasma membrane. Fusion with the membrane, facilitated by SNARE proteins, releases the nanoparticles into the extracellular space. This unique pathway is crucial for secreting cell wall components and bioactive molecules within the exosomes, and enables intercellular communication, influencing various physiological processes in plants.^{26–28} The multivesicular body (MVB) pathway, a conserved mechanism across eukaryotes, generates exosome-like nanoparticles through a series of steps within endosomes.^{29–31} Initially, intraluminal vesicles bud inward from the endosomal



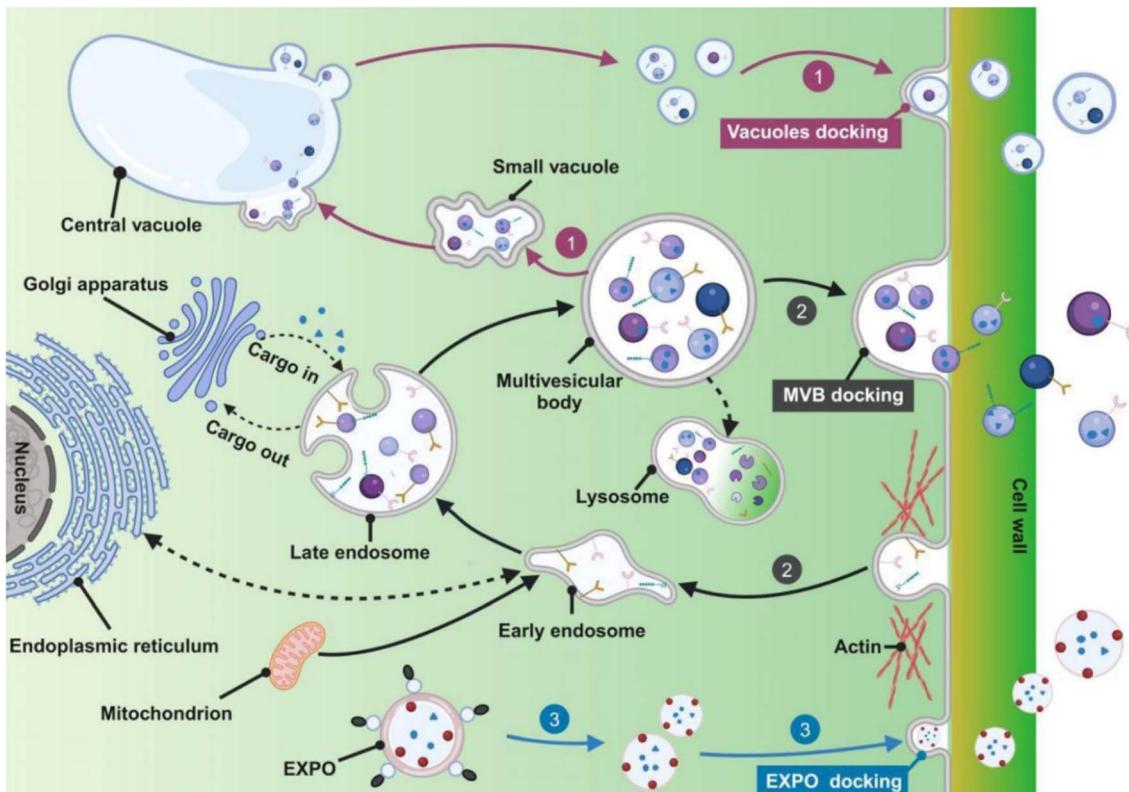


Fig. 2 Fruit exosome biogenesis (1) vacuolar pathway; (2) MVB pathway; (3) EXPO pathway (reproduced with permission under Creative Commons CC BY 4.0 license from ref. 36. Copyright © 2024 The Authors).

membrane, a process orchestrated by the endosomal sorting complex required for transport. This complex recognizes and incorporates ubiquitinated cargo proteins into the forming ILVs. Lipids, particularly phosphatidylinositol 3-phosphate (PI3P), play a crucial role by recruiting endosomal sorting complex required for transport (ESCRT) components to the endosomal membrane.^{32–34} Intraluminal vesicle (ILV)-containing endosomes mature into multivesicular bodies, which can fuse with the plasma membrane, a process regulated by Rab GTPases, to release ILVs as exosomes. These nanovesicles, rich in bioactive molecules, play crucial roles in intercellular communication and plant defence responses.^{29,30,35}

Additionally, the vacuolar pathway, another route for fruit exosome biogenesis, involves the budding of vesicles from the vacuolar membrane, a process potentially involving ESCRT machinery and lipids (PI3P and PI(3,5)P₂).^{26,28,29,37} While less understood than other pathways, it plays a crucial role in protein storage, nutrient recycling, and plant defence (using antimicrobial proteins). The fusion of vacuoles with the plasma membrane is regulated by small GTPases (like Rab and Ral proteins) and tethering/fusion factors (including the exocyst complex and SNARE proteins), facilitating exosome release into the extracellular space.^{28,30,35} While these pathways share some similarities with the biogenesis of mammalian-derived exosomes, they also exhibit unique characteristics due to their plant origin. However, further research is needed to fully elucidate its regulatory mechanisms and biotechnological

potential. Differentiation between fruit-derived exosomes and mammalian cell-derived exosomes is presented in Table 1.

3. Isolation and characterization fruit exosomes

The isolation and characterization of fruit exosomes necessitate rigorous and meticulous methodologies to fully elucidate and harness their therapeutic capabilities. Involving a delicate balance of separating these nanoscale vesicles from the fruit matrix without compromising their structural integrity or biological activity. Various methods exist for isolating these exosomes, all of which rely on leveraging their distinct physical characteristics.^{45–47} Differential ultracentrifugation remains a cornerstone technique for exosome isolation. This method exploits the principle of sedimentation, subjecting samples to sequential centrifugation steps at escalating gravitational forces (g). By targeting specific sedimentation coefficients, exosomes are effectively pelleted and separated from a heterogeneous mixture of larger vesicles and cellular debris.^{47,48} Density gradient centrifugation improves purity by separating exosomes based on their buoyant density ($1.13\text{--}1.18\text{ g mL}^{-1}$) within a sucrose or iodixanol gradient.⁴⁹ Tangential flow filtration (TFF) is a scalable technique designed for processing large volumes, utilizing membranes with defined pore sizes (typically 50–100 nm) to selectively retain exosomes while allowing smaller molecules to pass through.^{50,51} Ultrafiltration operates

Table 1 Comparison of fruit-derived exosomes and mammalian cell-derived exosomes

Characteristics	Fruit-derived exosomes	Mammalian cell-derived exosomes	Reference
Origin	Fruits, plant origin	Mammalian cells (e.g., mesenchymal stem cells, dendritic cells, etc.)	25 and 36
Size (nm)	30–800 nm	30–150 nm	36 and 38–40
Zeta potential (mV)	–70 to approximately neutral	–34.3 to –6.3	18, 40 and 41
Cargo	- Proteins - Nucleic acids - Secondary metabolites (unique to plants)	- Proteins (highly abundant) - Nucleic acids - Lipids	25, 36 and 42
Protein content	Typically lower protein concentration; proteins are mainly cytosolic with few membrane proteins present	Higher protein diversity, including membrane proteins and signaling molecules	18, 25, 40, 43 and 44
miRNA content	Contains various miRNAs, but the diversity and functional roles may be limited compared to mammalian exosomes	Rich in diverse miRNAs that play significant roles in intercellular communication and regulation	36, 40, 42 and 43
Lipid composition	Lipid profiles are simpler, primarily consisting of phospholipids and some bioactive lipids; may lack certain lipid species found in mammalian exosomes	Complex lipid composition, including sphingolipids and cholesterol, which are crucial for membrane stability and function	25, 36, 43 and 44
Immunogenicity	Lower risk of immunogenic response due to lack of mammalian proteins	Potential for immunogenic response due to the presence of foreign proteins	25, 36, 42 and 44
Scalability	Potentially easier and more cost-effective to isolate from abundant plant sources	Can be challenging to isolate large quantities from mammalian cells	25, 36, 43 and 44
Means of internalization	Clathrin/caveolae-mediated endocytosis, macropinocytosis, and phagocytosis	Phagocytosis and endocytosis	40, 42 and 44
Common route of administration	Oral administration	Intravenous administration	25, 36, 40 and 44

on a similar principle but employs centrifugal force to drive samples through the membrane, making it particularly suitable for smaller volumes.^{46,52} Precipitation-based methods, frequently available as commercial kits, take advantage of the altered solubility of exosomes in the presence of water-excluding polymers like polyethylene glycol (PEG), resulting in their precipitation.^{46,53} Immunoaffinity capture provides high specificity by utilizing antibodies immobilized on beads or surfaces to selectively bind exosomes that express specific surface markers.^{54,55} Microfluidic isolation employs miniaturized devices that integrate various separation mechanisms such as filtration, affinity capture, and acoustic trapping for high-throughput and automated isolation.^{52,56,57} Acoustic fluid handling manipulates exosomes based on their acoustic properties, using sound waves to focus and separate them.⁵⁸ Magnetophoresis involves magnetic beads coated with antibodies against exosomal markers, facilitating efficient capture and release of exosomes through magnetic fields.⁵⁹ Deterministic lateral displacement (DLD) exploits the size and deformability of exosomes, separating them based on their trajectory in microfluidic channels with asymmetrically arranged obstacles.^{60,61} Lastly, field-flow fractionation separates particles in a thin channel under external fields—such as flow or electric fields—based on their differential migration.^{62–65} Characterizing exosomes involves a variety of techniques aimed at elucidating

their biophysical and biochemical properties.⁶⁶ One widely used method is nanoparticle tracking analysis (NTA), which visualizes and monitors the Brownian motion of individual exosomes, utilizing light scattering to determine their size distribution and concentration.^{67,68} Dynamic light scattering (DLS) also employs light scattering but focuses on measuring the fluctuations in scattered intensity caused by Brownian motion, providing insights into hydrodynamic size and polydispersity.^{65,69} Electron microscopy, including both transmission and scanning electron microscopy, offers high-resolution images that reveal the morphology, size, and structural characteristics of exosomes.⁶⁶ Atomic force microscopy (AFM) provides nanoscale topographical data by scanning a sharp tip across the exosome surface, allowing for detailed examination of size, shape, and surface features.^{66,70} Western blotting remains a fundamental technique for detecting specific proteins within exosome lysates, thereby confirming the presence of known exosomal markers and target proteins.^{48,71} Flow cytometry can analyze individual exosomes for size, granularity, and expression of surface markers using fluorescently labeled antibodies; however, this method faces challenges due to the small size of exosomes.^{53,72} Enzyme-linked immunosorbent assays (ELISA) quantify specific proteins or other molecules in exosome samples through antibody-based detection, offering high sensitivity and specificity.^{47,66} Raman spectroscopy analyzes the



inelastic scattering of light from molecules within exosomes to provide information about their biochemical composition and structure.⁷³ Mass spectrometry serves to identify and quantify proteins, lipids, and metabolites in exosome samples, yielding a comprehensive molecular profile.^{62,74,75} RNA sequencing examines the RNA content within exosomes, revealing mRNA, miRNA, and other RNA species that provide insights into their functional roles and origins.⁶⁶ Lipidomics focuses on characterizing the lipid composition of exosomes to enhance understanding of their membrane structure and function.^{74,76} Lastly, the Integrated Magnetic-Electrochemical Exosome (iMEX) sensor is a rapid and sensitive approach for quantifying exosome surface markers, demonstrating its effectiveness in detecting varying numbers of extracellular vesicles spiked into human plasma.⁶⁶ In the exosome isolation and profiling process, the combination of nanotechnology and nanomaterials promotes it as a next-generation promising precision medicine development platform.⁷⁷⁻⁸⁰

4. Exosomes and cancer

The tumor microenvironment (TME) consists of a complex network of cellular and non-cellular components that facilitate cancer progression and metastasis.⁸¹⁻⁸³ A defining feature of the TME is the hypoxic condition, characterized by inadequate blood supply and oxygen availability, which enables cancer cells to activate survival pathways mediated by hypoxia-inducible factors (HIFs).^{8,84-87} This adaptability allows malignant cells to proliferate uncontrollably and divert essential resources for growth.⁸⁷⁻⁸⁹ Consequently, cancer cells disrupt the homeostasis

of surrounding normal tissues, creating a microenvironment conducive to malignant transformation and metastasis.^{84,90} Hypoxia also stimulates the secretion of exosomes from tumor cells, which are nanoscale vesicles that play crucial roles in intercellular communication within the TME.^{66,91} Tumor-derived exosomes (TEXs) facilitate angiogenesis by transferring pro-angiogenic factors and microRNAs to endothelial cells, enhancing their proliferation and migration.^{81,92,93} For example, exosomes from hypoxic tumor cells contain high levels of miR-210, promoting angiogenesis by targeting endothelial cell function genes.^{94,95} TEXs significantly influence immune cell behavior, contributing to immune evasion by shifting the immune response toward an immunosuppressive state.⁹⁶⁻⁹⁸ They can reprogram macrophages from an anti-tumorigenic M1 phenotype to a pro-tumorigenic M2 phenotype,^{99,100} with exosomal miR-934 facilitating this shift and enhancing metastasis.¹⁰⁰ Additionally, TEXs promote the differentiation of monocytes into myeloid-derived suppressor cells (MDSCs), further inhibiting T-cell responses through signaling molecules like prostaglandin E2 (PGE2) and transforming growth factor beta (TGF- β).¹⁰¹ Moreover, TEXs suppress T cell activity *via* cargo such as TGF- β and programmed death-ligand 1 (PD-L1), which inhibit T cell activation and promote regulatory T cell differentiation.¹⁰² Dendritic cells can also be converted into tolerogenic cells by TEXs, diminishing their ability to activate T cells effectively.¹⁰² This interplay underscores how tumors evade immune surveillance and promote growth. Exosomes are also implicated in metastasis through processes like the epithelial-mesenchymal transition (EMT), where epithelial cells acquire migratory properties. TEXs transfer bioactive molecules that

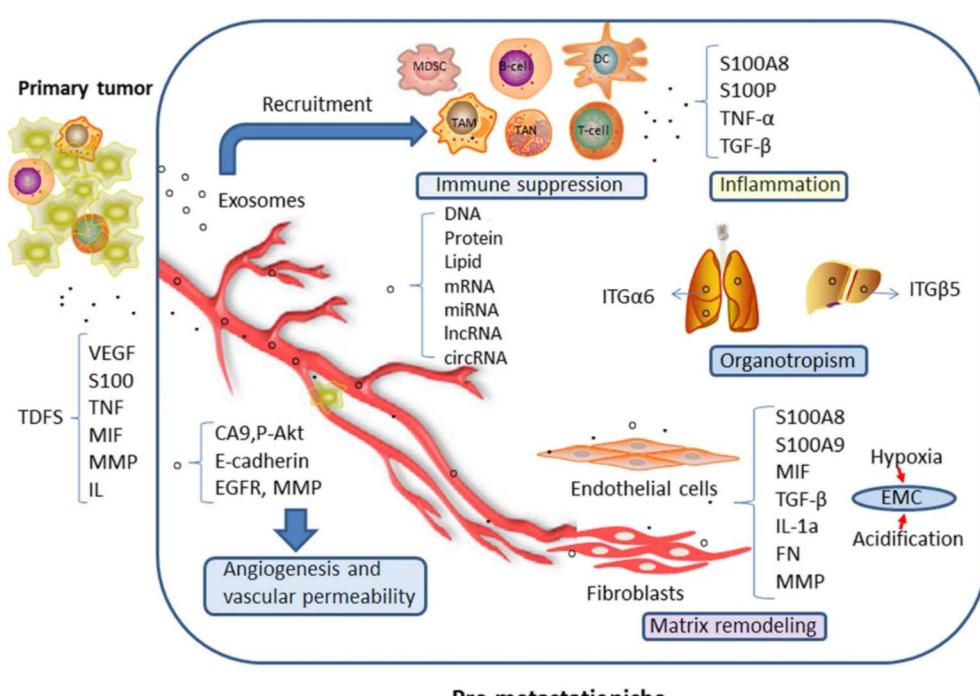


Fig. 3 Role of exosomes in cancer progression. (Reproduced with permission under Creative Commons CC BY 4.0 license from ref. 109 copyright © 2019 Springer publisher.)



facilitate the EMT and enhance metastatic potential.¹⁰³ For instance, exosomal microRNA-106b-5p has been linked to colorectal cancer metastasis by influencing interactions with M2 macrophages.¹⁰⁴ Furthermore, exosomal surface proteins such as tetraspanins and integrins play critical roles in organ-specific metastasis by mediating interactions between circulating tumor cells and target microenvironments.^{105,106} Exosomes also contribute to therapeutic resistance by transferring drug resistance genes and proteins. For example, exosomes from cisplatin-resistant lung cancer can transfer miRNA-100-5p, altering mTOR signaling to enhance survival during chemotherapy.¹⁰⁷ In breast cancer, exosomes can sequester human epidermal growth factor receptor 2 (HER2) targeted drugs or reprogram gene expression in recipient cells to reduce treatment susceptibility.¹⁰⁸ In summary, exosomes act as both facilitators of cancer progression and potential targets for

innovative therapeutic strategies. Their multifaceted roles in the TME highlight their significance in understanding cancer biology and developing new treatments. In the TME, TEX-mediated cancer promoting signature is described in Fig. 3.

5. Therapeutics application of fruit-derived exosomes

Exosome-based cancer therapeutic application presents a more advanced approach compared to traditional cell-based therapies.¹¹⁰ Fruit-derived exosomes are gaining traction as potential candidates for precision cancer therapies (Fig. 4).¹¹¹⁻¹¹³ This section will explore various anti-cancer therapeutic approaches utilizing these exosomes, examining their sources, mechanisms of action, therapeutic payloads, targeted therapies, efficacy in

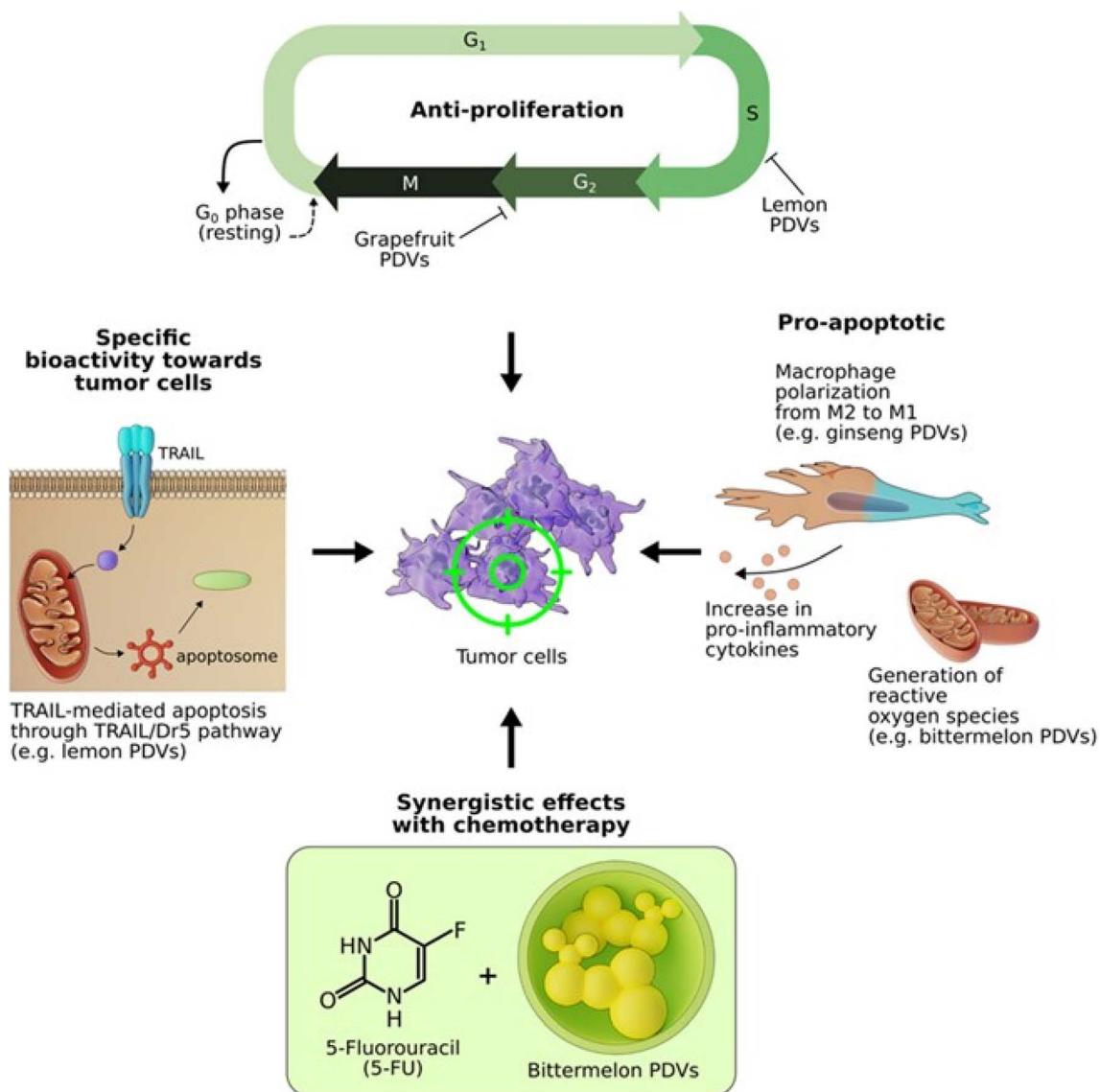


Fig. 4 Anti-cancer activity of plant-derived EVs (reproduced with permission under Creative Commons CC BY 4.0 license from ref. 18 copyright © 2022 The Authors). TRAIL-tumor necrosis factor-related apoptosis-inducing ligand, PDVs-plant derived extracellular vesicles, DR5-death receptor 5 is activated when it binds to TRAIL, which initiates apoptosis.





specific cancer types, and relevant research. Lemon-derived exosomes exhibit anti-cancer properties primarily through the induction of apoptosis in cancer cells, activating the TRAIL pathway to promote cell death in various cancer types.^{114,115} These exosomes contain small RNAs, including microRNAs, and show promise as natural anti-cancer agents against triple-negative breast cancer by inhibiting cell proliferation, migration, and phosphatidylinositol 3-kinase/protein kinase B (PI3K/AKT) and mitogen-activated protein kinase/extracellular signal-regulated kinase (MAPK/ERK) signaling pathways.¹¹⁶ They also target various cancer cell lines, including SW480 (human colorectal adenocarcinoma cell line), LAMA84 (human chronic myeloid leukemia cell line) and A549 (human lung carcinoma cell line) by downregulating anti-apoptotic proteins and upregulating pro-apoptotic factors. Studies have shown that lemon-derived exosomes inhibit proliferation in colorectal cancer cells *in vitro* and suppress tumour growth in xenograft models of chronic myeloid leukemia.^{40,114-116} Strawberry-derived exosome-like nanoparticles exhibit anti-cancer properties by preventing oxidative stress-induced damage in human mesenchymal stromal cells, modulating cellular differentiation, and promoting apoptosis in tumor cells through the delivery of bioactive compounds such as vitamin C, anthocyanins, and small RNAs.^{40,117} These exosomes may primarily target breast cancer cells, where they exert protective effects against oxidative stress and induce apoptosis, as indicated by research showing that they significantly reduce oxidative stress levels in cancer cell lines.¹¹⁷⁻¹¹⁹ Grapefruit-derived exosomes exhibit anti-cancer properties by inducing cell cycle arrest and apoptosis in cancer cells through the modulation of signaling pathways such as PI3K/AKT. These nanovesicles exhibit anti-tumor activity due to their rich cargo, which includes polyphenols, flavonoids, various miRNAs, and an enrichment of specific metabolites like alpha hydroxy acids (glycolic and citric acids), the amino acids (leucine and isoleucine), and the fatty acids (palmitic acid and doconexent).^{40,120} They primarily target breast, lung and melanoma cells and impair the proliferation of cancer cells, inhibiting metastasis and promoting apoptosis, as demonstrated by

significant anti-cancer effects *in vitro* and reduced tumor sizes in animal models.^{114,120} Treatment with apple-derived exosomes not only effectively suppressed tumor growth by increasing the levels of miR-30a-5p, which in turn regulates cell cycle progression, but also proved to be completely safe. This anti-tumor effect was linked to the regulation of specific proteins like E2F transcription factor 7 (E2F7) in gallbladder/esophageal cancer and Glucose-Regulated Protein 78 (GRP78) in renal carcinoma. Additionally, these exosomes demonstrated anti-inflammatory properties by boosting the production of miRNA-146a in M2 macrophages.¹²¹⁻¹²³ Bitter melon-derived exosomes (BMEV) demonstrate both anti-tumor and anti-inflammatory properties against oral squamous cell carcinoma (OSCC). These exosomes trigger cell death through a mechanism involving ROS production and JUN protein upregulation, while surprisingly downregulating the inflammatory nucleotide-binding domain and leucine-rich repeat protein 3 (NLRP3) pathway. This NLRP3 downregulation, potentially mediated by BMEV associated RNAs, enhances the sensitivity of OSCC cells to the chemotherapy drug 5-fluorouracil (5-FU), suggesting a promising synergistic therapeutic strategy.^{124,125} Lemon exosome-based anti-cancer therapeutic activity is very effective.¹²⁶

Grape-derived exosomes show great promise as oral drug delivery vehicles due to their biocompatibility, biodegradability, and remarkable stability in challenging environments such as the gastrointestinal tract. These nanovesicles, with an average size of 205.1 nm, significantly boosted leucine-rich repeat-containing G-protein-coupled receptor 5 (Lgr5) gene expression in rats, suggesting a potential role in promoting intestinal stem cell activity and regeneration. Since Lgr5 is a marker of intestinal stem cells, implicated in colon cancer development, grape-derived exosomes may offer a novel therapeutic strategy for targeting this disease.^{125,127} Though still in their infancy, fruit-derived exosomes offer a promising new avenue for cancer precision therapies, potentially providing a biologically safer and more effective treatment approach to combat this devastating disease and improve patient outcomes Table 2.

Table 2 Fruit-derived exosomes-applications in drug delivery

Fruit exosome source	Therapeutic carried	Target	Reference
Grape	Fisetin	MOLT-4 cell	128
Grapefruit	Doxorubicin, heparin nanoparticles	Glioma	129
	HSP70, variants of BSA	Colon cancer cells	130
	Doxorubicin-(si)RNA co-delivery	MDR LoVo colon cancer cells	131
	HSP70	Glioma cells	44
	miRNA17	Brain tumor	115
	Doxorubicin	Breast cancer	132
Apple, orange, pomegranate	ath-miR159a	Caco-2 cells	133
	ath-miR162a-3p		
	ath-miR166b-3p		
	ath-miR396b-5p		
Watermelon	Therapeutic miRNA mimic (hsa-miR146a-5p)	Ovarian cancer cells (ID8, A2780, and OVCAR8 models)	134
Bitter melon	5-Fluorouracil	Oral squamous cell carcinoma	124
Lemon	Doxorubicin	Ovarian cancer	126

6. Toxicity of fruit-derived exosomes

Fruit-derived exosomes have exhibited a promising safety profile, supporting their potential for clinical translation. The inherent absence of zoonotic or human pathogens in fruit-derived exosomes contributes to their favourable biocompatibility, as evidenced by *in vitro* and *in vivo* studies across various administration routes.³⁶ Oral administration of fruit-derived exosomes, even at relatively high doses (e.g., grapefruit-derived exosomes at 10 mg kg⁻¹ in mice for 7 days), has not been associated with adverse effects on systemic inflammatory markers like serum interferon-gamma (IFN- γ) levels, liver function (liver enzymes), or hepatocellular injury (AST/ALT levels).^{36,38} Similarly, other routes of administration, including intraperitoneal^{36,135} and intranasal,^{36,136} have not raised significant safety concerns. Intravenous administration of fruit-derived exosomes, particularly those derived from grapefruit, and lemon,^{36,126} has generally been well-tolerated in preclinical models. However, given the inherent diversity in composition and biophysical properties of fruit-derived exosomes from different fruit sources, comprehensive safety evaluations remain indispensable before considering their intravenous use in clinical settings.

7. Clinical trials

Exosomes are gaining notable recognition and global research interest due to their promising therapeutic potential for various diseases including cancer, to address significant clinical challenges precisely and effectively. Numerous pre-clinical trials and laboratory investigations have explored the potential of exosomes, especially those derived from various human cells and plants (including various fruits, vegetables, and other plant parts), as a therapeutic tool for cancer treatment.^{137,138} Meanwhile, exosome-mediated drug delivery is emerging as a cutting-edge approach in clinical trials, holding tremendous potential for tackling cancer and other global health concerns.¹³⁹ Clinical trials are crucial for translational medicines in modern oncology practices. In recent times, those have been investigated in the clinical trial stage to examine their potential, efficacy, side effects, and limitations further.¹⁴⁰ In the current decade, scientists around the world are studying plant-derived exosomes, particularly those from fruits, as candidates for clinical trials due to their low toxicity, high biocompatibility, and the effectiveness of their molecular cargos especially phytochemicals.^{38,141} A recent clinical study (clinical trial ID: NCT01668849) investigated the anti-inflammatory properties of grape-derived exosomes, administered in grape powder form, to determine their effectiveness in reducing oral mucositis in head and neck cancer patients undergoing chemo-radiation therapy, including their impact on cytokine production, immune responses to tumor exosomal antigens, and associated metabolic and molecular markers (<https://clinicaltrials.gov/>). Moreover, exosomes show remarkable advancements in medicine and cancer research, with substantial improvements made in recent years. The clinical journey of exosomes is not without obstacles. Plant-derived exosomes, including those

from fruits, hold therapeutic promise, but their clinical application faces significant hurdles. Notably, the translation of exosome-based clinical trials must conform to rigorous good manufacturing practices (GMP), addressing the numerous challenges that arise.^{142,143} As research continues to uncover the benefits of exosome-based therapies, future research efforts should prioritize conducting comprehensive clinical trials to explore the therapeutic efficacy and drug delivery potency of exosomes, with a specific focus on plant and fruit-derived sources, which are currently underrepresented in clinical investigations.

8. Future perspectives and challenges

Fruit-derived exosomes show great promise in targeted cancer treatment because they are naturally compatible with the body, unlikely to trigger an immune response, and capable of delivering various therapeutic molecules like miRNAs, proteins, and bioactive molecules. Their therapeutic potential is evident in their anti-tumor effects, due to their unique biomolecular cargoes.^{12,115,144} Fruit-derived exosomes offer a renewable and abundant source for large-scale production, addressing a key limitation associated with mammalian exosomes.^{12,145,146} A major hurdle is the lack of standardized definitions, nomenclature, and practices for characterizing and classifying these plant-derived vesicles.^{36,147} Establishing a robust framework with rigorous physicochemical characterization and well-defined biological pathways is crucial for advancing exosome research. Moreover, fruit-derived exosomes with their inherent heterogeneity,¹⁴⁸ further compounded by the limitations of current isolation techniques, pose a significant hurdle.^{11,147} Optimizing extraction protocols to enhance fruit-derived exosome yield while maintaining purity and structural integrity is crucial for advancing preclinical and facilitating clinical translation. A deeper understanding of the molecular mechanisms underlying the anti-tumor effects of fruit-derived exosomes is also crucial.^{12,115} Identifying the specific miRNAs, proteins, and small molecules responsible for these effects, as well as their target pathways in cancer cells, will be essential for developing more targeted and effective therapies. Exploring potential synergistic interactions between different fruit-derived exosomal components may also uncover novel therapeutic strategies.^{115,149,150} To facilitate clinical translation, extensive *in vivo* studies are necessary to evaluate the safety, biodistribution, and therapeutic efficacy of fruit-derived exosomes in relevant animal models of cancer.^{12,145} Developing strategies to enhance their tumor-homing ability, such as surface modification with targeting ligands, may improve their specificity and therapeutic index. Additionally, investigating their potential as carriers for existing chemotherapeutic agents or in combination with other treatment modalities, such as immunotherapy, could lead to more effective cancer treatment approaches.^{36,151,152} In summary, while fruit-derived exosomes demonstrate significant potential for precision cancer therapy, addressing challenges related to standardization, yield optimization, mechanistic understanding, and clinical translation is pivotal to fully unlocking their therapeutic promise. Collaborative efforts



among researchers, clinicians, and industry partners will be crucial to drive progress in this field and ultimately improve patient outcomes in cancer treatment.

9. Conclusion

Fruit-derived exosomes stand as a testament to the powerful synergy between nature's bounty and cutting-edge medical science. These nanoscale vesicles, derived from fruits, hold immense promise for revolutionizing cancer treatment by ushering in a new era of precision therapy. Their inherent biocompatibility, low immunogenicity, and remarkable ability to navigate biological barriers position them as exceptional candidates for targeted drug delivery, offering a potent weapon in the fight against cancer. While the field is still in its nascent stages, the therapeutic potential of fruit-derived exosomes is undeniable. Early research has illustrated their ability to effectively encapsulate and deliver a diverse array of therapeutic agents, including chemotherapeutics, small interfering RNAs, and microRNAs, directly to cancer cells with remarkable precision. This targeted approach minimizes off-target effects, reducing the often debilitating side effects associated with conventional cancer treatments. However, like any evolving field, challenges remain. Standardizing extraction and isolation methods is crucial to ensure consistent purity, yield, and ultimately, therapeutic efficacy. The lack of standardized protocols currently hinders the ability to compare results across studies and establish a clear consensus on the clinical potential of fruit-derived exosomes. Furthermore, a deeper understanding of fruit-derived exosome biogenesis, cargo loading mechanisms, and their intricate interactions with recipient cells is essential to fully unlock their therapeutic potential. Despite these challenges, the future of fruit exosome-based cancer therapy is ripe with opportunities. Advancements in nanotechnology and molecular engineering hold the key to further enhancing their targeting capabilities, allowing for the development of even more precise delivery systems tailored to specific cancer types and molecular subtypes. This will let us breathe in a future where cancer treatment is not only more effective but also significantly less toxic, improving the quality of life for countless individuals battling this devastating disease. The journey towards realizing the full potential of these nanoscale vesicles in precision cancer therapy requires a collaborative, interdisciplinary approach. By uniting the expertise of plant biologists, nanotechnologists, pharmacologists, and oncologists, we can overcome the current hurdles and pave the way for a future where nature's own delivery vehicles revolutionize cancer treatment. The seeds of progress have been sown, and with continued dedication and innovation, we can cultivate a future where exosomes blossom into a powerful weapon in the fight against cancer.

Data availability

Data sharing is not applicable to this article as no datasets were generated or analyzed during the current study.

Conflicts of interest

The authors of this article declare no conflicts of interest.

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