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Chemoenzymatic synthesis planning guided by synthetic potential scores

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Computer-aided chemoenzymatic synthesis planning integrates the advantages of enzymatic and organic reactions to design efficient hybrid synthesis routes for a target molecule. Existing tools rely on either a step-by-step strategy or a bypass strategy. Here we introduce a synthetic potential score (SPScore) to unify these two strategies. This score is developed by training a multilayer perceptron on existing reaction databases to evaluate the potential of enzymatic or organic reactions for synthesis of a molecule. We systematically evaluate the effectiveness of the SPScore in both single-step and multi-step hybrid retrosynthesis, demonstrating its strong ability to prioritize promising reaction types. In benchmarking various chemoenzymatic retrosynthesis algorithms guided by the SPScore, we find that an asynchronous search algorithm named ACERetro yields higher efficiency and robustness that can find hybrid synthesis routes to 46% more molecules compared with the state-of-the-art tool using a test dataset consisting of 1001 molecules. We then apply ACERetro to design efficient chemoenzymatic synthesis routes for 4 FDA-approved drugs. We anticipate that the application of the SPScore will provide a new avenue for computer-aided chemoenzymatic synthesis planning, thereby advancing the synthesis of functional molecules.

Introduction

Enzymatic and organic reactions span distinct reaction spaces in terms of designing synthesis routes for molecules of interest due to their different characteristics.¹ Enzymatic reactions typically exhibit excellent selectivity (stereo-, chemo-, or regio-), while organic reactions are advantageous due to their broad substrate scope, different types of reactions, and numerous well-studied cases. Combining these two reaction types can capitalize on their unique advantages to build more efficient chemoenzymatic synthesis routes to many compounds.^{2–6} A prominent example is the use of an engineered ribosyl-1-kinase for the synthesis of molnupiravir, an antiviral drug, which shortened the original synthesis route by 70% and achieved a sevenfold higher yield.⁷

Computer-aided synthesis planning (CASP) enables massive search and design of synthesis routes for a target molecule by integrating template-based^{8,9} or template-free^{10,11} single-step

retrosynthesis predictors with a search algorithm.^{12,13} To achieve computer-aided chemoenzymatic synthesis planning, currently there are two distinct strategies to build a search algorithm (Fig. 1A) including (i) step-by-step^{14,15} and (ii) bypass.^{16,17} The step-by-step strategy combines the results from single step enzymatic/organic reaction precursor predictors to build a hybrid synthesis route,^{14,15} whereas the bypass strategy identifies alternative reaction types in an existing or predicted synthesis route, *i.e.* identify enzymatic reactions as bypasses to chemical syntheses or *vice versa*.^{16,17} Levin *et al.*'s tool combines precursor prediction results from two reaction template prioritizers trained on separate reaction databases,¹⁴ but template prioritizers cannot heuristically identify the bypass without proper alignment as the two prioritizer models are trained separately. Similarly, Sankaranarayanan *et al.*'s tool employs an exhaustive search to identify biocatalytic opportunities for intermediates in predicted synthesis routes,¹⁶ but this tool is challenging to scale in an exponentially growing search space. More recently, Zeng *et al.*'s step-by-step strategy tool predicts reaction types (organic or enzymatic reaction) with a template-free precursor predictor,¹⁵ while Li *et al.*'s bypass strategy tool builds a reaction type score (RTscore) to distinguish synthesis reactions from decomposition reactions,¹⁷ highlighting the importance of heuristic methods for advancing computer-aided chemoenzymatic retrosynthesis. However, Zeng *et al.*'s method and Li *et al.*'s method can only be applied within their own step-by-step or bypass strategies. Finding a simple and effective way to unify the step-by-step strategy and the bypass strategy can

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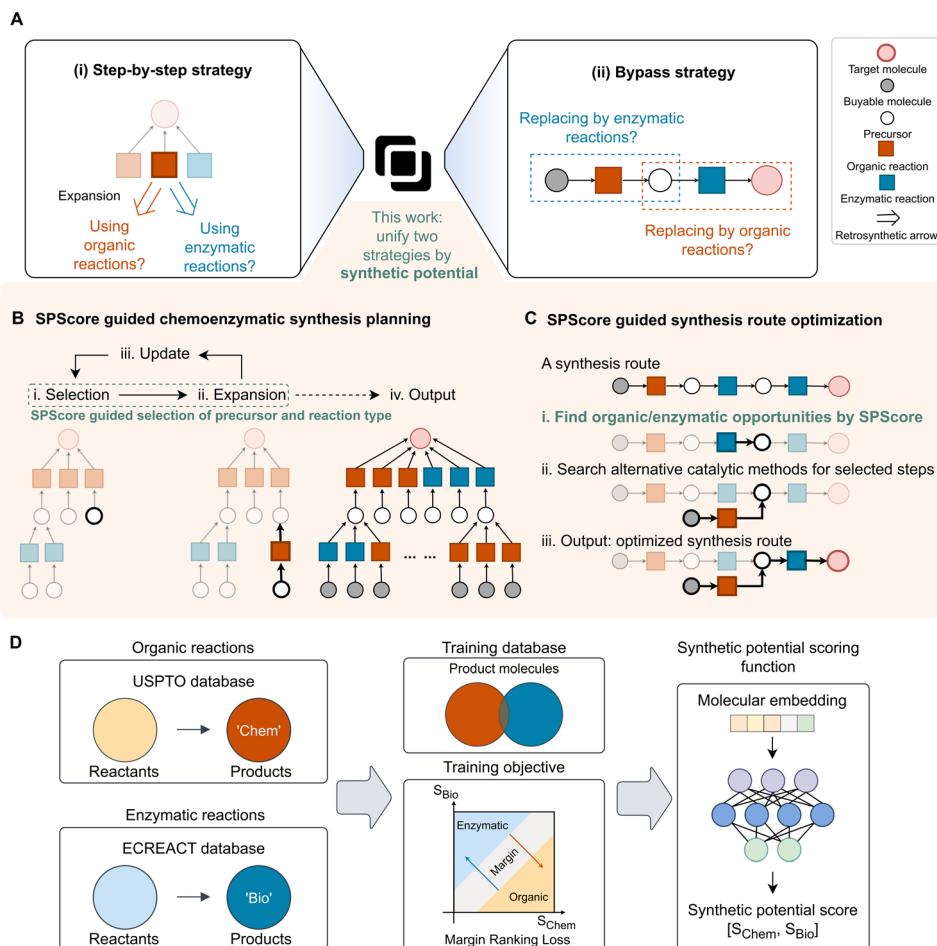


Fig. 1 Chemoenzymatic synthesis planning guided by a synthetic potential score. (A) Workflow of chemoenzymatic retrosynthesis strategies and representative work. (B) Workflow of the SPScore guided chemoenzymatic synthesis planning process. The target molecule is labeled by a red circle, and organic and enzymatic reactions are labeled by red squares and blue squares, respectively. (i) Selection: the molecule with the lowest score in the priority queue is selected. (ii) Expansion: the retrosynthesis tool using the reaction type inferred by the SPScore is used to predict reactions and precursors for the selected molecule. (iii) Update: the expansion results are added to the search tree. Precursors are scored and appended to the priority queue. (iv) Output: steps i, ii, and iii are executed recursively until a termination condition is met. When the search process is terminated, synthesis routes to the target molecule that started with buyable molecules (gray circles) are returned. (C) Workflow of the SPScore guided synthesis route optimization. (i) Identify steps with opportunities for improvement using the SPScore. For a given synthesis route, the SPScore of each molecule is computed. The selected steps (bold) are determined based on their predicted SPScores that deviate significantly from the actual reaction type used in the existing route. (ii) Search alternative reaction types for the selected steps. The synthesis planning algorithm in (B) is used to search synthesis routes with alternative reaction types for the selected steps. (iii) Output. The promising search results are appended to the original route, and the optimized route is returned. (D) Development of the synthetic potential scoring function. Reaction product molecules were extracted from USPTO (organic reactions) and ECRACT (enzymatic reactions), respectively. A neural network model is trained to infer the promising reaction type for a given molecule through the predicted SPScore.

help us further understand the principles behind computer-aided chemoenzymatic synthesis planning and promote the development of efficient hybrid synthesis planning tools. One precedent is that the introduction of synthetic complexity in retrosynthesis can help synthesis planning tools efficiently find concise and feasible synthesis routes.^{18,19} The context of chemoenzymatic retrosynthesis motivates us to introduce a synthetic potential score (SPScore)—a hypothetical metric describing the suitability of enzymatic and organic reactions for synthesizing a molecule. This score not only bridges the gap between the step-by-step strategy and the bypass strategy but also enhances the step-by-step strategy algorithm, transforming

it from a mere replica of traditional retrosynthesis algorithms designed for single reaction types into a more versatile and innovative approach.

In this work, we integrate the above-mentioned two strategies by using the SPScore to prioritize the reaction type (enzymatic or organic) in step-by-step chemoenzymatic synthesis planning (Fig. 1B) and identify alternative reaction types in given synthesis routes (Fig. 1C). This score was developed by training a multilayer perceptron on existing reaction corpora (Fig. 1D). We evaluated the performance of the SPScore on both single-step and multi-step retrosynthesis and developed a SPScore guided asynchronous search algorithm for

chemoenzymatic synthesis planning. The resulting strategy named the asynchronous chemoenzymatic retrosynthesis planning algorithm (ACERetro) can identify chemoenzymatic synthesis routes for 46% more molecules compared to the state-of-the-art tool when using 1001 molecules as a test dataset. To demonstrate its utility, we applied ACERetro to design promising chemoenzymatic synthesis routes for two FDA-approved drugs, ethambutol and Epidiolex. In addition, we applied ACERetro to optimize the synthesis routes of two additional FDA-approved drugs, rivastigmine and *(R,R)*-formoterol. A user-friendly web interface of ACERetro could be accessed at <https://aceretro.platform.moleculemaker.org/search-routes>.

Results

Development of the synthetic potential score

The synthetic potential of a molecule in the organic or enzymatic reactions is influenced by its structure and the current knowledge of synthetic chemistry and biochemistry. As a proof of concept, we demonstrated that synthetic potential can be learned from reaction data by using molecular fingerprints, which capture substructures, combined with a multilayer perceptron (MLP). The method is based on the premise that if a molecule has documented reactions for its synthesis, the reaction type of these reported reactions is the molecule's promising reaction type. A dataset comprising reaction products was extracted from two primary sources: USPTO 480K,²⁰ which contains 484 706 organic reactions, and ECREACT,²¹ which contains 62 222 enzymatic reactions. After removing duplicates and molecules that could not be converted into valid molecular fingerprints for each respective reaction type, the resulting dataset comprised 437 781 molecules in organic reactions and 37 939 molecules in enzymatic reactions, while 515 molecules were present in both reaction types.

Molecules were represented by ECFP4 (ref. 22) (extended connectivity fingerprint, up to four bonds) and MAP4 (ref. 23) (MinHashed Atom Pair fingerprint, diameter $d = 4$) with three different lengths (length = 1024, 2048, and 4096) and used to train several MLP models. Rather than predicting a binary label indicating the preferred catalysis type, the MLP is trained to generate two continuous values: the synthetic potential score for organic reactions (S_{Chem}) and for enzymatic reactions (S_{Bio}). These two scores are directly output by the MLP and reflect how favourable each reaction type is for a given molecule. Because the reaction corpus may not cover all possible transformations, formulating this task as a binary classification is not ideal. Instead, we use margin ranking loss as the training objective, which encourages the model to rank the more promising reaction type higher based on relative differences between S_{Chem} and S_{Bio} (see Methods). This loss function is well-suited for tasks where the goal is to learn a preference between two options. In our case, it allows the model to rank one catalytic option over the other, which better aligns with the decision-making nature of hybrid retrosynthetic planning. The SPScores range from 0 to 1, so they can act as the probability of a molecule being promisingly synthesized by each reaction type. When the difference between two SPScores of a molecule is

within the margin, both reaction types are considered promising for the synthesis of that molecule. If a molecule's SPScore of one reaction type is greater than the other, and the difference is greater than the margin, the reaction type with the larger SPScore is more suitable for the synthesis of the molecule. In the training process, a margin of 0.15 was used, which helps ensure that the three regions have similar areas. A margin that is neither too severe nor too trivial benefits subsequent adjustments to user preferences without the need of model retraining on a different margin. An excessively large number of epochs will cause the model to overfit the distribution of the training data.¹⁸ Since reactions sourced from the USPTO are confined to patents and differ in distribution from those documented in the literature,²⁴ the number of epochs is also considered in the evaluation criteria. Therefore, the best model was obtained by a comprehensive evaluation of precision, recall, and F_1 on the validation dataset, as well as the number of epochs (Fig. S2). As a result, the model that used ECFP4 with

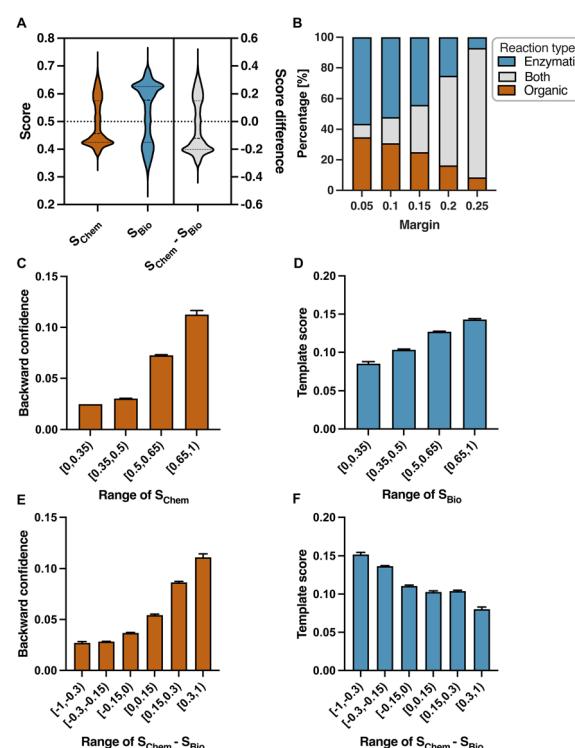


Fig. 2 Analysis of the SPScore on molecules from the ZINC "in vitro" subset. (A) The distribution of S_{Chem} , S_{Bio} and the score difference ($S_{\text{Chem}} - S_{\text{Bio}}$). (B) The percentage of predicted type of catalysis of molecules versus different margin settings. (C) The mean backward confidence with SEM (the standard error of the mean) versus the range of S_{Chem} . In organic reactions, RXN4Chemistry is used to predict retrosynthetic reactions for the molecules. The average backward confidence of the top-5 predictions is sorted by the range of molecules' SPScore in organic reactions. (D) The mean template score with SEM versus the range of S_{Enzy} . In enzymatic reactions, Levin *et al.*'s enzymatic templates are used to predict enzymatic reactions for the molecules. The average template score of the top-5 predictions is sorted by the range of molecules' SPScore in enzymatic reactions. (E) The mean backward confidence with SEM versus the range of $S_{\text{Chem}} - S_{\text{Bio}}$. (F) The mean template score with SEM versus the range of $S_{\text{Chem}} - S_{\text{Bio}}$.



a length of 4096 as molecular embedding will be used for the subsequent tasks.

Benchmarking SPScores on single-step retrosynthesis

To evaluate the performance of SPScores, we assessed the benchmark of our scoring model on a dataset comprising 11 003 molecules randomly selected from the “*in vitro*” subset of ZINC15 (ref. 25) that were not in the training dataset. Subsequently, their corresponding SPScores were calculated. The distribution of two SPScores and the difference of SPScores are shown in Fig. 2A. Although the margin used to train the model is fixed, a flexible user-defined margin can be adopted for different tolerance of reaction types during application. The use of margin also provides an intuitive perspective to understand the applications of SPScores. With the margin increasing from 0.05 to 0.25, more molecules are located in the region where molecules can be promisingly synthesized by both reaction types (Fig. 2B).

Since there is no definitive ground truth for the synthetic potential of a given molecule, the evaluation of the SPScore relies on indirect evidence on retrosynthesis results to demonstrate its practicality and robustness. To further explore whether the SPScore effectively predicts a molecule's promising reaction type, we conducted one-step retrosynthesis in each reaction type by employing RXN4Chemistry²⁶ for organic reactions and Levin *et al.*'s enzymatic templates¹⁴ for enzymatic reactions. For a given target molecule, RXN4Chemistry predicts possible organic reactions ranked by a confidence score, namely backward confidence, while Levin *et al.*'s enzymatic templates predict possible enzymatic reactions ranked by the template score. The average backward confidence of the top-5 predictions increases with the mean synthetic potential score in organic reactions predicted by our scoring model (Fig. 2C). A similar trend between the average template score and the mean synthetic potential score in enzymatic reactions is observed (Fig. 2D). This suggests that as the predicted probability of molecules being synthesized by a specific reaction type increases, the corresponding retrosynthesis tool's confidence in its predictions also tends to increase.

To assess whether the relative value of S_{Chem} and S_{Bio} can help identify the dominant reaction type, we analyzed both the average backward confidence and the average template score for top-5 predictions *versus* the mean SPScore difference ($S_{\text{Chem}} - S_{\text{Bio}}$) (see Fig. 2E and F). The result reveals that when S_{Bio} is larger than S_{Chem} , molecules tend to have a relatively high template score to be synthesized by enzymatic reactions and low backward confidence to be synthesized by organic reactions. Collectively, these trends shown in Fig. 2 suggest that our scoring function exhibits good ability to deduce the promising reaction type for molecules.

Benchmarking SPScores on multi-step retrosynthesis

A multi-step synthesis route dataset is used to evaluate the performance of SPScores on multi-step retrosynthesis. Due to the limited availability of databases containing multi-step synthesis routes, particularly for hybrid synthesis, the

synthesis routes of 493 target molecules identified by Levin *et al.*'s hybrid planner within three minutes were used for this *in silico* benchmarking study.¹⁴ The SPScore prediction of molecules is compared with the reaction used in the synthesis routes. Out of 397 040 synthesis routes linked to the 493 target molecules, 26 741 distinct product molecules with their reaction types were identified. In particular, 9162 (34.3%) molecules were synthesized exclusively by organic reactions, 10 211 (38.2%) molecules were synthesized exclusively by enzymatic reactions, and 7368 (27.5%) molecules were synthesized by both organic reactions and enzymatic reactions and are in the overlap part. Furthermore, from the 1531 shortest synthesis routes related to the 493 target molecules, 1544 unique product molecules with their respective reaction types were identified: 788 (51.0%) in organic reactions, 481 (31.2%) in enzymatic reactions, and 275 (17.8%) spanning both fields.

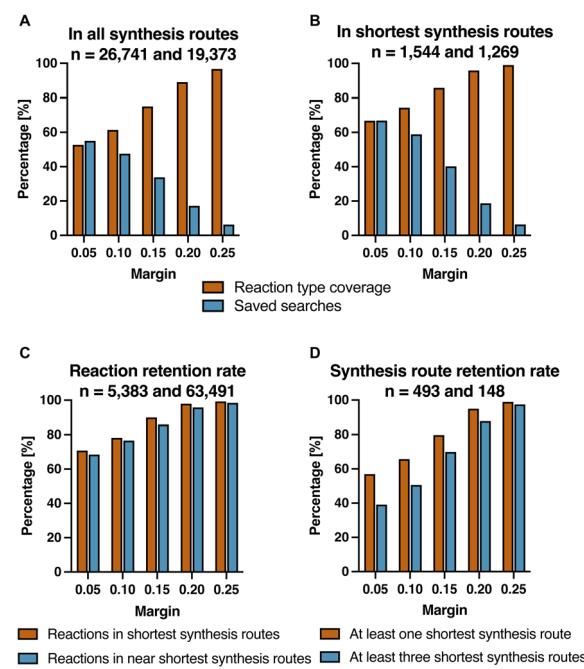


Fig. 3 Analysis of SPScores on synthesis routes. Reaction products are extracted from reactions in all synthesis routes and molecules' shortest synthesis routes predicted by Levin *et al.*'s tool on 493 molecules. Molecules are assigned with the ground truth label of reaction type based on the extracted reaction set. The amount of “reaction type coverage” is counted when the SPScore gives the correct prediction or predicts it as “both”. The “saved searches” means that the SPScore gives the correct reaction type prediction. The percentage of reaction type coverage out of all molecules (red) and saved searches out of non-“both” molecules (blue) are calculated against different margins for product molecules (A) in all synthesis routes and (B) in shortest synthesis routes. (C) The reaction retention rate for the shortest synthesis routes (red) and near shortest synthesis routes (blue) against different margin settings. The reactions from shortest synthesis routes and the near shortest synthesis routes (where the route length \leq shortest route length + 2) are extracted. The amount of retained reactions is counted when the SPScore gives the correct reaction type prediction for the product molecule or predicts it as “both”. (D) The shortest synthesis route retention rate against different margin settings. The amount of retained synthesis routes is counted when all the reactions in the synthesis route can be retrained by the SPScore's prediction.



When using SPScores to guide the search where the margin is set as 0.15, 85.8% of molecules' synthesis field in shortest synthesis routes and 75.0% of molecules' synthesis field in all synthesis routes can be covered. By this way, it can save 40.2% searches in shortest synthesis routes and 33.8% searches in all synthesis routes because SPScores can give the correct prediction that matches the reaction type in the original synthesis routes (Fig. 3A and B). The observed trend indicates that as the margin expands, the reaction type of a greater number of molecules is encompassed. However, this comes at the expense of a reduced number of saved searches.

The reaction retention rate is defined as the proportion of reactions where the actual reaction type matches the predicted preferred reaction type for the product molecule, as determined by the SPScore (see Methods and SI). When the margin is set as 0.15, 89.9% of reactions in the shortest synthesis routes can be covered, and 86.0% of reactions in the near shortest synthesis routes (route length \leq shortest route length + 2) can be covered (Fig. 3C). Next, we investigate whether the SPScore can provide guidance for finding the shortest synthesis route and discovering the diversity of shortest routes. Of 493 target molecules, the route retention rate is determined by counting the number of molecules that at least one shortest synthesis route whose actual reaction types can be covered by SPScores' prediction. In scenarios with the same margin of 0.15, 393 (79.7%) of the molecules have at least one shortest synthesis route that can be fully retained, while 109 (73.6%) molecules out of 148 have at least three shortest synthesis routes that can be retained (Fig. 3D). The results indicate that in the context of multi-step retrosynthesis, utilizing SPScores as a guide enables the retention of majority of favorable synthesis routes.

Searching for chemoenzymatic synthesis routes

Searching for chemoenzymatic synthesis routes requires predicting precursors in organic and enzymatic reactions.

Synchronous methods, like Levin *et al.*'s tool,¹⁴ search for precursors in enzymatic and organic reactions simultaneously and then combine the search results. In this study, we report a SPScore guided asynchronous method, ACERetro, which prioritizes the search of the most promising reaction type for a given molecule. To evaluate the performance of ACERetro (Fig. 4C), we conducted a self-benchmark study using two simplified versions of the algorithm: a fully hybrid synchronous algorithm (FHSync, Fig. 4A) and a SPScore-guided synchronous algorithm (SPSync, Fig. 4B). These two variations serve as ablation studies to assess the contribution of each component within ACERetro. FHSync does not incorporate SPScores or asynchronous search, while SPSync includes SPScore-based guidance but uses a synchronous (non-asynchronous) search strategy. The single-step precursor prediction tools previously used in the "in vitro" subset of ZINC15, namely RXN4Chemistry and Levin *et al.*'s enzymatic templates, were respectively used for organic reactions and enzymatic reactions. The fully hybrid search algorithm (FHSync) without SPScores directly searches both organic reactions and enzymatic reactions, and the results are combined in each step, while the SPScore guided synchronous hybrid search algorithm (SPSync) only searches for a promising reaction type predicted by the SPScore. In the SPScore guided asynchronous hybrid search algorithm (ACERetro), the SPScore is used to guide separate search processes for each reaction type (see details in Methods). By running these searches asynchronously, ACERetro can prioritize the more promising reaction type while still exploring the alternative. This setup improves the robustness of the planning process, allowing the algorithm to recover and switch paths if the initially favoured reaction type turns out to be less viable.

To explore search spaces of these three algorithms, we conducted a comparative study on a set of 1001 molecules from ZINC, which Levin *et al.*'s tool had explored under identical boundary conditions including search time and buyable dataset

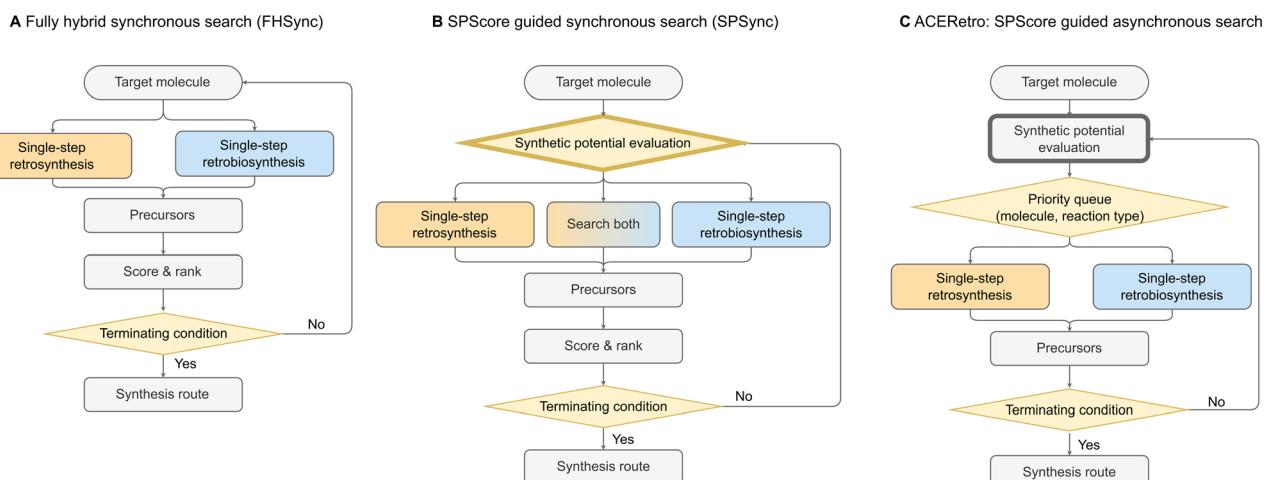


Fig. 4 Hybrid search algorithms for designing chemoenzymatic synthesis routes. (A) Fully hybrid synchronous search algorithm (FHSync). (B) SPScore guided synchronous search algorithm (SPSync). (C) ACERetro: SPScore guided asynchronous search algorithm. FHSync and SPSync are included as ablation studies to evaluate the components of ACERetro. FHSync represents a version without synthetic potential scoring and without asynchronous search, while SPSync includes synthetic potential scoring but does not use asynchronous search. Modules that incorporate SPScores are indicated with bold edges in the diagrams.



(see Methods). FHSync found synthesis routes to 597 molecules, SPSync found synthesis routes to 683 molecules, and ACERetro found synthesis routes to 720 molecules (Fig. 5A). Compared to Levin *et al.*'s tool, which found synthesis routes to only 493 molecules, FHSync can find synthesis routes to additional 104 (21.1%) molecules. This improvement is mainly attributed to the incorporation of the template-free model, RXN4Chemistry, in organic reactions. Moreover, the SPSync and ACERetro found synthesis routes to 190 (38.5%) and 227 (46.0%) more molecules

compared with Levin *et al.*'s tool, respectively. These results underscore that the efficiency of ACERetro surpasses that of the state-of-the-art method.

In a self-benchmarking analysis, the hybrid search algorithms with SPScore guidance (SPSync and ACERetro) outperform the algorithm without SPScore guidance (FHSync), which could find synthesis routes to 86 and 123 more molecules, emphasizing the pivotal role of SPScore in optimizing search efficiency. In the comparison between SPSync and ACERetro,

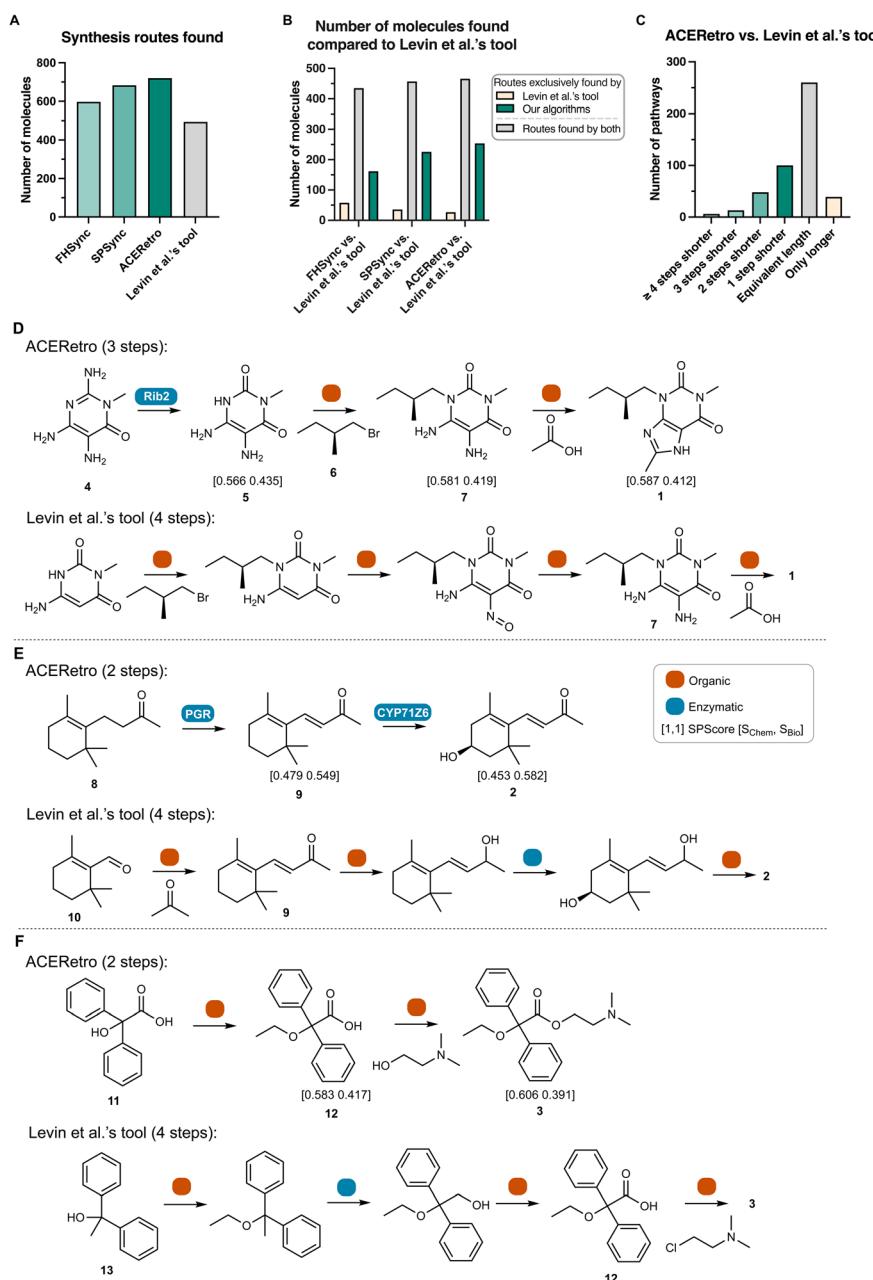


Fig. 5 Comparison of synthesis routes found by hybrid search algorithms. (A) Number of molecules for which synthesis routes were found out of 1001 molecules. (B) Number of molecules whose synthesis routes can be found by Levin *et al.*'s tool (red) only, both (grey), and ours (blue). (C) Comparison of the number of steps in the shortest synthesis route found by ACERetro compared to the shortest synthesis route found by Levin *et al.*'s tool for molecules for which synthesis routes were found by both (466 total) from the ZINC15 "boutique" subset. The example synthesis routes of (S)-verofylline (**1**), (3S)-3-hydroxy-β-ionone (**2**), and dimenoxadol (**3**) are shown in (D–F). All product molecules, except for **9**, do not appear in the training set of the SPScore. Cofactors and some non-primary reactants are ignored. Rib2: 2,5-diamino-6-(5-phospho-D-ribityl-amino)-pyrimidin-4(3H)-one deaminase, PGR: 13,14-dehydro-15-oxoprostaglandin 13-reductase, CYP71Z6: ent-isokaurene C2-hydroxylase.



ACERetro could find synthesis routes to 37 more molecules, which indicates that the asynchronous search is more efficient than the synchronous search. Unlike the synchronous search, which drops the search for molecules' suboptimal reaction type, the asynchronous search keeps all suboptimal reaction type of molecules in the queue for later exploration. The algorithm will start to search suboptimal reaction type of molecules based on a comprehensive consideration including SPScores, search depth, and molecular complexity (see Methods).

Variations in search spaces and strategies across synthesis planners lead to the prediction of different synthesis routes for molecules. A proficient planner can discover synthesis routes to a greater number of molecules than other planners are able to find. Thus, we conducted a comparative analysis to evaluate the number of molecules whose synthesis routes were exclusively identified by the three algorithms in comparison to Levin *et al.*'s tool (Fig. 5B). It was observed that each algorithm has the capability to discover synthesis routes for molecules that Levin *et al.*'s tool did not identify. In particular, out of 1001 molecules, synthesis routes to 466 could be found by both ACERetro and Levin *et al.*'s tool. While ACERetro exclusively identified synthesis routes to 254 molecules, Levin *et al.*'s tool could exclusively find routes to only 28 molecules, indicating that ACERetro discovered approximately 26 times more exclusive molecules than Levin *et al.*'s tool. These findings imply that ACERetro achieves an expanded search space and better heuristic search strategy than the state-of-the-art tool.

The search quality of the synthesis planning tools can be evaluated from the number of reactions in the predicted synthesis routes through limited context that synthesis planning tools can provide, albeit it is not an exhaustive metric.²⁷ A smaller number of steps usually imply the use of fewer reagents and fewer purification steps.²⁸ We compared the length of the shortest synthesis route to 466 molecules found by both ACERetro and Levin *et al.*'s tool. ACERetro found optimized shortest synthesis route to 167 (35.8%) molecules and the shortest synthesis route of equivalent length for 260 (55.8%) molecules (Fig. 5C). This indicates that ACERetro can predict more optimized synthesis routes than Levin *et al.*'s tool.

To further study the difference in search space between ACERetro and Levin *et al.*'s tool, we compared the synthesis routes to (*S*)-verofylline (**1**), (*3S*)-3-hydroxy- β -ionone (**2**), di-menoxadol (**3**) (Fig. 5D–F), and other 7 syntheses (Fig. S8). In the synthesis of **1**, ACERetro predicted a three-step hybrid synthesis route including one enzymatic reaction, while the shortest synthesis route predicted by Levin *et al.*'s tool included four reactions in organic reactions (Fig. 5D). The route predicted by ACERetro first uses an enzymatic reaction to synthesize **5** from **4**. The recommended enzyme is 2,5-diamino-6-(5-phospho-D-ribitylamino)-pyrimidin-4(3*H*)-one deaminase (Rib2; EC number 5.4.99.28). **5** is subsequently alkylated with **6** containing a chiral center to form **7**. The final step constructs an imidazole ring using acetic acid with **7** to produce **1**. Note that the Levin *et al.*'s tool route uses the same strategy to introduce the chiral center and construct the imidazole ring to **1**, but the difference in starting materials makes the route longer. In the synthesis routes of **2**, ACERetro predicted a two-step enzymatic

synthesis route, while Levin *et al.*'s tool predicted a four-step hybrid synthesis route (Fig. 5E). The former first uses a reductase to get the double bond starting with dihydro- β -ionone (**8**) to form β -ionone (**9**). Next, a hydroxylase is used to introduce the chiral hydroxyl group for **9** to form **2**. Recommended enzymes are 13,14-dehydro-15-oxoprostaglandin 13-reductase (PGR; EC number 1.3.1.48) and *ent*-isokaurene C2-hydroxylase (CYP71Z6; EC number 1.14.14.76), respectively. The latter uses a different starting material, β -cyclocitral (**10**) to form **9**, and three steps to form **2** from **9**. In the synthesis routes of **3**, ACERetro predicted a two-step synthesis route including only chemical reactions, while Levin *et al.*'s tool predicted a four-step hybrid synthesis route (Fig. 5F). The former first constructs ether from benzilic acid (**11**) to form **12** and then constructs ester to form **3**. The latter uses a similar reaction to form the final product from **12**. However, it uses 1,1-diphenylethanol (**13**) as the starting material to synthesize **12** via a three-step hybrid synthesis route.

The routes of **1**, **2**, and **3** predicted by ACERetro cover three scenarios: hybrid approach, purely organic approach, and purely enzymatic approach. The results show that ACERetro can often find shortcuts to synthesize compounds compared to Levin *et al.*'s tool, such as the synthesis of intermediate **7** in the synthesis route of **1**, the synthesis route from **9** to **2**, and the synthesis of **12** in the synthesis route of **3**. For the predicted enzyme reactions, although those enzymes have not been reported to use molecules in the predicted routes as substrates, the predicted reactions still provide effective guidance for future enzyme discovery and engineering. Among all routes predicted by ACERetro, the SPScore of each product except **5** is consistent with the corresponding reaction type in the synthesis route. However, note that S_{Bio} of **5** is higher than that of all other products in the route, and its $S_{\text{Chem}} - S_{\text{Bio}}$ has the smallest value, which indicates that **5** has higher potential to be synthesized by enzymatic reactions compared to other product molecules in the synthesis routes.

Applying ACERetro for synthesis planning

Ethambutol is a drug used in the treatment of tuberculosis (TB). The (*S,S*)-enantiomer, ((*S,S*)-ethambutol; (*S,S*)-**14**), is the most active antimycobacterial agent compared with other three isomers.^{29–31} Wilkinson *et al.* first reported the synthesis route for (*S,S*)-**14**, utilizing (2*S*)-2-aminobutan-1-ol (**15**) as the starting material.²⁹ Likewise, the synthesis routes of (*S,S*)-**14** developed by Butula *et al.*³² and Stauffer *et al.*³³ also used starting materials containing chiral centers (**15** and **18**) directly. Trost *et al.* used palladium catalyzed stereoselective epoxide (**16**) opening on phthalimide (**17**) to construct the chiral center,³⁴ while Kotkar *et al.* reported a synthesis route using proline-catalyzed α -aminoxylation on butyraldehyde (**19**)³⁵ (see Fig. 6A–E).

We conducted retrosynthesis planning on (*S,S*)-ethambutol by using ACERetro. The search parameters are the same as those used in the above-mentioned benchmarking study, except that the maximum search depth is set to 5 based on existing routes. The most promising predicted synthesis route connecting to buyable compounds is shown in Fig. 6F. The



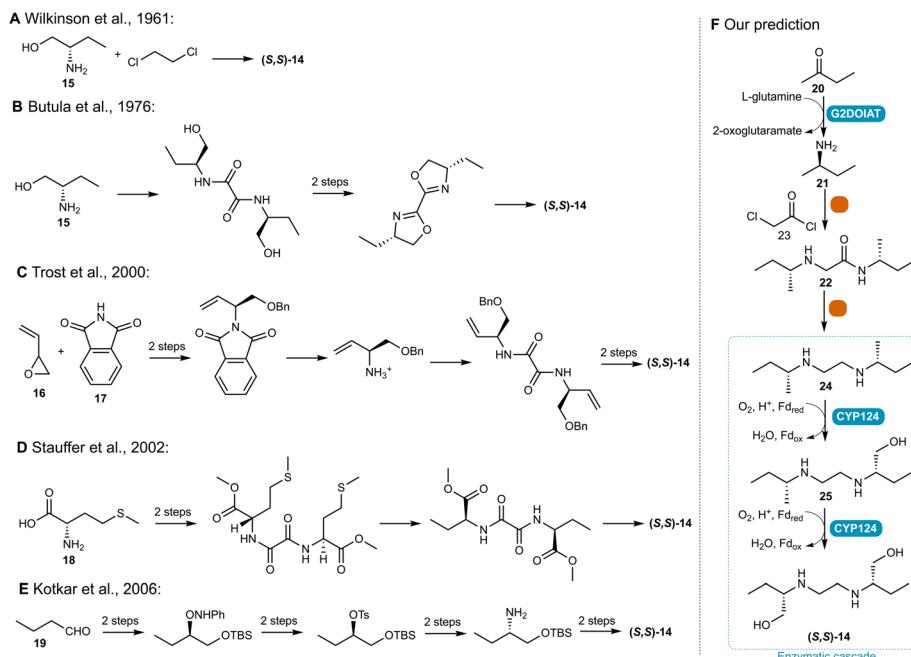


Fig. 6 Synthesis routes of ethambutol. (A–E) Published synthesis routes of ethambutol. (F) Predicted synthesis route of ethambutol. Ethambutol does not appear in the training set of the enzymatic model. G2DOIAT: L-glutamine:2-deoxy-scyllo-inosose aminotransferase, CYP124: CYP124 family of cytochrome P450 enzymes

synthesis route first builds the chiral center through an enzymatic reaction of aminotransferase from cheap starting material 2-butanone (**20**) to form *(2R*)-butan-2-amine (**21**). **22** is synthesized by the acylation reaction of **23** and **21**, followed by reduction to form **24**. Two steps of symmetrical hydroxylation catalyzed by the same enzyme are used to complete the synthesis of **12**.

The predicted route effectively employs a single enzymatic reaction to construct the chiral portion of the molecule. Compared to chemical methods reported in the literature, the enzymatic reaction conditions are milder. The enzyme recommended for this step is L-glutamine:2-deoxy-scyllo-inosose aminotransferase (G2DOIAT; EC number 2.6.1.100). The subsequent two symmetric hydroxylation reactions form a cascade, and a one-pot method can be employed to minimize the number of purifications. The CYP124 family of cytochrome P450 enzymes (CYP124; EC number 1.14.15.14) is recommended for this cascade. Moreover, introducing the hydroxyl group in the final step avoids side reactions during the acylation process and reduces the use of protecting groups. Although the predicted enzymatic reactions have not been experimentally verified for these substrates, the prediction still provides valuable guidance for future enzyme discovery and engineering.

Epidiolex is the brand name for $(-)$ -cannabidiol ($(-)$ -26), which is used for the treatment of epilepsy disorders. Kobayashi *et al.* developed the synthesis route using olivetol dimethyl ether (27) and 30 as the starting materials.³⁶ The chirality is constructed through the nucleophilic addition of 28 and 31 to form 29. Another synthesis route designed by Shultz *et al.* uses Ireland-Claisen rearrangements to build chirality starting from olivetol 32.³⁷ Gong *et al.* used the Friedel-Crafts reaction to

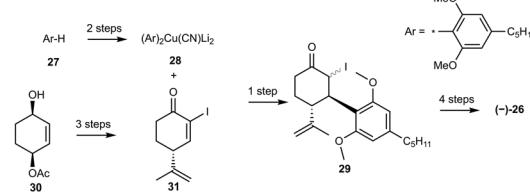
build chirality starting with phloroglucinol (35) and *cis*-isolimonenol (36).³⁸ The biosynthetic route of (–)-cannabidiol using hexanoyl-CoA as the starting material has also been reported.³⁹ The cannabidiolic acid synthase (CBDAS; EC number 1.21.3.8) uses cannabigerolic acid as the substrate to close the ring and introduce stereochemistry (see Fig. 7A–D).

The predicted route by ACERetro with a maximum search depth set to 4 and ignoring geometric isomerism in the buyable molecule database is shown in Fig. 7E. The prediction provides a concise synthesis route, starting with the alkylation of olivetol 32 with geraniol 39 to form cannabigerol 40. Then an enzymatic step is used to form the final product (–)-26 with stereoisomerism. The first alkylation reaction has literature to support it,⁴⁰ whereas the recommended enzyme for the second step, CBDAS, has not been proven to work using 40 as the substrate. However, the high similarity between 40 and 38 points to the possibility of finding enzyme mutants that allow the reaction to occur.

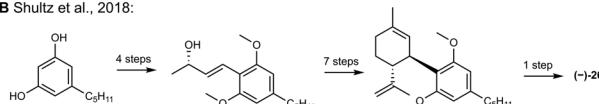
Applying ACERetro for optimizing synthesis routes

The SPSScore can also be used to optimize given synthesis routes by finding steps with opportunities for improvement that can be catalyzed by alternative reaction types in given routes. The steps with opportunities for improvement are selected based on the deviation between the SPSScore predicted reaction type and the reaction type in the original route. ACERetro is then used to search alternative synthesis routes for the selected steps. The promising alternative synthesis route is appended to its original synthesis route to form a new optimized synthesis route for a given route. The utility of SPSScore and ACERetro has been examined in the previous sections. To further showcase their

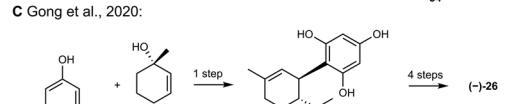
A Kobayashi et al., 2006:



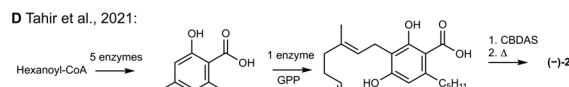
B Shultz et al., 2018:



C Gong et al., 2020:



D Tahir et al., 2021:



E Our prediction:

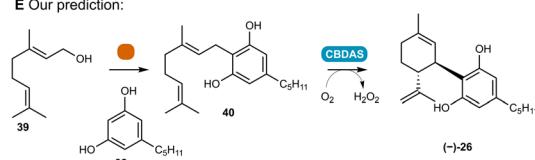


Fig. 7 Synthesis routes of Epidiolex. (A–D) Published synthesis routes of Epidiolex. (E) Predicted synthesis route of ethambutol. Epidiolex does not appear in the training set of the enzymatic model. CBDAS: cannabidiolic acid synthase.

combined ability to optimize synthesis routes, we present case studies on the synthesis route of rivastigmine (**41**) reported in the literature⁴¹ and a synthesis route for (*R,R*)-formoterol (**42**) reported by a synthesis planning tool.¹⁴ In the four-step organic synthesis route of the dementia drug rivastigmine **41**, the S_{Chem} of each molecule is greater than its S_{Bio} . Therefore, the intermediate **44** with the largest SPScore difference (“optimization score”) was selected to search possible synthesis routes using enzymatic reactions. ACERetro with the same parameters was used for the search, and the maximum search depth was set to 1 because the intermediate **44** in the original synthesis route only takes one step to reach the commercially available molecule. The search results show that an enzymatic reaction can be found using the same starting material **43** (Fig. 8A). This enzymatic reaction has been validated in the literature,⁴² proving the effectiveness of SPScores in finding steps with opportunities for improvement and then optimizing the route.

The chemoenzymatic synthesis route for (*R,R*)-formoterol **42** was predicted by Levin *et al.*’s tool. Top 3 steps with opportunities for improvement (**46**, **48**, and **49**) were identified where their predicted SPScores are far away from their reaction type in the original route. In particular, the S_{Chem} values of intermediates **46**, **48**, and **49** are larger than their corresponding S_{Bio} , yet enzymatic reactions were employed in the original route which causes a high optimization score. The new organic synthesis

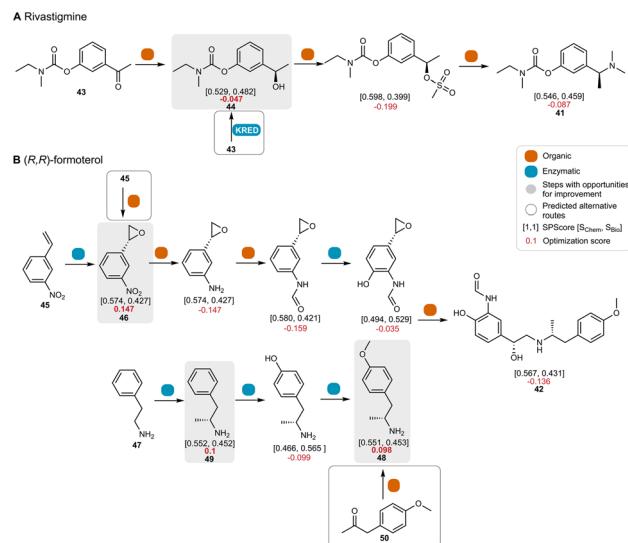


Fig. 8 SPScore guided synthesis route optimization. (A) Synthesis route optimization of rivastigmine. (B) Synthesis route optimization of (*R,R*)-formoterol. Steps with opportunities for improvement do not appear in the training set of the SPScore. (+)-Phenylethylamine, which is available in the buyable database, and other reagents are not shown in the diagram. The predicted bypasses have literature support. KRED: ketoreductase.

route for intermediate **46**, utilizing **45** as the starting compound, was predicted by ACERetro with a search depth capped at 1. Given that intermediates **49** and **48** are in the same branch, only the analysis for **48** is shown in Fig. 8B, which was undertaken by ACERetro with a maximum search depth of 2. The proposed route employs one-step chemical reaction to synthesize **48**, taking **50** and (+)-phenylethylamine as the precursor, which reduces the original three-step synthesis strategy to a single step. These predicted reactions for intermediates **46** and **48** have been corroborated by the literature.^{43,44}

Discussion

In this work, we identified the underlying principle between the step-by-step strategy and the bypass strategy by emphasizing the role of synthesis potential. We developed a SPScore guided asynchronous chemoenzymatic synthesis planning algorithm named ACERetro for designing chemoenzymatic synthesis routes for target molecules. When considering the evaluation of synthetic potential of molecules in each reaction type for computer-assisted chemoenzymatic synthesis planning, our heuristic search algorithm can prioritize the exploration of the most promising reaction type for a molecule. By leveraging the SPScore, we can also diagnose and then optimize existing synthesis routes through the identification of alternative bypasses. Consequently, the evaluated synthetic potential of molecules constructs a bridge between step-by-step synthesis planning and synthesis route optimization in the design of chemoenzymatic synthesis routes. Performing asynchronous retrosynthetic searches in between the organic reactions and enzymatic reactions can significantly improve search efficiency



and bolster the algorithm's robustness. This allows ACERetro to effectively address the challenge faced by the existing hybrid synthesis planners, which tend to be worse than single model planners in terms of efficiency and performance.

In addition, we capitalize on the characteristics of current organic reaction and enzymatic reaction databases. A sufficiently large organic reaction database can support the training of retrosynthesis tools based on language models, whereas a smaller-scale enzymatic reaction database is more suitable for rule-based reaction templates. Accordingly, we employ a template-free retrosynthesis tool, RXN4Chemistry, for organic reactions and a template-based retrosynthesis tool, ASKCOS, for enzymatic reactions. Free from the limitations imposed by a template prioritization system, ACERetro guided by the SPScore possesses the capability to integrate seamlessly with any existing retrosynthesis tool.

By comparing the confidence of single-step retrosynthesis and single-step retrobiosynthesis of 11 003 molecules with the trend of SPScore distribution, it is shown that SPScores can effectively predict promising reaction types for molecules. The performance of SPScores in multi-step retrosynthesis was further verified by reaction type coverage, reaction retention rate, and route retention rate among predicted synthesis routes of 493 molecules. In the benchmarking study on 1001 molecules, ACERetro incorporating two single-step precursor prediction tools outperformed Levin *et al.*'s tool, a state-of-the-art method. Through a comparative analysis of the results obtained from FHSync, SPSync, and ACERetro, self-benchmarking reveals that the incorporation of the template-free model, the implementation of SPScores, and the adoption of asynchronous search methodologies each contribute to enhancing the performance of synthesis planning.

Examples of synthesis routes for (S)-verofylline, (3S)-3-hydroxy-β-ionone, and dimenoxadol reveal that our method can identify shortest synthesis routes with higher quality, and the predictions include not only hybrid synthesis routes, but also chemical reaction only synthesis routes and enzymatic reaction only synthesis routes. The case studies on synthesis planning of ethambutol and Epidiolex demonstrate that our approach can effectively design hybrid synthesis routes for complex molecules and find potential enzyme candidates to perform the predicted enzymatic reactions. The complementarity of the two reaction types will further broaden the scope for designing efficient synthesis routes for molecules of interest. The case studies on synthesis route optimization for rivastigmine and (R,R)-fomoterol illustrate that SPScores can be effectively applied to optimize existing synthesis routes. Existing synthesis tools are often inadequate for lengthy synthesis steps. Finding steps with opportunities for improvement that may be optimized in existing synthesis routes and then conducting retrosynthetic analysis can simplify the search process and make full use of existing parts of the synthesis routes that have been experimentally verified.

The concept underlying SPScores involves inferring the most promising reaction type for a molecule based on existing catalysis data in a reaction database, employing a data-driven approach. This approach aims to differentiate the distinct

reaction spaces of organic reactions and enzymatic reactions. In this work, we performed a simplified but fruitful verification of the synthetic potential with molecular fingerprint and MLP. Combining more complex models such as molecular graphs and reinforcement learning to predict the SPScore will be explored in a follow-up study. Utilizing SPScore in chemoenzymatic synthesis planning can expedite the search process by avoiding less promising reaction types. However, there remains a risk that the model might overlook viable reactions in the reaction types it avoids. Consequently, a comprehensive and high-quality dataset encompassing various types of reactions is crucial to ensure optimal model performance. It is noteworthy that the reaction spaces of organic reactions and enzymatic reactions are dynamic. The unique reaction space of each may expand or contract with the discovery of new catalysts or enzymes. In ACERetro, the SPScore is first used to identify the promising reaction type before conducting a retrosynthetic analysis. An alternative improvement strategy could be first conducting retrosynthetic analysis to identify all potential deconstruction sites and intermediates and then selecting the appropriate reaction type for each step.

In summary, this study transforms chemoenzymatic synthesis planning from a fragmented process into a unified framework by combining the concept of synthetic potential with practical algorithmic design. ACERetro overcomes the limitations of existing methods by integrating both template-based and template-free strategies, enabling comprehensive synthesis planning across diverse organic and enzymatic reaction databases. Its ability to design efficient synthesis routes and identify alternative pathways highlights its potential as a powerful tool in the field. We believe that computer-aided chemoenzymatic synthesis planning will expand the synthesis space by leveraging the complementary strengths of enzymatic reactions and organic reactions. This approach can accelerate the adoption of enzymes as eco-friendly catalysts, facilitating enzyme screening and engineering for improved catalytic performance.

Methods

Training the synthetic potential scoring model

The USPTO 480K database comprises 484 706 organic chemistry reactions from patents, and the ECREACT database comprises 62 222 enzymatic reactions from Rhea,⁴⁵ BRENDA,⁴⁶ PathBank,⁴⁷ and MetaNetX.⁴⁸ After deduplication and excluding molecules with infeasible fingerprints (as detailed in the SI), we extracted 437 781 molecules from USPTO 480K (labeled with $y = 1$) and 37 939 molecules from ECREACT (labeled with $y = -1$). 515 overlapping molecules, found in both reaction types, are labeled with $y = 0$. An MLP model is trained to generate two continuous synthetic potential scores for each molecule: one for organic reactions (S_{Chem}) and one for enzymatic reactions (S_{Bio}). The model takes molecular fingerprints as input and outputs both scores through a final sigmoid activation layer, which bounds the values between 0 and 1. Importantly, the model is not trained to directly predict reaction type labels. Instead, it is optimized using a margin ranking loss based on ground truth



labels $y \in \{-1, 0, 1\}$, which indicate that the molecule's reaction type is enzymatic, organic, or compatible with both. This training approach encourages the model to assign a higher score to the more suitable reaction type for each molecule, allowing it to capture relative preferences rather than making hard classification decisions. The margin uses the relative value of S_{Chem} and S_{Bio} to divide the output space to three areas corresponding to three scenarios of reaction type as shown in Fig. 1A. To evaluate the MLP, we compute accuracy by checking whether the predicted scores satisfy the correct ranking implied by the ground truth label $y \in \{-1, 0, 1\}$. In particular:

- If $y = 1$ (*i.e.*, the reaction type is organic), we consider the prediction correct if $S_{\text{Chem}} > S_{\text{Bio}} + \text{margin}$.
- If $y = -1$ (*i.e.*, enzymatic), the prediction is correct if $S_{\text{Bio}} > S_{\text{Chem}} + \text{margin}$.
- If $y = 0$ (*i.e.*, overlap), the prediction is considered correct if the absolute difference between the two scores is less than the margin, *i.e.*, $|S_{\text{Chem}} - S_{\text{Bio}}| < \text{margin}$.

A weighted margin ranking loss, $\text{loss}(S_{\text{Chem}}, S_{\text{Bio}}, y) = \text{weight}_i \cdot \max(0, -y(S_{\text{Chem}} - S_{\text{Bio}}) + \text{margin})$ if $y = \pm 1$ and $\text{loss}(S_{\text{Chem}}, S_{\text{Bio}}, y) = \text{weight}_i \cdot \max(0, |S_{\text{Chem}} - S_{\text{Bio}}| - \text{margin})$ if $y = 0$, is applied to compute a criterion only when the prediction is out of the area of molecules' true labels. The weight is calculated based on the reciprocal of the ratio of each label.

The dataset was randomly split into a training, validation, and test set (80%, 10% and 10%, respectively). We used a grid search to tune the hyperparameters including the type of the molecular fingerprints (ECFP4 and MAP4), the length of the molecular fingerprints (1024, 2048, or 4096), and the number of hidden layers (1, 3, or 5). The accuracy, F_1 , and recall are calculated on the validation set. To mitigate the risk of overfitting, the number of epochs is incorporated into the evaluation function to select the optimal models (see the SI). The optimal model, which utilizes ECFP4 embedding of 4096 length and comprises 3 hidden layers, trained for 10 epochs, was employed for all subsequent tasks.

Benchmarking the synthetic potential score

11 003 molecules were randomly selected from the "*in vitro*" subset of ZINC15. The SPScore was calculated for each molecule. RXN4Chemistry was employed for one-step retrosynthesis in organic reactions. Each predicted reaction is accompanied by a corresponding backward confidence score. Levin *et al.*'s enzymatic templates were employed for single-step precursor prediction in enzymatic reactions. Each predicted reaction is accompanied by a corresponding template score. The search parameters for RXN4Chemistry and Levin *et al.*'s enzymatic templates are listed in the SI. For molecules within different S_{Chem} intervals, we calculated the average confidence scores for top-5 predictions, and an analogous procedure was undertaken for S_{Bio} intervals and score difference ($S_{\text{Chem}} - S_{\text{Bio}}$) intervals.

Multi-step hybrid synthesis routes were derived from the retrosynthetic predictions for 493 molecules conducted by Levin *et al.*'s tool within a three-minute timeframe. Out of 493 target molecules, we enumerated 26 741 distinct product molecules in 397 040 synthesis routes. All the synthesis routes

with the shortest length for each target molecule were collected, which contained 1544 distinct product molecules. The reaction type of a molecule (denoted as "Chem", "Bio", or "Both") is assigned based on whether the molecule has been synthesized by an organic chemical reaction or an enzymatic reaction. Reaction type coverage out of all molecules counts the molecule whose SPScore-predicted reaction type includes the actual reaction type out of all molecules. Saved searches out of "Chem" and "Bio" molecules count the molecule whose SPScore-predicted reaction type exactly matches the actual reaction type for these molecules labeled with "Chem" or "Bio", so the search algorithm does not need to search the alternative reaction type. The reaction retention rate measures how often the actual reaction type used in a pathway agrees with the preferred reaction type predicted by SPScore for the product of that reaction. In particular, for each reaction in a given dataset, we check whether the reaction type (*e.g.*, chemical or enzymatic) matches one of the reaction types that the SPScore predicts as favorable for the reaction's product molecule. The retention rate is calculated as the percentage of reactions that meets this criterion across the entire dataset (see the SI for the formulae). The near shortest synthesis routes include synthesis routes whose lengths are less than or equal to the shortest synthesis route length plus two. Synthesis route retention rate counts the synthesis route whose reactions can be all retained (see the SI for the formulae).

Development and evaluation of ACERetro

1001 compounds from the "boutique" subset of the ZINC15 database are used in the benchmarking study. Three search algorithms used the identical search parameters of RXN4Chemistry and Levin *et al.*'s enzymatic templates as described in the previous section. All three search algorithms—FHSync, SPSync, and ACERetro—are built on a tree search framework that follows an iterative process of selection, expansion, and update. In the selection mode, the molecule that has the lowest score in the priority queue and is not in the buyable database will be selected.

In the expansion step, the behavior differs across the three methods. In FHSync, both organic and enzymatic reaction models are applied to the selected molecule. RXN4Chemistry²⁶ and Levin *et al.*'s enzymatic templates¹⁴ are used to predict single-step precursors for the selected molecule. The precursors generated from both reaction types are merged, and all precursors which are not in the buyable database are scored based on the molecular complexity function (denoted as $f(P)$) and the depth with a depth exploration factor (denoted as d). In SPSync, the algorithm uses the SPScore to determine which reaction type is more promising for the selected molecule. Only the predicted reaction type is used for precursor generation, and scoring is performed in the same way as FHSync.

In ACERetro, the SPScore is used to guide an asynchronous search between the two reaction types. For each selected molecule, scores are calculated for both the organic and enzymatic pathways using the formula:



$$\text{Score}_i = (1 - c \cdot \text{SPScore}_i) \text{depth}^d \cdot f(P) \quad i \in [\text{Chem, Bio}]$$

where c is reaction type exploration factor. A molecule will have two scores associated with two reaction types. This approach allows ACERetro to flexibly favour the more promising catalytic route based on the SPScore, while still retaining the ability to switch paths if needed.

In the update step, all newly generated precursors, along with their associated scores, are added to the priority queue, which is then re-ranked before the next iteration begins. This shared architecture enables a fair comparison on how the integration of the SPScore and asynchronous search affects planning performance.

The maximum search depth and the expansion time were 10 and 180 s respectively. For a fair comparison, the above parameters together with commercially available compound database from the vendors eMolecules and Sigma-Aldrich are consistent with those used in Levin *et al.*'s tool (additional parameters in the SI). When the search reaches the time limit, all synthesis routes from buyable molecules to the target molecule are returned.

Case studies on synthesis planning

In the synthesis planning of (*S,S*)-ethambutol and Epidiolex, ACERetro is used to search synthesis routes with a maximum search depth set to 5 and 4, respectively. Because the buyable compound database does not contain complete geometric isomerism information of molecules, when searching in the buyable database, geometric isomerism of molecules is ignored, and optical isomerism is retained. All other parameters of ACERetro are the same as those used in the benchmarking tools. After all pathways and precursors are predicted, the enzyme used in each biocatalytic reaction is selected based on the similarity of products under the same reaction template.

Case studies on synthesis route optimization

For a given synthesis route, the SPScore is calculated for all intermediate molecules (*i.e.*, all except the starting material). To identify steps with potential for optimization, we compare the reaction type used in the current route with the reaction type preferred by the SPScore for each molecule. The top- n molecules with the highest potential for improvement are selected using the following expression:

$$I_n = \text{argsort}(y_i(\text{SPScore}_{\text{Chem}}^i - \text{SPScore}_{\text{Bio}}^i))$$

Here, i is the index of each molecule in the pathway, and n is the number of molecules we wish to identify as most optimizable. The term $y_i = -1$ if the molecule's reaction type in the given synthesis route is an organic reaction (labeled as "Chem"), and $y_i = 1$ if the molecule's reaction type in the given synthesis route is an enzymatic reaction ("Bio"). The equation aims to find top- n molecules with the largest SPScore difference away from the molecule's reaction type ("optimization score", $y_i(\text{SPScore}_{\text{Chem}}^i - \text{SPScore}_{\text{Bio}}^i)$). ACERetro is used to search the synthesis route of steps with opportunities for improvement. The search depth

to a molecule is set to the number of steps from the starting molecules to this molecule in the original synthesis route. For molecule **44** and **46**, the search depth is set as 1. The search depth to molecule **48** is set as 2. All other parameters of ACERetro are the same as those used in the case studies for synthesis planning.

Author contributions

Conceptualization: XL and HZ. Methodology: XL. Investigation: XL and HL. Supervision: HZ. Writing—original draft: XL. Writing—review & editing: XL, HL, and HZ.

Conflicts of interest

There are no conflicts to declare.

Data availability

The scripts for training scoring function, benchmarking, and synthesis planning in this manuscript are available at <https://github.com/Zhao-Group/ACERetro>. The corresponding data and codes have been archived on Zenodo at <https://doi.org/10.5281/zenodo.10578664>. RXN4Chemistry²⁶ is used for single-step precursor prediction of organic reactions, which is the intellectual property of IBM and is also accessible through IBM RXN for Chemistry website. Levin *et al.*'s enzymatic templates¹⁴ are used for single-step precursor prediction of biocatalytic reactions.

Supplementary information is available with training parameters and results, benchmark result analysis, synthesis route examples, and searching parameters. See DOI: <https://doi.org/10.1039/d5dd00008d>.

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Notes and references

- 1 C. Zhang and A. A. Lapkin, Reinforcement learning optimization of reaction routes on the basis of large, hybrid organic chemistry-synthetic biological, reaction network data, *React. Chem. Eng.*, 2023, **8**(10), 2491–2504.
- 2 S. Simić, E. Zukić, L. Schermund, K. Faber, C. K. Winkler and W. Kroutil, Shortening synthetic routes to small molecule active pharmaceutical ingredients employing biocatalytic methods, *Chem. Rev.*, 2022, **122**(1), 1052–1126.



- 3 F. Kaspar and A. Schallmey, Chemo-enzymatic synthesis of natural products and their analogs, *Curr. Opin. Biotechnol.*, 2022, **77**, 102759.
- 4 S. Chakrabarty, E. O. Romero, J. B. Pyser, J. A. Yazarians and A. R. H. Narayan, Chemoenzymatic total synthesis of natural products, *Acc. Chem. Res.*, 2021, **54**(6), 1374–1384.
- 5 J. Li, A. Amatuni and H. Renata, Recent advances in the chemoenzymatic synthesis of bioactive natural products, *Curr. Opin. Chem. Biol.*, 2020, **55**, 111–118.
- 6 F. Rudroff, M. D. Mihovilovic, H. Gröger, R. Snajdrova, H. Iding and U. T. Bornscheuer, Opportunities and challenges for combining chemo- and biocatalysis, *Nat. Catal.*, 2018, **1**(1), 12–22.
- 7 J. A. McIntosh, T. Benkovics, S. M. Silverman, M. A. Huffman, J. Kong, P. E. Maligres, *et al.*, Engineered ribosyl-1-kinase enables concise synthesis of molnupiravir, an antiviral for COVID-19, *ACS Cent. Sci.*, 2021, **7**(12), 1980–1985.
- 8 S. Szymkuć, E. P. Gajewska, T. Klucznik, K. Molga, P. Dittwald, M. Startek, *et al.*, Computer-assisted synthetic planning: the end of the beginning, *Angew. Chem., Int. Ed.*, 2016, **55**(20), 5904–5937.
- 9 W. Finnigan, L. J. Hepworth, S. L. Flitsch and N. J. Turner, RetroBioCat as a computer-aided synthesis planning tool for biocatalytic reactions and cascades, *Nat. Catal.*, 2021, **4**(2), 98–104.
- 10 X. Wang, Y. Li, J. Qiu, G. Chen, H. Liu, B. Liao, *et al.*, RetroPrime: a diverse, plausible and transformer-based method for single-step retrosynthesis predictions, *Chem. Eng. J.*, 2021, **420**, 129845.
- 11 U. V. Ucak, I. Ashyrmamatov, J. Ko and J. Lee, Retrosynthetic reaction pathway prediction through neural machine translation of atomic environments, *Nat. Commun.*, 2022, **13**(1), 1186, available from: <https://chemrxiv.org/engage/chemrxiv/article-details/6117ed347117505183e94d93>.
- 12 B. Chen, C. Li, H. Dai and L. Song, Retro*: Learning Retrosynthetic Planning with Neural Guided A* Search, in *Proceedings of the 37th International Conference on Machine Learning*, PMLR, 2020, pp. 1608–1616, available from: <https://proceedings.mlr.press/v119/chen20k.html>.
- 13 S. Hong, H. H. Zhuo, K. Jin, G. Shao and Z. Zhou, Retrosynthetic planning with experience-guided Monte Carlo tree search, *Commun. Chem.*, 2023, **6**(1), 1–14.
- 14 I. Levin, M. Liu, C. A. Voigt and C. W. Coley, Merging enzymatic and synthetic chemistry with computational synthesis planning, *Nat. Commun.*, 2022, **13**(1), 7747.
- 15 T. Zeng, Z. Jin, S. Zheng, T. Yu and R. Wu, Developing BioNavi for hybrid retrosynthesis planning, *JACS Au*, 2024, **4**(7), 2492–2502.
- 16 K. Sankaranarayanan and K. F. Jensen, Computer-assisted multistep chemoenzymatic retrosynthesis using a chemical synthesis planner, *Chem. Sci.*, 2023, **14**(23), 6467–6475.
- 17 H. Li, X. Liu, G. Jiang and H. Zhao, Chemoenzymatic Synthesis Planning Guided by Reaction Type Score, *J. Chem. Inf. Model.*, 2024, **64**(24), 9240–9248.
- 18 C. W. Coley, L. Rogers, W. H. Green and K. F. Jensen, SCscore: synthetic complexity learned from a reaction corpus, *J. Chem. Inf. Model.*, 2018, **58**(2), 252–261.
- 19 P. Schwaller, R. Petraglia, V. Zullo, V. H. Nair, R. A. Haeuselmann, R. Pisoni, *et al.*, Predicting retrosynthetic pathways using transformer-based models and a hyper-graph exploration strategy, *Chem. Sci.*, 2020, **11**(12), 3316–3325.
- 20 C. W. Coley, W. Jin, L. Rogers, T. F. Jamison, T. S. Jaakkola, W. H. Green, *et al.*, A graph-convolutional neural network model for the prediction of chemical reactivity, *Chem. Sci.*, 2019, **10**(2), 370–377.
- 21 D. Probst, M. Manica, Y. G. Nana Teukam, A. Castrogiovanni, F. Paratore and T. Laino, Biocatalysed synthesis planning using data-driven learning, *Nat. Commun.*, 2022, **13**(1), 964.
- 22 D. Rogers and M. Hahn, Extended-connectivity fingerprints, *J. Chem. Inf. Model.*, 2010, **50**(5), 742–754.
- 23 A. Capecchi, D. Probst and J. L. Reymond, One molecular fingerprint to rule them all: drugs, biomolecules, and the metabolome, *J. Cheminf.*, 2020, **12**(1), 43.
- 24 A. Thakkar, T. Kogej, J. L. Reymond, O. Engkvist and E. J. Bjerrum, Datasets and their influence on the development of computer assisted synthesis planning tools in the pharmaceutical domain, *Chem. Sci.*, 2019, **11**(1), 154–168.
- 25 T. Sterling and J. J. Irwin, ZINC 15 – ligand discovery for everyone, *J. Chem. Inf. Model.*, 2015, **55**(11), 2324–2337.
- 26 A. Toniato, A. C. Vaucher, P. Schwaller and T. Laino, Enhancing diversity in language based models for single-step retrosynthesis, *Digital Discovery*, 2023, **2**(2), 489–501.
- 27 Y. Mo, Y. Guan, P. Verma, J. Guo, M. E. Fortunato, Z. Lu, *et al.*, Evaluating and clustering retrosynthesis pathways with learned strategy, *Chem. Sci.*, 2021, **12**(4), 1469–1478.
- 28 P. Cornwall, L. J. Diorazio and N. Monks, Route design, the foundation of successful chemical development, *Bioorg. Med. Chem.*, 2018, **26**(14), 4336–4347.
- 29 R. G. Wilkinson, R. G. Shepherd, J. P. Thomas and C. Baughn, Stereospecificity in a new type of synthetic antituberculous agent, *J. Am. Chem. Soc.*, 1961, **83**(9), 2212–2213.
- 30 R. G. Shepherd and R. G. Wilkinson, Antituberculous agents. II. N,N'-diisopropylethylenediamine and analogs, *J. Med. Pharm. Chem.*, 1962, **5**(4), 823–835.
- 31 R. G. Wilkinson, M. B. Cantrall and R. G. Shepherd, Antituberculous agents. III. (+)-2,2 -(ethylenediamino)-di-1-butanol and some analogs, *J. Med. Pharm. Chem.*, 1962, **5**(4), 835–845.
- 32 I. Butula and G. Karlović, Katalytische Hydrierung von stickstoffhaltigen heterocyclen, IV1) Hydrogenolyse von verbrückten oxazolinen und oxazolidinen, *Justus Liebigs Ann. Chem.*, 1976, **1976**(7–8), 1455–1464.
- 33 C. S. Stauffer and A. Datta, Efficient synthesis of (S,S)-ethambutol from L-methionine, *Tetrahedron*, 2002, **58**(49), 9765–9767.
- 34 B. M. Trost, R. C. Bunt, R. C. Lemoine and T. L. Calkins, Dynamic kinetic asymmetric transformation of diene monoepoxides: a practical asymmetric synthesis of vinylglycinol, vigabatrin, and ethambutol, *J. Am. Chem. Soc.*, 2000, **122**(25), 5968–5976.

35 S. P. Kotkar and A. Sudalai, Enantioselective synthesis of (S,S)-ethambutol using proline-catalyzed asymmetric α -aminoxylation and α -amination, *Tetrahedron:Asymmetry*, 2006, **17**(11), 1738–1742.

36 Y. Kobayashi, A. Takeuchi and Y. G. Wang, Synthesis of cannabinoids via alkenylation of cyclohexenyl monoacetate, *Org. Lett.*, 2006, **8**(13), 2699–2702.

37 Z. P. Shultz, G. A. Lawrence, J. M. Jacobson, E. J. Cruz and J. W. Leahy, Enantioselective total synthesis of cannabinoids - a route for analogue development, *Org. Lett.*, 2018, **20**(2), 381–384.

38 X. Gong, C. Sun, M. A. Abame, W. Shi, Y. Xie, W. Xu, *et al.*, Synthesis of CBD and its derivatives bearing various C4'-side chains with a late-stage diversification method, *J. Org. Chem.*, 2020, **85**(4), 2704–2715.

39 J. M. Stout, Z. Boubakir, S. J. Ambrose, R. W. Purves and J. E. Page, The hexanoyl-CoA precursor for cannabinoid biosynthesis is formed by an acyl-activating enzyme in *Cannabis sativa* trichomes, *Plant J.*, 2012, **71**(3), 353–365.

40 P. Seccamani, C. Franco, S. Prott, A. Porta, A. Profumo, D. Caprioglio, *et al.*, Photochemistry of cannabidiol (CBD) revised. A combined preparative and spectrometric investigation, *J. Nat. Prod.*, 2021, **84**(11), 2858–2865.

41 P. C. Yan, G. L. Zhu, J. H. Xie, X. D. Zhang, Q. L. Zhou, Y. Q. Li, *et al.*, Industrial scale-up of enantioselective hydrogenation for the asymmetric synthesis of rivastigmine, *Org. Process Res. Dev.*, 2013, **17**(2), 307–312.

42 M. K. Sethi, S. R. Bhandya, N. Maddur, R. Shukla, A. Kumar and V. S. N. Jayalakshmi Mittapalli, Asymmetric synthesis of an enantiomerically pure rivastigmine intermediate using ketoreductase, *Tetrahedron:Asymmetry*, 2013, **24**(7), 374–379.

43 H. Agarwala, F. Ehret, A. D. Chowdhury, S. Maji, S. M. Mobin, W. Kaim, *et al.*, Electronic structure and catalytic aspects of [Ru(tpm)(bqdi)(Cl/H₂O)]_n, tpm = tris(1-pyrazolyl)methane and bqdi = o-benzoquinonediimine, *Dalton Trans.*, 2013, **42**(10), 3721–3734.

44 B. Laroche, H. Ishitani and S. Kobayashi, Direct reductive amination of carbonyl compounds with H₂ using heterogeneous catalysts in continuous flow as an alternative to N-alkylation with alkyl halides, *Adv. Synth. Catal.*, 2018, **360**(24), 4699–4704.

45 R. Alcántara, K. B. Axelsen, A. Morgat, E. Belda, E. Coudert, A. Bridge, *et al.*, Rhea—a manually curated resource of biochemical reactions, *Nucleic Acids Res.*, 2012, **40**(Database issue), D754–D760.

46 I. Schomburg, A. Chang and D. Schomburg, BRENDA, enzyme data and metabolic information, *Nucleic Acids Res.*, 2002, **30**(1), 47–49.

47 D. S. Wishart, C. Li, A. Marcu, H. Badran, A. Pon, Z. Budinski, *et al.*, PathBank: a comprehensive pathway database for model organisms, *Nucleic Acids Res.*, 2020, **48**(D1), D470–D478.

48 M. Ganter, T. Bernard, S. Moretti, J. Stelling and M. Pagni, MetaNetX.org: a website and repository for accessing, analysing and manipulating metabolic networks, *Bioinformatics*, 2013, **29**(6), 815–816.

