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Internal vibrational energy redistribution precedes energy dissipation into the solvent upon photoexcitation of heme proteins†

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Efficient energy dissipation is a fundamental property of proteins that has been challenging to observe. This study investigates the mechanism of energy flow from excited heme proteins into the surrounding solvent using optical-pump THz–probe spectroscopy. The results were compared with a computational model of the energy flow. We find that the experimentally observed time constants of 6.1 ± 0.3 ps and 7.3 ± 0.4 ps for cytochrome c and myoglobin are in excellent agreement with the computational model. This suggests a diffusive energy transfer for the release of excitation energy from the excited heme group through the protein as the dominant pathway of energy flow to the surrounding solvent in heme proteins. We also observe a notable isotope effect suggesting a complex coupling of the protein to the solvent.

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In nature, proteins require the ability to effectively dissipate energy. This is needed to avoid radiation damage in photosynthesis and allow communication between allosteric sites through directed energy transfer.¹ The capability of proteins to conduct energy efficiently is more fundamentally linked to the nature of proteins as highly flexible structures that continuously change conformation on timescales from picoseconds to microseconds.² This conformational flexibility naturally enables numerous energy dissipation pathways after photoexcitation. These dynamics can be selectively induced in heme proteins, by dissociating bound carbon monoxide, in a process denoted ‘protein quake’. This widely investigated phenomenon has been the focus of scientific discussion for many years.^{3–9} Despite extensive efforts, the molecular mechanism of the internal energy redistribution and the coupling to the solvent remains elusive.¹⁰ Ansari *et al.* postulated that the energy relaxation in the protein takes place in a quake-like manner *via* a hierarchical glass-like structure.¹¹ Due to the highly dynamic nature of proteins and their strong coupling to the solvent observations of the energy flow have been challenging and vary depending on the observation technique. Observing

the Soret band directly in transient absorption measurements, the Champion group was able to observe the oscillations in the intensity of the Soret band associated with a heme deformation called ‘doming’ when dissociating NO, lasting around 1 ps after excitation.⁹ In a follow-up study, they observed a 300 fs internal conversion after excitation of the heme group followed by a 1 ps relaxation time of the heme for myoglobin with bound oxygen.³

The heme group has been shown to be strongly dependent on the protein folding showing significant changes in UV-Vis absorption¹² as well as changes in its ability to transfer electrons.¹³

Mizutani and Kitagawa studied the anti-Stokes intensity of the ν_4 band in the heme group in myoglobin reporting a biphasic decay of the excited populations. The best fit to double exponentials yielded time constants of 1.9 ± 0.6 and 16 ± 9 ps for vibrational population decay in the heme group and 3.0 ± 1.0 and 25 ± 14 ps for the cooling of the photolyzed heme *via* energy flow to the water bath.⁶ In a study by the cooling of heme subsequent to photoexcitation of the carbon-monoxo myoglobin, heme was characterized by probing the spectral evolution of a porphyrin-iron charge-transfer transition near $13\,110\text{ cm}^{-1}$ (763 nm) that is sensitive to temperature. They deduced a single time constant of 6.2 ± 0.5 ps for the exponential cooling of the electronically relaxed heme.

On the structural dynamics side, advancements in X-ray free electron lasers (X-FEL) made it possible to ‘film’ molecular movies of ultrafast protein dynamics, and capture the protein quake in action.¹⁴ Time-resolved femtosecond crystallography, complemented by QM/MM simulations, indicated a collective response of the protein within 500 fs of photolysis of Mb that

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originates from the coupling of heme vibrations to global protein modes. Similarly, time-resolved solution phase small-angle (SAXS) and wide-angle X-ray scattering (WAXS) showed an increase in the radius of gyration (R_g) of Mb within 1 ps, followed by damped oscillations with a period of ~ 3.6 ps. The oscillations were later explained from the results of molecular dynamics (MD) simulations as arising from fluctuations in the density of water surrounding the protein¹⁵

Hochstrasser *et al.* observed the solvent directly, probing the bend-libration combination band of D_2O at 1800 cm^{-1} to study the heating of the solvent after photoexcitation at 580 nm. They observed a signal they attributed to a temperature change within 4 ps.¹⁶ The kinetics could be fitted to a model that consists of a fast and a slow component. The slow component (~ 20 ps), with 40% of the total amplitude, is attributed to energy transfer from heme to water through the protein *via* a classical diffusion process. The fast component with a time constant of 7.5 ± 2 and 8.5 ± 1.5 ps for Mb and Hb, was too fast to be described by classical diffusion and was assumed to be proceeding through collective motions of the protein.

Chergui *et al.* utilized transient absorption spectroscopy to observe the spectral region of the Soret band in cytochrome *c* and were able to pinpoint both the heme relaxation at 1.8 ± 0.5 ps and the relaxation of cytochrome *c* at 11 ± 0.5 ps for the oxidized protein.⁸ They were not able to resolve the relaxation time for the ferrous cytochrome *c* used in this study.

The open questions are: What are the modes responsible for the fast energy transfer in proteins? Is the energy flow governed by ballistic energy transfer or can it be described as diffusive? What are the time scales for energy dissipation into the solvent? Straub and co-workers investigated the molecular mechanisms of the transfer of excess heme energy to the solvents using MD simulations and identified three distinct relaxation pathways, which can contribute to the relaxation completing in ~ 5 ps: (1) “through projectile” due to interactions between the dissociated ligand and protein sidechains, (2) “through space” due to interactions between the heme and sidechains in the heme pocket, and (3) “through bond”, with the bond between the heme iron and the proximal histidine (His93). Based on a comparison of selected mutants evidence was found that the predominant channel for fast kinetic energy relaxation of the heme in native myoglobin is directed energy “funneling” through the heme side *via* electrostatic interaction of the propionate side chains to the surrounding solvent.¹⁷

In the present study, we want to focus on the energy flow of the excited heme into the solvent *via* a coupling of the protein dynamics to the solvent dynamics.^{18,19} Rapid transfer of energy requires an efficient coupling of the collective modes of proteins and solvent, from where it is dissipated by thermal diffusion or as a solvent quake.^{16,20–22} This energy transfer into the solvent should be correlated to the vibrational relaxation of the protein.¹

Using our sensitive optical pump THz probe (OPTP) set-up in Bochum, we report time-resolved THz measurements monitoring the energy flow into the low-frequency water modes subsequent to photoexcitation of heme, allowing us to observe the energy flow in a less congested spectral region. Upon

photoexcitation by a 400 nm laser pulse, a large amount of excess energy is selectively deposited in the heme group due to rapid internal conversion. This excess energy is redistributed to the surrounding protein moiety *via* intramolecular vibrational relaxation (IVR) and then to the water, where it can be probed selectively by optical pump THz probe measurements. We compare the results for myoglobin and cytochrome *c* with the predicted time constants and find an excellent agreement. We conclude that subsequent to photoexcitation, cooling first takes place *via* internal energy redistribution in the protein. Subsequent to this, the energy is transferred into the solvent primary diffusively, limited only by the thermal diffusivity of the protein matrix.

Methods

Our optical pump THz probe spectrometer is described in detail in ref. 23; briefly: 50 fs, 800 nm laser pulses were produced by a Ti:Sa amplified laser, which generated broad THz pulses *via* a two-color plasma filament.²⁴ The optical pump beam was frequency-doubled to 400 nm in a BBO crystal and attenuated to a pump fluence of 13 mJ cm^{-2} on a spot size slightly exceeding that of the THz, which had a diameter of $400\text{ }\mu\text{m}$ ($1/e^2$). The THz absorption was recorded as a function of pump–probe delay Δt between 0.25 ps and 300 ps. Under these conditions typical window materials with good THz transmission are prone to free carrier excitation, therefore we used a thin water jet instead.^{23,25–27} The thickness of the jet was determined to be $28\text{ }\mu\text{m}$ for cytochrome *c* and myoglobin in water and $25\text{ }\mu\text{m}$ for myoglobin in D_2O . For cytochrome *c* in D_2O , we used a nozzle with a thickness of $13\text{ }\mu\text{m}$ for the deuterated water jet. The myoglobin (equine) was obtained from Sigma Aldrich, the cytochrome *c* (equine) was purchased from Gerbu Biotechnik, both were used without further purification. The protein solutions (70 ml , 20.0 g L^{-1}) were prepared in potassium phosphate buffer (pH: 7.3, molarity: 0.05 M) and filtered with syringe filters of $20\text{ }\mu\text{m}$ pore size. To exclude protein degradation an SDS PAGE was performed, and CD spectra were measured before and after the measurement. The sample was circulated through the jet for 72 hours while OPTP spectra were recorded. To mitigate foam formation, defoamer BYK 019 was added to the sample reservoir, which was cooled just above $0\text{ }^\circ\text{C}$. The THz field is detected by electro-optic sampling (EOS) using a $100\text{ }\mu\text{m}$ -thick GaP crystal. The recorded field traces are analyzed by Fourier transformation, we plot the difference between optical pump off–optical pump on as ΔmOD (milli Optical Density) values. These values indicate the transmission change induced by optically pumping the sample, where a positive ΔmOD corresponds to reduced THz transmission at that frequency. Thus, all reported signals result from optical excitation. The reported errors are the standard error of 50 consecutive measurements.

Experimental results

Heme proteins are ideal molecules for studying the molecular mechanism of the energy flow in proteins. Vibrational excess energy as large as $25\text{ }000\text{ cm}^{-1}$ is deposited on the heme moiety



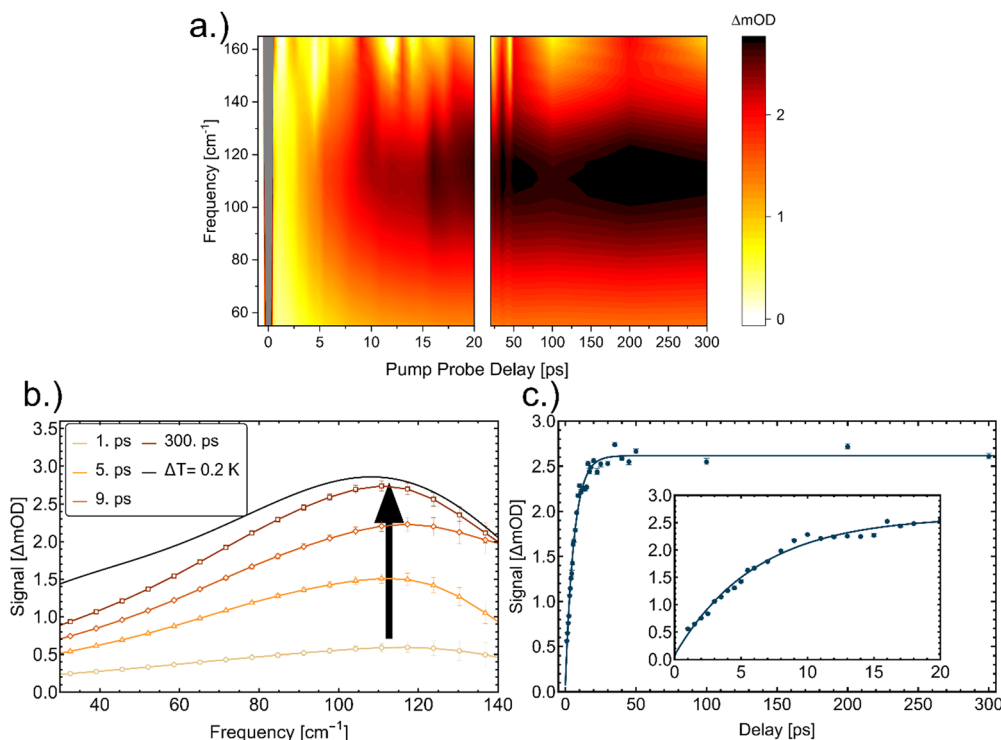


Fig. 1 (a) OPTP measurements of an aqueous cytochrome *c* solution as a function of time delay. (b) Transient THz spectra (vertical slices of the map) at representative pump–probe time delays (1, 5, 9, and 300 ps). The arrow serves as a guide to the eye. For comparison we plot in black the difference in absorption for bulk water upon a temperature change of 0.2 K. (c) Plot of the fitted maximum amplitude when fitting a Gaussian to the OPTP spectrum (ΔmOD) at a given time delay. The result of the fit (see eqn (1)) is shown as solid line, yielding a decay time of 6.09 ± 0.30 ps. The inset shows a zoom in for time delays between 1 ps and 20 ps.

immediately after photoexcitation with an optical pulse at 400 nm, resulting from extremely fast internal conversion in less than 100 fs *via* the Soret transition.¹²

In Fig. 1a we plot transient THz spectra after excitation of a cytochrome *c* solution as a function of time. Within the first 0.5 ps we expect an overlap of the optical pump and THz probe, therefore we cannot take this into account for further analysis.²⁸ This artifact signal is a convoluted response of the pump and probe beam related to a Raman response of the sample²⁹ that decays within 0.25 ps and is observed for all investigated samples.

The first transient spectra at 1 ps show a maximum centered around 120 cm^{-1} , then shifting to 110 cm^{-1} which increases between 1–20 ps, see Fig. 1c). For comparison, we also plot the change in absorption of bulk water when increasing the temperature by 0.2 K, see Fig. 1b.^{30–32} The timescales of the experiment do not allow for significant protein diffusion, thus we do not expect aggregation. For further analysis, we fitted the signal with a Gaussian function and plotted the amplitude of the Gaussian as a function of time. The amplitude increases until it reaches an equilibrium at around 30 ps, see Fig. 1c. For the fit of the data, we used eqn (1), with C_{offset} being the equilibrium value, τ the time constant, and A_1 the maximum amplitude. The result of the fit is shown in Fig. 1c.

$$A(t_{\text{pp}}) = C_{\text{offset}} - A_1 e^{-\frac{t_{\text{pp}}}{\tau}} \quad (1)$$

The deduced time constants are summarized in Table 1. We have repeated the measurements for two different proteins, cytochrome *c*

Table 1 Results of the fitted rise times for cytochrome *c* and myoglobin in H_2O and D_2O

Sample	Experimental time constant τ_{exp} [ps]	Predicted time constant τ_{calc} [ps]
Cytochrome <i>c</i>	6.1 ± 0.3	5.8
Cytochrome <i>c</i> in D_2O	7.1 ± 0.3	6.1
Myoglobin	7.3 ± 0.4	7.7
Myoglobin in D_2O	10.3 ± 0.7	8.2

and myoglobin as well as two solvents: H_2O and D_2O . For cytochrome *c* we obtain $\tau = 6.1 \pm 0.3$ ps and $\tau = 7.1 \pm 0.3$ ps in H_2O and D_2O , respectively. The results for the OPTP measurements of myoglobin are shown in Fig. 3. The fitted time constants are 7.3 ± 0.4 ps and 10.3 ± 0.7 ps in H_2O and D_2O , respectively. These time scales exceed the fast time scale of 2–3 ps observed for HPTS.²⁰

In order to avoid reflection at windows we use a liquid jet. The absolute amplitudes of the signals will depend on the thickness of the liquid jet which in turn depends on the viscosity of the sample and the specific nozzle (see ESI,[†] Table S1, and Fig. 2b), however, the rise times remain unaffected (ESI,[†] Fig. S1).

Experimentally we find that exchanging the solvent from H_2O to D_2O increases the time constant τ for both proteins, by a factor of 1.16 ± 0.06 in cytochrome *c* and by a factor of 1.41 ± 0.08 in myoglobin.

The different time scales for energy dissipation into the solvent are visualized in Fig. 4. We plotted the normalized



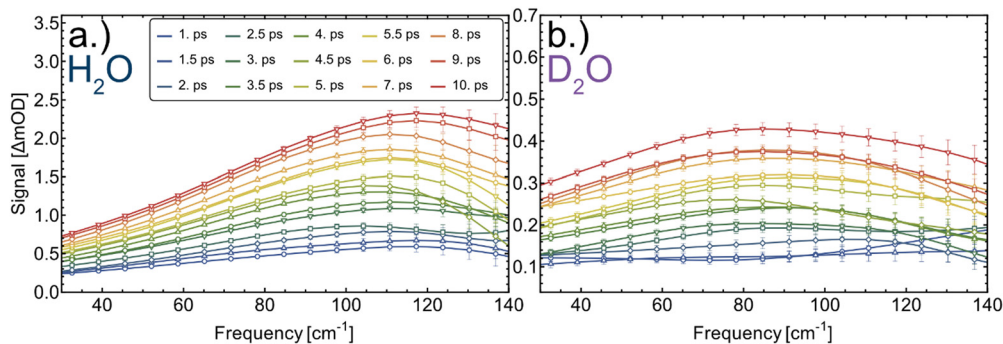


Fig. 2 Transient OPTP spectra at representative pump–probe time delays of 1–10 ps of cytochrome *c* in H₂O (a) and D₂O (b). The amplitude for cytochrome *c* in D₂O is smaller because it was measured in a thinner jet, decreasing the signal intensity.

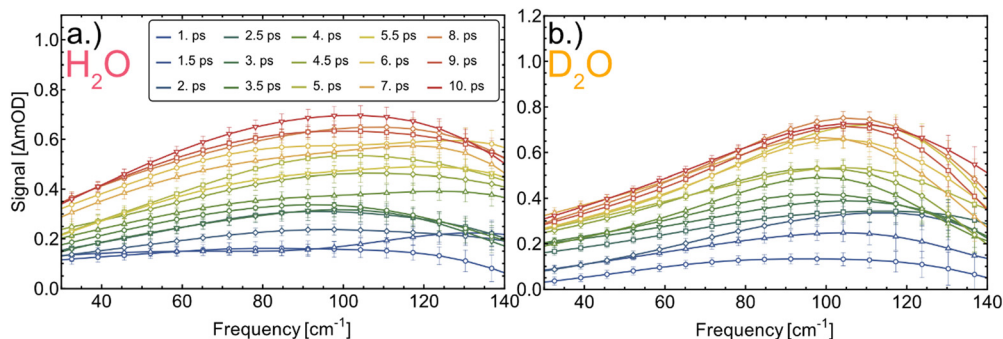


Fig. 3 Transient THz difference spectra at representative pump–probe time delays of 1–10 ps of myoglobin in H₂O (a) and D₂O (b).

difference to the equilibrium value on a logarithmic scale as a function of time. The expected exponential decrease, see eqn (2) should yield a linear line, with the slope being $-1/\tau$, see Fig. 4.

$$\frac{C_{\text{offset}} - A(t_{\text{pp}})}{A_1} = e^{-\frac{t_{\text{pp}}}{\tau}} \quad (2)$$

Modeling results

Thermal energy in a protein has been suggested to flow primarily diffusively in experimental²¹ and computational studies.^{1,33} To model the flow of thermal energy from the heme to the surrounding water we assume, for simplicity, a spherical protein (Fig. 5(a)) where the heat source lies at the center, a picture that has been proposed in the past.³⁴ We solve a diffusion equation for the temperature, $T(r,t)$, where r is the distance from the center and t is time, with heat source $g(r,t)$,

$$\frac{\partial T(r,t)}{\partial t} = D(r) \frac{1}{r} \frac{\partial^2}{\partial r^2} (rT(r,t)) + g(r,t). \quad (3)$$

$D(r)$ is the thermal diffusivity. For the heat source, we use the form $g(r,t) = \left(\frac{\Delta T}{\tau_{\text{H}}}\right) e^{-\frac{r^2}{2\sigma^2}} e^{-\frac{t}{\tau_{\text{H}}}}$. Similar models have been studied to examine photothermal effects on time-resolved IR spectroscopy following photoexcitation of a solute,³⁵ and thermal energy has been seen in computational studies to flow

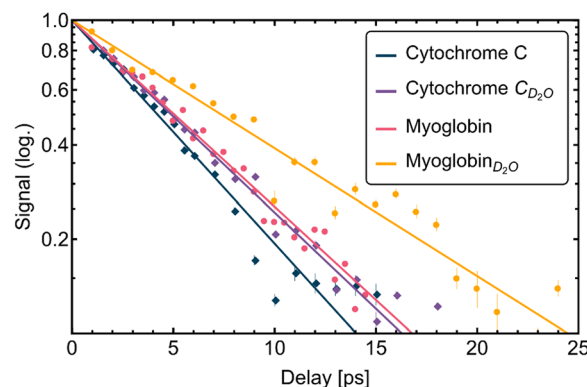


Fig. 4 Plot of the transient amplitude signals for all samples (cytochrome *c*, myoglobin in H₂O and D₂O) and the fits obtained *via* eqn (1). Data and fit were linearized with eqn (2) and the semi-logarithmic scale of the plot.

diffusively.³⁶ We take the initial temperature of the system to be 288 K, consistent with the temperature at which the experiments were done, and the increase in temperature of the heme due to photoexcitation, ΔT , to be 400 K, which corresponds to the rise in temperature of the heme due to optical excitation.^{37–39} The heating time of the surrounding solvent does not depend on the initial temperature or on ΔT , which was checked in the numerical analysis. The time constant for the decay of excess thermal energy from the heme into the protein, τ_{H} , is set to 1 ps,



and σ is taken to be an effective radius for the heating source (*i.e.*, the heme) and is set to 3.5 Å. The values for τ_{H} and σ are reasonable and modest variation does not significantly change the time constants for heating of water surrounding the protein.

The thermal diffusivity for myoglobin has been calculated to be $14.0 \text{ \AA}^2 \text{ ps}^{-1}$,¹² and we use this value for both myoglobin and cytochrome *c*. The thermal diffusivity for H₂O near room temperature and atmospheric pressure is $14.6 \text{ \AA}^2 \text{ ps}^{-1}$ and for D₂O it is $12.7 \text{ \AA}^2 \text{ ps}^{-1}$.⁴⁰ Protein–water thermal boundary conductance, G , has been computed to be of order $10^8 \text{ W m}^{-2} \text{ K}^{-1}$,²² and we use $3 \times 10^8 \text{ W m}^{-2} \text{ K}^{-1}$, consistent with computational results for the myoglobin–water⁴¹ and cytochrome *c*–water⁴² interface. The radius of the protein (r_1 in Fig. 5(a)) is taken to be 1.9 nm for myoglobin⁴³ and 1.4 nm for cytochrome *c*.⁴⁴ To account for the boundary conductance we replace $D(r)$ in eqn (3) at the protein–water interface with $\Delta r G/(\rho C_p)$ for a boundary width, Δr , of 4 Å, where C_p and ρ are the protein heat capacity and mass density, respectively. Small variation of Δr does not change the result. In the experiments, the concentration of myoglobin is 20.0 g L^{-1} , which corresponds to a shell of water that reaches $r_2 = 7 \text{ nm}$, where r_2 is the distance from the center of the protein to the edge of the hydration shell (Fig. 5(a)). The concentration of cytochrome *c* is 21.3 g L^{-1} , which corresponds to a shell of water that reaches $r_2 = 6 \text{ nm}$. Eqn (3) is solved numerically by finite difference.

We compute with eqn (3) the average temperature of the solvent surrounding the protein as a function of time. The results are plotted in Fig. 5(b) for myoglobin in H₂O and D₂O and Fig. 5(c) for cytochrome *c* in H₂O and D₂O. For both proteins, we see the rise in temperature to be higher in H₂O than D₂O and somewhat faster, due to the greater thermal diffusivity of H₂O compared to D₂O. To obtain the heating time, τ_{w} , of the water (H₂O or D₂O), we fit the data plotted in Fig. 5(b) and (c) to $T(t) = T_{\infty} + (T(0) - T_{\infty})e^{-t/\tau_{\text{w}}}$, where T_{∞} corresponds to the temperature at long times. For myoglobin, the heating time is obtained from a linear fit of $\ln(T_{\infty} - T(t))$ vs. t between 3 and 14 ps. For cytochrome *c*, with somewhat shorter heating times compared to myoglobin, the heating time is obtained from a linear fit of $\ln(T_{\infty} - T(t))$ vs. t between 2 and 13 ps. The fits are shown in Fig. 5(b) and (c) together with $\ln(T_{\infty} - T(t))$ for myoglobin and cytochrome *c*, respectively. In this way, we find for myoglobin the time constant, τ_{w} , for heating

H₂O to be 7.7 ps and D₂O to be 8.2 ps. For cytochrome *c* we find τ_{w} for heating H₂O to be 5.8 ps and D₂O to be 6.1 ps.

Discussion

For myoglobin in D₂O, we find good agreement between our experimental and predicted results of $10.3 \pm 0.7 \text{ ps}$ and 8.3 ps , respectively. These time constants also agree with the 7.5 ps time constant observed in the combination band of the bend and the libration of D₂O, as reported by the Hochstrasser group.¹⁶ In addition, they reported a slower component ($\sim 20 \text{ ps}$). In our data—within experimental uncertainty—we find no evidence for a slower component. This implies, that the energy transfer into the low-frequency modes of the protein and subsequent relaxation into the translational modes of the surrounding hydration water, (centered around $50\text{--}80 \text{ cm}^{-1}$) is the dominant relaxation pathway. The relaxation into the MIR modes and librational band proceeds slower, on the 20 ps timescale, visible in the experiments of the Hochstrasser group but not visible in our frequency range. This is also consistent with the relaxation process in the hydrogen bond network where the relaxation happens faster into the low-frequency modes than into the bending mode of water.⁴⁵

When myoglobin or cytochrome *c* are excited *via* absorption in the Soret band, the heme group is electronically excited and will undergo intramolecular vibrational relaxation, thereby releasing energy into the protein matrix within 1 ps.³ Here, we report new results on the molecular mechanism for the energy flow into the solvent. We want to note, that relaxation time of the larger myoglobin matrix is increased compared to the smaller cytochrome *c* protein. In the model, we set the time constant for the decay of excess thermal energy from the heme into the protein to 1 ps, and an effective radius for the heating source (*i.e.*, the heme) is set to 3.5 Å. For both proteins, we assume the same thermal diffusivity $D(r)$ with the only difference that the effective radius is assumed to be 1.9 nm for myoglobin⁴³ and 1.4 nm for cytochrome *c*.⁴⁴ For the thermal protein water boundary conductance, we used the previous computational results for the myoglobin–water⁴¹ and cytochrome *c*–water⁴² interface.

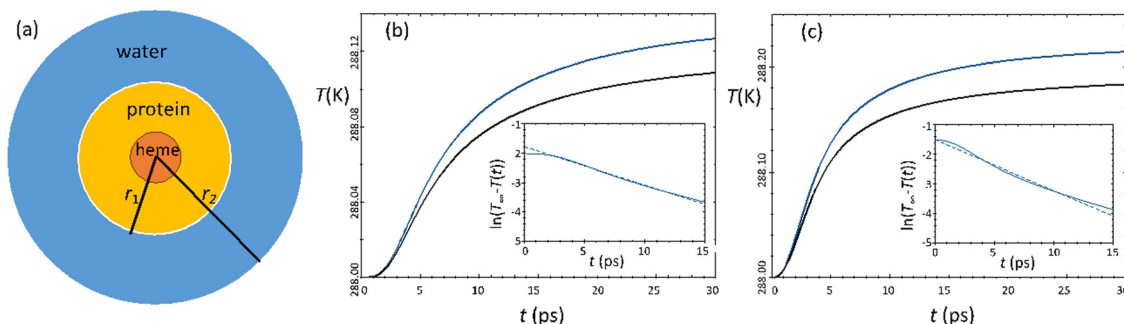


Fig. 5 (a) Schematic model for protein in solvent, with heat source (heme) taken as the center, where r_1 is the radius of the protein and r_2 is the distance from the center of the protein to the edge of the hydration shell. (b) Temperature computed for H₂O (blue) and D₂O (black) surrounding myoglobin; fitting to obtain the heating time, τ_{w} , of the solvent (see text for details) is shown in inset for H₂O (D₂O is similar), where T_{∞} is the temperature at long times. (c) Temperature computed for H₂O (blue) and D₂O (black) surrounding cytochrome *c*; fitting to obtain τ_{w} is shown in inset for H₂O (D₂O is similar).



We want to note that the predicted time scale of 5.8 ps for cytochrome *c* and 7.7 ps for myoglobin are in excellent agreement with the measured time scales of 6.1 ± 0.3 ps and 7.3 ± 0.4 ps, respectively. This confirms that the energy flow from the hot heme in the protein matrix is mostly diffusive and not directional or protein-specific.

We now want to come back to the previous assumption that fast kinetic energy relaxation of the heme in native myoglobin is directed energy “funneling” through the heme side *via* electrostatic interaction of the propionate side chains into the surrounding solvent.^{4,17}

In cytochrome *c* the propionate side chains are less exposed to the solvent as in the case of myoglobin, however, the measured relaxation time is shorter. As a consequence, our measurements suggest that the energy transfer time scales are determined by the intramolecular energy redistribution into the protein matrix, for both myoglobin and cytochrome *c*. We find indications for a difference in the relaxation mechanism between cytochrome *c* and myoglobin: when substituting the solvent from H₂O to D₂O, we observe a more pronounced isotope effect for myoglobin compared to cytochrome *c*. Thus, for cytochrome *c* the intramolecular vibrational relaxation is decisive, while in the case of myoglobin, with a larger solvent-exposed surface area, the protein–water conductance appears to play a larger role: The relaxation time constant is measured to increase from 7.3 ± 0.4 to 10.3 ± 0.7 ps in the case of myoglobin, and such a large increase in time is not reproduced by our simplified model. The model predicts a longer time constant for myoglobin in D₂O compared to H₂O, in qualitative agreement with the experiment, but the time constant is only about 15% larger for myoglobin in D₂O, whereas the measured value is about 40% larger. However, the simplified model does not take into account the molecular details of the coupling. This might depend on the relative energy match between the low frequency modes of the propionate groups *versus* those of D₂O/H₂O, thus affecting the time for energy transfer. We note that folding is not affected by deuteration, thus the protein folding dynamics remains unchanged when replacing H₂O with D₂O.⁴⁶

In summary, OPTP is able to sensitively probe the relaxation of the internal energy from the protein into the solvent and is complementary to more local intermolecular probes like Raman and IR. Combining OPTP measurements with theoretical modeling we can uncover the energy relaxation pathway in heme proteins: We can show that the energy relaxation in heme protein takes place *via* a hierarchical energy flow. While the first step occurs in about 1 ps from the hot heme into the protein matrix, the second step can be described by the thermal diffusivity of the protein matrix to the protein surface that is completed within 15 ps. The coupling to the solvent taken into account *via* the protein–water thermal boundary conductance seems to be more protein-specific. We propose that the difference in boundary conductance of the propionate side chains between H₂O and D₂O is responsible for the increase in the relaxation time by a factor of 1.41 ± 0.08 in myoglobin where the side chains are exposed, whereas the impact is smaller (factor of 1.16 ± 0.06) for cytochrome *c* where the side chains are not as exposed.

Data availability

All the data generated in this study has been deposited in our open-access repository: <https://doi.org/10.17877/RESOLV-2025-M7NAK2E7>.

Conflicts of interest

There are no conflicts to declare.

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