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## Interplay of hydrogen bonding and $\pi$ -stacking interactions in the solid-state architecture of pranoprofen: insights from X-ray crystallography and computational analyses

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We report the crystal structure of the anhydrous form of pranoprofen, a valuable non-steroidal anti-inflammatory drug, solved by direct space methodologies from synchrotron X-ray powder diffraction data. Through a detailed joint experimental and computational study we have explored the complex interplay of aromatic and hydrogen-bonding interactions present in the molecular arrangement of pranoprofen in the solid state. Cooperativity and reinforcement of  $\pi$ -stacking and hydrogen bonding interactions govern the singular crystal packing of pranoprofen, suggesting potential binding mechanisms with biological targets.

### Introduction

Pranoprofen ( $\alpha$ -methyl-5H-[1]-benzopyrano [2,3-*b*]-pyridine-7-acetic acid), is a non-steroidal anti-inflammatory drug which can be used as a safe and effective alternative anti-inflammatory treatment following strabismus and cataract surgery.<sup>1–3</sup> This drug has the beneficial effect of reducing the ocular signs and symptoms of dry eye and decreasing the inflammatory markers of conjunctival epithelial cells.<sup>4</sup> Its efficacy is equivalent to moderate-potency corticosteroids, but it has a better safety profile and it is considered for the treatment of chronic conjunctivitis of presumed nonbacterial origin.<sup>5</sup> It also can be used for the treatment of the acute and long term management of osteoarthritis and rheumatoid arthritis. Although this drug has shown high anti-inflammatory and analgesic efficiency and a minimal risk of side effects on gastrointestinal tract after oral administration<sup>6</sup> the pharmaceutical use of pranoprofen is limited due to its inadequate biopharmaceutical profile, with a short plasmatic

half-life, low water solubility and is unstable in aqueous solution, particularly when exposed to light.<sup>7,8</sup>

To improve these limitations, pranoprofen has been formulated in poly (lactic/glycolic) acid nanoparticles and pranoprofen-loaded in nanostructured lipid carriers as a means of exploring novel formulations to improve the biopharmaceutical profile of PF for topical administration.<sup>9–13</sup> As a cyclooxygenase (COX) inhibitor, pranoprofen works by suppressing the synthesis of prostaglandins, key mediators of inflammation and pain. Recent studies have also explored its potential in combination therapies and its role in managing oxidative stress and inflammation-related complications, further expanding its scope of use in modern medicine.

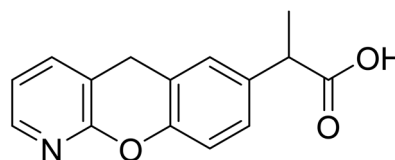
In this manuscript, we present the solid-state X-ray structure solved by direct space methodologies from X-ray synchrotron powder diffraction data of pranoprofen (Scheme 1) and analyzed its crystal packing, providing valuable insights into the predominant noncovalent interactions that shape its solid-state architecture. These findings can contribute to understanding the potential binding mechanisms of this drug with biological systems, such as the active sites of cyclooxygenase enzymes.

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**Scheme 1** Structure of pranoprofen.



Specifically, the H-bonding and  $\pi$ -stacking interactions identified in the solid state have been thoroughly examined using DFT calculations and various computational tools, including MEP, NCIPLOT, and QTAIM, with a particular focus on their cooperative effects.

## Materials and methods

### Materials

Pranoprofen® used in this study was kindly supplied by Alcon Cusi (Barcelona, Spain).

### X-Ray Powder Diffraction analysis (XRPD)

X-ray powder diffraction patterns of pranoprofen were obtained at 298 K using synchrotron radiation at ALBA's beam line BL04-MSPD<sup>14</sup> using Mythen detector<sup>15</sup> and a wavelength of 0.61939 Å. Capillary of 0.7 mm was used in rotation during data collection. Data acquisition time of 10 min per pattern was used, and the final treated data are the addition of ten acquisitions over the angular range of 0.5–43.6° (2 $\theta$ ).

### Computational details

The best solution from the direct space methodology conducted with the FOX software of anhydrous pranoprofen was subjected to geometry optimization by DFT<sup>16,17</sup> periodic calculations performed within the generalized gradient approximation (GGA),<sup>18</sup> as provided by the module CASTEP<sup>19</sup> in Materials Studio software,<sup>20</sup> using a basis set cutoff energy of 520 eV, ultrasoft pseudopotentials,<sup>21</sup> PBE functional, semi-empirical dispersion corrections (Grimme),<sup>22</sup> fixed unit cell and periodic boundary conditions. The validation of the structure<sup>23,24</sup> was conducted through a second calculation starting from the optimized structure obtained in the previous step by using the same DFT parameters but setting free the unit cell parameters. Atomic displacement RMSD values were calculated to assess the reliability of the optimization.

The X-ray geometry was used to analyze the energetic features of anhydrous pranoprofen at the PBE0-D3/def2-TZVP level of theory.<sup>25–27</sup> The positions of the H-atoms were optimized while keeping the heavy atoms fixed. Energetic calculations and wavefunction generation were performed using the Gaussian-16 program.<sup>28</sup> The QTAIM<sup>29</sup> and NCIPLOT<sup>30</sup> analyses were carried out using the AIMALL program<sup>31</sup> at the same computational level. Hydrogen bond energies were estimated using the values of the potential energy density and the methodology described in the literature.<sup>32</sup> Dimerization energies were calculated as the difference between the energy of the dimer and the sum of the monomer energies. Similarly, tetramers were computed as dimers. All interaction energies were corrected for basis set superposition error (BSSE) using the counterpoise method introduced by Boys and Bernardi.<sup>33</sup>

## Results and discussion

### Structure determination from space direct methods

The crystal structure determination of Pranoprofen was carried out by means of direct-space methodologies<sup>34</sup> and synchrotron high-resolution X-ray powder diffraction data obtained in the MSPD beam line in Alba (Fig. 1).

The powder diffractogram was indexed out to a monoclinic cell of about 1225 Å<sup>3</sup> by means of Dicvol04,<sup>35,36</sup> (figures of merit:  $M = 99$ ,  $F = 520$ ) and validated with a Le Bail fit of the data using FullProf<sup>37</sup> (goodness of fit:  $R_{wp}$ : 7.20,  $R_{exp}$ : 0.35;  $\chi^2 = 417$ ). The space group was deduced as  $P2_1/c$  from the systematic absences and confirmed with the SGAID program of the DAJUST<sup>38</sup> software. According to a typical estimated density value 1.4 Mg m<sup>-3</sup> the asymmetric unit was assumed to contain one molecule of pranoprofen ( $Z = 4$ ). The structure solution was carried out by direct space methodologies starting from a molecular model optimized by DFT with SPARTAN<sup>39</sup> by means of the program FOX<sup>40</sup> with the parallel tempering algorithm. The used background (estimated from a set of experimentally read points and interpolated) and the resulting cell, zero error and shape parameters of the Le Bail fit were used in the structure solution procedure with FOX. Several trials of 20 million runs were performed. The best solution (based on the  $R_{wp}$  value) was refined by the Rietveld method using FullProf,<sup>37</sup> in combination with DFT calculations in order to improve through geometry optimization the planarity of the aromatic rings and to locate the hydrogen atomic coordinates. Fig. 2 depicts the final Rietveld plot and the overlapped molecules

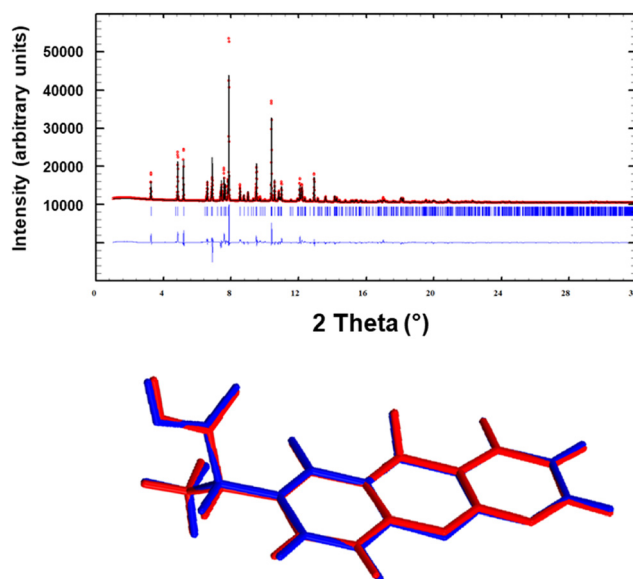
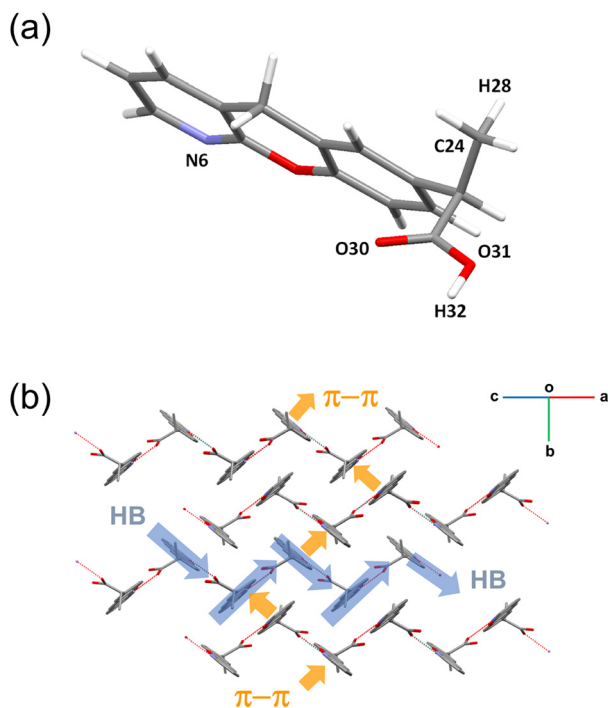


Fig. 1 Rietveld plot and comparative of asymmetric unit molecules after the Rietveld refinement (red) and after the validation procedure (blue) with DFT geometry optimization. The plot shows the experimental powder XRD profile (red marks), the calculated powder XRD profile (black solid line), the difference profile (blue, lower line) and Bragg positions (blue lines). Agreement factors:  $R_{wp} = 11.8\%$ ,  $R_{exp} = 0.34\%$ ;  $\chi^2 = 120$ .





**Fig. 2** (a) Numbering of atoms involved in hydrogen-bonding interactions. (b) Molecular packing, highlighting hydrogen bonds (blue arrows) and  $\pi$ -stacking interactions (orange arrows). Cell axes are also shown.

**Table 1** Crystallographic data from XRPD data of pranoprofen

|   |  |
|---|--|
| $T$ (K)                                 | 298                                      |
| System                                  | Monoclinic                               |
| Space group                             | $P2_1/c$                                 |
| $a$ (Å), $b$ (Å), $c$ (Å)               | 11.46400(16), 10.37828(14), 10.88732(18) |
| $\alpha$ (°), $\beta$ (°), $\gamma$ (°) | 90, 108.9328(10), 90                     |
| Volume (Å <sup>3</sup> )                | 1225.26(3)                               |
| $Z$                                     | 4  |
| $R$ (%)                                 | 11.8                                     |

of the asymmetric unit after the Rietveld refinement and the DFT validation procedure. A summary of crystal data with relevant refinement parameters is given in Table 1. Finally, the atomic coordinates (together with the cell parameters) were subjected to optimization by DFT calculations, aiming to validate the crystal structure (RMSD = 0.1027).<sup>41</sup> The hydrogen bond geometric data are summarized in Table 2.

### Structural description of pranoprofen crystal structure

Anhydrous pranoprofen crystallizes in the monoclinic  $P2_1/c$  space group and the crystal structure has one independent molecule in the asymmetric unit ( $Z = 4$ ). In the structure the

pranoprofen molecule establishes a strong intermolecular hydrogen bond between the carboxylic acid and the pyridine moieties. In addition, antiparallel orientated molecules are stacked through the establishment of aromatic interactions. Overall, the pranoprofen molecules are packed in a “zig-zag” pattern established by both hydrogen bond and aromatic interactions as depicted by blue and orange arrows in Fig. 2.

We have analyzed the most relevant intermolecular interactions in the structure by means of the Hirshfeld surface calculation<sup>42</sup> and the associated fingerprint plot<sup>43,44</sup> by using the Crystal Explorer software.<sup>45</sup>

Fig. 3 highlights the intermolecular hydrogen bonds on the Hirshfeld surface as red areas, together with the reciprocal  $H\cdots N$  and  $N\cdots H$  contacts as sharp spikes on the fingerprint plots. Weaker interactions like  $H\cdots O$ ;  $H\cdots H$ ;  $H\cdots C$  have been pointed out in the figure.

### DFT calculations

Fig. 4 provides a partial view of the solid-state structure of pranoprofen (**P**), highlighting the chromenopyridine scaffold of two **P** molecules engaged in antiparallel  $\pi$ -stacking interactions. In this arrangement, the pyridine ring of one molecule stacks over the phenyl ring of the adjacent molecule, and *vice versa*. Additionally, each molecule forms two very short  $OH\cdots N$  hydrogen bonds: one acting as a donor *via* the propionic acid arm and the other as an acceptor *via* the pyridine nitrogen atom. We propose that the latter interaction decreases the  $\pi$ -basicity of the pyridine ring, likely mitigating electrostatic repulsion between the  $\pi$ -electrons of the aromatic rings and thereby reinforcing the  $\pi$ -stacking interaction. This hypothesis is analyzed in detail below.

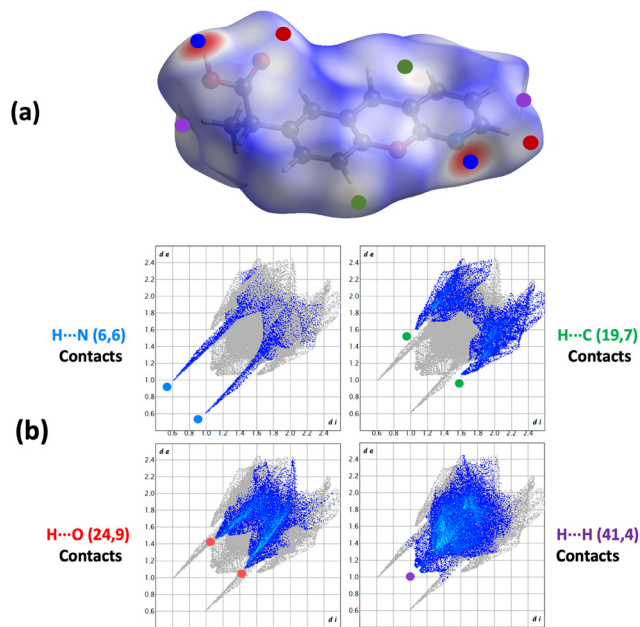
We initiated our study by calculating the MEP (molecular electrostatic potential) surfaces of pranoprofen (**P**) and its hydrogen-bonded dimer (**P**<sub>2</sub><sup>HB</sup>) to explore the electrophilic and nucleophilic regions of the molecule in the monomeric state and how these change upon dimer formation (Fig. 5). For the monomer, the MEP maximum is located at the carboxylic proton (+51.5 kcal mol<sup>-1</sup>), and the minimum is at the nitrogen atom of the pyridine ring (-39.5 kcal mol<sup>-1</sup>). This aligns with the very short  $POH\cdots N$  H-bond observed in the X-ray structure of **P** (see Fig. 3). Notably, the MEP values are also substantial and negative at the oxygen atom of the chromene unit and the carboxylic oxygen atom (-31.4 and -33.9 kcal mol<sup>-1</sup>, respectively). The MEP over the aromatic rings is modest and negative for both the phenyl and pyridine rings.

The MEP surface of the **P**<sub>2</sub><sup>HB</sup> dimer reveals significant changes in the molecule acting as the hydrogen bond acceptor. The MEP maximum increases to +54 kcal mol<sup>-1</sup>,

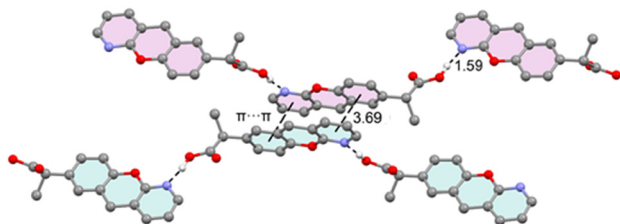
**Table 2** Hydrogen bond geometric data for pranoprofen

| Donor | Hydrogen | Acceptor | D-H (Å) | H $\cdots$ A (Å) | D $\cdots$ A (Å) | D-H $\cdots$ A (°) |
|-------|----------|----------|---------|------------------|------------------|--------------------|
| O31   | H32      | N6       | 1.02    | 1.59             | 2.59             | 165                |
| C24   | H28      | O30      | 1.10    | 2.59             | 3.49             | 138                |





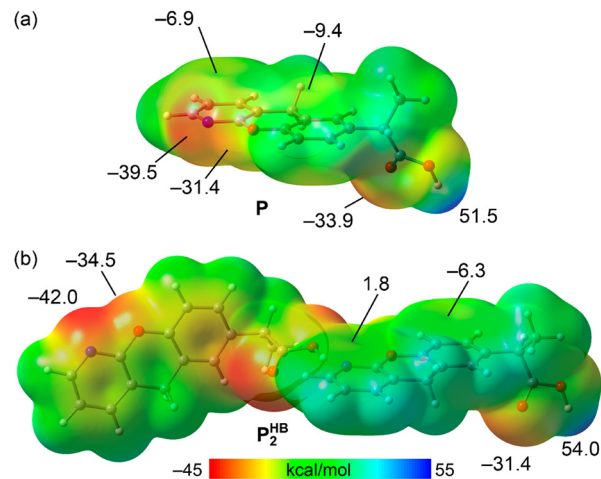
**Fig. 3** (a) Hirshfeld surface mapped on the pranoprofen molecule with  $d_{norm}$ . Relevant contacts are pointed out. (b) Fingerprint plots computed from Hirshfeld surface showing the percentage contribution of the individual types of interaction to the total Hirshfeld surface area. Main close contacts are highlighted: H...N (blue), H...O (red), H...C (green), H...H (purple).



**Fig. 4** X-ray solid state structure of P, with indication of the  $\pi$ -stacking, H-bonding interactions. Distances in Å. H-atoms omitted for clarity apart from the carboxylic proton.

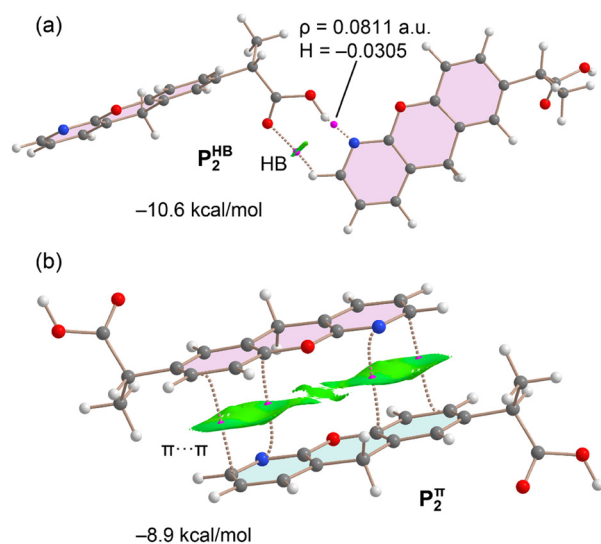
indicating an enhanced ability of the carboxylic group to establish hydrogen bonds. More importantly, the MEP over the pyridine ring transitions from negative ( $-6.9 \text{ kcal mol}^{-1}$ ) in the monomer to positive in the dimer ( $1.8 \text{ kcal mol}^{-1}$ ), providing clear evidence that the electronic properties of the chromenopyridine system are influenced by hydrogen bonding. Additionally, the MEP value at the N-atom of the pyridine ring in the monomer acting as a hydrogen bond donor becomes more negative ( $-42.0 \text{ kcal mol}^{-1}$ ), reflecting a strengthened H-bond acceptor capability.

This MEP analysis demonstrates that hydrogen bonding not only alters the polarity of the pyridine ring upon dimer formation but also enhances the dimer's overall ability to act as both a hydrogen bond donor and acceptor. Consequently, the infinite 1D hydrogen bond chains observed in the solid-state structure of P exhibit favorable cooperativity.



**Fig. 5** MEP surface analysis of P (a) and  $P_2^{HB}$  (b). Selected surface energies are given in  $\text{kcal mol}^{-1}$ .

**Fig. 6** presents the QTAIM/NCIplot analysis for the hydrogen-bonded and  $\pi$ -stacked dimers of P, designated as  $P_2^{HB}$  and  $P_2^{\pi}$ , respectively. The QTAIM analysis of the  $P_2^{HB}$  dimer indicates that the monomers are connected through two bond critical points (BCPs) and corresponding bond paths. One of these represents the short OH...N hydrogen bond, while the other corresponds to an ancillary CH...O interaction. The reduced density gradient (RDG) analysis reveals an isosurface coinciding with the BCP of the ancillary CH...O contact. Notably, the absence of an RDG isosurface for the OH...N hydrogen bond suggests a partial covalent character, supported by the high charge density at the BCP



**Fig. 6** Overlapped QTAIM and NCIplot analyses of the  $P_2^{HB}$  (a) and  $P_2^{\pi}$  (b) dimers. The dimerization energies are indicated. Bond critical points as fuchsia spheres and bond paths as dashed lines. Ring and cage critical points were omitted. For the NCIplot analysis, the following setting were used,  $S = 0.45$ ,  $\rho$  cut-off =  $0.04 \text{ a.u.}$ , color scale  $-0.035 \leq \text{sign}(\lambda_2)\rho \leq -0.035 \text{ a.u.}$  Only intermolecular interactions are represented.



(0.0811 a.u.) and the negative value of the total energy density. The interaction energy for the  $P_2^{HB}$  dimer is calculated to be  $-10.6 \text{ kcal mol}^{-1}$ , with the ancillary  $\text{CH}\cdots\text{O}$  interaction contributing only  $0.8 \text{ kcal mol}^{-1}$ .

The QTAIM/NCIplot analysis of the  $P_2^\pi$  dimer (Fig. 6b) highlights an extended green RDG isosurface, characterizing the  $\pi$ -stacking interaction. Intriguingly, this isosurface also partially extends over the non-aromatic ring of the chromenopyridine moiety. The  $\pi$ -stacking interaction is further defined by four BCPs and bond paths linking the phenyl and pyridine rings of the stacked molecules. The calculated interaction energy for the  $\pi$ -stacking is significant at  $-8.9 \text{ kcal mol}^{-1}$ , underscoring the critical role of this interaction in stabilizing the solid-state architecture of P.

Fig. 7 illustrates the combined QTAIM/NCIplot analysis of two different tetramers, both featuring the  $\pi$ -stacked dimer as a common core. These tetramers, labeled as  $P_4^A$  and  $P_4^B$ , differ in their hydrogen bonding roles: in  $P_4^A$ , the dimeric core ( $P_2^\pi$ ) acts as a hydrogen bond donor *via* the carboxylic groups, whereas in  $P_4^B$ , it serves as a hydrogen bond acceptor. In the tetramer  $P_4^A$ , no additional bond critical points or bond paths connect the monomers beyond those already observed in the isolated dimers  $P_2^{HB}$  and  $P_2^\pi$ . The interaction energy, calculated as the combination of two  $P_2^{HB}$  dimers, is  $-8.5 \text{ kcal mol}^{-1}$ —nearly identical to that of the isolated  $P_2^\pi$  dimer (see Fig. 4). This indicates that the monomers' participation in hydrogen bonding as donors does not influence the strength of the  $\pi$ -stacking interaction. In sharp contrast, the interaction energy for the  $P_4^B$  tetramer, also computed as the combination of two  $P_2^{HB}$  dimers, is significantly higher at  $-19.6 \text{ kcal mol}^{-1}$ , more than double the dimerization energy of  $P_2^\pi$ . QTAIM/NCIplot analysis reveals the formation of two symmetrically equivalent  $\text{CH}\cdots\text{O}$  interactions between the propyl group of one monomer and the carboxylate group of the adjacent monomer, marked as yellow circles in Fig. 7b. These interactions contribute an

energy of  $-2.8 \text{ kcal mol}^{-1}$  ( $-1.4 \text{ kcal mol}^{-1}$  per interaction). Consequently, the interaction energy attributed to the  $\pi$ -stacking within the tetramer increases to  $-16.8 \text{ kcal mol}^{-1}$ , nearly double that of the isolated  $\pi$ -stacking interaction.

This analysis highlights that the involvement of the pyridine N-atoms in hydrogen bonding interactions significantly enhances the  $\pi$ -stacking interaction, as anticipated by the MEP surface analysis. This interplay between hydrogen bonding and  $\pi$ -stacking could influence pranoprofen's behavior as an inhibitor in biological systems, where such interactions are prevalent in enzyme active sites. The mutual reinforcement of hydrogen bonding and  $\pi$ -stacking interactions could be leveraged to design more effective inhibitors or biomimetic materials, optimizing binding affinities and stability in target environments.

## Conclusions

The solid-state X-ray structure of pranoprofen reveals a complex interplay of  $\pi$ -stacking and hydrogen-bonding interactions that govern its molecular arrangement and stability. Computational analyses, including MEP, QTAIM, and NCIplot, provide a detailed understanding of the electronic and structural contributions to these interactions. The  $\pi$ -stacking interaction, supported by antiparallel aromatic ring alignment and mitigated electrostatic repulsion, plays a critical role in stabilizing the crystal structure. Moreover, the cooperative effects of hydrogen bonding, as evidenced by the enhanced polarity of key molecular regions upon dimer and tetramer formation, further reinforce the  $\pi$ -stacking interactions. These findings underline the importance of noncovalent interactions in shaping the solid-state architecture and offer insights into pranoprofen's potential binding mechanisms with biological targets, such as cyclooxygenase enzymes.<sup>46</sup> The observed mutual reinforcement of  $\pi$ -stacking and hydrogen bonding

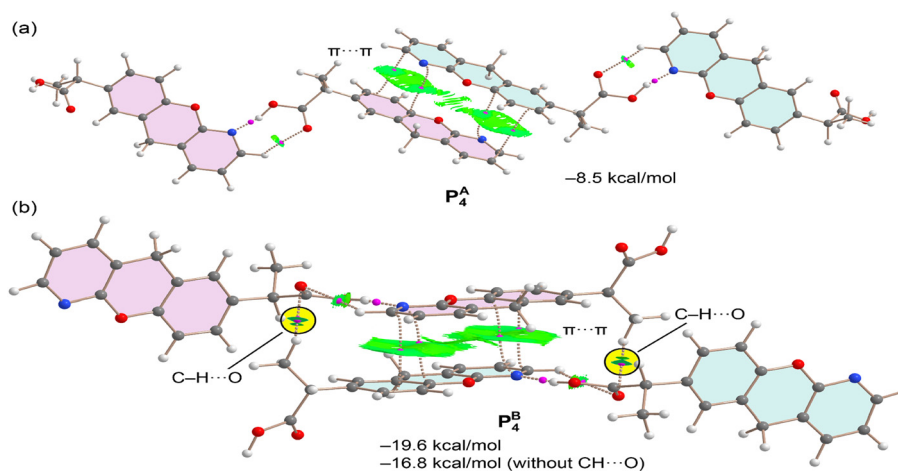


Fig. 7 Overlapped QTAIM and NCIplot analyses of the  $P_4^A$  (a) and  $P_4^B$  (b) dimers. The dimerization energies are indicated (the  $P_2^{HB}$  dimer is considered as monomer for the calculation). Bond critical points as fuchsia spheres and bond paths as dashed lines. Ring and cage critical points were omitted. For the NCIplot analysis, the following setting were used,  $S = 0.45$ ,  $\rho$  cut-off =  $0.04 \text{ a.u.}$ , color scale  $-0.035 \leq \text{sign}(\lambda_2\rho) \leq -0.035 \text{ a.u.}$  Only intermolecular interactions are represented.



interactions provides a valuable framework for designing biomimetic materials and improving drug efficacy through optimized molecular interactions.

## Data availability

Crystallographic data for anhydrous pranoprofen has been deposited at the CCDC under CCDC number 2410183 and can be obtained from <https://www.ccdc.cam.ac.uk>.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

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## Notes and references

- I. Akyol-Salman, D. Lece-Sertoz and O. Baykal, *J. Ocul. Pharmacol. Ther.*, 2007, **23**, 280–283.
- M. Sawa, K. Masuda and M. Nakashima, *Japanese Journal of Cataract and Refractive Surgery*, 1999, **13**, 193–200.
- A. Z. McColgin and J. S. Heier, *Curr. Opin. Ophthalmol.*, 2000, **11**, 3–6.
- X. Liu, S. Wang, A. A. Kao and Q. Long, *Cornea*, 2012, **31**, 1235–1239.
- R. Notivol, M. Martinez and M. M. Bergamini, *Am. J. Ophthalmol.*, 1994, **117**, 651–656.
- G. Kajal, C. Subrata and N. Arunabha, *Pharm. Sin.*, 2011, **2**, 152–168.
- M. Narashino, H. Ichikawa, S. Narita and A. S. Yotsukaido, *United States Pat.*, US005225206A, 1993.
- K. Kobe, H. Otsu, Y. Kobe and Y. Kakogawa, *United States Pat.*, US005856345A, 1999.
- G. Abrego, H. L. Alvarado, M. A. Egea, E. Gonzalez-Mira, A. C. Calpena and M. L. Garcia, *J. Pharm. Sci.*, 2014, **103**, 3153–3164.
- C. Cañadas, H. L. Alvarado, A. C. Calpena, A. M. Silva, E. B. Souto, M. L. García and G. Abrego, *Int. J. Pharm.*, 2016, **511**, 719–727.
- C. Cañadas, G. Abrego, H. L. Alvarado, A. C. Calpena and A. Boix-Montañes, *J. Pharm. Biomed. Anal.*, 2018, **160**, 109–118.
- M. Rincón, A. C. Calpena, M. J. Fabrega, M. L. Garduño-Ramírez, M. Espina, M. J. Rodríguez-Lagunas, M. L. García and G. Abrego, *Nanomaterials*, 2018, **8**, 1022–1050.
- M. Rincón, A. C. Calpena, B. Clares, M. Espina, M. L. Garduño-Ramírez, M. J. Rodríguez-Lagunas and G. Abrego, *Nanomedicine*, 2018, **13**, 2397–2413.
- F. Fauth, R. Boer, F. Gil-Ortiz, C. Popescu, O. Vallcorba, I. Peral, D. Fullà, J. Benach and J. Juanhuix, *Eur. Phys. J. Plus*, 2015, **130**, 160–173.
- F. Fauth, I. Peral, C. Popescu and M. Knapp, *Powder Diffr.*, 2013, **28**, S360–S370.
- P. Hohenberg and W. Kohn, *Phys. Rev. A*, 1964, **136**, B864.
- W. Kohn and L. J. Sham, *Phys. Rev. A*, 1965, **140**, A1133.
- J. P. Perdew, K. Burke and M. Ernzerhof, *Phys. Rev. Lett.*, 1997, **78**, 1396.
- S. J. Clark, M. D. Segall, C. J. Pickard, P. J. Hasnip, M. J. Probert, K. Refson and M. C. Payne, *Z. Kristallogr.*, 2005, **220**, 567–570.
- Materials Studio, version 7.0*, Accelrys Software, Inc., San Diego, CA, USA, 2013.
- D. Vanderbilt, *Phys. Rev. B: Condens. Matter Mater. Phys.*, 1990, **41**, 7892–7895.
- S. Grimme, *J. Comput. Chem.*, 2006, **27**, 1787–1799.
- J. van de Streek and M. A. Neumann, *Acta Crystallogr., Sect. B: Struct. Sci., Cryst. Eng. Mater.*, 2014, **70**, 1020–1032.
- J. van de Streek and M. A. Neumann, *Acta Crystallogr., Sect. B: Struct. Sci.*, 2010, **66**, 544–558.
- C. Adamo and V. Barone, *J. Chem. Phys.*, 1999, **110**, 6158–6170.
- S. Grimme, J. Antony, S. Ehrlich and H. Krieg, *J. Chem. Phys.*, 2010, **132**, 154104.
- F. Weigend, *Phys. Chem. Chem. Phys.*, 2006, **8**, 1057–1065.
- M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. V. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. J. Bearpark, J. J. Heyd, E. N. Brothers, K. N. Kudin, V. N. Staroverov, T. A. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. P. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman and D. J. Fox, *Gaussian 16, Revision C.01*, Gaussian, Inc., Wallingford CT, 2016.
- R. F. W. Bader, *Chem. Rev.*, 1991, **91**, 893–928.
- E. R. Johnson, S. Keinan, P. Mori-Sánchez, J. Contreras-García, A. J. Cohen and W. Yang, *J. Am. Chem. Soc.*, 2010, **132**, 6498–6506.
- T. A. Keith, *AIMAll (Version 19.10.12)*, TK Gristmill Software, Overland Park KS, USA, 2019, <https://aim.tkgristmill.com>.
- E. Espinosa, E. Molins and C. Lecomte, *Chem. Phys. Lett.*, 1998, **285**, 170–173.
- S. F. Boys and F. Bernardi, *Mol. Phys.*, 1970, **19**, 553–566.
- S. Ghosh, S. Islam, S. Pramanik and S. Kumar Seth, *J. Mol. Struct.*, 2022, **1268**, 133697.
- A. Boultif and D. Louër, *J. Appl. Crystallogr.*, 1991, **24**, 987–993.
- A. Boultif and D. Louër, *J. Appl. Crystallogr.*, 2004, **37**, 724–731.



- 37 J. Rodriguez–Carvajal, *Phys. B*, 1993, **192**, 55–69.
- 38 O. Vallcorba, J. Rius, C. Frontera, I. Peral and C. Miravittles, *J. Appl. Crystallogr.*, 2012, **45**, 844–848.
- 39 *Spartan'10*, Wavefunction Inc., Irvin, CA, USA, 2014.
- 40 V. Favre-Nicolin and R. Cerný, *J. Appl. Crystallogr.*, 2002, **35**, 734–743.
- 41 J. van de Streek and M. A. Neumann, *Acta Crystallogr., Sect. B: Struct. Sci., Cryst. Eng. Mater.*, 2014, **70**, 1020–1032.
- 42 M. A. Spackman and D. Jayatilaka, *CrystEngComm*, 2009, **11**, 19–32.
- 43 M. A. Spackman and J. J. Mckinnon, *CrystEngComm*, 2002, **4**, 378–392.
- 44 J. J. Mckinnon, D. Jayatilaka and M. A. Spackman, *Chem. Commun.*, 2007, 3814–3816.
- 45 M. J. Turner, J. J. McKinnon, S. K. Wolff, D. J. Grimwood, P. R. Spackman, D. Jayatilaka and M. A. Spackman, *CrystalExplorer17*, University of Western Australia, 2017, <https://hirshfeldsurface.net>.
- 46 S. Xu, D. J. Hermanson, S. Banerjee, K. Ghebreselasie, G. M. Clayton, R. M. Garavito and L. J. Marnett, *J. Biol. Chem.*, 2014, **289**, 6799–6808.

