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The synthesis of [18F]SynVesT-1 and [18F]SynVesT-2 from enantioenriched boronic esters

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Enantioenrcihed [18F]SynVesT-1 and [18F]SynVesT-2 are prepared from the corresponding enantioenriched arylboronic esters and [18F]fluoride mediated by (Py)₄Cu(OTf)₂. The ¹⁸F-reaction is optimised on an automated synthesiser to produce multi-GBq quantites of both radiotracers to support (pre)clinical imaging.

Positron emission tomography (PET) imaging is a non-invasive technology that can provide quantitative information on biochemical processes to diagnose and monitor diseases in living subjects.1 The loss or alteration of synaptic density is closely associated with neurodegenerative disorders including Alzheimer's and Parkinson's disease.^{2,3} The synaptic vesicle glycoprotein 2A (SV2A) has been shown to play an essential role in neurotransmission and levels of SV2A been validated as a biomarker for synaptic density.4 Several small molecule PET radiotracers to target SV2A have been developed allowing for the non-invasive quantification and visualisation of SV2A in-vivo with applications towards the diagnosis and development of treatments for neurological diseases. The PET radiotracer [11C]UCB-J has been shown to be a competent imaging agent of SV2A,⁵ but the relatively short half-life of 11 C ($t_{1/2} = 20.4$ minutes) prohibits the broader use of this radiotracer, particularly in sites without an on-site cyclotron, thus the development of PET radiotracer with fluorine-18 ($t_{1/2}$ = 110 minutes) is highly desirable. The presence of three aryl-fluoride bonds in UCB-J theoretically allows for a direct synthesis of the ¹⁸F isotopologue, however, to date attempts to radiolabel this compound with ¹⁸F have been met with limited success. ^{6b,7} This led to the development of the structurally related compounds $[^{18}\text{F}]\text{SynVestT-1}$ and $[^{18}\text{F}]\text{SynVesT-2}$ which can accessed in

- asymmetric synthesis of SynVesT-1 and SynVesT-2 radiolabelling precursors >99:1 e.r
- fully automated synthesis delivering GBq activity yields of SynVesT-1 and SynVesT-2
- improved stability profile of BEpin precursors compared to Bpin

Scheme 1: a) PET radiotracers for imaging SV2A; b) this work

PET imaging studies revealed the *R*-enantiomer of both SynVestT-1 and SynVestT-2, have superior imaging characteristics to the *S*-enantiomer,^{6e} yet to date most studies access the desired *R*-enantiomer by chiral HPLC separation of the ¹⁸F-racemate resulting in decreased RCYs. A single asymmetric synthesis of ¹⁸F-SynVest-1 aryl stannane radiolabelling precursor has been reported, while none for a SynVesT-2 radiolabelling precursor.⁸ We have recently demonstrated the translation of the copper-mediated ¹⁸F-fluorination of aryl boronic esters onto an automated synthesis platform resulting in a good manufacturing practice (GMP)

improved radiochemical yields (RCY) and molar activities (A_m) (Scheme 1a).⁶

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compatible method for [18 F]flumazenil, while Scott and Sanford have reported a similar protocol for [18 F]FDOPA. 9

We reasoned the development of an automated protocol for the production of ¹⁸F-SynVest radiotracers from aryl Bpin precursors would be beneficial due to the advantageous stability of these precursors and the ability to use tetrabutylammonium carbonate for the preparation of [¹⁸F]tetrabutylammonium fluoride (*vide infra*). To this end herein we describe the first asymmetric syntheses of aryl boronic precursors for both SynVesT-1 and SynVesT-2 and their automated syntheses (Scheme 1b).

To access the key aryl Bpin compounds we envisaged an asymmetric synthesis of an aryl bromide (5) would allow us to introduce the Bpin functionality as the final step for each of the SynVesT compounds. The start of our synthesis of SynVesT-1 began with the nitroalkene (1a), prepared from the commercially available 3-bromo-5-fluorobenzaldehyde. For the asymmetric Michael addition, we followed the protocol by Deng and co-workers that was successfully employed in the asymmetric synthesis of UCB-J.5,10 Treating 1a with 10 mol % of (+)-O-desmethyl quinidine and 3 equivalents of diethyl malonate in THF at room temperature gave 2a in 79 % yield. We were unable to determine the enantiomeric ratio (e.r) of 2a at this stage and so proceeded with a reductive cyclization using Fe/NH₄Cl in EtOH/H₂O which yielded 3a in 84 % yield as a 2:1 mixture of diastereoisomers (Scheme 2). Hydrolysis of the ester and sequential decarboxylation gave the butyrolactam(4a) in 77 % yield. Separation of the two enantiomers of (4a) was possible by chiral stationary phase HPLC analysis, giving an e.r of 82:18.

Scheme 2: Asymmetric synthesis of SynVesT-1 Bpin precursor

In an attempt to increase the e.r we took inspiration after the patent literature, and repeated the passymmetric of the patent literature, and the passymmetric of th

The absolute stereochemistry of the major enantiomer was assigned as the desired *R*-enantiomer based comparison of the optical rotation with literature values. ^{8,12} *N*-Alkylation of the butyrolactam with 4-(chloromethyl)-3-methylpyridine gave (5a) in 61 %. The target SynVesT-1 Bpin precursor (6a) was synthesised from B₂pin₂ (1.5 eq.), KOAc (5 eq.) and 10 mol % PdCl₂(dppf) in DMF at 80°C in 46 % yield. The same synthetic strategy was applied to the synthesis of SynVesT-2 Bpin precursor (6b) (Scheme 3). The addition of 20 mol % of DIPEA was found again to be necessary in the asymmetric Michael addition to achieve high levels of asymmetric induction (e.r =>99:1 with DIPEA vs. 90:10 in absence). In contrast to the previous route, here we were able to measure the e.r of (2b) directly, the assignment of the absolute stereochemistry was done based on analogy with SynVesT-1.

Having established synthetic routes to the two enantioenriched Bpin radiolabelling precursors for SynVesT-1 (6a) and SynVesT-2 (6b), our next consideration was optimisation of the ¹⁸F-fluorination on an automated radiochemical synthesizer.

From a safety, and good manufacturing practice (GMP) perspective, the synthesis ¹⁸F-radiotracers on an automated synthesis unit is a key challenge to address in method development. ¹³ Due to the higher dead-volumes, restrictions on

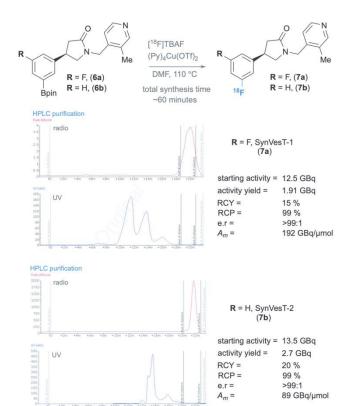
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reagents/solvents and the inability to work typical inert chemistry conditions this process can often be non-trivial and can require significant optimization.



Scheme 4: Automated synthesis of [18F]SynVesT-1 & [18F]SynVesT-2

Based on this protocol we employed the following conditions for the ¹⁸F-fluorination step; 12 mg Bpin precursor, 32 mg (Py)₄Cu(OTf)₂ in 0.5 ml dimethylformamide at 110 °C for 20 minutes. Following the fluorination step the crude mixture was purified by C18 HPLC purification and reformulation from a C18 cartridge. Chiral stationary phase HPLC analysis demonstrated that no racemisation of either precursor or ¹⁸F-product occurring during the reaction, giving both [18F]SynVesT-1, with an activity yield of 1.91 GBq (n=1) and [18F]SynVesT-2 with an activity yield of 2.7 GBq (n=1), with e.r >99:1 (Scheme 4).

Test strips confirmed that the purification protocol was successful in removing residual copper to levels below the ICH(Q3) limit of 340 μg/V for patient use. 12 During these studies we found that the purification of both Bpin precursors 6a and 6b was non-trivial and could often result in decomposition of the material. To address this limitation, we investigated the ¹⁸Ffluorination of a boronic ester derived from 3,4-diethylhexane-3,4-diol (Epin) which have been shown to be more stable towards purification but reactive in ¹⁸F-fluorination mediated by Cu.¹⁴ Precursor **6c** was synthesised from **5c** using PdCl₂(dppf) (5 mol %), B₂Epin₂ (1.1 equiv)and KOAc (3 equiv) in dioxane at 80 °C for 18 hours (Scheme 5a). Pleasingly compound 6c was easily purified by silica gel column chromatography and isolated in 48 % yield. Subjecting 6c to our automated ¹⁸F-conditions described above we were able to isolate 7c in 20 % RCY (n=2)

and in activity yields of 2.8 GBg and 1.5 GBg, starting from [18F]fluoride activities of 13.4 GBg and 8: 10Bg 39fe 66tively (Scheme 5b). As we observed for the Bpin precursors there was no loss in stereochemical integrity during the fluorination procedure, with 7a being isolated with an e.r > 99:1. This result further highlights that BEpin esters serve as practical precursors in Cu/18F radiochemistry for unstable and difficult to purify aryl boronic esters.

a) Synthesis of aryl BEpin precursor PdCl₂(dppf) (5 mol %) B₂Epin₂ (1.1 eq) KOAc (3 eq) 18 hrs Et Et EtEt b) Automated ¹⁸F-fluorination of BEpin precursor 18FITBAF (Py)₄Cu(OTf)₂ DMF. 110 °C total synthesis time ~60 minutes starting activity = 8-13.4 GBa activity yield = 1.5-2.8 GBq Et Et EtEt RCY = 20 % (n=2) RCP = 99 % (n=2) >99:1 Scheme 5: Automated synthesis of [18F]SynVesT-1 from BEpin

Herein, we have reported the asymmetric synthesis of Bpin precursors of both SynVesT-1 and SynVesT-2 in e.rs >99:1. Automated copper-mediated ¹⁸F-fluorination gave the corresponding enantiopure radiotracers with activity yields sufficient to support (pre)clinical imaging. The purification of the aryl Bpin precursors was found to be challenging but this could be overcome by use of aryl BEpin precursors that gave essentially identical yields under our optimised automated ¹⁸Fconditions. We believe this work demonstrates that these compounds are a useful alternative to the commonly used stannyl precursors.

Conflicts of interest

MT is an inventor on a patent (WO201510572A1) relating to a copper-mediated ¹⁸F-fluorination method.

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Data availability

The data underlying this study are available in the published article, in its Supporting Information, and openly available in the Cardiff

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University data catalogue https://doi.org/10.17035/cardiff.30196870.

Notes and references

- M. E. Phelps, Proc. Natl. Acad. Sci. U. S. A, 2000, 97, 9226.
- J. E. Hamos, L. J. DeGennaro, D. A. Drachman, Neurology, 1989, **39**, 355.
- S. E. Holmes, P. Honhar, S. Tinaz, M. Naganawa, A. T. Hilmer, J.-D. Gallezot, M. Dias, Y. Yang, T. Toyonaga, I. Esterlis, A. Mecca, C. Van Dyck, S. Henry, J. Ropchan, N. Nabulsi, E. D. Louis, R. Comley, S. J. Finnema, R. E. Carson, D. Matuskey, Npj Parkinsons Dis. 2024, 10, 42.
- S. J. Finnema, N. B. Nabulsi, T. Eid, K. Detynieccki, S.-F. Lin, M.-K. Chen, R. Dhaher, D. Matuskey, E. Baum, D. Holden, D. D. Spencer, J. Mercier, J. Hannestad, Y. Huang, R. E. Carson, Sci. Transl. Med. 2016, 8, 348ra96.
- N. B. Nabulsi, J. Mercier, D. Holden, S. Carré, S. Najafzadeh, M.-C. Vandergeten, S.-F. Lin, A. Deo, N. Price, M. Wood, T. Lara-Jaime, F. Montel, M. Laruelle, R. E. Carson, J. Hannestad, Y. Huang, J. Nucl. Med. 2016, 57, 777.
- (a) J. Mercier, L. Archen, V. Bollu, S. Carré, Y. Evrard, E. Jnoff, B. Kenda, B. Lallemand, P. Michel, F. Montel, F. Moureau, N. Price, Y. Quesnel, X. Sauvage, A. Valade, L. Provins, ChemMedChem, 2014, 9, 693; (b) S. Li, Z. Cai, W. Zhang, D. Holden, S.-F. Lin, S. J. Finnema, A. Shirali, J. Ropchan, S. Carre, J. Mercier, R. E. Carson, N. Nabulsi, Y. Huang, Eur. J. Nucl. Med. Mol. Imaging, 2019, 46, 1952; (c) C. C. Constantinescu, C. Tresse, M. Zheng, A. Gouasmat, V. M. Carroll, L. Mistico, D. Alagille, C. M. Sandiego, C. Papin, K. Marek, J. P. Seibyl, G. D. Tamagnan, O. Barret, Mol. Imaging Biol. 2019, **21**, 509; (d) S. Li, Z. Cai, X. Wu, D. Holden, R. Pracitto, M. Kapinos, H. Gao, D. Labaree, N. Nabulsi, R. E. Carson, Y. Huang, ACS Chem. Neurosci. 2019, 10, 1544. (e) K. Dahl, S. Larsson, P. Bonn, A. Wallin, O. Itsenko, M. Schöll, J. Label. Compod. Radiopharm. 2022, 65, 315; (f) L. R. Drake, Y. Wu, M. Naganawa, R. Asch, C. Zheng, S. Najafzadeh, R. Pracitto, M. Lindemann, S. Li, J. Ropchan, P. R. Emery, M. Dias, S. Henry, N. Nabulsi, D. Matuskey, A. T. Hillmer, J.-D. Gallezot, R. E. Carson, Z. Cai, Y. Huang, J. Nucl. Med. 2024, 65, 462; (g) L. Y. F. Haveman, A. M. T. de Kruijff, S. P. P. van Eeden, A. D. Windhorst, D. J. Vugts, Chem. Eur. J, 2025, 31, e202403127
- F. Sirindil, S. Maher, M. Schöll, K. Sander, E. Årstad, Int. J. Mol. Sci. 2022, 23, 15481.
- H. McErlain, E. B McLean, T. E. F. Morgan, V. K. Burianova, A. A. S. Tavares, A. Sutherland, J. Org. Chem. 2022, 87, 14443.
- (a) A. V. Mossine, S. S. Tanzey, A. F. Brooks, K. J. Makaravage, N. Ichiishi, J. M. Miller, B. D. Henderson, T. Erhard, C. Bruetting, M. B. Skaddan, M. S. Sanford, P. J. H. Scott, Nat. Protoc. 2020, 15, 1742; (b)T. Gendron, G. Destro, N. J. W. Straathof, J. B. I. Sap, F. Guibbal, C. Vriamont, C. Caygill, J. R. Atack, A. J. Watkins, C. Marshall, R. Hueting, C. Warnier, V. Gouvernuer, M. Tredwell, EJNMMI Radiopharm. Chem. 2022, **7**. 5.
- 10 (a) H. Li, Y. Wang, L. Tang, L. Deng, J. Am. Chem. Soc. 2004, 126, 9906; (b) H. Li, Y. Wang, L. Tang, F. Wu, X. Liu, C. Guo, B. M. Foxman, Li. Deng, Angew. Chem. Int. Ed. 2005, 44, 105.
- 11 P. Wang, P. Li, Q. Wei, Y. Liu, Process for preparation of gamma-aminobutyric acid chiral compounds. CN 104557583A, 2015.
- 12 See Supporting Information for details.
- 13 (a) J. Aerts, J. R. Ballinger, M. Behe, C. Decristoforo, P. H. Elsinga, A. Faivre-Chauvet, T. L. Mindt, P. K. Peitl, S. C. Todde, J. Koziorowski, J. Labelled Compd. Radiopharm. 2014, 57, 615; (b) J. I. Sachinidis, S. Poniger, H. J. Tochon-Danguy, Curr. Radiopharm. 2010, 3, 248.

14 a) A. Craig, F. J. Sachse, M. Laube, F. Brandt, K. Kopka, S. View Article Online Stadlbauer, Pharmaceutics, 2025, 17, 837; b) N39/D5CC05507E Hadjipaschalis, S. Ortalli, Z. Chen, R. S. Paton, J. Ford, M. Tredwell, V. Gouverneur, Org. Lett. 2025, 27, 6545-6550; c) S. Kaur, B. Wenzel, R. Oehme, C. Wiesner, K. Kopka, R.,-P. Moldovan, EJNMMI Radiopharm. Chem. 2025, 10, 60.

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Data availability

The data underlying this study are available in the published article, in its Supporting Information, and openly available in the Cardiff University data catalogue at https://doi.org/10.17035/cardiff.30196870.