


 Cite this: *Chem. Commun.*, 2025, 61, 1451

 Received 6th October 2024,  
 Accepted 28th October 2024

DOI: 10.1039/d4cc05254d

rsc.li/chemcomm

# Anion exchange polymer modified electrodes for detection of $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC): a potential electrochemical sensor for point-of-care and roadside testing†

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**The effect of a polybenzimidazolium anion exchange polymer on improving sensitivity and selectivity toward the electrochemical detection of  $\Delta^9$ -tetrahydrocannabinol ( $\Delta^9$ -THC) has been investigated. Herein we report a rapid, inexpensive and stable approach to detecting 10–1000 ng mL<sup>-1</sup> of  $\Delta^9$ -THC in buffered solutions and in human saliva.**

Cannabis, commonly known as marijuana, is increasing in use worldwide, with over 192 million users,<sup>1</sup> who use it not only for recreational but also for medicinal purposes. In 2023, 7.6% of the UK population, ~2.5 million people, reported using cannabis in the last 12 months.<sup>2</sup> Over one-third of people aged 16 to 59 years who used cannabis in the last year consumed the drug more than once a month, with 8.3% using it in their everyday life. The frequent use of cannabis has increased to over 8% since 2020 for those aged 30 to 34 years.

Several countries worldwide have legalised the use of cannabis for recreational purposes; however, its use is restricted in public parks, schools, and hospitals. Driving under the influence of cannabis is a major concern. A recent *meta*-analysis has found that cannabis increased impairment for lane control and reaction time, as well as decreased compensatory speed.<sup>3</sup> In the UK, among the deceased drivers between 2014 and 2021 in road accidents, cannabis was the most common illegal drug leading to road fatality.<sup>4</sup>

$\Delta^9$ -Tetrahydrocannabinol ( $\Delta^9$ -THC) is the psychoactive component in cannabis; because of its high lipophilicity it can easily cross over the blood–brain barrier and interact with the central nervous system (CNS) cannabinoid receptors. This leads to several psychological effects such as euphoria, relaxation, decreased concentration, somnolence, temporal distortions, and hallucinations.<sup>5,6</sup>

Therefore, multiple jurisdictions have attempted to set *per se*  $\Delta^9$ -THC limits in blood and saliva. In Canada, a roadside threshold of 5 ng mL<sup>-1</sup> when analysed in blood and 25 ng mL<sup>-1</sup> when measured in saliva has been adopted.<sup>3</sup> In England and Wales, a *per se* threshold of 2 ng mL<sup>-1</sup> in blood has been established.<sup>7</sup> In the US the *per se* limits or cutoffs vary among different states but range from THC blood levels of 1 to 5 ng mL<sup>-1</sup>  $\Delta^9$ -THC.<sup>8</sup> On-site testing for THC levels has proved problematic, and hence there is a need for rapid roadside detection of  $\Delta^9$ -THC that can be used to accurately screen drivers.

$\Delta^9$ -THC can be tested in urine, blood, breath, and saliva.<sup>9</sup> Urine tests, a commonly employed method for drug testing, can only detect  $\Delta^9$ -THC metabolites, which are only traceable in urine several hours after the intake of cannabis, far past the window of intoxication and impairment.<sup>10</sup> Blood testing is not necessarily a good approach for  $\Delta^9$ -THC detection probably due to its high log *P* and enterohepatic recirculation:  $\Delta^9$ -THC usually accumulates in the tissues for long periods of time and it can be detected in the blood up to 30 days post-consumption.<sup>6</sup>  $\Delta^9$ -THC is retained in the breath for short periods of time, no longer than 3 h, making it difficult to determine if the driver is still under its influence. Oral fluid appears to be preferable for  $\Delta^9$ -THC roadside monitoring, being quick and less invasive. Preliminary evidence exists that indicates that the concentration of  $\Delta^9$ -THC in saliva can be correlated with impairment.<sup>10</sup>

The  $\Delta^9$ -THC concentration in saliva has been found to be affected by the method of consumption. *Via* smoking, it is reported to take 0.17 h to detect THC's maximum concentration. The  $\Delta^9$ -THC level in the saliva of frequent smokers is on average 9  $\mu$ M (2.83  $\mu$ g mL<sup>-1</sup>) and 3  $\mu$ M (944 ng mL<sup>-1</sup>) for occasional users. For  $\Delta^9$ -THC consumed orally, it takes 0.33–0.41 h to detect the maximum salivary concentration presenting concentrations of 944 nM (297 ng mL<sup>-1</sup>) in frequent users and 642 nM (202 ng mL<sup>-1</sup>) in occasional users.<sup>11</sup> Hence, it is necessary to develop a detection method able to comply with current jurisdictions and capable of detecting a range of salivary  $\Delta^9$ -THC concentrations from 25 to 1000 ng mL<sup>-1</sup>.

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† Electronic supplementary information (ESI) available: Experimental details and addition information. See DOI: <https://doi.org/10.1039/d4cc05254d>



In recent years, multiple techniques for detecting  $\Delta^9$ -THC in saliva have been researched including the use of liquid chromatography with UV-vis or mass spectrometry detection, immunoassays, and surface enhanced Raman scattering.<sup>5,12</sup> Although these techniques can detect low levels of  $\Delta^9$ -THC, they rely on relatively costly, non-portable, or non-scalable instrumentation. For example, the current on-market roadside test device, Dräger DrugTest 5000, employs an immunoassay approach to detect  $\Delta^9$ -THC with a detection limit of 5 ng mL<sup>-1</sup> but at high cost (~5500 USD). Another device is the Druglizer LE5 Drug Testing System, based on a competitive lateral flow immunoassay with a  $\Delta^9$ -THC cut-off of 15 ng mL<sup>-1</sup>, provides qualitative test result costs in excess of 3195 USD plus cartridges. Other commercial roadside test devices are based on colour test strips, but do not provide quantitative information (Rapid STAT<sup>®</sup>, DrugWipe<sup>®</sup> S), which is a key requirement in jurisdictions where there is a  $\Delta^9$ -THC threshold.

Electrochemical sensors, on the other hand, offer the possibility of low cost, rapid, high selectivity and sensitivity, and low sample volume. Electrochemical sensors are simple and portable devices for the detection of  $\Delta^9$ -THC in biological fluids such as saliva.<sup>5,12</sup>

$\Delta^9$ -THC is detectable electrochemically due to the direct oxidation of the phenol moiety. The hydroxy group is deprotonated at high pH, leading to the formation of a phenoxide anion that can be oxidised under low potential values. The oxidation of the  $\Delta^9$ -THC has been proven to be favoured at carbon-based surfaces. However, the instability of phenoxy radical produced leads to subsequent chemical reactions (Fig. 1).<sup>13,14</sup>

Several electrochemical approaches for  $\Delta^9$ -THC detection have been published. However, some of the designs required lengthy electrodes or sample pre-treatments, using modified electrodes which have a short shelf-life (Table S1, ESI<sup>†</sup>).

In this context, the presented work explores the potential of a low-cost anion conductive polymer as a modification for screen printed carbon electrodes (SPCE). Anion exchange polymers, widely used as alkaline anion exchange membranes in fuel cells and electrolyzers, are positively charged, and are anticipated to result in the pre-concentration of  $\Delta^9$ -THC and facilitate its stabilisation by hindering inherent dimerization reactions. Pre-concentration of  $\Delta^9$ -THC is expected to enhance the signal to noise ratio, improving the sensitivity of the sensor. Anion exchange polymers can also be designed to possess hydrophobic domains leading to hydrophobic interactions with  $\Delta^9$ -THC, promoting its stabilisation and hindering the formation of by-products.

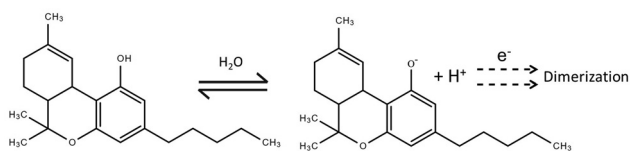


Fig. 1 Chemical structure of  $\Delta^9$ -THC and its electrochemical oxidation.

Lastly, anion exchange polymers repel positively charged interferences present in saliva, such as nicotine.

In this work, we demonstrate that anion exchange polymer, HMT-PMBI, meets all of the above and can be used to fabricate modified screen printed electrodes (SPEs), which can electrochemically detect  $\Delta^9$ -THC in concentrations as low as 10 ng mL<sup>-1</sup> in buffered solutions. Moreover, preliminary studies performed on untreated saliva are presented, wherein  $\Delta^9$ -THC is detected, opening the possibility in the near future, of rapidly identifying and quantifying  $\Delta^9$ -THC in the saliva samples of human subjects.

In order to achieve sensing detection, several experimental conditions were optimised. In the initial studies the goal was to voltammetrically evaluate screen printed electrodes prepared by the drop casting HMT-PMBI polymer solutions. Two different electrodes were investigated: Gii-Sense (pure 3D graphene foam sensing electrodes) and Metrohm C11L, details of which are presented in the ESI.<sup>†</sup> All the electrochemical cells employed were based on screen printed electrodes presenting a carbon working electrode, carbon counter electrode and Ag/AgCl reference electrode. For both Gii-Sense and Metrohm electrodes the presence of the anionic polymer film increases the anodic current density of  $\Delta^9$ -THC oxidation (Fig. S1, ESI<sup>†</sup>). In the presence of 10  $\mu$ g mL<sup>-1</sup>  $\Delta^9$ -THC, a typical unmodified Gii-Sense SPCE presents a peak current density of 5.6  $\mu$ A cm<sup>-2</sup> and unmodified Metrohm SPE, 14.6  $\mu$ A cm<sup>-2</sup>. These results were unexpected as it was thought that the hydrophobic  $\Delta^9$ -THC molecule would form hydrophobic interactions with the graphene foam leading to higher preconcentration and higher sensitivity. However, it is worth noting that the 3D foam structure of the Gii-Sense electrode presents higher capacitive currents, increases the background noise, possibly reducing sensitivity. The modification of SPEs with a thin polymer film leads to an increase in anodic current density, consistent with the hypothesis of preconcentration by the positively charged HMT-PMBI solid polymer electrolyte with the negatively charged  $\Delta^9$ -THC analyte. Furthermore,  $\Delta^9$ -THC also possesses a hydrophobic character which is complementary with the highly hydrophobic segments of the polymer (Fig. 1 and 2).<sup>15</sup> These segmented electrostatic and hydrophobic interactions are deemed responsible for the preconcentration of  $\Delta^9$ -THC and an increase in sensitivity of its detection. The polymer modified Metrohm electrodes exhibited a 4-fold increase in the anodic peak current (63.4  $\mu$ A cm<sup>-2</sup>), and the modified Gii-Sense electrodes yielded a 13 $\times$  response (71.2  $\mu$ A cm<sup>-2</sup>) compared to the pristine electrodes. The larger increase of the latter is believed to be due to its foam-like structure, presenting a

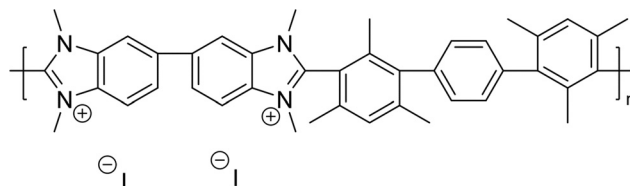


Fig. 2 HMT-PMBI structure in the iodide form.



higher electrode effective area and increased porosity, thus, facilitating the integration of the polymer within the foam during the drop-cast method and easing the diffusion of the electroactive species in comparison with packet carbon working electrodes.

Both electrodes were examined under conditions of low  $\Delta^9$ -THC concentrations ( $50 \text{ ng mL}^{-1}$ ) which are more akin to roadside saliva  $\Delta^9$ -THC concentrations (Fig. S2, ESI<sup>†</sup>). For  $50 \text{ ng mL}^{-1}$   $\Delta^9$ -THC concentrations, the polymer modified Metrohm electrodes presented a clearly visible oxidation peak, whereas modified Gii-Sense electrodes did not. No reproducible anodic peak was achievable with polymer modified Gii-Sense electrodes with  $\Delta^9$ -THC concentrations below  $500 \text{ ng mL}^{-1}$ . These results are consistent with a large capacitive current masking the low  $\Delta^9$ -current density of THC oxidation.

Differential pulse voltammetry (DVP) ( $E$  step:  $0.1 \text{ V}$ ,  $E$  pulse:  $0.2 \text{ V}$ ,  $T$  pulse:  $0.02 \text{ s}$ , scan rate:  $0.01 \text{ V s}^{-1}$ ) using Metrohm C11L electrodes provided greater consistency and smaller background, capacitive signals, resulting in enhanced  $\Delta^9$ -THC detection at low  $\Delta^9$ -THC concentrations.<sup>9</sup> Modified Gii-Sense electrodes on the other hand showed complementary sensitivity in the high concentration regime.

Taking into consideration the  $pK_a$  of  $\Delta^9$ -THC phenol (10.6), the effect of solution pH on electrochemical detection of  $\Delta^9$ -THC at polymer modified electrodes was examined using Metrohm electrodes (Fig. S3, ESI<sup>†</sup>). The electrochemical response to  $\Delta^9$ -THC was found to increase with increasing pH, which is deemed to be due to increasing deprotonation, rendering an anion which may diffuse into the cationic polymer film. Moreover, at higher pH values the  $\Delta^9$ -THC oxidation signal shifts to more negative potentials. This effect was previously observed by Williams *et al.*,<sup>9</sup> where a shift in the reaction quotient was suggested. The use of solutions of  $\text{pH} > 11$  were avoided as the high concentration of  $\text{OH}^-$  competes against  $\Delta^9$ -THC anions for polycation sites in the film, lowering the concentration of electrostatically bound  $\Delta^9$ -THC. In contrast, the anodic current decreases below  $\text{pH} 5.0\text{--}6.0$  due to protonation of  $\Delta^9$ -THC to form a cation, which is repelled by the polycationic film.<sup>12,16,17</sup> Therefore, the pH was optimized to 10 employing TRIS buffer.

The quantity of HMT-PMBI deposited on electrodes was found to have a direct impact on the anodic oxidation of  $\Delta^9$ -THC (Fig. S4, ESI<sup>†</sup>). The anodic current corresponding to  $\Delta^9$ -THC oxidation increased with increasing concentrations of drop cast HMT-PMBI solutions, up to 2% (w/v). However, the oxidative peak decreased when higher concentration polymer solutions were deposited, dropping to virtually no observable current for 10% (w/v) solutions, presumably due to high ionic resistance and long diffusion lengths imposed by the film. Consequently, 2% (w/v) of polymer was considered to be the optimal concentration for further study.

The rate of  $\Delta^9$ -THC uptake on the modified electrodes was investigated by immersing modified electrodes in a pH 10, TRIS buffered solution containing  $1000 \text{ ng mL}^{-1}$   $\Delta^9$ -THC (Fig. S5, ESI<sup>†</sup>). It was found that the electrochemical response of  $\Delta^9$ -THC increases with time, reaching a maximum after 45 minutes. 40 minutes was chosen as the pre-concentration period.

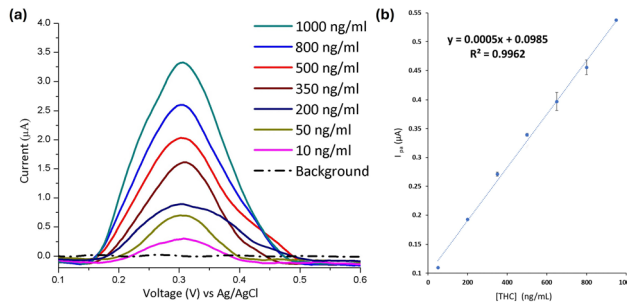


Fig. 3  $\Delta^9$ -THC signals from triplicate experiments performed using 2% (w/v) of HMT-PMBI coated on Metrohm C11L screen printed electrodes in TRIS buffered solution at pH 10 (a) and the linear fit of the  $\Delta^9$ -THC data presenting the corresponding error bars (b), each concentration was performed in triplicate.

With all the above parameters investigated, a  $\Delta^9$ -THC calibration curve was constructed (Fig. 3). Fig. 3a presents DVP curves of polymer modified electrodes recorded in buffered solutions of increasing  $\Delta^9$ -THC concentration, and Fig. 3b shows the plot of the anodic peak current vs.  $\Delta^9$ -THC concentration. A linear regressive response for  $\Delta^9$ -THC was found ( $I_{pa} (\mu\text{A}) = 0.0005X (\text{ng mL}^{-1}) + 0.0985$  ( $R^2 = 0.996$ )).  $\Delta^9$ -THC was detected at concentrations as low as  $10 \text{ ng mL}^{-1}$ . The calculated limit of detection (LOD) was  $5.28 \text{ ng mL}^{-1}$  obtained from the relation  $\text{LOD} = 3.3Sb/S$ .<sup>18</sup> Consequently, the calculated limit of quantification (LOQ), was  $16 \text{ ng mL}^{-1}$  (10 SD blank per slope).

Reproducibility of the response to  $1000 \text{ ng mL}^{-1}$   $\Delta^9$ -THC was examined by preparing 11 different modified electrodes (Fig. S6, ESI<sup>†</sup>). The mean of the anodic peaks was  $0.614 \mu\text{A}$  [std. dev.,  $0.0587$ ,  $9.5\%$  RSD, and  $0.614 \pm 0.0347$  ( $\pm 6.65\%$ ) margin of error at 95% confidence interval] (Fig. S6, ESI<sup>†</sup>). Moreover, the electrodes exhibited notable stability over time, when stored in a dry room and room temperature environment; no visible differences in their  $\Delta^9$ -THC detection capabilities were detected after several weeks of storage.

A variety of components are expected to be found in human saliva during a roadside testing. These include ascorbic acid (AA) or caffeine (CF).<sup>12</sup> The impact of these analytes on pristine and modified electrodes was therefore investigated (Fig. 4) by mixing  $1 \mu\text{g mL}^{-1}$  of  $\Delta^9$ -THC with increasing concentrations of AA and CF (concentrations ranging from  $10 \mu\text{g mL}^{-1}$  to  $100 \mu\text{g mL}^{-1}$ ) in TRIS buffer solutions (pH 10). The electrochemical responses of bare electrodes are significantly affected (up to 400% increase in the anodic current) by the presence of the AA and/or CF, indicating their clear interference.

In contrast, the presence of AA and CF did not influence the electrochemical response of  $\Delta^9$ -THC even for 100-fold mass ratios of AA and/or CF.

Selectivity is deemed to be due to the hydrophobic nature of the polymer structure repelling the relatively hydrophilic ascorbic acid and promoting the preconcentration of the hydrophobic  $\Delta^9$ -THC.

Preliminary studies of modified electrodes were performed on human saliva (Fig. 5) spiked with  $\Delta^9$ -THC.  $5 \mu\text{g mL}^{-1}$  of



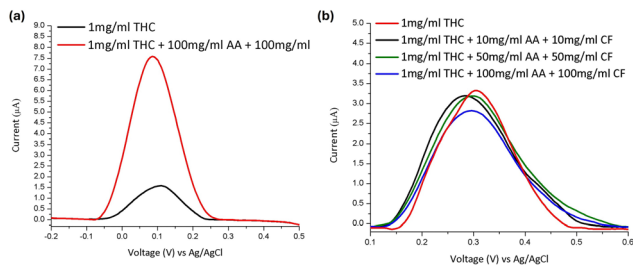


Fig. 4 Comparison of the selectivity of (a) unmodified Metrohm C11L screen printed electrode (b) 2% (w/v) of HMT-PMBI coated on Metrohm C11L screen printed electrode in the presence of ascorbic acid (AA) and caffeine (CF) as interferences (1  $\mu\text{g mL}^{-1}$   $\Delta^9$ -THC, TRIS buffer pH 10).

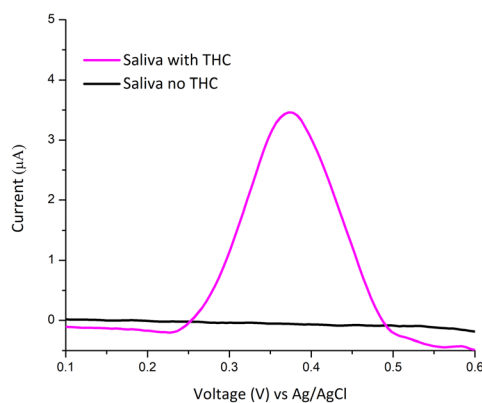


Fig. 5 Differential pulse voltammetry of  $\Delta^9$ -THC in spiked saliva using 2% (w/v) of HMT-PMBI coated on Metrohm C11L screen printed electrode. 1 mL of saliva was spiked with 5  $\mu\text{g mL}^{-1}$  of  $\Delta^9$ -THC and diluted to a 9 mL solution before testing (550  $\text{ng mL}^{-1}$ ).

$\Delta^9$ -THC was added to 1 mL of saliva and diluted to a 9 mL solution. Dilution of the saliva is recommended based on two conditions: (i) typical human saliva possesses kinematic viscosities of 1.40  $\text{mm}^2 \text{s}^{-1}$  whereas that of water is 1  $\text{mm}^2 \text{s}^{-1}$ ; high viscosities lower the absorption rate of electroactive species on the surface of the carbon working electrode. (ii) While saliva has neutral pH, detection is optimised at pH 10. Voltammetry on saliva alone did not reveal any anodic peak in the voltage range of interest; whereas the spiked saliva yielded a peak current of 0.39  $\mu\text{A}$ .

In summary, a simple, affordable, portable, and stable electrochemical method is presented for the detection of low concentrations of  $\Delta^9$ -THC (10–1000  $\text{ng mL}^{-1}$ ). An anion exchange polymer coated electrode is demonstrated to have sensitivity towards electrochemical oxidation of  $\Delta^9$ -THC, increasing the current density by 400%. The polycationic polymer films provides selectivity against interferences such as caffeine and ascorbic acid. These findings may prove important in the future design of low-cost, portable, roadside systems for  $\Delta^9$ -THC detection.

A.O. – investigation, validation, methodology, data curation; S.H.A. – funding acquisition, conceptualization, resources, formal analysis, project administration, writing – original draft, supervision; S.H. – investigation, synthesis of HMT-PMBI,

writing – review and editing. All authors have read and agreed to the published version of the manuscript.

This work was financially supported by the Royal Society of Chemistry through an RSC Research Fund grant (R21-8000 280413).

## Data availability

The data supporting this article have been included as part of the ESI.† All of the differential pulse voltammetry graphs produced in this research have been presented in the main article or in the ESI.† The calibration data obtained are also presented in Fig. 3. Raw data for the experiments not involving human participants are available by contacting the corresponding author. Data collected from human participants, described in Fig. 5, are not available for confidentiality reasons.

## Conflicts of interest

There are no conflicts to declare.

## Notes and references

- 1 N. S. Miller, R. Ipeku and T. Oberbarnscheidt, *Int. J. Environ. Res. Public Health*, 2020, **17**, 1578.
- 2 Office for National Statistics (ONS), Drug misuse in England and Wales: year ending March 2023, <https://www.ons.gov.uk/peoplepopulationandcommunity/crimeandjustice/articles/drugmisuseinenglandandwales/latest>, 2024.
- 3 P. Di Ciano, B. Brands, A. Fares, M. Wright, G. Stoduto, P. Byrne, M. McGrath, O. S. M. Hasan, B. Le Foll and C. M. Wickens, *Cannabis Cannabinoid Res.*, 2023, **8**, 408–413.
- 4 Department for Transport UK, Drugs in reported road fatalities in Great Britain, data to 2021: summary, <https://www.gov.uk/government/statistics/developing-drug-driving-statistics/drugs-in-reported-road-fatalities-in-great-britain-data-to-2021-summary>, 2024.
- 5 B. Zanfrognini, A. Monari, G. Foca, A. Ulrici, L. Pigani and C. Zanardi, *Microchem. J.*, 2022, **183**, 108108.
- 6 V. Ramzy and R. Priefer, *Talanta*, 2021, **222**, 121528.
- 7 Department for Transport UK, Changes to drug driving law, <https://www.gov.uk/government/collections/drug-driving>, 2024.
- 8 T. R. Arkell, T. R. Spindle, R. C. Kevin, R. Vandrey and I. S. McGregor, *Traffic Inj. Prev.*, 2021, **22**, 102–107.
- 9 K. J. Williams, C. J. White, A. P. Hunt and M. E. Meyerhoff, *Electroanalysis*, 2023, **35**, e202200451.
- 10 J.-R. Lee, J. Choi, T. O. Shultz and S. X. Wang, *Anal. Chem.*, 2016, **88**, 7457–7461.
- 11 M. J. Swortwood, M. N. Newmeyer, M. Andersson, O. A. Abulseoud, K. B. Scheidweiler and M. A. Huestis, *Drug Test. Anal.*, 2017, **9**, 905–915.
- 12 T. Pholsiri, W. Khamcharoen, S. Vimolmangkang, W. Siangproh and O. Chailapakul, *Sens. Actuators, B*, 2023, **383**, 133571.
- 13 K. Amini, A. Sepehrifard, A. Valinasabpour, J. Safruk, D. Angelone and T. de Campos Lourenco, *J. Cannabis Res.*, 2022, **4**, 12.
- 14 M. Renaud-Young, R. M. Mayall, V. Salehi, M. Golezdzinski, F. J. E. Comeau, J. L. MacCallum and V. I. Birss, *Electrochim. Acta*, 2019, **307**, 351–359.
- 15 A. G. Wright, J. Fan, B. Britton, T. Weissbach, H.-F. Lee, E. A. Kitching, T. J. Peckham and S. Holdcroft, *Energy Environ. Sci.*, 2016, **9**, 2130–2142.
- 16 S. Hernandez-Aldave, A. Tarat and P. Bertoncello, *Chemosensors*, 2021, **9**, 325.
- 17 M. Rees, A. G. Wright, S. Holdcroft and P. Bertoncello, *Sensors*, 2020, **20**, 443.
- 18 European Medicines Agency, Note for Guidance on Validation of Analytical Procedures: Text and Methodology (CPMP/ICH/381/95), EMEA 2006, Canary Wharf, London, 1994.

