



Cite this: *Chem. Commun.*, 2025, 61, 3093

Advances in magnetic nanoparticles for molecular medicine†

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Magnetic nanoparticles (MNPs) are highly versatile nanomaterials in nanomedicine, owing to their diverse magnetic properties, which can be tailored through variations in size, shape, composition, and exposure to inductive magnetic fields. Over four decades of research have led to the clinical approval or ongoing trials of several MNP formulations, fueling continued innovation. Beyond traditional applications in drug delivery, imaging, and cancer hyperthermia, MNPs have increasingly advanced into molecular medicine. Under external magnetic fields, MNPs can generate mechano- or thermal stimuli to modulate individual molecules or cells deep within tissue, offering precise, remote control of biological processes at cellular and molecular levels. These unique capabilities have opened new avenues in emerging fields such as genome editing, cell therapies, and neuroscience, underpinned by a growing understanding of nanomagnetism and the molecular mechanisms responding to mechanical and thermal cues. Research on MNPs as a versatile synthetic material capable of engineering control at the cellular and molecular levels holds great promise for advancing the frontiers of molecular medicine, including areas such as genome editing and synthetic biology. This review summarizes recent clinical studies showcasing the classical applications of MNPs and explores their integration into molecular medicine, with the goal of inspiring the development of next-generation MNP-based platforms for disease treatment.

Received 1st October 2024,
Accepted 20th January 2025

DOI: 10.1039/d4cc05167j

rsc.li/chemcomm

Introduction

Magnetic nanoparticles (MNPs), composed of crystals of magnetic metal elements or metal oxides, possess unique nanoscale magnetic properties, such as superparamagnetism, high magnetization, and rapid relaxation in the presence of external

magnetic fields. With dimensions similar to biomolecules, MNPs can translate magnetic signals into mechanical or thermal stimuli targeted at specific cell surface receptors, structural proteins, or subcellular organelles. This capability enables remote control of biological processes at cellular and molecular levels, making MNPs invaluable for both research and therapeutic applications.

The exploration of MNPs in biomedical research dates back to the 1960s, when magnetic attraction of iron carbonyl macroparticles was first demonstrated *in vivo* using experimental dogs.¹

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† Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d4cc05167j>



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This capacity for directing MNPs to specific tissues sparked numerous studies on magnetic targeting for drug and gene delivery. In parallel, MNPs were also identified as effective contrast agents in magnetic resonance imaging (MRI). Furthermore, the heat generated by MNPs under an alternating magnetic field (AMF) led to the development of deep tissue heating technology. Over the past four decades, targeted drug delivery, imaging, and thermal therapy have become the three primary areas of clinical research on MNPs.² Many studies have culminated in the clinical approval of MNPs for tumor imaging and thermal ablation therapy (Table 1).

During the same period, the synthesis of MNPs has seen major advancements (Fig. 1). The first generation synthesis relied on the rapid coprecipitation of iron salts in aqueous buffers, producing large quantities of iron oxide nanocrystals with varied shapes and broad size distributions.^{16,17} One notable product of this method is ferumoxytol, a formulation that was first approved for clinical use in 2009 and remains a prominent candidate in ongoing clinical trials (Fig. 1(A)). A breakthrough in the field was the development of the thermal decomposition method for nanocrystal synthesis.^{18,19} In this process, organic iron salts are heated to high temperatures in non-coordinating solvents, allowing their gradual decomposition into Fe–O precursors. This method allows for greater control over nanocrystal size, shape, and composition – critical factors influencing nanoscale magnetic responses (Fig. 1(B) through Fig. 1(L)).^{20–26} Compared to coprecipitation, which typically produces nanocrystals smaller than 10 nanometers, thermal decomposition can yield highly uniform nanocrystals ranging from a few nanometers to several hundred nanometers. Consequently, this enhances both the magnetic moment and heat generation, enabling novel approaches for manipulating molecular or cellular targets.^{25,27–31} A detailed review of nanosynthesis and magnetic properties of MNPs can be found elsewhere.^{2,32,33}

In the last decade, a wide range of innovative applications leveraging MNP-mediated mechanical or thermal stimulation have emerged, with the potential to significantly transform future medical interventions. For instance, our study demonstrated that MNPs can enhance the cellular entry of an insect viral vector, offering a valuable platform for targeted *in vivo* gene editing.³⁵ Other studies have shown that MNPs can mechanically stimulate cells, altering cytoskeleton, activating ion channels, and inducing cell polarization. Notably, Lee *et al.* used large clusters of MNPs (~ 100 s nm) to modulate neuronal activity *via* the mechanosensitive ion channel, Piezo1.^{29,30} Interestingly, separate research have indicated that synthetic and biogenic MNPs tethered to thermosensitive ion channels, TRPs, can activate neurons through heating.^{34,36–39} Although the precise mechanisms are still not fully understood,^{40–42} these findings could lead to groundbreaking advances in brain modulation.

Another important discovery is the ability of MNPs to catalyze the generation of hydroxyl free radicals *via* a heterogeneous Fenton reaction.^{43–45} The free radicals are key intermediates in chemo- and radiation therapies. Compared to small-molecule chemotherapeutic reagents, MNPs have improved availability to diseased tissue due to a long circulation half-life and targeting capability. Therefore, MNPs can not only serve as MRI detectable, targeted drug carriers but also an effective adjuvant to other drug molecules, providing new treatment avenues. This finding also shed light on mitigating potential adverse effects of MNPs.

In this review, we will briefly summarize the clinical studies illustrating the classical applications of MNPs, followed by an exploration of cutting-edge research on their mechanical, thermal, and enzymatic effects in recent years. We will highlight the studies utilizing MNPs as a nanoscale force or heat generators for targeted modulation of biological processes, as these will pave the way for future precision medicine.



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Table 1 Representative clinical applications and clinical trials of MNPs

	Agent	Condition	Identifier	Start date	Status
Magnetic targeting	Ferrofluids	Various cancers	—	1996	Phase I
	MTC-DOX	Liver cancer	NCT00034333	2002	Phase II/III, terminated
	Silica-gold iron-bearing NP ^{3,4}	Atherosclerotic lesions	NCT01270139	2007	Completed
	CD34 antibody labeled MNP ⁵	Leukemia	NCT01411904	2010	Withdrawn
Molecular imaging/cell tracking	MNP-labeled HCECs ⁶	Corneal edema	NCT04894110	2021	Phase I
	Endorem ^{®7}	Liver tumor	—	—	1996 FDA approval, currently discontinued
	SPIO ⁸	Melanoma	NCT00243594	1999	Phase I/II, completed
	Resovist [®]	Hepatic cancer	NCT00307866	2001	Phase III, completed
Thermal therapy	Resovist [®]	Liver tumor	—	2001	EMA approval, currently discontinued
	Lumirem [®]	GI track imaging	—	—	1996 FDA approval, 2001 EMA approval, currently discontinued
	Ferumoxtran-10	Prostate cancer	NCT00185029	2002	Phase IV
	Ferumoxtran-10	Breast cancer	NCT00107484	2005	Phase II, completed
Thermal therapy	Feridex ^{®9}	Multiple sclerosis and amyotrophic lateral sclerosis	NCT00781872	2006	Phase I/II, completed
	Sinerem ^{®10,11}	Prostate and bladder cancer	NCT00622973	2007	Phase III, completed
	USPIO	Infectious and degenerative diseases of the spine	NCT00372281	2007	Phase III
	Feraheme [®]	Pancreatic cancer	NCT00920023	2008	Phase IV, completed
Thermal therapy	USPIO ¹²	Aortic aneurysm instability	NCT00794092	2008	Phase II/III, completed
	Ferumoxytol	Brain tumor	NCT00978562	2009	Phase I, completed
	Feraheme [®]	Anemia associated with chronic renal failure, iron deficiency anemia	—	—	2009 FDA approval
	SPIO ¹³	Healthy volunteer	NCT00972946	2009	Completed
Thermal therapy	SPIO ¹³	Healthy volunteer with induced inflammation in the forearm	NCT01169935	2010	Completed
	Ferahem ^{®14}	Myocardial infarction	NCT01323296	2010	Completed
	Ferahem ^{®15}	Healthy volunteer	NCT01244282	2010	Phase I, completed
	SPIO	Myocardial infarction	NCT01127113	2010	Withdrawn
Thermal therapy	NanoTherm [®]	Glioblastoma multiforme	—	—	2010 EMA approval
	MNPs	Prostate cancer	NCT02033447	2013	Phase I, completed
	Iron nanoparticles	Prostate cancer	NCT05010759	2021	Terminated
	NanoTherm [®]	Recurrent glioblastoma multiforme	NCT06271421	2024	Recruiting

Abbreviations: MTC-DOX – magnetic targeted carriers absorbed with doxorubicin hydrochloride; HCEC – human corneal endothelial cell; NP – nanoparticle; MNP – magnetic nanoparticle; SPIO – superparamagnetic iron oxide particle; GI – gastrointestinal; USPIO – ultrasmall superparamagnetic iron oxide nanoparticle.

Current clinical investigations

MNPs are among the most extensively studied nanomaterials in clinical research and have received multiple approvals for medical use in both the US and Europe (Table 1, for a full list, see ESI,† Table S1). In clinical settings, only iron oxide nanoparticles, such as magnetite (Fe_3O_4) and maghemite ($\gamma-Fe_2O_3$), are used, owing to their well-established pharmacokinetic properties and biosafety profiles.^{46,47} Notably, ferumoxytol has been approved for the treatment of iron deficiency anemia, owing to its biocompatibility and biodegradability. While most clinical research has focused on cancer detection and treatment, recent studies have revealed the potential of MNPs in other disease conditions, including myocardial infarction (NCT01323296), atherosclerosis (NCT01270139), diabetes (NCT01521520), and retinal edema (NCT04894110). In this section, we summarize the key clinical applications of MNPs, emphasizing their magnetic force, imaging, and thermal properties.

Magnetic targeting

Magnetic nanoparticles (MNPs) were initially developed for magnetically targeting therapeutic agents to specific tissues. However, their clinical application has been complicated by the need to balance multiple factors essential for effective drug delivery, including drug payload, pharmacokinetics, targeting efficiency, and drug release. The efficiency of magnetic targeting depends on the relative magnitude of magnetic force *versus* fluid drag force exerted on MNPs in the blood circulation, which are body and surface forces, respectively. Larger MNPs are generally more favorable for magnetic targeting due to their smaller surface area-to-volume ratio.

Preclinical studies have demonstrated that magnetic targeting must be combined with intra-arterial (ia) infusion of drug-loaded MNPs *via* tumor-feeding vessels to achieve effective tumor inhibition.⁴⁸ In this approach, ia infusion increases the availability of MNPs in tumor tissue, while magnetic targeting enhances their local retention. Along these lines, several clinical trials for



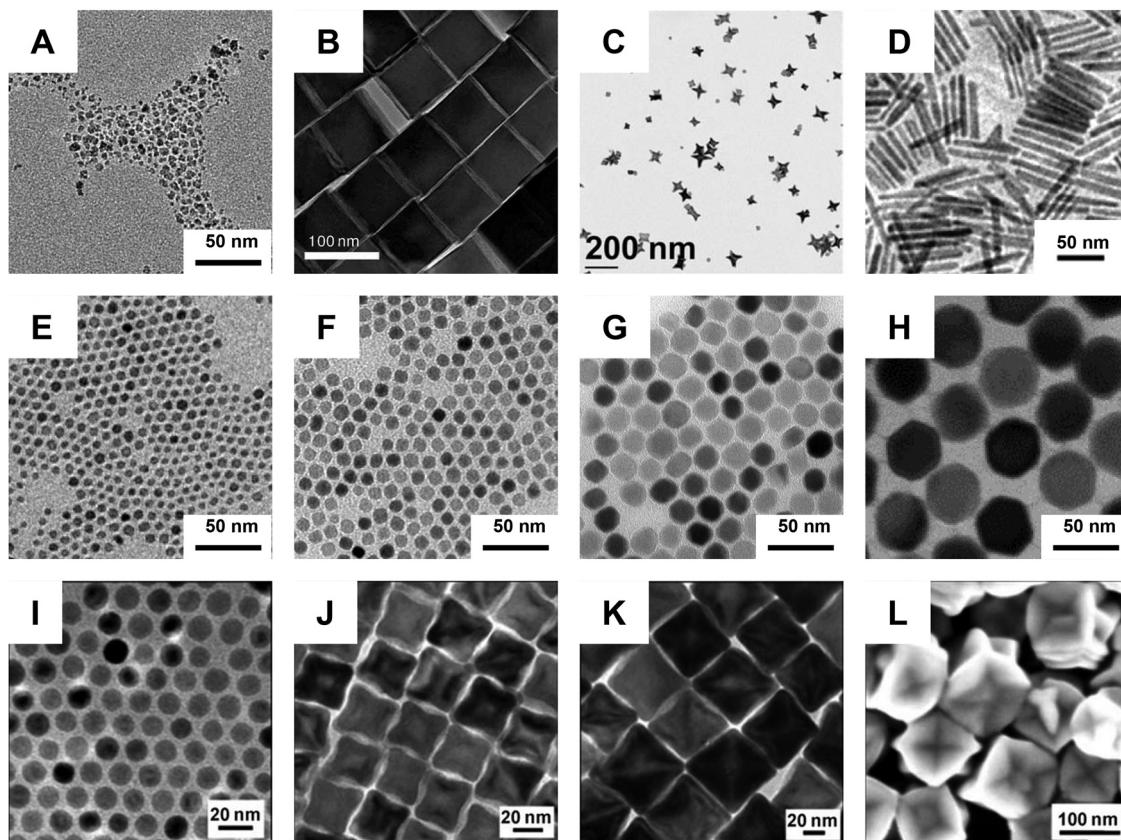


Fig. 1 Synthesis of iron oxide nanoparticles. (A) Ferumoxytol synthesized via co-precipitation. (Reproduced with permission from ref. 34, ©2023 John Wiley and Sons). B through L are iron oxide nanoparticles synthesized via thermodecomposition. (B) Magnetite nanocubes. (Reproduced with permission from ref. 22, ©2009 American Chemical Society). (C) Magnetite nanostars. (Reproduced with permission from ref. 23, ©2011 American Chemical Society). (D) Magnetite nanorods. (Reproduced with permission from ref. 24, ©2012 American Chemical Society). (E)–(H). Magnetite nanoparticles from 6 to 40 nm. (Reproduced with permission from ref. 25, ©2011 American Chemical Society). (I)–(L). Wüstite nanoparticles from 14 to 100 nm. (Reproduced with permission from ref. 26, ©2007 Wiley-VCH Verlag GmbH & Co. KGaA, Weinheim).

magnetic targeting were performed with magnetic targeted carriers (MTC) infused through tumor-feeding arteries in patients with advanced hepatocarcinoma.⁴⁹ MTCs are 0.5–5.0 μm iron particles bound to doxorubicin. While MRI confirmed enhanced accumulation of MTCs in liver tumors, the clinical trials were terminated due to a lack of therapeutic efficacy. It is likely that MTCs failed to penetrate the tumor tissue, as the dense extracellular matrix of solid tumors can form a barrier to the transport of macron-sized particles.^{50,51} The success of future magnetic drug delivery strategies will require a careful balance between particle size, drug payload, and magnetic traction.

An alternative approach utilizes MNPs to control cell movement in and out of the body. Clinical trials have explored this strategy for targeted delivery of endothelial cells to treat corneal edema in a minimally invasive way (NCT04894110), as well as for extracting circulating cancer cells or leukemia cells to reduce metastasis and detect minimal residual disease (NCT02025413 and NCT01411904).

Imaging applications

Since the 1980s, research has revealed that MNPs can alter the magnetic relaxation of nearby protons, generating imaging

contrast detectable through T_1 - or T_2 -weighted MRI. Compared to small-molecule contrast agents, such as GdDTPA, MNPs offer enhanced sensitivity in detecting functional changes associated with malignancy, significantly improving MRI detection capability. For example, liver tumors are identified by the loss of phagocytic activity in tumor tissue compared to normal liver tissue.⁵² The mechanism is also used to detect lymph node metastasis in various cancers.⁵³

Moreover, MNPs have been extensively investigated for detecting disease-associated inflammation, which increases vascular endothelial permeability or leads to the infiltration of phagocytes, both resulting in MNP accumulation in diseased tissues. Clinical studies have explored their use in monitoring diabetes progression, detecting atherosclerotic plaques, and assessing inflammation related to myocardial infarction and cerebral aneurysms (see ESI,† Table S1).

In cell therapy, evaluating the distribution of engineered cells in the body is critical to determining therapeutic efficacy. MNPs, with their low cytotoxicity, force-mediated endocytosis, and excellent detection sensitivity, provide a valuable tool for this purpose.^{54,55} Due to their entrapment in lysosomes and slow degradation, MNPs can label the cells for several days,



enabling real-time and long-term tracking of mesenchymal stem cells, dendritic cells, and mononuclear cells.⁵⁶ With the growing development of various cell therapies, the use of MNP-based cell tracking is expected to increase significantly in future clinical trials.

Thermal therapies

MNPs can convert electromagnetic energy into heat upon exposure to an AMF, a process commonly known as magnetic heating or magnetic hyperthermia in biomedical research. This ability enables heating of tissue deep within the body, as the magnetic field can penetrate the biological tissue with minimal adsorption and scattering. During magnetic heating, MNPs distributed within the tissue generate heat, which dissipates into the surrounding tissues. Since the biological tissue is a poor conductor of heat, regions containing MNPs can be heated to high temperatures, such as 50 °C, without significantly affecting nearby tissues.

A further advantage of magnetic heating is the distribution of MNPs in the body can be visualized with MRI. This enables precise thermal ablation of target tissue when combined with MRI-guided infusion. A key application of this technique is the treatment of brain tumors, which present therapeutic challenges due to their critical anatomical location and the presence of the blood brain barrier. Clinical studies have shown that MNP thermotherapy, in conjunction with reduced radiation doses, is a safe treatment for glioblastoma multiforme.⁵⁷ This combination therapy has achieved a median survival rate of 23.2 months, compared to 14.6 months in the reference group.^{57,58} This treatment has been approved for clinical use by EMA and is currently undergoing clinical trials for treating prostate cancer.

In summary, clinical studies enhance conventional imaging, targeted drug delivery, and thermal therapy by leveraging the

unique magnetic properties of MNPs to provide high imaging contrast, advanced targeting capabilities, and deep heating potential (Fig. 2). While much of the current research focuses on these areas, the following sections will highlight innovative applications that could shape the future of medicine.

Applications of magneto-mechanical force

MNPs, as nanosized magnets, experience a force in a nonuniform magnetic field along its gradient. The magnetic force (\mathbf{F}) is determined by both the magnetic moment (\mathbf{m}) of MNPs and the field gradient ($\nabla\mathbf{H}$) according to the following equation,

$$\mathbf{F} = \mu(\mathbf{m} \cdot \nabla)\mathbf{H} \quad (1)$$

where μ represents the magnetic permeability of the tissue. With a field gradient of 10 T m^{-1} , the magnetic force ranges from approximately 2 atto-Newton for a 10 nm magnetite nanocrystal to 2 femto-Newton as the size increases to 100 nm. In comparison, the shear force exerted on endothelial cells in microcirculation is on the order of hundreds of pico-Newton.⁵⁹ Additionally, the engagement of T cell receptors and agonist peptide–MHC complexes involves forces of approximately 10 pico-Newton.⁶⁰ Achieving biologically relevant benchmarks for magneto-mechanical applications often necessitates the use of high-gradient magnetic fields and large clusters of magnetic nanoparticles.

Most force-related research are conducted using small permanent magnets in superficial tissues of small rodent models. A notable advancement in this field is the magnetic particle imaging (MPI) instrument, designed to produce images of MNPs.⁶¹ The MPI instrument can generate a field gradient on the order of 10 T m^{-1} , facilitating magnetic navigation of MNPs

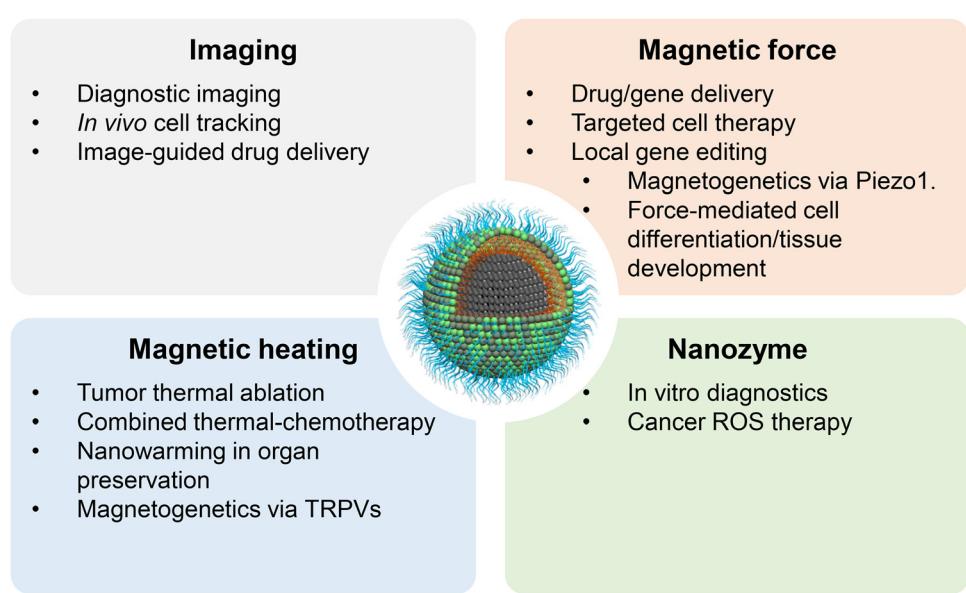


Fig. 2 Applications of magnetic nanoparticles. Applications of MNPs can be broadly categorized into four groups based on the physical or chemical properties emphasized.



within the body. A human brain-sized MPI instrument has been developed in the lab.⁶² However, the size and cost of the MPI instrument are comparable to an MRI machine, posing a considerable challenge to translational research.

Depending on its magnitude, the magnetic force can be used to direct circulating MNPs to specific tissues for targeted delivery, promote molecular interactions, or trigger biological responses by influencing cellular mechanosensors. These mechanisms have found impactful applications in the fields of drug/gene delivery, immunotherapy, regenerative medicine, gene editing, and neuroscience.

Magnetic force-mediated cellular/molecular interactions

The ability of MNPs to migrate or form aggregates in response to an external magnetic field can be utilized to control the distribution of biomolecules on the cell surface or within subcellular compartment. For instance, Etoc *et al.* created MNPs functionalized with canonical small Rho-GTPases, which regulate cell morphology through complex spatiotemporal activity patterns.⁶³ After injecting MNPs into the cells, a magnetic tip was used to displace them, triggering local remodeling of the actin cytoskeleton and morphological changes in the cells. Similarly, Li *et al.* demonstrated that fluorescent MNPs bound to molecules on the cell surface could be laterally pulled using a magnetic needle, offering a versatile approach for probing membrane processes.⁶⁴

T cell intratumoral infiltration, recognition of cancer cells, and activation are critical components of effective cancer immunotherapy. T cell activation depends not only on individual T cell receptor (TCR)-major histocompatibility complex (MHC) interactions but also on TCR clustering, which increases avidity and, consequently, sensitivity to cancer antigens. In this context, Perica *et al.* developed artificial antigen-presenting cells (nano-aAPCs) by conjugating MHC-Ig dimers and anti-CD28 antibodies to 50–100 nm iron-dextran nanoparticles.⁶⁵ After binding to the TCR, these nano-aAPCs could aggregate under a magnetic field, leading to larger TCR clusters and an increased number of activated antigen-specific T cells in a mouse melanoma model. In another approach, similar MNPs were conjugated with an immune checkpoint inhibitor (anti-PD-L1) and a T cell activator (anti-CD3).⁶⁶ The study showed that, following magnetic targeting, the concentration of MNPs in the tumor reached over 15% ID g⁻¹ at 24 hours post-injection – three times higher than in the reference group. The two surface ligands could bind to cancer cells and T cells, respectively, acting as a linker to enhance cell-to-cell interactions. Importantly, magnetic targeting improved the tumor selectivity of T cells, leading to prolonged survival while reducing the incidence of adverse events.

The past decade has seen rapid advancements in CRISPR gene editing. CRISPR employs Cas nucleases and guide RNAs (gRNAs) to modify specific genomic loci, rendering it a versatile tool for gene modification or regulation.^{12,13} Nonetheless, gRNAs can bind to DNA sequences with base mismatches, and Cas nucleases can tolerate base mismatches and DNA and RNA bulges, leading to off-target activities.⁶⁷ Similar to cancer

chemotherapy, *in vivo* therapeutic gene editing requires targeted delivery of the CRISPR system to enhance editing efficiency while minimizing genotoxicity due to off-target effects.⁶⁸ To address this challenge, we developed a novel gene delivery platform by modifying a baculoviral vector (BV) with MNPs conjugated to TAT peptides (Fig. 3).³⁵ BV, derived from an insect virus, offers good biocompatibility, and can effectively transduce a wide range of mammalian cells. However, its *in vivo* applications have been limited due to rapid inactivation by the complement system. Our study demonstrated that MNP-BV can be locally activated with a magnetic field, which enhances its entry into the cells and allows it to escape from complement inactivation through separation. The magnetic force also promotes actin filament polymerization, improving the localization of the viral genome to the cell nucleus. Meanwhile, MNP-BV outside the magnetic field is inactivated by the complement system. This dual mechanism of magnetic targeting and complement inactivation provides an on/off switch for MNP-BV, enabling spatially controlled gene editing.

Magnetic force-induced mechanosensing

The ability to detect and respond to mechanical cues in the surrounding environment is a fundamental function of mammalian cells.⁶⁹ Traditionally, research in this field has focused on cellular responses to fluid flow, mechanical stretching, and substrate stiffness. However, the versatility of conjugation chemistry enables MNPs to be functionalized for binding to cell surface proteins or accumulating in intracellular compartments, allowing for precise mechanical stimulation of specific mechanosensors or cytoskeletal components. Fernández-Sánchez *et al.* used an implanted magnet to exert magnetic force on MNP-loaded lysosomes in mesenchymal cells within the connective tissue surrounding colonic crypts.⁷⁰ This magnetic force mimicked the endogenous stress of early tumor growth without altering tissue stiffness, revealing previously unexplored mechanisms of tumor propagation based on β-catenin signaling pathways. Similarly, Abdel Fattah *et al.* created human neural organoids with MNP-labeled human pluripotent stem cells, where spatially defined magnetic force provides short-term mechanical tissue perturbations and long-term cytoskeleton remodeling in the organoids.⁷¹ The study demonstrates improved patterning in human neural tube organoids through precise, spatially defined MNP-driven actuation.

In our own study, we demonstrated that MNPs accumulated in endothelial cells could induce the relocation of lysosomes and the formation of actin filaments along the field gradient.^{59,72} Notably, with nearly 5.8×10^5 MNPs accumulated within each endothelial cell, the intracellular magnetic force reached 200 pN per cell.⁵⁹ The alteration of actin filaments further led to a reversible increase in vascular permeability by affecting the adherence junctions between endothelial cells, potentially providing a magnetic switch to enhance the extravasation of therapeutic agents. More recently, we also found that oscillating magnetic force applied to intracellular MNPs could stimulate several mechanosensing pathways associated with exosome biogenesis and release.⁷³ The cells experiencing



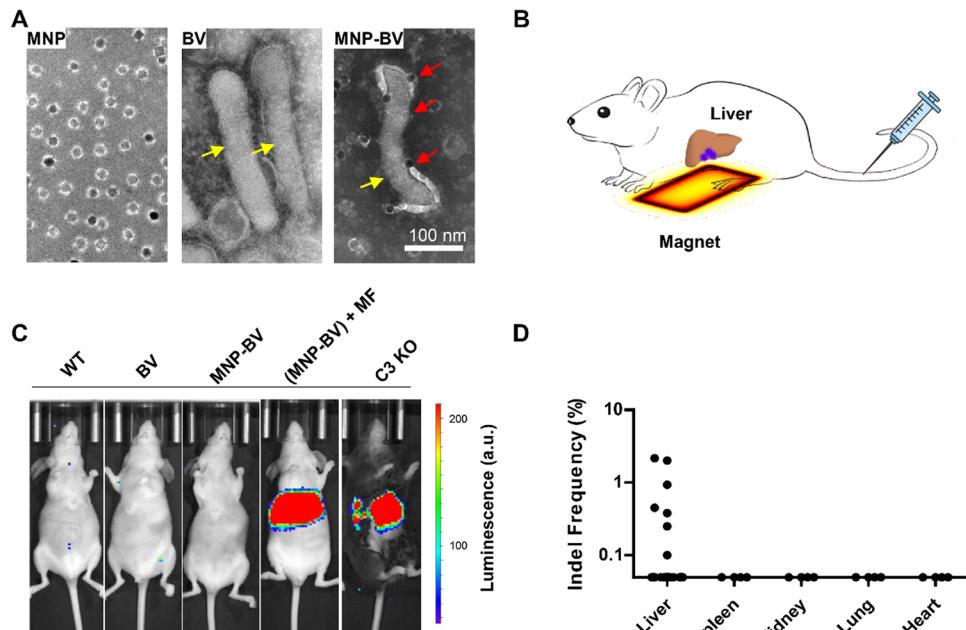


Fig. 3 Spatial control of *in vivo* gene editing. Baculoviral vectors (BV) labeled with MNPs facilitate localized gene editing through the combined effects of magnetic activation and complement inhibition. (A) TEM images of MNPs, BV, and MNP-BV hybrids. (B) Schematic illustration of MNP-BV-mediated transgene delivery to the mouse liver. (C) Bioluminescence analysis of transgene expression: mice injected with MNP-BV-LUC and exposed to a magnetic field exhibited strong luminescence in the liver. In contrast, no luminescence was observed in mice treated with BV-LUC alone or with MNP-BV-LUC without magnetic field application. C3 knockout mice served as controls. (D) Assessment of gene editing in vital organs. (Reproduced with permission from ref. 35, ©2018 Springer Nature).

intracellular magnetic force released more exosomes and exhibited increased intercellular communication compared with untreated cells.

MNPs can also trigger mechanosensing by interacting with cell surface protein sensors. Optogenetics, a widely used method for investigating brain functions, controls neuronal activity through light.⁷⁴ Central to this technology are light-sensitive ion channels, which can be activated by light to initiate action potentials in targeted neurons. However, the limited penetration of light in biological tissues necessitates the implantation of optical fibers through the skull to deliver light signals to cortical neurons. This invasive procedure can disrupt normal behavior and physiology. In contrast, a recent study introduced a non-contact neuron activation technique utilizing MNPs and mechanosensitive ion channels, such as Piezo1 (Fig. 4).^{29,30} By assembling 25 nm MNPs into 200 nm clusters, the researchers generated mechanical force of 2–10 pN force at a magnetic flux density of 20–50 mT. The study further employed a circular magnet array with an effective working range of approximately 70 cm, enabling consistent and reproducible neuromodulation in mice. This innovative approach holds potential for wireless, long-term studies of freely moving large animals.

Applications of magnetic heating

Magnetic heating arises from the relaxation of MNPs within an AMF. In biological media, this involves both Néel and Brownian

relaxation.⁷⁵ Néel relaxation refers to the internal reorientation of the magnetic domain constrained by the magnetic anisotropy energy barrier, while Brownian relaxation involves the rotation of the particle in the surrounding medium. The heating efficiency of MNPs is quantified by the specific loss power (SLP), also known as specific absorption rate (SAR), which represents the thermal energy generated per unit mass of MNPs. The SLP is influenced not only by the magnetic properties of the MNPs but also by the frequency (*f*), field strength (*H*), and wave form of the AMF. An increase in the frequency and field strength of the AMF leads to higher SLP. However, a safety limit of $H \times f < 4.85 \times 10^8 \text{ A m}^{-1} \text{ s}^{-1}$ for the whole body is imposed to prevent eddy currents in bodily fluids during clinical applications.⁷⁶ This restriction may be somewhat relaxed in the treatment of local diseases or in preclinical and *in vitro* studies.⁷⁷

Unlike the magneto-mechanical effect, there is still no analytical model that accurately predicts SLP during magnetic heating. One widely cited model of magnetic heating is the linear response theory (LRT).⁷⁸ According to LRT, under the assumptions that superparamagnetic MNPs have a uniaxial crystalline structure, a single magnetic domain, and a magnetic susceptibility linearly proportional to the inductive field, the SLP is given by,

$$\text{SLP} = \pi \mu_0 \chi_0 H^2 f \frac{2\pi f \tau}{1 + (2\pi f \tau)^2} \quad (2)$$

where μ_0 is the permeability of vacuum, χ_0 is the equilibrium susceptibility, and τ is the relaxation time.⁷⁸ The LRT predicts that

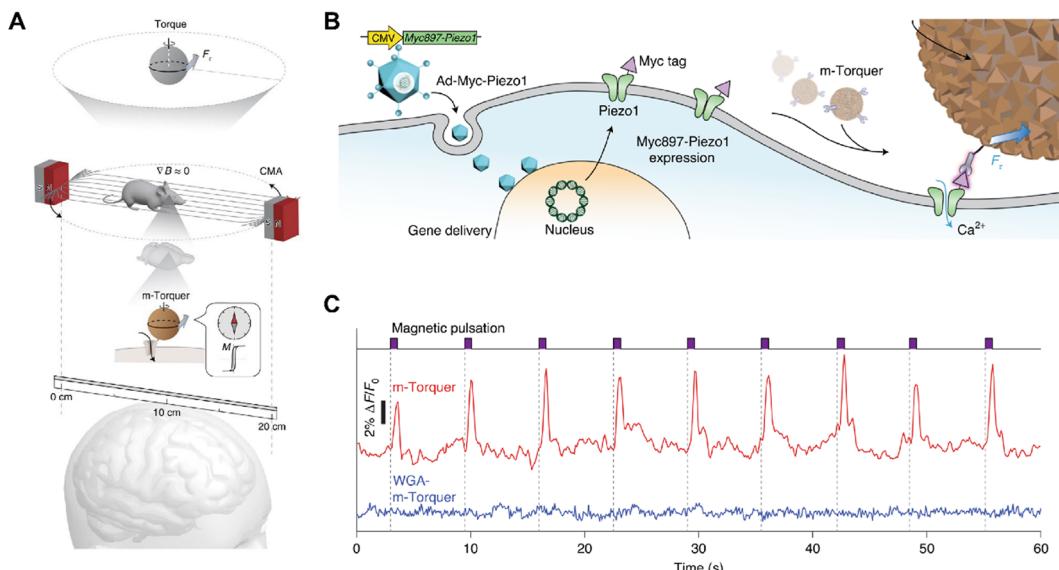


Fig. 4 Magnetic force-induced neuron activation. (A) Design of the nanoscale magnetic torquer system. The m-Torquer system acts as a nanoscale magnetic compass to generate the F_t needed for magnetomechanical signal transduction. (B) Schematic of genetic encoding of Piezo1 and its magnetomechanical gating with specifically targeted m-Torquer with Myc antibody. (C) Temporal repetitive stimulation of Piezo1 with m-Torquer-treated (red) neurons. WGA-m-Torquer-treated (blue) neurons were used as controls. (Reproduced with permission from ref. 30, ©2021 Springer Nature).

SLP reaches its maximum when the relaxation time matches the period of the AMF. Consequently, the optimal size for magnetic iron oxide nanoparticles is predicted to be approximately 14 nm when the frequency is 300 kHz. While some experimental results align with this prediction, our study, along with others, has found that SLR increases monotonically with particle size (Fig. 5).^{25,79,80} The discrepancy among experimental studies is likely due to variations in MNP dispersity and composition. It is important to note that many instruments used in these studies are not specifically designed for magnetic heating and are unable to produce a monochromatic AMF. Several new models of magnetic

heating have been proposed that incorporate more complex relaxation processes and realistic anisotropy energy profile.^{81–83} However, further experimental validation is needed to confirm these models' predictions.

The heat generation of MNPs is also known to depend on particle shape and composition. Experimental studies have demonstrated that SLP of cubic MNPs is twice that of spherical MNPs.^{84,85} Nano-octopods and nanorods have similarly shown higher SLP compared to spherical particles.^{86,87} Our study has further revealed that simply oxidizing magnetite nanoparticles to maghemite can more than double the SLP, likely due to an increase in magnetic anisotropy.⁸⁸ Additionally, tuning of MNP composition by incorporating metallic ions such as Mn^{2+} and Zn^{2+} has been shown to enhance the SLP as well.^{28,84}

The efficiency of magnetic heating typically falls within the range of hundreds of W g^{-1} MNP, which is considerably lower than other clinical heating techniques.²⁵ However, the use of radio frequency AMF enables deep tissue penetration with minimal absorption and scattering, making magnetic heating particularly valuable for thermal treatment of areas like the brain. Efficient heating traditionally requires a high concentration of MNPs, achievable only through local infusion. For instance, in the clinical application of NanoTherm, an MNP solution was infused into a brain tumor, achieving a final concentration of approximately 30 mg mL^{-1} within the tumor tissue.⁵⁷ Due to the low heating efficiency of magnetic heating, a long-standing debate exists around nano-scale heating effects. Some argue that magnetic heating results in a relatively smooth temperature increase across the affected tissue, as predicted by classical Fourier heat transfer theory.^{42,89} In contrast, others propose that individual MNPs can heat nearby molecules

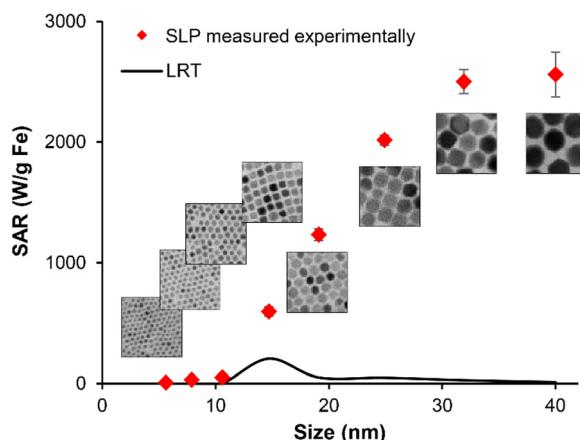


Fig. 5 Size-dependent heating of MNPs. SAR was measured for MNPs ranging from 6 to 40 nm dispersed in DI water, using an AMF at 325 kHz and 20.7 kA m^{-1} . The solid curve represents the theoretical SAR predictions according to the LRT model. (Reproduced with permission from ref. 25, ©2017 American Chemical Society).



without substantially increasing the surrounding medium's temperature, a concept linked to the emerging field of magnetogenetics.^{34,36–39}

To date, most research on magnetic heating has focused on cancer treatment. Beyond thermal ablation, magnetic heating has been extensively studied for tumor hyperthermia, which involves raising tissue temperature to approximate 41–45 °C.⁹⁰ Studies have shown that cells in the mitosis stage are more heat-sensitive, allowing hyperthermia to specifically target cancer cells due to their elevated proliferation rate.^{91,92} Consequently, magnetic hyperthermia has been combined with treatments such as chemotherapy and radiation. For example, in a study using MNPs loaded with doxorubicin, we observed a synergistic effect between mild hyperthermia at 43 °C and doxorubicin in reducing HeLa cell viability.⁹³ More recently, the development of high heating MNPs has opened up new applications for magnetic heating, particular in two emerging fields: organ cryopreservation and magnetogenetics.

Magnetic heating in organ cryopreservation

Organ transplantation is the last therapeutic option for patients with otherwise incurable diseases. However, a major public health challenge is the shortage of organs, primarily due to the lack of long-term organ preservation methods.^{94,95} Current organ preservation techniques can only maintain most organs

for 4–12 hours, creating significant stress for transportation, immune crossmatching, and surgical planning. For example, approximately 70% of donated hearts are discarded during processing.⁹⁴ Recently, cryopreservation *via* vitrification has emerged as a promising technology for long-term organ storage. This method preserves organs in a glass-like state to prevent structural damage and loss of cell viability caused by ice crystal formation.^{96–98} However, when these preserved organs are warmed to body temperature for transplantation, recrystallization in the cryoprotectant can occur if the warming rate is slower than the critical warming rate.^{99,100} Therefore, convective heating, which warms the organ from the surface, faces a critical size limitation of 1–3 mL due to slow heat conduction in the tissue, making it unsuitable for cryopreserving human organs.

Magnetic heating has emerged as a viable option for tissue warming, as it can deliver heat uniformly throughout the tissue. The nanowarming process involves perfusing the donor organ with a cryoprotectant containing MNPs, rapidly freezing it and storing at an ultralow temperature (<–140 °C), warming the organ to body temperature, and replacing the cryoprotectant with a blood-compatible solution (Fig. 6).¹⁰¹ Typically, the cryoprotectant contains high concentrations of organic and inorganic antifreeze compounds. For example, the common cryoprotectant, VS55, is composed of 3.1 M dimethyl sulfoxide,

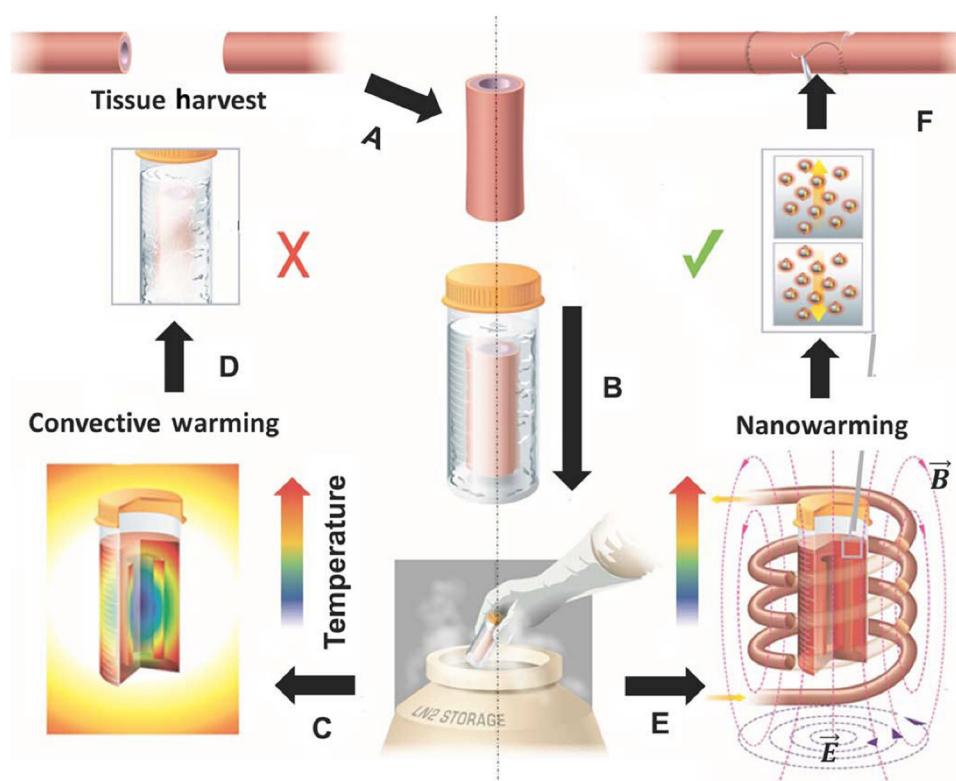


Fig. 6 Schematic illustrating tissue vitrification, convective warming, and nanowarming. (A) Tissues are harvested from a donor. (B) Tissues are loaded in a vial with CPA (VS55) and MNPs in a stepwise protocol, vitrified by standard convection, and stored at cryogenic temperatures. Warming by standard convection (C) leads to failure in larger 50-ml systems (D). Nanowarming in an alternating magnetic field, an inductive RF coil that stimulates nanoparticle heating (E), avoids warming failure and renders the tissue suitable for further testing or use (F). (Reproduced with permission from ref. 101, ©2017 The American Association for the Advancement of Science).



2.2 M propylene glycol, 3.1 M formamide, and 2.4 g L⁻¹ HEPES in EuroCollins solution. Ensuring MNP stability in the cryoprotectant during extreme temperature shifts is critical, as MNP aggregation can alter magnetic relaxation and decrease heating efficiency.¹⁰² Additionally, both the magnetic properties of MNPs and the medium viscosity vary considerably with temperature.¹⁰³

Several studies have demonstrated that the colloidal and thermal stability of MNPs in VS55 can be maintained through surface coating with resorcinol-formaldehyde resin or silica.¹⁰⁴⁻¹⁰⁷ Additionally, surface modification with poly(ethylene glycol) has been shown to reduce cellular interactions and thus cytotoxicity.^{106,107} Manuchehrabadi achieved nanowarming of porcine arteries and porcine aortic heart valve leaflet tissues using magnetic heating.¹⁰¹ The preserved tissues showed excellent viability with no change in biomechanical properties. Notably, uniform and rapid rewarming at a rate of >130 °C per minute was achieved in volumes up to 80 mL. A subsequent study demonstrated that vitrified rat kidneys recovered through nanowarming were suitable for transplantation and restored full renal function in nephrectomized recipients.¹⁰⁸ Although nanowarming technology is still in its infancy, these studies show great promise to transform the landscape of organ transplantation.

Magnetogenetics

Inspired by optogenetics, magnetogenetics utilizes MNPs under an AMF as magnetic switches to activate cellular functions. This emerging field holds great potential for remotely controlling individual neurons in the brain. In a pioneering study, Huang *et al.* demonstrated that synthetic ferrite nanoparticles, when bound to a temperature-sensitive ion channel (TRPV1), could activate this channel under a radio-frequency AMF.³⁹ Subsequently, similar effects were achieved using transient receptor potential (TRP) channels coupled with biogenic ferritin particles containing 5–12 nm iron oxide cores.¹⁰⁹ So far, studies using either synthetic or biogenic magnetic nanoparticles (MNPs) have shown the ability to remotely control brain activities in live mice (Fig. 7).^{36,38,110–112} Additionally, magnetic heating of MNPs near thermosensitive TRPA1-A channels, which regulate the “fruitless” driver in *Drosophila melanogaster*,

allows for controlled wing extension with sub-second response times.³⁴ When combined with synthetic genetic circuits, magnetogenetics has enabled applications in other areas, such as the remote regulation of glucose homeostasis and the targeted activation of engineered bacteria for cancer immunotherapy.^{109,113}

While experimental results support the feasibility of magnetogenetics, the underlying biophysical mechanisms remain unclear. It is generally assumed that the MNP-induced molecular activation is due to nanoscale heat confinement during magnetic nanoparticle heating, implying that the temperature on the surface of MNPs is significantly higher than that of the surrounding medium. As a result, the heat generated by MNPs under an AMF activates nearby TRP channels, leading to downstream cellular responses. However, analyses based on classical Fourier heat conduction theory and the properties of MNPs indicate that the temperature rise in MNPs under given conditions should be much lower than that required to activate TRPVs.^{40,42,89} Experimental attempts to measure the surface temperature of MNPs have yielded conflicting results, with both supporting and contradictory evidence.^{114–118} Moreover, recent publications have raised concerns that biogenic MNP-induced molecular activation may be artifactual.^{119–121}

In addition to magnetic heating, alternative hypotheses have been proposed to explain the mechanism of magnetogenetics, including the magnetic force/torque effect, magneto-caloric effects, and reactive oxygen species (ROS)-mediated molecular activation.^{41,122} It is important to note that the MNP-mediated cellular functions, such as the activation of TRPVs in neurons, can arise from sporadic and transient actions of individual MNPs.^{34,38,39,111} In contrast, magnetic nanoparticle heating is usually modeled as a classical equilibrium process. Additional evidence and theoretical analysis at the nanoscale are needed to provide a compelling explanation. Furthermore, it remains unknown what the working range of the nanoscale effect is. Since MNP-induced neural responses typically occur on the order of seconds, it is essential to explore the possibility of shortening the response time to milliseconds, which would be more meaningful for investigating neuronal functions. Despite ongoing debates and unclear mechanisms, magnetogenetics opens an exciting new frontier in neuroscience research. It offers

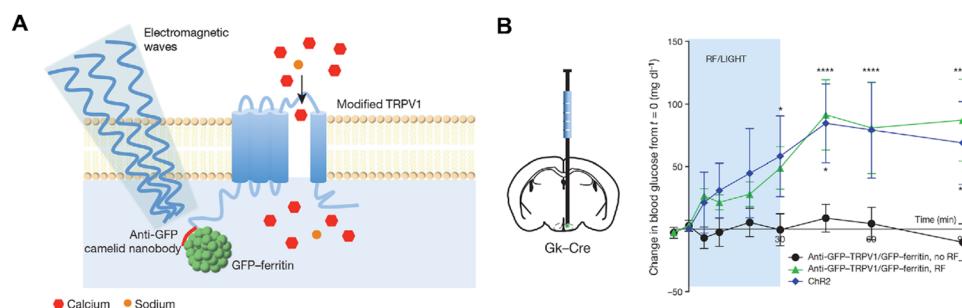


Fig. 7 Remote neural activation *in vivo* using radio waves. (A) Schematic of activation system. Ferritin-TRPV1 complexes were delivered to hypothalamic glucose-sensing neurons via viral transfection. (B) Treatment with a radio frequency AMF (465 kHz) elevated blood glucose levels in mice. (Reproduced with permission from ref. 38, © 2016 Springer Nature).



potential for brain–computer interfaces and introduces a novel class of therapeutic interventions by linking magnetic control to a range of physiological functions.

Functioning as nanozymes

Hydroxyl free radicals ($\cdot\text{OH}$) are highly reactive molecules capable of interacting with a wide range of biomolecules. Elevated levels of hydroxyl free radicals can damage cellular components such as DNA, proteins, and lipids, ultimately leading to cell death.^{123,124} Gao *et al.*'s pioneering work demonstrated that magnetite nanoparticles can catalyze the decomposition of hydrogen peroxide (H_2O_2) to hydroxyl free radicals through a heterogeneous Fenton reaction.⁴³ Since this discovery, MNPs have been extensively studied for their ability to induce cell polarization, apoptosis, and ferroptosis. Their functionalization potential for targeted and combination therapies gives them advantages over to small-molecule drugs.

The MNP-catalyzed Fenton reaction occurs on the surface of iron oxide nanocrystals. Thus, the reaction rate is influenced by the interface between the nanocrystal and its surrounding medium.^{125,126} Interestingly, our study revealed that the catalytic activity of MNPs is highly dependent on their composition, despite the tendency of the nanocrystals to oxidize and exhibit similar surface oxidative states (Fig. 8).⁴⁵ With comparable size, coating, and surface oxidation states, wüstite (FeO) nanoparticles demonstrated a 4.6 folds increase in catalytic activity compared to maghemite. Further analysis suggests that this increased catalytic activity is due to the presence of internal low-valence iron (Fe^0 and Fe^{2+}), which accelerates the recycling of surface Fe^{3+} back to Fe^{2+} *via* intraparticle electron transport.^{45,127}

Notably, the catalytic activity of MNPs is also pH-dependent. MNPs catalyze the production of hydroxyl free radicals only in acidic environments, such as endosomes or lysosomes, but act

as catalase, generating oxygen molecules instead of hydroxyl free radicals at neutral pH.^{128,129} This unique pH sensitivity enables targeting of specific cell populations based on cellular uptake and iron metabolism. For instance, Trujillo-Alonso *et al.* found that ferumoxytol exhibited cytotoxic effects on leukemia cells with low levels of ferroportin, a cellular iron exporter.¹³⁰ Moreover, Zanganeh *et al.* demonstrated that ferumoxytol could polarize macrophages toward a tumor-suppressing phenotype, inhibiting breast cancer growth and lung metastasis.⁴⁴

Research into the catalytic properties of MNPs also offers valuable insight into mitigating potential adverse effects in therapeutic applications. Wu *et al.* found that ultrasmall magnetite nanoparticles (2.3 nm) were highly toxic due to increased free radical production, whereas 9.3 nm magnetite nanoparticles showed no apparent toxicity in a mouse model.¹³¹ The size of MNPs can also influence the nature of cytotoxicity. Xie *et al.* discovered that 9 nm magnetite nanoparticles induced cellular mitochondrial dysfunction, while 14 nm magnetite nanoparticles primarily damaged cell membrane integrity and promoted lactate dehydrogenase leakage.¹³² Consequently, the catalytic activity of MNPs is a critical factor in designing MNP-based applications.

Outlook and future perspectives

Since the emergence of nanomedicine four decades ago, MNPs have become one of the most active and successful areas of research, driven by their exceptional biocompatibility and unique nanoscale magnetic properties. Clinical studies have primarily focused on imaging, thermal therapy, and magnetic targeting. While MNPs were initially approved for cancer medicine in the US and Europe, recent clinical research has expanded globally, with notable efforts in countries like China exploring their applications in treating a broader range of diseases.

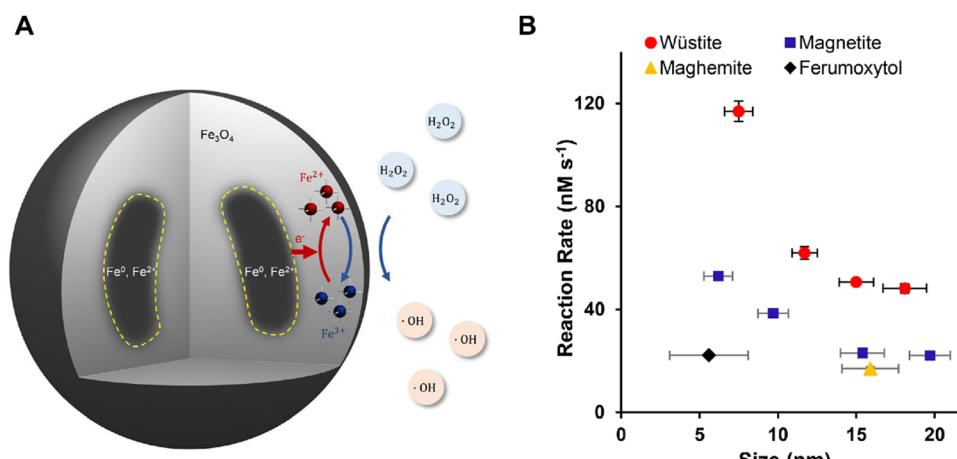


Fig. 8 MNP-mediated heterogeneous Fenton reaction. (A) Schematic of enhancing surface Fenton reaction *via* intraparticle electron transport. In a composite nanoparticle of magnetite (Fe_3O_4) shell and a core of low-valence iron (Fe^0 & Fe^{2+}), the low-valence iron can accelerate the catalysis by reducing the octahedral Fe^{3+} in the magnetite shell to Fe^{2+} through intraparticle electron transport. (B) Catalytic activities of MNPs of various compositions. (Reproduced with permission from ref. 45, ©2023 John Wiley and Sons).



A new frontier in MNP research is the controlled mechanical and thermal modulation of cells and tissues. Advances in synthesis techniques have enabled precise control over MNP size and composition, leading to the development of particles optimized for specific mechanical and thermal properties. In recent years, the use of MNPs to stimulate mechanosensitive receptors, ion channels, and other cellular components has opened novel avenues for manipulating cellular behavior. These research efforts are supported by a better understanding of mechano- and thermo-sensitive ion channels, such as the TRP and Piezo protein families, which play crucial roles in various aspects of cell biology. The significance of these proteins was recognized with the 2021 Nobel Prize.

The ability to remotely control cellular processes using MNPs offers exciting applications in regenerative medicine, tissue engineering, and neuromodulation. For instance, mechanical stimulation of stem cells *via* MNPs can influence differentiation pathways, while thermal activation of neurons through MNPs conjugated to thermosensitive ion channels has opened possibilities for non-invasive brain stimulation techniques. It is conceivable that MNP-derived nanotechnology will soon intersect with synthetic biology, forging new directions in biomedical applications. Remarkably, all these functions can be achieved using a single MNP, with its size, composition, and functionalization determining its specific properties. The inherent multifunctionality of MNPs thus provides a highly versatile platform for biomedical innovation.

Despite their promise, several challenges remain for the clinical translation of MNP applications in emerging fields. Over the past two decades, significant progress has been made in understanding the biocompatibility and safety of MNPs, particularly in cancer treatment clinical studies. However, as the MNP applications extend to vital organs in the context of non-terminal diseases, there is an urgent need to assess their biodistribution, clearance, and long-term toxicity. This is particularly important given the association between abnormal iron metabolism and diseases like Alzheimer's.¹³³ Additionally, regulatory considerations are crucial for the development of MNP functionalized with novel components such as targeting moieties and signaling molecules, as manufacturing practices, quality control, and reproducibility must meet stringent standards. The interdisciplinary nature of MNP-related research also presents challenges, especially since the magnetic properties of MNPs are heavily dependent on the applied magnetic field. Clinical translations will require the development of specialized electromagnetic instruments for human use. The widespread clinical use of MNPs in imaging is largely due to the availability of MRI machines in universities and major hospitals. Lastly, while the effects of magnetic fields on the human body have been extensively investigated, the impact of high-gradient, high-frequency magnetic fields on target organs needs to be carefully examined in new settings. The FDA has issued guidance for industry that provides a comprehensive summary of the major concerns associated with nanomaterials in clinical drug applications. Given the novelty of these research directions, it is likely that additional safety and ethical

considerations, beyond those encountered in conventional imaging and drug delivery studies, will emerge as the field continues to develop.

Looking forward, the continued convergence of nanotechnology, materials science, engineering, and biomedicine is likely to drive further innovations in MNP design and application. By harnessing their unique nanoscale magnetic properties and improving our understanding of their interactions with biological systems, MNPs are poised to revolutionize the landscape of nanomedicine and enable new approaches for disease treatment and management.

Data availability

No primary research results, software or code have been included and no new data were generated or analyzed as part of this review.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

This work was supported by the NIH/NHLBI funding (R21HL 166178 to ST), the NIH/NIAMS funding (R21AR078447 to ST), and the NSF funding (2342391 to ST).

References

- 1 P. H. Meyers, C. M. Nice Jr., G. R. Meckstroth, H. C. Becker, P. J. Moser and M. Goldstein, Pathologic studies following magnetic control of metallic iron particles in the lymphatic and vascular system of dogs as a contrast and isotopic agent, *Am. J. Roentgenol., Radium Ther. Nucl. Med.*, 1966, **96**, 913–921.
- 2 S. Tong, H. Zhu and G. Bao, Magnetic Iron Oxide Nanoparticles for Disease Detection and Therapy, *Mater. Today*, 2019, **31**, 86–99.
- 3 A. N. Kharlamov, A. E. Tyurnina, V. S. Veselova, O. P. Kovtun, V. Y. Shur and J. L. Gabinsky, Silica-gold nanoparticles for atheroprotective management of plaques: results of the NANOM-FIM trial, *Nanoscale*, 2015, **7**, 8003–8015.
- 4 A. N. Kharlamov, J. A. Feinstein, J. A. Cramer, J. A. Boothroyd, E. V. Shishkina and V. Shur, Plasmonic photothermal therapy of atherosclerosis with nanoparticles: long-term outcomes and safety in NANOM-FIM trial, *Future Cardiol.*, 2017, **13**, 345–363.
- 5 J. E. Jaetao, K. S. Butler, N. L. Adolphi, D. M. Lovato, H. C. Bryant, I. Rabinowitz, S. S. Winter, T. E. Tessier, H. J. Hathaway and C. Bergemann, *et al.*, Enhanced leukemia cell detection using a novel magnetic needle and nanoparticles, *Cancer Res.*, 2009, **69**, 8310–8316.
- 6 L. Charters, Magnetic cell therapy for corneal endothelial dysfunction, *Ophthalmol. Times Eur.*, 2024, **20a**, 34.
- 7 E. Senetierre, P. Taourel, Y. Bouvier, J. Pradel, B. Van Beers, J. P. Daures, J. Pringot, D. Mathieu and J. M. Bruel, Detection of hepatic metastases: ferumoxides-enhanced MR imaging versus unenhanced MR imaging and CT during arterial portography, *Radiology*, 1996, **200**, 785–792.
- 8 E. H. Arntzen, M. Srinivas, F. Bonetto, L. J. Cruz, P. Verdijk, G. Schreibelt, M. van de Rakt, W. J. Lesterhuis, M. van Riel and C. J. Punt, *et al.*, Targeting of ¹¹¹In-labeled dendritic cell human vaccines improved by reducing number of cells, *Clin. Cancer Res.*, 2013, **19**, 1525–1533.
- 9 D. Karassis, C. Karageorgiou, A. Vaknin-Dembinsky, B. Gowda-Kurkalli, J. M. Gomori, I. Kassis, J. W. Bulte, P. Petrou, T. Ben-Hur

and O. Abramsky, *et al.*, Safety and immunological effects of mesenchymal stem cell transplantation in patients with multiple sclerosis and amyotrophic lateral sclerosis, *Arch. Neurol.*, 2010, **67**, 1187–1194.

10 F. D. Birkhäuser, U. E. Studer, J. M. Froehlich, M. Triantafyllou, L. J. Bains, G. Petralia, P. Vermathen, A. Fleischmann and H. C. Thoeny, Combined ultrasmall superparamagnetic particles of iron oxide-enhanced and diffusion-weighted magnetic resonance imaging facilitates detection of metastases in normal-sized pelvic lymph nodes of patients with bladder and prostate cancer, *Eur. Urol.*, 2013, **64**, 953–960.

11 M. Triantafyllou, U. E. Studer, F. D. Birkhäuser, A. Fleischmann, L. J. Bains, G. Petralia, A. Christe, J. M. Froehlich and H. C. Thoeny, Ultrasmall superparamagnetic particles of iron oxide allow for the detection of metastases in normal sized pelvic lymph nodes of patients with bladder and/or prostate cancer, *Eur. J. Cancer*, 2013, **49**, 616–624.

12 J. M. Richards, S. I. Semple, T. J. MacGillivray, C. Gray, J. P. Langrish, M. Williams, M. Dweck, W. Wallace, G. McKillop and R. T. Chalmers, *et al.*, Abdominal aortic aneurysm growth predicted by uptake of ultrasmall superparamagnetic particles of iron oxide: a pilot study, *Circ. Cardiovasc. Imaging*, 2011, **4**, 274–281.

13 J. M. Richards, C. A. Shaw, N. N. Lang, M. C. Williams, S. I. Semple, T. J. MacGillivray, C. Gray, J. H. Crawford, S. R. Alam and A. P. Atkinson, *et al.*, In vivo mononuclear cell tracking using superparamagnetic particles of iron oxide: feasibility and safety in humans, *Circ. Cardiovasc. Imaging*, 2012, **5**, 509–517.

14 S. R. Alam, A. S. Shah, J. Richards, N. N. Lang, G. Barnes, N. Joshi, T. MacGillivray, G. McKillop, S. Mirsadraee and J. Payne, *et al.*, Ultrasmall superparamagnetic particles of iron oxide in patients with acute myocardial infarction: early clinical experience, *Circ. Cardiovasc. Imaging*, 2012, **5**, 559–565.

15 R. Baumgartner, W. Cho, A. Coimbra, C. Chen, Z. Wang, A. Struyk, N. Venketasubramanian, M. Low, C. Gargano and F. Zhao, *et al.*, Evaluation of an fMRI USPIO-based assay in healthy human volunteers, *J. Magn. Reson. Imaging*, 2017, **46**, 124–133.

16 R. Weissleder, A. Bogdanov, E. A. Neuwelt and M. Papisov, Long-circulating iron oxides for MR imaging, *Adv. Drug Delivery Rev.*, 1995, **16**, 321–334.

17 R. Weissleder, G. Elizondo, J. Wittenberg, C. A. Rabito, H. H. Bengele and L. Josephson, Ultrasmall superparamagnetic iron oxide: characterization of a new class of contrast agents for MR imaging, *Radiology*, 1990, **175**, 489–493.

18 S. Sun, H. Zeng, D. B. Robinson, S. Raoux, P. M. Rice, S. X. Wang and G. Li, Monodisperse MFe_2O_4 ($\text{M} = \text{Fe, Co, Mn}$) nanoparticles, *J. Am. Chem. Soc.*, 2004, **126**, 273–279.

19 J. Park, K. An, Y. Hwang, J. G. Park, H. J. Noh, J. Y. Kim, J. H. Park, N. M. Hwang and T. Hyeon, Ultra-large-scale syntheses of monodisperse nanocrystals, *Nat. Mater.*, 2004, **3**, 891–895.

20 A. Demortiere, P. Panissod, B. P. Pichon, G. Pourroy, D. Guillon, B. Donnio and S. Begin-Colin, Size-dependent properties of magnetic iron oxide nanocrystals, *Nanoscale*, 2011, **3**, 225–232.

21 J. H. Lee, Y. M. Huh, Y. W. Jun, J. W. Seo, J. T. Jang, H. T. Song, S. Kim, E. J. Cho, H. G. Yoon and J. S. Suh, *et al.*, Artificially engineered magnetic nanoparticles for ultra-sensitive molecular imaging, *Nat. Med.*, 2007, **13**, 95–99.

22 D. Kim, N. Lee, M. Park, B. H. Kim, K. An and T. Hyeon, Synthesis of uniform ferrimagnetic magnetite nanocubes, *J. Am. Chem. Soc.*, 2009, **131**, 454–455.

23 L. M. Bronstein, J. E. Atkinson, A. G. Malyutin, F. Kidwai, B. D. Stein, D. G. Morgan, J. M. Perry and J. A. Karty, Nanoparticles by Decomposition of Long Chain Iron Carboxylates: From Spheres to Stars and Cubes, *Langmuir*, 2011, **27**, 3044–3050.

24 H. Y. Sun, B. Chen, X. L. Jiao, Z. Jiang, Z. H. Qin and D. R. Chen, Solvothermal Synthesis of Tunable Electroactive Magnetite Nanorods by Controlling the Side Reaction, *J. Phys. Chem. C*, 2012, **116**, 5476–5481.

25 S. Tong, C. A. Quinto, L. Zhang, P. Mohindra and G. Bao, Size-Dependent Heating of Magnetic Iron Oxide Nanoparticles, *ACS Nano*, 2017, **11**, 6808–6816.

26 Y. Hou, Z. Xu and S. Sun, Controlled synthesis and chemical conversions of FeO nanoparticles, *Angew. Chem., Int. Ed.*, 2007, **46**, 6329–6332.

27 R. Chen, M. G. Christiansen and P. Anikeeva, Maximizing hysteric losses in magnetic ferrite nanoparticles *via* model-driven synthesis and materials optimization, *ACS Nano*, 2013, **7**, 8990–9000.

28 J. H. Lee, J. T. Jang, J. S. Choi, S. H. Moon, S. H. Noh, J. W. Kim, J. G. Kim, I. S. Kim, K. I. Park and J. Cheon, Exchange-coupled magnetic nanoparticles for efficient heat induction, *Nat. Nanotechnol.*, 2011, **6**, 418–422.

29 S. H. Choi, J. Shin, C. Park, J. U. Lee, J. Lee, Y. Ambo, W. Shin, R. Yu, J. Y. Kim and J. D. Lah, *et al.*, In vivo magnetogenetics for cell-type-specific targeting and modulation of brain circuits, *Nat. Nanotechnol.*, 2024, **19**, 1333–1343.

30 J. U. Lee, W. Shin, Y. Lim, J. Kim, W. R. Kim, H. Kim, J. H. Lee and J. Cheon, Non-contact long-range magnetic stimulation of mechanosensitive ion channels in freely moving animals, *Nat. Mater.*, 2021, **20**, 1029–1036.

31 R. J. Mannix, S. Kumar, F. Cassiola, M. Montoya-Zavala, E. Feinstein, M. Prentiss and D. E. Ingber, Nanomagnetic actuation of receptor-mediated signal transduction, *Nat. Nanotechnol.*, 2008, **3**, 36–40.

32 S. Laurent, D. Forge, M. Port, A. Roch, C. Robic, L. Vander Elst and R. N. Muller, Magnetic iron oxide nanoparticles: synthesis, stabilization, vectorization, physicochemical characterizations, and biological applications, *Chem. Rev.*, 2008, **108**, 2064–2110.

33 Y. W. Jun, J. W. Seo and J. Cheon, Nanoscaling laws of magnetic nanoparticles and their applicabilities in biomedical sciences, *Acc. Chem. Res.*, 2008, **41**, 179–189.

34 C. Sebesta, D. Torres Hinojosa, B. Wang, J. Asfouri, Z. Li, G. Duret, K. Jiang, Z. Xiao, L. Zhang and Q. Zhang, *et al.*, Subsecond multi-channel magnetic control of select neural circuits in freely moving flies, *Nat. Mater.*, 2022, **21**, 951–958.

35 H. Zhu, L. Zhang, S. Tong, C. M. Lee, H. Deshmukh and G. Bao, Spatial control of in vivo CRISPR-Cas9 genome editing *via* nanomagnets, *Nat. Biomed. Eng.*, 2019, **3**, 126–136.

36 S. A. Stanley, J. E. Gagner, S. Damantpour, M. Yoshida, J. S. Dordick and J. M. Friedman, Radio-wave heating of iron oxide nanoparticles can regulate plasma glucose in mice, *Science*, 2012, **336**, 604–608.

37 R. Chen, G. Romero, M. G. Christiansen, A. Mohr and P. Anikeeva, Wireless magnetothermal deep brain stimulation, *Science*, 2015, **347**, 1477–1480.

38 S. A. Stanley, L. Kelly, K. N. Latcha, S. F. Schmidt, X. Yu, A. R. Nectow, J. Sauer, J. P. Dyke, J. S. Dordick and J. M. Friedman, Bidirectional electromagnetic control of the hypothalamus regulates feeding and metabolism, *Nature*, 2016, **531**, 647–650.

39 H. Huang, S. Delikanli, H. Zeng, D. M. Ferkey and A. Pralle, Remote control of ion channels and neurons through magnetic-field heating of nanoparticles, *Nat. Nanotechnol.*, 2010, **5**, 602–606.

40 P. Anikeeva and A. Jasanoff, Problems on the back of an envelope, *eLife*, 2016, **5**, e19569.

41 M. Barbic, Possible magneto-mechanical and magneto-thermal mechanisms of ion channel activation in magnetogenetics, *eLife*, 2019, **8**, e45807.

42 M. Meister, Physical limits to magnetogenetics, *eLife*, 2016, **5**, e17210.

43 L. Gao, J. Zhuang, L. Nie, J. Zhang, Y. Zhang, N. Gu, T. Wang, J. Feng, D. Yang and S. Perrett, *et al.*, Intrinsic peroxidase-like activity of ferromagnetic nanoparticles, *Nat. Nanotechnol.*, 2007, **2**, 577–583.

44 S. Zanganeh, G. Hutter, R. Spitzer, O. Lenkov, M. Mahmoudi, A. Shaw, J. S. Pajarin, H. Nejadnik, S. Goodman and M. Moseley, *et al.*, Iron oxide nanoparticles inhibit tumour growth by inducing pro-inflammatory macrophage polarization in tumour tissues, *Nat. Nanotechnol.*, 2016, **11**, 986–994.

45 Z. Yi, X. Yang, Y. Liang, F. Chapelin and S. Tong, Enhancing ROS-Inducing Nanozyme through Intraparticle Electron Transport, *Small*, 2024, **20**, e2305974.

46 R. Weissleder, D. D. Stark, B. L. Engelstad, B. R. Bacon, C. C. Compton, D. L. White, P. Jacobs and J. Lewis, Superparamagnetic iron oxide: pharmacokinetics and toxicity, *AJR, Am. J. Roentgenol.*, 1989, **152**, 167–173.

47 A. Kunzmann, B. Andersson, T. Thurnherr, H. Krug, A. Scheynius and B. Fadeel, Toxicology of engineered nanomaterials: focus on biocompatibility, biodistribution and biodegradation, *Biochim. Biophys. Acta*, 2011, **1810**, 361–373.

48 C. Alexiou, W. Arnold, R. J. Klein, F. G. Parak, P. Hulin, C. Bergemann, W. Erhardt, S. Wagenpfeil and A. S. Lubbe,



Locoregional cancer treatment with magnetic drug targeting, *Cancer Res.*, 2000, **60**, 6641–6648.

49 M. W. Wilson, R. K. Kerlan Jr., N. A. Fidelman, A. P. Venook, J. M. LaBerge, J. Koda and R. L. Gordon, Hepatocellular carcinoma: regional therapy with a magnetic targeted carrier bound to doxorubicin in a dual MR imaging/conventional angiography suite—initial experience with four patients, *Radiology*, 2004, **230**, 287–293.

50 F. Yuan, A. Krol and S. Tong, Available space and extracellular transport of macromolecules: effects of pore size and connectedness, *Ann. Biomed. Eng.*, 2001, **29**, 1150–1158.

51 M. R. Dreher, W. Liu, C. R. Michelich, M. W. Dewhirst, F. Yuan and A. Chilkoti, Tumor vascular permeability, accumulation, and penetration of macromolecular drug carriers, *J. Natl. Cancer Inst.*, 2006, **98**, 335–344.

52 E. Seneterre, P. Taourel, Y. Bouvier, J. Pradel, B. Van Beers, J. P. Dauvès, J. Pringot, D. Mathieu and J. M. Bruel, Detection of hepatic metastases: ferumoxides-enhanced MR imaging versus unenhanced MR imaging and CT during arterial portography, *Radiology*, 1996, **200**, 785–792.

53 M. G. Harisinghani, J. Barentsz, P. F. Hahn, W. M. Deserno, S. Tabatabaei, C. H. van de Kaa, J. de la Rosette and R. Weissleder, Noninvasive detection of clinically occult lymph-node metastases in prostate cancer, *N. Engl. J. Med.*, 2003, **348**, 2491–2499.

54 L. Zhang, S. Hajebrahimi, S. Tong, X. Gao, H. Cheng, Q. Zhang, D. T. Hinojosa, K. Jiang, L. Hong and J. Huard, *et al.*, Force-Mediated Endocytosis of Iron Oxide Nanoparticles for Magnetic Targeting of Stem Cells, *ACS Appl. Mater. Interfaces*, 2023, **15**, 50574–50585.

55 N. Landazuri, S. Tong, J. Suo, G. Joseph, D. Weiss, D. J. Sutcliffe, D. P. Giddens, G. Bao and W. R. Taylor, Magnetic targeting of human mesenchymal stem cells with internalized superparamagnetic iron oxide nanoparticles, *Small*, 2013, **9**, 4017–4026.

56 I. J. de Vries, W. J. Lesterhuis, J. O. Barentsz, P. Verdijk, J. H. van Krieken, O. C. Boerman, W. J. Oyen, J. J. Bonenkamp, J. B. Boezeman and G. J. Adema, *et al.*, Magnetic resonance tracking of dendritic cells in melanoma patients for monitoring of cellular therapy, *Nat. Biotechnol.*, 2005, **23**, 1407–1413.

57 K. Maier-Hauff, F. Ulrich, D. Nestler, H. Niehoff, P. Wust, B. Thiesen, H. Orawa, V. Budach and A. Jordan, Efficacy and safety of intratumoral thermotherapy using magnetic iron-oxide nanoparticles combined with external beam radiotherapy on patients with recurrent glioblastoma multiforme, *J. Neurooncol.*, 2011, **103**, 317–324.

58 R. Stupp, W. P. Mason, M. J. van den Bent, M. Weller, B. Fisher, M. J. Taphoorn, K. Belanger, A. A. Brandes, C. Marosi and U. Bogdahn, *et al.*, Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma, *N. Engl. J. Med.*, 2005, **352**, 987–996.

59 Y. Qiu, S. Tong, L. Zhang, Y. Sakurai, D. R. Myers, L. Hong, W. A. Lam and G. Bao, Magnetic forces enable controlled drug delivery by disrupting endothelial cell–cell junctions, *Nat. Commun.*, 2017, **8**, 15594.

60 B. Liu, W. Chen, B. D. Evavold, D. Brian and C. Zhu, Accumulation of Dynamic Catch Bonds between TCR and Agonist Peptide-MHC Triggers T Cell Signaling, *Cell*, 2014, **157**, 357–368.

61 P. W. Goodwill, E. U. Saritas, L. R. Croft, T. N. Kim, K. M. Krishnan, D. V. Schaffer and S. M. Connolly, X-space MPI: magnetic nanoparticles for safe medical imaging, *Adv. Mater.*, 2012, **24**, 3870–3877.

62 M. Graeser, F. Thieben, P. Szwarcgulski, F. Werner, N. Gdaniec, M. Boberg, F. Giese, M. Moddel, P. Ludewig and D. van de Ven, *et al.*, Human-sized magnetic particle imaging for brain applications, *Nat. Commun.*, 2019, **10**, 1936.

63 F. Etoc, D. Lisse, Y. Bellaiche, J. Piehler, M. Coppey and M. Dahan, Subcellular control of Rac-GTPase signalling by magnetogenetic manipulation inside living cells, *Nat. Nanotechnol.*, 2013, **8**, 193–198.

64 J. H. Li, P. Santos-Otte, B. Au, J. Rentsch, S. Block and H. Ewers, Directed manipulation of membrane proteins by fluorescent magnetic nanoparticles, *Nat. Commun.*, 2020, **11**, 4259.

65 K. Perica, A. Tu, A. Richter, J. G. Bieler, M. Edidin and J. P. Schneck, Magnetic Field-Induced T Cell Receptor Clustering by Nanoparticles Enhances T Cell Activation and Stimulates Antitumor Activity, *ACS Nano*, 2014, **8**, 2252–2260.

66 C. S. Chiang, Y. J. Lin, R. Lee, Y. H. Lai, H. W. Cheng, C. H. Hsieh, W. C. Shyu and S. Y. Chen, Combination of fucoidan-based magnetic nanoparticles and immunomodulators enhances tumour-localized immunotherapy, *Nat. Nanotechnol.*, 2018, **13**, 746–754.

67 Y. N. Lin, T. J. Cradick, M. T. Brown, H. Deshmukh, P. Ranjan, N. Sarode, B. M. Wile, P. M. Vertino, F. J. Stewart and G. Bao, CRISPR/Cas9 systems have off-target activity with insertions or deletions between target DNA and guide RNA sequences, *Nucleic Acids Res.*, 2014, **42**, 7473–7485.

68 S. Tong, B. Moyo, C. M. Lee, K. Leong and G. Bao, Engineered materials for *in vivo* delivery of genome-editing machinery, *Nat. Rev. Mater.*, 2019, **4**, 726–737.

69 T. Iskratsch, H. Wolfenson and M. P. Sheetz, Appreciating force and shape—the rise of mechanotransduction in cell biology, *Nat. Rev. Mol. Cell Biol.*, 2014, **15**, 825–833.

70 M. E. Fernández-Sánchez, S. Barbier, J. Whitehead, G. Béalle, A. Michel, H. Latorre-Ossa, C. Rey, L. Fouassier, A. Clapéron and L. Brullé, *et al.*, Mechanical induction of the tumorigenic β -catenin pathway by tumour growth pressure, *Nature*, 2015, **523**, 92–95.

71 A. R. Abdel Fattah, N. Kolaitis, K. Van Daele, B. Daza, A. G. Rustandi and A. Ranga, Targeted mechanical stimulation *via* magnetic nanoparticles guides *in vitro* tissue development, *Nat. Commun.*, 2023, **14**, 5281.

72 H. Shen, S. Tong, G. Bao and B. Wang, Structural responses of cells to intracellular magnetic force induced by superparamagnetic iron oxide nanoparticles, *Phys. Chem. Chem. Phys.*, 2014, **16**, 1914–1920.

73 X. Yang, Z. Yi, Y. Liang and S. Tong, Magnetic Iron Oxide Nanoparticles Enhance Exosome Production by Upregulating Exosome Transport and Secretion Pathways, *ACS Appl. Mater. Interfaces*, 2024, **16**, 67235–67245.

74 V. Emiliani, E. Entcheva, R. Hedrich, P. Hegemann, K. R. Konrad, C. Luscher, M. Mahn, Z. H. Pan, R. R. Sims and J. Vierock, *et al.*, Optogenetics for light control of biological systems, *Nat. Rev. Methods Primers*, 2022, **2**, 55.

75 R. Kötitz, W. Weitschies, L. Trahms and W. Semmler, Investigation of Brownian and Néel relaxation in magnetic fluids, *J. Magn. Magn. Mater.*, 1999, **201**, 102–104.

76 I. A. Brezovich and R. F. Meredith, Practical aspects of ferromagnetic thermoseed hyperthermia, *Radiol. Clin. North Am.*, 1989, **27**, 589–602.

77 R. Hergt and S. Dutz, Magnetic particle hyperthermia—biophysical limitations of a visionary tumour therapy, *J. Magn. Magn. Mater.*, 2007, **311**, 187–192.

78 R. E. Rosensweig, Heating magnetic fluid with alternating magnetic field, *J. Magn. Magn. Mater.*, 2002, **252**, 370–374.

79 P. Guardia, A. Riedinger, S. Nitti, G. Pugliese, S. Marras, A. Genovese, M. E. Matera, C. Lefevre, L. Manna and T. Pellegrino, One pot synthesis of monodisperse water soluble iron oxide nanocrystals with high values of the specific absorption rate, *J. Mater. Chem. B*, 2014, **2**, 4426–4434.

80 L. Lartigue, C. Innocenti, T. Kalaivani, A. Awwad, M. Sanchez Duque Mdel, Y. Guari, J. Larionova, C. Guerin, J. L. Montero and V. Barragan-Montero, *et al.*, Water-dispersible sugar-coated iron oxide nanoparticles. An evaluation of their relaxometric and magnetic hyperthermia properties, *J. Am. Chem. Soc.*, 2011, **133**, 10459–10472.

81 J. Carrey, B. Mehdaoui and M. Respaud, Simple models for dynamic hysteresis loop calculations of magnetic single-domain nanoparticles: Application to magnetic hyperthermia optimization, *J. Appl. Phys.*, 2011, **109**, 083921.

82 E. M. Cubanova, R. A. Rylov and N. A. Usov, Dynamics of particles with cubic magnetic anisotropy in a viscous liquid, *J. Magn. Magn. Mater.*, 2022, **541**, 168494.

83 U. M. Engelmann, C. Shasha, E. Teeman, I. Slabu and K. M. Krishnan, Predicting size-dependent heating efficiency of magnetic nanoparticles from experiment and stochastic Néel-Brown Langevin simulation, *J. Magn. Magn. Mater.*, 2019, **471**, 450–456.

84 L. M. Bauer, S. F. Situ, M. A. Griswold and A. C. Samia, High-performance iron oxide nanoparticles for magnetic particle imaging-guided hyperthermia (hMPI), *Nanoscale*, 2016, **8**, 12162–12169.

85 E. C. Abenojar, S. Wickramasinghe, J. Bas-Concepcion and A. C. S. Samia, Structural effects on the magnetic hyperthermia



properties of iron oxide nanoparticles, *Prog. Nat. Sci.: Mater. Int.*, 2016, **26**, 440–448.

86 Z. Nemati, J. Alonso, L. M. Martinez, H. Khurshid, E. Garaio, J. A. Garcia, M. H. Phan and H. Srikanth, Enhanced Magnetic Hyperthermia in Iron Oxide Nano-Octopods: Size and Anisotropy Effects, *J. Phys. Chem. C*, 2016, **120**, 8370–8379.

87 R. Das, J. Alonso, Z. N. Porshokouh, V. Kalappattil, D. Torres, M. H. Phan, E. Garaio, J. A. García, J. L. S. Llamazares and H. Srikanth, Tunable High Aspect Ratio Iron Oxide Nanorods for Enhanced Hyperthermia, *J. Phys. Chem. C*, 2016, **120**, 10086–10093.

88 K. Y. Jiang, Q. B. Zhang, D. T. Hinojosa, L. L. Zhang, Z. Xiao, Y. Yin, S. Tong, V. L. Colvin and G. Bao, Controlled oxidation and surface modification increase heating capacity of magnetic iron oxide nanoparticles, *Appl. Phys. Rev.*, 2021, **8**, 031407.

89 Y. Rabin, Is intracellular hyperthermia superior to extracellular hyperthermia in the thermal sense?, *Int. J. Hyperthermia*, 2002, **18**, 194–202.

90 P. Moroz, S. K. Jones and B. N. Gray, Magnetically mediated hyperthermia: current status and future directions, *Int. J. Hyperthermia*, 2002, **18**, 267–284.

91 J. R. Lepock, Cellular effects of hyperthermia: relevance to the minimum dose for thermal damage, *Int. J. Hyperthermia*, 2003, **19**, 252–266.

92 B. Hildebrandt, P. Wust, O. Ahlers, A. Dieing, G. Sreenivas, T. Kerner, R. Felix and H. Riess, The cellular and molecular basis of hyperthermia, *Crit. Rev. Oncol. Hematol.*, 2002, **43**, 33–56.

93 C. A. Quinto, P. Mohindra, S. Tong and G. Bao, Multifunctional superparamagnetic iron oxide nanoparticles for combined chemotherapy and hyperthermia cancer treatment, *Nanoscale*, 2015, **7**, 12728–12736.

94 S. Giwa, J. K. Lewis, L. Alvarez, R. Langer, A. E. Roth, G. M. Church, J. F. Markmann, D. H. Sachs, A. Chandraker and J. A. Wertheim, *et al.*, The promise of organ and tissue preservation to transform medicine, *Nat. Biotechnol.*, 2017, **35**, 530–542.

95 J. K. Lewis, J. C. Bischof, I. Braslavsky, K. G. Brockbank, G. M. Fahy, B. J. Fuller, Y. Rabin, A. Tocchio, E. J. Woods and B. G. Wowk, *et al.*, The Grand Challenges of Organ Banking: Proceedings from the first global summit on complex tissue cryopreservation, *Cryobiology*, 2016, **72**, 169–182.

96 G. M. Fahy, B. Wowk, J. Wu, J. Phan, C. Rasch, A. Chang and E. Zendejas, Cryopreservation of organs by vitrification: perspectives and recent advances, *Cryobiology*, 2004, **48**, 157–178.

97 H. Huang, X. He and M. L. Yarmush, Advanced technologies for the preservation of mammalian biospecimens, *Nat. Biomed. Eng.*, 2021, **5**, 793–804.

98 R. Frederickson, Cryopreservation by vitrification, *Nat. Biotechnol.*, 2000, **18**, 250.

99 J. F. Peyridieu, A. Baudot, P. Boutron, J. Mazuer, J. Odin, A. Ray, E. Chapelier, E. Payen and J. L. Descotes, Critical cooling and warming rates to avoid ice crystallization in small pieces of mammalian organs permeated with cryoprotective agents, *Cryobiology*, 1996, **33**, 436–446.

100 J. B. Hopkins, R. Badeau, M. Warkentin and R. E. Thorne, Effect of common cryoprotectants on critical warming rates and ice formation in aqueous solutions, *Cryobiology*, 2012, **65**, 169–178.

101 N. Manuchehrabadi, Z. Gao, J. Zhang, H. L. Ring, Q. Shao, F. Liu, M. McDermott, A. Fok, Y. Rabin and K. G. Brockbank, *et al.*, Improved tissue cryopreservation using inductive heating of magnetic nanoparticles, *Sci. Transl. Med.*, 2017, **9**, eaah4586.

102 M. L. Etheridge, K. R. Hurley, J. Zhang, S. Jeon, H. L. Ring, C. Hogan, C. L. Haynes, M. Garwood and J. C. Bischof, Accounting for biological aggregation in heating and imaging of magnetic nanoparticles, *Technology*, 2014, **2**, 214–228.

103 M. L. Etheridge, Y. Xu, L. Rott, J. Choi, B. Glasmacher and J. C. Bischof, RF heating of magnetic nanoparticles improves the thawing of cryopreserved biomaterials, *Technology*, 2014, **2**, 229–242.

104 Z. Ye, Y. Tai, Z. Han, S. Liu, M. L. Etheridge, J. L. Pasek-Allen, C. Shastray, Y. Liu, Z. Li and C. Chen, *et al.*, Engineering Magnetic Nanoclusters for Highly Efficient Heating in Radio-Frequency Nanowarming, *Nano Lett.*, 2024, **24**, 4588–4594.

105 S. Liu, Z. Han, Z. Ye, M. Jiang, M. L. Etheridge, J. C. Bischof and Y. Yin, Magnetic-Nanorod-Mediated Nanowarming with Uniform and Rate-Regulated Heating, *Nano Lett.*, 2024, **24**, 11567–11572.

106 Z. Gao, H. L. Ring, A. Sharma, B. Namsrai, N. Tran, E. B. Finger, M. Garwood, C. L. Haynes and J. C. Bischof, Preparation of Scalable Silica-Coated Iron Oxide Nanoparticles for Nanowarming, *Adv. Sci.*, 2020, **7**, 1901624.

107 A. Chiu-Lam, E. Staples, C. J. Pepine and C. Rinaldi, Perfusion, cryopreservation, and nanowarming of whole hearts using colloidally stable magnetic cryopreservation agent solutions, *Sci. Adv.*, 2021, **7**, eabe3005.

108 Z. Han, J. S. Rao, L. Gangwar, B. E. Namsrai, J. L. Pasek-Allen, M. L. Etheridge, S. M. Wolf, T. L. Pruitt, J. C. Bischof and E. B. Finger, Vitrification and nanowarming enable long-term organ cryopreservation and life-sustaining kidney transplantation in a rat model, *Nat. Commun.*, 2023, **14**, 3407.

109 S. A. Stanley, J. Sauer, R. S. Kane, J. S. Dordick and J. M. Friedman, Remote regulation of glucose homeostasis in mice using genetically encoded nanoparticles, *Nat. Med.*, 2015, **21**, 92–98.

110 S. Rao, R. Chen, A. A. LaRocca, M. G. Christiansen, A. W. Senko, C. H. Shi, P. H. Chiang, G. Varnavides, J. Xue and Y. Zhou, *et al.*, Remotely controlled chemomagnetic modulation of targeted neural circuits, *Nat. Nanotechnol.*, 2019, **14**, 967–973.

111 S. A. Hescham, P. H. Chiang, D. Gregurec, J. Moon, M. G. Christiansen, A. Jahanshahi, H. Liu, D. Rosenfeld, A. Pralle and P. Anikeeva, *et al.*, Magnethermal nanoparticle technology alleviates parkinsonian-like symptoms in mice, *Nat. Commun.*, 2021, **12**, 5569.

112 M. A. Wheeler, C. J. Smith, M. Ottolini, B. S. Barker, A. M. Purohit, R. M. Grippo, R. P. Gaykema, A. J. Spano, M. P. Beenhakker and S. Kucenas, *et al.*, Genetically targeted magnetic control of the nervous system, *Nat. Neurosci.*, 2016, **19**, 756–761.

113 X. Ma, X. Liang, Y. Li, Q. Feng, K. Cheng, N. Ma, F. Zhu, X. Guo, Y. Yue and G. Liu, *et al.*, Modular-designed engineered bacteria for precision tumor immunotherapy via spatiotemporal manipulation by magnetic field, *Nat. Commun.*, 2023, **14**, 1606.

114 J. T. Dias, M. Moros, P. Del Pino, S. Rivera, V. Grazu and J. M. de la Fuente, DNA as a molecular local thermal probe for the analysis of magnetic hyperthermia, *Angew. Chem., Int. Ed.*, 2013, **52**, 11526–11529.

115 L. Polo-Corrales and C. Rinaldi, Monitoring iron oxide nanoparticle surface temperature in an alternating magnetic field using thermoresponsive fluorescent polymers, *J. Appl. Phys.*, 2012, **111**, 07B334.

116 A. Riedinger, P. Guardia, A. Curcio, M. A. Garcia, R. Cingolani, L. Manna and T. Pellegrino, Subnanometer local temperature probing and remotely controlled drug release based on azo-functionalized iron oxide nanoparticles, *Nano Lett.*, 2013, **13**, 2399–2406.

117 H. C. Davis, S. Kang, J. H. Lee, T. H. Shin, H. Puttermann, J. Cheon and M. G. Shapiro, Nanoscale Heat Transfer from Magnetic Nanoparticles and Ferritin in an Alternating Magnetic Field, *Biophys. J.*, 2020, **118**, 1502–1510.

118 S. Faure, N. Mille, S. S. Kale, J. M. Asensio, J. Marbaix, P. Farger, D. Stoian, W. van Beek, P. Fazzini and K. Soulantica, *et al.*, Internal Temperature Measurements by X-Ray Diffraction on Magnetic Nanoparticles Heated by a High-Frequency Magnetic Field, *J. Phys. Chem. C*, 2020, **124**, 22259–22265.

119 G. Wang, P. Zhang, S. K. Mendum, Y. Wang, Y. Zhang, X. Kang, B. N. Desai and J. J. Zhu, Revaluation of magnetic properties of Magneto, *Nat. Neurosci.*, 2020, **23**, 1047–1050.

120 K. Kole, Y. Zhang, E. J. R. Jansen, T. Brouns, A. Bijlsma, N. Calcini, X. Yan, A. D. S. Lantyer and T. Celikel, Assessing the utility of Magneto to control neuronal excitability in the somatosensory cortex, *Nat. Neurosci.*, 2020, **23**, 1044–1046.

121 F. X. Xu, L. Zhou, X. T. Wang, F. Jia, K. Y. Ma, N. Wang, L. Lin, F. Q. Xu and Y. Shen, Magneto is ineffective in controlling electrical properties of cerebellar Purkinje cells, *Nat. Neurosci.*, 2020, **23**, 1041–1043.

122 M. I. Brier, J. W. Mundell, X. Yu, L. Su, A. Holmann, J. Squeri, B. Zhang, S. A. Stanley, J. M. Friedman and J. S. Dordick, Uncovering a possible role of reactive oxygen species in magnetogenetics, *Sci. Rep.*, 2020, **10**, 13096.

123 P. T. Schumacker, Reactive oxygen species in cancer cells: live by the sword, die by the sword, *Cancer Cell*, 2006, **10**, 175–176.

124 L. Tong, C. C. Chuang, S. Wu and L. Zuo, Reactive oxygen species in redox cancer therapy, *Cancer Lett.*, 2015, **367**, 18–25.



125 T. Mai and J. Z. Hilt, Functionalization of iron oxide nanoparticles with small molecules and the impact on reactive oxygen species generation for potential cancer therapy, *Colloids Surf. A*, 2019, **576**, 9–14.

126 S. Fu, S. Wang, X. Zhang, A. Qi, Z. Liu, X. Yu, C. Chen and L. Li, Structural effect of $\text{Fe}(3)\text{O}(4)$ nanoparticles on peroxidase-like activity for cancer therapy, *Colloids Surf. B*, 2017, **154**, 239–245.

127 R. C. C. Costa, F. C. C. Moura, J. D. Ardisson, J. D. Fabris and R. M. Lago, Highly active heterogeneous Fenton-like systems based on $\text{Fe}^0/\text{Fe}_3\text{O}_4$ composites prepared by controlled reduction of iron oxides, *Appl. Catal. B*, 2008, **83**, 131–139.

128 Z. Yi, X. Yang, Y. Liang and S. Tong, Iron oxide nanozymes enhanced by ascorbic acid for macrophage-based cancer therapy, *Nanoscale*, 2024, **16**, 14330–14338.

129 Z. Chen, J. J. Yin, Y. T. Zhou, Y. Zhang, L. Song, M. Song, S. Hu and N. Gu, Dual enzyme-like activities of iron oxide nanoparticles and their implication for diminishing cytotoxicity, *ACS Nano*, 2012, **6**, 4001–4012.

130 V. Trujillo-Alonso, E. C. Pratt, H. Zong, A. Lara-Martinez, C. Kaittanis, M. O. Rabie, V. Longo, M. W. Becker, G. J. Roboz and J. Grimm, *et al.*, FDA-approved ferumoxytol displays anti-leukaemia efficacy against cells with low ferroportin levels, *Nat. Nanotechnol.*, 2019, **14**, 616–622.

131 L. Wu, W. Wen, X. Wang, D. Huang, J. Cao, X. Qi and S. Shen, Ultrasmall iron oxide nanoparticles cause significant toxicity by specifically inducing acute oxidative stress to multiple organs, *Part. Fibre Toxicol.*, 2022, **19**, 24.

132 Y. Xie, D. Liu, C. Cai, X. Chen, Y. Zhou, L. Wu, Y. Sun, H. Dai, X. Kong and P. Liu, Size-dependent cytotoxicity of Fe_3O_4 nanoparticles induced by biphasic regulation of oxidative stress in different human hepatoma cells, *Int. J. Nanomed.*, 2016, **11**, 3557–3570.

133 M. A. Smith, P. L. Harris, L. M. Sayre and G. Perry, Iron accumulation in Alzheimer disease is a source of redox-generated free radicals, *Proc. Natl. Acad. Sci. U. S. A.*, 1997, **94**, 9866–9868.

