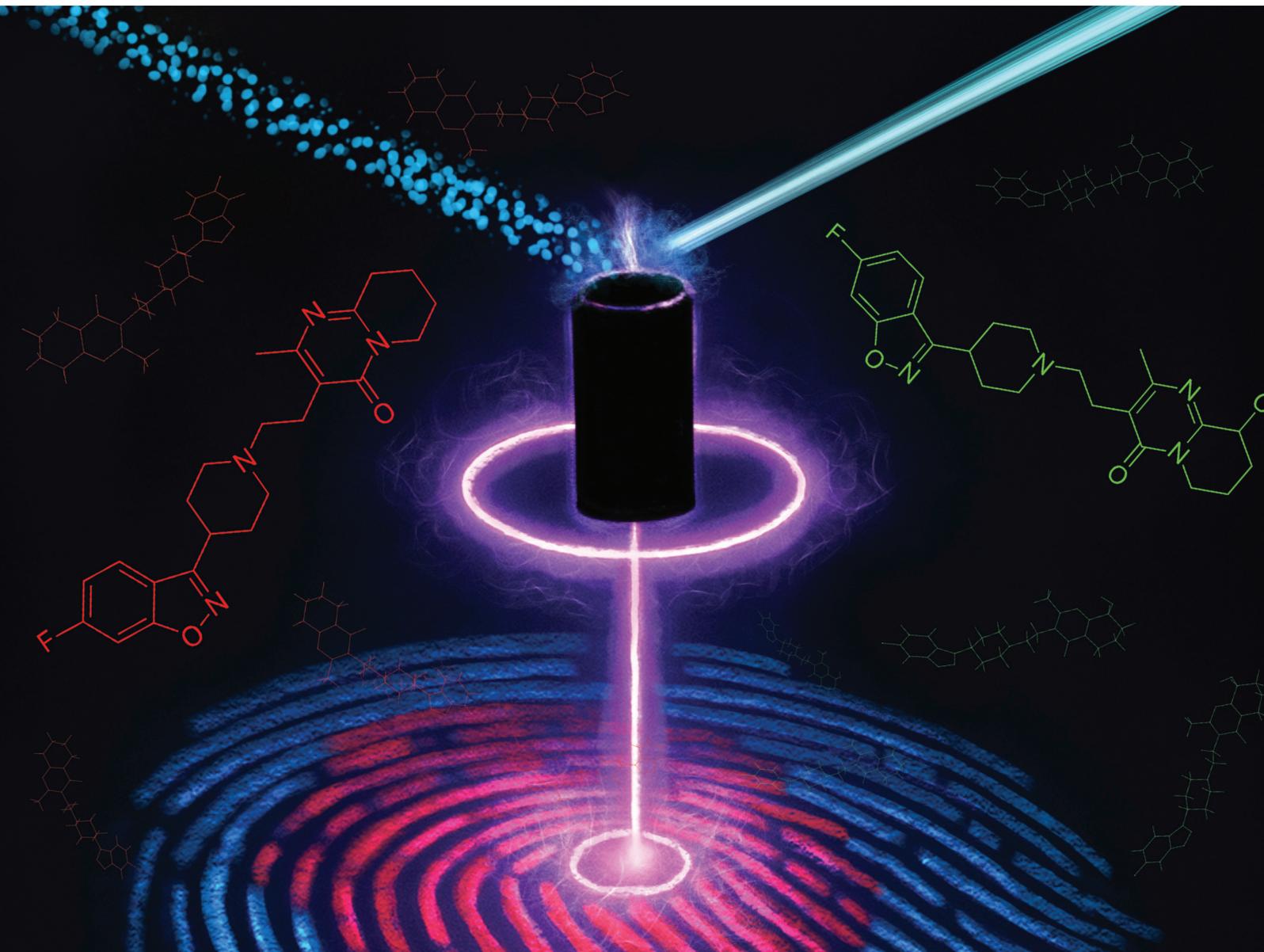


# Analyst

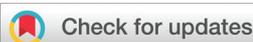
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**PAPER**

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A multi-modal mass spectrometry approach for the  
detection and mapping of date rape drugs in fingermarks



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## A multi-modal mass spectrometry approach for the detection and mapping of date rape drugs in fingerprints†

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Although drug facilitated sexual assault (DFSA) is an old issue, current statistics on the frequency of this crime have led to growing concerns. As these drugs are metabolised very quickly, toxicological evidence from biological fluids, corroborating the victim's statement, is challenging to recover, especially with late reports. We are proposing an additional method involving the analysis of the victim's fingerprints recovered at the scene(s) of the crime, which may contain the parent drug and its metabolite. As a case study, a multi-modal mass spectrometry-based approach has been developed and explored to detect and image both risperidone and its pharmacologically active metabolite, paliperidone, after contamination of a fingertip at very low concentrations and deposition of a fingerprint on a surface. In particular Matrix Assisted Laser Desorption Ionisation Mass Spectrometry Imaging (MALDI MSI), Desorption Electrospray Ionisation Mass Spectrometry Imaging (DESI MSI) and Soft Ionisation by Chemical Reaction In Transfer, (SICRIT®) have been used in different combinations to both detect the drugs and reconstruct the fingerprint ridge pattern; this approach enables the simultaneous provision of both chemical information (circumstances surrounding the crime) and biometric information. A forensic operational scenario has also been simulated whereby the contaminated fingerprint is deposited on paper and enhanced with a routine fingerprint enhancement technique prior to analysis *via* mass spectrometry imaging. Overall, our investigation indicates that this additional approach is feasible and is worth exploring further.

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### 1. Introduction

Drug facilitated sexual assault (DFSA) refers to sexual violence (SV) perpetrated against an individual under the influence of prescription, licit or illicit substances.<sup>1</sup> Proactive DFSA is the intentional incapacitation of the victim through the covert administration of such drugs to perpetrate SV.<sup>2</sup> Whilst the practice of incapacitating victims to sexually assault them is not at all a modern age issue,<sup>3,4</sup> proactive DFSA, through drink or needle spiking, has become an escalating concern due to the frequency with which it occurs and the long term mental health impact on the victims and their families, as well as the deriving social cost of drug-related harms and interventions.<sup>5,6</sup>

In 2022, in the UK, the Home Affairs Select Committee (HASC) issued a survey receiving more than 3000 spiking

victims or witnesses' responses,<sup>4</sup> though it was accepted that victims' numbers are probably higher due to under-reporting. HASC reports that between May 2022 and April 2023, the police received 6732 reports of spiking, averaging 561 reports a month.<sup>4</sup> In 2021, The Australian Bureau of Statistics (ABS) published a report on SV from which it emerged that in the previous 10 years, 43 900 women had experienced and reported DFSA, with an estimated lack of reporting of 321 800 cases.<sup>7</sup> In the US, a national study from 2007 found that 2.3% of surveyed women (2.6 million) reported alcohol/drug-facilitated rape after covert administration.<sup>8</sup> In a subsequent survey conducted between 2010 and 2012, including women and males subjected to SV through DFSA in their lifetime, 32% of women (7229) and 30.3% responders (5575) reported DFSA through drugs or a combination of drugs and alcohol.<sup>9</sup>

Sexual assaults are a challenging crime to prosecute due to several reasons including lack of/late reporting and independent witness testimony, difficulties in proving lack of consent, amnesia,<sup>10</sup> but also lack of solid physical evidence. In the case of DFSA, lack of toxicological evidence is also critical; the vast majority of DFSA drugs most commonly used for DFSA, also colloquially (and perhaps disingenuously) referred as to "date

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rape drugs", are rapidly metabolised; their short half-life time has resulted in false negatives and/or often renders their quantification in biological fluids, analysed days after the crime has occurred, a prohibitive challenge.<sup>4,11–13</sup>

Other types of biological specimens have therefore become desirable to attempt the detection of these substances; fingermarks (ridge pattern impressions accidentally deposited on a surface) may potentially be such a specimen. Through touch, the victim may in fact accidentally deposit their fingermarks on objects after the drug spiking, and/or during the subsequent sexual assault, when the drugs are being metabolised. The drug and their metabolites could therefore be excreted through sweat and transferred to the touched surface within deposited mark. It may be possible, as observed for other types of drugs and metabolites that, even if the recovery of the fingermark and analysis occurs after several days, these species can still be detected in this specimen.<sup>14,15</sup> The attention towards drugs is justified by the fact that whilst consumption of alcohol remains the most prevalent means to perpetrate SV,<sup>4,10</sup> a range of substances, covertly administered to the victim, has also been documented in the literature, including (in a non-exhaustive list) cannabinoids, opioids, benzodiazepines, amphetamines, GHB, antidepressants, cathinones and antipsychotics.<sup>4,16</sup>

This paper explores a multi-modal mass spectrometry approach for the detection and mapping of risperidone in fingermarks. Risperidone is a benzisoxazole and it is a widely prescribed atypical antipsychotic drug for the treatment of schizophrenia and other psychosis. However, its use has also been reported in DFSA.<sup>16,17–19</sup> The combination of its intended use as an antipsychotic and misuse as an DFSA drug, makes for an interesting case study due to (1) the potential transferability of the method to the detection and mapping of other DFSA drugs; and (2) the intelligence that is possible to provide around an individual to either narrow down the pool of suspects (fingermark of the perpetrator taking the prescribed medication) or to provide circumstantial or corroborating evidence assisting the victim's claims (from the fingermark of the victim covertly administered with this drug in the context of DFSA). Furthermore, risperidone has been detected in a large range of biological specimens<sup>20</sup> using LC MS/MS; interestingly, a non-invasive specimen, close to fingermarks, namely fingertip smears, was employed to detect a range of antipsychotics in patients in the context of drug adherence, through LC MS/MS.<sup>21</sup> However, the antipsychotics investigated did not include risperidone, the analytical specimen had no ridge detail and use of LC MS/MS required destruction of the fingermark sample.

The *in situ* mass spectrometric approaches employed in the work presented here have never been developed or applied to fingermarks with respect to the detection and mapping of risperidone and its main metabolite (individually and in mixture) and, in contrast to the application of LC MS/MS methods, they have the benefit of preserving, at large, the integrity of the fingermark. As such, they are particularly interesting from a forensic standpoint as well as from a drug adherence viewpoint

as they are potentially faster, not requiring extensive sample preparation.

Here we have employed an array of mass spectrometry techniques and developed methods for the detection and/or mapping of risperidone and its pharmacologically active metabolite 9-hydroxy-risperidone (paliperidone) in intact fingerprints (ridge impressions deliberately deposited on a surface), with some of these methods used in tandem to maximise the forensic intelligence retrievable. In particular, one vacuum and two ambient ionisation techniques were employed, namely Matrix Assisted Laser Ionisation Mass Spectrometry Imaging (vacuum MALDI MSI), Desorption Electrospray Ionisation Mass Spectrometry Imaging (DESI MSI) and thermal desorption soft ionisation through Chemical Reaction In Transfer, (SICRIT®), (Plasmion GmbH, Augsburg, Germany), based on a dielectric barrier discharge ionisation (DBDI) source.<sup>22</sup> The inclusion of SICRIT® in the workflow was due to documented success in detecting cocaine, heroin and fentanyl from fingerprints, in the picogram range.<sup>22</sup> In this study, MALDI MSI and DESI MSI were employed to detect and map risperidone and paliperidone in fingerprints, whereas SICRIT® was employed to preliminarily and quickly profile the presence of these species and to detect them at a higher sensitivity. Multi-modal workflows involved the sequential use (in separate instances) of (i) DESI MSI and MALDI MSI; and (ii) thermal ionisation profiling through SICRIT® followed by MALDI MSI or by DESI MSI and then MALDI MSI.

Finally, considering the operational perspective, risperidone and paliperidone-contaminated fingerprints were also preliminarily treated using fingermark enhancement techniques, following the protocols reported by the Fingermark Visualisation Manual (FVM)<sup>23</sup> prior to sequential analysis by MALDI MSI and DESI MSI, to demonstrate feasibility of these mass spectrometric techniques in an operational workflow. We have elected to visualise fingermarks on paper, as one of the possible surfaces of deposition scenarios; 1,2-indandione was initially used as it is a Cat A recommended technique for fingermark enhancement on paper (FVM<sup>3</sup>) and subsequently 4-dimethylaminocinnamaldehyde (DMAC) was employed as a Cat B process. We up-scaled the challenge by using red paper in addition to white paper as deposition surfaces; red paper has been reported as a challenging substrate for fingermark visualisation, both anecdotally, during research<sup>24</sup> and also in operational work.<sup>25</sup> This is because the colour and the background fluorescence of the substrate make it difficult to visualise marks with the commonly used reagents for porous surfaces, ninhydrin and indandione. Ninhydrin develops purple-red marks giving very low colour contrast with the coloured paper, and the red dyes used in the paper produce background fluorescence at the wavelengths used to examine 1,2-indandione.

Therefore, this experiment intended to demonstrate the compatibility of mass spectrometry imaging-based approaches with 1,2-indandione and DMAC, and the deriving biometric and chemical intelligence that can be provided.

Overall, this research has shown promise for an alternative approach in supporting DFSA allegations by detecting the



spiked/injected drug in the fingerprint of the victim that may be recovered after the incident. The detection and mapping of these species using *in situ* mass spectrometry-based methods, as standalone or as a multimodal approach, provide toxicological, biometric and circumstantial evidence to the investigators.

## 2. Materials and methods

### 2.1. Materials

Ultra-pure methanol (MeOH), acetonitrile (ACN), and formic acid (FA) for DESI MSI were sourced from ROMIL, UK Ltd. The Milli-Q water was obtained from an in-house Millipore water purification system (Merck KGaA, Darmstadt, Germany) for MALDI MS based analyses or from ROMIL for DESI MS based analyses. The MALDI matrix  $\alpha$ -CHCA, red phosphorus, trifluoroacetic acid and the standards for risperidone (RIS) and paliperidone (PAL) were sourced from Merck Life Science, UK Ltd. Double-sided conductive carbon tape was purchased from TAAB (Aldermaston, UK). ALUGRAM SIL G/UV254, silica pre-coated aluminium sheets were purchased from Macherey-Nagel GmbH, Germany. MeOH, zinc chloride and acetic acid for 1,2-indandione enhancement were obtained from Fisher Chemical (Loughborough, UK). 1,2-Indandione was supplied by BVDA, (Haarlem, Netherlands) and DMAC by Sigma (Gillingham, UK). Ethanol was purchased from JT Baker (Lutterworth, UK). Ethyl acetate was purchased from Acros Organics (Loughborough, UK) and petroleum ether (BP 40–60) from Honeywell (Muskegon, USA).

### 2.2. Instrumentation and instrumental settings

**2.2.1 DESI MS-based analysis.** DESI MS Imaging of risperidone and paliperidone was carried out on either a Waters Corporation tandem quadrupole mass spectrometer (XEVO TQ-XS) or on a SELECT SERIES Cyclic Ion Mobility mass spectrometer (cIMS) incorporating, in each case, a DESI XS ionisation source (Waters Corp. Wilmslow, UK).

The DESI XS source on the Xevo TQ-XS was operated in positive mode and as it follows: the capillary and the cone voltages were optimised and then set to 1.0 kV, and 30 V respectively. The positioning of the DESI source was set as follows: the *x*-axis and the *y*-axis at 2 a.u., the *z*-axis at 4 a.u. and the angle of the DESI emitter was set at 72°. The solvent composition used was 95 : 5 (MeOH : water, 0.1% FA) and the solvent flow rate was set at 2  $\mu\text{L min}^{-1}$ . For profiling experiments, ions were scanned over the *m/z* range 250–450 at a scan rate of 1 scan per second.

To confirm the presence of risperidone and paliperidone through MS/MS on the TQ-XS, the instrument was operated in MRM mode using the transitions at nominal *m/z* of 411  $\rightarrow$  191 and 427  $\rightarrow$  191, respectively. For MS/MS experiments, collision energy was optimised and set to 40 eV.

Imaging experiments were set up using the DESI method editor (Waters, Wilmslow, UK) using a scan rate of 10 scans per second and a step size of 50  $\mu\text{m} \times 50 \mu\text{m}$ . Collision gas

flow rate was set to 0.14  $\text{mL min}^{-1}$  for MS/MS profiling and imaging experiments.

The DESI XS source on the cIMS was operated in positive ionisation mode with the following optimised instrument settings: capillary voltage, 0.7 kV; cone voltage, 40 V; heated Transfer Line (HTL) temperature, 100 °C; source temperature, 120 °C; API gas pressure, 0.08 mPa; DESI emitter angle, 72°; solvent flow rate, 1.00  $\mu\text{L min}^{-1}$ , scan rate, 450  $\mu\text{m s}^{-1}$  and a step size of 50  $\mu\text{m} \times 50 \mu\text{m}$  for imaging experiments. The cIMS was operated in MS mode (sensitivity), with data acquired from *m/z* 50–1200. The manual quadrupole profile employed was (M1 = *m/z* 250, mass M2 = *m/z* 300, mass M3 = *m/z* 400). The solvent composition used was 95 : 5 (MeOH : water, 0.1% FA) including 100  $\text{pg } \mu\text{L}^{-1}$  LEU-ENK for continuous lock mass. The instrument was calibrated with polyalanine prior to every analysis with an average mean prediction error of 0.7 ppm.

**2.2.2 MALDI MS-based analyses.** MALDI MS and MSI measurements were carried out on the reflecting time-of-flight, the SELECT SERIES™ MRT (Waters Corp., Wilmslow, UK) equipped with a 2.5 kHz solid state Nd:YAG laser. The MALDI source on the MRT was operated in positive ionisation mode setting the laser intensity at 350 a.u., the lens focus, 6 a.u., the secondary attenuation ON and the MALDI gas flow at 350  $\text{mL min}^{-1}$ , following the systematic approach developed by Krishna *et al.*<sup>26</sup> The TOF was operated in full-flight path MRT mode (high resolution), with data acquired from *m/z* 50–2400.

A step size of 50  $\mu\text{m} \times 50 \mu\text{m}$  was used for imaging experiments. The manual quadrupole profile employed was (M1 = *m/z* 250, mass M2 = *m/z* 300, mass M3 = *m/z* 400). The instrument was calibrated with red phosphorus prior to every analysis with an average mean prediction error of 0.2 ppm.

**2.2.3 SICRIT® – based analyses.** The SICRIT ionisation source (Plasmion, Augsburg, Germany) was coupled to the SYNAPT G2 HDMS QToF mass spectrometer (Waters Corp., Wilmslow, UK) and operated at a temperature of 300 °C, with a frequency of plasma generation set at 15 kHz, amplitude at 1600 V and purge gas flow rate set at 3  $\text{L min}^{-1}$ . The SYNAPT G2 HDMS QToF mass spectrometer was operated in sensitivity mode, in the *m/z* range 50–1200 setting the capillary voltage at 2.6 kV, the acquisition rate at 1 scan per second. Sample solutions were introduced *via* injection module over 10 seconds, and fingerprint samples were analysed using the thermal desorption unit (TDU) with a data acquisition time of 30 seconds. For MS/MS experiments, the instrument was operated in pseudo-MRM using the transitions at nominal *m/z* 411  $\rightarrow$  191 (risperidone) and *m/z* 427  $\rightarrow$  207 (paliperidone), respectively. Collision energy of 30 eV for risperidone and 25 eV for paliperidone was used. The quadrupole profile was set in automatic mode (M1 = *m/z* 100, mass M2 = *m/z* 300, mass M3 = *m/z* 500).

**2.2.4 Data acquisition and processing.** Data were acquired using MassLynx™ (v4.2 SCN2021-Beta for Xevo TQ-XS, v4.2 SCN1016 for cIMS, v4.2 SCN991 for SYNAPT-G2, v4.2 SCN1026



for MRT), DESI Method Editor v.2.3.8 (Waters Corporation, Wilmslow UK), Omni Spray 2-D v2.1.0 (Prosolia Inc., Indiana, USA) for Xevo TQ-XS and the Quartz (v2.12.1 for cIMS, v2.7.1 for MRT) software.

Mass spectra were processed either directly in Masslynx v4.2 (respective releases as above) or opened in Masslynx and exported into mMass, an open-source mass spectral analysis platform.<sup>27,28</sup> MSI experiments were set up using DESI Method Editor v2.3.8 for the Xevo TQ-XS, and Quartz (v2.12.1, v2.7.1) for cIMS, and MRT respectively.

All the DESI-cIMS and MALDI-MRT imaging experiments were processed and visualised using HDI v1.8 (Waters Corporation, Wilmslow UK) with the following settings: MS resolution of (100 000 for cIMS, and 200 000 for MRT), *m/z* window of (0.02 for cIMS, and 0.005 for MRT), and the number of most intense peaks set to 1000. The DESI-Xevo TQ-XS MRM images were auto processed from Masslynx and visualised in HDI v1.8. Except for those generated by the Xevo TQ-XS, all the images were TIC (Total Ion Count), normalised unless specified otherwise. Relevant regions of interests (ROI) were exported to MassLynx™ v4.2 (respective releases as above) software for spectral analysis.

## 2.3 Methods

**2.3.1 Mapping of risperidone and paliperidone in fingerprints via DESI MSI.** Limits of detection were preliminarily assessed through 10-fold serial dilutions of both risperidone and paliperidone in 95% MeOH in Water (v/v%) ranging from  $5 \times 10^5$  ng mL<sup>-1</sup> to 5 ng mL<sup>-1</sup> in concentration. Samples were prepared by depositing 5  $\mu$ L of solution on a glass slide and leaving the solvent to evaporate completely for 10 min, prior to analysis. For MS and MS/MS profiling, 5  $\mu$ L of either drug at concentrations varying between 50  $\mu$ g mL<sup>-1</sup> and 50 ng mL<sup>-1</sup> were spotted on natural fingerprints preliminarily deposited on a glass slide and allowed to dry prior to analysis. For Imaging experiments, adapting the method of Groeneveld *et al.*,<sup>29</sup> either 20  $\mu$ L of Risperidone at a concentration of  $5 \times 10^3$  ng mL<sup>-1</sup> or 20  $\mu$ L of paliperidone at a concentration of 500 ng mL<sup>-1</sup> were deposited on a glass slide and allowed to evaporate. Subsequently a fingertip was dragged side-to-side on the location of the drug prior and then it was made to contact a clean glass slide to generate a drug-contaminated natural mark for analysis. To mimic detection in physiological conditions after administration of risperidone, a mixed solution of 100  $\mu$ L was prepared by mixing 50  $\mu$ L of  $5 \times 10^3$  ng mL<sup>-1</sup> risperidone and 50  $\mu$ L of  $5 \times 10^3$  ng mL<sup>-1</sup> paliperidone (1:1 ratio) and 10  $\mu$ L were deposited on a glass slide and allowed to evaporate prior to fingertip contamination and contaminated fingerprint generation as described above.

Risperidone and paliperidone were also analysed on a DESI XS source coupled to cIMS to compare the performance with that of the Xevo TQ-XS. A risperidone and paliperidone-contaminated fingerprint was prepared according to Groeneveld *et al.*<sup>29</sup> and as described for the same type of experiment on the DESI-Xevo TQ-XS, except 50  $\mu$ L of risperidone and paliperi-

done were mixed in a 1:1 ratio each at a concentration of  $1 \times 10^4$  ng mL<sup>-1</sup> rather than  $5 \times 10^3$  ng mL<sup>-1</sup>.

**2.3.2 Matrix spray deposition.** The HTX M<sup>3+</sup> Sprayer (HTX Imaging, North Carolina, USA) was used to matrix spray-coat the fingerprints prior to MALDI MSI. Samples were coated with 8 layers of 5 mg mL<sup>-1</sup>  $\alpha$ -CHCA prepared in 70:30 ACN:0.1% TFAaq. Following optimisation, the sprayer was operated as follows: nozzle temperature, 75 °C, pressure at 0.07 MPa (10 psi), flowrate 100  $\mu$ L min<sup>-1</sup> and a velocity of 1200 mm min<sup>-1</sup>.

**2.3.3 Mapping of risperidone and paliperidone in fingerprints via MALDI MSI.** In the standalone MALDI imaging workflow, two experiments were performed, namely (1) an imaging sensitivity test through risperidone and paliperidone spotting, to assess the limit of detection for these species in the presence of a fingerprint and (2) imaging of a risperidone and paliperidone-contaminated fingerprint. The ungroomed fingerprints were deposited onto aluminium slides which were then secured to the MALDI target plate using double-sided conductive tape, prior to imaging analysis. For drug spotting imaging, the risperidone and paliperidone standard solutions were individually prepared in a concentration range between 1000 ng mL<sup>-1</sup> to 0.1 ng mL<sup>-1</sup> in methanol and in a ten-fold dilution series. Each solution was mixed with 5 mg mL<sup>-1</sup> solution of  $\alpha$ -CHCA in 70:30 v/v ACN:0.1% TFAaq in a 1:1 ratio and 0.5  $\mu$ L of each resulting solution was spotted across the fingerprint. Risperidone and paliperidone-contaminated ungroomed fingerprints were prepared using the method of Groeneveld *et al.*,<sup>29</sup> as for the DESI-cIMS contaminated fingerprint experiment described in section 2.3.1., prior to be imaged.

**2.3.4 Thermal desorption of risperidone and paliperidone via SICRIT® ionisation.** Sensitivity MS profiling experiments were initially conducted for serial dilutions of risperidone and paliperidone in 100% MeOH spanning a range of concentrations from 0.5 ng mL<sup>-1</sup> to  $5 \times 10^5$  ng mL<sup>-1</sup>. One  $\mu$ L of each sample was manually injected into the ion source over a period of 10 seconds. The SICRIT was set as described in section 2.2. One natural drug-free fingerprint, two natural fingerprints contaminated with 5  $\mu$ L of  $5 \times 10^3$  ng mL<sup>-1</sup> of either risperidone or paliperidone, following the method of Groeneveld *et al.*<sup>29</sup> were subsequently analysed on glass slides. A further two fingerprints were deposited using the same method, by preliminarily spotting on a glass slide 5  $\mu$ L of 0.5  $\mu$ g mL<sup>-1</sup> of either risperidone or paliperidone. The same settings for analysing liquid solutions were employed except, this time, glass slides were inserted in the source *via* the heating cartridge. Data were acquired over 30 seconds at 1 scan per second. The temperature was preliminarily optimised between 50 and 300 °C.

**2.3.5 Risperidone and paliperidone contaminated fingerprint chemical enhancement on paper.** Risperidone and paliperidone-contaminated fingerprints were firstly produced following the method of Groeneveld *et al.*<sup>29</sup> and deposited by contacting the fingertip with both white and red paper to produce split fingerprints (half on white paper and half on red paper).



In the first enhancement process employed, the formulation for the working solution of indandione consisted of 0.25 g 1,2-indandione dissolved in 45 mL ethyl acetate, 45 mL methanol, 10 mL acetic acid, 1 L petroleum ether (BP 40–60), 1 mL zinc chloride stock solution, consisting of 0.1 g zinc chloride, 4 mL ethyl acetate and 1 mL Acetic acid. The paper samples were passed through a shallow tray containing the working solution, ensuring the paper was fully wetted, then allowed to dry in a fume cupboard. They were then placed into an DFO/Ninhydrin development chamber (Air Sciences Safedvelop, Ormskirk, UK) at 100 °C and 0% RH for 10 minutes before being removed for examination.

In the second process, the 4-dimethylaminocinnamaldehyde (DMAC) contact transfer process was used. DMAC impregnated sheets were first prepared by dipping 80gsm white copy paper in a solution of 0.25 g DMAC dissolved in 100 mL ethanol and allowing them to dry. These sheets were kept in plastic wallets in a freezer until ready for use. To visualise the fingermarks, the paper samples to be treated were placed between 2 sheets of impregnated paper, placed in a plastic wallet and left for 24 hours at room temperature.

Examination of the samples treated with both 1,2-indandione and DMAC was carried out using fluorescence examination using Crimelite 82S (Foster + Freeman, Evesham, UK) forensic light sources at 3 wavelengths namely (i) blue (420–470 nm) in conjunction with GG495 viewing goggles/camera filter, (ii) blue/green (445–510 nm) in conjunction with OG550 viewing goggles/camera filter and (iii) green (480–560 nm) in conjunction with OG590 viewing goggles/camera filter.

Photography of the fingermarks was carried out using a Canon EOS 800D DSLR camera with EF-S 18–55 mm lens.

**2.3.6 Sequential mass spectrometry based analysis of risperidone and paliperidone.** Risperidone (RIS)-, paliperidone (PAL)-and RIS/PAL-contaminated fingerprints were submitted to mass spectrometry analysis with or without prior chemical enhancement. In the latter case, control (CNT) marks (in which neither the drug nor the metabolite was present) have been included. A total of 8 combinations were trialled and they are summarised in Table 1.

**2.3.6.1 SICRIT ionisation → DESI MSI.** For this mass spectrometry based sequential experiment, a natural fingerprint was deposited on a glass slide after contamination of the fingertip with a dried solution of 10 µL of  $5 \times 10^3$  ng mL<sup>-1</sup> risperidone and 75 µL of  $5 \times 10^3$  ng mL<sup>-1</sup> paliperidone following the

method of Groeneveld *et al.*<sup>29</sup> The SICRIT source was set to 300 °C and once inserted in the heating cartridge, MS spectra were acquired for 30 seconds after which the glass slide was submitted to DESI MS/MS Imaging at 50 µm × 50 µm spatial resolution on a Xevo TQ-XS mass spectrometer using the settings described in section 2.2.

**2.3.6.2 Chemical enhancement → MALDI MSI/DESI MSI of risperidone and paliperidone in fingermarks.** Two sets of marks were prepared and submitted to this workflow. For the first set, a total of 6 marks, including 2 control, 2 risperidone- and 2 risperidone and paliperidone contaminated-fingermarks were prepared. Specifically, control marks were natural marks (*i.e.* deposited with no prior contamination of the fingertip with the drug(s)). Risperidone-contaminated marks were generated preliminarily by spotting a 10 µL of a  $1 \times 10^4$  ng mL<sup>-1</sup> solution of risperidone on a glass slide and risperidone/paliperidone-contaminated marks were generated spotting a mixed solution of 5 µL of risperidone  $1 \times 10^4$  ng mL<sup>-1</sup> and 5 µL of paliperidone  $1 \times 10^4$  ng mL<sup>-1</sup>. The fingerprints were generated by contacting the fingertip with both white and red paper to ensure that one half of the mark sat on white paper and the other half on red paper, as illustrated in Fig. S1.†

These marks were then enhanced using 1,2-indandione and photographed as described in section 2.3.5. Of these marks, a subset of 3 was imaged either *via* DESI MSI on a Xevo-TQ-XS mass spectrometer or *via* MALDI MSI on a SELECT SERIES MALDI MRT mass spectrometer.

The second set of marks consisted in 6 identically prepared fingerprints generated exclusively on red paper after dragging fingertips over a dried solution made out of 5 µL of  $1 \times 10^4$  ng mL<sup>-1</sup> risperidone and 5 µL of  $1 \times 10^4$  ng mL<sup>-1</sup> paliperidone on a glass slide. This set of drug-contaminated fingerprints was enhanced with DMAC, as described in section 2.3.5, prior to splitting the marks and subjecting each of the halves of any one fingermark to DESI or MALDI MSI. In this instance, DESI MSI was carried out using a cIMS.

### 3. Results and discussion

Sexual Violence (SV), assisted by the covert administration of incapacitating drugs (DFSA), is an increasing global and societal concern due to the long-term impact on the victims and the cost of intervention to society.<sup>5,6</sup> Proving DFSA is challenging and hampered by several factors, including, from an analytical perspective, the generally short half time of the

**Table 1** Combination of mass spectrometry techniques applied for the detection, confirmation and imaging of risperidone and paliperidone

Combination no.	Nature of the sequential process	CNT mark	RIS mark	RIS/PAL mark
1	SICRIT <sup>®</sup> Ionisation → DESI MSI	—	—	✓
2	SICRIT <sup>®</sup> ionisation → MALDI MSI	—	—	✓
3	SICRIT <sup>®</sup> ionisation → DESI MSI-MALDI MSI (with dry wet method <sup>29</sup> )	—	✓	✓
4	1,2-Indandione → DESI MSI	✓	✓	✓
5	1,2-Indandione → MALDI MSI	✓	✓	✓
6	DMAC → DESI MSI/MALDI MSI	—	—	✓



drugs combined with late reporting, which make it very difficult, if not impossible, to detect/quantify these species from biological fluids.<sup>4,11–13</sup> Fingermarks left by the victim, potentially containing the excreted “date rape drug” and/or its metabolite, could be an additional toxicological specimen of interest. Here we describe the development of a mass spectrometric-based approach for the detection of date rape drugs in fingermarks, using risperidone and its main and pharmacologically active metabolite 9-hydroxyrisperidone (paliperidone) as a case study.<sup>16,17–19</sup> MALDI MSI and DESI MSI, *in situ* techniques preserving the integrity of the sample, and Chemical Reaction In Transfer, SICRIT®, (Plasmion GmbH, Augsburg, Germany) were employed in this study both as standalone techniques and in combination.

### 3.1 Mapping of risperidone and paliperidone in fingermarks via MALDI MSI

A sensitivity imaging study was preliminarily conducted by spotting serial dilutions of either risperidone or paliperidone, in a concentration range between  $1 \times 10^3$  ng mL<sup>-1</sup> and 0.1 ng mL<sup>-1</sup>. These solutions were spotted on a natural fingerprint (Fig. S2†), generated without any prior cleaning or sebum enrichment of the fingertip. This experiment intended to assess the potential ion suppression exerted by the fingerprint chemical composition on these two species and determine the working concentration of these two species for subsequent fingerprint imaging analyses. It was found that the lowest concentration at which risperidone could be imaged in a mark was 1 ng mL<sup>-1</sup> (0.5 μL of the drug mixed with 0.5, μL of α-CHCA, total of 0.5 ng of risperidone) (Fig. S2A†), with a S/N of >5. Although for both 10 ng mL<sup>-1</sup> and 1 ng mL<sup>-1</sup> only scattered pixels are visible (Fig. S2A,† inset (i)), the presence of risperidone at these concentrations was confirmed through mass spectra extracted from the region of interest (ROI) shown in Fig. S2B.† For paliperidone the LOD was higher, as it could only be detected and visualised at a concentration of 10 ng mL<sup>-1</sup> (corresponding to 5 ng), (S/N 189) as confirmed by the ROI mass spectra in (Fig. S2D†) and by the S/N falling dramatically below 3 : 1 (0.63), with the 1 ng mL<sup>-1</sup> concentration. It is important to note that whilst other MALDI matrices were not tested, the α-CHCA composition and spray-coating conditions were preliminarily optimised. The selection of α-CHCA was due to the versatility of this matrix especially for small molecules in fingerprints so that a variety of different species could be detected; this allows for untargeted MALDI MSI analysis where casework is commissioned and molecular targets may not be known in advance.

Despite the LOD for risperidone and paliperidone at 1 ng mL<sup>-1</sup> and 10 ng mL<sup>-1</sup> respectively, the sensitivity when imaging a mark generated by dragging a fingertip over a 50 μL of a dried solution of the two species deposited on a glass slide, prior to touching the aluminium slide,<sup>29</sup> was lower. The visualisation of these species in a mixture was only possible in a 1 : 1 ratio of a final concentration  $5 \times 10^3$  ng mL<sup>-1</sup>, corresponding to a total amount of 250 ng per analyte (Fig. 1). The higher LOD in this instance is due to the chemical species

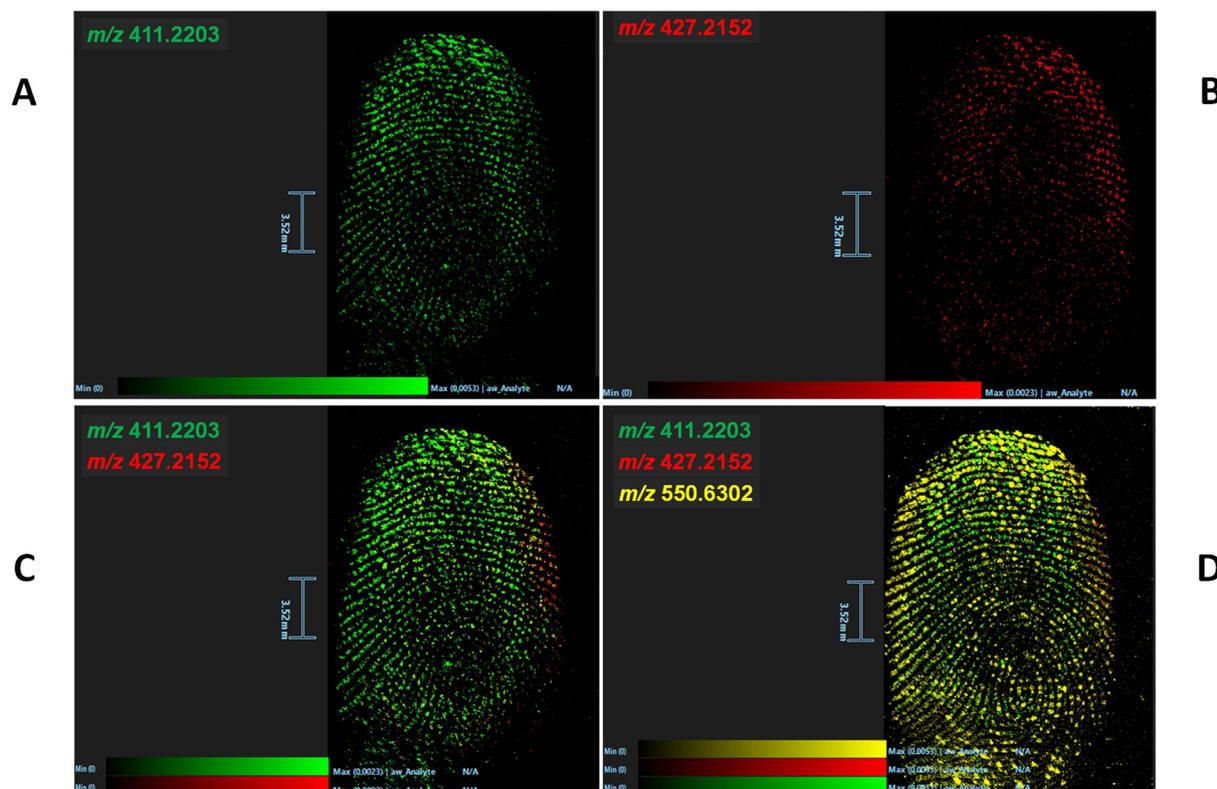
spreading on a bigger area (the fingerprint *versus* the spot) and dropping below the detection threshold of the mass spectrometer.

Fig. 1 shows the distributions of risperidone and paliperidone in a fingerprint (using a fingertip-contaminating solution of the two species, each at a final concentration of  $5 \times 10^3$  ng mL<sup>-1</sup>), providing chemical intelligence (Fig. 1A and B respectively), as well as in a combined image (Fig. 1C) to enhance the biometric information; in the latter instance, this overlay was also complemented by superimposing the image of distribution of a ubiquitous compound often found in fingerprints, namely dimethyldioctadecylammonium ion<sup>30–32</sup> (Fig. 1D). This image overlay contributes to the provision of biometric intelligence, through improving the ridge pattern continuity. The 1 : 1 risperidone/paliperidone ratio was selected according to the average concentrations in blood for the two species in previous studies<sup>20,33</sup> (with a ratio averaging 0.9 across stomach content, pericardial fluid, urine, kidney, cerebrum and liver tissues<sup>20</sup>). Whilst this MALDI MSI experiment has proven feasibility of the method, the approach needs to be further evaluated in fingerprints of individuals who have been administered with risperidone. Indeed, the amount of the two species in a fingerprint, just like in blood, may vary with the individual's metabolic rate, dosage, administration mode and sampling time. A study including a number of patients representative of the above conditions, would show in which instances, for example, only the metabolite is detectable, with the parent drug having undergone extensive metabolism and being no longer detectable.

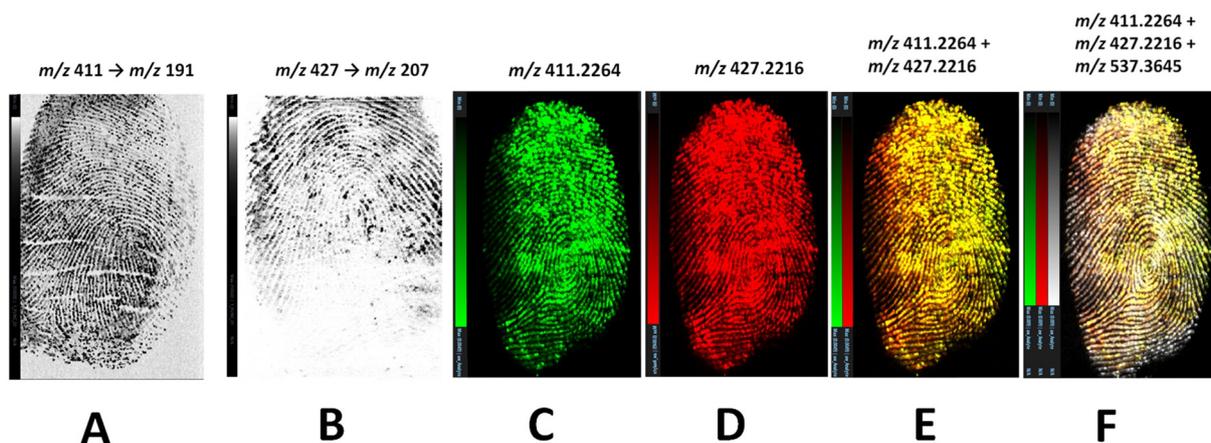
### 3.2 Mapping of risperidone and paliperidone in fingermarks via DESI MSI

Risperidone and paliperidone were initially analysed as standard solutions and in a serial dilution to assess feasibility of DESI MS for the detection and mapping of these species. Two DESI systems were used, namely a tandem quadrupole (Xevo TQ-S) and the DESI cIMS, both mounting a DESI XS source. Initial LOD experiments were performed on the DESI – XEVO TQ-XS using the transitions 411 → 191 (risperidone) and 427 → 207 (paliperidone) for drug standards yielding an LOD for Risperidone of 5 ng mL<sup>-1</sup> and a LOD for paliperidone of 500 ng mL<sup>-1</sup>. However, when a natural fingerprint was spotted with a serial dilution of either risperidone or paliperidone and profiled, a higher concentration of fingertip contaminating solution of  $5 \times 10^3$  ng mL<sup>-1</sup> was taken forward to imaging experiments, due to the ion suppression exerted by the molecular composition of the fingerprint. Under these concentration conditions, risperidone-, paliperidone- and risperidone/paliperidone contaminated-fingermarks were generated and analysed on either the DESI-Xevo TQ-XS mass spectrometer or the DESI-cIMS mass spectrometer. Using the grading scheme by Bandey,<sup>34</sup> which assesses the quality of the fingerprint from a biometric intelligence point of view, a grade 4 fingerprint image was yielded when visualising risperidone using the transition 411 → 191 (Fig. 2B) and a grade 2 image was instead obtained for paliperidone using the transition 427 →





**Fig. 1** MALDI MSI of risperidone/paliperidone contaminated fingerprint. (A) shows the image of distribution of risperidone (RIS) at  $m/z$  411.22046, from a spiking quantity of 250 ng. (B) shows the image of distribution of paliperidone (PAL) at  $m/z$  427.21576, from a spiking quantity of 0.25 ng and (C) the superimposition of the images of distribution of these two ions. (D) shows the superimposed images of distribution of RIS ( $m/z$  411.22046), PAL ( $m/z$  427.21576) and a ubiquitous contaminant at  $m/z$  550.63031 (dimethyldioctadecylammonium ion).



**Fig. 2** DESI MSI of risperidone and paliperidone in drug contaminated fingermarks on glass slides. Panels A and B show the DESI visualisation at  $50 \mu\text{m} \times 50 \mu\text{m}$ , on the Xevo-TQ-XS system, of risperidone and paliperidone at a fingertip contamination solution concentration of  $5 \mu\text{g mL}^{-1}$ . Panels C–F show the DESI images obtained on a DESI-cIMS in one experiment at  $50 \mu\text{m} \times 50 \mu\text{m}$ , using a fingertip contaminating solution containing both drugs at a final concentration of  $5 \times 10^3 \text{ ng mL}^{-1}$  each as for the MALDI MSI experiments; (C) risperidone at  $m/z$  411.2264 (green), (D) paliperidone at  $m/z$  427.2216 (red), (E) Risperidone/paliperidone superimposed ion images, (F) risperidone/paliperidone/fingerprint endogenous component ( $m/z$  537.3645) combined ion image.

207 (Fig. 2B) on the DESI-Xevo TQ-XS mass spectrometer. Observing Fig. 2B, there is no technology reasoning to explain that the mass spectrometer did not visualise paliperidone in the bottom half of the mark; rather, it can be speculated that

it could be due to either inhomogeneous rubbing of the fingertip on the dried paliperidone solution, or to important ion suppression exerted by molecules predominantly present in the bottom part of the (natural) mark, or to a combination of



both factors. Performing DESI imaging experiments on a tandem quadrupole mass spectrometer has the benefit of increased sensitivity and speed of analysis with minimum effect on sensitivity, as specific ions are targeted through SRM or MRM experiments. However, the molecular target, and dominant fragment ions, must be known and there is a finite number of parent ions and ion fragments that can be selected in any one experiment. If one aims for the visualisation of forensically interesting substances onto the ridge pattern, and additionally the visualisation and superimposition of all those ions (whether known or unknown) that provide completeness of the ridge pattern in a combined image, then MS imaging, rather than MS/MS imaging is more appropriate. Fig. 2C–F show the DESI MS images obtained on a cIMS mass spectrometer in one experiment, where the fingerprint was contaminated with a 1 : 1 ratio solution of risperidone ( $m/z$  411.2254) and paliperidone ( $m/z$  427.2216). Here, it can be seen that it is possible to obtain separate images from risperidone, paliperidone and their superimposition. Whilst it is possible to obtain these images in MRM on the DESI TQ-XS, using MS imaging on the cIMS, it has been possible to superimpose a third ion ( $m/z$  537.3645) (Fig. 2F), unknown and endogenous to the fingerprint, in order to improve the ridge pattern continuity and increase the ridge flow/*minutiae* thus increasing the grade of the mark obtained. In a real case scenario, the presence of the metabolite or the drug and the metabolite together in a mark would indicate consumption of the drug, whereas the presence of the drug only may indicate drug handling. However, it is important to be mindful of the fact that there are other reasons for detection or lack thereof of the drug/metabolite, which may affect interpretation of the findings, not only for DESI MSI but for MALDI MSI and any other detection/imaging technique. For example, lack of detection of the parent drug in the case of consumption, could be due to extensive individual metabolism, though the detection of metabolite still may indicate consumption activity. However, there is still the possibility that the detection of the parent drug and/or the metabolite could be due to secondary transfer. Therefore whilst the data presented thus far and further in the manuscript prove feasibility of the approaches for the provision of both the chemical and biometric information from an alternative specimen such as fingerprints, interpretation of the source level information (why the species are there or absent) requires a framework and a quantitative/objective approach that are not currently available.

### 3.3 Mass spectrometry using SICRIT ionisation source

In the view of rapidly detecting risperidone and paliperidone in fingerprints, SICRIT ionisation was explored.

Source parameters were optimised and both the amplitude and the temperature were observed to significantly impact ionisation. It was determined that an amplitude of 1600 V and a Temperature of 300 °C were optimal for analysis. Initially, standard solutions of either risperidone or paliperidone were analysed using the injection module to assess detectability and the limits of detection as described in section 2.3.3. In MS mode, an LOD of 100 ng mL<sup>-1</sup> (absolute amount of 500 pg),

for both the parent drug and paliperidone was observed. Compared to the LOD in DESI MS analysis on the Xevo-TQ-XS mass spectrometer, SICRIT ionisation showed to be slightly less sensitive for risperidone by a factor of 20 but slightly more sensitive for paliperidone by a factor of 5.

Risperidone and paliperidone were subsequently and separately spotted as 10 μL of a serial dilution from 1 × 10<sup>5</sup> ng mL<sup>-1</sup> to 1 ng mL<sup>-1</sup> on a natural fingerprint, to assess the impact of any ion suppression caused by the molecular composition of the fingerprint (Fig. 3). Risperidone and paliperidone were detected at  $m/z$  411.223 and  $m/z$  427.225 respectively, down to 100 ng mL<sup>-1</sup> (absolute amount 1 ng), with a signal to noise of >167 for risperidone and >119 for paliperidone. Compared with the SICRIT analysis of these species as standards through direct infusion, there is only a factor of 2 decrease in the ability to detect risperidone and paliperidone when analysed in spots on fingerprints.

Again, compared to the spotted fingerprint experiments in DESI MSI, SICRIT ionisation was more sensitive for risperidone by a factor of 50 and equally sensitive for paliperidone (better only by a factor of 2).

Compared to MALDI MSI fingerprint spotting experiments, where risperidone could be detected/mapped down to 1 ng mL<sup>-1</sup> for risperidone and to 10 ng mL<sup>-1</sup> for paliperidone, SICRIT ionisation was less sensitive by a factor of 100 for risperidone and by a factor of 1000 for paliperidone (2 and 3 orders of magnitude respectively).

When employing SICRIT ionisation for risperidone and paliperidone in fingerprints, ion suppression can be invoked. Fig. 3 shows (i) the interference/ion suppression exerted by fingerprint endogenous species at  $m/z$  411.350 and  $m/z$  411.403 (the latter likely squalene, theoretical  $m/z$  411.399) being problematic for risperidone and (ii) the interference/ion suppression of the ions at  $m/z$  427.261 and  $m/z$  427.406 affecting the paliperidone detection; these ion interferences indicate that, for these particular species, to fully exploit the potential of SICRIT ionisation, this source should be coupled with a higher mass resolution mass spectrometer (as in the work by Conway *et al.*<sup>22</sup>) in order to improve the detection limits.

MS/MS analyses for both analytes were performed at a concentration of 100 μg mL<sup>-1</sup> to investigate the formation of the most abundant ion fragments, which were observed to be at  $m/z$  191.117 (C<sub>11</sub>H<sub>15</sub>N<sub>2</sub>O) and  $m/z$  207.118 (C<sub>11</sub>H<sub>15</sub>N<sub>2</sub>O<sub>2</sub>) respectively (data not shown) confirming previous studies.<sup>35,36</sup>

### 3.4 Sequential imaging – SICRIT ionisation followed by mass spectrometry imaging

A multimodal approach was evaluated by employing SICRIT Ionisation as part of a sequential workflow. In one instance, SICRIT was followed by MALDI MSI and, in a subsequent experiment, it was followed by DESI MSI and then by MALDI MSI. The rationale behind trialling these approaches was that, provided that this ionisation source is coupled to HRMS (as in the work by Conway *et al.*<sup>22</sup>), these workflows would permit fast drug detection at a higher sensitivity, followed by the recovery of biometric intelligence using MSI.



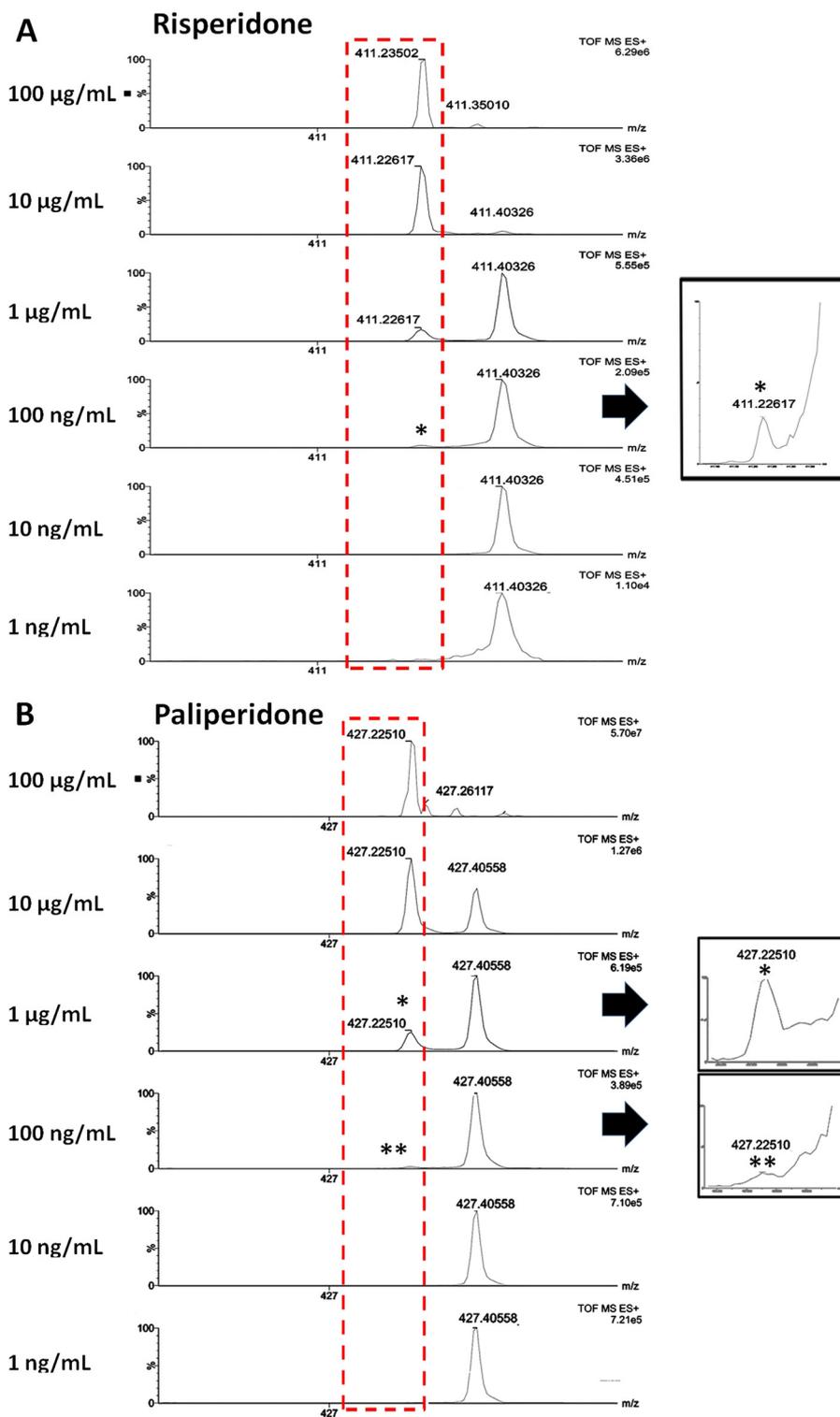


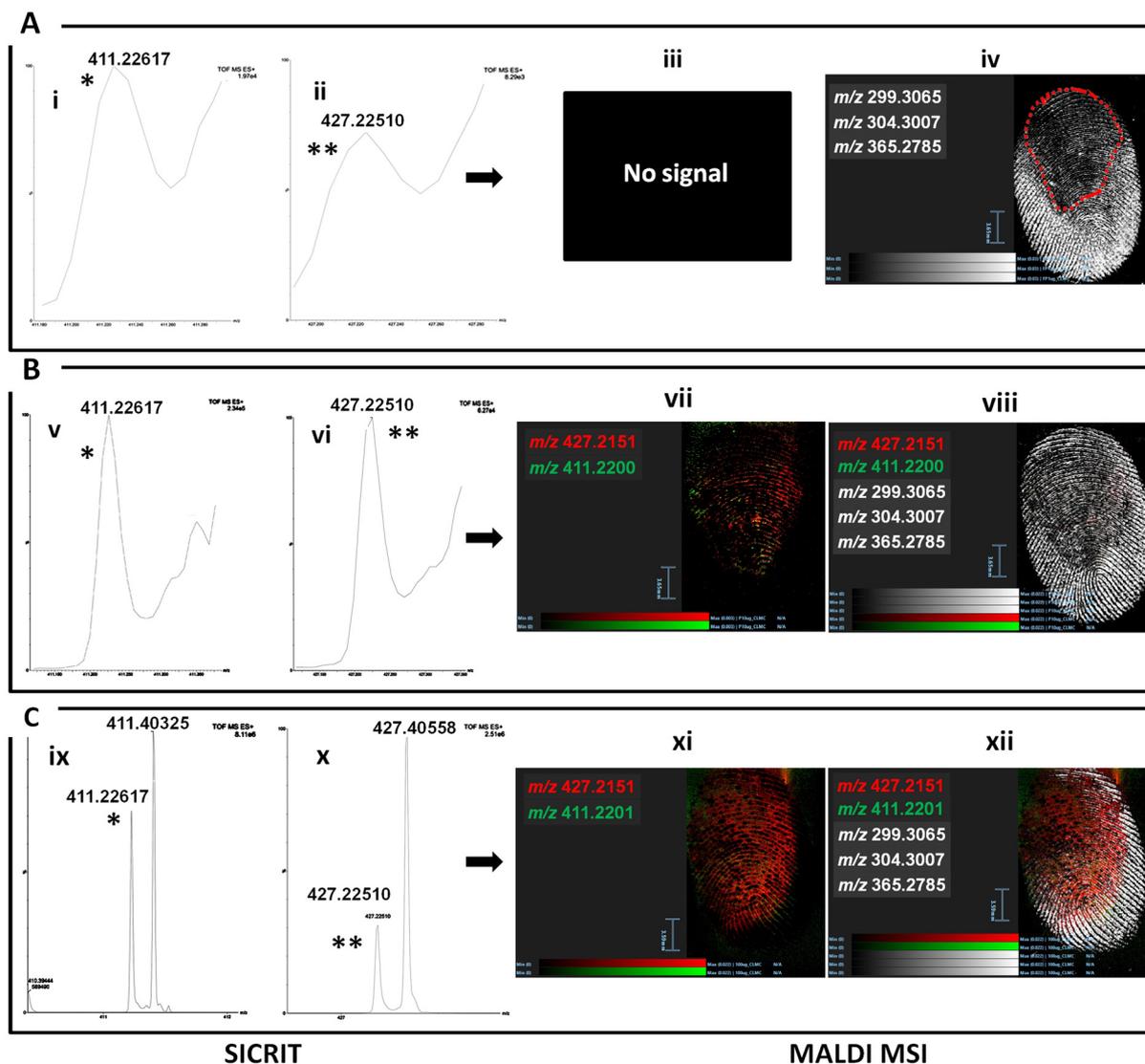
Fig. 3 SICRIT ionisation of serial dilutions of risperidone (A) and paliperidone (B) spotted in fingermarks. Both risperidone and paliperidone were detectable down to  $100 \text{ ng mL}^{-1}$ . The inset \* in both panels A and B show a viable risperidone and paliperidone ion signal, respectively.

### 3.5 SICRIT → MALDI MSI

For this workflow, drug-contaminated fingermarks were generated after preliminarily spotting on a glass slide  $50 \mu\text{L}$  of ris-

peridone and paliperidone mixing  $25 \mu\text{L}$  of each solution in equal concentrations varying between  $1 \times 10^5 \text{ ng mL}^{-1}$  and  $0.1 \text{ ng mL}^{-1}$ . Fig. 4 shows the MALDI MSI analyses for the drugs at an initial concentration of  $1 \times 10^3$ ,  $1 \times 10^4$  and  $1 \times 10^5 \text{ ng mL}^{-1}$ .





**Fig. 4** MALDI MSI analysis of a Risperidone/Paliperidone-contaminated fingerprint after SICRIT ionisation. Panel A shows the SICRIT mass spectra of risperidone at  $m/z$  411.226 (i) and paliperidone at  $m/z$  427.225 (ii) used to contaminate fingertips at initial concentrations of  $1 \times 10^3 \text{ ng mL}^{-1}$  and the subsequent MALDI MSI images showing no retrievable risperidone/paliperidone ion maps (iii) but a partial mark reconstructed by visualising the ions at  $m/z$  299.30627, 304.30057 and 365.27905 (iv). Image iv also shows the region delimited by a discontinuous red line where the SICRIT ion desorption has mostly occurred. Panel B shows the SICRIT mass spectra of risperidone at  $m/z$  411.226 (v) and paliperidone at  $m/z$  427.225 (vi) used to contaminate fingertips at initial concentrations of  $10 \mu\text{g mL}^{-1}$ , and the subsequent MALDI MS images showing an overlay of risperidone in green at  $m/z$  411.21973 and paliperidone in red (vii)  $m/z$  427.2151 and further reconstruction of the ridge pattern (viii) as for image iv. Finally, Panel C shows the SICRIT mass spectra of risperidone at  $m/z$  411.21973 (ix) and paliperidone at  $m/z$  427.21426 (x) used to contaminate fingertips at initial concentrations of  $100 \mu\text{g mL}^{-1}$ , and the subsequent MALDI MS images showing an overlay of risperidone in green at  $m/z$  411.21973 (xii) and paliperidone in red  $m/z$  427.21426 (xi) and further reconstruction of the ridge pattern (viii) as for images iv and viii. (\* and \*\* indicate in the SICRIT spectra the ion signals for risperidone and paliperidone, respectively).

$\text{mL}^{-1}$ . In the case of the drugs concentrations at  $1 \times 10^3 \text{ ng mL}^{-1}$ , the SICRIT ionisation yielded an ion signal for risperidone at  $m/z$  411.226 (i) (S/N 80) and paliperidone at  $m/z$  427.225 (ii) (S/N 33) (absolute amount  $<5 \text{ ng}$ ). However, when subsequently imaged by MALDI MSI, none of the two species yielded a viable molecular image and this may be due to either presence below the detection limits, or because to the way in which the encoded frequency pushing algorithm works.<sup>37</sup> It is also likely that the prior desorption of this drug *via* SICRIT

ionisation made risperidone detection in fingerprints fall below the detection limits on the MALDI MRT. Nonetheless the fingerprint ridge pattern could be reconstructed (Fig. 4A(iv)) through superimposition of the fingerprint ions at  $m/z$  299.30627, 304.30057 and 365.27905, thus showing the possibility to obtain both the chemical and the biometric information with a sequential workflow. Interestingly, Fig. 4A(iv) shows the region delimited in a discontinuous red line where most of the desorption of risperidone and other

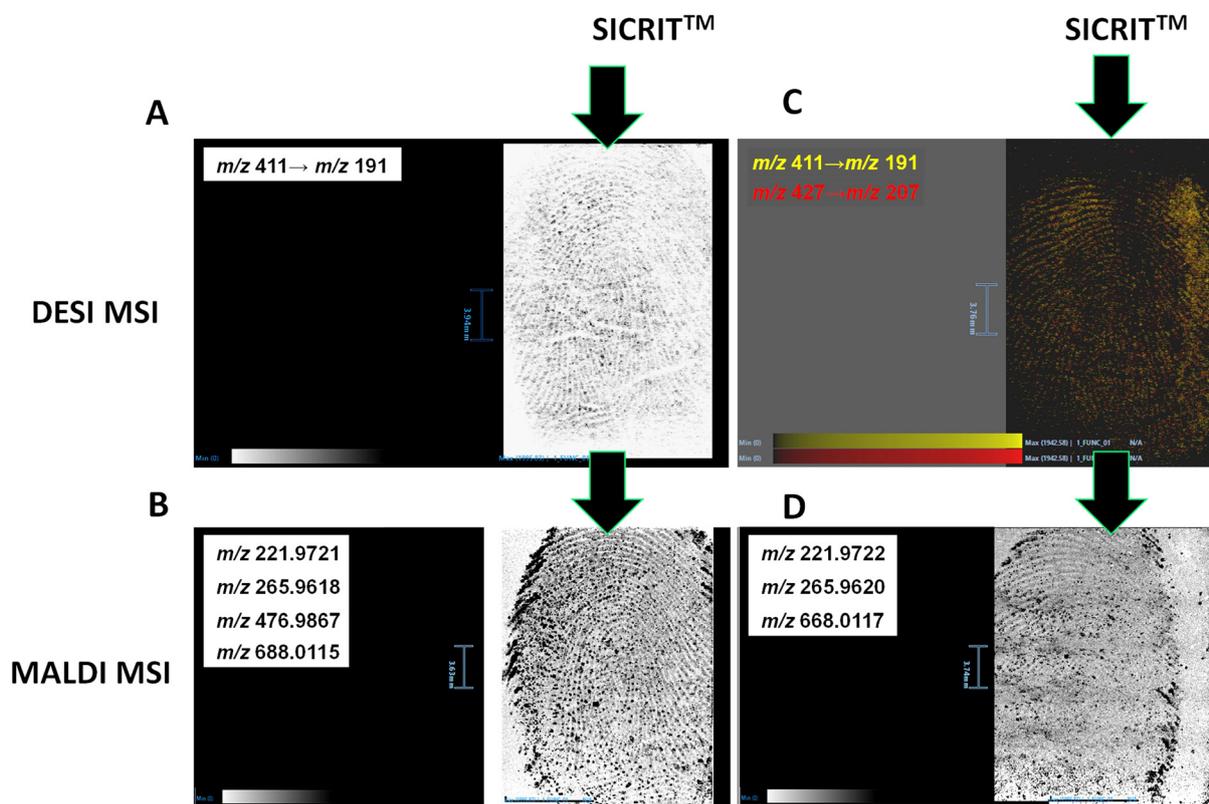


analytes in fingerprints appears to occur as a result of SICRIT ionisation prior to MALDI MSI. This region indicates an inhomogeneous SICRIT desorption process and perhaps the necessity to further engineer the source, to either enable a homogeneous desorption, or to confine the location of the desorption to a small region, in order to enhance the likelihood of both higher quality biometric information (more *minutiae* in a fingerprint) and the recovery of chemical intelligence (in this case risperidone detection). When this experiment was repeated with a fingerprint contaminated by mixing the two drugs' solutions at  $1 \times 10^4$  ng mL<sup>-1</sup> each, both risperidone and paliperidone were detected at the same  $m/z$  as previously (Fig. 4B(v-vi)). This fingerprint was then subjected to MALDI MSI and in this instance, the higher starting concentrations of the two species reduced the impact on MALDI MS visualisation by permitting both risperidone at  $m/z$  411.21973 and paliperidone at  $m/z$  427.21426 to be mapped in an overlaid fashion, onto the fingerprint ridge pattern (Fig. 4B(vii)). The further superimposition with ions at  $m/z$  299.30627, 304.30057 and 365.27905 enabled a much better ridge coverage with respect to the experiments with the drugs' concentrations at  $1 \times 10^3$  ng

mL<sup>-1</sup>, although the "SICRIT desorption region" is still visible and superimposable with that of the experiment conducted with the two drugs at  $1 \times 10^3$  ng mL<sup>-1</sup>. Finally the experiment was conducted with a fingerprint generated by initially mixing the two drugs' solutions at 100  $\mu$ g mL<sup>-1</sup> each; the higher concentrations of both drugs further improve both their ionisation and mass resolution using SICRIT, as well as the quality of the resulting MALDI MS image of the fingerprints reconstructed through the superimposition of these two ions (Fig. 4Cxi) and by superimposing further ions as in the previous examples (Fig. 4Cxi).

### 3.6 SICRIT $\rightarrow$ DESI MSI $\rightarrow$ MALDI MSI

For this sequential workflow, two experiments were performed, one with a fingerprint generated from fingertips contaminated with risperidone only at a concentration of 500 ng mL<sup>-1</sup>, and one with a fingerprint generated from fingertips contaminated with risperidone and paliperidone in a 1 : 1 ratio risperidone/paliperidone (each in a concentration of  $1 \times 10^4$  ng mL<sup>-1</sup>). DESI MS and MALDI MS images were generated in sequence following SICRIT ionisation (Fig. 5).



**Fig. 5** Sequential Imaging of Risperidone-contaminated fingerprint via SICRIT  $\rightarrow$  DESI MSI  $\rightarrow$  MALDI MSI. After MS profiling and detection of risperidone in a risperidone-contaminated fingerprint using SICRIT ionisation as per previous experiments (data not shown), risperidone was imaged via DESI – on the Xevo TQ-XS mass spectrometer. An MS image of risperidone, was obtained using the transition  $m/z$  411  $\rightarrow$  z 191 (A), reconstructing the fingerprint ridge pattern. The same fingerprint was imaged by MALDI MSI which yielded complementary ridge pattern reconstruction through the overlay of several fingerprint endogenous ions at  $m/z$  221.97234, 265.96210, 476.98682 and 688.01215 (B). The same workflow was applied to a risperidone/paliperidone contaminated fingerprint. After detection of these species through SICRIT ionisation (data not shown) both species were detected and imaged via DESI MSI using the transitions  $m/z$  411  $\rightarrow$  191 and  $m/z$  411  $\rightarrow$  207 (C). Having repeated this analysis twice, MALDI MSI did not yield any useful ridge detail (D).



In the first experiment, the risperidone-contaminated mark was preliminarily submitted to SICRIT Ionisation which did yield the risperidone ion signal at  $m/z$  411.219. This detection was expected as detection of risperidone in spotted marks was possible down to a concentration of  $100 \text{ ng mL}^{-1}$  with no ion interference, as Fig. 3 reported. This mark was then subjected to DESI MSI on the Xevo-TQ-XS mass spectrometer.

As Fig. 5A shows, the fingerprint ridge pattern could be reconstructed through the visualisation of risperidone using the transition  $m/z$  411  $\rightarrow$  191.

Previously, DESI MSI on the XEVO-TQ-XS yielded the images of distribution of risperidone in a drug-contaminated mark, using the same transition, with a fingertip contaminating solution of  $500 \text{ ng mL}^{-1}$ . However, this time, despite some risperidone being desorbed during SICRIT ionisation, it was still possible to detect the drug and reconstruct the fingerprint ridge pattern through visualising its MRM transition. The same mark was then submitted to MALDI MSI *via* the prior application and adaptation of the dry-wet method<sup>38,39</sup> in the attempt to provide complementary biometric information; here the mark was preliminarily powdered with ball milled  $\alpha$ -cyano 4 hydroxycinnamic acid average size of  $10 \mu\text{m}$  using the planetary ball mill (Retsch, GmbH), following the method of Ferguson *et al.*<sup>38</sup> and then it was spray coated with a  $5 \text{ mg mL}^{-1}$   $\alpha$ -CHCA solution in 70/30 ACN/0.1% TFA<sub>aq</sub> solution.<sup>39</sup> The mark was then imaged by MALDI MSI on the SELECT SERIES MALDI MRT.

At this stage, risperidone had been desorbed first *via* SICRIT Ionisation and then through DESI MSI. The lack of MALDI ion signal and ion image for this drug is therefore expected, considering also that the minimum concentration that could be imaged in a single drug process workflow using MALDI MSI was  $1 \times 10^3 \text{ ng mL}^{-1}$  (data not shown). However, the dry-wet method enabled the recovery of a combined MALDI MS ion image showing high quality level 1 and level 2 characteristics (ridge flow and local fingerprint features, respectively), thus adding to the biometric information (Fig. 5B). The MALDI image has been obtained through the superimposition of several ions at  $m/z$  221.97234, 265.96210, 476.98682 and 688.01215. In the second experiment, fingertips were contaminated with a 1 : 1 mixed solution of risperidone and paliperidone initially at a concentration of  $0.5 \mu\text{g mL}^{-1}$ , prior to contaminating a fingertip and depositing a drug/metabolite-contaminated fingerprint on a glass slide. The glass slide was inserted in the heated cartridge and subjected to SICRIT ionisation and subsequently to DESI MSI on the XEVO TQ-XS mass spectrometer. After SICRIT Ionisation, the imaging analysis still yielded the reconstruction of the ridge pattern through the superimposition of the ion images from risperidone and paliperidone using the transitions  $m/z$  411  $\rightarrow$  191 and  $m/z$  427  $\rightarrow$  207 (Fig. 5C). However, this time, the DESI analysis shown was a second acquisition. Whilst it was observed that viable signal was still detected and mapped onto the ridge pattern in this second DESI MSI acquisition, the subsequent MALDI MSI experiment did not yield viable ridge pattern (Fig. 5D) as evidently, too much of the fingerprint con-

stituents had been already desorbed. A 3-techniques multi-modal workflow would only be recommended if the first MSI technique (1) fails to visualise the drug/metabolite onto the ridge pattern and/or (2) does not provide sufficient biometric information for submission to biometric identification technology such as AFIS, for fingerprint comparison and match in a National Database.

### 3.7 Chemical enhancement $\rightarrow$ MALDI MSI/DESI MSI of risperidone and paliperidone in fingerprints

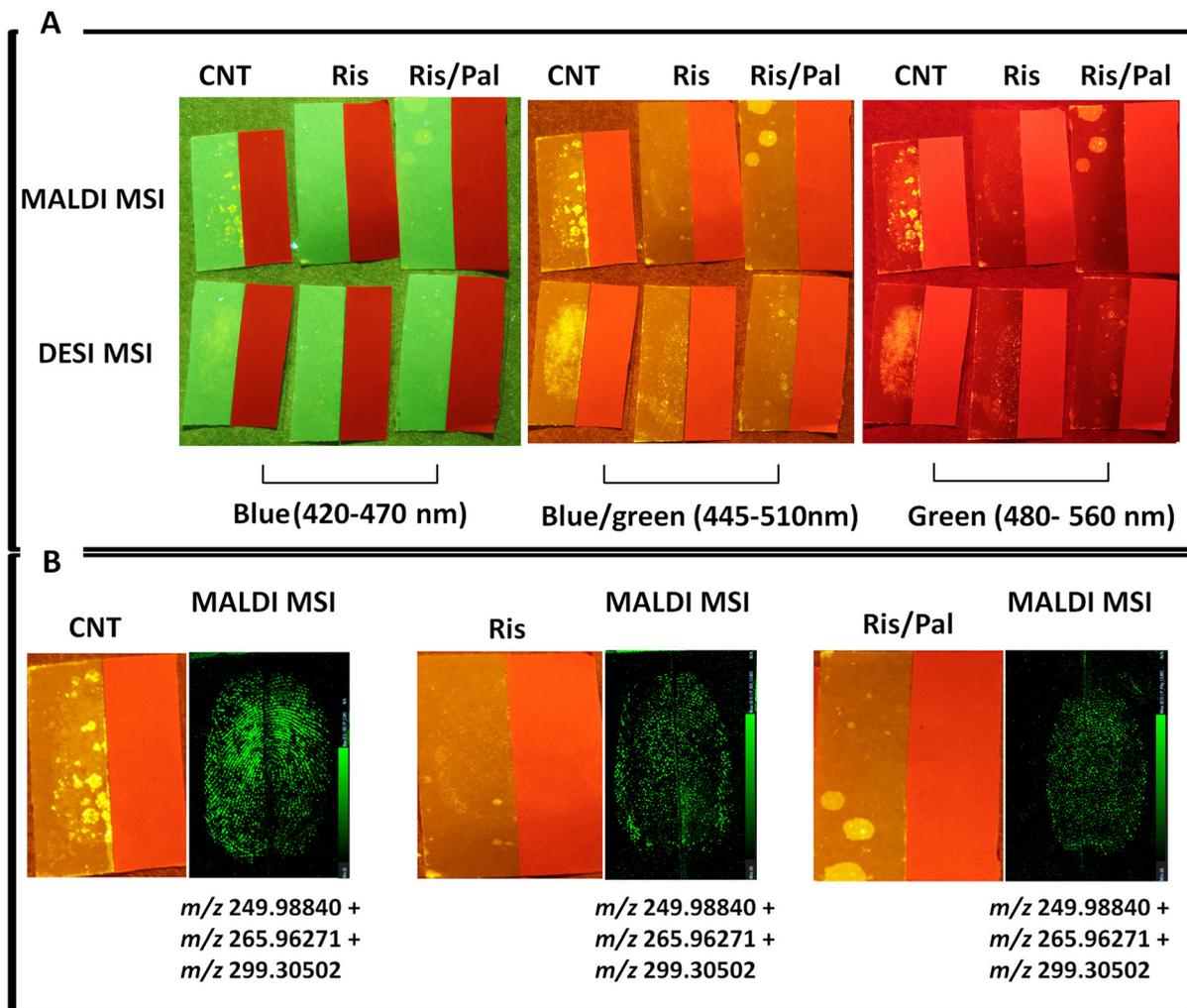
Chemical enhancement with 1,2-indandione was applied to split contaminated fingerprints as shown in Fig. 6. Six samples were prepared and chemically enhanced, three for the subsequent application of DESI MSI and 3 for the subsequent application of MALDI MSI. For each set, a control mark (with no risperidone or paliperidone contamination), one contaminated with risperidone only and one with a mixture of risperidone and paliperidone in a 1 : 1 ratio.

The application of 1,2-indandione and the visualisation through irradiation at 3 different wavelengths (Fig. 6A) yielded: (i) some level of visualisation on the white paper ranging for the two CNT marks and for only one of the risperidone only contaminated marks using blue light (420–470 nm); (ii) some level of visualisation ranging from grade 1 to 4 of the Home Office system for all the marks on white paper using blue/green light (445–510 nm) and (iii) same results as for the blue/green light were obtained using green light (480–560 nm), albeit the quality was slightly worse than the blue/green light. No visible fingerprint on red paper could be observed for any of the wavelengths used or different types of fingerprints.

The subsequent application of DESI MSI did not yield any fingerprint image (data not shown) and although MALDI MSI enabled visualisation of the fingerprint through its endogenous content, neither risperidone nor paliperidone were detected; Fig. 6B shows the superimposed MALDI MS images of the endogenous species at  $m/z$  249.98840, 265.96271, 299.30502 to reconstruct the ridge pattern reconstruction.

These observations demonstrated compatibility of MALDI MSI in the retrieval of biometric information after the application of this forensic enhancement technique, but not of the chemical intelligence, with respect to the drug and its metabolite. The absence of risperidone/paliperidone signal in the relevant marks on either white or red paper was ascribed to the use of a high impact chemical enhancement process requiring liquid immersion; furthermore this protocol is based on the Category A process formulation given in the UK Home Office Fingerprint Visualisation Manual (FVM),<sup>23</sup> but uses Petroleum ether as a 1 : 1 replacement for the recommended HFE7100 solvent (which ceases production in 2025); it is possible that the combination of liquid immersion and a solvent mixture in which both risperidone and paliperidone are soluble have led to these species being washed away, explaining absence of signal in both mass spectra and DESI/MALDI images. It is also interesting to note that from the chemical enhancement point of view, the use of 1,2-indandione produces the no or poor fingerprint visualisation (on





**Fig. 6** 1,2-Indandione enhancement of split marks on white and red paper followed by Mass Spectrometry Imaging. (A) It can be observed that, whilst the Blue/green wavelength (445–510 nm) yielded the best visualisation of all marks on white paper, in no circumstance could fingermarks be observed on red paper. (CNT, a latent mark; RIS mark contaminated with Risperidone; RIS/PAL mark contaminated with Risperidone and Paliperidone in a physiological ratio were deposited). (B) MALDI MS images of the CNT, RIS and RIS/PAL marks showing reconstruction of the ridge pattern through three the superimposition of distributions of three ions at  $m/z$  249.98840, 265.96271, and 299.30502 as neither RIS or PAL could be detected or mapped.

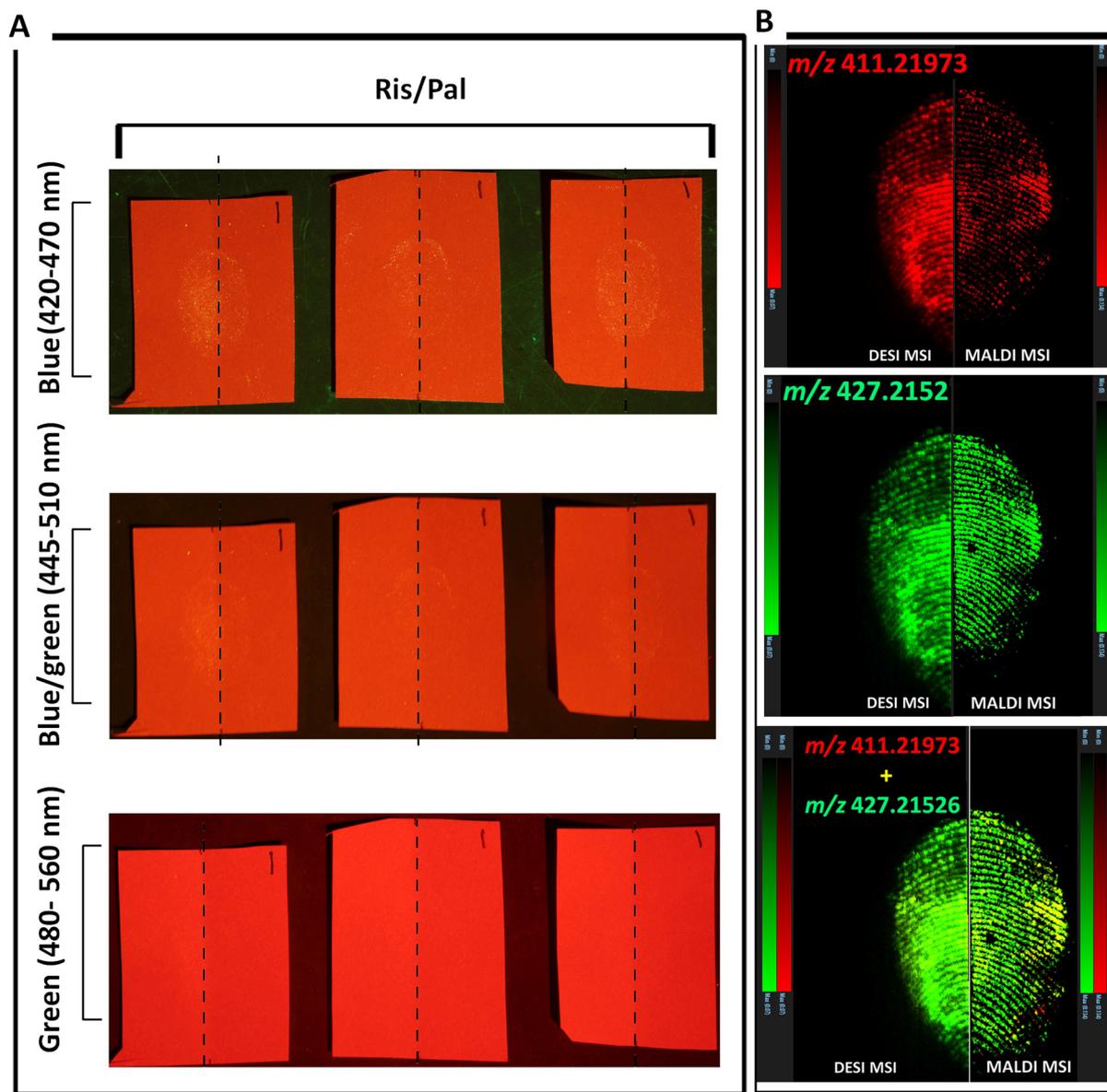
white paper) when Risperidone or Risperidone and Paliperidone are present. In consideration of the fact the red paper did not produce visualisation at any wavelength irradiation and that the hypothesised detrimental effect on MALDI MSI and DESI MSI signal caused by the liquid immersion process, another (lower impact) protocol was tested using a “drier” method consisting in the use of DMAC impregnated sheets (FVM, Cat B process<sup>23</sup>). The protocol was only applied to fingermarks on red paper to demonstrate the benefits of the subsequent application of mass spectrometry imaging techniques when the conventional forensic enhancement fails.

After enhancement and photography, the red paper was split and one-half mark was subjected to DESI MSI and the other half to MALDI MSI. Fig. 7 shows the chemical enhancement results with DMAC (Fig. 7A) and the subsequent visualisation using either DESI MSI or MALDI MSI (Fig. 7B).

This time, as ESI Fig. S3† shows with better clarity, the combination of DMAC enhancement and Blue light irradiation (420–470 nm) produced by and large the highest quality of visualisation (up to grade 3 for the control mark), though grade 1 marks could be observed with Blue/green wavelength (445–510 nm) but no mark at all was visible when using the green wavelength (480–560 nm), possibly due to the increased background fluorescence of the red paper at these wavelengths.

Following DMAC enhancement, all marks were split, and one half was imaged using DESI MSI on a cIMS mass spectrometer and the other half using MALDI MSI on the SELECT SERIES MALDI MRT (Fig. 7B). As a representative example, Fig. 7B shows 3 DESI/MALDI images of the first paper specimen from the left handside and ion images for both DESI MSI and MALDI MSI are shown for the ion at  $m/z$  411.21973 (risper-





**Fig. 7** Sequential Imaging of marks contaminated with risperidone/paliperidone (1 : 1) solution. Risperidone/paliperidone-contaminated fingerprint were deposited in triplicates on red paper, enhanced with DMAC and visualised under 3 wavelengths (A). As a representative example, (B) shows 3 DESI and 3 MALDI images from split marks on the first paper specimen from the left-hand side. Ion images for both DESI MSI and MALDI MSI are shown for the ion at  $m/z$  411.21973 (risperidone),  $m/z$  427.21526 (paliperidone) and the combined image resulting from the superimposition of the images for these two ions.

idone),  $m/z$  427.21526 (paliperidone) and the combined image resulting from the superimposition of the images for these two ions. Different from the enhancement with 1,2-indandione, this time the fingerprint ridge pattern could be reconstructed through visualising both drugs. This indicates that the immersion in a solvent in which the two species could be soluble, is too aggressive and supports the hypothesis that it may have washed risperidone and paliperidone away. The dry, vapour phase DMAC enhancement process will have preserved the presence of these two species, making possible the visualisation of their distribution through both DESI and MALDI. All

the DESI MS and MALDI MS images were TIC normalised. It was observed that whilst, generally the relevant ion signals were more intense in the DESI images, the quality of the ridge detail was better in the MALDI images, despite nominally using the same spatial resolution.

## 4. Conclusions

In this paper we report an exploration of mass spectrometry imaging-based approaches as single processes or in combi-



nation for the detection and imaging of “date rape drugs” in fingerprints, using risperidone and its pharmacologically active metabolite paliperidone as a case study. Both MALDI MSI and DESI MSI provided contextual chemical information (drug and metabolite detection mapped on the ridges) and biometric information (reconstruction of the ridge pattern), thus expanding the capabilities of “molecular fingerprinting”, particularly those offered by MALDI MSI. DESI MSI was trialled both coupled with a tandem quadrupole and with a Cyclic Ion Mobility Mass Spectrometer, with the former offering naturally higher sensitivity, due to its targeted approach exploiting specific MRM transitions. A number of multi-modal approaches were tested, including the combination with Chemical Reaction In Transfer Ionisation *via* SICRIT® which had been shown by others to detect drugs of abuse in the picogram range in fingerprints. It was generally possible to achieve a quick drug detection in the fingerprint using SICRIT, subsequent risperidone/paliperidone mapping *via* DESI MSI and finally, additional biometric details using MALDI MSI in a sequential approach.

The approach illustrated here is described as a means to corroborate the victim’s statement through the detection and imaging of the drug and metabolite in the victim’s fingerprints. However, the approach is equally important if applied to the suspect’s fingerprint; when not using needle spiking, in fact, the perpetrator would dissolve the drug in the drink and, as such, they may contaminate their fingertip and transfer the parent drug to their fingerprint upon contacting a surface. In this case too, it is possible to provide circumstantial evidence to be linked to an individual identity. In the case of the perpetrator, the amount of drug contaminating the fingertip is in the order of milligrams, and, as such, well within the detection limits afforded by MALDI MSI and DESI MSI using the cutting-edge instrumentation reported in this work. However, as for any detection/imaging analytical technique providing information at a source level, a word of caution is needed in relation to activity level propositions, especially with regards to the potential suspect. Secondary transfer could in fact be the reason why the drug is detected in someone’s fingerprints. Presently, the forensic and analytical community still generally lacks a solid framework to assess and distinguish quantitatively and objectively between different activity level propositions and, in this case, the cut off levels separating transfer from genuine handling of the substance in question; therefore the source level information must be treated with this in mind and evaluated in association with the intelligence gathered by classic police investigation and other obtained forensic evidence.

Finally, in order to show the possibility to integrate mass spectrometry-based imaging approaches in an operational workflow, drug/metabolite contaminated fingerprints were deposited on paper and enhanced with either 1,2-indandione or DMAC (conventional enhancement techniques applied to fingerprints on paper), prior to DESI or MALDI MSI. Whilst both enhancement techniques struggled to visualise fingerprints on red paper, the “dryer” process (DMAC) yielded, depending

on irradiation wavelengths, some ridge detail, and was compatible with both DESI and MALDI MSI. Both MSI techniques provided much better biometric information in terms of clarity and number of fingerprint details (*minutiae*), through drug/metabolite mapping, demonstrating integrability of the mass spectrometry imaging techniques within a particular operational forensic workflow.

## Data availability

Data for this article including mass spectrometry images and mass spectra are available in the Sheffield Hallam repository SHURDA at [URL – format <https://shurda.shu.ac.uk/id/eprint/234>].

## Conflicts of interest

Waters Corporation has matched-funded the PhD studentship awarded to Mr Rohith Krishna, enabling the generation of these data on Waters mass spectrometers. Notwithstanding there is no conflict of interest perceived.

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