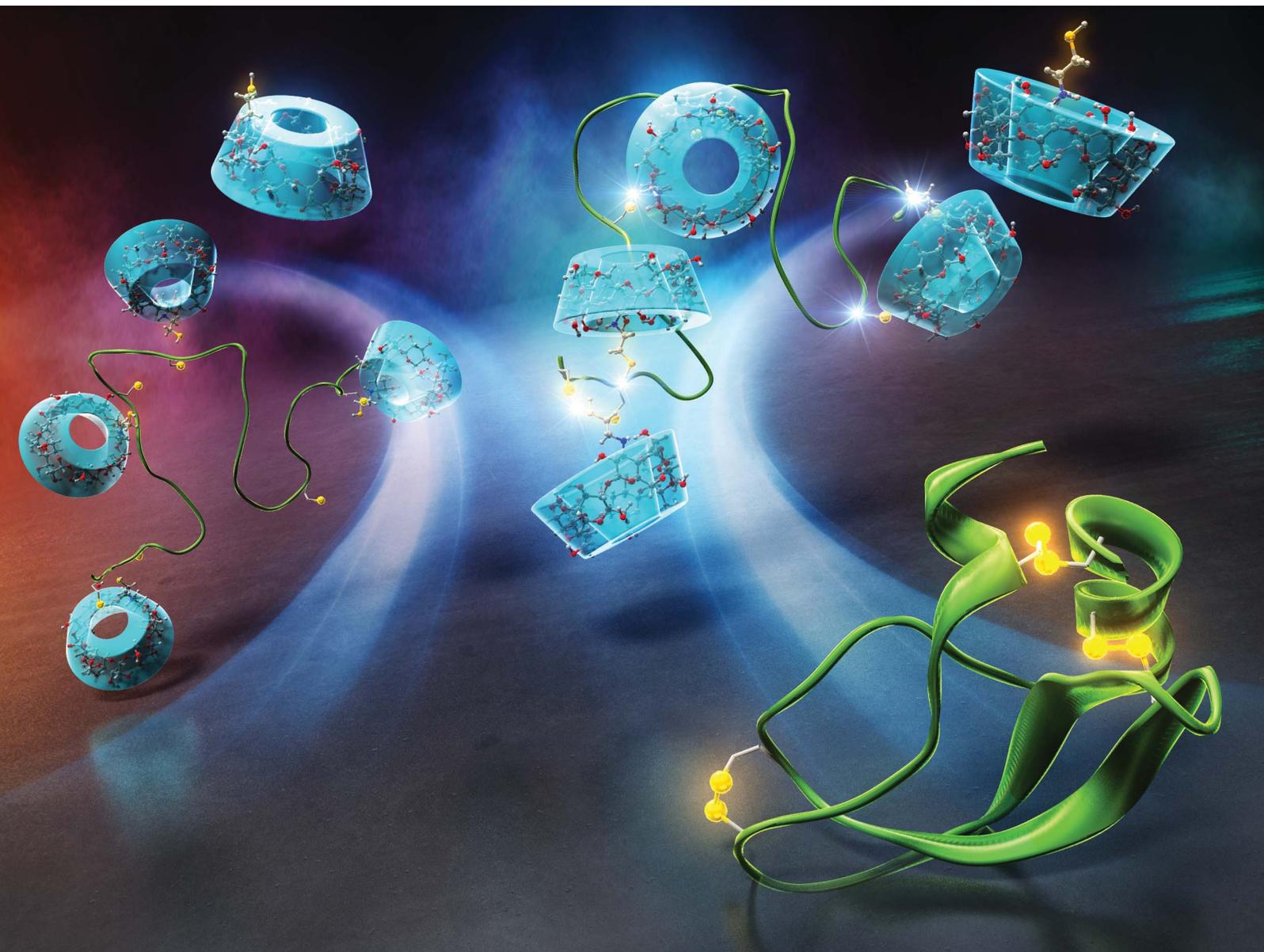


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EDGE ARTICLE

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Redox-active chemical chaperones exhibiting promiscuous binding promote oxidative protein folding under condensed sub-millimolar conditions†

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Proteins form native structures through folding processes, many of which proceed through intramolecular hydrophobic effect, hydrogen bond and disulfide-bond formation. *In vivo*, protein aggregation is prevented even in the highly condensed milieu of a cell through folding mediated by molecular chaperones and oxidative enzymes. Chemical approaches to date have not replicated such exquisite mediation. Oxidoreductases efficiently promote folding by the cooperative effects of oxidative reactivity for disulfide-bond formation in the client unfolded protein and chaperone activity to mitigate aggregation. Conventional synthetic folding promoters mimic the redox-reactivity of thiol/disulfide units but do not address client-recognition units for inhibiting aggregation. Herein, we report thiol/disulfide compounds containing client-recognition units, which act as synthetic oxidoreductase-mimics. For example, compound $\beta\text{CD}_w\text{SH/SS}$ bears a thiol/disulfide unit at the wide rim of β -cyclodextrin as a client recognition unit. $\beta\text{CD}_w\text{SH/SS}$ shows promiscuous binding to client proteins, mitigates protein aggregation, and accelerates disulfide-bond formation. In contrast, positioning a thiol/disulfide unit at the narrow rim of β -cyclodextrin promotes folding less effectively through preferential interactions at specific residues, resulting in aggregation. The combination of promiscuous client-binding and redox reactivity is effective for the design of synthetic folding promoters. $\beta\text{CD}_w\text{SH/SS}$ accelerates oxidative protein folding at highly condensed sub-millimolar protein concentrations.

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Introduction

Chemical reactions under highly condensed conditions often result in the formation of undesired kinetic byproducts due to multiple competitive factors. The promotion of condensed-phase reactions can increase the synthetic yield of drugs and materials, but currently there are few methodologies for

controlling the reactions of concentrated polypeptides due to their high propensities to aggregate in aqueous media. Protein folding is a biochemical pathway followed by each unique polypeptide sequence to construct the native structure of the folded protein. Many of these pathways proceed through cooperative intramolecular hydrophobic effect, and hydrogen bond and disulfide (SS) bond formation (Fig. 1a).¹ Under condensed conditions, protein folding competes with non-specific intermolecular interactions and SS bonding between multiple polypeptide chains to afford misfolded and aggregated proteins (Fig. 1a, Route A).² Over 20 members of the protein disulfide isomerase (PDI) family of oxidoreductases recognize nascent and unfolded reduced proteins such as proinsulin and immunoglobulin G and promote folding by redox reactions, thereby facilitating SS-bond formation. The chaperone activity of PDIs prevents protein aggregation in a cell (Fig. 1a, Route B).³ The interior of a cell is a typical condensed medium. Bio-inspired and enzyme-mimetic approaches could aid in the design of synthetic methodologies to regulate and promote condensed-phase polypeptide reactions.⁴

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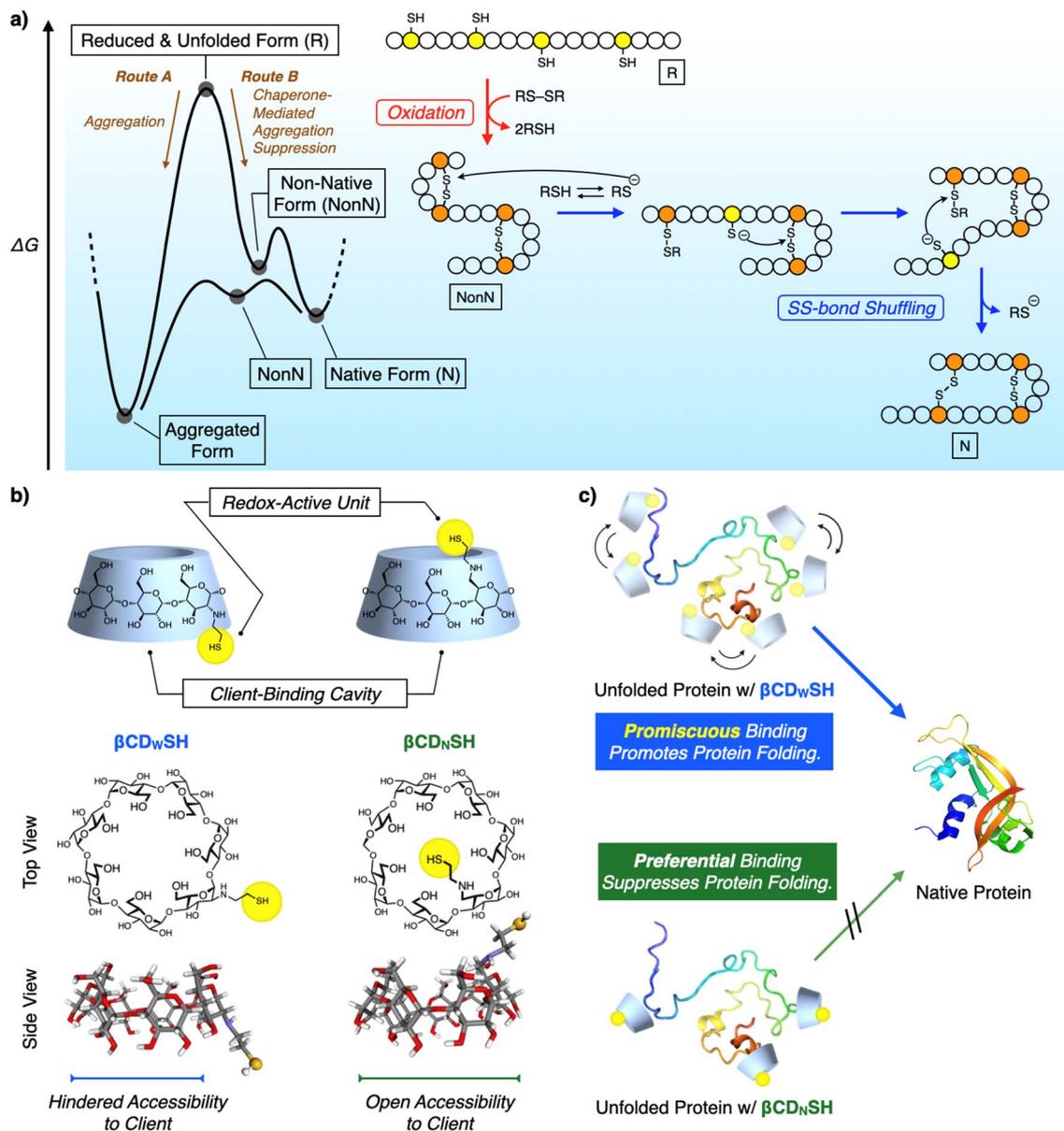


Fig. 1 (a) Schematic illustration of the oxidative folding of a polypeptide chain to form two disulfide bonds. Yellow and orange circles represent cysteine residues that are in the thiol/thiolate form and disulfide form, respectively. Open circles represent amino acid residues other than cysteine. Red arrow step: oxidation reaction from a reduced and unfolded form (R) to a non-native form (NonN). Blue arrow steps: disulfide bond isomerization from NonN to N by the nucleophilic attack of thiolates. (b) Molecular structures of β CD_WSH and β CD_NSH. Schematic images showing the redox-active units and client-binding cavities, and the molecular models calculated using DFT B3LYP 6-31G*. (c) The concept of (blue) the promiscuous binding property of β CD_WSH and (green) the preferential binding property of β CD_NSH with an unfolded protein. The promiscuous binding property of β CD_WSH is advantageous for promoting protein folding compared to the preferential binding property of β CD_NSH.

Enzymes are conventionally considered to be specific catalysts that recognize preferential substrates to provide single products.⁵ The well-defined structures of enzyme-substrate complexes have afforded important clues helpful for the design of synthetic mimics of enzyme recognition domains and active sites, a useful approach for developing drugs and catalysts.⁶ In contrast to such preferential recognition systems, enzymes that act as chaperones exhibit promiscuous binding.⁷ Oxidoreductases recognize unstructured proteins, and an active center

facilitates SS-bond formation in such unstructured proteins (called clients).³ The dynamic binding of promiscuous enzymes with a range of client unfolded polypeptides can address the conflicting demands for protein folding: namely, maintaining conformational mobility of the client polypeptide chains, allowing the formation of intramolecular covalent and non-covalent bonds, while simultaneously blocking intermolecular contacts. Such client-binding properties significantly influence the functionalities of folding catalysts. PDI and ERp46 are



representative oxidoreductases belonging to the PDI family. Both have thioredoxin-like domains as their active centers, but ERp46 promotes faster SS-bond formation in the client protein than does PDI.^{3c,8} The characteristic binding properties of the two enzymes are different, such as the promiscuous client-binding of ERp46 and the preferential recognition of a client's specific local structure by PDI. The different binding behavior of PDI and ERp46 likely leads to functional switches in their enzymatic activity, such as oxidative SS-bond formation and their anti-aggregation activity when acting as chaperones.⁹ Binding kinetics contribute to the functional characteristics of molecular chaperones such as SecB and trigger factor (TF). The higher k_{on} (client-binding rate constant) value for SecB than for TF enhances the holdase activity of SecB, while the moderate k_{on} value for TF allows it to exhibit both foldase and holdase activities, depending on the conditions affecting aggregation suppression.¹⁰ Such structure–activity analyses suggest the effectiveness of combining promiscuous recognition and redox activity to promote folding and suppress the aggregation of unfolded proteins.

Synthetic mimics of oxidoreductases have been developed to promote polypeptide folding into native structures. In conventional approaches, the mimic focuses on the enzymes' redox active centers for forming and rearranging the SS bonds of the client polypeptides. However, this approach often does not incorporate the binding units that prevent aggregation.¹¹ Protein folding using these approaches is generally conducted at micromolar concentrations, *i.e.*, dilute conditions to suppress the aggregation of unfolded and misfolded proteins. In contrast, here we report synthetic mimics of oxidoreductases that exhibit redox activity and recognize client unfolded polypeptides in a preferential and promiscuous manner (Fig. 1b and c). The enzyme-mimic with promiscuous binding ability significantly promoted folding and inhibited aggregation compared to the mimic exhibiting preferential binding. The former mimic promoted protein folding under condensed conditions, even at sub-millimolar concentrations.

Results and discussion

β -Cyclodextrin (β CD) is a representative synthetic receptor that binds with hydrophobic guest molecules *via* a concave pocket.¹² β CD also interacts with the side chains of amino acids.¹³ A thiol group was conjugated with β CD to introduce redox activity, and two structural isomers, β CD_NSH and β CD_WSH, bearing a thiol group at different positions were designed to investigate the steric effect of the thiol group on promoting protein folding and suppressing aggregation (Fig. 1b). The thiol group of β CD_WSH is located at the wide rim of β CD, while that of β CD_NSH is attached at the narrow rim of the concave structure. These geometrical differences in substituent placement would likely influence the ability of the β CD unit to access the client polypeptide, thus affecting the binding dynamics of the synthetic oxidoreductase-mimics.

Biochemical studies on oxidoreductases show that adding disulfide and thiol compounds to unfolded reduced proteins facilitates their folding. Disulfides act as an oxidant to prompt

SS-bond formation of the client polypeptide, whereas thiols react with SS bonds through nucleophilic attack, aiding recombination between the native cysteine pairs and driving folding towards the native structure (Fig. 1a). β CD_NSH and β CD_WSH were synthesized from the corresponding monotosylated β CDs (ESI†). Oxidizability and nucleophilicity were assessed from the redox potential E° and pK_{a} of β CD_NSH and β CD_WSH (Fig. S1–S3†). In the pK_{a} analysis, β CD_NSH and β CD_WSH showed characteristic sigmoidal curves with two inflexion points, which is likely assigned to the deprotonation processes of the thiol and ammonium groups (Fig. S2†). Based on the deprotonation process at the lower pH condition, pK_{a} values of the thiol groups of β CD_NSH and β CD_WSH were evaluated at 4.68 ± 0.20 and 6.69 ± 0.94 , respectively, which were significantly lower than that of reduced glutathione (GSH, pK_{a} 9.17) used as a standard.¹⁴ These pK_{a} values indicate that most percentages of β CD_NSH and β CD_WSH adopt the thiolate forms that possess nucleophilicity at the experimental conditions of the protein folding assays in this study (pH 7.5), and the lower pK_{a} value of the thiol group of β CD_NSH suggests higher nucleophilicity than that of β CD_WSH. E° values of the disulfide forms of β CD_NSH and β CD_WSH were comparable with each other as well as that of oxidized glutathione (GSSG),^{14c} indicating the oxidizability of the three compounds in the oxidized forms are comparable among one another.

The binding properties of β CD_WSH and β CD_NSH were investigated through molecular dynamics simulations. Three systems were constructed, each comprising one unfolded bovine pancreatic trypsin inhibitor (BPTI) molecule and five molecules of β CD, β CD_WSH or β CD_NSH in 300 mM NaCl solution. BPTI was used as a model to compare the simulation results with experimental assays. Each system underwent a 100 ns NPT run at 300 K and 1 atm. The time-averaged fraction of contacts between β CD and each BPTI residue revealed an inhomogeneous distribution of contacts for β CD and β CD_NSH with the BPTI residues compared to β CD_WSH (Fig. 2 and S4, ESI Movies 1–3†). Specifically, β CD_NSH exhibited higher contacts with specific residues, including F4–P8, I19–Y23, and L29–V34. These observed contact distribution trends for each type of β CD align reasonably well with our nuclear magnetic resonance (NMR) measurement results, as discussed in a later section. Furthermore, in comparison to β CD, both β CD_WSH and β CD_NSH showed fewer contacts (reported as percentages) on the side with the SH group—*i.e.*, on the wide side for β CD_WSH and the narrow side for β CD_NSH, likely due to steric hindrance by the SH group (Fig. S4 and S5†). These results suggest that specific contacts between β CD with unfolded BPTI residues are associated with contacts with the wide side of β CD. Steric hindrance by the SH group on the wide side can lead to promiscuous contacts with all residues (*i.e.*, β CD_WSH), while hindrance on the narrow side enhances the specificity of the wide side of β CD (*i.e.*, β CD_NSH).

The ability of β CD_NSH and β CD_WSH to promote folding was investigated using BPTI. Native (N) BPTI contains three disulfide bonds, between Cys5–Cys55, Cys14–Cys38 and Cys30–Cys51, and was used as a representative model protein for the following folding study. The entire folding pathway of BPTI was



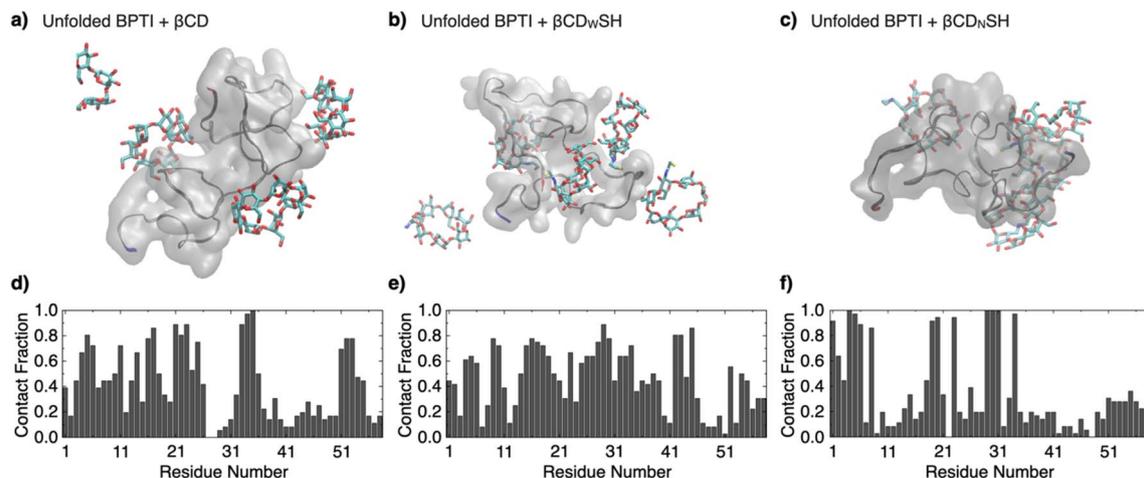


Fig. 2 Molecular dynamics simulations over 100 ns of one unfolded BPTI molecule in the presence of five molecules of (a and d) β CD, (b and e) β CD_wSH, and (c and f) β CD_NSH in 300 mM NaCl aq. (a–c) Representative snapshots. The N-terminus and C-terminus of BPTI are colored in blue and red, respectively. (d–f) Time-averaged fraction of contacts between the additives (β CD, β CD_wSH, or β CD_NSH, respectively) and each BPTI residue. See original movies in the ESI Movies (Movie 1: β CD, Movie 2: β CD_wSH, Movie 3: β CD_NSH†).

elucidated by Weissman and Kim.¹⁵ The folding of reduced (R) BPTI proceeds through the quasi-native intermediates N' and N*, which adopt native-like structures with two SS bonds prior to the formation of N (Fig. 3a). The folding assays were conducted using 30 μ M R-BPTI in the presence of 90 μ M disulfide compounds and 450 μ M thiol compounds, with essentially one equivalent of disulfide compound being added to the reduced-form of BPTI. The chosen ratio of disulfide and thiol compounds was based on a previous study.¹⁶ Reverse-phase

high-performance liquid chromatography (RP-HPLC) analysis of the oxidative folding of reduced BPTI in the presence of GSH and GSSG showed a gradual decrease in the R-fraction over a 60 min incubation period (Fig. 3b and e) whereas the N-fraction of BPTI emerged during the first 10 min of incubation to a final yield of 15% after 60 min incubation (Fig. 3f). Interestingly, the presence of β CD_wSH and its oxidized form β CD_wSS resulted in the rapid folding of reduced BPTI. The R-fraction decreased after 10 min incubation, and the N-fraction

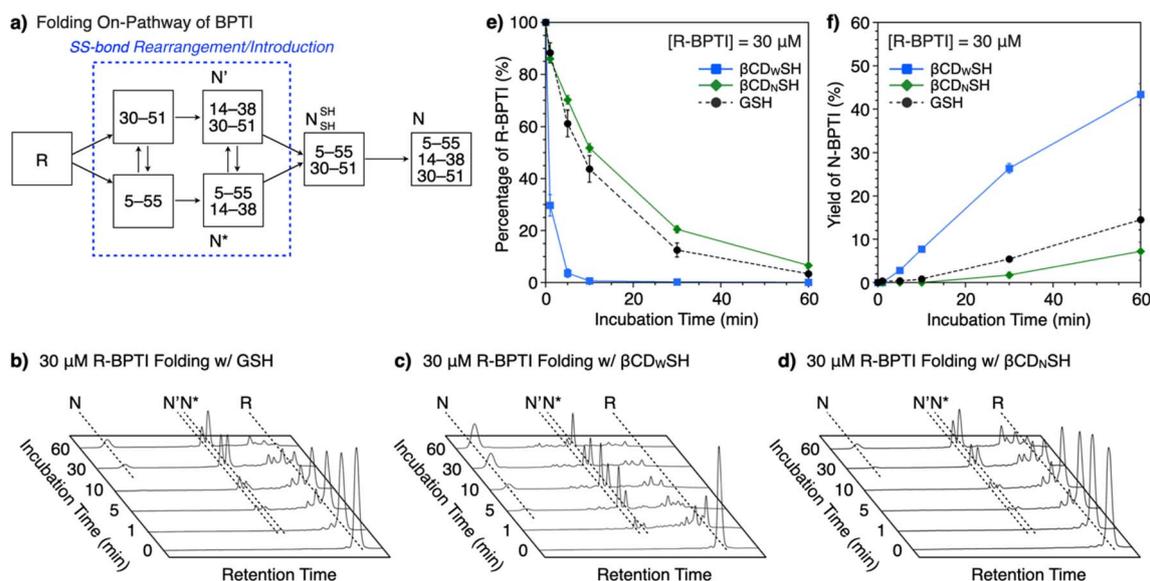


Fig. 3 Time course of BPTI oxidative folding as analyzed by RP-HPLC. (a) Folding pathway of BPTI. Time-course reverse-phase HPLC analyses of oxidative folding of BPTI (30 μ M) in the presence of (b) GSH and GSSG, (c) β CD_wSH and β CD_wSS, and (d) β CD_NSH and β CD_NSS, in the retention time range of 16 to 45 min. N and R denote the native and reduced forms of BPTI, respectively. N' and N* are the folding intermediates indicated in (a). [Thiols] = 450 μ M, [disulfides] = 90 μ M. Eluent buffers: water (containing 0.05% TFA) and CH₃CN (containing 0.05% TFA) with a linear gradient; flow rate: 1.0 mL min⁻¹; detection wavelength: 229 nm; temperature: 50 °C. Time course plots of (e) the percentage of R-BPTI and (f) the yield of N-BPTI in the presence of (black circles) GSH and GSSG, (blue squares) β CD_wSH and β CD_wSS, and (green diamonds) β CD_NSH and β CD_NSS. Error bars indicate the means \pm SEM of three independent experiments.



emerged after 5 min incubation (Fig. 3c). Sixty minutes incubation yielded 43% of the N-form (Fig. 3f). In sharp contrast, the addition of $\beta\text{CD}_\text{N}\text{SH}$ and $\beta\text{CD}_\text{W}\text{SH}$ to reduced BPTI improved folding only to a degree comparable to that obtained using the GSH/GSSG system, and the yield of N-form after 60 min incubation decreased (to 7%, Fig. 3f).

NMR spectroscopic studies were conducted to investigate the mechanism promoting folding and the binding properties of the cyclodextrin-conjugated thiols with unfolded BPTI. The interactions between unfolded BPTI and the cyclodextrin-conjugated thiols were visualized using an ^{15}N -labelled BPTI mutant in which all the cysteine residues were replaced with serine (^{15}N BPTI All-Ser) to provide a model of the R-form. ^1H - ^{15}N correlation spectra in the amide region were obtained following the addition of βCD using the selective optimized flip angle short transient (SOFAST) technique coupled to heteronuclear multiple quantum correlation (HMQC) experiments.¹⁷ Significant chemical shift perturbations to specific signals of ^{15}N BPTI All-Ser were observed (Fig. 4a and S6†). The bar graph showing the chemical shift differences ($\Delta\delta$) for each residue after adding βCD indicates that the resonances of some specific residues, including Y10, G12, Y35, and G37, shifted markedly, while most other residues showed much smaller chemical shift changes (Fig. 4d) consistent with significant changes in $\Delta\delta$ at the resonances of Y10, G12, Y35, and G37 upon the addition of $\beta\text{CD}_\text{N}\text{SH}$ (Fig. 4c and f). Interestingly, the addition of $\beta\text{CD}_\text{W}\text{SH}$ triggered smaller changes in $\Delta\delta$ relative to that resulting from the addition of βCD or $\beta\text{CD}_\text{N}\text{SH}$, but the resonances of most residues showed $\Delta\delta$ changes (Fig. 4b and e and S7†). These

results suggest that, while βCD or $\beta\text{CD}_\text{N}\text{SH}$ preferentially interacted with several specific residues with high selectivity and affinity, $\beta\text{CD}_\text{W}\text{SH}$ interacted promiscuously with many residues in ^{15}N BPTI All-Ser (Fig. 1c). Furthermore, the resonance of I19 showed a larger $\Delta\delta$ change upon the addition of $\beta\text{CD}_\text{W}\text{SH}$ than upon the addition of $\beta\text{CD}_\text{N}\text{SH}$, and the signal assigned to A48 shifted in opposite directions upon the addition of $\beta\text{CD}_\text{W}\text{SH}$ or $\beta\text{CD}_\text{N}\text{SH}$. These changing spectral profiles suggest that $\beta\text{CD}_\text{W}\text{SH}$ interacts with the unfolded protein not only in a weak and indiscriminate manner, but also *via* a different geometry than does $\beta\text{CD}_\text{N}\text{SH}$.

To directly monitor the interaction at the side chain of an amino acid residue with βCD s, we synthesized a peptide (BPTI7-13) consisting of the same amino acid sequence at 7–13 residues of BPTI. We plotted $\Delta\delta$ of the signal corresponding to Tyr10 of BPTI7-13 because the resonance of Tyr10 showed relatively larger shift of $\Delta\delta$ than the ones of other residues. As expected, titration with 0, 1, 3, and 5 mM of $\beta\text{CD}_\text{N}\text{SH}$ induced the largest $\Delta\delta$ and reached plateau, while the addition of $\beta\text{CD}_\text{W}\text{SH}$ elicited only small change (Fig. S8 and S9†). These analyses suggest that $\beta\text{CD}_\text{N}\text{SH}$ interacts with unfolded BPTI not only with the higher selectivity at some specific residues but also with the higher affinity than $\beta\text{CD}_\text{W}\text{SH}$. It is likely that the property of the high selectivity and affinity with the unfolded protein endows $\beta\text{CD}_\text{N}\text{SH}$ with the preferential client-binding character, while the promiscuous character of $\beta\text{CD}_\text{W}\text{SH}$ should be emerged by the lower selectivity and affinity property with the amino acid residues.

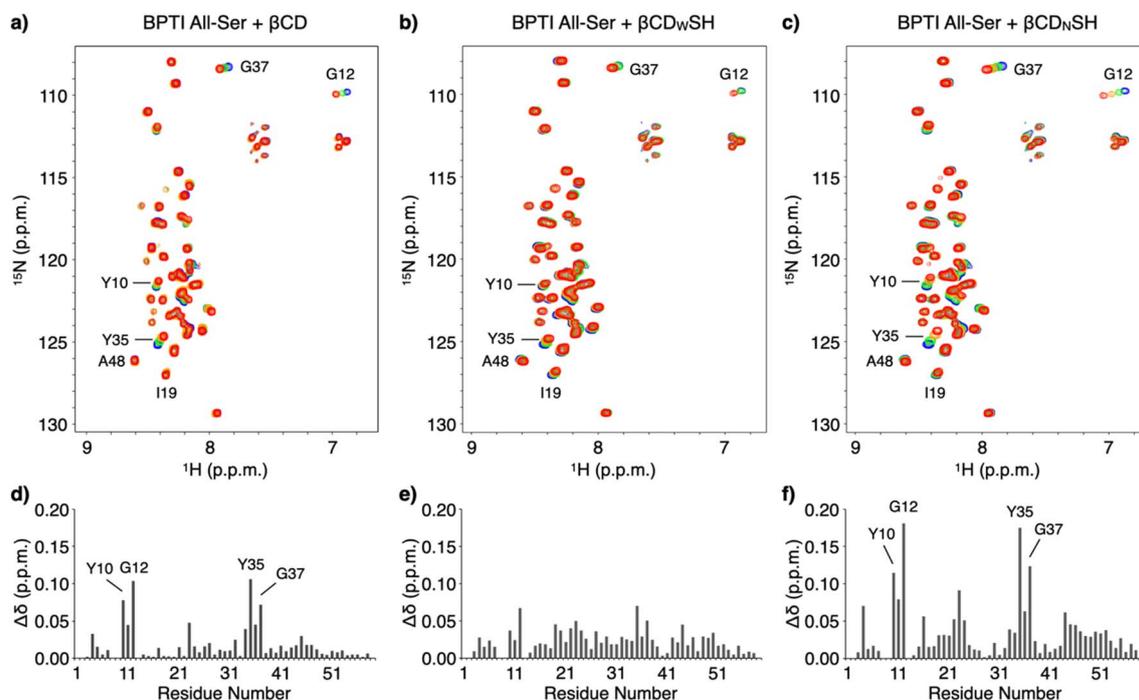


Fig. 4 ^1H - ^{15}N correlation SOFAST-HMQC spectra of ^{15}N BPTI All-Ser upon titration with (a) βCD , (b) $\beta\text{CD}_\text{W}\text{SH}$, and (c) $\beta\text{CD}_\text{N}\text{SH}$. [^{15}N BPTI All-Ser] = 100 μM , 50 mM HEPES (pH 7.0), 10 v/v% D_2O , 10 $^\circ\text{C}$, 500 MHz. Concentration of additives: 0 mM (blue), 1 mM (green), 3 mM (orange), and 5 mM (red). Chemical shift changes for ^{15}N BPTI All-Ser upon the addition of 5 mM (d) βCD , (e) $\beta\text{CD}_\text{W}\text{SH}$, and (f) $\beta\text{CD}_\text{N}\text{SH}$.



It would be of importance to discuss relationships between the BPTI primary structure and putative β CD binding sites such as Tyr and Phe (Fig. S10a and b†). The analysis indicates relatively larger $\Delta\delta$ at the signals of Tyr than those of Phe as an overall trend. Among the Tyr residues, the order of $\Delta\delta$ is as follows: (largest) Tyr10 > Tyr35 > Tyr23 > Tyr21 (smallest). Interestingly, the hydrophobic score analysis of each amino acid residue corresponds well with this order (Fig. S10c†). Namely, the Tyr residue with lower hydrophobic score showed larger $\Delta\delta$, and *vice versa*, suggesting that β CD and β CD_NSH interacts more effectively with the Tyr residues possessing relatively higher hydrophilicity. It is likely that the Tyr residue in a relatively hydrophilic region can expose out to the aqueous phase more favorably, which should be advantageous to the access of β CD and β CD_NSH. Meanwhile, the Tyr residue in the hydrophobic region might be buried inside of the polypeptide chain, which would inhibit the complexation with β CD and β CD_NSH.

Based on the interactions of β CD_WSH and β CD_NSH with unfolded proteins, we studied their aggregation suppression properties. Unfolded and reduced BPTI precipitates at 150 μ M, a highly condensed condition (Fig. 5a) and protein aggregation was apparently unaffected by β CD (Fig. 5b). Interestingly, the β CD_WSH/ β CD_WSS and β CD_NSH/ β CD_NSS systems acted oppositely to each other. The addition of β CD_NSH and β CD_NSS caused unfolded BPTI to aggregate, whereas little protein aggregation was observed in the presence of β CD_WSH or β CD_WSS (Fig. 5c and d). The ability of the β CD_WSH/ β CD_WSS system to suppress aggregation likely promotes folding of the reduced proteins under condensed conditions. We assayed the folding of unfolded and reduced 150 μ M BPTI upon addition of the cyclodextrin-conjugated thiols and disulfides ([thiols] = 1000 μ M, [disulfides] = 450 μ M as one equivalent for the full oxidation of R-BPTI). The concentration of the thiol compounds was limited to 1000 μ M due to their poor solubility. In the presence of β CD_WSH and β CD_WSS, the fraction corresponding to R-BPTI rapidly decreased during a 10 min incubation period, during which time a fraction assigned to N-BPTI became evident (Fig. 5e). Extending the incubation time to 300 min provided a fraction area of N-BPTI corresponding to a 45% yield (Fig. 5g). In contrast, the β CD_NSH/ β CD_NSS or GSH/GSSG system only slowly decreased the fraction area of R-BPTI, resulting in only 23% and 27% yields of N-BPTI, respectively (Fig. 5f and g, S11, and S12†).

The ability of β CD_NSH and β CD_WSH to promote folding was further investigated utilizing ribonuclease (RNase) A. Native RNase A contains four disulfide bonds, between Cys26–Cys84, Cys40–Cys95, Cys58–Cys110, and Cys65–Cys72. The folding of R-RNase A to the N-form proceeds through stepwise oxidations of 1SS, 2SS, and 3SS intermediates containing one, two, and three SS-bonds, respectively (Fig. 5h).¹⁸ As a side reaction, the 3SS intermediate can generate 4SS_U, a non-native form of fully oxidized RNase A containing four SS-bonds. SS-bond formation of R-RNase A at a high concentration was investigated in the presence of the chemical additives ([RNase A] = 100 μ M, [thiol] = 800 μ M, [disulfide] = 400 μ M). The progress of oxidation involving SS-bond formation was monitored by adding malPEG-

2000 (average M_n = 2000) to the reaction medium at selected time points (0, 1, 5, 10, 30, 60, 120, and 180 min) to quench the reaction. The maleimide moiety of malPEG-2000 reacts with the cysteine thiol groups of RNase A irreversibly, increasing its mass; therefore, RNase A bearing more thiol groups increases the mass of the protein correspondingly more. RNase A conjugated with different numbers of malPEG-2000 were separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) to quantitatively monitor the oxidation reactions. In the presence of GSH and GSSG, the band corresponding to fully oxidized RNase A containing N and 4SS_U appeared after 60 min incubation, and the percentage of N/4SS_U-RNase A increased to 31% after 180 min incubation (Fig. 5i and l). Interestingly, the β CD_WSH/ β CD_WSS system accelerated the oxidation of RNase A more effectively and rapidly than did the GSH/GSSG system, while the efficiency of the β CD_NSH/ β CD_NSS system was very similar to that of the GSH/GSSG system (Fig. 5j–l). Accordingly, the β CD_WSH/ β CD_WSS system supported the most rapid recovery of RNase A enzymatic activity, used to evaluate the formation of the N-form. After 180 min incubation, the β CD_WSH/ β CD_WSS system afforded a $38 \pm 1.1\%$ yield of N-RNase A. Under otherwise identical conditions, the β CD_NSH/ β CD_NSS and GSH/GSSG systems provided yields of N-form of up to $20 \pm 0.7\%$ and $14 \pm 1.1\%$, respectively (Fig. 5m).

As described above, β CD_WSS and β CD_NSS have similar oxidizability ($E^{\circ'}$), and their $E^{\circ'}$ values are also comparable to that of GSH/GSSG (Table 1). Based on the pK_a values, the nucleophilicity of β CD_WSH and β CD_NSH are higher than that of GSH, and particularly, the thiol group of β CD_NSH should have higher nucleophilicity than β CD_WSH. The higher nucleophilicity of the thiol group is generally advantageous for prompting SS-bond rearrangement. Nevertheless, the β CD_WSH/ β CD_WSS system facilitated oxidative SS-bond formation of reduced BPTI and RNase A more efficiently than did the β CD_NSH/ β CD_NSS system. Furthermore, folding to the native forms of BPTI and RNase A proceeded faster with the β CD_WSH/ β CD_WSS system than with the other systems, suggesting effective cooperation between the formation and rearrangement of the client protein's SS-bonds (Fig. 1a). The higher reactivities of the β CD_WSH/ β CD_WSS system for protein folding are likely due to the system's binding properties with the unfolded client protein, increasing the reaction rate at the thiol and disulfide unit of the cyclodextrin additive with the protein cysteine residues. Attaching the thiol group at the wide rim of β CD imparts a promiscuous and global binding property with unfolded proteins, likely due to steric hindrance at the client-binding pocket, as visualized by the MD simulation and supported by NMR measurements. Since the β CD_NSH/ β CD_NSS system, which binds with unfolded proteins tightly at specific domains, only weakly promoted protein folding and caused aggregation, it is likely that the promiscuous binding property of β CD_WSH/ β CD_WSS is advantageous for balancing polypeptide conformational mobility for forming intramolecular SS-bonds and non-covalent bonds required for folding, and for inhibiting polypeptide intermolecular contacts, thereby suppressing aggregation under condensed conditions. We suggest that the promiscuous binding behavior of β CD_WSH is analogous to the client-recognition behavior of ERp46, while



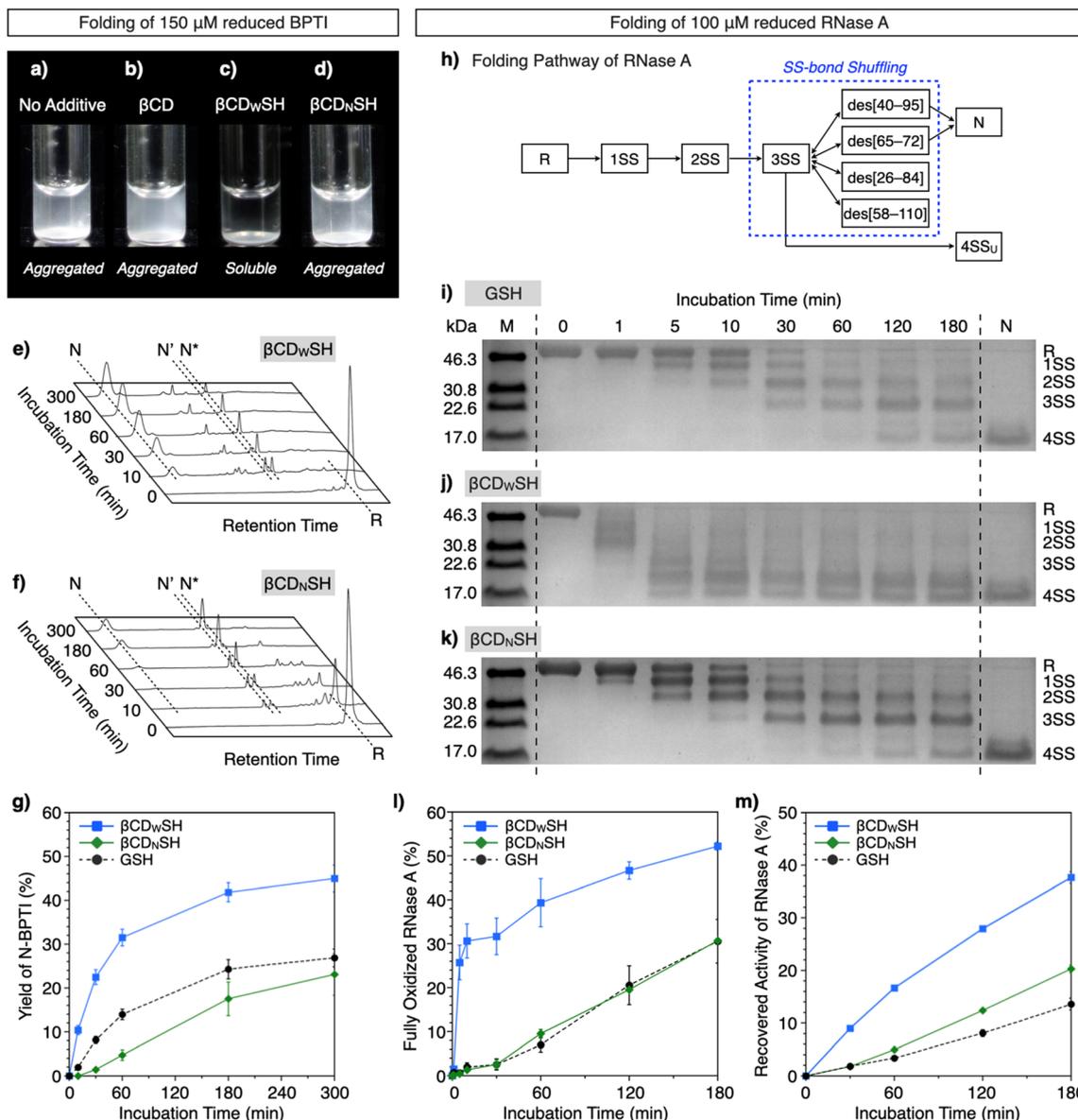


Fig. 5 BPTI and RNase A folding under condensed conditions ([BPTI] = 150 μM , [RNase A] = 100 μM). (a–d) Photographs of solutions of unfolded and reduced BPTI (150 μM) (a) without additives, and in the presence of (b) βCD , (c) $\beta\text{CD}_{\text{W}}\text{SH}/\beta\text{CD}_{\text{W}}\text{SS}$ and (d) $\beta\text{CD}_{\text{N}}\text{SH}/\beta\text{CD}_{\text{N}}\text{SS}$ ([additives] = 1000 μM). (e and f) Time-course reverse-phase HPLC analyses of the oxidative folding of BPTI (150 μM) in the presence of (e) $\beta\text{CD}_{\text{W}}\text{SH}$ and $\beta\text{CD}_{\text{W}}\text{SS}$ and (f) $\beta\text{CD}_{\text{N}}\text{SH}$ and $\beta\text{CD}_{\text{N}}\text{SS}$ in the retention time range of 16 to 45 min. N and R represent the native and reduced forms of BPTI, respectively. N' and N* are the folding intermediates. The fraction of N and N' in (e) corresponds to complexes between BPTI and $\beta\text{CD}_{\text{W}}\text{SH}$ based on MALDI-TOF MS analysis (Fig. S13†). Eluent buffers: water (containing 0.05% TFA) and CH_3CN (containing 0.05% TFA) with a linear gradient; flow rate: 1.0 mL min^{-1} ; detection wavelength: 229 nm; temperature: 50 $^\circ\text{C}$. (g) Time course plots of the yields of N-BPTI in the presence of (black circles) GSH and GSSG, (blue squares) $\beta\text{CD}_{\text{W}}\text{SH}$ and $\beta\text{CD}_{\text{W}}\text{SS}$, and (green diamonds) $\beta\text{CD}_{\text{N}}\text{SH}$ and $\beta\text{CD}_{\text{N}}\text{SS}$. (h) Folding pathway of RNase A. (i–k) SDS-PAGE gel images monitoring the oxidation of RNase A (100 μM) in the presence of (i) GSH and GSSG, (j) $\beta\text{CD}_{\text{W}}\text{SH}$ and $\beta\text{CD}_{\text{W}}\text{SS}$, and (k) $\beta\text{CD}_{\text{N}}\text{SH}$ and $\beta\text{CD}_{\text{N}}\text{SS}$ in buffer (50 mM Tris-HCl, 300 mM NaCl, pH 7.5) at 30 $^\circ\text{C}$. The folding reactions were quenched with malPEG-2000 after 1, 5, 10, 30, 60, 120, and 180 min of incubation. The leftmost and rightmost lanes contain molecular weight markers (M) and native RNase A (N), respectively. Time-course changes of (l) formation of fully oxidized RNase A (4SS forms) quantified by SDS-PAGE analyses and (m) recovered enzymatic activity of RNase A in the presence of GSH and GSSG, $\beta\text{CD}_{\text{W}}\text{SH}$ and $\beta\text{CD}_{\text{W}}\text{SS}$, and $\beta\text{CD}_{\text{N}}\text{SH}$ and $\beta\text{CD}_{\text{N}}\text{SS}$ in buffer (50 mM Tris-HCl, 300 mM NaCl, pH 7.5) at 30 $^\circ\text{C}$. Activity was evaluated by spectroscopic monitoring of the hydrolysis of cCMP to 3'-CMP at 30 $^\circ\text{C}$. Error bars indicate the means \pm SEM of three independent experiments.

the more preferential recognition character of $\beta\text{CD}_{\text{N}}\text{SH}$ corresponds to that of PDI, which binds to specific local regions of the client protein. Interestingly, $\beta\text{CD}_{\text{N}}\text{SH}$ and PDI recognize several common sites in unfolded BPTI, such as the residues

around G12 and G37, despite their large differences in size and chemical structure. Furthermore, consistent with the trend observed between ERp46 and PDI in the initial phase of the folding process, the ERp46-mimetic $\beta\text{CD}_{\text{W}}\text{SH}$ supported faster



Table 1 Chemical properties of $\beta\text{CD}_{\text{W}}\text{SH}$, $\beta\text{CD}_{\text{N}}\text{SH}$, and glutathione (GSH)

Compounds	$\text{p}K_{\text{a}}$	E° (mV)
$\beta\text{CD}_{\text{W}}\text{SH}$	6.69 ± 0.94 (SH) 11.09 ± 0.08 (NH_3^+)	-258 ± 1
$\beta\text{CD}_{\text{N}}\text{SH}$	4.68 ± 0.20 (SH) 10.84 ± 0.05 (NH_3^+)	-254 ± 1
GSH	9.17^a	-256^b

^a Ref. 14. ^b Ref. 11c.

SS-bond formation than the PDI-mimetic $\beta\text{CD}_{\text{N}}\text{SH}$.⁹ As observed in the BPTI folding process catalyzed by ERp46, off-pathway intermediate species were also observed in the presence of $\beta\text{CD}_{\text{W}}\text{SH}$.^{8b,9} These analogies between biological enzymes and the described chemical mimics demonstrate that the binding characteristics of chemical chaperones can be controlled by steric effects at the client-recognition site to deter aggregation. The bio-inspired design described here, combining a redox-active center and client-recognition unit with a tendency for promiscuous binding, is an effective approach for promoting the correct folding of error-prone and aggregation-prone polypeptide folding under condensed conditions.

In this study, one equivalent of disulfide compounds relative to the client proteins was added to the reaction solutions, indicating that the reaction systems are non-catalytic. Meanwhile, the native system of the oxidative protein folding is catalytic, in which the reduced forms of oxidoreductases were re-oxidized in a multi-step electron transfer reactions from O_2 , an oxidation source, to H_2O .^{3c} Installing such regeneration process of the oxidants should enable development of bio-mimetic catalytic reaction systems of the oxidative protein folding, which is an important subject in this research field to further enhance the production efficiency as high as that of the cellular system.

There is an increasing number of reports describing a link between disulfide chemistry and the pathogenesis of diseases that involve misfolded proteins.¹⁹ Over 20 members of the PDI family, which are disulfide catalysts, play key roles in maintaining protein homeostasis by both catalyzing oxidative folding and by chaperoning aggregation-prone clients, thereby likely decreasing the risk of misfolding-induced pathologies. The dysfunction of some disulfide catalysts thus may result in pathologies such as amyotrophic lateral sclerosis, Alzheimer's disease, Parkinson's disease, type 2 diabetes, and intellectual disability. There are few examples of the development of chemical chaperones to treat misfolding-related diseases,²⁰ and new approaches using thiol compounds in place of PDI family proteins are attractive for this purpose.

Conclusions

In this study, we developed a redox-active promiscuous binder, $\beta\text{CD}_{\text{W}}\text{SH}$, which bears a client-recognition unit and a redox-active center. $\beta\text{CD}_{\text{W}}\text{SH}$ promotes oxidative protein folding at sub-millimolar concentrations under condensed conditions.

The design of a sterically hindered client-binding pocket in βCD coupled with a redox-active group satisfies the conflicting demands for protein folding: to enhance the frequency of cysteine residue reactions, to maintain conformational mobility of the polypeptide chain, and to block intermolecular protein aggregation. The molecular design of redox-active chemical chaperones with promiscuous binding properties represents an important breakthrough for promoting protein folding under highly condensed conditions and is promising for producing functional and pharmaceutical proteins. This chemical approach can also find medical applications for the prevention of misfolding diseases caused by protein denaturation and aggregation.

Data availability

The data supporting this article have been included as part of the ESI.†

Author contributions

T. Mu. conceived the idea of this study. K. S., R. N., T. Ma., S. K., K. I. and H. K. performed the experiments. M. O., T. S., and T. Mu. supervised the conduct of this study. M. O., T. S. and T. M. wrote the manuscript. All authors critically reviewed and revised the manuscript draft and approved the final version for submission.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- (a) C. B. Anfinsen, Principles that govern the folding of protein chains, *Science*, 1973, **181**, 223–230; (b) M. J. Gething and J. Sambrook, Protein folding in the cell, *Nature*, 1992, **355**, 33–45; (c) T. E. Creighton, Protein folding coupled to disulphide bond formation, *Biol. Chem.*, 1997, **378**, 731–744; (d) C. M. Dobson, Protein folding and misfolding, *Nature*, 2003, **426**, 884–890; (e) F. Baneyx and M. Mujacic, Recombinant protein folding and misfolding in *Escherichia coli*, *Nat. Biotechnol.*, 2004, **22**, 1399–1408;



- (f) J. L. Arolas, F. X. Aviles, J.-Y. Chang and S. Ventura, Folding of small disulfide-rich proteins/clarifying the puzzle, *Trends Biochem. Sci.*, 2006, **31**, 292–301; (g) S. J. Kim, J. S. Yoon, H. Shishido, Z. Yang, L. A. Rooney, J. M. Barral and W. R. Skach, Translational tuning optimizes nascent protein folding in cells, *Science*, 2015, **348**, 444–448.
- 2 (a) O. B. Ptitsyn, V. E. Bychkova and V. N. Uversky, Kinetic and equilibrium folding intermediates, *Philos. Trans. R. Soc., B*, 1995, **348**, 35–41; (b) J. L. Sohl, S. S. Jaswal and D. A. Agard, Unfolded conformations of α -lytic protease are more stable than its native state, *Nature*, 1998, **395**, 817–819; (c) P. T. Lansbury Jr, Evolution of amyloid: What normal protein folding may tell us about fibrillogenesis and disease, *Proc. Natl. Acad. Sci. U. S. A.*, 1999, **96**, 3342–3344; (d) J. Ren, Y. Bi, J. R. Sowers, C. Hetz and Y. Zhang, Endoplasmic reticulum stress and unfolded protein response in cardiovascular diseases, *Nat. Rev. Cardiol.*, 2021, **18**, 499–521.
- 3 (a) J. S. Weissman and P. S. Kim, Efficient catalysis of disulphide bond rearrangements by protein disulphide isomerase, *Nature*, 1993, **365**, 185–188; (b) B. van den Berg, E. W. Chung, C. V. Robinson, P. L. Mateo and C. M. Dobson, The oxidative refolding of hen lysozyme and its catalysis by protein disulfide isomerase, *EMBO J.*, 1999, **18**, 4794–4803; (c) M. Okumura, H. Kadokura and K. Inaba, Structures and functions of protein disulfide isomerase family members involved in proteostasis in the endoplasmic reticulum, *Free Radical Biol. Med.*, 2015, **83**, 314–322; (d) R. A. Lumb and N. J. Bulleid, Is protein disulfide isomerase a redox-dependent molecular chaperone?, *EMBO J.*, 2002, **21**, 6763–6770; (e) M. Okumura, H. Kadokura, S. Hashimoto, K. Yutani, S. Kanemura, T. Hikima, Y. Hidaka, L. Ito, K. Shiba, S. Masui, D. Imai, S. Imaoka, H. Yamaguchi and K. Inaba, Inhibition of the functional interplay between endoplasmic reticulum (ER) oxidoreductin-1 α (Ero1 α) and protein-disulfide isomerase (PDI) by the endocrine disruptor bisphenol A, *J. Biol. Chem.*, 2014, **289**, 27004–27018; (f) M. Okumura, K. Noi, S. Kanemura, M. Kinoshita, T. Saio, Y. Inoue, T. Hikima, S. Akiyama, T. Ogura and K. Inaba, Dynamic assembly of protein disulfide isomerase in catalysis of oxidative folding, *Nat. Chem. Biol.*, 2019, **15**, 499–509; (g) M. Okumura, K. Noi and K. Inaba, Visualization of structural dynamics of protein disulfide isomerase enzymes in catalysis of oxidative folding and reductive unfolding, *Curr. Opin. Struct. Biol.*, 2021, **66**, 49–57; (h) M. Chinnaraj, R. Flaumenhaft and N. Pozzi, Reduction of protein disulfide isomerase results in open conformations and stimulates dynamic exchange between structural ensembles, *J. Biol. Chem.*, 2022, **298**, 102217.
- 4 T. Muraoka, M. Okumura and T. Saio, Enzymatic and synthetic regulation of polypeptide folding, *Chem. Sci.*, 2024, **15**, 2282–2299.
- 5 E. Rajakumara, S. Abhishek, K. Nitin, D. Saniya, P. Bajaj, U. Schwaneberg and M. D. Davari, Structure and cooperativity in substrate–enzyme interactions: Perspectives on enzyme engineering and inhibitor design, *ACS Chem. Biol.*, 2022, **17**, 266–280.
- 6 J.-M. Choi, S.-S. Han and H.-S. Kim, Industrial applications of enzyme biocatalysis: Current status and future aspects, *Biotechnol. Adv.*, 2015, **33**, 1443–1454.
- 7 (a) R. Rosenzweig, A. Sekhar, J. Nagesh and L. E. Kay, Promiscuous binding by Hsp70 results in conformational heterogeneity and fuzzy chaperone-substrate ensembles, *eLife*, 2017, **6**, e28030; (b) L. He and S. Hiller, Common patterns in chaperone interactions with a native client protein, *Angew. Chem., Int. Ed.*, 2018, **57**, 5921–5924.
- 8 (a) J. C. Edman, L. Ellis, R. W. Blacher, R. A. Roth and W. J. Rutter, Sequence of protein disulphide isomerase and implications of its relationship to thioredoxin, *Nature*, 1985, **317**, 267–270; (b) Y. Sato, R. Kojima, M. Okumura, M. Hagiwara, S. Masui, K. Maegawa, M. Saiki, T. Horibe, M. Suzuki and K. Inaba, Synergistic cooperation of PDI family members in peroxiredoxin 4-driven oxidative protein folding, *Sci. Rep.*, 2013, **3**, 2456; (c) R. Kojima, M. Okumura, S. Masui, S. Kanemura, M. Inoue, M. Saiki, H. Yamaguchi, T. Hikima, M. Suzuki, S. Akiyama and K. Inaba, Radically different thioredoxin domain arrangement of ERp46, an efficient disulfide bond introducer of the mammalian PDI family, *Structure*, 2014, **22**, 431–443.
- 9 T. Saio, K. Ishii, M. Matsusaki, H. Kumeta, S. Kanemura and M. Okumura, *bioRxiv*, 2024, preprint, DOI: [10.1101/2024.03.04.583432](https://doi.org/10.1101/2024.03.04.583432).
- 10 (a) T. Saio, X. Guan, P. Rossi, A. Economou and C. G. Kalodimos, Structural basis for protein antiaggregation activity of the trigger factor chaperone, *Science*, 2014, **344**, 1250494; (b) C. Huang, P. Rossi, T. Saio and C. G. Kalodimos, Structural basis for the antifolding activity of a molecular chaperone, *Nature*, 2016, **537**, 202–206.
- 11 (a) E. Welker, M. Narayan, W. J. Wedemeyer and H. A. Scheraga, Structural determinants of oxidative folding in proteins, *Proc. Natl. Acad. Sci. U. S. A.*, 2001, **98**, 2312–2316; (b) J. D. Gough, R. H. Williams, A. E. Donofrio and W. J. Lees, Folding disulfide-containing proteins faster with an aromatic thiol, *J. Am. Chem. Soc.*, 2002, **124**, 3885–3892; (c) J. Beld, K. J. Woycechowsky and D. Hilvert, Selenogluthione: Efficient oxidative protein folding by a diselenide, *Biochemistry*, 2007, **46**, 5382–5390; (d) W. J. Lees, Small-molecule catalysts of oxidative protein folding, *Curr. Opin. Chem. Biol.*, 2008, **12**, 740–745; (e) M. Okumura, M. Saiki, H. Yamaguchi and Y. Hidaka, Acceleration of disulfide-coupled protein folding using glutathione derivatives, *FEBS J.*, 2011, **278**, 1137–1144; (f) A. S. Patel and W. J. Lees, Oxidative folding of lysozyme with aromatic dithiols, and aliphatic and aromatic monothiols, *Bioorg. Med. Chem.*, 2012, **20**, 1020–1028; (g) J. C. Lukesh III, M. J. Palte and R. T. Raines, A potent, versatile disulfide-reducing agent from aspartic acid, *J. Am. Chem. Soc.*, 2012, **134**, 4057–4059; (h) P. S. Reddy and N. Metanis, Small molecule diselenide additives for in vitro oxidative protein folding, *Chem. Commun.*, 2016, **52**,



- 3336–3339; (i) K. Arai, H. Ueno, Y. Asano, G. Chakrabarty, S. Shimodaira, G. Mugesh and M. Iwaoka, Protein folding in the presence of water-soluble Cyclic diselenides with novel oxidoreductase and isomerase activities, *ChemBioChem*, 2018, **19**, 207–211; (j) S. Okada, M. Matsusaki, K. Arai, Y. Hidaka, K. Inaba, M. Okumura and T. Muraoka, Coupling effects of thiol and urea-type groups for promotion of oxidative protein folding, *Chem. Commun.*, 2019, **55**, 759–762; (k) S. Tsukagoshi, R. Mikami and K. Arai, Basic amino acid conjugates of 1,2-diselenan-4-amine with protein disulfide isomerase-like functions as a manipulator of protein quality control, *Chem.–Asian J.*, 2020, **15**, 2646–2656; (l) S. Okada, Y. Matsumoto, R. Takahashi, K. Arai, S. Kanemura, M. Okumura and T. Muraoka, Semi-enzymatic acceleration of oxidative protein folding by N-methylated heteroaromatic thiols, *Chem. Sci.*, 2023, **14**, 7630–7636.
- 12 G. Crini, Review: A history of cyclodextrins, *Chem. Rev.*, 2014, **114**, 10940–10975.
- 13 (a) J. M. Alexander, J. L. Clark, T. J. Brett and J. J. Stezowski, Chiral discrimination in cyclodextrin complexes of amino acid derivatives/ β -cyclodextrin/N-acetyl-L-phenylalanine and N-acetyl-D-phenylalanine complexes, *Proc. Natl. Acad. Sci. U. S. A.*, 2002, **99**, 5115–5120; (b) M. S. Semrau, G. Giachin, S. Covaceuszach, A. Cassetta, N. Demitri, P. Storici and G. Lolli, Molecular architecture of the glycogen-committed PP1/PTG holoenzyme, *Nat. Commun.*, 2022, **13**, 6199.
- 14 S.-S. Tang and G.-G. Chang, Nucleophilic aromatic substitution of glutathione and 1-chloro-2,4-dinitrobenzene in reverse micelles. A model system to assess the transition-state stabilization in glutathione transferase catalyzed conjugation, *J. Org. Chem.*, 1995, **60**, 6183–6185.
- 15 J. S. Weissman and P. S. Kim, Reexamination of the folding of BPTI: Predominance of native intermediates, *Science*, 1991, **253**, 1386–1393.
- 16 M. M. Lyles and H. F. Gilbert, Catalysis of the oxidative folding of ribonuclease A by protein disulfide isomerase: Dependence of the rate on the composition of the redox buffer, *Biochemistry*, 1991, **30**, 613–619.
- 17 P. Schanda and B. Brutscher, Very fast two-dimensional NMR spectroscopy for real-time investigation of dynamic events in proteins on the time scale of seconds, *J. Am. Chem. Soc.*, 2005, **127**, 8014–8015.
- 18 (a) A. Bierzynski, P. S. Kim and R. L. Baldwin, A salt bridge stabilizes the helix formed by isolated C-peptide of RNase A, *Proc. Natl. Acad. Sci. U. S. A.*, 1982, **79**, 2470–2474; (b) D. M. Rothwarf and H. A. Scheraga, Regeneration of bovine pancreatic ribonuclease A. 2. Kinetics of regeneration, *Biochemistry*, 1993, **32**, 2680–2689; (c) M. Iwaoka, D. Juminaga and H. A. Scheraga, Regeneration of three-disulfide mutants of bovine pancreatic ribonuclease A missing the 65-72 disulfide bond: Characterization of a minor folding pathway of ribonuclease A and kinetic roles of Cys65 and Cys72, *Biochemistry*, 1998, **37**, 4490–4501; (d) M. Narayan, E. Welker, W. J. Wedemeyer and H. A. Scheraga, Oxidative folding of proteins, *Acc. Chem. Res.*, 2000, **33**, 805–812.
- 19 (a) T. Uehara, T. Nakamura, D. Yao, Z. Q. Shi, Z. Gu, Y. Ma, E. Masliah, Y. Nomura and S. A. Lipton, S-Nitrosylated protein-disulphide isomerase links protein misfolding to neurodegeneration, *Nature*, 2006, **441**, 513–517; (b) D. Eletto, D. Eletto, D. Dersh, T. Gidalevitz and Y. Argon, Protein disulfide isomerase A6 controls the decay of IRE1 α signaling via disulfide-dependent association, *Mol. Cell*, 2014, **53**, 562–576; (c) O. B. Oka, M. van Lith, J. Rudolf, W. Tungku, M. A. Pringle and N. J. Bulleid, ERp18 regulates activation of ATF6 α during unfolded protein response, *EMBO J.*, 2019, **38**, e100990; (d) M. Matsusaki, S. Kanemura, M. Kinoshita, Y.-H. Lee, K. Inaba and M. Okumura, The protein disulfide isomerase family: from proteostasis to pathogenesis, *Biochim. Biophys. Acta, Gen. Subj.*, 2020, **1864**, 129338; (e) D. B. Medinas, P. Rozas and C. Hetz, Critical roles of protein disulfide isomerases in balancing proteostasis in the nervous system, *J. Biol. Chem.*, 2022, **298**, 102087.
- 20 (a) C. R. Brown, L. Q. Hong-Brown, J. Biwersi, A. S. Verkman and W. J. Welch, Chemical chaperones correct the mutant phenotype of the Δ F508 cystic fibrosis transmembrane conductance regulator protein, *Cell Stress Chaperones*, 1996, **1**, 117–125; (b) T. Okiyoneda, G. Veit, J. F. Dekkers, M. Bagdany, N. Soya, H. Xu, A. Roldan, A. S. Verkman, M. Kurth, A. Simon, T. Hegedus, J. M. Beekman and G. L. Lukacs, Mechanism-based corrector combination restores Δ F508-CFTR folding and function, *Nat. Chem. Biol.*, 2013, **9**, 444–454; (c) T. Kuramochi, Y. Yamashita, K. Arai, S. Kanemura, T. Muraoka and M. Okumura, Boosting the enzymatic activity of CxxC motif-containing PDI family members, *Chem. Commun.*, 2024, **60**, 6134–6137.

