

Cite this: *Chem. Sci.*, 2024, 15, 10121

All publication charges for this article have been paid for by the Royal Society of Chemistry

Received 21st March 2024

Accepted 20th May 2024

DOI: 10.1039/d4sc01897d

rsc.li/chemical-science

# Synthesis and biological evaluation of cleistocaltone A, an inhibitor of respiratory syncytial virus (RSV)<sup>†</sup>

Lorenz Wiese,<sup>a</sup> Sophie M. Kolbe,<sup>b</sup> Manuela Weber,<sup>a</sup> Martin Ludlow<sup>\*b</sup> and Mathias Christmann<sup>\*a</sup>

The first chemical synthesis of the phloroglucinol meroterpenoid cleistocaltone A (**1**) is presented. This compound, previously isolated from *Cleistocalyx operculatus* was reported to show promising antiviral properties. Based on a modified biosynthesis proposal, a synthetic strategy was devised featuring an intramolecular Diels–Alder reaction and an epoxidation/elimination sequence to generate the allyl alcohol handle in the side chain. The strategy was successfully executed and synthetic cleistocaltone A was evaluated against a contemporary RSV-A strain.

## Introduction

Viral infections pose a significant economic burden and societal threat worldwide.<sup>1–3</sup> The recent COVID-19 pandemic caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) has starkly highlighted the risks posed by respiratory viruses to vulnerable populations and the urgent need for effective antivirals. Respiratory Syncytial Virus (RSV) is also a potent respiratory pathogen and is responsible for high levels of acute lower respiratory tract infections in infants, the elderly and immunocompromised adults.<sup>4,5</sup> Despite the urgency, the current landscape of antiviral treatments is sparse, underscoring the critical need for further antiviral development.<sup>6</sup> Natural products have a long and successful history as privileged starting points in the search for novel therapeutic approaches to counter infectious diseases.<sup>7–9</sup> One effective strategy for navigating their vast chemical space is to deconvolute molecular compositions of herbal remedies described in traditional Chinese medicine (TCM), thus building upon a wealth of knowledge based on centuries of use.<sup>10</sup> For example, the genus *Cleistocalyx*, belonging to the *Myrtaceae* family, comprises various plant species, some of which are used in Southeast Asia to create medicinal teas. *Cleistocalyx operculatus* (Roxb.) Merr. and Perry, a tree native to southern China, has been traditionally used to treat cold, fever, and inflammation with extracts from its buds and leaves.<sup>11</sup> Cleistocaltone A (**1**, Fig. 1), a polymethylated phloroglucinol meroterpenoid (PPM) isolated from the buds of *C. operculatus* by

Ye, Wang, and co-workers in 2019, was demonstrated to possess promising *in vitro* activity against RSV.<sup>12</sup>

In nature, components from diverse classes of natural products frequently merge to form compounds with novel biological activities. Among these classes, phloroglucinol meroterpenoids stand out.<sup>13</sup> These compounds consist of a phloroglucinol segment and a terpene segment. The terpene portion typically originates from a geranyl group, which undergoes a series of biosynthetic transformations involving oxidation and cyclization. For instance, elodeoidol G (**2**, Fig. 1), an anti-inflammatory agent isolated by Luo, Kong, and coworkers, features a terpene moiety proposed to be biosynthesized from an oxidized geranyl building block through a series of epoxidations and an acid-catalyzed epoxide opening cascade.<sup>14</sup> Similarly, the cytotoxic compound hyphenrone J (**3**, Fig. 1), isolated by Qin, Xu, and coworkers, contains two prenyl groups and an oxidized geranyl side chain.<sup>15</sup>

Biosynthetically, the phloroglucinol portion of cleistocaltone A (**1**) is proposed to originate from champanone B (**4**, Fig. 1).

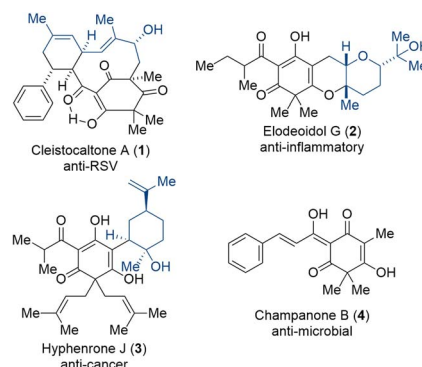
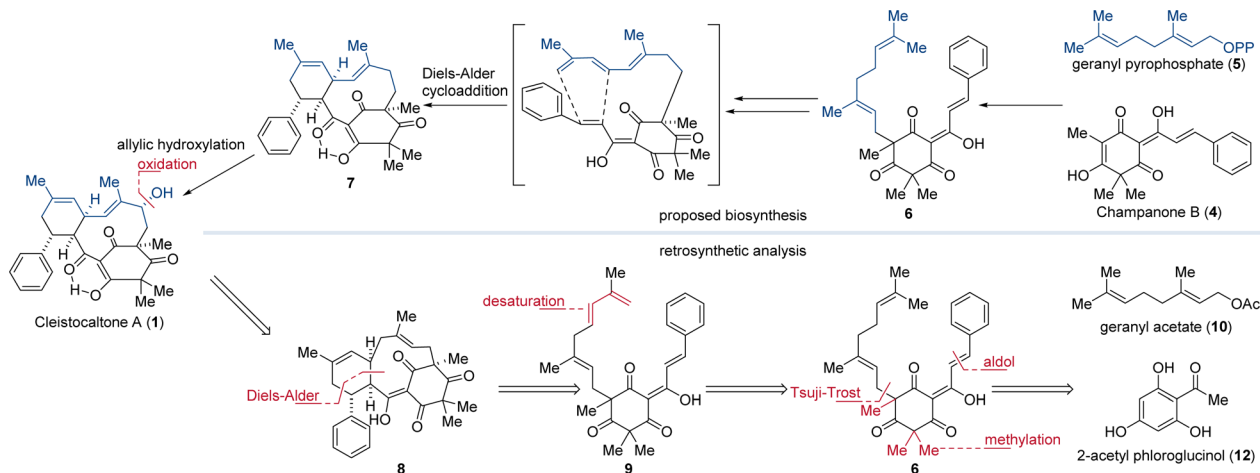


Fig. 1 Example for different phloroglucinol meroterpenoids with various biological activities.<sup>12,14–16</sup>

<sup>a</sup>Institute of Chemistry and Biochemistry, Freie Universität Berlin, 14195, Berlin, Germany. E-mail: mathias.christmann@fu-berlin.de

<sup>b</sup>Research Center for Emerging Infections and Zoonoses, University of Veterinary Medicine Hannover Foundation, 30559, Hanover, Germany

<sup>†</sup> Electronic supplementary information (ESI) available. See DOI: <https://doi.org/10.1039/d4sc01897d>



**Scheme 1** Biosynthesis of cleistocaltone A (**1**) proposed by Ye, Wang, and coworkers and retrosynthetic analysis both featuring an IMDA as the key step.<sup>12</sup>

Addition of geranyl pyrophosphate (GPP, **5**) then forms geranyl champanone B **6**. Subsequent desaturation of the geranyl terminus and isomerization of the homobenzylic double bond generates a triene, enabling an intramolecular Diels–Alder cycloaddition (IMDA) with the cinnamoyl dienophile (Scheme 1).

This reaction forges the characteristic 10-membered macrocycle found in **7** (Scheme 1).<sup>12</sup> Ye, Wang, and coworkers succeeded in isolating 24 mg of (±)-cleistocaltone A (**1**) from 15 kg of plant material, highlighting the scarcity of this compound from biological sources.<sup>12</sup> This underlines the significance of chemical synthesis to produce substantial quantities of (±)-cleistocaltone A (**1**) for further biological evaluation.

In recent years, biomimetic strategies have emerged as an intriguing and efficient approach for synthesizing complex natural products. These strategies elegantly mimic the pathways found in nature.<sup>17–19</sup> By harnessing the innate reactivity of molecules, biomimetic syntheses often minimize the need for protection groups, resulting in greater efficiency and sustainability.<sup>20</sup>

Mirroring the proposed biosynthetic pathway, we envisioned an IMDA as the pivotal step for constructing the 10-membered macrocycle of **8**. To generate the diene moiety in **9** for the IMDA, we planned a selective desaturation at the terminus of the geranyl group in **6**, obtained from geranyl acetate (**10**). This geranyl unit would be attached to the nucleophilic carbon atom of methylated phloroglucinol **11** using a Tsuji–Trost alkylation.<sup>21</sup> The phloroglucinol building block would be synthesized from commercially available 2-acetyl phloroglucinol (**12**) through trimethylation and an aldol condensation.

## Results and discussion

Our synthesis, as outlined in Scheme 2, commenced with the literature-known trimethylation of 2-acetyl phloroglucinol (**12**), employing sodium methoxide and methyl iodide.<sup>22</sup>

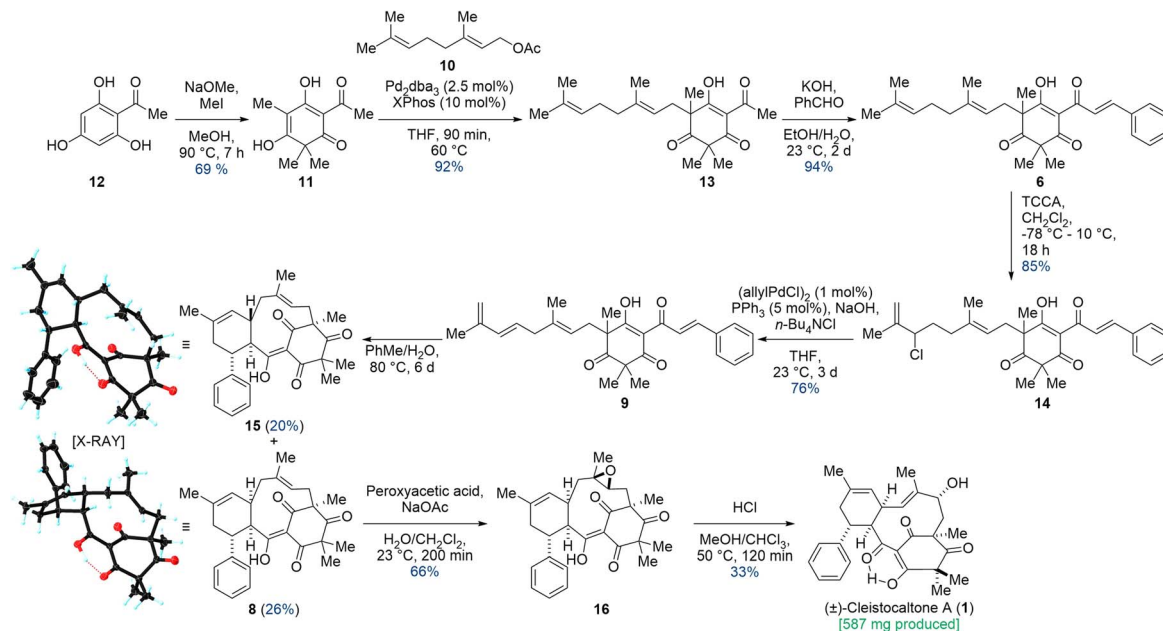
Following the neutralization of the reaction mixture, the desired product precipitated, allowing for straightforward separation *via* filtration. This workup efficiently removed

impurities and byproducts, including dimethylated 2-acetyl phloroglucinol, which stayed in solution. Consequently, we obtained trimethylated acetyl phloroglucinol **11** with a yield of 69%. The obtained trimethylated acetyl phloroglucinol **11** was then coupled with commercially available geranyl acetate (**10**) in a Tsuji–Trost allylic alkylation. Using Pd<sub>2</sub>dba<sub>3</sub> as a Pd(0) source and testing multiple ligands, XPhos proved superior, yielding the geranylated phloroglucinol **13** as a mixture of interconverting tautomers with an excellent yield of 92%. Rendering the coupling reaction enantioselective would be highly desirable. The single stereocenter formed in the Tsuji–Trost reaction ultimately dictates whether (+)- or (–)-cleistocaltone A (**1**) is synthesized due to the diastereoselectivity of subsequent steps. Unfortunately, despite rigorous screening, we could not identify a chiral ligand that induced significant enantiomeric excess. Consequently, we proceeded with the synthesis using a racemic mixture of geranylated acetyl phloroglucinol **13**.

In the subsequent aldol condensation, **13** was reacted with benzaldehyde under basic conditions. Due to a competing Cannizzaro reaction consuming benzaldehyde (yielding benzyl alcohol and benzoic acid),<sup>23</sup> a portion-wise addition of **6** equivalents of aldehyde was crucial for complete conversion. Then, the high solubility of **6** in cyclohexane was exploited for extraction, leaving benzoic acid and much of the benzyl alcohol in the aqueous phase. Unreacted benzaldehyde was removed under high vacuum, greatly streamlining the purification process. This yielded geranyl champanone B **6** as a mixture of tautomers in 94% yield.

The geranyl moiety in compound **6** was selectively chlorinated at its terminus using trichloroisocyanuric acid (TCCA) in CH<sub>2</sub>Cl<sub>2</sub>. This double bond exhibited the highest reactivity presumably due to a lack of negative steric (homoallylic quaternary center) and electronic influences (cinnamoyl group). Careful temperature control was crucial for this reaction. While the reaction proceeded smoothly on a milligram scale (85% yield of **14**, simple filtration), scaling up to gram quantities significantly decreased the yield to 50% and complicated the





Scheme 2 Linear synthesis of cleistocaltone A (**1**) starting from 2-acetyl phloroglucinol (**12**) and geranyl acetate (**10**).

column chromatography purification due to increased side-product formation. **14** was isolated as a diastereomeric mixture each consisting of interconverting tautomers. In the subsequent step, the allylic chloride in **14** was eliminated *via* a palladium-catalyzed  $\beta$ -hydride elimination to afford diene **9** as a mixture of two interconverting tautomers in 76% yield. The combination of sodium hydroxide and the phase-transfer catalyst tetrabutylammonium chloride ensured efficient removal of the liberated hydrochloric acid (HCl).<sup>24</sup>

With the required structural elements assembled, a thermal IMDA was performed in a biphasic mixture of water and toluene. This process afforded diastereomers **15** and **8** in 20% and 26% yields, respectively. Crystallization proved to be the most efficient purification method. Treatment of the crude product with pentane followed by cyclohexane preferentially dissolved the undesired IMDA diastereomer **15**, leaving the desired diastereomer **8** as a solid. While this purification was extremely convenient, some amount of **8** also dissolved, resulting in a reduced yield of only 26% (compared to a possible 32% as determined by <sup>1</sup>H-NMR). The relative configurations of **8** and **15** were confirmed by XRD analysis.

In exploring the impact of various solvents on the IMDA reaction's diastereomeric ratio, a 3 : 1 excess of the undesired diastereomer **15** was observed in both pyridine and *n*-butanol. Unfortunately, no other tested solvent yielded an excess of the desired diastereomer **8**. Consequently, a toluene/water mixture was chosen, allowing the reaction to proceed at a lower temperature (80 °C), which minimized side product formation and simplified purification.

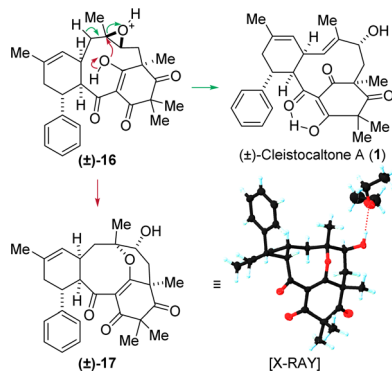
The epoxidation of **8** presented a challenge, as its two trisubstituted double bonds appeared electronically similar. We speculated that strain-release in the transition state would render the macrocyclic double bond significantly more reactive.

The facial preference was predicted from the X-ray crystal structure of the starting material. Treatment of **8** with peroxyacetic acid and sodium acetate in a biphasic water/dichloromethane mixture selectively furnished epoxide **16** as a single diastereomer in 66% yield.

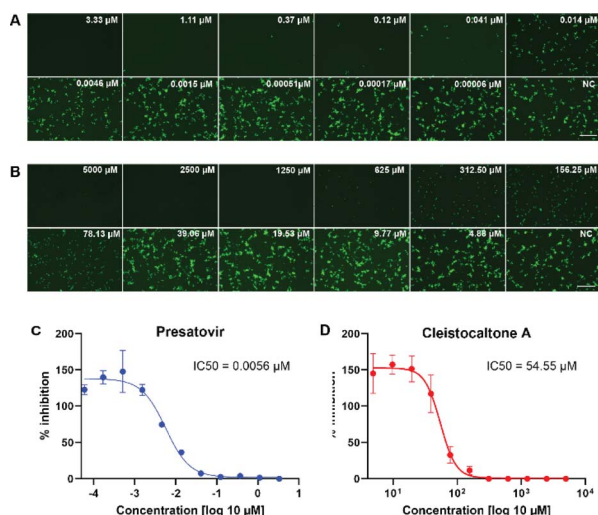
Various conditions for opening epoxide **16** were investigated. Lewis acidic conditions generally led to decomposition. Basic conditions, even using strong bases such as *n*-butyl lithium at room temperature, resulted in no conversion. This lack of reactivity is attributed to steric hindrance, with the macrocycle shielding the acidic  $\alpha$ -position adjacent to the epoxide. Fortunately, Brønsted acidic conditions proved successful. Treatment with HCl at 50 °C opened the epoxide, yielding the allylic alcohol and, ultimately, cleistocaltone A (**1**) in a 33% yield. Additionally, the acid-mediated opening of the epoxide in **16** resulted in an intriguing side product **17**, characterized by an unusual tetracyclic structure that includes an eight-membered cyclic ether. The structure was confirmed through XRD analysis. It is proposed that this product originates from a nucleophilic attack by the enol-oxygen on the tertiary carbon center, a process initiated by the protonation of the epoxide (Scheme 3).

We investigated the antiviral efficacy of synthetic cleistocaltone A (**1**) against a contemporary recombinant RSV-A strain (0594, genotype ON1)<sup>25</sup> using an *in vitro* model system (Scheme 4). Presatovir, a known selective and potent inhibitor of RSV-induced cell fusion,<sup>26</sup> served as a positive control. We confirmed Presatovir's high antiviral efficacy in infected Vero cells, with an IC<sub>50</sub> of 0.0056  $\mu$ M. Cleistocaltone A (**1**) also inhibited RSV infection, with a modestly higher IC<sub>50</sub> of 54.55  $\mu$ M. Although the IC<sub>50</sub> of synthetic ( $\pm$ )-cleistocaltone A (**1**) is about tenfold higher than that reported for the isolated racemic compound (IC<sub>50</sub> = 6.75  $\pm$  0.75  $\mu$ M),<sup>12</sup> it remains within a relevant range.





**Scheme 3** Proposed mechanism for the formation of (±)-cleistocaltone A (1) via an acid mediated epoxide opening and cyclic ether (±)-17 via a nucleophilic attack of the enol-OH in epoxide (±)-16. Racemic 17 crystallizes as a conglomerate. The configuration of the enantiomer present in the obtained crystal was opposite to that shown for (±)-17.



**Scheme 4** Assessment of the antiviral efficacy of cleistocaltone A (1) against RSV. (A and B) Vero cells were pretreated with three-fold serial dilutions of the fusion inhibitor Presatovir or two-fold serial dilutions of cleistocaltone A (1) and infected with rRSV-A-0594-EGFP (2000 TCID<sub>50</sub>/well) in the presence of each compound. Fluorescence photomicrographs showing EGFP fluorescence were obtained by UV microscopy at 48 hours post-infection. Scale bars, 500 μm. (C and D) Quantification of EGFP fluorescence in drug treated Vero cells was performed in 4% PFA fixed cells at 48 h.p.i. using a Tecan Infinite 2000 plate reader and normalized to untreated controls wells containing 0.1% DMSO. IC<sub>50</sub> values were determined by nonlinear regression analysis using GraphPad Prism 10. Error bars represent the standard deviations from two independent experiments (*n* = 8).

## Conclusion and outlook

We successfully executed a biomimetic gram-scale synthesis of cleistocaltone A (1) using an efficient eight-step route with an overall yield of 2.2%. Our approach enables further derivatization at various stages to obtain synthetic analogues for structure–activity-relationship (SAR) studies, paving the way for potential drug development.

## Biological samples

Vero and HEP-2 cells were obtained from the American Type Culture Collection (ATCC, Manassas, USA).

## Data availability

All data (experimental procedures and characterization) that support the findings of this study are available within the article and its ESI.† Crystallographic data for compounds 8, 15, and 17 have been deposited with the Cambridge Crystallographic Data Centre under CCDC2352922, CCDC2352924, and CCDC2352923.

## Author contributions

M. C. supervised the project. L. W. developed the synthesis and carried out all synthetic experiments. M. L. and S. K. developed and conducted all biological testing. M. W. performed all XRD measurements. L. W., M. C., and M. L. wrote the manuscript.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

The authors acknowledge the assistance of the Core Facility BioSupraMol supported by the DFG and Kristin Laudeley for excellent technical support.

## Notes and references

- 1 T. Szucs, *J. Antimicrob. Chemother.*, 1999, **44**, 11–15.
- 2 H. Bolek, L. Ozisik, Z. Caliskan and M. D. Tanriover, *J. Med. Virol.*, 2022, **95**, e28153.
- 3 A. M. Fendrick, A. S. Monto, B. Nightengale and M. Sarnes, *Arch. Intern. Med.*, 2003, **163**, 487–494.
- 4 M. Savic, Y. Penders, T. Shi, A. Branche and J. Y. Pirçon, *Influenza Other Respir. Viruses*, 2022, 1–10.
- 5 P. Srikantiah, P. Vora and K. P. Klugman, *Clin. Infect. Dis.*, 2021, **73**, S177–S179.
- 6 A. C. Langedijk and L. J. Bont, *Nat. Rev. Microbiol.*, 2023, **21**, 734–749.
- 7 G. M. Cragg and D. J. Newman, *Biochim. Biophys. Acta, Gen. Subj.*, 2013, **1830**, 3670–3695.
- 8 T. Rodrigues, D. Reker, P. Schneider and G. Schneider, *Nat. Chem.*, 2016, **8**, 531–541.
- 9 A. L. Harvey, *Drug Discovery Today*, 2008, **13**, 894–901.
- 10 O. Fidan, J. Ren and J. Zhan, *World J. Tradit. Chin. Med.*, 2022, **8**, 59–76.
- 11 C. Wang, P. Wu, S. Tian, J. Xue, L. Xu, H. Li and X. Wei, *J. Nat. Prod.*, 2016, **79**, 2912–2923.
- 12 J. G. Song, J. C. Su, Q. Y. Song, R. L. Huang, W. Tang, L. J. Hu, X. J. Huang, R. W. Jiang, Y. L. Li, W. C. Ye and Y. Wang, *Org. Lett.*, 2019, **21**, 9579–9583.





- 13 M. Nazir, M. Saleem, M. I. Tousif, M. A. Anwar, F. Surup, I. Ali, D. Wang, N. Z. Mamadalieva, E. Alshammari, M. L. Ashour, A. M. Ashour, I. Ahmed, Elizbit, I. R. Green and H. Hussain, *Biomolecules*, 2021, **11**, 1–61.
- 14 Q. J. Li, P. F. Tang, X. Zhou, W. J. Lu, W. J. Xu, J. Luo and L. Y. Kong, *Bioorg. Chem.*, 2020, **104**, 104275.
- 15 X. W. Yang, M. M. Li, X. Liu, D. Ferreira, Y. Ding, J. J. Zhang, Y. Liao, H. B. Qin and G. Xu, *J. Nat. Prod.*, 2015, **78**, 885–895.
- 16 A. Bonilla, C. Duque, C. Garzón, Y. Takaishi, K. Yamaguchi, N. Hara and Y. Fujimoto, *Phytochemistry*, 2005, **66**, 1736–1740.
- 17 A. J. E. Novak, C. E. Grigglesstone and D. Trauner, *J. Am. Chem. Soc.*, 2019, **141**, 15515–15518.
- 18 T. V. de Castro, D. M. Huang, C. J. Sumby, A. L. Lawrence and J. H. George, *Chem. Sci.*, 2023, **14**, 950–954.
- 19 S. A. French, C. J. Sumby, D. M. Huang and J. H. George, *J. Am. Chem. Soc.*, 2022, **144**, 22844–22849.
- 20 R. Bao, H. Zhang and Y. Tang, *Acc. Chem. Res.*, 2021, **54**, 3720–3733.
- 21 B. M. Trost, *Tetrahedron*, 2015, **71**, 5708–5733.
- 22 K. Nakagawa-Goto, J. H. Wu and K. H. Lee, *Synth. Commun.*, 2005, **35**, 1735–1739.
- 23 S. Cannizzaro, *Adv. Cycloaddit.*, 1853, **88**, 129–130.
- 24 K. Naoto and S. Shinzo, *European Pat.*, EP1072589A2, 2001, pp. 11–12.
- 25 W. K. Jo, A. Schadenhofer, A. Habierski, F. K. Kaiser, G. Saletti, T. Ganzenmueller, E. Hage, S. Haid, T. Pietschmann, G. Hansen, T. F. Schulz, G. F. Rimmelzwaan, A. D. M. E. Osterhaus and M. Ludlow, *Proc. Natl. Acad. Sci. U. S. A.*, 2021, **118**, e2026558118.
- 26 J. P. DeVincenzo, R. J. Whitley, R. L. Mackman, C. Scaglioni-Weinlich, L. Harrison, E. Farrell, S. McBride, R. Lambkin-Williams, R. Jordan, Y. Xin, S. Ramanathan, T. O'Riordan, S. A. Lewis, X. Li, S. L. Toback, S.-L. Lin and J. W. Chien, *N. Engl. J. Med.*, 2014, **371**, 711–722.

