# Reaction Chemistry & Engineering



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# COMMUNICATION

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Providing hydrogen peroxide  $(H_2O_2)$  for enzyme-catalyzed reactions requires a careful balance between sufficient reactivity and toxicity. Herein, we demonstrate the photocatalytic synthesis of  $H_2O_2$  in a continuous operation set-up using nitrogen-doped carbon nanodots (N-CNDs) and simultaneous determination of this *in situ* synthesis by continuous measurement with a novel developed  $H_2O_2$  sensor.

Hydrogen peroxide  $(H_2O_2)$  is often used as an oxidizing agent in a variety of applications, *e.g.* wastewater treatment, industrial bleaching,<sup>1</sup> or enzymatic bioreactors. In the latter,  $H_2O_2$  participates in the reactions of peroxidases and peroxygenases,<sup>2</sup> where two specific types of enzymes have received increased attention within recent years, namely lytic polysaccharide monooxygenases (LPMO),<sup>3,4</sup> and unspecific peroxygenases (UPO).<sup>5,6</sup>

A promising way to supply  $H_2O_2$  to biocatalyzed reactions is by generating it *in situ*. This has some advantages compared to an external supply, such as, no dilution of the reaction mixture, which might change reaction parameters. Furthermore, higher (local) concentrations of  $H_2O_2$  at the inlet of a reactor can cause damage to the enzymes and affect their activity and stability. Additionally, *in situ* generation of  $H_2O_2$  might present a more environmentally friendly and safe process compared to the conventional large scale production from oxidation of anthraquinone.<sup>2,7</sup>

# At-line monitoring of hydrogen peroxide released from its photocatalytic and continuous synthesis<sup>†</sup>

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> In situ generation of  $H_2O_2$  with a photocatalyst in a bioreactor is an elegant way to provide this delicate substrate on demand.  $H_2O_2$  is produced with a photocatalyst from dissolved oxygen, and is subsequently used as co-substrate in an enzymatic reaction. In situ generation of  $H_2O_2$  has previously been reported with enzymatic reactions,<sup>8</sup> bioelectrocatalysts,<sup>2,7</sup> or with photocatalysts, such as flavin,<sup>9</sup> titanium dioxide, or graphite carbon nitride.<sup>2,7</sup> In general, the use of light for biocatalysis and biocatalysis in flow are relatively new and expanding areas of research, and the advances in the field of photobiocatalysis,<sup>10</sup> and flow biocatalysis can be found elsewhere.<sup>11</sup>

> Here, we focused our efforts on nitrogen-doped carbon nanodots (N-CNDs) as a photocatalyst in dispersed phase.<sup>12,13</sup> They are used for *in situ* photocatalytic generation of  $H_2O_2$  in a glass capillary with a fixed illumination time, which serves as a model for a continuous plug-flow reactor. Thereby, the  $H_2O_2$  production can be determined by quantification of  $H_2O_2$  in the outlet.

Determination of  $H_2O_2$  is a challenging task, and which method to use depends on the specific application. A common way is with an enzymatic assay, such as horseradish peroxidase.<sup>14,15</sup> Though, these require sampling and are not suitable for continuous measurements. Alternatively, electrochemical sensors are suited for sustained measurements. However, they tend to lack selectivity due to a relatively high potential applied.<sup>16,17</sup> Recently, we published an  $H_2O_2$  sensor based on catalytic degradation and optical determination of produced oxygen (O<sub>2</sub>). The sensor can be operated *at-line*, and reliably down to one  $\mu M H_2O_2$ .<sup>18</sup>

In this short communication, we combined an *at-line*  $H_2O_2$  sensor and photocatalytic generation of  $H_2O_2$  in flow using N-CNDs. The synthesis of  $H_2O_2$  is measured at two flow rates and four different light intensities.

### Results and discussion

 $H_2O_2$  is synthesized in a glass capillary (50  $\mu L$  volume), which acts as a model for a plug-flow reactor, by illuminating an

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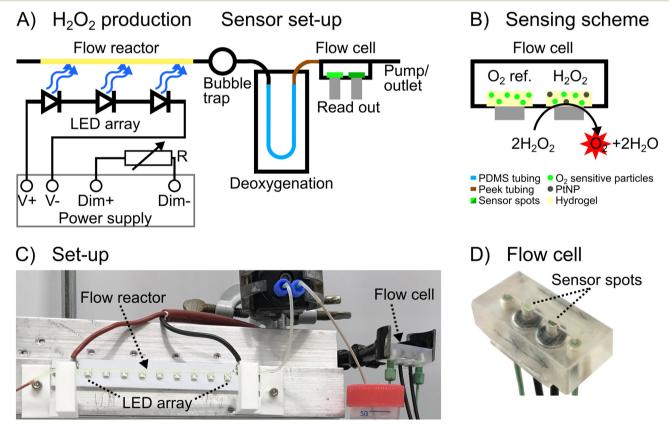
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N-CND solution flowing through the column.  $H_2O_2$  production is monitored with a connected flow cell with integrated luminescence sensor spots. The sensing scheme is based on the conversion of  $H_2O_2$  to  $O_2$  and  $H_2O$  using an immobilized inorganic catalysis (PtNP). The produced oxygen is determined by phosphorescent oxygen sensor particles, which are also embedded in the sensor spot (see Fig. 1B). The sample is deoxygenated before entering the sensor flow cell to increase the sensitivity of  $H_2O_2$  sensing as previously described.<sup>18</sup> A second sensor spot (without PtNP) is used as an  $O_2$  reference prior to the  $H_2O_2$  detection. The difference between the two sensor spots ( $\Delta pO_2$ ) is consequently the amount of  $O_2$  generated from the *in situ* synthesized  $H_2O_2$ .

The sensor flow cell is operated at flow rates of 30  $\mu$ L min<sup>-1</sup> or 50  $\mu$ L min<sup>-1</sup>. Increased H<sub>2</sub>O<sub>2</sub> concentrations cause increased  $\Delta pO_2$ , due to a higher production of O<sub>2</sub> at the H<sub>2</sub>O<sub>2</sub> sensor spot (*cf.* Fig. S1A and C<sup>†</sup>), and are directly proportional as indicated in the calibration curves (*cf.* Fig. S1B and D<sup>†</sup>). The H<sub>2</sub>O<sub>2</sub> sensor is fully reversible and shows no hysteresis. The response time of the sensor set-up (including retention time) is 5 or 3 min at flow rates of 30 or 50  $\mu$ L min<sup>-1</sup>, respectively (Fig. S1A and C<sup>†</sup>).

The sensor set-up is connected to an illuminated 50 µL glass capillary that serves as a model flow reactor with a 365 nm LED array.  $H_2O_2$  is synthesized in the flow reactor by illuminating the dispersed N-CNDs (2.5% or 5% w/v), Fig. 1A. The residence time is fixed to 35 seconds by covering parts of the capillary to control the length. All of the following results are based on a residence time of 35 s. The light intensity is regulated with resistors of 10, 22, 47, and 82 k $\Omega$ , corresponding to photon counts of 45.59, 93.30, 161.69, and 236.30  $\mu$ mol s<sup>-1</sup> m<sup>-2</sup>, respectively. The set-up is conditioned with N-CND solution prior to measurements, which are performed by switching the LED light source on/off in 15 min cycles, corresponding to 26 residence times. The sensor response to a measurement at 30 µL min<sup>-1</sup> with 2.5% w/v N-CNDs can be seen in Fig. 2A. Initially, the LED is off, and then switched on with different resistances, resulting in different light intensities (see above). This results in production of H<sub>2</sub>O<sub>2</sub>, which is measured by the sensor at 15 min. The H<sub>2</sub>O<sub>2</sub> production reaches a steady state within a few minutes, and the H<sub>2</sub>O<sub>2</sub> concentration returns to the background level of H<sub>2</sub>O<sub>2</sub> at approx. 30 min, at which time point the LED is switched off.



**Fig. 1** A) Illustration of the  $H_2O_2$  production and sensor set-up.  $H_2O_2$  is produced by a photocatalyst (N-CNDs) in a model of a flow reactor with an LED array. The produced  $H_2O_2$  solution is then passed through a deoxygenation set-up connected to a sensor flow cell. Here,  $O_2$  is measured with an  $O_2$  reference sensor spot, and  $H_2O_2$  is detected with an  $H_2O_2$  sensor spot. B)  $H_2O_2$  sensing scheme.  $H_2O_2$  is catalytically degraded to  $H_2O$  and  $O_2$  with PtNP. The amount of produced  $O_2$  is the difference between the two sensor spots.  $O_2$  is detected with  $O_2$  sensitive particles in both sensor spots. C) Photograph of the  $H_2O_2$  production and detection set-up. The image shows the flow reactor and LED array mounted on an aluminium cooling block, bubble trap, and deoxygenation set-up and sensor flow cell. All transparent tubing was covered with aluminium foil during measurements. D) Photograph of the sensor flow cell. Two sensor spots ( $O_2$  ref. and  $H_2O_2$ ) are seen in the cell. PEEK tubing (green) and optical fibres (black) for read-out are seen under the cell.

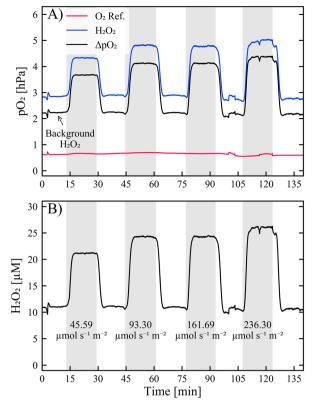


Fig. 2 A) Sensor response with 2.5% N-CND solution at 30  $\mu$ L min<sup>-1</sup>. 'O<sub>2</sub> ref.' (red line) is the amount of DO measured at the O<sub>2</sub> reference sensor spot, 'H<sub>2</sub>O<sub>2</sub>' is the amount of O<sub>2</sub> measured at the H<sub>2</sub>O<sub>2</sub> sensor spot, and ' $\Delta pO_2$ ' is the difference between the two sensor spots and the amount of produced O<sub>2</sub>. B) H<sub>2</sub>O<sub>2</sub> concentrations calculated from  $\Delta pO_2$  with the calibration curves in Fig. S1B and D.† The grey boxes indicate when the LED array is turned on, and the numbers are the photon counts in µmol photons per s m<sup>2</sup>.

 $\rm H_2O_2$  is detected in the N-CND solution without illumination (*cf.* Fig. 2). This is likely caused by a background production of  $\rm H_2O_2$  from ambient light, for instance, during purification or preparation of the solutions. The 'background  $\rm H_2O_2$ ' is approx. double the amount at double the concentration of photocatalyst N-CNDs, Fig. S3.†

The synthesized amount of  $H_2O_2$  (Fig. S3<sup>†</sup>) is calculated from the  $\Delta pO_2$  and sensor calibrations. The amount of photogenerated  $H_2O_2$  (Fig. 3) is obtained at each light intensity by subtracting the background  $H_2O_2$ . The general trend is that higher LED intensities produce larger amounts of  $H_2O_2$ . The difference in produced  $H_2O_2$  between lowest photon count of 45.59 and highest photon count of 236.30 µmol s<sup>-1</sup> m<sup>-2</sup> (resistance of 10 kOhm to 82 kOhm, respectively) is approx. 5 µM for all measurements, except for 2.5% w/v of N-CNDs, with a flow rate of 30 µL min<sup>-1</sup>, where the difference is approx. 2 µM  $H_2O_2$  (*cf.* Fig. 3).

There is little difference in the amount of  $H_2O_2$  produced at different flow rates, due to the similar residence time, (*cf.* Fig. 3). Whereas, there is a greater influence of the N-CND concentration. It should be noted that the synthesis of  $H_2O_2$ is not linear with the N-CND concentration or light intensity

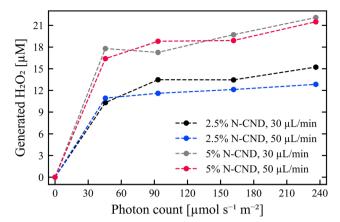


Fig. 3 Amount of  $H_2O_2$  produced with N-CND concentrations of 2.5 and 5%, and flow rates of 30 and 50  $\mu$ L min<sup>-1</sup>. The values at each light intensity are averages of produced  $H_2O_2$  with the background  $H_2O_2$  subtracted. Sensor responses can be seen in Fig. S2 and S3.†

(including zero values). Overall, this indicates that the  $H_2O_2$  synthesis is 'saturated' at the experimental conditions, which may also explain the relatively low effect of light intensity. When scaling up the system, however, the effect of light penetration depth should also be considered, as light intensity is inversely proportional to the square of the distance. Overall, the productivity of a photocatalytic system in flow set-up needs to be systematically analyzed to identify combinatory or conflicting effects of diverse operational conditions (catalyst amount, catalyst activity, residence time, wavelength of light, light intensity).<sup>19</sup>

The LED module develops heat during the experiments. This is confirmed by measuring the temperature with an optical temperature microsensor on the outside of the glass capillary. For that reason, we investigated whether the increased temperature causes autocatalytic synthesis of  $H_2O_2$ , by heating a 5% w/v N-CND solution in the absence of light (Fig. S4<sup>†</sup>). The increase in temperature did not cause synthesis of  $H_2O_2$ , since the  $\Delta pO_2$  is stable when the temperature increases up to 80 °C. Therefore, we concluded that the H<sub>2</sub>O<sub>2</sub> synthesis originates solely from the photocatalytic reaction catalyzed by N-CNDs. The N-CND solutions were further analyzed for bleaching during the measurements. UV-vis absorbance of the N-CND solutions was measured before and after light exposure (Fig. S4 and S5†). Neither the 2.5% w/v nor 5% w/v N-CND solutions showed photobleaching under the given conditions.

As mentioned by Battat, Weitz and Whitesides in a perspective article in 2022,<sup>20</sup> new technologies will be purchased and utilized if they offer a decisive advantage over older technologies. The sensor's small size and simplicity can ensure its widespread use in the future, even beyond the academic interest. Recently, fed-batch addition of  $H_2O_2$  for unspecific peroxygenase-catalyzed oxyfunctionalizations at 120 mL-scale reached the total turnover numbers up to 909 000, the highest productivity reported so far.<sup>21</sup> As consecutive steps of that technical demonstration, we believe that biotransformations using  $H_2O_2$  as a (co)substrate can

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soon be scaled up. Moving from fed-batch operation to continuous set-up can provide several advantages ranging from selectivity (byproduct-to-product) to mass and heat transfer.<sup>22</sup> In this context, precise monitoring of the inflow and outflow of  $H_2O_2$  would be essential to have full process control.

This case study is currently the first of its kind. For the first time, we have been able to show that simultaneous synthesis and concentration measurement of  $H_2O_2$  is possible. While some limitations and challenges need to be addressed in further studies, such as the restricted flow rate of a few milliliters per minute, this study marks a significant step forward in the field.

# Conclusions

We have demonstrated the photocatalytic synthesis and monitoring of  $H_2O_2$  in a continuously operated model flow reactor using N-CNDs as photocatalyst.

The synthesis of  $H_2O_2$  reaches a steady state within a few minutes of illumination with LED light sources and increases with light intensity and N-CND concentration. There is no effect of flow rate, with similar residence times between the two flow rates tested. Heating an N-CND solution in absence of light shows no production of  $H_2O_2$ , and it is therefore concluded that  $H_2O_2$  is synthesized from N-CND photocatalysis. The N-CNDs show no photobleaching during measurements.

In general, our presented hydrogen peroxide sensor is well suited for flow-chemistry set-ups, but can also be applied in conventional batch reactors to measure  $H_2O_2$  off-line or atline and is a convenient alternative to enzymatic assays. We see potential applications in determining activity of  $H_2O_2$ producing/consuming enzymes,  $H_2O_2$  as a reactive oxygen species, and  $H_2O_2$  in redox reactions. Our future experiments will be dedicated to coupling the developed set-up with UPOs for selective oxyfunctionalisations. We further envision the possibility of monitoring various bio- and non-bioprocesses relevant to both academia and industry.

In our future work, we will combine the photocatalytic system of *in situ*  $H_2O_2$  synthesis and measurement with biocatalysis, such as enzymatic oxyfunctionalization in microfluidics. This will optimize the process through precise  $H_2O_2$  monitoring and control.

# Author contributions

The paper is conceptualized by A. Ø. T. and L.-E. M. Experimental work and data analysis are carried out by A. Ø. T. and B. J. The paper is written by A. Ø. T., L.-E. M., and T. M. The project was supervised by S. K. and T. M. All authors have approved the final version of the paper.

# Conflicts of interest

There are no conflicts to declare.

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