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Discovery of pyrazole-based analogs as CDK2 inhibitors with apoptotic-inducing activity: design, synthesis and molecular dynamics study†

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The discovery of novel CDK2 inhibitors is crucial for developing targeted anticancer therapies. Thus, in this study, we aimed to design, synthesize, and evaluate a series of novel pyrazole derivatives (**2a–g**, **7a–d**, **8a** and **b**, **9**, and **10**) for their potential as CDK2/cyclin A2 enzyme inhibitors. The newly synthesized compounds were screened *in vitro* at 50 μ M for CDK2 inhibition, followed by IC₅₀ profiling of the most promising candidates. Compounds **4**, **7a**, **7d**, and **9** exhibited the strongest inhibition, with IC₅₀ values of 3.82, 2.0, 1.47, and 0.96 μ M, respectively. To assess their anti-proliferative effects, all target compounds were further screened against a panel of 60 National Cancer Institute (NCI) cell lines representing various carcinoma types. Among them, compound **4** demonstrated exceptional anti-proliferative activity with a mean growth inhibition (GI) of 96.47% across the panel, while compound **9** showed a mean GI of 65.90%. Additionally, compounds **2b** and **7c** exhibited notable inhibition against MCF7 breast cancer cells, with GI rates of 86.1% and 79.41%, respectively. Compound **4** was selected for further five-dose concentration evaluations, displaying a full-panel GI₅₀ value of 3.81 μ M, with a subpanel range of 2.36–9.17 μ M. Western blot analysis of compounds **4** and **9** in HCT-116 cell lines confirmed their inhibitory effects on CDK2. Furthermore, compound **4** induced significant cell cycle arrest at the G1 phase and promoted apoptosis. *In silico* molecular docking studies revealed that compounds **4**, **7a**, **7d**, and **9** adopt a similar binding mode as AT7519 (**I**) within the CDK2 binding site. Molecular dynamics simulations further validated the stability of these compounds within the catalytic domain of CDK2. ADME/TOPKAT analyses indicated their favorable pharmacokinetic profiles, which were confirmed by their low toxicity in normal cell lines. Based on these findings, it was concluded that the synthesized pyrazole derivatives, particularly compound **4**, show potent CDK2 inhibition and significant anticancer activity, with promising drug-like properties and minimal toxicity. This positions them as strong candidates for further development as CDK2-targeting anticancer agents.

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1. Introduction

Cell proliferation is regulated by an integrated network of proteins, which dictate the cell cycle events from order and timing perspectives. Cell cycle phases are controlled by a family of related serine/threonine proteins called cyclin-dependent kinases (CDKs), which become active upon association with their respective cyclin regulatory partners.¹ Cell cycle progression is controlled by the formation of a CDK/cyclin complex, which is responsible for the phosphorylation of target genes, such as the tumor suppressor protein retinoblastoma (Rb). A CDK/cyclin complex is activated by mitogenic signals and inhibited by the activation of cell-cycle checkpoints in response to DNA damage.² Cyclin-dependent kinase inhibitors (CKIs) negatively regulate CDKs/cyclin as the inhibitor of CDK4 (INK4)

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proteins (p16INK4a, p15INK4b, p18INK4c, and p19INK4d) and CDK2 (p21 and p27).³

CDK2-cyclin E is mainly responsible for the complete phosphorylation of retinoblastoma (Rb) in late G1, which allows the initiation of the S phase of the cell cycle, while CDK2-cyclin A eases S/G2 transition. Moreover, CDK2 plays an additional role in apoptosis, cell differentiation, immune response and the repair of normal DNA.⁴⁻⁷ Overexpression of CDK2 is predominant in many cancer types such as melanoma, glioblastoma, lymphoid tumor tissues and metastasis of prostate cancer. The irregular expression of CDK2 is often accompanied with the augmentation of its partner cyclins A and E in many human cancers as breast, endometrial, lung and thyroid carcinomas, melanoma, and osteosarcoma.^{8,9}

In recent years, computational methods have become invaluable tools in cancer diagnosis and treatment, enabling the identification of potential therapeutic targets and facilitating the design of inhibitors through molecular modeling and dynamic simulation studies. These techniques provide significant insights into the binding interactions and stability of drug candidates, accelerating the drug discovery process.¹⁰⁻¹³

In this study, pyrazole was chosen as a fundamental framework for drug exploration. This decision was driven by its molecular diversity, which accounts for the wide range of biological activities observed in different pyrazole derivatives, including antibacterial, anti-inflammatory, and anticancer activities.¹⁴⁻¹⁷ Fig. 1 illustrates a set of differently substituted pyrazole-containing compounds with outstanding CDK inhibitory activity.

AT7519 (I) is a multi CDK inhibitor for CDK 1, 2, 4, 6 and 9 isoforms with IC₅₀ values in the range of 10–210 nM.¹⁸ Interestingly, the imidazo-pyrimidine derivative **AZD5438 (II)** displayed potent CDK2 inhibition with an IC₅₀ value of 6 nM.¹⁹ Furthermore, the di-amino pyrazole derivative **CAN508 (III)** exhibited selective inhibition activity against CDK2 with an IC₅₀ value of 0.35 μ M.²⁰ Besides, 4-benzoylamino-1H-pyrazole-3-carboxamide derivative **DC-K2in212 (IV)** displayed 17-fold selectivity against CDK2 over CDK1 with an IC₅₀ value of 0.295 μ M.²¹

The binding mode of **AT7519 (I)** to the CDK2 ATP binding site disclosed the importance of the pharmacophoric pyrazole nucleus to occupy the adenine region of the ATP binding pocket. The H-bonding interactions mediated by the pyrazole

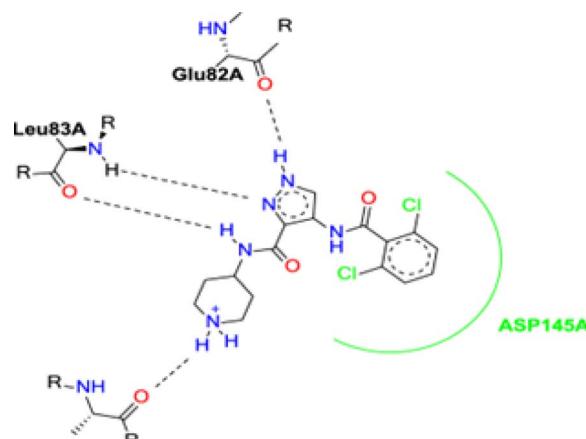


Fig. 2 Schematic of the binding mode of the lead compound **AT7519 (I)** to CDK2 ATP binding site.

core NH with Glu82 anchored **AT7519 (I)** tightly into the CDK2 hinge region. Both the carboxamide side chain N atom and pyrazole N displayed two H-bonding with Leu83. The 2,6-dichlorobenzamide moiety exhibited hydrophobic interactions with the Asp145 residue, as shown in Fig. 2.

Herein, the design of the target compounds with a pyrazole core, *i.e.*, **2a-g**, **4**, **7a-d**, **8a-b**, **9** and **10**, was derived from the structure optimization of the reference compound **AT7519 (I)** based on its reported structure–activity relationship (SAR), as follows.¹⁸

The pharmacophoric pyrazole core in the lead compound, which fills the adenine region of the ATP binding pocket is preserved. The H-bonding and hydrophobic interaction of our lead compound **AT7519 (I)** was furnished by the functional groups illustrated in Fig. 3.

Consequently, a nitrile group was introduced at position 4 in compounds **7a-d**, **8a** and **b**, **9**, and **10** to afford hydrogen bond acceptor functionality and form non-specific dipole interactions with the amino acid residues, promoting their binding with the sterically occluded protein kinases, and consequently enhancing their activity. The phenyl amino group at position 3 of the pyrazole ring acts as a hydrogen bond donor and provides occupation of a small hydrophobic pocket.

Enzyme inhibition *versus* CDK2 was performed for all the compounds. Furthermore, the targeted compounds were

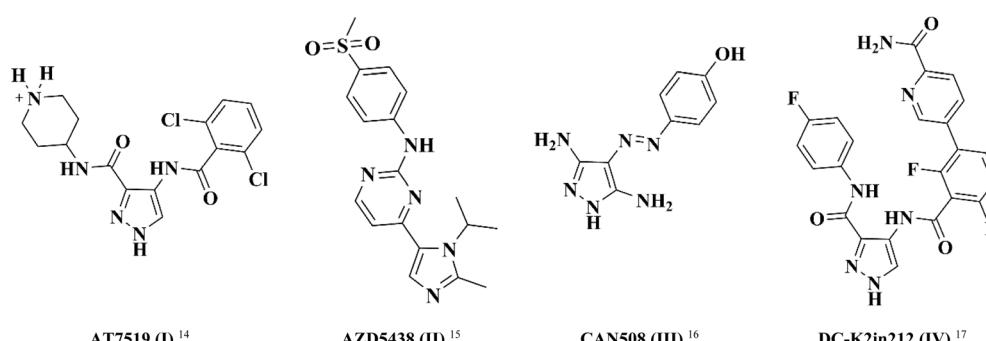


Fig. 1 Structure of clinically approved CDK2 inhibitors (I–IV).



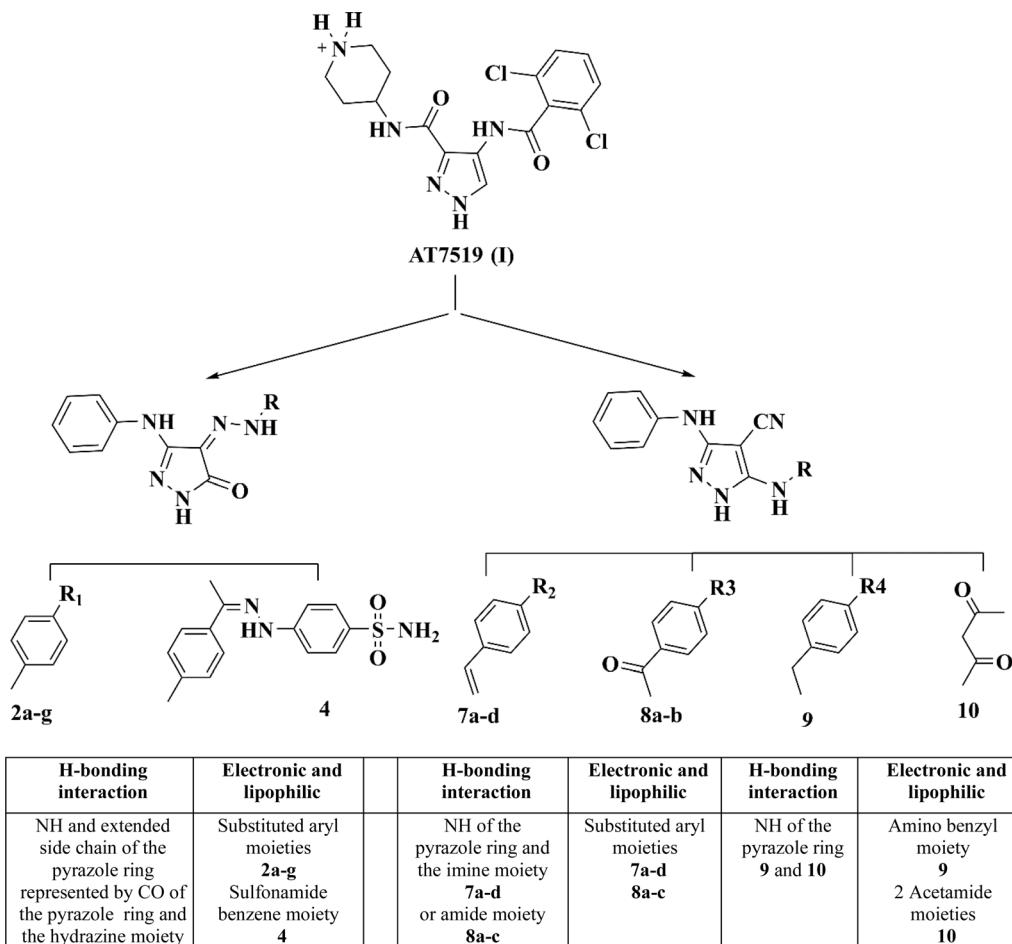


Fig. 3 Similarities of the pharmacophoric features between AT7519 (I) and the target compounds.

screened for their anticancer activity *versus* 60 human cancer cell lines from the NCI-USA. The most promising compounds were subjected to further assessment at five doses against a full NCI 60 cell panel assay. A molecular modeling study was also performed to explore their possible binding modes within the CDK2/ATP binding site. The active analogs were further investigated for their dynamic stability through a dynamic simulation process. Eventually, the ADMET computational parameters were assessed to predict the drug-likeness characteristics of the target compounds.

2. Results and discussion

2.1. Chemistry

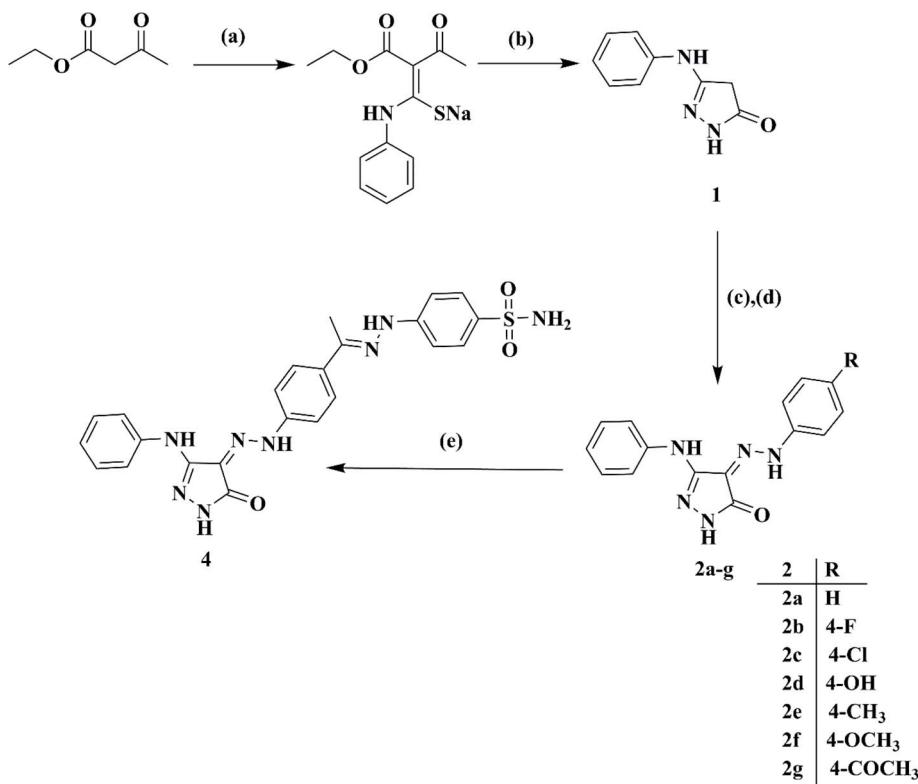
Ethyl acetoacetate was reacted with phenyl isothiocyanate in a solution of sodium methylate, followed by cyclization using hydrazine hydrate, resulting in 5-phenylamino-2,4-dihydro-pyrazol-3-one **1**.²² Subsequently, the target 4-(2-(4-un/substitutedphenyl)hydrazono)-5-(phenylamino)-2,4-dihydro-3*H*-pyrazol-3-ones **2a-g** were obtained *via* the coupling of **1** with the respective diazonium salts of different aromatic amines, as shown in Scheme 1.²³

The sulfonamide diazonium salt was reduced using a mild reducing agent, stannous chloride, to the corresponding 4-

hydrazineylbenzenesulfonamide **3**. The latter was condensed with acetyl derivative **2g** (generated from Scheme 1) in glacial acetic acid to attain pyrazole benzene sulfonamide derivative **4**.²⁴

The synthesis of 5-((4-un/substitutedbenzylidene)amino)-3-(phenylamino)-1*H*-pyrazole-4-carbonitriles **7a-d** was initiated *via* the preparation of the key 2-((methylthio)(phenylamino)methylene)malononitrile intermediate **5** by the nucleophilic addition reaction of C2 of malononitrile to phenyl isothiocyanate, followed by the methylation reaction using dimethyl sulfate.^{25,26} Subsequently, the latter was cyclized directly upon the fusion with hydrazine hydrate to afford the key intermediate 5-amino-3-phenylamino-1*H*-pyrazole-4-carbonitrile **6**,²⁷ which was refluxed with different aromatic aldehydes in glacial acetic acid to afford the final target compounds **7a-d**, as illustrated in Scheme 2.²⁸

Remarkably, 4-substituted-*N*-(4-cyano-3-(phenylamino)-1*H*-pyrazol-5-yl)benzamides **8a** and **b** were attained by the treatment of 5-amino-3-(phenylamino)-1*H*-pyrazole-4-carbonitrile **6** with the appropriate benzoyl chloride derivative with the aid of triethylamine as a catalyzing agent in dry benzene.^{29,30} Curiously, the reaction of compound **6** with benzyl chloride produced 5-(benzylamino)-3-(phenylamino)-1*H*-pyrazole-4-carbonitrile **9**, whereas the reaction of our intermediate of



Scheme 1 Synthetic route for target compounds **2a–g** and **4**. Reagents and conditions: (a) Na, CH₃OH, phenyl isothiocyanate, reflux, 30 min, cool to 50 °C. (b) Hydrazine hydrate, reflux, 1 h. (c) Ar–NH₂, HCl, sodium nitrite solution, stirring, 0–5 °C, 15 min. (d) Diazonium salt, 2 N HCl, sodium acetate, stirring, RT, 24 h. (e) **2g**, 4-hydrazinylbenzenesulfonamide **3**, C₂H₅OH, acetic acid, stirring, RT, 24 h.

interest **6** with a catalytic amount of acetic anhydride in acetic acid produced *N*-acetyl-*N*-(4-cyano-3-(phenylamino)-1*H*-pyrazol-5-yl)acetamide **10**, as illustrated in Scheme 2.³¹

The spectroscopic characteristics of compounds **2a–g**, **7a–d**, **8a** and **b**, **9**, and **10** confirmed their structures. In the case of compounds **2a–g**, the IR spectrum of compound **2d** revealed a broad OH band at the ν max of 3479 cm⁻¹, while the ¹H-NMR spectra of compounds **2e**, **2f** and **2g** revealed singlet bands corresponding to the Ar-CH₃, Ar-OCH₃ and Ar-COCH₃ protons at δ of 2.31, 3.78, 2.58 ppm, respectively. The ¹³C-NMR spectra of compounds **2f** and **2g** showed bands located at δ of 55.86 and 27.06 for Ar-OCH₃ and Ar-CH₃, respectively. The IR spectrum of compound **4** revealed bands for the expected new SO₂ group at 1338 and 1177 cm⁻¹, respectively. The ¹H-NMR spectrum of compound **4** confirmed the appearance of a singlet band for the aliphatic protons of CH₃ at δ of 4.39 ppm, protons of Ar-CH in the δ range of 6.93–7.9 ppm and identical singlet peak at δ of 6.94 ppm for the NH₂ protons. The ¹H NMR spectra of compounds **7a–d** showed a characteristic singlet signal assigned to the (N=CH) proton, which appeared in the δ range of 8.75–9.01 ppm. Compound **7c** showed a singlet signal at δ of 3.88 for OCH₃, where compound **7d** revealed a singlet signal at δ of 3.07 ppm for the 6 protons of 2CH₃. Alternatively, the ¹H NMR spectra of compounds **8a** and **b** revealed bands for NH and CO. Compound **8b** showed a singlet signal at δ of 3.89 for the OCH₃ protons. The ¹H NMR spectrum of compound **9** showed a singlet signal assigned to the benzyl proton at δ of 4.51 ppm.

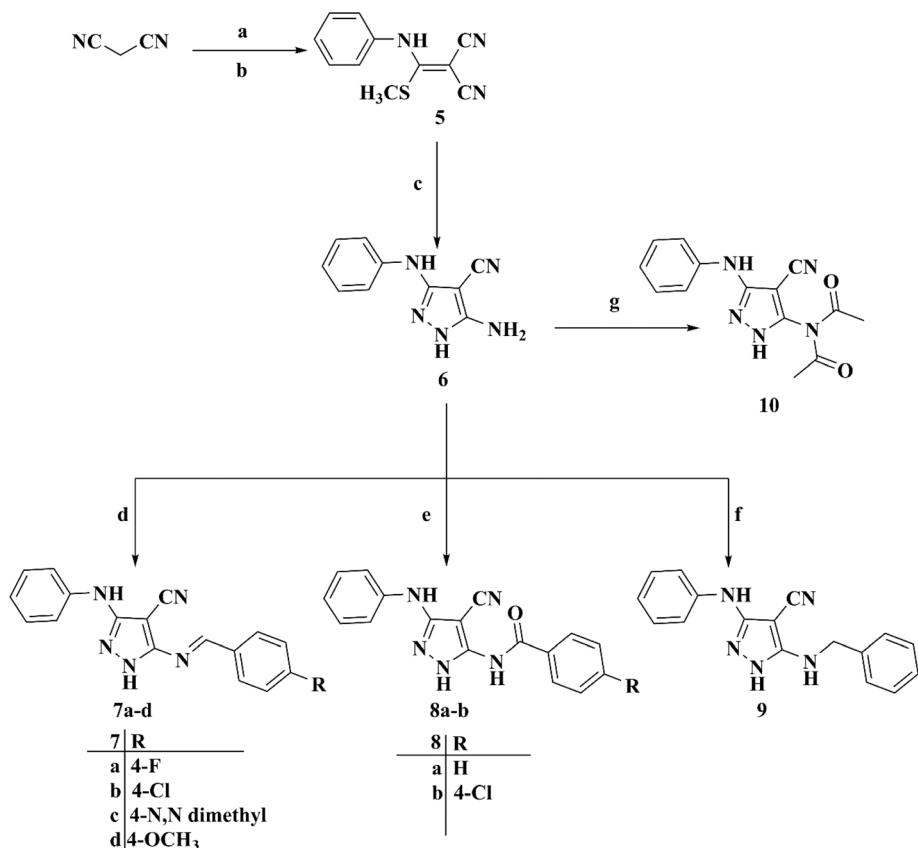
Compound **10** showed 2 extra CO bands. In contrast, compound **10** exhibited 2 singlet signals for the acetyl protons at 2.19 and 2.62 ppm. Finally, the mass spectra of all the compounds were harmonized with their calculated molecular weights.

2.2. Biological screening

2.2.1. *In vitro* cyclin-dependent kinase 2/cyclin A2 enzyme inhibition assay. The studied pyrazoles **2a–g**, **4**, **7a–d**, **8a** and **b**, **9**, and **10** were assessed for their *in vitro* CDK2/cyclin A2 enzyme inhibition assay at a concentration of 50 μ M using the Promega CDK2/cyclinA2 kinase enzyme system coupled with the ADP-Glo assay.^{32,33} The percentage enzyme activity was the lowest for five compounds, namely **4**, **7a**, **7d**, **8a**, and **9** with values of 24.7%, 17.5%, 11.5%, 27.1% and 14.5%, respectively. Following that, the IC₅₀ profiling of the promising CDK2/cyclin A2 inhibitors was tested ($n = 3$), which revealed that compound **9** has the best IC₅₀ against CDK2 of 0.96 μ M. Alternatively, compounds **7a** and **7d** showed IC₅₀ values of 2.01 and 1.47 μ M, respectively. Finally, compound **4** showed an IC₅₀ value of 3.82 μ M and compound **8b** exhibited moderate activity with an IC₅₀ value of 18.65 μ M, as shown in Fig. 4 and 5, respectively.

These results are consistent with the recent CDK2 findings,^{10–13} confirming the anticipated inhibitory potential of the selected pyrazole derivatives. The observed IC₅₀ values for compounds **9**, **7a**, **7d**, and **4**, particularly their strong activity against CDK2/cyclin A2, validate the structural modifications





Scheme 2 Synthetic route for target compounds **7a–d**, **8a** and **b**, **9**, and **10**. Reagents and reaction conditions: (a) KOH, DMF, phenyl isothiocyanate, stirring, RT, 24 h. (b) Dimethyl sulphate, stirring, RT, 8 h. (c) Hydrazine hydrate, W. B., 3–4 h. (d) Ar-CHO, acetic acid, reflux, 5 h. (e) Ar-CO-Cl, $\text{N}(\text{CH}_2\text{CH}_3)_3$, dry benzene, stirring, RT, 6 h. (f) Ar-CH₂-Cl, anhydrous K_2CO_3 , dry benzene, reflux, 6 h. (g) $(\text{CH}_3\text{CO})_2\text{O}$, acetic acid, stirring, RT, 24 h.

introduced during the design phase, further supporting their promising role as CDK2 inhibitors.

2.2.2. *In vitro* anticancer activity against 60 cell lines. The newly synthesized pyrazoles **2a–g**, **7a–d**, **8a** and **b**, **9**, and **10** were submitted to the National Cancer Institute (NCI) Developmental

Therapeutics Program (<http://www.dtp.nci.nih.gov>) for assessment of their potential antiproliferative activity against a panel of NCI 60 cancer cell lines. All compounds were approved for screening against the approved 60 cancer cell lines at a single dose of 10 μM . The human cancer cell lines

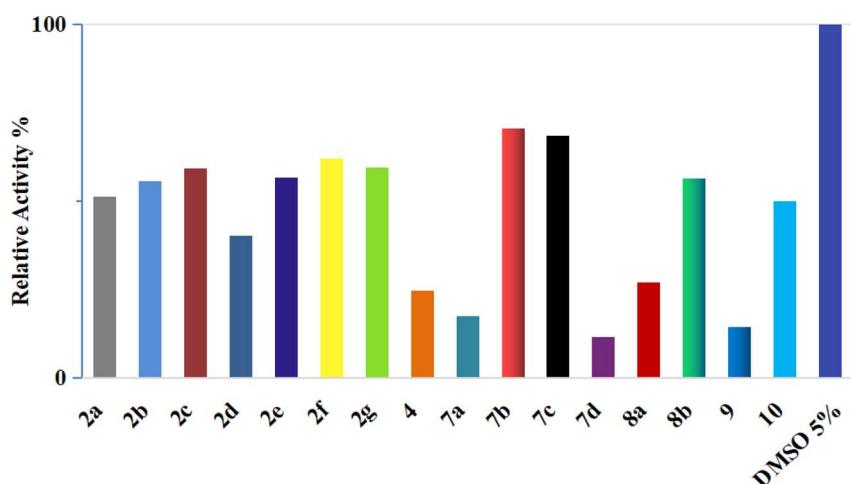


Fig. 4 Inhibition of CDK2/cyclin A2 by compounds **2a–g**, **4**, **7a–d**, **8a** and **b**, **9**, and **10** at a concentration of 50 μM .

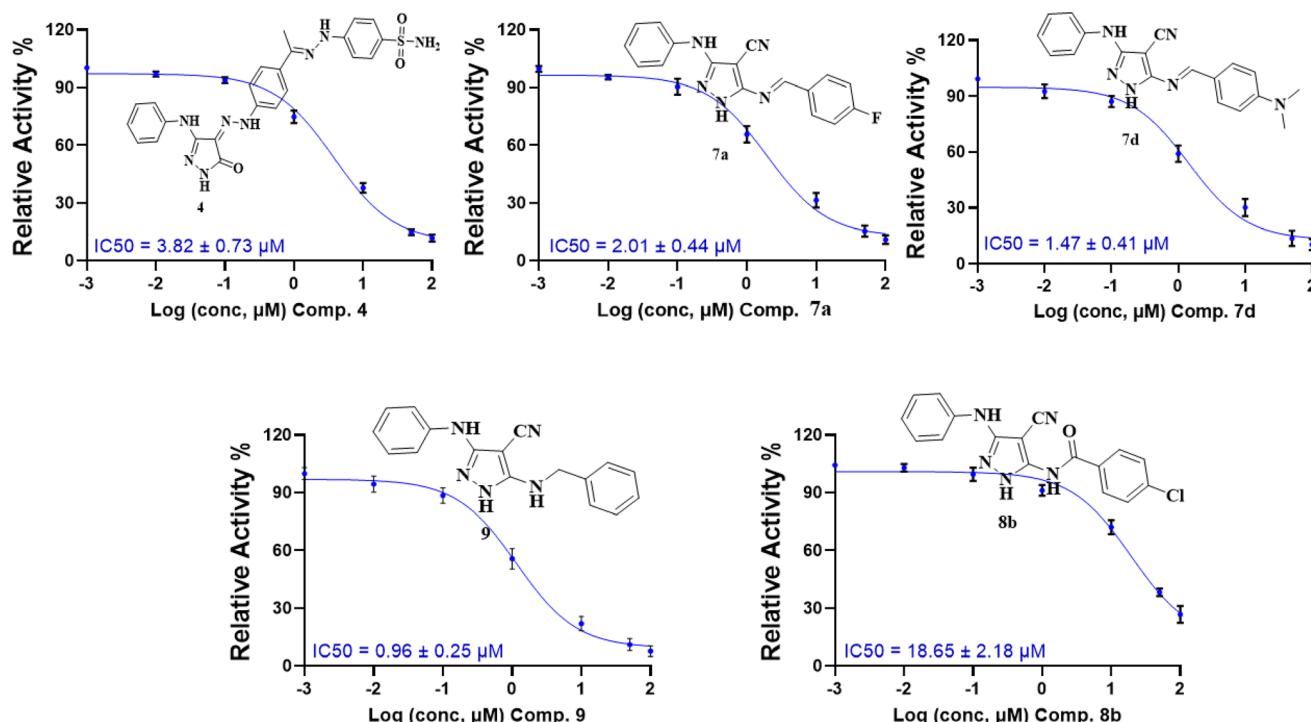


Fig. 5 Dose–response curves with the inhibitory concentration at 50% (IC_{50}) of compounds 4, 7a, 7d, 8b, and 9 using the CDK2/cyclin A2 protein kinase activity assay.

were derived from 9 variant cancer types including leukemia, melanoma, lung, colon, CNS, ovarian, renal, prostate and breast cancers. The results are presented as percentage of growth inhibition (GI%) of the screened compounds against the full panel of cell lines. The mean GI% of the screened compounds against the NCI 60 cell lines showed the remarkable activity of two compounds, 4 and 9, with the mean GI of 96.47% and 65.90%, respectively, as shown in Table 1.

A deep look at the results revealed that some derivatives of 2a–g revealed pronounced anticancer activity against some breast cancer cell lines such as compounds 2a, 2b and 2c, which

showed activity against MCF7 with GI% of 73.8%, 86.1% and 77.1%, respectively. Furthermore, analog 2b exhibited comparable anticancer activity against MDA-MB-468 with GI% of 72.25%. Alternatively, compound 7c demonstrated remarkable activity against the MCF7 breast cancer cell line with GI% of 79.41%. The achieved results revealed the influence of electronic characteristics of the side chain substituents on the anticancer activity of sulfonamide candidate 4. Therefore, the electron-withdrawing-substituted derivatives 2b and 2c and unsubstituted analog 2a exhibited pronounced anticancer activity against the breast cancer cell line. Alternatively, changing the electronic environment of the substituents to

Table 1 NCI *in vitro* testing results of compounds 4 and 9 with their corresponding GI% in μM

Panel line	Cell line	GI% compound 4	GI% compound 9
Leukemia	CCRF-CEM	96.41	81.65
	RPMI-8226	96.53	—
Melanoma cell lines	UACC-257	94.37	—
	SK-MEL-2	—	79.33
Breast cell lines	MALME-3M	—	78.10
	T-47D	—	82.76
Non-small cell lung cancer cell lines	NCI-H23	99.80	—
	NCI-H322M	90.78	—
	NCI-H460	91.13	—
Colon cancer cell lines	HT29	95.46	—
	KM12	96.31	—
	HCT-116	99.70	—
CNS cancer cell lines	SF-268	90.67	—
	SF-295	94.88	—



Table 2 NCI *in vitro* testing of compound 4 at five dose levels (GI₅₀, TGI, and LC₅₀ in μ M)

Panel line	Cell line	GI ₅₀ (μ M)	TGI (μ M)	LC ₅₀ (μ M)
Leukemia	CCRF-CEM	2.63	8.92	>100
	HL-60(TB)	2.86	0.171	60.5
	K-562	2.29	6.37	22.8
	MOLT-4	1.64	5.24	34.9
	RPMI-8226	3.36	28.2	>100
	SR	1.74	5.33	23.6
Non-small cell lung cancer	A549/ATCC	19.4	>100	>100
	EKVV	5.39	26.9	>100
	HOP-62	2.05	6	30.3
	HOP-92	1.3	6.55	30.7
	NCI-H226	3.31	12.3	35.1
	NCI-H23	2.54	6.53	23.10
	NCI-H322M	3.72	15.2	41.9
	NCI-H460	3.21	11.10	45.3
	NCI-H522	2.17	5.25	74.8
	COLO 205	3.28	12	34.6
Colon cancer	HCC-2998	2.29	6.08	21.3
	HCT-116	3.04	10.3	32
	HCT-15	5.29	18.4	42.9
	HT29	3.58	11.6	34
	KM12	2.66	8.95	29.9
	SW-620	3.18	12.6	58.8
	SF-268	2.22	6.23	31.5
CNS cancer	SF-295	3.67	13.6	38.7
	SF-539	1.58	3.68	8.56
	SNB-19	2.59	8.76	29.6
	U251	1.88	3.76	7.51
Melanoma	LOX IMVI	2.13	4.33	8.81
	MALME-3M	1.98	3.86	7.49
	M14	2.83	7.55	35.1
	MDA-MB-435	2.07	5.03	17.8
	SK-MEL-2	2.11	4.38	9.1
	SK-MEL-28	1.91	3.65	6.99
	SK-MEL-5	3.32	12.8	35.8
Ovarian cancer	UACC-257	3.97	16.7	75.5
	UACC-62	1.96	4.34	9.6
	IGROV1	4.5	17.6	43.2
	OVCAR-3	1.87	4.54	12.5
	OVCAR-4	2.84	9.26	40.7
	OVCAR-5	11	24.4	54.2
	OVCAR-8	2.59	7.81	29
Renal cancer	NCI/ADR-RES	>100	>100	>100
	SK-OV-3	2.34	6.47	23.7
	786-0	2.14	6.27	24.7
	A498	11.4	32.8	94.7
	ACHN	5.9	18.5	43
	CAKI-1	12.5	25.1	50.1
Prostate cancer	RXF 393	1.1	3.88	21.1
	SN12C	2.38	6.58	23.5
	TK-10	18.6	56.8	>100
	UO-31	19.4	>100	>100
	PC-3	3.06	11.3	35.8
Breast cancer	DU-145	3.21	10.4	32.2
	MCF7	2.4	7.34	27.4
	MDA-MB-231/ATCC	2	4.07	8.28
	HS 578T	3.31	19.4	>100
	BT-549	2.83	9.66	31.1
	T-47D	2.22	5.92	23.9
	MDA-MB-468	1.4	3	6.41



electron-donating groups led to a decrease in the anticancer activity, as seen in compounds **2d**, **2e**, and **2f** with GI% mean of 27.12%, 6.87% and 10.24%, respectively.

2.2.3. Five dose full NCI 60 cell panel assay. Owing to the promising preliminary screening results, compound **4** was remarkably chosen for assessment at an additional five doses (0.01, 0.1, 1, 10, and 100 μ M). The response parameters against several cell lines (GI_{50} , TGI and LC_{50}) were calculated and presented in Table 2, where GI_{50} indicates the compound concentration that causes a 50% decrease in the net cell growth, TGI (total growth inhibition) represents the cytostatic activity, and LC_{50} (lethal dose 50) represents the compound concentration that causes net 50% loss of the initial cells.³⁴

In addition, the mean graph midpoints (MG-MID) for the subpanel and full panel were calculated for the GI_{50} to demonstrate the average activity parameter for each compound. Commonly, compound **4** revealed obvious anticancer activity against almost the whole panel of cancer cell lines with GI_{50} values in the range of 1.1–5.9 μ M. Remarkably, compound **4** displayed noticeable cytostatic activity against the following cell lines: leukemia (HL-60(TB); TGI 0.171 μ M), breast cancer (MDA-MB-468; TGI 3 μ M), melanoma (SK MEL-28; TGI 3.65 μ M, MALME-3M; TGI 3.86 μ M), CNS cancer (SF-539; TGI 3.68 μ M and U251; TGI 3.76 μ M), and renal cancer (RXF 393; TGI 3.88 μ M).

Compound **4** exerted obvious antiproliferative activity against almost the whole NCI panel with a full panel GI_{50} (MG-MID) value of 3.81 μ M and subpanel GI_{50} (MG-MID) range of 2.6–9.17 μ M. Among the tested cancer subpanels, breast

Table 3 Mean graph mid-point values (MG-MID^a) for the GI_{50} parameter (μ M) of the subpanel cancer cell lines

Subpanel type	MG-MID	Selectivity index ^c
Leukemia	2.42	1.574
Non-small cell lung cancer	4.787778	0.795
Colon cancer	3.331429	1.143
CNS cancer	2.388	1.595
Melanoma	2.475556	1.539
Ovarian cancer	4.19	0.909
Renal cancer	9.1775	0.415
Prostate cancer	3.135	1.215
Breast cancer	2.36	1.614
Full panel MG-MID ^b	3.81	

^a MG-MID is the average activity parameter over individual subpanels/tested compound. ^b Full panel MG-MID is the average sensitivity over all cell lines (full panel)/tested compound. ^c Selectivity index was attained by dividing the full panel MG-MID (μ M) for each compound by its individual subpanel MG-MID (μ M).

^a MG-MID is the average activity parameter over individual subpanels/tested compound. ^b Full panel MG-MID is the average sensitivity over all cell lines (full panel)/tested compound. ^c Selectivity index was attained by dividing the full panel MG-MID (μ M) for each compound by its individual subpanel MG-MID (μ M).

cancer, CNS, leukemia, melanoma cancer, prostate cancer and colon cancer were the most susceptible subpanels to compound **4** with GI_{50} (MG-MID) values of 2.36, 2.388, 2.42, 2.47, 3.13 and 3.33 μ M, respectively, as shown in Table 3.

2.2.4. *In vitro* cytotoxicity against normal cells. The *in vitro* cytotoxicity of the most potent compounds **4** and **9** was evaluated against WI-38 normal cells using staurosporine as the reference standard. Compounds **4** and **9** demonstrated minimal cytotoxic effects on normal cells, with IC_{50} values of 41.16 and 56.78 μ M, respectively compared to the reference standard IC_{50} value of 28.87 μ M. These findings suggest that the newly synthesized compounds exhibit significantly lower cytotoxicity against normal cells, approximately half that observed in the cancer cell lines.

2.2.5. Flow cytometry cell cycle analysis and apoptosis study. Cell cycle analysis was conducted to explore the mechanism and the mode of action of the newly synthesized compounds as anticancer agents, typically inducing cytotoxicity by activating signaling pathways that trigger apoptosis.³⁵ The MTT cell viability assays were conducted using the mutant HCT-116 cell line, revealing that compounds **4** and **9** displayed potent cytotoxic effects, with IC_{50} values of 1.806 and 10.003 μ M, respectively. Compound **4** was further subjected to cell cycle analysis following the established protocols.³⁵ The flow cytometry results presented in Table 4 demonstrate that compound **4** decreased the percentage of cells in the G2/M phase from 6.82% to 3.79% and in the S phase from 33.81% to 23.05%, while increasing the percentage of cells in the G0/G1 phase from 59.37% to 73.16% compared to the control. The control cells exhibited a typical pattern of DNA content representing the pre-G1, S, G2/M, and G0/G1 phases of the cell cycle, as shown in Fig. 6. Conversely, the cells treated with compound **4** displayed an apoptosis pattern of DNA content, characterized by G0/G1 and G2/M phases, indicating apoptotic cell growth arrest occurring at the G1 phase, as shown in Table 5 and Fig. 7.

2.2.6. Gel electrophoresis and immuno-blot analysis of proteins (western blot). The method of sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) is employed to separate proteins based on their size. When combined with western blotting (immunoblotting), both techniques are commonly used to identify the presence and/or relative abundance of a specific protein within a sample containing a complex mixture of proteins.³⁶ In this process, each sample's total protein content is loaded onto a gel and separated by electrophoresis, allowing proteins to migrate through the gel matrix under the influence of an electric current. To facilitate protein migration, the proteins are initially denatured and given a negative charge by exposure to a detergent like SDS. A

Table 4 Flow cytometric analysis for cell cycle distribution of compound **4** on HCT-116 cells

Comp.	DNA content results				
	Conc. μ M	%G0/G1	%S	%G2/M	Results
Control	7.022	59.37	33.81	6.82	—
Comp. 4	1.806	73.16	23.05	3.79	Cell growth arrest at G1



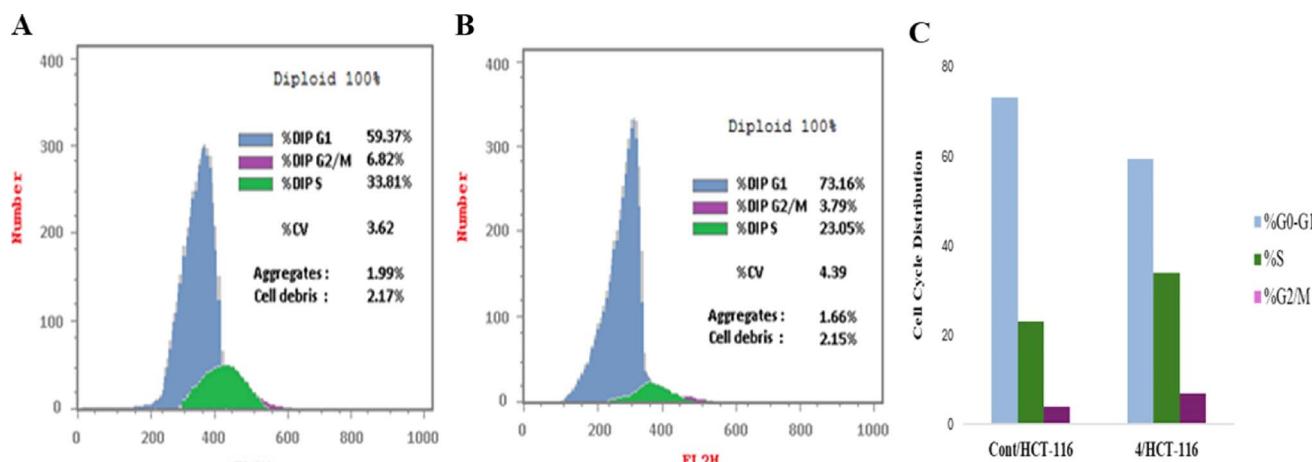


Fig. 6 Flow cytometric analysis for cell cycle distribution. (A) Control HCT-116, (B) compound 4, and (C) graphical representation for cell cycle distribution analysis among differently treated cells.

Table 5 Effect of compound 4 on apoptosis in HCT-116 cells

Apoptosis					
Comp	Conc. μ M	Total	Early	Late	Necrosis
Control	7.022	2.06	0.35	0.13	1.58
Comp. 4	1.806	27.65	4.63	15.81	7.21

molecular weight marker with known-sized bands assists in identifying proteins of interest. Once the protein components are adequately separated, they can be transferred onto a polyvinylidene fluoride (PVDF) membrane *via* electrophoresis, where they migrate out of the gel and onto the membrane. To detect a specific protein on the membrane, a primary antibody targeting that protein is added to form a protein–antibody complex, followed by the addition of a secondary antibody that binds to the complex *via* its antibody side. Typically, the

secondary antibody is linked to an enzyme that generates luminescence upon reacting with its substrate. The intensity of the luminescence, directly correlating with the amount of protein that interacted with the antibody, is captured by a Bio-Rad Imager.³⁶ HCT-116 cells were treated with either 3 μ M of compound 4 and 1 μ M of compound 9 and incubated 48 h. Treatment with compounds 4 and 9 resulted in inhibitions in the expression of CDK-2 compared to the vehicle treated cells that suggest their potential cellular inhibitory effect Fig. 8.

2.3. *In silico* studies

2.3.1. Molecular docking study. The docking patterns of compounds 4, 7a, and 9 were explored in detail, revealing the topmost CDK2 inhibition activity with IC_{50} values of 4.2, 3.98, and 0.97 μ M, respectively. Therefore, the X-ray crystal structure of CDK2 co-crystallized with AT7519 (**I**) (ref. 19) was downloaded from PDB (PDB code: 2VU3) (<http://www.rcsb.org/>).

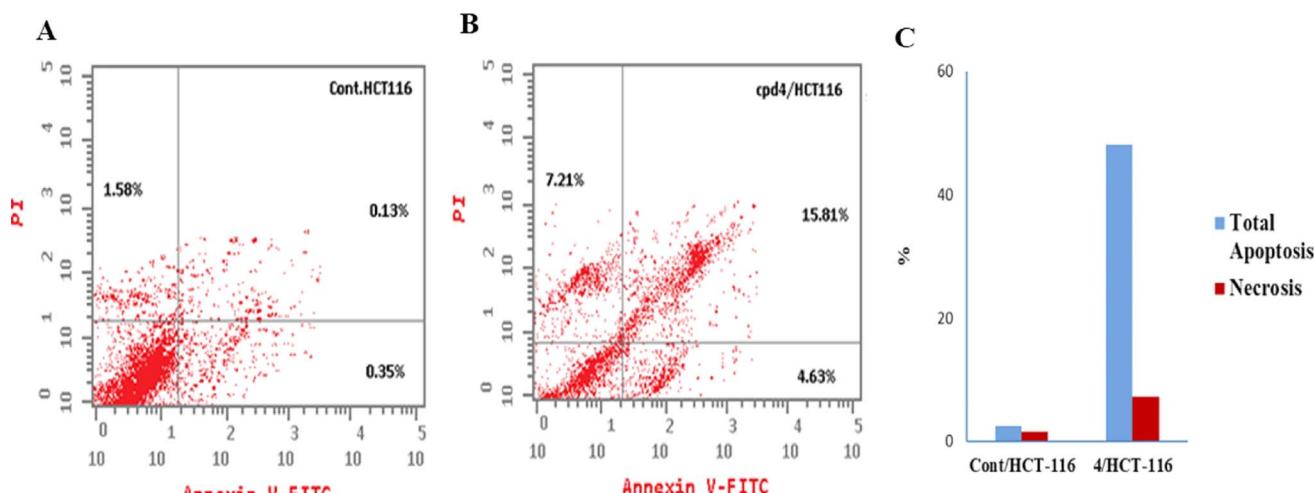


Fig. 7 Flow cytometric analysis of apoptosis among treated cells. (A) Control HCT-116, (B) compound 4, and (C) graphical illustration of apoptosis% among differently treated cells.



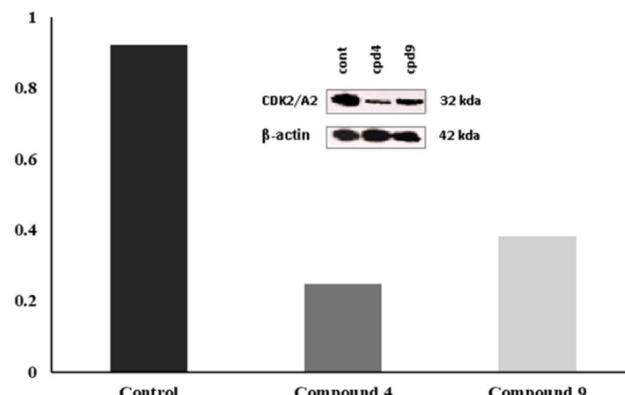


Fig. 8 Western blot assay for compounds 4 and 9.

The initial step of docking protocol was re-docking of the co-crystallized ligand to assess validation parameters as RMSD = 0.5227 Å, CDOCKER energy = $-12.8047 \text{ kcal mol}^{-1}$. Furthermore, the following step was the identification of binding mode of the native ligand which confirmed the validity of the applied docking protocol. Two H bonds with the essential Leu83 residue were mediated *via* the pyrazole core N atom and side chain NH. In addition to an extra hydrogen bond formed between His84 residue and NH piperidine. The 2,6-dichlorobenzamide moiety occupied almost all of the ATP binding region, whereby the pyrazole moiety accommodated the adenine region, as shown in Fig. 9.

Successively, we applied the same docking protocol for our compounds of interest. Most of the docked compounds presented a similar binding pattern to that of the reference AT7519 (I).

As shown in Fig. 10, the most potent CDK2 inhibitors 4 (IC_{50} of $4.2 \mu\text{M}$), 7a (IC_{50} of $3.98 \mu\text{M}$), and 9 (IC_{50} of $0.97 \mu\text{M}$) could

mediate the essential interactions with the enzyme binding site, in particular the H-bonding with the Leu83 residue *via* the C≡N group, phenylamino NH groups and the pyrazole NH; an observation that can rationalize their obtained activity. In contrast, docking of compound 4 (IC_{50} of $4.2 \mu\text{M}$) was significantly different from the reference AT7519 (I), given that it demonstrated H-bonding between Lys20 and NH_2SO_2 . The aforementioned binding pattern for compound 4 can explain its observed inhibitory activity.

2.3.2. Molecular dynamic simulation. The primary goal of the dynamic simulation and trajectory analysis is to determine whether the interaction between the docked molecules and CDK2 is stable or not, which affects their inhibition. The best active conformer of reference AT7519 (I) obtained from the docking study was selected for its dynamic stability inside the CDK2 active site and compared to that of the best active compounds 4, 7a, and 9. Fig. 11 shows the total energy *versus* the time in the range of 16–24 ps for the compounds upon interaction. The energy levels fluctuate between -8.820 and $-8.785 \text{ (kcal mol}^{-1}$), presenting their stable and preferred mode of interaction. The root mean square deviation (RMSD) of the docked complex measures how frequent the positions of the atoms in the drug-CDK2 complex change over time compared to their initial positions, as shown in Fig. 12. Here, the RMSD values are in the range of 0–4.25, except for compound 4 from 0–7.5, suggesting that the complex remains relatively stable with only minor deviations. This is a sign of a well-preserved interaction. Furthermore, the root mean square fluctuations (RMSF) were used to assess the flexibility during this simulation, as shown in Fig. 13. In this case, low RMSF values indicate stringent binding, which means that the interaction between the docked molecules and CDK2 is tight and inhibitory. In summary, in this study, dynamics simulation and various analyses were performed to demonstrate that the newly

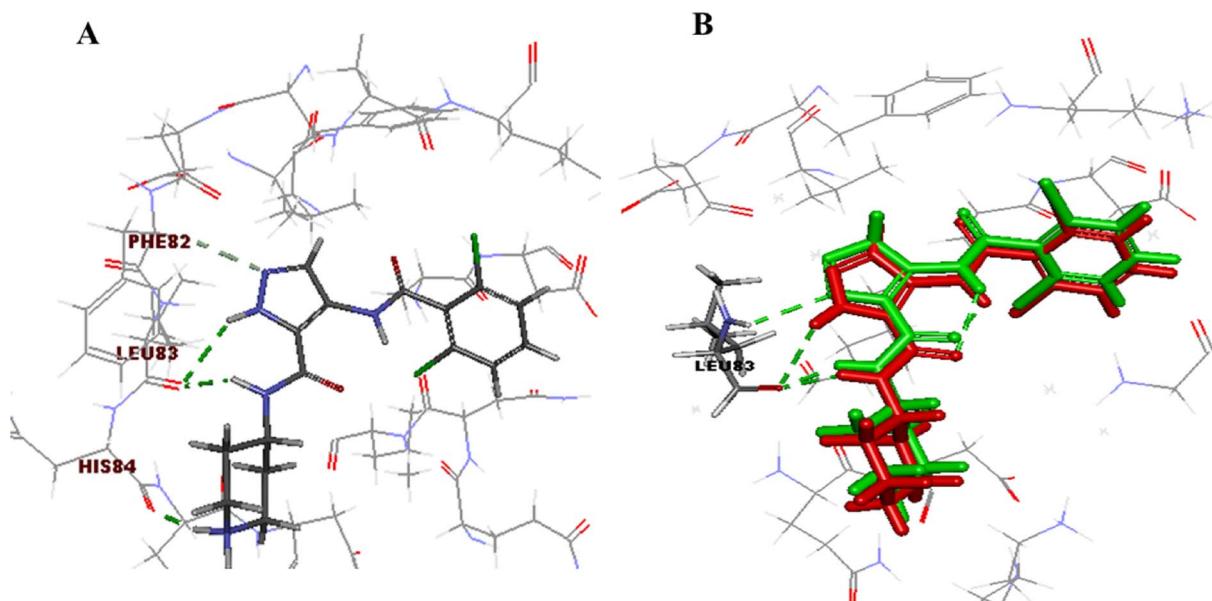


Fig. 9 2D ligand AT7519 (I) interaction diagram (A) and 3D representation (B) of superimposition of the native ligand (green) and its re-docked pose (red) in CDK2 ATP binding site.



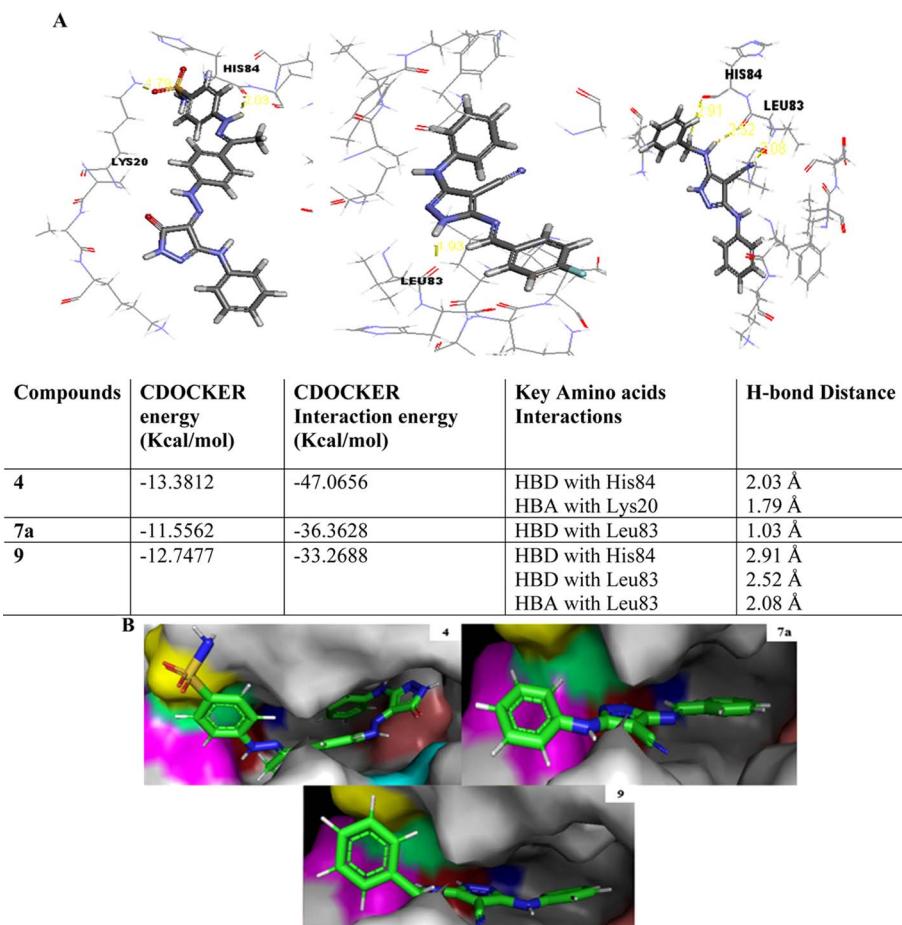


Fig. 10 Interaction diagrams presenting the best synthesized compounds 4, 7a and 9 in the binding site of the CDK2. (A) 2D representation showing H-bonding and bond distance with CDOCKER and CDOCKER interaction energies values and (B) 3D representation where yellow: Lys20, blue: Glu82, red: Leu83, pink: His84, green: Phe82, cyan: Gln131, and salmon: Asp145.

synthesized compounds form a stable and preferred interaction with CDK2, which inhibits its activity, suggesting their potential therapeutic implications.

2.3.3. Ramachandran plot. The verification of the predicted torsion angles within the targeted receptor was done using the Ramachandran plot. Discovery Studio 4.1 was used

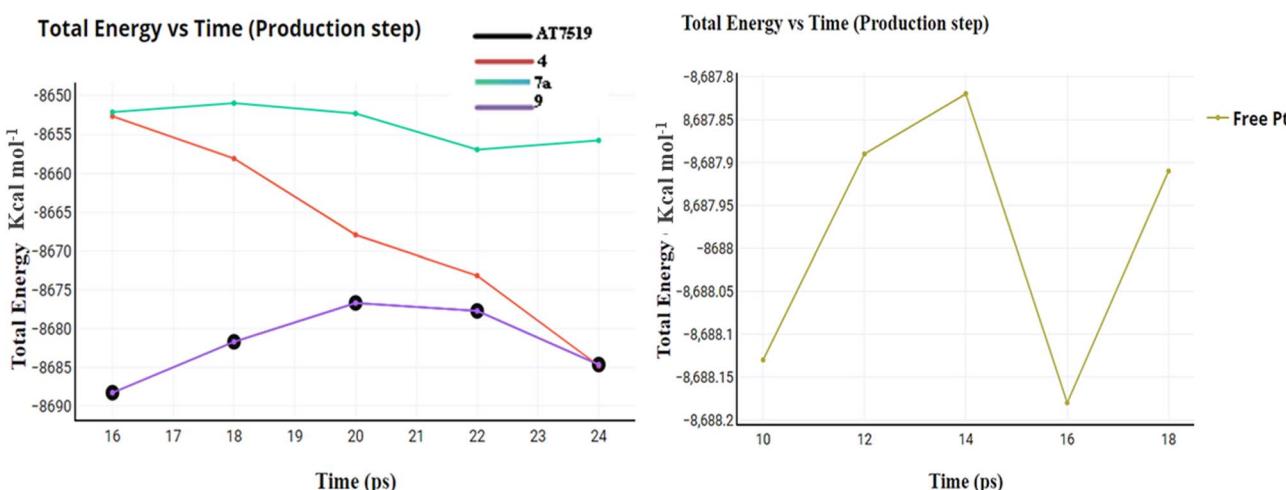


Fig. 11 Total energy versus time for compounds AT7519 (1), 4, 7a and 9 and the free protein itself.



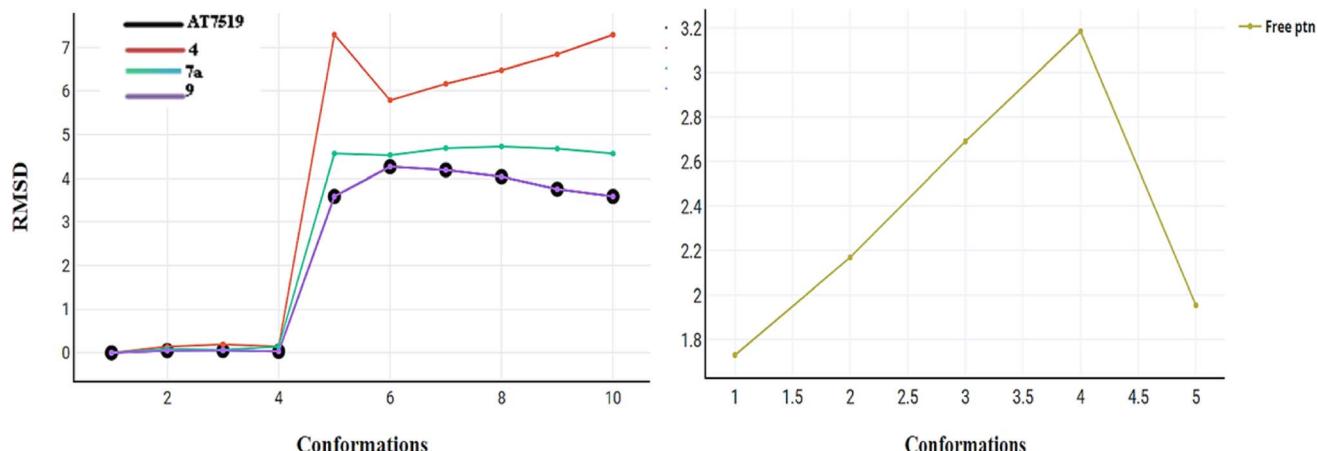


Fig. 12 RMSD for compounds AT7519 (l), 4, 7a and 9 and the free protein itself.

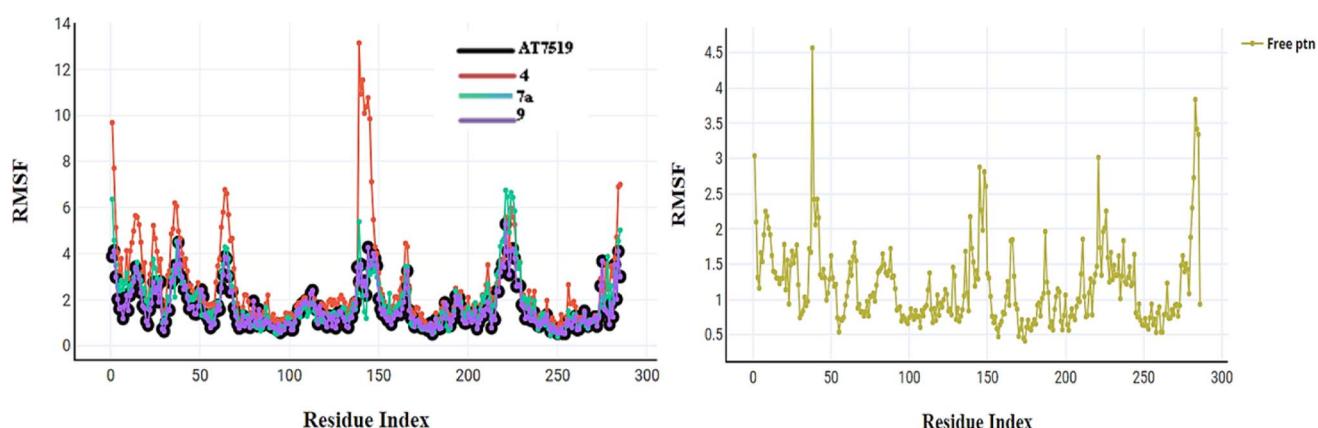


Fig. 13 RMSF for compounds AT7519 (l), 4, 7a, and 9 and the free protein itself.

before and after docking to authorize the interaction of our newly synthesized compounds in the correct binding sites and reveal the topological changes applied to the CDK2 protein. The conventional terms used to represent the torsion angles on either sides of the α carbons in peptides could be represented by the low energy conformations for ϕ (phi) and ψ (psi). The graphical representation in Fig. 14 displays the same number of favorable green areas during interaction with the docked molecules compared to the free protein, and thus ease of multiple conformations within the binding sites of the protein.

2.3.4. In silico ADMET study

2.3.4.1 *Pharmacokinetic and drug-likeness prediction.* Herein, we used the SwissADME web server (<http://www.swissadme.ch/>) to predict the drug-likeness and pharmacokinetics parameters of the novel compounds 2a–g, 7a–d, 8a and b, 9 and 10. In this work, we aimed to define the relationship between the chemical structure of our compounds of interest and definite parameters including blood–brain barrier (BBB) permeation, human gastrointestinal absorption (HIA), substrate or non-substrate for the permeability glycoprotein (P-gp), log k_p and interaction of molecules with cytochrome P450 isomers (CYP). The results are presented using the “BOILED-EGG chart (2D plot between

the calculated TPSA and log P properties of the target molecule). Thus, the GIT passive absorption probability, BBB penetration, effluxing by P-gp (PGP+) and no effluxing via P-gp (PGP–) are indicated as white area, yellow region, blue dots, and red dot, respectively.³⁷

Most of the target compounds are predicted to have no drug–drug interactions upon administration with no activity on cytochrome P450 isomers (CYP3A2 and CYP2D6).³⁸ The BOILED-Egg charts of the target compounds exhibited minimal CNS adverse effects, GIT passive absorption probability and no BBB permeability predicting. In addition, there is no possibility of tumor cell lines resistance through the efflux mechanism to the target compounds given that they may not be substrates for P-gp (PGP–).³⁹ Additionally, the target molecules are expected to have good bioavailability with score = 0.55 according to their five rule-based filters compliance⁴⁰ including Lipinski,⁴¹ Ghose,⁴² Veber,⁴³ Egan⁴⁴ and Muegge rules,⁴⁵ as shown in Fig. 15.

2.3.4.2 *Toxicity prediction.* The probable toxicities such as mutagenicity, carcinogenicity, tumorigenicity and teratogenicity were predicted by subjecting our compounds of interest to another virtual filter (Osiris Property Explorer (<http://>



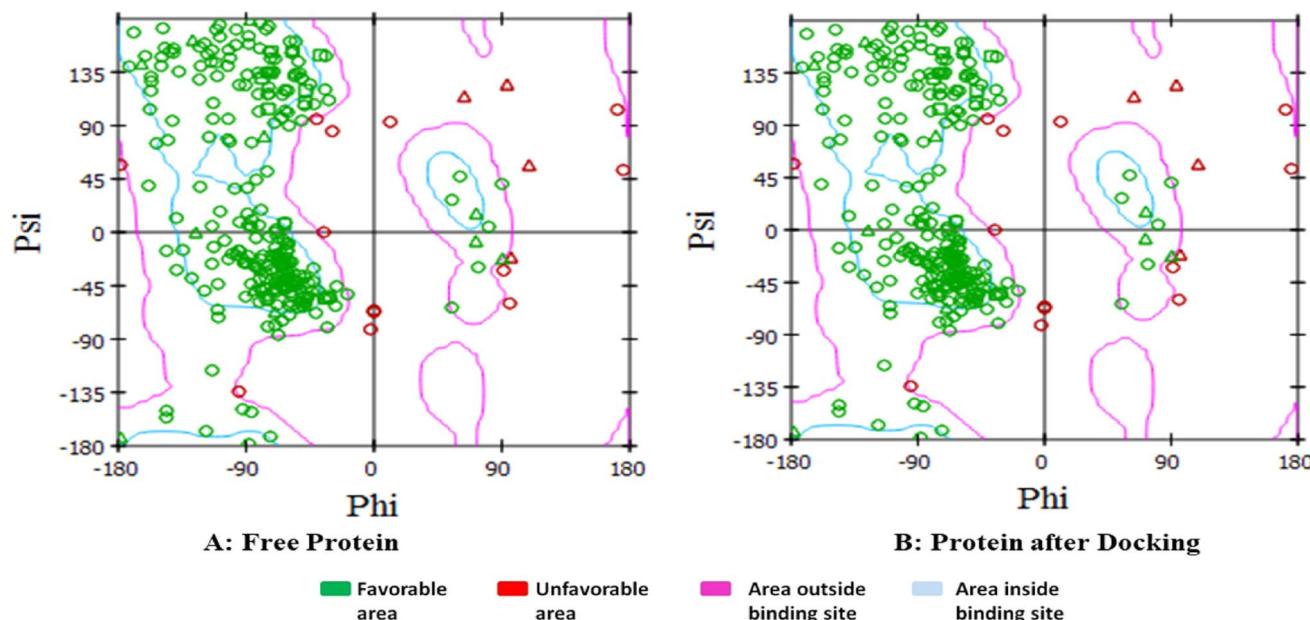


Fig. 14 Ramachandran plot presenting the torsional energy conformations for interaction between the newly synthesized molecules and CDK2, where (A) represents the free protein before docking while (B) represents the protein after docking.

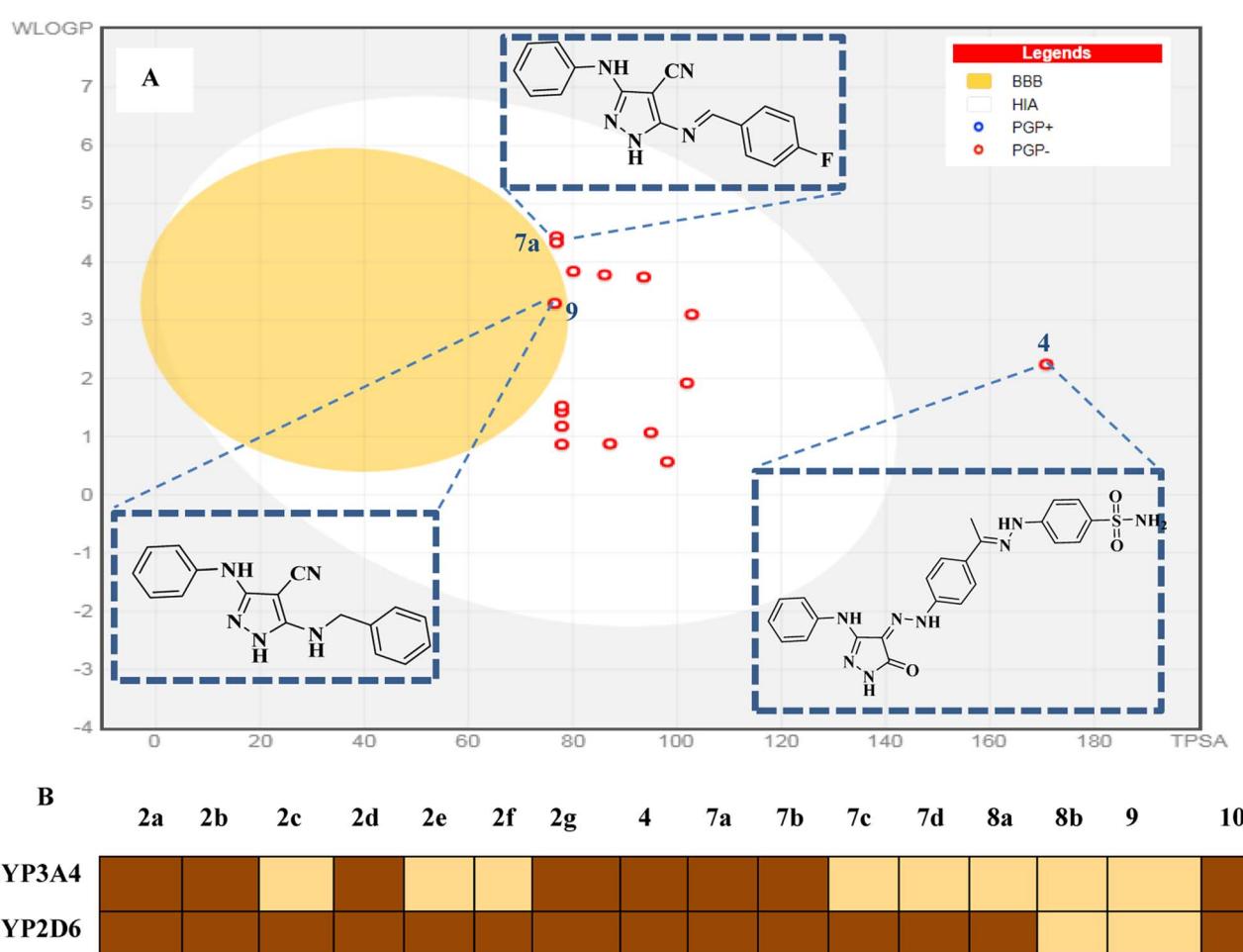


Fig. 15 Pharmacokinetic profile. (A) Boiled-egg diagram of all the synthesized compounds. White region: high probability of passive absorption by the gastrointestinal tract; yellow region (yolk): high probability of brain penetration. Yolk and the white region are not mutually exclusive. Blue point: P-gp substrate. Red point: no P-gp substrate. (B) Inhibitory capacity on the set of CYP450 enzymes.

Table 6 Computer-aided Osiris property explorer of all the studied compounds

Comp.	Probable toxicities
2a	High risk for irritant effect High risk of mutagenicity Moderate toxic effects on the reproductive system
2b	High risk for irritant effect Moderate toxic effects on the reproductive system
2c	High risk for irritant effect Moderate toxic effects on the reproductive system
2d	High risk for irritant effect Moderate toxic effects on the reproductive system
2e	High risk for irritant effect Moderate toxic effects on the reproductive system
2f	High risk for irritant effect Moderate tumorigenicity Moderate toxic effects on the reproductive system
2g	High risk of mutagenicity High risk for irritant effect Moderate toxic effects on the reproductive system
4	High risk for irritant effect Moderate toxic effects on the reproductive system
7a	Moderate risk of mutagenicity
7b	Moderate risk of mutagenicity
7c	Moderate risk of mutagenicity
7d	Moderate risk of mutagenicity High tumorigenicity
8a	Moderate risk of mutagenicity
8b	Moderate risk of mutagenicity
9	Moderate risk of mutagenicity
10	Moderate risk of mutagenicity Moderate tumorigenicity

www.organic-chemistry.org/prog/peo/). This online program compares the target compounds with *in vitro* and *in vivo* studied compounds within its database based on their functional group similarity. The color-coded results are red, green and yellow. Red color predicts high probability of toxicity, while yellow indicates moderate toxicity and green color means low toxic potential, as shown in Table 6.^{46,47} The results exhibited that some of the studied compounds are predicted to be safe and displayed low or no toxicity concerning tumorigenicity, mutagenicity, irritant effect and effect on the reproductive system.

3. Conclusion

The newly synthesized pyrazole derivatives **2a–g**, **7a–d**, **8a** and **b**, **9**, and **10** were assessed for their CDK2 inhibitory activity, and among them compounds **4**, **7a**, and **9** were found to be promising CDK2 inhibitors (IC_{50} of 4.2, 3.98, and 0.97 μ M, respectively). All the compounds were subjected to full panel screening for their anticancer activity against 60 cancer cell lines by NCI/USA. Two compounds, **4** and **9**, displayed promising growth inhibitory activity with mean GI% of 96.47 and 65.90, respectively. The NCI five dose assay of compound **4** exhibited its remarkable anticancer activity against almost the full panel (GI₅₀ range: 1.1–5.9 μ M) and (full panel GI₅₀ (MG-MID) of 3.81 μ M). Comparable binding interactions with the co-crystallized ligand **AT7519 (I)** (PDB code: 2VU3) were noticed

for the docked compounds in the CDK2 ATP binding site, especially binding with the Leu83 residue. Finally, the synthesized compounds showed good ADMET properties and toxicity profiles. As a future plan, these proposed compounds will serve as targeted scaffolds that require further optimization to enhance their therapeutic potential, including improving their selectivity, efficacy, and pharmacokinetic properties.

4. Experimental

4.1. Chemistry

The starting materials, reagents and solvents were purchased from Sigma-Aldrich (USA) or Alfa-Aesar Organics and used as received without further purification. The reactions were monitored by analytical thin layer chromatography (TLC), which was performed on pre-coated (0.25 mm) silica gel GF254 plates (E. Merck, Germany), and the compounds were detected with 254 nm UV lamp. Silica gel (60–230 mesh) was employed for routine column chromatography separation. Melting points (°C) were determined using the open capillary tube method on a Stuart SMP30 apparatus and are uncorrected. Mass spectroscopy was carried out on a direct inlet part to the mass analyzer in a Thermo Scientific GCMS model ISQ at the Regional Center for Mycology and Biotechnology (RCMB), Al-Azhar University, Nasr City, Cairo. ¹H NMR and ¹³C NMR spectra were recorded using a Bruker AVANCE III 400 MHz High-performance Digital FT-NMR spectrometer (Bruker Corporation, Germany) at the Microanalytical Unit, Faculty of Pharmacy, Cairo University and Mansoura University. ¹H/¹³C NMR spectra were run at 400/100 MHz, respectively, in DMSO-*d*6 as the solvent, and chemical shifts are quoted in δ as parts per million (ppm) downfield from tetramethylsilane (TMS) as the internal standard. Infrared spectra were determined using a Shimadzu Fourier-transform infrared spectrometer (IR-470, Shimadzu, Kyoto, Japan). All spectra were expressed as ν cm⁻¹. Compounds **1**,²² **5**,^{25,26} **3** (ref. 48) and **6** (ref. 27) were synthesized according to the reported methods.

4.1.1. General procedure for the synthesis of 4-(2-(4-un/ substitutedphenyl)hydrazono)-5-(phenylamino)-2,4-dihydro-3H-pyrazol-3-ones 2a–g. A solution of the appropriate aromatic amines (2.56 mmol) in 2 N hydrochloric acid (3 mL) was cooled to 0–5 °C in an ice bath. Subsequently, a solution of sodium nitrite (0.18 g, 2.56 mmol) in water (3 mL) was added dropwise while stirring. This mixture was stirred for 15 min at the same temperature. Next, a solution of 5-phenylamino-2,4-dihydro-pyrazol-3-one **1** (0.44 g, 2.56 mmol) in hydrochloric acid (2 N; 6 mL) was added while stirring. The solution was adjusted to neutral pH using sodium acetate and left to stir for 18–24 h at room temperature. The separated solid was collected, dried and crystallized from ethanol to give compounds **2a–g**.²³

4.1.1.1 5-(Phenylamino)-4-(2-phenylhydrazono)-2,4-dihydro-3H-pyrazol-3-one 2a. Dark-brick red powder; yield (83%); M. P.: 215–217 °C; IR: (ν max, cm⁻¹): 3460 (NH), 3150 (NH), 3050 (NH), 1598 (CO); ¹H-NMR (400 MHz, DMSO-*d*6) δ : 6.91 (t, 1H, *J* = 8, Ar-H), 7.17 (t, 1H, *J* = 8, Ar-H), 7.29 (t, 2H, *J* = 6, Ar-H), 7.44 (t, 2H, *J* = 8, Ar-H), 7.69 (d, 2H, *J* = 8, Ar-H), 7.76 (d, 2H, *J* = 8, Ar-H), 8.81 (s, 1H, NH, exchangeable by D₂O), 11.10 (s, 1H, NH,



exchangeable by D_2O), 13.06 (s, 1H, NH, exchangeable by D_2O); MS: (M. wt: 279.30): m/z , 279.11 (M^+ , [100%]); anal. calcd for $C_{15}H_{13}N_5O$; C, 64.51; H, 4.69; N, 25.07; found: C, 64.21; H, 4.75; N, 25.3.

4.1.1.2 4-(2-(4-Fluorophenyl)hydrazone)-5-(phenylamino)-2,4-dihydro-3H-pyrazol-3-one 2b. Dark-brown powder; yield (82%); M. P.: 239–240 °C; IR: (ν max, cm^{-1}): 3417 (NH), 3178 (NH), 3163 (NH), 1662 (CO); 1H -NMR (400 MHz, DMSO- d_6) δ : 6.92 (t, 1H, J = 6, Ar-H), 7.30 (dd, 4H, J = 6.6, Ar-H), 7.73–7.78 (m, 4H, Ar-H), 8.80 (s, 1H, NH, exchangeable by D_2O), 11.08 (s, 1H, NH, exchangeable by D_2O), 13.05 (s, 1H, NH, exchangeable by D_2O); MS: (M. wt: 297.29): m/z , 297.1 (M^+ , [100%]); anal. calcd for $C_{15}H_{12}FN_5O$; C, 60.60; H, 4.07; N, 23.56; found: C, 60.43; H, 4.29; N, 23.72.

4.1.1.3 4-(2-(4-Chlorophenyl)hydrazone)-5-(phenylamino)-2,4-dihydro-3H-pyrazol-3-one 2c. Dark-brick red powder; yield (86%); M. P.: 249–251 °C; IR: (ν max, cm^{-1}): 3450 (NH), 3250 (NH), 3190 (NH), 1672 (CO); 1H -NMR (400 MHz, DMSO- d_6) δ : 6.93 (t, 1H, J = 8, Ar-H), 7.31 (t, 2H, J = 6, Ar-H), 7.49 (d, 2H, J = 12, Ar-H), 7.75 (d, 4H, J = 8, Ar-H), 8.82 (s, 1H, NH, exchangeable by D_2O), 11.10 (s, 1H, NH, exchangeable by D_2O), 13.00 (s, 1H, NH, exchangeable by D_2O); MS: (M. wt: 313.74): m/z , 313.39 (M^+ , [25.3%]); anal. calcd for $C_{15}H_{12}ClN_5O$; C, 57.42; H, 3.86; N, 17.91; found: C, 57.60; H, 3.97; N, 22.18.

4.1.1.4 4-(2-(4-Hydroxyphenyl)hydrazone)-5-(phenylamino)-2,4-dihydro-3H-pyrazol-3-one 2d. Dark-red powder; yield (83%); M. P.: 257–259 °C; IR: (ν max, cm^{-1}): 3479 (OH), 3417 (NH), 3259 (NH), 3147 (NH), 1666 (CO); 1H -NMR (400 MHz, DMSO- d_6) δ : 6.83 (d, 2H, J = 8, Ar-H), 6.89 (t, 1H, J = 6, Ar-H), 7.28 (t, 2H, J = 8, Ar-H), 7.52 (d, 2H, J = 12, Ar-H), 7.73 (d, 2H, J = 8, Ar-H), 8.71 (s, 2H, NH, exchangeable by D_2O), 9.59 (s, 1H, OH, exchangeable by D_2O), 11.00 (s, 1H, NH, exchangeable by D_2O); MS: (M. wt: 295.30): m/z , 295.88 (M^+ , [104.6%]); anal. calcd for $C_{15}H_{13}N_5O_2$; C, 61.01; H, 4.44; N, 23.72; found: C, 60.89; H, 4.67; N, 23.9.

4.1.1.5 5-(Phenylamino)-4-(2-(p-tolyl)hydrazone)-2,4-dihydro-3H-pyrazol-3-one 2e. Reddish-brown powder; yield (86%); M. P.: 251–253 °C; IR: (ν max, cm^{-1}): 3302 (NH), 3151 (NH), 3024 (NH), 1670 (CO); 1H -NMR (400 MHz, DMSO- d_6) δ : 2.31 (s, 3H, CH_3), 6.89 (d, 1H, J = 3.86, Ar-H), 7.23–7.30 (m, 3H, Ar-H), 7.49 (d, 1H, J = 7.86, Ar-H), 7.58 (d, 1H, J = 8, Ar-H), 7.75 (d, 1H, J = 8, Ar-H), 8.78 (s, 1H, Ar-H), 9.17 (s, 1H, Ar-H), 10.37 (s, 1H, NH, exchangeable by D_2O), 11.07 (s, 1H, NH, exchangeable by D_2O), 13.08 (s, 1H, NH, exchangeable by D_2O); MS: (M. wt: 293.32): m/z , 293.13 (M^+ , [100%]); anal. calcd for $C_{16}H_{15}N_5O$; C, 65.52; H, 5.15; N, 23.88; found: C, 65.7; H, 5.34; N, 24.09.

4.1.1.6 4-(2-(4-Methoxyphenyl)hydrazone)-5-(phenylamino)-2,4-dihydro-3H-pyrazol-3-one 2f. Dark-brick red powder; yield (86%); M. P.: 240–242 °C; IR: (ν max, cm^{-1}): 3302 (NH), 3143 (NH), 3055 (NH), 1666 (CO); 1H -NMR (400 MHz, DMSO- d_6) δ : 3.78 (s, 3H, OCH_3), 6.90 (t, 1H, J = 8, Ar-H), 7.02 (d, 2H, J = 8, Ar-H), 7.28 (t, 2H, J = 6, Ar-H), 7.65 (d, 2H, J = 8, Ar-H), 7.75 (d, 2H, J = 8, Ar-H), 8.75 (s, 1H, NH, exchangeable by D_2O), 11.03 (s, 2H, NH, exchangeable by D_2O); ^{13}C -NMR (100 MHz, DMSO- d_6) δ : 55.86, 115.12, 115.45, 117.53, 117.61, 117.91, 118.15, 120.93, 121.69, 129.1, 129.35, 135.76, 141.33, 146.22, 157.25, 158.71; MS: (M. wt: 309.32): m/z , 309.12 (M^+ , [100%]); anal. calcd for

$C_{16}H_{15}N_5O_2$; C, 65.3; H, 4.79; N, 19.04; found: C, 62.35; H, 5.01; N, 22.87.

4.1.1.7 4-(2-(4-Acetylphenyl)hydrazone)-5-(phenylamino)-2,4-dihydro-3H-pyrazol-3-one 2g. Dark-reddish brown powder; yield (85%); M. P.: 265–267 °C; IR: (ν max, cm^{-1}): 3363 (NH), 3163 (NH), 3055 (NH), 1666 (CO), 1566 (CO); 1H -NMR (400 MHz, DMSO- d_6) δ : 2.58 (s, 3H, Ar-COCH₃), 6.91 (t, 1H, J = 6, Ar-H), 7.17 (t, 2H, J = 8, Ar-H), 7.29 (t, 2H, J = 6, Ar-H), 7.44 (t, 2H, J = 8, Ar-H), 7.69 (d, 1H, J = 8, Ar-H), 7.76 (d, 1H, J = 8, Ar-H), 8.91 (s, 1H, NH, exchangeable by D_2O), 11.18 (s, 1H, NH, exchangeable by D_2O), 13.06 (s, 1H, NH, exchangeable by D_2O); ^{13}C -NMR (100 MHz, DMSO- d_6) δ : 27.06, 115.63, 115.82, 118.15, 118.65, 121.31, 125.4, 125.82, 129.17, 129.5, 130.41, 133.02, 141.01, 146.05, 146.14, 158.15, 197; MS: (M. wt: 321.33): m/z , 321.72 (M^+ , [21.8%]); anal. calcd for $C_{17}H_{15}N_5O_2$; C, 63.54; H, 4.71; N, 21.79; found: C, 63.72; H, 4.89; N, 22.04.

4.1.2. Synthesis of 4-(2-(1-(4-(2-(5-oxo-3-(phenylamino)-1,5-dihydro-4H-pyrazol-4-ylidene)hydrazinyl)phenyl)ethylidene)hydrazinyl)benzene sulfonamide 4. To a solution of 4-(2-(4-acetylphenyl)hydrazone)-5-(phenylamino)-2,4-dihydro-3H-pyrazol-3-one **2g** (1.55 g, 10 mmol) in absolute ethanol (30 mL), a solution of 4-hydrazinylbenzenesulfonamide **3** (0.164 g, 10 mmol) in glacial acetic acid was added and the reaction mixture was stirred for 24 h at room temperature. Subsequently, the mixture was concentrated under reduced pressure and poured onto ice-cold water. The separated solid was filtered, washed with water, dried and crystallized from ethanol to give the title compound **4**.²⁴

Dark-brick red powder; yield (86%); M. P.: 249–251 °C; IR: (ν max, cm^{-1}): 3420 (NH), 3320 (NH), 3190 (NH), 1663 (CO), 1177 and 1338 (SO₂); 1H -NMR (400 MHz, DMSO- d_6) δ : 4.39 (t, 1H, J = 4.8, CH_3), 6.93 (t, 1H, Ar-H), 6.94 (s, 2H, NH₂, exchangeable by D_2O), 7.30 (d, 2H, J = 8.4, Ar-H), 7.34 (d, 2H, J = 5.6, Ar-H), 7.69 (d, 2H, J = 8.8, Ar-H), 7.73 (d, 2H, J = 8.8, Ar-H), 7.77 (d, 2H, J = 8, Ar-H), 7.90 (d, 2H, J = 8.8, Ar-H), 8.84 (s, 1H, NH, exchangeable by D_2O), 9.77 (s, 1H, NH, exchangeable by D_2O), 11.12 (s, 1H, NH, exchangeable by D_2O), 13.14 (s, 1H, NH, exchangeable by D_2O); ^{13}C -NMR (100 MHz, DMSO- d_6) δ : 13.56, 112.45, 115.97, 118.05, 121.13, 123.48, 127.03, 127.75, 129.15, 134.03, 135.58, 141.21, 141.98, 143.19, 146.15, 149.00, 158.54; MS: (M. wt: 490.54): m/z , 490.13 (M^+ , [50%]); anal. calcd for $C_{23}H_{22}N_8O_3S$; C, 56.31; H, 4.52; N, 22.84; S, 6.54; found: C, 56.47; H, 4.65; N, 23.19; S, 6.62.

4.1.3. General procedure for the synthesis of 5-((4-unsubstitutedbenzylidene)amino)-3-(phenylamino)-1H-pyrazole-4-carbonitriles **7a–d.** A mixture of 5-amino-3-(phenylamino)-1H-pyrazole-4-carbonitrile **6** (0.39 g, 2 mmol) and the appropriate aromatic aldehyde (2 mmol) in glacial acetic acid (5 mL) was refluxed for 4–5 h, the solvent was removed under vacuum and the residue was crystallized from ethanol²⁸ to give the titled compounds **7a–d**.

4.1.3.1 5-((4-Fluorobenzylidene)amino)-3-(phenylamino)-1H-pyrazole-4-carbonitrile **7a.** Dark-yellow powder; yield (81%); M. P.: 265–267 °C; IR: (ν max, cm^{-1}): 3317 (NH), 3190 (NH), 2222 (CN); 1H -NMR (400 MHz, DMSO- d_6) δ : 6.88–6.90 (m, 1H, Ar-H), 7.27 (t, 3H, J = 8, Ar-H), 7.43 (t, 3H, J = 8, Ar-H), 8.06–8.09 (m, 2H, Ar-H), 9.00 (s, 1H, imine-CH), 13.13 (s, 2H, NH, exchangeable by D_2O);



¹³C-NMR (75 MHz, DMSO-*d*6) δ : 114.66, 116.73, 116.95, 117.1, 120.96, 129.37, 132.24, 132.33, 142.24, 164.05, 166.55, 217.59; MS: (M. wt: 305.31): *m/z*, 305.11 (M^+ , [100%]); anal. calcd for C₁₇H₁₂FN₅; C, 66.88; H, 3.96; N, 22.94; found: C, 66.71; H, 4.15; N, 23.17.

4.1.3.2 5-((4-Chlorobenzylidene)amino)-3-(phenylamino)-1*H*-pyrazole-4-carbonitrile 7b. Orange powder; yield (80%); M. P.: 273–275 °C; IR: (ν max, cm^{−1}): 3344 (NH), 3190 (NH), 2218 (CN); ¹H-NMR (400 MHz, DMSO-*d*6) δ : 6.89–6.91 (m, 1H, Ar-H), 7.27 (t, 3H, J = 6, Ar-H), 7.67 (d, 2H, J = 8, Ar-H), 8.02 (d, 2H, J = 8, Ar-H), 9.01 (s, 2H, imine-CH + Ar-H), 13.18 (s, 2H, NH, exchangeable by D₂O); ¹³C-NMR (100 MHz, DMSO-*d*6) δ : 114.63, 117.12, 129.4, 129.81, 131.34, 131.68, 134.24; MS: (M. wt: 321.76): *m/z*, 321.08 (M^+ , [100%]), 323.08 (M^{+2} , [33.9%]); anal. calcd for C₁₇H₁₂ClN₅; C, 63.46; H, 3.76; N, 21.77; found: C, 63.19; H, 3.84; N, 21.98.

4.1.3.3 5-((4-Methoxybenzylidene)amino)-3-(phenylamino)-1*H*-pyrazole-4-carbonitrile 7c. Yellow powder; yield (85%); M. P.: 233–235 °C; IR: (ν max, cm^{−1}): 3332 (NH), 3190 (NH), 2214 (CN); ¹H-NMR (400 MHz, DMSO-*d*6) δ : 3.88 (s, 3H, OCH₃), 6.86–6.88 (m, 1H, Ar-H), 7.14 (d, 3H, J = 8, Ar-H), 7.26 (t, 2H, J = 8, Ar-H), 7.49 (s, 1H, NH, exchangeable by D₂O), 7.95 (d, 2H, J = 8, Ar-H), 8.91 (s, 2H, imine-CH + Ar-H), 13.05 (s, 1H, NH, exchangeable by D₂O); ¹³C-NMR (100 MHz, DMSO-*d*6) δ : 56.13, 114.94, 115.14, 115.73, 116.52, 116.98, 120.78, 127.97, 129.04, 129.37, 130.01, 131.9, 132.37, 142.24, 143.02, 163.66, 164.71, 192.02; MS: (M. wt: 317.34): *m/z*, 317.03 (M^+ , [11.3%]); anal. calcd for C₁₈H₁₅N₅O; C, 68.13; H, 4.76; N, 22.07; found: C, 68.4; H, 4.91; N, 22.33.

4.1.3.4 5-((4-(Dimethylamino)benzylidene)amino)-3-(phenylamino)-1*H*-pyrazole-4-carbonitrile 7d. Pale yellow powder; yield (83%); M. P.: 284–285 °C; IR: (ν max, cm^{−1}): 3317 (NH), 3205 (NH), 2214 (CN); ¹H-NMR (400 MHz, DMSO-*d*6) δ : 3.07 (s, 6H, 2CH₃), 6.83 (d, 3H, J = 8, Ar-H), 7.23–7.25 (m, 2H, Ar-H, NH, exchangeable by D₂O), 7.51–7.52 (m, 2H, Ar-H), 7.78 (d, 2H, J = 8, Ar-H), 8.71–8.75 (m, 1H, imine-CH + Ar-H), 12.85 (s, 1H, NH, exchangeable by D₂O); ¹³C-NMR (100 MHz, DMSO-*d*6) δ : 70.42, 111.54, 112.04, 115.33, 116.57, 116.76, 120.00, 122.33, 128.93, 129.08, 131.88, 142.82, 152.93, 154.01, 154.97, 164.71; MS: (M. wt: 330.39): *m/z*, 330.16 (M^+ , [100%]); anal. calcd for C₁₉H₁₈N₆; C, 69.07; H, 5.49; N, 25.44; found: C, 68.79; H, 5.62; N, 25.63.

4.1.4. General procedure for the synthesis of 4-substituted-N-(4-cyano-3-(phenylamino)-1*H*-pyrazol-5-yl)benzamides 8a and b. A mixture of 5-amino-3-(phenylamino)-1*H*-pyrazole-4-carbonitrile 6 (0.39 g, 2 mmol) and the appropriate benzoyl chloride derivative (2 mmol) and triethylamine (2 mmol) in dry benzene (5 mL) was stirred at room temperature for 6 h, and then the solvent was removed under vacuum, finally crystallizing the residue from ethanol to give the title compounds 8a and b.^{29,30}

4.1.4.1 4-Chloro-N-(4-cyano-3-(phenylamino)-1*H*-pyrazol-5-yl)benzamide 8a. Faint-yellow powder; yield (84%); M. P.: 169–171 °C; IR: (ν max, cm^{−1}): 3429 (NH), 3309 (NH), 3244 (NH), 2214 (CN), 1689 (CO); ¹H-NMR (400 MHz, DMSO-*d*6) δ : 6.88 (t, 1H, J = 6, Ar-H), 7.22 (t, 2H, J = 8, Ar-H), 7.54 (d, 2H, J = 8, Ar-H), 7.65 (d, 2H, J = 8, Ar-H), 8.06 (d, 2H, J = 8, Ar-H), 8.19 (s, 2H, NH, exchangeable by D₂O), 8.98 (s, 1H, NH, exchangeable by D₂O);

MS: (M. wt: 337.76): *m/z*, 337.07 (M^+ , [100%]), 339.07 (M^{+2} , [33.9%]); anal. calcd for C₁₇H₁₂ClN₅O; C, 60.45; H, 3.58; N, 20.73; found: C, 60.74; H, 3.8; N, 20.99.

4.1.4.2 N-(4-Cyano-3-(phenylamino)-1*H*-pyrazol-5-yl)-4-methoxybenzamide 8b. Yellow powder; yield (85%); M. P.: 233–235 °C; IR: (ν max, cm^{−1}): 3390 (NH), 3302 (NH), 3244 (NH), 2214 (CN), 1666 (CO); ¹H-NMR (400 MHz, DMSO-*d*6) δ : 3.89 (s, 3H, OCH₃), 6.89 (t, 1H, J = 8, Ar-H), 7.10 (d, 2H, J = 8, Ar-H), 7.24 (d, 2H, J = 8, Ar-H), 7.59 (d, 2H, J = 8, Ar-H), 8.12–8.15 (m, 4H, Ar-H + 2NH, exchangeable by D₂O), 8.96 (s, 1H, NH, exchangeable by D₂O); ¹³C-NMR (100 MHz, DMSO-*d*6) δ : 56.06, 56.14, 65.06, 113.66, 113.86, 114.24, 116.57, 118, 121.22, 123.62, 124.95, 129.08, 129.43, 133.61, 134.13, 134.23, 141.45, 151.38, 156.73, 163.24, 168.47; MS: (M. wt: 333.34): *m/z*, 333.03 (M^+ , [68.9%]); anal. calcd for C₁₈H₁₅N₅O₂; C, 64.86; H, 4.54; N, 21.01; found: C, 64.7; H, 4.72; N, 21.25.

4.1.5. Synthesis of 5-(benzylamino)-3-(phenylamino)-1*H*-pyrazole-4-carbonitrile 9. To a mixture of 5-amino-3-phenylamino-1*H*-pyrazole-4-carbonitrile 6 (0.39 g, 2 mmol) and anhydrous potassium carbonate (0.27 g, 2 mmol) in dry benzene (5 mL), an equimolar amount (0.25 g, 2 mmol) of benzyl chloride was added. The reaction mixture was directly heated at 80 °C for 6 h.³¹ After cooling to room temperature and upon pouring on ice-cold, a dark off white precipitate developed which was separated by filtration, then washed with water and recrystallized from ethanol to give the titled compound 9.

Dark-off white powder; yield (85%); M. P.: 136–137 °C; IR: (ν max, cm^{−1}): 3282 (NH), 3186 (NH), 3028 (NH), 2210 (CN); ¹H-NMR (400 MHz, DMSO-*d*6) δ : 4.52 (s, 2H, CH₂), 5.10 (s, 1H, NH, exchangeable by D₂O), 6.84–6.90 (m, 2H, Ar-H), 7.05 (d, 1H, J = 8, Ar-H), 7.22–7.33 (m, 8H, 7Ar-H, 1H, NH, exchangeable by D₂O), 8.75 (s, 1H, NH, exchangeable by D₂O); ¹³C-NMR (100 MHz, DMSO-*d*6) δ : 51.2, 52.51, 71.29, 115.26, 116.54, 121.23, 127.47, 127.52, 127.84, 128.25, 128.81, 128.86, 129.67, 137.01, 138.48, 142.65, 147.15, 156.57, 217.59; MS: (M. wt: 289.33): *m/z*, 289.69 (M^+ , [106.6%]); anal. calcd for C₁₇H₁₅N₅; C, 70.57; H, 5.23; N, 24.21; found: C, 70.68; H, 5.40; N, 24.39.

4.1.6 Synthesis of N-acetyl-N-(4-cyano-3-(phenylamino)-1*H*-pyrazol-5-yl)acetamide 10. A mixture of 5-amino-3-phenylamino-1*H*-pyrazole-4-carbonitrile 6 (0.39 g, 2 mmol), and catalytic amount of acetic anhydride in acetic acid (2 mL) was stirred at room temperature for 24 h and poured on ice/water.³¹ The separated solid was filtered, washed with water and recrystallized from ethanol to give the titled compound 10.

Off-white powder; yield (87%); M. P.: 173–175 °C; IR: (ν max, cm^{−1}): 3400 (NH), 3310 (NH), 2229 (CN), 1585 (CO), 1558 (CO); ¹H-NMR (400 MHz, DMSO-*d*6) δ : 2.19 (s, 3H, CH₃), 2.62 (s, 3H, CH₃), 6.89–6.97 (m, 1H, Ar-H), 7.25–7.32 (m, 1H, Ar-H), 7.67 (d, 1H, J = 8, Ar-H), 7.99 (s, 1H, Ar-H), 8.92 (s, 1H, Ar-H), 9.12 (s, 1H, NH, exchangeable by D₂O); 10.80 (s, 1H, NH, exchangeable by D₂O); ¹³C-NMR (100 MHz, DMSO-*d*6) δ : 23.7, 24.1, 65.07, 115.26, 116.54, 121.23, 127.47, 127.52, 127.84, 128.25, 128.81, 128.86, 129.67, 137.01, 138.48, 142.65, 147.15, 156.57, 217.59; MS: (M. wt: 283.29): *m/z*, 283.93 (M^+ , [21.4%]); anal. calcd for C₁₄H₁₃N₅O₂; C, 59.36; H, 4.63; N, 24.72; found: C, 59.54; H, 4.75; N, 24.98.



4.2. Biological assays

4.2.1. CDK2/cyclin A2 assay. The *in vitro* assay of CDK2/cyclin A2 protein kinase was carried out on all the synthesized compounds. The kinase assay was performed in a 96-well white plate with a reaction volume of 50 μ L. The reaction was performed in four steps as follows: 2.5 μ L of the test compounds or 5% DMSO and 5 μ L of CDK2/cyclin A2 enzyme (1.6 ng) was added, and then incubated at room temperature for 30 min. Then, 5 μ L of 50 μ M ATP, 0.1 μ g μ L⁻¹ histone H1 was added and incubated for 10 min. The reaction was stopped by adding 12.5 μ L of ADP-GloTM Reagent and incubated for 40 min. Then 25 μ L of kinase detection reagent was added to each well and incubated for 60 min before detection with luminescence (Integration time 0.5–1 s).^{25,32,33}

4.2.2. In vitro anti-proliferative activity. Under sterile conditions, the cell lines were grown in RPMI 1640 media (Gibco, NY, USA) supplemented with 10% fetal bovine serum (Biocell, CA, USA), and 5–10⁵ cell/mL was used to test the growth inhibition activity of the synthesized compounds. The compounds with concentrations ranging from 0.01 to 100 μ M were prepared in phosphate buffer saline. Each compound was initially solubilized in dimethyl sulfoxide (DMSO); however, each final dilution contained less than 1% DMSO. Solutions of different concentrations (0.2 mL) were pipetted into the separate well of a microtiter tray in duplicate. Cell culture (1.8 mL) containing a cell population of 6–104 cells per mL was pipetted into each well. Controls, containing only phosphate buffer saline and DMSO at identical dilutions, were also prepared in the same manner. These cultures were incubated in a humidified incubator at 37 °C. The incubator was supplied with 5% CO₂ atmosphere. After 48 h, the cells in each well were diluted 10 times with saline and counted using a Coulter counter. The counts were corrected for the dilution.³⁴

4.2.3. Flow cytometry cell cycle analysis. The cell cycle analysis protocol was performed on compound 4 on HCT-116 cells. This test is based on the content of DNA measured by staining using propidium iodide. Initially, the cells were washed in PBS before being kept at 4 °C for 3 min dropwise using the vortex addition of cold 70% ethanol to avoid cell clumping and to ensure fixation as well. Then, 50 mL of a stock of 100 mg mL⁻¹ ribonuclease was added to selectively stain only DNA. Finally, 200 mL of a stock solution of 50 mg mL⁻¹ of propidium iodide was added.³⁵

4.2.4. Flow cytometric analysis of apoptosis. For the detection of apoptosis in the treated cells, the Annexin V-FITC-apoptosis detection kit (PN IM3546) was used, followed by flow cytometric analysis according to the manufacturer's protocol. In this assay, HCT-116 cells were allowed to grow in a 25 cm³ flask until 70–80% confluence. Then, the HCT-116 cells were treated with compound 4 for 48 h, followed by washing with PBS, and suspended in binding buffer. To 100 mL of the cell suspension, 1 mL of annexin V-FITC solution and 5 mL of dissolved PI were added and incubated for 15 min in the dark. Then 400 mL of ice-cold binding buffer was added and mixed gently. The flow cytometric analysis of the percentage of apoptotic cells was

performed on a COULTER[®] EPICS[®] XLTM Flow Cytometer (USA).³⁵

4.2.5. Western blotting. HCT-116 cells were treated with either 3 μ M of compound 4 and 1 μ M of compound 9 and incubated 48 h. The cells were lysed by 1× RIPA buffer containing protease and phosphatase inhibitors to quantify the protein concentration by BCA protein assay. 20 μ g per well was loaded 12% SDS-PAGE and the protein was transferred into nitrocellulose membrane 0 and blocked with 5% BSA after that was propped against CDK2 [catalogue no. PA1547] overnight at 4 °C. The membrane was subjected to the corresponding IR-conjugated secondary antibodies. B-actin (catalogue no. P7718S) was used as a loading control. LiCOR Odyssey imager was utilized to take pictures to membrane using ImageJ software to analyze the membrane picture.³⁶

4.2.6. Molecular modeling studies. The molecular docking study was carried out using the CDOCKER protocol in the Discovery Studio 4.1 Software. The targeted compounds were docked into the CDK2 active site. The X-ray crystallographic structure of CDK2 complexed with AT7519 (I) (PDB ID: 2VU3) was downloaded from the PDB.¹⁹ The binding mode of the designed compounds was studied to explain their biological results and detect the essential hydrogen bonding with Leu83. The best pose out of ten for each compound was selected compared to the ligand binding mode. *In silico* ADMET studies using the Discovery Studio 4.1 Software and drug-likeness applying the Boiled-egg chart using <https://www.swissadme.ch/index.php> (ref. 37–45) were carried out to predict the pharmacokinetic properties of the targeted compounds, which helped in the structure requirement prediction of the observed anticancer activity.

Data availability

The data supporting this article have been included as part of the ESI.†

Conflicts of interest

The authors declare no conflict of interest.

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References

- 1 H. K. Matthews, C. Bertoli and R. A. M. de Bruin, Cell cycle control in cancer, *Nat. Rev. Mol. Cell Biol.*, 2022, 23, 74–88.
- 2 T. Otto and P. Sicinski, Cell cycle proteins as promising targets in cancer therapy, *Nat. Rev. Cancer*, 2017, 7, 93–115.
- 3 U. Asghar, A. K. Witkiewicz, N. C. Turner and E. S. Knudsen, The history and future of targeting cyclin-dependent kinases



in cancer therapy, *Nat. Rev. Drug Discovery*, 2015, **14**, 130–146.

4 N. Chunder, L. Wang, C. Chen, W. W. Hancock and A. D. Wells, Cyclin-dependent kinase 2 controls peripheral immune tolerance, *J. Immunol.*, 2012, **189**, 5659–5666.

5 P. Saurus, S. Kuusela, V. Dumont, E. Lehtonen, C. L. Fogarty, M. I. Lassenius, C. Forsblom, M. Lehto, M. A. Saleem, P. H. Groop and S. Lehtonen, Cyclin-dependent kinase 2 protects podocytes from apoptosis, *Sci. Rep.*, 2016, **6**, 21664.

6 F. Granes, M. B. Roig, H. J. Brady and G. Gil-Gomez, Cdk2 activation acts upstream of the mitochondrion during glucocorticoid induced thymocyte apoptosis, *Eur. J. Immunol.*, 2004, **34**, 2781–2790.

7 A. J. Deans, K. K. Khanna, C. J. McNees, C. Mercurio, J. Heierhorst and G. A. McArthur, Cyclin-dependent kinase 2 functions in normal DNA repair and is a therapeutic target in BRCA1-deficient cancers, *Cancer Res.*, 2006, **66**, 8219–8226.

8 X. F. Yin, J. Yu, Y. Zhou, C. Y. Wang, Z. M. Jiao, Z. N. Qian, H. Sun and B. H. Chen, Identification of CDK2 as a novel target in treatment of prostate cancer, *Future Oncol.*, 2018, **14**, 709–718.

9 P. Marion, P. Camille, P. Morgan and C. M. May, Targeting Cyclin-Dependent Kinases in Human Cancers: From Small Molecules to Peptide Inhibitors, *Cancers*, 2015, **7**, 179–237.

10 A. M. Shaker, M. I. Shahin, A. M. AboulMagd, H. M. Abdel-Rahman and D. A. Abou El Ella, Design, synthesis, and molecular docking of novel 1, 3, 4-triaryl pyrazole derivatives bearing methylsulfonyl moiety with anticancer activity through dual targeting CDK2 and COX-2 enzymes, *J. Mol. Struct.*, 2024, **1301**, 137323.

11 B. K. Chagaleti, B. S. Kumar, R. Rajagopal, A. Alfarhan, J. Arockiaraj, K. M. Kumaradoss and S. K. R. Namasivayam, Targeting cyclin-dependent kinase 2 CDK2: Insights from Molecular Docking and Dynamics Simulation-A systematic computational approach to discover novel cancer therapeutics, *Comput. Biol. Chem.*, 2024, 108134.

12 M. I. El-Gamal, S. O. Zaraei, M. M. Madkour and H. S. Anbar, Evaluation of substituted pyrazole-based kinase inhibitors in one decade (2011–2020): Current status and future prospects, *Molecules*, 2022, **27**(1), 330.

13 S. D. Packiapalavesam, V. Saravanan, A. A. Mahajan, M. H. Almutairi, B. O. Almutairi, J. Arockiaraj and S. K. R. Namasivayam, Identification of novel CA IX inhibitor: Pharmacophore modeling, docking, DFT, and dynamic simulation, *Comput. Biol. Chem.*, 2024, **110**, 108073.

14 S. I. Elewa, E. Mansour, I. F. Nassar and A. A. I. Mekawey, Synthesis of some new pyrazoline-based thiazole derivatives and evaluation of their antimicrobial, antifungal, and anticancer activities, *Russ. J. Bioorg. Chem.*, 2020, **46**, 382–392.

15 E. Eggert, R. C. Hillig, S. Koehr, D. Stockigt, J. Weiske, N. Barak, J. Mowat, T. Brumby, C. D. Christ, A. Ter Laak, T. Lang, A. E. Fernandez-Montalvan, V. Badock, H. Weinmann, I. V. Hartung, D. Barsyte-Lovejoy, M. Szewczyk, S. Kennedy, F. Li, M. Vedadi, P. J. Brown, V. Santhakumar, C. H. Arrowsmith, T. Stellfeld and C. Stresemann, Discovery and characterization of a highly potent and selective aminopyrazoline-based in vivo probe (BAY-598) for the protein lysine methyltransferase SMYD2, *J. Med. Chem.*, 2016, **59**, 4578–4600.

16 V. K. Mishra, M. Mishra, V. Kashaw and S. K. Kashaw, Synthesis of 1,3,5-trisubstituted pyrazolines as potential antimalarial and antimicrobial agents, *Bioorg. Med. Chem.*, 2017, **25**, 1949–1962.

17 M. A. Abdel-Sayed, S. M. Bayomi, M. A. El-Sherbeny, N. I. Abdel-Aziz, K. E. ElTahir, G. S. Shehatou and A. A. Abdel-Aziz, Synthesis, anti-inflammatory, analgesic, COX-1/2 inhibition activities and molecular docking study of pyrazoline derivatives, *Bioorg. Med. Chem.*, 2016, **24**, 2032–2042.

18 P. G. Wyatt, A. J. Woodhead, V. Berdini, J. A. Boulstridge, M. G. Carr, D. M. Cross, D. J. Davis, L. A. Devine, T. R. Early, R. E. Feltell, E. J. Lewis, R. L. McMenamin, E. F. Navarro, M. A. O'Brien, M. O'Reilly, M. Reule, G. Saxty, L. C. A. Seavers, D.-M. Smith, M. S. Squires, G. Trewartha, M. T. Walker and A. J.-A. Woolford, Identification of N-(4-Piperidinyl)-4-(2,6-dichlorobenzoylamino)-1H-pyrazole-3-carboxamide (AT7519), a Novel Cyclin Dependent Kinase Inhibitor Using Fragment-Based X-Ray Crystallography and Structure Based Drug Design, *J. Med. Chem.*, 2008, **51**, 4986–4999.

19 M. Anderson, D. M. Andrews, A. J. Barker, C. A. Brassington, J. Breed, K. F. Byth, J. D. Culshaw, M. R. V. Finlay, E. Fisher, H. H. J. McMiken, C. P. Green, D. W. Heaton, I. A. Nash, N. J. Newcombe, S. E. Oakes, R. A. Paupit, A. Roberts, J. J. Stanway, A. P. Thomas, J. A. Tucker, M. Walker and H. M. Weir, Imidazoles: SAR and development of a potent class of cyclin-dependent kinase inhibitors, *Bioorg. Med. Chem. Lett.*, 2008, **18**, 5487–5549.

20 C. Effects and M. Paprska, 4-Arylazo-3, 5-diamino-1 H-pyrazole CDK Inhibitors: SAR Study, Crystal Structure in Complex, *J. Med. Chem.*, 2006, **49**, 6500–6509.

21 T. Lin, J. Li, L. Liu, Y. Li, H. Jiang, K. Chen, P. Xu, C. Luo and B. Zhou, Design, synthesis, and biological evaluation of 4-benzoylamino-1H-pyrazole-3-carboxamide derivatives as potent CDK2 inhibitors, *Eur. J. Med. Chem.*, 2021, **215**, 113281.

22 H. D. P. Weissberger, Investigation of Pyrazole Compounds III: The Condensation of α Carbethoxyacetothioacetanilide with Hydrazines, *J. Am. Chem. Soc.*, 1943, **65**, 732–734.

23 S. M. Abou-seri, W. M. Eldehna, M. M. Ali, D. A. Abou and E. Ella, Piperazinylphthalazines as potential VEGFR-2 inhibitors and anticancer agents: Synthesis and *in vitro* biological evaluation, *Eur. J. Med. Chem.*, 2016, **215**, 165–179.

24 E. Z. Mohammed, W. R. Mahmoud, R. F. George, G. S. Hassan, F. A. Omar and H. H. Georgey, Synthesis, *in vitro* anticancer activity and *in silico* studies of certain pyrazole-based derivatives as potential inhibitors of cyclin dependent kinases (CDKs), *Bioorg. Chem.*, 2021, **116**, 105347.

25 S. Baumli, G. Lolli, E. D. Lowe, S. Troiani and L. A. N. Rusconi, The Structure of P-TEFb (CDK9/Cyclin

T1), its Complex with Flavopiridol and Regulation by Phosphorylation, *EMBO J.*, 2008, **27**, 1907–1918.

26 S. K. Hanks and T. Hunter, Protein Kinases 6. The Eukaryotic Protein Kinase Superfamily: Kinase (Catalytic) Domain Structure and Classification, *FASEB J.*, 1995, **9**, 576–596.

27 H. L. De Bondt, J. Rosenblatt, J. Jancarik, H. D. Jones, D. O. Morgan and D. O. Kim, Crystal Structure of Cyclin-Dependent Kinase 2, *Nature*, 1993, **363**, 595–602.

28 S. Baumli, A. J. Hole, L. Z. Wang, M. E. Noble and J. A. Endicott, The CDK9 Tail Determines the Reaction Pathway of Positive Transcription Elongation Factor B, *Structure*, 2012, **20**, 1788–1795.

29 J. Zhang, P. L. Yang and N. S. Gray, Targeting cancer with small molecule kinase inhibitors, *Nat. Rev. Cancer*, 2009, **9**, 28–39.

30 S. Baumli, G. Lolli, E. D. Lowe, S. Troiani and L. A. N. Rusconi, The Structure of P-TEFb (CDK9/Cyclin T1), its Complex with Flavopiridol and Regulation by Phosphorylation, *EMBO J.*, 2008, **27**, 1907–1918.

31 K. M. Amin, M. M. Hanna, H. E. Abo-Youssef and R. F. George, Synthesis, analgesic and anti-inflammatory activities evaluation of some bi-, tri- and tetracyclic condensed pyrimidines, *Eur. J. Med. Chem.*, 2009, **44**, 4572–4584.

32 S.-H. Lu, P.-L. Liu and F. F. Wong, Vilsmeier reagent-mediated synthesis of 6-[(formyloxy)methyl]-pyrazolopyrimidines *via* a one-pot multiple tandem reaction, *RSC Adv.*, 2015, **5**, 47098–47107.

33 J. Liu, H. Huang, X. Deng, R. Xiong, X. Cao, G. Tang, X. Wu, S. Xu and J. Peng, Design, synthesis and broad-spectrum Bcr-Abl inhibitory activity of novel thiazolamide-benzamide derivatives, *RSC Adv.*, 2019, **9**, 2092–2101.

34 S. M. Abou-seri, W. M. Eldehna, M. M. Ali, D. A. Abou and E. Ella, 1Piperazinylphthalazines as potential VEGFR-2 inhibitors and anticancer agents: Synthesis and *in vitro* biological evaluation, *Eur. J. Med. Chem.*, 2016, **107**, 165–179.

35 E. M. Azmy, M. Hagras, M. A. Ewida, A. S. Doghish, E. G. Khidr, A. A. El-Husseiny, M. H. Gomaa, H. M. Refaat, N. S. Ismail, I. F. Nassar and W. H. Lashin, Development of pyrrolo [2, 3-c] pyrazole, pyrrolo [2, 3-d] pyrimidine and their bioisosteres as novel CDK2 inhibitors with potent *in vitro* apoptotic anti-proliferative activity: Synthesis, biological evaluation and molecular dynamics investigations, *Bioorg. Med. Chem.*, 2023, **139**, 106729.

36 J. Zhang, Y. Gan, H. Li, J. Yin, X. He, L. Lin, S. Xu, Z. Fang, B. W. Kim, L. Gao and L. Ding, Inhibition of the CDK2 and Cyclin A complex leads to autophagic degradation of CDK2 in cancer cells, *Nat. Commun.*, 2022, **13**, 2835.

37 A. Daina and V. Zoete, A BOILED-Egg to Predict Gastrointestinal Absorption and Brain Penetration of Small Molecules, *ChemMedChem*, 2016, **11**, 1117–1121.

38 T. Lynch and A. Price, The effect of cytochrome P450 metabolism on drug response, interactions, and adverse effects, *Am. Fam. Physician*, 2007, **76**, 391–396.

39 M. C. O. Campos, D. B. Castro-Pinto, G. A. Ribeiro, M. M. Berredo-Pinho, L. H. F. Gomes, M. S. da Silva Bellieny, C. M. Goulart, A. Echevarria and L. L. Leon, P-glycoprotein efflux pump plays an important role in *Trypanosoma cruzi* drug resistance, *Parasitol. Res.*, 2013, **112**, 2341–2351.

40 A. Daina, O. Michelin and V. Zoete, SwissADME: A free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules, *Sci. Rep.*, 2017, **17**, 1–13.

41 C. A. Lipinski, F. Lombardo, B. W. Dominy and P. J. Feeney, Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings, *Adv. Drug Delivery Rev.*, 1997, **23**, 3–25.

42 A. K. Ghose, V. N. Viswanadhan and J. J. Wendoloski, A Knowledge-Based Approach in Designing Combinatorial or Medicinal Chemistry Libraries for Drug Discovery. A Qualitative and Quantitative Characterization of Known Drug Databases, *J. Comb. Chem.*, 1999, **1**, 55–68.

43 D. F. Veber, S. R. Johnson, H. Y. Cheng, B. R. Smith, K. W. Ward and K. D. Kopple, Molecular properties that influence the oral bioavailability of drug candidates, *J. Med. Chem.*, 2002, **45**, 2615–2623.

44 W. J. Egan, K. M. J. Merz and J. J. Baldwin, Prediction of drug absorption using multivariate statistics, *J. Med. Chem.*, 2000, **43**, 3867–3877.

45 I. Muegge, S. L. Heald and D. Brittelli, Simple Selection Criteria for Drug-like Chemical Matter, *J. Med. Chem.*, 2001, **44**, 1841–1846.

46 T. Sander, *Actelion's property explorer*, Pharm. Ltd, Gewerbestrasse, 2001, vol. 16, p. 4123.

47 T. Sander, J. Freyss, M. von Korff, J. R. Reich and C. Rufener, OSIRIS, an entirely in-house developed drug discovery informatics system, *J. Chem. Inf. Model.*, 2009, **49**, 232–246.

48 K. S. Abdelrahman, H. A. Hassan, S. A. Abdel-Aziz, A. A. Marzouk, R. shams, M. Tajiri, M. A. Aziz and H. Konno, Design, Synthesis, Molecular Docking and Biological Evaluation of Novel 1,5-Diarylpyrazole-N,O-Dimethyl Hydroxamate Derivatives as Antiproliferative agents, *J. Adv. Biomedical Pharm. Sci.*, 2021, **4**, 214–225.

