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Vitamin B₁₂-catalyzed coupling reaction of nitroalkanes and diazo compounds†

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Vitamin B₁₂ is a natural and environmentally friendly catalyst. When exposed to light or heat, central Co(ɪ) can react with electrophiles to obtain alkyl radicals, which can subsequently be used in complex processes. Herein, the vitamin B₁₂-catalyzed coupling reaction of nitroalkanes and diazo compounds is reported leading to substituted tertiary nitroalkanes in moderate yields. The reaction conditions were optimized, and the scope and limitations of the reaction were also investigated.

Introduction

Vitamin B₁₂ (cobalamin B₁₂, **1a**, Fig. 1) is the first known naturally occurring organometallic compound[1] synthesized by bacteria,[2] and plays a number of important roles in biological systems, participating in reactions such as enzymatic isomerization, methyl transfer and dehalogenation.[3] Vitamin B₁₂ is a highly functional and highly stable metal complex,[4] which can be classified into four compounds, cyanocobalamin, methylcobalamin, aquacobalamin and adenosylcobalamin, depending on the substituent group to which the central cobalt atom is attached,[5] which determines its solubility.[6] Since the central cobalt atom is easily transformed between nucleophilic reagent Co(ɪ), radical Co(ɪɪ), and electrophilic reagent Co(ɪɪɪ). In particular, the "super-nucleophilic" reagent Co(ɪ) with a lone pair of electrons reacts readily with electrophilic reagents to give alkyl cobaltamins, which undergo homolysis of the C–Co bond in the presence of light or heat to give the alkyl radical and thus complete a series of complex reactions,[7] and such natural compounds have proved to be useful catalysts for a number of organic reactions, such as alkene coupling and atom transfer radical polymerization reactions.[8] In general, vitamin B₁₂-catalysed reactions follow a radical mechanism,[9] including organic halides,[6] diazo compounds,[10] epoxides,[11] *etc.*, all of which are suitable radical sources.[7] Cobalamin has a wide appeal due to its remarkable stability, non-toxicity,[12] eco-friendly and cost-effective as compared to precious metal complexes.[13,14]

C–H bonds are ubiquitous structural units of organic molecules. Developing a capable C–H bond functionalization system has been challenging due to its chemically inert nature, although significant progress has been made using some transition metal catalysts or metallaphotoredox catalysis, such as Pd, Ru, Cu, Ag,

Co, and Fe complexes, *etc.*[15–22] Diazo compounds are commonly used as building blocks in organic synthesis due to their broad and tunable reactivity, additionally, as photocatalysis has advanced, photocatalysts reduce diazo compounds through a single electron transfer process, leading to the discovery of different novel radical reactivities.[23–25] In particular, α-carbonyl diazo compounds have a wide range of reactivities, they can be utilized in C–H functionalization, such as C(sp³)–H, aromatic C(sp²)–H, and C(sp)–H bonds insertion, in the presence of metal complexes.[26] In 2014, Frank Glorius[27] presented the first cobalt(ɪɪɪ)-catalyzed directed C–H functionalization of heteroaromatic compounds with diazo compounds (Scheme 1A). In 2015, Dorota Gryko and co-worker[28] took advantage of the high reactivity of α-carbonyl diazo compounds and used a natural non-toxic vitamin B₁₂ derivative as a catalyst to catalyze a novel alkylation reaction of alkene C(sp²)–H bond (Scheme 1B). The range of applications of vitamin B₁₂ was further broadened (Scheme 1).

In this process, ethyl diazoacetate (EDA) was reacted with cobalester in the presence of a reducing system (Zn/NH₄Cl) to give an alkyl cobalamin intermediate. Subsequently, the C–Co bond of the cobalt alkyl esters is cleaved to obtain alkyl radical, which in turn inserts into the C(sp²)–H bonds of the alkene. This reaction mechanism gave us a hint whether vitamin B₁₂ has the activity to catalyze the insertion of alkyl radicals into C(sp³)–H bonds under the stimulation of light or heat.

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Fig. 1 Structure of vitamin B₁₂ derivatives.

Cyanocobalamin, CNCbl: R = CN	**1a**
Methylcobalamin, MeCbl: R = CH₃	**1b**
Aquacobalamin, (H₂O)Cbl: R = H₂O	**1c**
Adenosylcobalamin, AdoCbl: R = 5'-deoxyadenosyl	**1d**

Scheme 1 Co(III)-catalyzed C–H functionalization with diazo compounds.
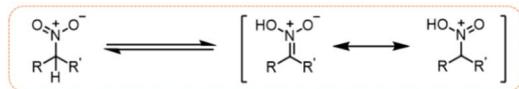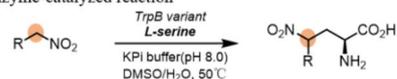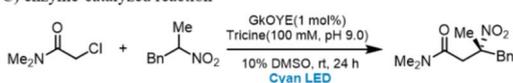
Scheme 2 Reactivity of nitroalkanes.

Nitroalkanes have been used as nucleophiles for example in Henry reaction. Even in the absence of base, these compounds readily tautomerize to a structure that is nucleophilic at carbon (Scheme 2A)[29–31] and, as such, have been used as substrates for C–C bond-forming reactions catalyzed by electrophile-activating enzymes (Scheme 2B).[31] In 2022, Todd Hyster has developed a highly chemo and stereoselective C-alkylation of nitroalkanes with alkyl halides catalyzed by an engineered flavin-dependent "ene"-reductase (ERED) (Scheme 2C).[32] Based on those reports, we envisioned that the diazo compounds can be transformed to an alkyl radical that can add to an *in situ*-generated tautomerized nitronate to form a new C–C bond.

Results

In our preliminary experiments, we found that under heating conditions, using cyanocobalamin as a catalyst and methanol as a solvent, nitrocyclohexanes and ethyl diazoacetate in the presence of the reducing agent Zn could achieve alkylation reaction of the $C(sp^3)$–H bond to give tertiary nitroalkanes (Scheme 3).

Using the reaction of nitrocyclohexane and EDA as a model reaction, we systematically performed optimization of the reaction conditions with regard to solvents, pH, reagent dosage, reaction time and temperature. When methanol or ethanol was
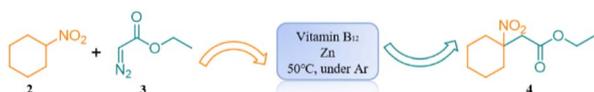
Scheme 3 Model reaction of nitrocyclohexane (**2**) with EDA (**3**).

Table 1 Optimization: the influence of solvent

Entry[a]	Solvent	Temp (°C)	Time (h)	Yield (%)
1	CH_3OH	50	24	7
2	CH_3CH_2OH	50	24	4
3	DMSO	50	24	36
4	$H_2O : DMSO = 1 : 1$	50	24	38
5	$H_2O : DMSO = 2 : 1$	50	24	36
6	$H_2O : DMSO = 1 : 2$	50	24	36
7	$(CH_3)_3COH : H_2O = 1 : 1$	50	24	13
8	$(CH_3)_2CHOH : H_2O = 1 : 1$	50	24	11
9	$THF : H_2O = 1 : 1$	50	24	7
10	$DMF : H_2O = 1 : 1$	50	24	14
11	1,4-Dioxane : $H_2O = 1 : 1$	50	24	14
12	$HFIP : H_2O = 1 : 1$	50	24	13

[a] Reaction conditions: nitrocyclohexane (0.25 mmol), EDA (2 equiv.), Zn (6 equiv.), vitamin B_{12} (2 mol%), solvent (2.5 mL), 24 h.

used, nitrocyclohexane can coupled with EDA gave traces of tertiary nitroalkanes, and when the solvent was changed into DMSO, the yields were greatly improved (Table 1, entries 1–3), the presence of water would slightly increase the reaction yield to 38%, which can improve the solubility of vitamin B_{12}. We next investigated the effects of mixed solvents with water, including isopropanol, 2-propanol, DMSO, THF, and 1,4-dioxane on the yield (Table 1, entries 4–12). When DMSO : $H_2O = 1 : 1$ was used as the reaction solvent, the yield increased to 38%. Moreover, when the aqueous solution was replaced with potassium phosphate buffer solutions of different pH, a decrease in the yield was observed (see ESI† for details). The reaction temperature has a strong influence on the yield, 50 °C is the optimal reaction temperature (see ESI† for more details).

The product yield strongly depended on the amount of catalyst and EDA, using an equal amount of EDA with 2.0 mol% of catalyst, tertiary nitroalkanes can be isolated with a yield of 32%. The use of higher concentrations of EDA resulted in an increase in yield, but a decrease in yield was observed when the amount of EDA exceeded 2.0 eq. (Table 2, entries 1–3). And then the catalyst was increased from 2.0 mol% to 8.0 mol% in the presence of 2.0 eq. EDA, only a small increase in yield can be observed (Table 2, entries 3–6), but from an economic point of view, the amount of catalyst of 2.0 mol% is chosen in this reaction. Furthermore, we determined 6 equiv. of Zn as the optimal reducing system loading (see ESI† for more details).

Table 2 Optimization: the influence of vitamin B_{12} and EDA

| Entry[a]	Vitamin B_{12} (mol%)	EDA (equiv.)	Yield (%)
1	2.0	1.0	32
2	2.0	2.0	38
3	2.0	3.0	29
4	4.0	2.0	40
5	6.0	2.0	40
6	8.0	2.0	42
7	10.0	2.0	38

[a] Reaction conditions: nitrocyclohexane (0.25 mmol), EDA (2 equiv.), Zn (6 equiv.), vitamin B_{12} (2 mol%), solvent (2.5 mL), 24 h.

The scope and limitation of the reaction were then investigated under the establishing the optimal reaction conditions above (Scheme 4). Among the series of diazo compounds (3a–3j), only diazoacetates 3b, 3c, 3d can give the corresponding products with relatively low yield. No product for 3a, 3e–3j was detected for other diazo compounds, probably because the higher steric effects prevented the progress of the reaction.

In addition, the reactions of ethyl diazoacetate and benzyl 2-diazoacetate with a series of nitro compounds were investigated (Scheme 5). 2-Nitropropane with 3 and 3d could produce the coupling product in 28% and 14% respectively. However, when phenyl nitromethanes were employed, no expected product was observed, presumably because the benzene is conjugated to the nitro group, making the formed carbanion not reactive enough to drive the reaction in progress.

In order to gain insight into the mechanism of the reaction, a variety of experiments were conducted (Scheme 6). Firstly,
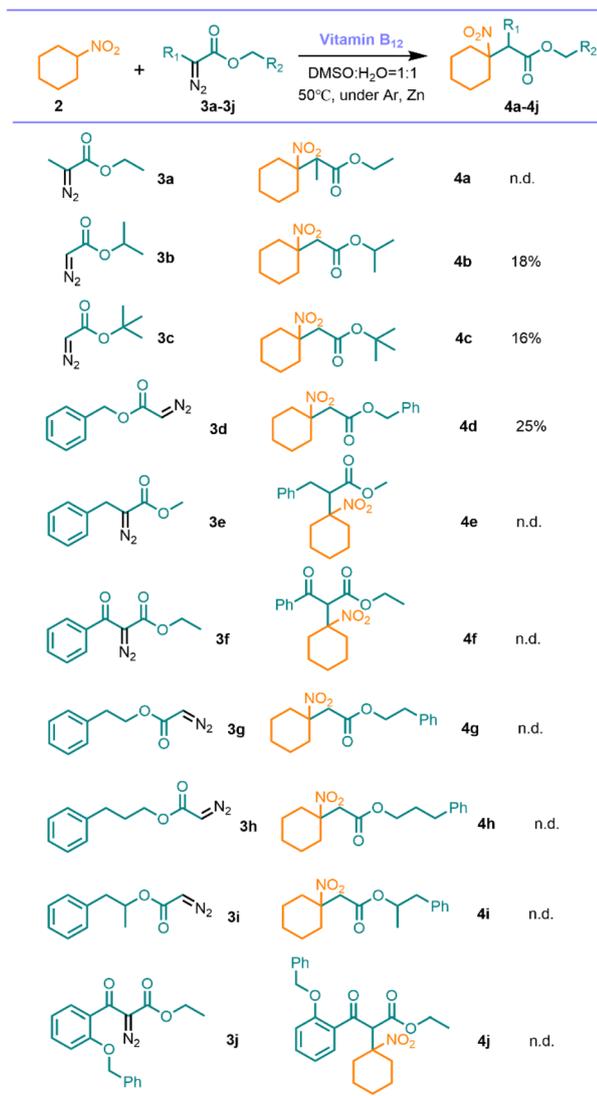

Scheme 4 Scope and limitation. Reaction conditions: nitro-cyclohexane (0.25 mmol), diazo compounds (2 equiv.), Zn (6 equiv.), vitamin B_{12} (2 mol%), solvent (2.5 mL), 24 h.
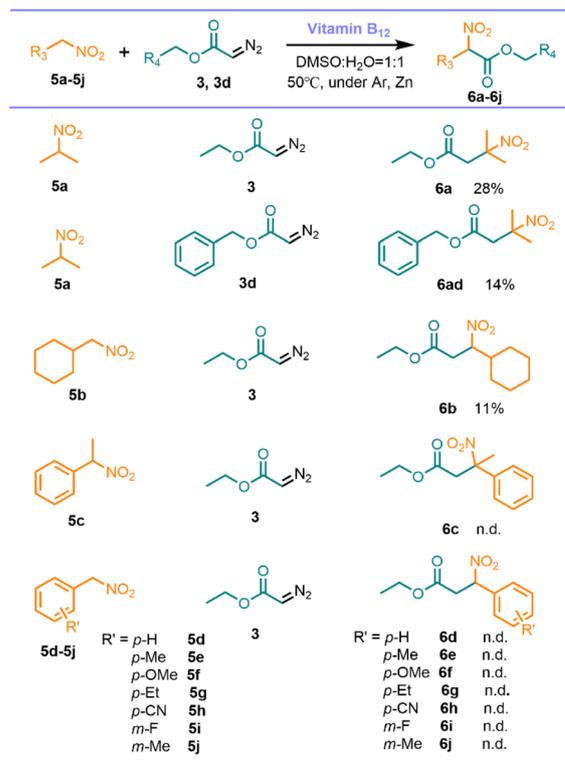
Scheme 5 Scope and limitation. Reaction conditions: nitro-cyclohexane (0.25 mmol), diazo compounds (2 equiv.), Zn (6 equiv.), vitamin B_{12} (2 mol%), solvent (2.5 mL), 24 h.

TEMPO, a radical scavenger, was added to the reaction mixture, which halted the reaction, suggesting a radical mechanism. Furthermore, when vitamin B_{12} was completely reduced, only EDA was added to the reaction at the room temperature, the ESI MS spectra of the crude reaction mixture showed a peak at 1415.60, confirming the generation of alkyl cobalamin intermediate (8). In this reaction, an additional proton in the product originates from H_2O, which was experimentally verified by H/D exchange experiments (see ESI† for detailed information).

On the basis of the experiments described above and previous studies,[28,32,33] we have proposed a mechanism for the formation
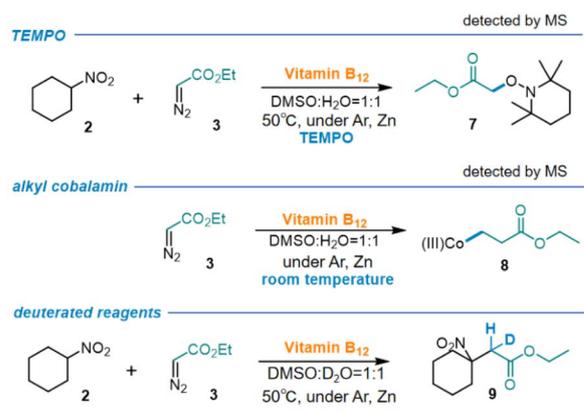
Scheme 6   Mechanistic studies.
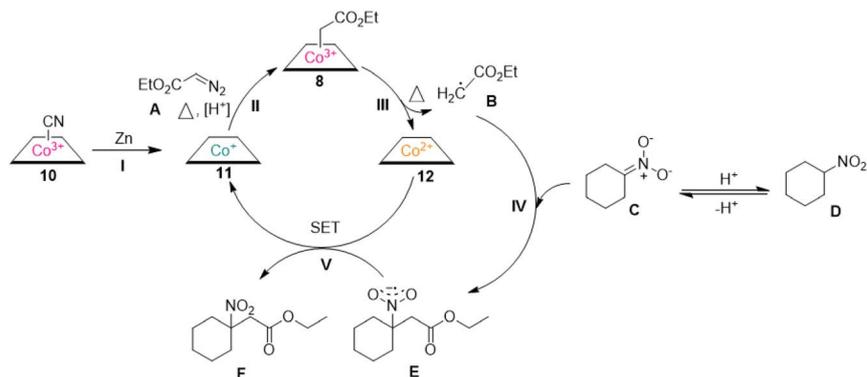
Scheme 7 Plausible mechanism for the vitamin B_{12} catalyzed coupling reaction.
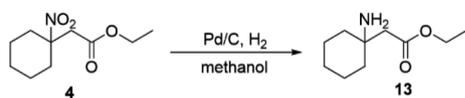
Scheme 8 Product derivatization.

of C–C bonds (Scheme 7). Zn acts as an additional reducing agent to reduce vitamin B_{12} to the catalytically active hypernucleophilic Co(I) (**11**), which reacted with EDA to produce an alkyl cobalt ester (**8**). The reaction is carried out under heating conditions at 50 °C, with sufficient energy to allow homolysis of the C–Co bond to give the alkyl radical (**B**). Subsequently, the alkyl radical can undergo an addition reaction with the *in situ*-generated nitronate (**C**) to give the nitro anion (**E**), while the construction of a new C–C bond is completed, leading to the tertiary nitroalkanes (**F**) by one-electron oxidation.

Nitro compounds are typical intermediates in organic chemistry that can be transformed directly into other functional groups. We use the product tertiary nitroalkane **4** as a precursor to be reduced to the corresponding tertiary amine **13** (Scheme 8), which is an important structural unit in the synthesis of many drugs and peptides. Thus, the coupling product can be used to create a broader range of compounds.

Conclusions

We have demonstrated for the first time that vitamin B_{12}, a natural and environmentally friendly catalyst, can effectively catalyze the C–H insert reaction of nitroalkanes with diazo compounds featuring a $C(sp^3)$–$C(sp^3)$ bond-forming step to construct tetrasubstituted centers. Under the developed conditions, diazo compounds form nucleophilic radicals and then react with nitroalkanes leading to the C–C bond formation. This methodology expands the chemical toolbox of transformations for EDA and nitroalkanes.

Methods

General information

All commercial reagents and solvents are to be used as received unless otherwise indicated. Reaction process monitoring using thin-layer chromatography (TLC) on Yantai Chemical Industry Research Institute silica gel (GF254, 0.23 mm thickness). Observations were made using UV light (254 nm) or potassium permanganate for color development. The crude product was separated and purified by column chromatography using Qingdao Ocean Chemical silica gel (200–300 mesh). ^{1}H and ^{13}C NMR spectra were recorded at ambient temperature on a Bruker 400 MHz instrument with TMS as an internal standard. Liquid chromatograph mass spectrums (LC-MS) data were obtained on an Water Xevo G2 QTof Liquid Chromatograph Mass Spectrometer and a SilGreen 100 C18 column.

General experimental procedure for the synthesis of the coupling products

The reactions were carried out under an argon atmosphere unless otherwise indicated. Vitamin B_{12} (2 mol%, 7 mg), Zn (196 mg, 6.0 mmol) were accurately weighed in a 10 mL two-necked flask using an electroanalytical balance. Degassed dimethyl sulfoxide and water $(DMSO : H_2O = 1 : 1, 2.5 \text{ mL})$ were added to the reaction flask, and the reaction mixture was stirred vigorously under argon gas at 50 °C until the vitamin B_{12} was completely reduced (colour changed from red to green). Nitroalkanes (0.25 mmol, 1.0 equiv.) and diazo compounds (0.50 mmol, 2.0 equiv.) dissolved in dimethyl sulfoxide were added to the reaction system with a syringe and the reaction was carried out for 24 h.

Zn needs to be activated before use. The activation process was as follows: (1) grinding in 10% HCl, (2) washing with water, methanol and water, (3) drying in vacuum. Activation of Zn has a great influence on the reaction yield.

2-(1-Nitrocyclohexyl) ethyl acetate (4). ^{1}H NMR (400 MHz, chloroform-d) δ 4.13 (q, 2H), 2.91 (s, 2H), 2.35–2.29 (m, 2H), 1.88–1.83 (m, 2H), 1.62–1.38 (m, 6H), 1.22 (t, 3H). ^{13}C NMR (101 MHz, chloroform-d) δ 168.62, 88.50, 61.02, 42.83, 34.27, 24.60, 22.29, 14.02. HRMS (ESI): $[M + H]^+$ $(C_{10}H_{18}NO_4)$ calculated: 216.1230, found: 216.1217.

Isopropyl 2-(1-nitrocyclohexyl) acetate (4b). ^{1}H NMR (400 MHz, chloroform-d) δ 5.04–4.94 (m, 1H), 2.89 (s, 2H), 2.37–2.31 (m, 2H), 1.90–1.84 (m, 2H), 1.61–1.41 (m, 6H), 1.22 (d, J = 6.3 Hz, 6H). ^{13}C NMR (101 MHz, chloroform-d) δ 168.13, 88.55, 68.75, 43.23, 34.30, 24.64, 22.30, 21.65. HRMS (ESI): $[M + H]^+$ $(C_{11}H_{20}NO_4)$ calculated: 230.1387, found: 230.1385.

Tert-butyl 2-(1-nitrocyclohexyl) acetate (4c). ^{1}H NMR (400 MHz, chloroform-d) δ 2.83 (s, 2H), 2.35–2.30 (m, 2H), 1.88–1.82 (m, 2H), 1.63–1.49 (m, 6H), 1.42 (s. 9H). ^{13}C NMR (101 MHz, chloroform-d) δ 167.82, 88.58, 81.88, 44.31, 34.32, 27.90, 24.67, 22.30. HRMS (ESI): $[M + H]^+$ ($C_{12}H_{22}NO_4$) calculated: 244.1543, found: 244.1548.

Benzyl 2-(1-nitrocyclohexyl) acetate (4d). ^{1}HNMR (400 MHz, chloroform-d) δ 7.36–7.32 (m, 5H), 5.11 (s, 2H), 2.99 (s, 2H), 2.37–2.30 (m, 2H), 1.91–1.84 (m, 2H), 1.60–1.41 (m, 6H). ^{13}C NMR (101 MHz, chloroform-d) δ 168.54, 135.26, 128.62, 128.46, 128.38, 88.52, 66.91, 42.65, 34.28, 24.60, 22.31. HRMS (ESI): $[M + H]^+$ ($C_{15}H_{20}NO_4$) calculated: 278.1387, found: 278.1386.

Ethyl 3-methyl-3-nitrobutanoate (6a). ^{1}H NMR (400 MHz, chloroform-d) δ 4.15 (q, $J = 7.1$ Hz, 2H), 2.97 (s, 2H), 1.69 (s, 6H), 1.25 (t, $J = 7.1$ Hz, 3H). ^{13}C NMR (101 MHz, chloroform-d) δ 168.83, 84.95, 61.05, 44.00, 26.31, 14.05.

3-Methyl-3-nitrobutyric acid benzyl ester (6ad). ^{1}H NMR (400 MHz, chloroform-d) δ 7.40–7.32 (m, 5H), 5.13 (s, 2H), 3.03 (s, 2H), 1.69 (s, 6H). ^{13}C NMR (101 MHz, chloroform-d) δ 168.70, 135.26, 128.64, 128.49, 128.36, 84.94, 66.86, 43.89, 26.33. HRMS (ESI): $[M + H]^+$ ($C_{12}H_{16}NO_4$) calculated: 238.1074, found: 238.1088.

Ethyl-3-cyclohexyl-3-nitropropanoate (6b). ^{1}H NMR (400 MHz, chloroform-d) δ 4.73 (ddd, 1H), 4.13 (q, 2H), 3.10 (dd, 1H), 2.66 (dd, 1H), 1.91–1.76 (m, 1H), 1.69–1.63 (m, 5H), 1.26–1.02 (m, 8H). ^{13}C NMR (101 MHz, chloroform-d) δ 169.75, 88.16, 61.24, 41.33, 34.37, 29.17, 28.53, 25.73, 14.00. HRMS (ESI): $[M + H]^+$ ($C_{11}H_{20}NO_4$) calculated: 230.1387, found: 230.1382.

Data availability

All data in this study are included in this article and its ESI.†

Author contributions

Z. Z. performed the study, and manuscript writing. M. C. conceived the project, performed the study and revised the manuscript. G. Z. supervised the project, approved and submitted the final manuscript. All authors have read and agreed to the published version of the manuscript.

Conflicts of interest

The authors declare no conflict of interest.

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